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Frontiers in broiler chicken welfare: adopting early detection of intestinal integrity loss in broiler welfare assessment protocols

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Current broiler welfare assessment protocols include indicators of impaired intestinal health, but these are non-specific. Loss of intestinal integrity is considered to be a major welfare consequence for broilers but the absence of specific, early indicators in welfare assessment protocols hampers early detection and treatment. Coccidiosis is one of the major threats to intestinal integrity in broiler chickens and taken as an example. We propose the development of specific biomarkers for detecting early onset of intestinal health deterioration. The genotype and external phenotype of organisms are linked by so-called internal phenotypes which are influenced by environmental conditions. We review the impact of coccidiosis on external and internal phenotypes in chickens. The change from the "homeostatic immune response" toward an inflammatory response to control infections is reflected in the change in feeling of comfort to the feeling of discomfort in broilers, in which the gut-brain axis likely plays a crucial role. With this change, a negative emotional state develops. Two routes of developing biomarkers are proposed that are interconnected. The first route is by enabling ~omics techniques for predominantly invasive biomarkers related to the internal phenotype of the broiler chickens during infection. The second approach involves using sensors and automated systems to monitor behavior, vocalizations, and fecal appearance for early disease detection at flock level. By linking these external indicators to invasive biomarkers, we can develop disease-specific biomarkers that enhance early diagnosis with precision and could add significant value to welfare assessment protocols. Research in this area should be encouraged.

KEYWORDS

broiler, sickness behavior, coccidiosis, emotional state, gut-brain axis, intestinal integrity, welfare

1 Introduction

Intestinal health in broiler chickens is both critical for absorption of nutrients and a prerequisite for broiler welfare. Antibiotic treatments have long been applied to prevent or treat intestinal infections, but concerns around antibiotic resistance and the implications for human and animal health resulted in moves toward antibiotic-free broiler production (1). However, as a consequence, intestinal health issues became more apparent (2), impairing broiler welfare and performance.

Coccidiosis is the leading cause of loss of intestinal integrity in broiler chickens, surpassing other contributing factors such as bacteria, viruses, other parasites, and nutritional imbalances (3). Therefore, this perspective paper will focus on coccidiosis as a primary example of intestinal health challenges in broilers. Coccidiosis, caused by the pathogenic Eimeria parasite, starts when chickens ingest sporulated oocysts present in their environment. Replication of the parasite takes place within the intestinal epithelial cells and parasite numbers increase rapidly within a few days resulting in shedding a large number of oocysts. The host will develop an adaptive immune response to the challenge, but until this response is fully developed there will be a non-specific innate immune response including excessive secretion of mucin. The innate immune response has high energy costs for the host and during infection the intestinal microbiome is disrupted, favoring replication of pathogenic bacteria and predisposing to secondary infections (3) (Figure 1). The extent of damage to the intestinal tract depends on the Eimeria species, the initial challenge dose, the cells that it replicates within and the coccidiosis control strategy. One of the several commercially available control strategies involve using live attenuated Eimeria based vaccines. For these vaccines to be successful the parasites would be required to complete their lifecycle, inherently causing some degree of epithelial damage and subsequent inflammation. Being that the vaccines must elicit an immune response to be effective, it is conceivable that this immune status change itself could be expected to have some potential negative impact on welfare.

Health, thus absence of disease, is one of the physical domains in the five domains model of animal welfare, with a clear link to the mental domain (4). With impaired health, animals may be in a negative affective state, while good health facilitates positive emotional states (4, 5). Therefore, indicators of a healthy or diseased state are included in welfare assessment protocols. According to the recent EFSA opinion on the welfare of broiler chickens on-farm (6), one of the identified welfare consequences for broiler chickens is "gastro-enteric disorders." The welfare consequence has been defined as "The animal experiences negative affective states such as discomfort, pain and/or distress due to impaired function of the gastrointestinal tract resulting from, for example nutritional deficiency and infectious, parasitic or toxigenic agents," and was identified in all broiler production systems and breeds (6). Existing indicators of impaired intestinal health welfare assessment protocols are often non-specific and relate to a severe state of disease. E.g., the Welfare Quality protocol includes "plumage cleanliness," "footpad dermatitis," and "mortality" (7). Another assessment protocol includes "sick," "terminally ill," "dirty," "small," and "dead" (8). EFSA (6) listed "plumage/body cleanliness," "footpad dermatitis," "cloacal temperature," "lethargy," "impaired growth rate," and "mortality" as possible indicators of gastro-intestinal disorders. A recent analysis of scientific literature related to welfare assessment under commercial conditions did not reveal other indicators in broiler chickens (9).

The listed welfare indicators, while important for assessing broiler welfare and acknowledging health, are not specific to intestinal health and may overlook subclinical disease. Early or subclinical intestinal disease may however be painful, cause malaise, and could impact welfare by hindering normal behavior and positive emotional states. Early detection encourages prompt intervention, preventing more serious consequences.

The connection between an organism's genotype and its external phenotype is mediated through several intermediary layers of so-called internal phenotypes (10). These layers, including the transcriptome, proteome, metabolome, and microbiome, represent key biochemical aspects of the central dogma of life. The transcriptome captures the direct effects of environmental factors on gene expression, while the proteome reflects the translation of these gene expressions into proteins. The metabolome represents complex profiles of metabolites, and the microbiome, particularly the gut microbiota, is influenced by both the host genome and environmental factors such as diet and pathogens. These internal phenotypic layers are interconnected and their combined profiles determine the external phenotype (11). Despite the known quantitative effects of the environment on external traits, the specific impacts on internal phenotypes are not yet fully understood. However, the use of ~omics technologies, each capturing distinct layers of internal phenotypes, plays a crucial role in identifying biomarkers that are key to understand and manage gastrointestinal health and diseases (12-14).

This perspective paper proposes developing specific biomarkers for early detection of intestinal health deterioration in chickens. We review coccidiosis's impact on external and internal phenotypes, particularly internal phenotypic layers. Developing such biomarkers could enable timely interventions helping to mitigate disease effects, enhance welfare, and reduce reliance on reactive treatments. Integrating these biomarkers into welfare assessments promotes proactive, sustainable poultry production and could help evaluate control strategies which impacts welfare, e.g., the innate immune response following vaccinations might have an underestimated impact on welfare. Figure 1 outlines the interrelationships between loss of intestinal integrity and broiler welfare that will be briefly discussed first, after which we will propose routes of detecting and developing promising biomarkers of impaired intestinal health.

2 The intricate interactions of intestinal barrier function components form the gut-brain axis and the gut-associated immune response

Figure 2 presents a schematic overview of elements of the intestinal barrier function, the gut associated immune response and their complex interaction during infection, which will be discussed here. The complex intestinal barrier function protects the birds from harmful infectious agents while still enabling absorption of nutrients. The intestinal barrier function components include the intestinal epithelium, the mucus layer(s), the microbiota, immune elements and biochemical elements. Disruption of this intestinal barrier function, often referred to as "leaky gut," can be caused by consumption of contaminated feedstuff containing mycotoxins (15) and other toxins, poorly digested dietary protein (16) and pathogens such as *Eimeria* (17–19).



Cells of the immune system are an integral part of the mucosal tissue of the gut. Resident cells of the myeloid linage, including macrophages and dendritic cells, play an important role in maintaining gut integrity. They communicate with other immune cells, epithelial and stroma cells as well as the gut associated neuronal network through cell-cell contact and soluble mediators, called cytokines and hormones. In addition, epithelial cells and immune cells interact in a bidirectional way with members of the microbiota (20). This communication supports development of the gut immune system and maintenance of gut integrity and barrier function, while preventing gut inflammation (21). Thus, in the absence of pathogenic challenges the gut immune system receives cues from the microbiota and the epithelial cells which induce an anti-inflammatory response pattern by tissue resident macrophages, a phenotype referred to as M2 macrophages (22). Invasion of the gut by viral, bacterial, or parasitic pathogens shifts this "homeostatic immune response" toward an inflammatory response to control infection. Epithelial cells and resident macrophages of the lamina propria are the first cells to detect pathogens and to activate defense mechanisms (23). This includes a change in the macrophage function toward an M1 phenotype characterized by the secretion of inflammatory cytokines and chemokines (Figure 2). The best characterized factors of this response are the cytokines interleukin-1 (IL-1), IL-6, IL-8, and tumor-necrosis factor- α (24). They orchestrate the attraction of monocytes and their differentiation to inflammatory macrophages and the immigration of heterophils (25), the predominant form of granulocytes in chickens. The resulting morphological pictures of a massive increase in cell numbers in the lamina propria is described as inflammation by histo-pathologists. The resulting cascade of events aims at eliminating the pathogen which is often associated with a disruption of the epithelial barrier and further amplification of the inflammatory response (26). During an Eimeria infection there is rapid infiltration of the mucosal tissue by heterophils and the activation of macrophages and T-cells which results in the secretion of inflammatory cytokines (27). These early responses do not clear the infection but lead to tissue damage and systemic responses. Frequently, secondary infections are a consequence of Eimeria induced damage to the gut barrier (28) exacerbating the inflammatory response and sickness behavior.

When produced in sufficient amounts, inflammatory cytokines may induce a systemic response called acute-phase-response



characterized by changes in liver metabolism and behavior. In the liver, acute-phase-proteins (APP) are synthesized in large amounts leading to an up to 1,000-fold increase in plasma concentrations (29). Quantification of APPs is a standard procedure in human medicine as an indicator of the severity of inflammation. In Chickens however (30), due to the requirement of blood sampling and subsequent analysis, their quantification as inflammatory markers is impractical. The production of large amounts of acutephase-proteins leads to a shift in the requirement of essential amino-acids, which will lead to muscle breakdown (31). Behavioral changes result from the effects of inflammatory cytokines on circumventricular organs in the brain and include an increase in body temperature [fever (32, 33)], inappetence, somnolence and apathy, called sickness behavior. Moreover, cytokine mediated stimulation of hypothalamic nuclei activates the hypothalamicpituitary-adrenal axis and the release of corticosterone from the adrenal gland which has potent immuno-suppressive effects (Figure 2).

A particular challenge for the immune system is the return to homeostatic conditions and restoration of gut integrity. Regulatory T-lymphocytes (Treg) are particularly important in this context. Currently, Treg cells and their cytokine repertoire are not well understood in chickens (34). They secrete a range of cytokines and chemokines which exert anti-inflammatory effect, best known are IL-10 and TGF- β . Other cytokines and chemokines regulate proliferation and differentiation of gut epithelial cells and help to strengthen the barrier.

The intestinal barrier components also play an important role in the gut-brain axis (Figure 2). The intestinal epithelium interacts with the brain via signaling molecules that traverse the intestinal epithelium and the circulatory system and cross the blood-brain barrier. While the mucus layer does not communicate directly with the brain, it forms a barrier that helps to maintain a healthy gut microbiota, which in turn communicates with the brain through the production of neurotransmitters and other metabolites. Immune cells in the gut produce cytokines and other signaling molecules that can influence brain function either directly or indirectly through interactions with the gut microbiota. Various molecules secreted into the lumen influence the gut microbiota and immune responses, which in turn influence the gut-brain axis. In summary, during bacterial colonization or Eimeria challenge, metabolites and/or biochemical elements are secreted into the gut lumen, which are then absorbed into the blood and transported to the brain. This influences hormone secretion via the gut-brain axis. Overall, the gut-brain axis involves complicated interactions between the gut, the nervous system, the endocrine system, the immune system and the gut microbiota. A disturbance of the gut-brain axis leads to a malfunction of the intestinal barrier and vice versa risking uncontrolled immunological reactions that can trigger low-grade inflammation of the mucous membranes and the brain, the first step toward triggering more permanent intestinal diseases. The gut-brain axis allows the brain to influence intestinal activity, including the activity of functional immune effector cells, and the gut to influence mood, behavior, cognition, and thus broiler welfare. Behavioral changes can relate to gut barrier dysfunction, as shown with feather pecking in laying hens (8). Studies underscore this, focusing on microbiota-mediated alteration of central serotonergic and dopaminergic systems (8) and the glutamatergic nervous system (35) in feather pecking. The maintenance of homeostatic state, as observed in clinically healthy birds with normal behavior, depends on the function of the intestinal epithelia and the resident microbiome. This relationship is mediated by biochemical elements that communicate with the brain via the enteric nervous system, the vagus nerve and the bloodstream, which transports effector molecules resulting from host-microbe interactions in the gut.

Thus, the change from the "homeostatic immune response" toward an inflammatory response is reflected in the change from comfort to discomfort in avians and mammals, in which the gutbrain axis likely plays a crucial role. With this change, welfare is impaired and a negative emotional state develops during intestinal infection. The massive increase in cells in the lamina propria, described as inflammation by histo-pathologists, could be seen as the first phase of the discomfort of being ill.

3 Discussion

As outlined above, when gut health is increasingly compromised, through the gut-brain axis multiple complex processes are activated that directly or indirectly affect a broiler chicken's emotional state and behavior. In current welfare assessment protocols indirect indicators of impaired intestinal health or indicators related to a very advanced stage of disease are included (7, 8). While, with respect to guaranteeing animal welfare, and to enable early intervention by farmers, biomarkers indicative of earlier stages of intestinal disease, and specifically highly prevalent diseases, are needed. This area clearly deserves further research, but there are promising areas of development.

It is widely assumed that internal phenotypes are interconnected and collectively shape the external phenotype (10, 35). However, many of these assumptions lack robust evidence and often oversimplify the complex processes influencing external phenotypes. This gap in understanding partly accounts for the challenges in bridging the knowledge divide between genotype and external phenotype. Recent efforts by researchers to elucidate connections between internal phenotypic layers have shown that integrating multi-scale quantitative, multiomics data using a regression-based approach holds significant promise (11). Such multi-omics data analyses approach has provided provisional insights into the interactions among the internal phenotypes or layers, including local (e.g., intestinal) and systemic components. Results generated from these integrated approaches can form hypothesis-driven research aimed at identifying causal relationships across biological scales, potentially reducing the knowledge gap between internal and external phenotypes. This general methodology can be adapted to various datasets and perturbations, including pathogens such as Eimeria. Beside the multi-omics data integration approach, "high-level" data integration of singular-omics representing one layer of the internal phenotype could be used to identify biomarkers. Omics technologies offer valuable insights into the complex mechanisms underlying the onset of coccidiosis by enabling the identification of specific biomarkers, as indicated in the left section of Figure 2 with the predominantly invasive biomarkers. These are more suitable for experimental conditions. However, by correlating these biomarkers with behavioral changes in chickens affected by coccidiosis, it is possible to develop more targeted preventive measures and improve disease management strategies in poultry.

Novel technologies offer opportunities for fine tuning noninvasive biomarkers of disease as indicated in the right section of Figure 2. Farmers and veterinarians use flock behavior, vocalizations and/or odor as indicators of flock health in their daily management (36). It is however difficult to detect very early stages of disease by visual inspections alone. Automated detection of broiler behavior, flock distribution and movement are promising, non-invasive tools for early detection of behavioral changes. It has been shown that optical flow patterns of flock activity can be used to predict Campylobacter infection of a flock in a very early stage, even earlier than by conventional microbiological methods (37). This suggests that certain deviations in behavior can be specific to certain infections, although further study is required to what extent such changes can also be linked to other infections. Behavioral changes have been proposed for early disease detection in dairy cattle (36), but it remains to be studied to what extent specific behaviors can be indicative of early intestinal challenge in broilers, and how these can be practically assessed. From a technical perspective it has already been demonstrated that individual behaviors and postures as well as group-level changes in activity and distribution can be detected in chickens (38, 39). Similarly, it has been shown that it is possible to detect abnormalities in fecal appearance by image analysis (40), early detection of respiratory diseases by sound analysis (41, 42) and a combination of visual and thermal images to detect fever in chickens (43). Although many of these automated detection technologies need further development for application in practice, these present opportunities for earlier and more specific disease detection in future welfare assessment schemes.

Despite significant progress in developing biomarkers for assessing intestinal health in broilers, the integration of these tools into standard welfare assessment protocols remains a future goal. The current biomarkers show promise in detecting early indicators of compromised intestinal integrity, which could allow for timely interventions and improved welfare outcomes. However, several challenges need to be addressed before these biomarkers can be reliably included in routine welfare assessments. These include validating biomarkers across diverse flock conditions, ensuring non- or minimally-invasive sampling, and establishing standardized thresholds. Additionally, further research is required to link specific biomarker responses to particular welfare-relevant outcomes, which will improve the specificity and relevance of these tools in practical applications. Overcoming these barriers will take time, yet the potential benefits underscore the importance of continued research and development in this area.

Data availability statement

The original contributions presented in the study are included in the article/supplementary material, further inquiries can be directed to the corresponding authors.

Author contributions

IdJ: Conceptualization, Funding acquisition, Visualization, Writing – review & editing, Writing – original draft. SK: Visualization, Writing – original draft, Writing – review & editing, Conceptualization. BK: Conceptualization, Writing – original draft, Writing – review & editing.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

1. Zhu QD, Sun P, Zhang BK, Kong LL, Xiao CP, Song ZG. Progress on gut health maintenance and antibiotic alternatives in broiler chicken production. *Front Nutr.* (2021) 8:692839. doi: 10.3389/fnut.2021.692839

2. Ducatelle R, Goossens E, Eeckhaut V, Van Immerseel F. Poultry gut health and beyond. *Animal Nutr.* (2023) 13:240–8. doi: 10.1016/j.aninu.2023.03.005

3. Parker CD, Lister SA, Gittins J. Impact assessment of the reduction or removal of ionophores used for controlling coccidiosis in the UK broiler industry. *Vet Rec.* (2021) 189:7. doi: 10.1002/vetr.513

4. Mellor DJ. Operational details of the five domains model and its key applications to the assessment and management of animal welfare. *Animals.* (2017) 7:60. doi: 10.3390/ani7080060

5. Webb LE, Veenhoven R, Harfeld JL, Jensen MB. What is animal happiness? *Ann NY Acad Sci.* (2019) 1438:62–76. doi: 10.1111/nyas.13983

6. EFSA Panel on Animal Health and Animal Welfare. Scientific Opinion on the welfare of broilers on farm. *EFSA J.* (2023) 21:236. doi: 10.2903/j.efsa. 2023.7788

7. Quality W. The Welfare Quality Assessment Protocol for Broiler Chickens and Laying Hens. Lelystad: The Welfare Quality Consortium. (2009).

8. BenSassi N, Vas J, Vasdal G, Averos X, Estevez I, Newberry RC. Onfarm broiler chicken welfare assessment using transect sampling reflects environmental inputs and production outcomes. *PLoS ONE.* (2019) 14:e0214070. doi: 10.1371/journal.pone.0214070

9. Paulović T, Jong I de, Ouweltjes W, Martin Valls GE, Llonch Obiols P, Ko H-L, et al. Development of a roadmap for action for the project More Welfare: towards new risk assessment methodologies and harmonised animal welfare data in the EU. *EFSA J.* (2024) 20:8566E. doi: 10.2903/sp.efsa.2024.EN-8566

10. Leuchter AF, Hunter AM, Krantz DE, Cook IA. Intermediate phenotypes and biomarkers of treatment outcome in major depressive disorder. *Dialogues Clin Neurosci.* (2014) 16:525–37. doi: 10.31887/DCNS.2014.16.4/aleuchter

11. Benis N, Kar SK, dos Santos V, Smits MA, Schokker D, Suarez-Diez M. Multi-level integration of environmentally perturbed internal phenotypes reveals key points of connectivity between them. *Front Physiol.* (2017) 8:388. doi: 10.3389/fphys.2017.00388

12. Beck LC, Granger CL, Masi AC, Stewart CJ. Use of omic technologies in early life gastrointestinal health and disease: from bench to bedside. *Expert Rev Proteomics*. (2021) 18:247–59. doi: 10.1080/14789450.2021.1922278

13. Goossens E, Dehau T, Ducatelle R, Van Immerseel F. Omics technologies in poultry health and productivity - part 2: future applications in the poultry industry. *Avian Pathol.* (2022) 51:418–23. doi: 10.1080/03079457.2022.20 85545

14. Nguyen TQ, Martínez-alvaro M, Lima J, Auffret MD, Rutherford KMD, Simm G, et al. Identification of intestinal and fecal microbial biomarkers using a porcine social stress model. *Front Microbiol.* (2023) 14:1197371. doi: 10.3389/fmicb.2023.1197371

15. Kang TH, Lee SI. Establishment of a chicken intestinal organoid culture system to assess deoxynivalenol-induced damage of the intestinal barrier function. *J Anim Sci Biotechnol.* (2024) 15:12. doi: 10.1186/s40104-023-00976-4

16. Gilbert MS, Ijssennagger N, Kies AK, van Mil SWC. Protein fermentation in the gut; implications for intestinal dysfunction in humans, pigs, and poultry. *Am J Physiol-Gastroint Liver Physiol.* (2018) 315:G159–G70. doi: 10.1152/ajpgi.00319.2017

17. Awad WA, Molnár A, Aschenbach JR, Ghareeb K, Khayal B, Hess C, et al. Campylobacter infection in chickens modulates the intestinal epithelial barrier function. *Innate Immun.* (2015) 21:151–60. doi: 10.1177/1753425914521648

18. Moran ET. Intestinal events and nutritional dynamics predispose Clostridium perfringens virulence in broilers. *Poult Sci.* (2014) 93:3028– 36. doi: 10.3382/ps.2014-04313

19. Chapman HD. Milestones in avian coccidiosis research: a review. *Poult Sci.* (2014) 93:501–11. doi: 10.3382/ps.2013-03634

20. Soderholm AT, Pedicord VA. Intestinal epithelial cells: at the interface of the microbiota and mucosal immunity. *Immunology.* (2019) 158:267–80. doi: 10.1111/imm.13117

21. Kayama H, Okumura R, Takeda K. Interaction between the microbiota, epithelia, and immune cells in the intestine. *Annu Rev Immunol.* (2020) 38:23-48. doi: 10.1146/annurev-immunol-070119-115104

22. Chen W, Liu D, Ren CH, Su XM, Wong CK, Yang RC, et al. Special network comprised of macrophages, epithelial cells, and gut microbiota for gut homeostasis. *Cells.* (2022) 11:307. doi: 10.3390/cells11020307

23. Qureshi MA, Heggen CL, Hussain I. Avian macrophage: effector functions in health and disease. *Dev Comp Immunol.* (2000) 24:103–19. doi: 10.1016/S0145-305X(99)00067-1

24. Hong YH, Lillehoj HS, Lillehoj EP, Lee SH. Changes in immune-related gene expression and intestinal lymphocyte subpopulations following *Eimeria maxima* infection of chickens. *Vet Immunol Immunopathol.* (2006) 114:259–72. doi: 10.1016/j.vetimm.2006.08.006

25. Kogut MH, Tellez GI, McGruder ED, Hargis BM, Williams JD, Corrier DE, et al. Heterophils are decisive components in the early responses of chickens to salmonellaenteritidis infections. *Microb Pathog.* (1994) 16:141–51. doi: 10.1006/mpat.1994.1015

26. Smith AL, Powers C, Beal R. The avian enteric immune system in health and disease. In: Kaspers B, Schat KA, Göbel T, Vervelde L, editors. *Avian Immunology*. New York: Acdemic press (2022). p. 303–26. doi: 10.1016/B978-0-12-818708-1.00030-0

27. Wang DN, Zhang Q, Zhang Z, Zhang Y, Wang S, Han YH, et al. Expression profile of Toll-like receptors and cytokines in the cecal tonsil of chickens challenged with *Eimeria tenella*. *Parasitol Res.* (2024) 123:12. doi: 10.1007/s00436-024-08371-2

28. Rose ME, Long PL, Bradley JWA. Immune-responses to infections with coccidia in chickens - gut hypersensitivity. *Parasitology*. (1975) 71:357. doi: 10.1017/S0031182000047132

29. Kushner I. The phenomenon of the acute phase response. *Ann NY Acad Sci.* (1982) 389:39–48. doi: 10.1111/j.1749-6632.1982.tb22124.x

30. Riva F, Eckersall PD, Chadwick CC, Chadwick LC, McKeegan DEF, Peinado-Izaguerri J, et al. Identification of novel biomarkers of acute phase response in chickens challenged with *Escherichia coli* lipopolysaccharide endotoxin. *BMC Vet Res.* (2024) 20:8. doi: 10.1186/s12917-024-04062-3

31. Barnes DM, Song Z, Klasing KC, Bottje W. Protein metabolism during an acute phase response in chickens. *Amino Acids*. (2002) 22:15-26. doi: 10.1007/s726-002-8198-6

32. Grabbe N, Kaspers B, Ott D, Murgott J, Gerstberger R, Roth J. Neurons and astrocytes of the chicken hypothalamus directly respond to lipopolysaccharide and chicken interleukin-6. *J Comp Physiol B-Biochem Syst Environ Physiol*. (2020) 190:75–85. doi: 10.1007/s00360-019-01249-1

33. Marais M, Maloney SK, Gray DA. Brain IL-6-and PG-dependent actions of IL-1β and lipopolysaccharide in avian fever. *Am J Physiol-Regul Integr Comp Physiol.* (2011) 301:R791–800. doi: 10.1152/ajpregu.00136.2011

34. Burkhardt NB, Elleder D, Schusser B, Krchlíková V, Göbel TW, Härtle S, et al. The discovery of chicken Foxp3 demands redefinition of avian regulatory T cells. *J Immunol.* (2022) 208:1128–38. doi: 10.4049/jimmunol.2000301

35. Fontanesi L. Metabolomics and livestock genomics: Insights into a phenotyping frontier and its applications in animal breeding. *Animal Frontiers*. (2016) 6:73–9. doi: 10.2527/af.2016-0011

36. Weary DM, Huzzey JM, von Keyserlingk MAG, Board-Invited Review. Using behavior to predict and identify ill health in animals. *J Anim Sci.* (2009) 87:770–7. doi: 10.2527/jas.2008-1297

37. Colles FM, Cain RJ, Nickson T, Smith AL, Roberts SJ, Maiden MCJ, et al. Monitoring chicken flock behaviour provides early warning of infection by human pathogen Campylobacter. *Proc R Soc B-Biol Sci.* (2016) 283:20152323. doi: 10.1098/rspb.2015.2323

38. Wurtz K, Camerlink I, D'Eath RB, Fernandez AP, Norton T, Steibel J, et al. Recording behaviour of indoor-housed farm animals automatically using machine vision technology: a systematic review. *PLoS ONE.* (2019) 14:e0226669. doi: 10.1371/journal.pone.0226669

39. Bhuiyan MR, Wree P. Animal behavior for chicken identification and monitoring the health condition using computer vision: a systematic review. *IEEE Access.* (2023) 11:126601-10. doi: 10.1109/ACCESS.2023.333 1092

40. Wang JT, Shen MX, Liu LS, Xu Y, Okinda C. Recognition and classification of broiler droppings based on deep convolutional neural network. *J Sensors*. (2019) 2019:3823515. doi: 10.1155/2019/3823515

41. Sun ZG, Tao WG, Gao MM, Zhang M, Song SL, Wang GT. Broiler health monitoring technology based on sound features and random forest. *Eng Applic Artif Intell*. (2024) 135:108849. doi: 10.1016/j.engappai.2024.108849

42. Liu LS, Li B, Zhao RQ, Yao W, Shen MX, Yang J. A novel method for broiler abnormal sound detection using WMFCC and HMM. *J Sensors*. (2020) 2020:2985478. doi: 10.1155/2020/2985478

43. Elmessery WM, Gutiérrez J, Abd El-Wahhab GG, Elkhaiat IA, El-Soaly IS, Alhag SK, et al. YOLO-based model for automatic detection of broiler pathological phenomena through visual and thermal images in intensive poultry houses. *Agric Basel.* (2023) 13:1527. doi: 10.3390/agriculture130 81527