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# Social behaviors as welfare indicators in teleost fish

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Animal welfare is a key issue not only for aquaculture industry and food production, but also for daily husbandry practices in research topics related to physiology in wild and farmed animals. In this context, teleost fish constitute interesting models to assess alternative welfare indicators because of their wide diversity in reproductive and social structures. Any framework for assessing teleost fish welfare needs to account for the physiological mechanisms involved in each species as a first step. A comprehensive approach should also take into account how these physiological and behavioral parameters can be altered by environmental enrichment considering the specific requirements in each case and identifying intrinsic biological characteristics of individual species. This review will show how cortisol and sex steroids regulate social behavior in teleost fish, and how different aspects of social behavior can be employed as welfare indicators according to specific characteristics in each case. This article will consider evidence in teleost fish, including cichlids, characids and cyprinids with different reproductive strategies and social structures (e.g., territorial social hierarchies or shoaling behavior). Neotropical species will be particularly emphasized. The main laboratory-based animal welfare indicators are cortisol, a classical stress hormone, together with sex steroids. Considering that the endocrine landscape is intrinsically related to social behavior, reproductive and agonistic behavioral traits such as aggression, anxiety and courtship are key elements to assess welfare under housing and culture conditions. This review highlights the importance of assessing physiological mechanisms and identifying behavioral characteristics in teleost fish, especially in Neotropical species, as a baseline to understand which environmental enrichment can improve animal welfare in each individual species.

#### KEYWORDS

social behavior, welfare indicators, cichlid fish, Cypriniformes, Characiformes, cortisol, sexual steroids

### 1. Introduction

Human activities involving live animals may affect welfare in different ways and, even if husbandry standards and housing requirements have been established for different groups, a better empirical understanding of animal welfare is continuously required. Many domestic animals or those used for food production are frequently kept under confined conditions in which basic behavioral and physiological needs are drastically affected. Taking into account the continually growing anthropocentric activities involving the use of animals for research, medicinal use and food production, animal welfare represents a central area of growing interest. An increasing number of human activities can potentially affect fish welfare, including environmental degradation, farming, commercial fisheries, aquaculture, ornamental fish, sports fishing, and scientific research (1). A better understanding of fish welfare will increase awareness in groups leading these activities and to this end a thorough revision of welfare indicators will help to understand the biological requirements for particular fish species.

Specific indicators need to assess potential welfare problems and give information on the internal state of animals exposed to human activities (2). While some welfare indicators can be easily sampled in natural environment and in farms, others can be more difficult to apply on site since they can involve laboratory analysis or a specific analytic facility (3). This way, behavioral welfare indicators are very good candidates to use "on-farm," since they are easy to observe and can be used as a non-invasive approach to assess any potential welfare problem (4). Behavior involves fish reaction to their perception of the environment and behavioral variables are key aspects to take into account when assessing their welfare and global internal state (5). Some specific behavioral traits are widely used as welfare indicators, such as food intake, changes in food anticipatory behavior, swimming activity, and ventilation rate (1). However, other behaviors are less used as welfare indicators, such as social, reproductive, and parental behaviors. Considering that behavior is modulated by physiology and motivational states of each individual fish, hormonal and neuroendocrine regulation play central roles in social and reproductive behaviors. Even if behavioral responses are very useful indicators, a robust and integrated welfare assessment also requires other factors such as endocrine, neurophysiology, pathological, and molecular tools (1). In this intertwined relationship between behavior, hormones and animal welfare, it is important to distinguish the endocrine mechanisms regulating reproductive and social behavior, in order to identify specific behavioral displays that can be used as welfare indicators. In this sense, sex steroids and cortisol play central roles modulating agonistic behaviors such as aggressive and submissive displays, courtship, mating, and parental behaviors (6, 7).

Taking into account that teleosts are the largest and most diverse group of vertebrates, with more than 30,000 species described (8), they represent very interesting biological models to assess welfare indicators. Teleost fish constitute the dominant evolutionary radiation of vertebrates in our planet, which is reflected not only in the vast array of colonized habitats, but also in a remarkable variety of ecological specializations, physiological mechanisms and behavioral characteristics. Considering the popular use of some specific teleost species in human activities, these examples illustrate the diversity in physiological mechanisms underlying behavior and the need to use this species-specific information as welfare indicators. It is estimated that 90% of the total volume of trade in ornamental fish corresponds to tropical freshwater fish (9). Considering this, this review will focus on specific teleost groups based not only on their relevance in human economic activities and the potential impact on their animal welfare, but also on the current knowledge on their behavioral strategies and biology.

Cichlid fish are a very appealing family to discuss the importance of behavioral traits as welfare indicators, since they show diverse social networks with hierarchical structures and different parental care or reproductive strategies. In most species with social hierarchies, dominants aggressively defend a spawning territory from subordinate, nonterritorial fish (10). While some species are important for food production and aquaculture activities, others are considered game fish and have been introduced to regions beyond their natural habitat, usually linked to escapes or releases from fish farms. Based on their native range, cichlids can be classified as African and Neotropical cichlids. Neotropical region is comprised by North, Central and South America and includes 67 described genus (11). While African cichlids usually inhabit lakes, Neotropical cichlids can be mainly found in lentic habitats such as rivers (12). Neotropical species have a central role in aquaculture and, as a consequence, their welfare can be deeply affected. Interestingly, the most studied species to date are the African cichlids, while Neotropical species are understudied despite of their economic relevance. Characiforms also include African and Neotropical species living in rivers and lakes. They also present different reproductive strategies and social behavior ranging from aggressive species such as piranhas to shoaling and/or schooling species such as tetras (13). Some characiforms are very popular in the aquarium trade (tetras, piranhas, and pencilfishes), while others have a commercial importance as sport-fishing species (the African tiger fish and the Neotropical dorados) or as food resource for human communities living near tropical rivers (the African citharinids and the Neotropical prochilodontids). Cypriniformes also includes species with very important scientific and commercial value in aquaculture, breeding, food industry and ornamental fisheries, such as carps, minnows, goldfish and zebrafish. Even if wild stocks can be found in different continents depending on each case, these species have been deeply introduced in diverse environments (14).

The aim of this review is to highlight the importance of studying behavior to assess the welfare of different teleost fish. With this main focus, we review physiological mechanisms underlying complex social behaviors, emphasizing the central role of cortisol as the main stress parameter affecting fish social structure and sex steroids regulating reproductive and parental behavioral strategies. Throughout this review we intend to highlight the need to understand the physiological mechanisms and the behavioral characteristics of different teleost fish according to their specific biological requirements, with the perspective of using behavioral traits as welfare indicators. Because of their behavioral curiosities and their intense use by human activities, we will focus on different examples of cichlid fish, characiforms, and cypriniforms.

# 2. Teleost fish as biological models for welfare indicators

Considering that multiple human activities involve teleost fish, there is a growing body of evidence suggesting that human activities can harm their welfare (1). However, the magnitude of these effects depends on the general characteristics or life-history of each group and with this in mind, it is important to revise behavioral and physiological mechanisms explaining how to analyze welfare indicators in each species.

Cichlid fish are very popular in aquaculture industry, and they constitute ideal models to understand reproductive physiology and neuroendocrine basis of social behavior. Considering their complex social structure, cichlids show different reproductive strategies and diverse forms of parental care, such as substrate guarding and mouthbrooding, and also an impressive variation in which sex provides parental care (e.g., biparental, female-only, or male-only

care). The African cichlid fish radiations are the most diverse animal radiations, with most species endemic to specific lakes such as the Lake Malawi, Lake Victoria, and Lake Tanganyika (15). The most popular cichlid models to study the neuroendocrine mechanisms regulating social behavior and reproduction are the African species Astatotilapia burtoni, Neolamprologus pulcher, and Oreochromis mossambicus. Considering that freshwater fish species dominate aquaculture fish production, with carps and cyprinids representing over 53% of fish production, it is important to emphasize that cichlids constitute 11% of this production (16). The most popular cichlids are tilapias, in particular Nile tilapia (Oreochromis niloticus) and its hybrids accounting for over 90% of production of tilapias (17). This species is not only widely used for neurobiological and endocrine studies, but it is also globally important as a cultured food fish since it is one of the most important fish species in worldwide aquaculture. Uncontrolled breeding of tilapias led to excessive recruitment, high densities, stunting and low percentage of marketable-size fish, until research on nutrition, culture systems and the development of hormonal sex-reversal techniques in 1970 allowed for monosex populations to be raised in uniform sizes (18). Consistently, most of the evidence assessing animal welfare in cichlids refers to O. niloticus, usually regarding welfare indicators of stress (19) or dietary impacts on improving the antioxidant status and general health in this species (20). However, evidence on welfare indicators in other cichlids is scarce. Therefore, it is very important to revise physiological mechanisms underlying the behavioral strategies of different species, in order to assess the importance of different behavioral traits as welfare indicators.

Cypriniformes includes more than 3,000 species and almost all of them are strictly freshwater and distributed across Eurasia, Africa, and North America (14). Besides carps and cyprinids, representing over 53% of fish production, the most important ornamental fishes are goldfish (Carassius auratus). Originally from southern Asia, they were later introduced in a broad range of freshwater habitats worldwide (21). As one of the most important cases in aquaculture, this species has been one of the first animals bred with an ornamental purpose (22) and it is widely used as a biological model in research on different fields such as endocrinology, developmental biology, fish pathology among others. Another species with economic importance is the common carp (Cyprinus carpio) native to Europe and Asia, but due to its invasive capacity it is found in many countries and freshwater habitats around the world, mainly ponds, lakes, and rivers (23). Because of its popularity as an ornamental fish, the commercial production of carp increased considerably (24), while it has also been used in scientific research in different fields such as genetic research, developmental biology, immunological systems, studies of tissue damage and the effect of different diets on growth rates and health. Zebrafish (Danio rerio) constitutes a particular case among Cypriniformes. Even if it is also traded as an ornamental fish, its main popularity worldwide is due to the fact that it is one of the most important non-mammalian biological models for scientific research in different fields such as genetics, behavioral genetics, endocrinology, memory and behavior, development, pathologies and as a model for psychiatric disorders and drug screening (25). Originally endemic to rivers and streams in India, Nepal, and Bangladesh (26), it has been easily introduced worldwide to breeding systems in laboratory conditions. Due to its importance as a model in experimental research, welfare indicators in this species were broadly studied. In particular,

the main focus has been placed on behaviors related to stress and anxiety (27), poor living conditions and behavioral displays (28).

Characiformes is a group of freshwater fish with about 18 families and almost 2,000 species, very popular in the ornamental fish trade and commonly farmed in different countries (29). They live in tropical waters from the north of Patagonia in Argentina to the south of the United States, and this wide variety of environments is reflected in their great ecological and morphological diversity (30). Characidae is the most diverse family among Neotropical fishes and the fourth most diverse worldwide (31). The best-known representatives of Characiformes are the tetras, whose cultured stocks are usually domesticated (32) and constitute one of the most popular fishes kept in modern aquariums. Tetras are sociable, non-aggressive species and represent no threat to other fish, making them ideal for a community aquarium (33).

Popularity of ornamental fish has grown by 14% annually since the 1970s, and nowadays more than one billion individual fish are internationally traded each year (9). For example, neon tetra (Paracheirodon innesi) and cardinal tetra (Paracheirodon axelrodi) are two species of great interest in the ornamental trade (34). P. innesi is more commercialized than P. axelrodi, since it is much less sensitive in terms of management and reproduction in captivity. These species are collected from the wild in South America (35) and between 12 and 15 million neon tetras are annually exported, which represents about 80% of the total ornamental fish market in the state of Amazonas (9). Considering that intensive fishing can unbalance the populations in the natural environment, this situation has raised deep concern. However, despite their economic relevance, ornamental fish still face critical welfare challenges, including inadequate maintenance conditions, poor water quality, handling, transportation, confinement, overcrowding, poor diet, or feeding methods and disease (36). Even though research on characiforms welfare is scarce, exploring different indicators in this order is crucial to improve the quality of life and health of fish, and to expand successful production in the ornamental trade.

#### 3. Social behavior

Social behavior, such as courtship, mating and aggression, involves individuals interacting with others and adjusting their behavior based upon previous experience in a constantly changing environment. Fish can present different social strategies, from shoaling and schooling to social hierarchies with dominant and subordinate individuals. Establishing dominance implies high levels of aggression and agonistic interactions before and during the development of social hierarchies. As a consequence, social stress during this process is one of the major welfare issues when rearing and keeping fish in captivity. Also reproductive behavior in fish includes very variable strategies such as monogamy (permanent or seasonal), polygamy (polygenic or polyandric) and specific systems like lek-breeding, harems and sneaking behaviors. This great variety of social behavioral strategies are represented in different examples of cichlid fish, Characiformes, and Cypriniformes.

These social displays and species-specific social behavioral strategies need to be taken into consideration when establishing requirements and housing conditions such as tank size, fish density, environmental enrichment, and diet. Many of these social behavioral traits are drastically affected when stocking densities are too low or too high, impacting directly on fish welfare. Since stocking density is a very important topic in aquaculture, an integrative approach should consider fish welfare and farmer's perspectives by integrating solid welfare assessment (operational welfare indicators) with good management practices (37). Moreover, considering that social behavior can be affected by the environmental enrichment and that in most cases this focuses on structural enrichment, it is important to emphasize that many other strategies are essential to improve farmed fish welfare, such as sensorial (visual, auditory, and chemical stimuli), occupational enrichment (aiming to introduce diverse challenges to the rearing environment to prevent monotony), social and dietary enrichment [reviewed in (38)].

Cichlids typically form hierarchical social systems with one or two dominant animals defending their status from other submissive and lower-ranked animals (39). African species have been widely studied as biological models to assess the neuroendocrine mechanisms regulating social behavior and reproductive strategies. In A. burtoni one dominant male mates with several females and defends the territory by aggressive displays (40). Parental care in this species is typically female mouthbrooding and even if females have been traditionally described as non-aggressive, female aggression has been observed in all-female communities (41) and also in recently collected stock of fish females taking care of their brood (42). Behavioral differences between A. burtoni stocks have been attributed to the fact that the majority of traditional research in this species has been focused on a single fish stock collected from Lake Tanganyika in 1977, which has been maintained in captivity under laboratory conditions for more than 30 years. Artificial selection probably occurred in laboratory stocks, and this can explain differences in aggression observed in female wild stocks (42). This constitutes a clear example of the importance of using social behavior as welfare indicators, and it emphasizes the relevance of assessing social behavior in freshly collected wild stocks.

Another widely studied African cichlid species is *O. mossambicus* (Mozambique tilapia), a maternal mouthbrooding cichlid in which dominant males dig nests and court females, while subordinates wait for opportunities for social ascension or sneaking fertilization (43). Interestingly, dominant males can also display courtship behavior toward subordinates, who performed typical female sexual behavior (43). This male–male courtship constitutes another clear example of the importance of studying social behavior can be argued to be an effect of crowding under captive conditions. In *O. niloticus* high levels of aggression have been described during hierarchy formation (44) and also in specific aquaculture and laboratory conditions, even leading to high mortalities (45). Interestingly, cold water reduces aggression and it can be applied to mitigate adverse social effects arising from hierarchy formation in captive conditions (46).

On the contrary, *N. pulcher* is a cooperative breeding species with a dominant breeding pair showing aggressive behaviors and subordinate helpers that assist in taking care of the offspring (47, 48). Even if this species does not constitute a relevant example in worldwide aquaculture, aggression has been assessed as a welfare indicator in terms of other anthropogenic activities and stressors in aquatic environments, suggesting that underwater noise produced by commercial shipping and recreational boating reduces digging behavior and defense against predators, while it increases the amount of aggressive behavior (49).

Neotropical cichlid species are also characterized by complex social behaviors. Interestingly, while most African species present uniparental care of eggs and larvae usually associated to mouthbrooding behavior, Neotropical cichlids usually present biparental behavior (50). This is the case of Cichlasoma dimerus, a South American species inhabitant of the Parana and Paraguay Rivers' basins that has been used as laboratory model to study the neuroendocrine basis of social behavior and reproduction (51). This is a serially monogamous species in which individuals display aggressive and submissive behaviors during the establishment and posterior maintenance of a very clear social hierarchy and, like most American cichlids, it is a substrate brooder with biparental behavior (6, 10). Aggressive behavior in this species has been characterized and quantified in different ethological contexts and, in all cases, males and females of this species show very high levels of aggressive and submissive displays, emphasizing the importance of this species as a biological model to study neuroendocrine basis of reproductive and social behavior (39).

Similar to *C. dimerus*, in Neotropical cichlids monogamous behavior and biparental care of the offspring are very common, so both males and females usually display aggressive behavior to defend their territory (52, 53). Moreover, the typical fighting behavior found in *C. dimerus* (54) is also observed in different Neotropical cichlid species, such as *Cichlasoma paranaense* (55) and *Astronotus ocellatus* (56). Even if these cases do not represent relevant commercial species, the study of social behavior results in a deeper knowledge on aggressive displays. The capacity of using this behavior as a welfare indicator is further discussed in the following section.

When assessing fish welfare, water renewal and temperature are key factors to take into account. Since chemical cues play a central role in communicating social status and regulating behavioral interactions among fish, water removal can wash out chemical substances and interfere in social rank signaling in highly social cichlids (57). In C. paranaense, water removal does not increase aggressive interactions, in contrast with evidence in P. scalare, suggesting interspecific differences on the aggressive response to chemical signaling in the social hierarchy (58, 59). Changes in water temperature can also alter aggressive behavior in a species-specific manner, since a raise in temperature increases aggressive displays in Apistogramma agassizii and Amatitlania nigrofasciata (60, 61), but C. paranaense reduces aggression in lower temperatures (62). These evidences show differences in how aggression can be interpreted as a welfare indicator and highlights species-specific requirements to ensure optimal social conditions.

Another relevant issue involving fish welfare is social isolation, which can be assessed in different cichlid species by studying aggression as a behavioral welfare indicator. Since cichlid fish are considered as very aggressive, they are usually kept in isolation in the aquaculture industry, such as *A. nigrofasciata* and the oscar *Astronotus ocellatus*. In these species, isolation increases the frequency of mouth fighting and the motivation for aggressive behavior (56, 63). However, isolation in cichlids impairs welfare not only in reference to aggression but in multiple ways, such as locomotor and feeding activity in *P. scalare* (64), the neuroendocrine system in *A. burtoni* (65) and also the cognitive performance in *C. paranaense* (55). This way, even if the consequences of isolation on welfare have been widely studied and

have been shown to affect aggression in these specific cases, this may be also important in other cichlids and further research needs to assess the effect of social isolation using aggression as a behavioral welfare indicator.

Cypriniformes also have complex social structures and different behavioral traits can be very useful to assess fish welfare. While common carps are highly social and show aggressive behaviors like bites or chases, evidence suggests that high density induces higher aggression, swimming and feeding activity, while it reduces resting time (66). Aggression can be also used as an indicator in reference to fish exposure to heavy metal and other toxic compounds such as polyethylene microplastics, glyphosate (67) and cadmium (68). Zebrafish is a highly social species with shoaling behavior, in which both sexes establish social status by aggressive and submissive displays (69). Social status not only regulates access to reproduction, but also affects physiology and health of individuals in different ways, since subordinate fish show signals of immunosuppression and altered plasma levels of sex-steroid hormones (70). Social behavior can be a useful welfare indicator to assess the effect of overcrowding, inflammatory response and the exposure to specific substances, since ethanol and certain metals trigger antisocial behaviors in zebrafish (71, 72). Furthermore, exposure to conspecific alarm substances or caffeine alter the normal shoaling behavior in this species (73).

Regarding the reproductive behavior of cypriniform species mentioned here, males usually compete with each other to fertilize eggs and females can choose their mates, such as female zebrafish preferring larger males (74). In common carp, zebrafish and goldfish courtship is similar, with males chasing and encircling the female until eggs are released and they can be fertilized by, in some cases, several males simultaneously (75-77). Most of these species do not present parental care and, in fact, filial cannibalism is observed in some species (78). In zebrafish, egg cannibalism has been reported in fieldbased semi-natural or in laboratory conditions, and it has been suggested that females preferring gravel as a spawning substrate is related to the fact that eggs falling between gravel can be protected from disturbance or cannibalism (79). In this context, the question on whether filial cannibalism can be indicative of fish welfare can be raised. Evidence suggests that even if in some teleost species there is a decrease in cannibalism in males with supplementary feeding (80), manipulative studies in other species fail to support the prediction that well-fed parents would show decreased filial cannibalism than starved ones [discussed in (81)]. This way, even if egg cannibalism may be a welfare indicator in some species, in particular referring to the nutritional state, in other cases this behavior can be related to natural or seminatural conditions and to other factors such as parental behavior.

Despite the extraordinary diversity on breeding systems of teleost fish, social and reproductive behavior in Characiformes remain poorly studied in contrast to the closely related Cypriniformes and Siluriformes. One of the few characiform species in which the reproductive behavior has been formally studied is the sabalo *Prochilodus lineatus*, a Neotropical migratory fish with shoaling behavior during reproduction. DNA and multilocus genotyping methods suggest that this species shows a monogamous genetic mating system in combination with polygamy (82, 83). This knowledge on mating strategies and reproductive behavior of Neotropical migratory fishes in nature constitutes relevant evidence for the management and conservation of such an important fishery resource. Within Characiformes, tetras are highly social species with schooling behavior. Fish aggregations can be classified as shoals, when there is a significant degree of cohesion among the individuals (e.g., social attraction), or as schools, when individuals perform synchronized behaviors in terms of speed and body orientation (84). In *P. innesi* robust schooling behavior is well reported (85), and evidence suggests that the individual optimal position in the group is context dependent since individual fish continuously modify their location in the school according to different external scenarios (13). In contrast to *P. innesi*, in *P. axelrodi* and the red-tailed tetra *Aphyocharax anisitsi* available information corresponds to collective knowledge from aquarists.

Considering that pheromones play a fundamental role in social communication, their importance in Characiformes has been assessed in two species. In swordtail characin *Corynopoma riisei*, evidence suggests that male sex pheromones from gill and anal fin glands induce behavioral changes in females, reducing stress and increasing female courtship behavior, thus facilitating reproduction (86, 87). In *A. anisitsi*, since gill glands are present in males with higher gonadosomatic index, they have been suggested as indicative of sex maturity (88). In this species evidence suggests that males respond to male and female chemical cues even in the absence of other sensory inputs (89). This evidence supports the possibility that pheromones participate in the chemical communication of *A. anisitsi* and is a clear example of how behavioral indicators can be used to assess not only social chemical cues in characids, but also exposure to aquatic environmental substances in characids (90).

# 4. Cortisol as classical stress parameter

Fish welfare can be affected by different common practices in aquaculture, such as overcrowding, handling, social isolation and transportation, all possible husbandry stressors affecting their quality of life (36, 91). Glucocorticoids are steroid hormones synthesized by the adrenal/interregnal gland, and cortisol is the main glucocorticoid in fish (92). Considering the stress-mediated increase in plasma glucocorticoids in all vertebrates, cortisol is the main component of the primary neuroendocrine stress response in fish. Stress can stimulate the hypothalamic-pituitary-interrenal (HPI) axis, inducing an increase in cortisol that affects brain functions, animals' growth, reproduction, immune system and behavior in fish (93, 94). Unfortunately, different phases of the distribution market may result stressful for fish, and it has been reported that handling fish for even very short period of time (30s) is enough to increase cortisol in several species (95). On the other hand, different physiological markers of stress have been studied to assess the effect of short and long transport, including blood glucose, lactate and cortisol (96). Even if cortisol can be used as an indicator of how fish perceive their environment, social structure and aggressive interactions trigger high social stress in fish (93, 97). As a consequence, the use of cortisol as a welfare indicator in husbandry practices and culture environment needs to be assessed with caution considering physiological levels and social behavior characteristics of each species.

In highly social fish, hierarchies are a source of physiological and physical stress, since lower-ranked individuals are chronically stressed (98). This is observed in different cichlids, with species-specific

peculiarities. In A. burtoni, non-territorial males show higher cortisol plasma levels than territorial males (99) and the same tendency is observed in C. dimerus (6, 100). However, in N. pulcher dominant males show higher cortisol than subordinates (101), while in O. niloticus territorial and non-territorial males present similar cortisol levels (102). This evidence illustrate the complex relationship between dominance and glucocorticoid levels in vertebrates, which varies among species but also depends on different characteristics of each social hierarchy, such as stability, environmental conditions, the type of breeding system and how the social rank is maintained in each case (103). It is important to emphasize that cortisol levels can also differ according to the ethological context, and this is related to the fact that aggressive interactions trigger social stress in fish (97). For example, in O. niloticus during initial fighting cortisol increases in both opponents (102), but after 5 days of social interactions fish can either show similar levels of stress in homogeneous-sized individuals, or higher levels in dominants than subordinates of heterogeneoussized group (104).

Taking these species-specific differences and social system characteristics into account, cortisol can be used as a stress indicator in husbandry or experimental conditions. For example, evidence suggests that social isolation during 24h induces an increase of cortisol levels in A. nigrofasciata (63). Environmental enrichment is usually used to improve welfare in housing conditions, creating a more complex and stimulating environment aimed at encouraging the development of natural species-specific behavior (105). In this sense, environment can also modulate isolation-induced stress, since in O. niloticus it has been reported that blue light prevents the confinement-induced cortisol response, when compared to green and white light (106). Interestingly, in O. mossambicus social isolation elicits a differential increase in cortisol depending on the previous social status of fish, suggesting that it is the perception of social isolation and not the objective isolation itself that triggers the hormonal response to isolation (107). On the other hand, in order to improve space efficiency, fish production usually counts with high stocking densities, which can cause stress in fish. Overcrowding can be identified as a stressor because it can increase aggressive interactions and a concomitant increase in cortisol (91, 97). However, it is important to emphasize that rearing density impacts the HPI axis in a species-specific way according to the social structure of each species. Evidence suggests that while in Atlantic salmon (Salmo sara) high-density reared fish show increased expression of stress-related markers and downregulation of innate immune gens, in O. niloticus this effect is observed in tilapias held in low densities probably due to increased aggressive interactions (19). Usually body tactile stimulation has a positive effect since it relieves stress and reduces cortisol on highly social animals, including some fish such as the cleaner-client coral-reef fish (108). However, the effect on body tactile stimulation has been tested in O. niloticus, reducing aggressive interaction but not plasma cortisol levels (109). This suggests that, even if environmental enrichment with tactile stimulation does not reduce stress in this territorial species, it has a positive effect because it reduces aggressive interactions and it can be used in further studies on fish welfare (109, 110).

Common carp reared in high densities show increased plasma cortisol, glucose and decreased humoral immune response than those in low densities (111). Interestingly, this increased in cortisol can be mitigated by diet enrichment with specific components such as arginine and dietary rosemary leaf powder (111, 112). Another key factor in fish welfare is an appropriate maintenance of water quality, which is essential to reduce stress during transport of ornamental fish. Farmers take different measures to control the stressing effect of transportation (96). Evidence in carps suggest that, even if plasma cortisol and glucose increase after they are distributed in plastic bags for 3 h, high levels of cortisol can be mitigated by diet enrichment with turmeric (113). Moreover, high levels of ammonia in overcrowded stocking are related to food left-overs and unwanted waste and they usually cause an increase in cortisol and glucose, these effects being mitigated by garlic-rich diet (114).

Another strategy to reduce stress responses through diet is by modifying the amino acid composition of the food. L-tryptophan enriched diet has been proven as an effective treatment to reduce stress increase in cortisol in different teleost species, such as *Labeo rohita* (Cypriniformes) (115), *Solea senegalensis* (Pleuronectiformes) (116) and *Totoaba macdonaldi* (Perciformes) (117). In the case of the rainbow trout *Oncorhynchus mykiss*, evidence suggests that L-tryptophan-induced reduction in plasma cortisol levels and suppression of aggressive behavior (118). Interestingly evidence in cichlids suggests not only that L-tryptophan reduces cortisol and aggression in *C. dimerus*, but also that there are no negative impacts on growth parameters (119, 120).

Another important factor affecting stress and fish welfare is light, and evidence in goldfish suggests that the absence of light-dark cycles and feeding cycles increases plasma cortisol levels (121). Regarding environmental enrichment in Cypriniformes, most studies focusing on housing conditions in zebrafish suggest that high cortisol related to stress can be mitigated by environmental enrichment with gravels, artificial plants and objects (122). Environmental enrichment also involves housing tank color. For example, zebrafish which were held in transparent and white tanks increased anxiety-like behaviors compared with zebrafish maintained in black or blue tanks, while blue tanks reduce cortisol levels (123). As already discussed earlier in this section for some cichlid species, isolation in zebrafish evokes anxiety like behaviors and an increase in cortisol levels. Even if anxiety-like behaviors can be mitigated by certain acoustic environmental enrichment, such as exposure to Antonio Vivaldi's music, isolationinduced cortisol increase persists (124). Considering that plasma or body cortisol measurements represent an invasive technique to fish, a very convenient alternative is to use cortisol dissolved in water as an indicator of fish stress, since there is a significant positive correlation between cortisol released to water and body cortisol (125). Evidence suggests that the pattern of changes in trunk cortisol of stressed zebrafish was also reflected in the water cortisol release rate pattern. Even if the waterborne cortisol release rate can be used as a noninvasive and reliable stress indicator, this methodology needs to be carefully assessed. For example, evidence in zebrafish suggests a higher magnitude of water cortisol release rates than other reported fish species, such as common dentex (Dentex dentex), common pandora (Pagellus erythrinus), sharpsnout sea bream (Diplodus puntazzo), meager (Argyrosomus regius), common carp, rainbow trout, common roach (Rutilus rutilus), and Atlantic salmon [reviewed in (125)]. Therefore, data on waterborne cortisol needs to be analyzed with caution, since a direct comparison among different studies on different species can be affected by differences in body size, water temperature, metabolic rate, cortisol clearance rate and ventilation rate (125).

Transport and market distribution are also key stressors for ornamental characid species, and efforts are focused on improving the water quality in different ways. In particular, the addition of sodium chloride is one of the most used stress-reducing substances added to transport water for many fish species, including characiforms (126). This is related to the fact electrolytes decrease during the recovery period after hauling, and the addition of salts lower the osmotic gradient between plasma and the environment thus reducing the energy cost of osmoregulation. For example, even if in Brycon amazonicum cortisol plasma levels increase immediately after transportation, this is avoided in presence of salt (127). Severe handling events can be also managed by sedating fish with different analgesics, but the appropriate dose is often not well established for different species. Interestingly, essential oils show anesthetic effects for transport (128), while probiotics help to increase fish health during stressful processes such as severe changes in environmental conditions (129). P. axelrodi is normally caught in small streams and later transported in plastic boxes before being exported to Europe, Asia, and North America (130). One of the major concerns in this process is mortality during local transportation, which can be attributed to bad management practices and poor water quality leading to disease outbreaks (131). Interestingly, even if cortisol levels increase after transport, this effect is decreased when fish are treated with watersoluble commercial probiotics (132). These results suggest that even if transportation is a major stressor in characiforms commercialization, available tools can help minimize this effect and assure fish welfare.

Similar to evidence discussed in cichlids and Cypriniformes, environmental enrichment and overcrowding also affects characids welfare. Environmental complexity was tested in the checker barb *Oliotius oligolepis*, showing a marked preference for a structured compartment enriched with plants and clay pots (133). Moreover, aggression and foraging activity are used as behavioral welfare indicators to assess the effect of environmental enrichment and group size in *Serrapinnus notomelas*, a Neotropical characin commonly used in the aquarium trade, suggesting that environmental enrichment facilitates foraging activity but not aggression (134). Interestingly, even if many species of Characiformes are most important as ornamental species, to the best of our knowledge there is still no evidence on cortisol as a welfare indicator of stress in captivity conditions.

#### 5. Sex steroids and agonistic behavior

The hormonal profile from each individual modulates reproductive and social behavior (e.g., aggression) and, in turn, social interactions with conspecifics also affect the hormonal landscape. In these intertwined and bidirectional effects, steroid levels are usually a result rather than the cause of acquiring a specific social status, since different social contexts modulate hormonal responses (135). In this context, sex steroids are intimately related to aggression and, together with reproductive behavior, they constitute useful information to assess fish welfare.

The social control of reproduction is present in different animal species to a higher or to a lesser extent. Since cichlids are highly social fish, they constitute ideal models to assess how reproduction can be related to different social structures and agonistic interactions (10). This way, dominant and subordinate social status are typically associated to differences in activity throughout the

hypothalamic-pituitary-gonadal axis. In C. dimerus, territorial dominant males show higher plasma 11-ketotestosterone (11-KT, an important fish androgen) and testosterone (T) levels, but reduced estradiol (E2) when compared with non-territorial subordinate males in a social hierarchy (51, 100). In a different ethological context, evidence in dyadic agonistic encounters suggest that winner females show higher  $E_2$  levels than losers (136). Higher circulating levels of androgens are also associated with reproduction and dominant status in males of O. niloticus (137) and N. pulcher (48). Similarly, in A. burtoni, dominant males present higher plasma T, 11-KT, E2 and progesterone (P4) (135). This is paralleled by subordinate males having a low gonadosomatic index, small testes, low fraction of motile sperm, spermatogenesis and sperm density when compared to dominants (138). It is important to emphasize that sex-steroid levels also depend on the ethological context analyzed. As a consequence, when subordinate A. burtoni males are given a chance to rise in rank, they rapidly increase plasma levels of T, 11-KT, E<sub>2</sub> and  $P_4$  (139). On the other hand, a rapid decrease in plasma androgen levels is observed during social descent from a dominant to subordinate social rank (135). These studies illustrate that social interactions and the establishment of dominance modulate sex-steroid levels.

A nature-based approach of fish welfare implies that when individuals are well, they are able to express their whole behavioral repertoire as if they were in a natural environment. Behavioral integrity can be used as a welfare indicator, since animals should be allowed to perform their natural repertoire of behavior to guarantee their overall welfare (140). To give an example, if reproduction succeeds it means that the artificial environment and housing conditions are adequate for fish welfare (141). As already explained in detail in section 3, cichlid fish have complex reproductive repertoire and, taking this behavioral variability into account, tank maintenance should provide optimal conditions for reproduction to occur, including an adequate sex-ratio, social hierarchy, breeding substrate and environmental enrichment that needs to be settled according to species-specific requirements. For example, while P. scalare chooses leaves as a spawning substrate in the natural environment, in captivity fish can lay their eggs in acrylic tubes, suggesting that environmental enrichment can provide an appropriate spawning substrate to facilitate reproduction (142). Water temperature also plays an important role in fish welfare and reproductive patterns. In O. niloticus plasma levels of E<sub>2</sub>, cortisol and vitellogenin were significantly reduced under high temperature conditions, suggesting that sex-hormones and steroidogenesis can also be used as welfare indicators to assess water temperature as a stressor (143). As a consequence, reproductive and parental care behavior can also be used as welfare indicators, also taking into account the physiological levels of sex hormones in each species.

Reproduction of ornamental species in captivity can represent an important challenge, and different protocols have been tested according to each species. In the common carp, when spawning in females is induced with Ovaprim or rGnRH, there is an increase in plasma levels of cortisol, T and  $20\beta$ -dihydroxy-4-pregnen-3-one (DHP), while E<sub>2</sub> levels are reduced (144). Taking into account that reproductive behavior can be usually induced by administration of sex steroids, further evidence in goldfish shows that administration of 11-KT and T can induce courtship masculinization in females, while treatment with prostaglandin induces feminization of reproductive

behavior in males (145). Unfortunately, teleost fish reproduction is drastically affected by endocrine disrupting chemicals present in the aquatic environment (146). Goldfish constitutes a clear example, since certain agents such as Bisphenol affect reproduction at different levels, by inhibiting the expression of genes related to the HPG axis, decreasing 11-KT plasma levels, and disrupting ovarian and testis maturation (147).

Similar to evidence in different cichlids, social behavior in cyprinids is also intimately regulated by sexual steroids. In zebrafish, E2 increases shoaling area and social preference, and decreases cortisol levels in male zebrafish (148), while androgens and progestins regulate aggression (149). Taking this into account, social and reproductive behavior can be used as welfare indicators of exposure to different environmental contaminants. There is broad literature revising how reproductive and social behavior are impaired by 17α-ethinylestradiol (EE2), an oral contraceptive that is found in wastewater and effluents (150). Exposure to EE2 has an anxiolytic effect in adult zebrafish (151) and reduces dominant male aggression, impairing dominance and their reproductive success in half of exposed fish (70). Moreover, EE2 reduces females' court behavior toward males and may have severe implications on the reproductive success of exposed zebrafish populations (152). These studies constitute specific examples on reproductive traits and agonistic behavior as welfare indicators in zebrafish, highlighting how vulnerable teleost fish are to environmental contaminants.

Considering Characiform ornamental species, reproduction in neon tetra P. innesi and cardinal tetra P. axelrodi is extremely sensitive to environmental conditions and represent interesting challenges for breeding systems. Both species usually prefer soft, slightly acidic water that resembles natural conditions, since water in the streams and rivers of the Amazonian region are poor in nutrients and minerals, but rich in humic acids (153). As a consequence, even if aquarists have been trying to breed them since the mid-20th century, reproduction of these species in captivity is a very difficult task. High market pressure stimulated the development of breeding and rearing methods for tetras (33) but because of its economic relevance there is only limited available data on such information. For example, reproduction in P. innesi was successful after a careful protocol with a spawning substrate, increasing temperature up to 25°C, darkness and a specific preparation of acidified water with humic acids (35). In this species, gametes are only produced during few initial spawning events, and even if fish continue to engage in spawning events, resulting offspring is not viable and gametes suffer significant damage after 15-20 days. Also, even if aquarist usually recommend maintaining temperature between 22 and 25°C for this species, evidence on spawning induction suggests that keeping bloodstocks at these temperatures for a prolonged time has a negative effect on spawning and reproduction (154). In P. axelrodi, spawning can be induced in laboratory conditions by controlling pH, electric conductivity, temperature at 26°C and the water level with artificial rain simulation (155). Despite various attempts by breeders or researchers to replicate conditions in this experiment, spawning induction was not achieved, probably because in the original protocol experiments were performed with rainwater from the natural habitat of P. axelrodi (155). In these species in which reproduction in captivity constitutes such a difficult challenge, the importance of diet on fish maintenance deserves special observation. In P. axelrodi, an omnivorous species with a tendency to carnivorous behavior (156), evidence suggests that growth parameters are improved in a diet with 53.5% total protein, and the same effect was reported in *P. innesi* (157). These species represent very extreme examples on the use of reproductive behavior as a welfare indicator of fish husbandry.

#### 6. Final remarks

Intensive production of teleost species with ornamental interest faces not only issues related to water quality, diet and sensitivity to environmental conditions, but also high crop densities and stress due to handling and transport, leading to the associated pathologies. Overall, popular practices used in the confinement and farming of these species are affecting reproduction in captivity. This highlights the importance of studying the basic biological requirements and welfare indicators of species with commercial interest, in doing so providing the optimal conditions required by individuals and especially these species whose reproduction is highly sensitive. Considering the above, efforts should focus on studying species-specific social behavior and behavioral indicators such as aggression, courtship and successful reproduction, and also on assessing individual stress and physiological state of the specimens with non-invasive methods, such as cortisol determinations in water. These behavioral and physiological analyses, in turn, can serve as indicators of health, welfare and sexual status of the specimens, helping as a diagnosis to distinguish which points should be further assessed and improved.

#### Author contributions

MFS conceived the original design and conception of this manuscript. MFS, LC, and LR wrote the manuscript. All authors contributed to the article and approved the submitted version.

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## Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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#### References

1. Huntingford F, Adams C, Braithwait VA, Kadri S, Pottinger TG, Sandøe P, et al. Current issues in fish welfare. *J Fish Biol.* (2006) 68:332–72. doi: 10.1111/j.0022-1112.2006.001046.x

2. Rousing T, Bonde M, Sørensen JT. Aggregating welfare indicators into an operational welfare assessment system: a bottom-up approach. *Acta Agric Scand Anim Sci Suppl.* (2001) 30:53–8. doi: 10.1080/090647001316923063

3. Noble C, Gismervik K, Iversen MH, Kolarevic J, Nilsson J, Stien LH, et al. *Welfare indicators for farmed Atlantic salmon: tools for assessing fish welfare*. Tromsø: Nofima (2018) Available at: http://hdl.handle.net/11250/2575780.

4. Martins CI, Galhardo L, Noble C, Damsgård B, Spedicato MT, Zupa W, et al. Behavioural indicators of welfare in farmed fish. *Fish Physiol Biochem*. (2012) 38:17–41. doi: 10.1007/s10695-011-9518-8

5. Huntingford FA. Fish Behaviour: Determinants and Implications for Welfare In: . *The Welfare of Fish. Animal Welfare*, eds. Kristiansen, T., Fernö, A., Pavlidis, M., van de Vis, H. vol 20. Springer, Cham. (2020). 73–110. doi: 10.1007/978-3-030-41675-1\_4

6. Ramallo MR, Morandini L, Alonso F, Birba A, Tubert C, Fiszbein A, et al. The endocrine regulation of cichlids social and reproductive behavior through the eyes of the chanchita, *Cichlasoma dimerus* (Percomorpha; Cichlidae). *J Physiol Paris*. (2014) 108:194–202. doi: 10.1016/j.jphysparis.2014.08.004

7. Maruska KP, Anselmo CM, King T, Mobley RB, Ray EJ, Wayne R. Endocrine and neuroendocrine regulation of social status in cichlid fishes. *Horm Behav.* (2022) 139:105110. doi: 10.1016/j.yhbeh.2022.105110

8. Froese R, Pauly D, editors. Fish base. World Wide Web Electronic Publication (2022). Available at: https://www.fishbase.org (Accessed March 02, 2022).

9. Evers HG, Pinnegar JK, Taylor MI. Where are they all from? – sources and sustainability in the ornamental freshwater fish trade. *J Fish Biol.* (2019) 94:909–16. doi: 10.1111/jfb.13930

10. Scaia MF, Cavallino L, Pandolfi M. Social control of spermatogenesis and steroidogenesis in cichlid fish: a comparative approach. *Reproduction*. (2020) 159:R31-43. doi: 10.1530/REP-18-0650

11. Ilves KL, Torti D. López-Fernández H exon-based phylogenomics strengthens the phylogeny of Neotropical cichlids and identifies remaining conflicting clades (Cichliformes: Cichlidae: Cichlinae). *Mol Phylogenet Evol.* (2018) 118:232–43. doi: 10.1016/j.ympev.2017.10.008

12. Kullander SO. Cichlidae (cichlids) In: RE Reis, SO Kullander and FerrarisCJ Jr, editors. *Checklist of the freshwater fishes of south and Central America*. Porto Alegre: Edipucrs (2003). 605–54.

13. Romano D, Stefanini C. Individual neon tetras (*Paracheirodon innesi*, Myers) optimise their position in the group depending on external selective contexts: lesson learned from a fish-robot hybrid school. *Biosyst Eng.* (2021) 204:170–80. doi: 10.1016/j. biosystemseng.2021.01.021

14. Nelson JS, Grande TC, Wilson MVH. Fishes of the world. 5th ed. Hoboken: John Wiley and Sons (2016). 752 p.

15. Seehausen O. African cichlid fish: a model system in adaptive radiation research. *Proc Biol Sci.* (2006) 273:1987–98. doi: 10.1098/rspb.2006.3539

16. Tacon AG. Trends in global aquaculture and aquafeed production: 2000–2017. *Rev Fish Sci Aquac.* (2020) 28:43–56. doi: 10.1080/23308249.2019.1649634

17. Senanan W, Bart AN. *The potential risks from farm escaped tilapias*, Sustainable Fisheries Partnership, United States (2009). 15 p.

18. Beveridge MCM, McAndrew BJ. Tilapias: biology and exploitation In: . *Fish and fisheries series*. Dordrecht: Springer (2000). 505.

19. Ellison AR, Webster TMU, Rodriguez-Barreto D, de Leaniz CG, Consuegra S, Orozco-ter Wengel P, et al. Comparative transcriptomics reveal conserved impacts of rearing density on immune response of two important aquaculture species. *Fish Shellfish Immunol.* (2020) 104:192–201. doi: 10.1016/j.fsi.2020.05.043

20. Abdel-Latif HM, Hendam BM, Shukry M, El-Shafai NM, El-Mehasseb IM, Dawood MA, et al. Effects of sodium butyrate nanoparticles on the hematoimmunological indices, hepatic antioxidant capacity, and gene expression responses in *Oreochromis niloticus. Fish Shellfish Immunol.* (2021) 119:516–23. doi: 10.1016/j. fsi.2021.10.039

21. Brown C, Wolfenden D, Sneddon L. Goldfish (*Carassius auratus*) In: J Yeates, editor. *Companion animal care and welfare: the UFAW companion animal handbook*. Hoboken, NJ: John Wiley & Sons (2018). 467–78.

organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

22. Wang SY, Luo J, Murphy RW, Wu SF, Zhu CL, Gao Y, et al. Origin of Chinese goldfish and sequential loss of genetic diversity accompanies new breeds. *PLoS One.* (2013) 8:e59571. doi: 10.1371/journal.pone.0059571

23. Barus V, Peaz M, Kohlmann K. *Cyprinus carpio* (Linnaeus, 1758) In: PM Banarescu and HJ Paepke, editors. *The freshwater fishes of Europe*, vol. 5 (2001). Wiebelsheim, Germany: Aula Verlag. 85–179.

24. Rahman MM. Role of common carp (*Cyprinus carpio*) in aquaculture production systems. *Front Life Sci.* (2015) 8:399–410. doi: 10.1080/21553769.2015.1045629

25. Norton WHJ. Screening for drugs to reduce aggression in zebrafish. *Neuropharmacology.* (2019) 156:107394. doi: 10.1016/j.neuropharm.2018.10.023

26. Sundin J, Morgan R, Finnøen MH, Dey A, Sarkar K, Jutfelt F. On the observation of wild zebrafish (*Danio rerio*) in India. *Zebrafish*. (2019) 16:546–53. doi: 10.1089/zeb.2019.1778

27. Egan RJ, Bergner CL, Hart PC, Cachat JM, Canavello PR, Elegante MF, et al. Understanding behavioral and physiological phenotypes of stress and anxiety in zebrafish. *Behav Brain Res.* (2009) 205:38–44. doi: 10.1016/j.bbr.2009.06.022

28. Lee CJ, Paull GC, Tyler CR. Improving zebrafish laboratory welfare and scientific research through understanding their natural history. *Biol Rev Camb Philos Soc.* (2022) 97:1038–56. doi: 10.1111/brv.12831

29. Nelson JS. Fishes of the world. 3rd ed. New York, NY: John Wiley & Sons (1994). 600 p.

30. Reis RE, Kullander SO, Ferraris CJ Jr. Checklist of the freshwater fishes of south and Central America. Edipucrs: Porto Alegre (2003).

31. Mirande JM. Combined phylogeny of ray-finned fishes (Actinopterygii) and the use of morphological characters in large-scale analyses. *Cladistics*. (2017) 33:333–50. doi: 10.1111/cla.12171

32. Balon EK. About the oldest domesticates among fishes. J Fish Biol. (2004) 65:1–27. doi: 10.1111/j.0022-1112.2004.00563.x

33. Kupren K, Kucharczyk D, Prusińska M, Krejszeff S, Targońska K, Mamcarz A. The influence of stocking density on survival and growth of Buenos Aires Tetra *Hemigrammus caudovittatus* larvae reared under controlled conditions. *Pol J Nat Sci.* (2008) 23:881–7. doi: 10.2478/v10020-008-0071-6

34. Weitzman SH, Fink WL. Neon tetra relationships and phylogenetic systematics. *Tropical Fish Hob.* (1987) 36:72–7.

35. Chapman FA, Colle DE, Rottmann RW, Shireman JV. Controlled spawning of the neon tetra. *Progr Fish Cult.* (1998) 60:32–7. doi: 10.1577/1548-8640(1998)060<0032:CS OTNT>2.0.CO;2

36. Saengsitthisak B, Punyapornwithaya V, Chaisri W, Mektrirat R, Bernard JK, Pikulkaew S. The current state of biosecurity and welfare of ornamental fish population in pet fish stores in Chiang Mai Province. *Thailand Vet Integr Sci.* (2021) 19:277–94. doi: 10.12982/VIS.2021.025

37. Saraiva JL, Rachinas-Lopes P, Arechavala-Lopez P. Finding the "golden stocking density": a balance between fish welfare and farmers' perspectives. *Front Vet Sci.* (2022) 9:930221. doi: 10.3389/fvets.2022.930221

38. Arechavala-Lopez P, Cabrera-Álvarez MJ, Maia CM, Saraiva JL. Environmental enrichment in fish aquaculture: a review of fundamental and practical aspects. *Rev Aquac.* (2022) 14:704–28. doi: 10.1111/raq.12620

39. Pandolfi M, Scaia MF, Fernandez MP. Sexual dimorphism in aggression: sexspecific fighting strategies across species. *Front Behav Neurosci*. (2021) 15:659615. doi: 10.3389/fnbeh.2021.659615

40. Fernald RD, Hirata NR. Field study of *Haplochromis burtoni*: quantitative behavioural observations. *Anim Behav.* (1977) 25:964–75. doi: 10.1016/0003-3472(77)90048-3

41. Renn SC, Fraser EJ, Aubin-Horth N, Trainor BC, Hofmann HA. Females of an African cichlid fish display male-typical social dominance behavior and elevated androgens in the absence of males. *Horm Behav.* (2012) 61:496–503. doi: 10.1016/j. yhbeh.2012.01.006

42. Renn SC, Carleton JB, Magee H, Nguyen MLT, Tanner AC. Maternal care and altered social phenotype in a recently collected stock of *Astatotilapia burtoni* cichlid fish. *Integr Comp Biol.* (2009) 49:660–73. doi: 10.1093/icb/icp085

43. Oliveira R, Almada V. Mating tactics and male-male courtship in the lek-breeding cichlid Oreochromis mossambicus. J Fish Biol. (1998) 52:1115–29.

44. Gonçalves-de-Freitas E, Bolognesi MC, Gauy ACS, Brandão ML, Giaquinto PC, Fernandes-Castilho M. Social behavior and welfare in Nile tilapia. *Aust Fish.* (2019) 4:23. doi: 10.3390/fishes4020023

45. Evans JJ, Pasnik DJ, Horley P, Kraeer K, Klesius PH. Aggression and mortality among Nile tilapia (*Oreochromis niloticus*) maintained in the laboratory at different densities. *J Anim Sci.* (2008) 2:57–64.

46. Barki A, Cnaani A, Biran J. How does temperature affect aggression during and after dominance hierarchy formation in Nile tilapia? *Appl Anim Behav Sci.* (2022) 247:105563. doi: 10.1016/j.applanim.2022.105563

47. Taborsky M, Limberger D. Helpers in fish. *Behav Ecol Sociobiol*. (1981) 8:143–5. doi: 10.1007/BF00300826

48. Taves MD, Desjardins JK, Mishra S, Balshine S. Androgens and dominance: sexspecific patterns in a highly social fish (*Neolamprologus pulcher*). *Gen Comp Endocrinol.* (2009) 161:202–7. doi: 10.1016/j.ygcen.2008.12.018

49. Bruintjes R, Radford AN. Context-dependent impacts of anthropogenic noise on individual and social behaviour in a cooperatively breeding fish. *Anim Behav.* (2013) 85:1343–9. doi: 10.1016/j.anbehav.2013.03.025

50. Balshine S, Abate ME. Parental care in cichlid fishes In: ME Abate and DLG Noakes, editors. *The behavior, ecology and evolution of cichlid fishes: a contemporary modern synthesis.* Dordrecht: Springer (2021). 541–86.

51. Ramallo MR, Birba A, Honji RM, Morandini L, Moreira RG, Somoza GM, et al. A multidisciplinary study on social status and the relationship between inter-individual variation in hormone levels and agonistic behavior in a Neotropical cichlid fish. *Horm Behav.* (2015) 69:139–51. doi: 10.1016/j.yhbeh.2015.01.008

52. Holder JL, Barlow GW, Francis RC. Differences in aggressiveness in the Midas cichlid fish (*Cichlasoma citrinellum*) in relation to sex, reproductive state and the individual. *Ethology*. (1991) 88:297–306.

53. Teresa FB, Goncalves-de-Freitas E. Reproductive behavior and parental roles of the cichlid fish *Laetacara araguaiae*. *Neotrop Ichthyol*. (2011) 9:355–62. doi: 10.1590/S1679-62252011005000018

54. Scaia MF, Morandini L, Noguera CA, Ramallo MR, Somoza GM, Pandolfi M. Fighting cichlids: dynamic of intrasexual aggression in dyadic agonistic encounters. *Behav Process.* (2018) 147:61–9. doi: 10.1016/j.beproc.2017.12.015

55. Brandão ML, Braithwaite VA, Gonçalves-de-Freitas E. Isolation impairs cognition in a social fish. *Appl Anim Behav Sci.* (2015) 171:204–10. doi: 10.1016/j.applanim.2015.08.026

56. Gonçalves-de-Freitas E, Mariguela TC. Social isolation and aggressiveness in the Amazonian juvenile fish Astronotus ocellatus. Braz J Biol. (2006) 66:233–8. doi: 10.1590/S1519-69842006000200007

57. Gonçalves-de-Freitas E, Teresa FB, Gomes FS, Giaquinto PC. Effect of water renewal on dominance hierarchy of juvenile Nile tilapia. *Appl Anim Behav Sci.* (2008) 112:187–95. doi: 10.1016/j.applanim.2007.07.002

58. Gauy ACDS, Boscolo CNP, Gonçalves-de-Freitas E. Less water renewal reduces effects on social aggression of the cichlid *Pterophyllum scalare*. *Appl Anim Behav Sci.* (2018) 198:121-6. doi: 10.1016/j.applanim.2017.10.003

59. Gauy ACDS, Bolognesi MC, Gonçalves-de-Freitas E. Unusual effect of chemical communication on social aggression in juvenile cichlid fish *Cichlasoma paranaense* (Cichliformes: Cichlidae). *Neotrop Ichthyol.* (2019) 17:e180159. doi: 10.1590/1982-0224-20180159

60. Kochhann D, Campos DF, Val AL. Experimentally increased temperature and hypoxia affect stability of social hierarchy and metabolism of the Amazonian cichlid *Apistogramma agassizii. Comp Biochem Physiol A Mol Integr Physiol.* (2015) 190:54–60. doi: 10.1016/j.cbpa.2015.09.006

61. Ratnasabapathi D, Burns J, Souchek R. Effects of temperature and prior residence on territorial aggression in the convict cichlid *Cichlasoma nigrofasciatum*. Aggress Behav. (1992)18:365–72. doi:10.1002/1098-2337(1992)18:5<365::AID-AB2480180506>3.0.CO;2-E

62. Brandão ML, Colognesi G, Bolognesi MC, Costa-Ferreira RS, Carvalho TB, Gonçalves-de-Freitas E. Water temperature affects aggressive interactions in a Neotropical cichlid fish. *Neotrop Ichthyol* (2018. (2018) 16:e170081. doi: 10.1590/1982-0224-20170081

63. Earley RL, Edwards JT, Aseem O, Felton K, Blumer LS, Karom M, et al. Social interactions tune aggression and stress responsiveness in a territorial cichlid fish (*Archocentrus nigrofasciatus*). *Physiol Behav.* (2006) 88:353–63. doi: 10.1016/j. physbeh.2006.04.002

64. Gómez-Laplaza LM, Morgan E. The influence of social rank in the angelfish, *Pterophyllum scalare*, on locomotor and feeding activities in a novel environment. *Lab Anim.* (2003) 37:108–20. doi: 10.1258/00236770360563741

65. Hannes RP, Franck D. The effect of social isolation on androgen and corticosteroid levels in a cichlid fish (*Haplochromis burtoni*) and in swordtails (*Xiphophorus helleri*). *Horm Behav.* (1983) 17:292–301. doi: 10.1016/0018-506X(83)90028-4

66. Mizory FA, Hussein NJ, Hussein NSASR. Effect of different stocking densities on behavior of common carp (*Cyprinus carpio*) in Duhok province, Kurdistan region, Iraq. *Int J Sci Res Biol Sci.* (2020) 7:01–6.

67. Chen J, Rao C, Yuan R, Sun D, Guo S, Li L, et al. Long-term exposure to polyethylene microplastics and glyphosate interferes with the behavior, intestinal microbial homeostasis, and metabolites of the common carp (*Cyprinus carpio* L). *Sci Total Environ.* (2022) 814:152681. doi: 10.1016/j.scitotenv.2021.152681

68. Awatif AS, Al-Khafaji BY, Mansour AJ. Behavioural effect of heavy metals on common carp (*Cyprinus carpio* L.) and Nile tilapia (*Oreochromis niloticus* L.). *Iran J Ichthyol.* (2022) 9:32–6.

69. Scaia MF, Akinrinade I, Petri G, Oliveira RF. Sex differences in aggression are paralleled by differential activation of the brain social decision-making network in zebrafish. *Front Behav Neurosci* (2022, (2022) 16:784835. doi: 10.3389/fnbeh.2022.784835

70. Filby AL, Paull GC, Bartlett EJ, Van Look KJ, Tyler CR. Physiological and health consequences of social status in zebrafish (*Danio rerio*). *Physiol Behav.* (2010) 101:576–87. doi: 10.1016/j.physbeh.2010.09.004

71. Saszik SM, Smith CM. The impact of stress on social behavior in adult zebrafish (*Danio rerio*). *Behav Pharmacol.* (2018) 29:53–9. doi: 10.1097/FBP.00000000000338

72. Kirsten K, Soares SM, Koakoski G, Kreutz LC, Barcellos LJG. Characterization of sickness behavior in zebrafish. *Brain Behav Immun.* (2018) 73:596–602. doi: 10.1016/j. bbi.2018.07.004

73. Rosa LV, Costa FV, Canzian J, Borba JV, Quadros VA, Rosemberg DB. Three-and bi-dimensional analyses of the shoaling behavior in zebrafish: influence of modulators of anxiety-like responses. *Prog Neuropsychopharmacol Biol Psychiatry*. (2020) 102:109957. doi: 10.1016/j.pnpbp.2020.109957

74. Nasiadka A, Clark MD. Zebrafish breeding in the laboratory environment. *ILAR J.* (2012) 53:161–8. doi: 10.1093/ilar.53.2.161

75. Sharma K, Shashank BN, Singh G. Studies on breeding and feeding patterns of the goldfish, *Carassius auratus* under captive conditions for sustainable ornamental fish hatchery management. *Livest Res Rural Dev.* (2011) 23

76. Haniffa MA, Benziger PA, Arockiaraj AJ, Nagarajan M, Siby P. Breeding behaviour and embryonic development of koi carp (*Cyprinus carpio*). Taiwania. (2007) 52:93–9.

77. Darrow KO, Harris WA. Characterization and development of courtship in zebrafish, Danio rerio. Zebrafish. (2004) 1:40-5. doi: 10.1089/154585404774101662

78. Lorenzoni M, Corboli M, Ghetti L, Pedicillo G. Carosi a "growth and reproduction of the goldfish *Carassius auratus*: a case study from Italy" In: F Gherardi, editor. *Biological invaders in inland waters: profiles, distribution, and threats. Invading naturespringer series in invasion ecology*, vol. 2. Dordrecht: Springer (2007). 259–73.

79. Spence R, Smith C, Ashton R. Oviposition decisions are mediated by spawning site quality in wild and domesticated zebrafish, *Danio rerio. Behaviour*. (2007) 144:953–66. doi: 10.1163/156853907781492726

80. Kvarnemo C, Svensson O, Forsgren E. Parental behaviour in relation to food availability in the common goby. *Anim Behav.* (1998) 56:1285–90. doi: 10.1006/anbe.1998.0899

81. Manica A. Filial cannibalism in teleost fish. *Biol Rev.* (2002) 77:261–77. doi: 10.1017/S1464793101005905

82. Ribolli J, Miño CI, Scaranto BMS, Reynalte-Tataje DA, Zaniboni FE. Genetic evidence supports polygamous mating system in a wild population of *Prochilodus lineatus*, a Neotropical shoal spawner fish. *Neotrop Ichthyol*. (2020) 18:e190123. doi: 10.1590/1982-0224-2019-0123

83. Jones AG, Ardren WR. Methods of parentage analysis in natural populations. *Mol Ecol.* (2003) 12:2511–23. doi: 10.1046/j.1365-294X.2003.01928.x

84. Delcourt J, Poncin P. Shoals and schools: back to the heuristic definitions and quantitative references. *Rev Fish Biol Fisheries*. (2012) 22:595–619. doi: 10.1007/s11160-012-9260-z

85. Saxby A, Adams L, Snellgrove D, Wilson RW, Sloman KA. The effect of group size on the behaviour and welfare of four fish species commonly kept in home aquaria. *Appl Anim Behav Sci.* (2010) 125:195–205. doi: 10.1016/j.applanim.2010.04.008

86. Weitzman SH, Menezes NA, Evers HG, Burns JR. Putative relationships among inseminating and externally fertilizing characids, with a description of a new genus and species of Brazilian inseminating fish bearing an anal-fin gland in males (Characiformes: Characidae). *Neotrop Ichtyol.* (2005) 3:329–60. doi: 10.1590/S1679-62252005000300002

87. Amcoff M, Hallsson LR, Winberg S, Kolm N. Male courtship pheromones affect female behavior in the swordtail characin (*Corynopoma riisei*). *Ethology*. (2014) 120:463–70. doi: 10.1111/eth.12217

88. Gonçalves T, Azevedo M, Malabarba L, Fialho C. Reproductive biology and development of sexually dimorphic structures in *Aphyocharax anisitsi* (Ostariophysi: Characidae). *Neotrop Ichtyol*. (2005) 3:433–8. doi: 10.1590/S1679-62252005000300012

89. Pintos S, Cavallino L, Vidal YA, Pandolfi M, Pozzi AG. Effects of intraspecific chemical cues on the behaviour of the bloodfin tetra *Aphyocharax anisitsi* (Ostariophysi: Characidae). *Behav Process.* (2021) 193:104533. doi: 10.1016/j.beproc.2021.104533

90. Sandoval-Herrera N, Mena F, Espinoza M, Romero A. Neurotoxicity of organophosphate pesticides could reduce the ability of fish to escape predation under low doses of exposure. *Sci Rep.* (2019) 9:10530. doi: 10.1038/s41598-019-46804-6

91. Eissa N, Wang HP. Transcriptional stress responses to environmental and husbandry stressors in aquaculture species. *Rev Aquac.* (2016) 8:61–88. doi: 10.1111/raq.12081

92. Mommsen TP, Vijayan MM, Moon TW. Cortisol in teleosts: dynamics, mechanisms of action, and metabolic regulation. *Rev Fish Biol Fisher*. (1999) 9:211–68. doi: 10.1023/A:1008924418720

93. Wendelaar-Bonga SE. The stress response in fish. *Physiol Rev.* (1997) 77:591–625. doi: 10.1152/physrev.1997.77.3.591

94. Winberg S, Höglund E, Øverli Ø. Variation in the neuroendocrine stress response In: CB Schreck, L Tort, AP Farrell and CJ Brauner, editors. *Fish physiology*. London: Academic Press (2016). 35–74.

95. Falahatkar B, Poursaeid S, Shakoorian M, Barton B. Responses to handling and confinement stressors in juvenile great sturgeon *Huso huso. J Fish Biol.* (2009) 75:784–96. doi: 10.1111/j.1095-8649.2009.02334.x

96. Sampaio FDF, Freire CA. An overview of stress physiology of fish transport: changes in water quality as a function of transport duration. *Fish Fish*. (2016) 17:1055-72. doi: 10.1111/faf.12158

97. Johnsson JI, Winberg S, Sloman KA. Social interactions In: KA Sloman, RW Wilson and S Balshine, editors. *Behaviour and physiology of fish*. Oxford: Elsevier Academic Press (2006). 151–96.

98. Gilmour KM, DiBattista JD, Thomas J. Physiological causes and consequences of social status in salmonid fish. *Integr Comp Biol.* (2005) 45:263–73. doi: 10.1093/ icb/45.2.263

99. Fox HE, White SA, Kao MHF, Fernald RD. Stress and dominance in a social fish. *J Neurosci*. (1997) 17:6463–9. doi: 10.1523/JNEUROSCI.17-16-06463.1997

100. Morandini L, Honji RM, Ramallo MR, Moreira RG, Pandolfi M. The interrenal gland in males of the cichlid fish *Cichlasoma dimerus*: relationship with stress and the establishment of social hierarchies. *Gen Comp Endocrinol.* (2014) 195:88–98. doi: 10.1016/j.ygcen.2013.10.009

101. Mileva VR, Fitzpatrick JL, Marsh-Rollo S, Gilmour KM, Wood CM, Balshine S. The stress response of the highly social African cichlid *Neolamprologus pulcher*. *Physiol Biochem Zool*. (2009) 82:720–9. doi: 10.1086/605937

102. Correa S, Fernandes M, Iseki K, Negrao J. Effect of the establishment of dominance relationships on cortisol and other metabolic parameters in Nile tilapia (*Oreochromis niloticus*). Braz J Med Biol Res. (2003) 36:1725–31. doi: 10.1590/ S0100-879X2003001200015

103. Creel S, Dantzer B, Goymann W, Rubenstein DR. The ecology of stress: effects of the social environment. *Funct Ecol.* (2013) 27:66–80. doi: 10.1111/j.1365-2435.2012.02029.x

104. Barreto TN, Boscolo CNP, Gonçalves-de-Freitas E. Homogeneously sized groups increase aggressive interaction and affect social stress in Thai strain Nile tilapia (*Oreochromis niloticus*). *Mar Freshw Behav Physiol*. (2015) 48:309–18. doi: 10.1080/10236244.2015.1070478

105. Williams TD, Readman GD, Owen SF. Key issues concerning environmental enrichment for laboratory-held fish species. *Lab Anim.* (2009) 43:107–20. doi: 10.1258/ la.2007.007023

106. Volpato GL, Barreto RE. Environmental blue light prevents stress in the fish Nile tilapia. *Braz J Med Biol.* (2001) 34:1041–5. doi: 10.1590/S0100-879X2001000800011

107. Galhardo L, Oliveira RF. The effects of social isolation on steroid hormone levels are modulated by previous social status and context in a cichlid fish. *Horm Behav.* (2014) 65:1–5. doi: 10.1016/j.yhbeh.2013.10.010

108. Soares MC, Oliveira RF, Ros AFH, Grutter AS, Bshary R. Tactile stimulation lowers stress in fish. *Nat Commun.* (2011) 2:534–5. doi: 10.1038/ncomms1547

109. Bolognesi MC, Dos Santos Gauy AC, Gonçalves-de-Freitas E. Tactile stimulation reduces aggressiveness but does not lower stress in a territorial fish. *Sci Rep.* (2019) 9:40. doi: 10.1038/s41598-018-36876-1

110. Gauy ACDS, Bolognesi MC, Martins GD, Gonçalves-de-Freitas E. Preference and motivation tests for body tactile stimulation in fish. *Animals.* (2021) 11:2042. doi: 10.3390/ani11072042

111. Hoseini SM, Yousefi M, Hoseinifar SH, Van Doan H. Effects of dietary arginine supplementation on growth, biochemical, and immunological responses of common carp (*Cyprinus carpio* L.), stressed by stocking density. *Aquaculture*. (2019) 503:452–9. doi: 10.1016/j.aquaculture.2019.01.031

112. Yousefi M, Hoseini SM, Vatnikov YA, Kulikov EV, Drukovsky SG. Rosemary leaf powder improved growth performance, immune and antioxidant parameters, and crowding stress responses in common carp (*Cyprinus carpio*) fingerlings. *Aquaculture*. (2019) 505:473–80. doi: 10.1016/j.aquaculture.2019.02.070

113. Hoseini SM, Gupta SK, Yousefi M, Kulikov EV, Drukovsky SG, Petrov AK, et al. Mitigation of transportation stress in common carp, *Cyprinus carpio*, by dietary administration of turmeric. *Aquaculture*. (2022) 546:737380. doi: 10.1016/j. aquaculture.2021.737380

114. Yousefi M, Vatnikov YA, Kulikov EV, Plushikov VG, Drukovsky SG, Hoseinifar SH, et al. The protective effects of dietary garlic on common carp (*Cyprinus carpio*) exposed to ambient ammonia toxicity. *Aquaculture*. (2020) 526:735400. doi: 10.1016/j. aquaculture.2020.735400

115. Khan NA, Sharma J, Chakrabarti R. The study of ameliorative effect of dietary supplementation of vitamin C, vitamin E, and tryptophan on *Labeo rohita* (Cyprinidae) fry exposed to intense light. *Fish Physiol Biochem.* (2019) 45:1153–65. doi: 10.1007/s10695-019-00626-5

116. Azeredo R, Machado M, Martos-Sitcha JA, Martínez-Rodríguez G, Moura J, Peres H, et al. Dietary tryptophan induces opposite health-related responses in the Senegalese sole (*Solea senegalensis*) reared at low or high stocking densities with implications in disease resistance. *Front Physiol.* (2019) 10:508. doi: 10.3389/fphys.2019.00508

117. Cabanillas-Gámez M, Bardullas U, Galaviz MA, Rodriguez S, Rodriguez VM, López LM. Tryptophan supplementation helps totoaba (*Totoaba macdonaldi*) juveniles to regain homeostasis in high-density culture conditions. *Fish Physiol Biochem*. (2020) 46:597–611. doi: 10.1007/s10695-019-00734-2

118. Lepage O, Larson ET, Mayer I, Winberg S. Serotonin but not melatonin, plays a role in shaping dominant-subordinate relationships and aggression in rainbow trout. *Horm Behav.* (2005) 48:233–42. doi: 10.1016/j.yhbeh.2005.02.012

119. Morandini L, Ramallo MR, Moreira RG, Höcht C, Somoza GM, Silva A, et al. Serotonergic outcome, stress and sexual steroid hormones, and growth in a south American cichlid fish fed with an L-tryptophan enriched diet. *Gen Comp Endocrinol.* (2015) 223:27–37. doi: 10.1016/j.ygcen.2015.10.005

120. Morandini L, Ramallo MR, Scaia MF, Höcht C, Somoza GM, Pandolfi M. Dietary L-tryptophan modulates agonistic behavior and brain serotonin in male dyadic contests of a cichlid fish. *J Comp Physiol A Neuroethol Sens Neural Behav Physiol.* (2019) 205:867–80. doi: 10.1007/s00359-019-01373-x

121. Saiz N, Gómez-Boronat M, De Pedro N, Delgado MJ, Isorna E. The lack of lightdark and feeding-fasting cycles alters temporal events in the goldfish (*Carassius auratus*) stress axis. *Animals*. (2021) 11:669. doi: 10.3390/ani11030669

122. Marcon M, Mocelin R, Benvenutti R, Costa T, Herrmann AP, de Oliveira DL, et al. Environmental enrichment modulates the response to chronic stress in zebrafish. *J Exp Biol.* (2018) 221:176735. doi: 10.1242/jeb.176735

123. de Abreu MS, Giacomini AC, Genario R, Dos Santos BE, Marcon L, Demin KA, et al. The impact of housing environment color on zebrafish anxiety-like behavioral and physiological (cortisol) responses. *Gen Comp Endocrinol.* (2020) 294:113499. doi: 10.1016/j.ygcen.2020.113499

124. Marchetto L, Barcellos LJ, Koakoski G, Soares SM, Pompermaier A, Maffi VC, et al. Auditory environmental enrichment prevents anxiety-like behavior, but not cortisol responses, evoked by 24-h social isolation in zebrafish. *Behav Brain Res.* (2021) 404:113169. doi: 10.1016/j.bbr.2021.113169

125. Pavlidis M, Digka N, Theodoridi A, Campo A, Barsakis K, Skouradakis G, et al. Husbandry of zebrafish, *Danio rerio*, and the cortisol stress response. *Zebrafish*. (2013) 10:524–31. doi: 10.1089/zeb.2012.0819

126. Carmichael GJ, Tomasso JR, Simco BA, Davis KB. Characterization and alleviation of stress associated with hauling largemouth bass. *Trans Am Fish Soc.* (1984) 113:778–85. doi: 10.1577/1548-8659(1984)113<778:CAAOSA>2.0.CO;2

127. Urbinati EC. Carneiro PCF "Práticas de manejo e estresse dos peixes em piscicultura intensiva" In: JEP Cyrino, EC Urbinati, DM Fracalossi and N Castagnolli, editors. *Tópicos especiais em piscicultura de água Doce tropicas intensiva*. São Paulo: TecArt (2004). 171–94.

128. Almeida APG, Correia TG, Heinzmann BM, Val AL, Baldisserotto B. Stressreducing and anesthetic effects of the essential oils of *Aloysia triphylla* and *Lippia alba* on *Serrasalmus eigenmanni* (Characiformes: Serrasalmidae). *Neotrop Ichtyol*. (2019) 17:e190021. doi: 10.1590/1982-0224-20190021

129. Taoka Y, Maeda H, Jo J, Jeon M, Bai SC, Lee W, et al. Growth stress tolerance and non-specific immune response of Japanese flounder *Paralichthy solivaceus* to probiotics in a closed recirculating system. *Fisher Sci.* (2006) 72:310–21. doi: 10.1111/j.1444-2906.2006.01152.x

130. Santos GMD, Santos ACMD. Sustentabilidade da pesca na Amazônia. *Estud Avanç*. (2005) 19:165–82. doi: 10.1590/S0103-40142005000200010

131. Waichman AV, Silva MP, Marcon JL. Water quality monitoring during commercialization of Amazonian ornamental fish In: LN Chao, P Petry, G Prang, L Sonneschein and M Tlusty, editors. *Conservation and management of ornamental fish resources of the Rio Negro Basin, Amazonia, Brazil (Project Piaba)*. Manaus: University of Amazonas Press (2001). 279–99.

132. Gómes LC, Brinn RP, Marcon JL, Dantas LA, Brandão FR, De Abreu JS, et al. Benefits of using the probiotic Efinol<sup>®</sup>L during transportation of cardinal tetra, *Paracheirodon axelrodi* (Schultz), in the Amazon. *Aquac Res.* (2009) 40:157–65. doi: 10.1111/j.1365-2109.2008.02077.x

133. Kistler C, Hegglin D, Wurbel H, Konig B. Preference for structured environment in zebrafish (*Danio rerio*) and checker barbs (*Puntius oligolepis*). *Appl Anim Behav Sci.* (2011) 135:318–27. doi: 10.1016/j.applanim.2011.10.014

134. Da Silva A, Lima MR, Meletti PC, Jerep FC. Impact of environmental enrichment and social group size in the aggressiveness and foraging activity of *Serrapinnus notomelas*. *Appl Anim Behav Sci*. (2020) 224:104943. doi: 10.1016/j.applanim.2020.104943

135. Maruska KP, Becker L, Neboori A, Fernald RD. Social descent with territory loss causes rapid behavioral endocrine and transcriptional changes in the brain. *J Exp Biol.* (2013) 216:3656–66. doi: 10.1242/jeb.088617

136. Scaia MF, Morandini L, Noguera C, Trudeau VL, Somoza GM, Pandolfi M. Can estrogens be considered as key elements of the challenge hypothesis? The case of intrasexual aggression in a cichlid fish. *Physiol Behav.* (2018) 194:481–90. doi: 10.1016/j. physbeh.2018.06.028

137. Pfennig F, Kurth T, Meißner S, Standke A, Hoppe M, Zieschang F, et al. The social status of the male Nile tilapia (*Oreochromis niloticus*) influences testis structure and gene expression. *Reproduction*. (2012) 143:71–84. doi: 10.1530/REP-11-0292

138. Maruska KP. Social regulation of reproduction in male cichlid fishes. *Gen Comp Endocrinol.* (2014) 207:2–12. doi: 10.1016/j.ygcen.2014.04.038

139. Maruska KP, Fernald RD. Behavioral and physiological plasticity: rapid changes during social ascent in an African cichlid fish. *Horm Behav.* (2010) 58:230–40. doi: 10.1016/j.yhbeh.2010.03.011

140. Würbel H. The state of ethological approaches to the assessment of animal suffering and welfare. *Appl Anim Behav Sci.* (2009) 118:105–7. doi: 10.1016/j.applanim.2009.02.021

141. Brandao ML, Dorigão-Guimarães F, Bolognesi MC, Gauy ACDS, Pereira AVS, Vian L, et al. Understanding behaviour to improve the welfare of an ornamental fish. *J Fish Biol.* (2021) 99:726–39. doi: 10.1111/jfb.14802

142. Cacho MDSR, Yamamoto ME, Chellappa S. Comportamento reprodutivo do acara bandeira, *Pterophyllum scalare* Cuvier & Valenciennes (Osteichthyes, Cichlidae). *Rev Bras Zool.* (1999) 16:653–64. doi: 10.1590/S0101-81751999000300006

143. Mhmoud S, Sabry A, Abdelaziz A, Shukry M. Deleterious impacts of heat stress on steroidogenesis markers, immunity status and ovarian tissue of Nile tilapia (*Oreochromis niloticus*). J Therm Biol. (2020) 91:102578. doi: 10.1016/j.jtherbio.2020.102578

144. Yeganeh S, Mohammadzadeh S, Moradian F, Milla S. The effects of recombinant GnRH with dopamine antagonist on reproduction performance, sex steroid levels, and stress response in female koi carp (*Cyprinus carpio*). Aquac Rep. (2022) 22:101001. doi: 10.1016/j.aqrep.2021.101001

145. Munakata A, Kobayashi M. Endocrine control of sexual behavior in teleost fish. Gen Comp Endocrinol. (2010) 165:456-68. doi: 10.1016/j.ygcen.2009.04.011

146. Senthilkumaran B. Pesticide-and sex steroid analogue-induced endocrine disruption differentially targets hypothalamo – hypophyseal – gonadal system during gametogenesis in teleosts – a review. *Gen Comp Endocrinol.* (2015) 219:136–42. doi: 10.1016/j.ygcen.2015.01.010

147. Wang Q, Yang H, Yang M, Yu Y, Yan M, Zhou L, et al. Toxic effects of bisphenol a on goldfish gonad development and the possible pathway of BPA disturbance in female and male fish reproduction. *Chemosphere.* (2019) 221:235–45. doi: 10.1016/j. chemosphere.2019.01.033

148. Moraes AB, Giacomini AC, Genario R, Marcon L, Scolari N, Bueno BW, et al. Prosocial and anxiolytic-like behavior following a single 24-h exposure to  $17\beta$ -estradiol in adult male zebrafish. *Neurosci Lett.* (2021) 747:135591. doi: 10.1016/j.neulet.2020.135591

149. Carver JJ, Carrell SC, Chilton MW, Brown JN, Yong L, Zhu Y, et al. Nuclear androgen and progestin receptors inversely affect aggression and social dominance in male zebrafish (*Danio rerio*). *Horm Behav.* (2021) 134:105012. doi: 10.1016/j. yhbeh.2021.105012

150. Liang YQ, Huang GY, Zhao JL, Shi WJ, Hu LX, Tian F, et al. Transcriptional alterations induced by binary mixtures of ethinylestradiol and norgestrel during the early development of zebrafish (*Danio rerio*). *Comp Biochem Physiol C Toxicol Pharmacol.* (2017) 195:60–7. doi: 10.1016/j.cbpc.2017.02.005

151. Tamagno WA, de Oliveira Sofiatti JR, Alves C, Sutorillo NT, Vanin AP, Bilibio D, et al. Synthetic estrogen bioaccumulates and changes the behavior and biochemical biomarkers in adult zebrafish. *Environ Toxicol Pharmacol.* (2022) 92:103857. doi: 10.1016/j.etap.2022.103857

152. Coe TS, Söffker MK, Filby AL, Hodgson D, Tyler CR. Impacts of early life exposure to estrogen on subsequent breeding behavior and reproductive success in zebrafish. *Environ Sci Technol.* (2010) 44:6481–7. doi: 10.1021/es101185b

153. Junk WJ, Soares MGM. Freshwater fish habitats in Amazonia: state of knowledge, management, and protection. *Aquat Ecosyst Health Manag*. (2001) 2001:437–51. doi: 10.1080/146349801317276107

154. Kucharczyk D, Targońska K, Żarski D, Krejszeff S, Kupren K, Łuczyński MJ, et al. The reproduction of neon tetra, *Paracheirodon innesi* (Myers, 1936), under controlled conditions. *Pol J Natur Sc.* (2010) 25:81–92. doi: 10.2478/ v10020-010-0006-x

155. Beltrão H, Anjos C. Biología reprodutiva e desenvolvimento embrionário e larval do cardinal tetra, *Paracheirodon axelrodi* Schultz, 1956 (Characiformes: Characidae), em laboratório. *Bol Inst Pesca*. (2006) 32:151–60.

156. Walker I. The food spectrum of the cardinal-tetra (*Paracheirodon axelrodi*, Characidae) in its natural habitat. *Acta Amazon*. (2004) 34:69–73. doi: 10.1590/S0044-59672004000100009

157. Luna-Figueroa J, Figueroa-Torres J, Soriano-Salazar M. Efecto de diferentes niveles de proteína de la dieta sobre el crecimiento de juveniles del pez neón *Paracheirodon innesi* (pisces: characidae). *Uniciencia*. (2001) 18:15–20.