

OPEN ACCESS

EDITED BY

Vladimír Jekl,
University of Veterinary and Pharmaceutical
Sciences Brno, Czechia

REVIEWED BY

Sokol Duro,
Agricultural University of Tirana, Albania
Bruno Alberigi,
Federal Rural University of Rio de
Janeiro, Brazil

*CORRESPONDENCE

Hakyoung Yoon
✉ hyyoon@jbnu.ac.kr

RECEIVED 28 December 2023

ACCEPTED 24 July 2024

PUBLISHED 08 August 2024

CITATION

Lee C, Kim M, Han J-I, Lee K and Yoon H
(2024) Evaluation of intrapulmonary
arteriovenous anastomoses before and after
oxygen supplementation, using transthoracic
agitated saline contrast echocardiography in
rescued Korean raccoon dogs.
Front. Vet. Sci. 11:1362363.
doi: 10.3389/fvets.2024.1362363

COPYRIGHT

© 2024 Lee, Kim, Han, Lee and Yoon. This is
an open-access article distributed under the
terms of the [Creative Commons Attribution
License \(CC BY\)](https://creativecommons.org/licenses/by/4.0/). The use, distribution or
reproduction in other forums is permitted,
provided the original author(s) and the
copyright owner(s) are credited and that the
original publication in this journal is cited, in
accordance with accepted academic practice.
No use, distribution or reproduction is
permitted which does not comply with these
terms.

Evaluation of intrapulmonary arteriovenous anastomoses before and after oxygen supplementation, using transthoracic agitated saline contrast echocardiography in rescued Korean raccoon dogs

Chang-eun Lee¹, Myeongsu Kim^{2,3}, Jae-Ik Han^{2,3}, Kichang Lee¹
and Hakyoung Yoon^{1,4*}

¹Department of Veterinary Medical Imaging, College of Veterinary Medicine, Jeonbuk National University, Iksan-si, Republic of Korea, ²Laboratory of Wildlife Medicine, College of Veterinary Medicine, Jeonbuk National University, Iksan-si, Republic of Korea, ³Jeonbuk Wildlife Center, Jeonbuk National University, Iksan-si, Republic of Korea, ⁴Biosafety Research Institute and College of Veterinary Medicine, Jeonbuk National University, Iksan-si, Republic of Korea

Introduction: Intrapulmonary arteriovenous anastomoses (IPAVAs) are defined as relatively large, dynamic shunt vessels that connect the pulmonary arterial and venous systems, thereby bypassing the pulmonary capillary system. IPAVAs lower elevated pulmonary arterial pressure; however, the presence of the shunt can result in impaired pulmonary gas exchange and paradoxical embolism. Furthermore, the prevalence and effects of IPAVAs in raccoon dogs remain unknown. This study aimed to determine the prevalence of IPAVA among rescued Korean raccoon dogs and evaluate the changes in IPAVA following oxygen supplementation.

Methods: Nineteen raccoon dogs rescued by the Jeonbuk Wildlife Centre between August 2022 and December 2023 were subjected to echocardiography. Sixteen healthy and three abnormal raccoon dogs were subjected to transthoracic agitated saline contrast echocardiography (bubble study) based on the echocardiography results. IPAVA was considered to be present if the left heart contrast was visualised after four cardiac cycles following the visualisation of the first right heart contrast. Bubble scores (BS0–5) were assigned based on the maximum number of microbubbles observed in the left ventricular lumen per frame of the ultrasound image. BS was assigned before and after supplementation with 100% oxygen for 5 min.

Results: IPAVA was detected in 12 of the 16 healthy raccoon dogs at rest (75%). The BS of the 15 IPAVA-positive raccoon dogs ranged from 1 to 4 points (BS1, 1; BS2, 4; BS3, 8; and BS4, 2). Blood flow through the IPAVA (Q_{IPAVA}) was reduced or absent in the 15 IPAVA-positive raccoon dogs after supplementation with 100% oxygen (BS0, 11; BS2, 4). Moreover, BS of the IPAVA showed a significant correlation with the cardiac output per body weight (BW).

Conclusion: The prevalence of IPAVA in healthy raccoon dogs at rest was 75%. Adequate oxygen supplementation was found to be effective in reducing Q_{IPAVA} , which may help prevent potential negative factors such as pulmonary gas exchange impairments and paradoxical embolism that can occur with IPAVA.

KEYWORDS

IPAVA, bubble study, hyperoxia, right to left shunt, transpulmonary shunt

1 Introduction

Intrapulmonary arteriovenous anastomoses (IPAVAs) are shunt vessels larger than the pulmonary capillaries that connect the pulmonary arterial system to the pulmonary venous system, thereby bypassing the pulmonary capillaries (1–4). The presence of these unusual blood vessels in humans was first suggested ~100 years ago (5); however, the pathological or physiological significance of these structures remains controversial and has not been fully elucidated (2, 6).

A characteristic feature of IPAVA is that it is a dynamic vessel that can open and close under the influence of specific factors. Oxygen concentration is one such factor. Blood flow through IPAVA (Q_{IPAVA}) in humans may increase under conditions of hypoxia (7–10) and decrease under conditions of hyperoxia (7, 11). A pronounced increase in Q_{IPAVA} during exercise (12–15) has also been observed in humans, dogs, and horses. Q_{IPAVA} increases the concentration of catecholamines, such as epinephrine, dopamine, and dobutamine, in humans (16–18). Q_{IPAVA} in humans has been shown to decrease in the lateral recumbent position compared with that in the upright position (19). Thus, posture, as well as endogenous and exogenous neurotransmitters, affect IPAVA. IPAVA is more prevalent during the foetal and neonatal periods than during adulthood in sheep (20). Similarly, it is less prevalent among older adults in humans (12).

It is unclear whether IPAVA is a physiological or pathological variation, and its clinical implications and relevance remain unknown (21). IPAVA may be a remnant of non-degenerating foetal blood vessels in the lungs. These vessels may originally be present during the foetal period when pulmonary gas exchange is not required and gradually disappear after birth (4, 20, 22). Pulmonary hypertension (PH) is less prevalent among patients with IPAVA, suggesting that IPAVA may play a physiological regulatory role by minimising the damaging effects of increased pressure and reducing right ventricular (RV) afterload (12, 22, 23). However, the presence of the IPAVA is associated with several complications. As Q_{IPAVA} bypasses pulmonary capillaries, it frees these vessels from their role in pulmonary gas exchange and

biological filtration. Studies conducted using microspheres have shown that $1.4 \pm 0.8\%$ of the cardiac output is shunted in dogs with IPAVA. Studies conducted using ^{99m}Tc -MAA have shown that $>5\%$ of the cardiac output is shunted in humans (8, 13). As even 2% of shunting results in a decrease in the alveolar-to-arterial oxygen difference, pulmonary gas exchange can be impaired by the increase in Q_{IPAVA} (2). Paradoxical embolism is another clinical manifestation of IPAVA wherein large substances filtered out by the pulmonary capillaries pass into the systemic circulation as the diameter of IPAVA (>25 or $50\ \mu\text{m}$) is much larger than that of the pulmonary capillaries ($7\text{--}10\ \mu\text{m}$) (3, 13, 24, 25). Emboli of $<100\ \mu\text{m}$ in size can lead to cerebral ischaemia and infarction in rats. Paradoxical emboli have been known to pass through a pulmonary arteriovenous malformation (PAVM), which is similar to IPAVA, resulting in ischaemic stroke in humans (26, 27). Thus, an increase in the Q_{IPAVA} may increase the risk of pulmonary gas exchange impairment and paradoxical embolism (8, 13, 14, 28, 29). Paradoxical embolism induced by IPAVA has been observed in several clinically important conditions in humans, such as stroke, transient ischaemic attack, migraine, and gas embolism during scuba diving (26, 29–31).

Several methods, such as microspheres, macroaggregated albumin (MAA), and agitated saline contrast using microbubbles (bubble study), have been used for the detection of IPAVA. These methods have several advantages and disadvantages. However, these methods use substances that are normally filtered or removed from the pulmonary capillaries to identify a right-to-left shunt. Microspheres are composed of polymer or glass. Consequently, these structures are not easily deformed and maintain a constant size (e.g., 25 and $50\ \mu\text{m}$). These properties have been used to determine the diameter of the vessel through which they pass. However, *in vivo* experiments are not feasible as biodegradable products are not yet commercially available. Consequently, microspheres are mainly used in *ex vivo* experiments, such as isolated lungs (1, 3, 13). Technetium-99 m labelled MAA (^{99m}Tc -MAA), an MAA tagged with radioactive material, has also been used to detect IPAVAs. Owing to its biodegradability, ^{99m}Tc -MAA is safe for use in humans. Moreover, it enables quantification of the shunt degree. However, ^{99m}Tc -MAA requires advanced equipment, such as a gamma camera, and the quantification of the shunt may be somewhat overestimated, as free ^{99m}Tc isolated from MAA passes through the pulmonary capillaries (1, 8, 28). Bubble study is performed by mixing normal saline with air to create microbubbles and injecting the mixed saline solution intravascularly. The presence of the bubbles is confirmed via echocardiography. This method does not quantify the extent of the shunt as well as microspheres and ^{99m}Tc -MAA. In addition, the resulting microbubbles are unevenly sized and fragile. However,

Abbreviations: IPAVAs, intrapulmonary arteriovenous anastomoses; BW, body weight; Q_{IPAVA} , blood flow through IPAVA; PH, pulmonary hypertension; RV, right ventricular; MAA, macroaggregated albumin; ^{99m}Tc -MAA, technetium-99 m labelled MAA; LPS, left parasternal sternal; RPS, right parasternal sternal; TR, tricuspid regurgitation; CW, continuous wave; BSA, body surface area; NS, normal saline; LV, left ventricle; ICCs, intraclass correlation coefficients; CIs, confidence intervals; VSD, ventricular septal defect; PS, pulmonary stenosis; ASD, atrial septal defect; VSD, ventricular septal defect; PFO, patent foramen ovale; PASP, pulmonary artery systolic pressure.

bubble study has enabled a qualitative assessment of Q_{IPAVA} and can be used to distinguish whether the shunt is intracardiac or extracardiac. Furthermore, it uses readily available materials, is safe for living organisms, and can be used on unanaesthetised animals. Thus, bubble study is an inexpensive, safe, and convenient method (1, 32, 33), and its use in research and clinical practise has increased in recent years (34–36).

IPAVAs have primarily been studied in humans (37), dogs, sheep, horses, and rats. However, most of these studies were *ex vivo* studies that used isolated lungs (10, 13, 15, 20). To the best of our knowledge, no previous study has focused on the prevalence of IPAVA in healthy animals at rest or whether hyperoxia reduces Q_{IPAVA} in animals, as it does in humans. Notably, IPAVAs have not been studied in raccoon dogs. Therefore, the present study aimed to determine the prevalence of IPAVA in healthy raccoon dogs at rest and evaluate the changes in IPAVA following supplementation with 100% oxygen.

2 Materials and methods

2.1 Animals

This prospective study included 19 wild Korean raccoon dogs (scientific name: *Nyctereutes procyonoides koreensis*) that were rescued or captured by the Jeonbuk Wildlife Centre between August 2022 and December 2023. The 19 raccoon dogs comprised 13 males and 6 females, with a mean (\pm standard deviation) weight of 3.5 ± 1.25 kg. All raccoon dogs underwent simple health examinations, such as physical examinations and blood tests. Thoracic radiography and echocardiography were performed subsequently to confirm the presence of cardiac disease. A bubble study was conducted while breathing room air [true inspired oxygen fraction (FiO_2): 0.21] and echocardiography was performed to confirm the presence and extent of IPAVA. The IPAVA-positive raccoon dogs were administered 100% oxygen via a mask at a rate of 5 L/min for 5 min. The FiO_2 was assumed to range from 0.5 to 0.7, as reported in previous similar studies, in accordance with the guidelines of the American Association of Respiratory Care (19, 38).

This study was approved by the Institutional Animal Care and Use Committee of the Jeonbuk National University (Approval No. JBNUNON2022-081-002).

2.2 Echocardiography and cardiac indicators

All ultrasound examinations were performed using Aplio i800 (Canon Medical Systems, Tokyo, Japan). A cardiac sector probe, PST-50BT (frequency, 3.0–8.2 MHz), was used to perform echocardiography. Raccoon dogs were placed in the left and right lateral recumbent positions and examined through the left (LPS) and right parasternal sternal (RPS) windows. Any abnormalities in morphology, movement, and blood flow through the heart and valves were evaluated to differentiate congenital cardiac diseases from acquired cardiac diseases.

The cardiac parameters were measured in each raccoon dog. The heart rate was determined using electrocardiography.

Tricuspid regurgitation (TR) velocity was measured using continuous wave (CW) Doppler to assess the pulmonary artery systolic pressure and PH. The stroke volume was calculated using the Teichholz formula with the systolic and diastolic diameters of the left ventricular lumen measured along the long or short axis of the RPS view (39). Cardiac output was calculated as the product of stroke volume and heart rate. Cardiac output was divided by the body surface area (BSA) and body weight (BW) to obtain the cardiac output index and cardiac output per BW, respectively. Other basic echocardiography indicators, such as the diameters of the left atrium, aorta, and main pulmonary artery; the thickness of the left ventricular wall; the e-peak and a-peak of the mitral valve flow; and the presence of regurgitation were also measured.

2.3 Bubble study (agitated saline contrast echocardiography)

The mixed saline solution, comprising 9 mL normal saline (NS), 0.5 mL room air, and 0.5 mL blood, was pushed back and forth between two 10 mL syringes connected perpendicularly to a three-way stopcock until the large visible air bubbles disappeared and the mixture became homogeneously turbid. Blood was added to lower the surface tension of the microbubbles and increase cohesion, thereby increasing the stability and longevity (34). Each raccoon dog was placed in the left lateral recumbent position and 2 mL of the agitated mixed saline solution was administered through a 23G intravascular catheter placed in the cephalic vein. Bubble contrast images of the right and left hearts were acquired immediately after the infusion using LPS apical four-chamber echocardiography. Imaging was continued until all intracardiac bubbles disappeared.

RadiAnt DICOM Viewer (Medixant, Poznan, Poland) was used to evaluate the contrast pattern in the acquired images. The emergence of the left heart bubble contrast within four cardiac cycles of the appearance of the first right heart bubble contrast was evaluated. An intracardiac shunt was considered to be present if the left heart bubble appeared within four cardiac cycles. In contrast, an extracardiac shunt, including IPAVA, was considered to be present if the left heart bubble appeared after four cardiac cycles. The criteria for categorising cardiac cycles for intracardiac and extracardiac shunts in the bubble study were based on those recently proposed in another human study (40). Bubble scores were assigned based on the maximum number of bubbles observed per frame within the left ventricle (LV). The bubble scores ranged from 0 to 5 points and this system has been used in many bubble studies (Figure 1): Bubble score (BS) 0, 0 microbubbles; BS1, 1–3 microbubbles; BS2, 4–12 microbubbles; BS3, >12 microbubbles; BS4, >12 microbubbles heterogeneously distributed throughout the left ventricle; and BS5, >12 microbubbles homogeneously distributed throughout the left ventricle (19, 41). Cases with BS0 and BS1–5 were classified as IPAVA negative and positive, respectively.

2.4 Statistical analysis

The Wilcoxon signed-rank test was used to evaluate significant differences in the BS and the maximum number of bubbles

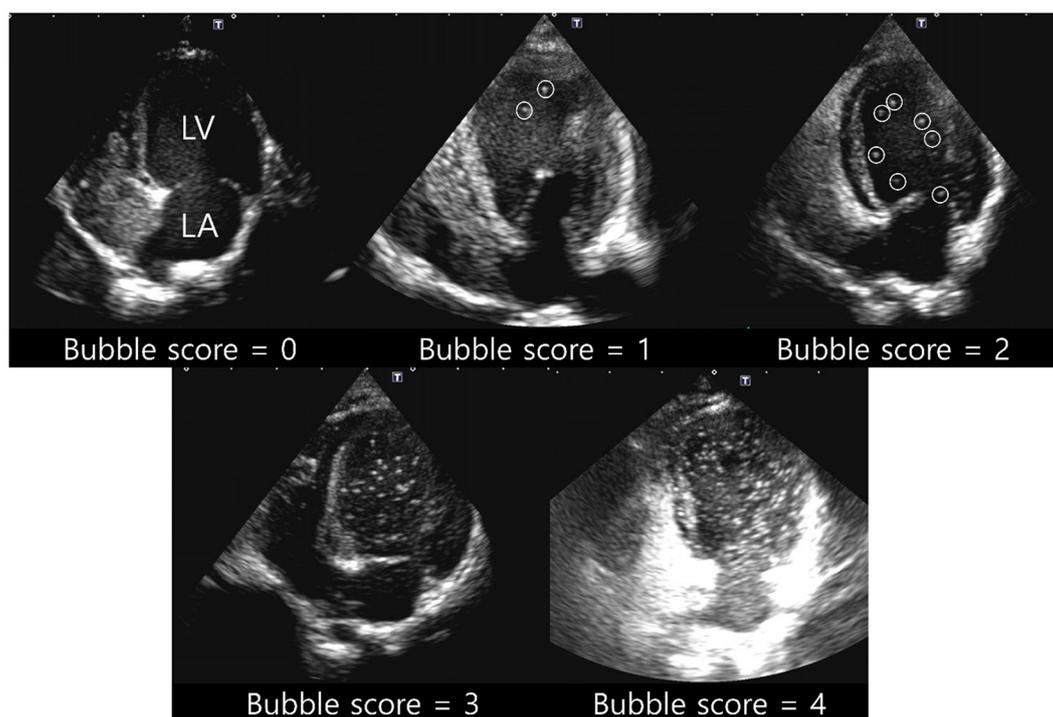


FIGURE 1

Representative pictures for bubble score (BS) of 0–4. BS0, 0 microbubbles; BS1, 1–3 microbubbles; BS2, 4–12 microbubbles; BS3, >12 microbubbles; BS4, >12 microbubbles heterogeneously distributed throughout the left ventricle; BS5, >12 microbubbles homogeneously distributed throughout the left ventricle. Circles of BS1 and BS2 indicate each bubble. BS5 was not observed in raccoon dogs. LA, left atrium; LV, left ventricle.

within the LV of IPAVA-positive raccoon dogs before and after supplementation with oxygen. Spearman's rank correlation test was used to evaluate the correlation between BS and the cardiac indicators. Raccoon dogs with cardiac disease were excluded owing to the presence of several variables that could be involved, including changes in haemodynamics. Only cardiac indicators from healthy raccoon dogs were used for the correlation analysis. Intraclass correlation coefficients (ICCs) with 95% confidence intervals (CIs) were calculated to determine the intra- and inter-observer reliability for the maximum number of bubbles in the left ventricle (excluding BS0). One author (Observer A) measured the items twice to determine the intra-observer reliability, whereas five observers, including the author (Observers A–E), measured the items to determine the inter-rater reliability. Statistical significance was set at p -values $<0.05^*$ or $<0.001^{**}$. All statistical tests were performed using IBM SPSS Statistics (version 20.0; IBM Corp., Armonk, NY, USA).

3 Results

3.1 Echocardiography

All 19 raccoon dogs underwent echocardiography. Sixteen healthy and three abnormal raccoon dogs with cardiac disease were identified (Figure 2). Mitral valve insufficiency was detected in two of the three abnormal raccoon dogs, whereas a ventricular septal defect (VSD) and pulmonary stenosis (PS) were detected in the third raccoon dog. All 16 healthy raccoon dogs had trivial to no TR.

3.2 IPAVA and bubble score

The appearance of left heart contrast after four cardiac cycles following the visualisation of the first right heart contrast was observed in 12 of the 16 healthy raccoon dogs (Figure 2). After excluding the four IPAVA-negative (BS0) raccoon dogs, one, three, seven, and one raccoon dog(s) had BS1, BS2, BS3, and BS4, respectively (Table 1). Thus, the prevalence of IPAVA in healthy raccoon dogs at rest was $\sim 75\%$ (12/16). The three abnormal raccoon dogs with cardiac disease were IPAVA-positive, with BS2, 3, and 4 (Table 1). BS5 was not observed in any of the bubble studies. The left heart contrast emerged after four cardiac cycles following the appearance of the first right heart contrast.

3.3 Oxygen supplementation

The BS and maximum number of bubbles in the LV decreased 5 min after supplementation with 100% oxygen in all 15 IPAVA-positive raccoon dogs (Table 1, Figure 3). A decrease in Q_{IPAVA} was observed after sufficient oxygenation in all 15 IPAVA-positive raccoon dogs (Figure 4). Before oxygenation, there were one, four, eight, and two cases with BS1, BS2, BS3, and BS4, respectively. After oxygenation, there were 11 and four cases with BS0 and BS2, respectively. Thus, a significant reduction in BS and maximum bubble count was observed after oxygenation ($p = 0.001$; Figure 4).

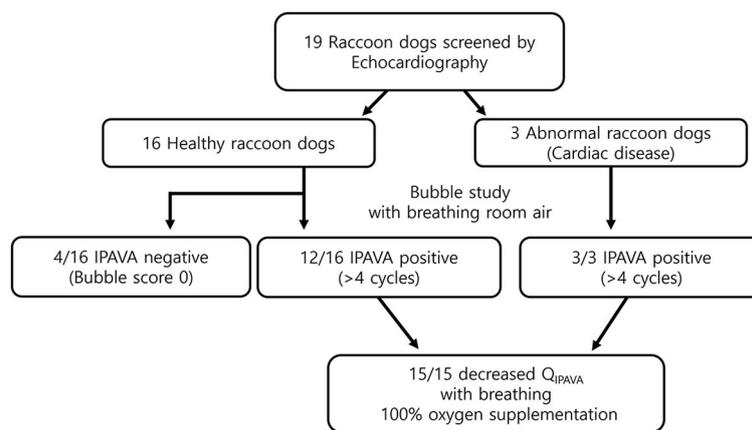


FIGURE 2 Flow chart of the study. Twelve healthy and three abnormal animals (i.e., with cardiac disease) were identified through echocardiography. The prevalence of IPAVA in healthy raccoon dogs was 75% (9/12). A decrease in the bubble score was observed in all IPAVA-positive animals (12/12) following supplementation with 100% oxygen for 5 min.

TABLE 1 Bubble scores and maximum numbers of bubbles in the IPAVA-positive raccoon dogs breathing room air and 100% oxygen.

Animal	Echocardiography	Bubble score (max. number of bubbles)	
		Room air	100% oxygen
1	Normal (healthy)	3 (25)	2 (10)
2		2 (6)	0 (0)
3		3 (25)	2 (7)
4		1 (3)	0 (0)
5		3 (20)	0 (0)
6		2 (5)	0 (0)
7		4 (55)	2 (8)
8		3 (25)	0 (0)
9		2 (8)	0 (0)
10		3 (14)	0 (0)
11		3 (13)	0 (0)
12		3 (14)	0 (0)
13	Cardiac disease	4 (67)	2 (6)
14		2 (7)	0 (0)
15		3 (21)	0 (0)

3.4 Correlation between the bubble score and the cardiac indicators

Table 2 presents the cardiac indicators of the 16 healthy raccoon dogs. No clear correlations were observed between BS and heart rate, stroke volume, cardiac output, or cardiac output index ($p > 0.05$); however, BS was correlated with cardiac output per BW. The p -value was 0.036 ($p < 0.05^*$), and Spearman’s rank correlation coefficient (ρ) was 0.526, indicating a moderate relationship.

3.5 Intra- and inter-observer reliability

Table 3 presents the intra- and inter-observer reliability for measuring the maximum number of bubbles observed in the LV during the bubble study. The intra-observer reliability, assessed based on the two measurements taken by Observer A (first author), was very high, with an ICC of 0.994 (95% CI, 0.986–0.998). The inter-observer reliability, assessed based on the measurements taken by the five observers (Observers A–E), was also very high, with an ICC of 0.995 (95% CI, 0.989–0.998).

4 Discussion

This study investigated the prevalence of IPAVA and changes in IPAVA after oxygenation supplementation in 19 wild raccoon dogs using a bubble study and echocardiography.

4.1 Bubble study: left heart bubble contrast indicates IPAVA after four cardiac cycles

The detection of IPAVA has become crucial owing to its clinical importance. A bubble study was used for the detection of IPAVA in the present study. Bubble studies are performed by creating microbubbles using agitated saline and are widely used for various purposes. The detection of right-to-left shunts of blood flow is its most notable purpose (25, 30, 34). Intravenously injected microbubbles are filtered or removed as they pass through the pulmonary capillaries. Consequently, they do not reach the left heart in normal cases (24). However, the presence of a right-to-left shunt pathway that bypasses the capillaries in the lungs, which serve as philtres, facilitates the passage of the microbubbles to the left side of the heart. This right-to-left shunt can be categorised as intracardiac or extracardiac. Extracardiac shunts are primarily observed in the lungs and include IPAVAs and

TABLE 2 Bubble scores, basic information, and cardiac indicators of healthy raccoon dogs breathing room air.

Animal	Sex	Bubble score (maximum number of bubbles)	Weight (kg)	Heart rate (beats/min)	Stroke volume (mL)	Cardiac output (L/min)	Cardiac output index (L/min/m ²)	Cardiac output per BW* (L/min/kg)
1	F	0 (0)	4.3	92	11.93	1.1	4.1	0.255
2	M	3 (25)	3.4	126	12.14	1.5	6.7	0.450
3	M	2 (6)	3.4	159	9.70	1.5	6.8	0.454
4	M	3 (25)	3.9	112	14.19	1.6	6.3	0.407
5	F	0 (0)	4.5	168	3.61	0.6	2.2	0.135
6	M	1 (3)	4.8	65	12.16	0.8	2.8	0.165
7	M	3 (20)	3.9	174	4.67	0.8	3.2	0.208
8	M	0 (0)	3.0	93	7.21	0.7	3.2	0.224
9	M	2 (5)	3.5	148	10.26	1.5	6.5	0.434
10	M	4 (55)	1.4	120	5.35	0.6	5.1	0.459
11	F	3 (25)	1.0	176	2.08	0.4	3.6	0.367
12	F	2 (8)	5.2	122	6.14	0.7	2.5	0.144
13	M	3 (14)	4.1	109	6.78	0.7	2.9	0.180
14	F	3 (13)	3.1	97	9.30	0.9	4.2	0.291
15	F	0 (0)	4.6	78	12.19	1.0	3.4	0.207
16	M	3 (14)	3.4	131	7.71	1.0	4.4	0.297
<i>p</i> -value		-	-	0.227	0.366	0.973	0.109	0.036*

The numbering of animals is not the same as that in Table 1.

Significant correlation with bubble score ($p < 0.05^$, $\rho = 0.526$).

TABLE 3 Intra-observer and inter-observer reliability for the maximum number of bubbles in the left ventricle using ICC and the 95% CI.

Parameter	Mean \pm SD (number of bubbles)	ICC	95% CI	<i>p</i> -value
Intra-observer (observer A)		0.994	0.986–0.998	$p < 0.001^{**}$
	First	17.89 \pm 17.61		
	Second	17.37 \pm 16.34		
Inter-observer		0.995	0.989–0.998	$p < 0.001^{**}$
	Observer A	17.63 \pm 16.93		
	Observer B	20.21 \pm 19.14		
	Observer C	18.26 \pm 16.09		
	Observer D	20.21 \pm 19.50		
	Observer E	19.53 \pm 17.15		

CI, confidence interval; ICC, intra-class correlation coefficient; SD, standard deviation.

Experimental values were considered significant at $p < 0.001^{**}$.

and other congenital septal defects, such as atrial septal defect (ASD) and ventricular septal defect (VSD). ASD and VSD can cause right-to-left shunts by increasing right heart pressure (19). Intra- and extracardiac right-to-left shunts can be differentiated using bubble studies. An intracardiac shunt is considered to be present if the left heart contrast is visualised within four cardiac cycles after the first visualisation of the right heart contrast. An extracardiac shunt is considered to be present if the left heart contrast is visualised after four cardiac cycles (40). This may be attributed to the fact that the microbubbles that appear in

the right heart reach the left heart relatively late if they travel through the lungs. VSD was detected in one raccoon dog via echocardiography just prior to the bubble study in the present study; however, the presence of an intracardiac shunt was unlikely as the left heart contrast did not appear within four cardiac cycles. Thus, bubble study can be used to distinguish extracardiac right-to-left shunts, including IPAVA, from intracardiac right-to-left shunts by evaluating how many cardiac cycles later the left heart bubble contrast is visualised after the first visualisation of the right heart contrast.

In addition to IPAVA, extracardiac shunts also include PAVMs, which are similar to IPAVA. PAVMs can be confused with IPAVA as the left heart contrast is visualised after four cardiac cycles after the first visualisation of the right heart contrast in the bubble study in patients with PAVM as well (4, 27, 40). However, unlike IPAVA, which may be a remnant of a foetal or neonatal structure, PAVM can appear in conjunction with genetic disorders, such as hereditary haemorrhagic telangiectasia, or be acquired due to trauma, infection, or iatrogenic causes (4, 20, 22). An accurate diagnosis of PAVM can be obtained using computed tomography angiography, especially in cases where in a large volume of shunting is observed, as the diameter of PAVM in humans is 1–5 cm, which is much higher than that of IPAVA (4, 27). However, unlike IPAVA, which exhibits dynamic opening and closing, PAVM is static and is assumed to remain open, thereby facilitating a constant flow of blood. Thus, the bubble score is not expected to change even under special conditions, such as oxygen supplementation, in cases with PAVM (1). A reduction in BS was observed in all IPAVA-positive subjects after oxygen supplementation in the present study, which was indicative of IPAVA rather than PAVM. Therefore, changes in the BS through bubble studies before and after oxygen supplementation can be used to differentiate IPAVA from PAVM.

In addition to a right-to-left shunt that bypasses the pulmonary capillaries, the passage of the microbubbles through the dilated pulmonary capillaries can also lead to the visualisation of LV bubble contrast. However, this seems unlikely, given the vessel diameters and bubble sizes reported in previous studies. Microbubbles in agitated saline are <30 μm in size; however, the sizes are irregular (42). In contrast, the diameter of pulmonary capillaries ranges from 7 to 10 μm in size in dogs and does not exceed 13–15 μm even under non-physiological pressure (43, 44). Therefore, the passage of microbubbles through the dilated pulmonary capillaries seems to be impossible. The diameter of IPAVA was found to range from 25 to 50 μm in an experiment using microspheres. Microbubbles can pass through this site easily (3, 13). These findings suggest that the microbubbles created in the bubble study cannot pass through normal pulmonary capillaries and that IPAVA is likely to be the cause if microbubbles appear dynamically in the left ventricle after four cardiac cycles following the visualisation of the right heart contrast.

The left heart contrast observed in raccoon dogs in the bubble study was considered to be indicative of IPAVA and not of an intracardiac shunt, PAVM, or dilated pulmonary vessels, as the left heart contrast appeared within four cardiac cycles after the first visualisation of the right heart contrast and showed dynamic changes depending on supplementation with oxygen. This bubble study method can be easily and conveniently used in the future to detect IPAVA in various species and predict the risks associated with right to left shunt.

4.2 IPAVA prevalence in healthy raccoon dogs at rest

The prevalence of IPAVA was ~75% (12/16) among the healthy raccoon dogs at rest in the present study, which is much higher than the previously reported prevalence of 28% in humans (19). Although studies related to IPAVA have been conducted in other

animal species, the prevalence of IPAVA in the resting state remains underexplored (10, 13, 15). Studies on horses and dogs have addressed the changes in IPAVA before and after exercise, and no changes in Q_{IPAVA} were observed in any of the subject before exercise (13, 15). Changes in Q_{IPAVA} were observed during normoxia and hypoxia in a previous study involving rats, and Q_{IPAVA} was not observed during normoxia in any of the rats (10). This suggests that the prevalence of IPAVA in healthy raccoon dogs at rest may be higher than that in other animal species. However, bubble studies were not used in the studies involving dogs and rats, and the sample size was <10 for all three species; therefore, direct comparison was difficult (10, 13, 15).

Several factors, such as catecholamine secretion, the lateral recumbent position during testing, and age, may have contributed to the increased prevalence of IPAVA in healthy raccoon dogs at rest observed in the present study. The raccoon dogs included in the present study were wild animals. Moreover, they were not sedated or anaesthetised. Thus, the possibility of elevated catecholamine levels in the body cannot be ruled out even in a resting (apparently stable) state without exercise. Intravenous administration of catecholamines, such as epinephrine, dopamine, and dobutamine, has been shown to increase Q_{IPAVA} by elevating the pulmonary artery systolic pressure (PASP) and/or cardiac output (16, 17). However, the catecholamine levels were not measured in the raccoon dogs in the present study; therefore, these causes cannot be substantiated. The lateral recumbent position assumed during bubble studies and echocardiograms may have also affected the IPAVA. Left heart contrast enhancement observed in human in the supine position disappears in the upright position, which may be attributed to altered pulmonary blood flow distribution due to a change in gravitational action on pulmonary blood flow (14, 19). Age is another factor that may have affected the IPAVA. The prevalence of IPAVA was found to be higher in foetal and neonatal lambs than that in adult lambs in a previous study (20). Similarly, the prevalence of IPAVA was found to be higher in younger individuals than in older individuals in human studies (12). The raccoon dogs included in this study were wild animal whose ages could not be estimated accurately; however, there were some young raccoon dogs weighing <1 kg. If a majority of the rescued raccoon dogs were young, this could explain the high prevalence of IPAVA. Further studies on the prevalence of IPAVA in raccoon dogs according to age, such as comparisons of immature and mature dogs, must be conducted in the future.

Other technical influences may also have had some effect on the bubble study. Unlike previous bubble studies, a small amount of blood from each raccoon dog was mixed in the solution during the preparation of the agitated saline contrast in the present study. This may have increased the stability and longevity of the microbubbles and facilitated their delivery to the left heart (34). Furthermore, differences in the ultrasound machine equipment and the manufacturer, product, and setting values of the probes used may have resulted in better detection of the microbubbles in the heart. Therefore, further studies must be conducted to compare the results of the bubble study of various microbubble manufacturing protocols and ultrasound machines.

Lastly, the high prevalence of IPAVA in raccoon dogs observed in the present study may be attributed to the species-specific nature of raccoon dogs. The prevalence of IPAVA may be higher among raccoon dogs compared with other species of animals or these

structures may open more easily in the resting state. Thus, the prevalence of IPAVA may vary between species. Identifying the causative factors would provide insights into IPAVA.

4.3 IPAVA in animals

IPAVAs have been studied extensively in humans. These structures are also observed in animals, and studies have been conducted in various species, such as dogs, horses, and rats (1, 10, 13, 15, 37). However, the experimental methods used in the present study involving raccoon dogs differs from those of previous animal studies. First, *in vivo* and *ex vivo* experiments (living dogs and isolated lungs) were conducted in previous studies involving dogs, and the presence or absence of microspheres in blood or tissue was confirmed before and after exercise through commercial microsphere injection rather than through the use of bubble study. Moreover, the primary objective of a previous canine study was to determine the prevalence of exercise-induced IPAVA (13). Changes in IPAVA after exercise have also been observed in Thoroughbred horses. The present study was similar to this previous study; in that, it used echocardiography to confirm right-to-left shunting. However, the mixed solution injected into the vein was succinylated gelatin in the previous study instead of normal saline mixed with room air (15). Echocardiography was performed in a standing position during the bubble study in a previous study on horses. In contrast, the raccoon dogs were placed in a lateral recumbent position in the present study. The IPAVA study involving rats used microspheres from living subjects and isolated lungs, similar to previous studies involving dogs; however, this study attempted to confirm the induction of IPAVA via hypoxia rather than exercise (10). These previous studies have suggested the requirement for continued research on IPAVA in humans and animal species. However, most studies have focused on IPAVA induced by abnormal conditions, such as exercise and hypoxia, and have not focused on IPAVA under normal resting conditions. Furthermore, whether supplemental oxygen lowers Q_{IPAVA} to the same extent in animals as that observed in humans has not been studied. IPAVA continues to be of clinical importance, and human studies have been conducted on the prevalence of IPAVA at rest rather than in specific conditions, such as hypoxia and exercise (19). Differences in various situations and detection methods for vascular injection substances related to IPAVA have been reported. It may be beneficial to compare IPAVA between animal species if a standardised method for IPAVA is continuously applied to various animal species in the future.

4.4 Oxygenation and the reduction in

Q_{IPAVA}

Q_{IPAVA} is altered by the concentration of oxygen inhaled. Q_{IPAVA} may increase under hypoxic conditions and decrease under hyperoxic condition (7–12, 45). However, the modulation of IPAVA in animals by high oxygen supplementation remains underexplored. A reduction in BS and the number of bubbles in the LV was observed in all 15 IPAVA-positive raccoon dogs in the present study following supplementation with 100% oxygen via a

mask at a rate of 5 L/min for 5 min. Similar findings have been reported in previous human studies (7, 11). Supplementation with 100% oxygen for 5 min was considered sufficient to change Q_{IPAVA} in the present study (11, 18). The statistical results also indicated that oxygenation was effective in reducing the BS and maximum number of bubbles in the LV.

It is unclear how hyperoxia achieved through oxygen supplementation reduces Q_{IPAVA} . It has been speculated that IPAVA behaviour is opposite to that of the pulmonary vessels and is more similar to that of the systemic vessels. Thus, it exhibits vasoconstriction during hyperoxia and vasodilation during hypoxia. This may be attributed to IPAVAs being a part of normal foetal circulation, similar to ductus arteriosus (11, 37). Identification of a clear cause for the decrease in Q_{IPAVA} in response to hyperoxia was not possible in the present study. However, as this phenomenon was also observed in animals other than humans, further studies focusing on animal IPAVA, such as comparisons of physiological changes before and after supplemental oxygen, may aid in understanding the phenomenon in humans.

The findings of the present study suggest that Q_{IPAVA} in raccoon dogs was effectively reduced after oxygen supplementation, similar to that observed in humans. Thus, adequate oxygenation effectively reduces blood flow bypassing the pulmonary capillaries. Moreover, it can also prevent pulmonary gas exchange impairment and paradoxical embolism, which are potential clinical manifestations of IPAVA (2, 8, 13, 26, 29, 31, 46). Therefore, the importance of oxygen supplementation can be emphasised in patients whose IPAVA is confirmed through bubble study.

4.5 Correlations of IPAVA with cardiac indicators

A higher BS indicates an increase in Q_{IPAVA} . The present study confirmed the relationship between BS and several cardiac indicators.

The first is the relationship between IPAVA and pulmonary artery systolic pressure (PASP). TR, which is proportional to PASP, was low or not measured using CW Doppler in most raccoon dogs with IPAVA in the present study. Q_{IPAVA} increases proportionally with PASP through exercise or drug infusion (12, 16). Furthermore, the presence of higher Q_{IPAVA} has been associated with a lower prevalence of PH. Thus, it can be inferred that Q_{IPAVA} increases in response to acutely elevated PASP and relieves PASP to some extent, suggesting that the presence of IPAVA may ameliorate or prevent PH (12, 22). This finding is consistent with the low likelihood of current PH observed in all IPAVA-positive raccoon dogs in the present study. However, determining whether the history of previous PH or the elevated PASP in the raccoon dogs was transient due to excitement during the bubble study is difficult. Furthermore, clear TR was not observed in the IPAVA-negative raccoon dogs; however, the number was small. Therefore, the relationship between IPAVA and PH remains ambiguous and requires further examination.

Q_{IPAVA} may be related to cardiac output rather than to PASP. Increased Q_{IPAVA} has been associated with increased cardiac output with or without increased PASP following the administration of

epinephrine in combination with atropine (18). Beta-receptor-expected mediate dilation of IPAVA, which indicates a direct effect on the smooth muscles in the IPAVA, was reported to be an unlikely phenomenon in a previous study. Thus, IPAVA is passively opened by increased cardiac output (16). The relationship between BS and cardiac indicators in healthy raccoon dogs was examined using echocardiography in the present study. The cardiac output per BW was found to be significantly correlated with BS, but not with cardiac output, heart rate, stroke volume, or cardiac output index (cardiac output per BSA). This may be attributed to the BWs of raccoon dogs varying widely in the present study, with the BWs of some raccoon dogs being more than 10-fold higher (BW, 0.5–5.2 kg). Hence, standardisation based on body type rather than the absolute amount of cardiac output would be more beneficial. The sample size of the present study was small. Future studies must consider cardiac output per BW as well as the cardiac output index when analysing correlations with cardiac output.

Several previous studies have investigated the direct factors affecting IPAVA; however, uncertainties still remain (16, 18, 45). The number of raccoon dogs included in this study was small, which limited the ability to clarify the correlations with cardiac indicators. Large-scale studies conducted in the future may yield different results. Therefore, further studies involving various animal species, including raccoon dogs, must be conducted in the future to investigate the correlation between IPAVA and cardiac indicators, such as PASP and cardiac output.

4.6 IPAVA based on the maximum number of bubbles in the LV

A BS grading (0–5) system capable of counting the number of bubbles in the LV is often used to determine the severity of IPAVA (19, 41). Although this scoring system is qualitative, it is correlated with Q_{IPAVA} and other indicators. Moreover, its efficacy has been verified (47). Nevertheless, the BS0–5 scoring system has not been formally established, and some studies have used different scoring criteria (4, 23, 40). The difference between consecutive grades of the BS0–5 system is not uniform, i.e., the difference between BS1 and BS2 was not equivalent to that between BS2 and BS3. The intra- and inter-observer reliability determined by counting the maximum number of bubbles in the LV was high in the present study, which is another criterion used to grade the severity of Q_{IPAVA} . In contrast to the scoring system, this method has facilitated the quantification of Q_{IPAVA} as it involves counting the number of bubbles individually. In addition, the method of counting bubbles was found to be significantly effective in monitoring the changes in Q_{IPAVA} after the administration of oxygen. Only a few cases had a large number of bubbles that may have led to adequately high deviations in counting (only two cases with >50 bubbles); thus, this consistent measurement method is promising for the quantitative assessment of Q_{IPAVA} using ultrasonography alone.

4.7 Limitations

This study has certain limitations. First, the sample size of the present study, including the raccoon dogs with cardiac disease,

was 19. Although further large-scale studies must be conducted, the sufficiently low p -values (≤ 0.001) of “change in BS before and after oxygenation” and “ICC of bubble count” appear to be meaningful enough to overcome the small number of subjects. Furthermore, the detailed medical history and age of the raccoon dogs could not be determined as these were wild animals. Age can affect the prevalence of IPAVA (12, 20). Moreover, additional upright or standing tests or the measurement of the catecholamine concentrations in relation to posture and excitement/strain could not be performed in the present study. This may have influenced the prevalence of IPAVA (16, 19). Following the result of the reduced Q_{IPAVA} of the raccoon dogs in this study, further study of raccoon dogs required for increasing IPAVA such as hypoxia, exercise, and catecholamine injection identified in previous studies of human or other animal species (9, 12, 13, 15, 16). A direct measurement of PASP, as opposed to an indirect measurement using TR, would have yielded more accurate results regarding its correlation with IPAVA. Similarly, arterial blood gas analysis would have provided further insights into the factors affecting IPAVA. The findings of the present study may differ from those of other bubble studies as the BS may differ depending on the microbubble creation protocol (34, 42). Although bubble studies were conducted to determine the presence of assess Q_{IPAVA} and perform a qualitative analysis, microspheres or ^{99m}Tc -MAA would have been suitable for the quantitative assessment and determination of the extent of shunting (1, 8, 13). However, microspheres are not yet applicable to live animals and ^{99m}Tc -MAA requires additional advanced equipment (1). Thus, bubble studies were conducted owing to the low cost, convenience, and safety.

5 Conclusion

The prevalence of IPAVA (75%) was observed in healthy raccoon dogs at rest, suggesting that the prevalence of IPAVA may vary across animal species. Adequate oxygen supplementation was effective in reducing the Q_{IPAVA} , which may help prevent potential negative factors such as pulmonary gas exchange impairments and paradoxical embolism that can occur with IPAVA.

Data availability statement

The original contributions presented in the study are included in the article/supplementary material, further inquiries can be directed to the corresponding author.

Ethics statement

The animal study was approved by Institutional Animal Care and Use Committee of the Jeonbuk National University. The study was conducted in accordance with the local legislation and institutional requirements.

Author contributions

CL: Conceptualisation, Data curation, Formal analysis, Investigation, Methodology, Project administration, Resources,

Software, Validation, Visualisation, Writing – original draught, Writing – review & editing. MK: Conceptualisation, Data curation, Resources, Writing – original draught, Formal analysis, Writing – review & editing. J-IH: Conceptualisation, Data curation, Resources, Supervision, Writing – original draught, Formal analysis, Writing – review & editing. KL: Conceptualisation, Data curation, Formal analysis, Investigation, Project administration, Supervision, Validation, Visualisation, Writing – original draught, Writing – review & editing. HY: Conceptualisation, Data curation, Formal analysis, Investigation, Methodology, Project administration, Resources, Software, Supervision, Validation, Visualisation, Writing – original draught, Writing – review & editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This study was supported by the National Institute of Wildlife Disease Control

and Prevention as a Specialised Graduate School Support Project for Wildlife Disease Specialists.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

- Duke JW, Elliott JE, Lovering AT. Clinical consideration for techniques to detect and quantify blood flow through intrapulmonary arteriovenous anastomoses: lessons from physiological studies. *Echocardiography*. (2015) 32:5195–204. doi: 10.1111/echo.12839
- Lovering AT, Eldridge MW, Stickland MK. Counterpoint: exercise-induced intrapulmonary shunting is real. *J Appl Physiol*. (2009) 107:994–7. doi: 10.1152/jappphysiol.91489.2008a
- Lovering AT, Stickland MK, Kelso AJ, Eldridge MW. Direct demonstration of 25- and 50- μ m arteriovenous pathways in healthy human and baboon lungs. *Am J Physiol Heart Circ Physiol*. (2007) 292:H1777–81. doi: 10.1152/ajpheart.01024.2006
- Chen A, Zhu J, Zhu L, Tang Y, Li Y, Zhang Q, et al. Neglected intrapulmonary arteriovenous anastomoses: a comparative study of pulmonary right-to-left shunts in patients with patent foramen ovale. *Front Cardiovasc Med*. (2023) 10:111818. doi: 10.3389/fcvm.2023.1111818
- Sapcey PC. *Traité danatomie descriptive*. Tours: Delahaye (1874).
- Hopkins SR, Olfert IM, Wagner PD. Point: Counterpoint: exercise-induced intrapulmonary shunting is imaginary vs. real. *J Appl Physiol*. (2009) 107:993–4. doi: 10.1152/jappphysiol.91489.2008
- Elliott JE, Choi Y, Laurie SS, Yang X, Gladstone IM, Lovering AT. Effect of initial gas bubble composition on detection of inducible intrapulmonary arteriovenous shunt during exercise in normoxia, hypoxia, or hyperoxia. *J Appl Physiol*. (2011) 110:35–45. doi: 10.1152/jappphysiol.00145.2010
- Bates ML, Farrell ET, Drezdon A, Jacobson JE, Perlman SB, Eldridge MW. Hypoxia and exercise increase the transpulmonary passage of 99mTc-labeled albumin particles in humans. *PLoS ONE*. (2014) 9:e101146. doi: 10.1371/journal.pone.0101146
- Laurie SS, Yang X, Elliott JE, Beasley KM, Lovering AT. Hypoxia-induced intrapulmonary arteriovenous shunting at rest in healthy humans. *J Appl Physiol*. (2010) 109:1072–9. doi: 10.1152/jappphysiol.00150.2010
- Bates ML, Fulmer BR, Farrell ET, Drezdon A, Pegelow DF, Conhaim RL, et al. Hypoxia recruits intrapulmonary arteriovenous pathways in intact rats but not isolated rat lungs. *J Appl Physiol*. (2012) 112:1915–20. doi: 10.1152/jappphysiol.00985.2011
- Lovering AT, Stickland MK, Amann M, Murphy JC, O'Brien MJ, Hokanson JS, et al. Hyperoxia prevents exercise-induced intrapulmonary arteriovenous shunt in healthy humans. *J Physiol*. (2008) 586:4559–65. doi: 10.1113/jphysiol.2008.159350
- Cameron Norris H, Mangum TS, Duke JW, Straley TB, Hawn JA, Goodman RD, et al. Exercise and hypoxia-induced blood flow through intrapulmonary arteriovenous anastomoses is reduced in older adults. *J Appl Physiol*. (2014) 116:1324–33. doi: 10.1152/jappphysiol.01125.2013
- Stickland MK, Lovering AT, Eldridge MW. Exercise-induced arteriovenous intrapulmonary shunting in dogs. *Am J Respir Crit Care Med*. (2007) 176:300–5. doi: 10.1164/rccm.200702-206OC
- Stickland MK, Welsh RC, Haykowsky MJ, Petersen SR, Anderson WD, Taylor DA, et al. Intra-pulmonary shunt and pulmonary gas exchange during exercise in humans. *J Physiol*. (2004) 561:321–9. doi: 10.1113/jphysiol.2004.069302
- La Gerche A, Daffy JR, Mooney DJ, Forbes G, Davie AJ. Transit of micro-bubbles through the pulmonary circulation of thoroughbred horses during exercise. *Res Vet Sci*. (2013) 95:644–7. doi: 10.1016/j.rvsc.2013.04.002
- Laurie SS, Elliott JE, Goodman RD, Lovering AT. Catecholamine-induced opening of intrapulmonary arteriovenous anastomoses in healthy humans at rest. *J Appl Physiol*. (2012) 113:1213–22. doi: 10.1152/jappphysiol.00565.2012
- Bryan TL, Van Diepen S, Bhutani M, Shanks M, Welsh RC, Stickland MK. The effects of dobutamine and dopamine on intrapulmonary shunt and gas exchange in healthy humans. *J Appl Physiol*. (2012) 113:541–8. doi: 10.1152/jappphysiol.00404.2012
- Elliott JE, Duke JW, Hawn JA, Halliwill JR, Lovering AT. Increased cardiac output, not pulmonary artery systolic pressure, increases intrapulmonary shunt in healthy humans breathing room air and 40% O₂. *J Physiol*. (2014) 592:4537–53. doi: 10.1113/jphysiol.2014.274829
- Elliott JE, Nigam SM, Laurie SS, Beasley KM, Goodman RD, Hawn JA, et al. Prevalence of left heart contrast in healthy, young, asymptomatic humans at rest breathing room air. *Resp Physiol Neurobi*. (2013) 188:71–8. doi: 10.1016/j.resp.2013.04.019
- McMullan DM, Hanley FL, Cohen GA, Portman MA, Riemer RK. Pulmonary arteriovenous shunting in the normal fetal lung. *J Am Coll Cardiol*. (2004) 44:1497–500. doi: 10.1016/j.jacc.2004.06.064
- Lovering AT, Riemer RK, Thébaud B. Intrapulmonary arteriovenous anastomoses. Physiological, pathophysiological, or both? *Ann Am Thorac Soc*. (2013) 10:504–8. doi: 10.1513/AnnalsATS.201308-265ED
- Matos JN, Malbon A, Dennler M, Glaus T. Intrapulmonary arteriovenous anastomoses in dogs with severe *Angiostrongylus vasorum* infection: clinical, radiographic, and echocardiographic evaluation. *J Vet Cardiol*. (2016) 18:110–24. doi: 10.1016/j.jvc.2015.10.008
- La Gerche A, Macisaac AI, Burns AT, Mooney DJ, Inder WJ, Voigt JU, et al. Pulmonary transit of agitated contrast is associated with enhanced pulmonary vascular reserve and right ventricular function during exercise. *J Appl Physiol*. (2010) 109:1307–17. doi: 10.1152/jappphysiol.00457.2010
- Butler B, Hills B. The lung as a filter for microbubbles. *J Appl Physiol*. (1979) 47:537–43. doi: 10.1152/jappphysiol.1979.47.3.537
- Lee H, Kim H, Han H, Lee M, Lee S, Yoo H, et al. Microbubbles used for contrast enhanced ultrasound and theragnosis: a review of principles to applications. *Biomed Eng Lett*. (2017) 7:59–69. doi: 10.1007/s13534-017-0016-5
- Cappa R, Du J, Carrera JF, Berthaud JV, Southerland AM. Ischemic stroke secondary to paradoxical embolism through a pulmonary arteriovenous malformation:

- case report and review of the literature. *J Stroke Cerebrovasc Dis.* (2018) 27:e125–e7. doi: 10.1016/j.jstrokecerebrovasdis.2018.02.015
27. Shovlin CL. Pulmonary arteriovenous malformations. *Am J Respir Crit Care Med.* (2014) 190:1217–28. doi: 10.1164/rccm.201407-1254CI
28. Lovering AT, Haverkamp HC, Romer LM, Hokanson JS, Eldridge MW. Transpulmonary passage of ^{99m}Tc macroaggregated albumin in healthy humans at rest and during maximal exercise. *J Appl Physiol.* (2009) 106:1986–92. doi: 10.1152/jappphysiol.01357.2007
29. Abushora MY, Bhatia N, Alnabki Z, Shenoy M, Alshaher M, Stoddard MF. Intrapulmonary shunt is a potentially unrecognized cause of ischemic stroke and transient ischemic attack. *J Am Soc Echocardiogr.* (2013) 26:683–90. doi: 10.1016/j.echo.2013.04.004
30. Woods TD, Harmann L, Purath T, Ramamurthy S, Subramanian S, Jackson S, et al. Small- and moderate-size right-to-left shunts identified by saline contrast echocardiography are normal and unrelated to migraine headache. *Chest.* (2010) 138:264–9. doi: 10.1378/chest.09-2797
31. Madden D, Lozo M, Dujic Z, Ljubkovic M. Exercise after SCUBA diving increases the incidence of arterial gas embolism. *J Appl Physiol.* (2013) 115:716–22. doi: 10.1152/jappphysiol.00029.2013
32. Droste DW, Kriete JU, Stypmann J, Castrucci M, Wichter T, Tietje R, et al. Contrast transcranial Doppler ultrasound in the detection of right-to-left shunts: comparison of different procedures and different contrast agents. *Stroke.* (1999) 30:1827–32. doi: 10.1161/01.STR.30.9.1827
33. Muskula PR, Main ML. Safety with echocardiographic contrast agents. *Circ Cardiovasc Imaging.* (2017) 10:e005459. doi: 10.1161/CIRCIMAGING.116.005459
34. Arndt JW, Oyama MA. Agitated saline contrast echocardiography to diagnose a congenital heart defect in a dog. *J Vet Cardiol.* (2008) 10:129–32. doi: 10.1016/j.jvc.2008.03.004
35. Kim Y, Kwon D, Kim SS, Lee K, Yoon H. Echocardiographic changes in the progress of reverse shunt and improvement to left-to-right shunt after medical treatment in dogs with bidirectional patent ductus arteriosus or ventricular septal defect: a report of two cases. *Vet Med Sci.* (2023) 9:1044–52. doi: 10.1002/vms3.1081
36. Ann S, Kim M, Yoon W, Suh S, Hyun C. Right to left atrial septum defect compounded with patent ductus arteriosus in a Maltese dog. *J Vet Clin.* (2017) 34:90–3. doi: 10.17555/jvc.2017.04.34.2.90
37. Lovering AT, Duke JW, Elliott JE. Intrapulmonary arteriovenous anastomoses in humans—response to exercise and the environment. *J Physiol.* (2015) 593:507–20. doi: 10.1113/jphysiol.2014.275495
38. Kallstrom TJ, Care AAFR. AARC Clinical Practice Guideline: oxygen therapy for adults in the acute care facility—2002 revision & update. *Respi Care.* (2002) 47:717–20.
39. Arora G, Morss AM, Piazza G, Ryan JW, Dinwoodey DL, Rofsky NM, et al. Differences in left ventricular ejection fraction using teichholz formula and volumetric methods by CMR: implications for patient stratification and selection of therapy. *J Cardiovasc Magnet Reson.* (2010) 12:1–2. doi: 10.1186/1532-429X-12-S1-P202
40. Bhatia N, Abushora MY, Donneyong MM, Stoddard MF. Determination of the optimum number of cardiac cycles to differentiate intra-pulmonary shunt and patent foramen ovale by saline contrast two- and three-dimensional echocardiography. *Echocardiography.* (2014) 31:293–301. doi: 10.1111/echo.12360
41. Lovering AT, Elliott JE, Beasley KM, Laurie SS. Pulmonary pathways and mechanisms regulating transpulmonary shunting into the general circulation: an update. *Injury.* (2010) 41:S16–23. doi: 10.1016/S0020-1383(10)70004-8
42. Tim M, Alerhand S, André D, Scott J. Point-of-care echocardiography for the evaluation of right-to-left cardiopulmonary shunts: a narrative review. *Can J Anesth.* (2020) 67:1824–38. doi: 10.1007/s12630-020-01813-2
43. Glazier J, Hughes J, Maloney J, West J. Measurements of capillary dimensions and blood volume in rapidly frozen lungs. *J Appl Physiol.* (1969) 26:65–76. doi: 10.1152/jappphysiol.1969.26.1.65
44. Rosenzweig DY, Hughes J, Glazier J. Effects of transpulmonary and vascular pressures on pulmonary blood volume in isolated lung. *J Appl Physiol.* (1970) 28:553–60. doi: 10.1152/jappphysiol.1970.28.5.553
45. Tremblay JC, Lovering AT, Ainslie PN, Stembridge M, Burgess KR, Bakker A, et al. Hypoxia, not pulmonary vascular pressure, induces blood flow through intrapulmonary arteriovenous anastomoses. *J Physiol.* (2015) 593:723–37. doi: 10.1113/jphysiol.2014.282962
46. Rapp JH, Pan XM, Yu B, Swanson RA, Higashida RT, Simpson P, et al. Cerebral ischemia and infarction from atheroemboli <100 μm in size. *Stroke.* (2003) 34:1976–80. doi: 10.1161/01.STR.0000083400.80296.38
47. Fischer CH, Campos O, Fernandes WB, Kondo M, Souza FL, De Andrade JL, et al. Role of contrast-enhanced transesophageal echocardiography for detection of and scoring intrapulmonary vascular dilatation. *Echocardiography.* (2010) 27:1233–7. doi: 10.1111/j.1540-8175.2010.01228.x