

THE SEARCH FOR BIOMARKERS IN PSYCHIATRY

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PUBLISHED IN: Frontiers in Psychiatry





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ISSN 1664-8714

ISBN 978-2-88971-301-1

DOI 10.3389/978-2-88971-301-1

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THE SEARCH FOR BIOMARKERS IN PSYCHIATRY

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Citation: Manzanares, J., Garcia-Gutierrez, M. S., Rueda, F. N., Gatecki, P., eds. (2021).
The Search for Biomarkers in Psychiatry. Lausanne: Frontiers Media SA.
doi: 10.3389/978-2-88971-301-1

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Editorial: The Search for Biomarkers in Psychiatry

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Keywords: biomarkers, central biomarkers, peripheral biomarkers, omics, psychiatry

Editorial on the Research Topic

The Search for Biomarkers in Psychiatry

Psychiatric disorders present a high level of complexity in many aspects. Contrary to other diseases, they are classified by diagnostic categories with a broad variety list of symptoms. This categorical organization results in a great clinical heterogeneity among patients diagnosed from the same psychiatric illness. Besides, the high rate of comorbidity among psychiatric disorders is an additional distinguishing factor that makes diagnosis complicated. On the top of that, the limited knowledge of the molecular mechanisms underlying mental disorders, greatly contribute to the reduced efficacy of current pharmacological treatments, being especially poor as the severity of the disease increases. This clinical situation stimulated the searching of biomarkers to improve prevention, diagnosis and treatment of psychiatric disorders. This special issue includes one review article of García-Gutiérrez et al., summarizing the concept and types of biomarkers in psychiatry and providing examples about the most promising results achieved sorted by categories (genetics, transcriptomics, proteomics, metabolomics, and epigenetics). The review includes a final conclusion that remarks the future challenges required to reach the goal of developing valid, reliable and broadly-usable biomarkers for psychiatric disorders and their treatment. Complementary, the opinion article of Vinberg further explored the main limitations and the future perspectives in the searching of peripheral biomarkers in psychiatry.

The potential role of key targets of the immune and endocannabinoid systems are studied in detail in three articles. The brief research report of Larsen et al. provide evidences about an association between alterations in different cytokines and motor activity in an acute psychiatric population, suggesting that some cytokines deserve further exploration as biomarkers for predicting and treating changes in motor activity. Additionally, the review article of Momtazmanesh et al., is a fantastic review summarizing the main cytokine alterations observed in schizophrenic patients, their potential link with certain symptoms and their potential clinical impact. Similarly, the review article of Navarrete et al., provide the most promising results achieved to date that support the potential role of different targets of the endocannabinoid system as biomarkers in several psychiatric conditions.

Besides, the review of Li Z. et al., is an excellent article containing the most recent studies supporting the potential use of circular RNA as biomarkers in two major psychiatric disorders, depression and schizophrenia. Finally, the review of Jurado-Barba et al., covers the potential clinical use of EEG as clinical assessment in alcohol dependence.

A total of seven articles are focused on the identification of biomarkers in major depressive disorder (MDD). Zhao et al., analyse the utility of CACNA1C rs1006737 polymorphism together with exposure to threatening life events as predictive biomarkers for MDD, pointing out this polymorphism as a target for new pharmacological treatments in this psychiatric disorder. Explore the use of high heart rate as biomarker for assessing the outcome of

OPEN ACCESS

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Specialty section:

This article was submitted to
Molecular Psychiatry,
a section of the journal
Frontiers in Psychiatry

Received: 04 June 2021

Accepted: 16 June 2021

Published: 12 July 2021

Citation:

Garcia-Gutierrez MS, Manzanares J
and Navarrete F (2021) Editorial: The
Search for Biomarkers in Psychiatry.
Front. Psychiatry 12:720411.
doi: 10.3389/fpsy.2021.720411

reinstatement at work in MDD patients who took a leave of absence, with promising results. Zhao et al., carried out the study of different single nucleotide polymorphisms of FoxO1, A2M, and TGF-B1 in MDD patients, demonstrating a potential association between some of them and the environment in MDD. Authors suggest that these SNPs may be useful for assessing MDD risk improving prevention.

An emerging type of biomarkers is metabolomic alterations. In this respect, Gao et al., focus on studying the role of 36 metabolic biomarkers in depression. The research sheds light on potential important metabolites and enzymes in the underlying molecular mechanisms of depression.

Troyan and Levada, wrote an exciting original article revealing the correlation between serum concentrations of the neurotrophin BDNF and IGF-1 and MDD. Interestingly, changes in these targets are also observed after 8 weeks of treatment with the antidepressant vortioxetine, supporting that both may be useful as biomarkers of diagnosis and treatment outcome in MDD. Similarly, Köhler-Forsberg et al., provide the results of an open label clinical trial in MDD patients supporting the utility of a panel of biomarkers (serotonin 4 receptor PET brain imaging, fMRI, cognitive-, EEG-and peripheral biomarkers) for MDD diagnosis and for predicting pharmacological efficacy. Ho et al., presented a comprehensive update regarding functional near-infrared spectroscopy (fNIRS) as a biomarker for guiding diagnosis and monitoring treatment response in MDD.

Kittel-Schneider et al., covered the emerging role of proteomics profile as a diagnostic tool for distinguish bipolar from unipolar depression. The results support the potential role of the multivariate predictive model proposed by authors as a predictive biomarker model useful to discriminate between these psychiatric entities.

Interestingly, Cross et al., presented a case report suggesting the clinical relevance to discard the presence of infectious diseases, such as Lyme borreliosis or PANDAS, in patients presenting neuropsychiatric symptoms. These results data, although preliminary, support the close connection between the immune system and the brain.

The original article of Li G. et al., point out lower serum uric acid concentrations as a biomarker for predicting depression in stroke patients. These interesting results support the development of additional longitudinal and clinical studies.

Taken together, these articles updated new advances about peripheral and central biomarkers using a variety of methods (proteomics, transcriptomics, genetics, and imaging) in psychiatric clinical conditions and samples to identify alterations with specific traits of the disease or with the outcome of the pharmacological treatments.

AUTHOR CONTRIBUTIONS

All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Cytokine Alterations in Schizophrenia: An Updated Review

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OPEN ACCESS

Edited by:

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Reviewed by:

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Specialty section:

This article was submitted to
Molecular Psychiatry,
a section of the journal
Frontiers in Psychiatry

Received: 08 September 2019

Accepted: 13 November 2019

Published: 06 December 2019

Citation:

Momtazmanesh S, Zare-Shahabadi A
and Rezaei N (2019) Cytokine
Alterations in Schizophrenia: An
Updated Review.
Front. Psychiatry 10:892.
doi: 10.3389/fpsy.2019.00892

Schizophrenia, a multisystem disorder with an unknown etiology, is associated with several immune dysfunctions, including abnormal levels of circulating cytokines. In this review, we investigated the changes of cytokines in schizophrenic patients, their connection with behavioral symptoms severity and their potential clinical implications. We also assessed the possible causative role of abnormal cytokine levels in schizophrenia pathogenesis. Based on meta-analyses, we categorized cytokines according to their changes in schizophrenic patients into four groups: (1) increased cytokines, including interleukin (IL)-6, tumor necrosis factor (TNF)- α , IL-1 β , IL-12, and transforming growth factor (TGF)- β , (2) non-altered cytokines, including IL-2, IL-4, and IL-17, (3) increased or non-altered cytokines, including IL-8 and interferon (IFN)- γ , and (4) IL-10 with increased, decreased, and non-altered levels. Notably, alterations in cytokines may be variable in four different categories of SP, including first-episode and drug-naïve, first-episode and non-drug-naïve, stable chronic, and chronic in acute relapse. Furthermore, disease duration, symptoms severity, incidence of aggression, and cognitive abilities are correlated with levels of certain cytokines. Clinical implications of investigating the levels of cytokine in schizophrenic patients include early diagnosis, novel therapeutic targets development, patient stratification for choosing the best therapeutic protocol, and predicting the prognosis and treatment response. The levels of IL-6, IL-8, IFN- γ , IL-2 are related to the treatment response. The available evidence shows a potential causative role for cytokines in schizophrenia development. There is a substantial need for studies investigating the levels of cytokines before disease development and delineating the therapeutic implications of the disrupted cytokine levels in schizophrenia.

Keywords: schizophrenia, cytokines, inflammation, behavioral symptoms, treatment outcome, antipsychotic agents

INTRODUCTION

Schizophrenia, a multisystem disorder with a global prevalence of 0.33–0.75%, is one of the top 15 causes of disability (1–4). The underlying etiology of this disease is controversial and not fully understood. Increased dopamine-based activities, together with decreased glutamatergic signaling, are the main suggested etiological hypotheses (5). Abnormalities in the immune system, which are associated with schizophrenia, are one of the other etiological hypotheses.

The immune system is composed of innate and adaptive immune responses. The innate immunity is a rapid-acting antigen-independent response, while the adaptive immunity is an antigen-dependent defense mechanism, with the ability to memorize the antigens. The immune response is mainly mediated by cytokines, which are mostly produced by a critical component of the adaptive immunity, T-lymphocytes. These mediators can be divided into 5 groups; (1) pro-inflammatory cytokines; interleukin (IL)-6, tumor necrosis factor (TNF)- α , IL-1 family, and IL-8, which are involved in initiation and aggravating inflammatory responses (6); (2) T-helper 1 cytokines; IL-2, interferon (IFN)- γ , and IL-12, which create a pro-inflammatory response and function in autoimmune diseases and defense against intracellular parasites; (3) T-helper 2 cytokines; IL-4, IL-5, and IL-13, which counterbalance the effects of T-helper1 cytokines; (4) T-helper 17 cytokines; IL-17, and IL-23, which are chiefly involve in pro-inflammatory processes and defense against extracellular pathogens; (5) T-regulatory cytokines; IL-10 and transforming growth factor (TGF)- β , which primarily suppress immune responses (7, 8).

The associated immune disorders in schizophrenia have been investigated for more than a century (9–11). These abnormalities include increased activity and density of microglia cells, abnormal profiles of peripheral leukocytes, serum cytokines, and cerebrospinal fluid (CSF) cytokines (12, 13). Based on genome-wide association studies, schizophrenia is also associated with specific major histocompatibility complex (MHC) region genes (14). Furthermore, this disease is significantly linked with enhancers having a strong role in the immune functions, even after excluding the MHC region genes (15). These findings support the clinical and genetic aspects of the connection between schizophrenia and the immune system.

One group of components of the immune system affecting the brain by several mechanisms are cytokines which are either produced outside of the central nervous system (CNS) or within the CNS. Peripheral cytokines, presented in the circulation, can access the CNS and affect it *via* four major ways: (1) binding to specific transporters, (2) stimulating afferent vagal fibers, (3) accessing areas such as circumventricular organs, and (4) passing the damaged blood-brain barrier (BBB) which has an increased permeability (16, 17). Notably, elevated levels of the peripheral markers of BBB damage, such as S100B, indicate BBB damage in schizophrenic patients (18). In addition to the peripheral cytokines, microglia, astrocytes, endothelial cells, and even neurons can produce different cytokines within the CNS (17, 19). Moreover, despite the prevailing view that the brain is an immune-privileged area, several studies have shown that immune responses can be established within CNS by several mechanisms, one of which is

transferring the immune cells located within the meninges, which are sources of different immune mediators, into the brain's parenchyma in a pathologic state. These cells physiologically transfer through the blood-meningeal barrier in order to pass the meninges as it is more permeable than BBB (20, 21).

These cytokines can have various roles in neurodevelopment, neuroendocrine activities, and neurotransmission. Their role in neurodevelopment is mainly through affecting microglia which are the chief cells responsible for this task (11, 16, 17, 19). Cell migration, water balance, body temperature regulation and synthesis and release of neurotransmitters can be influenced by these mediators (16).

A role for cytokines in schizophrenia was proposed almost three decades ago (22, 23). Ever since, an increasing number of studies investigated alterations of cytokine levels in schizophrenic patients, their changes following antipsychotic treatment, and their relationship with clinical manifestations. However, not only are the results of these studies controversial, but they also do not clearly answer whether the changes in the levels of these cytokines can have a causative role in the development of psychotic symptoms. Moreover, diagnostic and therapeutic implications of these alterations are not defined.

This review aims to answer four questions: A) What are the changes in the serum levels of cytokines in schizophrenic patients? B) What is the relationship between the levels of cytokines and the severity of clinical symptoms? C) How antipsychotics affect the baseline levels of cytokines? and D) What is the potential role of cytokines as predictors of treatment response?

Using the answers to these questions, we investigate whether abnormal cytokine levels are the culprit in the pathogenesis of schizophrenia and also provide clinical implications in terms of diagnosis and treatment.

ALTERATIONS IN THE LEVELS OF CYTOKINES IN SCHIZOPHRENIC PATIENTS

Alterations in the Levels of Pro-Inflammatory Cytokines

A considerable number of studies, including meta-analyses, found increased levels of IL-6 in different groups of patients including, first-episode and drug-naïve (FEDN) psychosis patients and cases with first-episode psychosis (FEP), majority of whom were using antipsychotics (24–28). Similar findings were observed in chronic patients, including those without any significant inflammation (29), patients in an acute relapse or recovering from it, and stable outpatients (27, 30–32). Recently, Hartwig et al. found increased levels of soluble IL-6 receptors in a clinical two-sample Mendelian randomization study, which can be explained as a compensatory response to the increased levels of IL-6 in schizophrenia (33).

Conversely, some studies found no significant changes in the levels of IL-6 in schizophrenic patients (34, 35).

Multiple studies, including meta-analyses, reported elevated levels of TNF- α , one of the other pro-inflammatory cytokines, in

FEDN patients (24, 25), (both adult and pediatric) FEP patients, majority of whom were using antipsychotics (26, 27, 36) and chronically ill patients using antipsychotics, regardless of their status in terms of acute relapse (24, 37). Higher levels of TNF- α have also been reported in chronic patients taking atypical antipsychotics having no major inflammation (29).

On the contrary, Potvin et al. conducted a meta-analysis and found no significant alteration in the levels of TNF- α in *in vivo* and *in vitro* studies (30). Their finding may be explained by the scarcity of studies until 2005. Decreased levels of TNF- α are seen in FEDN patients with a disease duration of under two years (37) and chronic patients with a disease duration of more than five years taking typical and atypical antipsychotics (38, 39).

Regarding the next pro-inflammatory cytokine, IL-8, despite meta-analyses supporting elevated levels of it in FEP patients (27) and chronic patients who are stable or are experiencing an acute relapse or are recovering from one (24, 32), a recent meta-analysis showed no significant alterations in the levels of IL-8 in FEP patients versus HC's (32). This is concordant with the findings of another study in FEP patients, most of whom were medicated (36). Concordantly, in a study in which less than a quarter of patients were taking benzodiazepine and others were drug-naïve, except for obese cases who had increased levels of IL-8, other patients showed no significant alterations in the IL-8 level (40).

IL-1 family is one of the major pro-inflammatory cytokines, one of the members of which is IL-1 β . A large number of studies, including several meta-analyses, found elevated levels of IL-1 β in FEDN patients (25) and adult and pediatric FEP patients, majority of whom were taking atypical antipsychotics (26, 28, 36), and chronically ill patients who were stable or were experiencing an acute relapse or were recovering from one (24, 27, 32, 37). Interestingly, it seems that the levels of IL-1 β mRNA do not change in FEP patients (36).

In contrast to a substantial number of studies suggesting an increase in the levels of IL-1 β , the meta-analysis conducted by Potvin et al. in 2008 found no significant alterations in the levels of IL-1 β in *in vivo* and *in vitro* studies (30). No significant elevation has been reported in chronic patients with a disease period of more than six years, either (41). Interestingly, a recent study has found decreased levels of IL-1 β in FEDN patients with a disease period of shorter than 2 years (37).

Other members of the IL-1 family may be elevated in schizophrenic patients as well. The levels of IL-1 α and its leukocyte mRNA were found to be higher in FEP patients who were mostly medicated (36). However, other investigators did not find the same pattern in long-term chronically ill patients (41). A recent study has reported increased levels of IL-33, one of the other members of IL-1 family, and its soluble receptor (sST2) in FEDN patients and patients in an acute relapse compared to patients in remission or HC's (42). Several studies, including a number of meta-analyses, reported increased levels of IL-1 receptor antagonist (IL-1RA) in patients with first-episode psychosis, whether most of the patients used antipsychotics (43) or whether they were drug-naïve (40), in chronic patients who were experiencing an acute relapse or had multiple episodes of schizophrenia (24, 27, 32), and in *in vitro* studies (30).

Alterations in the Levels of T-Helper 1 Cytokines

According to the majority of studies (including meta-analyses), the levels of IL-2, one of the cytokines produced by T-helper 1, do not alter in schizophrenic patients. These studies were performed on FEDN patients, FEP patients (with a history of using antipsychotics in many of them), and chronic patients who were experiencing an acute relapse, were recovering from it or were stable (24, 25, 27, 35, 36).

However, several studies found contradictory results. Some of these studies reported increased levels of IL-2 in FEDN patients with a normal BMI (44), FEP patients, majority of whom were using atypical antipsychotics (28), and chronic patients with stable antipsychotic medication regimens combined with a disease period of more than six years (41). Furthermore, some *in vitro* studies found a substantial decrease in the levels of IL-2 (30). Studies also found a decrease in the mRNA levels of this cytokine in the peripheral blood of chronically ill patients who were using antipsychotics for at least a year (45).

Moreover, several studies (including meta-analyses) found a significant increase in the levels of soluble IL-2 receptor (sIL-2R), levels of which might affect the level of IL-2 by binding with it, in FEDN patients and chronic patients who were stable or were experiencing an acute relapse or were recovering from one (24, 25, 27).

Alterations in IFN- γ , the next T-helper 1 cytokine, levels are very controversial. The correlation between IFN- γ and BMI, previously found in patients with first-episode schizophrenia, may explain this inconsistency (32). Due to this great controversy, the reported changes in the levels of this cytokine in each group of patients are discussed separately.

In FEDN patients, several studies, including a meta-analysis, found no significant alterations in the levels of IFN- γ (40, 46–49) while some other studies, including another meta-analysis, showed increased levels of IFN- γ (24, 50). Conversely, Reale et al. found decreased levels of IFN- γ in FEDN patients (51).

Studies showed elevated levels of this cytokine in adult or pediatric FEP patients, most of whom had a history of using antipsychotic medications (26–28). However, Di Nicola et al. found non-altered levels in adult FEP patients, most of whom were on atypical antipsychotics (36).

Several studies, including some meta-analyses, found elevated levels of IFN- γ in chronic schizophrenic patients who were stable or were experiencing an acute relapse (24, 27, 32). However, no significant disruption was found in patients recovering from an acute relapse (27, 46). Decreased levels of IFN- γ were also reported in patients with acute psychotic symptoms who were drug-naïve for at least 6 months (52) and in chronic patients. The details of the medications that were used were not reported in these studies (53, 54).

Investigation of the alterations in the levels of IL-12, one of the other T-helper 2 cytokines, showed that several studies (including meta-analyses) found elevated levels in FEP patients, whether drug-naïve or not, and chronic patients who were stable or were experiencing an acute relapse, or were recovering from one (24, 27, 55). In the study by Bedrossian

et al., the patients were treated with clozapine for at least one year (55).

In contrast to the studies reporting elevated levels of IL-12, some other studies reported non-altered levels of this cytokine in FEP (28, 43), and in chronic patients who had a history of using antipsychotic medications (32, 45).

Alterations in the Levels of T-Helper 2 Cytokines

Compared to other T-helper 2 cytokines, there are a larger number of studies performed on the alterations of IL-4. Several studies, including two meta-analyses, found no major differences in the levels of IL-4 between HC's and FEDN or FEP patients, majority of whom were taking atypical antipsychotics, or chronic patients who had schizophrenia for a long time (including treatment-resistant patients) (25, 27, 30, 36, 48, 56). However, like other cytokines, a small number of studies reported inconsistent results. Decreased levels of IL-4 were reported in chronic patients experiencing an acute relapse, during their treatment after the relapse (supported by a meta-analysis) (27), and in stable chronic patients (41, 57). In addition, elevated levels were reported in FEP pediatric patients taking antipsychotics (26) and in adult chronic patients taking clozapine (58).

Few studies have investigated other T-helper 2 cytokines, including IL-5 and IL-13. Increased levels of IL-5 were found in chronic adult patients with multiple episodes of unsuccessful treatment and FEP pediatric patients who were mostly taking antipsychotics (26, 59). Similarly, the levels of IL-13 have been reported to be elevated in adults with multiple episodes of schizophrenia (32, 59).

Alterations in the Levels of T-Helper 17 Cytokines

The literature is inconsistent about IL-17 alterations in schizophrenic patients. A recent meta-analysis found no significant changes in the levels of IL-17 in FEDN patients (60). Non-disturbed levels are also reported in chronic patients experiencing an acute relapse (46).

On the contrary, some studies reported increased levels of this cytokine in FEDN (50) and chronic hospitalized patients who were medication free for at least four weeks (61).

However, decreased levels have also been reported in FEDN patients (46) and in chronic patients using different antipsychotics (62).

One of the other main T-helper 17 cytokines is IL-23, which has been reported to be elevated in FEDN and chronic patients in an acute relapse (8, 61, 63).

Alterations in the Levels of T-Regulatory Cytokines

Goldsmith et al. found reduced levels of IL-10 in FEP patients and chronic patients who were in an acute relapse in their meta-analysis (27). Decreased levels are also reported in FEDN patients (64).

However, this meta-analysis showed that the levels of this cytokine do not change in stable chronic patients (27, 45). Other studies have reported no significant alterations in the levels of IL-10 in comparison with HC in FEDN (44), FEP (36), and chronic patients experiencing an acute relapse (65).

Conversely, many investigators have reported elevated levels of this cytokine in FEDN (49, 66), FEP (26), and chronic patients (41, 57, 67).

Several studies, including meta-analyses, found elevated levels of TGF- β in FEDN, FEP patients (27, 46), and in chronic patients experiencing an acute relapse (47).

However, some studies found other findings. Non-disturbed levels of TGF- β have been detected in FEDN and chronic patients who were medication free for four months before the study (18, 44). Interestingly, some studies found decreased levels of TGF- β in chronic treatment-resistant patients (56).

A summary of alterations in the levels of cytokines in schizophrenia based on the meta-analyses is shown in **Table 1**.

RELATIONSHIP BETWEEN CYTOKINE LEVELS AND SEVERITY OF CLINICAL SYMPTOMS

The levels of cytokines seem to be correlated with both the disease duration and symptom severity. Patients with elevated levels of IL-6, IL-8, and IL-4 tend to have a longer disease duration and longer hospitalizations (31, 65, 68).

Furthermore, higher levels of IL-6, IL-1 β , IL-33, and IL-17 are associated with more severe positive symptoms (28, 42, 50, 69, 70). In chronic patients using a stable dose of antipsychotics, decreased levels of TNF- α are similarly associated with more severe positive symptoms (38, 71), while no correlation has been found in FEDN patients (37).

TABLE 1 | Summary of alterations in serum levels of cytokines in schizophrenia, based on meta-analyses [18, 20, 21].

| Increased levels | | Non-altered levels | | Increased or non-altered | | Increased or decreased or non-altered | |
|------------------|------|--------------------|------|--------------------------|------|---------------------------------------|------|
| Cytokine | Type | Cytokine | Type | Cytokine | Type | Cytokine | Type |
| IL-6 | PI | IL-2 | TH1 | IL-8 | PI | IL-10 | TR |
| TNF- α | PI | IL-4 | TH2 | IFN- γ | TH1 | | |
| IL-1 β | PI | IL-17 | TH17 | | | | |
| IL-12 | TH1 | | | | | | |
| TGF- β | TR | | | | | | |

IL, interleukin; TNF, tumor necrosis factor; TGF, transforming growth factor; IFN, interferon; PI, pro-inflammatory cytokine; TH1, T-helper 1 cytokine; TH2, T-helper 2 cytokine; TR, T-regulatory cytokine; TH17, T-helper 17 cytokine.

Exacerbated negative symptoms are seen in patients with elevated levels of IL-6, TNF- α , IL-1 β , IL-8, IFN- γ , IL-4, and TGF- β as well as patients with decreased levels of IL-2 and IL-17 (50, 61, 64, 65, 72–75). Interestingly, the correlation between TNF- α and IL-1 β and negative symptoms is only seen in chronic patients and is not reported in FEDN patients (37). Additionally, in FEDN patients, the levels of IL-10 are negatively correlated with negative symptoms, while in chronic patients, they are positively correlated with these symptoms (54, 76).

Increased levels of IL-6, IL-33, sIL-2R, IL-17, and TGF- β are positively correlated with PANSS (positive and negative syndrome scale) general psychopathology sub-score (42, 50, 61, 65). PANSS is a widely used tool to determine the severity of the psychotic symptoms. It is a clinical interview assessing the severity of positive symptoms, negative symptoms, and general psychopathology in schizophrenic patients *via* 30 items. Higher scores indicate more severe conditions (77). The total PANSS score is positively correlated with the levels of IL-6, sIL-2R, IL-1 β , IFN- γ , IL-13, TGF- β 1 and IL-17 (61, 65, 78, 79). Interestingly, the levels of IL-6 and IL-17 correlate with the total score in both chronic and FEDN patients, while the levels of IFN- γ correlate with the total score only in FEDN patients (50). Moreover, only in chronic patients, decreased levels of TNF- α are associated with higher general and total sub-scores, and no association has been reported in FEDN patients (37, 38, 71).

Regarding the correlation between IL-8 and severity of symptoms, Dahan and his colleagues did not find any association between the levels of IL-8 and PANSS sub-scores. Instead, they reported that patients with higher levels of IL-8 had higher scores of the Clinical Global Impression (CGI) severity scale (a subjective assessment tool to determine the severity of the mental illness by clinicians) (65, 80). However, the levels of IL-8 are reported to correlate with the PANSS mean score positively (79).

The levels of cytokines may be relevant to behavior disorders as well. Surprisingly, worse cognitive abilities are associated with higher levels of IL-6, IL-1RA, IL-33, and IL-12 or lower levels of TNF- α in chronic patients, and with lower levels of IL-10 in FEDN patients (31, 38, 64, 81–86). Moreover, patients with higher levels of IL-10 had a higher total score of the RBANS

(Repeatable Battery for the Assessment of Neuropsychological Status) and a worse performance in the attention domain (83, 87). Furthermore, better performance on the memory and intelligence tests has been reported to be associated with higher levels of IL-2 (72). Aggressive behavior is more common among patients with higher levels of IL-17 and IL-10 (54, 61, 76). In addition, in FEDN schizophrenic patients, depressive behaviors are more prevalent among those who have higher levels of IL-4 and TNF- α (79).

Table 2 summarizes the relationship between levels of various cytokines and severity of clinical symptoms.

EFFECT OF ANTIPSYCHOTICS ON THE BASELINE LEVEL OF CYTOKINES

Altered Pro-Inflammatory Cytokines After Antipsychotic Treatment

Shortly (up to 2 months) after treatment with typical or atypical antipsychotics (such as risperidone), the levels of IL-6 and IL-1 β seem to decrease (16, 88, 89). However, in the long term, their levels either rise or do not change compared to the baseline levels (70, 90–92).

The increasing trend of IL-6 and IL-1 β can be explained by antipsychotics (especially atypical antipsychotics) side effects such as metabolic syndrome in the long period (93), as increased levels of IL-6, IL-1 β , TNF- α , IL-2, IFN- γ , and IL-4 have been reported in patients with metabolic syndrome (94, 95).

Unaltered Pro-Inflammatory Cytokines After Antipsychotic Treatment

After using typical or atypical antipsychotics for up to two months, the levels of TNF- α and IL-8 do not change (24, 70, 89, 91, 96). The IL-8 levels seem to remain unchanged even after three months of therapy with risperidone and haloperidol (90). However, one study found that taking typical or atypical antipsychotics or a combination of them in FEDN patients for seven months causes a significant decrease in the level of IL-8 while patients' BMI also increased (97). Furthermore, it has been reported that the levels of TNF- α significantly increase following

TABLE 2 | Relationship between cytokine levels and severity of clinical symptoms.

| Negative symptoms | | Positive symptoms | | Cognitive/intelligence abilities | | Total PANSS/RBANS/CGI score | | Incidence of depression | Incidence of aggressive behavior |
|----------------------|----------------------|----------------------|----------------------|----------------------------------|----------------------|---------------------------------------|--|-------------------------|----------------------------------|
| Positive correlation | Negative correlation | Positive correlation | Negative correlation | Positive correlation | Negative correlation | Positive correlation | Negative correlation | Positive correlation | Positive correlation |
| IL-6 | IL-2 (TH1) | IL-6 | TNF- α | IL-33 | IL-6 | IL-6 | TNF- α (only in chronic patients) | IL-4 | IL-17 |
| TNF- α | IL-17 | IL-1 β | | TNF- α | IL-RA | IL-8 | | | IL-10 |
| IL-8 | IL-10 | IL-33 | | IL-2 | IL-12 | sIL-2R | | | |
| IL-1 β (PI) | | IL-17 | | IL-10 | | IL-1 β | | | |
| IFN- γ (TH-1) | | | | | | IFN- γ (only in FEDN patients) | | | |
| IL-4(TH2) | | | | | | IL-13 | | | |
| TGF- β | | | | | | IL-10 | | | |
| IL-10 | | | | | | TGF- β 1 | | | |
| | | | | | | IL-17 | | | |

IL, interleukin; TNF, tumor necrosis factor; TGF, transforming growth factor; IFN, interferon; PANSS, The Positive and Negative Syndrome Scale; RBANS, The Repeatable Battery for the Assessment of Neuropsychological Status; CGI, Clinical Global Impression.

taking risperidone for more than three months (possibly because of its side effects resulting in induction of metabolic syndrome) (92), or after taking adjunct mood stabilizers with typical or atypical antipsychotics for an average of six weeks of treatment (98). Interestingly, Amisulpride seems to decrease the levels of TNF- α after six weeks of treatment (52).

The level of the anti-inflammatory cytokine, IL-1RA, is reported to decrease following 6 weeks of treatment with olanzapine or risperidone (48) or 8 weeks of antipsychotic therapy adjusted with patients' clinical status (99). Nonetheless, these studies are in contrast with meta-analyses that found no significant alterations in the levels of this cytokine after an average of eight weeks of treatment (70, 91).

Altered T-Helper 1 Cytokines After Antipsychotic Treatment

The levels of IL-2 seem to decrease in the first month after antipsychotic therapy (atypical or typical antipsychotics or mixed) (89). Particularly, olanzapine and haloperidol decrease the level of IL-2 significantly (70). However, studies with longer average treatment periods found no significant changes in the level of IL-2 (24, 70, 91).

Moreover, the levels of IL-12 seem to significantly increase following the use of risperidone for 7–8 weeks (24, 91). Similarly, an elevation in the levels of IL-12 has been reported after six weeks of treatment with olanzapine and haloperidol (100). Surprisingly, aripiprazole (a third-generation antipsychotic) seems to decrease IL-12 levels in chronic patients after four weeks of treatment (101). This is in contrast with the results of a meta-analysis of studies with a treatment period of 4–10 weeks using typical or atypical antipsychotics that found no significant changes in the level of IL-12 after medication (70).

Unaltered T-Helper 1 Cytokine After Antipsychotic Treatment

IFN- γ levels do not significantly change in the first month following antipsychotic treatment (89). However, findings after antipsychotic therapy for an average of approximately 2 months are inconsistent. Two meta-analyses found decreased levels following treatment with typical or atypical or mixed antipsychotics while a meta-analysis by Miller et al., in which more than half of the included studies had non-standardized antipsychotic treatment, suggested the levels of IFN- γ remained constant in this period (16, 70, 91). Olanzapine seems to be the main medication that decreases IFN- γ levels (70). Interestingly, assessing the effect of atypical antipsychotics for 3 months revealed increased levels of IFN- γ , which may be because of their side effects such as metabolic disorder (73).

T-Helper 2 Cytokines After Antipsychotic Treatment

Several meta-analyses have confirmed no disturbances in the level of IL-4 after an average of two months of antipsychotic treatment (70, 91). In contrast, some studies found that treatment with typical or atypical antipsychotics or a combination of them for 1 month or treatment with risperidone for 10 weeks led to

decreased levels of IL-4 (102, 46, 68). Thus, IL-4 might be a trait marker that decreases at first but then increases after a certain time due to the metabolic side effects of antipsychotics, which can result in normal levels in the long term. However, to the best we know, no meta-analysis has evaluated the short-term effects. Furthermore, it has been reported that treatment with atypical antipsychotics for eight weeks causes a significant reduction in the levels of IL-13 (78).

Unaltered T-Helper 17 Cytokines After Antipsychotic Treatment

IL-17 and IL-23 seem to be a trait marker whose level does not change after one month of treatment with typical or atypical antipsychotics or a combination of them compared to the baseline level (47, 63, 89). Assessing the effect of risperidone for 10 weeks has shown the same result (102). Interestingly, one study found that a 4-week treatment course with risperidone decreased the number of T-h17 cells while it had no significant effects on the IL-17 levels (50).

Unaltered T-Regulatory Cytokines After Antipsychotic Treatment

The levels of IL-10 do not change following an average of 8 weeks of treatment compared to the baseline levels (70, 91). However, it has been reported that in chronic patients, treatment with aripiprazole, risperidone, or clozapine for 4–6 weeks increased the IL-10 levels (101, 103). The results of treatment with atypical antipsychotics, particularly risperidone, in FEDN patients are different. In these patients, the levels of IL-10 are lower after treatment compared to baseline. Similarly, an average of eight weeks of treatment does not cause a significant alteration in the level of TGF- β (70, 91). However, a meta-analysis by Miller et al., in which more than half of the studies had non-standardized antipsychotic treatment, found decreased levels of TGF- β after 8 weeks of therapy (24). Four weeks of treatment with aripiprazole in chronic patients and 6 weeks of therapy, mostly with atypical antipsychotics in FEDN patients are associated with decreased levels of TGF- β as well (101, 104). However, elevated levels of TGF- β have been reported after four weeks of typical or atypical or mixed antipsychotic treatment in FEDN patients (46).

THE ROLE OF CYTOKINE LEVEL AS PREDICTORS OF TREATMENT RESPONSE

There is a growing body of evidence on the clinical implications of cytokines in schizophrenia. Considering the lack of predictor biomarkers of the treatment response in psychosis, one of the suggested applications is using cytokines to predict response to treatment (73). Furthermore, the relationship between cytokine levels and response to treatment can support the hypothesis that cytokines may play a role in schizophrenia pathogenesis.

Increased levels of IL-6 and IFN- γ are associated with treatment resistance (105, 73). Treatment resistance is defined by not achieving the remission criteria proposed by the Schizophrenia

Working Group Consensus (106) or Kane's criteria (107). Patients with higher levels of IL-8 and IL-2 experience less improvement in PANSS compared to other patients after twelve weeks of therapy with risperidone or haloperidol (90). Moreover, even though the level of TGF- β is not related to treatment response, TGF- β 1 polymorphism is reported to be associated with PANSS score improvement after antipsychotic treatment (108).

DISCUSSION

Abnormal Cytokine Profiles in Schizophrenic Patients

The results showed that schizophrenic patients had an inflammatory cytokine profile and imbalanced T-helper 1, T-helper 2, and regulatory cytokines. The severity of symptoms and abnormal behaviors in addition to antipsychotic therapies may affect abnormal cytokine levels in these patients. However, there were significant inconsistencies regarding the cytokine profile in the reviewed studies in schizophrenic patients. Six reasons may explain these controversies: 1) Diverse Patients' characteristics; 2) Different sampling methods; 3) Heterogeneous patient populations; 4) The difference between cytokine profile of plasma, serum or whole blood; 5) Different specifications of assay kits; 6) Small sample size.

1. Diverse Patients' characteristics: Several studies did not evaluate factors such as the BMI, diet, diurnal rhythm, smoking habits, psychological stress, and lifestyle in their patients, which may affect the cytokines profile (109–112). However, elevated levels of IL-6 are reported in a study excluding patients with a body mass index (BMI) > 25, which gave rise to the conclusion that the changes of IL-6 did not seem to be related to obesity (44).
2. Different sampling methods: The method and duration of storage and the used anticoagulant may affect the measurement of levels of cytokines. Moreover, as the levels of cytokines are influenced by the circadian pattern, the time of sampling is of great importance. Mornings are suggested to be the best time to take samples (113).
3. Heterogeneous patient populations: In some studies, FEDN patients were not separated from patients with a history of antipsychotic treatment. Not only do antipsychotics affect cytokine levels, but they can also cause weight gain, which is considered a low-grade inflammation (93). Thus, studies in a FEDN population are more reliable.
4. The difference between cytokine profile of plasma, serum or whole blood: The cytokine profile of each of these can be different from the other ones as the coagulation process may trigger release of some inflammatory cytokines (113).
5. Different specifications of assay kits: The quality of antibody used in ELISA kits, kit manufacture, and the operator's skills may affect the measurement (113). For example, Hope et al. indicated that different assay kits could affect the measurement of serum levels of different cytokines. In their study, the IL-6 level was higher than the detection limit of the kit in more than half of the samples (82).

6. Small sample size: A considerable number of the studies had small sample sizes (less than 30 participants in each of their groups) that limited their statistical power (54, 114).

Abnormal Cytokine Profile: Culprit, Consequence, or Simple Association?

The question is whether abnormalities in cytokine levels have a causative role in schizophrenia or are merely associated with the disease? We used the Bradford Hill criteria (115, 116) to answer this question. These criteria assess nine factors, including the strength of association, consistency, specificity, temporality, biological gradient, plausibility, coherence, analogy, and experimental effect to differentiate causality from association. As for the strength of association, the levels of several cytokines (specifically pro-inflammatory cytokines) are found to be significantly disturbed in schizophrenic patients. The second criterion, consistency, is not met in most of the cytokine level alterations. These changes are not specific either and can cause a wide variety of diseases. However, in the modern context, specificity is a less important factor in determining or refusing causality (116). Temporality, as the next criterion, was assessed by only a few studies. In two studies that measured the levels of cytokines before schizophrenia presentation, individuals who developed the disease in the following years had higher baseline levels of IL-6 compared to others (117, 118). This association is strengthened by the study of Khandaker et al. in 2018, finding a strong association between a genetic variant of IL-6 receptor relating to the levels of IL-6 and CRP and development of schizophrenia (119). Moreover, individuals with higher levels of inflammatory markers, including ESR (erythrocyte sedimentation rate) and CRP (C-reactive protein), were more likely to develop schizophrenia (120, 121). The severity of the psychotic symptoms correlated with abnormalities in the levels of cytokines, particularly pro-inflammatory cytokines. So, the biological gradient criterion was met. Regarding plausibility and coherence, there are three suggested major ways by which pro-inflammatory cytokines can contribute to schizophrenia development. First, they can increase kynurenic acid (a metabolite of tryptophan) formation. This metabolite function as an N-Methyl-D-aspartate (NMDA) receptor antagonist, which based on the glutamate hypothesis of schizophrenia (122), might play a causative role in schizophrenia together with decreased glutamatergic signaling. Second, these cytokines increase oxidative stress leading to increased neurodegeneration, which can be seen in schizophrenia. Third, pro-inflammatory cytokines may disturb neurodevelopment (particularly when there is a prenatal inflammation), increasing the risk of psychosis (12, 123). Furthermore, the interplay of cytokines and neurotransmitters may be one of the mechanisms by which cytokines can play a causative role in schizophrenia. Inflammatory cytokines can affect synthesis of monoamine neurotransmitters, increase reuptake of dopamine, serotonin, and norepinephrine, and influence on the release of neurotransmitters (16). Increased levels of soluble IL-6 receptors can be explained as a compensatory response to increased levels of IL-6 (33), and elevated levels of TNF- α mRNA suggest systemic blood immune cells as the main source

of higher levels of TNF- α (36). These two findings provide more experimental evidence for this causation. In terms of analogy, disrupted cytokine levels can be seen in depression and bipolar disorder (114). There is considerable evidence supporting the causal role of cytokines in depression (124). Finally, decreasing inflammation using adjuvant anti-inflammatory agents is reported to improve the symptoms in schizophrenic patients. Consequently, the experimental effect criterion was met as well (125, 126).

CLINICAL IMPLICATIONS

Early Diagnosis

The association between cytokine alterations and schizophrenia may have potential clinical implications. Currently, the diagnosis of schizophrenia is based on clinical symptoms, and there is no standard diagnostic biomarker that allows early recognition of this disease, while cytokines such as IL-6, TNF- α , IL-1 β , and IL-1RA can be used as potential biomarkers for early detection of at least a subgroup of schizophrenic patients (118, 127, 128). Furthermore, IL-6, sIL-2r, TNF- α , IL-1RA, and IL-4t can also be useful in detecting the acute-relapse phase of disease (27, 46).

Novel Therapeutic Horizons

Targeting inflammatory pathways may lead to new treatment options in schizophrenia. A growing body of evidence supports the effect of immune modulators on ameliorating the symptoms of schizophrenia. (12, 129). A recent meta-analysis showed that a variety of medications can reduce the severity of symptoms of schizophrenia. These medications are aspirin (by reducing inflammation by modifying cyclooxygenase-2 enzyme), estrogens (through immunomodulatory effects), minocycline (by inhibiting microglia), and N-acetylcysteine (as an anti-inflammatory agent) (130).

Prediction of Prognosis

Prediction of response to treatment is another application of cytokine levels. Patients with increased levels of IL-6, IL-8, IFN- γ , and soluble TNF- α receptor 1 and decreased levels of IL-2 exhibit less improvement after standard antipsychotic therapy (73, 90, 131). Moreover, patients with higher levels of CRP, an inflammatory marker, tend to have lower quality of life (132).

Patient Stratification for Choosing the Best Therapeutic Protocol

It is possible that patients who have higher levels of pro-inflammatory cytokines benefit more from adding specific drugs to the standard therapeutic regimen. Stratifying patients on this basis can potentially help the physicians choose the best treatment option. A randomized clinical trial study in treatment-resistant depressive patients supports this hypothesis. Depression is also associated with a disrupted cytokine profile. This study showed that in a subgroup of patients who had higher baseline inflammation, the use of infliximab—a TNF- α antagonist—led to better results after the treatment. However,

infliximab had no significant effects on another subgroup of patients (133).

Future Studies: Causation, Diagnosis, Prognostic, and Therapeutic Applications

Although current studies provide a wealth of information on the cytokines profile in schizophrenic patients, several major shortcomings cannot be overlooked. There is a substantial need for longitudinal studies investigating the levels of cytokines before the development of clinical manifestations of schizophrenia in individuals with a strong positive family history of schizophrenia. Moreover, the potential confounding factors such as age, sex, smoking, obesity, and individuals' diet should be rigorously controlled in the future studies (134).

More studies also need to be performed on the diagnostic and the prognostic applications of measurement of levels of cytokines. The relationship between severity of symptoms and levels of cytokines in FEDN patients can be one of the examples of the diagnostic applications. Identifying treatment-resistant patients based on their cytokines profile can be noted as the prognostic applications, on which more studies are needed. Last but not the least, future studies should be based on defined categories of schizophrenic patients.

Lastly, there is a significant shortcoming in the studies that investigate the therapeutic applications. As an example, selection of patients for adjuvant therapy with medications targeting immune modulatory pathways can be guided by inflammatory cytokine levels (135). Moreover, the role of adjuvant monoclonal antibody immunotherapy, which in contrast to NSAIDs only targets immune pathways, needs to be more investigated (134).

LIMITATIONS

This was a narrative review. Therefore, all the limitations of narrative reviews may apply to this study (136).

CONCLUSION

Schizophrenia is associated with several abnormalities in the levels of cytokines, which are the mediators of the immune system. A deeper understanding of this association can be useful in clinical practice in terms of early diagnosis and treatment.

AUTHOR CONTRIBUTIONS

SM developed the concept and design, collected the data, drafted the article, critically revised the manuscript for important intellectual content, and approved the final version. AZ-S critically revised the manuscript for important intellectual content and approved the final version. NR supervised the project, critically revised the manuscript for important intellectual content, and approved the final version.

REFERENCES

1. Saha S, Chant D, Welham J, Mcgrath JJPM. A systematic review of the prevalence of schizophrenia. *PloS Med* (2005) 2:e141. doi: 10.1371/journal.pmed.0020141
2. Pedersen CB, Mors O, Bertelsen A, Waltoft BL, Agerbo E, Mcgrath JJ, et al. A comprehensive nationwide study of the incidence rate and lifetime risk for treated mental disorders. *JAMA Psychiatry* (2014) 71:573–81. doi: 10.1001/jamapsychiatry.2014.16
3. Moreno-Kustner B, Martin C, Pastor L. Prevalence of psychotic disorders and its association with methodological issues. A systematic review and meta-analyses. *PloS One* (2018) 13:e0195687. doi: 10.1371/journal.pone.0195687
4. Pillinger T, D'ambrosio E, Mccutcheon R, Howes OD. Is psychosis a multisystem disorder? A meta-review of central nervous system, immune, cardiometabolic, and endocrine alterations in first-episode psychosis and perspective on potential models. *Mol Psychiatry* 24(6):776–794. doi: 10.1038/s41380-018-0275-2
5. Tamminga C. Schizophrenia and other psychotic disorders: Introduction and overview. In: Sadock VA, Sadock BJ, Ruiz P. Md, editors. *Kaplan and Sadock's Comprehensive Textbook of Psychiatry*, tenth ed. Wolters Kluwer Health. Philadelphia: Lippincott Williams & Wilkins (2017). 3613–7 p.
6. Dinarello CA. Proinflammatory cytokines. *Chest* (2000) 118:503–8. doi: 10.1378/chest.118.2.503
7. Warrington R, Watson W, Kim HL, Antonetti FR. An introduction to immunology and immunopathology. *Allergy Asthma Clin Immunol* (2011) 7(Suppl 1):S1. doi: 10.1186/1710-1492-7-S1-S1
8. Debnath M, Berk M. Functional implications of the IL-23/IL-17 immune axis in Schizophrenia. *Mol Neurobiol* (2017) 54:8170–8. doi: 10.1007/s12035-016-0309-1
9. Heath RG. *Proceedings of the annual meeting of the American Psychopathological Association*. In: *Schizophrenia: evidence of a pathologic immune mechanism*. United States: Baltimore Md: Johns Hopkins University Press (1969). 234–52 p.
10. Delisi L. Is immune dysfunction associated with schizophrenia? A review of the data. *Psychopharmacol Bull* (1984) 20:509–13.
11. Khandaker GM, Dantzer R. Is there a role for immune-to-brain communication in schizophrenia? *Psychopharmacol (Berl)* (2016) 233:1559–73. doi: 10.1007/s00213-015-3975-1
12. Khandaker GM, Cousins L, Deakin J, Lennox BR, Yolken R, Jones PB. Inflammation and immunity in schizophrenia: implications for pathophysiology and treatment. *Lancet Psychiatry* (2015) 2:258–70. doi: 10.1016/S2215-0366(14)00122-9
13. Muller N, Weidinger E, Leitner B, Schwarz MJ. The role of inflammation in schizophrenia. *Front Neurosci* (2015) 9:372. doi: 10.3389/fnins.2015.00372
14. Stefansson H, Ophoff RA, Steinberg S, Andreassen OA, Cichon S, Rujescu D, et al. Common variants conferring risk of schizophrenia. *Nature* (2009) 460:744–7. doi: 10.1038/nature08186
15. Schizophrenia Working Group of the Psychiatric Genomics, C. Biological insights from 108 schizophrenia-associated genetic loci. *Nature* (2014) 511:421–7. doi: 10.1038/nature13595
16. Miller AH, Haroon E, Raison CL, Felger JC. Cytokine targets in the brain: impact on neurotransmitters and neurocircuits. *Depress Anxiety* (2013) 30:297–306. doi: 10.1002/da.22084
17. Altamura AC, Buoli M, Pozzoli S. Role of immunological factors in the pathophysiology and diagnosis of bipolar disorder: comparison with schizophrenia. *Psychiatry Clin Neurosci* (2014) 68:21–36. doi: 10.1111/pcn.12089
18. Hong W, Zhao M, Li H, Peng F, Wang F, Li N, et al. Higher plasma S100B concentrations in schizophrenia patients, and dependently associated with inflammatory markers. *Sci Rep* (2016) 6:27584. doi: 10.1038/srep27584
19. Galic MA, Riazzi K, Pittman QJ. Cytokines and brain excitability. *Front Neuroendocrinol* (2012) 33:116–25. doi: 10.1016/j.yfrne.2011.12.002
20. Shechter R, London A, Schwartz M. Orchestrated leukocyte recruitment to immune-privileged sites: absolute barriers versus educational gates. *Nat Rev Immunol* (2013) 13:206–18. doi: 10.1038/nri3391
21. Louveau A, Plog BA, Antila S, Alitalo K, Nedergaard M, Kipnis J. Understanding the functions and relationships of the lymphatic system and meningeal lymphatics. *J Clin Invest* (2017) 127:3210–9. doi: 10.1172/JCI90603
22. Libikowa H, Stancek D, Wiedermann V, Hasto J, Breier S. Psychopharmacology and electroconvulsive therapy in relation to viral antibodies and interferon. Experimental and clinical study. *Arch Immunol Ther Exp (Warsz)* (1977) 25:641–9.
23. Smith RJMH. A comprehensive macrophage-T-lymphocyte theory of schizophrenia. *Med Hypotheses* (1992) 39:248–57. doi: 10.1016/0306-9877(92)90117-U
24. Miller BJ, Buckley P, Seabolt W, Mellor A, Kirkpatrick BJBP. Meta-analysis of cytokine alterations in schizophrenia: clinical status and antipsychotic effects. *Biol Psychiatry* (2011) 70:663–71. doi: 10.1016/j.biopsych.2011.04.013
25. Upthegrove R, Manzanares-Teson N, Barnes NMJSR. Cytokine function in medication-naïve first episode psychosis: a systematic review and meta-analysis. *Schizophr Res* (2014) 155:101–8. doi: 10.1016/j.schres.2014.03.005
26. Falcone T, Carlton E, Lee C, Janigro M, Fazio V, Forcen FE, et al. Does systemic inflammation play a role in pediatric psychosis? *Clin Schizophr Relat Psychoses* (2015) 9:65–78B. doi: 10.3371/CSRP.FACA.030813
27. Goldsmith D, Rapaport M, Miller BJMP. A meta-analysis of blood cytokine network alterations in psychiatric patients: comparisons between schizophrenia, bipolar disorder and depression. *Mol Psychiatry* (2016) 21:1696. doi: 10.1038/mp.2016.3
28. Lesh TA, Careaga M, Rose DR, Mcallister AK, Van De Water J, Carter CS, et al. Cytokine alterations in first-episode schizophrenia and bipolar disorder: relationships to brain structure and symptoms. *J Neuroinflammation* (2018) 15:165. doi: 10.1186/s12974-018-1197-2
29. Al-Hakeim HK, Al-Rammahi DA, Al-Dujaili AH. IL-6, IL-18, sIL-2R, and TNF α proinflammatory markers in depression and schizophrenia patients who are free of overt inflammation. *J Affect Disord* (2015) 182:106–14. doi: 10.1016/j.jad.2015.04.044
30. Potvin S, Stip E, Sepehry AA, Gendron A, Bah R, Kouassi EJPB. Inflammatory cytokine alterations in schizophrenia: a systematic quantitative review. *Biol Psychiatry* (2008) 63:801–8. doi: 10.1016/j.biopsych.2007.09.024
31. Frydecka D, Misiak B, Pawlak-Adamska E, Karabon L, Tomkiewicz A, Sedlaczek P, et al. Interleukin-6: the missing element of the neurocognitive deterioration in schizophrenia? The focus on genetic underpinnings, cognitive impairment and clinical manifestation. *Eur Arch Psychiatry Clin Neurosci* (2015) 265:449–59. doi: 10.1007/s00406-014-0533-5
32. Frydecka D, Krzystek-Korpacka M, Lubeiro A, Stramecki F, Stanczykiewicz B, Beszlej JA, et al. Profiling inflammatory signatures of schizophrenia: a cross-sectional and meta-analysis study. *Brain Behav Immun* (2018) 71:28–36. doi: 10.1016/j.bbi.2018.05.002
33. Hartwig FP, Borges MC, Horta BL, Bowden J, Smith GDJJP. Inflammatory biomarkers and risk of schizophrenia: a 2-sample mendelian randomization study. *JAMA Psychiatry* (2017) 74:1226–33. doi: 10.1001/jamapsychiatry.2017.3191
34. Hope S, Melle I, Aukrust P, Steen NE, Birkenaes AB, Lorentzen S, et al. Similar immune profile in bipolar disorder and schizophrenia: selective increase in soluble tumor necrosis factor receptor I and von Willebrand factor. *Bipolar Disord* (2009) 11:726–34. doi: 10.1111/j.1399-5618.2009.00757.x
35. Wei L, Du Y, Wu W, Fu X, Xia QJJOad. Elevation of plasma neutrophil gelatinase-associated lipocalin (NGAL) levels in schizophrenia patients. *J Affect Disord* (2018) 226:307–12. doi: 10.1016/j.jad.2017.10.002
36. Di Nicola M, Cattaneo A, Heggul N, Di Forti M, Aitchison KJ, Janiri L, et al. Serum and gene expression profile of cytokines in first-episode psychosis. *Brain Behav Immun* (2013) 31:90–5. doi: 10.1016/j.bbi.2012.06.010
37. Zhu F, Zhang L, Liu F, Wu R, Guo W, Ou J, et al. Altered serum tumor necrosis factor and interleukin-1 β in first-episode drug-naïve and chronic schizophrenia. *Front Neurosci* (2018) 12:296. doi: 10.3389/fnins.2018.00296
38. Lv MH, Tan YL, Yan SX, Tian L, Tan SP, Wang ZR, et al. Decreased serum TNF-alpha levels in chronic schizophrenia patients on long-term antipsychotics: correlation with psychopathology and cognition. *Psychopharmacol (Berl)* (2015) 232:165–72. doi: 10.1007/s00213-014-3650-y
39. Turhan L, Batmaz S, Kocbiyik S, Soygun AHJNJOP. The role of tumour necrosis factor alpha and soluble tumour necrosis factor alpha receptors in

- the symptomatology of schizophrenia. *Nord J Psychiatry* (2016) 70:342–50. doi: 10.3109/08039488.2015.1122079
40. Lin Y, Peng Y, He S, Xu J, Shi Y, Su Y, et al. Serum IL-1ra, a novel biomarker predicting olanzapine-induced hypercholesterolemia and hyperleptinemia in schizophrenia. *Prog Neuropsychopharmacol Biol Psychiatry* (2018) 84:71–8. doi: 10.1016/j.pnpbp.2018.01.020
41. Balóššev R, Koido K, Vasar V, Janno S, Kriisa K, Mahlapuu R, et al. Inflammatory, cardio-metabolic and diabetic profiling of chronic schizophrenia. *Eur Psychiatry* (2017) 39:1–10. doi: 10.1016/j.eurpsy.2016.05.010
42. Borovcanin MM, Janicijevic SM, Jovanovic IP, Gajovic N, Arsenijevic NN, Lukic ML. IL-33/ST2 Pathway and Galectin-3 as a new analytes in pathogenesis and cardiometabolic risk evaluation in psychosis. *Front Psychiatry* (2018) 9:271. doi: 10.3389/fpsy.2018.00271
43. Zhou Y, Peng W, Wang J, Zhou W, Zhou Y, Ying BJP, et al. Plasma levels of IL-1Ra is associated with schizophrenia. *Psychiatry Clin. Neurosci.* (2018) 73:109–15. doi: 10.1111/pcn.12794
44. Petrikis P, Voulgari PV, Tzallas AT, Archimandriti DT, Skapinakis P, Mavreas VJOPR. Cytokine profile in drug-naïve, first episode patients with psychosis. *J Psychosom Res* (2015) 79:324–7. doi: 10.1016/j.jpsychores.2015.06.011
45. Boerrigter D, Weickert TW, Lenroot R, O'donnell M, Galletly C, Liu D, et al. Using blood cytokine measures to define high inflammatory biotype of schizophrenia and schizoaffective disorder. *J Neuroinflammation* (2017) 14:188. doi: 10.1186/s12974-017-0962-y
46. Borovcanin M, Jovanovic I, Radosavljevic G, Dejanovic SD, Bankovic D, Arsenijevic N, et al. Elevated serum level of type-2 cytokine and low IL-17 in first episode psychosis and schizophrenia in relapse. *J Psychiatr Res* (2012) 46:1421–6. doi: 10.1016/j.jpsychores.2012.08.016
47. Borovcanin M, Jovanovic I, Radosavljevic G, Dejanovic SD, Stefanovic V, Arsenijevic N, et al. Antipsychotics can modulate the cytokine profile in schizophrenia: attenuation of the type-2 inflammatory response. *Schizophr Res* (2013) 147:103–9. doi: 10.1016/j.schres.2013.03.027
48. De Witte L, Tomasik J, Schwarz E, Guest PC, Rahmoune H, Kahn RS, et al. Cytokine alterations in first-episode schizophrenia patients before and after antipsychotic treatment. *Schizophr Res* (2014) 154:23–9. doi: 10.1016/j.schres.2014.02.005
49. Noto C, Ota VK, Santoro ML, Ortiz BB, Rizzo LB, Higuchi CH, et al. Effects of depression on the cytokine profile in drug naïve first-episode psychosis. *Schizophr Res* (2015) 164:53–8. doi: 10.1016/j.schres.2015.01.026
50. Ding M, Song X, Zhao J, Gao J, Li X, Yang G, et al. Activation of Th17 cells in drug naïve, first episode schizophrenia. *Prog Neuropsychopharmacol Biol Psychiatry* (2014) 51:78–82. doi: 10.1016/j.pnpbp.2014.01.001
51. Reale M, Patruno A, De Lutiis MA, Pesce M, Felaco M, Di Giannantonio M, et al. Dysregulation of chemo-cytokine production in schizophrenic patients versus healthy controls. *BMC Neurosci* (2011) 12:13. doi: 10.1186/1471-2202-12-13
52. Na K-S, Kim Y-KJN. Monocytic, Th1 and th2 cytokine alterations in the pathophysiology of schizophrenia. *Neuropsychobiology* (2007) 56:55–63. doi: 10.1159/000111535
53. Al-Asmari A, Khan MWJH, Toxicology E. Inflammation and schizophrenia: alterations in cytokine levels and perturbation in antioxidative defense systems. *Hum Exp Toxicol* (2014) 33:115–22. doi: 10.1177/0960327113493305
54. Das S, Deuri SK, Sarmah A, Pathak K, Baruah A, Sengupta S, et al. Aggression as an independent entity even in psychosis-the role of inflammatory cytokines. *J Neuroimmunol* (2016) 292:45–51. doi: 10.1016/j.jneuroim.2016.01.012
55. Bedrossian N, Haidar M, Fares J, Kobeissy FH, Fares YJFIMN. Inflammation and elevation of interleukin-12p40 in patients with schizophrenia. *Front Mol Neurosci* (2016) 9:16. doi: 10.3389/fnmol.2016.00016
56. Kartalci S, Erbay LGJTPD. IL-4, TGF- β , NF- κ B and MPO levels in patients with treatment resistant schizophrenia. *Turk Psikiyatri Derg* (2016) 27:170. doi: 10.5080/u13642
57. Noto C, Maes M, Ota VK, Teixeira AL, Bressan RA, Gadelha A, et al. High predictive value of immune-inflammatory biomarkers for schizophrenia diagnosis and association with treatment resistance. *World J Biol Psychiatry* (2015) 16:422–9. doi: 10.3109/15622975.2015.1062552
58. Eftekharian MM, Omrani MD, Arsang-Jang S, Taheri M, Ghafouri-Fard SJHA. Serum cytokine profile in schizophrenic patients. *Hum Antibodies* (2018) 27(1):1–7. doi: 10.3233/HAB-180344
59. Maxeiner H-G, Schneider EM, Kurfiss S-T, Brettschneider J, Tumani H, Bechter KJC. Cerebrospinal fluid and serum cytokine profiling to detect immune control of infectious and inflammatory neurological and psychiatric diseases. *Cytokine* (2014) 69:62–7. doi: 10.1016/j.cyto.2014.05.008
60. Fang X, Zhang Y, Fan W, Tang W, Zhang CJMN. Interleukin-17 alteration in first-episode psychosis: a meta-analysis. *Mol Neuropsychiatry* (2018) 3:135–40. doi: 10.1159/000481661
61. Li H, Zhang Q, Li N, Wang F, Xiang H, Zhang Z, et al. Plasma levels of Th17-related cytokines and complement C3 correlated with aggressive behavior in patients with schizophrenia. *Psychiatry Res* (2016) 246:700–6. doi: 10.1016/j.psychres.2016.10.061
62. Dimitrov DH, Lee S, Yantis J, Valdez C, Paredes RM, Braidia N, et al. Differential correlations between inflammatory cytokines and psychopathology in veterans with schizophrenia: potential role for IL-17 pathway. *Schizophr Res* (2013) 151:29–35. doi: 10.1016/j.schres.2013.10.019
63. Borovcanin M, Jovanovic I, Dejanovic SD, Radosavljevic G, Arsenijevic N, Lukic ML. Increase systemic levels of IL-23 as a possible constitutive marker in schizophrenia. *Psychoneuroendocrinology* (2015) 56:143–7. doi: 10.1016/j.psyneuen.2015.03.003
64. Xiu MH, Yang GG, Tan YL, Tan SP, Wang ZR, De Yang F, et al. Decreased interleukin-10 serum levels in first-episode drug-naïve schizophrenia: relationship to psychopathology. *Schizophr Res* (2014) 156:9–14. doi: 10.1016/j.schres.2014.03.024
65. Dahan S, Bragazzi NL, Yogev A, Bar-Gad M, Barak V, Amital H, et al. The relationship between serum cytokine levels and degree of psychosis in patients with schizophrenia. *Psychiatry Res* (2018) 268:467–72. doi: 10.1016/j.psychres.2018.07.041
66. Noto MN, Maes M, Nunes SOV, Ota VK, Rossaneis AC, Verri WAJr., et al. Activation of the immune-inflammatory response system and the compensatory immune-regulatory system in antipsychotic naïve first episode psychosis. *Eur Neuropsychopharmacol* (2019) 29:416–31. doi: 10.1016/j.euroneuro.2018.12.008
67. Fu G, Zhang W, Dai J, Liu J, Li F, Wu D, et al. Increased peripheral interleukin 10 Relate to white matter integrity in Schizophrenia. *Front Neurosci* (2019) 13:52. doi: 10.3389/fnins.2019.00052
68. Szymona K, Zdzisinska B, Karakula-Juchnowicz H, Kocki T, Kandefers-Szerszen M, Flis M, et al. Correlations of Kynurenic Acid, 3-Hydroxykynurenine, sIL-2R, IFN- α , and IL-4 with clinical symptoms during acute relapse of schizophrenia. *Neurotox Res* (2017) 32:17–26. doi: 10.1007/s12640-017-9714-0
69. Zhang XY, Zhou DF, Zhang PY, Wu GY, Cao LY, Shen YCJSR. Elevated interleukin-2, interleukin-6 and interleukin-8 serum levels in neuroleptic-free schizophrenia: association with psychopathology. *Schizophr Res* (2002) 57:247–58. doi: 10.1016/S0920-9964(01)00296-1
70. Romeo B, Brunet-Lecomte M, Martelli C, Benyamina AJIJON. Kinetics of cytokine levels during antipsychotic treatment in schizophrenia: a meta-analysis. *Int J Neuropsychopharmacol* (2018) 21:828–36. doi: 10.1093/ijnp/pyy062
71. Tian L, Tan Y, Chen D, Lv M, Tan S, Soares JC, et al. Reduced serum TNF alpha level in chronic schizophrenia patients with or without tardive dyskinesia. *Prog Neuropsychopharmacol Biol Psychiatry* (2014) 54:259–64. doi: 10.1016/j.pnpbp.2014.06.012
72. Asevedo E, Rizzo LB, Gadelha A, Mansur RB, Ota VK, Berberian AA, et al. Peripheral interleukin-2 level is associated with negative symptoms and cognitive performance in schizophrenia. *Physiol Behav* (2014) 129:194–8. doi: 10.1016/j.physbeh.2014.02.032
73. Mondelli V, Ciufolini S, Belvederi Murri M, Bonaccorso S, Di Forti M, Giordano A, et al. Cortisol and inflammatory biomarkers predict poor treatment response in first episode psychosis. *Schizophr Bull* (2015) 41:1162–70. doi: 10.1093/schbul/sbv028
74. Simsek S, Yildirim V, Çim A, Kaya SJJOC, Psychopharmacology A. Serum IL-4 and IL-10 levels correlate with the symptoms of the drug-naïve adolescents with first episode, early onset schizophrenia. *J Child Adolesc Psychopharmacol* (2016) 26:721–6. doi: 10.1089/cap.2015.0220
75. Goldsmith DR, Haroon E, Miller AH, Strauss GP, Buckley PF, Miller BJ. TNF-alpha and IL-6 are associated with the deficit syndrome and negative

- symptoms in patients with chronic schizophrenia. *Schizophr Res* (2018) 199:281–4. doi: 10.1016/j.schres.2018.02.048
76. Zhang Q, Hong W, Li H, Peng F, Wang F, Li N, et al. Increased ratio of high sensitivity C-reactive protein to interleukin-10 as a potential peripheral biomarker of schizophrenia and aggression. *Int J Psychophysiol* (2017) 114:9–15. doi: 10.1016/j.ijpsycho.2017.02.001
77. Kay SR, Fiszbein A, Opler LA. The positive and negative syndrome scale (PANSS) for schizophrenia. *Schizophr Bull* (1987) 13:261–76. doi: 10.1093/schbul/13.2.261
78. Pae C-U, Yoon C-H, Kim T-S, Kim J-J, Park S-H, Lee C-U, et al. Antipsychotic treatment may alter T-helper (TH) 2 arm cytokines. *Int Immunopharmacol* (2006) 6:666–71. doi: 10.1016/j.intimp.2005.10.004
79. Noto CS, Gadelha A, Belangero SI, Smith MaC, De Aguiar BW, Panizzuti B, et al. Association of biomarkers and depressive symptoms in schizophrenia. *Neurosci Lett* (2011) 505:282–5. doi: 10.1016/j.neulet.2011.10.042
80. Busner J, Targum SD. The clinical global impressions scale: applying a research tool in clinical practice. *Psychiatry (Edmont)* (2007) 4:28–37.
81. Hope S, Ueland T, Steen NE, Dieset I, Lorentzen S, Berg AO, et al. Interleukin 1 receptor antagonist and soluble tumor necrosis factor receptor 1 are associated with general severity and psychotic symptoms in schizophrenia and bipolar disorder. *Schizophr Res* (2013) 145:36–42. doi: 10.1016/j.schres.2012.12.023
82. Hope S, Hoseth E, Dieset I, Mørch RH, Aas M, Aukrust P, et al. Inflammatory markers are associated with general cognitive abilities in schizophrenia and bipolar disorder patients and healthy controls. *Schizophr Res* (2015) 165:188–94. doi: 10.1016/j.schres.2015.04.004
83. Xiu MH, Tian L, Chen S, Tan YL, Chen J, Chen N, et al. Contribution of IL-10 and its-592 A/C polymorphism to cognitive functions in first-episode drug-naïve schizophrenia. *Brain Behav Immun* (2016) 57:116–24. doi: 10.1016/j.bbi.2016.03.005
84. De Campos-Carli SM, Miranda AS, Dias IC, De Oliveira A, Cruz BF, Vieira EL, et al. Serum levels of interleukin-33 and its soluble form receptor (sST2) are associated with cognitive performance in patients with schizophrenia. *Compr Psychiatry* (2017) 74:96–101. doi: 10.1016/j.comppsy.2017.01.008
85. Kogan S, Ospina LH, Kimhy DJB. Behavior, and Immunity. Inflammation in individuals with schizophrenia-implications for neurocognition and daily function. *Brain Behav Immun* (2018) 74:296–9. doi: 10.1016/j.bbi.2018.09.016
86. Misiak B, Stanczykiewicz B, Kotowicz K, Rybakowski JK, Samochowiec J, Frydecka D. Cytokines and C-reactive protein alterations with respect to cognitive impairment in schizophrenia and bipolar disorder: a systematic review. *Schizophr Res* (2018) 192:16–29. doi: 10.1016/j.schres.2017.04.015
87. Randolph C, Tierney MC, Mohr E, Chase TN. The repeatable battery for the assessment of neuropsychological status (RBANS): preliminary clinical validity. *J Clin Exp Neuropsychol* (1998) 20:310–9. doi: 10.1076/jcen.20.3.310.823
88. Song XQ, Lv LX, Li WQ, Hao YH, Zhao JP. The interaction of nuclear factor-kappa B and cytokines is associated with schizophrenia. *Biol Psychiatry* (2009) 65:481–8. doi: 10.1016/j.biopsych.2008.10.018
89. Capuzzi E, Bartoli F, Crocamo C, Clerici M, Carrà GJN, Reviews B. Acute variations of cytokine levels after antipsychotic treatment in drug-naïve subjects with a first-episode psychosis: a meta-analysis. *Neurosci Biobehav Rev* (2017) 77:122–8. doi: 10.1016/j.neubiorev.2017.03.003
90. Zhang XY, Zhou DF, Cao LY, Zhang PY, Wu GY, Shen YCJTJOC. Changes in serum interleukin-2, -6, and -8 levels before and during treatment with risperidone and haloperidol: relationship to outcome in schizophrenia. *J Clin Psychiatry* (2004) 65:940–7. doi: 10.4088/JCP.v65n0710
91. Tourjman V, Kouassi E, Koué M-E, Rocchetti M, Fortin-Fournier S, Fusar-Poli P, et al. Antipsychotics' effects on blood levels of cytokines in schizophrenia: a meta-analysis. *Schizophr Res* (2013) 151:43–7. doi: 10.1016/j.schres.2013.10.011
92. Song X, Fan X, Li X, Zhang W, Gao J, Zhao J, et al. Changes in pro-inflammatory cytokines and body weight during 6-month risperidone treatment in drug naïve, first-episode schizophrenia. *Psychopharmacol (Berl)* (2014) 231:319–25. doi: 10.1007/s00213-013-3382-4
93. Dikec G, Arabaci LB, Uzunoglu GB, Mizrak SD. Metabolic side effects in patients using atypical antipsychotic medications during hospitalization. *J Psychosoc Nurs Ment Health Serv* (2018) 56:28–37. doi: 10.3928/02793695-20180108-05
94. Mirhafez SR, Pasdar A, Avan A, Esmaily H, Moezzi A, Mohebbati M, et al. Cytokine and growth factor profiling in patients with the metabolic syndrome. *Br J Nutr* (2015) 113:1911–9. doi: 10.1017/S0007114515001038
95. Srikanthan K, Feyh A, Visweshwar H, Shapiro JJ, Sodhi K. Systematic review of metabolic syndrome biomarkers: a panel for early detection, management, and risk stratification in the west virginian population. *Int J Med Sci* (2016) 13:25–38. doi: 10.7150/ijms.13800
96. Kubistova A, Horacek J, Novak TJA. Increased interleukin-6 and tumor necrosis factor alpha in first episode schizophrenia patients versus healthy controls. *Psychiatr Danub* (2012) 1:P2.
97. Haring L, Koido K, Vasar V, Leping V, Zilmer K, Zilmer M, et al. Antipsychotic treatment reduces psychotic symptoms and markers of low-grade inflammation in first episode psychosis patients, but increases their body mass index. *Schizophr Res* (2015) 169:22–9. doi: 10.1016/j.schres.2015.08.027
98. Dunjic-Kostic B, Jasovic-Gasic M, Ivkovic M, Radonjic NV, Pantovic M, Damjanovic A, et al. Serum levels of interleukin-6 and tumor necrosis factor-alpha in exacerbation and remission phase of schizophrenia. *Psychiatr Danub* (2013) 25:0–61.
99. Chu C-S, Li D-J, Chu C-L, Wu C-C, Lu TJPI. Decreased IL-1ra and NCAM-1/CD56 serum levels in unmedicated patients with schizophrenia before and after antipsychotic treatment. *Psychiatry Invest* (2018) 15(7):727–732. doi: 10.30773/pi.2017.11.10
100. Crespo-Facorro B, Carrasco-Marin E, Perez-Iglesias R, Pelayo-Teran JM, Fernandez-Prieto L, Leyva-Cobian F, et al. Interleukin-12 plasma levels in drug-naïve patients with a first episode of psychosis: effects of antipsychotic drugs. *Psychiatry Res* (2008) 158:206–16. doi: 10.1016/j.psychres.2006.08.005
101. Sobis J, Rykaczewska-Czerwinska M, Swietochowska E, Gorczyca PJPR. Therapeutic effect of aripiprazole in chronic schizophrenia is accompanied by anti-inflammatory activity. *Pharmacol Rep* (2015) 67:353–9. doi: 10.1016/j.pharep.2014.09.007
102. Noto C, Ota VK, Gouvea ES, Rizzo LB, Spindola L, Honda PH, et al. Effects of risperidone on cytokine profile in drug-naïve first-episode psychosis. *Int J Neuropsychopharmacol* (2014) 18(4):pyu042. doi: 10.1093/ijnp/pyu042
103. Ajami A, Abedian F, Hosseini SH, Akbarian E, Alizadeh-Navaei R, Taghipour MJIOI. Serum TNF- α , IL-10 and IL-2 in schizophrenic patients before and after treatment with risperidone and clozapine. *Iran J Immunol* (2014) 11:200–9. doi: 10.1016/j.ijiv.2013.06.006
104. Petrikis P, Voulgari PV, Tzallas AT, Boumba VA, Archimandriti DT, Zambetas D, et al. Changes in the cytokine profile in first-episode, drug-naïve patients with psychosis after short-term antipsychotic treatment. *Psychiatry Res* (2017) 256:378–83. doi: 10.1016/j.psychres.2017.07.002
105. Lin A, Kenis G, Bignotti S, Tura GJB, De Jong R, Bosmans E, et al. The inflammatory response system in treatment-resistant schizophrenia: increased serum interleukin-6. *Schizophr Res* (1998) 32:9–15. doi: 10.1016/S0920-9964(98)00034-6
106. Andreasen NC, Carpenter WJ Jr., Kane JM, Lasser RA, Marder SR, Weinberger DR. Remission in schizophrenia: proposed criteria and rationale for consensus. *Am J Psychiatry* (2005) 162:441–9. doi: 10.1176/appi.ajp.162.3.441
107. Kane J, Honigfeld G, Singer J, Meltzer H. Clozapine for the treatment-resistant schizophrenic. A double-blind comparison with chlorpromazine. *Arch Gen Psychiatry* (1988) 45:789–96. doi: 10.1001/archpsyc.1988.01800330013001
108. Lee H-Y, Kim Y-KJaN. Effect of TGF- β 1 polymorphism on the susceptibility to schizophrenia and treatment response to atypical antipsychotic agent. *Acta Neuropsychiatr* (2010) 22:174–9. doi: 10.1111/j.1601-5215.2009.00435.x
109. Rodrigues FMM, Ramos D, Xavier RF, Ito JT, De Souza AP, Fernandes RA, et al. Nasal and systemic inflammatory profile after short term smoking cessation. *Respir Med* (2014) 108:999–1006. doi: 10.1016/j.rmed.2014.04.020
110. Aziz N. Measurement of circulating cytokines and immune-activation markers by multiplex technology in the clinical setting: what are we really measuring? *For Immunopathol Dis Therap* (2015) 6:19–22. doi: 10.1615/ForumImmunDisTher.2015014162
111. Cox AJ, West NP, Cripps AWJTL. Endocrinology. Obesity, inflammation, and the gut microbiota. *Lancet Diabetes Endocrinol* (2015) 3:207–15. doi: 10.1016/S2213-8587(14)70134-2
112. Marsland AL, Walsh C, Lockwood K, John-Henderson NaJB. Behavior and Immunity. The effects of acute psychological stress on circulating and

- stimulated inflammatory markers: a systematic review and meta-analysis. *Brain Behav Immun* (2017) 64:208–19. doi: 10.1016/j.bbi.2017.01.011
113. Zhou X, Fragala MS, Mcelhaney JE, Kuchel GA. Conceptual and methodological issues relevant to cytokine and inflammatory marker measurements in clinical research. *Curr Opin Clin Nutr Metab Care* (2010) 13:541–7. doi: 10.1097/MCO.0b013e32833cf3bc
114. Manu P, Correll CU, Wampers M, Mitchell AJ, Probst M, Vancampfort D, et al. Markers of inflammation in schizophrenia: association vs. causation. *World Psychiatry* (2014) 13:189–92. doi: 10.1002/wps.20117
115. Hill AB. The environment and disease: association or causation? *Proc R Soc Med* (1965) 58:295–300. doi: 10.1177/003591576505800503
116. Fedak KM, Bernal A, Capshaw ZA, Gross S. Applying the Bradford Hill criteria in the 21st century: how data integration has changed causal inference in molecular epidemiology. *Emerg Themes Epidemiol* (2015) 12:14. doi: 10.1186/s12982-015-0037-4
117. Khandaker GM, Pearson RM, Zammit S, Lewis G, Jones PB. Association of serum interleukin 6 and C-reactive protein in childhood with depression and psychosis in young adult life: a population-based longitudinal study. *JAMA Psychiatry* (2014) 71:1121–8. doi: 10.1001/jamapsychiatry.2014.1332
118. Stojanovic A, Martorell L, Montalvo I, Ortega L, Monseny R, Vilella E, et al. Increased serum interleukin-6 levels in early stages of psychosis: associations with at-risk mental states and the severity of psychotic symptoms. *Psychoneuroendocrinology* (2014) 41:23–32. doi: 10.1016/j.psyneuen.2013.12.005
119. Khandaker GM, Zammit S, Burgess S, Lewis G, Jones PB. Association between a functional interleukin 6 receptor genetic variant and risk of depression and psychosis in a population-based birth cohort. *Brain Behav Immun* (2018) 69:264–72. doi: 10.1016/j.bbi.2017.11.020
120. Metcalf SA, Jones PB, Nordstrom T, Timonen M, Maki P, Miettinen J, et al. Serum C-reactive protein in adolescence and risk of schizophrenia in adulthood: a prospective birth cohort study. *Brain Behav Immun* (2017) 59:253–9. doi: 10.1016/j.bbi.2016.09.008
121. Kappelmann N, Khandaker GM, Dal H, Stochl J, Kosidou K, Jones PB, et al. Systemic inflammation and intelligence in early adulthood and subsequent risk of schizophrenia and other non-affective psychoses: a longitudinal cohort and co-relative study. *Psychol Med* (2019) 49:295–302. doi: 10.1017/S0033291718000831
122. Hu W, Macdonald ML, Elswick DE, Sweet RA. The glutamate hypothesis of schizophrenia: evidence from human brain tissue studies. *Ann N Y Acad Sci* (2015) 1338:38–57. doi: 10.1111/nyas.12547
123. Ratnayake U, Quinn T, Walker DW, Dickinson H. Cytokines and the neurodevelopmental basis of mental illness. *Front Neurosci* (2013) 7:180. doi: 10.3389/fnins.2013.00180
124. Felger JC, Lotrich FE. Inflammatory cytokines in depression: neurobiological mechanisms and therapeutic implications. *Neuroscience* (2013) 246:199–229. doi: 10.1016/j.neuroscience.2013.04.060
125. Sommer IE, Van Westrhenen R, Begemann MJ, De Witte LD, Leucht S, Kahn RS. Efficacy of anti-inflammatory agents to improve symptoms in patients with schizophrenia: an update. *Schizophr Bull* (2014) 40:181–91. doi: 10.1093/schbul/sbt139
126. Cho M, Lee TY, Kwak YB, Yoon YB, Kim M, Kwon JS. Adjunctive use of anti-inflammatory drugs for schizophrenia: a meta-analytic investigation of randomized controlled trials. *Aust N Z J Psychiatry* (2019) 53(8):742–759. doi: 10.1177/0004867419835028
127. Dubois T, Reynaert C, Jacques D, Lepiece B, Patigny P, Zdanowicz N. Immunity and psychiatric disorders: variabilities of immunity biomarkers are they specific? *Psychiatr Danub* (2018) 30:447–51.
128. Herron JW, Nerurkar L, Cavanagh J. Neuroimmune biomarkers in mental illness. In: Pratt J, Hall J (eds), Springer, Cham: Biomarkers in Psychiatry *Current Topics in Behavioral Neurosciences* (2018). vol 40. doi: 10.1007/7854_2018_45
129. Muller N. Inflammation in schizophrenia: pathogenetic aspects and therapeutic considerations. *Schizophr Bull* (2018) 44:973–82. doi: 10.1093/schbul/sby024
130. Cakici N, Van Beveren NJM, Judge-Hundal G, Koola MM, Sommer IEC. An update on the efficacy of anti-inflammatory agents for patients with schizophrenia: a meta-analysis. *Psychol Med* (2019) 49:2307–19. doi: 10.1017/S0033291719001995
131. Nishimori S, Ohnuma T, Takebayashi Y, Katsuta N, Takeda M, Nakamura T, et al. High serum soluble tumor necrosis factor receptor 1 predicts poor treatment response in acute-stage schizophrenia. *Prog Neuropsychopharmacol Biol Psychiatry* (2017) 76:145–54. doi: 10.1016/j.pnpbp.2017.03.006
132. Faugere M, Micoulaud-Franchi JA, Alessandrini M, Richieri R, Faget-Agius C, Auquier P, et al. Quality of life is associated with chronic inflammation in schizophrenia: a cross-sectional study. *Sci Rep* (2015) 5:10793. doi: 10.1038/srep10793
133. Raison CL, Rutherford RE, Woolwine BJ, Shuo C, Schettler P, Drake DF, et al. A randomized controlled trial of the tumor necrosis factor antagonist infliximab for treatment-resistant depression: the role of baseline inflammatory biomarkers. *JAMA Psychiatry* (2013) 70:31–41. doi: 10.1001/2013.jamapsychiatry.4
134. Miller BJ, Goldsmith DR. Towards an immunophenotype of schizophrenia: progress, potential mechanisms, and future directions. *Neuropsychopharmacology* (2017) 42:299–317. doi: 10.1038/npp.2016.211
135. Uptegrove R, Khandaker GM. Cytokines, oxidative stress and cellular markers of inflammation in schizophrenia. In: *Current topics in behavioral neurosciences*. Berlin, Heidelberg: Springer (2019). doi: 10.1007/7854_2018_88
136. Green BN, Johnson CD, Adams A. Writing narrative literature reviews for peer-reviewed journals: secrets of the trade. *J Chiropr Med* (2006) 5:101–17. doi: 10.1016/S0899-3467(07)60142-6

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Cytokines in Relation to Motor Activity in an Acute Psychiatric Population

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OPEN ACCESS

Edited by:

Francisco Navarrete Rueda,
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Reviewed by:

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Specialty section:

This article was submitted to
Molecular Psychiatry,
a section of the journal
Frontiers in Psychiatry

Received: 25 September 2019

Accepted: 19 November 2019

Published: 19 December 2019

Citation:

Larsen JB, Stunes AK, Iversen VC,
Vaaler AE and Reitan SK (2019)
Cytokines in Relation to Motor Activity
in an Acute Psychiatric Population.
Front. Psychiatry 10:920.
doi: 10.3389/fpsy.2019.00920

Background: Deviations in motor activity are important clinical features of several psychiatric disorders in an acute state. Immune activity is associated with several psychiatric disorders and may affect motor activity. We aimed to examine the association between immune activity measured as serum levels of cytokines and deviations in motor activity, in an acute psychiatric setting.

Methods: Data on motor activity and immune markers were available on 277 patients admitted to an acute psychiatric inpatient department. The degree of increased or decreased motor activity was clinically assessed at admission. Serum concentrations of the following immune markers were measured: interleukin (IL) -1 β , IL-4, IL-6, IL-10, tumor necrosis factor (TNF) - α , interferon (IFN) - γ , and transforming growth factor (TGF) - β .

Results: Scores of increased motor activity were negatively correlated with IFN- γ ($\rho = -0.128$, $p = 0.033$) in an acute psychiatric population. There was also a trend towards an association between motor activity and TGF- β ($\rho = 0.118$, $p = 0.050$). In a multiple-linear-regression model correcting for age, gender, and body-mass index (BMI, kg/m²), the association did not remain significant. No significant correlations between motor retardation and circulating cytokines were found.

Conclusions: After adjustment for potential confounders our study did not reveal any significant association between cytokines and motor activity. However, there is an indication of increased Th17 and decreased Th1 responses in relation to increased motor activity in line with the few previous reports in the field. The phenomenon however needs further exploration.

Keywords: cytokines, psychomotor retardation, agitation, depression, psychosis, acute psychiatric care

INTRODUCTION

Altered motor activity is gaining increased interest within psychiatric research and may be a prominent finding in an acute psychiatric setting (1, 2). Traditionally, increased or abnormal activity is seen in ADHD, tic disorders, affective disorders, anxiety, and schizophrenia (3). Motor symptoms may also characterize different subtypes of unipolar depression and predict treatment response (2, 4).

Evidence supports a role of immune activity in the etiology and pathogenesis of psychiatric disorders (5). Several studies have demonstrated altered systemic levels of cytokines in patients with schizophrenia, bipolar disorder, and unipolar depression compared to healthy controls (6). These cytokine alterations may also be more prominent in an acute psychiatric setting (6).

Immune activity often is classified into different profiles based on effector mechanisms and characterized by a set of cytokines promoting those effector mechanisms. Th1 profile is characterized by cytokines such as interferon (IFN) γ and tumor necrosis factor (TNF) α and mediates potent responses to viruses. Th2 profile is characterized by interleukin (IL) -4 and IL-10. Th2 mediates certain B-cell responses (e.g., immunoglobulin-E production) and opposes Th1. Th17 is characterized by cytokines such as IL-17 and TGF- β and mediates other effector mechanism in the immune system (7).

Cytokines may be the factor mediating altered motor activity in certain psychiatric conditions (8). One mechanism by which cytokines may influence motor activity, is through alterations in neural activity and dopamine metabolism in the basal ganglia (9). It is also shown that treatment with cytokines such as IFN- α induces psychomotor retardation and depressive symptoms in patients with hepatitis (10). Finally, an association between motor activity, psychomotor retardation, and cytokines in outpatients with major depression has been described (11). Agitation is an important clinical syndrome consisting of several symptoms and signs, including increased motor activity. In patients with Alzheimer's disease, a previous study demonstrated that increased IL-1 β was associated with agitation (12). However, few previous studies have investigated the association between increased motor activity only and immune markers.

The aim of this study, was to assess the association between circulation levels of cytokines, motor retardation, and increased motor activity in a sample of patients with a variety of severe mental disorders admitted to an acute psychiatric department. Because changes in motor activity are more common symptoms in certain diagnostic groups, unipolar depression and non-affective psychosis were chosen as subgroups.

MATERIALS AND METHODS

Setting and Participants

This cross-sectional study was conducted in the acute psychiatric inpatient wards of St. Olav's University Hospital, Trondheim, Norway. All acutely admitted inpatients between September 2011 and March 2012 were asked to participate. At the time of inclusion, the psychiatric department served a catchment area of 228,000 inhabitants (≥ 18 years old) and represented the only psychiatric inpatient acute unit in the area. Of the total 654 admitted patients in the inclusion period, 382 (58.4%) patients were included in the study. The study was approved by the regional committee for ethics (REC Central number 2011/137) and registered at ClinicalTrials.gov (NCT01415323). All patients

gave their written informed consent prior to inclusion. The inclusion process was conducted by specialists in clinical psychology or psychiatry in order to secure that all included patients had the mental capacity to give their consent. The study was conducted according to the Declaration of Helsinki.

Exclusion Criteria

The following exclusion criteria were applied: (1) chronic or ongoing infections, (2) comorbid autoimmune diseases, (3) C-reactive protein (CRP) levels above 35 mg/L, or (4) lack of patient consent. When patients had multiple admissions, we only included the first admission in our analyses.

Diagnostic Evaluation

Patients were diagnosed according to the International Classification of Diseases-10 (ICD-10) Criteria for Research (13). The diagnoses were set in a consensus meeting in the treatment staff, always including at least two senior psychiatrists of whom one had personally examined the patient. For subgroup analyses, we included patients with non-affective psychosis (ICD-10 F20–29) and unipolar depression (ICD-10 F32 and F33).

Assessments

Sociodemographic history, comorbid medical conditions, smoking status, substance abuse, and psychiatric symptoms were recorded after an interview by a staff member. In addition, participants were screened in a general medical examination and routine blood tests, including CRP and leukocyte count. Height and weight were measured for calculation of the body mass index (BMI, kg/m²).

The degree of motor retardation and increased motor activity was assessed by an experienced clinician using the Symptomatic Organic Mental Disorder Assessment Scale (SOMAS). SOMAS is a 5-item scale developed to assess atypical depressive symptoms. Item B rates the degree of motor retardation, and item C rates the degree of increased motor activity when the patient was most dysthymic during the previous 24 h (14). Both items are modified from the Positive and Negative Syndrome Scale (PANSS), where item B was modified from PANSS item "motor retardation" (general psychopathology scale, item G7), and item C was assessed from PANSS item "hyperactivity" (positive scale, item P4).

For analyses, patients were subdivided into two groups: with or without increased motor activity according SOMAS item C. If the patients were scored as ≥ 2 on SOMAS item C, they were grouped as motor active. Similarly, patients were separated into the two groups with or without motor retardation according to SOMAS item B. A score on SOMAS item B ≥ 3 was set to group the patients as motor retarded. In order to simplify the interpretation of findings on SOMAS item B, it was reverse-coded. Therefore, a higher score on both SOMAS item B and C would be interpreted as more severe symptoms of motor retardation or increased motor activity. Subgroup analyses were performed on patients with diagnoses non-affective psychosis group and unipolar depression.

Serum Analyses of Immune Biomarkers

Blood samples were collected on 9 ml serum tubes with SiO₂ without gel between 08.00 and 13.00 (median at 10:00) at the first working day after admission. Strict instructions regarding fasting were not given, though most patients would be fasting overnight. Samples were immediately cooled on ice, protected from daylight, and centrifuged within 30 min (15 min, 1,500 g, 4°C). Serum samples were stored at –80°C until further analysis in a registered Biobank (Biobank1, St. Olav's University Hospital, Trondheim, Norway). The following parameters were analyzed by multianalyte profiling Milliplex MAP assays: IL-1 β , IL-4, IL-6, IL-10, TNF- α , and IFN- γ (Millipore Corporation, Billerica, MA, US). TGF- β 1 was measured by a Bio-Plex Pro TGF- β Assay (Biorad Hercules, CA, US). Intra- and interassay coefficients of variance were less than 10%. The range of detected values was IL-1 β : 0.06–198.72 pg/ml; IL-4: 0.92–286.01 pg/ml; IL-6: 0.10–576.42 pg/ml; IL-10: 0.10–1125.33 pg/ml; TNF- α : 0.70–268.89 pg/ml; IFN- γ : 0.04–1529.70 pg/ml; and TGF- β : 13.07–415.61 ng/ml. The number of samples and percentage under the detection limit was as follows: IL-1 β : 236 (74.2%), IL-4: 242 (76.1%), IL-6: 177 (55.7%), IL-10: 177 (55.7%), TNF- α : 11 (3.5%), IFN- γ : 68 (21.4%), and TGF- β : 0.

Statistical Analyses

Statistical analyses were done using SPSS version 24.0 for Windows. The level of significance was set at $p \leq 0.05$, and all analyses were two-tailed. Significant findings were adjusted for multiple testing with the Bonferroni correction (α/k where k = the seven tested cytokines giving $\alpha/k = 0.007$). Data normality was assessed by using a Kolmogorov-Smirnov test. The distribution of all serum cytokines was skewed, and only TGF- β became normally distributed after logarithmic transformation. Descriptive statistics were calculated by using chi-square tests for categorical variables and student's independent samples t-tests or Mann-Whitney U test (depending on distribution) for

continuous variables. The Spearman correlation coefficient was calculated for the relationship between cytokines and SOMAS. Additionally, we examined the difference in cytokine levels between the groups with and without increased motor activity by using student's independent samples t-test or Mann-Whitney U test if the data were not normally distributed. The same statistical methods were applied when comparing cytokine levels between the groups with and without motor retardation.

RESULTS

Sociodemographic, Clinical, and Inflammatory Characteristics of the Sample

Of the total 382 patients included in the study, 24 were excluded due to infection or autoimmune diseases. This left us with 358 patients for whom serum samples were available, and cytokines were analyzed in 318 patients. For 277 of these 318 patients we also had complete measures for altered motor activity (increased or reduced motor activity). The main ICD-10 diagnostic categories in the 277 patients were unipolar depression (22.7%), substance-use disorders (15.9%), schizophrenia (9.7%), bipolar disorder (12.7%), neurotic, stress-related, and somatoform disorders (10.1%), and personality disorders (8.3%), and other diagnoses (20.6%).

The demographic, clinical, and immune data for the total study population, the non-affective psychosis group, and unipolar depression group are given in **Table 1**.

Relation Between Serum Inflammatory Markers and Measures of Motor Activity

When all patients were analyzed together, scores of increased motor activity were significantly negatively correlated with IFN- γ ($\rho = -0.128$, $p = 0.033$, **Table 2**). In addition, we found a trend

TABLE 1 | Demographic and clinical parameters.

| | All patients N = 358 | Non-affective psychosis N = 48 | Unipolar depression N = 73 | p-value ^a |
|--|-----------------------------|--------------------------------|----------------------------|--------------------------|
| Age (years), mean \pm SD | 38.9 \pm 14.8 | 40.1 \pm 11.8 | 40.1 \pm 14.9 | 0.756 ^b |
| Gender (female), N (%) | 174 (49) | 19 (40) | 42 (58) | 0.053 ^c |
| Smoking, N (%) | 175 (49) ^d | 25 (66) | 28 (25) | 0.045^c |
| BMI (kg/m ²), mean \pm SD | 25.5 \pm 5.9 ^e | 27.6 \pm 6.2 | 24.9 \pm 5.4 | 0.015^f |
| Higher education (above high school), N (%) | 50 (14) | 2 (4) | 14 (19) | 0.017^c |
| Unemployment (incl. sick leave), N (%) | 255 (71) | 41 (85) | 42 (58) | 0.002^c |
| Alcohol use upon admission, N (%) | 99 (28) | 9 (19) | 15 (21) | 0.808 ^c |
| Substance abuse, N (%) | 78 (22) | 8 (17) | 11 (15) | 0.813 ^c |
| Motor retardation score ^g , mean \pm SD | 1.6 \pm 0.7 | 1.5 \pm 0.8 | 1.7 \pm 0.9 | 0.205 ^f |
| Motor activity score ^g , mean \pm SD | 1.5 \pm 0.9 | 1.6 \pm 1.0 | 1.3 \pm 0.5 | 0.084 ^f |
| Number with blood samples, N (%) | 318 (89) | 40 (83) | 68 (93) | 0.088 ^c |

^aComparison between non-affective psychosis and unipolar depression. Significance with a p-value < 0.05 is indicated in bold text.

^bIndependent students' samples t-test.

^cChi-square test.

^dMissing 60 (10 within non-affective psychosis group and 11 within unipolar depression group).

^eMissing 89 (15 within non-affective psychosis group and 11 within unipolar depression) observed (data not shown).

^fMann-Whitney U test.

^gMotor retardation and motor activity were assessed by items from a Symptomatic Organic Mental Disorder Assessment Scale (SOMAS). Both items were scored on a scale from 1–5. BMI, body mass index; SD, standard deviation.

TABLE 2 | Correlation coefficients (rho) between serum cytokines, motor retardation, and motor activity^a.

| | IL-1 β | IL-6 | TNF- α | IFN- γ | IL-10 | IL-4 | TGF- β |
|--------------------------------|--------------|--------|---------------|---------------|--------|--------|--------------|
| <i>All patients</i> | | | | | | | |
| -Motor retardation | 0.112 | 0.066 | -0.027 | 0.090 | 0.061 | -0.043 | -0.042 |
| -Motor activity | 0.001 | -0.010 | -0.034 | -0.128* | -0.002 | 0.057 | 0.118 |
| <i>Non-affective psychosis</i> | | | | | | | |
| -Motor retardation | 0.154 | 0.165 | 0.002 | 0.191 | 0.194 | -0.181 | -0.153 |
| -Motor activity | 0.052 | 0.251 | 0.029 | 0.000 | 0.085 | 0.080 | -0.054 |
| <i>Unipolar depression</i> | | | | | | | |
| -Motor retardation | -0.089 | -0.003 | -0.047 | -0.007 | -0.056 | 0.059 | 0.173 |
| -Motor activity | -0.046 | -0.133 | -0.181 | -0.173 | -0.239 | -0.002 | 0.107 |

* $p \leq 0.05$.^aMotor retardation and motor activity were scored on a scale from 1–5.

IFN, interferon; IL, interleukin; TGF, transforming growth factor; TNF, tumor necrosis factor.

towards a positive correlation between TGF- β and increased motor activity ($\rho = 0.118$, $p = 0.050$). No significant correlations were found between cytokines, motor retardation, and increased motor activity analyzing the two subgroups non-affective psychosis and unipolar depression (Table 2). In a multiple-linear-regression model correcting for age, gender, and BMI, the associations between increased motor activity and IFN- γ ($\beta = -0.112$, $t = -1.726$, $p = 0.086$) and TGF- β ($\beta = 0.123$, $t = 1.904$, $p = 0.058$) did not remain significant. No findings remained significant after correcting for multiple testing with the Bonferroni correction.

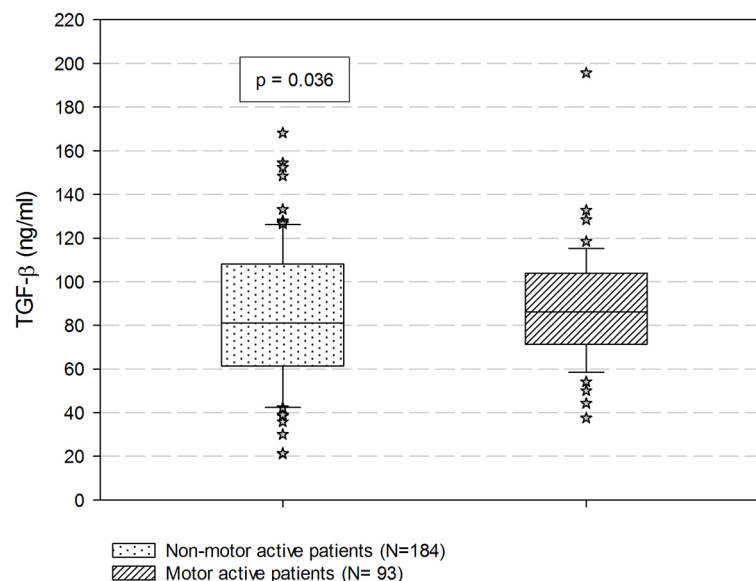
Comparisons of Cytokine Levels Based on the Presence or Absence of Increased Motor Activity and Motor Retardation

Mean log-transformed values of TGF- β were significantly higher in patients with increased motor activity compared to those with no increase in motor activity (11.31 ± 0.34 vs. 11.20 ± 0.45 , $t = -2.11$, $df = 236.41$, $ES = 0.016$, $p = 0.036$) (Figure 1). The

increased-motor-activity group had significantly lower levels of IFN- γ compared to the group without increased motor activity (2.27 ± 11.52 vs 6.00 ± 20.04 , $U = 7213$, $d = 0.13$, $p = 0.032$) (Figure 2). These findings were however not significant after the Bonferroni correction for multiple testing. No other differences in cytokine levels between groups reached the level of significance. When comparing cytokine levels between the two groups with and without motor retardation, no statistical differences were detected (data not shown).

DISCUSSION

After correcting for multiple testing and confounders, we did not find any significant association at the 0.05-level between motor activity and cytokines. However, a trend towards an association between increased motor activity and lower serum levels of IFN- γ and higher levels of TGF- β in patients admitted to an acute psychiatric ward was seen. No statistically significant association

**FIGURE 1** | Comparisons of serum TGF- β based on the prevalence of increased motor activity. P-value is estimated by student's independent samples t-test with log transformed values. Data are expressed as median with percentiles. TGF- β , transforming growth factor β .

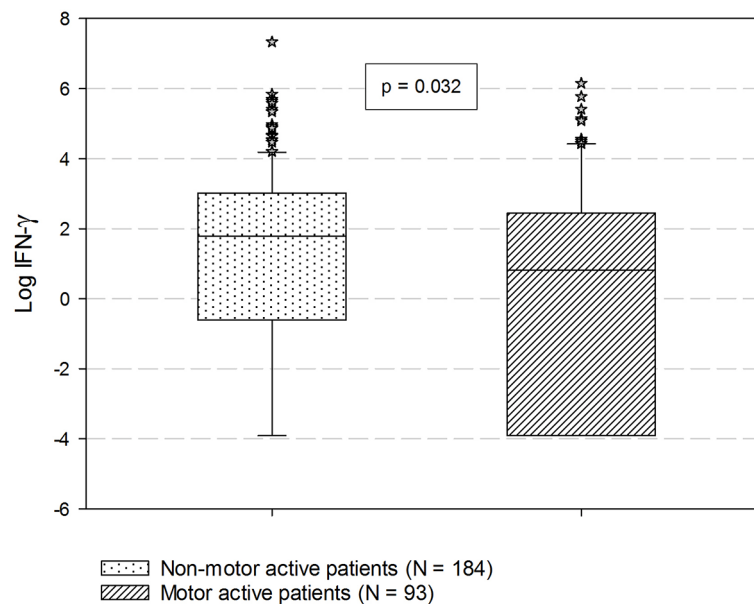


FIGURE 2 | Comparisons of serum IFN- γ based on the prevalence of increased motor activity. P-value is estimated by Mann-Whitney U test. Data are expressed as median with percentiles. IFN- γ , Interferon- γ .

between motor retardation and cytokines was seen. To our knowledge, this is the first report on the relation between immune markers and motor activity in an acute inpatient psychiatric population.

Aggression is a psychiatric sign associated with increased motor activity (15). In a recent study on inpatients with schizophrenia, aggressive behavior was associated with increased levels of Th17 cytokines TGF- β , IL-17, and IL-23 (16). In the present study, we examine motor activity only, but our finding of a trend towards increased TGF- β may be in line with the report on aggressive behavior. However, our findings did not remain significant after corrections for multiple testing and confounders. One might therefore also interpret this previous finding as not being in line with our study.

The present study did not show any significant difference in cytokine levels in relation to motor retardation. This may be somewhat surprising as other studies have indicated a relation between motor retardation and Th1 cytokines. Increased Th1 response has been demonstrated in association with low number of steps per day (17) as well as reduced psychomotor speed (11) in outpatients with major depression. Also, treatment with IFN- α increases the risk of depression and reduced psychomotor speed in patients with hepatitis C (18) and is associated with higher degree of motor retardation in depression (19). However, none of these studies were conducted in an acute psychiatric population, making our results not directly comparable. It is possible that other inflammatory factors are stronger in the acute population masking an association between Th1 and retardation.

There are several limitations and strengths to the current study. We did unfortunately not calculate power for this part of

the study. The degree of motor retardation and increased motor activity was assessed by items from SOMAS, which is validated for this purpose. However, the items are published in a previous study (14, 20), and is also shown to correspond with findings in actigraphy (2). For future studies actigraphy should be included. Finally, even though alterations in cytokines in relation to motor activity were influenced by age, gender, and BMI, cytokines still may be clinically important.

Several of the serum cytokines had a high percentage of samples below the detection limit. Although this was not the case for IFN- γ and TGF- β , it may have affected the analyses of other pro-inflammatory cytokines, increasing the risk for type-II error. Further, the results for subgroup analyses are limited by a relatively low number of participants in the subgroups non-affective psychosis and unipolar depression and by the lack of a healthy control group. However, it is interesting as it suggests that alterations in immune activity are more related to symptoms than diagnostic group.

The study population was also relatively heterogeneous with the possibility of confounding factors. Thus, all findings need replication and should be interpreted with care.

We were however able to include severely ill patients in acute states with a variety of psychiatric diagnoses. The blood samples were drawn during the first 24 h of the admission, the period in which the symptoms were most prominent. In addition, the clinic recruiting the study participants is the only acute psychiatry inpatient service in the catchment area, reducing the effect of socioeconomic factors. All patients in the area needing acute psychiatric services were admitted to this unit. Also, our total sample size is relatively large, compared to other studies in the field.

CONCLUSIONS

Our study comparing levels of immune markers in an acute setting did not reveal any significant associations between altered motor activity and cytokine levels. However, a trend towards a Th17 profile among patients with increased motor activity was seen. The finding should be further explored because it may have implications for predicting and treating deviations in motor activity.

DATA AVAILABILITY STATEMENT

All datasets generated for this study are included in the article/**Supplementary Material**.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by Regional Committees for Medical and Health Research Ethics (REC) Central Regional, Trondheim, Norway. The patients/participants provided their written informed consent to participate in this study.

AUTHOR CONTRIBUTIONS

JL did all statistical analyses, took a leading role in selection of analyses and interpretation of the results, and wrote the original draft for the manuscript. AS did all the laboratory analyses on cytokines. AS took an equal part in planning and discussing the choice of analyses, interpretation of results and writing of the manuscript. VI supervised on statistics and took an equal part in choice and discussion of the statistical analyses as well as writing

of the manuscript. AV initiated the study and headed the inclusion of patients in the clinic. AV took an equal part in interpretation of the results and writing of the manuscript. SR collaborated in initiation and performance of the study in the clinic. SR together with JL took a leading role in choice of laboratory analyses as well as interpretation of the data and in writing of the manuscript.

FUNDING

This work was supported by the Norwegian University of Science and Technology, Department of Mental Health, St. Olav's University Hospital, Division of Mental Health Care, and the Liaison Committee between the Central Norway Regional Health Authority. The funding organization had no role in the design, analysis, interpretation, or publication of the study.

ACKNOWLEDGMENTS

The authors highly appreciate the contribution from all the patients who have made the collection of the material possible. The staff at the acute inpatient department and at the laboratory at St. Olav's University Hospital Department of Østmarka are to be acknowledged.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fpsy.2019.00920/full#supplementary-material>

REFERENCES

- Cheniaux E, Filgueiras A, Silva Rde A, Silveira LA, Nunes AL, Landeira-Fernandez J. Increased energy/activity, not mood changes, is the core feature of mania. *J Affect Disord* (2014) 152–154:256–61. doi: 10.1016/j.jad.2013.09.021
- Krane-Gartiser K, Henriksen TE, Vaaler AE, Fasmer OB, Morken G. Actigraphically assessed activity in unipolar depression: a comparison of inpatients with and without motor retardation. *J Clin Psychiatry* (2015) 76 (9):1181–7. doi: 10.4088/JCP.14m09106
- Fasmer OB, Hauge E, Berle JO, Dilsaver S, Oedegaard KJ. Distribution of Active and Resting Periods in the Motor Activity of Patients with Depression and Schizophrenia. *Psychiatry Investig* (2016) 13(1):112–20. doi: 10.4306/pi.2016.13.1.112
- Todder D, Caliskan S, Baune BT. Longitudinal changes of day-time and night-time gross motor activity in clinical responders and non-responders of major depression. *World J Biol Psychiatry* (2009) 10(4):276–84. doi: 10.3109/15622970701403081
- Kiecolt-Glaser JK, Derry HM, Fagundes CP. Inflammation: depression fans the flames and feasts on the heat. *Am J Psychiatry* (2015) 172(11):1075–91. doi: 10.1176/appi.ajp.2015.15020152
- Goldsmith DR, Rapaport MH, Miller BJ. A meta-analysis of blood cytokine network alterations in psychiatric patients: comparisons between schizophrenia, bipolar disorder and depression. *Mol Psychiatry* (2016) 21 (12):1696–709. doi: 10.1038/mp.2016.3
- Li P, Spolski R, Liao W, Leonard WJ. Complex interactions of transcription factors in mediating cytokine biology in T cells. *Immunol Rev* (2014) 261 (1):141–56. doi: 10.1111/imr.12199
- Carvalho AF, Miskowiak KK, Hyphantis TN, Kohler CA, Alves GS, Bortolato B, et al. Cognitive dysfunction in depression - pathophysiology and novel targets. *CNS Neurol Disord Drug Targets* (2014) 13(10):1819–35. doi: 10.2174/1871527313666141130203627
- Felger JC, Treadway MT. Inflammation effects on motivation and motor activity: role of dopamine. *Neuropsychopharmacology* (2017) 42(1):216–41. doi: 10.1038/npp.2016.143
- Majer M, Welberg LA, Capuron L, Pagnoni G, Raison CL, Miller AH. IFN- α -induced motor slowing is associated with increased depression and fatigue in patients with chronic hepatitis C. *Brain Behav Immun* (2008) 22 (6):870–80. doi: 10.1016/j.bbi.2007.12.009
- Goldsmith DR, Haroon E, Woolwine BJ, Jung MY, Wommack EC, Harvey PD, et al. Inflammatory markers are associated with decreased psychomotor speed in patients with major depressive disorder. *Brain Behav Immun* (2016) 56:281–8. doi: 10.1016/j.bbi.2016.03.025
- Higuchi M, Hatta K, Honma T, Hitomi YH, Kambayashi Y, Hibino Y, et al. Association between altered systemic inflammatory interleukin-1 β and

- natural killer cell activity and subsequently agitation in patients with Alzheimer disease. *Int J Geriatr Psychiatry* (2010) 25(6):604–11. doi: 10.1002/gps.2381
13. WHO. *The ICD-10 classification of mental and behavioural disorders: diagnostic criteria for research*. Geneva: World Health Organization. (1993).
 14. Vaaler AE, Morken G, Iversen VC, Kondziella D, Linaker OM. Acute Unstable Depressive Syndrome (AUDS) is associated more frequently with epilepsy than major depression. *BMC Neurol* (2010) 10:67. doi: 10.1186/1471-2377-10-67
 15. Battaglia J. Pharmacological management of acute agitation. *Drugs* (2005) 65(9):1207–22. doi: 10.2165/00003495-200565090-00003
 16. Li H, Zhang Q, Li N, Wang F, Xiang H, Zhang Z, et al. Plasma levels of Th17-related cytokines and complement C3 correlated with aggressive behavior in patients with schizophrenia. *Psychiatry Res* (2016) 246:700–6. doi: 10.1016/j.psychres.2016.10.061
 17. Schmidt FM, Lichtblau N, Minkwitz J, Chittka T, Thormann J, Kirkby KC, et al. Cytokine levels in depressed and non-depressed subjects, and masking effects of obesity. *J Psychiatr Res* (2014) 55:29–34. doi: 10.1016/j.jpsychires.2014.04.021
 18. Haroon E, Felger JC, Woolwine BJ, Chen X, Parekh S, Spivey JR, et al. Age-related increases in basal ganglia glutamate are associated with TNF, reduced motivation and decreased psychomotor speed during IFN- α treatment: preliminary findings. *Brain Behav Immun* (2015) 46:17–22. doi: 10.1016/j.bbi.2014.12.004
 19. Capuron L, Fornwalt FB, Knight BT, Harvey PD, Ninan PT, Miller AH. Does cytokine-induced depression differ from idiopathic major depression in medically healthy individuals? *J Affect Disord* (2009) 119(1–3):181–5. doi: 10.1016/j.jad.2009.02.017
 20. Saether SG, Vaaler A, Evjenth A, Aune T, Holtje M, Ruprecht K, et al. Subtle phenotype differences in psychiatric patients with and without serum immunoglobulin G antibodies to synapsin. *Front In Psychiatry* (2019) 10:401. doi: 10.3389/fpsyt.2019.00401

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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CACNA1C rs1006737, Threatening Life Events, and Gene–Environment Interaction Predict Major Depressive Disorder

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OPEN ACCESS

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Specialty section:

This article was submitted to
Molecular Psychiatry,
a section of the journal
Frontiers in Psychiatry

Received: 09 September 2019

Accepted: 10 December 2019

Published: 22 January 2020

Citation:

Zhao M, Yang J, Qiu X, Yang X, Qiao Z,
Song X, Wang L, Zhao E, Yang Y and
Cao D (2020) CACNA1C rs1006737,
Threatening Life Events, and Gene–
Environment Interaction Predict Major
Depressive Disorder.
Front. Psychiatry 10:982.
doi: 10.3389/fpsy.2019.00982

Introduction: CACNA1C rs1006737 is a novel variant in discovery of replicable associations in major depressive disorder (MDD). However, there have been no specific studies considered effect of environmental pathogens to date examining its clinical significance. In this study we investigated the interaction effect between CACNA1C rs1006737 polymorphism and threatening life events (TLEs) in MDD and carried out a meta-analysis of published findings.

Methods: A total of 1,177 consecutive participants were genotyped. Information on exposure to TLEs, socio-demographic data, and history of psychological problems among first-degree relatives was collected. MDD was diagnosed according to the Chinese version of the 24-item Hamilton Rating Scale for Depression.

Results: There was a significant interaction effect between CACNA1C rs1006737 polymorphism and TLEs in MDD. A dose–response relationship was found between CACNA1C rs1006737 genotypes and TLEs in MDD. The results of the meta-analysis showed that CACNA1C rs1006737 genotypes interacted with TLEs in MDD.

Conclusion: CACNA1C rs1006737 genotype and previous exposure to TLEs interact to influence the risk of developing MDD. We propose that CACNA1C rs1006737 may represent a target for novel pharmacological therapies to prevent or treat MDD.

Keywords: CACNA1C, polymorphism, threatening life events, gene–environment interaction, major depressive disorder

INTRODUCTION

Predisposition to complex diseases is not solely conferred by genetic factors; it is also influenced by environmental exposure. Delineating their respective contributions is a major challenge in the study of complex diseases (1–5). Gene–environment interactions (G × E) are thought to account for a large fraction of the unexplained variance in heritability and disease risk (6, 7). However, disease risk due either to environmental exposure and/or its interactions with genotype remains poorly understood (8, 9).

Major depressive disorder (MDD) is an example of a complex disease for which G × E are likely important. An early study on G × E in MDD showed that a length polymorphism (*SLC6A4*) in the promoter region of the 5-*HTT* gene mediates the response to stressful life events (10). The *rs1006737* polymorphism in the third intron of the gene encoding the Cav1.2 subunit of the L-type voltage-gated calcium channel gene (*CACNA1C*) (Chromosome12:2345295), which is highly expressed throughout the forebrain (11), has been attributed to G × E. Cav1.2 couples a transient increase in membrane permeability to cell membrane depolarization and gene transcription and plays a critical role in dendritic development, neuronal survival, synaptic plasticity, memory formation, learning, and behavior (12–16). *In vitro* studies have shown that disease-associated increases in Ca²⁺ influx via Cav1.2 channels can alter gene expression (17) and contribute to activity-dependent dendrite retraction (18), which can occur in response to chronic stress (19).

Threatening life events (TLEs) precede the onset of depressive episodes more frequently than expected by chance (20); TLEs were shown to cluster before the onset of a depressive episode or an exacerbation of symptoms (21, 22). Although a single stressor may have relatively minor effects, the cumulative effects of multiple stressors (23) can lead to psychiatric disorders, consistent with a dose–response effect (24).

It was recently reported that the *CACNA1C rs1006737* polymorphism mediates the influence of TLEs on human MDD (25). However, this has been contradicted by another study (26). Neither of these investigations addressed the specificity of the *CACNA1C rs1006737* and dose–response effects of TLEs. Examining the perceived threat level of TLEs in the context of *CACNA1C rs1006737* genotype may provide more detailed insight into the nature of genetic effects on stress response.

Measurements of environmental risk can vary across studies; a meta-analysis is one tool for determining whether a result transcends inter-study variation, and is widely used in the field of psychiatric genetics, which has been plagued in recent years by non-reproducibility (27). By pooling data from several studies, a meta-analysis maximizes the power to detect significant effects and avoids overemphasizing estimates from any single study (28).

In this report, we investigated G × E effects between *CACNA1C rs1006737* genotype, TLEs, and MDD in a large clinical sample. A meta-analysis was also carried out to evaluate the current evidence for these interactions.

MATERIAL AND METHODS

Study Population

From November 2014 and December 2017, 590 patients with MDD (420 women and 170 men) were recruited for the study (mean age: 44.22 ± 13.45 years) along with 587 age- and sex-matched control subjects without a history of neuropsychiatric disorders. Both patients and controls subjects were from the same geographic area in Northern China and were of Chinese

Han ethnicity, and provided written, informed consent before participation in the study. The study was approved by the Ethics Committee of Harbin Medical University.

Independent Measures

Participants completed three questionnaires: a socio-demographic questionnaire, the Chinese version of the 24-item Hamilton Rating Scale for Depression (HRSD-24), and the Life Events Scale (LES). The socio-demographic questionnaire was used to collect detailed information about socioeconomic background and medical history including individual and family psychiatric history. The HRSD-24 is a reliable tool that has been used in several studies to assess depressive symptoms (29–31). Patients above the threshold (21 points) were included in the study. The LES was used to evaluate negative life events; this self-rating questionnaire consists of 48 items in three areas—i.e., family life (28 items), work-related problems (13 items), and social and other aspects (seven items) (32).

Genotyping

Genomic DNA was extracted from venous blood samples using the AxyPrep Blood Genomic DNA Miniprep kit (Axygen, Union City, CA, USA) and the single nucleotide polymorphism (SNP) *rs1006737* of the *CACNA1C* gene was detected by PCR amplification using primers designed with Primer 5.0 software, which had the following sequences: 5'-AAGTTCC ATTCCATCTCAGCCCGAA-3' (forward) and 5'-TGTT TTCAGAGCCGGAGACCTCAC-3' (reverse). SNP analysis was performed using SNaPshot according to the manufacturer's instructions.

Statistical Analysis

Data were analyzed using R Studio 1.1.423. The χ^2 test was used to evaluate differences in the distributions of independent variables. Genotype frequencies were tested for Hardy–Weinberg equilibrium. The Bonferroni method was used for multiple-testing correction of genetic association in univariate analysis and the significance level was set at $P < 0.01$ (0.05/5). G × E were examined with a logistic regression model. Four predictor variables were used: *CACNA1C rs1006737* genotype (GG, GA, or AA), sex, family history, and either the presence/absence or number of TLEs. The dependent variable was the onset of an episode of MDD. Study power was calculated with QUANTO 1.2.4 (<http://hydra.usc.edu/gxe/>).

For analyses incorporating TLEs, the TLEs were coded so that 0 represented no TLE occurrence and values of 1, 2, 3, or ≥4 represented the occurrence of TLEs that were minor, low-moderate, high-moderate, and severe, respectively. To simplify the interpretation of interactions, the number of TLEs was coded using four dummy variables (X1, X2, X3, and X4). If there was no TLE, all four were coded as zero. For example, if there was one, two, or three TLEs, X1, X2, and X3, respectively, were coded as 1. Thus, the coding for three TLEs was: X1 = 1, X2 = 1, X3 = 1, and X4 = 0. This method of coding dummy variables known as thermometer coding does not alter the model results but is simpler yet mathematically equivalent to contrasts (33); compared to typical indicator variables, it greatly simplifies the

model selection process. Removing a level of a standard indicator variable requires recoding the data and a likelihood ratio test; with thermometer coding, the task is not different from removing other independent variables.

We conducted a meta-analysis by pooling results from previous G × E studies of the *CACNA1C* rs1006737 polymorphism, TLEs, and MDD with findings from the present study. The Lipták–Stouffer z-score approach was used to obtain an aggregate value for the significance level of tests weighted by sample size, and a sensitivity analysis was conducted by recomputing effect size after systematically removing each study in turn. To gauge potential publication bias, we calculated fail-safe N and its ratio (34, 35).

RESULTS

Frequencies of Independent Variables

Demographic and genotypic data for the study population are shown in **Table 1**. About half of subjects had TLEs, and one in nine had a family history of psychological problems among first-degree relatives. Approximately half of subjects had the G/G genotype, one in three the G/A genotype, and the remaining subjects the A/A genotype. Genotype frequencies were in Hardy–Weinberg equilibrium among both cases and controls.

Associations With MDD

There were significant differences in genotype ($\chi^2 = 6.36$, $P = 0.04$), homozygosity ($\chi^2 = 5.47$, $P = 0.01$), TLEs ($\chi^2 = 64.27$, $P = 1.105 \times 10^{-14}$), and family history ($\chi^2 = 66.55$, $P = 3.408 \times 10^{-16}$) distribution between patients with MDD and controls (**Table 2**). TLEs ($P < 0.01$) and family history ($P < 0.01$) were still associated with MDD after Bonferroni correction. On the basis of sample size of the study, the power for association study of *CACNA1C* rs1006737 was 98.92%.

TABLE 1 | Summarized frequencies of socio-demographic and independent variables.

| Variables | Frequencies |
|---|-------------------|
| Socio-demographic variables | |
| Gender | |
| Female | 802 (68.1%) |
| Male | 375 (31.9%) |
| Mean age | 43.6 (s.d. 11.55) |
| Independent variables | |
| <i>CACNA1C</i> (rs1006737) genotypes | |
| G/G | 770 (65.4%) |
| G/A | 365 (31.1%) |
| A/A | 42 (3.5%) |
| Exposure to threatening experiences | |
| No TLE | 592 (50.2%) |
| 1 TLE | 284 (24.2%) |
| 2 or more TLEs | 301 (25.6%) |
| Family history of psychological problems among first-degree relatives | |
| FH+ | 102 (9%) |
| FH– | 1,075 (91%) |

FH, family history; s.d., standard deviation; TLE, threatening life events.

Interaction Between TLE Occurrence and *CACNA1C* rs1006737 Genotype in the Prediction of MDD

In our initial analyses, which considered only the presence or absence of TLEs, a full model was first generated including GG/GA/AA genotype, sex, family history, and the occurrence of TLEs. We then selected the optimal model based on the Akaike information criterion. The dominant mode of action that combined the effects of GA and AA genotypes showed an improvement in fit. This best-fit model suggested significant main effects of family history ($\beta = 8.56$, s.e. = 0.37, $P = 1.29 \times 10^{-8}$) and TLE occurrence ($\beta = 2.08$, s.e. = 0.15, $P = 1.26 \times 10^{-6}$) but not of sex ($\beta = 0.83$, s.e. = 0.13, $P = 0.17$) or genotype ($\beta = 0.91$, s.e. = 0.17, $P = 0.55$) for predicting MDD (**Table 3**). However, a significant genotype × TLE interaction was found ($\beta = 1.81$, s.e. = 0.27, $P = 0.02$). Estimates based on this model indicated that family history and TLE exposure could influence the prediction of MDD, and that genetics alone cannot predict MDD but can modify the risk effect conferred by exposure to TLEs. On the basis of sample size of the study, the power was 87.59% to detect a significant effect of rs1006737 × TLE interaction on MDD under dominant genetic model.

Interaction Between Number of TLEs and *CACNA1C* rs1006737 Genotype in the Prediction of MDD

Based on the evidence for an interaction between *CACNA1C* rs1006737 genotype and TLE exposure in the prediction of MDD, we explored how this polymorphism alters the dose-response relationship between number of TLEs and risk for MDD onset.

We generated a full model encompassing the dominant action model (GG vs. GA/AA), TLEs, sex, and family history. Two of the four possible interactions with genotype and number of TLEs were retained. The final model for the prediction of MDD included sex, family history, minor threat, low-moderate threat, high-moderate

TABLE 2 | Association between depression and genetic or environmental factors.

| | Case | Control | χ^2 | df | P |
|-------------------------|----------|----------|----------|----|------------|
| Genotypes | | | 6.36 | 2 | 0.04 |
| G/G | 384 (65) | 386 (66) | | | |
| G/A | 177 (30) | 188 (32) | | | |
| A/A | 29 (5) | 13 (2) | | | |
| Homozygous | | | 5.47 | 1 | 0.01 |
| G/* | 561 (95) | 574 (98) | | | |
| A/A | 29 (5) | 13 (2) | | | |
| Alleles | | | 1.08 | 1 | 0.29 |
| G | 945 (80) | 960 (82) | | | |
| A | 235 (20) | 214 (18) | | | |
| Threatening life events | | | 64.27 | 2 | 1.105e−14* |
| No | 229 (39) | 363 (62) | | | |
| 1 | 167 (28) | 117 (20) | | | |
| 2 or more | 194 (33) | 107 (18) | | | |
| Family history | | | 66.55 | 1 | 3.408e−16* |
| Negative | 499 (85) | 576 (98) | | | |
| Positive | 91 (15) | 11 (2) | | | |

* $P < 0.01$ after Bonferroni correction.

TABLE 3 | CACNA1C genotype interaction with threatening life experiences.

| | β (95% CI) | SE | P |
|----------------------------|-------------------|------|----------|
| Sex | 0.83 (0.63–0.99) | 0.13 | 0.17 |
| Family history | 8.56 (4.34–19.46) | 0.37 | 1.29e–08 |
| No/Any TLE (E) | 2.08 (1.55–2.81) | 0.15 | 1.26e–06 |
| CACNA1C genotype (G) | 0.91 (0.64–1.26) | 0.17 | 0.55 |
| Gene (G) × Environment (E) | 1.81 (1.06–3.012) | 0.27 | 0.02 |

threat, main effects of CACNA1C *rs1006737* genotypes, and the interaction between genotype and TLE values of 1 and 3 (Table 4).

The main effects of CACNA1C *rs1006737* genotypes ($\beta = 0.90$, s.e. = 0.17, $P = 0.55$), sex ($\beta = 0.84$, s.e. = 0.13, $P = 0.20$), and a TLE of 2 ($\beta = 0.85$, s.e. = 0.26, $P = 0.56$) were non-significant in this final model. Conversely, those of a TLE of 1 ($\beta = 0.181$, s.e. = 0.18, $P = 0.001$) or 3 ($\beta = 2.16$, s.e. = 0.31, $P = 0.01$) and family history ($\beta = 8.35$, s.e. = 0.37, $P = 2.05e^{-08}$) were significant. Importantly, we found that the CACNA1C *rs1006737* genotype interaction with a TLE of 1 ($\beta = 2.02$, s.e. = 0.17, $P = 0.04$, $\gamma = 2.37$) was significant, with individuals harboring the AA and GA genotypes showing greater sensitivity to the depression-inducing effects of a TLE of 1 than those with the GG genotype. We also observed a significant interaction between genotype and a TLE of 3 ($\beta = 0.28$, s.e. = 0.61, $P = 0.03$, $\gamma = 0.13$)—that is, high exposure to TLEs was associated with an increase in risk for MDD in all genotypes. Interaction coefficients (γ) ranging from a TLE of 1 to 3 indicated a low exposure–gene effect between exposure level and CACNA1C *rs1006737* polymorphism in MDD. In contrast, the interaction between CACNA1C *rs1006737* genotypes and a TLE of 2 ($\beta = 1.54$, s.e. = 0.52, $P = 0.40$) was non-significant in this final model (Table 4). Estimates based on this model indicated that family history and TLE exposure still have main effects on the prediction of MDD, and that genetics can modify the risk effect conferred by exposure to TLEs from a minor threat to a high-moderate threat.

Meta-Analysis

We included studies in our meta-analysis that met three criteria: the study had to be published in a peer-reviewed journal, and include genotypic information on the CACNA1C *rs1006737* gene as well as a measure of TLEs. After searching the PubMed, Wolters Kluwer, and Web of Science databases, we identified two previous studies that met all three criteria (8,9). The results of these studies were pooled with the present findings to assess the interaction between CACNA1C *rs1006737* polymorphism and TLEs in MDD

in a total of 8,728 subjects (Table 5 and Figure 1). The significance of the results remained robust in the sensitivity analysis when each study was removed in turn, with the exception of one study ($0.0001 < P < 0.01$). To render the outcome in the analysis non-significant ($P = 0.05$), an additional four unpublished or undiscovered studies with average sample size of $n = 2909$ and a non-significant result ($P = 0.50$) would be required. This yielded a fail-safe ratio of one study excluded for each one included in the meta-analysis.

DISCUSSION

Prior studies documenting the interaction effect between CACNA1C *rs1006737* genotypes and TLEs in MDD have reported a positive interaction effect (25) or contrary findings (26). However, neither of these studies focused on a single marker, resulting in low *a priori* probability and power. Additionally, the influence of the CACNA1C *rs1006737* polymorphism on the dose-response relationship between TLEs and risk for MDD was not reported in either study. Here we attempted to replicate the prior finding that CACNA1C *rs1006737* genotypes modified the depressogenic effects of TLEs. Our second goal was to clarify the dose-response relationship between TLEs and CACNA1C *rs1006737* genotypes in MDD. Finally, we carried out a meta-analysis of the interaction between CACNA1C *rs1006737* genotypes and TLEs in MDD to evaluate the current evidence.

Our main findings were that CACNA1C *rs1006737* genotypes and TLEs were independently associated with MDD, and that CACNA1C *rs1006737* genotypes significantly modified the risk conferred by TLEs for MDD; moreover, a dose-response relationship was found to exist between CACNA1C *rs1006737* genotypes and TLEs in MDD, with the meta-analysis confirming an interaction between these variables.

Previous association studies of MDD have suffered from low rates of reproducibility. Instead of a main effect of genotype on phenotype, G×E effects were reported. Since the latter are more difficult to detect than the former (36, 37), replications may be expected to be more rare and have a specific value when they do occur. We first analyzed the interaction between TLE occurrence and CACNA1C *rs1006737* genotypes in the prediction of MDD and found a better-fitting model after adjusting for potential confounds such as sex and family history. There is conflicting evidence regarding the impact of sex on depressive disorder, with

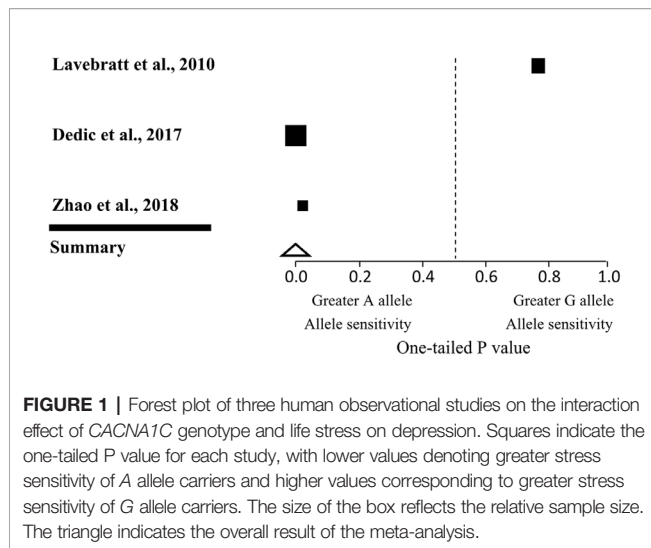
TABLE 4 | CACNA1C genotype interactions with threat exposure.

| | β (95% CI) | SE | P |
|---------------------------------|-------------------|------|----------|
| Sex | 0.84 (0.65–1.09) | 0.13 | 0.20 |
| Family history | 8.35 (4.22–19.01) | 0.37 | 2.05e–08 |
| Minor threat | 1.81 (1.26–2.59) | 0.18 | 0.001 |
| Low-moderate threat | 0.85 (0.51–1.45) | 0.26 | 0.56 |
| High-moderate threat | 2.16 (1.18–3.97) | 0.31 | 0.01 |
| CACNA1C genotype (G) | 0.90 (0.64–1.26) | 0.17 | 0.55 |
| Gene (G) × Minor threat | 2.02 (1.03–4.06) | 0.34 | 0.04 |
| Gene (G) × Low-moderate threat | 1.54 (0.56–4.39) | 0.52 | 0.40 |
| Gene (G) × High-moderate threat | 0.28 (0.08–0.93) | 0.61 | 0.03 |

TABLE 5 | Studies on the interaction between CACNA1C polymorphism, threatening life events, and depression included in the meta-analysis.

| Source, year | No. of Subjects | 1-Tailed P Value | P Value After Study Exclusion |
|-----------------------------|-----------------|------------------|-------------------------------|
| Lavebratt et al. (26) | 2,743 | 0.77 | 0.0001 |
| Dedic et al. (25) | 4,808 | 0.001 | 0.41 |
| Zhao et al. (34) | 1,177 | 0.01 | 0.01 |
| Total: | 8,728 | | |
| Average sample size: | 2909 | 0.003 | |

$P = 0.003$ stands for overall effect size which is significant.



some studies reporting it as valid for both sexes (38–41), and others suggesting an effect only in women (42, 43) or the inverse effect in men (44). We found a significant sex difference between patients with MDD and controls. Family history of psychological problems is associated with both exposure (45) and outcome (46); we also found a significant difference in family history between patients with MDD and controls, and therefore included this parameter in the model. In the regression analysis, family history remained significant but not for sex and TLEs, and had a main effect but not for genotype. Importantly, a genotype × TLE interaction was observed. These findings suggest that the *CACNA1C* *rs1006737* polymorphism does not have a main effect on MDD by itself, but does in combination with TLEs.

We then analyzed the interaction between number of TLEs and *CACNA1C* *rs1006737* genotypes in the prediction of MDD. Sex and family history were retained in the model, although the effect of sex was non-significant. Our results showed that individuals with the AA or GA genotype had greater sensitivity to the depressogenic effects of a TLE of 1 than those with the GG genotype, and that high exposure to TLEs was associated with a marked increase in the risk for MDD for all genotypes. The dose-response effect analysis revealed a low exposure-gene effect. Exposure levels were measured based on retrospective reporting by participants, and bias in this data could have influenced the dose-response effect. In many cases, cumulative measurements can be obtained by making repeated measurements over time, which enhances power to detect G × E (47). The dose-response effect in our study suggests that elucidating the mechanism underlying the progression from genetic variation to MDD requires more precise measures of environmental risk factors and stressful experiences.

A strength of this study is that it included a meta-analysis as well as an analysis of original data. The former provided evidence of a *CACNA1C* *rs1006737* genotype × TLE interaction effect in MDD. However, our previous meta-analysis of G × E (34, 35) showed that a subgroup analysis stratified by type of stressor, study design, or subjects' ancestry should be carried out wherever possible in order to reduce confounds for the G × E effect.

Our study also had some limitations. Firstly, data on environmental pathogens were collected from subjects' retrospective reports, which has risks such as forgetting, revisionist recall, and bias due to cognitive dysfunction or low mood (48). Secondly, mistreatment in childhood was not considered as an independent environmental risk factor separate from TLEs although it can affect the development of neural circuitry and is therefore a good candidate to study G × E effects in mental disorders (49, 50).

In conclusion, we provide evidence supporting an effect modification by the *CACNA1C* *rs1006737* genotype on the risk of MDD conferred by previous exposure to TLEs. Thus, *CACNA1C* *rs1006737* is an example of a gene that influences vulnerability to MDD not by a main effect on risk but rather by modulating sensitivity to the negative effects of the environment. Future work will include genome-wide association studies data to test for G × E interactions.

DATA AVAILABILITY STATEMENT

The datasets for this article are not publicly available because the datasets were also used in another study which is not published yet. Request to access the datasets should be directed to YY, yanjie1965@163.com.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by Ethics Committee of Harbin Medical University. The patients/participants provided their written informed consent to participate in this study. Written informed consent was obtained from the individual(s) for the publication of any potentially identifiable images or data included in this article.

AUTHOR CONTRIBUTIONS

MZ and JY conducted the statistical analyses and wrote the first draft of the manuscript. XQ and XY provided expertise in MDD search. ZQ and XS collected the data. LW and EZ did the experiment. YY and DC designed this study and provided expertise. All authors were involved in modifying the secondary-analysis design and editing the manuscript. All authors contributed to and have approved the final manuscript.

FUNDING

This study was supported by the National Natural Science Foundation of China (81473054, 81773536) to YY.

ACKNOWLEDGMENTS

We want to express our gratitude to all patients and healthy controls parting in the study, as well as to the psychiatrists for their help in the recruitment and identification of patients with major depressive disorder.

REFERENCES

- Ye CJ, Feng T, Kwon HK, Raj T, Wilson MT, Asinowski N, et al. Intersection of population variation and autoimmunity genetics in human T cell activation. *Sci (New York NY)* (2014) 345(6202):1254665. doi: 10.1126/science
- Wu S, Powers S, Zhu W, Hannun YA. Substantial contribution of extrinsic risk factors to cancer development. *Nature* (2016) 529(7584):43–7. doi: 10.1038/nature16166
- Nelson MR, Wegmann D, Ehm MG, Kessner D, St Jean P, Verzilli C, et al. An abundance of rare functional variants in 202 drug target genes sequenced in 14,002 people. *Sci (New York NY)* (2012) 337(6090):100–4. doi: 10.1126/science.1217876
- Grubert F, Zaugg JB, Kasowski M, Ursu O, Spacek DV, Martin AR, et al. Genetic control of chromatin states in humans involves local and distal chromosomal interactions. *Cell* (2015) 162(5):1051–65. doi: 10.1016/j.cell.2015.07.048
- Carr EJ, Dooley J, Garcia-Perez JE, Lagou V, Lee JC, Wouters C, et al. The cellular composition of the human immune system is shaped by age and cohabitation. *Nat Immunol* (2016) 17(4):461–8. doi: 10.1038/ni.3371
- Franks PW, Pearson E, Florez JC. Gene-environment and gene-treatment interactions in type 2 diabetes: progress, pitfalls, and prospects. *Diabetes Care* (2013) 36(5):1413–21. doi: 10.2337/dc12-2211
- Marigorta UM, Gibson G. A simulation study of gene-by-environment interactions in GWAS implies ample hidden effects. *Front In Genet* (2014) 5:225. doi: 10.3389/fgene.2014.00225
- Patel CJ, Ioannidis JP. Studying the elusive environment in large scale. *Jama* (2014) 311(21):2173–4. doi: 10.1001/jama.2014.4129
- Idaghdour Y, Quinlan J, Goulet JP, Berghout J, Gbeha E, Bruat V, et al. Evidence for additive and interaction effects of host genotype and infection in malaria. *Proc Natl Acad Sci U States America* (2012) 109(42):16786–93. doi: 10.1073/pnas.1204945109
- Caspi A, Sugden K, Moffitt TE, Taylor A, Craig IW, Harrington H, et al. Influence of life stress on depression: moderation by a polymorphism in the 5-HTT gene. *Sci (New York NY)* (2003) 301(5631):386–9. doi: 10.1126/science.1083968
- Hell JW, Westenbroek RE, Warner C, Ahljianian MK, Prystay W, Gilbert MM, et al. Identification and differential subcellular localization of the neuronal class C and class D L-type calcium channel $\alpha 1$ subunits. *J Cell Biol* (1993) 123(4):949–62. doi: 10.1083/jcb.123.4.949
- Shibasaki M, Kurokawa K, Ohkuma S. Upregulation of L-type Ca(v)1 channels in the development of psychological dependence. *Synapse (New York NY)* (2010) 64(6):440–4. doi: 10.1002/syn.20745
- White JA, McKinney BC, John MC, Powers PA, Kamp TJ, Murphy GG. Conditional forebrain deletion of the L-type calcium channel $\text{Ca}_v1.2$ disrupts remote spatial memories in mice. *Learn Memory (Cold Spring Harb. NY)* (2008) 15(1):1–5. doi: 10.1101/lm.773208
- Narayanan D, Xi Q, Pfeiffer LM, Jaggar JH. Mitochondria control functional $\text{CaV}1.2$ expression in smooth muscle cells of cerebral arteries. *Circ Res* (2010) 107(5):631–41. doi: 10.1161/CIRCRESAHA.110.224345
- Kobrinisky E, Duong SQ, Sheydina A, Soldatov NM. Microdomain organization and frequency-dependence of CREB-dependent transcriptional signaling in heart cells. *FASEB J: Off Publ Fed Am Societies Exp Biol* (2011) 25(5):1544–55. doi: 10.1096/fj.10-176198
- Wheeler DG, Barrett CF, Groth RD, Safa P, Tsien RW. CaMKII locally encodes L-type channel activity to signal to nuclear CREB in excitation-transcription coupling. *J Cell Biol* (2008) 183(5):849–63. doi: 10.1083/jcb.200805048
- Tian Y, Voineagu I, Pasca SP, Won H, Chandran V, Horvath S, et al. Alteration in basal and depolarization induced transcriptional network in iPSC derived neurons from Timothy syndrome. *Genome Med* (2014) 6(10):75. doi: 10.1186/s13073-014-0075-5
- Krey JF, Pasca SP, Shcheglovitov A, Yazawa M, Schwemberger R, Rasmusson R, et al. Timothy syndrome is associated with activity-dependent dendritic retraction in rodent and human neurons. *Nat Neurosci* (2013) 16(2):201–9. doi: 10.1038/nn.3307
- Erburu M, Cajaleon L, Guruceaga E, Venzala E, Munoz-Cobo I, Beltran E, et al. Chronic mild stress and imipramine treatment elicit opposite changes in behavior and in gene expression in the mouse prefrontal cortex. *Pharmacol. Biochem. Behav* (2015) 135:227–36. doi: 10.1016/j.pbb.2015.06.001
- Tennant C. Life events, stress and depression: a review of recent findings. *Aust New Z J Psychiatry* (2002) 36(2):173–82. doi: 10.1046/j.1440-1614.2002.01007.x
- Hosang GM, Korszun A, Jones L, Jones I, Gray JM, Gunasinghe CM, et al. Adverse life event reporting and worst illness episodes in unipolar and bipolar affective disorders: measuring environmental risk for genetic research. *Psychol Med* (2010) 40(11):1829–37. doi: 10.1017/S003329170999225X
- Hosang GM, Uher R, Maughan B, McGuffin P, Farmer AE. The role of loss and danger events in symptom exacerbation in bipolar disorder. *J Psychiatr Res* (2012) 46(12):1584–9. doi: 10.1016/j.jpsychires.2012.07.009
- Evans GW. The environment of childhood poverty. *Am Psychol* (2004) 59(2):77–92. doi: 10.1037/0003-066X.59.2.77
- Kolassa IT, Ertl V, Eckart C, Glockner F, Kolassa S, Papassotiropoulos A, et al. Association study of trauma load and SLC6A4 promoter polymorphism in posttraumatic stress disorder: evidence from survivors of the Rwandan genocide. *J Clin Psychiatry* (2010) 71(5):543–7. doi: 10.4088/JCP.08m04787blu
- Dedic N, Pohlmann ML, Richter JS, Mehta D, Czamara D, Metzger MW, et al. Cross-disorder risk gene CACNA1C differentially modulates susceptibility to psychiatric disorders during development and adulthood. *Mol Psychiatry* (2018) 23(3):533–43. doi: 10.1038/mp.2017.133
- Lavebratt C, Aberg E, Sjöholm LK, Forsell Y. Variations in FKBP5 and BDNF genes are suggestively associated with depression in a Swedish population-based cohort. *J Affect Disord* (2010) 125(1–3):249–55. doi: 10.1016/j.jad.2010.02.113
- Insel TR, Collins FS. Psychiatry in the genomics era. *Am J Psychiatry* (2003) 160(4):616–20. doi: 10.1176/appi.ajp.160.4.616
- Ioannidis JP, Ntzani EE, Trikalinos TA, Contopoulos-Ioannidis DG. Replication validity of genetic association studies. *Nat Genet* (2001) 29(3):306–9. doi: 10.1038/ng749
- Han D, Qiao Z, Chen L, Qiu X, Fang D, Yang X, et al. Interactions between the vascular endothelial growth factor gene polymorphism and life events in susceptibility to major depressive disorder in a Chinese population. *J Affect Disord* (2017) 217:295–8. doi: 10.1016/j.jad.2017.04.028
- Ma J, Xiao H, Yang Y, Cao D, Wang L, Yang X, et al. Interaction of tryptophan hydroxylase 2 gene and life events in susceptibility to major depression in a Chinese Han population. *J Affect Disord* (2015) 188:304–9. doi: 10.1016/j.jad.2015.07.041
- Ma J, Wang L, Yang Y, Qiao Z, Fang D, Qiu X, et al. GNB3 and CREB1 gene polymorphisms combined with negative life events increase susceptibility to major depression in a Chinese Han population. *PloS One* (2017) 12(2):e0170994. doi: 10.1371/journal.pone.0170994
- Yang DS, Zhang YL. Life Event Scale (LES). Rating Scales for Mental Health. In: Wang XD, Wang XL, Ma H, editors. *Chin. Ment. Health J, Beijing* (1999). pp. 101–106
- Schwartz M. Practical neural network recipes in C++-T. masters. *IEEE Trans Neural Networks* (1994) 9(4):479–90. doi: 10.1016/j.jda.2011.03.012
- Zhao M, Chen L, Yang J, Han D, Fang D, Qiu X, et al. BDNF Val66Met polymorphism, life stress and depression: a meta-analysis of gene-environment interaction. *J Affect Disord* (2018) 227:226–35. doi: 10.1016/j.jad.2017.10.024
- Zhao M, Yang J, Wang W, Ma J, Zhang J, Zhao X, et al. Meta-analysis of the interaction between serotonin transporter promoter variant, stress, and posttraumatic stress disorder. *Sci Rep* (2017) 7(1):16532. doi: 10.1038/s41598-017-15168-0
- Stallings MC, Hewitt JK. Conceptualization and measurement of organism-environment interaction. *Behav Genet* (1994) 24(1):103–4. doi: 10.1007/BF01067934
- Douglas W. Insensitivity of the analysis of variance to heredity-environment interaction. *Behav Brain Sci* (1990) 13(1):109–20. doi: 10.1017/s0140525x00077797
- Kaufman J, Yang BZ, Douglas-Palumberi H, Houshyar S, Lipschitz D, Krystal JH, et al. Social supports and serotonin transporter gene moderate depression in maltreated children. *Proc Natl Acad Sci U States America* (2004) 101(49):17316–21. doi: 10.1073/pnas.0404376101
- Kendler KS, Kuhn JW, Vittum J, Prescott CA, Riley B. The interaction of stressful life events and a serotonin transporter polymorphism in the

- prediction of episodes of major depression: a replication. *Arch Gen Psychiatry* (2005) 62(5):529–35. doi: 10.1001/archpsyc.62.5.529
40. Wilhelm K, Mitchell PB, Niven H, Finch A, Wedgwood L, Scimone A, et al. Life events, first depression onset and the serotonin transporter gene. *Br J Psychiatry: J Ment Sci* (2006) 188:210–5. doi: 10.1192/bjp.bp.105.009522
 41. Gillespie NA, Whitfield JB, Williams B, Heath AC, Martin NG. The relationship between stressful life events, the serotonin transporter (5-HTTLPR) genotype and major depression. *Psychol Med* (2005) 35(1):101–11. doi: 10.1017/s0033291704002727
 42. Eley TC, Sugden K, Corsico A, Gregory AM, Sham P, McGuffin P, et al. Gene-environment interaction analysis of serotonin system markers with adolescent depression. *Mol Psychiatry* (2004) 9(10):908–15. doi: 10.1038/sj.mp.4001546
 43. Grabe HJ, Lange M, Wolff B, Volzke H, Lucht M, Freyberger HJ, et al. Mental and physical distress is modulated by a polymorphism in the 5-HT transporter gene interacting with social stressors and chronic disease burden. *Mol Psychiatry* (2005) 10(2):220–4. doi: 10.1038/sj.mp.4001555
 44. Sjöberg RL, Nilsson KW, Nordquist N, Ohrvik J, Leppert J, Lindström L, et al. Development of depression: sex and the interaction between environment and a promoter polymorphism of the serotonin transporter gene. *Int J Neuropsychopharmacol* (2006) 9(4):443–9. doi: 10.1017/S1461145705005936
 45. Lima IVM, Sougey EB, Filho HPV. Genetics of affective disorders. *Rev Psiquiatria Clínica* (2004) 31(1):34–9. doi: 10.1016/0022-3956(92)90033-K
 46. McGuffin P, Katz R, Bebbington P. The camberwell collaborative depression study. III. Depression and adversity in the relatives of depressed probands. *Br J Psychiatry: J Ment Sci* (1988) 152:775–82. doi: 10.1192/bjp.152.6.775
 47. Wong MY, Day NE, Luan JA, Chan KP, Wareham NJ. The detection of gene-environment interaction for continuous traits: should we deal with measurement error by bigger studies or better measurement? *Int J Epidemiol* (2003) 32(1):51–7. doi: 10.1093/ije/dyg002
 48. Hardt J, Rutter M. Validity of adult retrospective reports of adverse childhood experiences: review of the evidence. *J Child Psychol Psychiatry Allied Disciplines* (2004) 45(2):260–73. doi: 10.1111/j.1469-7610.2004.00218.x
 49. De Bellis MD. Developmental traumatology: the psychobiological development of maltreated children and its implications for research, treatment, and policy. *Dev Psychopathol* (2001) 13(3):539–64. doi: 10.1017/s0954579401003078
 50. Flugge G, Van Kampen M, Mijster MJ. Perturbations in brain monoamine systems during stress. *Cell Tissue Res* (2004) 315(1):1–14. doi: 10.1007/s00441-003-0807-0

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Lower Serum Uric Acid Is Associated With Post-Stroke Depression at Discharge

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OPEN ACCESS

Edited by:

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Specialty section:

This article was submitted to
Molecular Psychiatry,
a section of the journal
Frontiers in Psychiatry

Received: 11 December 2019

Accepted: 21 January 2020

Published: 18 February 2020

Citation:

Li G, Miao J, Sun W, Song X, Lan Y,
Zhao X, Qiu X, Zhang C, Zhu Z and
Zhu S (2020) Lower Serum Uric Acid Is
Associated With Post-Stroke
Depression at Discharge.
Front. Psychiatry 11:52.
doi: 10.3389/fpsy.2020.00052

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Background: Serum uric acid (SUA) has been shown to play an important role in the pathophysiology of mood disorders including 3- and 6-month post-stroke depression (PSD). This study aimed to investigate whether SUA levels on admission were associated with PSD at discharge.

Methods: A total of 498 stroke patients were consecutively recruited from Tongji Hospital. Clinical and laboratory test data were collected on admission. They were categorized into equal tertiles according to the distribution of SUA and the number of patients. PSD status was evaluated by DSM-V criteria and 17-item Hamilton Rating Scale for Depression at discharge.

Results: The optimal cut-off points of SUA were: (T1) 80.00~300.80 $\mu\text{mol/L}$, (T2) 300.81~391.67 $\mu\text{mol/L}$, (T3) 391.68~710.0 $\mu\text{mol/L}$. A total of 232 patients (46.59%) were diagnosed as PSD at discharge. Significant differences were found between the PSD and non-PSD groups in SUA tertiles of patients ($P = 0.00$). After adjustment for conventional confounding factors, the odds ratios of PSD were 5.777 (95% CI = 3.463~9.637, $P = 0.00$) for the lowest tertile and 4.153 (95% CI = 2.492~6.921, $P = 0.00$) for the middle tertile of SUA, as compared with the highest tertile. In restricted cubic spline regression, continuous SUA showed linear relation with PSD risk at discharge after 300 $\mu\text{mol/L}$.

Conclusions: Lower SUA levels on admission were found to be associated with PSD at discharge and the threshold effect was also revealed. For stroke patients, doctors should pay attention to the baseline SUA for screening high-risk PSD at discharge in clinical practice.

Keywords: post-stroke depression, serum uric acid, restricted cubic spline regression, antioxidants, threshold effect

INTRODUCTION

Post-stroke depression (PSD) is the most frequent neuropsychiatric sequela after stroke, affecting about 40% of stroke patients (1, 2). It is well known that PSD was associated with reduced quality of life (QoL), poorer functional outcome, as well as increased cost of treatment, burden of family caregiver, and mortality (1, 3–5). Moreover, a recent meta-analysis reported that patients with early PSD had a mortality risk about 1.5 higher than non-depressed individuals, considering both short- and long-term mortality (6). The depression effects on cardiovascular risk, such as unhealthy lifestyle behaviors and lower adherence to treatment, the higher stroke severity, possible negative effects of a SSRI treatment on survival, and non-natural causes of death like suicide may be the potential interpretations (6). Although the importance of PSD has been well documented and there are validated screening tools for PSD, many PSD patients cannot be diagnosed and rate of refusal by busy stroke clinicians to recommend antidepressant treatment remain high. One reason is that there are no reliable objective biomarkers for diagnosing and predicting PSD.

To date, there is a lack of understanding of what the exactly underlying pathophysiological mechanisms of PSD are (7). Among these discovered biological factors, oxidative and nitrosative stress pathways are regarded as the most conclusive factors because there are low antioxidant levels and high metabolic rates and levels in the brain (8, 9). As we know, the brain is a site of excessive reactive oxygen species (ROS) production and especially vulnerable to oxidative stress, as it accounts for 20% oxygen consumed by the body (10). Hence, brain neurons damaged by oxidate stress may lead to altered membrane structure and function, which may further affect the expression of membrane receptors leading to increased risk of depression (11, 12). Serum uric acid (SUA) is an important antioxidant which is the end product in the degradation of the purine nucleotides adenine and guanine, accounting approximately 60% of the total antioxidant capacity in plasma (13). Meanwhile, SUA is a low-cost indicator which can be easily obtained on admission. A recent meta-analysis has shown that subjects with major depressive disorders (MDD) have levels of the antioxidant uric acid (UA) lower than healthy controls (14). Moreover, previous studies have shown that the lower SUA level is associated with both PSD and depression (9, 15). However, the assessment on the association between SUA and PSD only aimed at 3- and 6-month outcomes. It is still lacking objective and quantitative biological factors to assess PSD at discharge. When the early onset of PSD is ignored by clinicians, it will adversely affect the early rehabilitation after discharge, which is the most important stage for stroke rehabilitation. Thus, we need effective biological predictors in order to more efficiently identify and diagnose PSD at discharge, and then intervene to promote early recovery of stroke.

Therefore, considering that SUA is proven to be an effective predictor of 3- and 6-month PSD, we hypothesized that the SUA may also serve as a predictor of patients with PSD at discharge. The aim of this study was to assess the association between SUA on admission and the PSD outcome at discharge.

METHODS

Study Design

All first-ever stroke patients were consecutively recruited within 7 days of the onset of symptoms from the Tongji Hospital which is located in Wuhan City, Hubei Province, China, between May 2018 and October 2019. SUA was obtained within 24 h after admission. There are few clinical concerns about the decreased odds ratio (OR) value in PSD for each unit change of UA, which could be obtained by taking SUA as a continuous measure. Therefore, we divided the SUA into tertiles to observe whether any enhanced performance could be quantified while maintaining statistical effect in each category, according to the patients' amount and the skewed distribution of the raw SUA values (7, 16). All patients involved in this study or their family members gave written informed consents according to the Declaration of Helsinki. The approval of the study for experiments was obtained from the Ethics Committee of Tongji Medical College, Huazhong University of Science and Technology.

The registration number of this prospective cohort study was ChiCTR-ROC-17013993. The URL of the publicly accessible website on which this trial is registered is: <http://www.chictr.org.cn/index.aspx>. The original protocols used for this *post hoc* analysis did not include SUA as a potential predictor.

Inclusion and Exclusion Criteria

All suspected stroke patients were confirmed by magnetic resonance imaging or computerized tomography within 7 days after admission and the following inclusive criteria were used: (i) males and females, age ≥ 18 years old, first-ever diagnosed stroke patients, including ischemic and hemorrhagic stroke; (ii) hospitalized within 7 days after stroke onset; (iii) written informed consent was provided.

The exclusive criteria were as follows: (i) brain dysfunction caused by other non-vascular causes, such as primary brain tumors, subdural hematoma, paralysis after seizures, metastatic encephaloma, brain trauma, etc.; (ii) history of depression (previous treatment history or clinical diagnosis), dementia and/or other psychiatric illness; (iii) communication problems due to aphasia, dysarthria, disturbance of understanding or consciousness (a Mini-Mental State Examination score was <19 points, in particular the MMSE score of illiterate patients was <17 points); (iv) unable to complete the follow up; (v) Transient ischemic attack and subarachnoid hemorrhage; (vi) with other concomitant neuropsychiatric diseases, such as Parkinson's disease and epilepsy.

Serum samples were collected at room temperature on admission, then centrifuged at 3,500 r/min for 10 min, which could be used to measure the levels of serum biomarkers. Depressive symptoms were measured at discharge, while the baseline sociodemographic information, clinical characteristics, and routine laboratory indicators were collected on admission.

Data Collection and Follow-Up

Standard patient demographic data was collected with a case report form at baseline, covering gender, age, body mass index

(BMI), marital status, degree of education, and vascular risk factors, including smoking, drinking, and history of stroke, diabetes mellitus, hypertension, hyperlipidemia, coronary heart disease, and surgery. The stroke severity was assessed within 24 h of hospital admission by well-trained doctors using the National Institutes of Health Stroke Scale (NIHSS) score. Barthel Index (BI) score, mRS score, and hospitalization days were also included into the variables. The concentrations of SUA, serum albumin (ALB), homocysteine (Hcy), hypersensitive C-reactive protein (Hs-CRP) were measured by standard autoanalyzer techniques with a Roche automatic analyzer (cobas c 701) in clinical lab of Tongji Hospital. The inflammatory factors, including IL-1 β , IL-6, IL-10, IL-18, TNF- α , BDNF, and IFN- γ , were measured using a solid-phase sandwich enzyme-linked immunosorbent assay kit (CUSABIO, China) according to the manufacturer's specifications in Kindstar Company, Wuhan. To minimize assay variance, all samples were analyzed on the same day in duplicate in a random order by a technician blind to the clinical diagnoses; the intra-assay coefficients were <5%.

Psychological Measurement

All psychological evaluations were performed by two experienced psychiatrists (X.S. and W.S.) who were blinded to other clinical and laboratory findings after receiving standardized training. The interrater reliability reached an acceptable level. PSD was diagnosed by a psychiatrist at discharge according to DSM-V criteria. Seventeen-item Hamilton Rating Scale for Depression (HRSD) was used to measure the degree of PSD at discharge. The DSM-V diagnostic criteria (Depressive Disorder Due to Another Medical Condition) was met, and HRSD score ≥ 7 at discharge, which was regarded as the primary endpoint. Patients were divided into PSD group and non-PSD group according to whether they had PSD outcome or not. The validity and reliability of the Chinese HRSD version had been proven in previous studies (17).

Statistical Analysis

Results were expressed as percentages for categorical variables and as medians [interquartile range (IQR)] or means \pm standard deviation (S.D.) for the continuous variables, depending on the normal or nonnormal distribution of data by Kolmogorov-Smirnov test. Proportions were compared using the Chi-squared test, Student's *t*-test, and analysis of variance (ANOVA) were employed for the normally distributed variables, while the Mann-Whitney U-test was used for the asymmetrically distributed variables. Statistical comparisons among SUA stratification were assessed by Pearson's Chi-square test or Fisher's exact test for categorical variables, and continuous variables were evaluated by Kruskal-Wallis test or ANOVA. After adjusting for main baseline variables identified in the univariate logistic regression analysis and traditional confounders related to PSD, the OR values and 95% confidence intervals (95% CIs) for PSD risk were obtained by multivariate-adjusted binary logistic regression. The restricted cubic spline (RCS) regression was

used to test the linear association between PSD and SUA as a continuous measure with three knots.

All statistical analysis was performed with R version 3.5.2 and SPSS for Windows, version 22.0 (SPSS Inc., Chicago, IL, USA). Statistical significance was identified as a two-tailed *P* values less than 0.05 ($P < 0.05$). The R packages "rms", "Hmisc", "Formula" and "ggplot2" were applied.

RESULTS

Baseline Characteristics of all Patients in SUA Tertiles

A total of 707 stroke patients from Tongji Hospital between May 2018 and October 2019 were recruited in the study, and 567 patients were eligible for the research. By the time of discharge, there were 69 patients failed to be followed up. Ultimately, we included a total of 498 stroke patients, which consisted of 379 males (76.10%) and 119 females (23.90%). Their mean age was 57.17 ± 10.88 years (**Table 1**). We divided all cases into three groups according to tertiles of SUA levels, which ensured the most categories with adequate number of patients per subgroups between the range of 80.00 and 710.00 $\mu\text{mol/L}$ (T1, 166 patients; T2, 166 patients; T3, 166 patients).

The cut-off values for this stratification on the SUA level into tertiles were: (T1) 80.00~300.80 $\mu\text{mol/L}$, (T2) 300.81~391.67 $\mu\text{mol/L}$, (T3) 391.68~710.0. **Table 1** summarized the characteristics of the patients by the tertiles of SUA, including the sociodemographic, clinical, and laboratory characteristics. Patients with the low, moderate, and high SUA were significantly different in following sociodemographic and clinical variables: age, gender, BMI, smoking, drinking, history of diabetes and hypertension, HRSD score at discharge, baseline NIHSS score, BI score, and mRS score. The majority of patients had drinking (60.64%) and smoking habits (59.64%) and the lowest and middle SUA tertiles were significantly associated with higher HRSD score than the highest tertile ($P = 0.00$). And there were statistical differences observed for ALB, Hcy, Hs-CRP, IL-1 β , and IL-18 in the SUA tertiles of patients (all *P* values < 0.05). Some significant differences in baseline characteristics were reasonable because the patients were consecutively recruited.

Baseline Characteristics of Patients in PSD Group and Non-PSD Group

The baseline characteristics between PSD and non-PSD groups are presented in **Table 2**. In this study, 232 (46.59%) patients were diagnosed as PSD at discharge. Compared with non-PSD group, PSD patients were more likely to be older and with lower educational level and BI scores, higher proportion of coronary artery disease history, surgery history, HRSD scores, baseline NIHSS, and mRS scores.

We also divided the SUA levels into hyperuricemic and non-hyperuricemic groups, in which hyperuricemia is defined as 416.5 $\mu\text{mol/L}$ (7.0 mg/dl) or higher in males, and 357 $\mu\text{mol/L}$ (6.0 mg/dl) or higher in females. The association between

TABLE 1 | Baseline characteristics of patients with stroke according to SUA tertiles.

| Variables | All patients | SUA tertiles | | | P-value |
|---------------------------------------|-----------------------------|-------------------------------------|---------------------------------------|--------------------------------------|---------|
| | | Tertile 1, n = 166 (80.0–300.80) | Tertile 2, n = 166 (300.81–391.67) | Tertile 3, n = 166 (391.68–710.0) | |
| UA, median (IQR) | 328.0 (275.7, 410.00) | 253.00 (215.25, 276.00) | 328.00(328.00, 361.63) | 410.00 (410.00, 448.00) | 0.000 |
| Demographic parameters | | | | | |
| Age (years) | 57.17 ± 10.88 | 58.25 ± 9.88 | 58.01 ± 11.34 | 55.23 ± 11.17 | 0.019 |
| Females, n (%) | 119 (23.90) | 57 (34.34) | 34 (20.48) | 28 (16.87) | 0.000 |
| BMI (kg/m ²) | 24.96 ± 3.54 | 24.25 ± 3.27 | 25.05 ± 3.69 | 25.59 ± 3.53 | 0.002 |
| Married, n (%) | 483 (96.99) | 161 (96.99) | 160 (96.39) | 162 (97.59) | 0.814 |
| Education level | | | | | 0.111 |
| Junior middle school and below, n (%) | 306 (61.45) | 114 (68.67) | 99 (59.64) | 93 (56.02) | 0.051 |
| Senior high/polytechnic school, n (%) | 126 (25.30) | 38 (22.89) | 42 (25.30) | 46 (27.72) | 0.601 |
| Bachelor and above, n (%) | 66 (13.25) | 14 (8.43) | 25 (15.06) | 27 (16.27) | 0.077 |
| Vascular risk factors | | | | | |
| Smoking, n (%) | 297 (59.64) | 79 (47.59) | 96 (57.83) | 122 (73.49) | 0.000 |
| Drinking, n (%) | 302 (60.64) | 87 (52.41) | 104 (62.65) | 111 (66.87) | 0.021 |
| History of diabetes, n (%) | 106 (21.29) | 45 (21.11) | 41 (24.70) | 20 (12.05) | 0.002 |
| History of hypertension, n (%) | 273 (54.82) | 88 (53.01) | 103 (62.05) | 82 (49.40) | 0.058 |
| History of hyperlipidemia, n (%) | 109 (21.89) | 27 (16.27) | 45 (27.11) | 37 (22.29) | 0.057 |
| Coronary artery diseases, n (%) | 51 (10.24) | 17 (10.24) | 19 (11.45) | 15 (9.04) | 0.769 |
| History of previous stroke, n (%) | 92 (18.47) | 25 (15.06) | 32 (19.28) | 35 (21.08) | 0.349 |
| History of surgery, n (%) | 158 (31.73) | 61 (36.75) | 54 (32.53) | 43 (25.90) | 0.101 |
| Clinical characteristics | | | | | |
| NIHSS score, median (IQR) | 3 (2, 6.5) | 4 (2, 8) | 4 (2, 7) | 3 (1, 5) | 0.001 |
| BI score, median (IQR) | 85 (45, 100) | 72.5 (40, 100) | 80 (48.75, 100) | 95 (65, 100) | 0.000 |
| mRS score, median (IQR) | 2 (1, 4) | 3 (2, 4) | 3 (1, 4) | 2 (1, 3) | 0.000 |
| HRSD score, median (IQR) | 7 (4, 12) | 9 (5, 13.25) | 9.0 (4.75, 13.00) | 5 (3, 7) | 0.000 |
| Hospitalization days, median (IQR) | 9 (7, 16) | 9 (7, 14) | 10 (7, 16) | 9.5 (6, 16) | 0.926 |
| Serum biochemicals | | | | | |
| Albumin, median (IQR) | 41.07 (39.60, 42.60) | 40.65 (38.18, 42.63) | 41.07 (40.10, 42.80) | 41.07 (41.07, 42.13) | 0.013 |
| Hcy, median (IQR) | 15.10 (11.70, 15.3) | 12.60 (10.30, 15.10) | 15.10 (12.15, 15.10) | 15.10 (14.50, 16.95) | 0.000 |
| Hs-CRP, median (IQR) | 3.85 (1.10, 8.26) | 2 (0.60, 8.26) | 3.45 (1.10, 8.26) | 8.26 (1.98, 8.26) | 0.000 |
| IL-1 β , median (IQR) | 61.14 (18.62, 135.82) | 50.43 (14.27, 135.82) | 50.51 (18.77, 135.82) | 119.01 (28.25, 163.83) | 0.003 |
| IL-6, median (IQR) | 4.48 (1.93, 9.47) | 3.63 (1.86, 9.30) | 5.03 (2.08, 9.47) | 4.75 (1.88, 9.47) | 0.730 |
| IL-10, median (IQR) | 8.72 (2.30, 23.46) | 8.08 (2.10, 18.17) | 9.26 (2.27, 23.80) | 11.26 (2.48, 29.68) | 0.268 |
| IL-18, median (IQR) | 1,773.41 (568.82, 3,520.77) | 1,503.89 (397.79, 3,520.77) | 1,641.50 (575.27, 3,520.77) | 2,552.29 (780.13, 3,534.55) | 0.043 |
| TNF- α , median (IQR) | 36.73 (18.19, 49.53) | 36.75 (19.32, 51.51) | 38.53 (19.10, 54.59) | 33.32 (15.33, 43.33) | 0.062 |
| BDNF, median (IQR) | 3.60 (1.96, 5.78) | 3.21 (1.63, 5.92) | 3.67 (2.18, 5.72) | 3.86 (2.14, 6.13) | 0.251 |
| IFN- γ , median (IQR) | 4.43 (1.73, 9.72) | 4.50 (1.54, 8.99) | 4.41 (1.83, 8.40) | 4.32 (1.71, 9.79) | 1.000 |

BMI, body mass index; BI, Barthel Index; mRS, modified Rankin Scale; NIHSS, National Institutes of Health Stroke Scale; SUA, serum uric acid; Hcy, homocysteine; Hs-CRP, hyper-sensitive C-reactive protein; TNF, tumor necrosis factor; BDNF, brain derived neurotrophic factor; IFN, interferon; IL, interleukin.

hyperuricemic and PSD was explored by univariate and multivariate adjusted logistic regression models. We could speculate that hyperuricemia might protect from PSD from the regression results (OR \approx 0.42, 95% CI \approx 0.26–0.69, P = 0.001).

Moreover, we considered the ALB level as the representative index of overall nutritional status. There were differences in ALB levels across the SUA tertiles, as shown in **Table 1**. The *post hoc* test after Mann-Whitney U test declared that the ALB in lowest tertile SUA group was significantly less than the middle and the highest tertile SUA groups (both P = 0.032). Moreover, there was no difference in ALB levels between PSD and non-PSD groups (P = 0.140, **Table 2**). The subjects in the lowest tertile SUA group may be with worse nutritional status and decreased food intake. It reminds that there may be more risk factors, such as unhealthy

lifestyles rather than ALB itself, leading to PSD prior to admission in the lowest SUA tertile group.

Association Between the Level of SUA and PSD

Significant differences were found between the PSD and non-PSD groups in SUA tertiles of patients (all P < 0.05, **Table 3**). Indeed, the proportions of patients in the lowest tertile (80.00–300.80 $\mu\text{mol/L}$, P = 0.001) and the middle tertile (300.81–391.67 $\mu\text{mol/L}$, P = 0.00) were significantly higher in the PSD groups, whilst the proportion of patients in the highest tertile (391.68–710.00 $\mu\text{mol/L}$) was significantly lower in the PSD group (P = 0.00) (**Table 3**). In **Table 4**, with all patients taken as a whole, PSD occurrence taken as a dependent variable

TABLE 2 | Clinical and demographic characteristics of patients with PSD and non-PSD.

| Variables | PSD patients (n = 232) | Non-PSD patients (n = 266) | P-value |
|--|-----------------------------|-------------------------------|---------|
| SUA | 328 (266.08, 348.45) | 389 (295.5, 410) | 0.000 |
| Demographic parameters | | | |
| Age (years) | 58.34 ± 10.49 | 56.14 ± 11.13 | 0.024 |
| Females, n (%) | 59 (25.43) | 60 (22.56) | 0.453 |
| BMI (kg/m ²) | 24.83 ± 3.26 | 25.08 ± 3.76 | 0.426 |
| Married, n (%) | 225 (96.98) | 258 (96.99) | 0.995 |
| Education level | | | 0.018 |
| Junior middle school and below, n (%) | 155 (66.81) | 151 (56.77) | |
| Senior high/Polytechnic school, n (%) | 56 (24.14) | 70 (26.32) | |
| Bachelor/Junior college and above, n (%) | 21 (9.05) | 45 (16.92) | |
| Vascular risk factors | | | |
| Smoking, n (%) | 133 (57.33) | 164 (61.65) | 0.326 |
| Drinking, n (%) | 139 (59.91) | 163 (61.28) | 0.756 |
| History of diabetes, n (%) | 52 (22.41) | 54 (20.30) | 0.566 |
| History of hypertension, n (%) | 131 (56.47) | 142 (53.38) | 0.491 |
| History of hyperlipidemia, n (%) | 52 (22.41) | 57 (21.43) | 0.791 |
| Coronary artery diseases, n (%) | 31 (13.36) | 20 (7.52) | 0.032 |
| History of previous stroke, n (%) | 46 (19.83) | 46 (17.29) | 0.467 |
| History of previous surgery, n (%) | 84 (36.21) | 74 (27.82) | 0.045 |
| Clinical characteristics | | | |
| NIHSS score, median (IQR) | 5 (2, 8) | 2 (1, 4) | 0.000 |
| BI score, median (IQR) | 65 (35, 95) | 95 (68.75, 100) | 0.000 |
| mRS score, median (IQR) | 3 (2, 4) | 2 (1, 3) | 0.000 |
| HRSD score, median (IQR) | 12 (10, 16) | 4 (2, 6) | 0.000 |
| Hospitalization days, median (IQR) | 10 (7, 15) | 9 (6, 16) | 0.486 |
| Serum biochemicals | | | |
| Albumin, median (IQR) | 41.07 (39.20, 42.48) | 41.07 (40.30, 42.80) | 0.140 |
| Homocysteine, median (IQR) | 14.65 (11.73, 15.10) | 15.10 (11.70, 15.40) | 0.353 |
| Hs-CRP, median (IQR) | 3.50 (1.10, 8.26) | 4.45 (1.10, 8.26) | 0.712 |
| IL-1β, median (IQR) | 55.51 (18.90, 135.82) | 67.32 (18.19, 135.91) | 0.467 |
| IL-6, median (IQR) | 4.75 (1.84, 9.47) | 4.28 (1.97, 9.47) | 0.377 |
| IL-10, median (IQR) | 8.15 (1.95, 23.24) | 9.50 (2.50, 23.64) | 0.620 |
| IL-18, median (IQR) | 1,681.96 (503.52, 3,520.77) | 1,938.46 (680.12, 3,520.77) | 0.496 |
| TNF-α, median (IQR) | 36.84 (18.32, 50.95) | 36.04 (17.66, 48.50) | 0.710 |
| BDNF, median (IQR) | 3.34 (1.85, 5.75) | 3.80 (2.12, 5.93) | 0.393 |
| IFN-γ, median (IQR) | 4.56 (1.84, 9.73) | 4.21 (1.71, 8.71) | 0.357 |

BMI, body mass index; BI, Barthel Index; mRS, modified Rankin Scale; NIHSS, National Institutes of Health Stroke Scale; SUA, serum uric acid; Hcy, homocysteine; Hs-CRP, hyper-sensitive C-reactive protein; TNF, tumor necrosis factor; BDNF, brain-derived neurotrophic factor; IFN, interferon; IL, interleukin.

TABLE 3 | SUA tertiles of patients.

| Variable | PSD patients (n = 232) | Non-PSD patients (n = 266) | χ^2 | P-value |
|---|---------------------------|-------------------------------|----------|---------|
| SUA | | | 65.672 | 0.000 |
| Tertile 1, n = 166 (80~300.8 μmol/L) | 95 | 71 | 11.334 | 0.001 |
| Tertile 2, n = 166 (300.81~391.67 μmol/L) | 102 | 64 | 22.095 | 0.000 |
| Tertile 3, n = 166 (391.68~710 μmol/L) | 35 | 131 | 65.078 | 0.000 |

SUA, serum uric acid; PSD, post-stroke depression.

TABLE 4 | Multivariate adjusted odds ratios for the association between SUA levels and PSD at discharge.

| | Tertile | OR ^a | 95% CI | P-value |
|----------------------|----------|-----------------|-------------|---------|
| Unadjusted | Lowest | 5.965 | 3.667~9.704 | 0.00 |
| | Moderate | 5.008 | 3.089~8.120 | 0.00 |
| Model 1 ^b | Lowest | 6.009 | 3.682~9.808 | 0.00 |
| | Moderate | 4.776 | 2.937~7.767 | 0.00 |
| Model 2 ^c | Lowest | 6.009 | 3.674~9.829 | 0.00 |
| | Moderate | 4.808 | 2.949~7.838 | 0.00 |
| Model 3 ^d | Lowest | 5.777 | 3.463~9.637 | 0.00 |
| | Moderate | 4.153 | 2.492~6.921 | 0.00 |

OR, odds ratio; CI, confidence level; PSD, post-stroke depression.

^aReference OR (1.000) is the highest tertile of SUA for PSD.

^bModel 1: adjusted for age, gender, education levels, body mass index, smoking, drinking.

^cModel 2: adjusted for covariates from model 1 and further adjusted for medical history (coronary artery disease, diabetes mellitus, hyperlipidemia, hypertension, previous stroke, and surgery).

^dModel 3: adjusted for covariates from model 2 and further adjusted for baseline NIHSS scores, mRS scores, and Barthel Index scores.

and highest tertile taken as the reference was used for SUA in the unadjusted and multivariate adjusted logistic regression models. In unadjusted logistic regression model, the lowest tertile of SUA was independently consistent with a risk predictor of PSD with an unadjusted OR of 5.965 (95% CI = 3.667~9.704, $P = 0.00$). The middle tertile of SUA was independently consistent with a risk predictor of PSD with an unadjusted OR of 5.008 (95% CI = 3.089~8.120, $P = 0.00$).

After adjusting for conventional confounders including age, gender, education levels, BMI, smoking, drinking, history of coronary artery disease, diabetes mellitus, hyperlipidemia, hypertension, previous stroke and surgery, baseline NIHSS, mRS and BI scores, and the lowest tertile of SUA was remained significant independently associated with the prevalence of PSD (model 1: OR = 6.009, 95% CI = 3.682~9.808, $P = 0.00$; model 2: OR = 6.009, 95% CI = 3.674~9.829, $P = 0.00$; model 3: OR = 5.777, 95% CI = 3.463~9.637, $P = 0.00$), as compared with the highest tertile. Similarly, the middle tertile of SUA was remained significant independently associated with the prevalence of PSD (model 1: OR = 4.776, 95% CI = 2.937~7.767, $P = 0.00$; model 2: OR = 4.808, 95% CI = 2.949~7.838, $P = 0.00$; model 3: OR = 4.153, 95% CI = 2.492~6.921, $P = 0.00$).

Furthermore, we used RCS regression model to confirm the linear relationship between continuous SUA levels and PSD at discharge ($\chi^2 = 6.33$, $df = 2$, $P = 0.0119$ for nonlinearity,

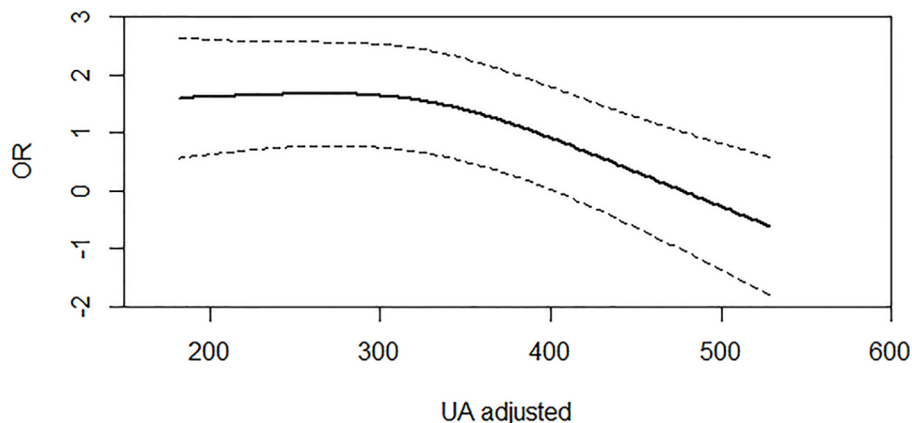


FIGURE 1 | Association of serum UA (uric acid) levels with risk of PSD. Dashed lines are 95% confidence intervals. Odds ratios and 95% confidence intervals derived from restricted cubic spline regression (P for nonlinear = 0.0119). Odds ratios were estimated using logistic regression modeling, adjusting for the same variables as model 3 in **Table 4**.

Figure 1). The RCS curve showed that there was nonlinear effect between SUA and PSD outcome. However, the linear association was found between SUA and PSD after the value of around 300 $\mu\text{mol/L}$. Coincidentally, the cut-off point of RCS curve (300 $\mu\text{mol/L}$) was approximately the same as the first tertile split points (300.8 $\mu\text{mol/L}$). It is a reasonable approach to take the tertiles of SUA levels to analyze the association between SUA and PSD risk. There is a protective effect in PSD when the SUA level was more than 480 $\mu\text{mol/L}$, while the 95% CI of other SUA values (≤ 480 $\mu\text{mol/L}$) is not completely above or below the value “1” of significance.

DISCUSSION

In this study, we have investigated the association between SUA levels on admission and the development of PSD outcome at discharge using a prospective cohort. Our results suggested that lower levels of SUA on admission were associated with the presence of PSD at discharge, and documented that the risk of PSD in patients significantly declined with the increasing SUA level after 300 $\mu\text{mol/L}$. Therefore, our findings revealed the SUA value could provide important information for predicting PSD patients at discharge. A total of 46.59% of stroke patients presented with PSD in this study, which is more than the incidence reported in previous studies. This could be potentially explained by that Tongji hospital was the highest-ranked large general 3A hospital in Central China and recruited more serious stroke patients whose the median NIHSS score (IQR) was 3 (2, 6.5) (7). Previous study has reported the higher levels of SUA on admission was associated with the occurrence of 3-month major PSD, while lower level of SUA on admission was closely related to the occurrence of major PSD between 3- and 6-month post-stroke. Six months after acute ischemic stroke, there was no relationship between major PSD and baseline SUA (15). The differences in time of assessment and diagnosis, subjects'

samples, and psychiatric assessment methods could explain the discrepancies between our study and their findings.

Compared with the patients without PSD at discharge, our results also demonstrated that the PSD patients had significantly increased stroke severity (higher NIHSS scores) and worse functional outcome (higher mRS scores and lower BI scores), which were consistent with the previous studies (18, 19). The patients from PSD group were also more likely to have history of coronary artery diseases and surgery which may potentially worsen the stroke patients' conditions and increase the risk of occurrence of PSD at discharge (20). Furthermore, the incidence of PSD was higher in older stroke patients, which was also consistent with the previous reports (21). This was probably because older patients were more likely to be involved in a depressive mood when dealing with the stress caused by stroke. The increased risk of PSD at discharge could also be caused by the worsening functional and cognitive impairment in the elderly (22). Previous studies have indicated that proinflammatory factors played important role in the development of PSD (23). However, there were no significant differences in proinflammatory factors between PSD and non-PSD groups which was potentially due to the different definitions of outcomes.

In recent years, the interest of psychiatrist in the association between oxidative stress and depression has grown stronger and relevant studies have shown that the depression is accompanied by increased oxidative stress and decreased antioxidant defenses (24, 25). Unlike previous studies, the present investigation focused on the occurrence of PSD at discharge and attempted to link this outcome with SUA levels on admission (15). Our study may provide an effective biomarker for psychiatrists to diagnose PSD at discharge.

Given that lower SUA had been found in major depressive and anxiety disorders (26), our study reasonably found PSD patients had lower SUA levels on admission. As a crucial central nervous system antioxidant, SUA had been proven to be an

effective predictor in the development of PSD (15). Individuals with lower baseline SUA levels may be more susceptible to oxidative damage in the brain, as a result of lower antioxidant defenses and larger oxygen consumption in brain. This damage could make individuals susceptible to developing depressive mood and has also been suggested as a potential mechanism in the relapse of depression (27). As a free radical scavenger, SUA contributes to over half antioxidant capacity in plasma. As neuronal cell membranes were composed by large amounts of polyunsaturated fatty acids with a large surface area and ROS often react with lipids, optimal neuronal cell function depends on sufficient antioxidants, such as SUA, to remove ROS and protect neuronal cells from oxidative damage. Nanetti et al. found that acute ischemic stroke could cause the strong oxidative stress and generation of free radicals, and that this progression of ischemic injury could be limited by the antioxidant capacity (28). Thus, higher SUA levels could potentially protect neuron integrity and function of stroke patients and decrease the risk of PSD at discharge. Previous studies have also reported that UA has neuroprotective effects due to its potent antioxidant capacity in other diseases, such as multiple sclerosis, Alzheimer's disease, and Parkinson's disease (29–31). In acute stage of stroke, UA may exert antioxidant protection against free radical damage. Previous study has shown a negative correlation between SUA levels and stroke outcomes (32). A randomized clinical trial also showed that UA therapy was related to an improved prognosis in patients with hyperglycemia in acute stage through decreasing glucose-driven oxidative stress (33). Moreover, a phase 2 clinical trial with inosine in Parkinson's disease demonstrated a potential efficacy of increased UA on mood disorders as evaluated on the Geriatric Depression Scale. We could speculate that SUA (>480 $\mu\text{mol/L}$) may exert protective effect in PSD (34).

Moreover, UA is also a marker of purine metabolism as the final product other than antioxidant. The purinergic system has been associated with the pathophysiology of depression and is thought to influence mood, appetite, sleep, cognition, and drive through the neuromodulator adenosine and the neurotransmitter adenosine triphosphate, both of which are upstream metabolites of UA (35). Increased levels of UA are associated with decreased adenosinergic transmission and accelerated purinergic transformation, which could limit the development of depressive disorders (36). Previous studies also suggested that lower SUA levels at the onset of ischemic stroke could indicate a worse prognosis, such as increased risk of PSD, by increasing the neurological damage levels (37, 38). On the other hand, previous study also reported that the lower UA in current MDD may be related to increased exposure to ROS through mitochondrial dysfunction which could deplete UA, if not caused by lifestyle factors (39). As we all know, mitochondrial dysfunction also plays an important role in disorders of purine metabolism which could not provide sufficient UA to counteract increased oxidative stress in PSD (40).

The main strengths of this study are the sufficient sample size, well-established psychiatric diagnoses, prospective cohort study nature, and adjustment for many confounders in different models. Though further longitudinal studies are needed to

confirm the association between the SUA level on admission and the risk of PSD at discharge, our study results revealed the threshold effect between them. Moreover, SUA presented affordable routine clinical biomarker which did not incur additional expense as compared with microRNAs which is limited to precise laboratory tests and not appropriate for extensive follow up and clinical application (18). Some limitations of our study should also be acknowledged. First, the stroke patients from a single center and exclusion of patients with alteration of consciousness level and severe speech disturbances may result in biases for the incidence of PSD at discharge. Second, it should be noted that the functioning of antioxidant defenses and oxidative stress is an ongoing dynamic process, the complexities of which cannot be reflected in the measurement of individual peripheral marker level at a single time point. The levels of a single biomarker of SUA may not be representative of the functioning of the whole redox homeostasis system, but nevertheless provide a clue that the system is associated with the psychopathology of PSD. Third, there were five indicators, ALB, Hcy, SUA, Hs-CRP, and hospitalization days, were collected and studied by our post-hoc analysis. The original study protocol was not designed to assess association between these five indicators and PSD risk at discharge. Given the *post hoc* nature of the analysis, the proffered mechanism would likely be speculative. Fourth, the SUA level may be influenced by metabolic profile and the information collection of glycemia, lipid profile, and metabolic syndrome were important issues (41, 42). There is no specific analysis on these issues in our study. Future longitudinal studies should further explore these associations, and SUA's potential as predictor for PSD at discharge.

In conclusion, our study suggested that lower SUA levels on admission were associated with PSD at discharge and the threshold effect was also found by RCS regression model. For stroke patients, doctors should pay attention to the baseline SUA levels for screening high risk PSD patients at discharge in clinical practice. SUA was the potential biomarker for PSD prediction. The limitation of the study includes the study biases and the results should be further confirmed in longitudinal and experimental studies.

DATA AVAILABILITY STATEMENT

The de-identified database used in the current study are available from the corresponding authors on reasonable request.

ETHICS STATEMENT

All patients involved in this study or their family members gave written informed consents according to the Declaration of Helsinki. The approval of the study for experiments was obtained from the Ethics Committee of Tongji Medical College, Huazhong University of Science and Technology.

AUTHOR CONTRIBUTIONS

SZ and ZZ led the study. GL performed the data analysis and implemented the methodology. XS, WS, JM, YL, XQ, XZ, and CZ collected the data. ZZ and GL prepared the original draft. SZ reviewed and edited the final manuscript.

FUNDING

This work was financially supported by the National Key R&D Program of China (grant number 2017YFC1310000), the Fundamental Research Funds for the Central Universities

(grant number 2018KFYXMPT015), and Hubei Technological Innovation Special Fund (grant number 2019ACA132). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript. ZZ and SZ had full access to all the data in the study and had final responsibility for the decision to submit for publication.

ACKNOWLEDGMENTS

We would like to acknowledge all participants of this project and investigators for collecting data.

REFERENCES

- Robinson RG, Jorge RE. Post-stroke depression: a review. *Am J Psychiatry* (2016) 173(3):221–31. doi: 10.1176/appi.ajp.2015.15030363
- Flaster M, Sharma A, Rao M. Poststroke depression: a review emphasizing the role of prophylactic treatment and synergy with treatment for motor recovery. *Top Stroke Rehabil* (2013) 20(2):139–50. doi: 10.1310/tsr2002-139
- Ayerbe L, Ayis S, Crichton SL, Rudd AG, Wolfe CD. Explanatory factors for the increased mortality of stroke patients with depression. *Neurology* (2014) 83(22):2007–12. doi: 10.1212/WNL.0000000000001029
- Ayerbe L, Ayis S, Wolfe CD, Rudd AG. Natural history, predictors and outcomes of depression after stroke: systematic review and meta-analysis. *Br J Psychiatry* (2013) 202(1):14–21. doi: 10.1192/bjp.bp.111.107664
- Cheng SY, Zhao YD, Li J, Chen XY, Wang RD, Zeng JW. Plasma levels of glutamate during stroke is associated with development of post-stroke depression. *Psychoneuroendocrinology* (2014) 47:126–35. doi: 10.1016/j.psyneuen.2014.05.006
- Bartoli F, Di Brita C, Crocamo C, Clerici M, Carra G. Early post-stroke depression and mortality: meta-analysis and meta-regression. *Front Psychiatry* (2018) 9:530. doi: 10.3389/fpsy.2018.00530
- Huang G, Chen H, Wang Q, Hong X, Hu P, Xiao M, et al. High platelet-to-lymphocyte ratio are associated with post-stroke depression. *J Affect Disord* (2019) 246:105–11. doi: 10.1016/j.jad.2018.12.012
- Moylan S, Berk M, Dean OM, Samuni Y, Williams LJ, O'Neil A, et al. Oxidative & nitrosative stress in depression: why so much stress? *J Neurosci Biobehav Rev* (2014) 45:46–62. doi: 10.1016/j.neubiorev.2014.05.007
- Wen S, Cheng M, Wang H, Yue J, Wang H, Li G, et al. Serum uric acid levels and the clinical characteristics of depression. *Clin Biochem* (2012) 45(1–2):49–53. doi: 10.1016/j.clinbiochem.2011.10.010
- Wium-Andersen MK, Kobylecki CJ, Afzal S, Nordestgaard BG. Association between the antioxidant uric acid and depression and antidepressant medication use in 96 989 individuals. *Acta Psychiatr Scand* (2017) 136(4):424–33. doi: 10.1111/acps.12793
- Maes M, Christophe A, Delanghe J, Altamura C, Neels H, Meltzer HY. Lowered omega3 polyunsaturated fatty acids in serum phospholipids and cholesteryl esters of depressed patients. *Psychiatry Res* (1999) 85(3):275–91. doi: 10.1016/S0165-1781(99)00014-1
- Maes M, Galecki P, Chang YS, Berk M. A review on the oxidative and nitrosative stress (O&NS) pathways in major depression and their possible contribution to the (neuro)degenerative processes in that illness. *Prog Neuropsychopharmacol Biol Psychiatry* (2011) 35(3):676–92. doi: 10.1016/j.pnpbp.2010.05.004
- Ames BN, Cathcart R, Schwiers E, Hochstein P. Uric acid provides an antioxidant defense in humans against oxidant- and radical-caused aging and cancer: a hypothesis. *Proc Natl Acad Sci USA* (1981) 78(11):6858–62. doi: 10.1073/pnas.78.11.6858
- Bartoli F, Trotta G, Crocamo C, Malerba MR, Clerici M, Carra G. Antioxidant uric acid in treated and untreated subjects with major depressive disorder: a meta-analysis and meta-regression. *Eur Arch Psychiatry Clin Neurosci* (2018) 268(2):119–27. doi: 10.1007/s00406-017-0817-7
- Gao J, Xu W, Han K, Zhu L, Gao L, Shang X. Changes of serum uric acid and total bilirubin in elderly patients with major postischemic stroke depression. *Neuropsychiatr Dis Treat* (2018) 14:83–93. doi: 10.2147/NDT.S149712
- Gu Y, Han B, Wang L, Chang Y, Zhu L, Ren W. Low Serum levels of uric acid are associated with development of poststroke depression. *Med (Baltimore)* (2015) 94(45):e1897. doi: 10.1097/MD.0000000000001897
- Zheng YP, Zhao JP, Phillips M, Liu JB, Cai MF, Sun SQ, et al. Validity and reliability of the chinese hamilton depression rating scale. *Br J Psychiatry* (1988) 152:660–4. doi: 10.1192/bjp.152.5.660
- Zhang Y, Cheng L, Chen Y, Yang GY, Liu J, Zeng L. Clinical predictor and circulating microRNA profile expression in patients with early onset post-stroke depression. *J Affect Disord* (2016) 193:51–8. doi: 10.1016/j.jad.2015.12.061
- de Man-van GJ, Hafsteinsdottir TB, Lindeman E, Ettema RG, Grobbee DE, Schuurmans MJ. In-hospital risk prediction for post-stroke depression: development and validation of the post-stroke depression prediction scale. *Stroke* (2013) 44(9):2441–5. doi: 10.1161/STROKEAHA.111.000304
- Sin NL, Kumar AD, Gehi AK, Whooley MA. Direction of association between depressive symptoms and lifestyle behaviors in patients with coronary heart disease: the heart and soul study. *Ann Behav Med* (2016) 50(4):523–32. doi: 10.1007/s12160-016-9777-9
- Linden T, Blomstrand C, Skoog I. Depressive disorders after 20 months in elderly stroke patients: a case-control study. *Stroke* (2007) 38(6):1860–3. doi: 10.1161/STROKEAHA.106.471805
- De Ryck A, Brouns R, Geurden M, Elseviers M, De Deyn PP, Engelborghs S. Risk factors for poststroke depression: identification of inconsistencies based on a systematic review. *J Geriatr Psychiatry Neurol* (2014) 27(3):147–58. doi: 10.1177/0891988714527514
- Fang M, Zhong L, Jin X, Cui R, Yang W, Gao S, et al. Effect of inflammation on the process of stroke rehabilitation and poststroke depression. *Front Psychiatry* (2019) 10:184. doi: 10.3389/fpsy.2019.00184
- Palta P, Samuel LJ, Miller ER, Szanton SL. Depression and oxidative stress: results from a meta-analysis of observational studies. *Psychosom Med* (2014) 76(1):12–9. doi: 10.1097/PSY.0000000000000009
- Black CN, Bot M, Scheffer PG, Cuijpers P, Penninx BW. Is depression associated with increased oxidative stress? A systematic review and meta-analysis. *Psychoneuroendocrinology* (2015) 51:164–75. doi: 10.1016/j.psyneuen.2014.09.025
- Black CN, Bot M, Scheffer PG, Snieder H, Penninx BW. Uric acid in major depressive and anxiety disorders. *J Affect Disord* (2018) 225:684–90. doi: 10.1016/j.jad.2017.09.003
- Moylan S, Maes M, Wray NR, Berk M. The neuroprogressive nature of major depressive disorder: pathways to disease evolution and resistance, and therapeutic implications. *Mol Psychiatry* (2013) 18(5):595–606. doi: 10.1038/mp.2012.33
- Nanetti L, Raffaelli F, Vignini A, Perozzi C, Silvestrini M, Bartolini M. Oxidative stress in ischaemic stroke. *Eur J Clin Invest* (2011) 41(12):1318–22. doi: 10.1111/j.1365-2362.2011.02546.x
- Moccia M, Lanzillo R, Costabile T, Russo C, Carotenuto A, Sasso G, et al. Uric acid in relapsing-remitting multiple sclerosis: a 2-year longitudinal study. *J Neurol* (2015) 262(4):961–7. doi: 10.1007/s00415-015-7666-y
- Kim TS, Pae CU, Yoon SJ, Jang WY, Lee NJ, Kim JJ. Decreased plasma antioxidants in patients with Alzheimer's disease. *Int J Geriatr Psychiatry* (2006) 21(4):344–8. doi: 10.1002/gps.1469

31. de Lau LM, Koudstaal PJ, Hofman A, Breteler MM. Serum uric acid levels and the risk of Parkinson disease. *Ann Neurol* (2005) 58(5):797–800. doi: 10.1002/ana.20663
32. Chamorro A, Obach V, Cervera A, Revilla M, Deulofeu R, Aponte JH. Prognostic significance of uric acid serum concentration in patients with acute ischemic stroke. *Stroke* (2002) 33(4):1048–52. doi: 10.1161/hs0402.105927
33. Amaro S, Llull L, Renu A, Laredo C, Perez B, Vila E, et al. Uric acid improves glucose-driven oxidative stress in human ischemic stroke. *Ann Neurol* (2015) 77(5):775–83. doi: 10.1002/ana.24378
34. Schwarzschild MA, Ascherio A, Beal MF, Cudkovic ME, Curhan GC, Hare JM, et al. Inosine to increase serum and cerebrospinal fluid urate in Parkinson disease: a randomized clinical trial. *JAMA Neurol* (2014) 71(2):141–50.
35. Ortiz R, Ulrich H, Zarate CJ, Machado-Vieira R. Purinergic system dysfunction in mood disorders: a key target for developing improved therapeutics. *Prog Neuropsychopharmacol Biol Psychiatry* (2015) 57:117–31. doi: 10.1016/j.pnpbp.2014.10.016
36. Dos Santos Oliveira PM, Santos V, Coroa M, Ribeiro J, Madeira N. Serum uric acid as a predictor of bipolarity in individuals with a major depressive episode. *Bipolar Disord* (2019) 21(3):235–43. doi: 10.1111/bdi.12708
37. Wang Z, Lin Y, Liu Y, Chen Y, Wang B, Li C, et al. Serum uric acid levels and outcomes after acute ischemic stroke. *Mol Neurobiol* (2016) 53(3):1753–9. doi: 10.1007/s12035-015-9134-1
38. Wu H, Jia Q, Liu G, Liu L, Pu Y, Zhao X, et al. Decreased uric acid levels correlate with poor outcomes in acute ischemic stroke patients, but not in cerebral hemorrhage patients. *J Stroke Cerebrovasc Dis* (2014) 23(3):469–75. doi: 10.1016/j.jstrokecerebrovasdis.2013.04.007
39. Gardner A, Boles RG. Beyond the serotonin hypothesis: mitochondria, inflammation and neurodegeneration in major depression and affective spectrum disorders. *Prog Neuropsychopharmacol Biol Psychiatry* (2011) 35(3):730–43. doi: 10.1016/j.pnpbp.2010.07.030
40. Ali-Sisto T, Tolmunen T, Toffol E, Viinamäki H, Mantyselkä P, Valkonen-Korhonen M, et al. Purine metabolism is dysregulated in patients with major depressive disorder. *Psychoneuroendocrinology* (2016) 70:25–32. doi: 10.1016/j.psyneuen.2016.04.017
41. Ford ES, Li C, Cook S, Choi HK. Serum concentrations of uric acid and the metabolic syndrome among US children and adolescents. *Circulation* (2007) 115(19):2526–32. doi: 10.1161/CIRCULATIONAHA.106.657627
42. Facchini F, Chen YD, Hollenbeck CB, Reaven GM. Relationship between resistance to insulin-mediated glucose uptake, urinary uric acid clearance, and plasma uric acid concentration. *JAMA* (1991) 266(21):3008–11. doi: 10.1001/jama.266.21.3008

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Proteomic Profiling as a Diagnostic Biomarker for Discriminating Between Bipolar and Unipolar Depression

OPEN ACCESS

Edited by:

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Specialty section:

This article was submitted to
Molecular Psychiatry,
a section of the journal
Frontiers in Psychiatry

Received: 06 December 2019

Accepted: 26 February 2020

Published: 17 April 2020

Citation:

Kittel-Schneider S, Hahn T,
Haenisch F, McNeill R, Reif A and
Bahn S (2020) Proteomic Profiling
as a Diagnostic Biomarker for
Discriminating Between Bipolar
and Unipolar Depression.
Front. Psychiatry 11:189.
doi: 10.3389/fpsy.2020.00189

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Introduction: Affective disorders are a major global burden, with approximately 15% of people worldwide suffering from some form of affective disorder. In patients experiencing their first depressive episode, in most cases it cannot be distinguished whether this is due to bipolar disorder (BD) or major depressive disorder (MDD). Valid fluid biomarkers able to discriminate between the two disorders in a clinical setting are not yet available.

Material and Methods: Seventy depressed patients suffering from BD (bipolar I and II subtypes) and 42 patients with major MDD were recruited and blood samples were taken for proteomic analyses after 8 h fasting. Proteomic profiles were analyzed using the Multiplex Immunoassay platform from Myriad Rules Based Medicine (Myriad RBM; Austin, Texas, USA). Human DiscoveryMAPTM was used to measure the concentration of various proteins, peptides, and small molecules. A multivariate predictive model was consequently constructed to differentiate between BD and MDD.

Results: Based on the various proteomic profiles, the algorithm could discriminate depressed BD patients from MDD patients with an accuracy of 67%.

Discussion: The results of this preliminary study suggest that future discrimination between bipolar and unipolar depression in a single case could be possible, using predictive biomarker models based on blood proteomic profiling.

Keywords: affective disorder, bipolar disorder, major depression (MD), major depressive disorder (MDD), proteome, biomarker, blood, machine learning

INTRODUCTION

Depressive episodes affect up to 322 million people worldwide (Depression and Other Common Mental Disorders. Global Health Estimates. Geneva: World Health Organization 2017, <https://apps.who.int/iris/bitstream/handle/10665/254610/WHO-MSD-MER-2017.2-eng.pdf>). People suffering from a depressive episode can be suffering from either unipolar depression (major depressive disorder; MDD) or bipolar affective disorder (BD) as the underlying cause, depending on whether previous (hypo)-manic episodes have occurred. Unfortunately, this distinction can only be made after the first (hypo)-manic episode has presented. Therefore the most appropriate treatment for the underlying disorder may not initially be prescribed, especially as BD and MDD require fundamentally different pharmacological approaches; BD requires mood stabilizing medication, whereas MDD is treated with antidepressant monotherapy as a first-line treatment (1–3). Patients suffering from BD are often misdiagnosed as MDD and therefore adequate treatment can be delayed for up to several years (4, 5). Inadequate and delayed treatment increases the direct and indirect economic cost of BD, augments individual suffering, and impairs the overall prognosis (6). However, despite distinct treatment approaches, BD and MDD appear to share common molecular pathomechanisms. The gradient of MDD polygenic risk score has been shown to slide across the mood disorder spectrum, demonstrating an inverse relationship to the mania polygenic risk score (7).

The development of fluid biomarkers that can discriminate between BD and MDD would be highly beneficial, but reliable biomarkers have so far remained elusive. Nonspecific findings have been obtained in many studies, which failed to detect disorder-specific alterations, instead identifying molecular mechanisms implicated in several different psychiatric conditions. For example, several studies have reported dysregulation of the nitrinergic system in BD, but also in ADHD and schizophrenia (8–11). Recent work has additionally suggested that nitric oxide may play a role in the pathophysiology of major depression (12). Another potential cross-disorder mechanism is a dysfunctional hypothalamic-pituitary-adrenal axis (HPA axis), which has been implicated in both BD and MDD (13–15). Moreover, inflammatory processes (including the glucocorticoid system) may also play a role in MDD and BD (16, 17).

Despite several shared neurobiological features of psychiatric disorders, combining different modalities or vast arrays of biomarkers (e.g. using proteomic profiling) has demonstrated potential for providing disorder-specific biomarkers. A previous own study defined a diagnostic panel consisting of 20 protein analytes suitable for the diagnosis of BD (18). Additionally, Chen et al. published a set of 20 differential urine metabolites that could discriminate between BD and MDD (19). Although these initial findings are encouraging, the use of univariate statistical inference does not provide sufficient information to determine discriminative power for individual patients, nor does it quantify generalisation to new data. These initial promising results therefore need to be replicated in additional samples, and more

importantly tested for personalized predictive power, before a diagnostic biomarker panel can be used in clinical routine. In addition, modern machine learning approaches may considerably increase model performance by considering multivariate patterns in the data, thereby improving upon classic univariate approaches. In this study, we investigated whether a multivariate machine learning approach using data from multiplexed proteomic assays could accurately be used to discriminate between BD and MDD.

MATERIALS AND METHODS

Study Participants

Bipolar and major depression patients were part of a naturalistic sample recruited from patients treated in our in- and outpatient clinics. The male and female participants were within the age range 18–78 years, had a body mass index (BMI) between 18 and 46 kg/m, and had a test negative for recreational drug screening at the time of sampling. Patients were diagnosed with BD or MDD by two trained psychiatrists (SKS, AR) according to criteria of the International Classification of Diseases–10 (ICD-10), while being treated as inpatients or outpatients at the Department of Psychiatry, Psychosomatic Medicine and Psychotherapy of the University Hospital of Würzburg. Diagnoses were confirmed by the Operational Criteria Checklist for Affective and Psychotic Illness (OPCRIT) (20). Severity of symptoms was assessed using the standard questionnaire-based rating scales Young Mania Rating Scale (YMRS) and Montgomery–Åsberg Depression Rating Scale (MADRS) (21, 22).

Both bipolar I and bipolar II disorder patients were recruited and were in depressed mood states at the time of sample collection. MDD patients also had an acute depressive episode at the time of sample collection. Exclusion criteria included a diagnosis of severe coronary heart disease or cardiac insufficiency (i.e. coronary stent, cardiac bypass surgery angina pectoris, and cardiac insufficiency NYHA>I), severe autoimmune disorders (Hashimoto's thyroiditis excluded), acute or chronic infections, treatment with immunosuppressive/immune-modulating drugs or antibiotics, other severe neuropsychiatric disorders, chronic terminal diseases affecting the brain (such as cancer or hepatic/renal insufficiency), and alcohol or drug addiction (self-reported or taken from hospital discharge letters/general practitioner's letters). Patients were fasting for at least 8 h prior to blood sample collection. For more demographic details as well as somatic disorders and medication taken at sampling point see **Table 1** and **Supplemental Table 2**.

Only study participants who gave written informed consent were enrolled in the study, which complied with the latest Declaration of Helsinki, and was approved by the Ethics Committee of the University of Würzburg.

Sample Collection

Patients were recruited over a total time period of 4 years (2009–2013), and therefore proteomic profiles were analyzed in four

TABLE 1 | Demographic data.

| Depressive episode | n=70 | n=42 |
|---|------------------|------------------|
| | Bipolar Disorder | Major Depression |
| BD I/BD II | 30/40 | N/A |
| Age (years, mean +/- SD) | 43.47 +/- 11.69 | 44.28 +/- 14.93 |
| Gender (female/male) | 44/28 | 25/16 |
| BMI | 27.50 +/- 5.72 | 27.78 +/- 6.27 |
| Disease duration (years, mean +/-SD) | 15.10 +/-11.32 | N/A |
| MADRS sum score (mean +/-SD) | 18.0 +/- 1.96 | 18.47 +/- 8.32 |
| YMRS sum score (mean +/-SD) | 2.0 +/- 0.22 | N/A |
| Medication | | |
| Lithium | 7 | 3 |
| Valproic acid | 5 | 0 |
| Other anticonvulsants | 1 | 0 |
| Antipsychotics | 17 | 9 |
| Lithium + Valproic acid | 4 | 0 |
| Valproic acid + antipsychotics | 7 | 0 |
| Other anticonvulsants + antipsychotics | 4 | 0 |
| Lithium + anticonvulsants + antipsychotics | 4 | 0 |
| Lithium + antipsychotics | 21 | 1 |
| Antidepressants only | 2 | 28 |

BD I, bipolar disorder type I; BD II, bipolar disorder type II; BMI, body mass index; MADRS, Montgomery-Åsberg Depression Scale; YMRS, Young Mania Rating Scale; N/A, not available.

batches. Proteomic analyses were completed in 2010, 2011, and 2013. The maximum storage time for each sample in -80°C prior to analysis was 2 years.

Sample Preparation

Blood samples were taken on the day of clinical assessment (± 24 h). Blood was obtained from the participants by venous puncture in the morning after fasting for 10–13 h (between 7 to 9 am). Serum was collected from fasting patients using Vacutainer (Becton-Dickinson, Franklin Lakes, NJ, USA). Blood clotting time was 2 h at room temperature prior to centrifugation for 15 min at $1.100 \times g$. Samples were stored in low binding Eppendorf reaction tubes (Hamburg, Germany) at -80°C . Sample shipment took place on dry ice.

Multiplex Immunoassay Analysis

Serum from all participants was profiled using the multiplex immunoassay platform at Myriad Rules Based Medicine (Myriad RBM; Austin, Texas, USA), which has been previously described in detail (23). The Human DiscoveryMAPTM was used to measure the plasma concentrations of different proteins, peptides and small molecules (collectively referred to as “analytes”), in a Clinical Laboratory Improved Amendments certified lab. The total number of analytes measured differed between batches, depending on when the study samples were profiled (total range: 190 to 257 analytes). The analytes measured are reported in **Supplemental Table 1** and the concentration of all the analytes for all participants are reported in **Supplemental Table 3** and **Supplemental Table 4**. The raw data of the four different multiplex assays were normalised for batch effects to reduce variability.

Statistical Analysis

Statistical analyses were performed in R (24). We pre-processed the analyte data by excluding analytes with greater than 20% missing values, and imputing missing data. Data points under the lowest limit of detection (LLD) were replaced by the minimum value above the LLD for the specific analyte, and values above the highest detectable limit were replaced with the maximum measured value within the detectable range. In total, 1.1% of data points were imputed. We \log_{10} -transformed data to stabilize the variance. Additional analyses were performed using SPSS (v25, IBM®). The two analytes that appeared to play a significant role in discriminating between unipolar and bipolar depression were analyzed separately using ANCOVA.

Machine Learning Algorithm

In order to discriminate between patients suffering from MDD and BD, we first scaled all features (i.e. analytes) to have zero mean and unit variance. Next, tree ensemble classification was performed using the scikit-learn implementation of the AdaBoost algorithm with default hyperparameters (25, 26). To facilitate training in this extensively imbalanced dataset, we additionally employed Random Oversampling to the training set.

To assess the generalizability of the classifier, we used 10-fold cross-validation. Tenfold cross-validation is the most common standard in the field which ensures low model bias (due to the fairly large training sample) and low variance (due to the reasonably sized test set). Finding a balance between training and test sample size in each iteration is important, particularly because of the fairly small sample size used (for an introduction to the issue of k-fold cross-validation in practice see Bengio et al. (27). In each fold, data from 90% of the sample is used to train the classifier. Categorization of the remaining 10%, which has so far not been seen by the algorithm, is subsequently calculated. This procedure is repeated 10 times, each time leaving out different, nonoverlapping 10% portions of the sample, yielding each subject's categorization. To ensure unbiased test performance estimates in this imbalanced sample, accuracy was computed by calculating the mean of sensitivity and specificity, yielding “balanced accuracy.”

To establish whether the observed test accuracy estimate is statistically significant, we ran the entire pipeline 1,000 times with randomly permuted labels and counted the number of permutations which achieved higher accuracy than the one observed with the true labels. The p-value was then calculated by dividing this number by 1,000. If none of the permutation accuracies exceeded accuracy obtained with the true labels, this is denoted as $p < .001$.

To quantify the contribution of each feature, we computed permutation importance scores, calculated as the mean decrease of test accuracy for all samples if a given feature is randomly shuffled 10 times. Generally, permutation importance as used here provides a measure of how much a feature contributed to classification performance while leaving all other features intact. All analyses were performed using the PHOTON framework (www.photon-ai.com).

RESULTS

Patients with MDD and BD did not differ in basic demographic variables (see **Table 1**). However, medication significantly differed, as the majority of BD patients were taking mood stabilizers and the majority of MD patients were taking antidepressants (**Table 1**). Using data from 105 analytes, which was the lowest common denominator of the four batches, a multivariate predictive model was constructed to discriminate between MDD and BD (combined BD I and BD II disorders). The algorithm could discriminate between these two groups on the basis of the proteomic profile with an accuracy of 67% ($p < .001$ with 1,000 permutations) (see **Figure 1**). The analytes which the algorithm used for discrimination and prediction are displayed in **Supplemental Tables 1, 3, and 4**. The two analytes Platelet-Derived Growth Factor BB (PDGF-BB) and Thrombospondin-1 (TSP-1) were identified as particularly important for discriminating between BD and MDD in our sample. A subanalysis examining only young BD patients (<35 years) did not increase the accuracy of discrimination (data not shown). Additionally, it was observed that including a set of covariates (such as symptom severity and BD subtype) did not lead to improved accuracy (data not shown). To assess the potential influence of medication, we performed a covariate analysis using PDGF-BB and TSP-1 and included diagnosis (bipolar depression vs. MDD), medication, age, gender and BMI as covariates. No significant differences were found ($p = 0.19$, $p = 0.47$ respectively; see **Table 2**).

DISCUSSION

Recent studies found that ~10% of patients suffering from a depressive episode subsequently develop BD (4). To date, there are no tests in clinical routine to determine the risk of developing BD in patients experiencing their first depressive episode. However, it would be of great clinical significance to be able to accurately predict the underlying disorder, as pharmacological treatment differs considerably between MDD and BD. In this study, we have demonstrated that a machine-learning algorithm was able to individually discriminate BD (acutely depressed) from MDD (acutely depressed) patients with a moderately good accuracy of 67%, based on their proteomic profile. Based on our data, PDGF-BB and TSP-1 appeared to play a prominent role in this discrimination. Along with other serum analytes, PDGF-BB has previously been reported as associated with lower fractional anisotropy, higher mean diffusivity, and higher radial diffusivity

in several brain regions in a sample of depressed BD patients (28). Furthermore, PDGF-BB was found to be increased in BD patients suffering a depressive episode after treatment with a combination of sleep deprivation, lithium and bright light therapy (29). PDGF-BB was also found to have low intra-individual variability when measured with different methods and in serum and plasma, suggesting that this marker may be technically reliable (30). Physiologically, PDGF receptors have been reported to play a role in glutamatergic signaling (31), which is thought to be dysregulated in subtypes of affective patients (32).

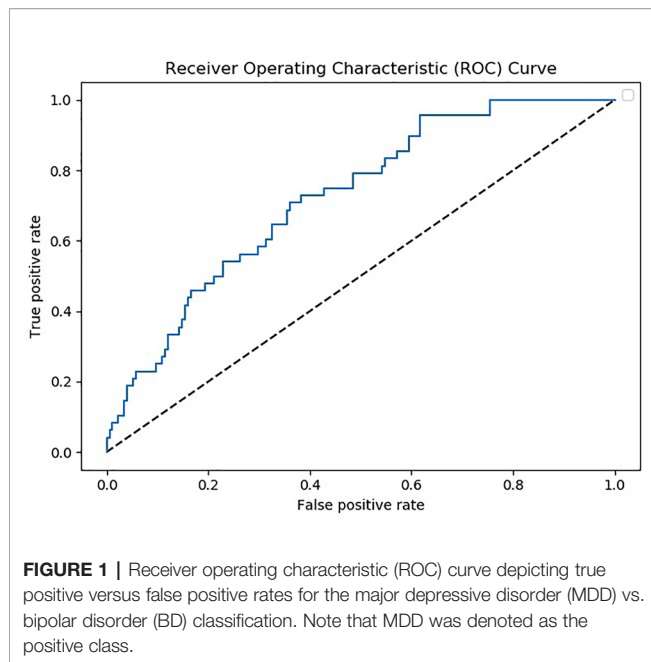
TSP-1 may be involved in synaptogenesis (33). Electroconvulsive therapy (ECT) is used to treat therapy-resistant MDD and BD depression, and was found to increase TSP-1 mRNA and protein expression in a rat model. However, chronic antidepressant treatment in this animal model appeared to have no effect on TSP-1 (34). Preclinical data has additionally suggested that TSP-1 may play a role in bidirectional neuron-astrocyte communication, dysregulation of which could be a pathomechanism for the development of mental illnesses (35). There are also several in vitro studies demonstrating that the mood stabilizer valproate can induce TSP-1 protein expression and thrombospondin-1 (*THBS-1*) gene expression in different cell and animal models (36–38). This is in contrast to our results, which showed no difference in TSP-1 expression between patients treated with valproate and patients treated with the other mood stabilizers and antidepressants. However, valproate may only exert its main effect on TSP-1 expression in the central nervous system, with its effects not detectable in the periphery. With regards to human in vivo data, a recent study reported decreased TSP-1 serum levels in female patients with MDD compared to healthy controls and male MDD patients. However, ECT treatment did not influence TSP-1 levels, leading the authors to conclude that serum TSP-1 may be a state marker of female MDD rather than a trait marker (39). Nonetheless, a technical issue in both our study and the one performed by Okada-Tsuchioka et al. is that TSP-1 concentration was measured in serum and not plasma. As thrombocytes release TSP-1 in high concentrations, the TSP-1 generated by other cells may be masked (40). Future studies should therefore measure plasma TSP-1.

Several studies have previously attempted to discriminate between MDD and BD patients using fluid biomarkers. A recent study from Chen and colleagues analyzed urinary metabolic phenotypes and demonstrated that a panel of six urinary metabolites could potentially be used to discriminate between the two disorders (19). In a study comparing cytokine

TABLE 2 | TSP-1 and PDGF-BB levels.

| | Disorder | N | Mean (μg/L) | Std. Deviation +/- | ANCOVA, p |
|---------|------------------|----|-------------|--------------------|-----------|
| PDGF-BB | Bipolar Disorder | 70 | 15,811.67 | 5,828.79 | 0.19 |
| | Major depression | 42 | 16,641.46 | 6,515.39 | |
| TSP-1 | Bipolar Disorder | 70 | 16,498.75 | 8,180.89 | 0.47 |
| | Major depression | 42 | 17,424.39 | 4,829.24 | |

PDGF-BB, platelet-derived growth factor BB; TSP-1, thrombospondin-1; ANCOVA, analysis were calculated including diagnosis, age, gender, and medication as variables.



concentrations between remitted BD and MDD patients and healthy controls, higher concentrations of soluble Interleukin-6 receptor (sIL-6R), C-reactive protein (CRP), soluble Tumor-Necrosis-Factor-receptor-1 (sTNF-R) and Monocyte-chemoattractant-protein -1 (MCP-1) were shown in BD compared to MDD (41). Frye et al. used a similar approach to our current and previous studies but with a smaller sample size of MDD patients, BD patients and healthy controls, and observed differences in several serum proteins between groups. To date, the best diagnostic accuracy (>0.8) for discriminating between BD I patients and healthy controls was shown by growth-differentiation factor 15 (GDF-15), retinol-binding protein (RBP-4) and transthyretin (TTR). However, in the same study no marker could be identified that accurately discriminated between BD and MDD, which would be the most clinically relevant diagnostic biomarker (42). GDF-15 and RBP-4 were not included as analytes in our studies and therefore no data comparisons can be made. However, Frye and colleagues also found six proteins to be significantly increased in depressed BD and MDD patients, and these were found to differ from those of our own previous multi-center study with the exception of MMP-7 (18). PDGF-BB and TSP-1 were also not found to be significant markers in our previous study, although our previous samples were derived from BD patients in all episodes (including euthymic), currently depressed and euthymic MDD patients, which could explain the differences in results. Whereas in this smaller sample we only compared acutely depressed bipolar patients vs. current major depression. This inconsistency could be therefore due to the differences in the samples which were investigated with respect to current episode and subtypes as well as to different proteins included in the analysis. Our current study additionally improves upon previous approaches, as it is the first to move beyond group-statistical inference to provide single-subject predictions. As individualized prediction is a key

requirement for clinical application, our results support the clinical utility of multivariate predictive analytics approaches in the field.

A current limitation of using fluid peripheral biomarkers is that concentrations measured in the periphery do not necessarily reflect pathophysiological processes in the central nervous system. However, our primary aim is to develop a biomarker that can discriminate between disorders, and not identify underlying disease pathomechanisms. We therefore believe that the most important factor is whether biomarker expression varies sufficiently between individuals to allow for discrimination between different disorders, and not whether the biomarker is directly involved in disease aetiology. However, for most single metabolites, the differences between groups are statistically significant but not great enough for single prediction [for examples, see (9, 39, 43, 44)]. We therefore suggest that currently the most promising approach for individual prediction is to measure several analytes simultaneously in the form of a biomarker panel, with additional machine learning, rather than measuring only a few select proteins.

Machine-learning algorithms in the development of diagnostic biomarkers have so far mainly been used in neuroimaging studies. There are several preliminary studies demonstrating the potential of this approach, for applications such as identifying individuals at high-risk of BD (45) and for defining subphenotypes of BD (46). However, machine learning may impair the algorithm's ability to derive a high performing model, and preclude the use of more sophisticated approaches, potentially rendering our results an artificially low estimate of the true accuracy. Despite these limitations, the sample size used for evaluation in machine learning entails fairly small test sets, potentially increasing variance of performance estimates (although we employed cross-validation).

To conclude, the initial results obtained from our study are promising. However, larger samples of patients are needed to replicate the results, thereby supporting the development of diagnostic biomarkers which can be used in clinical routine. We are aware that in this hypothesis generating study we could only examine a discovery sample. The necessary next step is to validate our findings in a second, independent dataset which is however not readily available. We are currently reaching out to conduct according replication studies which will finally be the touchstone whether or not the pilot data presented here holds true or not.

LIMITATION

The results of our study have several limitations. First, in the subanalysis, sample sizes were small. Second, as all patients were medicated an influence of mood stabilizing medication and antidepressants on serum proteins cannot be excluded. Further studies using increased sample sizes and including drug-naïve BD and MDD patients should be performed to overcome this methodological weakness. However, studies on drug-naïve patients are difficult to conduct due to ethical issues.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/**Supplementary Material**, further inquiries can be directed to the corresponding author(s).

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by the Ethics Committee of the University of Würzburg. The patients/participants provided their written informed consent to participate in this study.

AUTHOR CONTRIBUTIONS

SK-S and AR recruited the patients and collected the sample. SK-S wrote the paper draft and did parts of the analysis. FH and SB conducted the multiplex proteomic analysis. FH did the preanalysis of the data and added to the manuscript draft. TH conducted the machine learning analysis and added to the manuscript draft. SB, AR, and RM took part in writing the final manuscript. RM revised the language of the final revised manuscript.

REFERENCES

- Yatham LN, Kennedy SH, Schaffer A, Parikh SV, Beaulieu S, O'Donovan C, et al. Canadian Network for Mood and Anxiety Treatments (CANMAT) and International Society for Bipolar Disorders (ISBD) collaborative update of CANMAT guidelines for the management of patients with bipolar disorder: update 2009. *Bipolar Disord* (2009) 11(3):225–55. doi: 10.1111/j.1399-5618.0269881109102919
- Goodwin GM. Evidence-based guidelines for treating bipolar disorder: revised second edition—recommendations from the British Association for Psychopharmacology. *J Psychopharmacol* (2009) 23(4):346–88. doi: 10.1177/0269881109102919
- Harter M, Klesse C, Berger M, Bermejo I, Bschor T, Gensichen J, et al. [Evidence-based treatment of depression: what does the new S3- and national healthcare guideline Unipolar Depression really recommend?]. *Z Psychosom Med Psychother* (2010) 56(4):334–42. doi: 10.1007/s00115-010-3084-7
- Li CT, Bai YM, Huang YL, Chen YS, Chen TJ, Cheng JY, et al. Association between antidepressant resistance in unipolar depression and subsequent bipolar disorder: cohort study. *Br J Psychiatry* (2012) 200(1):45–51. doi: 10.1192/bjp.bp.110.086983
- Hu C, Xiang YT, Ungvari GS, Dickerson FB, Kilbourne AM, Si TM, et al. Undiagnosed bipolar disorder in patients treated for major depression in China. *J Affect Disord* (2012) 140(2):181–6. doi: 10.1016/j.jad.2012.02.014
- Jin H, McCrone P. Cost-of-illness studies for bipolar disorder: systematic review of international studies. *Pharmacoeconomics* (2015) 33(4):341–53. doi: 10.1007/s40273-014-0250-y
- Coleman JRI, Gaspar HA, Bryois J. Bipolar Disorder Working Group of the Psychiatric Genomics C, Major Depressive Disorder Working Group of the Psychiatric Genomics C, Breen G. The Genetics of the Mood Disorder Spectrum: Genome-wide Association Analyses of More Than 185,000 Cases and 439,000 Controls. *Biol Psychiatry* (2019). S0006–3223(19)31813–X. doi: 10.1016/j.biopsych.2019.10.015
- Selek S, Savas HA, Gergerlioglu HS, Bulut M, Yilmaz HR. Oxidative imbalance in adult attention deficit/hyperactivity disorder. *Biol Psychol* (2008) 79(2):256–9. doi: 10.1016/j.biopsycho.2008.06.005
- Kittel-Schneider S, Reuss M, Meyer A, Weber H, Gessner A, Leistner C, et al. Multi-level biomarker analysis of nitric oxide synthase isoforms in bipolar disorder and adult ADHD. *J Psychopharmacol* (2015) 29(1):31–8. doi: 10.1177/0269881114555251

FUNDING

This study has been supported by the BMBF (BipoLife, TPPI subproject to AR) and TH was supported by the German Research Foundation (DFG grants HA7070/2-2, HA7070/3, and HA7070/4) as well as LOEWE grant no 21000831. This publication was funded by the Goethe-University of Frankfurt.

ACKNOWLEDGMENTS

We like to thank Theresia Töpner and Nicole Döring for excellent technical support in the preprocessing of the samples and the presentation of the data in the supplemental tables. We thank the patients for participating in the study.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fpsy.2020.00189/full#supplementary-material>

- Reif A, Herterich S, Strobel A, Ehli AC, Saur D, Jacob CP, et al. A neuronal nitric oxide synthase (NOS-I) haplotype associated with schizophrenia modifies prefrontal cortex function. *Mol Psychiatry* (2006) 11(3):286–300. doi: 10.1038/sj.mp.4001779
- Yao JK, Leonard S, Reddy RD. Increased nitric oxide radicals in postmortem brain from patients with schizophrenia. *Schizophr Bull* (2004) 30(4):923–34. doi: 10.1093/oxfordjournals.schbul.a007142
- Baranyi A, Amouzadeh-Ghadikolai O, Rothenhauser HB, Theokas S, Robier C, Baranyi M, et al. Nitric Oxide-Related Biological Pathways in Patients with Major Depression. *PLoS One* (2015) 10(11):e0143397. doi: 10.1371/journal.pone.0143397
- Lackschewitz H, Huther G, Kroner-Herwig B. Physiological and psychological stress responses in adults with attention-deficit/hyperactivity disorder (ADHD). *Psychoneuroendocrinology* (2008) 33(5):612–24. doi: 10.1016/j.psyneuen.2008.01.016
- Juruena MF, Cleare AJ, Papadopoulos AS, Poon L, Lightman S, Pariante CM. Different responses to dexamethasone and prednisolone in the same depressed patients. *Psychopharmacol (Berl)*. (2006) 189(2):225–35. doi: 10.1007/s00213-006-0555-4
- Watson S, Thompson JM, Ritchie JC, Nicol Ferrier I, Young AH. Neuropsychological impairment in bipolar disorder: the relationship with glucocorticoid receptor function. *Bipolar Disord* (2006) 8(1):85–90. doi: 10.1111/j.1399-5618.2006.00280.x
- Raison CL, Rutherford RE, Woolwine BJ, Shuo C, Schettler P, Drake DF, et al. A randomized controlled trial of the tumor necrosis factor antagonist infliximab for treatment-resistant depression: the role of baseline inflammatory biomarkers. *JAMA Psychiatry* (2013) 70(1):31–41. doi: 10.1001/2013.jamapsychiatry.4
- Goldstein BI, Kemp DE, Soczynska JK, McIntyre RS. Inflammation and the phenomenology, pathophysiology, comorbidity, and treatment of bipolar disorder: a systematic review of the literature. *J Clin Psychiatry* (2009) 70(8):1078–90. doi: 10.4088/JCP.08r04505
- Haenisch F, Cooper JD, Reif A, Kittel-Schneider S, Steiner J, Leweke FM, et al. Towards a blood-based diagnostic panel for bipolar disorder. *Brain Behav Immun* (2015), 52:49–57. doi: 10.1016/j.bbi.2015.10.001
- Chen JJ, Zhou CJ, Liu Z, Fu YY, Zheng P, Yang DY, et al. Divergent Urinary Metabolic Phenotypes between Major Depressive Disorder and Bipolar Disorder Identified by a Combined GC-MS and NMR Spectroscopic

- Metabonomic Approach. *J Proteome Res* (2015) 14(8):3382–9. doi: 10.1021/acs.jproteome.5b00434
20. Rucker J, Newman S, Gray J, Gunasinghe C, Broadbent M, Brittain P, et al. OPCRIT+: an electronic system for psychiatric diagnosis and data collection in clinical and research settings. *Br J Psychiatry* (2011) 199(2):151–5. doi: 10.1192/bjp.bp.110.082925
 21. Schmidtke A, Fleckenstein P, Moises W, Beckmann H. [Studies of the reliability and validity of the German version of the Montgomery-Asberg Depression Rating Scale (MADRS)]. *Schweiz Arch Neurol Psychiatr* (1988) 139(2):51–65.
 22. Muhlbacher M, Egger C, Kaplan P, Simhandl C, Grunze H, Geretsegger C, et al. [Reliability and concordance validity of a German version of the Young Mania Rating Scale (YMRS-D)]. *Neuropsychiatr* (2011) 25(1):16–25.
 23. Bertenshaw GP, Yip P, Seshiah P, Zhao J, Chen TH, Wiggins WS, et al. Multianalyte profiling of serum antigens and autoimmune and infectious disease molecules to identify biomarkers dysregulated in epithelial ovarian cancer. *Cancer Epidemiol Biomarkers Prev* (2008) 17(10):2872–81. doi: 10.1158/1055-9965.EPI-08-0464
 24. R Core Team. R: a Language and Environment for Statistical Computing. R foundation for Statistical Computing, Vienna, Austria (2013). <http://www.R-project.org/>
 25. Freund Y, Schapire RE. A decision-theoretic generalization of on-line learning and an application to boosting. *J Comput Syst Sci* (1997) 55(1):119–39. doi: 10.1006/jcss.1997.1504
 26. Pedregosa F, Varoquaux G, Gramfort A, Michel V, Thirion B, Grisel O, et al. Scikit-learn: Machine Learning in Python. *J Mach Learn Res* (2011) 12:2825–30.
 27. Bengio Y, Grandvalet Y. No unbiased estimator of the variance of K-fold cross-validation. *J Mach Learn Res* (2004) 5:1089–105.
 28. Benedetti F, Poletti S, Hoogenboezem TA, Mazza E, Ambree O, de Wit H, et al. Inflammatory cytokines influence measures of white matter integrity in Bipolar Disorder. *J Affect Disord* (2016) 202:1–9. doi: 10.1016/j.jad.2016.05.047
 29. Benedetti F, Poletti S, Hoogenboezem TA, Locatelli C, Ambree O, de Wit H, et al. Stem Cell Factor (SCF) is a putative biomarker of antidepressant response. *J Neuroimmune Pharmacol* (2016) 11(2):248–58. doi: 10.1007/s11481-016-9672-y
 30. Belzeaux R, Lefebvre MN, Lazzari A, Le Carpentier T, Consoloni JL, Zendjidian X, et al. How to: Measuring blood cytokines in biological psychiatry using commercially available multiplex immunoassays. *Psychoneuroendocrinology* (2017) 75:72–82. doi: 10.1016/j.psychneuen.2016.10.010
 31. Lei S, Lu WY, Xiong ZG, Orser BA, Valenzuela CF, MacDonald JF. Platelet-derived growth factor receptor-induced feed-forward inhibition of excitatory transmission between hippocampal pyramidal neurons. *J Biol Chem* (1999) 274(43):30617–23. doi: 10.1074/jbc.274.43.30617
 32. Jun C, Choi Y, Lim SM, Bae S, Hong YS, Kim JE, et al. Disturbance of the glutamatergic system in mood disorders. *Exp Neurobiol* (2014) 23(1):28–35. doi: 10.5607/en.2014.23.1.28
 33. Christopherson KS, Ullian EM, Stokes CC, Mullen CE, Hell JW, Agah A, et al. Thrombospondins are astrocyte-secreted proteins that promote CNS synaptogenesis. *Cell* (2005) 120(3):421–33. doi: 10.1016/j.cell.2004.12.020
 34. Okada-Tsuchioka M, Segawa M, Kajitani N, Hisaoka-Nakashima K, Shibasaki C, Morinobu S, et al. Electroconvulsive seizure induces thrombospondin-1 in the adult rat hippocampus. *Prog Neuropsychopharmacol Biol Psychiatry* (2014) 48:236–44. doi: 10.1016/j.pnpbp.2013.10.001
 35. Nagai J, Rajbhandari AK, Gangwani MR, Hachisuka A, Coppola G, Masmanidis SC, et al. Hyperactivity with Disrupted Attention by Activation of an Astrocyte Synaptogenic Cue. *Cell* (2019) 177(5):1280–92 e20. doi: 10.1016/j.cell.2019.03.019
 36. Byler TK, Leocadio D, Shapiro O, Bratslavsky G, Stodgell CJ, Wood RW, et al. Valproic acid decreases urothelial cancer cell proliferation and induces thrombospondin-1 expression. *BMC Urol* (2012) 12:21. doi: 10.1186/1471-2490-12-21
 37. Causey MW, Salgar S, Singh N, Martin M, Stallings JD. Valproic acid reversed pathologic endothelial cell gene expression profile associated with ischemia-reperfusion injury in a swine hemorrhagic shock model. *J Vasc Surg* (2012) 55(4):1096–103 e51. doi: 10.1016/j.jvs.2011.08.060
 38. Chelluri R, Caza T, Woodford MR, Reeder JE, Bratslavsky G, Byler T. Valproic Acid Alters Angiogenic and Trophic Gene Expression in Human Prostate Cancer Models. *Anticancer Res* (2016) 36(10):5079–86. doi: 10.21873/anticancer.11077
 39. Okada-Tsuchioka M, Omori W, Kajitani N, Shibasaki C, Itagaki K, Takebayashi M. Decreased serum levels of thrombospondin-1 in female depressed patients. *Neuropsychopharmacol Rep* (2019). 40(1):39–45. doi: 10.1002/npr2.12088
 40. Barclay JL, Keshvari S, Whitehead JP, Inder WJ. Development of an enzyme-linked immunosorbent assay for thrombospondin-1 and comparison of human plasma and serum concentrations. *Ann Clin Biochem* (2016) 53(Pt 5):606–10. doi: 10.1177/0004563216628891
 41. Bai YM, Su TP, Li CT, Tsai SJ, Chen MH, Tu PC, et al. Comparison of pro-inflammatory cytokines among patients with bipolar disorder and unipolar depression and normal controls. *Bipolar Disord* (2015) 17(3):269–77. doi: 10.1111/bdi.12259
 42. Frye MA, Nassan M, Jenkins GD, Kung S, Veldic M, Palmer BA, et al. Feasibility of investigating differential proteomic expression in depression: implications for biomarker development in mood disorders. *Transl Psychiatry* (2015) 5:e689. doi: 10.1038/tp.2015.185
 43. Schroter K, Brum M, Brunkhorst-Kanaan N, Tole F, Ziegler C, Domschke K, et al. Longitudinal multi-level biomarker analysis of BDNF in major depression and bipolar disorder. *Eur Arch Psychiatry Clin Neurosci* (2019). 270(2):169–81. doi: 10.1007/s00406-019-01007-y
 44. Kittel-Schneider S, Weigl J, Volkert J, Gessner A, Schmidt B, Hempel S, et al. Further evidence for plasma progranulin as a biomarker in bipolar disorder. *J Affect Disord* (2014) 157:87–91. doi: 10.1016/j.jad.2014.01.006
 45. Hajek T, Cooke C, Kopecek M, Novak T, Hoschl C, Alda M. Using structural MRI to identify individuals at genetic risk for bipolar disorders: a 2-cohort, machine learning study. *J Psychiatry Neurosci* (2015) 40(5):316–24. doi: 10.1503/jpn.140142
 46. Wu MJ, Mwambi B, Bauer IE, Passos IC, Sanches M, Zunta-Soares GB, et al. Identification and individualized prediction of clinical phenotypes in bipolar disorders using neurocognitive data, neuroimaging scans and machine learning. *Neuroimage* (2016). 145(Pt B):254–64. doi: 10.1016/j.neuroimage.2016.02.016

Conflict of Interest: SK-S has received speaker's and author's honoraria from Medice and Takeda. AR has received speaker fees and honoraria (publications, advisory boards) from Medice, Shire/Takeda, Servier, neuraxpharm, Janssen and SAGE.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Endocannabinoid System Components as Potential Biomarkers in Psychiatry

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OPEN ACCESS

Edited by:

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Specialty section:

This article was submitted to
Molecular Psychiatry,
a section of the journal
Frontiers in Psychiatry

Received: 15 January 2020

Accepted: 30 March 2020

Published: 27 April 2020

Citation:

Navarrete F, García-Gutiérrez MS, Jurado-Barba R, Rubio G, Gasparyan A, Austrich-Olivares A and Manzanares J (2020) Endocannabinoid System Components as Potential Biomarkers in Psychiatry. *Front. Psychiatry* 11:315. doi: 10.3389/fpsy.2020.00315

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The high heterogeneity of psychiatric disorders leads to a lack of diagnostic precision. Therefore, the search of biomarkers is a fundamental aspect in psychiatry to reach a more personalized medicine. The endocannabinoid system (ECS) has gained increasing interest due to its involvement in many different functional processes in the brain, including the regulation of emotions, motivation, and cognition. This article reviews the role of the main components of the ECS as biomarkers in certain psychiatric disorders. Studies carried out in rodents evaluating the effects of pharmacological and genetic manipulation of cannabinoid receptors or endocannabinoids (eCBs) degrading enzymes were included. Likewise, the ECS-related alterations occurring at the molecular level in animal models reproducing some behavioral and/or neuropathological aspects of psychiatric disorders were reviewed. Furthermore, clinical studies evaluating gene or protein alterations in *post-mortem* brain tissue or *in vivo* blood, plasma, and cerebrospinal fluid (CSF) samples were analyzed. Also, the results from neuroimaging studies using positron emission tomography (PET) or functional magnetic resonance (fMRI) were included. This review shows the close involvement of cannabinoid receptor 1 (CB1r) in stress regulation and the development of mood disorders [anxiety, depression, bipolar disorder (BD)], in post-traumatic stress disorder (PTSD), as well as in the etiopathogenesis of schizophrenia, attention deficit hyperactivity disorder (ADHD), or eating disorders (*i.e.* anorexia and bulimia nervosa). On the other hand, recent results reveal the potential therapeutic action of the endocannabinoid tone manipulation by inhibition of eCBs degrading enzymes, as well as by the modulation of cannabinoid receptor 2 (CB2r) activity on anxiolytic, antidepressive, or antipsychotic associated effects. Further clinical research studies are needed; however, current evidence suggests that the components of the ECS may

become promising biomarkers in psychiatry to improve, at least in part, the diagnosis and pharmacological treatment of psychiatric disorders.

Keywords: endocannabinoid system, cannabinoid receptor (CB1r, CB2r), endocannabinoid, biomarker, psychiatry, diagnosis, treatment

INTRODUCTION

Psychiatric disorders are one of the main causes of disability in the general population (1). According to a recent estimation, psychiatric disorders account for 32.4% of years lived with disability (YLDs) and 13% of disability adjusted life-years (DALYs), leading the global burden of disease (2). Despite this, we still have a great lack of knowledge about its neurobiological basis, and clinically applicable biomarkers have been elusive. During the last decades, an increasing effort has been made in the search of biomarkers in psychiatry to help in the diagnosis and prediction of disease progression or treatment response. However, a clinical biomarker should be validated, sensitive, specific, feasible, and easily reproducible, characteristics that make difficult the implementation in this field (3–5).

Abbreviations: ECS, endocannabinoid system; eCBs, endocannabinoids; CB1r, cannabinoid receptor 1; CB2r, cannabinoid receptor 2; CSF, cerebrospinal fluid; PET, positron emission tomography; fMRI, functional magnetic resonance; BD, bipolar disorder; PTSD, posttraumatic stress disorder; ADHD, attention deficit hyperactivity disorder; YLDs, years lived with disability; DALYs, disability adjusted life-years; CNS, central nervous system; PFC, prefrontal cortex; NAC, nucleus accumbens; Hipp, hippocampus; Amy, amygdala; PVN, paraventricular nucleus; Hyp, hypothalamus; VTA, ventral tegmental area; AEA, anandamide; 2-AG, 2-arachidonoylglycerol; NAPE-PLD, N-acylphosphatidylethanolamine specific phospholipase D; FAAH, fatty acid amide hydrolase; GPCRs, Gq protein-coupled receptors; DAGL, diacylglycerol; MAGL, monoacylglycerol lipase; ABHD, alpha/beta-hydrolase domain; DSM-5, diagnostic and statistical manual of mental disorders, 5th version; TRPV1, transient receptor potential cation channel subfamily V member 1; dIPAG, dorsolateral periaqueductal gray matter; NO, nitric oxide; WT, wild-type; DMH, dorsomedial hypothalamus; BLA, basolateral amygdala; EPM, elevated plus maze; LHb, lateral habenula; LC, locus coeruleus; DRN, dorsal raphe nucleus; NA, noradrenaline; 5-HT, serotonin; HPA axis, hypothalamus–pituitary–adrenal axis; CRH, corticotropin-releasing hormone; CRHR1, corticotropin-releasing hormone type 1 receptor; 5HTT, serotonin transporter; HDAC, histone deacetylase; CUS, chronic unpredictable stress; CB2xP, transgenic mice overexpression CB2r in the brain; BCP, beta-caryophyllene; THC, tetrahydrocannabinol; MDD, major depressive disorder; BDNF, brain-derived neurotrophic factor; TST, tail suspension test; FST, forced swimming test; CMS, chronic mild stress; FSL, Flinders sensitive line rats; WKY, Wistar Kyoto rats; cKO, conditional knockout; SSRIs, serotonin selective reuptake inhibitors; 5HTT, serotonin transporter; rTMS, repeated transcranial magnetic stimulation; ECT, electroconvulsive therapy; ACC, anterior cingulate cortex; PPI, prepulse inhibition; NMDAR, N-methyl-D-aspartate receptors; MAM, methylazoxymethanol; SHR, spontaneously hypertensive rat; CNR1P1, cannabinoid receptor interacting protein; Nrg1, neuregulin 1; DLPFC, dorsolateral prefrontal cortex; STG, superior temporal gyrus; qRT-PCR, quantitative real time polymerase chain reaction; FEP, first episode psychosis; PBMCs, peripheral blood mononuclear cells; PANSS, positive and negative syndrome scale; SUD, substance use disorder; CBD, cannabidiol; ACTH, adrenocorticotropin hormone; CS, corticosterone; SPS, single prolonged stress; DLS, dorsolateral striatum; PEA, palmitoylethanolamide; OEA, oleoylethanolamide; SEA, stearoylethanolamide; AN, anorexia nervosa; BN, bulimia nervosa; NPY, neuropeptide Y; POMC, proopiomelanocortin; CART, cocaine-amphetamine-regulated transcript; ARC, arcuate nucleus; FID, food intake disorders; BMI, body mass index.

The endocannabinoid system (ECS) components (receptors, ligands, synthesizing and degrading enzymes) have gained a special interest because of their critical neuromodulatory involvement in a plethora of functional mechanisms in the central nervous system (CNS), including emotional regulation, motivational behavior, and cognitive function (6, 7). The wide distribution of ECS in the brain, together with the effects derived from its pharmacological modulation on mood or cognition with exogenous cannabinoid compounds, mainly those contained or derived from the *Cannabis sativa* plant, suggests that the identification of the functional role of ECS elements in certain psychiatric disorders could be a breakthrough to improve diagnosis and treatment (8–11).

Therefore, this review summarizes the findings regarding the potential involvement of ECS components as biomarkers, mainly in terms of the discovery of new therapeutic approaches, but also from the point of view of its diagnostic, prognostic and predictive application. For that purpose, studies on animal models and patients have been collected focusing on the most prevalent psychiatric conditions, including anxiety disorders (3.8%) (12), depressive disorders (3.4%) (12), schizophrenia (0.3%) (12), bipolar disorder (0.6%) (12), post-traumatic stress disorder (7.8%) (13), attention-deficit hyperactivity disorder (2.2%) (14), and eating disorders (0.2%) (12).

A BRIEF OVERVIEW OF THE ENDOCANNABINOID SYSTEM COMPONENTS

ECS regulates a number of physiological functions and mediates the crosstalk between different neurotransmitter systems, therefore representing a key player in the control of behavioral responses (15, 16). ECS is a ubiquitous lipid signaling system distributed throughout the organism that participates in multiple intracellular signaling pathways (17, 18). Cannabinoid receptors, endogenous ligands or endocannabinoids (eCBs), and their synthesizing and degrading enzymes are the main components of the ECS (**Figure 1**) present in the central and peripheral nervous system (15, 19) and in many other peripheral tissues regulating distinct functions (20).

The CB1 receptor (CB1r) is the most abundant G protein-coupled receptor in the brain (21). Physiological actions of endocannabinoids in the CNS are mediated by the activation of CB1r (22). Their expression in the CNS is widespread and heterogeneous and has crucial roles regulating brain function and disease processes (23–25). CB1r is abundant in the basal ganglia, cerebellum, in corticolimbic regions including the prefrontal cortex (PFC), nucleus accumbens (Nac), and hippocampus (Hipp), and in

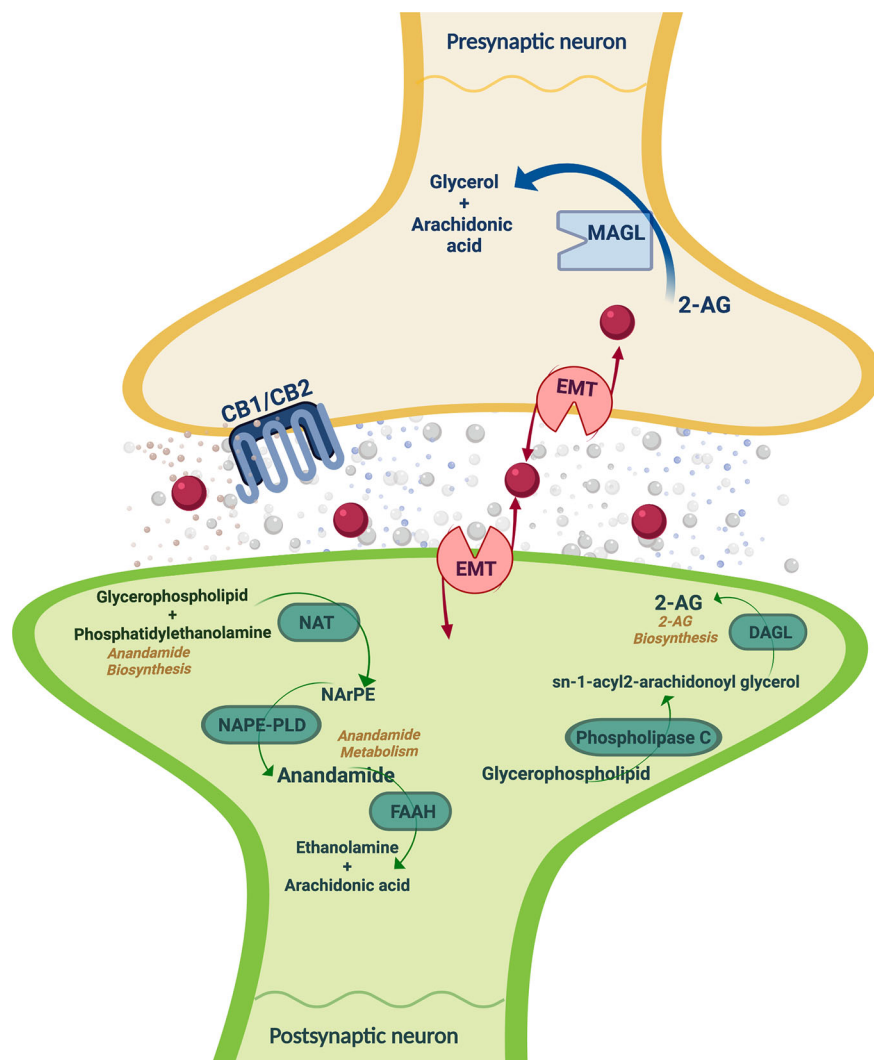


FIGURE 1 | Schematic representation of the main ECS components, including the metabolizing routes of the eCBs. CB1/CB2, cannabinoid receptors 1 and 2; 2-AG, 2-arachidonoylglycerol; FAAH, fatty acid amide hydrolase; MAGL, monoacylglycerol lipase; DAGL, EMT: endocannabinoid membrane transporter; NAT, N-acyl transferase; NArPE, N-arachidonoyl phosphatidylethanolamine; NAPE-PLD, N-acylphosphatidylethanolamine specific phospholipase D; DAGL, diacylglycerol lipase. Image created with BioRender.

brain areas related to stress responses, such as the central amygdala (Amy) and the paraventricular nucleus (PVN) of the hypothalamus (Hyp) (21, 26, 27). Furthermore, CB1r is also located in terminals of peripheral neurons and glial cells, as well as in the reproductive system (*i.e.* uterus, ovary, testis, prostate), some glandular systems (adrenal gland), adipose tissue, heart, liver, lung, bone marrow, thymus, and the microcirculation (20, 26, 28–33).

CB2 cannabinoid receptor (CB2r) was initially considered as a peripheral cannabinoid receptor due to its high expression in the rat spleen (34) and leukocyte subpopulation in humans (32), participating in the regulation of the immune system (35). The first findings identified the presence of CB2r in the CNS only under pathological conditions such as in senile plaques in Alzheimer's disease (36), activated microglial cells/

macrophages in multiple sclerosis, spinal cord in amyotrophic lateral sclerosis (37) and in the vicinity of tumors (38). However, Van Sickle and colleagues revealed that CB2r is expressed in neurons of the brainstem of mice, rats, and ferrets under normal conditions (39). This finding was key to increase the interest of CB2r in the regulation of brain function. Different studies identified CB2r in several brain regions including the frontal cortex, striatum, basal ganglia, Amy, Hipp, and the ventral tegmental area (VTA) (40–44). Interestingly, in some of these brain regions, CB2r was detected not only in the microglia (45) but also in the neurons (44, 46, 47).

The eCBs are lipid messengers acting as paracrine, autocrine, and probably endocrine mode, because their lipid nature allows them to diffuse and cross membranes (15, 17, 18, 48, 49). eCBs are agonists of

CB1r and CB2r that are not accumulated in secretory vesicles but rather synthesized under tonic or phasic (on demand) modes, and released to the extracellular space following physiological and pathological stimuli (50). The two main eCBs are derivatives of polyunsaturated fatty acids, N-arachidonylethanolamine (anandamide, AEA) (51), and 2-arachidonoylglycerol (2-AG), being the most abundant eCBs in the brain (52). Firstly, AEA synthesis is produced by the N-acylphosphatidylethanolamine specific phospholipase D (NAPE-PLD) that hydrolyzes N-arachidonoyl phosphatidylethanolamine localized in cell membranes (49, 53). The AEA half-life is very short because of its quick uptake by a high affinity AEA membrane transporter distributed in the neurons and glia (54). AEA is inactivated by fatty acid amide hydrolase (FAAH) present in many organs and in the brain at postsynaptic location (55, 56). FAAH is a serine-hydrolase enzyme bound to intracellular membranes that metabolizes AEA into arachidonic acid and ethanolamine (57). Secondly, 2-AG participates in the CB1r-dependent retrograde signaling and is an intermediate metabolite for lipid synthesis providing arachidonic acid for prostaglandin synthesis (57). Neuronal membrane depolarization or the activation of Gq protein-coupled receptors (GPCRs) triggers the synthesis of 2-AG (49). The diacylglycerol precursors come from the hydrolysis of membrane phosphatidylinositol by phospholipase C, β or δ . The degradation of these precursors by diacylglycerol lipases (DAGL- α and DAGL- β) drives 2-AG synthesis (58, 59). The DAGL α isoform synthesizes the greatest amount of 2-AG, whereas DAGL β synthesizes 2-AG under certain circumstances (54). Monoacylglycerol lipase (MAGL) is a serine-hydrolase enzyme mainly found in presynaptic terminals that catalyzes 2-AG into arachidonic acid and glycerol (55, 60). Also, the α/β -hydrolase domain 6 (ABHD6) and domain 12 (ABHD12) degrade 2-AG (49, 57).

THE ENDOCANNABINOID SYSTEM IN PSYCHIATRY: SEARCHING FOR POTENTIAL BIOMARKERS

The ECS is one of the most widely distributed neurotransmitter systems in the human brain, with a critical neuromodulatory role that motivates the interaction with other neurotransmitter and neurohormonal systems (61). Accumulating evidence points out the pivotal role of the ECS in the regulation of cognitive and behavioral functioning, suggesting its therapeutic potential in psychiatry (9, 11, 62). Furthermore, it is worth to mention that psychiatric disorders are accompanied by disturbances in the ECS components, as detailed below. Taken together, these facts suggest the potential usefulness of cannabinoid receptors, endocannabinoid ligands and degrading or synthesizing enzymes as biomarkers to move towards improved diagnostic criteria and therapeutic approaches in psychiatry.

The literature review consisted of an exhaustive search for scientific information in the Medline database (PubMed), which was always focused on the following ECS components as potential biomarkers in psychiatry: CB1r, CB2r, AEA, 2-AG, FAAH and MAGL. A total of seven search boxes were employed

according to the total of psychiatric conditions included in the review: anxiety, depression, schizophrenia, bipolar disorder, post-traumatic stress disorder, attention-deficit and hyperactivity disorder, and eating disorders. These terms were combined with the term 'cannabinoid' by the Boolean operator 'AND'. All the results for each search were critically analyzed by the authors to decide the inclusion or exclusion of each reference according to the adequacy of its content with the subject matter of the study. Finally, no PubMed filters were applied to maximize the selection of all the available and appropriate information.

Anxiety Disorders

According to the Diagnostic and Statistical Manual of Mental Disorders (DSM-5), anxiety disorders share features of excessive fear and anxiety and related behavioral disturbances. Fear is the emotional response to real or perceived imminent threat, whereas anxiety is an emotional anticipatory response to future potential threatening or stressful situations, triggering symptoms of negative affective, somatic, behavioral and cognitive components (63). The ECS plays a prominent role in the stress response and anxiety, as it is widely documented mainly by animal studies (64–67). However, our knowledge on the precise molecular mechanisms of the ECS signaling in humans is insufficient (68, 69). In the last years, compelling evidence for the involvement of ECS in anxiety has been accumulated that suggests new therapeutic leads through the discovery of potential biomarkers.

Animal Studies

A large body of literature supports the involvement of CB1r as a potential biomarker in anxiety disorders (70–72). CB1r is widely distributed in brain areas associated with emotional regulation and stress responsiveness such as PFC, Hipp, Amy, and Hyp (19). Previous pharmacological studies evaluated the effects of different cannabinoid compounds after either systemic or intracerebral administration in rodents exposed to several animal models of anxiety (73, 74). In addition, it is important to highlight the pivotal role of CB1r in the effects of anxiolytic drugs such as benzodiazepines. Indeed, our group demonstrated that the CB1r antagonist, AM251, completely abolished the anxiolytic effects and significantly reduced the amnesic and the sedative actions induced by alprazolam (75). A very similar result was recently obtained regarding the AM251-induced blockade of the anxiolytic effects of alprazolam (76). On the other hand, the enhancement of CB1r-mediated endocannabinoid function increases the anxiolytic action of diazepam (77).

Accumulated evidence points out that CB1r manipulation produces a bidirectional effect on anxiety-related behavior (78, 79). CB1r activation decreases anxiety at lower doses (80), whereas anxiogenic effects occur at higher doses or after CB1r blockade (70, 81–87). However, several factors could modify this general assumption such as regional endogenous tone, age, sex, species differences, type of test, previous exposure to stressful situations, or dosage of cannabinoid receptor agonists or antagonists. In addition, the underlying mechanisms involved in the bidirectional effects of CB1r pharmacological modulation remain poorly understood. Among the available evidences

addressing this aspect, one study revealed that CB1r in the cortical glutamatergic neurons mediates the anxiolytic effect of CP-55,940 cannabinoid agonist at low doses, whereas anxiogenic actions of higher doses are related with CB1r and GABA_B receptors in GABAergic terminals (88). A growing body of evidence also suggests that the anxiogenic effects of moderate to high cannabinoid doses appear to be mediated by the interaction between endocannabinoid and endovanilloid systems, specifically through the activation of transient receptor potential cation channel subfamily V member 1 (TRPV1) vanilloid receptors (89). In this regard, the combination of high WIN-55,212 doses in the dorsolateral periaqueductal gray matter (dlPAG) with the TRPV1 antagonist capsazepine abolished the anxiogenic effect (90). Furthermore, the anxiolytic effects of high doses of the cannabinoid agonist ACEA combined with an antagonist of TRPV1 in the rat prelimbic medial prefrontal cortex (PL) suggested the critical interaction between both systems (91). Moreover, the co-administration of intra-dlPAG AEA at higher doses with a nitric oxide (NO) scavenger (carboxy-PTIO) restored the anxiolytic profile, leading to the hypothesis that the increase in anxiety-like behavior mediated by TRPV1 receptors is due to subsequent NO formation (92).

Deletion of *CNR1* gene in mice (CB1^{-/-} mice) has been another important tool to elucidate the role of this cannabinoid receptor in anxiety. Many studies have demonstrated the clear anxiety-like behavior of male CB1^{-/-} mice (70–72, 83, 93, 94), although there are some negative results (95). Among the multiple mechanisms involved in the anxious phenotype shown by CB1^{-/-} mice, significant age-dependent alterations in the metabolism of endocannabinoids could be pointed out (96). Interestingly, CB1^{-/-} female mice do not have an anxious phenotype in comparison with female wild-type (WT) subjects. This finding supports an interaction between sex and the ECS at early stages of development that is critical for establishing adult anxiety-like behavior (97). Indeed, these sex-specific effects were also described under pharmacological blockade of CB1r (98). Furthermore, our group described that the effects of the anxiolytic drugs bromazepam and buspirone were missing in CB1^{-/-} mice (99), suggesting a critical role of CB1r that was related with the control of GABAergic responses mediated by GABA_A and GABA_B receptors (100).

Recent studies provide relevant information regarding the specific brain regional involvement of CB1r-mediated anxiolytic actions. In this sense, the intra-dlPAG administration of AEA, ACEA (selective CB1r agonist) or AM404 (AEA reuptake inhibitor) induced anxiety-like responses that were blocked by AM251 (CB1r antagonist) (101). Similarly, AEA-mediated CB1r activation produces anxiolytic-like actions in the dlPAG employing a panic-like animal model (102, 103) or the Vogel conflict test (104). In addition, facilitation of 2-AG-mediated signaling in the dorsomedial hypothalamus (DMH) significantly reduced panic-like responses in Wistar rats, an effect that was reversed by the CB1r antagonist AM251 (105). Furthermore, activation of CB1r by 2-AG in the basolateral amygdala (BLA) has a critical role in the effects of stress-induced glucocorticoid

release on suppression of synaptic GABAergic inhibition (106). Interestingly, pharmacologically-induced elevations of AEA or 2-AG in the BLA decrease anxiety in the elevated plus maze (EPM) test under conditions of low emotional arousal while are ineffective when the level of emotional arousal increased (107). Moreover, electron microscopy revealed CB1r expression in the rat lateral habenula (LHb), mediating the actions of increased 2-AG levels after acute stress exposure, while its blockade by SR141716 (rimonabant) significantly reduced anxiety-like behavior (108). In another study, WIN-55,212 was locally administered in the lateral septum (LS) of male Wistar rats, producing a CB1r-mediated anxiogenic response in the EPM paradigm since AM251 blocked this effect (109). Also, the role of CB1r functional manipulation in anxiety behavior regulation and the effects on subsequent signaling pathways in relevant corticolimbic areas such as PFC, AMY, NAc, and Hipp (110–116) have been evaluated.

A better understanding of the functional connections of the ECS with other neurotransmitter or neurohormonal systems is relevant to understand the role of ECS components as potential biomarkers in psychiatry. According to previous studies, CB1r is located in the locus coeruleus (LC) and in the dorsal raphe nucleus (DRN), and it regulates noradrenaline (NA) and serotonin (5HT) release, respectively, by the modulation of GABAergic and glutamatergic terminals (117, 118). In addition, the dopaminergic and opiodergic systems of the Amy may also be involved in the anxiolytic-like effects induced by the activation of CB1r (119, 120). Furthermore, the involvement of the ECS in the regulation of the hypothalamus–pituitary–adrenal (HPA) axis after stress exposure attracted special attention in the last years (121). Gray and cols. recently found that the stress-related neuropeptide corticotropin-releasing hormone (CRH), acting through the CRH type 1 receptor (CRHR1), reduces AEA levels in the PFC and the Amy by increasing the hydrolysis of FAAH and by increasing 2-AG levels. These data suggest that stress-related elevations in CRH signaling induce persistent changes in eCB function, impairing its tonic regulation on stress and enhancing anxiety responses (122, 123).

Genetic studies pointed out interesting results regarding the involvement of polymorphisms or epigenetic modifications of *CNR1* as susceptibility/risk biomarkers to develop anxiety disorders. Lazary and cols. analyzed the interaction of the promoter regions of the serotonin transporter (5HTT; SLC6A4) and *CNR1* genes on anxiety. Specific constellations of CB1r and 5HTT promoters were closely associated with high or low synaptic 5HT concentrations, which could result critically in the vulnerability to experience an anxiety disorder (124). Hay and cols. employed CRISPR/CAS9 technology to disrupt a highly conserved regulatory sequence (ECR1) of the gene encoding CB1r (*CNR1*). This manipulation significantly reduced *CNR1* expression in the Hipp, but not in the Hyp, and induced a sex-dependent anxiogenic effect (125). In addition, a connection between ECS and epigenetic mechanisms was proposed. The exposure to immobilization stress increases anxiety-like behavior, an effect blocked by histone deacetylase (HDAC) inhibitors. Interestingly, the CB1r antagonist rimonabant

attenuated the anxiolytic-like effects of the HDAC inhibitors, suggesting an association between epigenetic mechanisms and ECS signaling (126). Furthermore, in mice exposed to a chronic unpredictable stress (CUS) there were reduced levels of histone H3K9 acetylation (H3K9ac) associated with CB1r encoding gene (127).

Since the direct pharmacological modulation of CB1r has provided some disappointing results, in recent years much attention has been paid to the therapeutic role of functional manipulation of the endogenous cannabinoid ligands AEA and 2-AG by inhibiting enzymatic degradation (FAAH and MAGL, respectively) or blocking reuptake (128–132). AEA plays a crucial role in emotional control (133). Inhibition of its degradation by FAAH or its reuptake induces a robust anxiolytic effect (134–142). Indeed, stress exposure induces anxiety-like behavior and reduces AEA brain levels (143) by increasing FAAH activity in the Amy (144), a brain region closely involved in AEA-mediated emotional regulation (145). According to the effects observed in FAAH *knockout* mice (FAAH^{-/-} mice), as well as with the administration of URB597 (FAAH inhibitor), preservation of CB1r function regulating GABA transmission in the striatum may be one of the mechanisms involved in the anxiolytic actions of FAAH inhibition (146, 147). Environmental experimental conditions are critical to observe the anxiolytic effect of FAAH inhibition, only present under high stressful or aversive stimuli (148). Interestingly, the dual blockade of FAAH and TRPV1 represents another therapeutic approach to reduce anxiogenic behavior (149). In addition, co-administration of an ineffective dose of URB597 with an ineffective dose of diazepam led to a synergistic anxiolytic action (77).

The endocannabinoid 2-AG also presents a close involvement in emotional regulation linked with signaling in hippocampal glutamatergic neurons (150). In the last years, several evidences support the anxiolytic actions associated with the inhibition of 2-AG enzymatic degradation by means of MAGL (151–153). A link with the HPA axis has been proposed, since the elevation of 2-AG levels was accompanied by a dramatic increase in plasma corticosterone, effect that is probably mediating its anxiolytic actions (154). In addition, increased 2-AG levels in the NAc of mice previously exposed to chronic social defeat stress are associated with an anxiolytic effect and the enhancement of synaptic plasticity (155). Furthermore, the enhancement of 2-AG levels in the dlPAG by the local injection of 2-AG or the hydrolysis inhibitor, URB602, prevented NMDA-induced panic-like response in Wistar rats (156). Interestingly, genetic deletion of MAGL in mice induced an anxiety-like phenotype (157), whereas mice lacking DAGL α showed a high anxiety-like phenotype, strengthening the critical involvement of 2-AG in emotional regulation (158, 159).

Another relevant endocannabinoid biomarker is the CB2r. The first studies demonstrating the role of this receptor in the regulation of anxiety-like behavior were performed in our laboratory employing transgenic animals overexpressing CB2r in the brain (CB2xP mice). Increased expression of CB2r was significantly correlated with reduced anxiogenic-related behaviors. Interestingly, CB2xP mice presented an impaired HPA-axis response to restraint stress, as well as increased

GABA_{A α 2} and GABA_{A γ 2} gene expression probably accounting for the lack of anxiolytic action of alprazolam in these animals (160). Furthermore, a pharmacological approach to evaluate acute and chronic effects of the activation (JWH133, CB2r selective agonist) or blockade (AM630, CB2r selective antagonist) of CB2r revealed opposite effects. Importantly, chronic CB2r blockade induced a significant anxiolytic effect that was associated with an upregulation of CB2r, GABA_{A α 2} and GABA_{A γ 2} in the cortex and the amygdala (160). In line with these results, the acute activation of CB2r by the administration of β -caryophyllene (BCP) induced an anxiolytic effect that was completely abolished by AM630-mediated CB2r blockade (161). Recently, Robertson and cols. described that CB2r gene expression is rapidly increased in the Hipp after social stress exposure (social defeat) (162). Genetic manipulation experiments allowed the deepening in the cell-specific functional involvement of CB2r in the Hipp, dissecting the effects of CB2r gene expression disruption in hippocampal neurons or microglia on the regulation of anxiety behavior (163). Moreover, the functional role of CB2r in VTA dopaminergic neurons was also explored. Surprisingly, deletion of CNR2 in VTA dopaminergic neurons induced a very significant anxiolytic effect (47).

Human Studies

In 1981, Fabre and McLendon published the first evidences regarding the anxiolytic properties of cannabinoid compounds. In this study, the synthetic cannabinoid nabilone was administered to 25 patients, producing a significant improvement in anxiety (164). Nowadays, there is a large body of evidence regarding cannabis consumption and regulation of anxiety behavior (165), although the underlying mechanisms are poorly understood. A recent study addressed this issue by combining fMRI and positron emission tomography (PET) in 14 patients following an oral dose of delta-9-tetrahydrocannabinol (THC) while they were performing a fear-processing task. The results suggested that the acute effects of cannabis on anxiety in males are mediated by the modulation of amygdalar function by THC and the extent of these effects are related to local availability of CB1r (166). On the other hand, several clinical trials using rimonabant to treat obesity showed psychiatric side effects such as increased anxiety behavior, depression or even suicidality (167). In spite of the presence of important confounding factors that probably were not appropriately taken into consideration (e.g. psychiatric comorbidity in obese patients), rimonabant was withdrawn from the market, and the enthusiasm in its therapeutic usefulness significantly decreased. Interestingly, a recent report suggested that rimonabant increases anxiety only under an aversive/anxiogenic situation (public speaking), without modifying baseline anxiety behavior (168). Alternative pharmacological approaches to modulate CB1r are now under investigation. Neutral antagonists, peripherally restricted ligands, and allosteric modulators may provide promising results [for a recent review (169)].

The elucidation of genetic variations of different endocannabinoid components involved in the vulnerability to develop anxiety-related disorders has recently gained great

interest. In this regard, Gonda and cols. evaluated the interaction between four categories of stressful life events and specific genetic variations in the CNR1 rs7766029 polymorphism, for the development of depression and anxiety. The results suggested that CNR1 rs7766029 interacted significantly with financial but not with other types of life events to increase the vulnerability to develop depression and anxiety (170). In addition, allelic variants of the gene encoding FAAH have been involved in the regulation of anxiety-related behaviors. First, the disturbances of FAAH genetic variation in AEA hydrolysis appear related with alterations in frontolimbic circuits (171), with an age-dependent effect accounting for differences between the adolescence and childhood life stages (172). Second, an interaction between genetic variations of FAAH and corticotropin-releasing hormone receptor type 1 (CRHR1) has been described in relation with the risk to develop anxiety disorders (173, 174). Third, reduced FAAH activity in patients carrying the A allele of the FAAH rs324420 (C385A) polymorphism significantly increases the vulnerability to develop anxiety and depression when exposed to repetitive childhood trauma (175). Moreover, a functional variant of gene encoding CB2r (Cnr2) appears to interact with FAAH gene, increasing the sensitivity for childhood trauma when both are dysfunctional (176).

Depressive Disorders

Major depressive disorder (MDD) has been one of the leading causes of years lived with disability (YLD) during the last three decades (1). According to the World Health Organization estimation for 2015, the number of people living with depression in the world is 322 million, and it is a major contributor to suicide deaths (177). DSM-5 states that the common feature of depressive disorders is the presence of sad, empty, or irritable mood, accompanied by somatic and cognitive changes that significantly affect the individual's capacity to function. MDD is being characterized by distinct changes in affect, cognition, and neurovegetative functions with episodes lasting for at least 2 weeks. Additionally, five or more symptoms have to be present during the same episode, with at least one of the symptoms being either depressed mood or anhedonia (63). Nowadays, pharmacological treatment of MDD entails relevant limitations such as delayed onset of antidepressive actions and appearance of important side effects. The limited success of drug discovery in the context of depression is ultimately linked to an inadequate understanding of the underlying biology of this disorder. In this sense, there is evidence to suggest that the ECS is impaired in MDD providing a unique opportunity to identify potential diagnostic and therapeutic biomarkers.

Animal Studies

Martin and cols. employed the CB1^{-/-} mice and exposed them to the CUS procedure. Their findings showed that CB1^{-/-} were more vulnerable to CUS-induced depressive-like responses and presented an increase susceptibility to develop anhedonia (94). Some years later, it was shown that the increased despair behavior in CB1^{-/-} mice was critically associated with down-

regulated brain-derived neurotrophic factor (BDNF) levels in the Hipp. Also, local administration of BDNF in the Hipp of these animals reversed the depressive-like phenotype (178). A complete genetic screening by mRNA microarray hybridization revealed a differential gene expression pattern related to the high depressive-like behavior of CB1^{-/-} mice at basal conditions (179). According to the results derived from the studies employing CB1^{-/-} mice, it was proposed that CB1^{-/-} mice could represent a validated and appropriated model to evaluate depressive-like disorders (180).

In the tail suspension test (TST) and forced swimming test (FST), acute AM251 injection induced an antidepressant effect, decreasing the immobility time in both behavioral paradigms (181). Similar results were obtained by both the acute and chronic administration of rimonabant in Wistar rats and BALB/c mice employing the FST and the chronic mild stress (CMS) paradigms, respectively (182). However, other results reveal that the activation of CB1r mediates antidepressant effects (183–187), and even that chronic rimonabant administration produces a depressogenic effect (188). Interestingly, McLaughlin and cols. showed that CB1r located in the dentate gyrus of the Hipp was responsible for the antidepressant effects of the CB1r agonist HU-210 (189). In addition, a very recent study elegantly discovered a circuit-specific CB1r-mediated modulation of glutamatergic transmission that shapes the information flow from BLA to the NAc (190). In this study, the authors consider if the reduction of CB1r in the NAc may be used as a biomarker for MDD diagnosis and point out that this aspect needs to be further determined by evaluation of CB1r levels in the NAc of MDD patients (190).

The evaluation of ECS components disturbances in animal models of depressive disorders provided relevant information. Hill and cols. showed that male Long-Evans rats exposed to the CUS presented increased CB1r binding site density in the PFC while decreased in the Hipp, Hyp and Nac, and lower levels of AEA were found in all these brain regions (191). Furthermore, sex-dependent effects of CUS were analyzed in Sprague-Dawley rats, obtaining lower and higher CB1r protein expression in males and females, respectively, whereas increased FAAH levels were present in both sexes (192). In addition, further studies employing the CUS procedure specifically focused on CB1r-mediated signaling, revealing significant loss of function disturbances in the NAc (193) and in the LHb (194). Moreover, apart from stress-related animal models, Flinders Sensitive Line (FSL) or Wistar Kyoto (WKY) rats are well-known genetic rat models of depression that were recently used to exhaustively analyze disturbances in different components of the ECS in specific brain regions and plasma (195, 196).

Enhancement of endocannabinoid signaling has been postulated as a new promising pharmacological strategy in the treatment of stress-related disorders (e.g. anxiety or depression) (197). Accordingly, a significant reduction in depressive-like behavior was found after the administration of the FAAH inhibitors URB597 (196, 198, 199) or PF3845 (200). In addition, the inhibition of MAGL by the administration of

JZL194 also yielded similar antidepressant effects in the CUS animal model of depression. Interestingly, JZL194-mediated effects may be related with an enhancement of adult neurogenesis and long-term synaptic plasticity in the dentate gyrus of the Hipp, probably activating mTOR signaling pathway (201). Furthermore, a recent study evaluated the effects of JZL195, a dual inhibitor of FAAH and MAGL, in WKY rats. JZL195 elevated the endocannabinoids and BDNF levels in the ventral striatum and reduced the depressive-like phenotype in female WKY rats (202).

In spite of the limited available results, CB2r is also critically involved in emotional regulation (203). Probably, the first evidence suggesting the role of CB2r in depression was a significant reduction of these receptors in the striatum, midbrain, and Hipp in an animal model of depression (204). Afterwards, a study showed the antidepressant effects of the CB2r-selective agonist GW405833 in rats (205). Interestingly, our group further evaluated CB2r involvement in depressive-like behavior regulation using genetic and pharmacological approaches. Mice overexpressing CB2r (CB2xP) presented decreased depressive-like behaviors under basal conditions or after the exposure to a CUS procedure. In addition, the chronic administration of AM630 blocked the CUS-induced depressogenic effect in stressed mice, effect associated with an upregulation of CB2r and BDNF in the Hipp (43). Recently, similar results were obtained by CB2r functional activation through the administration of the CB2r agonists JWH133 (206) and β -caryophyllene (207). Furthermore, the specific deletion of CB2r in midbrain DA neurons in DAT-Cnr2 conditional knockout (cKO) mice significantly increased depressive-like behavior (47).

Crosstalk of ECS with other neurotransmitter or neurohormonal systems plays a pivotal role in the effects produced by antidepressant drugs. In this regard, interactions with the serotonergic system represent a critical point due to the widely recognized clinical therapeutic usefulness of antidepressants targeting serotonin (e.g. serotonin selective reuptake inhibitors, SSRIs). SSRIs fluoxetine and escitalopram modify the concentrations of different ECS components under basal conditions (208–210) or in an animal model of depression (211). Furthermore, low doses of WIN-55,212 produced antidepressant-like actions that appeared to be mediated by 5HT (212), and CB1^{-/-} mice have decreased levels of 5HT transporter (5HTT) (213). Moreover, co-administration of a subeffective dose of fluoxetine potentiated the effect of subeffective doses of AEA, AM404 or URB597 (214). In addition, ECS also interacts with other systems involved in emotional and stress regulation such as the HPA axis (215), glutamatergic (216), opioidergic (217), and cholinergic (218) systems.

On the other hand, it is relevant to highlight that nonpharmacological approaches such as repeated transcranial magnetic stimulation (rTMS) improve depressive-like behavior, at least in part, by modulating the ECS. Recent studies performed in rodents exposed to CUS and subsequently treated with rTMS revealed that: 1) rTMS increases BDNF production and hippocampal cell proliferation to protect against CUS-induced changes through its effect on CB1r (219); 2) rTMS antidepressant effects are at least partly mediated by increasing hippocampal 2-AG and CB1 receptor expression levels (220); and 3) high-frequency

rTMS induces its antidepressant effect by upregulating DAGL α and CB1r (221). In addition, electroconvulsive therapy (ECT) significantly reduced AEA content and FAAH activity in the PFC of Sprague-Dawley rats, as well as decreased and enhanced binding site density of the CB1r in the PFC and Amy, respectively (222).

Human Studies

Besides the preclinical clues supporting the critical role of ECS in depression, currently there is a broad body of evidence available from clinical studies. Among them, those evaluating alterations in different ECS components in *post-mortem* brain tissue or plasma samples have provided compelling results. The first evidence revealed that CB1r protein expression was decreased in the anterior cingulate cortex (ACC) of patients with major depression (223). Furthermore, Choi and cols. showed that CB1r mRNA levels were higher in the PFC of major depression patients (224). However, in a recent study a lack of CB1r protein expression differences was found between depressive subjects and paired control patients (225).

In the last years, an increasing effort has been made to elucidate the alterations of ECS components (mainly the endocannabinoids AEA and 2-AG) in blood samples of patients with depression, to identify possible trait, prognosis or monitoring biomarkers that could improve the therapeutic approach. In a cohort of 28 women with diagnostic criteria for clinical depression and without medication, serum 2-AG content was significantly decreased, and this decrease was negatively correlated with duration of the depressive episode (226). Similarly, basal serum concentrations of AEA and 2-AG were significantly lower in women with nontreated major depression, and the exposure to a stressful situation significantly increased 2-AG concentrations without modifying AEA (227). However, another study described increased plasma concentrations of both AEA and 2-AG in depressed patients, and the elevation of 2-AG was significantly associated with SSRI antidepressant therapy (228). Interestingly, the antidepressant-related effects or physical exercise on eCBs levels were also analyzed. Intense exercise in control healthy patients induced a significant increase in AEA serum levels that was correlated with higher BDNF levels, whereas 2-AG concentrations remained stable (229). On the contrary, moderate exercise in women with MDD produced significant elevations in AEA but not in 2-AG, although both eCBs presented significant moderate negative associations between serum changes and mood states (230). Finally, ECT significantly elevated AEA and 2-AG levels in the cerebrospinal fluid (CSF) of patients with major depression (231).

The ECS-related polymorphic gene variant study results are relevant because of the potential diagnostic and therapeutic implications. Regarding the CNR1 and the single nucleotide polymorphism (SNP) rs1049353 (G1359A) that may contribute to the susceptibility to mood disorders (232), G-allele has been associated with higher depressive-related symptomatology (233) and increased risk of antidepressant treatment resistance in women with comorbid anxiety disorder (234). However, it provides a better response to citalopram in male depressive patients (235), whereas A-allele decreased risk to develop depression because of childhood physical abuse (236).

Furthermore, several CNR1 polymorphisms appeared to be related with high neuroticism and low agreeableness personality traits, increasing the risk to develop depression (237). On the other hand, the presence of 2 long alleles of the polymorphic triplet (AAT)_n of CNR1 gene was associated with reduced prevalence of depression in Parkinson's disease patients (238). In addition, the minor C allele of the CNR1 rs2023239 polymorphism may confer a protective effect against lifetime development of MDD in methadone-maintained patients (239). Despite the previous findings, a recent meta-analysis points out that CNR1 rs1049353 or AAT triplet repeat polymorphism had no association with susceptibility to depression (240).

Other relevant gene polymorphisms of the ECS are those related with FAAH and CB2r. First, variants of the FAAH gene may be related with susceptibility to mood disorders such as major depression (232). In fact, genetically reduced FAAH activity in A allele carriers of FAAH rs324420 (C385A) polymorphism constitutes a risk factor to develop anxiety and depression in patients exposed to repetitive childhood trauma. Interestingly, the authors noted that this genotype could entail pharmacogenomic consequences, namely ineffectiveness or adverse effects of FAAH inhibitors in this subpopulation (175). Second, polymorphisms of CNR2 were first studied by Onaivi and cols. in Japanese depressed patients, revealing a high incidence of Q63R but not H316Y polymorphism (204, 241). Recently, the R allele of Q63R CNR2 polymorphism, together with the A allele of FAAH C385A polymorphism were associated with increased sensitivity for childhood trauma and subsequent expression of anxious and depressive phenotypes (176). Finally, according to the previously mentioned recent meta-analysis performed by Kong and cols., CNR2 rs2501432 polymorphism might be closely associated with depression (240).

Schizophrenia

According to the DMS-5 schizophrenia is a psychotic disorder associated with a myriad of signs including positive symptoms (delusion, hallucinations, disorganized speech or grossly disorganized or catatonic behavior), negative symptoms (lack of motivation and social withdrawal), and cognitive symptoms (reduced attention and altered speech) (63, 242, 243). An extensive body of literature supports the role of ECS in schizophrenia neuropathology, a fact that is mainly sustained by the psychotic effects derived from cannabis consumption and attributed to the exogenous cannabinoid THC (244). Therefore, a great interest has been posed in the identification of specific biomarkers related with ECS functioning for preventive, diagnostic, or therapeutic purposes.

Animal Models

Preclinical research that focused on the role of ECS in schizophrenia relies on the evaluation of sensorimotor gating deficits by the prepulse inhibition (PPI) paradigm (245, 246). Among all the components of the ECS, CB1r is critically involved in schizophrenia. In fact, results of studies using pharmacological approaches showed that CB1r activation induces psychotic-like effects, while blockade of CB1r presents opposite actions.

Decreases in startle responses together with PPI disruption were achieved by CP-55,940 administration, and rimonabant completely reversed these effects (247). A similar experiment was carried out in which CP-55,940 decreased startle response and impaired PPI, and rimonabant significantly reversed CP-55,940-induced deficits in PPI only at the lower prepulse intensity (248). This CB1r-mediated auditory gating disruption was further confirmed by measuring neuronal network oscillations in the Hipp and entorhinal cortex of Sprague-Dawley rats. CP-55,940 significantly impaired sensory gating and neuronal oscillation, an effect that was reversed by AM251 (249).

After learning that the modulation of the CB1r produced sensorimotor alterations, different animal models of schizophrenia were used to find out if CB1r blockade could be a strategy with therapeutic potential. Blockade of N-methyl-D-aspartate (NMDA) receptors (NMDAr) was used to simulate schizophrenia-like symptoms in rodents (250). Interestingly, the administration of AM251 significantly abolished phencyclidine-induced disruption of PPI in a similar way to clozapine (251), as well as impairments in recognition memory or increased behavioral despair in the FST (252). Another NMDAr antagonist used to model schizophrenia-like behavior is MK-801. The administration of the CB1r antagonist AVE1625 reversed MK801-induced cognitive impairments and decreased catalepsy and weight gain induced by clinically used antipsychotic drugs (haloperidol, olanzapine) (253). Furthermore, AM251 attenuated amnesic effects and hyperactivity induced by MK-801 (254). Therefore, it appears that blockade of CB1r may have relevant therapeutic applications for the treatment of schizophrenia.

In the so-called 'three-hit' animal model of schizophrenia, CB1r binding and cannabinoid agonist-mediated G-protein activation decreases in the cortical, subcortical, and cerebellar brain regions (255). In a neurodevelopmental animal model of schizophrenia induced by the gestational administration of methylazoxymethanol (MAM), CB1r mRNA levels were lower in the PFC and higher in the dorsolateral striatum of adult MAM-treated Sprague-Dawley rats relative to the control group (256). Moreover, in the spontaneously hypertensive rat (SHR) strain, partially reproducing some schizophrenia-like behavioral aspects, CB1r immunoreactivity was significantly increased in the PL, cingulate cortex, and CA3 region of the Hipp (257). Recently, two studies found decreased methylation of the cannabinoid receptor interacting protein (CNR1P1) DNA promoter in the ventral Hipp (vHipp) of rats exposed to the MAM model (258, 259). CNR1P1 is an intracellular protein that interacts with the C-terminal tail of CB1r and regulates its intrinsic activity. Interestingly, a lentivirus-mediated overexpression of CNR1P1 in the vHipp of Sprague-Dawley rats induced significant schizophrenia-like cognitive and social interaction impairments, together with an increase of dopamine neuron population activity in the VTA (260).

Apart from the many preclinical studies supporting the pivotal role of CB1r, animal models of schizophrenia provided interesting results about eCBs brain level alterations. In this regard, a significant increase in 2-AG levels in the PFC of PCP-treated Lister-Hooded rats was reversed by treatment with THC, which in turn induces a large reduction of AEA in the same

region (261). Furthermore, in Sprague-Dawley rats exposed to a bilateral olfactory bulbectomy, considered as an animal model of depression and schizophrenia, a significant decrease of AEA and 2-AG levels was found in the ventral striatum (262). In addition, mice with a heterozygous deletion of neuregulin 1 (Nrg 1 HET mice), a well-accepted and characterized animal model of schizophrenia (263), displayed relevant alterations in eCBs levels (264).

Finally, CB2r has also been recently involved in schizophrenia. Ishiguro and cols. studied the effects of the pharmacological blockade of CB2r in two animal models of schizophrenia induced by the administration of MK-801 or metamphatamine. The CB2r antagonist AM630 significantly exacerbated the MK-801- or metamphatamine-induced hyperlocomotion and PPI disruption, suggesting that CB2r was mediating these actions (265). Our group analyzed exhaustively the behavioral profile of CB2^{-/-} mice to evaluate the implication of CB2r in schizophrenia-like behavior. The phenotype showed by CB2^{-/-} mice resembled some relevant features of schizophrenia such as increased sensitivity to motor effects of cocaine, anxiety- and depressive-like behavior, disrupted short- and long-term memory consolidation and impaired PPI. These behavioral alterations were accompanied by gene expression changes in different targets from dopaminergic, noradrenergic, and serotonergic systems. Interestingly, the atypical antipsychotic risperidone significantly improved PPI disruption induced by CB2r deletion and differentially modulated some of the neurochemical disturbances compared with WT mice (266). In addition, the activation of CB2r by the agonist JWH015 reversed PPI disruptions of the MK-801-induced animal model of schizophrenia, and this effect was specifically mediated by CB2r since only AM630 but not AM251 abolished PPI improvement (267). Furthermore, activation of CB2r (JWH133) and blockade (AM630) increased MK-801-induced hyperlocomotion, although this effect was much more evident and pronounced with AM630 (268). Therefore, these results strongly suggest that CB2r functional regulation is significantly involved in schizophrenia-like behavior.

Human Studies

To date, an extensive and great effort has been made to elucidate the role that CB1r plays in schizophrenia. Accumulated clinical data clearly shows significant alterations of CB1r protein and gene expression levels, as well as certain CNR1 polymorphisms correlations, especially in the brain but also in the peripheral blood cells from schizophrenic patients in comparison with healthy control subjects. The information reviewed and detailed below provides important clues to further investigate the application of CB1r-related measures as potential trait, state, prognostic or even therapeutic biomarkers.

Several published studies analyzed CB1r protein and gene expression levels in different *post-mortem* brain regions from schizophrenic patients. Several studies examined quantitative autoradiography to evaluate CB1r availability through the binding of different radioligands. A significant increase in CB1r availability was shown in the dorsolateral prefrontal cortex

(DLPFC) (269–271), although this increase was only present in paranoid schizophrenic patients (272). Interestingly, a recent study failed to show differences in CB1r-mediated functional coupling to G-proteins in the PFC of schizophrenic and control patients (273). Furthermore, higher CB1r binding levels were shown in the left ACC (274) and in superficial layers of the posterior cingulate cortex (PCC) (275), whereas no changes were found in the superior temporal gyrus (STG) (276) from schizophrenic patients. In contrast, some authors reported lower CB1r protein levels measured by immunocytochemistry (277) or Western blot (278) and decreased CB1r gene expression analyzed by *in situ* hybridization (277) or quantitative real time polymerase chain reaction (qRT-PCR) (273) in the PFC from schizophrenic patients compared with control subjects. Volk and cols. specifically addressed this apparent discrepancy between CB1r binding and protein or gene expression levels. In a cohort of 21 schizophrenic patients presenting lower levels of both CB1r mRNA and protein in the PFC, relative to matched healthy comparison subjects, they obtained an increased CB1r binding (271).

Neuroimaging experiments were recently carried out to obtain an *in vivo* approximation of the disturbances related with CB1r in schizophrenia. In this regard, PET studies yielded dissimilar results. Wong and cols. studied CB1r binding employing the novel PET tracer [¹¹C]-OMAR (JHU 75528) in schizophrenic patients and matched controls. CB1r binding was higher in several brain regions of patients with schizophrenia, only reaching statistical significance in the pons. Interestingly, a significant correlation was found between CB1r binding and schizophrenia-related symptomatology (279). In addition, Ceccarini and cols. also showed a significant increase of CB1r binding in the NAc, insula, cingulate cortex, inferior frontal cortex, parietal and mediotemporal lobes in schizophrenic patients compared with controls measured with [¹⁸F]-MK-9470 PET. It is relevant to highlight that in the nontreated schizophrenia patients, CB1r binding was negatively correlated to negative symptoms and to depression scores, especially in the NAc (280). On the contrary, Rangathan and cols. obtained an opposite result with lower CB1r availability levels ([¹¹C]-OMAR PET) in the Amy, caudate, PCC, Hipp, Hyp, and insula of schizophrenic patients (281). An interesting commentary on these discrepancies was published, in which several confounding factors such as symptom severity, sex, age, PET tracer, statistical analysis method or comorbid nicotine use are discussed. Overall, it could be concluded that CB1r has an important but yet complex and poorly understood role in schizophrenia (282). Finally, a very recent study examined CB1r availability by [¹⁸F]-FMPEP-d2 or [¹¹C]-MePPEP PET, in first episode psychosis (FEP). Significant lower CB1r availability was found in patients with schizophrenia, independently of antipsychotic medication treatment. Greater reduction in CB1r availability was associated with greater symptom severity and poorer cognitive functioning (283).

The possible association between CNR1 polymorphisms and schizophrenia has been explored. In this sense, negative results were obtained with a single-base polymorphism within the first

exon of the CNR1 (284), the polymorphism rs1049353 1359G/A at codon 453 in the coding region of CNR1 (285–288), or other CNR1 polymorphisms such as rs6454674 (287), AL136096 (287), rs806368 (288, 289), rs806379 (288), rs806380 (288), rs806376 (289), and rs806366 (289). However, significant associations of CNR1 polymorphisms rs7766029, rs806366, and rs1049353 were described (290). Regarding (AAT)_n triplet repeat in the promoter region of the CNR1 gene, discrepant results were reported since Tsai and cols. suggested that this polymorphism was not directly involved in the pathogenesis of schizophrenia in a Chinese population (291), whereas it was significantly associated with the hebephrenic or disorganized subtype of schizophrenia (285). Interestingly, some relevant associations were identified between specific CNR1 polymorphisms and therapeutic response. Hamdani and cols. described increase G-allele frequency of the rs1049353 polymorphism in responsive schizophrenic patients, with a dose effect of the G allele. Thus, the authors proposed that the G allele of CNR1 rs1049353 polymorphism could represent a “psychopharmacogenetic” biomarker to take into consideration for the treatment of schizophrenia (288). In addition, in 65 FEP patients, TT genotype of the CNR1 rs2023239 polymorphism was associated with a better improvement of negative and positive symptoms (292). Similarly, in another group of patients with FEP, carriers of rs7766029 CC genotype presented significantly higher improvement in verbal memory and attention while carriers of rs12720071 AG genotype showed a better improvement in executive functions (293). Furthermore, minor alleles of CNR1 polymorphisms rs6928499, rs1535255, and rs2023239 might be associated with a lower risk to develop antipsychotic-induced metabolic syndrome. These relevant data could result in potential pharmacogenetic applications to optimize drug management of schizophrenic patients (294). On the contrary, one study showed that G allele carriers of the CNR1 rs1049353 (G1359A) polymorphism might be associated with a poorer therapeutic response (233).

Gene and protein analysis of CB1r in peripheral blood cells from schizophrenic patients attracted much attention. Peripheral cell (e.g. lymphocytes) changes could be mirroring, at least in part, some of the neuropathological hallmarks of the disorder. In this regard, the first published study did not detect changes in the CB1r mRNA levels in peripheral blood mononuclear cells (PBMCs) between schizophrenia and control patients (295). Similarly, no differences were observed in CB1r levels of peripheral immune cells by flow cytometry between control and schizophrenic patients, although a positive correlation between CB1r expression on monocytes and cognitive impairment was detected (296). However, an opposite result revealed an increase of CB1r in PBMCs of schizophrenic patients also evaluated by flow cytometry (297). Furthermore, there is an increase of CB1r mRNA levels in PBMCs of schizophrenic patients (298, 299) that may correlate with a reduced DNA methylation of CNR1 promoter region (299). Moreover, CB1r gene expression was correlated positively with positive and negative syndrome scale (PANSS) total symptom severity and negatively with cognitive functioning measures (298).

Besides the extensive literature evaluating the role of CB1r in schizophrenia, some efforts were done to complete the picture regarding the involvement of eCBs and its degrading and synthesizing enzymes as biomarkers. Leweke and cols reported a significant increase of AEA levels in the CSF of schizophrenic patients (300). In antipsychotic naïve first-episode paranoid schizophrenic patients, there was an eightfold increase in AEA levels in the CSF, whereas no alteration was present in patients treated with typical but not atypical antipsychotics. Furthermore, AEA levels were negatively correlated with psychotic symptoms in nonmedicated acute schizophrenics (301). Similarly, blood AEA levels were higher in patients with acute schizophrenia and were normalized with the clinical remission (295). Increased AEA levels were also detected in the CSF of schizophrenic patients who used cannabis. Interestingly, the increase of AEA was more than 10-fold higher in low-frequency compared with high-frequency cannabis users (302). In addition, higher AEA serum levels were obtained in twin-pairs discordant for schizophrenia (303), or in schizophrenic patients with substance use disorder (SUD) comorbidity, considering that baseline AEA predicted endpoint SUD scores (304). However, other studies showed different results such as no changes in serum AEA levels (305), increased 2-AG and decreased AEA in the cerebellum, Hipp, and PFC of schizophrenic patients (306).

With regard to degrading or synthesizing eCBs enzymes, the relationship of some FAAH or NAPDE-PLD polymorphisms with schizophrenia was studied, but no significant associations were obtained (290, 307). In addition, FAAH and MAGL mRNA levels were similar while FAAH activity was higher in the PFC of schizophrenic patients compared to controls (273). Interestingly, a reduction of FAAH mRNA levels correlated with clinical remission in schizophrenic patients (295). Furthermore, in FEP patients, some interesting correlations were detected between peripheral FAAH and DAGL expression and short-term verbal memory, NAPE-PLD expression and working memory, and MAGL expression and attention. Accordingly, the authors suggested the use of these ECS elements as biomarkers or pharmacological targets for FEP (308). Finally, mRNA levels of the 2-AG metabolizing enzyme, α - β -hydrolase domain 6 (ABHD6), were significantly increased in patients with schizophrenia (309).

Notwithstanding the scarce literature exploring the role of CB2r in schizophrenia, some important findings suggest its involvement and draw attention to research on its therapeutic potential. Perhaps, de Marchi and cols. published the first evidence measuring CB2r mRNA levels by semi-quantitative RT-PCR in PBMCs from schizophrenic patients in their acute phase, and when clinical remission was achieved after antipsychotic treatment with olanzapine. CB2r gene expression significantly decreased in PBMCs from patients in clinical remission (295). In FEP patients, CB2r protein expression was significantly down-regulated together with reduced levels of eCBs synthesizing enzymes (NAPE-PLD and DAGL) (310). Interestingly, increased CB2r gene expression was found in schizophrenic patients' PBMCs (298), correlating with PANSS and cognitive performance severity (296, 298), and in cells of the

innate immune system (297). On the other hand, Ishiguro and cols. evaluated the implication of specific CNR2 polymorphisms in schizophrenia. R63 allele of rs2501432 (R63Q), C allele of rs12744386, and the haplotype of the R63-C allele were significantly increased in patients with schizophrenia in comparison with control subjects. Apparently, these polymorphic alterations of CNR2 are associated with loss of function. A lower response to CB2r ligands was found in cultured CHO cells transfected with the R63 allele. Reduced CB2r mRNA and protein expression levels were found in the DLPFC of schizophrenic patients independently of the diagnosis (265). In addition, the association between three CNR2 polymorphisms (rs2501432C/T, rs2229579C/T, rs2501401G/A), and schizophrenia was explored (311). However, other CNR2 polymorphisms (rs6689530 and rs34570472) were not associated with schizophrenia in a Korean population (289).

Bipolar Disorder

Bipolar disorder (BD) is a debilitating, lifelong neuropsychiatric illness characterized by unsteady mood states alternating from (hypo)mania to depression. According to the DSM-5, for a diagnosis of BD it is necessary to meet specific criteria for a manic episode that may be followed by hypomanic or major depressive episodes (63). Despite the availability of effective pharmacological agents, BD is inadequately treated in a subset of patients, so the identification of new therapeutic targets is necessary. In this sense, the close implication of ECS in mood regulation suggested its involvement in BD (312). This assumption is supported by the observation of the effects of high doses of cannabis and THC in healthy patients, producing psychosis, sometimes with marked hypomanic features (313). In addition, THC and cannabidiol (CBD), the main components of *Cannabis sativa* plant, may present mood stabilizing properties. Therefore, there is an increasing interest to evaluate ECS implication in BD.

First studies were focused on the evaluation of polymorphisms of CNR1 gene in BD pathophysiology. One study carried out in patients with BD within a Turkish population investigated the implication of three types of polymorphisms of CNR1 in this disease, demonstrating that only one of them (rs6454674) could be correlated with BD. In addition, the mean of the yearly manic attacks was statistically higher in patients presenting heterozygote rs6454674 T/G polymorphisms compared to those with homozygote polymorphism (314). In addition, the association of CNR1 rs1049353 (1359 G/A) and FAAH rs324420 SNP (cDNA 385C to A) polymorphisms with BD was assessed in a Caucasian population. Here, the authors concluded that the distribution of CNR1 1359 G/A genotypes and alleles did not differ between BD and healthy patients, whereas the frequency of the AC genotype of FAAH (cDNA 385C to A) polymorphism was slightly higher in BD patients (232).

Nevertheless, other studies did not identify differences in ECS components between BD and healthy controls. Indeed, no differences were obtained between BD patients and healthy controls in DNA methylation of the CNR1 gene promotor region (299). Furthermore, a polymorphism of CNR1

promotor region was evaluated in another study, and no changes were observed in BD patients, concluding that it was not likely to relate with BD (315). Koethe and cols. carried out a study with *post-mortem* brain samples from BD patients and controls, evaluating numerical density of neurons and immunopositive glial cells for CB1r. No changes were found in these patients (223). Furthermore, in another study evaluating polymorphisms of CNR1 and FAAH, no significant differences or association were observed in BD patients (316).

Because of these contradictory results, some authors shifted their attention to the implication of CB2r in BD, with limited but promising findings. A genetic association was observed in patients with BD and CNR2 rs41311993 (524C/A) polymorphism, but not SNPs of rs2229572 (1073C/T) or rs2501432 (315A/G), suggesting that CB2r may play a role in BD (317). In addition, a genome-wide association study carried out in a population from the UK biobank, identified the association of a locus in CNR2 with distressing psychotic experiences, providing support for a shared genetic liability with BD and other neuropsychiatric disorders (318).

In summary, there is limited information about the implication of ECS in the pathophysiology of BD. Thus, more preclinical and clinical studies are needed to explore further its role in the development of this neuropsychiatric disorder and its usefulness as a therapeutic target to improve BD management.

Post-Traumatic Stress Disorder

Post-traumatic stress disorder (PTSD) is a chronic and disabling mental disease caused by the exposure to stressful, frightening or distressing events, and is included in the category of trauma- and stressor-related disorders in the DSM-5 (63). PTSD patients experience intrusion symptoms, persistent avoidance of any stimuli associated with the traumatic event, negative alterations in cognition and mood, and disturbances in arousal and reactivity, that must last more than 1 month and produce distress or functional impairment (63). The neurobiological mechanisms underlying PTSD-related symptomatology are not completely understood, being a limiting factor to identify new therapeutic targets. In this regard, a relevant association between ECS and PTSD was suggested providing interesting results about the potential development of new pharmacological approaches. Indeed, preclinical and clinical findings point out the involvement of certain ECS components in PTSD symptomatology, such as CB1r or FAAH, suggesting its potential role as biomarkers for PTSD (319, 320).

Animal Studies

The involvement of CB1r in PTSD is supported by the presence of this receptor in brain areas regulating the response to stress and to changes observed in different animal models of PTSD. For instance, using a shock and reminder model of PTSD, higher mRNA levels of CB1r were detected in the BLA (133), and increased CB1r protein expression was found in the BLA and the CA1 region of the Hipp (321) of exposed mice as well. On the other hand, in a predator exposure-based PTSD model, anxiety-like behavior was negatively correlated with CB1r gene expression in the PFC and the amygdaloid complex, whereas

no changes were observed in the Hipp (322). In addition, Xing and cols. reported in young Sprague-Dawley rats, that the exposure to an unpredictable electric shock model of PTSD induced a down-regulation of CB1r gene expression in comparison with nonstressed rats. Interestingly, the authors showed sex differences in the stress-related regulation of CB1r, showing that females presented higher mRNA levels of CB1r, as well as greater CB1r inactivation by phosphorylation. The authors concluded that these sex-related differences could lead to increased susceptibility to stress-related anxiety disorders, including PTSD, in females (323). A genetic approach was used to evaluate further the involvement of CB1r in the regulation of stress response. Repeated exposure to an acoustic stressor (high intensity bell sound) did not produce changes in adrenocorticotropin hormone (ACTH) or corticosterone (CS) levels in CB1^{-/-} mice. These results suggested that the presence of CB1r is essential in the regulation of the stress response, and that CB1^{-/-} mice may result appropriate to model some forms of PTSD (324).

Pharmacological manipulation approaches of the CB1r were also explored in several rodent models of PTSD and its potential usefulness as a therapeutic biomarker. The blockade of CB1r with rimonabant increased freezing behavior in a PTSD model of shock and reminder during cued expression/extinction training (325). On the other hand, several authors evaluated the effects of cannabinoid activation by WIN-55,212-2 administration into hippocampal CA1 region. The results showed a normalization of shock-induced upregulation of CB1r in the PFC and CA1 region of the Hipp (321, 326) and facilitation of inhibitory avoidance extinction in a fear-related inhibitory avoidance paradigm (327). All these effects were blocked by AM251 administration. Furthermore, WIN-55,212 administration into the BLA normalized stress-induced effects on inhibitory avoidance and acoustic startle response and facilitated fear extinction in a single prolonged stress (SPS) model of PTSD. These effects were blocked by AM251 (321, 328, 329). Similarly, the injection of WIN-55,212 in the NAc of rats exposed to a shock and reminder model of PTSD significantly facilitated the fear extinction process (330). Interestingly, Goodman and Packard demonstrated that systemic or intradorsolateral striatum (DLS) administration of WIN-55,212 could impair the consolidation of stimulus-response memory, suggesting relevant consequences for neuropsychiatric disorders such as PTSD (331). However, the intra-PFC administration of WIN-55,212 did not modulate fear extinction disturbances induced by the exposure to the SPS model (329). Finally, according to the results obtained with WIN-55,212, Reich and cols. studied the effects of a CB1r selective agonist, ACEA, in rats exposed to 3 weeks of a chronic-mild-unpredictable protocol followed by fear conditioning evaluation. In this study, ACEA administration significantly reduced freezing behavior in stressed rats, enhancing long-term extinction of fear-related memories (332).

In order to validate that cannabinoid activation improves disturbances induced by stress- or trauma-related stimuli, the effects of pharmacological endocannabinoid signaling facilitation were analyzed. In a fear conditioning paradigm, the

administration of AM404 led to a dose-dependent enhancement in fear extinction, as well as a decreased shock-induced reinstatement of fear. Interestingly, the administration of rimonabant together with AM404 reversed the improvement of extinction, suggesting that AM404 effects were related to an increase in CB1r activation during extinction training (333). In addition, the injection of the FAAH inhibitor, URB597, normalized the upregulation of CB1r in the CA1 of Hipp and BLA of rats exposed to a shock and reminder model of PTSD (334) and attenuated startle response and anxiety-like behavior in a predator exposure animal model of PTSD (335). Interestingly, these effects were abolished by CB1r blockade, suggesting the implication of CB1r on URB597 effects (335, 335). Similar results were obtained with the administration of URB597 into CA1 (Hipp) and BLA brain regions, showing a facilitation of extinction processes and attenuation of startle response, anxiety- and depression-like behaviors mediated by CB1r activation (133, 336, 337). Furthermore, URB597 administration additionally prevented the increase of CB1r levels in CA1 and BLA after rodent exposure to shock and reminder model of PTSD (321).

FAAH inhibition significantly facilitates CB1r-mediated signaling of AEA and can produce a greater beneficial spectrum of biological effects than those caused by direct CB1r activation. Interestingly, the role of FAAH in learning and memory was evaluated by using another FAAH inhibitor, OL-135. The administration of this drug increased acquisition and extinction rates in mice exposed to fixed platform water maze test. Rimonabant blocked OL-135-induced effects on both acquisition and extinction levels (338). In the same study, the authors revealed that FAAH^{-/-} mice phenotype was similar to that obtained after OL-135 administration, suggesting that the increase in AEA levels facilitates extinction processes, and that CB1r would be critically involved (338). FAAH inhibition and the consequent increase of AEA in the brain regions involved in the regulation of stress and anxiety seem to restore dysfunctional homeostasis of AEA signaling because of stress exposure. Thus, FAAH must be strongly considered as a target for PTSD management (139).

Human Studies

According to the involvement of ECS components in several behavioral traits of PTSD in animal models, various studies explored alterations in different biological samples (*post-mortem* brain tissue, blood, hair) collected from PTSD patients and adequately paired controls. At the peripheral level, some authors studied the possible correlation between CNR1 polymorphisms and PTSD symptoms. The rs1049353 polymorphism of CNR1 was studied in PTSD patients to correlate specific alleles or genotypes with fear and/or dysphoric symptoms of PTSD. This study suggested that rs1049353 polymorphism interacts with childhood physical abuse to increase fear but not dysphoric symptoms in PTSD (339). In another study carried out in a Caucasian population, the association between variants of CNR1 gene haplotypes and diagnosis of PTSD was studied. The authors reported that the

variant C-A was more common in PTSD cases compared to non-PTSD controls, and the variant C-G was less common in PTSD compared to non-PTSD patients (340).

A different approach was the measurement of plasmatic eCBs in a selected cohort of patients that suffer the terroristic attacks of the World Trade Center in 2001 and met the diagnostic criteria for PTSD. 2-AG, AEA, and cortisol concentrations were measured. Only 2-AG was significantly reduced in PTSD patients, while no significant differences were found in AEA or cortisol concentrations (341). Another study showed reduced AEA and cortisol concentrations in PTSD patients compared to healthy controls with lifetime clinical histories of trauma (342). Despite these contradictory results, the fluctuations in plasmatic concentrations of eCBs may affect the reproducibility of the evaluation. Thus, the assessment of eCBs alterations in hair samples provides a more stable measurement. Hair concentrations of PEA (palmitoylethanolamide), OEA (oleoylethanolamide) and SEA (stearoylethanolamide) were measured in war survivors with and without PTSD. A regression analysis revealed a strong negative relationship between these endocannabinoids and the severity of PTSD symptoms. OEA concentrations were significantly reduced in hair samples from PTSD patients (343).

Only one human study analyzed CB1r binding using the CB1r-selective radioligand [¹¹C]OMAR by PET. Results showed elevated CB1r binding values, especially in women, together with lower AEA and cortisol in PTSD patients. The authors suggested that abnormal CB1r-mediated AEA signaling is involved in the etiology of PTSD (342). In addition, fMRI was also used to evaluate FAAH implication in PTSD symptomatology. A common SNP (C385A) in the human FAAH gene was correlated with the quicker habituation of amygdala reactivity to threat and lower score on stress-reactivity. This variant reduced FAAH activity and possibly increased AEA-induced endocannabinoid signaling (344–346). Furthermore, Rabinak and cols. conducted an fMRI study with healthy volunteers and patients receiving acute dronabinol (synthetic THC) oral administration in a standard Pavlovian fear extinction paradigm. Interestingly, dronabinol enhanced extinction learning, providing the first evidence about the feasibility of pharmacological enhancement of extinction learning in humans using cannabinoid system modulators (347, 348). Some clinical trials with PTSD patients suggested the usefulness of dronabinol for improving the global PTSD symptom severity, sleep quality, frequency of nightmares, and PTSD hyperarousal symptoms (349). Similar results were obtained with nabilone, since its administration to PTSD patients improved insomnia, PTSD symptoms, and global assessment of functioning, reducing the frequency and intensity of nightmares (350, 351). Nevertheless, more randomized and controlled clinical trials are needed to confirm dronabinol or nabilone potential therapeutic application in the management of PTSD.

Attention-Deficit/Hyperactivity Disorder

ADHD is a neuropsychiatric disorder characterized by persistent pattern of inattention and/or hyperactivity-impulsivity that interferes or reduces the quality of social, academic, or occupational functioning in accordance with DSM-5 (63). In

the last years, the identification of different components of the ECS that are potentially involved in ADHD pathophysiological mechanisms has attracted much attention as shown below.

Animal Studies

An experiment carried out in SHR rats (an animal model reproducing some features of ADHD) evaluated the modulating effects of the cannabinoid system on impulsivity, using a delay reinforcement task and the administration of WIN55212-2 or AM251 (352). This study concluded that treatment with WIN55212-2 decreased whereas AM251 increased the choices of the large reward, suggesting that CB1r plays a relevant role in impulsive behavior. Furthermore, basal gene and protein expression of CB1r in the brainstem of SHR rats was significantly lower in comparison with their normotensive counterpart, Wistar rats (353). Moreover, the overexpression of four genes, between them CNR1, was strongly associated with overall poor performance on mice during their gestational growth because of a malnutrition *via* high-fat or low-protein diets on the dam (354). These abnormal disturbances on diet in the gestational period are linked to the etiology of multiple neurodevelopmental disorders, including ADHD (355).

The psychostimulant drug amphetamine is often prescribed to treat ADHD. The administration of amphetamine increases monoamine neurotransmission in the brain regions as NAc and medial PFC. Accumulating reports supported the role of CB1r in the regulation of monoamine release, suggesting its possible involvement in ADHD. The administration of rimonabant did not affect monoamine release whereas dose-dependently abolished amphetamine-induced dopamine release in the NAc. This result suggested that CB1r is essential to reach the therapeutic effect of amphetamine, mediated at least in part, by the enhancement of dopaminergic signaling in the mesolimbic system in the NAc (356).

Human Studies

As previously stated, there is large available evidence regarding the role of different variants of CNR1 gene in psychiatry. In relation to ADHD, SNP variants at the CNR1 gene were tested on a family-based sample of trios (an ADHD child and their parents) and on an unselected adolescent sample from Northern Finland. The study detected a significant association of a SNP haplotype (C-G) with ADHD suggesting a greater risk in males than females (340). Another study reported the interaction of the two most studied CNR1 polymorphisms, rs806379 and rs1049353, that are involved with early psychosocial adversity (357). These polymorphisms of the CB1 receptor are highly associated with impulsivity representing an usual phenotype involved in ADHD (358).

Eating Disorders

The most common eating disorders are anorexia nervosa (AN) and bulimia nervosa (BN). According to DSM-5, AN is characterized by distorted body image and excessive dieting leading to severe weight loss with a pathological fear of becoming fat, whereas BN is characterized by recurrent episodes of binge eating alternated with recurrent

inappropriate compensatory behavior to prevent weight gain (63). Importantly, ECS plays a major regulatory role on feeding behaviors and energy balance (359) that has drawn attention to its relationship with ADHD neurobiology. This can be confirmed by rodents with diet-induced or genetic obesity promoting an increase of endocannabinoid hypothalamic levels (360).

Animal Studies

The participation of CB1r in the regulation of feeding behavior is well established. The acute administration of rimonabant decreased food intake and body weight gain and reduced CB1r gene expression in the PVN of male Wistar rats. However, a chronic treatment led to tolerance to the hypophagic effects of CB1r blockade without changes in food intake, body weight, or hypothalamic mRNA gene expression (361). Another study on rimonabant-treated male Sprague-Dawley rats showed that the colocalized Fos labeling of hypothalamic regions with anorexigenic and orexigenic peptides had decreased neuropeptide Y (NPY) levels (362). More recently, another study was performed in CB1r conditional and CB2^{-/-} mice. The hypothalamic neuropeptide expression pattern displayed a marked decrease of proopiomelanocortin (POMC) and cocaine-amphetamine-regulated transcript (CART) expression in the arcuate nucleus of the hypothalamus (ARC), both neuropeptides involved on anorexigenic and behavioral changes in food intake (363).

C57BL/6J mice were treated with naltrexone (opioid receptor antagonist), rimonabant, and BD-1063 (sigma-1 receptor antagonist) on an intermittent maladaptive feeding animal model. All the treatments reduced overconsumption of a palatable food (364). In addition, the administration of other cannabinoid compounds such as CBD or CB1r antagonists significantly reduced food intake and body weight gain (365–368). Recently, a study based on the activity-based anorexia (ABA) model reproducing key aspects of human AN, measured levels of AEA, 2-AG, and the CB1r in different brain regions of female ABA Sprague-Dawley rats. 2-AG significantly decreased in various brain areas but not in the caudate putamen, whereas no changes were observed in AEA. Density of CB1r was reduced in the dentate gyrus of Hipp and in the lateral Hyp (369). These results suggested that ECS is involved in the contribution and maintenance of some aspects in the pathophysiology of AN.

Human Studies

Although there is some progress in the understanding of the mechanisms underlying eating disorders and body weight regulation, there is still lack of information to suggest cannabinoid related treatment for patients with AN and BN. However, some studies suggest that the ECS, primarily CB1r, plays a key role in the reward areas and metabolic patterns involved in food intake and weight gain. In this regard, a PET study on 54 patients with food intake disorders (FID, including AN and BN) revealed an inverse association between regional CB1r availability and body mass index (BMI) in the Hyp and brainstem areas in both patients with FID and healthy individuals. However, FID patients negatively correlated with BMI throughout

the mesolimbic reward system (370). Also, global CB1r availability is significantly increased in the cortical and subcortical brain areas in AN patients compared with healthy controls, maybe due to a compensatory mechanism of an underactive ECS in these patients (371). Finally, eating disorder female patients presented lower CB1r mRNA levels in PBMCs (372).

CONCLUSIONS

The close involvement of the ECS in the etiology and neuropathology of neuropsychiatric disorders is undeniable. Considering the urgent need to identify new and better biomarkers in psychiatry, the evidence included in this review provides an overview of the opportunities that cannabinoid receptors (**Table 1**), endogenous cannabinoid ligands (**Table 2**), or their metabolizing enzymes (**Table 3**) offer as potential biomarkers in the clinical setting. The large number of pharmacological studies with various cannabinoid compounds, mainly conducted in animal models, reported interesting and promising information to design new therapeutic strategies that, alone or in combination with the drugs currently used in psychiatry, may improve the efficacy and safety of psychiatric disorders treatment.

According to the information gathered in the present review from pharmacological and genetic approaches mainly performed in rodents, some general conclusions can be drawn regarding the usefulness of ECS components as therapeutic biomarkers (**Figure 2**). The blockade or genetic deletion of CB1r is closely associated with the worsening of emotional behavioral traits, as revealed principally in animal models of anxiety, depression or PTSD, whereas CB1r pharmacological activation induces an improvement effect. On the other hand, CB1r activation induces psychotic symptoms, while CB1r blockade presents antipsychotic effects. Regarding CB2r, its pharmacological activation or its overexpression by means of genetic manipulation or chronic treatment induced upregulation improving anxiety- and depressive-like behaviors, as well as schizophrenia-like traits. In contrast, all these behaviors worsen by CB2r blockade or gene deletion. Interestingly, the strengthening of the endocannabinoid tone, by the inhibition of enzymatic degradation or the blockade of reuptake mechanisms, is closely related with an improvement, particularly in emotional regulation as explored in animal models of anxiety, depression, and PTSD.

Therefore, the available evidence points out that the functional manipulation of the ECS components presents a great therapeutic potential. However, the close interaction of the ECS with other neurotransmitter or neurohormonal systems, as well as the specific and differential neuroanatomical distribution of the ECS components, provides a complex scenario not only from a therapeutic point of view but also considering the occurrence of side effects. In this sense, some aspects should be critically addressed, especially from a pharmacological perspective. The dosing, duration, and mechanism of action involved in the manipulation of the ECS are crucial to reach an improvement and limit adverse reactions.

TABLE 1 | Main findings from human studies supporting the role of CB1r and CB2r as biomarkers in psychiatric disorders.

| CB1r | | | | | |
|---|---|--|---|--|-----------------------------|
| Subjects/Diagnosis | Sample/Intervention | Method | Measurement | Results | References |
| Healthy controls | THC (10 mg) p.o. | [¹¹ C]MePPEP PET | CB1r availability in amygdala | ↑ CB1r | Bhattacharyya et al. (166) |
| Healthy controls | Rimonabant (90 mg) p.o. | Visual Analogue Mood Scale | Anxiety level | ↑ anxiety | Bergamaschi et al. (168) |
| AD/DD | Buccal mucosa cells | DNA Genotyping | CNR1 rs7766029 polymorphism | ↑ frequency financial-related anxiety and depression | Gonda et al. (170) |
| DD | PMBT – anterior cingulate cortex | Immunohistochemistry | Density of CB1r immunopositive glial cells | ↓ CB1r | Koethe et al. (223) |
| DD | PMBT – dorsolateral prefrontal cortex | Quantitative polymerase chain reaction (qPCR) | CB1r relative gene expression | ↑ CB1r | Choi et al. (224) |
| DD | Blood | DNA Genotyping | CNR1 rs1049353 (1359 G/A) polymorphism | ↑ frequency | Monteleone et al. (232) |
| DD/SCZ | Blood | DNA Genotyping | CNR1 rs1049353 (1359 G/A) polymorphism | ↑ depressive symptoms in G-allele carriers | Schennach et al. (233) |
| DD | Blood | DNA Genotyping | CNR1 rs1049353 (1359 G/A) polymorphism | ↑ treatment resistance in G-allele carriers | Domschke et al. (234) |
| DD | Blood | DNA Genotyping | CNR1 rs1049353 (1359 G/A) polymorphism | ↑ citalopram response in GG genotype male carriers | Mitjans et al. (235) |
| Missouri Adolescent Female Twin Study (MOAFTS) participants | Blood | DNA Genotyping | CNR1 rs1049353 (1359 G/A) polymorphism | ↓ risk for anhedonia/DD in A-allele carriers with childhood trauma | Agrawal et al. (236) |
| DD in Parkinson's disease | Blood | DNA Genotyping | CNR1 (AAT)n triplet polymorphism | ↓ risk for DD in 2 long alleles carriers | Barrero et al. (238) |
| DD in methadone-maintained patients | Blood | DNA Genotyping | CNR1 rs2023239 polymorphism | ↓ risk for DD in C-allele carriers | Idick et al. (239) |
| SCZ | PMBT – dorsolateral prefrontal cortex | In situ [³ H]CP-55940 radioligand binding | CB1r binding | ↑ CB1r | Dean et al. (269) |
| SCZ | PMBT – dorsolateral prefrontal cortex | In situ [³ H]MePPEP radioligand binding | CB1r binding | ↑ CB1r | Jenko et al. (270) |
| SCZ | PMBT – prefrontal cortex | In situ [³ H]OMAR radioligand binding | CB1r binding | ↑ CB1r | Volk et al. (271) |
| SCZ | PMBT – anterior cingulate cortex | In situ [³ H]SR141716A radioligand binding | CB1r binding | ↑ CB1r | Zavitsanou et al. (274) |
| SCZ | PMBT – posterior cingulate cortex | In situ [³ H]CP-55,940 radioligand binding | CB1r binding | ↑ CB1r | Newell et al. (275) |
| SCZ | PMBT – posterior cingulate cortex | In situ hybridization | CB1r mRNA | ↓ CB1r | Eggan et al. (277) |
| SCZ | PMBT – prefrontal cortex | Immunohistochemistry | CB1r protein | ↓ CB1r | Urigüen et al. (278) |
| | | Western Blot | CB1r protein | ↓ CB1r in antipsychotic-treated patients | |
| SCZ | PMBT – prefrontal cortex | Quantitative polymerase chain reaction (qPCR) | CB1r relative gene expression | ↓ CB1r | Muguruzaf et al. (273) |
| SCZ | <i>In vivo</i> neuroimaging (several brain regions) | [¹¹ C]-OMAR PET | CB1r binding | ↑ CB1r (only in the pons) | Wong et al. (279) |
| SCZ | <i>In vivo</i> neuroimaging (several brain regions) | [¹⁸ F]-MK-9470 PET | CB1r binding | ↑ CB1r | Ceccarini et al. (280) |
| SCZ | <i>In vivo</i> neuroimaging (several brain regions) | [¹¹ C]-OMAR PET | CB1r binding | ↓ CB1r | Ranganathan et al. (281) |
| FEP | <i>In vivo</i> neuroimaging (several brain regions) | [¹⁸ F]-FMPEP-d2 or [¹¹ C]-MePPEP PET | CB1r binding | ↓ CB1r ↔ severity | Borgan et al. (283) |
| SCZ | Blood | DNA genotyping | CNR1 rs1049353, rs7766029, rs806366 polymorphisms | Nominal association | Costa et al. (290) |
| SCZ | Blood | DNA genotyping | CNR1 (AAT)n triplet polymorphism | 9 and 17 repeat alleles ↔ ↑ susceptibility disorganized SCZ | Ujike et al. (285) |
| SCZ | Blood | DNA genotyping | CNR1 rs1049353 (1359 G/A) polymorphism | ↑ treatment response in G-allele carriers | Hamdani et al. (290) |
| SCZ | Blood | DNA genotyping | CNR1 rs2023239 polymorphism | ↑ better improvement in TT genotype carriers | Suárez-Pinilla et al. (292) |

(Continued)

TABLE 1 | Continued

| CB1r | | | | | |
|---|---|---|--|--|------------------------------|
| Subjects/Diagnosis | Sample/Intervention | Method | Measurement | Results | References |
| FEP | Blood | DNA genotyping | CNR1 rs7766029, rs12720071 polymorphisms | ↑ better improvement in rs7766029 CC genotype or rs12720071 AG genotype | Kuzman, R. et al. (293) |
| SCZ | Blood | DNA genotyping | CNR1 rs6928499, rs1535255, rs2023239 polymorphisms | ↓ risk metabolic syndrome in minor alleles carriers | Yu et al. (294) |
| SCZ | Blood | DNA genotyping | CB1r relative gene expression | ↑ treatment response in G-allele carriers | Schennach et al. (233) |
| SCZ | Blood – PBMCs | Flow cytometry | CNR1 rs1049353 (1359 G/A) polymorphism | ↑ CB1r | De Campos-Carli et al. (297) |
| SCZ | Blood - PBMCs | Quantitative polymerase chain reaction (qPCR) | CB1r expression | ↑ CB1r | Chase et al. (298) |
| BD | Blood | DNA genotyping | CNR1 rs6454674 polymorphism | ↑ severity in T/G heterozygotes | Alpak et al. (314) |
| Detroit Neighborhood Health Study (DNHS) participants | Blood | DNA genotyping | CNR1 rs1049353 polymorphism | ↑ risk for PTSD-related symptoms in A-allele carriers with childhood abuse | Mota et al. (339) |
| PTSD | <i>In vivo</i> neuroimaging (several brain regions) | [¹¹ C]-OMAR PET | CB1r binding | ↑ CB1r | Neumeister, A. et al. (342) |
| ADHD | Blood | DNA genotyping | SNP variants at the CNR1 gene | ↑ frequency SNP haplotype (C-G) | Lu et al. (340) |
| ADHD (alcoholic patients) | Blood | DNA genotyping | CNR1 (AAT)n triplet polymorphism | ↑ frequency longer form of alleles | Ponce et al. (358) |
| AN/BN | <i>In vivo</i> neuroimaging (several brain regions) | [¹⁸ F]-MK-9470 PET | CB1r binding | ↑ CB1r | Gérard et al. (371) |
| AN/BN | Blood | Quantitative polymerase chain reaction (qPCR) | CB1r relative gene expression | ↓ CB1r in AN/BN women with self-injurious behavior | Schroeder et al. (372) |
| CB2r | | | | | |
| Subjects/Diagnosis | Sample | Method | Measurement | Results | References |
| General population | Buccal mucosa | DNA genotyping | CNR2 rs2501432 (R63Q) polymorphism | ↑ risk for AD/DD in rs2501432 R-allele carriers with childhood trauma | Lazary et al. (176) |
| DD | Blood | DNA genotyping | CNR2 rs2501431 polymorphism | ↑ depressive symptoms in G-allele carriers | Mitjans et al. (235) |
| DD | Blood | DNA genotyping | CNR2 rs2501432 (R63Q) polymorphism | ↑ frequency R63Q polymorphism | Onaivi et al. (249, 241) |
| SCZ | Blood – PBMCs | Quantitative polymerase chain reaction (qPCR) | CB2r relative gene expression | ↓ CB2r with clinical remission | De Marchi et al. (295) |
| SCZ | Blood – PBMCs | Quantitative polymerase chain reaction (qPCR) | CB2r relative gene expression | ↑ CB2r | Chase et al. (298) |
| SCZ | Blood – PBMCs | Flow cytometry | CB2r expression | ↑ CB2r | De Campos-Carli et al. (297) |
| SCZ | Blood – PBMCs | Western Blot | CB2r expression | ↓ CB2r | Bioque et al. (310) |
| SCZ | Blood | DNA genotyping | CNR2 rs2501432 (R63Q), rs12744386 polymorphism | ↑ frequency in rs2501432 R63 and rs12744386 C alleles | Ishiguro et al. (265) |
| SCZ | Blood | DNA genotyping | CNR2 rs2501432C/T polymorphism | ↑ risk for SCZ in T-allele carriers | Tong et al. (311) |
| BD | Blood | DNA genotyping | CNR2 rs41311993 (524C/A) polymorphism | ↑ frequency 524C/A polymorphism | Minocci et al. (317) |

AD, anxiety disorder; DD, depressive disorder; SCZ, schizophrenia; FEP, first episode psychosis; BD, bipolar disorder; PTSD, post-traumatic stress disorder; ADHD, attention-deficit hyperactivity disorder; AN, anorexia nervosa; BN, bulimia nervosa; PMBT, post-mortem brain tissue; PET, positron emission tomography; PBMCs, peripheral blood mononuclear cells. ↓: decrease and ↑: increase.

TABLE 2 | Main findings from human studies supporting the role of AEA and 2-AG as biomarkers in psychiatric disorders.

| AEA & 2-AG | | | | | |
|------------------------|------------------------|---|-----------------------------|--|-----------------------------|
| Subjects/ Diagnosis | Sample | Method | Measurement | Results | References |
| DD | Serum | Chemical ionization liquid chromatography-mass spectrometry (LC-APCI-MS) | AEA and 2-AG quantification | ↑ AEA in minor depression ↓ 2-AG in major depression | Hill et al. (226) |
| DD | Serum | Chemical ionization liquid chromatography-mass spectrometry (LC-APCI-MS) | AEA and 2-AG quantification | ↓ AEA ↓ 2-AG | Hill et al. (227) |
| DD | Plasma | Chromatography-coupled tandem mass spectrometry system | AEA and 2-AG quantification | ↑ AEA | Romero-Sanchiz et al. (228) |
| DD | Serum | Electrospray ionization liquid chromatography-mass spectrometry (LC-ESI-MS-MS) | AEA and 2-AG quantification | ↑ AEA = 2-AG after moderate exercise | Meyer et al. (230) |
| DD | CSF | Liquid chromatography-multiple reaction monitoring (LC/MRM) | AEA and 2-AG quantification | ↑ AEA ↑ 2-AG after ECT | Kranaster et al. (231) |
| SCZ | CSF | High pressure liquid chromatography-gas chromatography/mass spectrometry (HPLC-GC/MS) | AEA quantification | ↑ AEA | Leweke et al. (300) |
| SCZ | CSF | High pressure liquid chromatography-mass spectrometry (HPLC-MS) | AEA quantification | ↑ AEA in antipsychotics-naïve patients = AEA in typical antipsychotics-treated patients | Giuffrida et al. (301) |
| SCZ | Blood | Chemical ionization liquid chromatography-mass spectrometry (LC-APCI-MS) | AEA quantification | ↑ AEA ↓ AEA with clinical remission | De Marchi et al. (295) |
| SCZ | Plasma | Liquid chromatography-mass spectrometry (LC-MS) | AEA quantification | ↑ AEA | Koethe, D. et al. (303) |
| SCZ | PMBT (several regions) | Liquid chromatography coupled with triple quadrupole mass spectrometry (LC/MS/MS) | AEA and 2-AG quantification | ↓ AEA ↑ 2-AG | Muguruza et al. (306) |
| PTSD | Plasma | Chemical ionization liquid chromatography-mass spectrometry (LC-APCI-MS) | AEA and 2-AG quantification | = AEA ↓ 2-AG | Hill et al. (341) |
| AN | Plasma | Chemical ionization liquid chromatography-mass spectrometry (LC-APCI-MS) | AEA and 2-AG quantification | ↑ AEA = 2-AG | Monteleone et al. (370) |

DD, depressive disorder; SCZ, schizophrenia; PTSD, post-traumatic stress disorder; AN, anorexia nervosa; CSF, cerebrospinal fluid; PMBT, post-mortem brain tissue; AEA, anandamide; 2-AG, 2-arachidonylglycerol.

↓: decrease and ↑: increase.

TABLE 3 | Main findings from human studies supporting the role of FAAH as a biomarker in psychiatric disorders.

| FAAH | | | | | |
|---|--------------------------|---|---|--|-------------------------|
| Subjects/Diagnosis | Sample | Method | Measurement | Results | References |
| Healthy controls | Blood | DNA Genotyping | FAAH rs324420 (C385A) polymorphism | = anxiety-related self-reports in A-allele and C/C genotypes | Gärtner et al. (171) |
| PING study participants | Saliva | DNA Genotyping | FAAH rs324420 (C385A) polymorphism | ↓ anxiety level in A-allele adolescent carriers | Gee et al. (172) |
| Project FRONTIER participants | Blood | DNA Genotyping | FAAH rs324420 (C385A) & CRFR1 minor alleles polymorphisms | ↑ anxiety level in FAAH A-allele and CRFR1 non-minors alleles carriers | Harris et al. (173) |
| Duke Neurogenetics Study (DNS) participants | Saliva | DNA Genotyping | FAAH rs324420 (C385A) and CRHR1 rs110402 polymorphisms | ↑ risk for AD in FAAH A-allele and CRHR1 A-allele carriers | Demers et al. (174) |
| General population | Buccal mucosa | DNA Genotyping | FAAH rs324420 (C385A) polymorphism | ↑ risk for AD/DD in A-allele carriers with childhood trauma | Lazary et al. (175) |
| General population | Buccal mucosa | DNA Genotyping | FAAH rs324420 (C385A) polymorphism | ↑ risk for AD/DD in A-allele carriers with childhood trauma | Lazary et al. (176) |
| DD/BD | Blood | DNA Genotyping | FAAH rs324420 (C385A) polymorphism | ↑ frequency AC genotype carriers | Monteleone et al. (232) |
| SCZ | PMBT – prefrontal cortex | Enzymatic assay – scintillation counting | FAAH activity | ↑ FAAH activity | Muguruza et al. (273) |
| SCZ | Blood | Quantitative polymerase chain reaction (qPCR) | FAAH relative gene expression | ↓ FAAH with clinical remission | De Marchi et al. (295) |
| BD | Blood | DNA Genotyping | FAAH rs324420 (C385A) polymorphism | ↑ frequency AC genotype carriers | Monteleone et al. (232) |

AD, anxiety disorder; DD, depressive disorder; SCZ, schizophrenia; BD, bipolar disorder; PMBT, post-mortem brain tissue; FAAH, fatty acid amido hydrolase.

↓: decrease and ↑: increase.

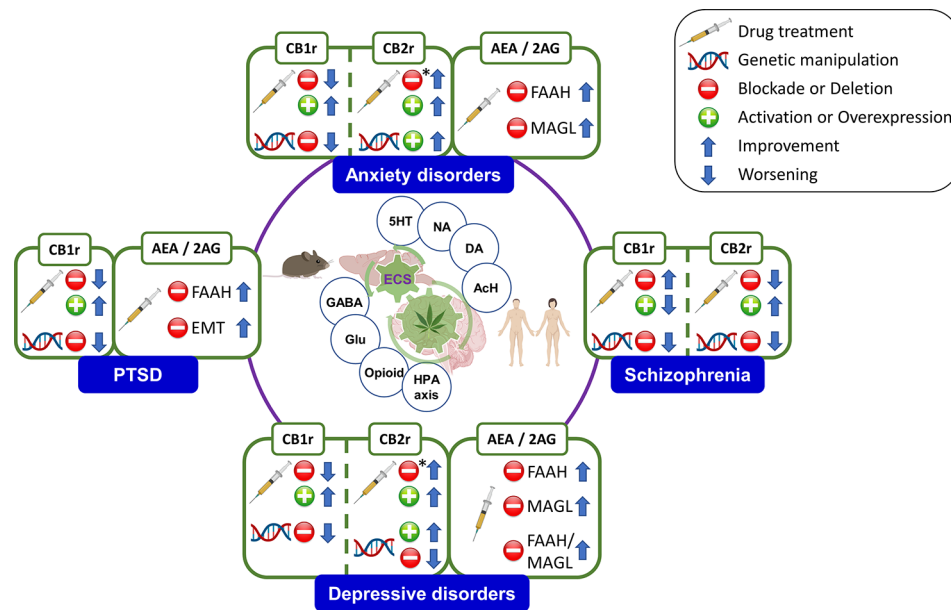


FIGURE 2 | Main findings regarding the therapeutic potential of the functional manipulation of the endocannabinoid system (ECS) components by pharmacological and genetic approaches in anxiety, depression, schizophrenia, and post-traumatic stress disorder (PTSD). CB1r, cannabinoid receptor 1; CB2r, cannabinoid receptor 2; AEA, anandamide; 2-AG, 2-arachidonoylglycerol; FAAH, fatty acid amide hydrolase; MAGL, monoacylglycerol lipase; 5-HT, serotonin; NA, noradrenaline; DA, dopamine; AcH, acetylcholine; GABA, gamma-aminobutyric acid; Glu, glutamate; HPA axis, Hypothalamus–Pituitary–Adrenal axis; *, chronic treatment.

Apart from the widely explored role of CB1r, it should be noted that in recent years an increasing emphasis is being placed on the design of strategies to regulate endogenous cannabinoid tone, through inhibitors of the degradation or reuptake of eCBs. Since this approach provided negative results, particularly regarding the inhibition of FAAH (373), current trends focused on the combination of different mechanisms of action to enhance the endocannabinoid tone (374). Moreover, the pharmacological modulation of CB2r has also attracted much attention given its safety profile and the wide range of properties attributed to it, including mood and cognitive regulation. Thus, future priorities for both human and animal research would be the potentiation of both endocannabinoid tone and CB2r-mediated actions.

The trials carried out on patients show alterations of the ECS components at different levels, which in certain cases are related to risk or predictive factors regarding the evolution of the disease, or the degree of response to drug treatment. It is relevant to highlight the underlying sex-dependent effects in terms of sexual dimorphism of the ECS (375) and sex differences in prevalence rates and presentation of the psychiatric disorders (376). These could be involved not only in the changes of the ECS components to provide sex-related diagnostic or prognostic biomarkers, but also in the pharmacological actions derived from the treatment with cannabinoid compounds (377). Finally, it is important to note that more *in vivo* clinical studies are recently being carried out employing blood samples (PBMCs, plasma) or neuroimaging techniques (PET, fMRI) to identify ECS-related alterations, providing very relevant data. In this sense, a higher effort is

required to design and perform more clinical studies, especially increasing the sample sizes to achieve greater significance, representativeness and reproducibility, finally making possible to identify some of the ECS components as useful biomarkers applicable to clinical practice in psychiatry.

AUTHOR CONTRIBUTIONS

FN and JM designed the sections and contents of the review manuscript. FN oversaw the organization to distribute the writing tasks among the authors and participated in manuscript writing. MG-G, RJ-B, GR, AG, and AA-O perform the literature searches and participated in the manuscript writing. All the authors critically reviewed and approved the final version of the manuscript.

ACKNOWLEDGMENTS

The preparation of the manuscript was supported by ‘Instituto de Salud Carlos III, Fondos FEDER, Red de Trastornos Adictivos’ (RTA, RD16/0017/0014 to J.M. and RD16/0017/0017 to G.R.) and ‘Ministerio de Sanidad, Delegación del Gobierno para el Plan Nacional Sobre Drogas’ (PNSD, 2019/012 to J.M.).

REFERENCES

- G.B.D. Disease, I. Injury, and C. Prevalence. Global, regional, and national incidence, prevalence, and years lived with disability for 354 diseases and injuries for 195 countries and territories, 1990–2017: a systematic analysis for the Global Burden of Disease Study 2017. *Lancet* (2018) 392:1789–858. doi: 10.1016/S0140-6736(18)32279-7
- Vigo D, Thornicroft G, Atun R. Estimating the true global burden of mental illness. *Lancet Psychiatry* (2016) 3:171–8. doi: 10.1016/S2215-0366(15)00505-2
- Krystal JH, State MW. Psychiatric disorders: diagnosis to therapy. *Cell* (2014) 157:201–14. doi: 10.1016/j.cell.2014.02.042
- Scarr E, Millan MJ, Bahn S, Bertolino A, Turck CW, Kapur S, et al. Biomarkers for Psychiatry: The Journey from Fantasy to Fact, a Report of the 2013 CINP Think Tank. *Int J Neuropsychopharmacol* (2015) 18:pyv042. doi: 10.1093/ijnp/pyv042
- Venkatasubramanian G, Keshavan MS. Biomarkers in Psychiatry - A Critique. *Ann Neurosci* (2016) 23:3–5. doi: 10.1159/000443549
- Mechoulam R, Parker LA. The endocannabinoid system and the brain. *Annu Rev Psychol* (2013) 64:21–47. doi: 10.1146/annurev-psych-113011-143739
- Ashton JC, Dowie MJ, Glass M. *The endocannabinoid system and human brain functions: insight from memory, motor, and mood pathologies, The endocannabinoid system: genetics, biochemistry, brain disorders, and therapy*. Cambridge (Massachusetts), USA (2017) p. 115–86.
- Manzanares J, Uriguen L, Rubio G, Palomo T. Role of endocannabinoid system in mental diseases. *Neurotox Res* (2004) 6:213–24. doi: 10.1007/BF03033223
- Marco EM, Garcia-Gutierrez MS, Bermudez-Silva FJ, Moreira FA, Guimaraes F, Manzanares J, et al. Endocannabinoid system and psychiatry: in search of a neurobiological basis for detrimental and potential therapeutic effects. *Front Behav Neurosci* (2011) 5:63. doi: 10.3389/fnbeh.2011.00063
- Parolaro D, Realini N, Vigano D, Guidali C, Rubino T. The endocannabinoid system and psychiatric disorders. *Exp Neurol* (2010) 224:3–14. doi: 10.1016/j.expneurol.2010.03.018
- Katzman MA, Furtado M, Anand L. Targeting the Endocannabinoid System in Psychiatric Illness. *J Clin Psychopharmacol* (2016) 36:691–703. doi: 10.1097/JCP.0000000000000581
- Ritchie H, Roser M. (2020). Published online at OurWorldInData.org. Retrieved from: <https://ourworldindata.org/mental-health> Online Resource.
- Bromet E, Karam E, Koenen K, Stein D. *The Global Epidemiology of Trauma Exposure and Posttraumatic Stress Disorder*. Cambridge: Cambridge University Press (2018).
- Fayyad J, Sampson NA, Hwang I, Adamowski T, Aguilar-Gaxiola S, Al-Hamzawi A, et al. The descriptive epidemiology of DSM-IV Adult ADHD in the World Health Organization World Mental Health Surveys. *Atten Defic Hyperact Disord* (2017) 9:47–65. doi: 10.1007/s12402-016-0208-3
- Katona I, Freund TF. Multiple functions of endocannabinoid signaling in the brain. *Annu Rev Neurosci* (2012) 35:529–58. doi: 10.1146/annurev-neuro-062111-150420
- Atkinson DL, Abbott JK. *Cannabinoids and the brain: the effects of endogenous and exogenous cannabinoids on brain systems and function, The complex connection between cannabis and schizophrenia*. Cambridge (Massachusetts), USA (2018) p. 37–74.
- Piomelli D. The molecular logic of endocannabinoid signalling. *Nat Rev Neurosci* (2003) 4:873–84. doi: 10.1038/nrn1247
- Zou S, Kumar U. Cannabinoid Receptors and the Endocannabinoid System: Signaling and Function in the Central Nervous System. *Int J Mol Sci* (2018) 19: 833. doi: 10.3390/ijms19030833
- Mackie K. Distribution of cannabinoid receptors in the central and peripheral nervous system. *Handb Exp Pharmacol* (2005), 168: 299–325. doi: 10.1007/3-540-26573-2_10
- Rodriguez de Fonseca F, Del Arco I, Bermudez-Silva FJ, Bilbao A, Cippitelli A, Navarro M. The endocannabinoid system: physiology and pharmacology. *Alcohol Alcohol* (2005) 40:2–14. doi: 10.1093/alcal/agh110
- Tsou K, Brown S, Sanudo-Pena MC, Mackie K, Walker JM. Immunohistochemical distribution of cannabinoid CB1 receptors in the rat central nervous system. *Neuroscience* (1998) 83:393–411. doi: 10.1016/S0306-4522(97)00436-3
- Matsuda LA, Lolait SJ, Brownstein MJ, Young AC, Bonner TI. Structure of a cannabinoid receptor and functional expression of the cloned cDNA. *Nature* (1990) 346:561–4. doi: 10.1038/346561a0
- Gutierrez-Rodriguez A, Puente N, Elezgarai I, Ruehle S, Lutz B, Reguero L, et al. Anatomical characterization of the cannabinoid CB1 receptor in cell-type-specific mutant mouse rescue models. *J Comp Neurol* (2017) 525:302–18. doi: 10.1002/cne.24066
- Piazza PV, Cota D, Marsicano G. The CB1 Receptor as the Cornerstone of Exostasis. *Neuron* (2017) 93:1252–74. doi: 10.1016/j.neuron.2017.02.002
- Busquets-Garcia A, Bains J, Marsicano G. CB1 Receptor Signaling in the Brain: Extracting Specificity from Ubiquity. *Neuropsychopharmacology* (2018) 43:4–20. doi: 10.1038/npp.2017.206
- Howlett AC, Bidaut-Russell M, Devane WA, Melvin LS, Johnson MR, Herkenham M. The cannabinoid receptor: biochemical, anatomical and behavioral characterization. *Trends Neurosci* (1990) 13:420–3. doi: 10.1016/0166-2236(90)90124-S
- Hu SS, Mackie K. Distribution of the Endocannabinoid System in the Central Nervous System. *Handb Exp Pharmacol* (2015) 231:59–93. doi: 10.1007/978-3-319-20825-1_3
- Batkai S, Jarai Z, Wagner JA, Goparaju SK, Varga K, Liu J, et al. Endocannabinoids acting at vascular CB1 receptors mediate the vasodilated state in advanced liver cirrhosis. *Nat Med* (2001) 7:827–32. doi: 10.1038/89953
- Wagner JA, Varga K, Ellis EF, Rzigalinski BA, Martin BR, Kunos G. Activation of peripheral CB1 cannabinoid receptors in haemorrhagic shock. *Nature* (1997) 390:518–21. doi: 10.1038/37371
- Pertwee RG. Pharmacology of cannabinoid CB1 and CB2 receptors. *Pharmacol Ther* (1997) 74:129–80. doi: 10.1016/S0163-7258(97)82001-3
- Pertwee RG, Ross RA. Cannabinoid receptors and their ligands. *Prostaglandins Leukot Essent Fatty Acids* (2002) 66:101–21. doi: 10.1054/plf.2001.0341
- Galiegue S, Mary S, Marchand J, Dussosoy D, Carriere D, Carayon P, et al. Expression of central and peripheral cannabinoid receptors in human immune tissues and leukocyte subpopulations. *Eur J Biochem* (1995) 232:54–61. doi: 10.1111/j.1432-1033.1995.tb20780.x
- Guindon J, Hohmann AG. The endocannabinoid system and pain. *CNS Neurol Disord Drug Targets* (2009) 8:403–21. doi: 10.2174/187152709789824660
- Munro S, Thomas KL, Abu-Shaar M. Molecular characterization of a peripheral receptor for cannabinoids. *Nature* (1993) 365:61–5. doi: 10.1038/365061a0
- Cabral GA, Griffin-Thomas L. Emerging role of the cannabinoid receptor CB2 in immune regulation: therapeutic prospects for neuroinflammation. *Expert Rev Mol Med* (2009) 11:e3. doi: 10.1017/S1462399409000957
- Benito C, Nunez E, Tolon RM, Carrier EJ, Rabano A, Hillard CJ, et al. Cannabinoid CB2 receptors and fatty acid amide hydrolase are selectively overexpressed in neuritic plaque-associated glia in Alzheimer's disease brains. *J Neurosci* (2003) 23:11136–41. doi: 10.1523/JNEUROSCI.23-35-11136.2003
- Yiangou Y, Facer P, Durrenberger P, Chessell IP, Naylor A, Bountra C, et al. COX-2, CB2 and P2X7-immunoreactivities are increased in activated microglial cells/macrophages of multiple sclerosis and amyotrophic lateral sclerosis spinal cord. *BMC Neurol* (2006) 6:12. doi: 10.1186/1471-2377-6-12
- Guzman M, Sanchez C, Galve-Roperh I. Control of the cell survival/death decision by cannabinoids. *J Mol Med (Berl)* (2001) 78:613–25. doi: 10.1007/s001090000177
- Van Sickle MD, Duncan M, Kingsley PJ, Mouihate A, Urbani P, Mackie K, et al. Identification and functional characterization of brainstem cannabinoid CB2 receptors. *Science* (2005) 310:329–32. doi: 10.1126/science.1115740
- Gong JP, Onaivi ES, Ishiguro H, Liu QR, Tagliaferro PA, Brusco A, et al. Cannabinoid CB2 receptors: immunohistochemical localization in rat brain. *Brain Res* (2006) 1071:10–23. doi: 10.1016/j.brainres.2005.11.035
- Onaivi ES. Neuropsychobiological evidence for the functional presence and expression of cannabinoid CB2 receptors in the brain. *Neuropsychobiology* (2006) 54:231–46. doi: 10.1159/000100778

42. Onaivi ES, Ishiguro H, Gong JP, Patel S, Perchuk A, Meozzi PA, et al. Discovery of the presence and functional expression of cannabinoid CB2 receptors in brain. *Ann N Y Acad Sci* (2006) 1074:514–36. doi: 10.1196/annals.1369.052
43. Garcia-Gutierrez MS, Perez-Ortiz JM, Gutierrez-Adan A, Manzanares J. Depression-resistant endophenotype in mice overexpressing cannabinoid CB(2) receptors. *Br J Pharmacol* (2010) 160:1773–84. doi: 10.1111/j.1476-5381.2010.00819.x
44. Zhang HY, Gao M, Liu QR, Bi GH, Li X, Yang HJ, et al. Cannabinoid CB2 receptors modulate midbrain dopamine neuronal activity and dopamine-related behavior in mice. *Proc Natl Acad Sci U S A* (2014) 111:E5007–15. doi: 10.1073/pnas.1413210111
45. Cabral GA, Raborn ES, Griffin L, Dennis J, Marciano-Cabral F. CB2 receptors in the brain: role in central immune function. *Br J Pharmacol* (2008) 153:240–51. doi: 10.1038/sj.bjp.0707584
46. Garcia-Gutierrez MS, Navarrete F, Navarro G, Reyes-Resina I, Franco R, Lanciego JL, et al. Alterations in Gene and Protein Expression of Cannabinoid CB2 and GPR55 Receptors in the Dorsolateral Prefrontal Cortex of Suicide Victims. *Neurotherapeutics* (2018) 15:796–806. doi: 10.1007/s13311-018-0610-y
47. Liu QR, Canseco-Alba A, Zhang HY, Tagliaferro P, Chung M, Dennis E, et al. Cannabinoid type 2 receptors in dopamine neurons inhibits psychomotor behaviors, alters anxiety, depression and alcohol preference. *Sci Rep* (2017) 7:17410. doi: 10.1038/s41598-017-17796-y
48. Pertwee RG, Howlett AC, Abood ME, Alexander SP, Di Marzo V, Elphick MR, et al. Cannabinoid receptors and their ligands: beyond CB(1) and CB (2). *Pharmacol Rev* (2010) 62:588–631. doi: 10.1124/pr.110.003004
49. Kano M, Ohno-Shosaku T, Hashimoto-dani Y, Uchigashima M, Watanabe M. Endocannabinoid-mediated control of synaptic transmission. *Physiol Rev* (2009) 89:309–80. doi: 10.1152/physrev.00019.2008
50. Alger BE, Kim J. Supply and demand for endocannabinoids. *Trends Neurosci* (2011) 34:304–15. doi: 10.1016/j.tins.2011.03.003
51. Devane WA, Hanus L, Breuer A, Pertwee RG, Stevenson LA, Griffin G, et al. Isolation and structure of a brain constituent that binds to the cannabinoid receptor. *Science* (1992) 258:1946–9. doi: 10.1126/science.1470919
52. Mechoulam R, Ben-Shabat S, Hanus L, Ligumsky M, Kaminski NE, Schatz AR, et al. Identification of an endogenous 2-monoglyceride, present in canine gut, that binds to cannabinoid receptors. *Biochem Pharmacol* (1995) 50:83–90. doi: 10.1016/0006-2952(95)00109-D
53. Okamoto Y, Morishita J, Tsuboi K, Tonai T, Ueda N. Molecular characterization of a phospholipase D generating anandamide and its congeners. *J Biol Chem* (2004) 279:5298–305. doi: 10.1074/jbc.M306642200
54. Di Marzo V, Stella N, Zimmer A. Endocannabinoid signalling and the deteriorating brain. *Nat Rev Neurosci* (2015) 16:30–42. doi: 10.1038/nrn3876
55. Ueda N. Endocannabinoid hydrolases. *Prostaglandins Other Lipid Mediat* (2002) 68-69:521–34. doi: 10.1016/S0090-6980(02)00053-9
56. Egertova M, Cravatt BF, Elphick MR. Comparative analysis of fatty acid amide hydrolase and cb(1) cannabinoid receptor expression in the mouse brain: evidence of a widespread role for fatty acid amide hydrolase in regulation of endocannabinoid signaling. *Neuroscience* (2003) 119:481–96. doi: 10.1016/S0306-4522(03)00145-3
57. Fezza F, Bari M, Florio R, Talamonti E, Feole M, Maccarrone M. Endocannabinoids, related compounds and their metabolic routes. *Molecules* (2014) 19:17078–106. doi: 10.3390/molecules191117078
58. Gao Y, Vasilyev DV, Goncalves MB, Howell FV, Hobbs C, Reisenberg M, et al. Loss of retrograde endocannabinoid signaling and reduced adult neurogenesis in diacylglycerol lipase knock-out mice. *J Neurosci* (2010) 30:2017–24. doi: 10.1523/JNEUROSCI.5693-09.2010
59. Tanimura A, Yamazaki M, Hashimoto-dani Y, Uchigashima M, Kawata S, Abe M, et al. The endocannabinoid 2-arachidonoylglycerol produced by diacylglycerol lipase alpha mediates retrograde suppression of synaptic transmission. *Neuron* (2010) 65:320–7. doi: 10.1016/j.neuron.2010.01.021
60. Dinh TP, Carpenter D, Leslie FM, Freund TF, Katona I, Sensi SL, et al. Brain monoglyceride lipase participating in endocannabinoid inactivation. *Proc Natl Acad Sci U S A* (2002) 99:10819–24. doi: 10.1073/pnas.152334899
61. Marsicano G, Lutz B. Neuromodulatory functions of the endocannabinoid system. *J Endocrinol Invest* (2006) 29:27–46.
62. Ibarra-Lecue I, Pilar-Cuellar F, Muguruza C, Florensa-Zanuy E, Diaz A, Uriguen L, et al. The endocannabinoid system in mental disorders: Evidence from human brain studies. *Biochem Pharmacol American Psychiatric Association* (2018) 157:97–107. doi: 10.1016/j.bcp.2018.07.009
63. American Psychiatric Association. (2013). *Diagnostic and Statistical Manual of Mental Disorders (DSM), 5th edition*.
64. Marco EM, Echeverry-Alzate V, Lopez-Moreno JA, Gine E, Penasco S, Viveros MP. Consequences of early life stress on the expression of endocannabinoid-related genes in the rat brain. *Behav Pharmacol* (2014) 25:547–56. doi: 10.1097/FBP.0000000000000068
65. Boero G, Pisu MG, Biggio F, Muredda L, Carta G, Banni S, et al. Impaired glucocorticoid-mediated HPA axis negative feedback induced by juvenile social isolation in male rats. *Neuropharmacology* (2018) 133:242–53. doi: 10.1016/j.neuropharm.2018.01.045
66. Sutt S, Raud S, Areda T, Reimets A, Koks S, Vasar E. Cat odour-induced anxiety—a study of the involvement of the endocannabinoid system. *Psychopharmacol (Berl)* (2008) 198:509–20. doi: 10.1007/s00213-007-0927-4
67. Cagni P, Barros M. Cannabinoid type 1 receptor ligands WIN 55,212-2 and AM 251 alter anxiety-like behaviors of marmoset monkeys in an open-field test. *Behav Brain Res* (2013) 240:91–4. doi: 10.1016/j.bbr.2012.11.018
68. Morena M, Patel S, Bains JS, Hill MN. Neurobiological Interactions Between Stress and the Endocannabinoid System. *Neuropsychopharmacology* (2016) 41:80–102. doi: 10.1038/npp.2015.166
69. Lutz B, Marsicano G, Maldonado R, Hillard CJ. The endocannabinoid system in guarding against fear, anxiety and stress. *Nat Rev Neurosci* (2015) 16:705–18. doi: 10.1038/nrn4036
70. Haller J, Bakos N, Szirmay M, Ledent C, Freund TF. The effects of genetic and pharmacological blockade of the CB1 cannabinoid receptor on anxiety. *Eur J Neurosci* (2002) 16:1395–8. doi: 10.1046/j.1460-9568.2002.02192.x
71. Haller J, Varga B, Ledent C, Freund TF. CB1 cannabinoid receptors mediate anxiolytic effects: convergent genetic and pharmacological evidence with CB1-specific agents. *Behav Pharmacol* (2004) 15:299–304. doi: 10.1097/01.fbp.0000135704.56422.40
72. Haller J, Varga B, Ledent C, Barna I, Freund TF. Context-dependent effects of CB1 cannabinoid gene disruption on anxiety-like and social behaviour in mice. *Eur J Neurosci* (2004) 19:1906–12. doi: 10.1111/j.1460-9568.2004.03293.x
73. Witkin JM, Tzavara ET, Nomikos GG. A role for cannabinoid CB1 receptors in mood and anxiety disorders. *Behav Pharmacol* (2005) 16:315–31. doi: 10.1097/00008877-200509000-00005
74. Valverde O. Participation of the cannabinoid system in the regulation of emotional-like behaviour. *Curr Pharm Des* (2005) 11:3421–9. doi: 10.2174/138161205774370780
75. Garcia-Gutierrez MS, Manzanares J. The cannabinoid CB1 receptor is involved in the anxiolytic, sedative and amnesic actions of benzodiazepines. *J Psychopharmacol* (2010) 24:757–65. doi: 10.1177/0269881109106910
76. Batista LA, Moreira FA. Cannabinoid CB1 receptors mediate the anxiolytic effects induced by systemic alprazolam and intra-periaqueductal gray 5-HT1A receptor activation. *Neurosci Lett* (2019) 703:5–10. doi: 10.1016/j.neulet.2019.03.010
77. Naderi N, Haghighparast A, Saber-Tehrani A, Rezaei N, Alizadeh AM, Khani A, et al. Interaction between cannabinoid compounds and diazepam on anxiety-like behaviour of mice. *Pharmacol Biochem Behav* (2008) 89:64–75. doi: 10.1016/j.pbb.2007.11.001
78. Viveros MP, Marco EM, File SE. Endocannabinoid system and stress and anxiety responses. *Pharmacol Biochem Behav* (2005) 81:331–42. doi: 10.1016/j.pbb.2005.01.029
79. Komaki A, Hashemi-Firouzi N, Shojaei S, Sourai Z, Heidari S, Shahidi S. Study the Effect of Endocannabinoid System on Rat Behavior in Elevated Plus-Maze. *Basic Clin Neurosci* (2015) 6:147–53.
80. Lisboa SF, Niraula A, Resstel LB, Guimaraes FS, Godbout JP, Sheridan JF. Repeated social defeat-induced neuroinflammation, anxiety-like behavior and resistance to fear extinction were attenuated by the cannabinoid receptor agonist WIN55,212-2. *Neuropsychopharmacology* (2018) 43:1924–33. doi: 10.1038/s41386-018-0064-2
81. Arevalo C, de Miguel R, Hernandez-Tristan R. Cannabinoid effects on anxiety-related behaviours and hypothalamic neurotransmitters. *Pharmacol Biochem Behav* (2001) 70:123–31. doi: 10.1016/S0091-3057(01)00578-0

82. Navarro M, Hernandez E, Munoz RM, del Arco I, Villanua MA, Carrera MR, et al. Acute administration of the CB1 cannabinoid receptor antagonist SR 141716A induces anxiety-like responses in the rat. *Neuroreport* (1997) 8:491–6. doi: 10.1097/0001756-199701200-00023
83. Degroot A, Nomikos GG. Genetic deletion and pharmacological blockade of CB1 receptors modulates anxiety in the shock-probe burying test. *Eur J Neurosci* (2004) 20:1059–64. doi: 10.1111/j.1460-9568.2004.03556.x
84. Genn RF, Tucci S, Marco EM, Viveros MP, File SE. Unconditioned and conditioned anxiogenic effects of the cannabinoid receptor agonist CP 55,940 in the social interaction test. *Pharmacol Biochem Behav* (2004) 77:567–73. doi: 10.1016/j.pbb.2003.12.019
85. Hill MN, Gorzalka BB. Enhancement of anxiety-like responsiveness to the cannabinoid CB(1) receptor agonist HU-210 following chronic stress. *Eur J Pharmacol* (2004) 499:291–5. doi: 10.1016/j.ejphar.2004.06.069
86. Rodgers RJ, Evans PM, Murphy A. Anxiogenic profile of AM-251, a selective cannabinoid CB1 receptor antagonist, in plus-maze-naïve and plus-maze-experienced mice. *Behav Pharmacol* (2005) 16:405–13. doi: 10.1097/00008877-200509000-00013
87. Sink KS, Segovia KN, Sink J, Randall PA, Collins LE, Correa M, et al. Potential anxiogenic effects of cannabinoid CB1 receptor antagonists/inverse agonists in rats: comparisons between AM4113, AM251, and the benzodiazepine inverse agonist FG-7142. *Eur Neuropsychopharmacol* (2010) 20:112–22. doi: 10.1016/j.euroneuro.2009.11.002
88. Rey AA, Purrio M, Viveros MP, Lutz B. Biphasic effects of cannabinoids in anxiety responses: CB1 and GABA(B) receptors in the balance of GABAergic and glutamatergic neurotransmission. *Neuropsychopharmacology* (2012) 37:2624–34. doi: 10.1038/npp.2012.123
89. Faraji N, Komaki A, Salehi I. Interaction Between the Cannabinoid and Vanilloid Systems on Anxiety in Male Rats. *Basic Clin Neurosci* (2017) 8:129–37. doi: 10.18869/nirp.bcn.8.2.129
90. Campos AC, Guimaraes FS. Evidence for a potential role for TRPV1 receptors in the dorsolateral periaqueductal gray in the attenuation of the anxiolytic effects of cannabinoids. *Prog Neuropsychopharmacol Biol Psychiatry* (2009) 33:1517–21. doi: 10.1016/j.pnpbp.2009.08.017
91. Fogaca MV, Aguiar DC, Moreira FA, Guimaraes FS. The endocannabinoid and endovanilloid systems interact in the rat prelimbic medial prefrontal cortex to control anxiety-like behavior. *Neuropharmacology* (2012) 63:202–10. doi: 10.1016/j.neuropharm.2012.03.007
92. Batista PA, Fogaca MV, Guimaraes FS. The endocannabinoid, endovanilloid and nitergic systems could interact in the rat dorsolateral periaqueductal gray matter to control anxiety-like behaviors. *Behav Brain Res* (2015) 293:182–8. doi: 10.1016/j.bbr.2015.07.019
93. Litvin Y, Phan A, Hill MN, Pfaff DW, McEwen BS. CB1 receptor signaling regulates social anxiety and memory. *Genes Brain Behav* (2013) 12:479–89. doi: 10.1111/gbb.12045
94. Martin M, Ledent C, Parmentier M, Maldonado R, Valverde O. Involvement of CB1 cannabinoid receptors in emotional behaviour. *Psychopharmacol (Berl)* (2002) 159:379–87. doi: 10.1007/s00213-001-0946-5
95. Thiemann G, Watt CA, Ledent C, Molleman A, Hasenohr RU. Modulation of anxiety by acute blockade and genetic deletion of the CB(1) cannabinoid receptor in mice together with biogenic amine changes in the forebrain. *Behav Brain Res* (2009) 200:60–7. doi: 10.1016/j.bbr.2008.12.035
96. Maccarrone M, Valverde O, Barbaccia ML, Castane A, Maldonado R, Ledent C, et al. Age-related changes of anandamide metabolism in CB1 cannabinoid receptor knockout mice: correlation with behaviour. *Eur J Neurosci* (2002) 15:1178–86. doi: 10.1046/j.1460-9568.2002.01957.x
97. Bowers ME, Ressler KJ. Sex-dependence of anxiety-like behavior in cannabinoid receptor 1 (Cnr1) knockout mice. *Behav Brain Res* (2016) 300:65–9. doi: 10.1016/j.bbr.2015.12.005
98. Simone JJ, Baumbach JL, McCormick CM. Sex-specific effects of CB1 receptor antagonism and stress in adolescence on anxiety, corticosterone concentrations, and contextual fear in adulthood in rats. *Int J Dev Neurosci* (2018) 69:119–31. doi: 10.1016/j.ijdevneu.2018.07.011
99. Uriguen L, Perez-Rial S, Ledent C, Palomo T, Manzanares J. Impaired action of anxiolytic drugs in mice deficient in cannabinoid CB1 receptors. *Neuropharmacology* (2004) 46:966–73. doi: 10.1016/j.neuropharm.2004.01.003
100. Uriguen L, Garcia-Gutierrez MS, Manzanares J. Decreased GABAA and GABAB receptor functional activity in cannabinoid CB1 receptor knockout mice. *J Psychopharmacol* (2011) 25:105–10. doi: 10.1177/0269881109358204
101. Moreira FA, Aguiar DC, Guimaraes FS. Anxiolytic-like effect of cannabinoids injected into the rat dorsolateral periaqueductal gray. *Neuropharmacology* (2007) 52:958–65. doi: 10.1016/j.neuropharm.2006.10.013
102. Batista LA, Bastos JR, Moreira FA. Role of endocannabinoid signalling in the dorsolateral periaqueductal grey in the modulation of distinct panic-like responses. *J Psychopharmacol* (2015) 29:335–43. doi: 10.1177/0269881114566259
103. Viana TG, Hott SC, Resstel LB, Aguiar DC, Moreira FA. Anti-aversive role of the endocannabinoid system in the periaqueductal gray stimulation model of panic attacks in rats. *Psychopharmacol (Berl)* (2015) 232:1545–53. doi: 10.1007/s00213-014-3793-x
104. Lisboa SF, Resstel LB, Aguiar DC, Guimaraes FS. Activation of cannabinoid CB1 receptors in the dorsolateral periaqueductal gray induces anxiolytic effects in rats submitted to the Vogel conflict test. *Eur J Pharmacol* (2008) 593:73–8. doi: 10.1016/j.ejphar.2008.07.032
105. Viana TG, Bastos JR, Costa RB, Hott SC, Mansur FS, Coimbra CC, et al. Hypothalamic endocannabinoid signalling modulates aversive responses related to panic attacks. *Neuropharmacology* (2019) 148:284–90. doi: 10.1016/j.neuropharm.2019.01.022
106. Di S, Itoga CA, Fisher MO, Solomonow J, Roltsch EA, Gilpin NW, et al. Acute Stress Suppresses Synaptic Inhibition and Increases Anxiety via Endocannabinoid Release in the Basolateral Amygdala. *J Neurosci* (2016) 36:8461–70. doi: 10.1523/JNEUROSCI.2279-15.2016
107. Morena M, Leiti KD, Vecchiarelli HA, Gray JM, Campolongo P, Hill MN. Emotional arousal state influences the ability of amygdalar endocannabinoid signaling to modulate anxiety. *Neuropharmacology* (2016) 111:59–69. doi: 10.1016/j.neuropharm.2016.08.020
108. Berger AL, Henricks AM, Lugo JM, Wright HR, Warrick CR, Sticht MA, et al. The Lateral Habenula Directs Coping Styles Under Conditions of Stress via Recruitment of the Endocannabinoid System. *Biol Psychiatry* (2018) 84:611–23. doi: 10.1016/j.biopsych.2018.04.018
109. Hajizadeh Moghaddam A, Bigdellu R, Fatemi Tabatabaei SR, Roohbakhsh A. Cannabinoid system of the lateral septum in the modulation of anxiety-like behaviors in rats. *Arch Iran Med* (2013) 16:711–6. doi: 10.131612/AIM.006
110. Rubino T, Sala M, Vigano D, Braida D, Castiglioni C, Limonta V, et al. Cellular mechanisms underlying the anxiolytic effect of low doses of peripheral Delta9-tetrahydrocannabinol in rats. *Neuropsychopharmacology* (2007) 32:2036–45. doi: 10.1038/sj.npp.1301330
111. Rubino T, Realini N, Castiglioni C, Guidali C, Vigano D, Marras E, et al. Role in anxiety behavior of the endocannabinoid system in the prefrontal cortex. *Cereb Cortex* (2008) 18:1292–301. doi: 10.1016/j.neuropharm.2007.06.024
112. Hartmann A, Fassini A, Scopinho A, Correa FM, Guimaraes FS, Lisboa SF, et al. Role of the endocannabinoid system in the dorsal hippocampus in the cardiovascular changes and delayed anxiety-like effect induced by acute restraint stress in rats. *J Psychopharmacol* (2019) 33:606–14. doi: 10.1177/0269881119827799
113. Kochenborger L, Levone BR, da Silva ES, Taschetto AP, Terenzi MG, Paschoalini MA, et al. The microinjection of a cannabinoid agonist into the accumbens shell induces anxiogenesis in the elevated plus-maze. *Pharmacol Biochem Behav* (2014) 124:160–6. doi: 10.1016/j.pbb.2014.05.017
114. Lisboa SF, Borges AA, Nejo P, Fassini A, Guimaraes FS, Resstel LB. Cannabinoid CB1 receptors in the dorsal hippocampus and prelimbic medial prefrontal cortex modulate anxiety-like behavior in rats: additional evidence. *Prog Neuropsychopharmacol Biol Psychiatry* (2015) 59:76–83. doi: 10.1016/j.pnpbp.2015.01.005
115. Remmers F, Lange MD, Hamann M, Ruehle S, Pape HC, Lutz B. Addressing sufficiency of the CB1 receptor for endocannabinoid-mediated functions through conditional genetic rescue in forebrain GABAergic neurons. *Brain Struct Funct* (2017) 222:3431–52. doi: 10.1007/s00429-017-1411-5
116. Rubino T, Guidali C, Vigano D, Realini N, Valenti M, Massi P, et al. CB1 receptor stimulation in specific brain areas differentially modulate anxiety-related behaviour. *Neuropharmacology* (2008) 54:151–60. doi: 10.1016/j.neuropharm.2007.06.024
117. Mendiguren A, Aostri E, Pineda J. Regulation of noradrenergic and serotonergic systems by cannabinoids: relevance to cannabinoid-induced effects. *Life Sci* (2018) 192:115–27. doi: 10.1016/j.lfs.2017.11.029

118. Haring M, Enk V, Aparisi Rey A, Loch S, Ruiz de Azua I, Weber T, et al. Cannabinoid type-1 receptor signaling in central serotonergic neurons regulates anxiety-like behavior and sociability. *Front Behav Neurosci* (2015) 9:235. doi: 10.3389/fnbeh.2015.00235
119. Zarrindast MR, Mahboobi S, Sadat-Shirazi MS, Ahmadi S. Anxiolytic-like effect induced by the cannabinoid CB1 receptor agonist, arachydonilcyclopropylamide (ACPA), in the rat amygdala is mediated through the D1 and D2 dopaminergic systems. *J Psychopharmacol* (2011) 25:131–40. doi: 10.1177/0269881110376688
120. Zarrindast MR, Sarahroodi S, Arzi A, Khodayar MJ, Taheri-Shalmani S, Rezayof A. Cannabinoid CB1 receptors of the rat central amygdala mediate anxiety-like behavior: interaction with the opioid system. *Behav Pharmacol* (2008) 19:716–23. doi: 10.1097/FBP.0b013e3283123c83
121. Lee TT, Gorzalka BB. Evidence for a Role of Adolescent Endocannabinoid Signaling in Regulating HPA Axis Stress Responsivity and Emotional Behavior Development. *Int Rev Neurobiol* (2015) 125:49–84. doi: 10.1016/b.sirn.2015.09.002
122. Gray JM, Vecchiarelli HA, Morena M, Lee TT, Hermanson DJ, Kim AB, et al. Corticotropin-releasing hormone drives anandamide hydrolysis in the amygdala to promote anxiety. *J Neurosci* (2015) 35:3879–92. doi: 10.1523/JNEUROSCI.2737-14.2015
123. Gray JM, Wilson CD, Lee TT, Pittman QJ, Deussing JM, Hillard CJ, et al. Sustained glucocorticoid exposure recruits cortico-limbic CRH signaling to modulate endocannabinoid function. *Psychoneuroendocrinology* (2016) 66:151–8. doi: 10.1016/j.psyneuen.2016.01.004
124. Lazary J, Lazary A, Gonda X, Benko A, Molnar E, Hunyady L, et al. Promoter variants of the cannabinoid receptor 1 gene (CNR1) in interaction with 5-HTTLPR affect the anxious phenotype. *Am J Med Genet B Neuropsychiatr Genet* (2009) 150B:1118–27. doi: 10.1002/ajmg.b.31024
125. Hay EA, McEwan A, Wilson D, Barrett P, D'Agostino G, Pertwee RG, et al. Disruption of an enhancer associated with addictive behaviour within the cannabinoid receptor-1 gene suggests a possible role in alcohol intake, cannabinoid response and anxiety-related behaviour. *Psychoneuroendocrinology* (2019) 109:104407. doi: 10.1016/j.psyneuen.2019.104407
126. Hayase T. Putative Epigenetic Involvement of the Endocannabinoid System in Anxiety- and Depression-Related Behaviors Caused by Nicotine as a Stressor. *PLoS One* (2016) 11:e0158950. doi: 10.1371/journal.pone.0158950
127. Lomazzo E, König F, Abassi L, Jelinek R, Lutz B. Chronic stress leads to epigenetic dysregulation in the neuropeptide-Y and cannabinoid CB1 receptor genes in the mouse cingulate cortex. *Neuropharmacology* (2017) 113:301–13. doi: 10.1016/j.neuropharm.2016.10.008
128. Bedse G, Hartley ND, Neale E, Gauden AD, Patrick TA, Kingsley PJ, et al. Functional Redundancy Between Canonical Endocannabinoid Signaling Systems in the Modulation of Anxiety. *Biol Psychiatry* (2017) 82:488–99. doi: 10.1016/j.biopsych.2017.03.002
129. Bedse G, Bluett RJ, Patrick TA, Romness NK, Gauden AD, Kingsley PJ, et al. Therapeutic endocannabinoid augmentation for mood and anxiety disorders: comparative profiling of FAAH, MAGL and dual inhibitors. *Transl Psychiatry* (2018) 8:92. doi: 10.1038/s41398-018-0141-7
130. Chicca A, Nicolussi S, Bartholomaeus R, Blunder M, Aparisi Rey A, Petrucci V, et al. Chemical probes to potently and selectively inhibit endocannabinoid cellular reuptake. *Proc Natl Acad Sci U S A* (2017) 114:E5006–15. doi: 10.1073/pnas.1704065114
131. Kinsey SG, O'Neal ST, Long JZ, Cravatt BF, Lichtman AH. Inhibition of endocannabinoid catabolic enzymes elicits anxiolytic-like effects in the marble burying assay. *Pharmacol Biochem Behav* (2011) 98:21–7. doi: 10.1016/j.pbb.2010.12.002
132. Busquets-García A, Puighermanal E, Pastor A, de la Torre R, Maldonado R, Ozaita A. Differential role of anandamide and 2-arachidonoylglycerol in memory and anxiety-like responses. *Biol Psychiatry* (2011) 70:479–86. doi: 10.1016/j.biopsych.2011.04.022
133. Zimmermann T, Bartsch JC, Beer A, Lomazzo E, Guggenhuber S, Lange MD, et al. Impaired anandamide/palmitoylethanolamide signaling in hippocampal glutamatergic neurons alters synaptic plasticity, learning, and emotional responses. *Neuropsychopharmacology* (2019) 44:1377–88. doi: 10.1038/s41386-018-0274-7
134. Aisenberg N, Serova L, Sabban EL, Akirav I. The effects of enhancing endocannabinoid signaling and blocking corticotropin releasing factor receptor in the amygdala and hippocampus on the consolidation of a stressful event. *Eur Neuropsychopharmacol* (2017) 27:913–27. doi: 10.1016/j.euroneuro.2017.06.006
135. Bortolato M, Campolongo P, Mangieri RA, Scattoni ML, Frau R, Trezza V, et al. Anxiolytic-like properties of the anandamide transport inhibitor AM404. *Neuropsychopharmacology* (2006) 31:2652–9. doi: 10.1038/sj.npp.1301061
136. Campos AC, Ferreira FR, Guimaraes FS, Lemos JJ. Facilitation of endocannabinoid effects in the ventral hippocampus modulates anxiety-like behaviors depending on previous stress experience. *Neuroscience* (2010) 167:238–46. doi: 10.1016/j.neuroscience.2010.01.062
137. Duan T, Gu N, Wang Y, Wang F, Zhu J, Fang Y, et al. Fatty acid amide hydrolase inhibitors produce rapid anti-anxiety responses through amygdala long-term depression in male rodents. *J Psychiatry Neurosci* (2017) 42:230–41. doi: 10.1503/jpn.160116
138. El-Alfy AT, Abourashed EA, Patel C, Mazhari N, An H, Jeon A. Phenolic compounds from nutmeg (*Myristica fragrans* Houtt.) inhibit the endocannabinoid-modulating enzyme fatty acid amide hydrolase. *J Pharm Pharmacol* (2019) 71:1879–89. doi: 10.1111/jphp.13174
139. Griebel G, Stemmelin J, Lopez-Grancha M, Fauchey V, Slowinski F, Pichat P, et al. The selective reversible FAAH inhibitor, SSR411298, restores the development of maladaptive behaviors to acute and chronic stress in rodents. *Sci Rep* (2018) 8:2416. doi: 10.1038/s41598-018-20895-z
140. Kathuria S, Gaetani S, Fegley D, Valino F, Duranti A, Tontini A, et al. Modulation of anxiety through blockade of anandamide hydrolysis. *Nat Med* (2003) 9:76–81. doi: 10.1038/nm803
141. Marco EM, Rapino C, Caprioli A, Borsini F, Laviola G, Maccarrone M. Potential Therapeutic Value of a Novel FAAH Inhibitor for the Treatment of Anxiety. *PLoS One* (2015) 10:e0137034. doi: 10.1371/journal.pone.0137034
142. Scherma M, Medalie J, Fratta W, Vadiel SK, Makriyannis A, Piomelli D, et al. The endogenous cannabinoid anandamide has effects on motivation and anxiety that are revealed by fatty acid amide hydrolase (FAAH) inhibition. *Neuropharmacology* (2008) 54:129–40. doi: 10.1016/j.neuropharm.2007.08.011
143. Bluett RJ, Gamble-George JC, Hermanson DJ, Hartley ND, Marnett LJ, Patel S. Central anandamide deficiency predicts stress-induced anxiety: behavioral reversal through endocannabinoid augmentation. *Transl Psychiatry* (2014) 4:e408. doi: 10.1038/tp.2014.53
144. Hill MN, Kumar SA, Filipowski SB, Iverson M, Stühr KL, Keith JM, et al. Disruption of fatty acid amide hydrolase activity prevents the effects of chronic stress on anxiety and amygdalar microstructure. *Mol Psychiatry* (2013) 18:1125–35. doi: 10.1038/mp.2012.90
145. Morena M, Aukema RJ, Leitl KD, Rashid AJ, Vecchiarelli HA, Josselyn SA, et al. Upregulation of Anandamide Hydrolysis in the Basolateral Complex of Amygdala Reduces Fear Memory Expression and Indices of Stress and Anxiety. *J Neurosci* (2019) 39:1275–92. doi: 10.1523/JNEUROSCI.2251-18.2018
146. Rossi S, De Chiara V, Musella A, Sacchetti L, Cantarella C, Castelli M, et al. Preservation of striatal cannabinoid CB1 receptor function correlates with the antianxiety effects of fatty acid amide hydrolase inhibition. *Mol Pharmacol* (2010) 78:260–8. doi: 10.1124/mol.110.064196
147. Moreira FA, Kaiser N, Monory K, Lutz B. Reduced anxiety-like behaviour induced by genetic and pharmacological inhibition of the endocannabinoid-degrading enzyme fatty acid amide hydrolase (FAAH) is mediated by CB1 receptors. *Neuropharmacology* (2008) 54:141–50. doi: 10.1016/j.neuropharm.2007.07.005
148. Haller J, Barna I, Barsvari B, Gyimesi Pelczér K, Yasar S, Panlilio LV, et al. Interactions between environmental aversiveness and the anxiolytic effects of enhanced cannabinoid signaling by FAAH inhibition in rats. *Psychopharmacol (Berl)* (2009) 204:607–16. doi: 10.1007/s00213-009-1494-7
149. Micale V, Cristino L, Tamburella A, Petrosino S, Leggio GM, Drago F, et al. Anxiolytic effects in mice of a dual blocker of fatty acid amide hydrolase and transient receptor potential vanilloid type-1 channels. *Neuropsychopharmacology* (2009) 34:593–606. doi: 10.1038/npp.2008.98
150. Guggenhuber S, Romo-Parra H, Bindila L, Leschik J, Lomazzo E, Remmers F, et al. Impaired 2-AG Signaling in Hippocampal Glutamatergic Neurons: Aggravation of Anxiety-Like Behavior and Unaltered Seizure Susceptibility. *Int J Neuropsychopharmacol* (2015) 19:pyv091. doi: 10.1093/ijnp/pyv091

151. Almeida-Santos AF, Gobira PH, Rosa LC, Guimaraes FS, Moreira FA, Aguiar DC. Modulation of anxiety-like behavior by the endocannabinoid 2-arachidonoylglycerol (2-AG) in the dorsolateral periaqueductal gray. *Behav Brain Res* (2013) 252:10–7. doi: 10.1016/j.bbr.2013.05.027
152. Almeida-Santos AF, Moreira FA, Guimaraes FS, Aguiar DC. 2-Arachidonoylglycerol endocannabinoid signaling coupled to metabotropic glutamate receptor type-5 modulates anxiety-like behavior in the rat ventromedial prefrontal cortex. *J Psychopharmacol* (2017) 31:740–9. doi: 10.1177/0269881117704986
153. Sciolino NR, Zhou W, Hohmann AG. Enhancement of endocannabinoid signaling with JZL184, an inhibitor of the 2-arachidonoylglycerol hydrolyzing enzyme monoacylglycerol lipase, produces anxiolytic effects under conditions of high environmental aversiveness in rats. *Pharmacol Res* (2011) 64:226–34. doi: 10.1016/j.phrs.2011.04.010
154. Aliczki M, Zelena D, Mikics E, Varga ZK, Pinter O, Bakos NV, et al. Monoacylglycerol lipase inhibition-induced changes in plasma corticosterone levels, anxiety and locomotor activity in male CD1 mice. *Horm Behav* (2013) 63:752–8. doi: 10.1016/j.yhbeh.2013.03.017
155. Bosch-Bouju C, Larrieu T, Linders L, Manzoni OJ, Laye S. Endocannabinoid-Mediated Plasticity in Nucleus Accumbens Controls Vulnerability to Anxiety after Social Defeat Stress. *Cell Rep* (2016) 16:1237–42. doi: 10.1016/j.celrep.2016.06.082
156. Gobira PH, Almeida-Santos AF, Guimaraes FS, Moreira FA, Aguiar DC. Role of the endocannabinoid 2-arachidonoylglycerol in aversive responses mediated by the dorsolateral periaqueductal grey. *Eur Neuropsychopharmacol* (2016) 26:15–22. doi: 10.1016/j.euroneuro.2015.11.014
157. Imperatore R, Morello G, Luongo L, Taschler U, Romano R, De Gregorio D, et al. Genetic deletion of monoacylglycerol lipase leads to impaired cannabinoid receptor CB(1)R signaling and anxiety-like behavior. *J Neurochem* (2015) 135:799–813. doi: 10.1111/jnc.13267
158. Jenniches I, Ternes S, Albayram O, Otte DM, Bach K, Bindila L, et al. Anxiety, Stress, and Fear Response in Mice With Reduced Endocannabinoid Levels. *Biol Psychiatry* (2016) 79:858–68. doi: 10.1016/j.biopsych.2015.03.033
159. Shonesy BC, Bluett RJ, Ramikie TS, Baldi R, Hermanson DJ, Kingsley PJ, et al. Genetic disruption of 2-arachidonoylglycerol synthesis reveals a key role for endocannabinoid signaling in anxiety modulation. *Cell Rep* (2014) 9:1644–53. doi: 10.1016/j.celrep.2014.11.001
160. Garcia-Gutierrez MS, Manzanares J. Overexpression of CB2 cannabinoid receptors decreased vulnerability to anxiety and impaired anxiolytic action of alprazolam in mice. *J Psychopharmacol* (2011) 25:111–20. doi: 10.1177/0269881110379507
161. Bahi A, Al Mansouri S, Al Memari E, Al Ameri M, Nurulain SM, Ojha S. beta-Caryophyllene, a CB2 receptor agonist produces multiple behavioral changes relevant to anxiety and depression in mice. *Physiol Behav* (2014) 135:119–24. doi: 10.1016/j.physbeh.2014.06.003
162. Robertson JM, Achua JK, Smith JP, Prince MA, Staton CD, Ronan PJ, et al. Anxious behavior induces elevated hippocampal Cb2 receptor gene expression. *Neuroscience* (2017) 352:273–84. doi: 10.1016/j.neuroscience.2017.03.061
163. Li Y, Kim J. Distinct roles of neuronal and microglial CB2 cannabinoid receptors in the mouse hippocampus. *Neuroscience* (2017) 363:11–25. doi: 10.1016/j.neuroscience.2017.08.053
164. Fabre LF, McLendon D. The efficacy and safety of nabilone (a synthetic cannabinoid) in the treatment of anxiety. *J Clin Pharmacol* (1981) 21:377S–82S. doi: 10.1002/j.1552-4604.1981.tb02617.x
165. Crippa JA, Zuardi AW, Martin-Santos R, Bhattacharyya S, Atakan Z, McGuire P, et al. Cannabis and anxiety: a critical review of the evidence. *Hum Psychopharmacol* (2009) 24:515–23. doi: 10.1002/hup.1048
166. Bhattacharyya S, Egerton A, Kim E, Rosso L, Riano Barros D, Hammers A, et al. Acute induction of anxiety in humans by delta-9-tetrahydrocannabinol related to amygdalar cannabinoid-1 (CB1) receptors. *Sci Rep* (2017) 7:15025. doi: 10.1038/s41598-017-14203-4
167. Christensen R, Kristensen PK, Bartels EM, Bliddal H, Astrup A. Efficacy and safety of the weight-loss drug rimonabant: a meta-analysis of randomised trials. *Lancet* (2007) 370:1706–13. doi: 10.1016/S0140-6736(07)61721-8
168. Bergamaschi MM, Queiroz RH, Chagas MH, Linares IM, Arrais KC, de Oliveira DC, et al. Rimonabant effects on anxiety induced by simulated public speaking in healthy humans: a preliminary report. *Hum Psychopharmacol* (2014) 29:94–9. doi: 10.1002/hup.2374
169. Nguyen T, Thomas BF, Zhang Y. Overcoming the Psychiatric Side Effects of the Cannabinoid CB1 Receptor Antagonists: Current Approaches for Therapeutics Development. *Curr Top Med Chem* (2019) 19:1418–35. doi: 10.2174/1568026619666190708164841
170. Gonda X, Petschner P, Eslari N, Sutori S, Gal Z, Koncz S, et al. Effects of Different Stressors Are Modulated by Different Neurobiological Systems: The Role of GABA-A Versus CB1 Receptor Gene Variants in Anxiety and Depression. *Front Cell Neurosci* (2019) 13:138. doi: 10.3389/fncel.2019.00138
171. Gartner A, Dorfel D, Diers K, Witt SH, Strobel A, Brocke B. Impact of FAAH genetic variation on fronto-amygdala function during emotional processing. *Eur Arch Psychiatry Clin Neurosci* (2019) 269:209–21. doi: 10.1007/s00406-018-0944-9
172. Gee DG, Fetcho RN, Jing D, Li A, Glatt CE, Drysdale AT, et al. Individual differences in frontolimbic circuitry and anxiety emerge with adolescent changes in endocannabinoid signaling across species. *Proc Natl Acad Sci U S A* (2016) 113:4500–5. doi: 10.1073/pnas.1600013113
173. Harris BN, Hohman ZP, Campbell CM, King KS, Tucker CA, Garrison Institute on A. FAAH genotype, CRFR1 genotype, and cortisol interact to predict anxiety in an aging, rural Hispanic population: A Project FRONTIER study. *Neurobiol Stress* (2019) 10:100154. doi: 10.1016/j.ynstr.2019.100154
174. Demers CH, Drabant Conley E, Bogdan R, Hariri AR. Interactions Between Anandamide and Corticotropin-Releasing Factor Signaling Modulate Human Amygdala Function and Risk for Anxiety Disorders: An Imaging Genetics Strategy for Modeling Molecular Interactions. *Biol Psychiatry* (2016) 80:356–62. doi: 10.1016/j.biopsych.2015.12.021
175. Lazary J, Eslari N, Juhasz G, Bagdy G. Genetically reduced FAAH activity may be a risk for the development of anxiety and depression in persons with repetitive childhood trauma. *Eur Neuropsychopharmacol* (2016) 26:1020–8. doi: 10.1016/j.euroneuro.2016.03.003
176. Lazary J, Eslari N, Juhasz G, Bagdy G. A functional variant of CB2 receptor gene interacts with childhood trauma and FAAH gene on anxious and depressive phenotypes. *J Affect Disord* (2019) 257:716–22. doi: 10.1016/j.jad.2019.07.083
177. World Health Organization. (2017). *Depression and other common mental disorders. Global health estimates*. World Health Organization.
178. Aso E, Ozaita A, Valdizan EM, Ledent C, Pazos A, Maldonado R, et al. BDNF impairment in the hippocampus is related to enhanced despair behavior in CB1 knockout mice. *J Neurochem* (2008) 105:565–72. doi: 10.1111/j.1471-4159.2007.05149.x
179. Aso E, Ozaita A, Serra MA, Maldonado R. Genes differentially expressed in CB1 knockout mice: involvement in the depressive-like phenotype. *Eur Neuropsychopharmacol* (2011) 21:11–22. doi: 10.1016/j.euroneuro.2010.06.007
180. Valverde O, Torrens M. CB1 receptor-deficient mice as a model for depression. *Neuroscience* (2012) 204:193–206. doi: 10.1016/j.neuroscience.2011.09.031
181. Shearman LP, Rosko KM, Fleischer R, Wang J, Xu S, Tong XS, et al. Antidepressant-like and anorectic effects of the cannabinoid CB1 receptor inverse agonist AM251 in mice. *Behav Pharmacol* (2003) 14:573–82. doi: 10.1097/00008877-200312000-00001
182. Griebel G, Stemmelin J, Scatton B. Effects of the cannabinoid CB1 receptor antagonist rimonabant in models of emotional reactivity in rodents. *Biol Psychiatry* (2005) 57:261–7. doi: 10.1016/j.biopsych.2004.10.032
183. Adamczyk P, Golda A, McCreary AC, Filip M, Przegalinski E. Activation of endocannabinoid transmission induces antidepressant-like effects in rats. *J Physiol Pharmacol* (2008) 59:217–28.
184. Ebrahimi-Ghiri M, Nasehi M, Zarrindast MR. Anxiolytic and antidepressant effects of ACPA and harmaline co-treatment. *Behav Brain Res* (2019) 364:296–302. doi: 10.1016/j.bbr.2019.02.034
185. Elbatsh MM, Moklas MA, Marsden CA, Kendall DA. Antidepressant-like effects of Delta(9)-tetrahydrocannabinol and rimonabant in the olfactory bulbectomised rat model of depression. *Pharmacol Biochem Behav* (2012) 102:357–65. doi: 10.1016/j.pbb.2012.05.009
186. Haj-Mirzaian A, Amini-Khoei H, Haj-Mirzaian A, Amiri S, Ghesmati M, Zahir M, et al. Activation of cannabinoid receptors elicits antidepressant-like effects in a mouse model of social isolation stress. *Brain Res Bull* (2017) 130:200–10. doi: 10.1016/j.brainresbull.2017.01.018
187. Segev A, Rubin AS, Abush H, Richter-Levin G, Akirav I. Cannabinoid receptor activation prevents the effects of chronic mild stress on emotional learning and LTP in a rat model of depression. *Neuropsychopharmacology* (2014) 39:919–33. doi: 10.1038/npp.2013.292

188. Beyer CE, Dwyer JM, Piesla MJ, Platt BJ, Shen R, Rahman Z, et al. Depression-like phenotype following chronic CB1 receptor antagonism. *Neurobiol Dis* (2010) 39:148–55. doi: 10.1016/j.nbd.2010.03.020
189. McLaughlin RJ, Hill MN, Morrish AC, Gorzalka BB. Local enhancement of cannabinoid CB1 receptor signalling in the dorsal hippocampus elicits an antidepressant-like effect. *Behav Pharmacol* (2007) 18:431–8. doi: 10.1097/FBP.0b013e3282ee7b44
190. Shen CJ, Zheng D, Li KX, Yang JM, Pan HQ, Yu XD, et al. Cannabinoid CB1 receptors in the amygdalar cholecystokinin glutamatergic afferents to nucleus accumbens modulate depressive-like behavior. *Nat Med* (2019) 25:337–49. doi: 10.1038/s41591-018-0299-9
191. Hill MN, Carrier EJ, McLaughlin RJ, Morrish AC, Meier SE, Hillard CJ, et al. Regional alterations in the endocannabinoid system in an animal model of depression: effects of concurrent antidepressant treatment. *J Neurochem* (2008) 106:2322–36. doi: 10.1111/j.1471-4159.2008.05567.x
192. Reich CG, Taylor ME, McCarthy MM. Differential effects of chronic unpredictable stress on hippocampal CB1 receptors in male and female rats. *Behav Brain Res* (2009) 203:264–9. doi: 10.1016/j.bbr.2009.05.013
193. Wang W, Sun D, Pan B, Roberts CJ, Sun X, Hillard CJ, et al. Deficiency in endocannabinoid signaling in the nucleus accumbens induced by chronic unpredictable stress. *Neuropsychopharmacology* (2010) 35:2249–61. doi: 10.1038/npp.2010.99
194. Park H, Rhee J, Lee S, Chung C. Selectively Impaired Endocannabinoid-Dependent Long-Term Depression in the Lateral Habenula in an Animal Model of Depression. *Cell Rep* (2017) 20:289–96. doi: 10.1016/j.celrep.2017.06.049
195. Kirkedal C, Elfving B, Muller HK, Moreira FA, Bindila L, Lutz B, et al. Hemisphere-dependent endocannabinoid system activity in prefrontal cortex and hippocampus of the Flinders Sensitive Line rodent model of depression. *Neurochem Int* (2019) 125:7–15. doi: 10.1016/j.neuint.2019.01.023
196. Vinod KY, Xie S, Psychoyos D, Hungund BL, Cooper TB, Tejani-Butt SM. Dysfunction in fatty acid amide hydrolase is associated with depressive-like behavior in Wistar Kyoto rats. *PLoS One* (2012) 7:e36743. doi: 10.1371/journal.pone.0036743
197. Mangieri RA, Piomelli D. Enhancement of endocannabinoid signaling and the pharmacotherapy of depression. *Pharmacol Res* (2007) 56:360–6. doi: 10.1016/j.phrs.2007.09.003
198. Bortolato M, Mangieri RA, Fu J, Kim JH, Arguello O, Duranti A, et al. Antidepressant-like activity of the fatty acid amide hydrolase inhibitor URB597 in a rat model of chronic mild stress. *Biol Psychiatry* (2007) 62:1103–10. doi: 10.1016/j.biopsych.2006.12.001
199. Realini N, Viganò D, Guidali C, Zamberletti E, Rubino T, Parolaro D. Chronic URB597 treatment at adulthood reverted most depressive-like symptoms induced by adolescent exposure to THC in female rats. *Neuropharmacology* (2011) 60:235–43. doi: 10.1016/j.neuropharm.2010.09.003
200. Wang Y, Zhang X. FAAH inhibition produces antidepressant-like effects of mice to acute stress via synaptic long-term depression. *Behav Brain Res* (2017) 324:138–45. doi: 10.1016/j.bbr.2017.01.054
201. Zhang Z, Wang W, Zhong P, Liu SJ, Long JZ, Zhao L, et al. Blockade of 2-arachidonoylglycerol hydrolysis produces antidepressant-like effects and enhances adult hippocampal neurogenesis and synaptic plasticity. *Hippocampus* (2015) 25:16–26. doi: 10.1002/hipo.22344
202. Dong B, Shilpa BM, Shah R, Goyal A, Xie S, Bakaljian MJ, et al. Dual pharmacological inhibitor of endocannabinoid degrading enzymes reduces depressive-like behavior in female rats. *J Psychiatr Res* (2020) 120:103–12. doi: 10.1016/j.jpsychires.2019.10.010
203. Chen DJ, Gao M, Gao FF, Su QX, Wu J. Brain cannabinoid receptor 2: expression, function and modulation. *Acta Pharmacol Sin* (2017) 38:312–6. doi: 10.1038/aps.2016.149
204. Onaivi ES, Ishiguro H, Gong JP, Patel S, Meozzi PA, Myers L, et al. Brain neuronal CB2 cannabinoid receptors in drug abuse and depression: from mice to human subjects. *PLoS One* (2008) 3:e1640. doi: 10.1371/journal.pone.0001640
205. Hu B, Doods H, Treede RD, Ceci A. Depression-like behaviour in rats with mononeuropathy is reduced by the CB2-selective agonist GW405833. *Pain* (2009) 143:206–12. doi: 10.1016/j.pain.2009.02.018
206. Wang S, Sun H, Liu S, Wang T, Guan J, Jia J. Role of hypothalamic cannabinoid receptors in post-stroke depression in rats. *Brain Res Bull* (2016) 121:91–7. doi: 10.1016/j.brainresbull.2016.01.006
207. Hwang ES, Kim HB, Lee S, Kim MJ, Kim KJ, Han G, et al. Antidepressant-like effects of beta-caryophyllene on restraint plus stress-induced depression. *Behav Brain Res* (2019) 380:112439. doi: 10.1016/j.bbr.2019.112439
208. Smaga I, Bystrowska B, Gawlinski D, Pomierny B, Stankowicz P, Filip M. Antidepressants and changes in concentration of endocannabinoids and N-acylethanolamines in rat brain structures. *Neurotox Res* (2014) 26:190–206. doi: 10.1007/s12640-014-9465-0
209. Hill MN, Ho WS, Hillard CJ, Gorzalka BB. Differential effects of the antidepressants tranylcypromine and fluoxetine on limbic cannabinoid receptor binding and endocannabinoid contents. *J Neural Transm (Vienna)* (2008) 115:1673–9. doi: 10.1007/s00702-008-0131-7
210. Smaga I, Zaniewska M, Gawlinski D, Faron-Gorecka A, Szafranski P, Cegla M, et al. Changes in the cannabinoid receptors in rats following treatment with antidepressants. *Neurotoxicology* (2017) 63:13–20. doi: 10.1016/j.jneuro.2017.08.012
211. Rodriguez-Gaztelumendi A, Rojo ML, Pazos A, Diaz A. Altered CB receptor-signaling in prefrontal cortex from an animal model of depression is reversed by chronic fluoxetine. *J Neurochem* (2009) 108:1423–33. doi: 10.1111/j.1471-4159.2009.05898.x
212. Bambico FR, Katz N, Debonnel G, Gobbi G. Cannabinoids elicit antidepressant-like behavior and activate serotonergic neurons through the medial prefrontal cortex. *J Neurosci* (2007) 27:11700–11. doi: 10.1523/JNEUROSCI.1636-07.2007
213. Burokas A, Martin-Garcia E, Gutierrez-Cuesta J, Rojas S, Herance JR, Gispert JD, et al. Relationships between serotonergic and cannabinoid system in depressive-like behavior: a PET study with [¹¹C]-DASB. *J Neurochem* (2014) 130:126–35. doi: 10.1111/jnc.12716
214. Umathe SN, Manna SS, Jain NS. Involvement of endocannabinoids in antidepressant and anti-compulsive effect of fluoxetine in mice. *Behav Brain Res* (2011) 223:125–34. doi: 10.1016/j.bbr.2011.04.031
215. Hill MN, Ho WS, Sinopoli KJ, Viau V, Hillard CJ, Gorzalka BB. Involvement of the endocannabinoid system in the ability of long-term tricyclic antidepressant treatment to suppress stress-induced activation of the hypothalamic-pituitary-adrenal axis. *Neuropsychopharmacology* (2006) 31:2591–9. doi: 10.1038/sj.npp.1301092
216. Khakpai F, Ebrahimi-Ghiri M, Alijanpour S, Zarrindast MR. Ketamine-induced antidepressant like effects in mice: A possible involvement of cannabinoid system. *BioMed Pharmacother* (2019) 112:108717. doi: 10.1016/j.biopha.2019.108717
217. Ostadhadhi S, Haj-Mirzaian A, Nikoui V, Kordjazy N, Dehpour AR. Involvement of opioid system in antidepressant-like effect of the cannabinoid CB1 receptor inverse agonist AM-251 after physical stress in mice. *Clin Exp Pharmacol Physiol* (2016) 43:203–12. doi: 10.1111/1440-1681.12518
218. Kruk-Slomka M, Michalak A, Biala G. Antidepressant-like effects of the cannabinoid receptor ligands in the forced swimming test in mice: mechanism of action and possible interactions with cholinergic system. *Behav Brain Res* (2015) 284:24–36. doi: 10.1016/j.bbr.2015.01.051
219. Wang HN, Wang L, Zhang RG, Chen YC, Liu L, Gao F, et al. Anti-depressive mechanism of repetitive transcranial magnetic stimulation in rat: the role of the endocannabinoid system. *J Psychiatr Res* (2014) 51:79–87. doi: 10.1016/j.jpsychires.2014.01.004
220. Fang G, Wang Y. Effects of rTMS on Hippocampal Endocannabinoids and Depressive-like Behaviors in Adolescent Rats. *Neurochem Res* (2018) 43:1756–65. doi: 10.1007/s11064-018-2591-y
221. Xue SS, Xue F, Ma QR, Wang SQ, Wang Y, Tan QR, et al. Repetitive high-frequency transcranial magnetic stimulation reverses depressive-like behaviors and protein expression at hippocampal synapses in chronic unpredictable stress-treated rats by enhancing endocannabinoid signaling. *Pharmacol Biochem Behav* (2019) 184:172738. doi: 10.1016/j.pbb.2019.172738
222. Hill MN, Barr AM, Ho WS, Carrier EJ, Gorzalka BB, Hillard CJ. Electroconvulsive shock treatment differentially modulates cortical and subcortical endocannabinoid activity. *J Neurochem* (2007) 103:47–56. doi: 10.1111/j.1471-4159.2007.04688.x
223. Koethe D, Llenos IC, Dulay JR, Hoyer C, Torrey EF, Leweke FM, et al. Expression of CB1 cannabinoid receptor in the anterior cingulate cortex in schizophrenia, bipolar disorder, and major depression. *J Neural Transm (Vienna)* (2007) 114:1055–63. doi: 10.1007/s00702-007-0660-5

224. Choi K, Le T, McGuire J, Xing G, Zhang L, Li H, et al. Expression pattern of the cannabinoid receptor genes in the frontal cortex of mood disorder patients and mice selectively bred for high and low fear. *J Psychiatr Res* (2012) 46:882–9. doi: 10.1016/j.jpsychires.2012.03.021
225. Rodriguez-Munoz M, Sanchez-Blazquez P, Callado LF, Meana JJ, Garzon-Nino J. Schizophrenia and depression, two poles of endocannabinoid system deregulation. *Transl Psychiatry* (2017) 7:1291. doi: 10.1038/s41398-017-0029-y
226. Hill MN, Miller GE, Ho WS, Gorzalka BB, Hillard CJ. Serum endocannabinoid content is altered in females with depressive disorders: a preliminary report. *Pharmacopsychiatry* (2008) 41:48–53. doi: 10.1055/s-2007-993211
227. Hill MN, Miller GE, Carrier EJ, Gorzalka BB, Hillard CJ. Circulating endocannabinoids and N-acyl ethanolamines are differentially regulated in major depression and following exposure to social stress. *Psychoneuroendocrinology* (2009) 34:1257–62. doi: 10.1016/j.psychneuen.2009.03.013
228. Romero-Sanchiz P, Nogueira-Arjona R, Pastor A, Araos P, Serrano A, Boronat A, et al. Plasma concentrations of oleylethanolamide in a primary care sample of depressed patients are increased in those treated with selective serotonin reuptake inhibitor-type antidepressants. *Neuropharmacology* (2019) 149:212–20. doi: 10.1016/j.neuropharm.2019.02.026
229. Heyman E, Gamelin FX, Goekint M, Piscitelli F, Roelands B, Leclaire E, et al. Intense exercise increases circulating endocannabinoid and BDNF levels in humans—possible implications for reward and depression. *Psychoneuroendocrinology* (2012) 37:844–51. doi: 10.1016/j.psychneuen.2011.09.017
230. Meyer JD, Crombie KM, Cook DB, Hillard CJ, Koltyn KF. Serum Endocannabinoid and Mood Changes after Exercise in Major Depressive Disorder. *Med Sci Sports Exerc* (2019) 51:1909–17. doi: 10.1249/MSS.0000000000002006
231. Kranaster L, Hoyer C, Aksay SS, Bumb JM, Leweke FM, Janke C, et al. Electroconvulsive therapy enhances endocannabinoids in the cerebrospinal fluid of patients with major depression: a preliminary prospective study. *Eur Arch Psychiatry Clin Neurosci* (2017) 267:781–6. doi: 10.1007/s00406-017-0789-7
232. Monteleone P, Bifulco M, Maina G, Tortorella A, Gazerro P, Proto MC, et al. Investigation of CNR1 and FAAH endocannabinoid gene polymorphisms in bipolar disorder and major depression. *Pharmacol Res* (2010) 61:400–4. doi: 10.1016/j.phrs.2010.01.002
233. Schennach R, Zill P, Obermeier M, Hauer D, Dehning S, Ceroveck A, et al. The CNR1 gene in depression and schizophrenia - is there an association with early improvement and response? *Psychiatry Res* (2012) 196:160. doi: 10.1016/j.psychres.2011.11.021
234. Domschke K, Dannlowski U, Ohrmann P, Lawford B, Bauer J, Kugel H, et al. Cannabinoid receptor 1 (CNR1) gene: impact on antidepressant treatment response and emotion processing in major depression. *Eur Neuropsychopharmacol* (2008) 18:751–9. doi: 10.1016/j.euroneuro.2008.05.003
235. Mitjans M, Gasto C, Catalan R, Fananas L, Arias B. Genetic variability in the endocannabinoid system and 12-week clinical response to citalopram treatment: the role of the CNR1, CNR2 and FAAH genes. *J Psychopharmacol* (2012) 26:1391–8. doi: 10.1177/0269881112454229
236. Agrawal A, Nelson EC, Littlefield AK, Bucholz KK, Degenhardt L, Henders AK, et al. Cannabinoid receptor genotype moderation of the effects of childhood physical abuse on anhedonia and depression. *Arch Gen Psychiatry* (2012) 69:732–40. doi: 10.1001/archgenpsychiatry.2011.2273
237. Juhasz G, Chase D, Pegg E, Downey D, Toth ZG, Stones K, et al. CNR1 gene is associated with high neuroticism and low agreeableness and interacts with recent negative life events to predict current depressive symptoms. *Neuropsychopharmacology* (2009) 34:2019–27. doi: 10.1038/npp.2009.19
238. Barrero FJ, Ampuero I, Morales B, Vives F, de Dios Luna Del Castillo J, Hoenicka J, et al. Depression in Parkinson's disease is related to a genetic polymorphism of the cannabinoid receptor gene (CNR1). *Pharmacogenomics J* (2005) 5:135–41. doi: 10.1038/sj.tpj.6500301
239. Ickick R, Peoc'h K, Karsinti E, Ksouda K, Hajj A, Bloch V, et al. A cannabinoid receptor 1 polymorphism is protective against major depressive disorder in methadone-maintained outpatients. *Am J Addict* (2015) 24:613–20. doi: 10.1111/ajad.12273
240. Kong X, Miao Q, Lu X, Zhang Z, Chen M, Zhang J, et al. The association of endocannabinoid receptor genes (CNR1 and CNR2) polymorphisms with depression: A meta-analysis. *Med (Baltimore)* (2019) 98:e17403. doi: 10.1097/MD.00000000000017403
241. Onaivi ES, Ishiguro H, Gong JP, Patel S, Meozzi PA, Myers L, et al. Functional expression of brain neuronal CB2 cannabinoid receptors are involved in the effects of drugs of abuse and in depression. *Ann N Y Acad Sci* (2008) 1139:434–49. doi: 10.1196/annals.1432.036
242. Owen MJ, Sawa A, Mortensen PB. Schizophrenia. *Lancet* (2016) 388:86–97. doi: 10.1016/S0140-6736(15)01121-6
243. Marder SR, Cannon TD. Schizophrenia. *N Engl J Med* (2019) 381:1753–61. doi: 10.1056/NEJMra1808803
244. Sherif M, Radhakrishnan R, D'Souza DC, Ranganathan M. Human Laboratory Studies on Cannabinoids and Psychosis. *Biol Psychiatry* (2016) 79:526–38. doi: 10.1016/j.biopsych.2016.01.011
245. Powell SB, Zhou X, Geyer MA. Prepulse inhibition and genetic mouse models of schizophrenia. *Behav Brain Res* (2009) 204:282–94. doi: 10.1016/j.bbr.2009.04.021
246. Braff DL. Prepulse inhibition of the startle reflex: a window on the brain in schizophrenia. *Curr Top Behav Neurosci* (2010) 4:349–71. doi: 10.1007/7854_2010_61
247. Mansbach RS, Rovetti CC, Winston EN, Lowe 3. , Effects of the cannabinoid CB1 receptor antagonist SR141716A on the behavior of pigeons and rats. *Psychopharmacol (Berl)* (1996) 124:315–22. doi: 10.1007/BF02247436
248. Martin RS, Secchi RL, Sung E, Lemaire M, Bonhaus DW, Hedley LR, et al. Effects of cannabinoid receptor ligands on psychosis-relevant behavior models in the rat. *Psychopharmacol (Berl)* (2003) 165:128–35. doi: 10.1007/s00213-002-1240-x
249. Hajos M, Hoffmann WE, Kocsis B. Activation of cannabinoid-1 receptors disrupts sensory gating and neuronal oscillation: relevance to schizophrenia. *Biol Psychiatry* (2008) 63:1075–83. doi: 10.1016/j.biopsych.2007.12.005
250. Lee G, Zhou Y. NMDAR Hypofunction Animal Models of Schizophrenia. *Front Mol Neurosci* (2019) 12:185. doi: 10.3389/fnmol.2019.00185
251. Ballmaier M, Bortolato M, Rizzetti C, Zoli M, Gessa G, Heinz A, et al. Cannabinoid receptor antagonists counteract sensorimotor gating deficits in the phencyclidine model of psychosis. *Neuropsychopharmacology* (2007) 32:2098–107. doi: 10.1038/sj.npp.1301344
252. Guidali C, Vigano D, Petrosino S, Zamberletti E, Realini N, Binelli G, et al. Cannabinoid CB1 receptor antagonism prevents neurochemical and behavioural deficits induced by chronic phencyclidine. *Int J Neuropsychopharmacol* (2011) 14:17–28. doi: 10.1017/S1461145710000209
253. Black MD, Stevens RJ, Rogacki N, Featherstone RE, Senyah Y, Giardino O, et al. AVE1625, a cannabinoid CB1 receptor antagonist, as a co-treatment with antipsychotics for schizophrenia: improvement in cognitive function and reduction of antipsychotic-side effects in rodents. *Psychopharmacol (Berl)* (2011) 215:149–63. doi: 10.1007/s00213-010-2124-0
254. Kruk-Slomka M, Budzyska B, Slomka T, Banaszkiewicz I, Biala G. The Influence of the CB1 Receptor Ligands on the Schizophrenia-Like Effects in Mice Induced by MK-801. *Neurotox Res* (2016) 30:658–76. doi: 10.1007/s12640-016-9662-0
255. Szucs E, Dvoracko S, Tomboly C, Buki A, Kekesi G, Horvath G, et al. Decreased CB receptor binding and cannabinoid signaling in three brain regions of a rat model of schizophrenia. *Neurosci Lett* (2016) 633:87–93. doi: 10.1016/j.neulet.2016.09.020
256. Gomes FV, Edelson JR, Volk DW, Grace AA. Altered brain cannabinoid 1 receptor mRNA expression across postnatal development in the MAM model of schizophrenia. *Schizophr Res* (2018) 201:254–60. doi: 10.1016/j.schres.2018.04.030
257. Almeida V, Levin R, Peres FF, Suiaima MA, Vendramini AM, Santos CM, et al. Role of the endocannabinoid and endovanilloid systems in an animal model of schizophrenia-related emotional processing/cognitive deficit. *Neuropharmacology* (2019) 155:44–53. doi: 10.1016/j.neuropharm.2019.05.015
258. Neary JL, Perez SM, Peterson K, Lodge DJ, Carless MA. Comparative analysis of MBD-seq and MeDIP-seq and estimation of gene expression changes in a rodent model of schizophrenia. *Genomics* (2017) 109:204–13. doi: 10.1016/j.ygeno.2017.03.004
259. Perez SM, Aguilar DD, Neary JL, Carless MA, Giuffrida A, Lodge DJ. Schizophrenia-Like Phenotype Inherited by the F2 Generation of a Gestational Disruption Model of Schizophrenia. *Neuropsychopharmacology* (2016) 41:477–86. doi: 10.1038/npp.2015.169

260. Perez SM, Donegan JJ, Boley AM, Aguilar DD, Giuffrida A, Lodge DJ. Ventral hippocampal overexpression of Cannabinoid Receptor Interacting Protein 1 (CNRI1) produces a schizophrenia-like phenotype in the rat. *Schizophr Res* (2019) 206:263–70. doi: 10.1016/j.schres.2018.11.006
261. Vigano D, Guidali C, Petrosino S, Realini N, Rubino T, Di Marzo V, et al. Involvement of the endocannabinoid system in phencyclidine-induced cognitive deficits modelling schizophrenia. *Int J Neuropsychopharmacol* (2009) 12:599–614. doi: 10.1017/S1461145708009371
262. Eisenstein SA, Clapper JR, Holmes PV, Piomelli D, Hohmann AG. A role for 2-arachidonoylglycerol and endocannabinoid signaling in the locomotor response to novelty induced by olfactory bulbectomy. *Pharmacol Res* (2010) 61:419–29. doi: 10.1016/j.phrs.2009.12.013
263. Karl T. Neuregulin 1: a prime candidate for research into gene-environment interactions in schizophrenia? Insights from genetic rodent models. *Front Behav Neurosci* (2013) 7:106. doi: 10.3389/fnbeh.2013.00106
264. Clarke DJ, Stuart J, McGregor IS, Arnold JC. Endocannabinoid dysregulation in cognitive and stress-related brain regions in the Nrg1 mouse model of schizophrenia. *Prog Neuropsychopharmacol Biol Psychiatry* (2017) 72:9–15. doi: 10.1016/j.pnpbp.2016.08.006
265. Ishiguro H, Horiuchi Y, Ishikawa M, Koga M, Imai K, Suzuki Y, et al. Brain cannabinoid CB2 receptor in schizophrenia. *Biol Psychiatry* (2010) 67:974–82. doi: 10.1016/j.biopsych.2009.09.024
266. Ortega-Alvaro A, Aracil-Fernandez A, Garcia-Gutierrez MS, Navarrete F, Manzanares J. Deletion of CB2 cannabinoid receptor induces schizophrenia-related behaviors in mice. *Neuropsychopharmacology* (2011) 36:1489–504. doi: 10.1038/npp.2011.34
267. Khella R, Short JL, Malone DT. CB2 receptor agonism reverses MK-801-induced disruptions of prepulse inhibition in mice. *Psychopharmacol (Berl)* (2014) 231:3071–87. doi: 10.1007/s00213-014-3481-x
268. Kruk-Slomka M, Banaszkiewicz I, Biala G. The Impact of CB2 Receptor Ligands on the MK-801-Induced Hyperactivity in Mice. *Neurotox Res* (2017) 31:410–20. doi: 10.1007/s12640-017-9702-4
269. Dean B, Sundram S, Bradbury R, Scarr E, Copolov D. Studies on [3H]-CP-55940 binding in the human central nervous system: regional specific changes in density of cannabinoid-1 receptors associated with schizophrenia and cannabis use. *Neuroscience* (2001) 103:9–15. doi: 10.1016/S0306-4522(00)00552-2
270. Jenko KJ, Hirvonen J, Henter ID, Anderson KB, Zoghbi SS, Hyde TM, et al. Binding of a tritiated inverse agonist to cannabinoid CB1 receptors is increased in patients with schizophrenia. *Schizophr Res* (2012) 141:185–8. doi: 10.1016/j.schres.2012.07.021
271. Volk DW, Eggen SM, Horta AG, Wong DF, Lewis DA. Reciprocal alterations in cortical cannabinoid receptor 1 binding relative to protein immunoreactivity and transcript levels in schizophrenia. *Schizophr Res* (2014) 159:124–9. doi: 10.1016/j.schres.2014.07.017
272. Dalton VS, Long LE, Weickert CS, Zavitsanou K. Paranoid schizophrenia is characterized by increased CB1 receptor binding in the dorsolateral prefrontal cortex. *Neuropsychopharmacology* (2011) 36:1620–30. doi: 10.1038/npp.2011.43
273. Muguruza C, Morentin B, Meana JJ, Alexander SP, Callado LF. Endocannabinoid system imbalance in the postmortem prefrontal cortex of subjects with schizophrenia. *J Psychopharmacol* (2019) 33:1132–40. doi: 10.1177/0269881119857205
274. Zavitsanou K, Garrick T, Huang XF. Selective antagonist [3H]-SR141716A binding to cannabinoid CB1 receptors is increased in the anterior cingulate cortex in schizophrenia. *Prog Neuropsychopharmacol Biol Psychiatry* (2004) 28:355–60. doi: 10.1016/j.pnpbp.2003.11.005
275. Newell KA, Deng C, Huang XF. Increased cannabinoid receptor density in the posterior cingulate cortex in schizophrenia. *Exp Brain Res* (2006) 172:556–60. doi: 10.1007/s00221-006-0503-x
276. Dong R, Liu X, Liu Y, Deng Z, Nie X, Wang X, et al. Enrichment of epidermal stem cells by rapid adherence and analysis of the reciprocal interaction of epidermal stem cells with neighboring cells using an organotypic system. *Cell Biol Int* (2007) 31:733–40. doi: 10.1016/j.cellbi.2007.01.007
277. Eggen SM, Hashimoto T, Lewis DA. Reduced cortical cannabinoid 1 receptor messenger RNA and protein expression in schizophrenia. *Arch Gen Psychiatry* (2008) 65:772–84. doi: 10.1001/archpsyc.65.7.772
278. Uriguen L, Garcia-Fuster MJ, Callado LF, Morentin B, La Harpe R, Casado V, et al. Immunodensity and mRNA expression of A2A adenosine, D2 dopamine, and CB1 cannabinoid receptors in postmortem frontal cortex of subjects with schizophrenia: effect of antipsychotic treatment. *Psychopharmacol (Berl)* (2009) 206:313–24. doi: 10.1007/s00213-009-1608-2
279. Wong DF, Kuwabara H, Horta AG, Raymont V, Brasic J, Guevara M, et al. Quantification of cerebral cannabinoid receptors subtype 1 (CB1) in healthy subjects and schizophrenia by the novel PET radioligand [11C]-OMAR. *Neuroimage* (2010) 52:1505–13. doi: 10.1016/j.neuroimage.2010.04.034
280. Ceccarini J, De Hert M, Van Winkel R, Peuskens J, Bormans G, Kranaster L, et al. Increased ventral striatal CB1 receptor binding is related to negative symptoms in drug-free patients with schizophrenia. *Neuroimage* (2013) 79:304–12. doi: 10.1016/j.neuroimage.2013.04.052
281. Ranganathan M, Cortes-Briones J, Radhakrishnan R, Thurnauer H, Planeta B, Skosnik P, et al. Reduced Brain Cannabinoid Receptor Availability in Schizophrenia. *Biol Psychiatry* (2016) 79:997–1005. doi: 10.1016/j.biopsych.2015.08.021
282. Mihov Y. Positron Emission Tomography Studies on Cannabinoid Receptor Type 1 in Schizophrenia. *Biol Psychiatry* (2016) 79:e97–9. doi: 10.1016/j.biopsych.2016.04.015
283. Borgan F, Laurikainen H, Veronese M, Marques TR, Haaparanta-Solin M, Solin O, et al. In Vivo Availability of Cannabinoid 1 Receptor Levels in Patients With First-Episode Psychosis. *JAMA Psychiatry* (2019). 76:1074–84. doi: 10.1001/jamapsychiatry.2019.1427
284. Leroy S, Griffon N, Bourdel MC, Olie JP, Poirier MF, Krebs MO. Schizophrenia and the cannabinoid receptor type 1 (CB1): association study using a single-base polymorphism in coding exon 1. *Am J Med Genet* (2001) 105:749–52. doi: 10.1002/ajmg.10038
285. Ujike H, Takaki M, Nakata K, Tanaka Y, Takeda T, Kodama M, et al. CNR1, central cannabinoid receptor gene, associated with susceptibility to hebephrenic schizophrenia. *Mol Psychiatry* (2002) 7:515–8. doi: 10.1038/sj.mp.4001029
286. Zammit S, Spurlock G, Williams H, Norton N, Williams N, O'Donovan MC, et al. Genotype effects of CHRNA7, CNR1 and COMT in schizophrenia: interactions with tobacco and cannabis use. *Br J Psychiatry* (2007) 191:402–7. doi: 10.1192/bjp.bp.107.036129
287. Seifert J, Ossege S, Emrich HM, Schneider U, Stuhmann M. No association of CNR1 gene variations with susceptibility to schizophrenia. *Neurosci Lett* (2007) 426:29–33. doi: 10.1016/j.neulet.2007.08.008
288. Hamdani N, Tabeze JP, Ramoz N, Ades J, Hamon M, Sarfati Y, et al. The CNR1 gene as a pharmacogenetic factor for antipsychotics rather than a susceptibility gene for schizophrenia. *Eur Neuropsychopharmacol* (2008) 18:34–40. doi: 10.1016/j.euroneuro.2007.05.005
289. Bae JS, Kim JY, Park BL, Kim JH, Kim B, Park CS, et al. Genetic association analysis of CNR1 and CNR2 polymorphisms with schizophrenia in a Korean population. *Psychiatr Genet* (2014) 24:225–9. doi: 10.1097/YPG.0000000000000047
290. Costa M, Squassina A, Congiu D, Chillotti C, Niola P, Galderisi S, et al. Investigation of endocannabinoid system genes suggests association between peroxisome proliferator activator receptor- α gene (PPARA) and schizophrenia. *Eur Neuropsychopharmacol* (2013) 23:749–59. doi: 10.1016/j.euroneuro.2012.07.007
291. Tsai SJ, Wang YC, Hong CJ. Association study of a cannabinoid receptor gene (CNR1) polymorphism and schizophrenia. *Psychiatr Genet* (2000) 10:149–51. doi: 10.1097/00041444-200010030-00008
292. Suarez-Pinilla P, Roiz-Santanez R, Ortiz-Garcia de la Foz V, Guest PC, Ayasa-Arriola R, Cordova-Palomera A, et al. Brain structural and clinical changes after first episode psychosis: Focus on cannabinoid receptor 1 polymorphisms. *Psychiatry Res* (2015) 233:112–9. doi: 10.1016/j.psychres.2015.05.005
293. Rojnic Kuzman M, Bosnjak Kuharic D, Ganoci L, Makaric P, Keki I, Rossini Gajsak L, et al. Association of CNR1 genotypes with changes in neurocognitive performance after eighteen-month treatment in patients with first-episode psychosis. *Eur Psychiatry* (2019) 61:88–96. doi: 10.1016/j.eurpsy.2019.07.004
294. Yu W, De Hert M, Moons T, Claes SJ, Correll CU, van Winkel R. CNR1 gene and risk of the metabolic syndrome in patients with schizophrenia. *J Clin Psychopharmacol* (2013) 33:186–92. doi: 10.1097/JCP.0b013e318283925e
295. De Marchi N, De Petrocellis L, Orlando P, Daniele F, Fezza F, Di Marzo V. Endocannabinoid signalling in the blood of patients with schizophrenia. *Lipids Health Dis* (2003) 2:5. doi: 10.1186/1476-511X-2-5
296. Ferretjans R, de Campos SM, Ribeiro-Santos R, Guimaraes FC, de Oliveira K, Cardoso AC, et al. Cognitive performance and peripheral endocannabinoid

- system receptor expression in schizophrenia. *Schizophr Res* (2014) 156:254–60. doi: 10.1016/j.schres.2014.04.028
297. de Campos-Carli SM, Araujo MS, de Oliveira Silveira AC, de Rezende VB, Rocha NP, Ferretjans R, et al. Cannabinoid receptors on peripheral leukocytes from patients with schizophrenia: Evidence for defective immunomodulatory mechanisms. *J Psychiatr Res* (2017) 87:44–52. doi: 10.1016/j.jpsychires.2016.12.001
 298. Chase KA, Feiner B, Rosen C, Gavin DP, Sharma RP. Characterization of peripheral cannabinoid receptor expression and clinical correlates in schizophrenia. *Psychiatry Res* (2016) 245:346–53. doi: 10.1016/j.psychres.2016.08.055
 299. D'Addario C, Micale V, Di Bartolomeo M, Stark T, Pucci M, Sulcova A, et al. A preliminary study of endocannabinoid system regulation in psychosis: Distinct alterations of CNR1 promoter DNA methylation in patients with schizophrenia. *Schizophr Res* (2017) 188:132–40. doi: 10.1016/j.schres.2017.01.022
 300. Leweke FM, Giuffrida A, Wurster U, Emrich HM, Piomelli D. Elevated endogenous cannabinoids in schizophrenia. *Neuroreport* (1999) 10:1665–9. doi: 10.1097/00001756-199906030-00008
 301. Giuffrida A, Leweke FM, Gerth CW, Schreiber D, Koethe D, Faulhaber J, et al. Cerebrospinal anandamide levels are elevated in acute schizophrenia and are inversely correlated with psychotic symptoms. *Neuropsychopharmacology* (2004) 29:2108–14. doi: 10.1038/sj.npp.1300558
 302. Leweke FM, Giuffrida A, Koethe D, Schreiber D, Nolden BM, Kranaster L, et al. Anandamide levels in cerebrospinal fluid of first-episode schizophrenic patients: impact of cannabis use. *Schizophr Res* (2007) 94:29–36. doi: 10.1016/j.schres.2007.04.025
 303. Koethe D, Pahlisch F, Hellmich M, Rohleder C, Mueller JK, Meyer-Lindenberg A, et al. Familial abnormalities of endocannabinoid signaling in schizophrenia. *World J Biol Psychiatry* (2019) 20:117–25. doi: 10.1080/15622975.2018.1449966
 304. Potvin S, Kouassi E, Lipp O, Bouchard RH, Roy MA, Demers MF, et al. Endogenous cannabinoids in patients with schizophrenia and substance use disorder during quetiapine therapy. *J Psychopharmacol* (2008) 22:262–9. doi: 10.1177/0269881107083816
 305. Desfosses J, Stip E, Bentaleb LA, Lipp O, Chiasson JP, Furtos A, et al. Plasma Endocannabinoid Alterations in Individuals with Substance Use Disorder are Dependent on the “Mirror Effect” of Schizophrenia. *Front Psychiatry* (2012) 3:85. doi: 10.3389/fpsy.2012.00085
 306. Muguruza C, Lehtonen M, Aaltonen N, Morentin B, Meana JJ, Callado LF. Quantification of endocannabinoids in postmortem brain of schizophrenic subjects. *Schizophr Res* (2013) 148:145–50. doi: 10.1016/j.schres.2013.06.013
 307. Morita Y, Ujike H, Tanaka Y, Uchida N, Nomura A, Ohtani K, et al. A nonsynonymous polymorphism in the human fatty acid amide hydrolase gene did not associate with either methamphetamine dependence or schizophrenia. *Neurosci Lett* (2005) 376:182–7. doi: 10.1016/j.neulet.2004.11.050
 308. Bioque M, Cabrera B, Garcia-Bueno B, MacDowell KS, Torrent C, Saiz PA, et al. Dysregulated peripheral endocannabinoid system signaling is associated with cognitive deficits in first-episode psychosis. *J Psychiatr Res* (2016) 75:14–21. doi: 10.1016/j.jpsychires.2016.01.002
 309. Volk DW, Siegel BI, Verrico CD, Lewis DA. Endocannabinoid metabolism in the prefrontal cortex in schizophrenia. *Schizophr Res* (2013) 147:53–7. doi: 10.1016/j.schres.2013.02.038
 310. Bioque M, Garcia-Bueno B, Macdowell KS, Meseguer A, Saiz PA, Parellada M, et al. Peripheral endocannabinoid system dysregulation in first-episode psychosis. *Neuropsychopharmacology* (2013) 38:2568–77. doi: 10.1038/npp.2013.165
 311. Tong D, He S, Wang L, Jin L, Si P, Cheng X. Association of single-nucleotide polymorphisms in the cannabinoid receptor 2 gene with schizophrenia in the Han Chinese population. *J Mol Neurosci* (2013) 51:454–60. doi: 10.1007/s12031-013-0062-0
 312. Arjmand S, Behzadi M, Kohlmeier KA, Mazhari S, Sabahi A, Shabani M. Bipolar disorder and the endocannabinoid system. *Acta Neuropsychiatr* (2019) 31:193–201. doi: 10.1017/neu.2019.21
 313. Ashton CH, Moore PB. Endocannabinoid system dysfunction in mood and related disorders. *Acta Psychiatr Scand* (2011) 124:250–61. doi: 10.1111/j.1600-0447.2011.01687.x
 314. Alpak G, Copoglu S, Geyik E, Unal A, Igci M, Igci Y, et al. Rs6454674, Rs806368 and Rs1049353 CNR1 Gene Polymorphisms in Turkish Bipolar Disorder Patients: A Preliminary Study. *Dis Mol Med* (2014) 2:4. doi: 10.5455/dmm.20140428011918
 315. Tsai SJ, Wang YC, Hong CJ. Association study between cannabinoid receptor gene (CNR1) and pathogenesis and psychotic symptoms of mood disorders. *Am J Med Genet* (2001) 105:219–21. doi: 10.1002/ajmg.1259
 316. Pisanu C, Congiu D, Costa M, Sestu M, Chillotti C, Ardau R, et al. No association of endocannabinoid genes with bipolar disorder or lithium response in a Sardinian sample. *Psychiatry Res* (2013) 210:887–90. doi: 10.1016/j.psychres.2013.09.025
 317. Minocci D, Massei J, Martino A, Milanti M, Piz L, Di Bello D, et al. Genetic association between bipolar disorder and 524A>C (Leu133Ile) polymorphism of CNR2 gene, encoding for CB2 cannabinoid receptor. *J Affect Disord* (2011) 134:427–30. doi: 10.1016/j.jad.2011.05.023
 318. Legge SE, Jones HJ, Kendall KM, Pardinas AF, Menzies G, Bracher-Smith M, et al. Association of Genetic Liability to Psychotic Experiences With Neuropsychotic Disorders and Traits. *JAMA Psychiatry* (2019) 76:1256–65. doi: 10.1001/jamapsychiatry.2019.2508
 319. Berardi A, Schelling G, Campolongo P. The endocannabinoid system and Post Traumatic Stress Disorder (PTSD): From preclinical findings to innovative therapeutic approaches in clinical settings. *Pharmacol Res* (2016) 111:668–78. doi: 10.1016/j.phrs.2016.07.024
 320. Bassir Nia A, Bender R, Harpaz-Rotem I. Endocannabinoid System Alterations in Posttraumatic Stress Disorder: A Review of Developmental and Accumulative Effects of Trauma. *Chronic Stress (Thousand Oaks)* (2019) 3: 1–23. doi: 10.1177/2470547019864096
 321. Shoshan N, Akirav I. The effects of cannabinoid receptors activation and glucocorticoid receptors deactivation in the amygdala and hippocampus on the consolidation of a traumatic event. *Neurobiol Learn Mem* (2017) 144:248–58. doi: 10.1016/j.nlm.2017.08.004
 322. Campos AC, Ferreira FR, da Silva WA Jr., Guimaraes FS. Predator threat stress promotes long lasting anxiety-like behaviors and modulates synaptophysin and CB1 receptors expression in brain areas associated with PTSD symptoms. *Neurosci Lett* (2013) 533:34–8. doi: 10.1016/j.neulet.2012.11.016
 323. Xing G, Carlton J, Zhang L, Jiang X, Fullerton C, Li H, et al. Cannabinoid receptor expression and phosphorylation are differentially regulated between male and female cerebellum and brain stem after repeated stress: implication for PTSD and drug abuse. *Neurosci Lett* (2011) 502:5–9. doi: 10.1016/j.neulet.2011.05.013
 324. Fride E, Suris R, Weidenfeld J, Mechoulam R. Differential response to acute and repeated stress in cannabinoid CB1 receptor knockout newborn and adult mice. *Behav Pharmacol* (2005) 16:431–40. doi: 10.1097/00008877-200509000-00016
 325. Bowers ME, Ressler KJ. Interaction between the cholecystokinin and endogenous cannabinoid systems in cued fear expression and extinction retention. *Neuropsychopharmacology* (2015) 40:688–700. doi: 10.1038/npp.2014.225
 326. Korem N, Akirav I. Cannabinoids prevent the effects of a footshock followed by situational reminders on emotional processing. *Neuropsychopharmacology* (2014) 39:2709–22. doi: 10.1038/npp.2014.132
 327. Abush H, Akirav I. Cannabinoids modulate hippocampal memory and plasticity. *Hippocampus* (2010) 20:1126–38. doi: 10.1002/hipo.20711
 328. Ganon-Elazar E, Akirav I. Cannabinoids prevent the development of behavioral and endocrine alterations in a rat model of intense stress. *Neuropsychopharmacology* (2012) 37:456–66. doi: 10.1038/npp.2011.204
 329. Ganon-Elazar E, Akirav I. Cannabinoids and traumatic stress modulation of contextual fear extinction and GR expression in the amygdala-hippocampal-prefrontal circuit. *Psychoneuroendocrinology* (2013) 38:1675–87. doi: 10.1016/j.psyneuen.2013.01.014
 330. Korem N, Lange R, Hillard CJ, Akirav I. Role of beta-catenin and endocannabinoids in the nucleus accumbens in extinction in rats exposed to shock and reminders. *Neuroscience* (2017) 357:285–94. doi: 10.1016/j.neuroscience.2017.06.015
 331. Goodman J, Packard MG. Peripheral and intra-dorsolateral striatum injections of the cannabinoid receptor agonist WIN 55,212-2 impair consolidation of stimulus-response memory. *Neuroscience* (2014) 274:128–37. doi: 10.1016/j.neuroscience.2014.05.007
 332. Reich CG, Iskander AN, Weiss MS. Cannabinoid modulation of chronic mild stress-induced selective enhancement of trace fear conditioning in

- adolescent rats. *J Psychopharmacol* (2013) 27:947–55. doi: 10.1177/0269881113499207
333. Chhatwal JP, Davis M, Maguschak KA, Ressler KJ. Enhancing cannabinoid neurotransmission augments the extinction of conditioned fear. *Neuropsychopharmacology* (2005) 30:516–24. doi: 10.1038/sj.npp.1300655
334. Danandeh A, Vozella V, Lim J, Oveisi F, Ramirez GL, Mears D, et al. Effects of fatty acid amide hydrolase inhibitor URB597 in a rat model of trauma-induced long-term anxiety. *Psychopharmacol (Berl)* (2018) 235:3211–21. doi: 10.1007/s00213-018-5020-7
335. Fidelman S, Mizrahi Zer-Aviv T, Lange R, Hillard CJ, Akirav I. Chronic treatment with URB597 ameliorates post-stress symptoms in a rat model of PTSD. *Eur Neuropsychopharmacol* (2018) 28:630–42. doi: 10.1016/j.euroneuro.2018.02.004
336. Maymon N, Mizrahi Zer-Aviv T, Sabban EL, Akirav I, Neuropeptide Y, and cannabinoids interaction in the amygdala after exposure to shock and reminders model of PTSD. *Neuropharmacology* (2020) 162:107804. doi: 10.1016/j.neuropharm.2019.107804
337. Segev A, Korem N, Mizrahi Zer-Aviv T, Abush H, Lange R, Sauber G, et al. Role of endocannabinoids in the hippocampus and amygdala in emotional memory and plasticity. *Neuropsychopharmacology* (2018) 43:2017–27. doi: 10.1038/s41386-018-0135-4
338. Varvel SA, Wise LE, Niyuhire F, Cravatt BF, Lichtman AH. Inhibition of fatty-acid amide hydrolase accelerates acquisition and extinction rates in a spatial memory task. *Neuropsychopharmacology* (2007) 32:1032–41. doi: 10.1038/sj.npp.1301224
339. Mota N, Sumner JA, Lowe SR, Neumeister A, Uddin M, Aiello AE, et al. The rs1049353 polymorphism in the CNR1 gene interacts with childhood abuse to predict posttraumatic threat symptoms. *J Clin Psychiatry* (2015) 76:e1622–3. doi: 10.4088/JCP.15110084
340. Lu AT, Ogdie MN, Jarvelin MR, Moilanen IK, Loo SK, McCracken JT, et al. Association of the cannabinoid receptor gene (CNR1) with ADHD and post-traumatic stress disorder. *Am J Med Genet B Neuropsychiatr Genet* (2008) 147B:1488–94. doi: 10.1002/ajmg.b.30693
341. Hill MN, Blier LM, Makotkine I, Golier JA, Galea S, McEwen BS, et al. Reductions in circulating endocannabinoid levels in individuals with post-traumatic stress disorder following exposure to the World Trade Center attacks. *Psychoneuroendocrinology* (2013) 38:2952–61. doi: 10.1016/j.psyneuen.2013.08.004
342. Neumeister A, Normandin MD, Pietrzak RH, Piomelli D, Zheng MQ, Gujarrro-Anton A, et al. Elevated brain cannabinoid CB1 receptor availability in post-traumatic stress disorder: a positron emission tomography study. *Mol Psychiatry* (2013) 18:1034–40. doi: 10.1038/mp.2013.61
343. Wilker S, Pfeiffer A, Elbert T, Ovuga E, Karabatsiakos A, Krumbholz A, et al. Endocannabinoid concentrations in hair are associated with PTSD symptom severity. *Psychoneuroendocrinology* (2016) 67:198–206. doi: 10.1016/j.psyneuen.2016.02.010
344. Hariri AR, Gorka A, Hyde LW, Kimak M, Halder I, Ducci F, et al. Divergent effects of genetic variation in endocannabinoid signaling on human threat- and reward-related brain function. *Biol Psychiatry* (2009) 66:9–16. doi: 10.1016/j.biopsych.2008.10.047
345. Gunduz-Cinar O, MacPherson KP, Cinar R, Gamble-George J, Sugden K, Williams B, et al. Convergent translational evidence of a role for anandamide in amygdala-mediated fear extinction, threat processing and stress-reactivity. *Mol Psychiatry* (2013) 18:813–23. doi: 10.1038/mp.2012.72
346. Dincheva I, Drysdale AT, Hartley CA, Johnson DC, Jing D, King EC, et al. FAAH genetic variation enhances fronto-amygdala function in mouse and human. *Nat Commun* (2015) 6:6395. doi: 10.1038/ncomms7395
347. Rabinak A, Angstadt M, Sripada CS, Abelson JL, Liberzon I, Milad MR, et al. Cannabinoid facilitation of fear extinction memory recall in humans. *Neuropharmacology* (2013) 64:396–402. doi: 10.1016/j.neuropharm.2012.06.063
348. Rabinak CA, Angstadt M, Lyons M, Mori S, Milad MR, Liberzon I, et al. Cannabinoid modulation of prefrontal-limbic activation during fear extinction learning and recall in humans. *Neurobiol Learn Mem* (2014) 113:125–34. doi: 10.1016/j.nlm.2013.09.009
349. Roitman P, Mechoulam R, Cooper-Kazaz R, Shalev A. Preliminary, open-label, pilot study of add-on oral Delta9-tetrahydrocannabinol in chronic post-traumatic stress disorder. *Clin Drug Invest* (2014) 34:587–91. doi: 10.1007/s40261-014-0212-3
350. Jetly R, Heber A, Fraser G, Boisvert D. The efficacy of nabilone, a synthetic cannabinoid, in the treatment of PTSD-associated nightmares: A preliminary randomized, double-blind, placebo-controlled cross-over design study. *Psychoneuroendocrinology* (2015) 51:585–8. doi: 10.1016/j.psyneuen.2014.11.002
351. Cameron C, Watson D, Robinson J. Use of a synthetic cannabinoid in a correctional population for posttraumatic stress disorder-related insomnia and nightmares, chronic pain, harm reduction, and other indications: a retrospective evaluation. *J Clin Psychopharmacol* (2014) 34:559–64. doi: 10.1097/JCP.0000000000000180
352. Leffa DT, Ferreira SG, Machado NJ, Souza CM, Rosa FD, de Carvalho C, et al. Caffeine and cannabinoid receptors modulate impulsive behavior in an animal model of attentional deficit and hyperactivity disorder. *Eur J Neurosci* (2019) 49:1673–83. doi: 10.1111/ejn.14348
353. Haspula D, Clark MA. Heterologous regulation of the cannabinoid type 1 receptor by angiotensin II in astrocytes of spontaneously hypertensive rats. *J Neurochem* (2016) 139:523–36. doi: 10.1111/jnc.13776
354. Grissom NM, Herdt CT, Desilets J, Lidsky-Everson J, Reyes TM. Dissociable deficits of executive function caused by gestational adversity are linked to specific transcriptional changes in the prefrontal cortex. *Neuropsychopharmacology* (2015) 40:1353–63. doi: 10.1038/npp.2014.313
355. Van Lieshout RJ, Taylor VH, Boyle MH. Pre-pregnancy and pregnancy obesity and neurodevelopmental outcomes in offspring: a systematic review. *Obes Rev* (2011) 12:e548–59. doi: 10.1111/j.1467-789X.2010.00850.x
356. Kleijn J, Wiskerke J, Cremers TI, Schoffeleer AN, Westerink BH, Pattij T. Effects of amphetamine on dopamine release in the rat nucleus accumbens shell region depend on cannabinoid CB1 receptor activation. *Neurochem Int* (2012) 60:791–8. doi: 10.1016/j.neuint.2012.03.002
357. Buchmann AF, Hohm E, Witt SH, Blomeyer D, Jennen-Steinmetz C, Schmidt MH, et al. Role of CNR1 polymorphisms in moderating the effects of psychosocial adversity on impulsivity in adolescents. *J Neural Transm (Vienna)* (2015) 122:455–63. doi: 10.1007/s00702-014-1266-3
358. Ponce G, Hoenicka J, Rubio G, Ampuero I, Jimenez-Arriero MA, Rodriguez-Jimenez R, et al. Association between cannabinoid receptor gene (CNR1) and childhood attention deficit/hyperactivity disorder in Spanish male alcoholic patients. *Mol Psychiatry* (2003) 8:466–7. doi: 10.1038/sj.mp.4001278
359. Bermudez-Silva FJ, Cardinal P, Cota D. The role of the endocannabinoid system in the neuroendocrine regulation of energy balance. *J Psychopharmacol* (2012) 26:114–24. doi: 10.1177/0269881111408458
360. Quarta C, Mazza R, Obici S, Pasquali R, Pagotto U. Energy balance regulation by endocannabinoids at central and peripheral levels. *Trends Mol Med* (2011) 17:518–26. doi: 10.1016/j.molmed.2011.05.002
361. Rorato R, Miyahara C, Antunes-Rodrigues J, Elias LL. Tolerance to hypophagia induced by prolonged treatment with a CB1 antagonist is related to the reversion of anorexigenic neuropeptide gene expression in the hypothalamus. *Regul Pept* (2013) 182:12–8. doi: 10.1016/j.regpep.2012.12.004
362. Verty AN, Boon WM, Mallet PE, McGregor IS, Oldfield BJ. Involvement of hypothalamic peptides in the anorectic action of the CB receptor antagonist rimobant (SR 141716). *Eur J Neurosci* (2009) 29:2207–16. doi: 10.1111/j.1460-9568.2009.06750.x
363. Lage R, Parisi C, Seoane-Collazo P, Ferno J, Mazza R, Bosch F, et al. Lack of Hypophagia in CB1 Null Mice is Associated to Decreased Hypothalamic POMC and CART Expression. *Int J Neuropsychopharmacol* (2015) 18:pyv011. doi: 10.1093/ijnp/pyv011
364. Moore CF, Schlain GS, Mancino S, Sabino V, Cottone P. A behavioral and pharmacological characterization of palatable diet alternation in mice. *Pharmacol Biochem Behav* (2017) 163:1–8. doi: 10.1016/j.pbb.2017.10.013
365. Riedel G, Fadda P, McKillop-Smith S, Pertwee RG, Platt B, Robinson L. Synthetic and plant-derived cannabinoid receptor antagonists show hypophagic properties in fasted and non-fasted mice. *Br J Pharmacol* (2009) 156:1154–66. doi: 10.1111/j.1476-5381.2008.00107.x
366. Wiley JL, Burston JJ, Leggett DC, Alekseeva OO, Razdan RK, Mahadevan A, et al. CB1 cannabinoid receptor-mediated modulation of food intake in mice. *Br J Pharmacol* (2005) 145:293–300. doi: 10.1038/sj.bjp.0706157
367. Sofia RD, Knobloch LC. Comparative effects of various naturally occurring cannabinoids on food, sucrose and water consumption by rats. *Pharmacol Biochem Behav* (1976) 4:591–9. doi: 10.1016/0091-3057(76)90202-1

368. Wierucka-Rybak M, Wolak M, Bojanowska E. The effects of leptin in combination with a cannabinoid receptor 1 antagonist, AM 251, or cannabidiol on food intake and body weight in rats fed a high-fat or a free-choice high sugar diet. *J Physiol Pharmacol* (2014) 65:487–96.
369. Collu R, Scherma M, Piscitelli F, Giunti E, Satta V, Castelli MP, et al. Impaired brain endocannabinoid tone in the activity-based model of anorexia nervosa. *Int J Eat Disord* (2019) 52:1251–62. doi: 10.1002/eat.23157
370. Monteleone P, Matias I, Martiadis V, De Petrocellis L, Maj M, Di Marzo V. Blood levels of the endocannabinoid anandamide are increased in anorexia nervosa and in binge-eating disorder, but not in bulimia nervosa. *Neuropsychopharmacology* (2005) 30:1216–21. doi: 10.1038/sj.npp.1300695
371. Gerard N, Pieters G, Goffin K, Bormans G, Van Laere K. Brain type 1 cannabinoid receptor availability in patients with anorexia and bulimia nervosa. *Biol Psychiatry* (2011) 70:777–84. doi: 10.1016/j.biopsych.2011.05.010
372. Schroeder M, Eberlein C, de Zwaan M, Kornhuber J, Bleich S, Frieling H. Lower levels of cannabinoid 1 receptor mRNA in female eating disorder patients: association with wrist cutting as impulsive self-injurious behavior. *Psychoneuroendocrinology* (2012) 37:2032–6. doi: 10.1016/j.psyneuen.2012.03.025
373. Dider S, Ji J, Zhao Z, Xie L. Molecular mechanisms involved in the side effects of fatty acid amide hydrolase inhibitors: a structural phenomics approach to proteome-wide cellular off-target deconvolution and disease association. *NPJ Syst Biol Appl* (2016) 2:16023. doi: 10.1038/npsba.2016.23
374. Toczek M, Malinowska B. Enhanced endocannabinoid tone as a potential target of pharmacotherapy. *Life Sci* (2018) 204:20–45. doi: 10.1016/j.lfs.2018.04.054
375. Rubino T, Parolaro D. Sexually dimorphic effects of cannabinoid compounds on emotion and cognition. *Front Behav Neurosci* (2011) 5:64. doi: 10.3389/fnbeh.2011.00064
376. Green T, Flash S, Reiss AL. Sex differences in psychiatric disorders: what we can learn from sex chromosome aneuploidies. *Neuropsychopharmacology* (2019) 44:9–21. doi: 10.1038/s41386-018-0153-2
377. Cooper ZD, Craft RM. Sex-Dependent Effects of Cannabis and Cannabinoids: A Translational Perspective. *Neuropsychopharmacology* (2018) 43:34–51. doi: 10.1038/npp.2017.140

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Diagnostic and Predictive Applications of Functional Near-Infrared Spectroscopy for Major Depressive Disorder: A Systematic Review

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OPEN ACCESS

Edited by:

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Specialty section:

This article was submitted to
Mood and Anxiety Disorders,
a section of the journal
Frontiers in Psychiatry

Received: 01 December 2019

Accepted: 15 April 2020

Published: 06 May 2020

Citation:

Ho CSH, Lim LJH, Lim AQ,
Chan NHC, Tan RS, Lee SH and
Ho RCM (2020) Diagnostic
and Predictive Applications of
Functional Near-Infrared
Spectroscopy for Major Depressive
Disorder: A Systematic Review.
Front. Psychiatry 11:378.
doi: 10.3389/fpsy.2020.00378

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Introduction: Major depressive disorder (MDD) is a global psychiatric disorder with no established biomarker. There is growing evidence that functional near-infrared spectroscopy (fNIRS) has the ability to aid in the diagnosis and prediction of the treatment response of MDD. The aim of this review was to systematically review, and gather the evidence from existing studies that used fNIRS signals in the diagnosis of MDD, correlations with depression symptomatology, and the monitoring of treatment response.

Methods: PubMed, EMBASE, ScienceDirect, and Cochrane Library databases were searched for published English articles from 1980 to June 2019 that focused on the application of fNIRS for (i) differentiating depressed versus nondepressed individuals, (ii) correlating with depression symptomatology, and in turn (iii) monitoring treatment responses in depression. Studies were included if they utilized fNIRS to evaluate cerebral hemodynamic variations in patients with MDD of any age group. The quality of the evidence was assessed using the Newcastle–Ottawa quality assessment scale.

Results: A total of 64 studies were included in this review, with 12 studies being longitudinal, while the rest were cross-sectional. More than two-thirds of the studies ($n = 49$) had acceptable quality. fNIRS consistently demonstrated attenuated cerebral hemodynamic changes in depressed compared to healthy individuals. fNIRS signals have also shown promise in correlating with individual symptoms of depression and monitoring various treatment responses.

Conclusions: This review provides comprehensive updated evidence of the diagnostic and predictive applications of fNIRS in patients with MDD. Future studies involving larger

sample sizes, standardized methodology, examination of more brain regions in an integrative approach, and longitudinal follow-ups are needed.

Keywords: diagnostic, prediction, functional near-infrared spectroscopy, major depressive disorder, systematic review

INTRODUCTION

Major depressive disorder (MDD) is a global mental illness which is increasingly prevalent in modern societies. As per the World Health Organization, MDD affected approximately 322 million people of all ages globally. The total number of people likely to have depression increased by 18.4% between 2005 and 2015, and this number is expected to increase exponentially over time (1). Symptoms of depression include low mood, decreased energy, poor attention, memory problems, disturbed appetite and sleep, anhedonia, feelings of guilt, and worthlessness (2). In severe cases, depression may present with psychotic symptoms, suicidal thoughts and increase the possibility of unnatural death (3). MDD can also cause significant disability. Depression is one of the leading cause of world disability in the year 2020, and in 10 years' time, it is anticipated to be the biggest cause of global disease burden overtaking cardiovascular diseases (1). Despite its gaining prevalence, MDD is still considerably undertreated and under diagnosed, especially in primary care settings (4).

Depressive disorders have been correlated with problems in the limbic, thalamic and cortical areas (5). MDD has also been correlated with neuropsychological deficiencies in numerous cognitive areas, involving attention, language, memory, and executive function (6). To diagnose patients with MDD, clinicians conventionally refer to the International Statistical Classification of Diseases and Related Health Problems—10th revision (ICD-10) or the Diagnostic and Statistical Manual of Mental Disorders, 5th Edition (DSM-5) classifications as guides to confirm the diagnosis. However, the accuracy of reaching a diagnosis of MDD relying on history taking remains debated. Diagnosis is often based on the subjective assessment and clinical experience of the clinicians. Some patients may also not be forthcoming about their symptoms, especially suicidal ideation. Furthermore, many of the psychiatric symptoms are polymorphous and may overlap in various psychiatric disorders, thereby making diagnosis all the more challenging.

MDD is a multifaceted and varied illness in which up to two thirds of patients could experience treatment resistance that protracts and worsens the episodes (7). Only approximately 33% of patients with MDD attain remission, despite being treated with optimal medications based on measurement-based care and consensus guideline. Furthermore, the probability of treatment response seems to decrease with each new treatment option (8). Treatment-resistant depression (TRD) is related to increased morbidity and mortality, with recurring and chronic periods in the long run (9). Hence, it would be useful if there are ways to ascertain improvements in treatment at any stage of the illness, as it would offer wider benefits for global management of depression.

Biomarkers offer a conceivable target for assisting in the diagnosis and identifying predictors of response to various interventions (10). Biomarkers may come in various forms, such as inflammatory markers, endogenously produced hormones and brain imaging. In recent years, brain-imaging techniques such as electroencephalography (EEG), functional magnetic resonance imaging (fMRI), positron emission tomography (PET), and magnetoencephalography (MEG) existed to be used as adjuncts to help clinicians diagnose MDD. According to some neuroimaging findings using PET and fMRI to investigate brain function in patients with depression (11–13), they found blood flow reduction in the prefrontal cortex to be associated with a decline in activity within the cingulate cortex. There had been reports using neuroimaging techniques recording hemodynamic response relating to brain activity during cognitive stimulation on depressed patients. One such study using fMRI performed by Okada et al. revealed decreased left prefrontal activation and reduced task performance in depressed patients utilizing the verbal fluency task (VFT) (11). However, these tests are expensive to conduct. Additionally, the patients were required to place themselves in an awkward posture, i.e., lying supine in a narrow space with the head fixed during the investigation (14).

In 2009, functional near-infrared spectroscopy (fNIRS) was sanctioned in Japan. It was classified to be an advanced medical technology for differentiation of psychiatric illness (15). Additionally, in 2013, fNIRS obtained medical insurance coverage for being an adjunct diagnostic tool. fNIRS examinations have since been utilized for psychophysiological assessment of cognitive function.

fNIRS is a form of spectroscopy that utilizes light sources between a spectral window of 650 to 1000 nm which penetrates organic tissues. Oxygenated hemoglobin (oxy-Hb) variations are then computed using the variance in absorbance using the modified Beer–Lambert law. This is a noninvasive technique that can detect cortical oxygenation levels of hemoglobin (16), and low cortical oxygenation levels have been correlated with depressive illness. The advantages of using fNIRS are that it is a relatively inexpensive procedure, portable, and easy to set up, and it does not involve nonionizing radiation. Hence, it may be repeated multiple times on an as-needed and when-needed basis for patients. In fact, fNIRS has been applied in many other areas of the medical field, such as cognition and preoperative functional assessment (17). With regard to psychiatric illness, in addition to MDD, there are studies pertaining to its use for patients with schizophrenia and bipolar disorder among other disorders (18) (**Table 1**). Many fNIRS studies involve tasks that help activate brain activity in the subject, such as VFT or passively viewing photographs to trigger an emotional response.

TABLE 1 | Studies using fNIRS to assess different psychiatric disorders.

| Study | Psychiatric disorder | Key finding |
|-------------------------|---|---|
| Noda T. et al. (19) | Schizophrenia | Prefrontal and temporal region oxy-Hb uptake during the post-verbal fluency task (VFT) period was associated with working memory deficits in patients with schizophrenia. |
| Hirose T. et al. (20) | Bipolar disorder | Suicide risk in patients with bipolar disorder was correlated with delayed activation timing of NIRS signal during the VFT in the prefrontal region. |
| Katzorke A. et al. (21) | Dementia/cognitive impairment | Patients with mild cognitive impairment had decreased hemodynamic response in the inferior frontotemporal cortex as compared to healthy controls using VFT. |
| Ueda S. et al. (22) | Attention-deficit hyperactivity disorder (ADHD) | Adult ADHD patients had reduced prefrontal hemodynamic response during the Stroop Color-Word Task compared to healthy controls, and this response was similar to pediatric studies. |

Oxy-Hb, oxygenated hemoglobin; fNIRS, functional near-infrared spectroscopy.

To date, most of the studies on the diagnostic and predictive applications of fNIRS for MDD have been conducted in Japan. Furthermore, most studies have been conducted with a relatively small number of participants. To date, there is only one meta-analysis, published in 2015, looking at fNIRS for differentiating patients with depression from healthy subjects (23). Since then, more studies have been conducted on the matter as the use of fNIRS on depressed individuals to help in diagnosis and monitoring treatment response has gained acceptance over the last few years. Through a meta-analysis, Zhang et al. found that MDD patients had considerably decreased prefrontal cortex activation when undertaking cognitive tasks relative to controls. Patients with MDD, as opposed to controls, were associated with reduced rise in oxy-Hb in prefrontal regions during cognitive stimulation. This distinctive pattern of blood oxygen variations in the prefrontal cortex in MDD patients may be used as an objective diagnostic instrument for MDD. However, with such a low quantity of studies in the first meta-analysis, we decided to evaluate the use of fNIRS as a diagnostic biomarker for the diagnosis of MDD and its distinction between different stages of the illness, and to discuss its usefulness as a monitoring biomarker for treatment response in MDD patients, as based on a systematic review of the latest available literature. This paper aims to cover (i) the use of fNIRS to differentiate depressed from healthy individuals, (ii) correlation of fNIRS signals with depression symptomatology, and (iii) how it can be applied to monitor treatment response.

METHODS

Data Sources and Search Strategy

This study was conducted on the basis of the Preferred Reporting Items for Systematic Reviews and Meta-Analysis (PRISMA). A systematic review was completed with published English-language literature from 1980 to June 2019, focusing on the

utility of fNIRS for (i) differentiating depressed versus nondepressed individuals, (ii) correlating with depression symptomatology, and (iii) monitoring treatment responses in depression. The four electronic databases searched were PubMed, EMBASE, ScienceDirect, and Cochrane Library. The search terms used were “Spectroscopy” or “Near-infrared” or “near-infrared spectroscopy” or “fNIRS” or “optical topography” and “Depression” or “depressed” or “depressive disorder” or “mood disorder” or “affective disorder.” Terms were searched as both text words and subject headings. The team scanned for related publications, conference proceedings and bibliographies of papers gathered manually. This systematic review was funded by the National University of Singapore iHeathtech Other Operating Expenses (R-722-000-004-731).

Eligibility Criteria and Data Collection

Studies were short-listed if the authors utilized fNIRS to measure cerebral hemodynamic variations in patients with MDD of any age group. All titles and abstracts retrieved from the databases were independently reviewed by two reviewers (LL and NC). Where appropriate, full-text papers were extracted for further inspection. Studies which were selected by either reviewer and fulfilled the inclusion criteria proceeded on to full-text review, whereby the study characteristics and results were extracted. Any discrepancies in the study selection were brought to the attention of the third reviewer (CH) and resolved by discussion. Studies were divided into the three categories that were most appropriate for the three questions set out in the review. The data review form consisted of the subsequent data: authors and publication year, country, sample characteristics, diagnostic criteria used, type of NIRS device, paradigm used, and main findings of the study. As much data as possible were obtained from the articles, and efforts were made to contact the authors if supplementary data were needed.

Quality Assessment of the Articles

The articles included in the full-text review were evaluated by utilizing the Newcastle–Ottawa quality assessment scale (NOS) (24), with the subsequent generation of a table with star scores for each study. There is no overall score that determines whether a study is “good” or “bad,” but a star is awarded for meeting each criterion involving selection and outcome, with the exception of compatibility where two stars can be awarded. Two reviewers (LL and NC) independently assessed the papers, and discrepancies were brought to the third reviewer (CH) to resolve by discussion.

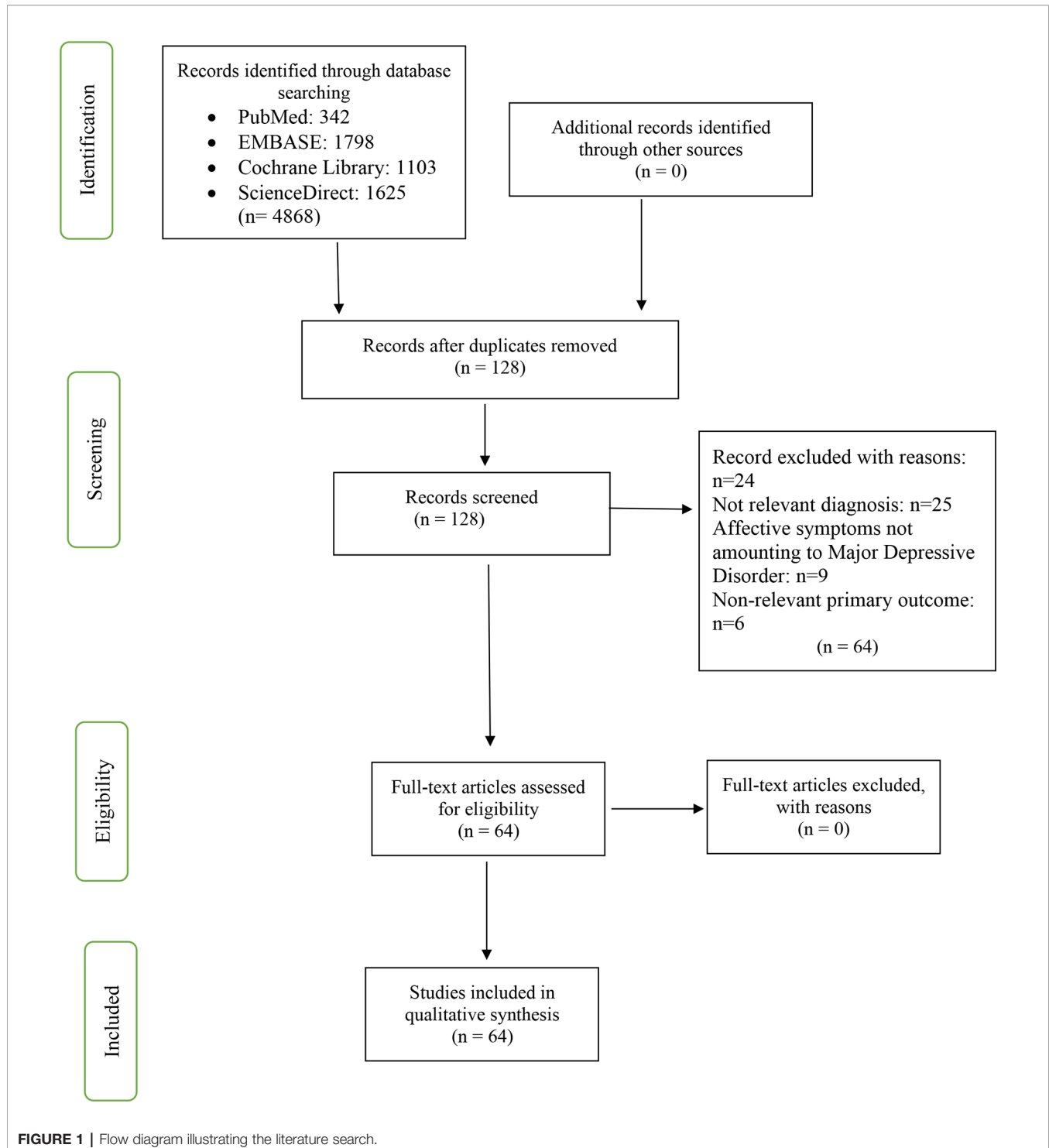
RESULTS

Study Selection

A total of 4868 citations were identified from our database search, with 342 from PubMed, 1798 from EMBASE, 1625 from ScienceDirect, and 1103 from Cochrane Library. After reviewing the titles, abstracts, and removing duplicated publications, 128 articles were selected. Of these, 64 studies met the inclusion criteria and were encompassed in this

analysis. The selection process is displayed in **Figure 1**, constructed according to the PRISMA statement. Forty-seven studies were from Japan, eight studies from China, six studies from Germany, one study from the UK, one study from the USA, and one study from Uzbekistan. These studies were further divided into categories based on which of the three questions they addressed: (i) differentiation of depressed from healthy

individuals, (ii) correlation with depression symptomatology, and (iii) assessment of treatment response. Fifty-one studies addressed the first question, 31 studies addressed the second question, and 16 studies addressed the third question. Twelve studies were longitudinal, while the rest were cross-sectional studies. Seven studies were extracted from conference proceedings, while the remaining were full-text articles.



Use of fNIRS to Differentiate Depressed From Healthy Individuals

We identified 51 papers that used fNIRS to distinguish depressed patients from healthy controls (HCs) (**Table 2**). The bulk of the studies were conducted in Japan ($n = 38$), followed by China ($n = 7$), Germany ($n = 4$), and the United Kingdom ($n = 1$). The pooled sample across the studies comprised a total of 2094 unipolar depressed patients and 2457 HCs. All the depressed patients in the respective studies fulfilled the ICD-10 or DSM-IV diagnostic criteria for MDD, with the exception of two studies that used the DSM-5 and one study that used the DSM-III as the diagnostic criteria instrument. The psychopathology measure used for the studies was mostly the Hamilton Depression Rating Scale (HAM-D). With the exception of six studies that did not include an indication of the medications that were consumed by the depressed patients, three studies that included medication-naïve patients and two studies that included antidepressant-naïve patients, the depressed patients generally received antidepressants of different types and dosages with or without other types of medication. The majority of studies utilized the VFT as the active paradigm, while others adopted the verbal repetition task, hyperventilation and paper-bag breathing, visuospatial task, Stroop task, Sternberg's task, Tower of Hanoi, CO₂ inhalation task, working memory (WM) task, word generation tasks (WGT), emotional Stroop task, facial emotion recognition task, image-recall task, mirror drawing task (MDT), trail-making test (TMT), or a combination of tasks [e.g., word fluency task (WFT) with right-finger-tapping task]. The fNIRS instruments that were used and the number of channels utilized, with a large proportion using the 52-channel ETG-4000, varied across studies. Most of the studies measured the concentration of oxy-Hb and deoxygenated hemoglobin (deoxy-Hb) during various tasks for distinguishing the depressed from the healthy. The probe is normally positioned at the frontal (usually prefrontal) and/or temporal areas, with the exception of two papers that measured the parietal area as well and one paper that measured only the left and right hemispheres in general. For the papers that included the assessment of the parietal brain areas, one of them included subjects suffering from Alzheimer's disease, and the parietal dysfunctions were linked to the initial phase of dementia (30). Other studies such as Rosenbaum et al. (72) examined the frontoparietal networks of patients with early- and late-life depression (LLD), which is associated with cognitive control. The study discovered that the frontoparietal networks appeared to be essential in LLD as patients suffering from LLD and memory impairment are shown to be at elevated risk for having dementia (75).

The overall results showed that the depressed patients showed a smaller oxy-Hb increase than the controls, with a smaller increase in the frontal (especially the prefrontal) and temporal activation (appreciably lower activation in the depressed compared to HCs) while performing the WM task, VFT, WGT, and TMT. Kito et al. (30) measured both the frontal and parietal brain areas, and the results showed decreased cortical activation throughout the VFT in patients with depression and the HCs. With the same brain areas being

measured, the depressed patients and HCs showed noteworthy dissimilarities in functional connectivity (FC) both in the resting state and during TMT performance, with depressed patients showing a decrease, and HCs showing a surge, in FC from the resting state to TMT performance (72). When subjects were asked to hyperventilate, there was a significant decrease in oxy-Hb levels, and deoxy-Hb was considerably elevated in the depressed and HCs, with the depressed patients showing a lesser reduction in oxy-Hb than healthy individuals (26, 31). During paper-bag breathing, Matsuo et al. (31) revealed that oxy-Hb significantly increased in both groups, whereas deoxy-Hb significantly decreased in the depressed juxtaposed to the healthy. Matsuo et al. (26) established no noteworthy alterations in oxy-Hb and deoxy-Hb in depressed patients or in HCs. Using the emotional Stroop task, Matsubara et al. found that the HCs compared to the depressed patients showed notably increased oxy-Hb during the happy-word trials, but the decreases in oxy-Hb during the threat-word trials were comparable in both depressed patients and HCs (32). However, Matsubara et al. and Nishizawa et al. concluded that the depressed when comparing with HCs showed notable increase in oxy-Hb within the left middle frontal region while performing the threat task, whereas depressed patients compared to HCs showed no noteworthy variation in oxy-Hb while performing the happy task (42, 60). For the CO₂ inhalation task, the vasomotor reactivity was significantly decreased in the depressed patients than controls (35). When subjects were performing right-finger-tapping tasks, the increase in oxy-Hb was higher in the depressed patients when comparing with HCs (18). In the image-recall task, the change in oxy-Hb in the HCs was considerably more than in the depressed while experiencing unpleasant conditions (53).

Almost all of the studies did not indicate the specificity and sensitivity of differentiating between depressed patients and HCs, with the exception of one that indicated an 80% sensitivity but with its specificity being unspecified (29). One study had indicated a sensitivity of 71.5% and a specificity of 70% for differentiating depressed patients from those with Alzheimer's disease (30), and another study mentioned a sensitivity of 0.71 and a specificity of 0.46 to distinguish between the euthymic, unipolar and bipolar depressive patients (37). Last, there was one study that indicated different sensitivities and specificities, which were dependent on the integral values of the two regions of interest (region 1 consisted of frontopolar and dorsolateral prefrontal cortical regions, while region 2 consisted of the middle and superior temporal cortical regions and the ventrolateral prefrontal cortex) (39).

Correlation of fNIRS Signals With Depression Symptomatology

We identified 31 eligible papers reporting on fNIRS studies in which cerebral hemodynamic changes were correlated with depression symptomatology in a total of 1424 patients (**Table 3**). These studies were generally in those with MDD, but some were inclusive of patients with other mental health illness, for example: bipolar disorder, affective disorder, and

TABLE 2 | Summary of fNIRS studies differentiating depressed patients from healthy controls.

| Source | Country | Sample size (male/female) | Age (mean \pm standard deviation) | Diagnostic criteria (instrument) | Psychopathology measure | Medication | NIRS device | Paradigm | Brain area | Main findings |
|--------------------------|---------|--|---|--|----------------------------|-----------------------------------|-----------------------------------|--|---------------|--|
| Pu et al., (25) | Japan | MDD: 24 (12/12) HC: 26 (8/18) | MDD: 47.9 \pm 13.9 HC: 42.4 \pm 9.3 | DSM-IV-TR (MINI) | BDI HAMD | All on antidepressants | 52-Channel NIRS (ETG-4000) | 2-Back task with blocked periodic baseline, activation | PF T | – MDD group had \downarrow response sensitivity and accuracy than HC. |
| Matsuo et al. (26) | Japan | UNI: 8 (1/7) BP: 1 (1/0) HC: 10 (0/10) | UNI+BP: 65.6 \pm 6.4 HC: 59.5 \pm 5.9 | DSM-IV (unspecified) | HAMD | All on medication | HEO-200 | Verbal repetition task, VFT, Hyperventilation, Paper-bag breathing | F | – VFT: Oxy-Hb \uparrow and deoxy-Hb \downarrow in HC group but no noteworthy changes in depressed group. – Hyperventilation: oxy-Hb \downarrow while deoxy-Hb \uparrow |
| Herrmann et al. (27) | Germany | MDD: 9 (5/4) HC: 9 (5/4) | MDD: 37.3 \pm 13.8 HC: 35.1 \pm 5.5 | ICD-10 | BDI | All on medication | 2-Channel NIRO- 300 monitor | VFT | PF | – Oxy-Hb \uparrow in HC. – MDD had significantly \downarrow activation. |
| Shoji et al. (28) | Japan | MDD: 26 HC: 32 | NA | ICD-10 | HAMD | NA | 44-channel ETG- 4000 (Hitachi) | Word generation tasks | PF | – Oxy-Hb variations in MDD were appreciably lesser than HC in all word tasks. |
| Kinoshita et al. (29) | Japan | MDD: 17 | 44.2 \pm 12.2 | DSM-IV (SCID) | HAMD | Majority on antidepressants | 22-Channel ETG- 4000 (Hitachi) | DEX/CRH test VFT | F | – Results did not fit well with the diagnostic criteria (DSM or ICD). |
| Kito et al. (30) | Japan | MDD: 30 (9/21) HC: 33 (11/22) | MDD: 71.1 \pm 6.8 HC: 69.6 \pm 5.5 | DSM-IV | HAMD | All on medication | FOIRE-3000 (Shimadzu) | VFT | F, P | – Cortical activation in the VFT in MDD \downarrow compared to HC. |
| Matsuo et al. (31) | Japan | MDD: 14 (4/10) HC: 21 (3/18) | MDD: 56.1 \pm 17.3 HC: 50.3 \pm 12.6 | DSM-IV | HAMD | All on medication | Single channel HEO-200 (Omron) | VFT Hyperventilation Paper-bag breathing | F | – VFT: \uparrow in oxy-Hb was lower in MDD compared to HC. – Hyperventilation: MDD demonstrated an appreciably smaller reduction in oxy-Hb than HC. – Paper-bag breathing: oxy-Hb \uparrow in both groups while deoxy-Hb \downarrow in MDD. |
| Matsubara et al. (32) | Japan | MDD: 10 HC: 10 | NA | NA | NA | NA | 52-Channel ETG- 4000 (Hitachi) | Emotional Stroop task | T | – HC showed \uparrow in oxy-Hb in the fronto-temporal regions as opposed to MDD. |
| Koike et al. (33) | Japan | MDD: 405 HC: 369 | NA | NA | NA | NA | fNIRS | VFT | PF | – The intensity of signals was smaller in MDD. – The duration of time taken to complete the task was later in MDD. |
| Azechi et al. (34) | Japan | MDD: 30 HC: 30 | NA | NA | NA | NA | 2-Channel NIRS | VFT, TOH, SBT Stroop Task | F | – Task performances of the VFT and TOH were \downarrow in MDD than in HC. |
| Matsuo et al. (35) | Japan | MDD: 10 (5/5) HC: 10 (6/4) | MDD: 62.2 \pm 4.8 HC: 58.7 \pm 5.8 | DSM-IV (MINI) | HAMD | Only 4 patients had medication | 24-Channel ETG- 100 (Hitachi) | VFT WRT CO ₂ inhalation | PF | – During cognitive task, there was \downarrow activation of PF cortex in MDD. – Negative association between \downarrow PF activation while performing the cognitive task and degree of hyperintensity in the periventricular region or left F cortex in MDD. – CO ₂ inhalation causing vasomotor reactivity was \downarrow in MDD than HC. |
| Ohta et al. (36) | Japan | MDD: 17 (5/12) HC: 24 (12/12) | MDD: 42.8 \pm 18.2 HC: 36.2 \pm 16.5 | DSM-IV (MINI) | HAMD | All on medication | 52-Channel ETG- 4000 (Hitachi) | WFT | F | – Improvement of oxy-Hb values in the bilateral F cortices, – MDD showed attenuated \uparrow in oxy-Hb while doing WFT in the bilateral F regions. – Hypofrontality in MDD is most notable in the left medial inferior F lobe. |

(Continued)

TABLE 2 | Continued

| Source | Country | Sample size (male/female) | Age (mean ± standard deviation) | Diagnostic criteria (instrument) | Psychopathology measure | Medication | NIRS device | Paradigm | Brain area | Main findings |
|--------------------------|---------|---|--|--|--|--|--|--|---------------|---|
| Suto et al. (18) | Japan | MDD: 10 (9/1) HC: 16 (12/4) | MDD: 47.9 ± 12.8 HC: 42.9 ± 4.6 | DSM-IV | HAMD | All on medication | 24-Channel ETG- 100 (Hitachi) | WFT Right-finger- tapping task | F, T | <ul style="list-style-type: none"> – Enhancement of oxy-Hb was not as notable on the right-sided channels in MDD. – During first half of task period, MDD had ↓ oxy-Hb increase than HC. ↑ were seen in the anterior lower T and lower F channels. – Finger tapping tasks in MDD patients caused ↑ in oxy-Hb compared to HC. |
| Shimodera et al. (37) | Japan | UNI: 39 (19/20) BP: 14 (7/7) HC: 24 (13/11) | UNI: 56.9 ± 12.6 BP: 51.4 ± 14.0 HC: 40.9 ± 10.6 | DSM-IV-TR | HAMD | All on medication | 52-Channel OMM- 3000/16 (Shimadzu) | VFT | PF | <ul style="list-style-type: none"> – Both UNI and BP showed ↓ area under curves (AUCs) than HC. – MDD showed significantly ↓ weighted center (WC) than BP or HC. |
| Ma et al. (38) | China | Menopausal MD: 30 (0/30) MDD: 30 (0/30) HC: 30 (0/30) | MD: 51.17 ± 6.06 MDD: 37.50 ± 10.60 HC: 34.83 ± 8.77 | DSM-IV | HAMD | Only 20 patients took medication | 45-Channel FOIRE- 3000 (Shimadzu) | VFT | PF | <ul style="list-style-type: none"> – MD and MDD both showed ↓ oxy-Hb activation in bilateral DLPFC than HC, but involving different channels. – Atypical hemodynamics of the left DLPFC can discriminate HC from MDD using NIRS. |
| Takizawa et al. (39) | Japan | UNI: 153 (76/ 77) BP: 134 (65/69) HC: 590 (276/ 314) | UNI: 43.8 ± 12.7 BP: 44.0 ± 14.9 HC: 43.9 ± 15.7 | DSM-IV SCID | HAMD | 10 drug-free patients with UNI | 52-Channel ETG- 4000 (Hitachi) | VFT | PF, T | <ul style="list-style-type: none"> – NIRS can differentiate HC from patients, and differentiate UNI from BP. |
| Liske et al. (40) | Germany | MDD: 41 HC: 46 | NA | NA | SIMS | NA | NA | Motor task– pressing button | P | <ul style="list-style-type: none"> – In HC, a noteworthy contrast in the NIRS signal resulting from the left P brain regions but this was weaker in MDD. |
| Zhu et al. (41) | China | UNI: 35 (11/24) BP: 39 (19/20) HC: 36 (18/18) | UNI: 35.9 ± 13.2 BP: 37.0 ± 12.9 HC: 33.6 ± 10.3 | DSM-IV-TR (MINI) | HAMD | All on medication | 52-Channel ETG- 4000 (Hitachi) | 1-Back version of the n-back WMT | PF, T | <ul style="list-style-type: none"> – Compared to HCs, UNI and BP ↓ activation of oxy-Hb in the inferior PF region during WMT. – Distinct prefrontal activation patterns in the Broca's area and left frontopolar region, underline BD and UD. |
| Matsubara et al. (42) | Japan | UNI: 16 (8/8) BP: 16 (8/8) HC: 20 (10/10) | UNI: 45.4 ± 12.2 BP: 44.1 ± 17.5 HC: 41.4 ± 8.5 | DSM-IV-TR (MINI) | HAMD | All on medication | 52-Channel ETG- 4000 (Hitachi) | Emotional Stroop task | PF | <ul style="list-style-type: none"> – While doing the threat task, depressed patients revealed ↑ oxy-Hb in the left middle frontal region. – When performing happy task, depressed patients, did not display any meaningful changes in oxy-Hb. |
| Akashi et al. (43) | Japan | MDD: 52 (32/ 20) Subdivided into with/without discrepancy HC: 48 (21/27) | MDD: 41.8 ± 12.7 HC: 38.9 ± 9.5 | DSM-IV (MINI) | Structured interview guide for HAMD (SIGH-D) | Most of the patients taking medications | 52-Channel ETG- 4000 (Hitachi) | VFT | F, T | <ul style="list-style-type: none"> – In the FT regions, upsurge in mean oxy-Hb were ↓ in MDD than in HC. |
| Gao et al. (44) | China | MDD: 27 (7/20) HC: 24 (11/13) | MDD: 40.78 ± 13.42 HC: 43.13 ± 11.28 | DSM-IV | HAMD | All were medication free for at least 4 weeks | CW-NIRS | Facial emotion recognition | PF | <ul style="list-style-type: none"> – Hemodynamic variations between the bilateral PF cortex and left PF cortex may provide dependable predictors for diagnosing depression |

(Continued)

TABLE 2 | Continued

| Source | Country | Sample size (male/female) | Age (mean ± standard deviation) | Diagnostic criteria (instrument) | Psychopathology measure | Medication | NIRS device | Paradigm | Brain area | Main findings |
|---------------------------------------|---------|---|---|--|----------------------------|---|--|--|---------------|---|
| Schecklmann et al. (45) | Germany | UNI: 16 (9/7) BP: 14 (3/11) HC: 15 (7/8) | UNI: 43.4 ± 9.8 BP: 40.8 ± 10.2 HC: 40.9 ± 8.0 | ICD-10 | BDI-II | All on medication | 52-Channel ETG- 4000 (Hitachi) | WMT | PF, F | Results discovered unspecific deficits that inhibited discrimination between bipolar and unipolar depression in domains of working memory. |
| Tomioka et al. ^{3,4} (46) | Japan | MDD: 25 (3/22) HC: 62 (14/48) | MDD: 51.9 ± 16.6 HC: 51.7 ± 17.2 | DSM-IV | HAMD | Medication- naive | 52-Channel ETG- 4000 (Hitachi) | VFT | PF, T | – MDD showed ↓ oxy-Hb values when comparing to with HC in the bilateral F and T cortices at baseline. – Hypofrontality response to VFT may represent a potential trait marker for depression |
| Ohtani et al. ^{3,4} (47) | Japan | UNI: 10 (4/6) BP: 18 (9/9) HC: 14 (7/7) | UNI: 39.2 ± 12.1 BP: 39.7 ± 9 HC: 33.6 ± 8.3 | DSM-IV-TR | HAMD | All on medication | 52-Channel ETG- 4000 (Hitachi) | VFT | PF, T | – UNI and BP showed ↓ activation than HCs in the bilateral ventrolateral PF cortex and the anterior part of the T cortex. |
| Masuda et al. ^{3,4} (48) | Japan | MDD: 47 Response group to SSRIs: 28 (15/13) Nonresponse group: 19 (6/ 13) HC: 63 (35/28) | Response group: 48.9 ± 2.9 Nonresponse group: 43.2 ± 3.3 HC: 41.7 ± 1.4 | DSM-IV-TR | HAMD | Medication- naive | 47-Channel ETG- 7100 (Hitachi) | VFT | PF, T | – In FT region, hemodynamic responses were ↓ in patients with response and nonresponse than in HC prior to treatment. – In the medial F region, hemodynamic responses were ↑ in patients with response to prior treatment. |
| Feng et al. ⁴ (49) | China | MDD: 15 (7/8) HC: 15 (6/9) | MDD: 30.93 ± 13.47 HC: 30.87 ± 10.11 | DSM-5 SCI | HAMD | NA | 45-Channel FOIRE- 3000 (Shimadzu) | VFT | PF | – After the music treatment, average active oxy-Hb values of some channels were ↑ in both HC and MDD. – After music therapy, patients with MDD demonstrated substantial activation in the OFC, DLPFC, and VMPFC. |
| Hirano et al. ^{3,4} (50) | Japan | MDD: 30 (11/ 19) HC: 108 (45/ 63) | MDD: 59.4 ± 14.2 HC: 58.9 ± 13 | ICD-10 | MADRS QIDS-SR | All on medication | 52-Channel ETG- 4000 (Hitachi) | VFT | PF, T | – MDD exhibited ↓ oxy-Hb values in the bilateral F cortex while doing VFT than HC. |
| Downey et al. ⁴ (51) | UK | MDD: 18 HC: 51 | NA | NA | NA | NA | MiniNTS 4 detectors/24 sources (UCL) | VFT, N-back working memory tasks | PF | – MDD had ↓ bilateral PF cortex oxy-Hb responses to VFT unlike HC, and this was additionally ↓ after 4 ECT sessions. – On WM task, MDD showed PF cortex inhibition at baseline and a different time course of oxy- and deoxy-Hb following 4 ECT. |
| Rosenbaum et al. ³ (52) | Germany | MDD: 60 HC: 24 | MDD: 40 ± 14.79 HC: 33 ± 11.45 | DSM-IV (SCI) | PHQ-9 MADRS | 32% of patients treated with antidepressant medication | 52-Channel ETG- 4000 (Hitachi) | 7-min resting phase, VAS, rumination response scale | P | – MDD as compared to HC, showed ↓ functional connectivity in parts of the DMN. |

(Continued)

TABLE 2 | Continued

| Source | Country | Sample size (male/female) | Age (mean ± standard deviation) | Diagnostic criteria (instrument) | Psychopathology measure | Medication | NIRS device | Paradigm | Brain area | Main findings |
|--|---------|---|--|--|----------------------------|--|---|---|--------------------|---|
| Kondo et al. ³ (53) | Japan | MDD: 25 (17/8) HC: 25 (18/7) | MDD: 36 ± 8.91 HC: 34.1 ± 10.1 | DSM-IV-TR SCI | HAMD | All medicated with antidepressants | 44-Channel ETG- 4000 (Hitachi) | Image-recall task | F, T | – The oxy-Hb in HC was ↑ compared to MDD in bilateral F region. – The severity of depression was related to ↓ in oxy-Hb in left F lobe. |
| Zhu et al. ³ (54) | China | Affective disorder (AD): 28 (8/20) HC: 30 (21/9) | AD: 23.32 ± 5.01 HC: 23.60 ± 2.03 | DSM-IV | SDS | 13 patients were medicated. 15 free of any medicine | 42-Channel FOIRE-3000 (Shimadzu) | Resting state measurements | F | – Relative to HC, AD demonstrated ↓ in intraregional and symmetrical interhemispheric connectivity in the PFC, revealed ↓ locally functional connectivity in the right IFG, and ↓ long-distance connectivity concerning the bilateral IFG. |
| Okada et al. ³ (55) | Japan | MDD: 36 (24/12) HC: 36 (21/12) | MDD (male): 23.3 ± 2.5 MDD (female): 21.3 ± 1.1 HC (male): 23.9 ± 2.4 HC (female): 23.6 ± 2.1 | DSM-III-R | HAMD | 13 patients received antidepressant medication. 23 were medication free for a minimum of 3 months | Multichannel near- IR spectrophotometry (NIRS) | MDT | LT, RT brain | – Nearly half of patients revealed a “nondominant hemisphere response pattern” – The supposedly “nondominant” hemisphere may convert to being dominant during the depression |
| Uemura et al. ³ (56) | Japan | MDD: 13 (6/7) HC: 67 (28/39) | MDD: 74.5 ± 5.8 HC: 73.8 ± 5.3 | NA | GDS | All subjects were medicated. | 8-Channel FOIRE- 3000; (Shimadzu) | Trail-making test, part B (TMT-B; tablet version) | PF | – Oxy-Hb activation when performing the TMT-B was ↓ in MDD in both the right and left PF cortex. – ↓ PF activation in the elderly with depressive symptomatology may cause deterioration in executive function. |
| Kinou et al. ³ (57) | Japan | MDD: 32 (15/ 17) HC: 32 (15/17) | MDD: 44.8 ± 9.8 HC: 45.7 ± 13.5 | DSM-IV | H | All, except 3 patients | 52-Channel ETG- 4000 (Hitachi) | VFT | PF | – MDD revealed ↓ oxy-Hb activation during the task. |
| Yamagata et al. ³ (58) | Japan | Early-onset depression (EOD): 11 (2/9) Late-onset (LOD): 12 (3/9) HC: 13 (8/5) | EOD: 68.4 ± 5.6 LOD: 70.2 ± 1.9 HC: 70.3 ± 4.4 | DSM-IV | HAMD | All patients were taking one prescribed antidepressant. | 52-Channel ETG- 4000 (Hitachi) | WFT | F, T | – HC demonstrated ↑ in oxy-Hb vs. surges in oxy-Hb being minimally ↓ in EOD and extremely ↓ in LOD, bilaterally throughout the F cortices and T areas. – Attenuated activation in the left lateral PF and T areas may help to differentiate LOD and EOD. |
| Noda et al. ³ (59) | Japan | MDD: 30 (14/ 16) HC: 30 (14/16) | MDD: 36.7 ± 11.6 HC: 35.1 ± 9.4 | DSM-IV (SCI) | GRID-HAMD | All patients medicated with antidepressants. | 52-Channels ETG- 4000 (Hitachi) | VFT | F, T | – Oxy-Hb increases while performing task was meaningfully ↓ in MDD. – ↓ Right F-T activation on NIRS during VFT is related to the MDD severity. |
| Nishizawa et al. ³ (60) | Japan | MDD: 14 (7/7) HC: 20 (13/7) | MDD: 38.2 ± 12.9 HC: 29.0 ± 5.7 | DSM-IV-TR (SCID) | HAMD | All taking antidepressants except for four patients. | 22-Channel ETG- 4000 (Hitachi) | Emotional Stroop task | F, T | – Hyperactivated oxy-Hb was witnessed in the left F cortex on contact to unfavorable stimuli, but no noteworthy dissimilarity was established between MDD and HC on exposure to favorable stimuli. |

(Continued)

TABLE 2 | Continued

| Source | Country | Sample size (male/female) | Age (mean ± standard deviation) | Diagnostic criteria (instrument) | Psychopathology measure | Medication | NIRS device | Paradigm | Brain area | Main findings |
|-------------------------------------|---------|---|--|--|----------------------------|---|--------------------------------------|--|---------------|--|
| Akiyama et al. ³ (61) | Japan | MDD: 177 (73/ 104) HC: 50 (40/10) | MDD: 47.2 ± 15.1 HC: 32.7 ± 7.5 | DSM-IV-TR | HAMD | All patients were on antidepressants | 52-Channel ETG- 4000 (Hitachi) | VFT | PF, T | – Significant hypoactivation in bilateral F-T regions was observed in MDD compared with HC. |
| Ohi et al. ³ (62) | Japan | UNI: 26 (17/9) BP: 22 (13/9) HC: 51 (33/18) | UNI: 41.1 ± 12.7 BP: 39.9 ± 12.5 HC: 35.7 ± 11.9 | DSM-V | HAMD | Chlorpromazine equivalents of total antipsychotics | 52-Channel ETG-4000 (Hitachi) | VFT | PF | – UNI and BP groups had ↓ PF activity than HC. – Patients with and without family history had ↓ PF activity than HC subjects. – Demonstrate connection of more serious PF dysfunction with higher genetic loading for disease. |
| Takei et al. ³ (63) | Japan | UNI: 29 (14/15) BP: 31 (14/17) HC: 31 (11/20) | UNI: 34.5 ± 9.0 BP: 34.9 ± 6.6 HC: 33.6 ± 10.0 | DSM-IV | HAMD | Nearly all patients were on medication. | 52-Channel ETG-4000 (Hitachi) | Conversation task and control task | F, T | – Both UNI and BP showed ↓ of continuous activation in the left DLPFC and left FPC, and decreased rapid change in bilateral FPC activation. – F activation while conversing ↓ in equally in UNI and BP. – Pathophysiological character of UNI and BP are reflected in continuous activation and rapid change change Impaired adaptive ability in UNI may be related to ↓ amount of rapid change in right FPC. |
| Pu et al. ³ (64) | Japan | Late-onset depression (LOD): 24 (6/18) HC: 30 (14/16) | LOD: 72.3 ± 5.5 HC: 72.0 ± 4.7 | DSM-IV (MINI) | BDI HAMD | Antidepressant- naive | 52-Channel ETG-4000 (Hitachi) | VFT | PF, T | – LOD had ↓ activation in both PF and superior T cortices than HC. – ↓ frontopolar cortical activation was linked with social functioning impairment in LOD. |
| Tsuji et al. ³ (65) | Japan | MDD: Suicide attempters (SAs): 30 (8/22) Nonattempters (NAs): 38 (16/ 22) HC: 40 (15/25) | MDD (SAs): 37.6 ± 10.0 MDD (NAs): 38.8 ± 9.7 HC: 38.2 ± 10.5 | DSM-IV (MINI) | HAMD | All on medication. | 52-Channel ETG- 4000 (Hitachi) | VFT | F, T | – MDD had significantly ↓ activation in the bilateral FT regions compared to HCs. – SAs demonstrated ↓ hemodynamic response in the left precentral gyrus than HCs and NAs. – Aggression and hopelessness were negatively associated with hemodynamic responses in the right middle F gyrus in SAs but not in HCs and NAs. |
| Pu et al. ³ (66) | Japan | Late-onset depression (LOD): 36 (9/27) HC: 35 (11/24) | LOD: 71.8 ± 5.1 HC: 70.9 ± 4.3 | DSM-IV (MINI) | BDI HAMD | Antidepressant- naive | 52-Channel ETG- 4000 (Hitachi) | Working memory (WM) task | PF, T | – LOD was correlated with ↓ PF and T activation when comparing with HC. – Hemodynamic response in PF and T regions when performing WM task could correlate with social functioning in LOD. |
| Tsuji et al. ³ (67) | Japan | MDD with melancholia (MDD-MF): 30 (15/15) | MDD-MF: 42.2 ± 11.8 MDD-NMF: 40.6 ± | DSM-IV (MINI) | HAMD | All on medication | 52-Channel ETG- 4000 (Hitachi) | VFT | F, T | – Both MDD groups demonstrated ↓ hemodynamic responses in the F-T regions. |

(Continued)

TABLE 2 | Continued

| Source | Country | Sample size (male/female) | Age (mean ± standard deviation) | Diagnostic criteria (instrument) | Psychopathology measure | Medication | NIRS device | Paradigm | Brain area | Main findings |
|---------------------------------------|---------|--|--|--|----------------------------|--------------------------------|---|---|---------------|---|
| Liu et al. ³ (68) | China | MDD without -melancholia (MDD-NMF): 52 (18/34) HC: 68 (32/36) MDD: 30 (12/ 18) HC: 30 (16/14) | 11.7 HC: 40.5 ± 10.6 MDD: 38.38 ± 12.8 HC: 33.2 ± 10.5 | DSM-IV-TR | HAMD | Free of medication | 52-Channel FOIRE- 3000 (Shimadzu) | VFT | PF | <ul style="list-style-type: none"> – ↓ Activation in lateral and lower PFC in MDD. – Antero-medial PFC and bilateral PFC were correlated with the degree of depressive symptoms. – MDD patients with obsession-compulsion symptoms and anxiety exhibited a PFC ↓ activation state in NIRS. |
| Wang et al. ³ (69) | China | First-episode MDD (fMDD): 36 (15/21) Recurrent MDD: 34 (11/ 23) HC: 37 (22/15) | fMDD: 38.75 ± 13.86 Recurrent MDD: 43.26 ± 13.85 HC: 35.70 ± 11.39 | DSM-IV | HAMD | All were on antidepressants | 52-Channel ETG- 4000 (Hitachi) | VFT | F, T | <ul style="list-style-type: none"> – In comparison with HC and fMDD, chronic MDD had significantly ↓ brain activation over right PF and superior T cortices. – Variation in activations in bilateral F and T regions. HC: more channels in left than right hemisphere; fMDD: more channels in right than left hemisphere; recurrent MDD: only channels in left hemisphere. |
| Tsujii et al. ³ (70) | Japan | MDD with melancholia (MDD-MF): 32 (16/16) MDD without melancholia (MDD-NMF): 28 (15/13) HC: 24 (11/13) | MDD-MF: 40.8 ± 15.3 MDD-NMF: 38.9 ± 11.8 HC: 38.6 ± 9.2 | DSM-IV (MINI) | SIGH-D BDI-II | All on medication | 52-Channel ETG- 4000 (Hitachi) | VFT | F, T | <ul style="list-style-type: none"> – Noteworthy disparities were witnessed in mean oxy-Hb fluctuations of MDD-MF in 25 channels and in those with MDD-NMF in 12 channels compared to HC. |
| Nishida et al. ³ (71) | Japan | MDD: 14 (7/7) HC: 15 (8/7) | MDD: 46.2 ± 11.9 HC: 45.5 ± 10.9 | DSM-IV-TR (MINI) | HAMD | All on medication | 52-Channel ETG- 4000 (Hitachi) | VFT | PF, T | <ul style="list-style-type: none"> – MDD revealed a ↓ oxy-Hb activation than HC, predominantly in ventrolateral PF and T cortex regions. |
| Rosenbaum et al. ³ (72) | Germany | Depressed: 49 Nondepressed: 51 2 participants diagnosed with bipolar disorder and eating disorder. | Depressed: 64.08 ± 7.06 Nondepressed: 64.16 ± 6.14 | NA | BDI GDS | 54% took medication. | 38-Channel ETG-4000 (Hitachi) | Adapted trail- making test (TMT-A, TMT-B, and TMT-C) | F, P | <ul style="list-style-type: none"> – Depressed and nondepressed revealed substantial differences in functional connectivity (FC) while doing the task performance and at rest. – During task performance, depressed patients exhibited ↓ FC in a left frontopolar cortical network, and ↑ FC in a left frontoparietal cortical network at the resting state and altered FC and network organization during different mental states. |

(Continued)

TABLE 2 | Continued

| Source | Country | Sample size (male/female) | Age (mean ± standard deviation) | Diagnostic criteria (instrument) | Psychopathology measure | Medication | NIRS device | Paradigm | Brain area | Main findings |
|--------------------------------|---------|--|--|--|----------------------------|---------------------------|--------------------------------------|----------|---------------|---|
| Pu et al. ³ (73) | Japan | MDD: 67 (29/ 38) MDD with suicidal ideation: 31 (11/20) MDD without suicidal ideation: 36 (18/18) HC: 67 (29/38) | MDD: 58.1 ± 16.0 MDD with suicidal ideation: 57.3 ± 15.7 MDD without suicidal ideation: 58.7 ± 16.5 HC: 58.1 ± 17.8 | DSM-IV (MINI) | HAMD | All on antidepressants | 52-Channel ETG- 4000 (Hitachi) | VFT | PF, T | – Regional hemodynamic changes were considerably ↓ in MDD than in HCs in PF and T regions. |
| Pu et al. ³ (74) | Japan | MDD: 26 (11/ 15) HC: 30 (12/18) | MDD: 47.9 ± 19.2 HC: 50.5 ± 19.7 | DSM-IV-TR (MINI) | BDI HAMD | All on antidepressants | 52-Channel ETG- 4000 (Hitachi) | VFT | PF, T | – Regional hemodynamic changes were appreciably ↓ in MDD than in HC in PF and T regions, and was correlated positively with task-oriented coping (adaptive coping) in the bilateral ventrolateral and dorsolateral prefrontal cortex, and the midline frontopolar and bilateral orbitofrontal cortex regions. |

³This article can also be found in the summary in **Table 3**.

⁴This article can also be found in the summary in **Table 4**.

MDD, major depressive disorder; HC, healthy control; BDI, Beck depression inventory; HAMD, Hamilton depression rating scale; UNI, unipolar depression; BP, bipolar disorder; VFT, verbal fluency task; PF, prefrontal; T, temporal; F, frontal; NA, not available; TOH, Tower of Hanoi; SBT, Sternberg's task; WRT, word repetition task; MINI, Mini-international neuropsychiatric interview; DSM, Diagnostic and Statistical Manual; ICD, International Classification of Disease; SCID, structured clinical interview for DSM-IV; SIMS, structured inventory of malingered symptomatology; WMT, working memory task; IFG, inferior frontal gyrus; MDT, mirror drawing task; DLPFC, dorsolateral prefrontal cortex; PFC, prefrontal cortex; SIGH-D, structured interview guide for the Hamilton depression rating scale.

TABLE 3 | Summary of fNIRS studies correlating cerebral hemodynamic changes with depression symptomatology.

| Source | Country | Sample size (male/female) | Age (mean \pm standard deviation) | Diagnostic criteria instrument | Psychopathology measure for symptomatology | Specific symp- tomatology | NIRS device/no. of channels | Paradigm | Brain area | Main finding |
|--------------------------------------|---------|---|---|--------------------------------------|--|--|--|-----------------------------------|------------|---|
| Kawano et al. (76) | Japan | N: 25 *included other disorders | 44.1 \pm 9.3 | DSM-IV | HAMD | Depressive symptoms | 22-Channel ETG- 4000 (Hitachi) | VFT | F, T | – The integral value of blood flow in frontal lobe was negatively correlated with the degree of depression |
| Onishi et al. ⁴ (77) | Japan | N: 10 (5/5) | 71.0 \pm 6.0 | DSM-IV | HAMD MMSE | Depressive symptoms Cognitive functioning | 48-Channel ETG- 4000 (Hitachi) | Rock, paper, scissors (RPS) | PF | – Negative correlation between ratio of HAMD and oxy-Hb was observed. |
| Hirano et al. ^{2,4} (50) | Japan | N: 30 (11/19) *inclusive of bipolar patients | 59.4 \pm 14.2 | ICD-10 | MADRS QIDS-SR MMSE | Depressive symptoms Cognitive functioning | 52-Channel ETG- 4000 (Hitachi) | VFT | PF, T | – After ECT, the reduction of degree of depression was associated with increase in oxy-Hb values in the right ventrolateral PF cortex. – Changes in oxy-Hb is significantly interrelated with MADRS score \downarrow but not significantly correlated with \downarrow in total QIDS-SR total scores. |
| Masuda et al. ^{2,4} (48) | Japan | N: 47 Response group to SSRIs: 28 (15/ 13) Nonresponse group: 19 (6/ 13) | Response group: 48.9 \pm 2.9 Nonresponse group: 43.2 \pm 3.3 | DSM-IV | POMS STAI DACS | Anxiety symptoms Depressive symptoms | 47-Channel ETG- 7100 (Hitachi) | VFT | PF, T | – Hemodynamic responses in medial F region were significantly greater before treatment in patients with a response to SSRIs than those with no response. |
| Kondo et al. ² (53) | Japan | N: 25 (17/8) | 36 \pm 8.91 | DSM-IV | HAMD | Depressive symptoms | 44-Channel ETG- 4000 (Hitachi) | Image-recall task | F, T | – A noteworthy negative correlation between oxy-Hb and HAMD score was seen in left F region during the unpleasant condition. – \downarrow in oxy-Hb in left F lobe was associated to degree of depression. |
| Koseki et al. (78) | Japan | MDD: 75 (39/ 36) | 39.23 \pm 12.49 | DSM-IV | HAMD ATQ-R NART STAI | Depressive symptoms Automatic thoughts State/trait anxiety | 52-Channel ETG- 4000 (Hitachi) | VFT | PF, T | – Activation in right superior temporal gyrus was associated to deviation to negative of the proportion of negative and positive thought. |
| Zhu et al. ² (54) | China | Affective disorder (AD): 28 (8/20) | 23.32 \pm 5.01 | DSM-IV | SDS | Depressive symptoms | 42-Channel FOIRE-3000 (Shimadzu) | Resting state | F | – Degree of self-reported symptoms of depression was negatively associated with strength of intraregional and symmetrically interhemispheric connectivity in the PFC. |
| Uemura et al. ² (56) | Japan | N: 13 (6/7) | 74.5 \pm 5.8 | NA | GDS MMSE | Depressive symptoms | 8-Channel FOIRE- 3000 (Shimadzu) | TMT-B | PF | – Both oxy-Hb activation in left and right hemisphere were significantly negatively associated with GDS. |
| Noda et al. ² (59) | Japan | N: 30 (14/16) | 36.7 \pm 11.6 | DSM-IV (SCID-I) | GRID-HAMD | Depressive symptoms | 52-Channels ETG- 4000 (Hitachi) | VFT | F, T | – Average increase in oxy-Hb during the task revealed a significant negative association with the HAMD total scores. |

(Continued)

TABLE 3 | Continued

| Source | Country | Sample size (male/female) | Age (mean \pm standard deviation) | Diagnostic criteria instrument | Psychopathology measure for symptomatology | Specific symp- tomatology | NIRS device/no. of channels | Paradigm | Brain area | Main finding |
|---------------------------------------|---------|---|--|--------------------------------------|--|--|--|--------------------------------|------------|---|
| Akiyama et al. ² (61) | Japan | N: 177 (73/ 104) | 47.2 \pm 15.1 | DSM-IV-TR | HAMD PHQ-9 | Depressive symptoms | 52-Channel ETG- 4000 (Hitachi) | VFT | PF, T | – Left lateral F-T activation was significantly \downarrow in the group with depressed mood or anhedonia. |
| Tomioka et al. ^{2,4} (46) | Japan | N: 25 (3/22) | 51.9 \pm 16.6 | DSM-IV | HAMD | Depressive symptoms | 52-Channel ETG- 4000 (Hitachi) | VFT | PF, T | – Patients with MDD who revealed \uparrow baseline oxy-Hb activation while performing VFT in the inferior F and middle T regions showed more improvements in depressive symptoms after being treated. |
| Rosenbaum et al. ² (72) | Germany | N: 49 *inclusive of other disorders | 64.08 \pm 7.06 | NA | BDI GDS | Depressive symptoms Cognitive functioning | 38-Channel ETG-4000 (Hitachi) | TMT-A TMT- B TMT-C | F, P | – Depressed patients revealed \downarrow FC in a left frontopolar cortical network while doing task performance \uparrow FC in a left frontoparietal cortical network at the resting state. |
| Yamagata et al. ² (58) | Japan | Early-onset depression (EOD): 11 (2/9) Late-onset depression (LOD): 12 (3/9) n = 48 | EOD: 68.4 \pm 5.6 LOD: 70.2 \pm 1.9 | DSM-IV | HAMD MMSE | Depressive symptoms | 52-Channel ETG- 4000, NIRS system (Hitachi) | WFT | F, T | – HAMD score exhibited negative correlations with oxy-Hb for two channels. |
| Akashi et al. (79) | Japan | n = 48 | NA | NA | NA | Inhibitory deficit | 47-Channel NIRS | Stop-signal task | F | – MDD showed the variations of brain dysfunctions correlated in the inhibitory controls. |
| Satomura et al. (80) | Japan | Initial (T0)–N: 65 After 1.5 years (T1.5)–N: 45 | 39.8 \pm 11.8 | DSM-IV (SCID-I) | HAMD GAF | Depressive symptoms Global functioning | 52-Channel ETG- 4000 (Hitachi) | VFT | PF, T | – Brain activation in the bilateral MFG and right IFG as calculated by NIRS may differentially denote clinical severity and trait-related anomalies in MDD. |
| Ohtani et al. ^{2,4} (47) | Japan | N: 10 (4/6) | 39.2 \pm 12.1 | DSM-IV | SASS HAMD | Social adaptation Depressive symptoms | 52-Channel ETG- 4000 (Hitachi) | VFT | PF, T | – Longitudinal changes in SASS scores were positively related with magnitude of change in the right VLPFC/aTC activation in MDD group. |
| Pu et al. ² (64) | Japan | Late-onset MDD (LOD): N: 24 (6/18) | 72.3 \pm 5.5 | DSM-IV (MINI) | BDI HAMD SASS MMSE | Depressive symptoms Social functioning | 52-Channel ETG-4000 (Hitachi) | VFT | PF, T | – Average oxy-Hb changes in LOD patients had a significantly positive association with SASS scores. |
| Pu et al. ² (66) | Japan | Late-onset depression (LOD): 36 (9/ 27) | 71.8 \pm 5.1 | DSM-IV (MINI) | BDI HAMD SASS MMSE | Depressive symptoms Social functioning | 52-Channel ETG-4000 (Hitachi) | Working memory (WM) task | PF, T | – Reduced activation in PF and T regions was significantly associated to \downarrow scores on the SASS in patient group and might act as a biological marker of social functioning in LOD patients. |
| Pu et al. ² (74) | Japan | N: 26 (11/15) | 47.9 \pm 19.2 | DSM-IV-TR (MINI) | HAMD BDI CISS | Depressive symptoms Coping styles | 52-Channel ETG-4000 (Hitachi) | VFT | PF, T | – Regional hemodynamic changes were \downarrow in MDD group compared to the control group in PF and T areas, and positively interrelated with task-oriented coping (adaptive coping) in the various PF regions. |

(Continued)

TABLE 3 | Continued

| Source | Country | Sample size (male/female) | Age (mean \pm standard deviation) | Diagnostic criteria instrument | Psychopathology measure for symptomatology | Specific symptomatology | NIRS device/no. of channels | Paradigm | Brain area | Main finding |
|------------------------------------|---------|--|---|--------------------------------|--|--|----------------------------------|------------------------------|------------|--|
| Pu et al. ² (81) | Japan | N: 67, 31 with suicidal ideation and 36 without | 58.1 \pm 16.0 | DSM-IV (MINI) | HAMD | Suicidal ideation | 52-Channel ETG-4000 (Hitachi) | VFT | PF, T | – Hemodynamic variations correlated negatively with the degree of suicidal thinking in OFC, FPC, and DLPFC. |
| Nishida et al. ² (71) | Japan | N: 14 (7/7) | 46.2 \pm 11.9 | DSM-IV-TR (MINI) | HAMD PSQI ESS | Depressive symptoms Sleep quality | 52-Channel ETG-4000 (Hitachi) | VFT | PF, T | – Significant negative correlations between average oxy-Hb variations during VFT and PSQI scores. – Mean oxy-Hb changes showed no significant correlations with ESS scores. – No significant association between average oxy-Hb variations during the task and sleep variables. |
| Tsuji et al. ² (70) | Japan | MDD with melancholia (MDD-M): 32 (16/16) MDD without melancholia (MDD-NM): 28 (15/13) | MDD-MF: 40.8 \pm 15.3 MDD-NMF: 38.9 \pm 11.8 | DSM-IV (MINI) | BDI-II SIGH-D GAF | Depressive symptoms Psychomotor retardation | 52-Channel ETG-4000 (Hitachi) | VFT | F, T | – No meaningful associations between average oxy-Hb changes and GAF/HAMD/BDI-II total scores. – Psychomotor retardation on HAMD indicated noteworthy positive correlation with average oxy-Hb changes in right T areas in MDD-MF but revealed significant negative association with average oxy-Hb changes in the middle to left T region in MDD-NMF. |
| Tsuji et al. ² (67) | Japan | MDD with melancholia (MDD-M): 30 (15/15) MDD without melancholia (MDD-NM): 52 (18/34) | MDD-M: 42.2 \pm 11.8 MDD-NM: 40.6 \pm 11.7 | DSM-IV (MINI) | HAMD SF-36 | Depressive symptoms Quality of life | 52-Channel ETG-4000 (Hitachi) | VFT | F, T | MDD-M patients reveal qualitatively dissimilar prefrontal dysfunction patterns correlated with emotional role functioning as compared to MDD-NM patients. |
| Wang et al. ² (69) | China | First-episode MDD (fMDD): 36 (15/21) Recurrent MDD (rMDD): 34 (11/23) | fMDD: 38.75 \pm 13.86 rMDD: 43.26 \pm 13.85 | DSM-IV | HAMD | Depressive symptoms | 52-Channel ETG-4000 (Hitachi) | VFT | F, T | -rMDD group had \downarrow increases in oxy-H comparing to the fMDD group. |
| Liu et al. ² (68) | China | N: 30 (12/18) | 38.38 \pm 12.8 | DSM-IV-TR | HAMD HAMA Y-BOCS | Depressive, anxiety, obsessive-compulsive symptoms | 52-Channel FOIRE-3000 (Shimadzu) | VFT | PF | – Average oxy-Hb changes revealed significant positive correlation with HAMD scores. – No statistical relationship was witnessed on the degree of obsessive-compulsive symptoms. |
| Rosenbaum et al. ² (52) | Germany | N: 60 | 40 \pm 14.79 | DSM-IV (SCI) | PHQ-9 MADRS | Depressive symptom State and trait | 52-Channel ETG-4000 (Hitachi) | RRS VAS Self-report on | P | – Subjects who are depressed revealed \downarrow functional connectivity in parts of the DMN compared to HCs. – mind-wandering revealed positive |

(Continued)

TABLE 3 | Continued

| Source | Country | Sample size (male/female) | Age (mean ± standard deviation) | Diagnostic criteria instrument | Psychopathology measure for symptomatology | Specific symp- tomatology | NIRS device/no. of channels | Paradigm | Brain area | Main finding |
|-----------------------------------|---------|---|---|--------------------------------------|--|---|---|--|----------------------------------|---|
| Okada et al. ² (55) | Japan | N: 36 (24/12) | Male: 23.3 ± 2.5 Female: 21.3 ± 1.1 | DSM-III-R | HAMD | Depressive symptoms | Multichannel near- IR spectrophotometry | Mirror drawing task (MDT) | Left and right hemispheres | associations, whereas rumination was negatively associated with FC in the cortical parts of the DMN – Nearly half of the patients revealed a “nondominant hemisphere response pattern,” which was not witnessed in normal subjects during the MDT. – During the course of depression, the supposedly “nondominant” hemisphere may become dominant. – Lower Global Functioning scores were correlated with ↓ hemodynamic responses. |
| Kinou et al. ² (57) | Japan | N: 32 (15/17) *included HC, MDD, schizophrenia | 44.8 ± 9.8 | DSM-IV | HAMD Global assessment of functioning | Depressive symptoms | 52-Channel ETG-4000 (Hitachi) | VFT | PF | – Prompt change in activation was positively associated with GAF scores in the MDD patients. |
| Takei et al. ² (63) | Japan | N: 29 (14/15) | 34.5 ± 9.0 | DSM-IV | HAMD GAF | Depressive symptoms Cognitive functioning | 52-Channel ETG-4000 (Hitachi) | Conversation task and control task | F, T | – Illustration of the association of significantly more severe PF dysfunction with higher genetic loading in major mental illnesses. |
| Ohi et al. ² (62) | Japan | N: 26 (17/9) | 41.1 ± 12.7 | DSM-5 | HAMD Clinical interview for family history | Depressive symptoms Family history/ familial loadings | 52-Channel ETG-4000 (Hitachi) | VFT | PF | – SAs revealed smaller hemodynamic response in the left precentral gyrus than NAs and HCs. – Hemodynamic responses in the right middle F gyrus were negatively correlated with aggression and hopelessness in SAs. |
| Tsuji et al. ² (65) | Japan | MDD: suicide attempters (SAs): 30 (8/ 22) nonattempters (NAs): 38 (16/ 22) | MDD (SAs): 37.6 ± 10.0 MDD (NAs): 38.8 ± 9.7 | DSM-IV (MINI) | HAMD Barratt impulsiveness scale Buss– Perry aggression questionnaire Beck hopelessness scale | Depressive symptoms Impulsivity Aggression Hopelessness | 52-Channel ETG-4000 (Hitachi) | VFT | F, T | |

²This article can also be found in the summary in **Table 2**.

⁴This article can also be found in the summary in **Table 4**.

MDD, major depressive disorder; HC, healthy control; HAMD, Hamilton depression rating scale; VFT, verbal fluency task; PF, prefrontal; T, temporal, F, frontal; NA, not available; MINI, Mini-international neuropsychiatric interview; DSM, Diagnostic and Statistical Manual; ICD, International Classification of Disease; MMSE, Mini-Mental State Examination; SASS, school and staffing survey; POMS, profile of mood states; STAI, state-trait anxiety inventory; MADRS, Montgomery–Asberg depression rating scale; QIDS-SR, quick inventory of depressive symptomatology-self report; DSRS, dementia severity rating scale; HAMA, Hamilton anxiety rating scale; GDS, geriatric depression scale; PHQ, patient health questionnaire; BDI, Beck depression inventory; TMT, trail-making test; SCID, structured clinical interview for DSM-IV; IFG, inferior frontal gyrus; MFG, medial frontal gyrus; DLPFC, dorsolateral prefrontal cortex; OFC, orbitofrontal cortex; FPC, frontopolar cortex; PSQI, Pittsburgh sleep quality index; ESS, Epworth sleepiness scale; SIGH-D, structured interview guide for the Hamilton depression rating scale; Y-BOCS, Yale–Brown obsessive–compulsive scale; RRS, rumination response scale; VAS, visual analog scale; DMN, default mode network; MDT, mirror drawing task; GAF, global assessment of functioning; ATQR, Automatic thoughts questionnaire-revised.

TABLE 4 | Summary of fNIRS studies assessing antidepressant/treatment response.

| Source. | Country | Sample size (male/female) | Age (mean ± standard deviation) | Diagnostic criteria instrument | Psychopathology measure for treatment response | Treatment outcome | Medication (mg)/ treatment | NIRS device/no. of channels | Duration/no. of follow-up | Paradigm | Brain area | Main finding |
|---------------------------------------|---------|---|---|--------------------------------------|---|--|--|--|--|--------------------------------------|---------------|--|
| Tomioka et al. ^{2,3} (46) | Japan | MDD: 25 (3/22) | 51.9 ± 16.6 | DSM-IV | HAMD | Depressive symptoms | Imipramine (118.7 ± 67.3) | 52-Channel ETG-4000 (Hitachi) | 12 weeks | VFT | PF, T | – NIRS signals before initiation of treatment could foretell patients' clinical response upon being treated |
| Onishi et al. ³ (77). | Japan | MDD: 10 (5/5) | 71.0 ± 6.0 | DSM-IV | HAMD MMSE | Depressive symptoms Cognitive functioning | Mianserin (2), Sodium valproate (2), Paroxetine (2), Lithium (2), Amoxapine (1), Olanzapine (3), Milnacipran (5), Maprotiline (1) | 48-Channel ETG-4000 (Hitachi) | 2 follow-ups (1 day following improvement in depressive symptoms after treatment, and another day >4 weeks later) | Rock, paper, scissors (RPS) | PF | – The more left PF cortical activity tended to ↑, symptoms of depression ↓ |
| Ohtani et al. ^{2,3} (47) | Japan | MDD: 10 (4/6) | 39.2 ± 12.1 | DSM-IV | HAMD SASS | Social functioning | Chlorpromazine (1), Imipramine (3), Diazepam (4) | 52-Channel ETG-4000 (Hitachi) | 6 months | VFT | PF, T | – Longitudinal variations in SASS results were correlated positively with degree of change in the right ventrolateral PF cortex and the anterior part of the T cortex activation in MDD. |
| Yamagata et al. (82) | Japan | MDD: 11 (5/6) | 36.3 ± 11.2 | DSM-IV | HAMD | Depressive symptoms | Sertraline; week 4: (29.5 ± 10.1), week 8: (61.4 ± 20.4), week 12: (65.9 ± 23.1) | 52-Channel ETG-4000 (Hitachi) | 12 weeks, 3 follow-ups (weeks 4, 8, 12) | VFT | PF, T | – NIRS may be a biological marker in MDD patients for predicting clinical response to Sertraline. |
| Masuda et al. ^{2,3} (48) | Japan | MDD: 47 Response group to SSRIs: 28 (15/13) Nonresponse group: 19 (6/13) MDD: 15 (7/8) | Response (48.9 ± 2.9) Nonresponse (43.2 ± 3.3) | DSM-IV | POMS STAI DACS | Overall functioning for response Group | Escitalopram (33), Paroxetine (7), Sertraline (5), Fluvoxamine (2) | 47-Channel ETG-7100 (Hitachi) | 12 weeks, weekly or biweekly follow-up | VFT | PF, T | Response to SSRI in MDD is predicted by different hemodynamic activities in the frontotemporal cortex. |
| Feng et al. ² (49) | China | MDD: 15 (7/8) | 30.9 ± 13.5 | DSM-V | HAMD | Depressive symptoms | Music therapy, either “creative” (composing music) or “receptive” (listening to music) | 45-Channel FOIRE-3000 (Shimadzu) | 10 days, one session (60 min) a day | VFT, | PF | Music therapy could activate frontal cortex areas to improve mood and cognitive abilities |
| Aoki et al. (83) | Japan | MDD: 2 (1/1) | 24.0 ± 2.0 | ICD-10 | NA | NA | Animal-assisted therapy (AAT) with medication | 42-Channel FOIRE-3000 (Shimadzu) | Pretest and posttest design | VFT | PF | AAT helps to stimulate prefrontal activity in MDD and the effects of AAT can be evaluated by NIRS |
| Hirano et al. ^{2,3} (50) | Japan | MDD = 30 (11/19) | 59.4 ± 14.2 | ICD-10 | MADRS QIDS-SR MMSE | Reduction in MADRS and QIDS- | Electroconvulsive therapy (ECT) | 52-Channel ETG-4000 (Hitachi) | 3x per week, till stable response | VFT | PF, T | Acute therapeutic effects of ECT on MDD patients is correlated to |

(Continued)

TABLE 4 | Continued

| Source. | Country | Sample size (male/female) | Age (mean \pm standard deviation) | Diagnostic criteria instrument | Psychopathology measure for treatment response | Treatment outcome | Medication (mg)/treatment | NIRS device/no. of channels | Duration/no. of follow-up | Paradigm | Brain area | Main finding |
|---------------------------------|------------|---------------------------|-------------------------------------|--------------------------------|--|--|---|--|------------------------------------|----------------------------------|------------|--|
| | | | | | | SR scores (improved functioning) | | | | | | recovery from abnormal functional responses to cognitive tasks in frontal brain regions. |
| Takamiya et al. (84) | Japan | MDD: 33 (17/16) | 46.4 \pm 11.7 | DSM-IV | HAMD | Differences between low-dose/high-dose groups in HAMD scores | High-dose group (> 1 defined daily dose, N = 10), low-dose group (< 1 defined daily dose, N = 23) | 52-Channel ETG-4000 (Hitachi) | Cross-sectional study | VFT | PF, T | The dose-dependent influence of antidepressants on NIRS signals should be considered while deciphering NIRS data. |
| Shinba et al. (85) | Japan | MDD: 15 (11/4) | 45.4 \pm 10.8 | DSM-IV | MADRS | Improved functioning | Transcranial magnetic stimulation (TMS), Fluvoxamine (89.6 \pm 85.8) | NIRO-3000 (Hamamatsu) | 6 weeks, 5 sessions a week | NA | PF | Maintenance of frontal activation [measured by frontal hemoglobin concentration (fHbC)] during TMS stimulation is related to effectiveness of treating MDD patients. |
| Usami et al. (86) | Japan | MDD: 10 (1/9) | 12.9 \pm 0.9 | DSM-IV | DSRS | Depressive symptoms Global functioning | NA | 2-Channel Spectratech | 6 weeks | VFT | PF | Concentration of oxy-Hb could be utilized as a state marker for changes in depressed children. |
| Payzieva & Maxmudova (87) | Uzbekistan | MDD: 5 | NA | NA | NA | NA | NA | OxyPrem (BORL, Switzerland) | Pretest and posttest design | Mental arithmetic task | PF | Computerized cognitive exercises may help in improve cognition of MDD patients and NIRS can be used to monitor cognitive functions. |
| Pu et al. (81) | Japan | MDD: 29 (7/22) | 72.4 \pm 5.7 | DSM-IV | HAMD SASS | Depression symptoms Social functioning | Paroxetine (10–40 mg, N = 15), Milnacipran (50–150 mg, N = 14) | 52-Channel ETG-4000 (Hitachi) | 8 weeks | VFT | PF, T | Social functioning improvements were superior in late-onset depression with initial \downarrow NIRS activation in the right ventrolateral PF area. |
| Downey et al. ² (51) | UK | MDD: 18 | NA | NA | NA | NA | Ketamine | 4 detectors and 24 sources MiniNTS (UCL) | NA | VFT | PF | PF cortical responses appear to be \downarrow in the severely depressed and additionally suppressed by ECT treatment |
| Schiffer et al. (88) | US | MDD: 10 (5/5) | 35.1 \pm 7.1 | DSM-IV | HAMD HAMA | Depression and anxiety symptoms | 4-min near-infrared (NIR) light photobiomodulation | INVOS system (Somanetics) | 4 weeks, 2 follow-ups (weeks 2, 4) | Lateral visual field stimulation | PF | NIR-PBM may have uses for depression treatment |

(Continued)

TABLE 4 | Continued

| Source | Country | Sample size (male/female) | Age (mean \pm standard deviation) | Diagnostic criteria instrument | Psychopathology measure for treatment response | Treatment outcome | Medication (mg)/ treatment | NIRS device/no. of channels | Duration/no. of follow-up | Paradigm | Brain area | Main finding |
|---------------------------|---------|------------------------------|---|--------------------------------------|---|------------------------|---|-----------------------------------|---|--|---------------|--|
| Eschweiler et al. (89) | Germany | MDD: 12 (4/8) | 57.0 \pm 8.0 | DSM-IV | HAMD BDI | Depressive symptoms | (PBM) treatment to left/right forehead Repetitive transcranial magnetic stimulation (rTMS) | Four-site NIRS | 4 weeks, 4 follow-ups (weeks 1, 2, 3, 4) | Arithmetic and mirror- tracing tasks | PF | Low local hemodynamic responses predict clinical benefits of rTMS. |

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³This article can also be found in the summary in **Table 3**.

MDD: major depressive disorder; HC, healthy control; HAMD, Hamilton depression rating scale; VFT, verbal fluency task; PF, prefrontal; T, temporal; F, frontal; NA, not available; MINI, Mini-International Neuropsychiatric Interview; DSM, Diagnostic and Statistical Manual; ICD, International Classification of Diseases; MMSE, Mini-Mental State Examination; SASS, school and staffing survey; POMS, profile of mood states; STAI, state-trait anxiety inventory; MADRS, Montgomery-Asberg depression rating scale; QIDS-SR, quick inventory of depressive symptomatology-self report; DSRS, dementia severity rating scale; HAMA, Hamilton anxiety rating scale.

schizophrenia. The majority of the studies adopted the VFT, except for 12 studies that utilized paradigms such as WFT, RPS task, image-recall task, resting state measurements, TMT, stop-signal task, WM task, MDT, conversation task, emotional Stroop task, and the use of self-report measures such as the rumination response scale (RRS) and visual analog scales (VAS). The NIRS instruments utilized markedly differed from study to study.

According to our assessment of the authors' conclusions, almost every study discovered that oxy-Hb concentration values were negatively correlated with the degree of total depressive symptoms [HAMD, geriatric depression scale (GDS), Montgomery-Asberg depression rating scale (MADRS), quick inventory of depressive symptomatology-self rated (QIDS-SR), etc.] and Uemura et al. even demonstrated that such a significant negative correlation was still present after correcting for confounding factors such as gender, age, and educational history (56). Such a negative correlation was further supported by Wang et al. who illustrated that the recurrent MDD group had notably diminished increases in oxy-Hb compared to first-episode MDD patients (69). Additionally, Nishizawa et al. utilized the emotional Stroop task to evaluate brain regions associated with the severity of depression. They found that hyperactivated oxy-Hb was seen in the left frontal cortex upon contact to adverse stimuli, but no meaningful differences were established between the depressed compared with the healthy upon exposure to favorable stimuli (60). Additionally, the degree of depression was contrariwise associated with an evoked wave in the left upper frontal cortex following exposure to favorable stimuli. Akiyama et al. also found that there was significant decrease in left lateral frontotemporal activation in active depressed patients as compared to those who still had residual depressive symptoms despite MDD remission (61), further suggesting the possibility of NIRS's ability to differentiate depression of varying severity.

While such a noteworthy negative correlation was established in many papers, there were some studies that observed no significant correlations (70) and even a significant positive association (68) between HAMD scores and oxy-Hb concentrations. Such results could have been due to methodological limitations that were faced by these studies. For example, in the study by Tsujii et al. (70), the sample size was small. Thus, such slight difference in the hemodynamic response in the left temporal region among the two groups was not statistically different. A type II error might be a possibility if the size of the sample was enlarged, and the dissimilarity between the healthy and depressed in the left channels might be meaningful. Additionally, Liu et al. (68) argued that such inconsistency in results could have been due to the dynamic process of psychiatric disease development; therefore, there are some findings that may be momentary markers and not persistent markers.

Apart from the general negative association between the oxy-Hb concentration values and the degree of total depressive symptoms, many of the studies looked into a variety of individual depressive symptoms, such as suicidal thoughts, sleep quality, psychomotor retardation, obsessive-compulsive

symptoms, cognitive functioning [assessed by the Mini-Mental State Examination (MMSE), Consortium to Establish a Registry for Alzheimer's Disease (CERAD) test battery, etc.], and many others.

Suicidality is often observed in MDD patients. Suicidal ideation usually precedes a suicide attempt. Pu et al. investigated individuals prior to any attempt (73), while Tsujii et al. explored responses after an attempt (65). Pu et al. (73) found that the hemodynamic variations in the OFC, FPC, and DLPFC negatively correlated to the degree of suicidal thinking in depressed individuals; Tsujii et al. (65) revealed that the suicide attempters (SAs) revealed a reduced hemodynamic response in the left precentral gyrus compared to the nonattempters (NAs) and HCs, and the hemodynamic responses in the right middle frontal gyrus were negatively correlated with aggression and hopelessness in the SAs but not in the HCs and NAs.

Nishida et al. examined the relationship between sleep quality and oxy-Hb concentrations (71). They detected a negative relationship between average oxy-Hb concentration variations while performing the task and Pittsburgh sleep quality index (PSQI) scores. This illustrated that self-rated sleep disturbances were related to a reduced left prefrontal reactivity when performing VFT in MDD patients, thus indicating that the reactivity of the prefrontal region was vulnerable to sleep disorders.

Psychomotor retardation is another classic symptom of MDD. In Tsujii et al., psychomotor retardation on the HAM-D showed noteworthy affirmative association with average oxy-Hb changes in the right temporal region in the MDD patients with melancholic features (MDD-MF) but displayed a suggestive negative association with average oxy-Hb changes in the middle to left temporal region in the MDD patients with non-melancholic features (MDD-NMF) (70). These findings were consistent with earlier functional neuroimaging studies that determined that MDD with psychomotor retardation is related to reduced blood flow in patients with MDD-NMF, unlike those suffering from MDD-MF. Thus, this suggested that the pathophysiology of non-melancholic MDD is distinctive from melancholia. Specifically, the pathophysiology of melancholia could be linked to the right temporal region. MDD patients often showed impaired inhibitory control. In the study by Akashi et al. fluctuations in oxy-Hb concentrations in MDD patients in the superior frontal probes were associated with impairments in performance in inhibitory controls (79).

With regard to patients' functioning, while no statistical significance was established with the degree of obsessive-compulsive symptoms (68), average oxy-Hb concentrations were significantly positively interrelated with the global assessment of functioning (GAF) scores (57, 63) and SASS scores (47, 64, 66). For SASS in particular, observational changes in bilateral ventrolateral prefrontal cortex activity and the anterior portion of the temporal cortex (VLPFC/aTC) were associated with gains in social adaptability in MDD patients, although the MDD groups revealed less activation than the HCs in the VLPFC/aTC. Additionally, when Tsujii et al. assessed quality of life (QOL) using the Medical Outcomes Study 36-item

short-form health survey (SF-36), hemodynamic responses in the prefrontal region were positively associated with the role emotional domain scores for the MDD-MF patients. Comparatively, MDD-NMF patients showed no noteworthy correlations (67). This indicated that while lower functioning was associated with lower oxy-Hb concentrations, patients with MDD-MF had qualitatively distinctive prefrontal impairment features associated with emotional role functioning unlike patients with MDD-NMF.

Pu et al. assessed coping styles with the coping inventory for stressful situations (CISS) and discovered that MDD patients were more possibly using an emotion-oriented coping style as compared to using task-orientated and avoidance-orientated coping. Emotion-oriented coping style was also positively associated with subjective assessments of the degree of depression (74). Task-orientated coping was positive correlated with regional hemodynamic changes in various prefrontal regions. The findings suggested that the hemodynamic responses in the various prefrontal regions when performing VFT imply patients who suffer from MDD utilize task-orientated coping when depressed.

Apart from the use of oxy-Hb concentration measures, three papers also assessed FC (52, 54, 72). First, in patients with affective disorders, Zhu et al. (54) showed altered patterns of FC, representing far greater disorganized patterns of correlation maps, by outlining and examining four FC types: the intraregional connectivity (SC-I), the intrahemispheric connectivity (SC-II), the symmetrically interhemispheric connectivity (LC-I), and the asymmetrical interhemispheric connectivity (LC-II). This showed that patients suffering from affective disorders indicated considerably weaker intraregional connectivity and symmetrical interhemispheric connectivity in the IFG as compared to HCs and that such disruptions in the right IFG are correlated with problems in emotional reactions and reducing negative thinking in affective disorder patients. Second, similar to previous results found by many others, Rosenbaum et al. (72) also found that the subjects who were depressed revealed increased FC in a left frontoparietal cortical network at the resting state and decreased FC in a left frontopolar cortical network while doing a performance task. However, when these relations were calculated on behalf of the separate diagnostic groups, these correlations were congruent only in the non-depressed group and varied in the depressed group, thus implying that these results might be more related to depression status than to the degree of symptoms per se. In comparison to controls, the depressed individuals indicated an exceedingly connected network with a left hemispheric focus during the resting-state condition, and this result was inferred to be related to rumination. This relationship was further examined by Rosenbaum et al. (52). Though ruminative thoughts were negatively associated with FC in the cortical areas of the default mode network (DMN), mind-wandering displayed positive correlations with FC.

Moreover, Okada et al. illustrated hemodynamic changes using brain response patterns (55). The response patterns were cerebral responses to the MDT. During the MDT, the overall response areas were analyzed by using a planimeter as the areas

demarcated by the total Hb response curves and the baseline. The area deviation ratio was calculated by dividing the difference between the blood volume response areas of the right and left hemispheres by the hemisphere blood volume response area showing a lower response. When the ratios were higher (range 4–44), response patterns were classified as dominant hemispheric response patterns in normal subjects or non-dominant hemispheric response patterns in MDD patients. They demonstrated that about 50% of patients (12 out of 24 males and 4 out of 12 females) showed a “non-dominant hemisphere response pattern,” which was not seen in healthy subjects, and this “non-dominant” hemisphere could become dominant during depression. Furthermore, significant negative associations were found between the oxy-Hb and the age of onset in six channels but there were no noteworthy associations between age and oxy-Hb at the period of assessment.

Use of fNIRS to Monitor Treatment Response

All 16 studies included in **Table 4** were case-control studies. Eleven studies were performed in Japan, one study in China, one study in Uzbekistan, one study in United Kingdom, one study in the United States, and one study in Germany. The combined sample throughout the 16 studies comprised of 282 MDD patients. Every patient met the MDD criteria of DSM-IV, DSM-5 or ICD-10. The majority of the studies examined the correlation of cerebral hemodynamic changes with improvements in the depressive symptoms of patients, while five studies (47, 48, 50, 77, 81) looked at either social and/or cognitive functioning of the patients as the treatment outcome.

Almost all patients responded favorably to treatment, as the articles reported significant improvement in social functioning or a decrease in depressive symptoms. However, the patients in the study by Takamiya et al. showed no meaningful differences in depressive scores following treatment in both the high- and low-dose groups (84). In group comparisons, there were significant outcomes on NIRS signals for high doses of antidepressants compared with low doses. This highlighted that the dose-dependent influence of antidepressants on the NIRS signal ought to be considered while understanding NIRS data. The most commonly used paradigm to stimulate cognitive activity was the VFT. One study (77) used a rock, paper, scissors (RPS) task. Two studies (87, 89) used mental arithmetic tasks, and another study (88) utilized a lateral visual field simulation. The prefrontal brain regions were examined in all of these studies.

Measuring NIRS signals before starting treatment has been shown to predict treatment progress in medication-naïve patients suffering from MDD (46). In the paper by Tomioka et al. (46), the average oxy-Hb concentration values in the prefrontal and temporal regions during the VFT were significantly correlated with improvements in HAMD scores after treatment with antidepressants. The group comparisons revealed that the mean oxy-Hb concentration variations in the temporal and prefrontal cortices while performing VFT were considerably reduced in the depression group compared to HC group. This result concurred among other studies (18) that explored the uses of NIRS for the treatment of MDD.

Similar to the study by Tomioka et al. (46), Yamagata et al. also conducted a study on the application of NIRS in medication-naïve participants with MDD (82). The latter study highlighted the noninvasive nature of NIRS as they conducted repeated measurements at short intervals (three follow-ups at 4-week intervals). According to their findings, they found a substantial adverse association between mean oxy-Hb concentration values in the significant cluster (at week 4) and differences in HAMD scores from weeks 4 to 8 ($r = -.73$) and from weeks 8 to 12 ($r = -.63$). As such, they also concluded that NIRS could be used in predicting MDD patients' response to antidepressant treatment.

Ohtani et al. looked at social adaptation in their study, and in their 6-month follow-up, they found that NIRS signals could be a predictor for the longitudinal assessment of social adaptation (47). Longitudinal increases in temporal and prefrontal regions were shown to be correlated with improvements in social adaptation for patients, although there was no significant change in the severity of their clinical symptoms.

On the other hand, Pu et al. investigated patients' social functioning in their study (81). The pretreatment NIRS activation in the prefrontal region was found to be positively associated with pretreatment social adaptation self-evaluation scale (SASS) scores and negatively correlated with increase in SASS scores after 8 weeks. Their discoveries suggested that NIRS signals would be useful in evaluating social functioning in patients afflicted by late-onset depression (prior to treatment) and predicting the improvement in their social functioning after treatment. Furthermore, fNIRS can be used to predict the treatment response of not only antidepressants but also other novel treatments, such as neurostimulation (50, 85, 89), music therapy (49), and animal-assisted therapy (83).

Quality Assessment of the Included Studies

The NOS was used to assess the risk of bias in the included studies, and the results are shown in **Table 5**. No study had a complete score of eight stars. More than two-thirds of the studies had acceptable quality, of which 49 studies were awarded four or more stars, with 11 studies given seven stars, 14 studies given six stars, 19 studies given five stars, and 5 studies given four stars. Fifteen studies fared more poorly in quality with less than four stars, of which 3 studies were given three stars, 11 studies were given two stars, and 1 study was given one star.

DISCUSSION

Currently, there is no specific test or biomarker for diagnosing and monitoring the progression of depression. MRI studies have shown reduced size in the medial and lateral prefrontal cortex, amygdala, hippocampus, and striatum; increased functional activity in the medial prefrontal cortex, amygdala, and hippocampus; and decreased activity in the lateral prefrontal cortex and striatum in depressed subjects compared with HCs (90). Nevertheless, the applicability and utility of MRI in clinical practice is limited due to the high cost, need to stay immobile, presence of noise while scanning, long scanning duration, and

TABLE 5 | Risks of bias within studies.

| Study sources | Selection | | | Comparability | | Exposure | |
|-------------------------|----------------------------------|-------------------------|-----------------------|------------------------|---------------------------|--|-------------------|
| | Is the case definition adequate? | Representative of cases | Selection of controls | Definition of controls | Determination of exposure | Same method for determining cases and controls | Nonresponse rates |
| Suto et al. (18) | ★ | | | ★ | ★ | ★ | |
| Usami et al. (86) | ★ | | | | ★ | | |
| Takei et al. (63) | ★ | | ★ | ★ | ★★ | ★ | ★ |
| Shimodera et al. (37) | ★ | | ★ | ★ | ★ | | |
| Ma et al. (38) | ★ | | ★ | ★ | ★★ | ★ | ★ |
| Takizawa et al. (39) | ★ | | ★ | ★ | ★★ | ★ | ★ |
| Zhu et al. (54) | ★ | | ★ | ★ | ★★ | ★ | ★ |
| Matsubara et al. (42) | ★ | | ★ | ★ | ★★ | ★ | ★ |
| Pu et al. (81) | ★ | | | | ★ | ★ | ★ |
| Akashi et al. (43) | ★ | | | ★ | ★ | ★ | ★ |
| Downey et al. (51) | ★ | | | | ★ | ★ | ★ |
| Gao et al. (44) | ★ | | ★ | ★ | ★ | ★ | ★ |
| Pu et al. (64) | ★ | | ★ | ★ | ★★ | ★ | ★ |
| Tsujii et al. (65) | ★ | | ★ | ★ | ★★ | ★ | ★ |
| Pu et al. (74) | ★ | | ★ | ★ | ★ | ★ | ★ |
| Schecklmann et al. (45) | ★ | | | ★ | ★ | ★ | ★ |
| Tsujii et al. (67) | ★ | | | ★ | ★ | ★ | ★ |
| Liu et al. (68) | ★ | | ★ | ★ | ★★ | ★ | ★ |
| Wang et al. (69) | ★ | | | ★ | ★ | ★ | ★ |
| Tsujii et al. (70) | ★ | | ★ | ★ | ★ | ★ | ★ |
| Satomura et al. (80) | ★ | | | | ★ | ★ | ★ |
| Nishida et al. (71) | ★ | | | ★ | ★ | ★ | ★ |
| Rosenbaum et al. (72) | ★ | | ★ | ★ | ★ | ★ | ★ |
| Pu et al. (73) | ★ | | ★ | ★ | ★ | ★ | ★ |
| Koseiki et al. (78) | ★ | | | | ★ | | |
| Pu et al. (74) | ★ | | ★ | ★ | ★ | ★ | ★ |
| Schiffer et al. (88) | ★ | | | | ★ | | |
| Eschweiler et al. (89) | ★ | | ★ | ★ | ★ | ★ | ★ |
| Tomioka et al. (46) | ★ | | ★ | ★ | | | |
| Pu et al. (25) | ★ | ★ | | ★ | ★★ | ★ | ★ |
| Matsuo et al. (26) | ★ | | | ★ | ★★ | ★ | ★ |
| Onishi et al. (77) | ★ | | | | ★ | | |
| Rosenbaum et al. (52) | ★ | | | ★ | ★ | ★ | ★ |
| Ohtani et al. (47) | ★ | | | ★ | ★★ | ★ | ★ |

(Continued)

TABLE 5 | Continued

| Study sources | Selection | | Comparability | | Exposure | | |
|-----------------------|----------------------------------|-------------------------|-----------------------|------------------------|---------------------------|--|-------------------|
| | Is the case definition adequate? | Representative of cases | Selection of controls | Definition of controls | Determination of exposure | Same method for determining cases and controls | Nonresponse rates |
| Herrmann et al. (27) | ★ | ★ | | ★ | ★ | ★ | |
| Yamagata et al. (82) | ★ | | | | ★ | | |
| Kondo et al. (53) | ★ | | | ★ | ★ | ★ | ★ |
| Shoji et al. (28) | ★ | | | ★ | ★ | ★ | |
| Kinoshita et al. (29) | ★ | | | | ★ | | |
| Kito et al. (30) | ★ | | | ★ | ★ | ★ | |
| Kawano et al. (76) | ★ | ★ | | | ★ | | |
| Matsuo et al. (31) | ★ | ★ | | | ★ | | |
| Zhu et al. (54) | ★ | | ★ | ★ | ★ | ★ | |
| Okada et al. (55) | ★ | | | ★ | ★ | ★ | |
| Uemura et al. (56) | ★ | | | | ★ | | |
| Matsubara et al. (32) | ★ | | | ★ | ★ | ★ | |
| Masuda et al. (48) | ★ | | | | ★ | | |
| Kinou et al. (57) | ★ | | | ★ | ★ | ★ | |
| Yamagata et al. (58) | ★ | | | ★ | ★ | ★ | |
| Koike et al. (33) | ★ | | | | ★ | | |
| Feng et al. (49) | ★ | | ★ | ★ | ★ | ★ | |
| Aoki et al. (83) | ★ | | | | ★ | | |
| Noda et al. (59) | ★ | | | ★ | ★ | ★ | |
| Hirano et al. (50) | ★ | | | ★ | ★ | ★ | |
| Azechi et al. (34) | ★ | | | ★ | ★ | ★ | |
| Nishizawa et al. (60) | ★ | | | ★ | ★ | ★ | |
| Takamiya et al. (84) | ★ | | | | ★ | | |
| Matsuo et al. (35) | ★ | | | ★ | ★★ | ★ | |
| Ohta et al. (36) | ★ | | | ★ | ★ | ★ | |
| Akiyama et al. (61) | ★ | | ★ | ★ | ★ | ★ | |
| Ohi et al. (62) | ★ | | | ★ | ★ | ★ | |
| Shinba et al. (85) | ★ | | | | ★ | | |
| Akashi et al. (79) | ★ | | | | | | |
| Liske et al. (40) | ★ | | ★ | ★ | ★ | | ★ |

risk of inducing claustrophobia. These considerations may be especially relevant for depressed patients who may be emotionally sensitive and thus find it harder to tolerate being confined to a tight space with recurrent noise for a prolonged period. Compared to fMRI, fNIRS is less expensive, fast to operate, quiet, and more portable and, thus, able to perform the scan in the clinic or ward directly and allows body movement in a naturalistic environment (91). However, some of its inherent limitations may influence the accuracy and eventual applicability in clinical practice. For instance, its spatial resolution and penetration depth are limited, which reduces access to the subcortical regions that have a pertinent role in psychiatric disorders. As fNIRS is established on the principles of neurovascular coupling, it is affected by blood pressure, hemoglobin level, blood circulation, vasculature, and carbon dioxide concentration (92, 93). Nevertheless, a meticulous study design, enhanced fNIRS techniques, and statistical processing could potentially mitigate these shortfalls.

This is the first systematic review on the application of fNIRS to depression that intended to provide an overview of the up-to-date information of 3 pertinent clinical questions: (i) the usefulness of fNIRS in differentiating depressed from healthy individuals, (ii) the correlation of fNIRS signals with depression symptomatology, and in turn (iii) monitoring treatment response. Answers to these questions are essential in ascertaining the applicability and utility of fNIRS as a tool to assist the diagnosing of depression, predicting clinical symptoms of patients, monitoring the treatment response and disease progression, and for prognosticating the disease. Based on the combined findings of the reviewed papers, most of the studies were conducted in Japan, although there has been an increasing trend for studies being conducted in China, Germany and other countries. The number of fNIRS studies has also exponentially increased year on year, suggesting an increased recognition of this technology in mainstream science. The most commonly used active paradigm is the VFT. This is not surprising as the VFT is quite a common bedside neuropsychological test that has been extensively utilized to ascertain executive function and language content (94) and is easy to perform within a short period, though it is imperative to realize that the performance on the VFT can be influenced by culture. Participants are obliged to state as many unique words that begin with a particular letter or are from a semantic category in a restricted time (up to a minute usually). Cognitive dysfunction occurs in patients who are depressed, of which impairments in executive ability is one of the core domains, in addition to learning/memory, attention/concentration, and processing speed (95). As a validated test to ascertain executive function, the VFT is able to elicit distinct differences in performance and neuroimaging responses between depressed and healthy people (23).

Based on the findings of this systematic review, the fNIRS signals show promise as an ideal biomarker in addressing our review's three fundamental questions: do fNIRS signals differentiate depressed from nondepressed individuals, correlate with specific depressive symptomatology, and in turn aid the monitoring of treatment responses (regardless of whether

the treatment is a medication or another treatment modality)? The findings of our systematic review demonstrated a consistent pattern of attenuated pre-frontal stimulation in patients with depression while conducting cognitive tasks using the NIRS method, and cerebro-hemodynamic changes are associated with particular symptoms encountered by patients and treatment responses. However, most studies did not include sensitivity and specificity data that compared depressed patients from HCs, which would be important to elucidate the validity of fNIRS as a diagnostic tool. Thus, future studies may need to consider standardizing their analysis such that comparisons across studies can be made. The correlation of brain signals with clinical symptoms such as suicidal ideation, sleep quality, and psychomotor retardation may facilitate more in-depth personalized profiling of the individual's depression and may be able to better identify risk issues in management (e.g., self-harm, suicide, neglect). Symptom identification can be especially helpful when patients are not forthcoming about their symptoms, which makes it difficult to ascertain their risks. However, the majority of the selected studies had small sample populations, thereby reducing the power of the study, and there were variations in study methodologies, including the device and paradigm used, which limited the ability to effectively combine or compare results. One important aspect of fNIRS as a biomarker would be its potential to prognosticate the disease, i.e., whether the patient will improve or worsen with time. Nevertheless, the existing studies were either cross-sectional or had short longitudinal follow-ups, which made it difficult to address this issue. Longer longitudinal studies of at least 6 months to a year would be beneficial, considering that most depressive episodes last for at least a few months. Furthermore, longitudinal studies can help to elucidate whether the hemodynamic responses recorded by NIRS are a state- or trait-dependent marker of depression, as the current evidence is conflicting and confounded by several reasons.

Most studies scanned their subjects 2 weeks to 1 month after starting antidepressants, and then scans were performed monthly. Having more frequent fNIRS measurements in longitudinal studies (e.g., weekly) can provide us with a better understanding of brain dynamics and minimize the influence of confounding factors. The majority of studies to date have examined only the temporal and frontal regions. It may thus be worthwhile to expand the assessment to other brain regions, such as the occipital and parietal regions, especially because these two regions have also been implicated in depression (96, 97). Furthermore, most studies examined hemodynamic changes in discrete brain regions, with only a few studies investigating FC aberrations and even fewer ascertaining effective connectivity across brain regions (97). Connectivity studies allow a more holistic approach to how various brain areas interactively function and may represent another means of examining the disease state. Most of the studies involved patients on medication, and those with medication-naïve patients had small sample sizes. Thus, we are unable to exclude the possibility that antidepressants interfered with the signal results. Studies have highlighted that antidepressants can affect

the NIRS signals (84). Therefore, it may be worthwhile to have more studies involving scans of medication-naïve patients, though this can be practically challenging at the ground level.

An encouraging application is combining different neuroimaging modalities to allow a more accurate and comprehensive surveillance of the neurophysiological changes. This can include combining fNIRS with EEG, fMRI, PET, or diffusion tensor imaging (DTI) to complement fNIRS. This can in turn improve the spatial and temporal resolution and thereby increase sensitivity and specificity. Nevertheless, the applicability of combining modalities such as fMRI and PET is limited due to the immobile and bulky apparatus, and although EEG is portable, use of EEG increases the setup time, and the increased noise-signal ratio with artifacts may reduce its accuracy. On the other hand, integrative approaches that combine various modalities of biomarkers such as genetic, neuroimaging, and neuropsychological data that are analyzed using high-dimensional multivariate statistical methods are gaining prominence and may more comprehensively facilitate disease diagnostics and prediction (98).

Our review has a number of limitations. First, we selected observational studies that had small sample sizes, thereby increasing the risk of selective and confounding biases and reduced power due to small effect sizes. However, based on our search, there were no randomized controlled trials of patients with depression using fNIRS. Second, we only included studies in English and searched in four databases. We recognize that there are an increasing number of fNIRS studies conducted in China and other countries that may not be published in English and indexed in these four databases. Thus, this potentially led to

publication bias. Third, we also included conference proceedings, and several details of these datasets were unavailable from the source. However, it was essential to include them to reflect the spectrum of studies being performed in the current fNIRS research landscape. Regardless, this review has strengths in being the first to combine three pertinent questions pertaining to fNIRS as a biomarker, and there has not been an updated systematic review of fNIRS in depression since 2015. This study is a timely update of the current landscape of using fNIRS on depressed patients, given the increasing availability of this technology worldwide.

In conclusion, there is a good amount of evidence in the current literature to suggest fNIRS as a diagnostic and predictive tool for MDD, and it has shown consistent hemodynamic patterns in depressed patients compared to healthy individuals. There is also an increasing amount of studies on fNIRS in depression, indicating the increased recognition of this technique in the study of depression. Future studies involving larger sample sizes, standardized methodology, examination of more brain regions in an integrative approach, and longitudinal follow-ups are needed to advance the use of fNIRS in psychiatric clinical practice and research.

AUTHOR CONTRIBUTIONS

Conceptualization: CH. Data extraction and review: LL, AL, NC, RT, SL. Writing: CH. Review and manuscript amendment: LL, AL, NC, RT, SL, RH.

REFERENCES

- World Health Organization. (2017). *Depression and Other Common Mental Disorders: Global Health Estimates*. World Health Organization [cited 2019 Oct 13]; Available from: <https://apps.who.int/iris/bitstream/handle/10665/254610/WHO-MSD-MER-2017.2-eng.pdf?sequence=1>.
- Lim GY, Tam WW, Lu Y, Ho CS, Zhang MW, Ho RC. Prevalence of Depression in the Community from 30 Countries between 1994 and 2014. *Sci Rep* (2018) 128(1):2861. doi: 10.1038/s41598-018-21243-x
- Choo CC, Harris KM, Ho RC. Prediction of Lethality in Suicide Attempts: Gender Matters. *Omega*. (2019) 80(1):87–103. doi: 10.1177/0030222817725182
- Sheehan DV. Depression: underdiagnosed, undertreated, underappreciated. *Manag Care Langhorne Pa* (2004) 13(6 Suppl Depression):6–8.
- Mayberg HS. Modulating dysfunctional limbic-cortical circuits in depression: towards development of brain-based algorithms for diagnosis and optimised treatment. *Br Med Bull* (2003) 65:193–207. doi: 10.1093/bmb/65.1.193
- Ottowitz WE, Dougherty DD, Savage CR. The neural network basis for abnormalities of attention and executive function in major depressive disorder: implications for application of the medical disease model to psychiatric disorders. *Harv Rev Psychiatry* (2002) 10(2):86–99. doi: 10.1080/10673220216210
- Fava M. Diagnosis and definition of treatment-resistant depression. *Biol Psychiatry* (2003) 53(8):649–59. doi: 10.1016/S0006-3223(03)00231-2
- Gaynes BN, Warden D, Trivedi MH, Wisniewski SR, Fava M, Rush AJ. What did STAR*D teach us? Results from a large-scale, practical, clinical trial for patients with depression. *Psychiatr Serv Wash DC*. (2009) 60(11):1439–45. doi: 10.1176/ps.2009.60.11.1439
- Olchanski N, McInnis Myers M, Halseth M, Cyr PL, Bockstedt L, Goss TF, et al. The economic burden of treatment-resistant depression. *Clin Ther* (2013) 35(4):512–22. doi: 10.1016/j.clinthera.2012.09.001
- Strawbridge R, Young AH, Cleare AJ. Biomarkers for depression: recent insights, current challenges and future prospects. *Neuropsychiatr Dis Treat* (2017) 13:1245–62. doi: 10.2147/NDT.S114542
- Okada G, Okamoto Y, Yamashita H, Ueda K, Takami H, Yamawaki S. Attenuated prefrontal activation during a verbal fluency task in remitted major depression. *Psychiatry Clin Neurosci* (2009) 63(3):423–5. doi: 10.1111/j.1440-1819.2009.01952.x
- Meyer JH, Houle S, Sagrati S, Carella A, Hussey DF, Ginovart N, et al. Brain serotonin transporter binding potential measured with carbon 11-labeled DASB positron emission tomography: effects of major depressive episodes and severity of dysfunctional attitudes. *Arch Gen Psychiatry* (2004) 61(12):1271–9. doi: 10.1001/archpsyc.61.12.1271
- Drevets WC. Neuroimaging studies of mood disorders. *Biol Psychiatry* (2000) 48(8):813–29. doi: 10.1016/S0006-3223(00)01020-9
- Lai CYY, Ho CSH, Lim CR, Ho RCM. Functional near-infrared spectroscopy in psychiatry. *BJPsych Adv* (2017) 23(5):324–30. doi: 10.1192/apt.bp.115.015610
- Fukuda M. Near-infrared spectroscopy in psychiatry. *Brain Nerve Shinkei Kenkyu No Shinpo*. (2012) 64(2):175–83.
- Scholkmann F, Kleiser S, Metz AJ, Zimmermann R, Mata Pavia J, Wolf U, et al. A review on continuous wave functional near-infrared spectroscopy and imaging instrumentation and methodology. *NeuroImage* (2014) 85 (Pt 1):6–27. doi: 10.1016/j.neuroimage.2013.05.004
- Boas DA, Elwell CE, Ferrari M, Taga G. Twenty years of functional near-infrared spectroscopy: introduction for the special issue. *NeuroImage* (2014) 85 Pt 1:1–5. doi: 10.1016/j.neuroimage.2013.11.033
- Suto T, Fukuda M, Ito M, Uehara T, Mikuni M. Multichannel near-infrared spectroscopy in depression and schizophrenia: cognitive brain activation study. *Biol Psychiatry* (2004) 55(5):501–11. doi: 10.1016/j.biopsych.2003.09.008

19. Noda T, Nakagome K, Setoyama S, Matsushima E. Working memory and prefrontal/temporal hemodynamic responses during post-task period in patients with schizophrenia: A multi-channel near-infrared spectroscopy study. *J Psychiatr Res* (2017) 95:288–98. doi: 10.1016/j.jpsychires.2017.09.001
20. Hirose T, Tsujii N, Mikawa W, Shirakawa O. Delayed hemodynamic responses associated with a history of suicide attempts in bipolar disorder: a multichannel near-infrared spectroscopy study. *Psychiatry Res Neuroimaging*. (2018) 280:15–21. doi: 10.1016/j.pscychresns.2018.08.003
21. Katzorke A, Zeller JBM, Müller LD, Lauer M, Polak T, Deckert J, et al. Decreased hemodynamic response in inferior frontotemporal regions in elderly with mild cognitive impairment. *Psychiatry Res Neuroimaging*. (2018) 274:11–8. doi: 10.1016/j.pscychresns.2018.02.003
22. Ueda S, Ota T, Iida J, Yamamuro K, Yoshino H, Kishimoto N, et al. Reduced prefrontal hemodynamic response in adult attention-deficit hyperactivity disorder as measured by near-infrared spectroscopy. *Psychiatry Clin Neurosci* (2018) 72(6):380–90. doi: 10.1111/pcn.12643
23. Zhang H, Dong W, Dang W, Quan W, Tian J, Chen R, et al. Near-infrared spectroscopy for examination of prefrontal activation during cognitive tasks in patients with major depressive disorder: a meta-analysis of observational studies. *Psychiatry Clin Neurosci* (2015) 69(1):22–33. doi: 10.1111/pcn.12209
24. Wells GA, Shea B, O'Connell D, Peterson J, Welch V, Losos M. (2000). *The Newcastle–Ottawa Scale (NOS) for assessing the quality of nonrandomized studies in meta-analyses*. [Internet]. [cited 2019 Oct 13]. Available from: http://www.ohri.ca/programs/clinical_epidemiology/oxford.asp.
25. Pu S, Yamada T, Yokoyama K, Matsumura H, Kobayashi H, Sasaki N, et al. A multi-channel near-infrared spectroscopy study of prefrontal cortex activation during working memory task in major depressive disorder. *Neurosci Res* (2011) 70(1):91–7. doi: 10.1016/j.neures.2011.01.001
26. Matsuo K, Kato T, Fukuda M, Kato N. Alteration of Hemoglobin Oxygenation in the Frontal Region in Elderly Depressed Patients as Measured by Near-infrared Spectroscopy. *J Neuropsychiatry Clin Neurosci* (2000) 12(4):465–71. doi: 10.1176/jnp.12.4.465
27. Herrmann MJ, Ehliis A-C, Fallgatter AJ. Bilaterally Reduced Frontal Activation During a Verbal Fluency Task in Depressed Patients as Measured by Near-Infrared Spectroscopy. *J Neuropsychiatry Clin Neurosci* (2004) 16(2):170–5. doi: 10.1176/jnp.16.2.170
28. Shoji Y, Morita K, Yanagimoto H, Fujiki R, Ishii Y, Uchimura N. Characteristics of the single event related [Oxy-Hb] changes in patients with depressive disorder.
29. Kinoshita S, Kanazawa T, Kikuyama H, Yoneda H. Clinical application of DEX/CRH test and multi-channel NIRS in patients with depression. *Behav Brain Funct BBF*. (2016) 12(1):25. doi: 10.1186/s12993-016-0108-x
30. Kito H, Ryokawa A, Kinoshita Y, Sasayama D, Sugiyama N, Ogihara T, et al. Comparison of alterations in cerebral hemoglobin oxygenation in late life depression and Alzheimer's disease as assessed by near-infrared spectroscopy. *Behav Brain Funct BBF* (2014) 10:8. doi: 10.1186/1744-9081-10-8
31. Matsuo K, Kato N, Kato T. Decreased cerebral haemodynamic response to cognitive and physiological tasks in mood disorders as shown by near-infrared spectroscopy. *Psychol Med* (2002) 32(6):1029–37. doi: 10.1017/S0033291702005974
32. Matsubara T, Matsuo K, Harada K, Nakashima M, Nakano M, Hirotsu M, et al. Different Fronto-Temporal Activation During an Emotional Word Task in Patients with Unipolar and Bipolar Depression: A Functional Near-Infrared Spectroscopy Study (2015). *Biol Psychiatry* (2015) 77:1S–444S.
33. Koike S, Sakakibara E, Satomura Y, Sakurada H, Yamagishi M, Matsuoka J, et al. Differentiation between Schizophrenia, Bipolar Disorder, and Major Depression Using the Prefrontal Brain Activity and the Evaluation of Ultra-high Risk for Psychosis: A Large-Sample Functional Near-infrared Spectroscopy Study. *Early Interv Psychiatry* (2018) 12(Suppl. 1):104–232. doi: 10.1111/eip.12724
34. Azechi M, Iwase M, Ishii R, Ikezawa K, Canuet L, Kurimoto R, et al. P27-5 Frontal lobe dysfunction and regional hemodynamic changes in major depression: A near infrared spectroscopy study. *Clin Neurophysiol* (2010) 121:S264. doi: 10.1016/S1388-2457(10)61079-6
35. Matsuo K, Onodera Y, Hamamoto T, Muraki K, Kato N, Kato T. Hypofrontality and microvascular dysregulation in remitted late-onset depression assessed by functional near-infrared spectroscopy. *NeuroImage*. (2005) 26(1):234–42. doi: 10.1016/j.neuroimage.2005.01.024
36. Ohta H, Yamagata B, Tomioka H, Takahashi T, Yano M, Nakagome K, et al. Hypofrontality in panic disorder and major depressive disorder assessed by multi-channel near-infrared spectroscopy. *Depress Anxiety* (2008) 25 (12):1053–9. doi: 10.1002/da.20463
37. Shimodera S, Imai Y, Kamimura N, Morokuma I, Fujita H, Inoue S, et al. Near-infrared spectroscopy of bipolar disorder may be distinct from that of unipolar depression and of healthy controls: NIRS of bipolar vs unipolar disorder. *Asia-Pac Psychiatry* (2012) 4(4):258–65. doi: 10.1111/j.1758-5872.2012.00218.x
38. Ma X-Y, Wang Y-J, Xu B, Feng K, Sun G-X, Zhang X-Q, et al. Near-Infrared Spectroscopy Reveals Abnormal Hemodynamics in the Left Dorsolateral Prefrontal Cortex of Menopausal Depression Patients. *Dis Markers* (2017) 2017:1695930. doi: 10.1155/2017/1695930
39. Takizawa R, Fukuda M, Kawasaki S, Kasai K, Mimura M, Pu S, et al. Neuroimaging-aided differential diagnosis of the depressive state. *NeuroImage* (2014) 85 (Pt 1):498–507. doi: 10.1016/j.neuroimage.2013.05.126
40. Liske BCJ, Ackermann PH, Münch D, Florian H, Ehliis A-C, Stevens A, et al. New electrophysiological findings in the detection of malingering attention deficits in subjects with an episode of depression compared to healthy subjects. *Clin Neurophysiology* (2015) 126:e63–e170. doi: 10.1016/j.clinph.2015.04.178
41. Zhu Y, Quan W, Wang H, Ma Y, Yan J, Zhang H, et al. Prefrontal activation during a working memory task differs between patients with unipolar and bipolar depression: A preliminary exploratory study. *J Affect Disord* (2018) 225:64–70. doi: 10.1016/j.jad.2017.07.031
42. Matsubara T, Matsuo K, Nakashima M, Nakano M, Harada K, Watanuki T, et al. Prefrontal activation in response to emotional words in patients with bipolar disorder and major depressive disorder. *NeuroImage* (2014) 85 (Pt 1):489–97. doi: 10.1016/j.neuroimage.2013.04.098
43. Akashi H, Tsujii N, Mikawa W, Adachi T, Kirime E, Shirakawa O. Prefrontal cortex activation is associated with a discrepancy between self- and observer-rated depression severities of major depressive disorder: a multichannel near-infrared spectroscopy study. *J Affect Disord* (2015) 174:165–72. doi: 10.1016/j.jad.2014.11.020
44. Gao L, Cai Y, Wang H, Wang G, Zhang Q, Yan X. Probing prefrontal cortex hemodynamic alterations during facial emotion recognition for major depression disorder through functional near-infrared spectroscopy. *J Neural Eng*. (2019) 16(2):026026. doi: 10.1088/1741-2552/ab0093
45. Schecklmann M, Dresler T, Beck S, Jay JT, Febres R, Haeussler J, et al. Reduced prefrontal oxygenation during object and spatial visual working memory in unipolar and bipolar depression. *Psychiatry Res* (2011) 194(3):378–84. doi: 10.1016/j.pscychresns.2011.01.016
46. Tomioka H, Yamagata B, Kawasaki S, Pu S, Iwanami A, Hirano J, et al. A Longitudinal Functional Neuroimaging Study in Medication-Naïve Depression after Antidepressant Treatment. *PloS One* (2015) 10(3):e0120828. doi: 10.1371/journal.pone.0120828
47. Ohtani T, Nishimura Y, Takahashi K, Ikeda-Sugita R, Okada N, Okazaki Y. Association between longitudinal changes in prefrontal hemodynamic responses and social adaptation in patients with bipolar disorder and major depressive disorder. *J Affect Disord* (2015) 176:78–86. doi: 10.1016/j.jad.2015.01.042
48. Masuda K, Nakanishi M, Okamoto K, Kawashima C, Oshita H, Inoue A, et al. Different functioning of prefrontal cortex predicts treatment response after a selective serotonin reuptake inhibitor treatment in patients with major depression. *J Affect Disord* (2017) 214:44–52. doi: 10.1016/j.jad.2017.02.034
49. Feng K, Shen C-Y, Ma X-Y, Chen G-F, Zhang M-L, Xu B, et al. Effects of music therapy on major depressive disorder: A study of prefrontal hemodynamic functions using fNIRS. *Psychiatry Res* (2019) 275:86–93. doi: 10.1016/j.pscychres.2019.03.015
50. Hirano J, Takamiya A, Yamagata B, Hotta S, Miyasaka Y, Pu S, et al. Frontal and temporal cortical functional recovery after electroconvulsive therapy for depression: A longitudinal functional near-infrared spectroscopy study. *J Psychiatr Res* (2017) 91:26–35. doi: 10.1016/j.jpsychires.2017.02.018
51. Downey D, Sabrina B, Liam T, Rebecca E, Elwell C, Richard M-W, et al. Prefrontal cortex haemodynamic responses in severe major depression and the effects of ECT: an fNIRS study. *Eur Neuropsychopharmacol* (2016), S304.
52. Rosenbaum D, Haipt A, Fuhr K, Haeussinger FB, Metzger FG, Nuerk H-C, et al. Aberrant functional connectivity in depression as an index of state and trait rumination. *Sci Rep* (2017) 7(1):2174. doi: 10.1038/s41598-017-02277-z

53. Kondo A, Shoji Y, Morita K, Sato M, Ishii Y, Yanagimoto H, et al. Characteristics of oxygenated hemoglobin concentration change during pleasant and unpleasant image-recall tasks in patients with depression: Comparison with healthy subjects. *Psychiatry Clin Neurosci* (2018) 72 (8):611–22. doi: 10.1111/pcn.12684
54. Zhu H, Xu J, Li J, Peng H, Cai T, Li X, et al. Decreased functional connectivity and disrupted neural network in the prefrontal cortex of affective disorders: A resting-state fNIRS study. *J Affect Disord* (2017) 221:132–44. doi: 10.1016/j.jad.2017.06.024
55. Okada F, Takahashi N, Tokumitsu Y. Dominance of the “nondominant” hemisphere in depression. *J Affect Disord* (1996) 37(1):13–21. doi: 10.1016/0165-0327(95)00040-2
56. Uemura K, Shimada H, Doi T, Makizako H, Park H, Suzuki T. Depressive symptoms in older adults are associated with decreased cerebral oxygenation of the prefrontal cortex during a trail-making test. *Arch Gerontol Geriatr*. (2014) 59(2):422–8. doi: 10.1016/j.archger.2014.07.003
57. Kinou M, Takizawa R, Marumo K, Kawasaki S, Kawakubo Y, Fukuda M, et al. Differential spatiotemporal characteristics of the prefrontal hemodynamic response and their association with functional impairment in schizophrenia and major depression. *Schizophr Res* (2013) 150(2–3):459–67. doi: 10.1016/j.schres.2013.08.026
58. Yamagata B, Tomioka H, Takahashi T, Isomura AJ, Kobayashi H, Mimura M. Differentiating early and late-onset depression with multichannel near-infrared spectroscopy. *Psychogeriatrics*. (2008) 8(2):79–87. doi: 10.1111/j.1479-8301.2008.00232.x
59. Noda T, Yoshida S, Matsuda T, Okamoto N, Sakamoto K, Koseki S, et al. Frontal and right temporal activations correlate negatively with depression severity during verbal fluency task: a multi-channel near-infrared spectroscopy study. *J Psychiatr Res* (2012) 46(7):905–12. doi: 10.1016/j.jpsychires.2012.04.001
60. Nishizawa Y, Kanazawa T, Kawabata Y, Matsubara T, Maruyama S, Kawano M, et al. fNIRS Assessment during an Emotional Stroop Task among Patients with Depression: Replication and Extension. *Psychiatry Investig* (2019) 16 (1):80–6. doi: 10.30773/pi.2018.11.12.2
61. Akiyama T, Koeda M, Okubo Y, Kimura M. Hypofunction of left dorsolateral prefrontal cortex in depression during verbal fluency task: A multi-channel near-infrared spectroscopy study. *J Affect Disord* (2018) 231:83–90. doi: 10.1016/j.jad.2018.01.010
62. Ohi K, Shimada T, Kihara H, Yasuyama T, Sawai K, Matsuda Y, et al. Impact of Familial Loading on Prefrontal Activation in Major Psychiatric Disorders: A Near-Infrared Spectroscopy (NIRS) Study. *Sci Rep* (2017) 7:44268. doi: 10.1038/srep44268
63. Takei Y, Suda M, Aoyama Y, Sakurai N, Tagawa M, Motegi T, et al. Near-infrared spectroscopic study of frontopolar activation during face-to-face conversation in major depressive disorder and bipolar disorder. *J Psychiatr Res* (2014) 57:74–83. doi: 10.1016/j.jpsychires.2014.06.009
64. Pu S, Matsumura H, Yamada T, Ikezawa S, Mitani H, Adachi A, et al. Reduced frontopolar activation during verbal fluency task associated with poor social functioning in late-onset major depression: Multi-channel near-infrared spectroscopy study. *Psychiatry Clin Neurosci* (2008) 62(6):728–37. doi: 10.1111/j.1440-1819.2008.01882.x
65. Tsujii N, Mikawa W, Tsujimoto E, Adachi T, Niwa A, Ono H, et al. Reduced left precentral regional responses in patients with major depressive disorder and history of suicide attempts. *PLoS One* (2017) 12(4):e0175249. doi: 10.1371/journal.pone.0175249
66. Pu S, Yamada T, Yokoyama K, Matsumura H, Mitani H, Adachi A, et al. Reduced prefrontal cortex activation during the working memory task associated with poor social functioning in late-onset depression: multi-channel near-infrared spectroscopy study. *Psychiatry Res* (2012) 203(2–3):222–8. doi: 10.1016/j.psychres.2012.01.007
67. Tsujii N, Mikawa W, Tsujimoto E, Akashi H, Adachi T, Kirime E, et al. Relationship between prefrontal hemodynamic responses and quality of life differs between melancholia and non-melancholic depression. *Psychiatry Res Neuroimaging*. (2016) 253:26–35. doi: 10.1016/j.psychres.2016.04.015
68. Liu X, Sun G, Zhang X, Xu B, Shen C, Shi L, et al. Relationship between the prefrontal function and the severity of the emotional symptoms during a verbal fluency task in patients with major depressive disorder: a multi-channel NIRS study. *Prog Neuropsychopharmacol Biol Psychiatry* (2014) 54:114–21. doi: 10.1016/j.pnpbp.2014.05.005
69. Wang J, Lv B, Quan W, Wydel TN, Tian J, Wang P, et al. Right fronto-temporal activation differs between Chinese first-episode and recurrent Major Depression Disorders during a verbal fluency task: A near-infrared spectroscopy study. *Psychiatry Res Neuroimaging*. (2017) 264:68–75. doi: 10.1016/j.psychres.2017.03.013
70. Tsujii N, Mikawa W, Akashi H, Tsujimoto E, Adachi T, Kirime E, et al. Right temporal activation differs between melancholia and nonmelancholic depression: a multichannel near-infrared spectroscopy study. *J Psychiatr Res* (2014) 55:1–7. doi: 10.1016/j.jpsychires.2014.04.003
71. Nishida M, Kikuchi S, Matsumoto K, Yamauchi Y, Saito H, Suda S. Sleep complaints are associated with reduced left prefrontal activation during a verbal fluency task in patients with major depression: A multi-channel near-infrared spectroscopy study. *J Affect Disord* (2017) 207:102–9. doi: 10.1016/j.jad.2016.09.028
72. Rosenbaum D, Hagen K, Deppermann S, Kroczeck AM, Haeussinger FB, Heinzl S, et al. State-dependent altered connectivity in late-life depression: a functional near-infrared spectroscopy study. *Neurobiol Aging*. (2016) 39:57–68. doi: 10.1016/j.neurobiolaging.2015.11.022
73. Pu S, Nakagome K, Yamada T, Yokoyama K, Matsumura H, Yamada S, et al. Suicidal ideation is associated with reduced prefrontal activation during a verbal fluency task in patients with major depressive disorder. *J Affect Disord* (2015) 181:9–17. doi: 10.1016/j.jad.2015.04.010
74. Pu S, Nakagome K, Yamada T, Yokoyama K, Matsumura H, Mitani H, et al. The relationship between the prefrontal activation during a verbal fluency task and stress-coping style in major depressive disorder: a near-infrared spectroscopy study. *J Psychiatr Res* (2012) 46(11):1427–34. doi: 10.1016/j.jpsychires.2012.08.001
75. Alexopoulos GS. Depression in the elderly. *Lancet Lond Engl* (2005) 365 (9475):1961–70. doi: 10.1016/S0140-6736(05)66665-2
76. Kawano M, Kanazawa T, Kikuyama H, Tsutsumi A, Kinoshita S, Kawabata Y, et al. Correlation between frontal lobe oxy-hemoglobin and severity of depression assessed using near-infrared spectroscopy. *J Affect Disord* (2016) 205:154–8. doi: 10.1016/j.jad.2016.07.013
77. Onishi Y, Kikuchi S, Watanabe E, Kato S. Alterations in prefrontal cortical activity in the course of treatment for late-life depression as assessed on near-infrared spectroscopy. *Psychiatry Clin Neurosci* (2008) 62(2):177–84. doi: 10.1111/j.1440-1819.2008.01752.x
78. Koseki S, Noda T, Yokoyama S, Kunisato Y, Ito D, Suyama H, et al. The relationship between positive and negative automatic thought and activity in the prefrontal and temporal cortices: a multi-channel near-infrared spectroscopy (NIRS) study. *J Affect Disord* (2013) 151(1):352–9. doi: 10.1016/j.jad.2013.05.067
79. Akashi H, Noa T, Sakai S, Mikawa W, Kirime E, Shirakawa O. Inhibitory controls in bipolar and major depressive disorder: a NIRS study. *Bipolar Disord* (2012) 14:52.
80. Satomura Y, Sakakibara E, Takizawa R, Koike S, Nishimura Y, Sakurada H, et al. Severity-dependent and -independent brain regions of major depressive disorder: A long-term longitudinal near-infrared spectroscopy study. *J Affect Disord* (2019) 243:249–54. doi: 10.1016/j.jad.2018.09.029
81. Pu S, Nakagome K, Yamada T, Yokoyama K, Matsumura H, Nagata I, et al. Prefrontal activation predicts social functioning improvement after initial treatment in late-onset depression. *J Psychiatr Res* (2015) 62:62–70. doi: 10.1016/j.jpsychires.2015.01.009
82. Yamagata B, Yamanaka K, Takei Y, Hotta S, Hirano J, Tabuchi H, et al. Brain functional alterations observed 4-weekly in major depressive disorder following antidepressant treatment. *J Affect Disord* (2019) 252:25–31. doi: 10.1016/j.jad.2019.04.001
83. Aoki J, Iwahashi K, Ishigooka J, Fukumauchi F, Numajiri M, Ohtani N, et al. Evaluation of cerebral activity in the prefrontal cortex in mood [affective] disorders during animal-assisted therapy (AAT) by near-infrared spectroscopy (NIRS): A pilot study. *Int J Psychiatry Clin Pract* (2012) 16 (3):205–13. doi: 10.3109/13651501.2011.644565
84. Takamiya A, Hirano J, Ebuchi Y, Ogino S, Shimegi K, Emura H, et al. High-dose antidepressants affect near-infrared spectroscopy signals: A retrospective study. *NeuroImage Clin* (2017) 14:648–55. doi: 10.1016/j.nicl.2017.02.008

85. Shinba T, Kariya N, Matsuda S, Matsuda H, Obara Y. Increase of frontal cerebral blood volume during transcranial magnetic stimulation in depression is related to treatment effectiveness: A pilot study with near-infrared spectroscopy. *Psychiatry Clin Neurosci* (2018) 72(8):602–10. doi: 10.1111/pcn.12680
86. Usami M, Iwada Y, Kodaira M, Watanabe K, Saito K. Near Infrared Spectroscopy Study of the Frontopolar Hemodynamic Response and Depressive Mood in Children with Major Depressive Disorder: A Pilot Study. Yoshikawa T, editor. *PLoS One* (2014) 9(1):e86290. doi: 10.1371/journal.pone.0086290
87. Payzieva S, Maxmudova D. NIRS Study of the Effects of Computerized Brain Training Games for Cognitive Rehabilitation of Major Depressive Disorder Patients in Remission: A Pilot Study. *Stud Health Technol Inform*. (2014) 199:163–7.
88. Schiffer F, Johnston AL, Ravichandran C, Polcari A, Teicher MH, Webb RH, et al. Psychological benefits 2 and 4 weeks after a single treatment with near infrared light to the forehead: a pilot study of 10 patients with major depression and anxiety. *Behav Brain Funct BBF*. (2009) 5:46. doi: 10.1186/1744-9081-5-46
89. Eschweiler GW, Wegerer C, Schlotter W, Spandl C, Stevens A, Bartels M, et al. Left prefrontal activation predicts therapeutic effects of repetitive transcranial magnetic stimulation (rTMS) in major depression. *Psychiatry Res* (2000) 99(3):161–72. doi: 10.1016/S0925-4927(00)00062-7
90. Arnone D, McIntosh AM, Ebmeier KP, Munafò MR, Anderson IM. Magnetic resonance imaging studies in unipolar depression: systematic review and meta-regression analyses. *Eur Neuropsychopharmacol J Eur Coll Neuropsychopharmacol* (2012) 22(1):1–16. doi: 10.1016/j.euroneuro.2011.05.003
91. Ho CSH, Zhang MWB, Ho RCM. Optical Topography in Psychiatry: A Chip Off the Old Block or a New Look Beyond the Mind-Brain Frontiers? *Front Psychiatry* (2016) 7:74. doi: 10.3389/fpsy.2016.00074
92. Kirilina E, Jelzow A, Heine A, Niessing M, Wabnitz H, Brühl R, et al. The physiological origin of task-evoked systemic artefacts in functional near infrared spectroscopy. *NeuroImage*. (2012) 61(1):70–81. doi: 10.1016/j.neuroimage.2012.02.074
93. Caldwell M, Scholkman F, Wolf U, Wolf M, Elwell C, Tachtsidis I. Modelling confounding effects from extracerebral contamination and systemic factors on functional near-infrared spectroscopy. *NeuroImage*. (2016) 143:91–105. doi: 10.1016/j.neuroimage.2016.08.058
94. Hanly JR, Dewick HC, Davies ADM, Playeer J, Turnbull C. Verbal fluency in parkinson's disease. *Neuropsychologia*. (1990) 28(7):737–41. doi: 10.1016/0028-3932(90)90129-C
95. Zuckerman H, Pan Z, Park C, Brietzke E, Musial N, Shariq AS, et al. Recognition and Treatment of Cognitive Dysfunction in Major Depressive Disorder. *Front Psychiatry* (2018) 9:655. doi: 10.3389/fpsy.2018.00655
96. Freedman M. Frontal and parietal lobe dysfunction in depression: delayed alternation and tactile learning deficits. *Neuropsychologia* (1994) 32(8):1015–25. doi: 10.1016/0028-3932(94)90050-7
97. Li J, Xu C, Cao X, Gao Q, Wang Y, Wang Y, et al. Abnormal activation of the occipital lobes during emotion picture processing in major depressive disorder patients. *Neural Regener Res* (2013) 8(18):1693–701.
98. Mas S, Gassó P, Morer A, Calvo A, Bargalló N, Lafuente A, et al. Integrating Genetic, Neuropsychological and Neuroimaging Data to Model Early-Onset Obsessive Compulsive Disorder Severity. *PLoS One* (2016) 11(4):e0153846. doi: 10.1371/journal.pone.0153846

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Circular RNA in Schizophrenia and Depression

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OPEN ACCESS

Edited by:

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Universidad Miguel Hernández de
Elche, Spain

Reviewed by:

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Specialty section:

This article was submitted to
Molecular Psychiatry,
a section of the journal
Frontiers in Psychiatry

Received: 05 November 2019

Accepted: 17 April 2020

Published: 07 May 2020

Citation:

Li Z, Liu S, Li X, Zhao W, Li J
and Xu Y (2020) Circular RNA in
Schizophrenia and Depression.
Front. Psychiatry 11:392.
doi: 10.3389/fpsy.2020.00392

Schizophrenia (SZ) and depression (DEP) are two common major psychiatric disorders that are associated with high risk of suicide. These disorders affect not only physical and mental health, but they also affect the social function of the individual. However, diagnoses of SZ and DEP are mainly based on symptomatic changes and the clinical experience of psychiatrists. These rather subjective measures can induce misdiagnoses and missed diagnoses. Therefore, it is necessary to further explore objective indexes for improving the early diagnoses and prognoses of SZ and DEP. Current research indicates that non-coding RNA (ncRNA) may play a role in the occurrence and development of SZ and DEP. Circular RNA (circRNA), as an important component of ncRNA, is associated with many biological functions, especially post-transcriptional regulation. Since circRNA is easily detected in peripheral blood and has a high degree of spatiotemporal tissue specificity and stability, these attributes provide us with a new idea to further explore the potential value for the diagnosis and treatment of SZ and DEP. Here, we summarize the classification, characteristics, and biological functions of circRNA and the most significant results of experimental studies, aiming to highlight the involvement of circRNA in SZ and DEP.

Keywords: schizophrenia (SZ), depression (DEP), circular RNA (circRNA), biological function, expression, epigenetic characteristics

INTRODUCTION

Among the great quantity molecular regulatory factors affecting gene expression, non-coding RNA (ncRNA) is known to be a class of important regulatory factors. NcRNA refers to a kind of RNAs that do not encode proteins, but can participate in genetic regulatory processes at multiple levels, including interactions with DNA, RNA, and proteins through a variety of mechanisms (1). Gene regulation of ncRNAs *via* epigenetics plays an important role in many major diseases, such as schizophrenia (SZ) and depression (DEP) (2–4). At present, SZ is considered to represent an umbrella of genetic diseases that are mainly characterized by a certain number of genetic-interaction networks and their differential clinical symptoms (5). For DEP, studies have proposed a pathogenic model that involves interactions among the environment, genetics, and epigenetics (6). Moreover,

the high rate of comorbidity observed in psychiatric disorders indicates a large genetic correlation between each other, and the existence of shared molecular mechanisms involved in the occurrence and development of diseases (7). Sha Liu et al.'s genome-wide association study (GWAS) of seven psychiatric disorders (8), including SZ, DEP, bipolar disorder (BD), autism spectrum disorder (ASD), attention-deficit/hyperactivity disorder (ADHD), anxiety disorder (ANX), and neuroticism, also confirmed this. And they further found that chromosomes 5q14.3, 11q23.2, and 7p22.3 are the three genomic regions with the highest pleiotropic effects, suggesting that genetic factors have significant influence on SZ and DEP. Although few in-depth studies have been published on specific signaling pathways and networks regulated by ncRNAs in SZ and DEP, especially in terms of circular RNAs (circRNAs), it is possible that further elucidation of ncRNAs may be useful for better understanding the pathogenesis of SZ and DEP.

CircRNA, as an important type of ncRNA, forms an annular closed structure by undergoing reverse splicing, which was first discovered in plants in the 1970s (9). Since it was originally found only in a few transcriptional genes, circRNAs were considered to be a product of splicing errors from low-abundance transcriptional processes. However, this hypothesis has since been refuted. CircRNAs are widely present in human cells, some of which are expressed at as much as 10-fold higher levels compared to those of their linear isomer counterparts (10–12). CircRNA differs from linear RNA in that it is directly joined together to form a covalently closed structure without a 3' tail and 5' cap element (13–15). Due to the circular structure of circRNA, it is not easily degraded by RNA enzymes and also exhibits a high stability within cells (16, 17), enabling it to stably perform cellular functions over a longer period of time.

At present, the diagnostic criteria for SZ and DEP are still based solely on symptomology. Although diagnostic criteria—such as from the Diagnostic and Statistical Manual of Mental Disorders (Fifth Edition) (DSM-5) of American Psychiatric Association and the International Classification of Diseases (ICD-11)—are currently available, diagnoses and evaluations of treatment efficacies for SZ and DEP still primarily rely on clinical meetings with psychiatrists and lack objective physiological, biochemical, and pathological indicators. Hence, this more subjective diagnostic system can easily cause misdiagnoses and missed diagnoses. The above characteristics of circRNAs provide us with a novel approach to explore the biological basis for the occurrence and development of SZ and DEP. Therefore, the focus of this review is on circRNAs and their possible involvement in both SZ and DEP.

METHODS

We searched PubMed and GeenMedical for publications over the period of 1976 to 2019 and included the following the key phrases during searching: “circular RNA and neuropsychiatric

disorders” “circular RNA and schizophrenia” “circular RNA and depression” and “circular RNA and biological function”. Only papers in English were used in the preparation of this review. The references obtained were screened by the authors to determine which ones would be included for discussion in the present review.

OVERVIEW OF CIRCRNAS

Classifications and Looping Mechanisms of circRNAs

CircRNAs are divided into the following four categories: (1) whole-exonic-type circRNAs (EcircRNA); (2) circRNAs with introns and exons (EIcircRNA) (18); (3) lasso-type circRNAs composed of introns (ciRNA); and (4) circRNAs produced by cyclization of viral RNA genomes, ribosomal RNAs, small-nuclear RNAs, and transfer RNAs (tricRNA) (**Figure 1**). Based on their multiple biological mechanisms, circRNAs can retain the ability to either stay inside the nucleus or undergo nuclear export. This selective retention is important for determining the biological roles of different circRNAs and further provides us with research ideas for exploring the potential of circRNAs in the occurrence and development of SZ, DEP, and many other diseases. CircRNA is generally considered to be the result of reverse splicing. This indicates that the downstream 5' donor site is covalently cyclized with the upstream 3' receptor site (19). At present, there are three main looping mechanisms of circRNAs.

Cable-Tail-Insertion Cyclization Dependent on a Shearing Body

In eukaryotic cells, the formation of circRNAs by exon cable-tail-insertion cyclization (20, 21) is the earliest and most common way of looping, and more than 80% of circRNAs contain exons. Exon cable-tail-insertion cyclization relies on a typical shearing-body mechanism. On the mRNA precursor, the downstream 5' donor site of the exon is catalyzed to link to the upstream 3' receptor site by successively assembling small-nuclear ribosomal proteins to form cable-tail-insertion cyclization. Eventually, RNA is formed by further shearing. However, the specific mechanism underlying this process remains unclear.

Promotion of circRNA Formation by Cyclic-Acting Elements

Partial introns on both sides of circRNA exons contain reverse complementary sequences that can form RNA double chains side by side at shear sites (22). Finally, two kinds of different circRNAs with or without introns are formed by variable shearing. In general, a nucleotide sequence of 30–40 nt in length can promote the cyclization of circRNAs. Furthermore, exons inside can also compete for RNA pairing and ultimately form different types of circRNAs *via* variable shearing.

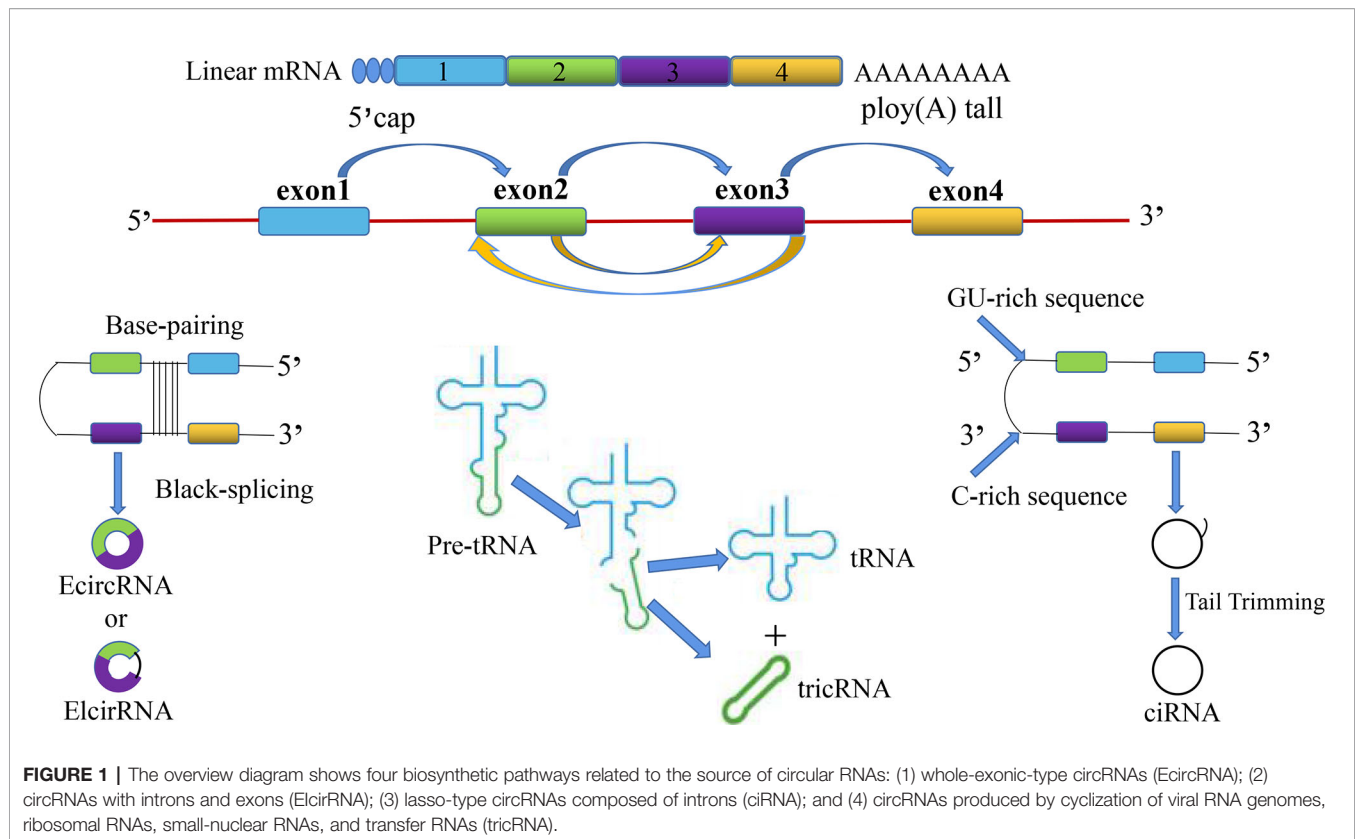


FIGURE 1 | The overview diagram shows four biosynthetic pathways related to the source of circular RNAs: (1) whole-exonic-type circRNAs (EcircRNA); (2) circRNAs with introns and exons (ElcirRNA); (3) lasso-type circRNAs composed of introns (ciRNA); and (4) circRNAs produced by cyclization of viral RNA genomes, ribosomal RNAs, small-nuclear RNAs, and transfer RNAs (tricRNA).

Regulation of circRNA Formation by RNA-Binding Proteins (RBPs)

In *Drosophila melanogaster*, MBL protein (23) can promote the formation of circRNAs by binding to the introns flanking an exon. Some studies (24) have shown that hundreds of circRNAs are regulated during the process of the epithelial-mesenchymal transition (EMT), and the looping formations of more than one-third of circRNAs are dynamically regulated by the splicing factor, Quaking (QKI). The effect of QKI on circRNA abundance depends on the binding motif of the QKI intron. Critically, the addition of a QKI motif is sufficient to re-induce the formation of circRNA from a linear splicing transcript. In contrast to the effect of QKI protein, high expression of ADAR1 protein (22) can inhibit the formation of circRNAs by destroying the RNA pairing flanking the exon.

Some exon-derived circRNAs are localized in the cytoplasm and contain both ribosome-binding sites and evolutionarily conserved termination codons. Exon-derived circRNAs also share the start codon of the host mRNA, which indicates that hundreds of endogenous circRNAs have translation potential (25), and some of their translation products can carry out functions. Previous studies have confirmed that many (31 out of 132) proteins encoded in ribosomal-related circRNAs (ribo-circRNAs) contain at least one identifiable protein domain. Furthermore, circRNA, such as circMbl, can be translated in a cap-independent manner *in vitro* (26). The same phenomenon is found for circ-ZNF609, which is associated with multiple

ribosomes. Circ-ZNF609 translates protein in a splicing-dependent and cap-independent manner. This gene controls the proliferation of primary myoblasts and is the key regulatory factor of their growth (27).

Molecular Biological Functions of circRNAs

Efficient microRNA Sponges

One of the most important molecular biological functions of circRNAs is that they indirectly affect the expression of target gene mRNAs by binding to microRNAs (miRNAs). As an important regulator at the post-transcriptional level, the function of miRNAs is mainly to downregulate or hinder the translation of target genes (mRNAs) by binding to the complementary 3'-end non-coding region located on the target mRNA (28, 29). As miRNA sponges, most circRNAs play an important role in transcriptional and translational regulation of mRNAs by competing for intracellular miRNAs (30, 31).

Among these miRNA sponges, Cdr1as circRNA is highly expressed and stable. It contains more than 70 conserved miRNA response elements (MREs) of miR-7 and one MRE of miR-671, which are then widely combined with AGO2, a key element of gene silencing. Bezzi and colleagues (32) provided evidence to support the functional association among Cdr1as, miR-7, and its target gene. A plasmid expressing Cdr1as was injected into zebrafish embryos to knockdown the expression of miR-7. This effect was partially rescued by injecting miR-7 precursor. At the

same time, miR-671 indirectly influenced the expression level of the miR-7 by triggering the degradation of Cdr1as. The interaction between circRNAs and miRNAs plays an important role in tumors, such as breast cancer, liver cancer and cervical cancer (31–35). Han et al. found that circMTO1 was significantly downregulated in human hepatocellular carcinoma (HCC) (36). Silencing of circMTO1, a miRNA-9 sponge, in patients with HCC downregulated the expression of the miRNA9 target gene, p21, thereby promoting the proliferation and invasion of HCC cells. Furthermore, the survival time of HCC cells with low expression of circMTO1 was reduced. This study also found that miRNA9 inhibitors could block the silencing effect of circMTO1. These findings suggest that circMTO1 may be used as an objective evaluation index of survival prognosis of HCC and may be a potential target for liver cancer treatment. In addition to circMTO1 acting as a sponge for miRNA-9 in HCC, hsa-circRNA-103809 (37) was also found to participate in the pathological mechanism of HCC. It could be used as a sponge for miR-377-3p to increase the expression of its target gene [fibroblast growth factor receptor 1 (FGFR1)] and promoted the proliferation and invasion of HCC cells. Moreover, hsa-circRNA-103809 short-hairpin RNA could act as a tumor inhibitor by downregulating FGFR1 in HCC.

Templates for Translating Proteins

N6-methyladenosine (m6A) is the most abundant base modification of RNA. A previous study found (38) that the m6A motif was enriched in circRNAs, and that a single m6A site was sufficient to drive the initiation of circRNA protein translation in human cells. However, this protein translation of circRNA driven by m6A did not function without the following:

the activation factor, eIF4G2; the m6A reader, YTHDF 3; enhancement of the methyltransferase, METTL 3/14; inhibition and upregulation of demethylase during heat shock, FTO. Interestingly, further analysis indicated that m6A-driven circRNA translation is widespread.

Interactions With RBPs

Interactions between circRNAs and RBPs are not only related to their own biological occurrences, but they are also involved in the post-transcriptional regulation of RNAs (39). In general, the cell-cycle protein, cyclin-dependent kinase 2 [also known as cell-division protein kinase 2 (CDK2)], interacts with the cell-cycle proteins A and E to facilitate the cell cycle. However, cyclin-dependent kinase inhibitor 1 (p21) can inhibit these interactions and prevent cell-cycle progression. Du et al. (40) found that circ-Foxo3 was highly expressed in non-cancer cells, forming a ternary complex to inhibit the cell cycle by binding to CDK2 and p21. Additionally, silencing of endogenous circ-Foxo3 promoted cell proliferation. Meanwhile, Zhu et al. (41) showed that circZKSCAN1 exerted its inhibitory effect by competitively binding to the RNA-binding protein (RBP), FMRP. This mechanism blocked the binding of FMRP to the β -catenin-binding protein cell cycle and apoptosis regulator 1 (CCAR1) mRNA, and subsequently inhibited the transcriptional activity of the Wnt signal transduction pathway. In addition, downregulation of the RNA-splicing protein, Quaking5, in HCC tissue caused a decrease of circZKSCAN1. These findings suggest the formation of a Qki5-circZKSCAN1-FMRP-CCAR1-Wnt signaling pathway, which may be useful as a potential therapeutic target for HCC (Figure 2).

The above results indicate that circRNAs act not only as miRNA sponges by competing with intracellular miRNA to

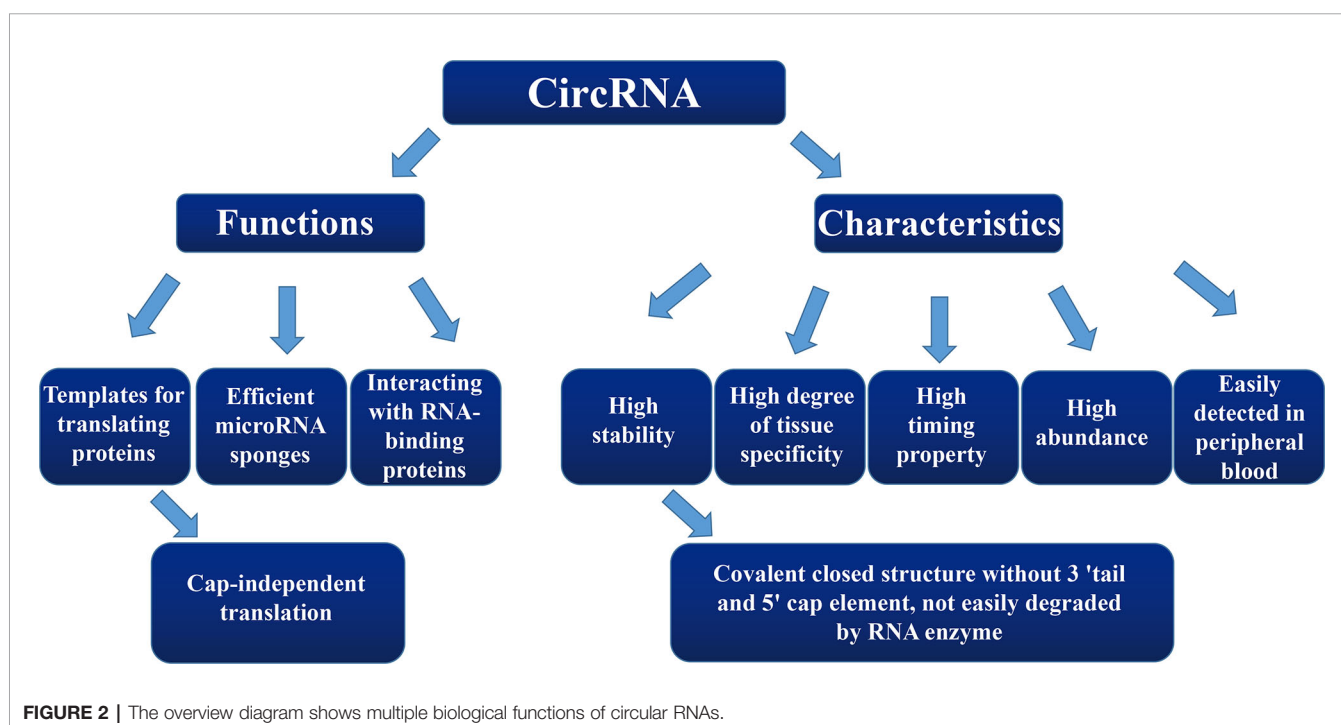


TABLE 1 | Molecular biological function of circRNAs

| Author | Main findings |
|----------------------|---|
| Chen and Sarnow (25) | Hundreds of endogenous circRNAs with ribosome binding sites shared the same start codon of their hosting mRNA, which meant they had translation potential. |
| Bezzi et al. (32) | Cdr1as knocked down the expression of miR-7, and this effect was partially rescued by injecting miR-7 precursor. At the same time, miR-671 indirectly influenced the expression level of miR-7 by triggering the degradation of Cdr1as. |
| Yang et al. (38) | N6-methyladenosine (m6A) motif was enriched in circRNA, and a single m6A site was sufficient to drive the initiation of circRNA protein translation in human cells. |
| Han et al. (36) | Silencing of circMTO1, a miRNA-9 sponge, in patients with HCC could downregulate the expression of miRNA9 target gene p21, thereby promoting the proliferation and invasion of HCC cells. |
| Legnini et al. (27) | Circ-ZNF609 translated protein in a splicing-dependent and cap-independent manner, which controlled the proliferation of primary myoblasts and was the key regulatory factor of their growth. |
| Zhu et al. (41) | CircZKSCAN1 exerted its inhibitory effect by competitively binding to the RNA binding protein (FMRP). Therefore, the Qki5-circZKSCAN1-FMRP-CCAR1-Wnt signal axis could be used as a potential therapeutic target for HCC treatment. |
| Zhan et al. (37) | Hsa-circRNA-103809 could be used as a sponge for miR-377-3p to increase the expression of its target gene [fibroblast growth factor receptor 1 (FGFR1)] and promoted the proliferation and invasion of HCC cells. |

CircRNAs can act not only as miRNA sponges by competing with intracellular miRNA to influence the expression of its target genes but also as a translation templates for proteins. They can also combine with RBPs directly and realize the regulation of post-transcriptional RNA, which plays an important role in the occurrence and development of diseases.

influence the expression of target genes, but they also act as translational templates for proteins. More importantly, circRNAs lead to altered gene expression outcomes by splicing their corresponding precursor transcripts, which may play an important role in the occurrence and development of various diseases (Table 1).

CIRCRNAS AND SZ

SZ is one of the most common mental disorders, with a global prevalence of 1%, which brings a heavy burden to affected families and society (42). According to the latest research conducted by Huang et al. (43), the prevalence of SZ is 0.559% in China. Symptoms of SZ are divided into positive, negative, and cognitive categories (44, 45). The diagnosis of SZ is mainly based on symptomatic changes and the clinical experience of psychiatrists. However, these rather subjective measures can cause misdiagnoses and missed diagnoses. Therefore, it is necessary to further explore objective indexes to improve the early diagnosis and prognosis of SZ. Since circRNAs are easily detected in peripheral blood and have a high degree of spatiotemporal tissue specificity and stability (46, 47), these properties provide us with a novel approach to further explore the mechanisms underlying the occurrence and development of SZ.

Molecular Mechanisms of circRNAs in SZ

As representative antisense nucleic-acid sponge sequences, circRNAs can competitively bind to miRNAs *via* MREs, which negatively influence the regulatory effects of miRNAs at the post-transcriptional level. MiRNA-320a-3p, miRNA-320b, miRNA-181b-5p, 21-5p, 195-5p, 137, 346, 34a-5p, and hsa-miRNA-206 (48–50) have all been confirmed to play important roles in the occurrence and development of SZ. Wei et al. (51) found that eight miRNAs were upregulated in patients with SZ, as compared to corresponding levels in control participants. Through quantitative reverse-transcription polymerase chain reaction (qRT-PCR) assays and a follow-up study on 400 patients with SZ who received regular atypical antipsychotic treatment for 12 months, this study ultimately showed that miR-130b and miR-193a-3p were upregulated in SZ, and the possibility of time-dependent changes were ruled out. Hence, miR-130b and miR-193a-3p may be used as state-independent biomarkers for SZ. A recent study (52) has confirmed that the expression of the network axis composed of miR-30a-5p, its transcription factor, EGR1 and target gene, NEUROD1, changed with the disease state in the patients with SZ before and after treatment. Further receiver operating characteristic (ROC) curve analysis revealed that compared with the single miR-30a-5p, the EGR1-miR-30a-5p-NEUROD1 molecular regulatory axis had higher diagnostic value for predicting SZ. Hence, these findings suggest a possibility for the miRNA-TF-gene regulatory network/axis as a novel diagnostic marker for SZ.

Due to their circular structure, circRNAs are highly conserved across species. Some studies have confirmed that circRNAs are highly expressed in the brain and are differentially expressed in distinct tissues and disease states (53, 54). Moreover, circRNAs exhibit dynamic expression levels during neural development, including neural differentiation and maturation. For instance, circRNAs are upregulated during neuronal differentiation, and compared with their mRNA homologues, they generally exhibit differential expression (55–57). SZ often occurs in late adolescence and early adulthood. Previous studies have suggested that excessive pruning of synapses in the brain during growth and development may contribute to the pathogenesis of SZ (58).

Some studies have suggested synaptic localization of circRNAs. Wolf et al. (56) found that circRNAs in the nervous system were generally upregulated compared to those in the thyroid, liver, and muscle, and that expression varied among different brain regions. Especially in the cerebellum, the expression ratio of circular to linear RNAs was significantly increased, which correlated with a larger number of neurons in the cerebellum compared to that in other brain regions. Furthermore, by measuring high-purity synaptosomal components, this study found that brain-expressed circRNAs had a strong enriching effect at synapses. Another study found that circStau2a was mainly located at synapses, whereas its linear transcript, mRNAStau2, was almost completely localized to the cytoplasm (56). Similarly, circRNA derived from the known neuronal differentiation regulator, RMST, exhibits a high synaptic enrichment rate (59). It has also been found that

during the development of the hippocampus, upregulated circRNAs are produced by the gene locus that simultaneously encodes proteins rich in synapse-related functions. Conversely, no enrichment of encoding of any other functional class of proteins was found in the gene locus that produced a downregulated dynamic expression pattern of circRNAs (29, 60, 61). In summary, the expression levels of circRNAs are regulated by neuronal development. Many circRNAs also change the structures of neurons, which are largely independent of the function of their linear transcripts.

Expression of circRNAs in Postmortem SZ Brains

Our literature review showed that the most extensive studies of circRNAs and SZ have involved the analysis of the expression profiles of circRNAs based on control studies of patients with SZ and the healthy controls matched to them. Mahmoudi et al. used circRNA enrichment sequencing to analyze the expression profiles of circRNAs in the cerebral cortex (BA46) of 35 postmortem patients with SZ as well as those of healthy controls (62). They found that more than 95,000 types of circRNAs in the human dorsolateral prefrontal cortex (DLPFC) showed significant diversity, and that half of them had not previously been reported, such as circHomer1, circKhlh2, circMpped1, and circNell2 (63, 64). A differential analysis of these circRNAs revealed that 390 circRNAs were downregulated and 240 were completely deleted in patients with SZ. In addition to the reduced overall level of gene expression in circRNAs, the total numbers of unique circRNAs found in SZ were also reduced compared to those in healthy controls, and there was a large overlap between them. For instance, circNELL2, which is upregulated after synaptic plasticity induction, was downregulated in patients with SZ. The researchers also explored the correlation between differentially expressed circRNAs and their linear isoforms. Surprisingly, the expression levels of more than half of the circRNAs showed an inverse relationship with those of their linear RNAs, suggesting that both forms of transcripts may sometimes antagonize each other's biosynthesis. Zimmerman et al. (65) also discovered circHomer1a, which is highly expressed and neuron-enriched in the frontal cortex. Interestingly, in the prefrontal cortex (PFC) and induced pluripotent stem cell-derived neuronal cultures in patients with SZ and BD, circHomer1a was found to be significantly downregulated. Additionally, changes in circHomer1a expression in the DLPFC and orbitofrontal cortex (OFC) were positively correlated with the age of onset of SZ. In addition, circHomer1a was found to interact with RNA-binding protein, HuD, through animal-level verification, which was involved in regulating synaptic gene expression and cognitive flexibility. These findings are of great significance for exploring the molecular mechanisms underlying the pathogenesis of mental disorders.

The above results suggest that the overall expression levels of circRNAs are downregulated in patients with SZ, while there are still some unique and significantly differentially expressed circRNAs that differ from this overall trend. These circRNAs

may play an important role in the clinical phenotypes of SZ by regulating corresponding cellular metabolic pathways underlying this mental disorder. Furthermore, there is a strong possibility of abnormal regulation of circRNAs in other mental illnesses and behavioral disorders.

Expression of circRNAs in Peripheral Blood of Patients With SZ

CircRNA plays a particularly important role in the regulation of gene expression in SZ (62, 66, 67). Previous studies (68) have identified that differentially expressed circRNAs from peripheral blood perhaps contribute on the development of the diagnosis and treatment of SZ.

A case-control study carried out by Yao and colleagues (69) screened nine kinds of significantly expressed circRNAs. Further verification by qRT-PCR revealed that three circRNAs were downregulated and two circRNAs were upregulated in the SZ group. The expression levels of circRNAs in patients with SZ after 4 and 8 weeks of conventional antipsychotic treatment were then re-quantified to see if there was any change. ROC curve analysis showed that three circRNAs (hsa circRNA 103704, hsa circRNA 101836, and hsa circRNA 104597) were of diagnostic significance, with a sensitivity of 87.25% and specificity of 85.44%. In particular, the sensitivity and specificity of hsa circRNA 104597 were 84.31% and 86.41%, respectively, indicating that it may be useful in the diagnostic and therapeutic evaluation of SZ.

The above results suggest that circRNAs may provide novel ideas for exploring objective biomarkers for SZ diagnosis. However, there has only been one study on this topic, which had some limitations. For instance, the sample size of screening differentially expressed circRNAs was relatively small. Since patients with SZ had been treated with standardized antipsychotic drugs for a relatively short period of time, changes in the expression level of circRNAs before and after treatment may also be due to their own time-dependent changes. Furthermore, only one type of circRNA was found to may represent a novel objective diagnostic value for SZ. Finally, there are still many significantly expressed circRNAs related to SZ that require further exploration (69–71).

CIRC RNAs AND DEPRESSION

Depression (DEP) is one of the most common mental disorders in humans. The World Health Organization (WHO) has reported that more than 800 million people worldwide suffer from DEP (72, 73). Approximately 40–60% of patients with DEP show suicidal ideation or behavior, and the suicide rate has been reported to be as high as 15% (74, 75). The disease burden of DEP is increasing year by year. In 1990, DEP ranked fifth in terms of its global burden as a disease. However, DEP is expected to become the top disease burden in China by 2030 (76). The process of diagnosing this disease is similar to that for SZ and, as with SZ, it can easily lead to misdiagnoses and missed diagnoses. Therefore, it is necessary to further explore the pathological

mechanisms and specific molecular markers related to DEP to improve the early diagnosis and prognosis of DEP.

Expression of circRNAs in Peripheral Blood in Depression

Cui and colleagues randomly selected peripheral blood mononuclear cells (PBMCs) from five patients with DEP and five healthy controls to analyze the expression profile of circRNAs. First, 15 differentially expressed circRNAs (including four upregulated and 11 downregulated circRNAs) were screened (77). Then, four significantly differentially expressed circRNAs were identified through verification in 100 DEP patients, 103 healthy controls, and 30 randomly selected DEP patients after 4 and 8 weeks of antidepressant therapy. However, only hsa-circRNA-103636 showed significant changes in DEP patients after 8 weeks of antidepressant treatment. Further ROC curve analysis showed that the sensitivity and specificity of this gene were 0.73 and 0.65, respectively. Therefore, hsa-circRNA-103636 may have potential diagnostic value of DEP.

Many studies have shown that type-2 diabetes (T2DM) is closely related to the occurrence and development of DEP (78, 79). Tian et al. collected venous plasma from five patients with T2DM and five cases of T2DM with DEP to determine the expression profiles of ncRNAs (including lncRNAs, circRNAs, and mRNAs) in order to discover specific molecular markers associated with DEP. The results of screening identified 28 lncRNAs, 107 circRNAs (including 75 upregulated and 32 downregulated circRNAs), and 89 mRNAs in the differential expression profiles of patients with DEP (80). Compared with that in the T2DM group, circRNA-TFRC had a significantly higher expression level in the DEP group. This circRNA is related to insulin resistance, which indicates that patients with DEP may have more severe insulin resistance symptoms. Some studies (81) have found that the expression level of TFRC is associated with mental disorders, suggesting that TFRC may be useful as a molecular target for studying the pathogenesis of DEP. Another upregulated circRNA-TNIK [Traf 2- and Nck-interacting kinase (TNIK)] is also associated with DEP. TNIK is a serine/threonine kinase that exhibits a high expression level in the brain, which has a good effect on the development and synaptic transmission of dendritic cells. A previous study (80) found that compared with a T2DM group, the expression level of TNIK was significantly increased in the DEP group. This suggests that the function of TNIK merits further investigation.

Jiang and colleagues (82) used microarrays to analyze plasma samples from seven patients with T2DM and seven patients with T2DM and DEP. Compared with the T2DM group, 183 circRNAs were upregulated and 64 circRNAs were downregulated in T2DM patients with DEP. Among them, the upregulated gene, DCP2 (hsa-circRNA-001520), may be used as a type of de-capping enzyme and plays an important role in neuronal development and mental retardation by degrading mRNAs. Another downregulated gene, CSGALNACT1 (hsa-circRNA-001781), is associated with an antidepressant response (83). In addition, Zhang et al. demonstrated that circDYM was also significantly decreased in MDD patients and two depressive-like mouse models [induced *via* chronic unpredictable pressure (CUS) and lipopolysaccharide (LPS)]. Recovery of circDYM levels significantly ameliorated

CUS- or LPS-induced depressive-like behavior, which suggests that circDYM may be a novel therapeutic target for DEP (84).

According to the target genes of circRNAs related to DEP predicted by TargetScan and Miranda, 18 miRNAs and 529 mRNAs were found to interact with 4 circRNAs. Interestingly, hsa-miR-761 and hsa-miR-298 are common targets for hsa-circRNA-003251, hsa-circRNA-005019, and hsa-circRNA-015115. A previous study found that hsa-circRNA-003251 and hsa-circRNA-015115 may play important roles in the circRNA-

TABLE 2 | Expression of circRNAs in schizophrenia and depression

| Author | Main findings |
|----------------------|--|
| Ng et al. (59) | CircRNA derived from the known neuronal differentiation regulator RMST had a very high synaptic enrichment rate. |
| You et al. (60) | During the development of the hippocampus, the upregulated circRNA is produced by the gene locus that simultaneously encodes protein rich in synapse-related functions. |
| Hanan et al. (57) | CircRNA in the nervous system is generally upregulated compared to the thyroid, liver, and muscle. Especially in the cerebellum, the expression ratio of circular and linear RNA is significantly increased, and the brain-expressed circRNAs, such as circStau2a, have a strong enrichment effect on synaptic-neurons. |
| Mahmoudi et al. (62) | A total of 390 circRNAs, such as circNEIL2, were down-regulated and 240 were completely deleted in patients with SZ. In addition to the reduced overall level of circRNA expression, the total numbers of unique circRNAs found in SZ were also reduced. More than half of circRNAs might sometimes antagonize linear RNA biosynthesis. |
| Yao et al. (68) | Three circRNAs (hsa circRNA 103704, hsa circRNA 101836, and hsa circRNA 104597) were of diagnostic significance, with a sensitivity of 87.25% and a specificity of 85.44%. In particular, the sensitivity and specificity of hsa circRNA 104597 were 84.31% and 86.41%, respectively, indicating that it might be a new potential biomarker for the diagnostic and therapeutic evaluation of SZ. |
| Abasolo et al. (81) | The expression level of TFRC is associated with mental disorders, suggesting that TERC could use as a molecular target for studying the pathogenesis of DEP. |
| Cui et al. (77) | Fifteen differentially expressed circRNAs (including 4 upregulated and 11 downregulated circRNAs) were screened out by a case-control study. However, only hsa-circRNA-103636 showed significant changes in depressed patients after 8 weeks of antidepressant treatment, indicating that it could be used as a new potential biomarker for the diagnosis and treatment of DEP. |
| Jiang et al. (82) | Compared with the T2DM group, 183 circRNAs were upregulated and 64 circRNAs were downregulated in T2DM patients with DEP. Hsa-circRNA-003251 and hsa-circRNA-015115 might play important roles in the circRNA-miRNA-mRNA network associated with DEP by acting as hsa-miR-761 sponges. In addition to the above two RNAs, hsa-circRNA-100918 and hsa-circRNA-001520 might be involved in the Wnt signaling pathway of thyroid hormone. |
| An (80) | Compared with the T2DM group, circRNA-TFRC had a higher expression level and circRNA-TNIK was significantly increased in the DEP group. TFRC was related to insulin resistance, which indicated that patients with DEP had more severe insulin resistance symptoms. TNIK had a good effect on the development and synaptic transmission of dendritic cells. |

CircRNAs have specific expression profiles in brain regions and peripheral venous blood. Based on these, researchers used case-control studies to explore significantly expressed circRNAs in schizophrenia and depression in order to find the molecular bases of these two diseases.

miRNA-mRNA network associated with DEP by acting as an hsa-miR-761 sponge. Hsa-miR-761 (85, 86) has been shown to be involved in the regulation of mitochondrial networks and to promote the generation of learning and memory. Furthermore, overexpression of hsa-miR-761 inhibits the p38-MAPK signaling pathway (87). Therefore, hsa-miR-761 may participate in the pathological mechanisms of DEP. In addition, KEGG pathway analysis predicted that upregulation of hsa-circRNA-003251, hsa-circRNA-015115, hsa-circRNA-100918, and hsa-circRNA-001520 might be involved in the Wnt signaling pathway implicated in thyroid hormones. Another study showed that thyroid hormones affected mood and promoted signal pathways in the brain (88). These findings suggest that the occurrence and development of DEP might also be related to changes and metabolic disorders involving thyroid hormones (Table 2).

Based on the above findings, circRNA is a kind of biological signal that is measurable in physiological, pathological, and therapeutic process and perhaps contribute on predicting risk assessments, early diagnoses, and monitoring of responses in patients with SZ or DEP. At present, although some studies on the specific expression levels of circRNAs in various diseases have emerged, most of them have been at the level of expression profiles. Therefore, further studies are needed to better elucidate the precise associations between circRNAs and clinical symptoms in various diseases.

LIMITATIONS AND FUTURE RESEARCH

At present, an increasing number of studies on circRNAs related to SZ and DEP are emerging. However, there are still

many questions to be answered, such as the following: whether changes in a single circRNA or among a group of circRNAs are specific to SZ and DEP; whether levels of circRNAs change with age; whether the expression of circRNAs are affected by smoking or drug abuse; and whether the circRNAs in cellular compartments have any differences in terms of their molecular bases of SZ and DEP (e.g., extra-vesicular circRNAs). These limitations indicate fruitful directions for future studies that may help elucidate the potential diagnostic and therapeutic value of circRNAs for SZ and DEP.

AUTHOR CONTRIBUTIONS

YX designed and supervised the study. ZL and SL drafted the manuscript. XL, WZ, and JL collected some literature.

FUNDING

This work was supported by the National Natural Science Foundation of China, (81701326 and 81971601), the Support Program of the Youth Sanjin Scholars, and the 136 Medical Rejuvenation Project of Shanxi Province.

ACKNOWLEDGMENTS

The authors are grateful to the National Natural Science Foundation of China for research funding.

REFERENCES

- Stefanska B, MacEwan DJ. Epigenetics and pharmacology. *Br J Pharmacol* (2015) 172(11):2701–4. doi: 10.1111/bph.13136
- Wang Q, Roy B, Dwivedi Y. Co-expression network modeling identifies key long non-coding RNA and mRNA modules in altering molecular phenotype to develop stress-induced depression in rats. *Transl Psychiatry* (2019) 9(1):125. doi: 10.1038/s41398-019-0448-z
- Nestler EJ, Peña CJ, Kundakovic M, Mitchell A, Akbarian S. Epigenetic Basis of Mental Illness. *Neuroscientist* (2016) 22(5):447–63. doi: 10.1177/1073858415608147
- Perkins DO, Jeffries C, Sullivan P. Expanding the ‘central dogma’: the regulatory role of nonprotein coding genes and implications for the genetic liability to schizophrenia. *Mol Psychiatry* (2005) 10(1):69–78. doi: 10.1038/sj.mp.4001577
- Arnedo J, Svrakic DM, Del Val C, Romero-Zaliz R, Hernández-Cuervo H, Fanous AH, et al. Uncovering the hidden risk architecture of the schizophrenias: confirmation in three independent genome-wide association studies. *Am J Psychiatry* (2015) 172(2):139–53. doi: 10.1176/appi.ajp.2014.14040435
- Mill J, Petronis A. Molecular studies of major depressive disorder: the epigenetic perspective. *Mol Psychiatry* (2007) 12(9):799–814. doi: 10.1038/sj.mp.4001992
- Kessler RC, Chiu WT, Demler O, Merikangas KR, Walters EE. Prevalence, severity, and comorbidity of 12-month DSM-IV disorders in the National Comorbidity Survey Replication. *Arch Gen Psychiatry* (2005) 62(6):617–27. doi: 10.1001/archpsyc.62.6.617
- Liu S, Rao S, Xu Y, Li J, Huang H, Zhang X, et al. Identifying common genome-wide risk genes for major psychiatric traits. *Hum Genet* (2020) 139(2):185–98. doi: 10.1007/s00439-019-02096-4
- Sanger HL, Klotz G, Riesner D, Gross HJ, Kleinschmidt AK. Viroids are single-stranded covalently closed circular RNA molecules existing as highly base-paired rod-like structures. *Proc Natl Acad Sci U S A* (1976) 73(11):3852–6. doi: 10.1073/pnas.73.11.3852
- Jeck WR, Sorrentino JA, Wang K, Slevin MK, Burd CE, Liu J, et al. Circular RNAs are abundant, conserved, and associated with ALU repeats. *RNA* (2013) 19(2):141–57. doi: 10.1261/rna.035667.112
- Wang PL, Bao Y, Yee MC, Barrett SP, Hogan GJ, Olsen MN, et al. Circular RNA is expressed across the eukaryotic tree of life. *PLoS One* (2014) 9(6):e90859. doi: 10.1371/journal.pone.0090859
- Jeck WR, Sharpless NE. Detecting and characterizing circular RNAs. *Nat Biotechnol* (2014) 32(5):453–61. doi: 10.1038/nbt.2890
- Capel B, Swain A, Nicolis S, Hacker A, Walter M, Koopman P, et al. Circular transcripts of the testis-determining gene Sry in adult mouse testis. *Cell* (1993) 73(5):1019–30. doi: 10.1016/0092-8674(93)90279-Y
- Danan M, Schwartz S, Edelheit S, Sorek R. Transcriptome-wide discovery of circular RNAs in Archaea. *Nucleic Acids Res* (2012) 40(7):3131–42. doi: 10.1093/nar/gkr1009
- Puttaraju M, Been MD. Group I permuted intron-exon (PIE) sequences self-splice to produce circular exons. *Nucleic Acids Res* (1992) 20(20):5357–64. doi: 10.1093/nar/20.20.5357
- Salzman J, Gawad C, Wang PL, Lacayo N, Brown PO. Circular RNAs are the predominant transcript isoform from hundreds of human genes in diverse cell types. *PLoS One* (2012) 7(2):e30733. doi: 10.1371/journal.pone.0030733

17. Vincent HA, Deutscher MP. Substrate recognition and catalysis by the exoribonuclease RNase R. *J Biol Chem* (2006) 281(40):29769–75. doi: 10.1074/jbc.M606744200
18. Li Z, Huang C, Bao C, et al. Exon-intron circular RNAs regulate transcription in the nucleus. *Nat Struct Mol Biol* (2015) 22(3):256–64. doi: 10.1038/nsmb.2959
19. Wilusz JE, Sharp PA. Molecular biology. A circuitous route to noncoding RNA. *Science* (2013) 340(6131):440–1. doi: 10.1126/science.1238522
20. Lu T, Cui L, Zhou Y, Zhu C, Fan D, Gong H, et al. Transcriptome-wide investigation of circular RNAs in rice. *RNA* (2015) 21(12):2076–87. doi: 10.1261/rna.052282.115
21. Barrett SP, Wang PL, Salzman J. Circular RNA biogenesis can proceed through an exon-containing lariat precursor. *Elife* (2015) 4:e07540. doi: 10.7554/eLife.07540
22. Ivanov A, Memczak S, Wyler E, Torti F, Porath HT, Orejuela MR, et al. Analysis of intron sequences reveals hallmarks of circular RNA biogenesis in animals. *Cell Rep* (2015) 10(2):170–7. doi: 10.1016/j.celrep.2014.12.019
23. Ashwal-Fluss R, Meyer M, Pamudurti NR, Ivanov A, Bartok O, Hanan M, et al. circRNA biogenesis competes with pre-mRNA splicing. *Mol Cell* (2014) 56(1):55–66. doi: 10.1016/j.molcel.2014.08.019
24. Conn SJ, Pillman KA, Toubia J, Conn VM, Salamanidis M, Phillips CA, et al. The RNA binding protein quaking regulates formation of circRNAs. *Cell* (2015) 160(6):1125–34. doi: 10.1016/j.cell.2015.02.014
25. Chen CY, Sarnow P. Initiation of protein synthesis by the eukaryotic translational apparatus on circular RNAs. *Science* (1995) 268(5209):415–7. doi: 10.1126/science.7536344
26. Pamudurti NR, Bartok O, Jens M, Ashwal-Fluss R, Stottmeister C, Ruhe L, et al. Translation of CircRNAs. *Mol Cell* (2017) 66(1):9–21.e7. doi: 10.1016/j.molcel.2017.02.021
27. Legnini I, Di Timoteo G, Rossi F, Morlando M, Briganti F, Sthandier O, et al. Circ-ZNF609 Is a Circular RNA that Can Be Translated and Functions in Myogenesis. *Mol Cell* (2017) 66(1):22–37.e9. doi: 10.1016/j.molcel.2017.02.017
28. Kim VN. MicroRNA biogenesis: coordinated cropping and dicing. *Nat Rev Mol Cell Biol* (2005) 6(5):376–85. doi: 10.1038/nrm1644
29. Bartel DP. MicroRNAs: target recognition and regulatory functions. *Cell* (2009) 136(2):215–33. doi: 10.1016/j.cell.2009.01.002
30. Salmena L, Poliseno L, Tay Y, Kats L, Pandolfi PP. A ceRNA hypothesis: the Rosetta Stone of a hidden RNA language. *Cell* (2011) 146(3):353–8. doi: 10.1016/j.cell.2011.07.014
31. Memczak S, Jens M, Elefsinioti A, Torti F, Krueger J, Rybak A, et al. Circular RNAs are a large class of animal RNAs with regulatory potency. *Nature* (2013) 495(7441):333–8. doi: 10.1038/nature11928
32. Bezzi M, Guarnerio J, Pandolfi PP. A circular twist on microRNA regulation. *Cell Res* (2017) 27(12):1401–2. doi: 10.1038/cr.2017.136
33. Hansen TB, Jensen TI, Clausen BH, Bramsen JB, Finsen B, Damgaard CK, et al. Natural RNA circles function as efficient microRNA sponges. *Nature* (2013) 495(7441):384–8. doi: 10.1038/nature11993
34. Tang W, Ji M, He G, Yang L, Niu Z, Jian M, et al. Silencing CDR1as inhibits colorectal cancer progression through regulating microRNA-7. *Oncotargets Ther* (2017) 10:2045–56. doi: 10.2147/OTT.S131597
35. Piwecka M, Glažar P, Hernandez-Miranda LR, Memczak S, Wolf SA, Rybak-Wolf A. Loss of a mammalian circular RNA locus causes miRNA deregulation and affects brain function. *Science* (2017) 357(6357). doi: 10.1126/science.aam8526
36. Han D, Li J, Wang H, Su X, Hou J, Gu Y, et al. Circular RNA circMTO1 acts as the sponge of microRNA-9 to suppress hepatocellular carcinoma progression. *Hepatology* (2017) 66(4):1151–64. doi: 10.1002/hep.29270
37. Zhan W, Liao X, Chen Z, et al. Circular RNA hsa_circRNA_103809 promoted hepatocellular carcinoma development by regulating miR-377-3p/FGFR1/ERK axis. *J Cell Physiol* (2019) 235(2):1733–45. doi: 10.1002/jcp.29092
38. Yang Y, Fan X, Mao M, Song X, Wu P, Zhang Y, et al. Extensive translation of circular RNAs driven by N6-methyladenosine. *Cell Res* (2017) 27(5):626–41. doi: 10.1038/cr.2017.31
39. Janga SC, Mittal N. Construction, structure and dynamics of post-transcriptional regulatory network directed by RNA-binding proteins. *Adv Exp Med Biol* (2011) 722:103–17. doi: 10.1007/978-1-4614-0332-6_7
40. Du WW, Yang W, Liu E, Yang Z, Dhaliwal P, Yang BB. Foxo3 circular RNA retards cell cycle progression via forming ternary complexes with p21 and CDK2. *Nucleic Acids Res* (2016) 44(6):2846–58. doi: 10.1093/nar/gkw027
41. Zhu YJ, Zheng B, Luo GJ, Ma XK, Lu XY, Lin XM, et al. Circular RNAs negatively regulate cancer stem cells by physically binding FMRP against CCAR1 complex in hepatocellular carcinoma. *Theranostics* (2019) 9(12):3526–40. doi: 10.7150/thno.32796
42. Cohen CI, Meesters PD, Zhao J. New perspectives on schizophrenia in later life: implications for treatment, policy, and research. *Lancet Psychiatry* (2015) 2(4):340–50. doi: 10.1016/S2215-0366(15)00003-6
43. Huang Y, Wang Y, Wang H, Liu Z, Yu X, Yan J, et al. Prevalence of mental disorders in China: a cross-sectional epidemiological study. *Lancet Psychiatry* (2019) 6(3):211–24. doi: 10.1016/S2215-0366(18)30511-X
44. Schultz SK, Andreasen NC. Schizophrenia. *Lancet* (1999) 353(9162):1425–30. doi: 10.1016/S0140-6736(98)07549-7
45. Bassett AS, Collins EJ, Nuttall SE, Honer WG. Positive and negative symptoms in families with schizophrenia. *Schizophr Res* (1993) 11(1):9–19. doi: 10.1016/0920-9964(93)90033-F
46. Xia S, Feng J, Lei L, Hu J, Xia L, Wang J, et al. Comprehensive characterization of tissue-specific circular RNAs in the human and mouse genomes. *Brief Bioinform* (2017) 18(6):984–92.
47. Maass PG, Glažar P, Memczak S, Dittmar G, Hollfinger I, Schreyer L, et al. A map of human circular RNAs in clinically relevant tissues. *J Mol Med (Berl)* (2017) 95(11):1179–89. doi: 10.1007/s00109-017-1582-9
48. Liu S, Zhang F, Wang X, Shugart YY, Zhao Y, Li X, et al. Diagnostic value of blood-derived microRNAs for schizophrenia: results of a meta-analysis and validation. *Sci Rep* (2017) 7(1):15328. doi: 10.1038/s41598-017-15751-5
49. Wang Y, Wang J, Guo T, Peng Y, Wang K, Bai K, et al. Screening of schizophrenia associated miRNAs and the regulation of miR-320a-3p on integrin $\beta 1$. *Med (Baltimore)* (2019) 98(8):e14332. doi: 10.1097/MD.00000000000014332
50. Du Y, Yu Y, Hu Y, Li X-W, Wei Z-X, Pan R-Y, et al. Genome-Wide, Integrative Analysis Implicates Exosome-Derived MicroRNA Dysregulation in Schizophrenia. *Schizophr Bull* (2019) 45(6):1257–66. doi: 10.1093/schbul/sby191
51. Wei H, Yuan Y, Liu S, Wang C, Yang F, Lu Z, et al. Detection of circulating miRNA levels in schizophrenia. *Am J Psychiatry* (2015) 172(11):1141–7. doi: 10.1176/appi.ajp.2015.14030273
52. Liu S, Zhang F, Shugart YY, Yang L, Li X, Liu Z, et al. The early growth response protein 1-miR-30a-5p-neurogenic differentiation factor 1 axis as a novel biomarker for schizophrenia diagnosis and treatment monitoring. *Transl Psychiatry* (2017) 7(1):e998. doi: 10.1038/tp.2016.268
53. Ebbesen KK, Kjems J, Hansen TB. Circular RNAs: Identification, biogenesis and function. *Biochim Biophys Acta* (2016) 1859(1):163–8. doi: 10.1016/j.bbagr.2015.07.007
54. Meng X, Li X, Zhang P, Wang J, Zhou Y, Chen M. Circular RNA: an emerging key player in RNA world. *Brief Bioinform* (2017) 18(4):547–57.
55. Venø MT, Hansen TB, Venø ST, Clausen BH, Grebing M, Finsen B, et al. Spatio-temporal regulation of circular RNA expression during porcine embryonic brain development. *Genome Biol* (2015) 16:245. doi: 10.1186/s13059-015-0801-3
56. Rybak-Wolf A, Stottmeister C, Glažar P, Jens M, Pino N, Giusti S, et al. Circular RNAs in the Mammalian Brain Are Highly Abundant, Conserved, and Dynamically Expressed. *Mol Cell* (2015) 58(5):870–85. doi: 10.1016/j.molcel.2015.03.027
57. Hanan M, Soreq H, Kadener S. CircRNAs in the brain. *RNA Biol* (2017) 14(8):1028–34. doi: 10.1080/15476286.2016.1255398
58. Kumra S, Charles Schulz S. Editorial: research progress in early-onset schizophrenia. *Schizophr Bull* (2008) 34(1):15–7. doi: 10.1093/schbul/sbm123
59. Ng SY, Bogu GK, Soh BS, Stanton LW. The long noncoding RNA RMST interacts with SOX2 to regulate neurogenesis. *Mol Cell* (2013) 51(3):349–59. doi: 10.1016/j.molcel.2013.07.017
60. You X, Vlatkovic I, Babic A, Will T, Epstein I, Tushev G, et al. Neural circular RNAs are derived from synaptic genes and regulated by development and plasticity. *Nat Neurosci* (2015) 18(4):603–10. doi: 10.1038/nn.3975
61. Ahmad R, Sportelli V, Ziller M, Spengler D, Hoffmann A. Tracing Early Neurodevelopment in Schizophrenia with Induced Pluripotent Stem Cells. *Cells* (2018), 7(9). doi: 10.3390/cells7090140

62. Mahmoudi E, Fitzsimmons C, Geaghan MP, Shannon Weickert C, Atkins JR, Wang X, et al. Circular RNA biogenesis is decreased in postmortem cortical gray matter in schizophrenia and may alter the bioavailability of associated miRNA. *Neuropsychopharmacology* (2019) 44(6):1043–54. doi: 10.1038/s41386-019-0348-1
63. Glažar P, Papavasileiou P, Rajewsky N. circBase: a database for circular RNAs. *RNA* (2014) 20(11):1666–70. doi: 10.1261/rna.043687.113
64. Liu Z, Ran Y, Tao C, Li S, Chen J, Yang E. Detection of circular RNA expression and related quantitative trait loci in the human dorsolateral prefrontal cortex. *Genome Biol* (2019) 20(1):99. doi: 10.1186/s13059-019-1701-8
65. Zimmerman AJ, Hafez AK, Amoah SK, Rodriguez BA, Dell'Orco M, Lozano E. A psychiatric disease-related circular RNA controls synaptic gene expression and cognition. *Mol Psychiatry* (2020). doi: 10.1038/s41380-020-0653-4
66. Yang Q, Wu J, Zhao J, Xu T, Zhao Z, Song X, et al. Circular RNA expression profiles during the differentiation of mouse neural stem cells. *BMC Syst Biol* (2018) 12(Suppl 8):128. doi: 10.1186/s12918-018-0651-1
67. Zaghlool A, Ameer A, Wu C, Westholm JO, Niazi A, Manivannan M, et al. Expression profiling and in situ screening of circular RNAs in human tissues. *Sci Rep* (2018) 8(1):16953. doi: 10.1038/s41598-018-35001-6
68. Yao G, Niu W, Zhu X, et al. hsa_circRNA_104597: a novel potential diagnostic and therapeutic biomarker for schizophrenia. *Biomark Med* (2019) 13(5):331–40. doi: 10.2217/bmm-2018-0447
69. Werfel S, Nothjunge S, Schwarzmayr T, Strom TM, Meitinger T, Engelhardt S. Characterization of circular RNAs in human, mouse and rat hearts. *J Mol Cell Cardiol* (2016) 98:103–7. doi: 10.1016/j.yjmcc.2016.07.007
70. Zhang S, Zhu D, Li H, Li H, Feng C, Zhang W. Characterization of circRNA-Associated-ceRNA Networks in a Senescence-Accelerated Mouse Prone 8 Brain. *Mol Ther* (2017) 25(9):2053–61. doi: 10.1016/j.ymthe.2017.06.009
71. Floris G, Zhang L, Follesa P, Sun T. Regulatory Role of Circular RNAs and Neurological Disorders. *Mol Neurobiol* (2017) 54(7):5156–65. doi: 10.1007/s12035-016-0055-4
72. Smith K. Mental health: a world of depression. *Nature* (2014) 515(7526):181. doi: 10.1038/515180a
73. Harris EC, Barraclough B. Excess mortality of mental disorder. *Br J Psychiatry* (1998) 173:11–53. doi: 10.1192/bjp.173.1.11
74. Courtet P, Lopez-Castroman J. Antidepressants and suicide risk in depression. *World Psychiatry* (2017) 16(3):317–8. doi: 10.1002/wps.20460
75. Ponsoni A, Branco LD, Cotrena C, Shansis FM, Grassi-Oliveira R, Fonseca RP. Self-reported inhibition predicts history of suicide attempts in bipolar disorder and major depression. *Compr Psychiatry* (2018) 82:89–94. doi: 10.1016/j.comppsych.2018.01.011
76. Holden C. Mental health. Global survey examines impact of depression. *Science* (2000) 288(5463):39–40. doi: 10.1126/science.288.5463.39
77. Cui X, Niu W, Kong L, He M, Jiang K, Chen S, et al. hsa_circRNA_103636: potential novel diagnostic and therapeutic biomarker in Major depressive disorder. *Biomark Med* (2016) 10(9):943–52. doi: 10.2217/bmm-2016-0130
78. Danna SM, Graham E, Burns RJ, Deschênes SS, Schmitz N. Association between Depressive Symptoms and Cognitive Function in Persons with Diabetes Mellitus: A Systematic Review. *PLoS One* (2016) 11(8):e0160809. doi: 10.1371/journal.pone.0160809
79. Katon W. Depression and diabetes: unhealthy bedfellows. *Depress Anxiety* (2010) 27(4):323–6. doi: 10.1002/da.20683
80. An T, Zhang J, Ma Y, Lian J, Wu YX, Lv BH, et al. Relationships of Non-coding RNA with diabetes and depression. *Sci Rep* (2019) 9(1):10707. doi: 10.1038/s41598-019-47077-9
81. Abasolo N, Torrell H, Roig B, Moyano S, Vilella E, Martorell L. RT-qPCR study on post-mortem brain samples from patients with major psychiatric disorders: reference genes and specimen characteristics. *J Psychiatr Res* (2011) 45(11):1411–8. doi: 10.1016/j.jpsychires.2011.06.001
82. Jiang G, Ma Y, An T, Pan Y, Mo F, Zhao D, et al. Relationships of circular RNA with diabetes and depression. *Sci Rep* (2017) 7(1):7285. doi: 10.1038/s41598-017-07931-0
83. Hunter AM, Leuchter AF, Power RA, Muthén B, McGrath PJ, Lewis CM, et al. A genome-wide association study of a sustained pattern of antidepressant response. *J Psychiatr Res* (2013) 47(9):1157–65. doi: 10.1016/j.jpsychires.2013.05.002
84. Zhang Y, Du L, Bai Y, Han B, He C, Gong L. CircDYM ameliorates depressive-like behavior by targeting miR-9 to regulate microglial activation via HSP90 ubiquitination. *Mol Psychiatry* (2018). doi: 10.1038/s41380-018-0285-0
85. Cohen JE, Lee PR, Fields RD. Systematic identification of 3'-UTR regulatory elements in activity-dependent mRNA stability in hippocampal neurons. *Philos Trans R Soc Lond B Biol Sci* (2014) 369(1652). doi: 10.1098/rstb.2013.0509
86. Long B, Wang K, Li N, Murtaza I, Xiao JY, Fan YY, et al. miR-761 regulates the mitochondrial network by targeting mitochondrial fission factor. *Free Radic Biol Med* (2013) 65:371–9. doi: 10.1016/j.freeradbiomed.2013.07.009
87. Xu Y, Zhao C, Sun X, Liu Z, Zhang J. MicroRNA-761 regulates mitochondrial biogenesis in mouse skeletal muscle in response to exercise. *Biochem Biophys Res Commun* (2015) 467(1):103–8. doi: 10.1016/j.bbrc.2015.09.113
88. Henley WN, Koehnle TJ. Thyroid hormones and the treatment of depression: an examination of basic hormonal actions in the mature mammalian brain. *Synapse* (1997) 27(1):36–44. doi: 10.1002/(SICI)1098-2396(199709)27:1<36::AID-SYN4>3.0.CO;2-E

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Biomarkers in Psychiatry: Concept, Definition, Types and Relevance to the Clinical Reality

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OPEN ACCESS

Edited by:

Helge Frieeling,
Hannover Medical School, Germany

Reviewed by:

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University of Ulm, Germany
Bhaskar Roy,
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Specialty section:

This article was submitted to
Molecular Psychiatry,
a section of the journal
Frontiers in Psychiatry

Received: 15 January 2020

Accepted: 28 April 2020

Published: 15 May 2020

Citation:

García-Gutiérrez MS, Navarrete F,
Sala F, Gasparyan A,
Austrich-Olivares A and Manzanares J
(2020) Biomarkers in Psychiatry:
Concept, Definition, Types and
Relevance to the Clinical Reality.
Front. Psychiatry 11:432.
doi: 10.3389/fpsy.2020.00432

During the last years, an extraordinary effort has been made to identify biomarkers as potential tools for improving prevention, diagnosis, drug response and drug development in psychiatric disorders. Contrary to other diseases, mental illnesses are classified by diagnostic categories with a broad variety list of symptoms. Consequently, patients diagnosed from the same psychiatric illness present a great heterogeneity in their clinical presentation. This fact together with the incomplete knowledge of the neurochemical alterations underlying mental disorders, contribute to the limited efficacy of current pharmacological options. In this respect, the identification of biomarkers in psychiatry is becoming essential to facilitate diagnosis through the developing of markers that allow to stratify groups within the syndrome, which in turn may lead to more focused treatment options. In order to shed light on this issue, this review summarizes the concept and types of biomarkers including an operational definition for therapeutic development. Besides, the advances in this field were summarized and sorted into five categories, which include genetics, transcriptomics, proteomics, metabolomics, and epigenetics. While promising results were achieved, there is a lack of biomarker investigations especially related to treatment response to psychiatric conditions. This review includes a final conclusion remarking the future challenges required to reach the goal of developing valid, reliable and broadly-usable biomarkers for psychiatric disorders and their treatment. The identification of factors predicting treatment response will reduce trial-and-error switches of medications facilitating the discovery of new effective treatments, being a crucial step towards the establishment of greater personalized medicine.

Keywords: biomarkers, neuropsychiatry, personalized medicine, lymphocytes, peripheral biomarkers, central biomarkers

INTRODUCTION

According to World Health Organization mental illness presented devastating rates of prevalence, mortality, morbidity and disability. Suffering a serious mental illness reduces average life expectancy in 13 to 32 years (1, 2). Aside from mortality, in most Western countries, mental disorders are the leading cause of disability, responsible for 30-40% of chronic sick leave and costing approximately a

4% of gross domestic product (3). Besides, for all types of mental illness, pharmacological treatment options are scarce and present limited efficacy. Several studies highlighted that, in terms of recovery and remission, current pharmacological interventions showed significant limitations. A series of effectiveness trials sponsored by the National Institute of Mental Health (NIMH) in USA provided relevant data in this regard. In CATIE (Clinical Antipsychotic Trials of Intervention Effectiveness) study, 74% of patients suffering from chronic schizophrenia (SCZ) experienced problems of treatment adherence within 18 months (4). In addition, in the Sequenced Treatment Alternatives to Relieve Depression (STAR*D) study only 31% of patients with major depressive disorder (MDD) were in remission after being treated with a selective serotonin reuptake inhibitor for a total of 14 weeks (5). An additional study carried out in patients diagnosed with bipolar disorder (BD) (STEP-BD, Systematic Treatment Enhancement Program for Bipolar Disorder) revealed that only 24% of patients experienced a remission of depression during 8 consecutive weeks, outcome similar to those observed in the vehicle group (6).

Several factors contribute to this clinical reality. On one hand, the heterogeneity/complexity of mental disorders. Patients suffering from a mental illness displayed several symptoms related with behavior, thinking, feelings and/or social interaction. To facilitate the diagnoses, mental disorders are classified by diagnostic categories with a broad variety list of symptoms according to the Diagnostic and Statistical Manual of Mental Disorders, Fifth Edition (DSM-IV), or International Statistical Classification of Diseases and Related Health Problems, Tenth Edition (ICD-11). Consequently, patients diagnosed with the same psychiatric illness present a great heterogeneity in their clinical presentation. In addition, several mental illnesses present symptoms in common, that can often make the diagnosis difficult.

On the other hand, psychiatric diseases present high comorbidity. Approximately 85%–90% of patients with depression also experience symptoms of anxiety, and vice versa (7, 8). Among schizophrenic patients, psychiatric comorbidities are common. Around 50% of patients suffer from depression and more than 47% have a lifetime diagnosis of comorbid substance use disorders (9–11). The simultaneous presence of two or more psychiatric diseases are associated with greater severity, worse response to the pharmacological treatment and have a greater risk of suicide than either condition alone.

Despite these sobering facts, progress in human brain research and the advent of new technologies, such as ‘omics’ technologies, offers the opportunity for change mental health treatment and outcomes in a near future. In this respect, the identification of biomarkers has become a new promising tool for guiding diagnosis, predicting clinical outcome and, therefore, improving the understanding of the pathophysiology of mental disorders. This review provides an overview about the current state of biomarkers in neuropsychiatry, with the ultimate aim of remarking some goals achieved up to date and the future challenges needed to develop valid, reliable and broadly-usable

biomarkers for psychiatric disorders and their treatment. For this purpose, the review includes a definition of biomarker’s concept throughout history, describes the different types of biomarkers and their potential role in clinical practice, and emphasizes the samples and techniques commonly used. The role of ‘omics’ is described in greater detail due to its huge progress in the recent years. A final conclusion remarks the difficulties and limitations of current biomarkers strategies in neuropsychiatry and the future challenges needed to progress in this field.

WHAT ARE BIOMARKERS? EVOLUTION OF BIOMARKERS THROUGH HISTORY

During the last 50 years, the definition of biomarker has been modified according to scientific and clinical progress. The term “biomarker” was used for the first time in 1973 to indicate the presence or absence of biological material. However, the concept is older, referenced as a “biochemical marker” in 1949 (12) and “biological marker” in 1957 (13).

In 2000, the Biomarker Definition Working Group, supported by the U.S. National Institute of Health (NIH), defined a biomarker as “a characteristic that is objectively measured and evaluated as an indication of normal biological processes, pathogenic processes, or pharmacologic responses to a therapeutic intervention” (14). This definition has two major limitations. The first one lies in the fact that sometimes a biomarker is measured by subjective parameters. The second one is the fact that additional processes or responses beyond those covered by the definition are excluded.

In 2016, Fitzgerald and colleagues redefined the concept of biomarker as “a functional variant or quantitative index of a biological process that predicts or reflects the evolution of or predisposition to a disease or a response to a therapy” (15). Nevertheless, this description lacks the consideration of structural variants and qualitative index as potential biomarkers.

In order to harmonize the term of biomarker, the Food and Drug Administration (FDA) in collaboration with the NIH Joint Leadership Council convened the FDA-NIH Biomarker Working Group in 2016. This group simplified the biomarker definition being considered as “a defined characteristic that is measured as an indicator of normal biological processes, pathogenic processes or responses to an exposure or intervention” (16). This definition, clearer and more concise, defines a biomarker specifying its principal applications without any unnecessary complexity or contradictory information. Besides, to ensure its clinical use, a good biomarker should be measured with high reproducibility, present a sizeable signal-to-noise ratio and, more importantly, meet the condition of being modified in a dynamic and reliable way as the clinical condition progress. In addition, a biomarker should be accessible for its detection and measurement, as would be the case of a plasmatic parameter or a genetic marker, or being detected by histological or image/neuroimaging techniques (17).

Types and Role of Biomarkers in the Clinical Practice

According to their applications, biomarkers can provide complementary information about the disease or the intervention under consideration. Biomarkers may be identified at any event occurring since the pathogenesis, the onset of first clinical manifestations, diagnosis, treatment outcome or recovery. The FDA-NIH Biomarker Working Group distinguished several types of biomarkers based on their main clinical application: diagnostic, monitoring, pharmacodynamic/response, predictive, prognostic, safety, and susceptibility/risk biomarkers (**Figure 1**). A biomarker may meet multiple criteria for different uses or present specific features that enable its particular use (18).

Diagnostic Biomarker

Encompasses a variety of biomarkers used to detect or confirm the presence of a disease or medical condition. This type of biomarker can be used to identify disease subtypes. The advent of the era of the precision medicine emphasizes the fact that diagnostic biomarkers are useful not only to identify patients with a disease, but also to redefine its classification. This is an

important feature, because many diseases have subtypes with different prognosis or treatment responses. Thus, diagnostic biomarkers would contribute to improve personalized medicine increasing the effectiveness of the therapeutic response. These biomarkers may play also a critical role as *prognostic biomarkers* or *predictive treatment outcome biomarkers* (16, 17). An example could be the concentrations of A β 42 and total tau (T-tau) in cerebrospinal fluid of patients with dementia as diagnostic biomarker for Alzheimer's disease (19) (**Figure 2**).

Monitoring Biomarker

This category includes biomarkers that are analyzed at different time points to monitor the status of a disease or medical condition, and as a marker of the response to an intervention, including exposure to a medical product or an environmental agent (**Figure 2**) (16). Changes in biomarkers values are considered as indicators of the progression of the clinical condition and as measurements of the pharmacological response and other types of clinical interventions (17). An example of a monitoring biomarker is the elevation of serum creatinine and/or potassium concentrations after a pharmacological or medical intervention,

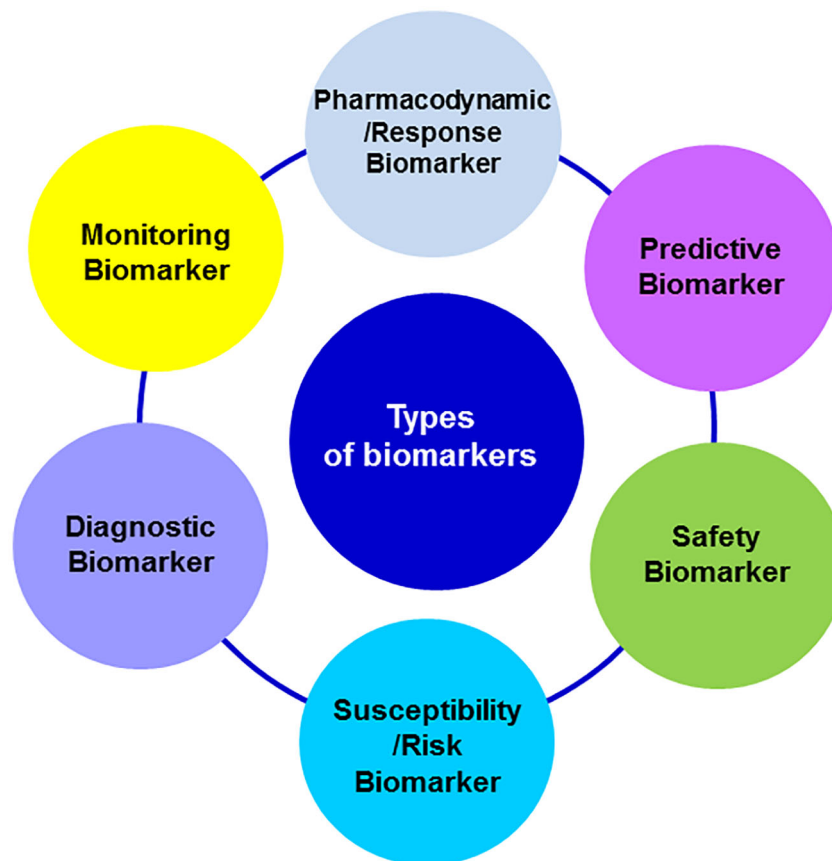


FIGURE 1 | Classification of biomarkers based on its main clinical application.

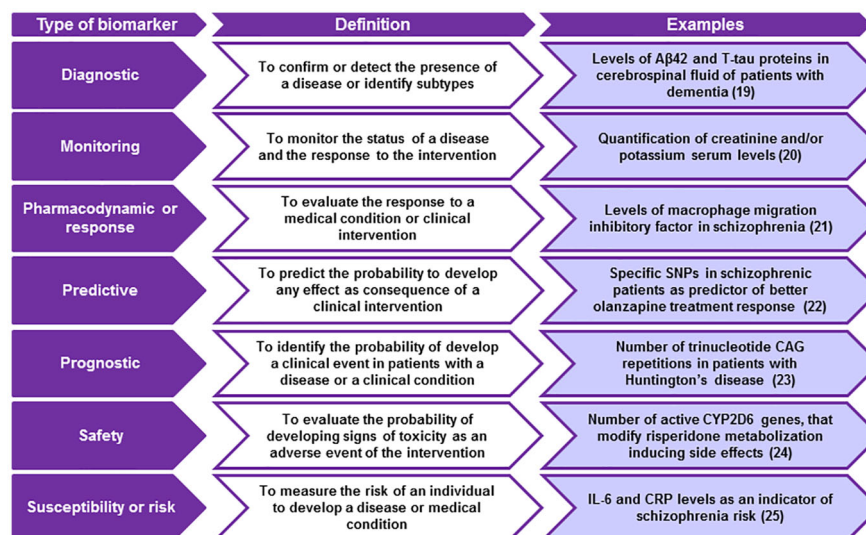


FIGURE 2 | Aims and examples of the main types of biomarkers. SNPs, single nucleotide polymorphisms; CRP, c-reactive protein; IL-6, interleukin 6.

TABLE 1 | Potential role of monitoring biomarkers in neuropsychiatry.

| Type of intervention | Utility | References |
|---------------------------------|---|------------|
| Clinical care or clinical trial | To evaluate patient's clinical situation during treatment or at the end of the intervention | (21) |
| Before treatment initiation | To detect signs and/or symptoms of a disease or medical condition as an indicative parameter of the prognosis | (22) |
| | To determine the need for prompt treatment | |
| Medical product development | To provide information about the safety and effectiveness of a drug | (23) |
| Public health | To provide information about the risk of developing any disease or medical condition among the population | (24) |
| Pharmacodynamics studies | To provide evidences about therapeutic response | (25) |

parameters that are commonly used as an indicator of the probability to develop side effects (20). Monitoring biomarkers can be applied in different situations including clinical care or clinical trials, at the beginning of a treatment, for medical product development purposes, as a measure of the risk of developing a disease, or to evaluate the pharmacodynamics of a clinical intervention (Table 1).

Pharmacodynamic or Response Biomarker

Proposed to be a potential useful tool in clinical practice providing useful information for patient management. A pharmacodynamic biomarker is modified in response to a medical condition or clinical intervention, including drug treatments (16). Because of the serial nature of their assessment, this type of biomarker is frequently considered as a monitoring biomarker (20). The main utility of this biomarker is to guide the clinical management, providing crucial information for deciding whether or not to continue the treatment. Thus, pharmacodynamic biomarkers determine the progression of the treatment (26) (Figure 2).

Another area in which these biomarkers are of special interest is in early therapeutic drug development, being useful to establish

the proof that a drug induces pharmacodynamic changes in humans related to its clinical benefit and to guide dose-response studies (18).

Predictive Biomarker

A marker is considered as a predictive biomarker when its presence or modification allows predicting which patient or group of patients are more likely to experience an effect as consequence of being exposed to a medical product or environmental agent (16). This effect could be a symptomatic benefit, an increase in survival rates or an adverse event. These biomarkers are frequently used in randomized controlled clinical trials of new therapies. In this context, the biomarker is used to select patients for participation or to stratify them into intervention groups. If the biomarker predicts a favorable outcome, its presence may indicate a greater effect of the new therapy compared to the control therapy (20). Thus, the use of predictive biomarkers facilitates the selection of specific patients more likely to respond or not to therapy (Figure 2). An example of a predictive biomarker is the presence of 12 single nucleotide polymorphisms (SNPs) in Had Chinese schizophrenic

population, that were correlated with greater olanzapine effectiveness (27).

Prognostic Biomarker

Commonly used to identify the probability of developing a clinical event in patients diagnosed with a disease or medical condition (16). These events include death, disease progression or recurrence, or the development of a new medical condition. In clinical trials, prognostic biomarkers are used to identify patients more likely to develop a clinical event or disease progression, allowing to identify populations at higher risk. In this context, prognostic biomarkers are used as inclusion or exclusion criteria (17). An example of a prognostic biomarker is the number of trinucleotide CAG repetitions in patients with Huntington's disease. A high number of CAG nucleotides repetitions are correlated with a greater threshold of disease's severity (**Figure 2**) (28).

An additional utility of prognostic biomarkers is in treatment selection. They can provide information about treatment safety, guiding patient hospitalization or their entrance in intensive care units.

Several factors influence the clinical outcome, including the clinical condition severity, the effects induced by all treatments and the intrinsic characteristics of patients. Some of these characteristics may be used as a prognostic biomarker, allowing to identify patients more likely to experience a clinical event, disease recurrence or progression, and any effect (favorable or unfavorable) induced by a medical product or environmental agent (16, 20).

Safety Biomarker

Is any measure that can be assessed before and after the exposure to a medical intervention, or an environmental agent, allowing to identify the probability of developing signs of toxicity as an adverse event, to detect the presence of toxicity, and for monitoring its extension (**Figure 2**) (16).

For many therapies, monitoring hepatic, renal and cardiovascular functions are critical to detect toxicity ensuring the safety of the therapy under study. All safety biomarkers have in common its ability to detect or predict toxicity prior to the onset of clinical signs and before irreversible damage. The toxicity can be determined by the detection or changes in the biomarker level.

Another usefulness of safety biomarkers is the identification of patients in which particular therapies should not be initiated because of significant safety risks. For example, genetic variations in CYP2D6 enzymes modify the response to certain drugs commonly used in psychiatry such as almost 50% of antipsychotics drugs. Alterations in the metabolism of drugs can modify its effectiveness, decreasing the response to the treatment or enhancing toxicity risk in patients (15). In case of the antipsychotic risperidone, there is a correlation between the number of active CYP2D6 genes and its cardiac toxicity. QTc interval is longer in subjects with one active CYP2D6 gene compared to patients with two. The study revealed that the number of CYP2D6 active genes was related with the corrected plasma concentration of risperidone (29). Safety biomarkers are

used with this purpose in public health or in epidemiological interventions aimed to control or mitigate risk exposure.

Susceptibility or Risk Biomarker

Is used as a risk measure to develop a disease or medical condition (8). An example is a genetic biomarker that can be detected many years or decades before the onset of clinical signs and/or symptoms of the disease (**Figure 2**) (10). Susceptibility/risk biomarkers are essential for the development of epidemiological studies aimed to evaluate the risk of developing a disease, contributing to establish preventive strategies in clinical practice. In this line, some studies suggested a potential correlation between interleukin-6 (IL-6) and C-reactive protein (CRP) levels and the risk of developing SCZ. Lower CRP levels together with the blockade of IL-6 signaling significantly increase SCZ risk, being proposed as a potential susceptibility/risk biomarkers for this neuropsychiatric disorder (30).

SAMPLES AND TECHNIQUES USED FOR THE SEARCHING OF BIOMARKERS

Biomarkers should be easily measurable, in easily accessible samples and using affordable techniques to ensure its inclusion in the routine clinical practice. Historically, plasma together with tissues obtained from biopsies were one of the most common samples used in the searching for biomarkers. Besides, based on the disease of interest, additional body fluids readily available in large amounts as urine, saliva, tear fluid, sweat, amniotic, cerebrospinal and pleural fluids, cervicovaginal secretion and wound efflux can be used for this purpose (31).

In the case of diseases of the central nervous system (CNS), such as psychiatric and neurological disorders, access to brain samples is of particular interest. In this respect, brain human post-mortem samples, usually provided by brain banks, play a crucial role. However, systematic biochemical investigations using these samples are scarce, limited and unrealistic mainly to the fact that the course of the disease cannot be monitored. In this respect, the progress of functional neuroimaging has allowed to study some neuronal functions including alterations of local cerebral flow, energy metabolism and neurotransmitter receptor density and occupation over the course of disease. Nevertheless, functional neuroimaging fails to provide information at cellular biochemistry level and the access to this technique is limited due to its high economic costs.

In this context, blood lymphocytes have gained special attention in the searching of peripheral biomarkers (32). Lymphocytes can be isolated easily from blood samples and studied on a daily basis allowing to monitor the course of the disease. This is possible due to the fact that receptor properties and transduction processes of lymphocytes are similar to those observed in the CNS. Several studies pointed out a close bidirectional interaction between the CNS and the immune system, in particular with lymphocytes (33). For instance, peripheral cytokines released by lymphocytes modify CNS functions including its autonomic control as well as

neuroendocrine and behavioral responses. Besides, several evidences suggested that alterations in neurotransmitters and hypothalamic-pituitary-adrenal (HPA) axis in the CNS are concomitant with alterations in the function and metabolism of lymphocytes.

To date, some genes such as *c-fos*, interleukins (IL-2, IL-4, IL-6, IL-10), nerve growth factor (NGF), brain-derived neurotrophic factor (BDNF), cannabinoid receptors, acetylcholine, GABA_A receptors, B₂-adrenergic receptors, glucocorticoid receptors, mineralocorticoid receptors, D₃ dopaminergic receptor, and serotonin receptors have been analyzed in lymphocytes from psychiatric patients, such as schizophrenic and depressive patients, with promising results as peripheral biomarkers (34–41). Thus, gene expression studies in lymphocytes of psychiatric patients at different stages of the disease, that may reflect alterations in the CNS, would allow to further characterize the mechanisms underlying the pathogenesis of the disease and may contribute to predict the pharmacological treatment response (biomarkers of treatment outcome) (Figure 3).

Another crucial factor for the searching of biomarkers is the techniques used. These techniques should have a high-throughput for the application of analytical data through robust dimensional data obtained in high performance tests (42). In this respect, 'omics' technologies, including genomics, proteomics, transcriptomics, metabolomics, and epigenetics, have contributed to the rapid discovery of many potential biomarkers.

'OMICS' BIOMARKERS AND NEUROPSYCHIATRY

This section summarizes the main advantages of each 'omics' technology in the search of biomarkers for assessing risk,

diagnosis, monitoring progression and prediction of treatment response in neuropsychiatry disorders.

Genomic Biomarkers

Genomic biomarkers are expanding knowledge for the understanding of disease pathogenesis providing new targets for disease characterization, early diagnosis, and better-targeted treatment (drug discovery, drug development and adverse drug responses) to direct patients towards a more likely benefit based on their unique profile (43). According to the European Medicines Agency (EMA), a genomic biomarker is defined as "a measurable DNA and/or RNA characteristic that is an indicator of normal biologic processes, pathogenic processes, and/or response to therapeutic or other interventions" (44). These measurable features include the expression, function and regulation of a particular gene. In the DNA, these features can be characterized by single nucleotide polymorphisms (SNPs), variability of short sequence repeats, haplotypes, deletions or insertions of (a) single nucleotide (s), copy number variations and cytogenetic rearrangements (translocations, duplications, deletions or inversions) (45). The use of genetic techniques allowed the analysis of candidate genes, genome-wide studies and polygenic risk score analysis to understand multiple psychiatric disorders such as SCZ (46, 47). These techniques include Comparative Genomic Hybridization (CGH), microarray, exome sequencing, and whole genome sequence. Specifically, pharmacogenomics is crucial to identify genetic polymorphisms in drug-metabolizing enzymes, transporters, receptors, and other drug targets, being essential to drug discovery and drug therapy optimization [for review (48, 49)] (Figure 4).

Genome-wide association studies have allowed the identification of potential genomic biomarkers in different

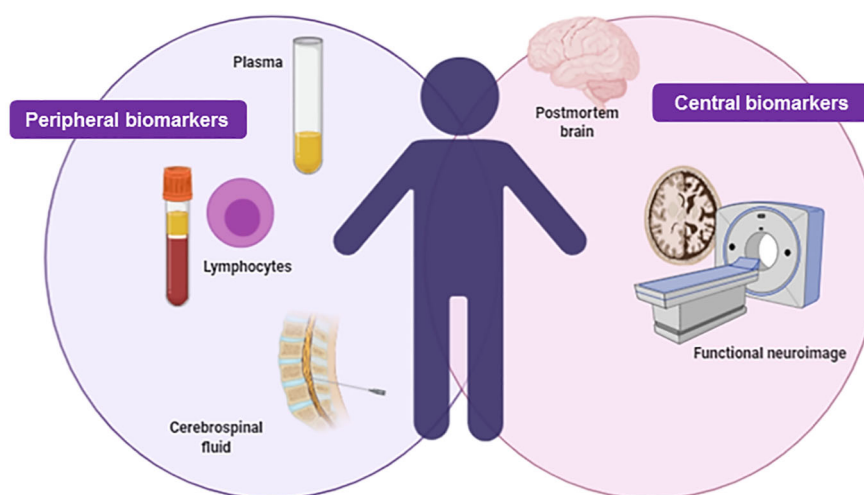


FIGURE 3 | Integrative figure regarding the main samples used in the searching of biomarkers in neuropsychiatry: peripheral and central biomarkers. Created with BioRender.com.

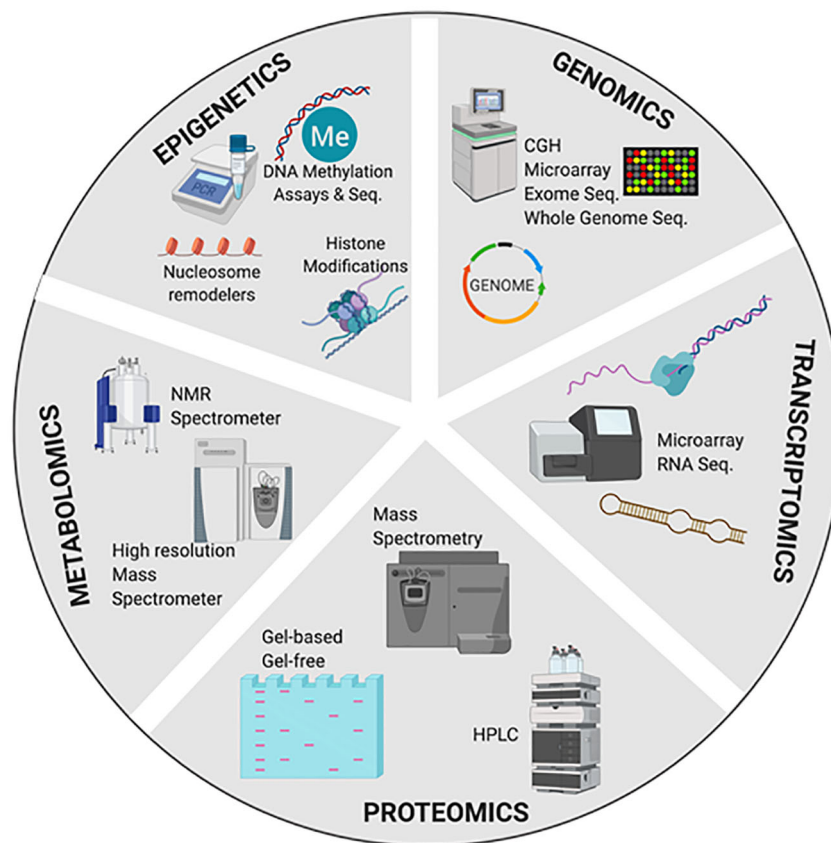


FIGURE 4 | Multi-omics approach for the discovery and validation of biomarkers to probe multidimensional phases of the disease. CGH, comparative genomic hybridization; Seq, sequencing; qRT-PCR, quantitative real time PCR; qPCR, semiquantitative real time PCR; HPLC, high performance liquid chromatography; NMR, nuclear magnetic resonance spectroscopy. Created with BioRender.com.

neuropsychiatric (50–52) and drug-use disorders (53). For example, the Collaborative Study of the Genetics of Alcoholism (COGA) have correlated the SNPs rs4780836 [A > C; chromosome 16:19974071 (GRCh38.p12)], rs2605140 [A > G; chromosome 17: 18253061 (GRCh38.p12)], rs11690265 [chromosome 2: C > T; chr2:27418655 (GRCh38.p12)], rs692854 [non-functional Se (FUT2) gene; alleles C > A; chromosome 19: 48706207 (GRCh38.p12)], and rs13380649 (alleles A > G; chromosome 16: 19999778 (GRCh38.p12)] with greater vulnerability and predisposition to develop alcohol use disorders (AUD) in European American and African American. Besides, the study has demonstrated that there is a correlation between these SNPs and alterations in electroencephalograms, such as lower posterior gamma, higher slow wave connectivity (delta, theta, alpha), higher frontal gamma ratio and higher beta correlation in the parietal area of patients with AUD (54).

In bipolar disorder (BD), the SNP rs17026688 in the gene encoding glutamate decarboxylase-like protein 1 (GADL1) has been associated with the response to lithium in Chinese patients (55). This SNP has been related to immune dysfunctions in BD patients, such as higher percentages of total T cells, CD4⁺T cells, activated B cells and monocytes. Besides, treatment of BD

patients-derived peripheral blood mononuclear cells (PBMCs) with lithium *in vitro* increases the immune response (CD4⁺ cells). These findings suggest that the immune imbalance might not only be a biomarker for diagnosis but also a biomarker of the disease progression and therapeutic response in BD.

In addition, a large study carried out through Europe, North America and Australia identified 30 genome-wide significant loci for BD in patients of European descent. These loci contain genes that encode for neurotransmitters transporters, synaptic components, and ion channels, including calcium voltage-gated channel subunit alpha1 C (CACNA1C) and other voltage-gated calcium channel genes. Among the 30 loci identified in BD patients, eight have also been described in SCZ patients (56–58); however, conditional analyses performed in this study suggested that BD and SCZ associations are independent for three of the eight shared loci, providing information that may be useful for understanding the genetics mechanisms underlying these psychiatric disorders that in some cases present symptoms in common that make its diagnosis difficult. Furthermore, the BD subtype polygenic risk score analyses performed in the study supported the nosological distinction between bipolar I (BD1) and bipolar II disorder (BD2) and the importance of psychosis

beyond DSM subtypes. One limitation of the study is the genetic heterogeneity of the samples that may contribute to inconsistent replication in some of the results (59).

Besides, DNA genomic biomarkers as a useful indicator of the state of the disease (severity) also presented relevant consequences for the clinical management of neuropsychiatric diseases. In this respect, a recent study revealed a close relationship between the SNPs rs1360780 in the FKBP5 gene and rs17689918 in the corticotrophin-releasing hormone receptor 1 (CRHR1) gene and greater severity of the disease in posttraumatic stress disorder (PTSD) patients (60).

Transcriptomic Biomarkers in Neuropsychiatry

The transcriptome is defined as the complete set of all RNA molecules in one cell or a population of cells at a specific developmental stage or physiological condition (61). Thus, transcriptome is dynamic and reflects the cellular state. Measuring the expression of an organism's genes as a snapshot in different tissues, conditions, or time points provides information on how genes are regulated and would contribute to a better understanding of human diseases and their pharmacological treatment, allowing to identify potential therapeutic biomarkers when variations in treatment outcomes occur (62, 63). Although first studies for transcriptome began in the early 1990s, technological advances have spread throughout this time. There are two key contemporary techniques in the field: microarrays, which quantify a set of predetermined sequences, and RNA sequencing (RNA-Seq), which uses high-throughput sequencing to capture all RNA sequences (63).

For instance, transcriptomics studies identified that the efficacy of antidepressants is related to gene expression changes at transcriptome-wide scale. In a microarray study, alterations of *MMO28* and *KXD1* genes encoding for matrix metalloproteinase 28 and KxDL motif-containing protein 1, respectively, were associated with better response to nortriptyline in depressive patients (64). This data could contribute to improve the characterization of the molecular pathways underlying the efficacy of antidepressants.

In addition, a clinical study associated new miRNAs (miR-146a-5p, miR-146b-5p, miR-24-3p, and miR-425-3p) with the effectiveness of antidepressant drugs such as duloxetine, escitalopram, and nortriptyline, in patients with MDD. These miRNAs are ubiquitously expressed and highly correlated in blood and in brain tissue, playing an important role in the regulation of the mitogen-activated protein kinase (*MAPK*) and *Wnt* signaling pathways, closely related with stress response and MDD (65). Interestingly, an additional study revealed that MDD patients responders to antidepressant treatment present a significant reduction of miR-1202 baseline levels compared to non-responders. Moreover, miR-1202 increases as the efficacy of the antidepressant treatment is observed (66).

Besides, a total of 25 miRNAs have been modified in the amygdala of rats exposed to the learned helpless animal model of

depression being the miR-128-3p the most affected. Also, a reduction of *Wnt* signaling genes has been also detected. Accordingly, an increase of miR-128-3p expression along with a significant downregulation of key target genes from *Wnt* pathway signaling (*WNT5B*, *DVL*, and *LEF1*) has been identified in the AMY of MDD patients (67).

Especially, RNA studies provide promising results in the searching for biomarkers of suicide. Alterations of RNA editing on the cyclic nucleotide phosphodiesterase (PDE, particularly PDE8A involved in the hydroxylation of cAMP and cGMP) were found in the dorsolateral prefrontal cortex (DLPFC) and the anterior cingulate cortex (ACC) of suicide completers. These alterations have been proposed as a biomarker of risk for attempting suicide in patients with depressive symptoms (68).

Other example of the potential role of these biomarkers in neuropsychiatry is a genome-wide expression study in patients who met the DSM-IV criteria for methamphetamine dependence. The results revealed that treatment with topiramate significantly modified the gene expression of specified genes *GRINA*, *PRKACA*, *PRKCI*, *SNAP23* and *TRAK2* involved in severe pathways underlying drug addiction and other relevant physiological functions, including neuronal function/synaptic plasticity, signal transduction, cardiovascular function, and inflammation/immune response (69).

Likewise, the microRNA-124 (miR-124) and microRNA-181 (miR-181) were pointed out as potential biomarkers for cocaine use disorder (CUD) (70). The study revealed that these two microRNAs were upregulated in the blood samples of females CUD compared with healthy female controls.

Proteomic Biomarkers in Neuropsychiatry

Proteomics approaches using blood, plasma or serum constitutes a highly desired method for biomarker profiling of psychiatric disorders, due to the fact that these biological samples are used for routine diagnostic analyses in clinical practice, making easier to obtain samples. Besides, in neuropsychiatry, the cerebrospinal fluid (CSF) is a sample of particular interest for the identification of potential proteomic biomarkers due to its proximity to the brain. Although its collecting is very complex, due to the invasive procedure involved, it contains much less proteins than plasma. Thus, the "buffering" of protein composition is much weaker and tend to lead in a reduction of chances to identify potential proteomic biomarkers.

In proteomics, the separation of proteins using gel-based or gel free techniques, commonly followed by mass spectrometry are the mainly techniques used. The strategies for obtaining biological samples are diverse, but it is recommended to reduce the complexity of the sample, and sometimes to employ enrichment techniques improving the levels of certain subcellular fractions of interest or for specific types of proteins (glycoproteins, membrane, secreted, nuclear matrix and phosphorylated proteins) (71).

Diagnostic complications and timely treatment in neuropsychiatric disorders are frequent. Such is the case of SCZ, diagnosed by certain signs and symptoms but not by measurable and identifiable biological characteristics. In this

respect, proteome studies carried out in blood plasma, serum and postmortem brain tissue from SCZ patients identified alterations in proteins that play a significant role in neuronal transmission and synaptic function, calcium homeostasis and signalling, energy metabolism, oxidative stress, cytoskeleton and in immune system and inflammation. These proteins have been proposed as biomarker candidates for prognosis, diagnosis, and medication monitoring in SCZ (72, 73). One of these proteins is zinc finger protein 729 that was found significantly down-regulated in patients with SCZ compared to healthy individuals and patients diagnosed with depression or BD (74). Another example is the study that showed reduced plasma levels of glia maturation factor beta (GMF- β), the brain-derived neurotrophic factor (BDNF), and the 115-kDa isoform of the Rab3 GTPase-activating protein catalytic subunit (RAB3GAP1) in SCZ patients. These biological markers have been proposed as potential biomarkers in this pathology (75).

Besides, the acetyl-L-carnitine (LAC) has been proposed as a proteomic biomarker in MDD. LAC plays an important role in several behavioral features. The reduction of LAC concentrations was associated with abnormal hippocampal glutamatergic function and plasticity. Such alterations suggested that the degree of LAC deficiency was directly proportional with the severity, the age of MDD onset, and the clinical history of treatment-resistant depression (TRD). These findings suggest that LAC may be useful as a diagnostic and prognosis biomarker for MDD (76).

Recently, neurofilaments light chains (NF-L) have been proposed as potential biomarkers for neuronal damage in certain psychiatric diseases. In the plasma of female patients affected by anorexia nervosa, levels of NF-L were significantly elevated, being associated with the neuronal damage observed in AN patients, that partially normalizes with weight recovery (77). An additional study pointed out the potential role of NF-L as a discriminative biomarker between primary psychiatric disorders and neurodegenerative clinical conditions with wide-ranging of behavioral, psychiatric, and cognitive symptoms (78). Interestingly, a reduction of NF-L has been identified in the hippocampus of rodents exposed to an animal model of depression (inescapable stress). In this study, treatment with valproic acid reduces depressive-like behavior and reverses NF-L reduction (79). Besides, elevated concentrations of NF-L have been observed in the CSF of BD patients. Authors demonstrated that there is a positive correlation between CSF NF-L levels and the response to antipsychotics and lithium (80). However, another prospective study failed to observe any association between the high baseline NF-L levels in CSF and clinical outcomes in BD (81). Further studies are needed to identify the role of neurofilaments as biomarkers for psychiatric disorders.

In conclusion, to consider a potential proteomic biomarker, it is necessary to evaluate its sensitivity, specificity and positive or negative predictive values upon the disease of interest (31). In this respect, modern proteomics workflows that enable high throughput studies with large cohorts of well-defined samples represent the opportunity to solve the limited reproducibility of past proteomic workflows (73).

Metabolomics Biomarkers in Neuropsychiatry

Metabolomics biomarkers for drug development are growing. This technology focuses on the presence of small molecules metabolites in various complex matrices like CSF, blood, urine, saliva, and other human fluids. The metabolome is inherently more dynamic and time sensitive than proteome and genome, providing a direct functional measure of cellular activity and physiological status (82, 83). Changes in metabolome are the consequence of the interaction between lifestyle, environmental, genetic, developmental, and pathological factors. Consequently, metabolomics are of particular interest because, in contrast to genomics, captures the dynamic nature of the disease, and in contrast to proteomics, metabolomics measure the final products produced by complex interactions between proteins, signalling cascades and cellular environments.

Metabolomics biomarkers are not characterized by one single metabolite. Rather, they are a set of correlated metabolites defining a specific state of disease or the response to a clinical or pharmacological intervention (84). Currently, gas chromatography-mass spectrometry (GC-MS), liquid chromatography-mass spectrometry (LC-MS), and nuclear magnetic resonance (NMR) are the main types of analytical platforms used in the searching for metabolomics biomarkers.

Several studies have focused on the identification of potential metabolomics biomarkers in different psychiatric diseases [for review see (85–88)]. For example, urinary metabolomics have been proposed to be potential useful tools for the identification of pathways that may be involved in the mechanism of action of specific treatments. Such is the case of the study in which 77 urinary metabolites were identified in children with autism spectrum disorder treated with sulforaphane, a supplement that significantly improved the social responsiveness. Some of these metabolites play a role in the regulation of neurotransmitters, hormones, oxidative stress, amino acid/gut microbiome, and sphingomyelin metabolism (89).

Recently, a study conducted in patients with symptoms of depression, tried to find a predictor or a biological correlation of depression recovery after the administration of certain antidepressants including escitalopram, bupropion-escitalopram or venlafaxine-mirtazapine combinations. An increase of phosphatidylcholine C38:1 baseline plasma concentrations was associated with poorer outcome in patients. In contrast, an increased ratio in hydroxylated sphingomyelins after 8 weeks of treatment was linked to symptoms recovery (90).

However, few metabolomics biomarkers, especially in neuropsychiatry, have passed the regulatory standards for their use in clinical practice, mainly due to the lack of robust assays for the routine quantification of potential biomarkers and the heterogeneity of studies. The reduced sample size, particularly of some clinical subgroups, and the limited quantitative power of current mass spectroscopy technology, hampered the identification of robust metabolomics biomarkers, making necessary their validation through additional assays. Besides, the complexity of samples, such as urine or blood, also contributes to that reality. In this respect, the use of chromatographic

techniques for separation is needed to reduce the potential interferences associated with the complexity of human samples (91).

Epigenetic Biomarkers

Dynamic variations in the structure of chromatin, which do not change the sequence of DNA itself but modified the expression of genes, have been paid attention due to its potential implication in the development of human diseases, including psychiatric disorders (92, 93). Accordingly, epigenetics may provide a functional interface between genotype, environmental exposure and phenotype (94).

To date, different forms of epigenetic regulation have been identified, such as direct methylation of DNA, histone modifications (as methylation and acetylation ubiquitination), exchanges of histone molecules with related isoforms, modification on chromatin by nucleosome remodelers that modify the access to DNA, and additional mechanisms like non-coding RNAs, non-genic DNAs, and differential exosome expression (95). In this way, identifying the aberrant changes in the epigenetic scenery associated to neuropsychiatry diseases and the factors that promote such alterations may allow the identification of potential new biomarkers (96).

An epigenetic biomarker is defined as “any epigenetic mark or altered epigenetic mechanism that can be measured in the body fluids or tissues defining a disease (detection); predicts the outcome of disease (prognostic), responds to therapy (predictive); monitors responses to therapy or medication (therapy monitoring) and predicts risk of future disease development (risk)” (97). So far, several techniques have been designed to analyze not only epigenetic processes at the level of specific genes but also epigenetic changes that occur in defined regions of the genome by epigenome-wide association studies. DNA methylation assays and DNA methylation sequencing are the most employed techniques, but not exclusively. Novel epigenetic techniques, such as those provided by CRISPR/Cas9 system, represent new opportunities in the searching for epigenetic biomarkers (98).

Many of the findings achieved thus far are encouraging, revealing significant associations with epigenetic modulations of genes regulating neurotransmission, neurodevelopment, and immune function in psychiatric diseases (99). One example is the hypermethylation of BDNF gene identified in brain and peripheral blood samples of MDD, SCZ and BD patients (100, 101). Another similar example is the hypermethylation of FKBP5 gene, an important modulator of stress response, detected in peripheral blood samples of PTSD patients (102). In panic disorder, hypomethylation of monoamine oxidase A (MAOA) and glutamate decarboxylases 1 (GAD1) genes have been evident in recent studies (103).

In suicide, advances in epigenetic techniques have allow to characterize epigenetic alterations in key elements of the hypothalamus-pituitary-adrenal axis (HPA-axis), neurotrophic factors, serotonergic and GABAergic systems, that have been proposed as epigenetic biomarkers for suicide, suicide ideation and suicide attempt (104).

Interestingly, epigenetic biomarkers have been pointed out as potential biomarkers for guiding treatment. Thus, antipsychotic drugs, such as olanzapine, induced DNA methylations alterations through the brain in SCZ patients, changes related with its efficacy (105, 106). For instance, reduced response to antidepressants has been associated with the absence of methylation at a specific CpG site in exon 4 of BDNF in MDD patients (107). Consequently, BDNF exon 4 methylation, and circulating BDNF protein levels may be used together as a predictive tool to personalize treatment of MDD (108).

More interestingly, histone deacetylases (HDAC), that have been demonstrated to control epigenetic programming associated with the modulation of behaviour and cognition, appears to be crucial for reversing dysfunctional epigenetic regulation induced by early life events exposure in preclinical models (109, 110). Additional studies have supported the potential role of HDAC as promising new therapeutic targets for the treatment of MDD (111). In this context, HDAC inhibitors, alone or in combination with current antidepressant drugs, are currently being explored (112–114).

Altogether, epigenetic studies highlight the importance of epigenetic mechanisms on controlling genes or gene complexes. In neuropsychiatry, despite huge advances were achieved, there are still far for providing a clear molecular mechanism underlying these disorders and effective treatment options. The heterogeneity of the techniques and methods used, with a range in sensitivity for detecting effects (115–117); the lack of adjusting the genome-wide results to account for cell specificity (118, 119); the confounding factors such as patient's treatment, population origin and phenotypes included (105, 120); and the lack of further studies to demonstrate the concordance between brain-blood data have hampered the clinical use of epigenetic biomarkers (121).

SUMMARY AND CONCLUSIONS

As set out in this review, there are several proteins, metabolites and genes that have been linked with certain neuropsychiatric diseases mainly due to the advance in ‘omics’ technologies. However, none of them have demonstrated to be a real and useful biomarker in clinical practice.

Despite each ‘omic’ presents its limitations and challenges (122–124), three essential key targets are in common to advance in the searching of biomarkers in neuropsychiatry: 1) accurate selection of the clinical population, 2) shortened sampling time and 3) standardization of procedures for sample processing. These items can be applied for any diseases, but are of special interest for psychiatric disorders. The broad spectrum of phenotypes in patients diagnosed from the same psychiatric disorder and the overlapping of some traits or clusters in different neuropsychiatric disorders, which can often make diagnosis difficult, increases the heterogeneity of the clinical population analyzed. To overcome this issue, emerged ‘omics’ studies have focused on the identification of potential biomarkers for specific traits. However, the reduced number of samples

analyzed per trait/phenotype has made difficult to achieve robust conclusions about the potential clinical use of the proposed biomarkers. In this respect, modern ‘omics’ workflows that enable high throughput studies with large cohorts of well-defined samples can solve this problem.

Besides, the heterogeneity of procedures for sample processing along with the differences in power and sensitivity of each ‘omics’ technologies have contribute to that reality. In this respect, new ‘omics’ with better quantity power and sensitivity would contribute to find robust and realistic biomarkers.

One of the major challenges still lying ahead is the way to integrate the plethora of data obtained from each ‘omics’ to reach the holistic realization of a ‘systems biology’ understanding the biological question (125). In this context, bioinformatics tools have been designed to understand the potential of ‘omics’ technology (126).

Another concern is that current biomarker validation is a lengthy and complex process. In essence, this process includes the validation of the method, determined by the characteristics of the assay employed, and the clinical validation, to provide evidences that the biomarker is linked specifically with the

disease or clinical end point under consideration. Is in this aspect in which future longitudinal integrative ‘omics’ studies can be crucial to provide a rigorously biomarkers validation ensuring its sensitivity, specificity, predictive value, and likelihood ratio, by its assessment in a large cohort (normal clinical population). It is expected that in the following years considerable breakthroughs will occur in these regards.

AUTHOR CONTRIBUTIONS

MG-G and JM conceived the presented idea. MG-G took the lead in writing the manuscript. FN, FS, AG, and AA-O wrote the manuscript in consultation with MG-G. All authors provided critical feedback and helped shape the research, analysis, and manuscript.

ACKNOWLEDGMENTS

We thank all participants in this study. We greatly appreciated to Biorender for helping to create **Figures 3 and 4**.

REFERENCES

- Ilyas A, Chesney E, Patel R. Improving life expectancy in people with serious mental illness: should we place more emphasis on primary prevention? *Br J Psychiatry* (2017) 211(4):194–7. doi: 10.1192/bjp.bp.117.203240
- Chesney E, Goodwin GM, Fazel S. Risks of all-cause and suicide mortality in mental disorders: a meta-review. *World Psychiatry* (2014) 13(2):153–60. doi: 10.1002/wps.20128
- OECD, Safety OfEC-oadECfHaF. *Health at a glance: Europe 2018. State of health in the EU cycle*. Paris: OECD Publishing (2018).
- Swartz MS, Stroup TS, McEvoy JP, Davis SM, Rosenheck RA, Keefe RS, et al. What CATIE found: results from the schizophrenia trial. *Psychiatr Serv* (2008) 59(5):500–6. doi: 10.1176/ps.2008.59.5.500
- Sinyor M, Schaffer A, Levitt A. The sequenced treatment alternatives to relieve depression (STAR*D) trial: a review. *Can J Psychiatry* (2010) 55(3):126–35. doi: 10.1177/070674371005500303
- Parikh SV, LeBlanc SR, Ovanessian MM. Advancing bipolar disorder: key lessons from the Systematic Treatment Enhancement Program for Bipolar Disorder (STEP-BD). *Can J Psychiatry* (2010) 55(3):136–43. doi: 10.1177/070674371005500304
- Gorman JM. Comorbid depression and anxiety spectrum disorders. *Depression Anxiety* (1996) 4(4):160–8. doi: 10.1002/(SICI)1520-6394(1996)4:4<160::AID-DA2>3.0.CO;2-J
- Angst J, Merikangas K. The depressive spectrum: diagnostic classification and course. *J Affect Disord* (1997) 45(1-2):31–9; discussion 9–40. doi: 10.1016/s0165-0327(97)00057-8
- Pincus HA, Tew JD, First MB. Psychiatric comorbidity: is more less? *World Psychiatry* (2004) 3(1):18–23.
- Green AI, Canuso CM, Brenner MJ, Wojcik JD. Detection and management of comorbidity in patients with schizophrenia. *Psychiatr Clinics North America* (2003) 26(1):115–39. doi: 10.1016/s0193-953x(02)00014-x
- Buckley PF, Miller BJ, Lehrer DS, Castle DJ. Psychiatric comorbidities and schizophrenia. *Schizophr Bull* (2009) 35(2):383–402. doi: 10.1093/schbul/sbn135
- Mundkur BD. Evidence excluding mutations, polysomy, and polyploidy as possible causes of non-mendelian segregations in *Saccharomyces*. *Ann Missouri Botanical Garden* (1949) 36(3):23. doi: 10.2307/2394394
- Porter KA. Effect of homologous bone marrow injections in x-irradiated rabbits. *Br J Exp Pathol* (1957) 38(4):401–12.
- Biomarkers Definitions Working G. Biomarkers and surrogate endpoints: preferred definitions and conceptual framework. *Clin Pharmacol Ther* (2001) 69(3):89–95. doi: 10.1067/mcp.2001.113989
- FitzGerald GA. Measure for Measure: Biomarker standards and transparency. *Sci Trans Med* (2016) 8(343):343fs10. doi: 10.1126/scitranslmed.aaf8590
- Biomarker Working Group F-N. BEST (Biomarkers, Endpoints, and other Tools) Resource. In: Spring S, editor. *BEST (Biomarkers, Endpoints, and other Tools) Resource*. Silver Spring (MD): FDA-NIH (2016).
- Aronson JK, Ferner RE. Biomarkers-A General Review. *Curr Protoc Pharmacol* (2017) 76:9 23:1–9 17. doi: 10.1002/cpph.19
- Cagney DN, Sul J, Huang RY, Ligon KL, Wen PY, Alexander BM. The FDA NIH Biomarkers, EndpointS, and other Tools (BEST) resource in neuro-oncology. *Neuro Oncol* (2018) 20(9):1162–72. doi: 10.1093/neuonc/nox242
- Olsson B, Lautner R, Andreasson U, Ohrfelt A, Portelius E, Bjerke M, et al. CSF and blood biomarkers for the diagnosis of Alzheimer's disease: a systematic review and meta-analysis. *Lancet Neurol* (2016) 15(7):673–84. doi: 10.1016/S1474-4422(16)00070-3
- Califf RM. Biomarker definitions and their applications. *Exp Biol Med (Maywood)* (2018) 243(3):213–21. doi: 10.1177/1535370217750088
- Kraus VB. Biomarkers as drug development tools: discovery, validation, qualification and use. *Nat Rev Rheumatol* (2018) 14(6):354–362. doi: 10.1038/s41584-018-0005-9
- Prata J, Santos SG, Almeida MI, Coelho R, Barbosa MA. Bridging Autism Spectrum Disorders and Schizophrenia through inflammation and biomarkers - pre-clinical and clinical investigations. *J Neuroinflammation* (2017) 14(1):179. doi: 10.1186/s12974-017-0938-y
- Pu S, Setoyama S, Noda T. Association between cognitive deficits and suicidal ideation in patients with major depressive disorder. *Sci Rep* 2017 7(1):11637. doi: 10.1038/s41598-017-12142-8. PMID: 28912439
- Rosen C, Zetterberg H. Cerebrospinal fluid biomarkers for pathological processes in Alzheimer's disease. *Curr Opin Psychiatry* 2013 26(3):276–282. doi: 10.1097/YCO.0b013e32835f6747
- Wiecki TV, Antoniadou CA, Stevenson A, Kennard C, Borowsky B, Owen G, et al. A Computational Cognitive Biomarker for Early-Stage Huntington's Disease. *PLoS One* 2016 11(2):e0148409. doi: 10.1371/journal.pone.0148409
- Okazaki S, Hishimoto A, Otsuka I, Watanabe Y, Numata S, Boku S, et al. Increased serum levels and promoter polymorphisms of macrophage

- migration inhibitory factor in schizophrenia. *Prog Neuropsychopharmacol Biol Psychiatry* (2018) 83:33–41. doi: 10.1016/j.pnpbp.2018.01.001
27. Zhou W, Xu Y, Lv Q, Sheng YH, Chen L, Li M, et al. Genetic Association of Olanzapine Treatment Response in Han Chinese Schizophrenia Patients. *Front Pharmacol* (2019) 10:177. doi: 10.3389/fphar.2019.00177
 28. Ghosh R, Tabrizi SJ. Huntington disease. *Handb Clin Neurol* (2018) 147:255–78. doi: 10.1016/B978-0-444-63233-3.00017-8
 29. Llerena A, Berecz R, Dorado P, de la Rubia A. QTc interval, CYP2D6 and CYP2C9 genotypes and risperidone plasma concentrations. *J Psychopharmacol* (2004) 18(2):189–93. doi: 10.1177/0269881104042618
 30. Hartwig FP, Borges MC, Horta BL, Bowden J, Davey Smith G. Inflammatory Biomarkers and Risk of Schizophrenia: A 2-Sample Mendelian Randomization Study. *JAMA Psychiatry* (2017) 74(12):1226–33. doi: 10.1001/jamapsychiatry.2017.3191
 31. Lescuyer P, Hochstrasser D, Rabilloud T. How shall we use the proteomics toolbox for biomarker discovery? *J Proteome Res* (2007) 6(9):3371–6. doi: 10.1021/pr0702060
 32. Gladkevich A, Kauffman HF, Korf J. Lymphocytes as a neural probe: potential for studying psychiatric disorders. *Prog Neuropsychopharmacol Biol Psychiatry* (2004) 28(3):559–76. doi: 10.1016/j.pnpbp.2004.01.009
 33. Quan N, Herkenham M. Connecting cytokines and brain: a review of current issues. *Histol Histopathol* (2002) 17(1):273–88. doi: 10.14670/HH-17.273
 34. Elenkov IJ, Chrousos GP, Wilder RL. Neuroendocrine regulation of IL-12 and TNF-alpha/IL-10 balance. Clinical implications. *Ann New York Acad Sci* (2000) 917:94–105. doi: 10.1111/j.1749-6632.2000.tb05374.x
 35. Peng S, Li W, Lv L, Zhang Z, Zhan X. BDNF as a biomarker in diagnosis and evaluation of treatment for schizophrenia and depression. *Discovery Med* (2018) 26(143):127–36.
 36. Chu CS, Chu CL, Wu CC, Lu T. Serum nerve growth factor beta, brain- and glial-derived neurotrophic factor levels and psychopathology in unmedicated patients with schizophrenia. *J Chin Med Assoc : JCMSA* (2018) 81(6):577–81. doi: 10.1016/j.jcma.2017.11.010
 37. Annunziata P, Cioni C, Mugnaini C, Corelli F. Potent immunomodulatory activity of a highly selective cannabinoid CB2 agonist on immune cells from healthy subjects and patients with multiple sclerosis. *J Neuroimmunol* (2017) 303:66–74. doi: 10.1016/j.jneuroim.2016.12.009
 38. de Campos-Carli SM, Araujo MS, de Oliveira Silveira AC, de Rezende VB, Rocha NP, Ferretjans R, et al. Cannabinoid receptors on peripheral leukocytes from patients with schizophrenia: Evidence for defective immunomodulatory mechanisms. *J Psychiatr Res* (2017) 87:44–52. doi: 10.1016/j.jpsychires.2016.12.001
 39. Ilani T, Ben-Shachar D, Strous RD, Mazor M, Sheinkman A, Kotler M, et al. A peripheral marker for schizophrenia: Increased levels of D3 dopamine receptor mRNA in blood lymphocytes. *Proc Natl Acad Sci U S A* (2001) 98(2):625–8. doi: 10.1073/pnas.021535398
 40. Lima L, Urbina M. Serotonin transporter modulation in blood lymphocytes from patients with major depression. *Cell Mol Neurobiol* (2002) 22(5-6):797–804. doi: 10.1023/a:1021869310702
 41. Hernandez E, Lastra S, Urbina M, Carreira I, Lima L. Serotonin, 5-hydroxyindoleacetic acid and serotonin transporter in blood peripheral lymphocytes of patients with generalized anxiety disorder. *Int Immunopharmacol* (2002) 2(7):893–900. doi: 10.1016/s1567-5769(02)00025-5
 42. Matsui S. Genomic biomarkers for personalized medicine: development and validation in clinical studies. *Comput Math Methods Med* (2013) 2013:865980. doi: 10.1155/2013/865980
 43. Novelli G, Ciccacci C, Borgiani P, Papalucci Amati M, Abadie E. Genetic tests and genomic biomarkers: regulation, qualification and validation. *Clin cases Miner Bone Metab* (2008) 5(2):149–54.
 44. European Medicines Agency (EMA). ICH Topic E15 Definitions for genomic biomarkers, pharmacogenomics, pharmacogenetics, genomic data and sample coding categories. (2007).
 45. European Medicines Agency (EMA). Definitions for genomic biomarkers, pharmacogenomics, pharmacogenetics, genomic data and sample coding categories. (2007).
 46. Jiang W, King TZ, Turner JA. Imaging Genetics Towards a Refined Diagnosis of Schizophrenia. *Front Psychiatry* (2019) 10:494. doi: 10.3389/fpsy.2019.00494
 47. Martin AR, Daly MJ, Robinson EB, Hyman SE, Neale BM. Predicting Polygenic Risk of Psychiatric Disorders. *Biol Psychiatry* (2019) 86(2):97–109. doi: 10.1016/j.biopsych.2018.12.015
 48. van Westrhenen R, Aitchison KJ, Ingelman-Sundberg M, Jukic MM. Pharmacogenomics of Antidepressant and Antipsychotic Treatment: How Far Have We Got and Where Are We Going? *Front Psychiatry* (2020) 11:94. doi: 10.3389/fpsy.2020.00094
 49. Corponi F, Fabbri C, Serretti A. Pharmacogenetics in Psychiatry. *Adv Pharmacol* (2018) 83:297–331. doi: 10.1016/bs.apha.2018.03.003
 50. Baresic A, Nash AJ, Dahoun T, Howes O, Lenhard B. Understanding the genetics of neuropsychiatric disorders: the potential role of genomic regulatory blocks. *Mol Psychiatry* (2020) 25(1):6–18. doi: 10.1038/s41380-019-0518-x
 51. Maul S, Giegling I, Fabbri C, Corponi F, Serretti A, Rujescu D. Genetics of resilience: Implications from genome-wide association studies and candidate genes of the stress response system in posttraumatic stress disorder and depression. *Am J Med Genet Part B Neuropsychiatr Genet : Off Publ Int Soc Psychiatr Genet* (2020) 183(2):77–94. doi: 10.1002/ajmg.b.32763
 52. Ikeda M, Saito T, Kondo K, Iwata N. Genome-wide association studies of bipolar disorder: A systematic review of recent findings and their clinical implications. *Psychiatry Clin Neurosci* (2018) 72(2):52–63. doi: 10.1111/pcn.12611
 53. Sanchez-Roige S, Palmer AA, Clarke TK. Recent Efforts to Dissect the Genetic Basis of Alcohol Use and Abuse. *Biol Psychiatry* (2020) 87(7):609–18. doi: 10.1016/j.biopsych.2019.09.011
 54. Kinreich S, Meyers JL, Maron-Katz A, Kamarajan C, Pandey AK, Chorlian DB, et al. Predicting risk for Alcohol Use Disorder using longitudinal data with multimodal biomarkers and family history: a machine learning study. *Mol Psychiatry* (2019). doi: 10.1038/s41380-019-0534-x
 55. Wu TN, Lee CS, Wu BJ, Sun HJ, Chang CH, Chen CY, et al. Immunophenotypes associated with bipolar disorder and lithium treatment. *Sci Rep* (2019) 9(1):17453. doi: 10.1038/s41598-019-53745-7
 56. Schizophrenia Working Group of the Psychiatric Genomics Consortium. Biological insights from 108 schizophrenia-associated genetic loci. *Nature* (2014) 511(7510):421–7. doi: 10.1038/nature13595
 57. Ripke S, O'Dushlaine C, Chambert K, Moran JL, Kahler AK, Akterin S, et al. Genome-wide association analysis identifies 13 new risk loci for schizophrenia. *Nat Genet* (2013) 45(10):1150–9. doi: 10.1038/ng.2742
 58. Goes FS, McGrath J, Avramopoulos D, Wolyniec P, Pirooznia M, Ruczinski I, et al. Genome-wide association study of schizophrenia in Ashkenazi Jews. *Am J Med Genet Part B Neuropsychiatr Genet : Off Publ Int Soc Psychiatr Genet* (2015) 168(8):649–59. doi: 10.1002/ajmg.b.32349
 59. Stahl EA, Breen G, Forstner AJ, McQuillin A, Ripke S, Trubetskoy V, et al. Genome-wide association study identifies 30 loci associated with bipolar disorder. *Nat Genet* (2019) 51(5):793–803. doi: 10.1038/s41588-019-0397-8
 60. Jaksic N, Sabic Dzanovic E, Aukst Margetic B, Rudan D, Cima Franc A, Bozina N, et al. A Candidate Gene Association Study of FKBP5 and CRRH1 Polymorphisms in Relation to War-Related Posttraumatic Stress Disorder. *Psychiatr Danubina* (2019) 31(2):269–75. doi: 10.24869/psyd.2019.269
 61. Wang Z, Gerstein M, Snyder M. RNA-Seq: a revolutionary tool for transcriptomics. *Nat Rev Genet* (2009) 10(1):57–63. doi: 10.1038/nrg2484
 62. Karuturi SAJGaRKM. Transcriptome Analysis. In: Ranganathan S, Nakai K, Schonbach C editors. *Encyclopedia of Bioinformatics and Computational Biology*. Elsevier (2019) p. 792–805.
 63. Lowe R, Shirley N, Bleackley M, Dolan S, Shafee T. Transcriptomics technologies. *PLoS Comput Biol* (2017) 13(5):e1005457. doi: 10.1371/journal.pcbi.1005457
 64. Hodgson K, Tansey KE, Powell TR, Coppola G, Uher R, Zvezdana Dernovsek M, et al. Transcriptomics and the mechanisms of antidepressant efficacy. *Eur Neuropsychopharmacol* (2016) 26(1):105–12. doi: 10.1016/j.euroneuro.2015.10.009
 65. Lopez JP, Fiori LM, Cruceanu C, Lin R, Labonte B, Cates HM, et al. MicroRNAs 146a/b-5 and 425-3p and 24-3p are markers of antidepressant response and regulate MAPK/Wnt-system genes. *Nat Commun* (2017) 8:15497. doi: 10.1038/ncomms15497
 66. Fiori LM, Lopez JP, Richard-Devantoy S, Berlin M, Chachamovich E, Jollant F, et al. Investigation of miR-1202, miR-135a, and miR-16 in Major Depressive Disorder and Antidepressant Response. *Int J Neuropsychopharmacol / Off Sci J*

- Collegium Internationale Neuropsychopharmacol (CINP)* (2017) 20(8):619–23. doi: 10.1093/ijnp/pyx034
67. Roy B, Dunbar M, Agrawal J, Allen L, Dwivedi Y. Amygdala-Based Altered miRNome and Epigenetic Contribution of miR-128-3p in Conferring Susceptibility to Depression-Like Behavior via Wnt Signaling. *Int J Neuropsychopharmacol / Off Sci J Collegium Internationale Neuropsychopharmacol (CINP)* (2020) 23(3):165–77. doi: 10.1093/ijnp/pyz071
 68. Chimienti F, Cavarec L, Vincent L, Salvétat N, Arango V, Underwood MD, et al. Brain region-specific alterations of RNA editing in PDE8A mRNA in suicide decedents. *Trans Psychiatry* (2019) 9(1):91. doi: 10.1038/s41398-018-0331-3
 69. Li MD, Wang J, Niu T, Ma JZ, Seneviratne C, Ait-Daoud N, et al. Transcriptome profiling and pathway analysis of genes expressed differentially in participants with or without a positive response to topiramate treatment for methamphetamine addiction. *BMC Med Genomics* (2014) 7:65. doi: 10.1186/s12920-014-0065-x
 70. Viola TW, Heberle BA, Zaparte A, Sanvicente-Vieira B, Wainer LM, Fries GR, et al. Peripheral blood microRNA levels in females with cocaine use disorder. *J Psychiatr Res* (2019) 114:48–54. doi: 10.1016/j.jpsychires.2019.03.028
 71. Frantzi M, Bhat A, Latosinska A. Clinical proteomic biomarkers: relevant issues on study design & technical considerations in biomarker development. *Clin Transl Med* (2014) 3(1):7. doi: 10.1186/2001-1326-3-7
 72. Nascimento JM, Martins-de-Souza D. The proteome of schizophrenia. *NPJ Schizophr* (2015) 1:14003. doi: 10.1038/npschz.2014.3
 73. Comes AL, Papiol S, Mueller T, Geyer PE, Mann M, Schulze TG. Proteomics for blood biomarker exploration of severe mental illness: pitfalls of the past and potential for the future. *Trans Psychiatry* (2018) 8(1):160. doi: 10.1038/s41398-018-0219-2
 74. Xu R, Liang J, Luo Y, Wan X, Li K, Qi L, et al. Mass spectrometry identification of potential biomarker proteins in the 150-kD electrophoretic band in patients with schizophrenia. *Med (Baltimore)* (2018) 97(51):e13553. doi: 10.1097/MD.00000000000013553
 75. Rodrigues-Amorim D, Rivera-Baltanas T, Vallejo-Curto MDC, Rodriguez-Jamardo C, de Las Heras E, Barreiro-Villar C, et al. Proteomics in Schizophrenia: A Gateway to Discover Potential Biomarkers of Psychoneuroimmune Pathways. *Front Psychiatry* (2019) 10:885. doi: 10.3389/fpsyt.2019.00885
 76. Nasca C, Bigio B, Lee FS, Young SP, Kautz MM, Albright A, et al. Acetyl-L-carnitine deficiency in patients with major depressive disorder. *Proc Natl Acad Sci U S A* (2018) 115(34):8627–32. doi: 10.1073/pnas.1801609115
 77. Nilsson IAK, Millischer V, Karrenbauer VD, Jureus A, Salehi AM, Norring C, et al. Plasma neurofilament light chain concentration is increased in anorexia nervosa. *Trans Psychiatry* (2019) 9(1):180. doi: 10.1038/s41398-019-0518-2
 78. Katisko K, Cajanus A, Jaaskelainen O, Kontkanen A, Hartikainen P, Korhonen VE, et al. Serum neurofilament light chain is a discriminative biomarker between frontotemporal lobar degeneration and primary psychiatric disorders. *J Neurol* (2020) 267(1):162–7. doi: 10.1007/s00415-019-09567-8
 79. Ferrero AJ, Cereseto M, Sifonios LL, Reines A, Peixoto E, Rubio MC, et al. Cytoskeleton of hippocampal neurons as a target for valproic acid in an experimental model of depression. *Prog Neuropsychopharmacol Biol Psychiatry* (2007) 31(7):1419–28. doi: 10.1016/j.pnpbp.2007.06.014
 80. Jakobsson J, Bjerke M, Ekman CJ, Sellgren C, Johansson AG, Zetterberg H, et al. Elevated concentrations of neurofilament light chain in the cerebrospinal fluid of bipolar disorder patients. *Neuropsychopharmacology* (2014) 39(10):2349–56. doi: 10.1038/npp.2014.81
 81. Isgren A, Sellgren C, Ekman CJ, Holmen-Larsson J, Blennow K, Zetterberg H, et al. Markers of neuroinflammation and neuronal injury in bipolar disorder: Relation to prospective clinical outcomes. *Brain Behav Immun* (2017) 65:195–201. doi: 10.1016/j.bbi.2017.05.002
 82. Quinones MP, Kaddurah-Daouk R. Metabolomics tools for identifying biomarkers for neuropsychiatric diseases. *Neurobiol Dis* (2009) 35(2):165–76. doi: 10.1016/j.nbd.2009.02.019
 83. Martins-de-Souza D. Proteomics, metabolomics, and protein interactomics in the characterization of the molecular features of major depressive disorder. *Dialogues Clin Neurosci* (2014) 16(1):63–73.
 84. Marchand CR, Farshidfar F, Rattner J, Bathe OF. A Framework for Development of Useful Metabolomic Biomarkers and Their Effective Knowledge Translation. *Metabolites* (2018) 8(4):59. doi: 10.3390/metabo8040059
 85. Shih PB. Metabolomics Biomarkers for Precision Psychiatry. *Adv Exp Med Biol* (2019) 1161:101–13. doi: 10.1007/978-3-030-21735-8_10
 86. Grinton KE, Elsea SH. Untargeted Metabolomics for Autism Spectrum Disorders: Current Status and Future Directions. *Front Psychiatry* (2019) 10:647. doi: 10.3389/fpsyt.2019.00647
 87. Konjevod M, Tudor L, Svob Strac D, Nedic Erjavec G, Barbas C, Zarkovic N, et al. Metabolomic and glycomic findings in posttraumatic stress disorder. *Prog Neuropsychopharmacol Biol Psychiatry* (2019) 88:181–93. doi: 10.1016/j.pnpbp.2018.07.014
 88. Davison J, O'Gorman A, Brennan L, Cotter DR. A systematic review of metabolite biomarkers of schizophrenia. *Schizophr Res* (2018) 195:32–50. doi: 10.1016/j.schres.2017.09.021
 89. Bent S, Lawton B, Warren T, Widjaja F, Dang K, Fahey JW, et al. Identification of urinary metabolites that correlate with clinical improvements in children with autism treated with sulforaphane from broccoli. *Mol Autism* (2018) 9:35. doi: 10.1186/s13229-018-0218-4
 90. Czyst AH, South C, Gadad BS, Arning E, Soyombo A, Bottiglieri T, et al. Can targeted metabolomics predict depression recovery? Results from the CO-MED trial. *Trans Psychiatry* (2019) 9(1):11. doi: 10.1038/s41398-018-0349-6
 91. Khamis MM, Adamko DJ, El-Anead A. Strategies and Challenges in Method Development and Validation for the Absolute Quantification of Endogenous Biomarker Metabolites Using Liquid Chromatography-Tandem Mass Spectrometry. *Mass Spectrometry Rev* (2019). doi: 10.1002/mas.21607
 92. Margueron R, Reinberg D. Chromatin structure and the inheritance of epigenetic information. *Nat Rev Genet* (2010) 11(4):285–96. doi: 10.1038/nrg2752
 93. Kular L, Kular S. Epigenetics applied to psychiatry: Clinical opportunities and future challenges. *Psychiatry Clin Neurosci* (2018) 72(4):195–211. doi: 10.1111/pcn.12634
 94. van Os J, Rutten BP, Poulton R. Gene-environment interactions in schizophrenia: review of epidemiological findings and future directions. *Schizophr Bull* (2008) 34(6):1066–82. doi: 10.1093/schbul/sbn117
 95. Focking M, Doyle B, Munawar N, Dillon ET, Cotter D, Cagney G. Epigenetic Factors in Schizophrenia: Mechanisms and Experimental Approaches. *Mol Neuropsychiatry* (2019) 5(1):6–12. doi: 10.1159/000495063
 96. Dupont C, Armand DR, Brenner CA. Epigenetics: definition, mechanisms and clinical perspective. *Semin Reprod Med* (2009) 27(5):351–7. doi: 10.1055/s-0029-1237423
 97. Garcia-Gimenez JL, Seco-Cervera M, Tollefsbol TO, Roma-Mateo C, Peiro-Chova L, Lapunzina P, et al. Epigenetic biomarkers: Current strategies and future challenges for their use in the clinical laboratory. *Crit Rev Clin Lab Sci* (2017) 54(7-8):529–50. doi: 10.1080/10408363.2017.1410520
 98. Xie N, Zhou Y, Sun Q, Tang B. Novel Epigenetic Techniques Provided by the CRISPR/Cas9 System. *Stem Cells Int* (2018) 2018:7834175. doi: 10.1155/2018/7834175
 99. Smigielski L, Jagannath V, Rossler W, Walitza S, Grunblatt E. Epigenetic mechanisms in schizophrenia and other psychotic disorders: a systematic review of empirical human findings. *Mol Psychiatry* (2020). doi: 10.1038/s41380-019-0601-3
 100. Ikegame T, Bundo M, Murata Y, Kasai K, Kato T, Iwamoto K. DNA methylation of the BDNF gene and its relevance to psychiatric disorders. *J Hum Genet* (2013) 58(7):434–8. doi: 10.1038/jhg.2013.65
 101. Angelucci F, Brene S, Mathe AA. BDNF in schizophrenia, depression and corresponding animal models. *Mol Psychiatry* (2005) 10(4):345–52. doi: 10.1038/sj.mp.4001637
 102. Kang JL, Kim TY, Choi JH, So HS, Kim SJ. Allele-specific DNA methylation level of FKBP5 is associated with post-traumatic stress disorder. *Psychoneuroendocrinology* (2019) 103:1–7. doi: 10.1016/j.psyneuen.2018.12.226

103. Kim EJ, Kim YK. Panic disorders: The role of genetics and epigenetics. *AIMS Genet* (2018) 5(3):177–90. doi: 10.3934/genet.2018.3.177
104. Cheung S, Woo J, Maes MS, Zai CC. Suicide epigenetics, a review of recent progress. *J Affect Disord* (2020) 265:423–38. doi: 10.1016/j.jad.2020.01.040
105. Ovenden ES, McGregor NW, Emsley RA, Warnich L. DNA methylation and antipsychotic treatment mechanisms in schizophrenia: Progress and future directions. *Prog Neuropsychopharmacol Biol Psychiatry* (2018) 81:38–49. doi: 10.1016/j.pnpbp.2017.10.004
106. Melka MG, Castellani CA, Rajakumar N, O'Reilly R, Singh SM. Olanzapine-induced methylation alters cadherin gene families and associated pathways implicated in psychosis. *BMC Neurosci* (2014) 15:112. doi: 10.1186/1471-2202-15-112
107. Tadic A, Muller-Engling L, Schlicht KF, Kotsiari A, Dreimuller N, Kleimann A, et al. Methylation of the promoter of brain-derived neurotrophic factor exon IV and antidepressant response in major depression. *Mol Psychiatry* (2014) 19(3):281–3. doi: 10.1038/mp.2013.58
108. Lieb K, Dreimuller N, Wagner S, Schlicht K, Falter T, Neyazi A, et al. BDNF Plasma Levels and BDNF Exon IV Promoter Methylation as Predictors for Antidepressant Treatment Response. *Front Psychiatry* (2018) 9:511. doi: 10.3389/fpsy.2018.00511
109. Machado-Vieira R, Ibrahim L, Zarate CA Jr. Histone deacetylases and mood disorders: epigenetic programming in gene-environment interactions. *CNS Neurosci Ther* (2011) 17(6):699–704. doi: 10.1111/j.1755-5949.2010.00203.x
110. Zhao WN, Ghosh B, Tyler M, Lalonde J, Joseph NF, Kosaric N, et al. Class I Histone Deacetylase Inhibition by Tianeptiline Modulates Neuroplasticity and Enhances Memory. *ACS Chem Neurosci* (2018) 9(9):2262–73. doi: 10.1021/acscchemneuro.8b00116
111. Schroeder M, Hillemecher T, Bleich S, Frieling H. The epigenetic code in depression: implications for treatment. *Clin Pharmacol Ther* (2012) 91(2):310–4. doi: 10.1038/clpt.2011.282
112. Kv A, Madhana RM, Js IC, Lahkar M, Sinha S, Naidu VGM. Antidepressant activity of vorinostat is associated with amelioration of oxidative stress and inflammation in a corticosterone-induced chronic stress model in mice. *Behav Brain Res* (2018) 344:73–84. doi: 10.1016/j.bbr.2018.02.009
113. Deussing JM, Jakovcevski M. Histone Modifications in Major Depressive Disorder and Related Rodent Models. *Adv Exp Med Biol* (2017) 978:169–83. doi: 10.1007/978-3-319-53889-1_9
114. Fuchikami M, Yamamoto S, Morinobu S, Okada S, Yamawaki Y, Yamawaki S. The potential use of histone deacetylase inhibitors in the treatment of depression. *Prog Neuropsychopharmacol Biol Psychiatry* (2016) 64:320–4. doi: 10.1016/j.pnpbp.2015.03.010
115. Olova N, Krueger F, Andrews S, Oxley D, Berrens RV, Branco MR, et al. Comparison of whole-genome bisulfite sequencing library preparation strategies identifies sources of biases affecting DNA methylation data. *Genome Biol* (2018) 19(1):33. doi: 10.1186/s13059-018-1408-2
116. Walker DL, Bhagwate AV, Baheti S, Smalley RL, Hilker CA, Sun Z, et al. DNA methylation profiling: comparison of genome-wide sequencing methods and the Infinium Human Methylation 450 Bead Chip. *Epigenomics* (2015) 7(8):1287–302. doi: 10.2217/EPI.15.64
117. Timmons JA, Szkop KJ, Gallagher JJ. Multiple sources of bias confound functional enrichment analysis of global -omics data. *Genome Biol* (2015) 16:186. doi: 10.1186/s13059-015-0761-7
118. Kinoshita M, Numata S, Tajima A, Ohi K, Hashimoto R, Shimodera S, et al. Aberrant DNA methylation of blood in schizophrenia by adjusting for estimated cellular proportions. *Neuromol Med* (2014) 16(4):697–703. doi: 10.1007/s12017-014-8319-5
119. Montano C, Taub MA, Jaffe A, Briem E, Feinberg JL, Trygvaldottir R, et al. Association of DNA Methylation Differences With Schizophrenia in an Epigenome-Wide Association Study. *JAMA Psychiatry* (2016) 73(5):506–14. doi: 10.1001/jamapsychiatry.2016.0144
120. Rahmani E, Shenav L, Schweiger R, Yousefi P, Huen K, Eskenazi B, et al. Genome-wide methylation data mirror ancestry information. *Epigenet Chromatin* (2017) 10:1. doi: 10.1186/s13072-016-0108-y
121. Edgar RD, Jones MJ, Meaney MJ, Turecki G, Kobor MS. BECon: a tool for interpreting DNA methylation findings from blood in the context of brain. *Trans Psychiatry* (2017) 7(8):e1187. doi: 10.1038/tp.2017.171
122. Kim SH, Weiss C, Hoffmann U, Borggrete M, Akin I, Behnes M. Advantages and Limitations of Current Biomarker Research: From Experimental Research to Clinical Application. *Curr Pharmaceut Biotechnol* (2017) 18(6):445–55. doi: 10.2174/1389201018666170601091205
123. Lippolis JD, Powell EJ, Reinhardt TA, Thacker TC, Casas E. Symposium review: Omics in dairy and animal science-Promise, potential, and pitfalls. *J dairy Sci* (2019) 102(5):4741–54. doi: 10.3168/jds.2018-15267
124. Jiang D, Armour CR, Hu C, Mei M, Tian C, Sharpton TJ, et al. Microbiome Multi-Omics Network Analysis: Statistical Considerations, Limitations, and Opportunities. *Front Genet* (2019) 10:995. doi: 10.3389/fgene.2019.00995
125. Misra BB, Langefeld CD, Olivier M, Cox LA. Integrated Omics: Tools, Advances, and Future Approaches. *J Mol Endocrinol* (2018). doi: 10.1530/JME-18-0055
126. Liang Y, Kelemen A. Dynamic modeling and network approaches for omics time course data: overview of computational approaches and applications. *Briefings Bioinf* (2018) 19(5):1051–68. doi: 10.1093/bib/bbx036

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Association Between *FoxO1*, *A2M*, and *TGF-β1*, Environmental Factors, and Major Depressive Disorder

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OPEN ACCESS

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Specialty section:

This article was submitted to
Mood and Anxiety Disorders,
a section of the journal
Frontiers in Psychiatry

Received: 01 January 2020

Accepted: 29 June 2020

Published: 10 July 2020

Citation:

Zhao M, Chen L, Qiao Z, Zhou J,
Zhang T, Zhang W, Ke S, Zhao X,
Qiu X, Song X, Zhao E, Pan H, Yang Y
and Yang X (2020) Association
Between *FoxO1*, *A2M*, and *TGF-β1*,
Environmental Factors, and Major
Depressive Disorder.
Front. Psychiatry 11:675.
doi: 10.3389/fpsy.2020.00675

Introduction: Investigations of gene-environment (G×E) interactions in major depressive disorder (MDD) have been limited to hypothesis testing of candidate genes while poly-gene-environmental causation has not been adequately address. To this end, the present study analyzed the association between three candidate genes, two environmental factors, and MDD using a hypothesis-free testing approach.

Methods: A logistic regression model was used to analyze interaction effects; a hierarchical regression model was used to evaluate the effects of different genotypes and the dose-response effects of the environment; genetic risk score (GRS) was used to estimate the cumulative contribution of genetic factors to MDD; and protein-protein interaction (PPI) analyses were carried out to evaluate the relationship between candidate genes and top MDD susceptibility genes.

Results: Allelic association analyses revealed significant effects of the interaction between the candidate genes *Forkhead box (Fox)O1*, *α2-macroglobulin (A2M)*, and *transforming growth factor (TGF)-β1* genes and the environment on MDD. Gene-gene (G×G) and gene-gene-environment (G×G×E) interactions in MDD were also included in the model. Hierarchical regression analysis showed that the effect of environmental factors on MDD was greater in homozygous than in heterozygous mutant genotypes of the *FoxO1* and *TGF-β1* genes; a dose-response effect between environment and MDD on genotypes was also included in this model. Haplotype analyses revealed significant global and individual effects of haplotypes on MDD in the whole sample as well as in subgroups. There was a significant association between GRS and MDD ($P = 0.029$) and a GRS and environment interaction effect on MDD ($P = 0.009$). Candidate and top susceptibility genes were connected in PPI networks.

Conclusions: *FoxO1*, *A2M*, and *TGF-β1* interact with environmental factors and with each other in MDD. Multi-factorial G×E interactions may be responsible for a higher explained variance and may be associated with causal factors and mechanisms that could inform new diagnosis and therapeutic strategies, which can contribute to the personalized medicine of MDD.

Keywords: *FoxO1*, *A2M*, *TGF-β1*, depression, G×E interaction, polymorphism

INTRODUCTION

Major depressive disorder (MDD) is the most common psychiatric disorder and is associated with high morbidity, mortality, costs, and risk of suicide (1–3). The heritability of MDD is approximately 30%–40% (4) and may be greater for recurrent, early-onset disease (5). Establishing the contribution of genetics to MDD can lead to more accurate and timely diagnosis and clinical management, which has significant public health implications.

Studies on main genetic effects in MDD have reported conflicting findings (6). One reason for this is that risk of MDD is polygenic, with many susceptibility genes exerting small effects (7). Additionally, genotypic variations among individuals may increase the risk of MDD only upon exposure to adverse environmental factors, a phenomenon known as gene-environment (G×E) interaction (8). The association between candidate susceptibility genes and disease occurrence has mainly been predicated on the observation that genetic variants not only increase MDD risk but can also explain the underlying biological and molecular mechanisms (9). However, such hypothesis-driven approaches to the study of MDD etiology can only detect a fraction of genetic variants (8). On the other hand, a hypothesis-free approach requires enormous datasets that include both exposure [e.g., stressful life events (10, 11) and childhood adversities (9)] and phenotype (MDD) data. To this end, only three genome-wide environmental interaction studies has been carried out (12–14). In a recent report, a novel omics-based approach was used to identify genetic variants for genome-wide G×E interaction studies based on cross-species and -tissue biological prioritization strategies. However, genomics approaches usually cannot identify causal variants or genes (15). Integrating multiple omics approaches can provide a more comprehensive view of the biology of MDD.

Three candidate MDD susceptibility genes [*Forkhead box* (Fox) O1, *transforming growth factor* (TGF)- β 1, and α 2 *macroglobulin* (A2M)] that are ideal candidates for G×E interaction analyses were previously identified using an omics-based approach. It has been suggested that environmental and genetic factors contribute equally to the development of mental illness (16). Distal environmental risk factors such as family history (FH) and culture are important because they increase vulnerability to proximal factors; (17) however, the latter are more relevant for G×E investigations since they are more likely to meet the criteria for risk factors and lend themselves to biologically plausible hypotheses regarding their effects on specific neural systems that underlie psychopathological symptoms.

Genome-wide association studies (GWAS) have identified more than 100 genetic variants associated with MDD (18). Each of these has a small effect and the number of associated genetic markers increases with sample size. However, the utility of individual genetic markers for MDD risk prediction is uncertain (19). A multilocus genetic risk score (GRS)-based analysis has been proposed for combining the relatively small effects of single genes to better assess the complex relationship between genetic markers and MDD (20). GRS analyses have

shown that including more weakly associated genetic variants improves the prediction of mental illness risk (21).

Proteins interact to mediate many cellular processes; disrupting one subunit of a protein complex can lead to direct and indirect functional consequences in various diseases and physiological conditions (22). For a specific illness, interactions between proteins encoded by susceptibility genes tend to be more frequent than those between random proteins, as revealed by protein-protein interaction (PPI) network analyses (22).

In the present study, we investigated the association between *FoxO1*, *TGF- β 1*, and *A2M* genes and MDD in a large population. G×E, G×G, G×G×E, and environment × environment (E×E) interactions were analyzed in the context of MDD. We also assessed haplotypes of these three genes and the relationship between haplotype and environment. We used a GRS approach to calculate the combined effects of these three genes on the prediction of MDD and the GRS-environment interaction (GRS×E) in MDD. Finally, we examined *FoxO1*, *TGF- β 1*, and *A2M* PPI networks to clarify their interactions with each other and with other MDD-related proteins.

MATERIALS AND METHODS

Study Population

From November 2014 to December 2018, 800 MDD patients (564 women and 236 men) were recruited for the study (mean age: 45.64 ± 14.10 years) along with 800 age- and sex-matched control subjects from the same geographic area in Northern China. All subjects were of Chinese Han ethnicity and provided written, informed consent before participating in the study. The study protocol was approved by the ethics committee of Harbin Medical University.

Independent Measures

Participants completed three questionnaires: a socio-demographic questionnaire, the Chinese version of the 24-item Hamilton Rating Scale for Depression (HRSD-24), and the Life Events Scale (LES). The self-rating socio-demographic questionnaire was adapted from the version developed by the Epidemiology Department of Harbin Medical University and was used to collect detailed background information on family psychiatric history, childhood trauma history, and socioeconomic background. The HRSD-24 (23) is widely used to measure depression symptoms; (24–26) a number of patients above the threshold (21 points) were included in the study. The LES questionnaire for measuring negative life events contained 48 items in three dimensions: family life (28 items), work-related problems (13 items), and social and other aspects (seven items). The LES was scored based on the occurrence/absence (1 and 1, respectively) and frequency (0 or more) of SLEs (26, 27).

Genotyping

Single nucleotide polymorphisms (SNPs) in the *FoxO1* (rs2297626, rs7319021, rs28553411, rs17592468, and rs17592371), *A2M* (rs10492115, rs226415, rs10842849, rs11048839, rs10842847, and

rs669), and *TGF-β1* (rs2317130, rs1800469, rs12983775, rs12462166, and rs2241715) genes were selected for genotyping, the candidate SNPs comprised most of the allelic variants with $r^2 > 0.8$ in the Asian population. Genomic DNA was extracted from venous blood samples using the AxyPrep Blood Genomic DNA Miniprep kit (Axygen, Union City, CA, USA). Polymerase chain reaction (PCR) was performed in a reaction volume of 2 μl DNA, 7.5 μl 2× PCR mix, 2 μl primer mix, 0.2 μl *ExoI* enzyme (Fermentas, Burlington, ON, Canada), 0.7 μl *ExoI* buffer (Fermentas), and 0.8 μl FastAP enzyme (Fermentas). Amplification conditions were as follows: at 95°C for 3 min; 35 cycles of 94°C for 15 s, 55°C for 15 s, and 72°C for 30 s; and 72°C for 3 min. PCR products were purified by incubation with *ExoI* and FastAP at 37°C for 15 min and at 80°C for 15 min. Extension conditions were as follows: 96°C for 1 min, and 30 cycles of 96°C for 10 s, 52°C for 5 s, and 60°C for 30 s. Extension products after denaturation at 95°C for 3 min were analyzed by DNA sequencing (Applied Biosystems, Foster City, CA, USA).

Statistical Analysis

Haploview v.4.0 software was used to assess Hardy-Weinberg Equilibrium (HWE) and pairwise linkage disequilibrium (LD) and calculate minimal allele frequency (MAF) for genotyped polymorphisms (28). The χ^2 test/Fisher's exact test was used to analyze differences in the distributions of independent variables. The Bonferroni method was adopted for multiple-testing correction. Associations between phenotype and independent variables were analyzed by multivariable logistic regression, with polymorphisms scored as 0, 1, or 2 depending on the carrier status of the minor allele, sex, family history (FH), occurrence/absence or number of SLEs and CAs, and interaction effects ($E \times E/G \times E/G \times G/G \times G \times E/GRS \times E$). Allele-carrier status and dose-response effect of environmental factors were also determined. Hierarchical regression analysis was performed using RStudio v.1.1.423 software. Alpha levels were corrected with the number of polymorphisms of candidate genes (five polymorphisms for *FoxO1* and *TGF-β1*; six polymorphisms for *A2M*), so that the p values correction was $0.05/5 = 0.01$ for *FoxO1* and *TGF-β1*, $0.05/6 = 0.0083$ for *A2M*. Study power was calculated with QUANTO 1.2.4 (<http://hydra.usc.edu/gxe/>).

Haplotype analysis was performed using UNPHASED v.3.0.11 software (29). Maximum likelihood haplotype frequencies in the study population were assessed with the expectation maximization algorithm. Rare haplotypes with a frequency < 1% were excluded from analyses. The global association and effect of individual haplotypes on phenotype were analyzed in the total sample and in four subgroups classified according to positive/negative environmental factors (SLEs/CAs). The significance level of the global association was determined with the likelihood ratio test. Individual effects of each haplotype, i.e., the difference in effect between a haplotype and all others pooled together, were computed with the score test. A permutation analysis was conducted to assess the reliability of the results; 1,000 random permutations were set to generate empirical P values. Permuted $P < 0.05$ was considered significant in the haplotype analyses.

GRS Analysis

To estimate the cumulative contribution of genetic factors to MDD in an individual by taking into account the reported risk alleles, we used the predictABEL package in RStudio software to compute weighted GRS with the following formula:

$$GRS = \sum_{i=1}^k \beta_i N_i$$

where GRS is the sum of the effect estimates, k is the number of independent genetic variants with strong association as risk predictors, β_i is the weighted coefficient from logistic regression analysis, and N_i is the number of risk alleles for each locus. The association between GRS and MDD was evaluated by logistic regression analysis of FH, sex, GRS, SLEs/CAs, and the interaction between GRS and SLEs/CAs.

Population Stratification Analysis

Population stratification analysis was performed to eliminate the possibility of false-positive associations using STRUCTURE v.2.3.4 software (<http://pritch.bsd.uchicago.edu/structure.html>). *FoxO1*, *A2M*, and *TGF-β1* genotype data were used for population stratification analysis of the third-stage sample set. Sixteen SNPs in these genotype datasets were obtained from 311 samples derived from the 1000 Genomes Project, including 99 samples from the Yoruba population of Ibadan, Nigeria (YRI), 109 samples from Chinese Han in Beijing (CHB), and 103 samples from northern and western Europe and the United States (CEU) (1000 Genomes Project Phase 3 at www.internationalgenome.org/data-portal/sample). STRUCTURE assumes that there are K populations in the dataset. The admixture and correlated frequency models had a burn-in length of 10,000 and 10,000 Markov chain Monte Carlo repeats and took into consideration immigration and geography-based genetic isolation. The program was run several times at each K value from 2 to 6 to obtain consistent results.

PPI Analysis

We investigated whether *FoxO1*, *A2M*, and *TGF-β1* genes interact with each other and are involved in the PPI network containing protein products of the top MDD susceptibility genes identified by a recent GWAS (30–32). The PPI network comprises nodes and edges representing protein and physical interactions, respectively. Proteins encoded by MDD susceptibility genes were used as seed proteins. STRING v.11.0 software (<https://string-db.org/cgi/input.pl>) (33) was used to reconstruct the PPI network.

RESULTS

Descriptive Statistics

Demographic information on the study population is shown in **Table 1**. Genetic markers were successfully genotyped at a rate > 95%. No significant deviation from HWE was observed, and MAF was > 5% for each polymorphism (**Supplemental Table S1**). Pairwise LD D' values are shown in **Figure 1**.

TABLE 1 | Summarized frequencies of demographic and independent variables.

| Variables | Frequencies |
|---|--------------------|
| Gender | |
| Female | 1,023 (63.9%) |
| Male | 578 (36.1%) |
| Mean age | 44.28 (s.d. 11.97) |
| Exposure to CAs | |
| No | 1,412 (88%) |
| Yes | 189 (12%) |
| Exposure to SLEs | |
| No | 820 (51%) |
| Yes | 781 (49%) |
| 1 | 554 (70%) |
| 2 | 74 (9%) |
| 3 or more | 153 (21%) |
| Family history of psychological problems among first-degree relatives | |
| FH- | 1,440 (90%) |
| FH+ | 161 (10%) |

FH, family history.

Association Between Independent Variables and MDD

Two polymorphisms of *FoxO1* (rs17592371 and rs2297626), two of *A2M* (rs669 and rs226415), and one of *TGF-β1* (rs12462166) were significantly associated with MDD (Table 2). Family history of MDD ($\chi^2 = 54.823$, $P < 0.0001$), occurrence ($\chi^2 = 406.993$, $P < 0.0001$), and number ($\chi^2 = 781.522$, $P < 0.0001$) of SLEs were related to MDD. The results remained robust after Bonferroni correction (Table 2). The power for association study of *FoxO1* (rs17592371 and rs2297626) was 87.43% and 90.45% and the power of *TGF-β1* (rs12462166) was 86.55%.

Interaction Between SLEs and Candidate Gene Genotypes in MDD

The two *FoxO1* (rs17592371 and rs28553411), three *A2M* (rs10842847, rs10842849, and rs226415), and two *TGF-β1* (rs12462166 and rs12983775) gene polymorphisms showed significant interactions with environmental factors (Table 3). In the hierarchical regression analysis, the effect of SLEs on MDD

TABLE 2 | Association of Study Variables with MDD.

| Variable | Subgroup | Number | | χ^2 Value | P Value | P-Bonferroni |
|---------------|------------|---------|---------|----------------|---------|--------------|
| FoxO1 | | Case | Control | | | |
| | rs17592371 | CC | 293 359 | 9.790 | 0.007 | 0.035 |
| | | CT | 387 318 | | | |
| | | TT | 120 123 | | | |
| rs2297626 | AA | 293 356 | 11.424 | 0.003 | 0.015 | |
| | AG | 382 321 | | | | |
| | GG | 125 123 | | | | |
| A2M | | | | | | |
| | rs669 | AA | 694 644 | Fisher's | 0.001 | 0.006 |
| | | AG | 101 152 | | | |
| | | GG | 5 4 | | | |
| rs226415 | AA | 6 15 | 10.395 | 0.005 | 0.03 | |
| | AG | 168 158 | | | | |
| | GG | 626 627 | | | | |
| TGF-β1 | | | | | | |
| | rs12462166 | CC | 132 187 | 13.679 | 0.001 | 0.005 |
| | | CT | 454 438 | | | |
| | | TT | 214 175 | | | |
| FH | | | | | | |
| | Yes | 125 36 | 54.823 | <0.001 | <0.001 | |
| | No | 675 765 | | | | |
| SLEs presence | | | | | | |
| | Yes | 592 189 | 406.993 | <0.001 | <0.001 | |
| | No | 208 612 | | | | |
| SLEs amount | | | | | | |
| | 0 | 15 613 | 781.522 | <0.001 | <0.001 | |
| | 1 | 438 116 | | | | |
| | 2 | 38 36 | | | | |
| | 3 or more | 110 36 | | | | |

FH, family history; SLEs, stressful life events.

was stronger for the TT genotype ($\beta = 0.612$, $SE = 0.25$, $z = 2.450$, $P = 0.014$) than for the TC genotype ($\beta = 0.508$, $SE = 0.176$, $z = 2.878$, $P = 0.003$) of *FoxO1* rs17592371; and for the GG genotype ($\beta = 0.692$, $SE = 0.291$, $z = 2.379$, $P = 0.017$) as compared to the GT genotype ($\beta = 0.542$, $SE = 0.166$, $z = 3.258$, $P = 0.001$) of *TGF-β1* rs12462166 (Figure 2A). A dose-response effect of the interaction between number of SLEs and genotypes in MDD was only observed for *A2M* rs10842848, which did not interact with an SLE of 0 ($\beta = 0.099$, $SE = 0.170$, $z = 0.586$, $P = 0.558$) but

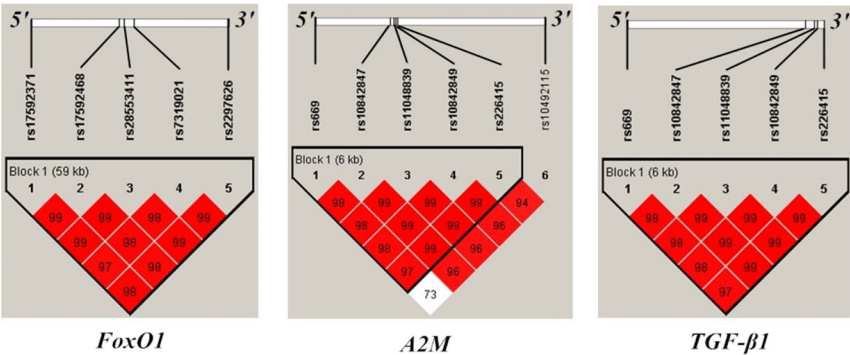


FIGURE 1 | Position and linkage disequilibrium (LD) map of genotyped polymorphisms in *FoxO1*, *A2M*, and *TGF-β1* gene. Pairwise LD statistics in examined genes were calculated with Haploview. Squares are bright red if the $|D'|$ value is high, that is, LD is strong.

TABLE 3 | Effect of Candidate Genes, Environment, and Their Interaction on MDD.

| Dependent Variables | β | SE | z | p Value |
|--------------------------------|---------|-------|--------|---------|
| FoxO1 | | | | |
| rs17592371 | -0.051 | 0.079 | -0.651 | 0.515 |
| CAs | 0.521 | 0.235 | 2.215 | 0.026 |
| rs17592371xCAs | 0.589 | 0.250 | 2.352 | 0.018 |
| rs17592371 | -0.156 | 0.121 | -1.285 | 0.198 |
| SLEs | 1.686 | 0.170 | 9.905 | <0.001 |
| rs17592371xSLEs | 0.468 | 0.171 | 2.739 | 0.006 |
| rs28553411 | -1.210 | 0.148 | -8.136 | <0.001 |
| CAs | 1.622 | 0.223 | 7.247 | <0.001 |
| rs28553411xCAs | 0.616 | 0.202 | 3.047 | 0.002 |
| rs17592371 | 1.881 | 0.393 | 4.781 | <0.001 |
| rs28553411 | -1.746 | 0.421 | -4.141 | <0.001 |
| rs17592371xrs28553411 | 0.275 | 0.108 | 2.547 | 0.010 |
| rs17592371 | 1.133 | 0.679 | 1.668 | 0.095 |
| rs28553411 | -1.219 | 0.679 | -1.796 | 0.072 |
| CAs | -0.083 | 0.202 | -0.413 | 0.679 |
| rs17592371xrs28553411xCAs | 0.321 | 0.109 | 2.922 | 0.003 |
| rs17592371 | 1.104 | 0.721 | 1.531 | 0.125 |
| rs28553411 | -1.226 | 0.721 | -1.700 | 0.089 |
| SLEs | 1.704 | 0.143 | 11.842 | <0.001 |
| rs17592371xrs28553411xSLEs | 0.211 | 0.087 | 2.409 | 0.016 |
| A2M | | | | |
| rs10842847 | -0.142 | 0.120 | -1.181 | 0.238 |
| CAs | -0.133 | 0.183 | -0.725 | 0.468 |
| rs10842847xCAs | 0.894 | 0.371 | 2.409 | 0.016 |
| rs10842847 | -0.206 | 0.176 | -1.169 | 0.242 |
| SLEs | 1.152 | 0.124 | 9.273 | <0.001 |
| rs10842847xSLEs | 0.711 | 0.253 | 2.811 | 0.004 |
| rs10842849 | -0.166 | 0.121 | -1.372 | 0.170 |
| CAs | -0.129 | 0.184 | -0.704 | 0.481 |
| rs10842849xCAs | 0.988 | 0.374 | 2.639 | 0.008 |
| rs10842849 | -0.172 | 0.176 | -0.978 | 0.327 |
| SLEs | 1.283 | 0.124 | 10.309 | <0.001 |
| rs10842849xSLEs | 0.608 | 0.254 | 2.397 | 0.016 |
| rs226415 | 2.310 | 0.194 | 1.201 | 0.023 |
| SLEs | 2.310 | 0.134 | 17.136 | <0.001 |
| rs226415xSLEs | -0.697 | 0.251 | -2.775 | 0.005 |
| rs10842849 | 0.052 | 0.615 | 0.085 | 0.932 |
| rs226415 | 0.700 | 0.673 | 1.039 | 0.298 |
| rs10842849xrs226415 | -0.676 | 0.289 | -2.341 | 0.019 |
| rs10842849 | -0.123 | 0.625 | -0.197 | 0.843 |
| rs226415 | -0.060 | 0.631 | -0.095 | 0.924 |
| CAs | -0.141 | 0.183 | -0.768 | 0.442 |
| rs10842849xrs226415xCAs | 1.132 | 0.378 | 2.989 | 0.002 |
| TGF-β1 | | | | |
| rs12461266 | -0.206 | 0.082 | -2.493 | 0.012 |
| CAs | -0.159 | 0.277 | -0.574 | 0.565 |
| rs12461266xCAs | 0.629 | 0.202 | 3.106 | 0.001 |
| rs12461266 | -0.276 | 0.124 | -2.218 | 0.026 |
| SLEs | 1.488 | 0.193 | 7.667 | <0.001 |
| rs12461266xSLEs | 0.527 | 0.167 | 3.146 | 0.001 |
| rs12983775 | 0.066 | 0.126 | 0.520 | 0.603 |
| SLEs | 2.363 | 0.204 | 11.563 | <0.001 |
| rs12983775xSLEs | -0.479 | 0.167 | -2.857 | 0.004 |

CAs, childhood adversities; SLEs, stressful life events.

showed interactions with an SLE of 1 ($\beta = 0.420$, SE = 0.179, $z = 2.339$, $P = 0.019$), 2 ($\beta = 0.647$, SE = 0.278, $z = 2.330$, $P = 0.019$), and 3 or more ($\beta = 0.749$, SE = 0.295, $z = 2.535$, $P = 0.011$). That is, high exposure to SLEs was related to an increase in MDD risk (Figure 2B).

We evaluated the G×G interaction between *FoxO1* rs17592371 and rs28553411, between the *TGF- β 1* rs12461266 and rs12983775, and among *A2M* rs10842847, rs10842849, and rs226415 as well as the three-way interaction between the two genetic markers that were significant in the G×G interaction analysis and LES (Table 3). The interaction between the two polymorphisms of the *FoxO1* gene was significant ($\beta = 0.275$, SE = 0.108, $z = 2.547$, $P = 0.010$), as was the interaction between those of the *A2M* gene ($\beta = -0.676$, SE = -0.289, $z = -2.341$, $P = 0.019$). The effects of three-way interaction (G×G×E) on MDD were also significant for *FoxO1* (G×G×SLE: $\beta = 0.211$, SE = 0.087, $z = 2.409$, $P = 0.016$; G×G×CA: $\beta = 0.321$, SE = 0.109, $z = 2.922$, $P = 0.003$) and *A2M* (G×G×CA: $\beta = 1.132$, SE = 0.378, $z = 2.989$, $P = 0.002$) (Table 3). These results remained significant after controlling for family history and sex in the regression model. The effect of E×E interaction on MDD was significant ($\beta = 0.746$, SE = 0.312, $z = 2.392$, $P = 0.016$).

Haplotype Analysis

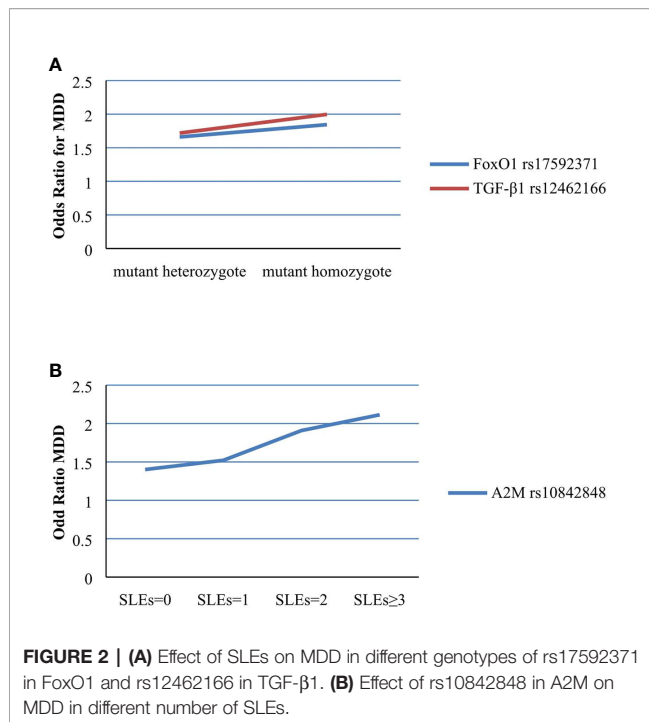
Haplotype analysis was performed for the total sample and subgroups. In the former, 12 haplotypes of five *FoxO1* gene polymorphisms, 15 haplotypes of five *TGF- β 1* gene polymorphisms, and 14 haplotypes of *A2M* gene polymorphisms had a frequency > 1%. The haplotypes of the *FoxO1* gene showed a significant global association ($\chi^2 = 19.732$, df = 13, $P_{\text{global}} < 0.001$) and the less common haplotype (G-A-G-C-T-A) of the *A2M* gene was significantly associated with MDD ($\chi^2 = 4.931$, $P_{\text{effect}} = 0.026$). In subgroup samples, haplotypes of the *A2M* gene within the SLE occurrence group ($\chi^2 = 21.267$, df = 11, $P_{\text{global}} = 0.030$) and haplotypes of *FoxO1* ($\chi^2 = 41.792$, df = 11, $P_{\text{global}} < 0.001$) and *TGF- β 1* ($\chi^2 = 34.531$, df = 14, $P_{\text{global}} = 0.001$) genes within the CA occurrence group were significant in the global association. Two less common haplotypes (G-A-A-C-G-G and G-A-G-C-T-A) of the *A2M* gene within the SLE occurrence group ($\chi^2 = 5.815$, $P_{\text{effect}} = 0.015$; $\chi^2 = 5.814$, $P_{\text{effect}} = 0.015$) and a less common *A2M* haplotype (G-A-G-C-T-A) in the CA occurrence group ($\chi^2 = 3.947$, $P_{\text{effect}} = 0.046$) were associated with MDD.

GRS Analysis

After controlling for FH and sex, logistic regression revealed that the 16 SNP GRSs comprising risk variants were associated with MDD ($\beta = 0.022$, SE = 0.010, $z = 2.180$, $P = 0.029$). Interaction analyses indicated that GRSs interacted with SLEs in MDD ($\beta = 0.054$, SE = 0.021, $z = 2.577$, $P = 0.009$).

Population Stratification Analysis

The combined population of YRI, CHB, and CEU exhibited obvious stratification (Supplemental Figure S1A). In the triangle chart with $K = 3$, each angle represented a potentially independent ancestry and the different colored dots represented individuals in assumed population components that did not cluster with the same color in the triangle, indicating that there was no significant stratification in our sample (Supplemental Figure S1B). Consistent results were obtained with K values ranging from two to six. Thus, population stratification was unlikely to be a confounding factor in this study.



PPI Analysis

We detected interactions among FoxO1, A2M, and TGF-β1 proteins and the protein products of multiple MDD risk genes. The top susceptibility genes except for the *calcium voltage-gated channel subunit α1 C* gene, identified in GWASs of MDD (30–32) formed a densely interconnected PPI network (Figure 3) that

included FoxO1, A2M, and TGF-β1. Interestingly, A2M was found to directly interact with FoxO1 and TGF-β1. TGF-β1 directly interacted with FOS, JUN, cyclic AMP response element-binding protein (CREB) 1, brain-derived neurotrophic factor, AKT1, and Mothers against decapentaplegic homolog (SMAD) 2; and FoxO1 directly interacted with FOS, JUN, CREB binding protein, SMAD2, AKT1, AKT2, and sirtuin 1. These factors have been implicated in MDD genetic risk studies (34–38).

DISCUSSION

The interaction between genetic markers and environment determines the risk of MDD (8). However, most hypothesis-driven studies of MDD to date have not produced reproducible findings. In this study, we investigated candidate MDD risk genes using a hypothesis-free, omics-based, cross-tissue and -species approach. We found that *FoxO1* rs17592371 and rs2297626, *A2M* rs669 and rs226415, and *TGF-β1* rs12462166 alleles were associated with MDD. We also found that *FoxO1* rs17592371 and rs28553411, *A2M* rs10842847, rs10842849, and rs226415, and *TGF-β1* rs12462166 and rs12983775 interacted with the environment in MDD, suggesting a potential relationship between these genes and the etiology of MDD, which could be a bona fide association or the result of linkage. A known allelic association between a gene and a disorder may reflect a G×E interaction, but the absence of such an association does not disqualify a gene as a candidate for disease risk. Some evidences showed that *FoxO1* could express in hippocampus and corpus striatum, *FoxO1* may contribute to the pathological

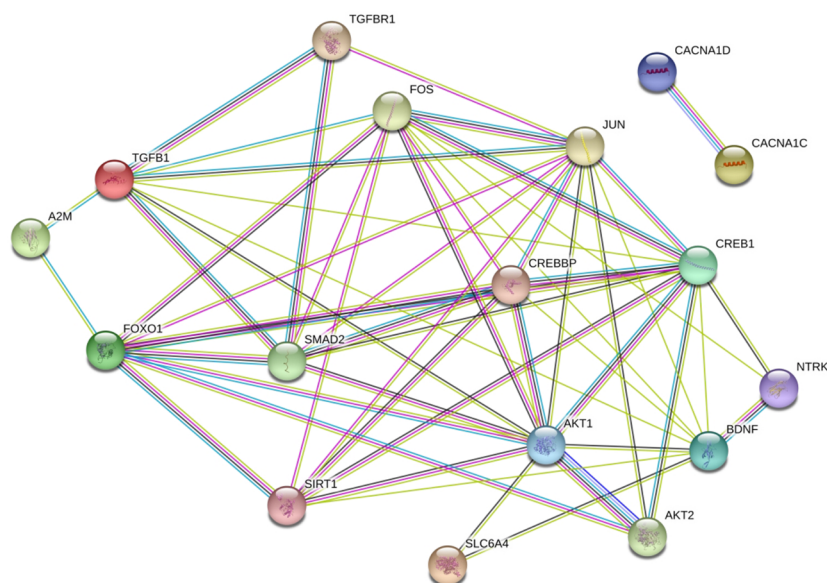


FIGURE 3 | Proteins encoded by FoxO1, A2M, and TGF-β1 in a densely interconnected protein-protein interaction (PPI) network formed by susceptibility genes of MDD. (33) The amaranthine lines indicate known PPIs that have been experimentally determined, and the light blue lines indicate known PPIs from curated databases. The green lines indicate predicted interactions from gene neighborhood, the dark blue lines indicate predicted interactions through gene co-occurrence and the black lines indicate known PPIs from co-expression. The light green lines indicate predicted PPIs through text mining.

process of MDD or other psychiatric disorders (18, 39). It was suggested that immune system associated with MDD and *A2M* played an important role in the immune system, which indicated that *A2M* may associate with MDD. Some studies found that the expression of *A2M* was higher in MDD patients comparing healthy people (21, 40). *TGF- β 1* can restrain autoimmune response and maintain immune system stability, some studies found that *TGF- β 1* could moderate the imbalance of proinflammatory cytokines and anti-inflammatory cytokines in MDD patients (15, 16). All of these evidences above were consistent with our findings.

The carrier status of the minor allele and number of SLEs were separately used to test different genotypic effects on MDD and the dose-response relationship between SLEs and MDD. The effect of SLEs on MDD was more highly significant for the TT than for the TC genotype of *FoxO1* rs17592371 and for the GG as compared to the GT genotype of *TGF- β 1* rs12462166. These results indicated that mutant homozygous genotypes may be more significant than the heterozygous genotype for the effect of SLEs on MDD, as previously suggested (41, 42). We also found that exposure to SLEs was related to an increased risk for MDD, which is consistent with previous reports (42, 43), indicating that although the effect of a single environmental factor may be quite small, the cumulative effect of multiple factors may be large and that strong effects typically result from a chain of related events rather than a single factor.

Because gene variants can interact, we examined the SNPs that showed significant G×E interactions to investigate G×G interactions. We found that two polymorphisms in the *FoxO1* gene and the *A2M* gene showed significant interactions and that the G×G×E interaction was also significant. This was similar to a previous finding that *serotonin transporter-linked polymorphic region (5-HTTLPR)* gene and SNP rs140700 both interacted with the environment and with each other in MDD, and that the 5-HTTLPR×rs140700×E interaction was significant (41). Much of the genetic variation associated with a complex trait can be explained by the joint contribution of multiple genetic markers as well as environmental influence (44). G×G and G×G×E interaction analyses are considered as biologically relevant and explain the role of heritability (45–48). We also found that the E×E (CA × SLE) interaction in MDD was significant. CA increases the likelihood of SLE occurrence (49) that is, early negative experiences increase vulnerability to such events later in life, resulting in a sequential E×E interaction. For instance, mistreatment in childhood caused sensitization to the effects of specific types of SLE in adulthood (49). The present study is the first report of an E×E interaction in MDD.

Although direct associations between haplotype and psychiatric disorders have been reported (50, 51), there have been no studies demonstrating an interaction between haplotype and environmental factors. In our study, the global effect of haplotype on MDD was significant for the *FoxO1* and *TGF- β 1* genes in the total sample and in subjects that had experienced CA. A significant individual effect and global effect of haplotype on MDD was found for the *A2M* gene in both the total and subgroup samples. The global effect of haplotype on MDD was

always significant in combination with environmental factors. Interactions existed not only between the environment and genes, but also between the environment and haplotype in MDD.

GRS analyses have shown that the prediction of mental illness is improved by including more weakly associated genetic variants, suggesting that these influence the risk of mental disorder (21, 40). Our results showed that the association between 16 SNP GRS and MDD was significant, with a significant interaction between GRS and SLEs. Some of the 16 SNPs did not show a significant conditional or interaction effect. GRSs may detect the effects of weaker associated genetic variants (52). The results of the PPI analysis showed that *FoxO1*, *A2M*, and *TGF- β 1* were connected, suggesting that a wide variety of cellular processes are involved in MDD. This could explain the reason for the additive effects of *FoxO1*, *A2M*, and *TGF- β 1* gene variants indicated by the GRSs. In addition, several top MDD susceptibility genes were included in this PPI network, although the clinical significance of these associations requires further investigation. The disturbances of certain cellular processes or pathways contributing to the risk of MDD has been emerging and gained evidences supporting (22, 53, 54). Constructing highly interconnected PPI networks between *FoxO1*, *A2M*, and *TGF- β 1* protein and proteins of multiple defined risk genes for MDD may reveal the underlying biological mechanisms. *FoxO1*, *A2M*, and *TGF- β 1* protein could participate in this PPI network, indicating their potential involvement in the common molecular network modulating the pathogenesis of MDD. It is noteworthy that *A2M* protein directly interacted with *TGF- β 1* protein, indicating their connection in the biological process relevant to MDD. A recent study found that *A2M* as an inflammatory fluid proteinase scavenger could bind to a plethora of cytokines, including *TGF- β 1*. (55)

We investigated the potential influence of population stratification in our samples using STRUCTURE software v.2.3.4 and the third-stage sample set, but did not find any evidence of stratification, which confirmed that the results were not affected by this confounding factor.

Our study had three major limitations. First, we did not use standard questionnaires that have confirmed reliability and validity to measure CA. Second, we did not analyze additional SNPs in our samples and the third-stage sample set, which can improve the reliability of population stratification analysis. Lastly, including a gene expression microarray analysis in our study would have allowed us to compare the transcriptomes of subjects exposed to SLEs/CAs and non-exposed individuals, which could provide more information for predicting MDD risk.

In conclusion, our data indicated that the hypothesis-free approach is useful for identifying novel genes contributing to the G×E interaction in MDD. *FoxO1*, *A2M*, and *TGF- β 1* genes not only interact with environmental factors but also associate with multiple other genes in the MDD G×E interaction. *FoxO1*, *A2M*, and *TGF- β 1* genes may serve the clinical diagnose and treatment of MDD by providing biomarkers, which can contribute to personalized medicine. Future studies will need to focus on the complexity of poly-gene–environmental causation to obtain more detailed insight into the etiology of MDD.

DATA AVAILABILITY STATEMENT

All datasets for this study are included in the article/**Supplementary Material**.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by ethics committee of Harbin Medical University. The patients/participants provided their written informed consent to participate in this study.

AUTHOR CONTRIBUTIONS

MZ and LC conducted the statistical analyses and wrote the first draft of the manuscript. ZQ and JZ provided expertise in MDD search. TZ, WZ, and SK did the experiment. XZ, EZ, and XS collected the data. XQ and HP revised the manuscript. YY and XY designed this study and provided expertise. All authors were involved in modifying the secondary-analysis design and editing the manuscript. All authors contributed to the article and approved the submitted version.

REFERENCES

- Wittchen HU, Jacobi F, Rehm J, Gustavsson A, Svensson M, Jonsson B, et al. The size and burden of mental disorders and other disorders of the brain in Europe 2010. *Eur Neuropsychopharmacol* (2011) 21(9):655–79. doi: 10.1016/j.euroneuro.2011.07.018
- Ferrari AJ, Charlson FJ, Norman RE, Patten SB, Freedman G, Murray CJ, et al. Burden of depressive disorders by country, sex, age, and year: findings from the global burden of disease study 2010. *PloS Med* (2013) 10(11):e1001547. doi: 10.1371/journal.pmed.1001547
- Murray CJ, Vos T, Lozano R, Naghavi M, Flaxman AD, Michaud C, et al. Disability-adjusted life years (DALYs) for 291 diseases and injuries in 21 regions, 1990–2010: a systematic analysis for the Global Burden of Disease Study 2010. *Lancet* (2012) 380(9859):2197–223. doi: 10.1016/S0140-6736(12)61690-0
- Sullivan PF, Neale MC, Kendler KS. Genetic epidemiology of major depression: review and meta-analysis. *Am J Psychiatry* (2000) 157(10):1552–62. doi: 10.1176/appi.ajp.157.10.1552
- Viktorin A, Meltzer-Brody S, Kuja-Halkola R, Sullivan PF, Landen M, Lichtenstein P, et al. Heritability of Perinatal Depression and Genetic Overlap With Nonperinatal Depression. *Am J Psychiatry* (2016) 173(2):158–65. doi: 10.1176/appi.ajp.2015.15010085
- Bosker FJ, Hartman CA, Nolte IM, Prins BP, Terpstra P, Posthuma D, et al. Poor replication of candidate genes for major depressive disorder using genome-wide association data. *Mol Psychiatry* (2011) 16(5):516–32. doi: 10.1038/mp.2010.38
- Hyman S. Mental health: depression needs large human-genetics studies. *Nature* (2014) 515(7526):189–91. doi: 10.1038/515189a
- Heim C, Binder EB. Current research trends in early life stress and depression: review of human studies on sensitive periods, gene-environment interactions, and epigenetics. *Exp Neurol* (2012) 233(1):102–11. doi: 10.1016/j.expneurol.2011.10.032
- Caspi A, Sugden K, Moffitt TE, Taylor A, Craig IW, Harrington H, et al. Influence of life stress on depression: moderation by a polymorphism in the 5-HTT gene. *Science* (2003) 301(5631):386–9. doi: 10.1126/science.1083968
- Kendler KS, Kuhn JW, Vittum J, Prescott CA, Riley B. The interaction of stressful life events and a serotonin transporter polymorphism in the

FUNDING

This study was supported by the National Natural Science Foundation of China (81773536) to YY, the Fundamental Research Funds for State Universities of Heilongjiang Province (2017JCZX23) to EZ, and China Postdoctoral Science Foundation Grant (2019M651311) to JZ.

ACKNOWLEDGMENTS

We want to express our gratitude to all patients and healthy controls parting in the study, as well as to the psychiatrists for their help in the recruitment and identification of patients with major depressive disorder.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fpsy.2020.00675/full#supplementary-material>

- prediction of episodes of major depression: a replication. *Arch Gen Psychiatry* (2005) 62(5):529–35. doi: 10.1001/archpsyc.62.5.529
- Stephan R, Benjamin MN, Aiden C, James TRW, Kai-How F, Peter AH, et al. Biological insights from 108 schizophrenia-associated genetic loci. *Nature* (2014) 511(7510):421–7. doi: 10.1038/nature13595
- Otowa T, Kawamura Y, Tsutsumi A, Kawakami N, Kan C, Shimada T, et al. The First Pilot Genome-Wide Gene-Environment Study of Depression in the Japanese Population. *PloS One* (2016) 11(8):e0160823. doi: 10.1371/journal.pone.0160823
- Dunn EC, Wiste A, Radmanesh F, Almli LM, Gogarten SM, Sofer T, et al. Genome-Wide Association Study (GWAS) And Genome-Wide By Environment Interaction Study (GWEIS) Of Depressive Symptoms In African American And Hispanic/Latina Women. *Depress Anxiety* (2016) 33(4):265–80. doi: 10.1002/da.22484
- Cattaneo A, Cattane N, Malpighi C, Czamara D, Suarez A. FoxO1, A2M, and TGF-beta1: three novel genes predicting depression in gene X environment interactions are identified using cross-species and cross-tissues transcriptomic and miRNomic analyses. *Mol Psychiatry* (2018) 23(11):2192–208. doi: 10.1038/s41380-017-0002-4
- Visscher PM, Brown MA, McCarthy MI, Yang J. Five years of GWAS discovery. *Am J Hum Genet* (2012) 90(1):7–24. doi: 10.1016/j.ajhg.2011.11.029
- Uher R, Zwickler A. Etiology in psychiatry: embracing the reality of poly-gene-environmental causation of mental illness. *World Psychiatry* (2017) 16(2):121–9. doi: 10.1002/wps.20436
- Costello EJ, Compton SN, Keeler G, Angold A. Relationships between poverty and psychopathology: a natural experiment. *JAMA* (2003) 290(15):2023–9. doi: 10.1001/jama.290.15.2023
- Hyde CL, Nagle MW, Tian C, Chen X, Paciga SA, Wendland JR. Identification of 15 genetic loci associated with risk of major depression in individuals of European descent. *Nat Genet* (2016) 48(9):1031–6. doi: 10.1038/ng.3623
- Karg K, Burmeister M, Shedden K, Sen S. The serotonin transporter promoter variant (5-HTTLPR), stress, and depression meta-analysis revisited: evidence of genetic moderation. *Arch Gen Psychiatry* (2011) 68(5):444–54. doi: 10.1001/archgenpsychiatry.2010.189
- Dudbridge F. Polygenic Epidemiology. *Genet Epidemiol* (2016) 40(4):268–72. doi: 10.1002/gepi.21966

21. Wray NR, Lee SH, Mehta D, Vinkhuyzen AA, Dudbridge F, Middeldorp CM. Research review: Polygenic methods and their application to psychiatric traits. *J Child Psychol Psychiatry* (2014) 55(10):1068–87. doi: 10.1111/jcpp.12295
22. Luo X, Huang L, Jia P, Li M, Su B, Zhao Z, et al. Protein-protein interaction and pathway analyses of top schizophrenia genes reveal schizophrenia susceptibility genes converge on common molecular networks and enrichment of nucleosome (chromatin) assembly genes in schizophrenia susceptibility loci. *Schizophr Bull* (2014) 40(1):39–49. doi: 10.1093/schbul/sbt066
23. Hamilton M. A rating scale for depression. *J Neurol Neurosurg Psychiatry* (1960) 23:56–62. doi: 10.1136/jnnp.23.1.56
24. Assmann N, Schramm E, Kriston L. Moderating effect of comorbid anxiety disorders on treatment outcome in a randomized controlled psychotherapy trial in early-onset persistently depressed outpatients. *Depres Anxiety* (2018) 35(10):1001–8. doi: 10.1002/da.22839
25. Erkens N, Schramm E, Kriston L, Hautzinger M, Harter M, Schweiger U, et al. Association of comorbid personality disorders with clinical characteristics and outcome in a randomized controlled trial comparing two psychotherapies for early-onset persistent depressive disorder. *J Affect Disord* (2018) 229:262–8. doi: 10.1016/j.jad.2017.12.091
26. Ma J, Wang L, Yang Y, Qiao Z, Fang D, Qiu X, et al. GNB3 and CREB1 gene polymorphisms combined with negative life events increase susceptibility to major depression in a Chinese Han population. *PLoS One* (2017) 12(2): e0170994. doi: 10.1371/journal.pone.0170994
27. Han D, Qiao Z, Chen L, Qiu X, Fang D, Yang X, et al. Interactions between the vascular endothelial growth factor gene polymorphism and life events in susceptibility to major depressive disorder in a Chinese population. *J Affect Disord* (2017) 217:295–8. doi: 10.1016/j.jad.2017.04.028
28. Barrett JC, Fry B, Maller J, Daly MJ. Haploview: analysis and visualization of LD and haplotype maps. *Bioinformatics* (2005) 21(2):263–5. doi: 10.1093/bioinformatics/bth457
29. Dudbridge F. Pedigree disequilibrium tests for multilocus haplotypes. *Genet Epidemiol* (2003) 25(2):115–21. doi: 10.1002/gepi.10252
30. Chang LC, Jaimain S, Lin CW, Rujescu D, Tseng GC, Sibille E. A conserved BDNF, glutamate- and GABA-enriched gene module related to human depression identified by coexpression meta-analysis and DNA variant genome-wide association studies. *PLoS One* (2014) 9(3):e90980. doi: 10.1371/journal.pone.0090980
31. Xiao X, Zhang C, Grigoriu-Serbanescu M, Wang L, Li L, Zhou D, et al. The cAMP responsive element-binding (CREB)-1 gene increases risk of major psychiatric disorders. *Mol Psychiatry* (2018) 23(9):1–11. doi: 10.1038/mp.2017.243
32. Haenisch B, Herms S, Mattheisen M, Steffens M, Breuer R, Strohmaier J, et al. Genome-wide association data provide further support for an association between 5-HTTLPR and major depressive disorder. *J Affect Disord* (2013) 146(3):438–40. doi: 10.1016/j.jad.2012.08.001
33. Falush D, Stephens M, Pritchard JK. Inference of population structure using multilocus genotype data: linked loci and correlated allele frequencies. *Genetics* (2003) 164(4):1567–87.
34. Burokas A, Arbolea S, Moloney RD, Peterson VL, Murphy K, Clarke G, et al. Targeting the Microbiota-Gut-Brain Axis: Prebiotics Have Anxiolytic and Antidepressant-like Effects and Reverse the Impact of Chronic Stress in Mice. *Biol Psychiatry* (2017) 82(7):472–87. doi: 10.1016/j.biopsych.2016.12.031
35. Xu F, Yang J, Chen J, Wu Q, Gong W, Zhang J, et al. Differential co-expression and regulation analyses reveal different mechanisms underlying major depressive disorder and subsyndromal symptomatic depression. *BMC Bioinform* (2015) 16:112. doi: 10.1186/s12859-015-0543-y
36. Noto C, Ota VK, Santoro ML, Gouvea ES, Silva PN, Spindola LM, et al. Depression, Cytokine, and Cytokine by Treatment Interactions Modulate Gene Expression in Antipsychotic Naïve First Episode Psychosis. *Mol Neurobiol* (2016) 53(8):5701–9. doi: 10.1007/s12035-015-9489-3
37. Dow AL, Russell DS, Duman RS. Regulation of activin mRNA and Smad2 phosphorylation by antidepressant treatment in the rat brain: effects in behavioral models. *J Neurosci* (2005) 25(20):4908–16. doi: 10.1523/JNEUROSCI.5155-04.2005
38. Kim HD, Hesterman J. SIRT1 Mediates Depression-Like Behaviors in the Nucleus Accumbens. *J Neurosci* (2016) 36(32):8441–52. doi: 10.1523/JNEUROSCI.0212-16.2016
39. Howard DA-O, Adams MA-O, Shirali MA-O, Clarke TK, Marioni RE, Davies G, et al. Genome-wide association study of depression phenotypes in UK Biobank identifies variants in excitatory synaptic pathways. *Nat Commun* 9(1):1470. doi: 10.1038/s41467-018-03819-3
40. Dudbridge F. Power and predictive accuracy of polygenic risk scores. *PLoS Genet* (2013) 9(3):e1003348. doi: 10.1371/journal.pgen.1003348
41. Lazary J, Lazary A, Gonda X, Benko A, Molnar E, Juhasz G, et al. New evidence for the association of the serotonin transporter gene (SLC6A4) haplotypes, threatening life events, and depressive phenotype. *Biol Psychiatry* (2008) 64(6):498–504. doi: 10.1016/j.biopsych.2008.03.030
42. Evans GW. The environment of childhood poverty. *Am Psychol* (2004) 59(2):77–92. doi: 10.1037/0003-066X.59.2.77
43. Sameroff AJ, Seifer R, Bartko WT. Environmental perspectives on adaptation during childhood and adolescence. In Luthar S. S., Burack J. A., Cicchetti D., Weisz J. R. (Eds.), *Developmental Psychopathology: Perspectives on Adjustment, Risk, and Disorder*. (Cambridge England; New York, NY, USA: Cambridge University Press) (1997). pp. 507–526.
44. Li S, Cui Y. Gene-centric gene-gene interaction: A model-based kernel machine method. *Ann Appl Stat* (2012) 6(2012):1134–61. doi: 10.1214/12-AOAS545
45. Lunzer M, Golding GB, Dean AM. Pervasive cryptic epistasis in molecular evolution. *PLoS Genet* (2010) 6(10):e1001162. doi: 10.1371/journal.pgen.1001162
46. Huang W, Richards S, Carbone MA, Zhu D, Anholt RR, Ayroles JF, et al. Epistasis dominates the genetic architecture of Drosophila quantitative traits. *Proc Natl Acad Sci U.S.A.* (2012) 109(39):15553–9. doi: 10.1073/pnas.1213423109
47. Zuk O, Hechter E, Sunyaev SR, Lander ES. The mystery of missing heritability: Genetic interactions create phantom heritability. *Proc Natl Acad Sci U S A.* (2012) 109(4):1193–8. doi: 10.1073/pnas.1119675109
48. Hemani G, Shakhbazov K, Westra HJ, Esko T, Henders AK, McRae AF, et al. Detection and replication of epistasis influencing transcription in humans. *Nature* (2014) 508(7495):249–53. doi: 10.1038/nature13005
49. Starr LR, Hammen C, Conway CC, Raposa E, Brennan PA. Sensitizing effect of early adversity on depressive reactions to later proximal stress: Moderation by polymorphisms in serotonin transporter and corticotropin releasing hormone receptor genes in a 20-year longitudinal study. *Dev Psychopathol* (2014) 26(4 Pt 2):1241–54. doi: 10.1017/S0954579414000996
50. Brookes KJ, Mill J, Guindalini C, Curran S, Xu X, Knight J, et al. A common haplotype of the dopamine transporter gene associated with attention-deficit/hyperactivity disorder and interacting with maternal use of alcohol during pregnancy. *Arch Gen Psychiatry* (2006) 63(1):74–81. doi: 10.1001/archpsyc.63.1.74
51. Todd RD, Neuman RJ. Gene-environment interactions in the development of combined type ADHD: evidence for a synapse-based model. *Am J Med Genet B Neuropsychiatr Genet* (2007) 144b(8):971–5. doi: 10.1002/ajmg.b.30640
52. Demirkan A, Penninx BW, Hek K, Wray NR, Amin N, Aulchenko YS, et al. Genetic risk profiles for depression and anxiety in adult and elderly cohorts. *Mol Psychiatry* (2011) 16(7):773–83. doi: 10.1038/mp.2010.65
53. Gilman SR, Chang J, Xu B, Bawa TS, Gogos JA, Karayiorgou M, et al. Diverse types of genetic variation converge on functional gene networks involved in schizophrenia. *Nat Neurosci* (2012) 15(12):1723–8. doi: 10.1038/nn.3261
54. Network, Pathway Analysis Subgroup of Psychiatric Genomics C. Psychiatric genome-wide association study analyses implicate neuronal, immune and histone pathways. *Nat Neurosci* (2015) 18(2):199–209. doi: 10.1038/nn.3922
55. Hope C, Mettenberg J, Gonias SL, DeKosky ST, Kamboh MI, Chu CT. Functional analysis of plasma alpha(2)-macroglobulin from Alzheimer's disease patients with the A2M intronic deletion. *Neurobiol Dis* (2003) 14(3):504–12. doi: 10.1016/j.nbd.2003.08.005

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The reviewer JM declared a past co-authorship with several of the authors YY, ZQ, EZ, XS, XY to the handling Editor.

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Searching for the Needles in a Haystack; Is It Needless? The Search for Peripheral Biomarkers in Psychiatry

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Keywords: biomarkers, severe mental illness (SMI), transdiagnostic, standard operating procedure (SOP), translation

OPEN ACCESS

Edited by:

Francisco Navarrete Rueda,
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Reviewed by:

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Specialty section:

This article was submitted to
Molecular Psychiatry,
a section of the journal
Frontiers in Psychiatry

Received: 06 January 2020

Accepted: 30 June 2020

Published: 15 July 2020

Citation:

Vinberg M (2020) Searching for
the Needles in a Haystack;
Is It Needless? The Search for
Peripheral Biomarkers in Psychiatry.
Front. Psychiatry 11:689.
doi: 10.3389/fpsy.2020.00689

INTRODUCTION

Psychiatric diagnoses rely on syndrome description based on experienced symptoms as reported by patients and further transformed into diagnoses based on a professional's (generally a doctor's) education, knowledge and, not least, clinical experience. This is a major challenge and also an Achilles heel in psychiatry. The lack of more precise biological measures creates exposure to criticism against the use of psychiatric diagnoses: on the one hand, that psychiatric diagnoses are non-existent and, on the other hand, the challenge of demarking severe mental illness (SMI) from many minor psychiatric diagnoses that seem less biologically driven. Nevertheless, research methodologies such as proteomics, transcriptomics, genomics, and brain imaging have advanced the pathophysiological understanding of SMI (e.g. schizophrenia, bipolar disorder, and major depressive disorders) in particular (1–3). Although these efforts may seem promising, we face a translational gap in clarifying to what extent such approaches can prove useful in the clinic and help diagnostics to support a targeted treatment choice and optimize treatment overall. In particular, blood, plasma, and serum are untapped sources of possible clinical and research-useful biomarkers (1). Despite an increasing number of studies on biomarkers, so far the area has not contributed to solid clinically improvements in diagnostics or clinical care.

DISCUSSION

This paucity of progress is likely to be due to several factors that, potentially, could be overcome. First, our categorical diagnostic classification systems—the International Classification of Diseases, 10th revision (ICD-10), and the Diagnostic and Statistical Manual of Mental Disorders, 5th revision (DSM-V)—lump very heterogenic syndromes together based on empirical experience and these categories are not directed towards capturing the underlying biology. Consequently, most clinical research anchored in current diagnostic systems will, by nature, create narrow research questions such as: Do we believe that there are significant biological differences on comparing two severely depressed patients, one with unipolar disorder and one with bipolar disorder? One way to circumvent these limitations is to use initiatives such as the Research Domain Criteria (RDoC)

framework (2), which has been created to integrate the observed behavioral information with neurobiological measures that incorporate multiple dimensions (behavior, thought patterns, neurobiological measures, and genetics), with the overall aim of generating a classification system that can be linked to the underlying dysfunctional pathways. Thus, future studies must look beyond the categorical diagnoses and use a transdiagnostic approach that includes a transdiagnostic assessment procedure for describing the individual behavior thoroughly. Along these lines, a systematic review (3) including the most promising peripheral biomarkers (BDNF, TNF-alpha, IL-6, C-reactive protein, and cortisol) concluded that these were most likely to be non-specific as diagnostic biomarkers (3). However, these biomarkers seem useful as an expression of illness activity and severity, thereby potentially being useful for treatment monitoring (3).

Second, the proteomic biomarker area must recognize that the earliest studies in particular are characterized by significant method limitations (4), such as low study quality (lack of consistency concerning the time of day when the biological samples are withdrawn; laboratory technique with a risk of prolonged storage before the biological samples have proceeded), poorly characterized samples, small sample sizes, and substantial unexplained between-study heterogeneity. Furthermore, limitations such as potential bias in individual studies, indications of publication bias, and lack of standard operating procedures for all aspects of the individual assessment can lead to non-replication and failed studies. These factors point to a need to improve the quality of future research. That said, knowledge from all these studies is essential because it can critically inform future warranted high-quality large-scale studies.

Third, it can be a problem to integrate animal models of SMI. However, close cooperation and infrastructures linking findings from animal models to clinical settings are beneficial (e.g. studying one biomarker in a mouse model and also observing the same potential biomarker in humans for both cases and controls). Integrating animal models is necessary to provide direct insight into the cellular metabolites that are produced during psychiatric processes. In addition, the influence on biomarkers due to short- or long-term medication can be observed under controlled conditions. Thus, a combination of representative animal models and human studies are a promising pathway to improve the potential use of biomarkers in psychiatry (5).

Fourth, there are huge commercial interests in this area. Companies are tempted to promise that their specially designed biomarker will capture the early signs of SMI and lead to better treatment results. This could be overly optimistic and will confuse the area, not least the patients. Thus, a combination of industry and academic groups that need to be funded can lead to being overly optimistic without a solid scientific basis for their promises should also be considered. Nevertheless, the impact of private companies' knowledge, ambitious research, and technology is indispensable and well-described transparent cooperation between university-driven research and industry is necessary.

Fifth, another reason for the many non-replicated findings seems to be the inclusion of very different patient populations: it is difficult to compare biomarkers in a sample of newly diagnosed patients with patients having a late-stage disorder characterized by many admissions, polypharmacy, etc. Furthermore, the present illness state is important because biomarkers often change according to the present state (e.g. depressed, manic, psychotic, or in remission). Therefore, it is mandatory to characterize each illness state and illness course (onset of the disorder, number of episodes) as biomarkers change over time and are sensitive to the impact of environmental factors (smoking, substance use, alcohol use), co-existent physical disorder, age and gender; characterization of the menstrual cycle in females should also be taken into account.

Sixth, the impact of psychotropic medication on biomarker measures is complicated and a major confounder in most studies. This problem is difficult to solve but in larger studies it is possible to include at-risk individuals who do not receive medication (e.g. first-degree relatives to patients with SMI). Twin studies can further help to distinguish between potential environmental and genetic origins of the biomarker signatures (6).

Finally, measuring peripheral biomarkers at only one time point and using a case-control design is not an optimal design. Much more useful information is available if the trajectories of the biomarker are considered; for example, it may help to discriminate trait and state and whether a potential change can be replicated over time. Repeated observations over time including trajectory information are of major interest and also when using biomarkers as monitors of treatment response. Unfortunately, most studies on peripheral molecular biomarkers are cross-sectional studies using one time point to compare patients with controls (3). However, studying biomarkers at several time points makes it possible to evaluate their potential as state and treatment monitors and their use in the prediction of treatment response.

PERSPECTIVES

Despite the above-described obstacles, overall there are, as described, promising peripheral biomarkers (3) that potentially will add to our future clinical tools and improve/impact clinical care. In particular, combining individual biomarkers across tissues and molecular systems seems to be a promising avenue for research in biomarker models (7, 8). However, instead of searching for the needles in the haystack, we need to collaborate and concentrate on describing all the elements in the haystack, integrating the inter-individual and intra-individual variability. At present, there is data and laboratory capacity and extended knowledge spread over multiple international research sites and industrial companies. However, creating well-structured multicenter studies is warranted, including mandatory structures for data sharing and integrated health information systems. Creating infrastructures and pipelines will move the field into the next stage of identifying pragmatically useful clinical biomarkers. This can be achieved by using innovative approaches and advanced technology (e.g. the R-

LiNK initiative aimed at optimizing response to lithium treatment through personalized evaluation of individuals with bipolar I disorder) (9).

Evaluating the potential use of biomarkers or clusters of biomarkers as clinical tools in prevention, to assist in developing personalized medicine and in treatment monitoring, with an overall goal to facilitate more and better treatments for patients with SMI, is clearly needed. Researchers are encouraged to initiate a strategic research agenda (e.g. across a diagnostic biobank consortium) that will require the use of precise biomarker protocols (e.g. in line with the CONSORT 2010 checklist of information to include when reporting randomized trials) (10). Before initiation, a checklist stating a range of criteria, including gold standards, in all aspects

using standardized operational procedures for all included studies, should be completed. This could also facilitate harmonization and joint databases, as in other areas of medicine (e.g. the Biomarker for Cardiovascular Risk Assessment across Europe consortium) (11). Overall, improving the opportunities for identifying the needles in the haystack.

AUTHOR CONTRIBUTIONS

The author confirms being the sole contributor of this work and has approved it for publication.

REFERENCES

- Comes AL, Papiol S, Mueller T, Geyer PE, Mann M, Schulze TG, et al. Proteomics for blood biomarker exploration of severe mental illness: Pitfalls of the past and potential for the future. *Transl Psychiatry* (2018) 8:160. doi: 10.1038/s41398-018-0219-2
- Insel TR. Translating scientific opportunity into public health impact: A strategic plan for research on mental illness. *Arch Gen Psychiatry* (2009) 66:128–33. doi: 10.1001/archgenpsychiatry.2008.540
- Pinto JV, Moulin TC, Amaral OB. On the transdiagnostic nature of peripheral biomarkers in major psychiatric disorders: A systematic review. *Neurosci Biobehav Rev* (2017) 83:97–108. doi: 10.1016/j.neubiorev.2017.10.001
- Munkholm K, Vinberg M, Kessing LV. Peripheral blood brain-derived neurotrophic factor in bipolar disorder: A comprehensive systematic review and meta-analysis. *Mol Psychiatry* (2016) 21:216–28. doi: 10.1038/mp.2015.54
- Humer E, Probst T, Pieh C. Metabolomics in psychiatric disorders: What we learn from animal models. *Metabolites* (2020) 10(2):72. doi: 10.3390/metabo10020072
- Ottesen NM, Meluken I, Frikke-Schmidt R, Plomgaard P, Scheike T, Fernandes BS, et al. Are remitted affective disorders and familial risk of affective disorders associated with metabolic syndrome, inflammation and oxidative stress? – a monozygotic twin study. *Psychol Med* (2019) 4:1–10. doi: 10.1017/S003329171900182X
- Munkholm K, Vinberg M, Pedersen BK, Poulsen HE, Ekstrøm EC, Kessing LV, et al. A multisystem composite biomarker as a preliminary diagnostic test in bipolar disorder. *Acta Psychiatr Scand* (2019) 139:227–36. doi: 10.1111/acps.12983
- Maes M, Carvalho AF. The compensatory immune-regulatory reflex system (CIRS) in depression and bipolar disorder. *Mol Neurobiol* (2018) 55:8885–903. doi: 10.1007/s12035-018-1016-x
- Scott J, Hidalgo-Mazzei D, Strawbridge R, Young A, Resche-Rigon M, Etain B, et al. Prospective cohort study of early biosignatures of response to lithium in bipolar-I-disorders: Overview of the H2020-funded R-LiNK initiative. *Int J Bipolar Disord* (2019) 7:20. doi: 10.1186/s40345-019-0156-x
- Schulz KF, Altman DG, Moher D the CONSORT group. CONSORT 2010 Statement: Updated guidelines for reporting parallel group randomised trials. *BMC Med* (2010) 8:18. doi: 10.1186/1741-7015-8-18
- Zeller T, Hughes M, Tuovinen T, et al: BiomarcCaRE: Rationale and design of the European BiomarcCaRE project including 300,000 participants from 13 European countries. *Eur J Epidemiol* (2014) 29:777–90. doi: 10.1007/s10654-014-9952-x

Conflict of Interest: MV has received consultancy fees from Lundbeck, Sunovion and Janssen/Cilag in the past three years.

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A Novel Network Pharmacology Strategy to Decode Metabolic Biomarkers and Targets Interactions for Depression

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Specialty section:

This article was submitted to
Mood and Anxiety Disorders,
a section of the journal
Frontiers in Psychiatry

Received: 03 December 2019

Accepted: 26 June 2020

Published: 15 July 2020

Citation:

Gao Y, Xu T, Zhao Y-X, Ling-Hu T,
Liu S-B, Tian J-S and Qin X-M (2020)
A Novel Network Pharmacology
Strategy to Decode Metabolic
Biomarkers and Targets Interactions
for Depression.
Front. Psychiatry 11:667.
doi: 10.3389/fpsy.2020.00667

Depression is one of the most prevalent and serious mental disorders with a worldwide significant health burden. Metabolic abnormalities and disorders in patients with depression have attracted great research attention. Thirty-six metabolic biomarkers of clinical plasma metabolomics were detected by platform technologies, including gas chromatography–mass spectrometry (GC–MS), liquid chromatography–mass spectrometry (LC–MS) and proton nuclear magnetic resonance (¹H-NMR), combined with multivariate data analysis techniques in previous work. The principal objective of this study was to provide valuable information for the pathogenesis of depression by comprehensive analysis of 36 metabolic biomarkers in the plasma of depressed patients. The relationship between biomarkers and enzymes were collected from the HMDB database. Then the metabolic biomarkers–enzymes interactions (MEI) network was performed and analyzed to identify hub metabolic biomarkers and enzymes. In addition, the docking score-weighted multiple pharmacology index (*DSWMP*) was used to assess the important pathways of hub metabolic biomarkers involved. Finally, validated these pathways by published literature. The results show that stearic acid, phytosphingosine, glycine, glutamine and phospholipids were important metabolic biomarkers. Hydrolase, transferase and acyltransferase involve the largest number of metabolic biomarkers. Nine metabolite targets (TP53, IL1B, TNF, PTEN, HLA-DRB1, MTOR, HRAS, INS and PIK3CA) of potential drug proteins for treating depression are widely involved in the nervous system, immune system and endocrine system. Seven important pathways, such as PI3K-Akt signaling pathway and mTOR signaling pathway, are closely related to the pathology mechanisms of depression. The application of important biomarkers and pathways in clinical practice may help to improve the diagnosis of depression and the evaluation of antidepressant effect, which provides important clues for the study of metabolic characteristics of depression.

Keywords: metabolic biomarkers, depression, network pharmacology, drug-target network, docking score-weighted multiple pharmacology index (*DSWMP*)

INTRODUCTION

Depression is one of the most prevalent and serious mental disorders. In recent years, the number of patients with depression has increased dramatically. In the world's population, the lifetime prevalence of depressed patients is about 17% with a significant burden of disease (1). Previous research reports have found that in the United States, more than 19 million adults suffer from depression in the USA and spend more than 30 billion annually, directly or indirectly (2). In addition, the incidence of depression is about 3 to 5% and currently 26 million Chinese people suffer from depression (3). Depression is a major cause of neuropsychiatric disability worldwide and the accurate diagnosis of depression before treatment is a hub change (4). Increased studies have reported the serotonin and norepinephrine dysfunction in the central nervous system of patients with depression (5). In addition, studies have also reported that hypothalamic pituitary adrenal (HPA) axis is one of the largest neuroendocrine findings of depression (6). Abnormal changes of inflammatory cytokines and endogenous metabolites are also involved in the molecular mechanism of pathology of depression (7, 8). The occurrence and development of depression is a complicated process, the etiology and pathogenesis mechanism of depression represent challenging issues in scientific and medical research.

A new psychological immune neuroendocrine (PINE) network model on depression has been proposed to provide an in-depth understanding of the pathogenesis of depression and the treatment of the disease with antidepressant drugs (1). The PINE network model is composed of three parts of the central nervous system, immune, and endocrine molecular networks, and the three networks are interconnected (9). The three molecular networks consist of many nodes and edges. The nodes in the network can be small molecules with different properties, such as genes, proteins or metabolites. If different nodes are related in biological function, it can be connected by edges. How to determine the key underlying molecular mechanism from PINE network that plays leading roles in the depression is a difficult problem due to the high complexity of the network composition and the incompletely understanding the complex multi-targets mechanism of depression.

Network pharmacology is constructed by integrating pharmacological data and network analysis methods to provide a comprehensive approach to explain the disease pathogenesis and drug treatment mechanism (10). The network pharmacology technology is considered to be one of the next frontiers of new drug research (11). Recently, network pharmacology has been

widely used in the pathogenesis of complex diseases and the mechanism of drug action. For example, Huang decoded the mechanism of traditional Chinese medicine in treating depression by analyzing drug target networks and disease target networks (12).

Increased studies have reported metabolic disorders or abnormal metabolic pathways in the plasma of depressed patients (13, 14). Thirty-six metabolic biomarkers of clinical plasma metabolomics were detected by platform technologies, including gas chromatography–mass spectrometry (GC–MS), liquid chromatography–mass spectrometry (LC–MS) and proton nuclear magnetic resonance ($^1\text{H-NMR}$), combined with multivariate data analysis techniques in previous our work (13, 14). These metabolic biomarkers are distributed in disordered metabolic pathways and maybe potential diagnostic biomarkers or therapeutic markers. Therefore, how to use the network pharmacology method to mine the existing markers is of great significance to the study of the metabolic mechanism of depression.

The main purposes of this study were to comprehensively analyze plasma metabolites by network pharmacology method and provide valuable information for the pathogenesis of depression. During this process, the relationship between biomarkers and enzymes were collected from the HMDB database and the metabolic biomarkers–enzymes interactions (MEI) network was performed and analyzed to identify hub metabolic biomarkers and enzymes. In addition, the interactions between each MB and each target of the nervous system, immune system and endocrine system was calculated by systemsDock, and then the docking score-weighted multiple pharmacology index (*DSWMP*) was used to assess the importance pathways of hub metabolic biomarkers involved. Finally, the important pathways were verified through published literature. The application of important biomarkers and pathways in clinical practice may help to improve the diagnosis of depression and the evaluation of antidepressant effect, which provides important clues for the study of metabolic characteristics of depression.

METHODS

Metabolites Data Collection and Processing

For the research object, only the metabolic biomarkers of clinical plasma metabolomics in our consideration, because the excavation of metabolites is more meaningful in the same clinical sample. Candidate metabolites refer to the statistical significance found in the original study. Unique metabolites were obtained by removing duplicates. We obtained biological function information and structural data of identified metabolic biomarkers from the Human Metabolome Database (HMDB) (15). The structures of these compounds were downloaded from PubChem (16). Enzymes related to the metabolic biomarkers were summarized from HMDB, while the functional categories of the relevant proteins were found from the UniProt database (17). The target proteins related to the nervous system (**Table S1**), immune system (**Table S2**) and endocrine system (**Table S3**) were retrieved from several Published online databases. The FDA-approved drugs for the nervous system, immune system

Abbreviations: $^1\text{H-NMR}$, Proton nuclear magnetic resonance; APT-2, Acyl-protein thioesterase 2; ATP, Adenosine triphosphate; CMS, Chronic mild stress; CNS, Central nervous system; CoA, Acetyl-coenzyme A; CPLA2, Cytosolic phospholipase A2; EPA, Eicosapentaenoic acid; DSWP, Docking score-weighted polypharmacological index; HMDB, Human Metabolome Database; HPA, Hypothalamic pituitary adrenal; HRAS, HRas Proto-Oncogene; LC–MS, Liquid chromatography–mass spectrometry; GC–MS, Gas chromatography–mass spectrometer; MEI, Metabolic Biomarker–enzyme interactions; PTEN, Phosphatase and tensin homolog; MTI, Metabolic Biomarker–target interactions; MTOR, Rapamycin Kinase; PIK3CA, Phosphatidylinositol 4, 5-bisphosphate 3-kinase catalytic subunit alpha isoform; PINE, Psycho-immune-neuroendocrine; TP53, Tumor Protein P53; TNF, Tumor necrosis factor.

and endocrine system with their targets were collected from DrugBank (18). The depression-related proteins were retrieved from Genecards (19), OMIM (20) and TTD (21). Finally, we carefully checked the target proteins in the literature. Integrate all target proteins and divide them into three categories, including the nervous system, immunity and endocrine.

Molecular Docking

Molecular docking was used to investigate the affinity between metabolic biomarkers and targets. The protein 3D structures were achieved from the RCSB protein data bank (<http://www.rcsb.org>) (22). We chose structures with more complete peptide chains, higher resolution, and better ligands as the selection criteria for the most appropriate protein structure.

SystemsDock (<http://systemsdock.unit.oist.jp/>) (23) was used for network pharmacology prediction and analysis, including four specific steps, selecting proteins by different parameters, defining binding sites by interactive molecular visualizer, preparing metabolic biomarkers for the test, and performing docking simulation and evaluating the result. The docking score calculated by systemsDock was used to assess the interactions between the metabolic biomarkers and the target proteins.

Mapping Metabolic Biomarkers–Target/Enzyme Network

Metabolic biomarker–enzyme relationships were collected from HMDB and visualized by using Cytoscape 3.4.0 (24). The network topological properties of nodes (metabolic biomarkers and enzymes) were calculated by the NetworkAnalyzer Cytoscape plugin (25). In an undirected network, the degree centrality of a node denotes the number of edges connected to this node is commonly used to measure the importance of a node in a single-layer network (26, 27). Degree centrality is a key indicator in analyzing the network, thereby reflecting the importance and influence of a metabolome biomarker or an enzyme in the metabolic biomarkers–enzyme interactions (MEI) network.

The threshold of the docking score was set to 5.52 (pK_d), which was equal to the dissociation constant (K_d) of 3 μ M. A docking score greater than the threshold was considered to be a very good correlation between the metabolic biomarker and the target protein (23). Based on previously reported works and experimental findings, a high accuracy level (80–83%) was found to evaluate the binding of a metabolomic biomarker and a protein, when the threshold score was in the range of 4.82 to 6.11 (pK_d). The metabolic biomarkers–target interactions (MTI) network was visualized by Cytoscape software.

Docking Score-Weighted Multiple Pharmacology Index

The target-related pathways were collected from Kyoto Encyclopedia of Genes and Genomes (KEGG) (28). One metabolic biomarker can be combined with many targets, and one target can be enriched to function in many different biological pathways. Therefore, a metabolic biomarker involves multiple different biological pathways. At present, there is no method to directly confirm that the metabolic biomarkers are

directly enriched in the pathway through the targets, so we have defined a docking score-weighted multiple pharmacology index (*DSWMP*):

$$DSWMP(P_k) = \frac{\sum_i^N \sum_j^M DS_{BiTj}}{Count(B_i)}, T_j \in TP_k \quad (1),$$

where DS_{BiTj} is the affinity between metabolic biomarker B_i and target T_j , and TP_k is a group of targets involved in pathway P_k . N and M are the numbers of metabolic biomarkers and targets, respectively.

Experimental Evidence of Important Pathways

To find experimental evidence for the predicted important pathways of depression by *DSWMP* calculation, the published literature will be identified by searching Pubmed, Embase, Cochrane Central Register of Controlled Trials and Web of Science. In the end, we chose the literature to verify the important pathways we found and provide references for the next step of research.

RESULTS AND DISCUSSION

In this report, a novel network pharmacology strategy was designed to detect the important pathway and elucidate the molecular mechanisms of depression (**Figure 1**). Firstly, the relationship between biomarkers and enzymes were collected from the HMDB database and the MEI network was performed and analyzed to identify hub metabolic biomarkers and enzymes. Secondly, the interactions between each MB and each target were calculated by systemsDock, and then the *DSWMP* was used to assess the important pathways of hub metabolic biomarkers involved. Thirdly, the important pathways verified through published literature.

Metabolic Biomarker–Enzyme Interactions (MEI) Network Analysis

The information of 36 metabolomic biomarkers and 350 enzymes are shown in **Table S4**. Most enzymes related to metabolic biomarkers belong to hydrolases, transferases and acyltransferases, which is consistent with our previous results. These enzymes mainly regulate metabolites such as energy metabolism, amino acid metabolism, gut microbe metabolism, glycerophospholipid metabolism and sphingolipid metabolism (13, 14). These enzymes play an important role in the regulation of metabolic pathways, which affect the molecular mechanism of depression. Overall, the results show that functional studies of enzymes related to metabolic biomarkers can reflect the molecular mechanism of depression, which also provides a reliable reference for the next network analysis.

The MEI network was used to characterize the relationship between enzymes and metabolic biomarkers, as shown in **Figure 2**. In MEI network, the node represents a metabolic biomarker or an enzyme, and the edges represent the interactions between them originated from HMDB. It turns out that the concentrations of some biomarkers in depression patients are up-regulated, while

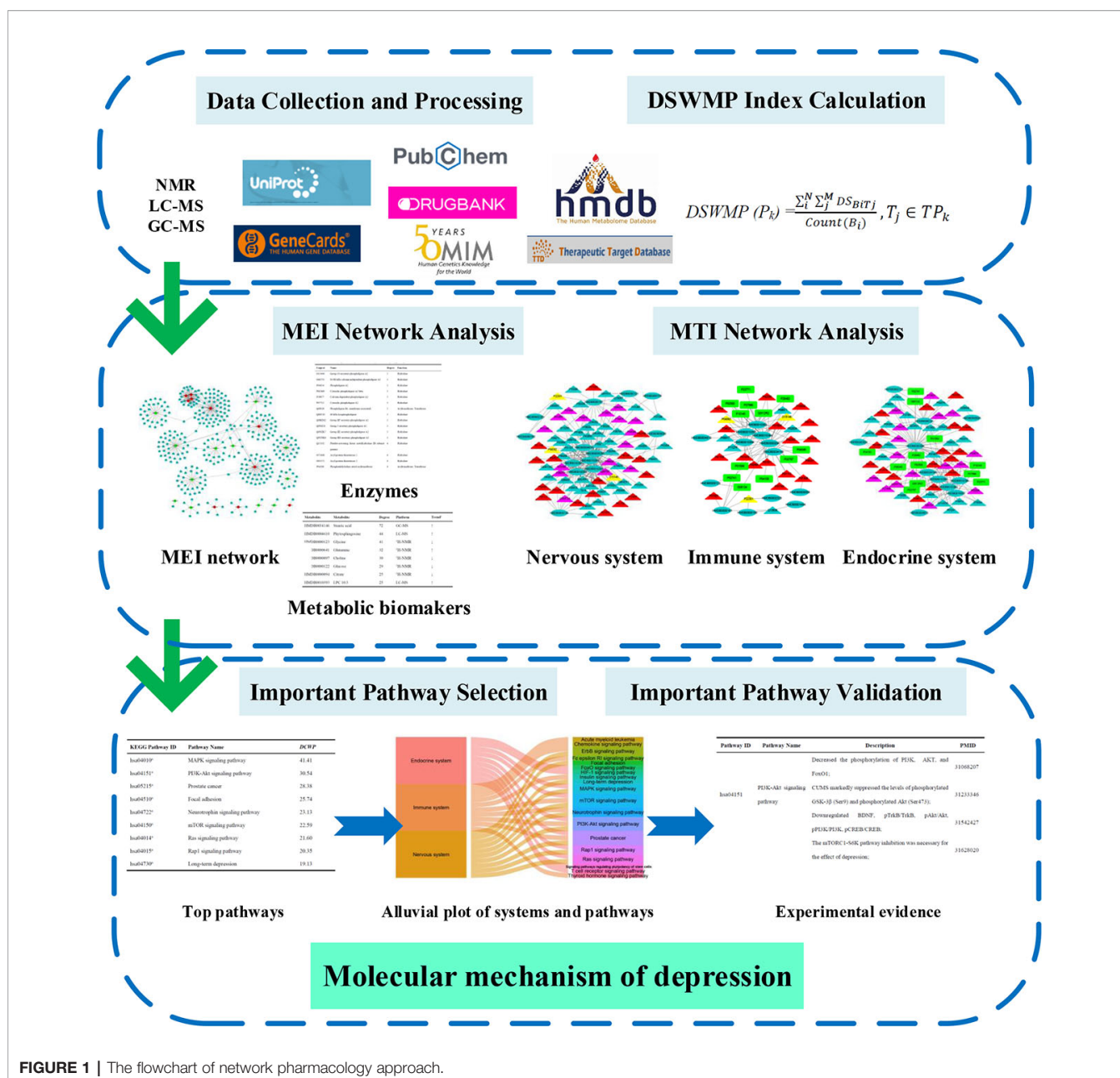


FIGURE 1 | The flowchart of network pharmacology approach.

others are down-regulated. Among these regulated biomarkers, their related enzymes form a particularly large cluster around them, suggesting that multiple enzymes regulate the same metabolite. **Tables 1 and 2** show the important enzymes and biomarkers identified by the network pharmacology approach, respectively. Among these data, previous literatures reported that cytosolic phospholipase A2 (cPLA2) is an important enzyme for PUFA metabolism and PGE 2 synthesis, which is the pivotal to the mode action of mood stabilizers in animal studies (29, 30). In addition, in peripheral blood cells of patients with depression, the mRNA expression of the gene encoding COX-2 increases sharply, which plays an important role in the pathogenesis of depression (31).

Among these metabolite-related enzymes, the phospholipase A2 family is related to many metabolites, so it is also important to study the function of depression. Previous research reports suggest that different phospholipase A2 types are associated with somatic symptoms of depression (32). It is believed that genetic variation of the phospholipase A2 gene increases the risk factor of depression induced by IFN-α (32). In addition, phospholipase A2 inhibitors have potential therapeutic effects in treating inflammation-related diseases, such as depression (33). Moreover, eicosapentaenoic acid (EPA) could up-regulate the expression of cytosolic phospholipase A2 gene and play an antidepressant effect in clinic, which shows the superiority of EPA antidepressant effect (34). Another research group also found that the same G allele of



TABLE 1 | Important enzymes identified by network topology analysis of MEI network.

| Uniprot | Name | Degree | Function |
|---------|---|--------|--|
| O15496 | Group 10 secretory phospholipase A2 | 5 | Hydrolase |
| O60733 | 85/88 kDa calcium-independent phospholipase A2 | 5 | Hydrolase |
| P04054 | Phospholipase A2 | 5 | Hydrolase |
| P0C869 | Cytosolic phospholipase A2 beta | 5 | Hydrolase |
| P39877 | Calcium-dependent phospholipase A2 | 5 | Hydrolase |
| P47712 | Cytosolic phospholipase A2 | 5 | Hydrolase |
| Q6P1J6 | Phospholipase B1, membrane-associated | 5 | Acyltransferase, Transferase |
| Q86U10 | 60 kDa lysophospholipase | 5 | Hydrolase |
| Q9BZM2 | Group IIF secretory phospholipase A2 | 5 | Hydrolase |
| Q9NZ20 | Group 3 secretory phospholipase A2 | 5 | Hydrolase |
| Q9NZK7 | Group IIE secretory phospholipase A2 | 5 | Hydrolase |
| Q9UNK4 | Group IID secretory phospholipase A2 | 5 | Hydrolase |
| Q15102 | Platelet-activating factor acetylhydrolase IB subunit gamma | 4 | Hydrolase |
| O75608 | Acyl-protein thioesterase 1 | 4 | Hydrolase |
| O95372 | Acyl-protein thioesterase 2 | 4 | Hydrolase |
| P04180 | Phosphatidylcholine-sterol acyltransferase | 4 | Acyltransferase, Transferase |
| P68402 | Platelet-activating factor acetylhydrolase IB subunit beta | 4 | Hydrolase |
| Q13093 | Platelet-activating factor acetylhydrolase | 4 | Hydrolase |
| Q6P1A2 | Lysophospholipid acyltransferase 5 | 4 | Acyltransferase, Transferase |
| Q7L5N7 | Lysophosphatidylcholine acyltransferase 2 | 4 | Acyltransferase, Transferase |
| Q8NCC3 | Group XV phospholipase A2 | 4 | Acyltransferase, Hydrolase, Transferase |
| Q8NF37 | Lysophosphatidylcholine acyltransferase 1 | 4 | Acyltransferase, Transferase |
| Q99487 | Platelet-activating factor acetylhydrolase 2, cytoplasmic | 4 | Hydrolase |
| Q14032 | Bile acid-CoA:amino acid N-acyltransferase | 2 | Acyltransferase, Hydrolase, Serine esterase, Transferase |
| P06276 | Cholinesterase | 2 | Hydrolase, Serine esterase |
| Q13510 | Acid ceramidase | 2 | Hydrolase |
| Q16773 | Kynurenine-oxoglutarate transaminase 1 | 2 | Aminotransferase, Lyase, Transferase |
| Q5QUJ3 | Alkaline ceramidase 2 | 2 | Hydrolase |
| Q8TDN7 | Alkaline ceramidase 1 | 2 | Hydrolase |
| Q969I3 | Glycine N-acyltransferase-like protein 1 | 2 | Acyltransferase, Transferase |
| Q9HCG7 | Non-lysosomal glucosylceramidase | 2 | Glycosidase, Hydrolase |

TABLE 2 | Important metabolic biomarkers identified by network topology analysis of MEI network.

| Metabolite | Metabolite | Degree | Platform | Trend ^a |
|-------------|--------------------------|--------|--------------------------|--------------------|
| HMDB0034146 | Stearic acid | 72 | GC-MS | ↑ |
| HMDB0004610 | Phytosphingosine | 44 | LC-MS | ↑ |
| HMDB0000123 | Glycine | 41 | ¹ H-NMR | ↓ |
| HMDB0000641 | Glutamine | 32 | ¹ H-NMR | ↑ |
| HMDB0000097 | Choline | 30 | ¹ H-NMR | ↓ |
| HMDB0000122 | Glucose | 29 | ¹ H-NMR | ↓ |
| HMDB0000094 | Citrate | 25 | ¹ H-NMR | ↓ |
| HMDB0010393 | LPC 10:3 | 25 | LC-MS | ↑ |
| HMDB0010383 | LPC 16:1 | 25 | LC-MS | ↑ |
| HMDB0010396 | LPC 21:4 | 25 | LC-MS | ↑ |
| HMDB0010384 | LPC 18:0 | 25 | LC-MS | ↑ |
| HMDB0000161 | Alanine | 21 | ¹ H-NMR | ↓ |
| HMDB0000159 | Phenylalanine | 19 | ¹ H-NMR | ↓ |
| HMDB0000294 | Urea | 12 | GC-MS | ↓ |
| HMDB0000883 | Valine | 7 | ¹ H-NMR/GC-MS | ↓ |
| HMDB0003681 | 4-Acetamidobutanoic acid | 6 | LC-MS | ↓ |
| HMDB0000190 | Lactate | 5 | ¹ H-NMR | ↑ |
| HMDB0000925 | TMAO | 5 | ¹ H-NMR | ↑ |
| HMDB0002329 | Oxalic acid | 2 | GC-MS | ↑ |
| HMDB0000099 | L-Cystathionine | 2 | LC-MS | ↓ |
| HMDB0001494 | Acetylphosphate | 2 | LC-MS | ↓ |
| HMDB0060348 | 4-oxohex-2-enedioic acid | 1 | LC-MS | ↓ |

^aThe fold change was calculated as the ratio of the concentration of metabolite of the depression to that of healthy control.

the PLA2 BanI polymorphism is one of the risk factors for depression in Korean populations (35). The expression levels of acyl protein thioesterase 2 (APT-2) and oleamide are increased in chronic mild stress (CMS). This provides a reference for the treatment of depression by targeting these proteins (36). At present, there is no literature on the relationship between acetylhydrolase and depression, which may be a novel research point.

Fourteen metabolic biomarkers are associated with more than ten enzymes in MEI network (Table 2). The concentrations of most MB were down-regulated, while stearic acid, phytosphingosine, glutamine and phospholipids were up-regulated. It is well known that fatty acids are an important source of human body production and storage. Acetyl-coenzyme A (CoA) produced by oxidation of β -fatty acids can produce adenosine triphosphate (ATP) in the TCA cycle and could be converted to ketone bodies for storage in the kidney and liver (37). There are reports showing that the concentration of stearic acid in plasma of patients with depression is significantly increased, which may lead to blockage of fatty acid transport and inhibition of TCA cycle, which is consistent with the previous research results (38, 39). Moreover, the AUCs for oxalic acid and stearic acid were >0.7 , indicating a great clinical diagnostic value. Therefore, stearic acid may be a diagnostic indicator of depression. These results indicate that the metabolic biomarkers in MEI network are involved in the molecular mechanism of depression.

In addition, most metabolic biomarkers belong to long-chain fatty acids and phytosphingosine, which indicates that the lipid

metabolism disorder plays a crucial role in the mechanism of depression. These metabolic biomarkers are involved in glycerophospholipid metabolism and sphingolipid metabolism. Among them, sphingosine involves various biological processes, including cell-cell interactions, cell proliferation, differentiation and apoptosis. Studies have reported that elevated levels of hemolytic phosphatidylcholine also increase oxidative stress, which is a key factor in the onset of depression (40). The results indicate that the long-chain fatty acids and phytosphingosine in MEI network is involved in the pathological mechanism of depression.

Glutamine and its related enzymes form an independent part of the MEI network, suggesting that glutamine may play an unusual role in the pathogenesis mechanism of depression. Glutamate is associated with the neurobiology of depression and can cause neurotoxicity if over-released (40). In addition, glutamine and glutamate can be converted between neurons and astrocytes, which is necessary for the steady state of the glutamine-glutamic acid cycle (41). Therefore, the increase in glutamine in the plasma of depression patients may be a compensatory adaptation to glutamate-induced neurotoxicity. The results show that the glutamine in the MEI network is involved in the pathological mechanism of depression.

Metabolic Biomarker-Target Interactions (MTI) Network Analysis

Tables S1–S3 show the nervous system, immune system and endocrine system target proteins, and the nervous system,

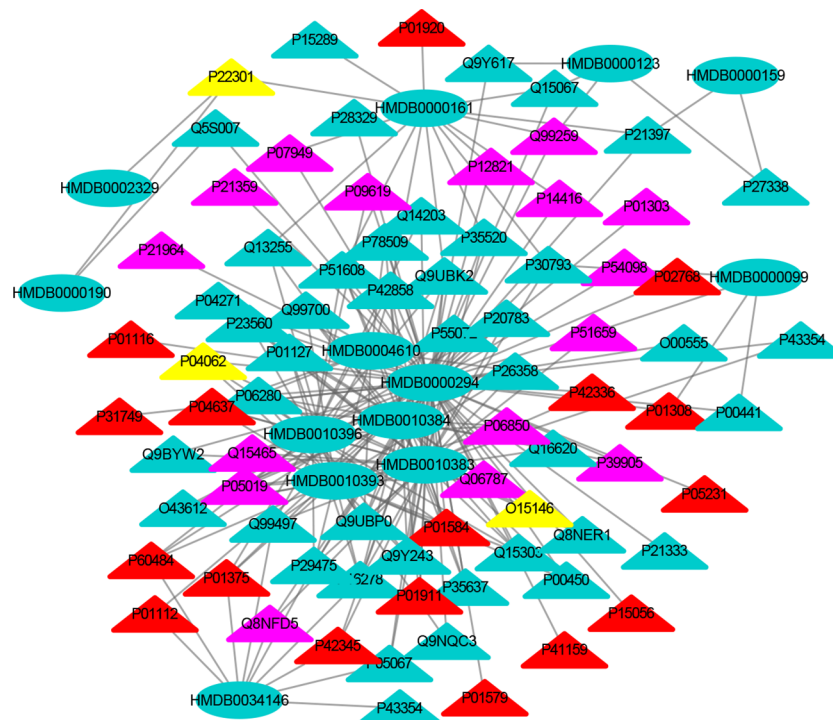


FIGURE 3 | The MTI network of the nervous system. The ellipses refer to metabolic biomarkers, and triangles refer to targets, respectively. The color of the nodes indicates that the three systems connected to depression are different, including nervous system, immune system and endocrine system. The colors of the nodes are blue, yellow, and red, indicating that the target is connected to one system, two systems, and three systems, respectively.

immune system and endocrine system target numbers are 155, 60 and 125, respectively. The interactions between each metabolic biomarker and the target protein were performed by molecular docking. The MTI network of nervous system, immune system and endocrine system were visualized by Cytoscape, respectively (**Figures 3–5**). In MTI network, the node represents a metabolic biomarker or a target, and the edges represent the interactions between them from the docking score meeting the set threshold. The metabolic biomarker links the target to a highly interconnected cluster. In order to analyze the correlation between metabolic biomarkers and targets and to mine important molecules, we calculated the topological parameter of the network, namely degree centrality.

The MTI network of the nervous system (**Figure 3**) contains 13 metabolic biomarkers and 78 targets, resulting in 232 edges associations between them. The average number of targets for each metabolic biomarker is 17.85 and the average number of metabolic biomarkers for each target is 2.97. Similarly, the MTI network of the immune system (**Figure 4**) contains 13 metabolic biomarkers and 37 targets, resulting in 107 edges associations between them. The average number of targets for each metabolic

biomarker is 8.23 and the average number of metabolic biomarkers for each target is 2.89. Moreover, the MTI network of the endocrine system (**Figure 5**) contains 11 metabolic biomarkers and 67 targets, resulting in 78 edges associations between them. The average number of targets for each metabolic biomarker is 15.91 and the average number of metabolic biomarkers for each target is 2.61. This indicates that the molecular mechanism of depression in the nervous system, immune system and endocrine system has the characteristics of multiple metabolic biomarkers and multiple targets.

In these three MTI networks, the three metabolic biomarkers of urea, LPC 16:1 and LPC 18:0 have a higher degree centrality in the network. The number of targets that the three metabolic biomarkers could bind to in the depression-related nervous system, immune system and endocrine system are 68, 33, 61; 40, 18, 25; 29, 13, 20, respectively. The role of these metabolic biomarkers in the pathogenesis of depression is worthy of our in-depth study, because these metabolic biomarkers play a crucial role in the three networks. Among these metabolic biomarkers, urea, LPC 16:1 and LPC 18:0 can be classified as organic carbonic acids or glycerophospholipids, and further research is needed on their role in the pathogenesis of depression.

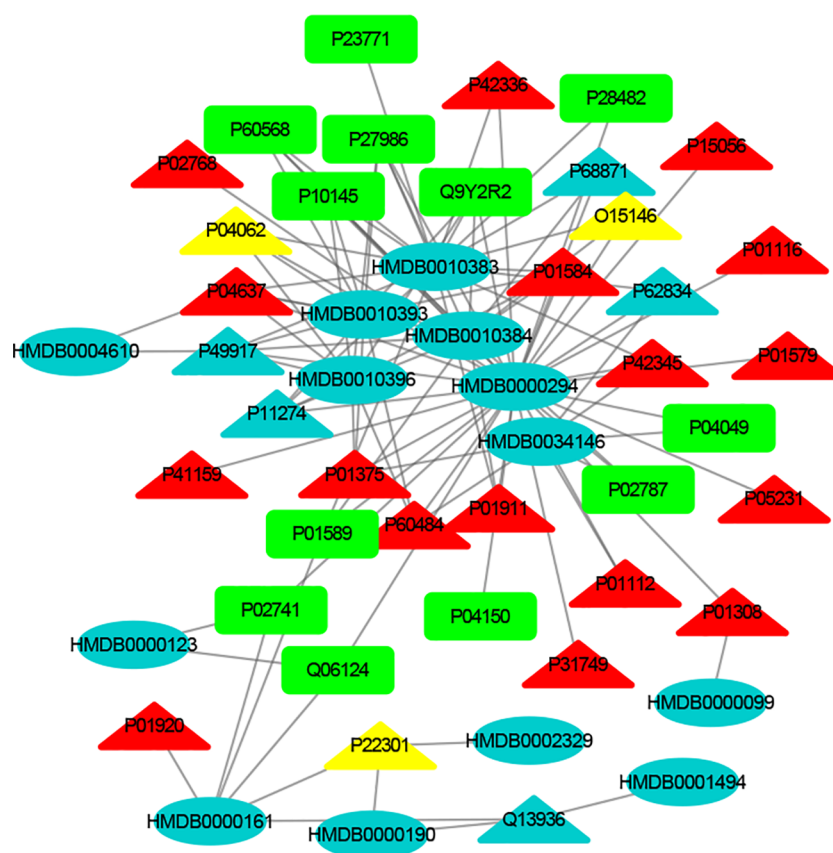


FIGURE 4 | The MTI network of the immune system. The ellipses refer to metabolic biomarkers, and triangles refer to targets, respectively. The color of the nodes indicates that the three systems connected to depression are different, including nervous system, immune system and endocrine system. The colors of the nodes are blue, yellow, and red, indicating that the target is connected to one system, two systems, and three systems, respectively.

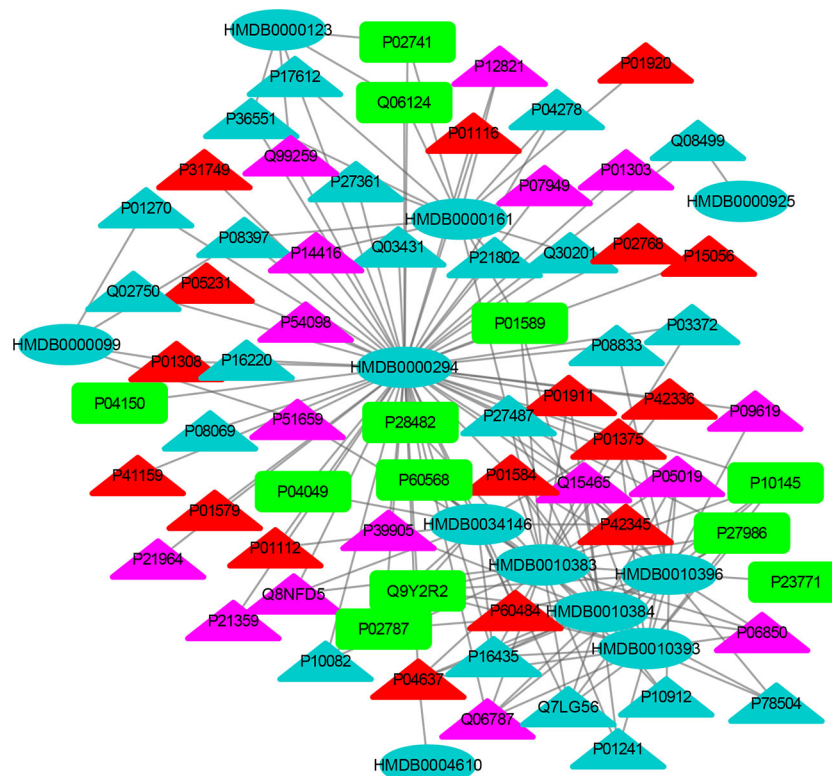


FIGURE 5 | The MTI network of the endocrine system. The ellipses refer to metabolic biomarkers, and triangles refer to targets, respectively. The color of the nodes indicates that the three systems connected to depression are different, including nervous system, immune system and endocrine system. The colors of the nodes are blue, yellow, and red, indicating that the target is connected to one system, two systems, and three systems, respectively.

There are nine identical targets in these three systems, which simultaneously play an important role in the three MTI networks involved in the pathogenesis of depression. Among these targets related to metabolic biomarkers, P04637 (Tumor Protein P53), P01584 (Interleukin 1 Beta), P01375 (Tumor Necrosis Factor), P60484 (Phosphatase And Tensin Homolog), P01911 (Major Histocompatibility Complex, Class II, DR Beta 1), P42345 (Mechanistic Target of Rapamycin Kinase), P01112 (HRas Proto-Oncogene, GTPase), P01308 (Insulin), P42336 (Phosphatidylinositol-4,5-Bisphosphate 3-Kinase Catalytic Subunit Alpha), were widely involved in the nervous system, immune system and endocrine system.

The minor allele 72C of the tumor protein P53 (TP53) gene plays a protective role in the occurrence of depression. It participates in the pathological mechanism of depression through the cell survival and death regulation (42). Previous studies have reported that proinflammatory cytokines such as IL-1 beta (IL1B) (43) and tumor necrosis factor (TNF) (44) have been implicated in the pathogenesis molecular mechanism of depression. The evidence provided by Liu suggests that the rs701848, rs2735343 and rs112025902 polymorphisms in the phosphatase and tensin homologous gene (PTEN) genes may be related to the risk of depression in Chinese (45). Major Histocompatibility Complex (HLA-DRB1) plays a significant

role in the immune system by presenting peptides derived from extracellular proteins (46). HRas Proto-Oncogene (HRAS) (46) can encode some genes in signal transduction pathways. These proteins can be linked to GTP and GDP, and they have their own GTPase activity. In addition, rapamycin kinase (mTOR) is a serine/threonine kinase that regulates cell proliferation (47). Li research found that the activation of mTOR in the prefrontal cortex of rats was one of the important mechanisms by which ketamine exerts antidepressant effects (48). Some related studies have also found that acute ketamine administration will activate mTOR in the peripheral blood of patients with depression (49). The antioxidant alpha lipoic acid has been shown to increase insulin (INS) sensitivity and has been used to treat diabetic patients. INS also plays an important role in the pathogenesis of depression. Therefore, the nutrient alpha lipoic acid should be clinically tested as an adjunct treatment for depression. Therefore, some scholars suggested that the nutrient alpha lipoic acid should be used as an adjunct treatment for depression (50). Phosphatidylinositol 4, 5-bisphosphate 3-kinase catalytic subunit alpha isoform (PIK3CA) inhibitors used in bipolar disease and depression (51). Consequently, these results demonstrated that the crucial roles of nine identical targets in the treatment of depression and further confirmed that drug works in a multi-targets manner to treat depression.

TABLE 3 | Top ten pathways for the nervous system, immune system and endocrine system.

| KEGG Pathway ID | Pathway Name | DCWP |
|-----------------------|--|-------|
| hsa04010 ^a | MAPK signaling pathway | 41.41 |
| hsa04151 ^a | PI3K-Akt signaling pathway | 30.54 |
| hsa05215 ^a | Prostate cancer | 28.38 |
| hsa04510 ^a | Focal adhesion | 25.74 |
| hsa04722 ^a | Neurotrophin signaling pathway | 23.13 |
| hsa04150 ^a | mTOR signaling pathway | 22.59 |
| hsa04014 ^a | Ras signaling pathway | 21.60 |
| hsa04015 ^a | Rap1 signaling pathway | 20.35 |
| hsa04730 ^a | Long-term depression | 19.13 |
| hsa04012 ^a | ErbB signaling pathway | 18.50 |
| hsa04151 ^b | PI3K-Akt signaling pathway | 25.21 |
| hsa04062 ^b | Chemokine signaling pathway | 23.30 |
| hsa05215 ^b | Prostate cancer | 22.58 |
| hsa04150 ^b | mTOR signaling pathway | 22.29 |
| hsa04919 ^b | Thyroid hormone signaling pathway | 19.97 |
| hsa05221 ^b | Acute myeloid leukemia | 18.45 |
| hsa04012 ^b | ErbB signaling pathway | 18.45 |
| hsa04664 ^b | Fc epsilon RI signaling pathway | 18.41 |
| hsa04722 ^b | Neurotrophin signaling pathway | 18.10 |
| hsa04910 ^b | Insulin signaling pathway | 17.49 |
| hsa04151 ^c | PI3K-Akt signaling pathway | 37.33 |
| hsa04150 ^c | mTOR signaling pathway | 25.73 |
| hsa04068 ^c | FoxO signaling pathway | 25.71 |
| hsa05215 ^c | Prostate cancer | 25.11 |
| hsa04660 ^c | T cell receptor signaling pathway | 25.07 |
| hsa04066 ^c | HIF-1 signaling pathway | 23.17 |
| hsa04015 ^c | Rap1 signaling pathway | 23.16 |
| hsa04014 ^c | Ras signaling pathway | 23.14 |
| hsa04010 ^c | MAPK signaling pathway | 21.95 |
| hsa04550 ^c | Signaling pathways regulating pluripotency of stem cells | 20.65 |

^a, ^b, and ^c: the pathway was related to target of nervous system, immune system and endocrine system, respectively.

Important Pathways Selection and Validation

In order to improve the therapeutic effect while reducing side effects, drug treatment of diseases usually go through a variety of target methods (52). These targets involve multiple different pathways of disease pathology. The target-pathway approach is a disease-specific research module, which has been widely used in disease pathology exploration and drug development. To find the important pathological mechanism of depression, the strategy of *DSWMP* index was applied and calculated to select crucial pathway, which provides a methodological reference for the research and development of disease. The *DSWMP* index is an indicator to evaluate the importance of the pathway. The size of the *DSWMP* index is determined by the number of metabolic biomarkers binding targets and the binding energy between them. In other words, the *DSWMP* index method can be used to evaluate the importance of metabolic biomarkers in pathways.

There are 105, 113 and 125 metabolic biomarkers-related targets involved in the nervous system pathways immune system and endocrine system of depression, respectively. In each function, the top 10 pathways of *DSWMP* index are shown in **Table 3** and **Figure 6**. Among these pathways, the pathways of hsa04151 (PI3K-Akt signaling pathway) and hsa04150 (mTOR signaling pathway) are top-ranked in the pathological mechanism of depression among the nervous system, immune system and endocrine system, indicating that these two pathways play an important role in the pathogenesis of depression. The hsa04151 has been reported involved in the inhibition of apoptosis, cell proliferation and expression of inflammatory cytokines (53, 54). The hsa04150 pathway connects intracellular and extracellular signaling communication, and plays an important role in the protein

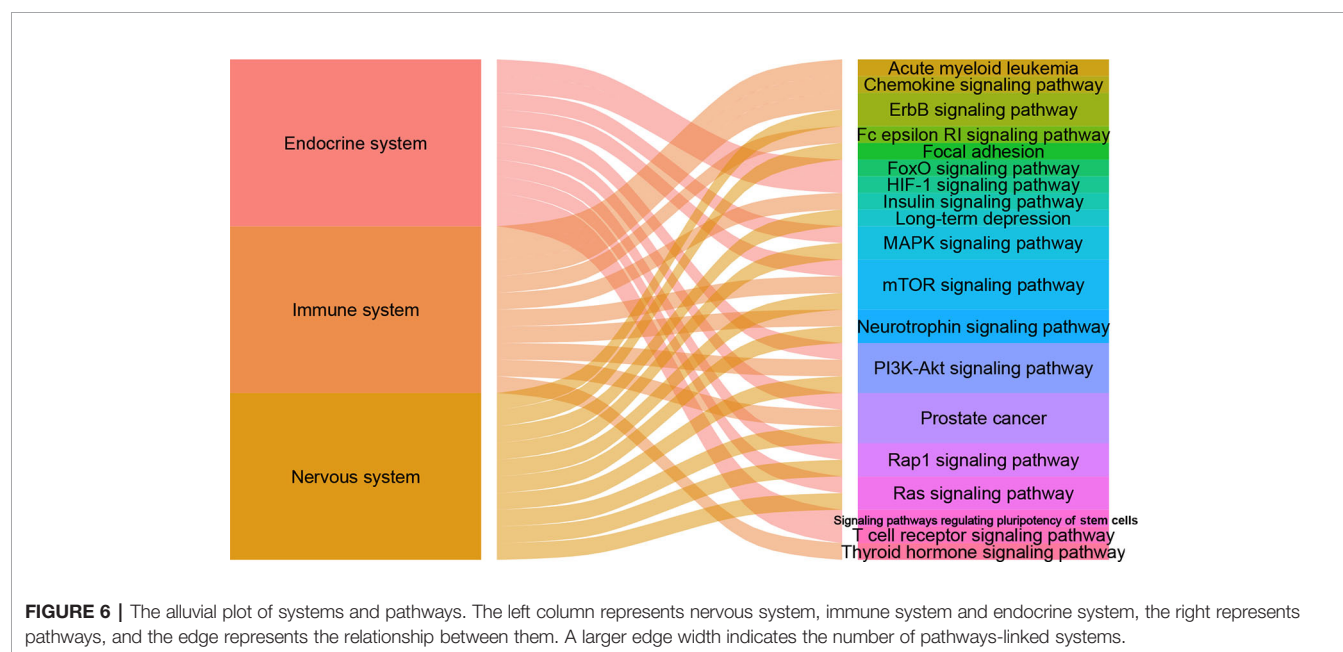


TABLE 4 | Experimental evidence for the seven important signaling pathways with most significant therapeutic relationships of depression.

| Pathway ID | Pathway Name | Description | PMID |
|------------|--------------------------------|---|----------------------------------|
| hsa04151 | PI3K-Akt signaling pathway | Decreased the phosphorylation of PI3K, AKT, and FoxO1; CUMS markedly suppressed the levels of phosphorylated GSK-3 β (Ser9) and phosphorylated Akt (Ser473); Downregulated BDNF, pTrkB/TrkB, pAkt/Akt, pPI3K/PI3K, pCREB/CREB; | 31068207 31233346 31542427 |
| hsa04150 | mTOR signaling pathway | The mTORC1-S6K pathway inhibition was necessary for the effect of depression; Reduction in the levels of mature BDNF and mTOR (Ser2448) phosphorylation in the hippocampus; UCMS-induced reductions of p70S6K and post-synaptic density 95 (PSD-95) mRNA levels, and of phospho-mTOR and phospho-4EBP1 in the prefrontal cortex, hippocampus, hypothalamus, and olfactory bulb; | 31628020 31078612 27374162 |
| hsa04010 | MAPK signaling pathway | Downregulated expression of the mitogen activated protein kinase (MAPK) phosphatase 1 (MKP-1) and the downregulated phosphorylation of extracellular signal-regulated kinase (pERK) in the anterior cingulate cortex (ACC) of mice; CTRP3 may be an innovative therapeutic target for treating patients with depression through regulating p38 and JNK signaling; | 32109506 31629950 |
| hsa04012 | ErbB signaling pathway | The MAPK/ERK signaling pathway has been shown to be involved in the pathogenesis of MDD and the rapid onset of action of antidepressant therapies; The stressed rats showed elevated expression of NRG1 and phosphorylated ErbB4 (pErbB4) in the myocardium, whereas ErbB2 and pErbB2 were inhibited; | 30859414 27133902 |
| hsa04722 | Neurotrophin signaling pathway | The stressed rats displayed elevated expression of NRG1 and phosphorylated ErbB4 (pErbB4) in the prefrontal cortex, whereas ErbB2 and pErbB2 were inhibited; Decreased neurotrophic factors expression and neurotrophin receptors signaling have repeatedly been reported in association with stress, depression, and neurodegenerative disorders; | 26626816 28315978 |
| hsa04015 | Rap1 signaling pathway | Reduced synaptic markers in hippocampus, demonstrated by reductions in β III-tubulin (neuronal marker), PSD-95, SNAP-25, and neurotrophin-3; p75 neurotrophin receptor/nerve growth factor signaling and innate immune toll-like receptor signaling in MDD; Rap1-MKK3/6-p38 MAPK pathway in the induction of mGluR-dependent long term depression (LTD) by directly coupling to receptor trafficking machineries to facilitate the loss of synaptic AMPA receptors; | 31889537 28451885 14709549 |
| hsa04014 | Ras signaling pathway | The small G-protein Rap and the transcription factor STAT-3 are also involved since reducing the levels of Rap1 (using small interfering RNA) or STAT-3 (using dominant negative STAT3) significantly blocks 5-HT1A-receptor-mediated neurite outgrowth; Blood mononuclear cell proteome suggests integrin and Ras signaling as critical pathways for antidepressant treatment response; | 15925428 24607422 |
| | | Ras-GRF proteins contribute to forms of synaptic plasticity that are required specifically for mature hippocampal function; RAS-GRF1 mediates NMDA-type glutamate receptor (NMDAR)-induction of long term depression in the CA1 region of the hippocampus of mice | 16467520 23766509 |

synthesis process of new synaptic connections (55). Recent research supports the hypothesis that major depression may be the result of disruption of mTOR-dependent translational regulation (13, 14). This result indicates that this pathway plays a crucial role in the molecular mechanism of depression. In addition, there are another five pathways, including hsa04010 (MAPK signaling pathway), hsa04012 (ErbB signaling pathway), hsa04722 (Neurotrophin signaling pathway), hsa04015 (Rap1 signaling pathway) and hsa04014 (Ras signaling pathway), were involved in two of the nervous system, immune system and endocrine system related to depression. Finally, the experimental evidence for the seven important signaling pathways with the most significant therapeutic relationships of depression is shown in **Table 4**. These pathways could have closely related to the pathological process of depression and need to research in depth.

It is undeniable that there are several limitations in this study. First, it is not sufficient to study based on the data currently available, because the technique of identifying metabolites for depression is still a continuous improvement process. Second, although we have used published literature to verify some results, validation of molecular mechanisms based on clinical samples are necessary to analysis the complex pathogenesis of depression. Our results provide good ideas and methods for studying the pathogenesis of depression.

CONCLUSION

We identified 36 metabolic biomarkers of clinical plasma metabolomics using NMR and MS. The relationship between biomarkers and enzymes were collected from the HMDB database. The results show that stearic acid, phytosphingosine, glycine, glutamine and phospholipids were important metabolic biomarkers. Hydrolase, transferase and acyltransferase involve the largest number of metabolic biomarkers. The important metabolites and enzymes screened by the topology of the network may play a key role in the underlying molecular mechanism of depression.

The nervous system, immune system and endocrine system are mainly involved in the underlying pathological mechanism of depression. The *DSWMP* index was used to assess the importance pathways of hub metabolic biomarkers involved. Nine proteins (TP53, IL1B, TNF, PTEN, HLA-DRB1, MTOR, HRAS, INS and PIK3CA) are widely involved in the nervous system, immune system and endocrine system. These targets may be important targets for antidepressants in the treatment of depression. Seven important pathways, such as PI3K-Akt signaling pathway and mTOR signaling pathway, are closely related to the pathogenesis molecular mechanisms of depression and require further investigation. A combination of network

pharmacology strategy and metabolomics approach has great potentials in comprehensively and deeply understanding the molecular mechanism of depression. The application of important biomarkers and pathways in clinical practice may help to improve the diagnosis of depression and the evaluation of antidepressant effect, which provides important clues for the study of metabolic characteristics of depression.

DATA AVAILABILITY STATEMENT

All datasets generated for this study are included in the article/**Supplementary Material**.

AUTHOR CONTRIBUTIONS

X-MQ and J-ST provided the concept and designed the study. YG, TX, Y-XZ, TL-H and S-BL conducted the analyses and wrote

the manuscript. YG, TX, Y-XZ, TL-H and S-BL participated in data analysis. X-MQ and J-ST contributed to revise the manuscript. All authors contributed to the article and approved the submitted version.

FUNDING

This work was supported by the National S&T Major Projects for “Major New Drugs Innovation and Development” (2017ZX09301047), the Science and Technology of Shanxi Province (No. 201701D121137 and No. 201903D321210).

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fpsy.2020.00667/full#supplementary-material>

REFERENCES

1. Stapelberg NJC, Pratt R, Neumann DL, Shum DHK, Brandis S, Muthukkumarasamy V, et al. From feedback loop transitions to biomarkers in the psycho-immune-neuroendocrine network: Detecting the critical transition from health to major depression. *Neurosci Biobehav Rev* (2018) 90:1–15. doi: 10.1016/j.neubiorev.2018.03.005
2. Baglioni C, Battagliese G, Feige B, Spiegelhalter K, Nissen C, Voderholzer U, et al. Insomnia as a predictor of depression: a meta-analytic evaluation of longitudinal epidemiological studies. *J Affect Disord* (2011) 135(1–3):10–9. doi: 10.1016/j.jad.2011.01.011
3. Peng GJ, Tian JS, Gao XX, Zhou YZ, Qin XM. Research on the Pathological Mechanism and Drug Treatment Mechanism of Depression. *Curr Neuropsychopharmacol* (2015) 13(4):514–23. doi: 10.2174/1570159x1304150831120428
4. Lopez AD, Mathers CD. Measuring the global burden of disease and epidemiological transitions: 2002–2030. *Ann Trop Med Parasitol* (2006) 100(5–6):481–99. doi: 10.1179/136485906x97417
5. Nemeroff CB. Recent advances in the neurobiology of depression. *Psychopharmacol Bull* (2002) 36 Suppl 2:6–23. doi: 10.1016/S0074-7742(06)73005-7
6. Heim C, Newport DJ, Mletzko T, Miller AH, Nemeroff CB. The link between childhood trauma and depression: insights from HPA axis studies in humans. *Psychoneuroendocrinology* (2008) 33(6):693–710. doi: 10.1016/j.psyneuen.2008.03.008
7. Gao XX, Cui J, Zheng XY, Li ZY, Choi YH, Zhou YZ, et al. An investigation of the antidepressant action of xiaoyaosan in rats using ultra performance liquid chromatography-mass spectrometry combined with metabolomics. *Phytother Res : PTR* (2013) 27(7):1074–85. doi: 10.1002/ptr.4805
8. Köhler O, Benros ME, Krogh J. Anti-inflammatory Intervention in Depression—Reply. *JAMA Psychiatry* (2015) 72(5):512–3. doi: 10.1001/jamapsychiatry.2014.3186
9. Stapelberg NJC, Neumann DL, Shum D, Headrick JP. Health, pre-disease and critical transition to disease in the psycho-immune-neuroendocrine network: Are there distinct states in the progression from health to major depressive disorder? *Physiol Behav* (2019) 198:108–19. doi: 10.1016/j.physbeh.2018.10.014
10. Hopkins AL. Network pharmacology. *Nat Biotechnol* (2007) 25(10):1110–1. doi: 10.1038/nbt1007-1110
11. Hopkins AL. Network pharmacology: the next paradigm in drug discovery. *Nat Chem Biol* (2008) 4(11):682–90. doi: 10.1038/nchembio.118
12. Huang C, Zheng C, Li Y, Wang Y, Lu A, Yang L. Systems pharmacology in drug discovery and therapeutic insight for herbal medicines. *Briefings Bioinf* (2014) 15(5):710–33. doi: 10.1093/bib/bbt035
13. Liu CC, Wu YF, Feng GM, Gao XX, Zhou YZ, Hou WJ, et al. Plasma-metabolite-biomarkers for the therapeutic response in depressed patients by the traditional Chinese medicine formula Xiaoyaosan: A (1)H NMR-based metabolomics approach. *J Affect Disord* (2015) 185:156–63. doi: 10.1016/j.jad.2015.05.005
14. Liu X, Liu C, Tian J, Gao X, Li K, Du G, et al. Plasma metabolomics of depressed patients and treatment with Xiaoyaosan based on mass spectrometry technique. *J Ethnopharmacol* (2020) 246:112219. doi: 10.1016/j.jep.2019.112219
15. Wishart DS, Jewison T, Guo AC, Wilson M, Knox C, Liu Y, et al. HMDB 3.0—The Human Metabolome Database in 2013. *Nucleic Acids Res* (2013) 41(Database issue):D801–7. doi: 10.1093/nar/gks1065
16. Kim S, Thiessen PA, Bolton EE, Chen J, Fu G, Gindulyte A, et al. PubChem Substance and Compound databases. *Nucleic Acids Res* (2016) 44(D1):D1202–13. doi: 10.1093/nar/gkv951
17. Apweiler R, Bairoch A, Wu CH, Barker WC, Boeckmann B, Ferro S, et al. UniProt: the Universal Protein knowledgebase. *Nucleic Acids Res* (2004) 32(Database issue):D115–9. doi: 10.1093/nar/gkh131
18. Wishart DS, Feunang YD, Guo AC, Lo EJ, Marcu A, Grant JR, et al. DrugBank 5.0: a major update to the DrugBank database for 2018. *Nucleic Acids Res* (2018) 46(D1):D1074–d82. doi: 10.1093/nar/gkx1037
19. Safran M, Dalah I, Alexander J, Rosen N, Iny Stein T, Shmoish M, et al. GeneCards Version 3: the human gene integrator. *Database : J Biol Database Curation* (2010) 2010:baq020. doi: 10.1093/database/baq020
20. Amberger JS, Bocchini CA, Schiettecatte F, Scott AF, Hamosh A. OMIM.org: Online Mendelian Inheritance in Man (OMIM®), an online catalog of human genes and genetic disorders. *Nucleic Acids Res* (2015) 43(Database issue):D789–98. doi: 10.1093/nar/gku1205
21. Zhu F, Shi Z, Qin C, Tao L, Liu X, Xu F, et al. Therapeutic target database update 2012: a resource for facilitating target-oriented drug discovery. *Nucleic Acids Res* (2012) 40(Database issue):D1128–36. doi: 10.1093/nar/gkr797
22. Berman HM, Kleywegt GJ, Nakamura H, Markley JL. The Protein Data Bank archive as an open data resource. *J computer-aided Mol Design* (2014) 28(10):1009–14. doi: 10.1007/s10822-014-9770-y
23. Hsin KY, Matsuoka Y, Asai Y, Kamiyoshi K, Watanabe T, Kawaoka Y, et al. systemsDock: a web server for network pharmacology-based prediction and analysis. *Nucleic Acids Res* (2016) 44(W1):W507–13. doi: 10.1093/nar/gkw335

24. Shannon P, Markiel A, Ozier O, Baliga NS, Wang JT, Ramage D, et al. Cytoscape: a software environment for integrated models of biomolecular interaction networks. *Genome Res* (2003) 13(11):2498–504. doi: 10.1101/gr.1239303
25. Assenov Y, Ramírez F, Schelhorn SE, Lengauer T, Albrecht M. Computing topological parameters of biological networks. *Bioinf (Oxford England)* (2008) 24(2):282–4. doi: 10.1093/bioinformatics/btm554
26. Janga SC, Tzakos A. Structure and organization of drug-target networks: insights from genomic approaches for drug discovery. *Mol Biosyst* (2009) 5(12):1536–48. doi: 10.1039/B908147j
27. Gu J, Luo F, Chen L, Yuan G, Xu X. A systematic study of chemogenomics of carbohydrates. *Mol Biosyst* (2014) 10(3):391–7. doi: 10.1039/c3mb70534j
28. Kanehisa M, Goto S, Sato Y, Furumichi M, Tanabe M. KEGG for integration and interpretation of large-scale molecular data sets. *Nucleic Acids Res* (2012) 40(Database issue):D109–14. doi: 10.1093/nar/gkr988
29. Bosetti F, Rintala J, Seemann R, Rosenberger TA, Contreras MA, Rapoport SI, et al. Chronic lithium downregulates cyclooxygenase-2 activity and prostaglandin E(2) concentration in rat brain. *Mol Psychiatry* (2002) 7(8):845–50. doi: 10.1038/sj.mp.4001111
30. Rao JS, Ertley RN, DeMar JCR, Rapoport SI, Bazinet RP, Lee HJ. Dietary n-3 PUFA deprivation alters expression of enzymes of the arachidonic and docosahexaenoic acid cascades in rat frontal cortex. *Mol Psychiatry* (2007) 12(2):151–7. doi: 10.1038/sj.mp.4001887
31. Galecki P, Galecka E, Maes M, Chamielec M, Orzechowska A, Bobińska K, et al. The expression of genes encoding for COX-2, MPO, iNOS, and sPLA2-IIA in patients with recurrent depressive disorder. *J Affect Disord* (2012) 138(3):360–6. doi: 10.1016/j.jad.2012.01.016
32. Su KP, Huang SY, Peng CY, Lai HC, Huang CL, Chen YC, et al. Phospholipase A2 and cyclooxygenase 2 genes influence the risk of interferon-alpha-induced depression by regulating polyunsaturated fatty acids levels. *Biol Psychiatry* (2010) 67(6):550–7. doi: 10.1016/j.biopsych.2009.11.005
33. Magrioti V, Kokotos G. Phospholipase A2 inhibitors as potential therapeutic agents for the treatment of inflammatory diseases. *Expert Opin Ther patents* (2010) 20(1):1–18. doi: 10.1517/13543770903463905
34. Su KP, Yang HT, Chang JP, Shih YH, Guu TW, Kumaran SS, et al. Eicosapentaenoic and docosahexaenoic acids have different effects on peripheral phospholipase A2 gene expressions in acute depressed patients. *Prog Neuropsychopharmacol Biol Psychiatry* (2018) 80(Pt C):227–33. doi: 10.1016/j.pnpbp.2017.06.020
35. Pae CU, Yu HS, Kim JJ, Lee CU, Lee SJ, Lee KU, et al. BanI polymorphism of the cytosolic phospholipase A2 gene and mood disorders in the Korean population. *Neuropsychobiology* (2004) 49(4):185–8. doi: 10.1159/000077364
36. Ge L, Zhu MM, Yang JY, Wang F, Zhang R, Zhang JH, et al. Differential proteomic analysis of the anti-depressive effects of oleamide in a rat chronic mild stress model of depression. *Pharmacol Biochem Behav* (2015) 131:77–86. doi: 10.1016/j.pbb.2015.01.017
37. Lin L, Huang Z, Gao Y, Chen Y, Hang W, Xing J, et al. LC-MS-based serum metabolic profiling for genitourinary cancer classification and cancer type-specific biomarker discovery. *Proteomics* (2012) 12(14):2238–46. doi: 10.1002/pmic.201200016
38. Gao X, Zheng X, Li Z, Zhou Y, Sun H, Zhang L, et al. Metabonomic study on chronic unpredictable mild stress and intervention effects of Xiaoyaosan in rats using gas chromatography coupled with mass spectrometry. *J Ethnopharmacol* (2011) 137(1):690–9. doi: 10.1016/j.jep.2011.06.024
39. Tian JS, Peng GJ, Gao XX, Zhou YZ, Xing J, Qin XM, et al. Dynamic analysis of the endogenous metabolites in depressed patients treated with TCM formula Xiaoyaosan using urinary (1)H NMR-based metabolomics. *J Ethnopharmacol* (2014) 158(Pt A):1–10. doi: 10.1016/j.jep.2014.10.005
40. Nunes SO, Vargas HO, Prado E, Barbosa DS, de Melo LP, Moylan S, et al. The shared role of oxidative stress and inflammation in major depressive disorder and nicotine dependence. *Neurosci Biobehav Rev* (2013) 37(8):1336–45. doi: 10.1016/j.neubiorev.2013.04.014
41. Schousboe A, Scafidi S, Bak LK, Waagepetersen HS, McKenna MC. Glutamate metabolism in the brain focusing on astrocytes. *Adv Neurobiol* (2014) 11:13–30. doi: 10.1007/978-3-319-08894-5_2
42. Mahmood S, Evinová A, Škereňová M, Ondrejka I, Lehotský J. Association of EGF, IGFBP-3 and TP53 Gene Polymorphisms with Major Depressive Disorder in Slovak Population. *Cent Eur J Public Health* (2016) 24(3):223–30. doi: 10.21101/cejph.a4301
43. Ovakainen Y, Koponen H, Jokelainen J, Keinänen-Kiukaanniemi S, Kumpusalo E, Vanhala M. Depressive symptomatology is associated with decreased interleukin-1 beta and increased interleukin-1 receptor antagonist levels in males. *Psychiatry Res* (2009) 167(1–2):73–9. doi: 10.1016/j.psychres.2007.12.004
44. Mak A, Tang CS, Ho RC. Serum tumour necrosis factor-alpha is associated with poor health-related quality of life and depressive symptoms in patients with systemic lupus erythematosus. *Lupus* (2013) 22(3):254–61. doi: 10.1177/0961203312471872
45. Liu LJ, Zhu C, Tian HJ, Zheng TS, Ye MJ, Li H. Correlations of PTEN genetic polymorphisms with the risk of depression and depressive symptoms in a Chinese population. *Gene* (2016) 595(1):77–82. doi: 10.1016/j.gene.2016.09.034
46. Ortega-Hernandez OD, Cuccia M, Bozzini S, Bassi N, Moscavitich S, Diaz-Gallo LM, et al. Autoantibodies, polymorphisms in the serotonin pathway, and human leukocyte antigen class II alleles in chronic fatigue syndrome: are they associated with age at onset and specific symptoms? *Ann New Y Acad Sci* (2009) 1173:589–99. doi: 10.1111/j.1749-6632.2009.04802.x
47. Abelaira HM, Réus GZ, Neotti MV, Quevedo J. The role of mTOR in depression and antidepressant responses. *Life Sci* (2014) 101(1–2):10–4. doi: 10.1016/j.lfs.2014.02.014
48. Li N, Lee B, Liu RJ, Banasr M, Dwyer JM, Iwata M, et al. mTOR-dependent synapse formation underlies the rapid antidepressant effects of NMDA antagonists. *Sci (New Y NY)* (2010) 329(5994):959–64. doi: 10.1126/science.1190287
49. Yu JJ, Zhang Y, Wang Y, Wen ZY, Liu XH, Qin J, et al. Inhibition of calcineurin in the prefrontal cortex induced depressive-like behavior through mTOR signaling pathway. *Psychopharmacology* (2013) 225(2):361–72. doi: 10.1007/s00213-012-2823-9
50. Salazar MR. Alpha lipoic acid: a novel treatment for depression. *Med Hypotheses* (2000) 55(6):510–2. doi: 10.1054/mehy.2000.1103
51. Pilot-Storck F, Chopin E, Rual JF, Baudot A, Dobrokhoto P, Robinson-Rechavi M, et al. Interactome mapping of the phosphatidylinositol 3-kinase-mammalian target of rapamycin pathway identifies deformed epidermal autoregulatory factor-1 as a new glycogen synthase kinase-3 interactor. *Mol Cell Proteomics : MCP* (2010) 9(7):1578–93. doi: 10.1074/mcp.M900568-MCP200
52. Zhang W, Bai Y, Wang Y, Xiao W. Polypharmacology in Drug Discovery: A Review from Systems Pharmacology Perspective. *Curr Pharm Design* (2016) 22(21):3171–81. doi: 10.2174/1381612822666160224142812
53. Shi HS, Zhu WL, Liu JF, Luo YX, Si JJ, Wang SJ, et al. PI3K/Akt signaling pathway in the basolateral amygdala mediates the rapid antidepressant-like effects of trefoil factor 3. *Neuropsychopharmacology* (2012) 37(12):2671–83. doi: 10.1038/npp.2012.131
54. Cunha MP, Budni J, Ludka FK, Pazini FL, Rosa JM, Oliveira Á, et al. Involvement of PI3K/Akt Signaling Pathway and Its Downstream Intracellular Targets in the Antidepressant-Like Effect of Creatine. *Mol Neurobiol* (2016) 53(5):2954–68. doi: 10.1007/s12035-015-9192-4
55. Laplante M, Sabatini DM. mTOR signaling at a glance. *J Cell Sci* (2009) 122(Pt 20):3589–94. doi: 10.1242/jcs.051011

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Neuropsychophysiological Measures of Alcohol Dependence: Can We Use EEG in the Clinical Assessment?

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OPEN ACCESS

Edited by:

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Specialty section:

This article was submitted to
Molecular Psychiatry,
a section of the journal
Frontiers in Psychiatry

Received: 27 December 2019

Accepted: 29 June 2020

Published: 17 July 2020

Citation:

Jurado-Barba R, Sion A,
Martínez-Maldonado A,
Domínguez-Centeno I,
Prieto-Montalvo J, Navarrete F,
García-Gutierrez MS,
Manzanares J and Rubio G (2020)
Neuropsychophysiological Measures
of Alcohol Dependence: Can We Use
EEG in the Clinical Assessment?
Front. Psychiatry 11:676.
doi: 10.3389/fpsy.2020.00676

Addiction management is complex, and it requires a bio-psycho-social perspective, that ought to consider the multiple etiological and developmental factors. Because of this, a large amount of resources has been allocated to assess the vulnerability to dependence, i.e., to identify the processes underlying the transition from substance use to dependence, as well as its course, in order to determine the key points in its prevention, treatment, and recovery. Consequently, knowledge from neuroscience must be taken into account, which is why different initiatives have emerged with this objective, such as the “Research Domain Criteria” (RDoC), and the “Addiction Neuroclinical Assessment” (ANA). Particularly, neuropsychophysiological measures could be used as markers of cognitive and behavioral attributes or traits in alcohol dependence, and even trace clinical change. In this way, the aim of this narrative review is to provide an overview following ANA clinical framework, to the most robust findings in neuropsychophysiological changes in alcohol dependence, that underlie the main cognitive domains implicated in addiction: incentive salience, negative emotionality, and executive functioning. The most consistent results have been found in event-related potential (ERP) analysis, especially in the P3 component, that could show a wide clinical utility, mainly for the executive functions. The review also shows the usefulness of other components, implicated in affective and substance-related processing (P1, N1, or the late positive potential LPP), as well as event-related oscillations, such as theta power, with a possible use as vulnerability or clinical marker in alcohol dependence. Finally, new tools emerging from psychophysiology research, based on functional connectivity or brain graph analysis could help toward a better understanding of altered circuits in alcohol dependence, as well as communication efficiency and effort during mental operations. This review concludes with an examination of these tools as possible markers in the clinical field and discusses methodological differences, the need for more replicability studies and incipient lines of work. It also uses consistent findings in

psychophysiology to draw possible treatment targets and cognitive profiles in alcohol dependence.

Keywords: alcohol dependence, electroencephalogram, endophenotypes, incentive salience, negative emotionality, executive dimension, event-related potential

INTRODUCTION

Addictions continue to be a public health problem, despite the continued efforts by different types of professionals. That is why great efforts are being made from a preventive and therapeutic point of view, although it seems that these efforts are not being effective. Both preventive and therapeutic strategies include information from a psychosocial approach, and more recently from the neurosciences, not only from the field of psychopharmacology but also genetics and neuroimaging (1).

However, the results of these procedures, although highly evidenced and confirmed, are quite heterogeneous, possibly in relation to the clinical and methodological characteristics of the studies available in this field (1). For instance, different types of patients, type of consumption, withdrawal periods, and of course different type of interventions. In fact, treatment does not bring with certainty the abstinence maintenance and a great number of patients go through relapse in the first follow-up year (2–4).

To cope with this complexity, there is a consensus that in order to improve the outcomes of behavioral interventions in alcohol use disorders (AUD) it is necessary to understand the mechanisms underlying the behavioral change in effective treatments. From this standpoint, building a strong foundation for alcohol dependence treatment includes answering the question of why and how, not just whether, a treatment is effective (5).

Research on the mechanisms of effective AUD treatments that underlie behavior change has been focusing on cognitive neuroscience. Dysfunctional processes that maintain AUD, such as craving, withdrawal, lapse, and relapse are understood by studying the functioning of cognitive processes, such as attention or motivation, and the underlying neural systems.

The Research Domain Criteria (RDoC) is a framework that allows the study of psychiatric disorders from this point of view since its objective is the analysis of what may cause the symptoms, rather than the symptoms themselves. It even allows the search and validation of biomarkers (6). It takes into consideration five dimensions for all possible pathologies: negative valence systems, positive valence systems, cognitive systems, social systems, and activation supervising systems (7). Therefore, patients could be evaluated in each one of these dimensions at different levels of analysis, from basic disciplines involving genetics, molecules, cells or neuroanatomical circuits, including physiology to clinical or behavioral measures, such as neuropsychological assessment, self-informed measures or behavioral paradigms (8, 9).

This approach makes it possible to characterize the psychological attributes by being analyzed in cognitive, social, emotional, and behavioral terms, with measures coming from basic science and clinical area (10).

Following this framework, the Addiction Neuroclinical Assessment (ANA) considers the same levels of analysis, selecting three domains as essential; these are the incentive salience, negative emotionality, and executive abilities (8). Therefore, each level of analysis within the dimensions proposed by ANA could be considered as a follow-up measure of the course of substance dependence, perhaps understanding it as an intermediate endophenotype. The disorder is understood as an active process, not an endpoint, and each of the measures obtained at different levels of analysis would be considered a marker of this process (7, 11). In this way, in this study we summarize the relationship between the pathological consumption of alcohol and the use of EEG as a measure of cognitive components assessment, according to ANA dimensions.

Besides the changes produced by moderate alcohol consumption in the brain electrical activity (12–14), in alcohol dependence EEG measures have been employed as markers of structural and functional changes that rise as a consequence of continuous and pathological consumption. Hence, there is considerable bibliography available with respect to these changes (15–17). By using the brain electrical activity, we can obtain different types of information, useful for understanding alcohol dependence processes; the most frequent one in the scientific literature is the one provided by event related potentials, in the sensorial, cognitive or motor modality. They are represented by the average electrical activity appearing after each event, and are composed by different components, which are named according to their polarity (positive or negative) and the moment when they appear (in milliseconds). For instance, the P300 component would reflect a positive deflection approximately 300 ms after stimulus presentation (16). A different type of analysis is the one brought by brain oscillations, their pattern would reflect the sum of postsynaptic potentials generated by a neuronal field close to an electrode. In this way, the number of oscillations (measured in Hz) determines the traditionally known frequency bands (e.g., delta, theta, alpha, beta, and gamma bands). All of them have been explored in alcohol dependent patients, however, the most frequent findings point to beta rhythms (12–28 Hz) alterations, observed even during the abstinence period and their offspring (15). According to these basic concepts different conditions of EEG recordings arise (e.g., resting-state or event-related activity), as well as different methods of analysis, which are addressed later in this work, such as the synchrony in brain rhythms between different locations.

The measures obtained by means of neuropsychophysiological assessment are an example of possible biomarkers that reflect, in an indirect but objective way, the cognitive processes that give rise to the problematic behavior of alcohol consumption. Several

benefits could be drawn from these type of measures: 1) It allows the reflection of the related brain activity, with a great temporal, “instantaneous and continuous” precision, in words of Campanella (18); 2) When the employed paradigm is well-characterized from the cognitive and methodological points of view, the neural pattern can reflect the psychological features that contribute to the disorder, even when no observation was made from the psychopathological point of view (18, 19), 3) In addition, it is a measure that escapes the patient’s voluntary control, for example, it reflects the lack of inhibitory control or craving, even when the patient is not aware of it (20).

The suggestion that neuropsychophysiological measures, namely, event-related potentials (ERP) could be biomarkers, e.g., the P300 component, or other intermediate endophenotypes, such as N170 or N200, is a well-established line of work, from the research standpoint. Nonetheless, these are not consistently used in clinical evaluation. The aim of this revision is to provide an overview, through a narrative revision of the literature, of the most solid results in the different neuropsychophysiological measures that can be obtained by using electroencephalographic (EEG) measures.

DIMENSIONS OF ADDICTION NEUROCLINICAL ASSESSMENT AND NEUROPSYCHOPHYSIOLOGICAL EVALUATION

Incentive Salience

Incentive salience is one of the main objectives of evaluation inside ANA frameworks. As defined by early theories of addiction and I-RISA model (21), incentive salience can be understood as the narrowing of the attentional focus on the substance and the related contextual cues, that gain in motivational value and appetitive properties, in detriment of natural reinforcers, changing the entire reward system. A sensitization to the substance takes place, based on associative and conditioned reinforcement mechanisms, where the mere presence of related cues can drive substance-seeking behavior (22, 23), by changing activation and craving states (24, 25). In alcohol dependence, alterations in the affective information processing and preferential attention toward substance cues have been described (26, 27) as well as attentional bias and interference control effects (28–30). Moreover, cue exposure is related to a higher desire and urgency consumption and increased expectancies regarding alcohol effects, as well as dependence severity (26, 27). Changes in the reward system are also observed, resulting in difficulties regarding the delay of gratification and poor loss and gain evaluation (20, 31–34). Thereby, the incentive salience construct, together with cognitive and motivational processes involved seem essential in the search for clinical markers of change through the dependence process and as vulnerability factors involved in maladaptive behaviors. These can be measured through a great variety of tools, including self-informed measures, neuropsychological

tests, and behavioral paradigms, sometimes combined with neuroimaging and psychophysiological techniques. ANA model proposes specific materials for evaluating the incentive salience domain, including behavioral paradigms such as the dot-probe attentional bias task (29, 35, 36), cue-reactivity task (37) or the monetary incentive delay task (38).

Nonetheless, initiatives like ANA or RDoC encourage the use of neuroimaging tools, like psychophysiological measures. Taking into consideration that the motivational and attentional allocation changes toward substance-related cues are produced in parallel with structural and functional changes of reward and salience circuitry of the brain, the neural underpinnings of these processes become relevant in the discovery for new markers of clinical changes and vulnerability related to the dependence process. In this way, neuropsychophysiological measures enable the assessment of early stimuli evaluation (perceptive and preattentional operations), together with distractibility and the attentional biased produced by substance-related stimuli, as well as mental operations during reward processing and delay of gratification.

Early visual components, such as P1, N1, or P2, as well as the late positive potential (LPP) are ERP components usually related to early stimulus evaluation and attentional processes that can be found altered in AUD in cue reactivity tasks or attentional bias paradigms. One of the most frequent ways to evaluate the influence of alcohol cues on neuropsychophysiological activity is by using cue reactivity tasks that usually consist in the presentation of visual substance-related stimuli (images or words), and even olfactory or gustatory cues. These can be passively visualized (e.g., a bottle of whiskey or neutral content stimuli, such as a book or a pen) or can be part of a classification or a discrimination task while the electrical brain activity is being recorded.

There is evidence in the literature for an early preferential processing of substance-related cues in AUDs, shown by greater N1 amplitudes (39), an exogenous and automatic attention index, indicating a preferential attentional processing of alcohol cues, in line with motivated attention (40) and theories of addiction (41). P3 to alcohol cues in an oddball task is also found increased in alcohol dependence, indicating the alteration of higher-order processing such as controlled attentional allocation (42). This preferential processing of alcohol cues also appear in individuals at risk, ERP changes being observed in social drinkers (43) with heavy use (44), that show increased P1 latencies and LPP amplitudes toward alcohol images, and in individuals with a low sensitivity to alcohol effects, that have increased P3 amplitudes (45, 46). In this way, since early stages of alcohol consumption and in individuals at risk (low alcohol sensitivity), substance-related cues seem to be processed faster and with a high motivational salience, and ERP monitoring could help predict substance-related problems.

Alcohol salience can even dampen other demanding processes in course, heavy drinkers (47) and recently detoxified individuals (48) showing higher N2 amplitudes toward substance cues during inhibitory processes (during NoGo conditions). Thus, alcohol salience can be so powerful that other important

executive processes could be affected, such as inhibitory control, essential for adaptive behaviors.

Regarding the attentional bias produced by substance-related cues, a common task is the dot-probe paradigm, that consists in the displaying of pairs of images (substance-related and neutral ones) appearing side by side, usually for 500 ms, prior to the presentation of a probe (a dot or an asterisk) replacing one of the pictures (on the left or right side of the screen). The subject must respond by indicating as quickly as possible, on which side the point has appeared, pressing one key for the right side and another for the left side. The attentional bias is typically calculated using the differences in reaction times when the dot is presented on the same side where alcohol-related pictures were (congruent trials) and when it is on the opposite side (incongruent trials). ERP analysis, in this case, would help to elucidate what happens with brain activity at several stages of the cue processing, that is, sensorial filtering, attention orientation and re-orientation, stimuli salience and arousal, and also more controlled attentional processes, even in absence of behavioral changes in reaction times or self-report measures. In the same way as cue tasks, visual probe paradigms display in individuals at risk (low alcohol sensitivity) a preferential early attention orientation, indicated by larger P1 amplitudes, and difficulties reorienting attention away from alcohol cues, reflected by larger negativity between 220 and 280 ms (49).

With respect to reward system changes, monetary incentive or choice tasks are a usual tool of evaluation of motivational decision making. This type of task usually consists in gambling tasks where participants take low or high risks in order to gain a prize (generally money). The subject must decide between gambling alternatives with higher rewards and losses (higher risk) or with lower rewards and losses (lower risk). These tasks allow us to evaluate not only risk-taking and the capacity of delaying gratification, but also the evaluation of the own decisions and responses through the task. Alcohol-dependent individuals (50) and youth at risk with families densely affected by alcoholism (51) seem to show lower amplitudes of the P3 component and lower activity in frontal areas (e.g., cingulate gyrus), during both gain and loss conditions of gambling tasks. There even seems to be a relation between risk-taking features and impulsivity (50). In this way, ERP analysis of risk and outcome evaluation could help in the discovery of vulnerability markers. Even recent alcohol intoxication seems to affect neuropsychophysiological activity during reward processing, reducing ERP positivity in 250 to 400 time-window (52), indicating an affectation of performance monitoring and feedback that drives the decision-making process during the task. This is also exposed in binge drinkers, that show alterations in automatic error processing (larger error-related negativity [ERN]) in a Go-NoGo task and in motivational processing (delayed error positivity P_e) in a risk task (53). This would indicate an affectation of controlled processes such as monitoring self-behavior since early stages of consumption.

Taking into consideration the importance of the incentive salience and its effects across the dependence course, its evaluation through ERP measures can bring more light upon

the specific processes that underlie to appetitive changes, attentional interference produced by alcohol cues and craving, that motivationally drive the decision making. Moreover, changes in psychophysiological activity appear early in the course of continuous consumption, indicated by the early attentional and motivational capture by alcohol stimuli and by changes in the reward system in acute effects of alcohol administration and social drinkers. Some studies even find these changes in population at risk, namely individuals with low sensitivity to alcohol and individuals with families densely affected by alcohol dependence. Hence, early informational processing components, such as P1 or N2 or those involved in controlled attention, N2 and P3 could be assessed as possible vulnerability markers or as markers of cognitive efficacy change through time.

Negative Emotionality

Within the ANAs framework, negative emotionality refers to the propensity to experience and react with negative emotions, such as sadness, anxiety, fear, and anger to environmental cues (54). In fact, there is enough evidence that shows how dependent individuals have clear difficulties within the emotional regulation process, namely a diminished emotional awareness and a reduced acknowledgment of other people's emotions and their own, giving rise to an inadequate emotional adjusting in relation to environmental demands (55, 56). Furthermore, tolerating negative emotional states might become difficult and even bring people to act impulsively, making them to engage in behaviors that they find rewarding in the short-term without fully considering its risks, in order to diminish the experience of this negative affect. These negative reactions to environmental cues seem to be highly related to alcohol consumption. So much that different theoretical models consider that negative emotionality is present during different stages of the alcohol consumption cycle.

Among them, classical theoretical approaches (57) propose that deregulation in the reward system is characterized by the transition of the experience of pleasant sensations every time an individual consumes alcohol, to a progressive shift toward feelings of relief when he takes the substance, that is, taking alcohol with the whole purpose of avoiding the negative emotionality states experience during withdrawal stages (58). This leads to further consumption as a way to avoid this negative state, reinforcing the cycle. Additionally, neuroadaptations seem to persist even after prolonged abstinence, increasing the risk of relapse (59).

There is also broad evidence of the existence of negative emotionality prior to the development of AUD. This seems to play a key role in the engagement of problematic consumption as a self-regulation mechanism. Hagan et al. (60) found that a higher negative emotionality in children predicts future alcohol consumption, stress, and internalizing symptoms during adulthood. Additionally, there is evidence that the increase of alcohol consumption in adolescents is highly related to a reduction of the positive affect and an increased negative affect, giving rise to the usual anhedonia symptoms in this type of

patients (61). Implying then, that negative emotionality can also be considered an intermediate endophenotype of alcoholism.

This negative emotionality is reflected at a biological level, hence psychophysiological changes can be assessed with EEG measurements. The use of EEG techniques in affective process evaluation can be challenging, due to its complexity. However, the correct implementation of EEG measures ensures a direct measurement of the affective processing, providing key temporal information of all stages of emotional processing, allowing then a better understanding of some of the alterations found in AUD patients. However, the number of studies of emotional alterations in AUDs with EEG measures is scarce.

ERP analysis is one of the most used EEG measures to study emotional processing with psychophysiological techniques. Among the ERPs related to emotional processing, we can mention the Early Posterior Negativity (EPN), P1, P2 and the LPP (62). The modulation of these components by affective information is reliable and systematically observed (63–65). LPP is characterized by a positive centro-parietal deflection, starting around 200 ms after the stimulus onset, and it is prolonged several milliseconds in time. It is modulated by the emotional content of stimuli, showing an increase in comparison with its activity under neutral stimuli. However, cognitive reappraisal related to positive emotional regulation seems to reduce LPP amplitude (66–68). In this type of tasks, subjects passively visualize images with high emotional content, and they are asked to classify them according to three dimensions: valence, arousal, and dominance. The emotional content of these images is usually related to positive appetitive stimuli (e.g., sex-related), negative threatening ones (e.g., aggressions) and sometimes motivationally relevant stimuli related to the substance (e.g., a beer).

Studies with paradigms of viewing of neutral and affective images found that alcohol consumption selectively reduces the processing of negative cues, specifically there is a decreased amplitude of LPP during the viewing of negative images, and this has been seen in healthy population after the intake of a small doses of alcohol (69) and in population at risk, such as binge drinkers (70). A reduced LPP amplitude would be indicating an early effect of alcohol consumption on the impact of negative-valence content in later information processing stages. This could be in line with theories regarding negative affect evaluation and processing alterations in alcohol dependence.

Prolonged alcohol consumption is also related to alterations in the recognition of emotional facial expressions. In emotion recognition tasks, the person is asked to identify emotional expressions on faces, usually without any context. When AUDs had to determine an emotional face expression they presented a distinct neuropsychophysiological response, indexed by a decreased amplitude of early (P1, N10, N170) and late (LPP) ERPs (71). This alteration persists after a prolonged abstinence (59, 72). These difficulties for recognizing emotional facial expressions are highly related to more interpersonal difficulties, probably leading to social isolation and to an increase of their negative emotionality (59).

In summary, both early processing (P1, N1, N170) and later (LPP) components can be used as potential markers when evaluating negative emotionality aspects, such as affective processing, emotional recognition, and appraisal. For instance, alcohol seems to affect LPP from the beginning of consumption and together with N1, they are affected even in prolonged abstinence, indicating a possible role for these components as possible biomarkers. Moreover, considering its modularity by re-appraisal, LPP could be measured through time, in relation to cognitive and emotional regulation therapy.

Executive Dimension

Under the ANA framework, executive functions would be included within the executive domain, which comprises those higher-order processes mainly involved in the organization of behaviors, aimed at achieving future objectives (8, 73). Specifically, this domain is focused both on those processes of temporary organization of behavior such as attention, inhibition of response, planning, working memory and behavioral flexibility, as well as evaluation of future events (8). For an adequate measurement of the functions included within this domain, the authors of the ANA propose a series of assessment tests that can be widely used by both researchers and clinicians (8). However, in addition to the behavioral and self-reporting tests proposed by the ANA authors, they also recommend supplementing the use of these tests with other measures from neuroscience (8), but without specifying on any particular measure. This is because the evaluation of the executive dimension with behavioral and self-reporting tests exclusively may not identify aspects underlying these measures, such as inefficient brain functioning (74). This inefficient brain functioning may not manifest itself behaviorally and/or consciously in controlled contexts (e.g., attentional evaluation in clinical consultation), while in everyday contexts it does, putting at risk the maintenance of abstinence. In this case, psychophysiology can be very useful, because it can be sensitive to this type of information. Along these lines, quite interesting results have been found by combining different evaluation tests and different types of analysis of the electrophysiological signal.

One of the most studied tasks with the objective of obtaining a neuropsychophysiological marker of attentional control in alcohol addiction is the oddball paradigm. This task consists in the presentation of a series of infrequent stimuli (targets) during the presentation of stimuli in a frequent way (standards), allowing to evaluate the attentional processing from bottom-up as well as from top-down (75). The analysis of the electrophysiological activity recorded during the performance of this task that has been mostly carried out is that of ERPs (76–81). The task leads to the generation of different electrophysiological components, being P3a and P3b the most studied ones (76–81). These two electrophysiological components appear between 300 and 700 ms after the stimulus, and differ mainly in the cause that generates them and in the topographic distribution that they have. The first of these is the P3a component, whose evocation is produced by the

absence of explicit instruction to attend to the infrequent stimulus, and has a frontal topographical distribution (17). This component would reflect the bottom-up attentional processing, because there is no controlled processing of the stimuli presented. The second component is the P3b, which has a parietal topographic distribution, and whose evocation is produced by the explicit instruction to attend to the infrequent stimulus (17). In this case, this component would reflect the top-down attentional processing, since, unlike the P3a component, here there is a controlled processing of the stimuli presented in the task.

A large number of studies using this oddball paradigm in alcohol-dependent people have found that both the P3a and P3b components have a reduced amplitude compared to healthy controls (42, 79, 80, 82–84). A result that is replicated in healthy children of people with alcohol dependence (78, 85). This alteration of the P3 component both in people with alcohol dependence and in their offspring would reflect an attentional deficit from both the bottom-up and top-down processes, which seems to be produced by basal brain hyperexcitability (17, 76, 86). The literature proposes that this abnormal brain functioning is produced by an alteration in the mechanisms of cortical inhibition, and not by the consumption of alcohol per se (although it also contributes to the general impairment of brain functioning), being a previous vulnerability that they present (17, 76, 86). Because of this, the amplitude reduction of this component is proposed as an electrophysiological marker for alcohol addiction development. However, in other studies comparing this component between people with alcohol addiction and healthy controls, such as those performed by Bauer et al. (87), Fein et al. (83) and Malone et al. (88), these differences are not found when other variables such as the presence of a life history of major depression are taken into account. The discrepancy of results between studies may be reflecting the effect of the absence of differentiation between people who develop alcohol addiction due to the presence of different previous vulnerabilities (e.g., genetics), and people who develop it secondarily as a consequence of the presence of a particular set of symptoms (e.g., social anxiety) (81, 88, 89).

An example of this idea is reflected by Fein et al. (83). These authors study if there are differences in the P3b component generated by the oddball paradigm between people with alcohol addiction in abstinence with and without major depression. They found that those with alcohol addiction and major depression had no difference in the amplitude of the P3b component compared to the healthy controls group. They conclude that the absence of differences is due to the fact that the development of alcohol dependence, in this case, was caused by excessive consumption of the substance with self-medication as the main objective (83). Along the same lines, Malone et al. (88) propose that this does not only occur with the presence of concurrent major depression, but that this effect is also observed in people diagnosed with major depression throughout life. These two studies seem to reflect, by way of example, that the P3 component evoked during the performance of the oddball paradigm could help to identify those people with altered

attentional functioning as a consequence of a previous vulnerability, having clear repercussions in the choice of the most appropriate therapeutic line (e.g., pharmacological and neuropsychological vs. neuropsychological) (90–94).

Besides the oddball paradigm, the Go-NoGo paradigm is another task that has also been quite studied with the aim of obtaining other types of electrophysiological markers. This task has been used in different versions, varying some of its parameters, such as the type of stimuli presented [e.g., geometric figures (74), neutral stimuli or alcohol-related stimuli (95)]. However, the basic design of the task consists of presenting a series of Go stimuli more frequently to which the participant has to give some kind of response, interspersing NoGo stimuli less frequently, where the participant has to inhibit his response. Although apparently both the oddball and the Go-NoGo paradigms are very similar, the main difference is that the Go-NoGo paradigm requires greater involvement in the task by the participant, while in the oddball paradigm the necessary involvement is lower. Precisely because of this, the cognitive process that allows us to evaluate this task is the inhibitory capacity and the influence of different stimulation conditions on it.

With the use of the Go-NoGo paradigm, and as with the oddball paradigm, the analysis of ERPs has been the most carried out, which has provided the most consistent results around the N2 and P3 components (96–99). On the one hand, the negative component N2 appears around 250 ms after the presentation of the NoGo stimulus with a front-central distribution, with greater amplitude in the frontal region (98). The literature proposes that this component reflects the subject's ability to recognize the need to inhibit the response to a stimulus of these characteristics, where a greater amplitude of the component would reflect a better recognition of this need to inhibit (98, 100). On the other hand, the positive component P3 appears between 300 and 600 ms after the presentation of the NoGo stimulus with a fronto-centro-parietal distribution, with greater amplitude in the central region (96). In this case, the literature proposes that this component reflects the subject's ability to carry out an effective inhibition of the motor response, where a greater amplitude of the component would reflect a better inhibition of the response (101). Specifically, in people with alcohol addiction, the literature shows that both N2 and P3 can be reduced compared to healthy participants, reflecting an inhibitory deficit at one or both levels even when behavioral outcomes do not reflect this deficit (96–99).

In summary, the tasks with more clinical evidence for the evaluation of the executive dimension with electrical brain activity are the oddball and Go-NoGo paradigms. In the case of the oddball paradigm, the cognitive process evaluated is the bottom-up and top-down attentional processing. The neuropsychophysiological components obtained with this task with greater clinical utility are P3a and P3b, allowing to identify those people with alterations of the attentional functioning as a consequence of a previous vulnerability. In the case of the Go-NoGo paradigm, the cognitive processes evaluated are both the inhibitory capacity and the ability to identify the need to inhibit.

In this case, the neuropsychophysiological components obtained with greater clinical utility are N2 and P3, allowing the assessment of this capacity with greater sensitivity than behavioral data.

EVIDENCE IN OTHER NEUROPSYCHOPHYSIOLOGICAL MEASURES

Research lines regarding the search for neuropsychophysiological markers of neurocognitive alterations of alcohol dependence are rising as the possibilities for signal analyses increase. This allows us to evaluate EEG signal obtained with more complex paradigms from the methodological and cognitive point of view. By employing different types of analysis of the brain electrical activity, such as frequency analysis methods (102), event-related oscillations (103, 104) and functional connectivity analysis (102).

Frequency-Based Analysis

In addition to electrical activity related to events, brain oscillations analysis has been used across the literature. Most frequency bands have been evaluated during resting-state recordings in alcohol dependence, although with more diffuse results than those found in ERP studies. As an example, results in resting delta (0, 1–4 Hz) are considered inconclusive (15). With respect to theta band (4–8 Hz), various outcomes have been found (15, 103), but it is fundamentally evidenced as an increase in tonic theta in frontal, central, and parietal areas (105), with greater values for those patients that experience relapse (106). Other studies, however, find a decrease of theta power related to greater cortical damage (107). Changes in theta could be indicating the cortical imbalance in the excitation-inhibition homeostasis (15, 105). Moreover, theta has been related to inhibitory and motor responses (108).

With respect to alpha band (8–12 Hz), that predominates through the resting-state, it seems to be reduced in alcohol-dependent individuals (15, 109) and it has been associated with cortical activations, alert mechanisms, and active inhibition (110, 111), possibly indicating the activation of compensation mechanisms.

For beta band (14–30 Hz), the most frequent result indicates that patients have a greater beta power in fronto-central areas, similar to their offspring, where this increase is produced prior to developing an alcohol use problem (112). There is even evidence of a greater desynchronization of beta in patients that relapse comparing to those that remain abstinent. Porjesz and colleagues support the theory that a greater beta power at frontal sources reflects the disbalance between excitatory and inhibitory neurons, that could underlie to AUDs vulnerability (112–114). In this manner, while findings in beta have been considered as a trait marker, outcomes in theta have been thought as a state marker.

Event-Related Oscillations

Evoked response oscillations analysis allows us to know in a detailed manner characteristic of brain rhythms during a task

performance and their alterations. Within the study of the cognitive function, research groups have mainly found changes in theta band in several behavioral paradigms (34, 115–117).

In relation to incentive salience and motivational-related tasks, alcohol-dependent individuals show a decreased theta power during reward processing and weaker activity in prefrontal sources during the loss condition (34), the latter indicating difficulties in the risk evaluation process. Although a few number of studies are available regarding theta oscillations in substance-cue or reward processing, theta power is found altered in other ERP paradigms, namely visual oddball or conflict tasks in alcohol-dependent individuals (13, 118–120) and their offspring (103). Theta rhythm is thought to reflect frontal activity, implicated in attentional and monitoring processes (121) and it could be implicated in several cognitive operations, such as attentional control, inhibitory processes, and conflict responses.

Theta frequency band has been found to be modulated by stimuli valence. Aftanas et al. (122) measure theta synchronization and desynchronization while healthy subjects are exposed to stimuli with different emotional content, and find a time-locked synchronization theta at anterior and posterior sites, at 200 to 500 ms post stimuli presentation. This frequency band is useful as a neuropsychophysiological marker of emotional processing. In particular, theta power at frontal areas during the processing of affective information can be useful to assess emotional regulation or control. While theta at occipital sites can be used as a measurement of early emotional assessment. Regarding alcohol effects on theta band in emotional tasks, binge drinkers seem to have an attenuated theta power in an affective appraisal task, during both early appraisal and later integrative processes (115). This lower theta responsivity to emotions can suggest that binge drinkers already present some of the characteristic anhedonia of AUD patients.

In relation to the executive control dimension, for example, Kamarajan et al. (118) studied the evoked power of brain signals in alcoholics during the performance of a Go-NoGo task. The results showed that alcoholics had lower power in the delta and theta frequency bands compared to the control group in the NoGo condition (118). In this same line, Pandey et al. (117) found that alcoholics had lower power in the delta, theta, and alpha frequency bands compared to the control group, although behavioral differences were found only in the Go condition. Both studies reflect the presence of a neurocognitive deficit in both the execution and suppression of the motor response (117), where, in addition, Kamarajan et al. (118) suggest that oscillatory correlates during cognitive processing may be used as endophenotypic markers in alcoholism. Theta has already been suggested as indicative of attentional and executive processes (121), hence, these results might indicate a role for this oscillatory activity in detecting and characterizing alterations of these processes during the course of AUDs.

In summary, we could think of brain oscillatory rhythms as possible markers for cognitive processes, such as attentional allocation and affective processing, as well as behavioral monitoring and risk-taking, and executive inhibitory processes,

deeply involved in the dependence course and vulnerability toward maladaptive behaviors. In particular, theta and beta rhythms at rest and during cognitive operations could be of interest in the search for clinical or stable biomarkers

Functional Connectivity of the Brain

Nowadays research focuses on more detailed and global characterization of brain functioning, functional connectivity (FC) and graph theory-based analysis bringing information upon the wiring and effective communication of the brain during several cognitive and emotional processes. In this way, fMRI studies find connectivity changes in the brain affected by chronic alcohol consumption, with an alteration of white matter tracts (123), and changes in attentional and salience networks, as well as reward-related and executive ones (124–126). Coherence analysis or phase-synchrony in several frequency bands between pair of brain regions are frequent ways of studying FC in neuropsychophysiological measures (127). There are several outcomes related to alcohol effects in the brain communication in resting-state. In this way, some studies show a reduced connectivity in theta and alpha, as well as beta band (109, 128). Others show an increase in theta (112, 129–131) and interhemispheric alpha and beta (132). Despite the quite diverse outcomes, there seems to be an agreement upon their role in the brain, and some of them have even been proposed as possible markers of alcohol effects in the brain or as vulnerability markers, such as changes in theta connectivity (131). In this manner, alterations in FC of theta might indicate changes in the inhibitory neuronal system, related to GABAergic and cholinergic neurotransmission (112), as well as emotional and motivational processing (133), whereas beta and alpha FC might reflect supervision and coordination processes involved in brain activation and deactivation systems (134).

Although reduced, there are some studies carried out in active states in EEG regarding alcohol or other substances' effects on functional connectivity study of the brain. In incentive salience, an fMRI study was carried out in a cue-exposure paradigm, as well as a resting-state EEG recording of alcohol-dependent individuals (130). Results showed fMRI changes in FC in frontal and limbic regions, highly implicated in motivated attention toward alcohol cues, as well as an increased connectivity in theta band in resting EEG in similar regions. Theta is considered to reflect emotional and motivational processes, and this result might underlie to the alterations present in the reward evaluation system in alcohol dependence. The authors of the mentioned study think that theta hyperconnectivity might have a relationship with craving, even hypothesizing the existence of a “central craving network”, where different regions are in charge of appetitive and motivational aspects involved in incentive salience (130). Moreover, this idea is supported by a study with smokers, where theta coherence is increased toward nicotine cues in frontal and parieto-occipital sites, and it also predicts changes in craving (135). Alcohol-related cues could have a similar effect in brain connectivity, in fact, one of our studies (136) showed a relationship between resting-state beta connectivity and the approximation index

toward aversive contexts related to the substance in a modified Alcohol Approach-Avoidance task (AAT). Specifically, a higher beta connectivity was related to a greater avoidance of aversive alcohol-related contexts, possibly indicating beta synchronization role in motivational and salience circuits.

Regarding reward-related processing, such as monetary tasks, no studies using EEG or MEG measures were found in FC in alcohol dependence. However, a study carried out in healthy population indicates changes in FC (133), reflected by increases in theta synchronization between frontal region and mostly parietal areas during the loss condition and greater theta FC within posterior regions during the gain condition. Moreover, this study is carried out in co-twins and they find a genetic heritability of fronto-parietal connectivity in theta during the loss condition, indicating that reward evaluation of negative outcomes could be genetically transmitted. Taking this into consideration, it would be interesting to evaluate theta connectivity in incentive or reward tasks and find out its possible role as a vulnerability marker for alterations in reward evaluation and its relationship with maladaptive behavior.

Neuropsychophysiological connectivity measures could also be applied to the assessment of negative emotionality. In healthy population, increased long-range connections between frontal parietal and temporal areas in beta and gamma bands are related to negative and positive emotional information processing (137). In this way, fast rhythms communication through the brain could play a role in emotional processing and could be of use in the research of neural markers of alterations in AUD.

In executive functioning, FC analysis can have a special relevance in the knowledge of electrical brain functioning in resting state and its relation to different executive aspects (e.g., impulsivity). Herrera-Diaz et al. found that alcohol-dependent individuals present, together with an increased beta power, a reduced FC at fronto-central and occipito-parietal regions in alpha and beta bands, comparing to healthy controls. What is more, resting FC in alpha band between anterior and central regions seems to have an inverse relationship with BIS-11 non-planned impulsivity scores (102). Authors propose that results reflect the existence of alterations in the brain's electrical signal at rest in alcohol-dependent individuals, indicating a possible association to psychopathological characteristics of the addictive behavior (102).

Brain Graph Characteristics

Research in brain connectivity and dynamics has been increasingly developed during the last years, revealing the importance of neural integration and segregation of communication between brain areas, as well as communication patterns, hubs, and efficiency of the information flow. These particular measures can be useful in different pathologies where functional brain alterations are evidenced (138, 139), as in addictions (140, 141). These measures rely on graph theory, a mathematical model used in several fields of science, particularly useful in neuroscience, more specifically in brain functional and effective connectivity. It comprehends brain functioning as a network, composed by vertices or nodes, e.g., brain areas or

channels and edges, e.g., connections (measured by statistical dependence) between pairs of areas (142). The functioning or communication between these nodes is characterized in terms of several elements, such a distance or path length between two areas, the number of connections received by a node (degree) or the number of connections forming a triangle around a node (clustering coefficient) (143). Moreover, the efficacy of this flow of communication depends on the distance between nodes, being inversely related to this index. These characteristics can be calculated at both local (segregation measures) and global (integration measures) levels (144). Segregation refers to the functioning of specialized areas and local networks in the brain, and it can be measured through parameters such as degree, local clustering or local efficiency. Whereas integration involves the coordination of neural populations giving rise to cognitive states, and it is generally measured by indexes such as the average shortest distance (characteristic path length) between nodes, global efficiency and global clustering level. These two principles create diverse and complex patterns of communication in the brain, allowing a balance between flow efficiency and costs, at both resting and during active states. In this way, brain connectivity tends toward a balance in energetic cost and to a maximization of the network (145). This is known as the “small-worldness” of a network and it implies an optimal functioning of local and global communication, characterized by a high global efficiency and clustering level, as well as a short characteristic path length (146).

So, how can these brain graph measures help us in the search for neuroscience-based clinical tools in alcohol dependence? Small-world attributes and characterization of neuropsychophysiological activity patterns can bring answers upon the mapping of functional connections in the altered networks in alcohol dependence. Information processing can be put together with a precise characterization and mapping of the communication flow through the brain, by detecting key nodes or hubs of connections, as well as possible changes in the network organization (147). In other words, we could detect local and global network deficiencies as well as possible compensation mechanisms, with extended communication to other areas or networks. In fact, the available literature upon this theme is still short, but it points toward this type of results. For example, in a fMRI study, individuals with policonsumption show a lower efficiency and a reduced small-worldness of the brain network (140), reflecting a loss of interregional communication in the brain. In alcohol dependence, a smaller global efficiency and clustering level have been related to a greater consumption severity (148), indicating their possible role in identifying neural markers for clinical severity. There is also data in EEG studies, low doses of alcohol in social consumers producing higher global efficiency and an increased density in resting alpha, as well as a short characteristic path (12). Similar observations were made in alcohol-dependent individuals during a working memory task (149), where smaller characteristic paths, reduced clustering, and an increased global efficiency were found in low beta band. These results might be indicating an altered functioning of network efficiency under the effects of

alcohol, as well as a compensating mechanism in response to task demands, in order to carry out cognitive operations. Hence, graph-based measures of brain connectivity using neuropsychophysiological measures could help explain in detail neural communication alterations that happen through the course of dependence.

DISCUSSION

Brain electrical activity has been used with some degree of evidence to evaluate several of the cognitive processes that are contemplated in the dimensions of ANA. These cognitive components may describe alcohol dependence as an active process, in which event-related potentials or oscillations serve as a measure of the progression of the disorder. For instance, in relation to stimuli relevance evaluation, a greater alcohol salience can be observed in early visual ERP components, reflecting motivational processes. A common finding is a greater amplitude and sometimes latency of P1, N1, or P2 components, reflecting a preferential attention toward alcohol and related cues. This preferential attention can affect other cognitive processes, such as the inhibitory capacity, reflected by greater N2 amplitudes when alcohol-dependent individuals carry out an inhibitory process in presence of alcohol cues, dampening behavioral control. Moreover, incentive salience is also evidenced through higher P3 amplitudes in frontal regions and a reduced power of theta band, during loss and gain conditions in decision-making tasks. This supports the behavioral results that indicate difficulties within the proper evaluation of behavioral alternatives in AUDs. In the clinical context, patients with more altered values in this dimension put alcohol at the center of their thinking, without attending to other things in the environment. As a consequence, the risk of relapse in situations where alcohol is present is greater.

Patients are not always aware of this attentional bias, so a good therapeutic proposal would be the training in attentional control and avoidance of risky situations at least at the beginning of treatment. As treatment progresses, work through associative learning could be targeted to establish new contingencies, new stimulus-response-reinforcement relationships. At this point the patient could address a re-evaluation of the way he makes decisions, to value the new reinforcements and train the delay of the reward (see **Table 1**).

Although the evidence in negative emotionality is scarcer and at the time being is still far from being used as a possible endophenotype for AUDs, it could give rise to promising results. In this manner, we could consider neuropsychophysiological activity under affective processes as markers for processes we know as altered in alcohol dependence. To this point, the bibliography considers the use of P1, N1, P2 and LPP components as evidence for the deficient processing of affective content. In addition, there is a promising line in which there is evidence for theta frequency band. Theta seems to be less modulated by affective content in binge drinkers (115), possibly indicating a reduced sensitivity to negative and

TABLE 1 | Behavioral and ERP/ERO paradigms within ANA framework (Incentive salience).

| ANA dimension | Paradigm | Psychophysiological variable | Cognitive processes assessed | Population | Treatment target | Vulnerability | Severity | Outcome | Course |
|--------------------|--|--|---|---|--|---------------|----------|---------|--------|
| Incentive salience | <i>Cue reactivity task</i> | N1: higher amplitude | Preferential attentional processing of substance cues | Alcohol dependence | Attentional bias/ Associative learning | ? | +++ | ? | ? |
| | | P1: higher latency | Faster early attentional processing of alcohol cues | Social drinkers | | ? | +++ | ? | ? |
| | <i>Dot-probe attentional bias task</i> | P1: higher amplitude and latency | Preferential early attention orientation | Individuals at risk (low alcohol sensitivity) | Attentional bias/ Associative learning | + | ? | ? | ? |
| | | Larger negativity between 220 and 280 ms | Difficulties reorienting attention away from alcohol cues | Individuals at risk (low alcohol sensitivity) | | + | ? | ? | ? |
| | <i>Monetary incentive delay task</i> | P300: decreased amplitude | Task demands processing and context updating | Alcohol dependence and offspring | Cognitive evaluation process and decision making | + | + | + | ++ |
| | | Theta: reduced power | Attentional resource allocation Decision making Reward processing | Alcohol dependence | | + | + | + | ? |

This table contains a brief summary of main ERP components, ERO activity and their alterations related to alcohol effects, within ANA's domain Incentive salience. It also makes a small mention to main cognitive functions that can be targeted inside psychotherapeutic programs. Shadowed columns refer to the evidence gathered by each neuropsychophysiological marker in the evaluation of cognitive and motivational processes of alcohol dependence in four steps: vulnerability toward alcohol use disorders, dependence severity, outcome and course. "<++++>" indicates consistent outcomes in the literature, "<+>" indicates the presence of some studies, although inconclusive, "<+>" implies an incipient line of study, with preliminary results and finally, "<?>" indicates a future line of work.

Potential clinical use and treatment targets.

positive affect. In this way, those people with less recognition of the emotional content, and less adjustment to it, evidenced by the electrical activity, will have greater difficulties in interpersonal relationships (59). Difficulty in recognizing one's own emotions and those of others can lead on the one hand to interpersonal conflicts, which are considered to be one of the main risk factors for relapses (150, 151). There is also evidence of how the presence of negative affect can impair performance in other areas. For example escape drinkers, that is, individuals motivated to drink in order to avoid negative emotions, show greater N2 amplitudes, implicated in more

controlled attentional operations, indicating a greater attentional bias toward alcohol cues (39). Hence, the presence of negative affect seems to lead to greater attention being paid to alcohol-related stimuli. Moreover, it could lead to the use of alcohol as the most effective mechanism for emotional self-regulation. Thus, a treatment goal for this patient profile should first impact on the identification of both negative and positive emotions and the acceptance that negative emotions are adaptive. Once the patient is aware of his or her emotional state, the therapeutic objective could be emotional regulation and adaptive coping (see Table 2).

TABLE 2 | Behavioral and ERP/ERO paradigms within ANA framework (Negative emotionality).

| ANA dimension | Paradigm | Psychophysiological variable | Cognitive processes assessed | Population | Treatment target | Vulnerability | Severity | Outcome | Course |
|-----------------------|------------------------------------|--|--|--|------------------------------|---------------|----------|---------|--------|
| Negative emotionality | <i>Affective images processing</i> | LPP: decreased amplitude to negative images. | Emotional cues classification and processing | Acute alcohol intake Binge drinkers | Affect regulation and coping | + | ? | ? | ? |
| | <i>Appraisal</i> | Theta: decreased power | Emotional processing | Binge drinkers | Re-appraisal | + | ? | ? | ? |
| | <i>Emotional Face expressions</i> | P100, N100, and N170: decreased amplitude. | Recognizing emotional face expressions | Alcohol dependence | Conscious affect processing | ? | ? | ? | ? |

This table contains a brief summary of main ERP components, ERO activity and their alterations related to alcohol effects, within ANA's domain negative emotionality. It also makes a small mention to main cognitive functions that can be targeted inside psychotherapeutic programs. Shadowed columns refer to the evidence gathered by each neuropsychophysiological marker in the evaluation of cognitive and motivational processes of alcohol dependence in four steps: vulnerability toward alcohol use disorders, dependence severity, outcome and course. "<++++>" indicates consistent outcomes in the literature, "<+>" indicates the presence of some studies, although inconclusive, "<+>" implies an incipient line of study, with preliminary results and finally, "<?>" indicates a future line of work.

Potential clinical use and treatment targets.

TABLE 3 | Behavioral and ERP/ERO paradigms within ANA framework (Executive dimension).

| ANA dimension | Paradigm | Psychophysiological variable | Cognitive processes assessed | Population | Treatment target | Vulnerability | Severity | Outcome | Course |
|---------------------|-------------------------|---|----------------------------------|--|---|---------------|----------|---------|--------|
| Executive Dimension | <i>Oddball Paradigm</i> | P3a and P3b: reduced amplitude. P3: reduced Delta, theta: decreased power | Attentional resource allocation | Alcohol dependence Offspring Offspring | Attentional control | +++ | +++ | +++ | + |
| | <i>Go-NoGo</i> | N2: reduced amplitude | Inhibition of the motor response | Alcohol dependence | Executive behavior: Identification of high-risk situations. Generation of alternative responses | + | ++ | ++ | + |
| | | P300: reduced amplitude | Inhibition of the motor response | Alcohol dependence and offspring | | ++ | +++ | +++ | ? |
| | | Delta | Attention and task demand | | | + | ? | ? | ? |
| | | Theta | Attention and inhibition | Alcohol dependence | | ++ | ? | ? | ? |
| | | Alpha | Inhibition | | | + | ? | ? | ? |

This table contains a brief summary of main ERP components, ERO activity and their alterations related to alcohol effects, within ANA's domain executive function. It also makes a small mention to main cognitive functions that can be targeted inside psychotherapeutic programs. Shadowed columns refer to the evidence gathered by each neuropsychophysiological marker in the evaluation of cognitive and motivational processes of alcohol dependence in four steps: vulnerability toward alcohol use disorders, dependence severity, outcome and course. "<+++>" indicates consistent outcomes in the literature, "<+>" indicates the presence of some studies, although inconclusive, "<+>" implies an incipient line of study, with preliminary results and finally, "<?>" indicates a future line of work.

Potential clinical use and treatment targets.

Finally, the most conclusive results are in the P3 component (see **Table 3**), where both alcohol-dependent patients and their offspring show alterations. Which represents a clear example of a possible endophenotype. This component reflects controlled processes of conscious attention, context updating processing and executive attention (15, 152), that are clearly comprised inside the executive dimension of ANA. In this dimension, there is also evidence in N2, and the delta, theta and alpha frequency bands, reflecting alterations in inhibitory control. The patient with alterations in this cognitive domain will be especially vulnerable to relapses, as it will be difficult to stop automatic behaviors in benefit of more controlled processes, such as rejecting the tendency to consume alcohol one they are inside a context of risk (153).

Taking into account what has been reviewed so far, the combination of information from neuropsychology, clinical psychology, and neuropsychophysiological markers could lead to differentiated clinical profiles, based not on the final behavior of pathological consumption, but on the underlying cognitive processes. The objective for a future line of research would be to adjust the treatment to the deficient processes and to the preserved ones. In addition, the development of this line of work, perhaps can lead us to explain the clinical heterogeneity

that we find in daily practice. Thus, as an example for illustrative purpose, a patient with an A profile (see **Table 4**), who manifests both moderate levels of incentive salience and negative emotionality, and a mild affectation of the executive component, could find himself in situations of risk when exposed to the context of consumption, or in moments of emotional conflict. But he would only relapse when his executive control capacity would fail. Possibly his relapse would be characterized by a consumption not necessarily intense and with a determined duration until he could recover the executive control. However, the patient with a B profile (see **Table 4**), under the same risk conditions, but with severe impairment of the executive function, would hypothetically relapse more easily, leading to a relapse of a greater intensity and prolonged in time.

This future line of work would help design long-term treatment strategies, which considers the starting point, but also the evolution of these cognitive processes within the process of change caused by treatment and abstinence, and therefore it would be individualized (1, 2). Thus, within the framework of techniques that already have proven efficacy, such as contingency management, relapse prevention, motivational interviewing, or pharmacotherapy, the A-profile patient could follow a standardized treatment as a basis,

TABLE 4 | Examples of cognitive profiles in AUD based on cognitive and ERP/ERO evidence.

| Domain | Low impairment | Moderate impairment | Severe impairment |
|-----------------------|----------------|---------------------|-------------------|
| Incentive Salience | + | ++ | +++ |
| Negative emotionality | + | ++ | +++ |
| Executive control | + | ++ | +++ |

Hypothetical deterioration profiles in the three dimensions of the ANA framework. The crosses indicate the severity of the alteration in each dimension analyzed. "<+>" indicates low impairment, "<+>" moderate impairment, and "<+++>" severe impairment.

but with an emphasis on recognizing risk situations and formulating alternative plans for consumption. While the B-profile patient, in addition to following the standardized treatment, could reinforce the therapy, by carrying out in the initial moments of the treatment a good management of contingencies, avoiding the situations related to alcohol, and then working explicitly with techniques of compensation or substitution of the inhibitory control.

This line of work has results on specific components such as the attention bias, approximation trends toward alcohol, and the underlying neuropsychophysiological measures such as alpha power or P3 amplitude, which seem to change after treatment (136, 154, 155), being good markers of interindividual change. Therefore, there could be evidence of greater homogeneity when used to assess the clinical course of patients. Since they show a great sensitivity to change along the treatment process (155, 156). Possibly because they reflect the integrity of some cognitive components that we could consider as intermediate endophenotypes, or as “stepping stones” to the disorder, but also toward the clinical recovery (8). This is a very promising line of work.

Notwithstanding, given the variability of the revised measures, the applicability of this proposal is still developing, in order to be accessible for the different health care professionals. Patrick et al. revise the specific needs for implementing these types of measures in our daily clinical practice (10), highlighting several of them. The first one has to do with the need for technological resources: although EEG equipment is habitual in hospital units, they are not always prepared for a complex event-related activity measurement. Additionally, this evaluation supposes an extensive methodological effort, this is probably the greatest difficulty encountered for results replicability and their employment. The second important need has to do with the generalization of the results and their consistency. A great diversity of cognitive and behavioral paradigms is available for cognitive and EEG evaluation, with different types of stimuli, diverse presentation times and even required responses from individuals. This could explain part of the variability found in research, thus a need for studies that verify validity and reliability of the proposed measures is becoming relevant. In this way, standardization and validation processes of behavioral event-related potentials paradigms would be enabled, similar to those carried out to develop neuropsychological assessment (10).

Despite the variability, most of the groups working in this line of research agree in highlighting several advantages of including electrophysiological variables as objective measures of the cognitive process, since their alteration may be present in patients whether or not the final behavior is successful, reflecting the lack of efficiency of the process (10). In this way, the results of neuropsychological and behavioral evaluation sometimes do not identify the high effort that the patient makes to reach successful behavior. Therefore, they do not identify either the lack of resources to cope with more demanding situations, since resources are destined to frequent events. Leaving the patient unable to adjust to more difficult or novel situations. That may make more difficult reinsertion into

the different life spheres for the patient with alcohol dependence and in the final recovery achievement (11, 157). Thus, the employment of these types of measures brings a certain type of information that would otherwise pass unnoticed in the clinical context and that could be relevant in order to obtain a more objective profile of the three dimensions proposed by ANA. In the end, this would help into the development of adequate individualized treatment designs (1).

In conclusion, the neuropsychophysiological evaluation through event-related potentials is already set for being employed in the clinical context, although a more extensive diffusion and a process of standardization of paradigms are still necessary, so that professionals can acknowledge its use and for a greater applicability inside personalized interventional programs.

Future lines of work can focus on the use of novel measures of brain communication in AUD characterization. Functional and effective connectivity analyses of the brain can bring light upon the synchronization and coordination between regions and networks that take place during cognitive processing. Moreover, brain graph measures can help us observe specific brain areas that may be acting as hubs of communication, as well as network efficiency at local and global levels in several cognitive functions. In this way, we could observe specific network functioning and interacting from preattentive processes to more controlled and complex mental operations. This is interesting for AUD alterations, since several cognitive domains present deficiencies, considering the early salience of alcohol cues and problems in the affective processing and the executive function. To this date, functional connectivity studies in psychiatric conditions are mostly exploratory and have the purpose of characterizing the neural functioning of the brain. In AUD, functional connectivity studies using EEG have found alterations in brain synchronization in several frequency bands (alpha, beta or gamma) (132, 149, 158) and they seem to present a reduced efficiency of communication, although results are somehow scarce and diverse. Nonetheless, prospective studies should be able to offer clearer hypotheses on brain alterations in psychiatric patients, such as AUD, and to help find out if they persist in time or are modulated by different clinical factors.

AUTHOR CONTRIBUTIONS

RJ-B, JP-M, and GR performed the *Introduction* and the *Discussion* sections. AS performed the *Incentive Salience* section. ID-C performed the *Negative Emotionality* section. AM-M performed the *Executive Dimension*. RJ-B, GR, AS, ID-C, and AM-M contributed to the performance of the *Other Psychophysiological Evidences*, *Brain Functional Connectivity*, and *Network Characteristics* sections. FN, MG-G, and JM have collaborated in the manuscript planning and reviewing, as well as the writing supervision in terms of relevance for the reader. JM contributed by selecting the most relevant information for the aim of the publication. FN and MG-G have made a revision of the English version of the manuscript.

ACKNOWLEDGMENTS

The preparation of the manuscript was supported by: 'Instituto de Salud Carlos III, Fondos FEDER, Red de Trastornos Adictivos' (RTA, RD16/0017/0014 to JM and RD16/0017/0017 to GR); 'Ministerio de Sanidad, Delegación del Gobierno para el

Plan Nacional Sobre Drogas' (PNSD, 2019I012 to JM and PNSD,2016-Rubio to GR, AS, JP-M and RJ-B); 'Ministerio de Ciencia e Innovación, Fondo de Investigaciones Sanitarias' (FIS15/00463 to GR and AM-M; FIS, PI18/00576 to JM); 'Ministerio de Economía y Competitividad' (PSI2015-68851-P to RJ-B and ID).

REFERENCES

- Campbell EJ, Lawrence AJ, Perry CJ. New steps for treating alcohol use disorder. *Psychopharmacol (Berl)* (2018) 235:1759–73. doi: 10.1007/s00213-018-4887-7
- Litten RZ, Ryan ML, Falk DE, Reilly M, Fertig JB, Koob GF. Heterogeneity of alcohol use disorder: Understanding mechanisms to advance personalized treatment. *Alcohol Clin Exp Res* (2015) 39:579–84. doi: 10.1111/acer.12669
- Heinz A, Beck A, Grusser SM, Grace AA, Wrase J. Identifying the neural circuitry of alcohol craving and relapse vulnerability. *Addict Biol* (2009) 14:108–18. doi: 10.1111/j.1369-1600.2008.00136.x
- Kelly JF, Greene MC, Bergman BG, White WL, Hoepfner BB. How Many Recovery Attempts Does it Take to Successfully Resolve an Alcohol or Drug Problem? Estimates and Correlates From a National Study of Recovering U.S. Adults. *Alcohol Clin Exp Res* (2019) 43:1533–44. doi: 10.1111/acer.14067
- Kazdin AE. Mediators and Mechanisms of Change in Psychotherapy Research. *Annu Rev Clin Psychol* (2007) 3:1–27. doi: 10.1146/annurev.clinpsy.3.022806.091432
- Insel T, Cuthbert B, Garvey M, Heinssen R, Pine DS, Quinn K, et al. Research Domain Criteria (RDoC): Toward a New Classification Framework for Research on Mental Disorders. *Am J Psychiatry* (2010) 167:748–51. doi: 10.1176/appi.ajp.2010.09091379
- Kwako LE, Bickel WK, Goldman D. Addiction Biomarkers: Dimensional Approaches to Understanding Addiction. *Trends Mol Med* (2018) 24(2):206–20. doi: 10.1016/j.molmed.2017.12.007
- Kwako LE, Momenan R, Litten RZ, Koob GF, Goldman D. Addictions Neuroclinical Assessment: A Neuroscience-Based Framework for Addictive Disorders. *Biol Psychiatry* (2016) 80(3):179–89. doi: 10.1016/j.biopsych.2015.10.024
- National Institute of Mental Health. Research Domain Criteria (RDoC). Available at: <https://www.nimh.nih.gov/research/research-funded-by-nimh/rdoc/units/index.shtml>
- Patrick CJ, Iacono WG, Venables NC. Incorporating neurophysiological measures into clinical assessments: Fundamental challenges and a strategy for addressing them. *Psychol Assess* (2019) 31:1512–29. doi: 10.1037/pas0000713
- Yücel M, Oldenhof E, Ahmed SH, Belin D, Billieux J, Bowden-Jones H, et al. A transdiagnostic dimensional approach towards a neuropsychological assessment for addiction: an international Delphi consensus study. *Addiction* (2019) 114(6):1095–109. doi: 10.1111/add.14424
- Lithari C, Klados MA, Pappas C, Albani M, Kapoukranidou D, Kovatsi L, et al. Alcohol affects the brain's resting-state network in social drinkers. *PLoS One* (2012) 7:e48641. doi: 10.1371/journal.pone.0048641
- Harper J, Malone SM, Iacono WG. Impact of alcohol use on EEG dynamics of response inhibition: a cotwin control analysis. *Addict Biol* (2018) 23:256–67. doi: 10.1111/adb.12481
- Schwarz E, Kielholz P, Hobi V, Goldberg L, Gilsdorf U, Hofstetter M, et al. Alcohol-induced biphasic background and stimulus-elicited EEG changes in relation to blood alcohol levels. *Int J Clin Pharmacol Ther Toxicol* (1981) 19:102–11.
- Kamarajan C, Porjesz B. Advances in Electrophysiological Research. *Alcohol Res* (2015) 37:53–87.
- Campanella S, Noël X, Tomberg C. Cognitive Event-Related Potentials and Alcoholism. *J Psychophysiol* (2010) 24:231–9. doi: 10.1027/0269-8803/a000036
- Porjesz B, Rangaswamy M, Kamarajan C, Jones KA, Padmanabhapillai A, Begleiter H. The utility of neurophysiological markers in the study of alcoholism. *Clin Neurophysiol* (2005) 116:993–1018. doi: 10.1016/j.clinph.2004.12.016
- Plan Nacional Sobre Drogas' (PNSD, 2019I012 to JM and PNSD,2016-Rubio to GR, AS, JP-M and RJ-B); 'Ministerio de Ciencia e Innovación, Fondo de Investigaciones Sanitarias' (FIS15/00463 to GR and AM-M; FIS, PI18/00576 to JM); 'Ministerio de Economía y Competitividad' (PSI2015-68851-P to RJ-B and ID).
- Campanella S, Schroder E, Kajosch H, Noel X, Kornreich C. Why cognitive event-related potentials (ERPs) should have a role in the management of alcohol disorders. *Neurosci Biobehav Rev* (2018) 106:234–44. doi: 10.1016/j.neubiorev.2018.06.016
- Stacy AW, Wiers RW. Implicit Cognition and Addiction: A Tool for Explaining Paradoxical Behavior. *Annu Rev Clin Psychol* (2010) 6:551–75. doi: 10.1146/annurev.clinpsy.121208.131444
- Aragues M, Jurado R, Quinto R, Rubio G. Laboratory paradigms of impulsivity and alcohol dependence: a review. *Eur Addict Res* (2011) 17:64–71. doi: 10.1159/000321345
- Goldstein RZ, Volkow ND. Drug addiction and its underlying neurobiological basis: neuroimaging evidence for the involvement of the frontal cortex. *Am J Psychiatry* (2002) 159:1642–52. doi: 10.1176/appi.ajp.159.10.1642
- Everitt BJ, Belin D, Economidou D, Pelloux Y, Dalley JW, Robbins TW. Neural mechanisms underlying the vulnerability to develop compulsive drug-seeking habits and addiction. *Philos Trans R Soc B: Biol Sci.* (2008) 363(1507):3125–35. doi: 10.1098/rstb.2008.0089
- O'Brien CP, Childress AR, Ehrman R, Robbins SJ. Conditioning factors in drug abuse: can they explain compulsion? *J Psychopharmacol* (1998) 12:15–22. doi: 10.1177/026988119801200103
- Kalivas PW, Volkow ND. The neural basis of addiction: a pathology of motivation and choice. *Am J Psychiatry* (2005) 162:1403–13. doi: 10.1176/appi.ajp.162.8.1403
- Myrick H, Anton RF, Li X, Henderson S, Drobos D, Voronin K, et al. Differential Brain Activity in Alcoholics and Social Drinkers to Alcohol Cues: Relationship to Craving. *Neuropsychopharmacology* (2004) 29:393–402. doi: 10.1038/sj.npp.1300295
- Filbey FM, Claus E, Audette AR, Niculescu M, Banich MT, Tanabe J, et al. Exposure to the taste of alcohol elicits activation of the mesocorticolimbic neurocircuitry. *Neuropsychopharmacology* (2008) 33:1391–401. doi: 10.1038/sj.npp.1301513
- Reid MS, Flammino F, Starosta A, Palamar J, Franck J. Physiological and subjective responding to alcohol cue exposure in alcoholics and control subjects: evidence for appetitive responding. *J Neural Transm* (2006) 113:1519–35. doi: 10.1007/s00702-005-0439-5
- Duka T, Townshend JM. The priming effect of alcohol pre-load on attentional bias to alcohol-related stimuli. *Psychopharmacol* (2004) 176:353–61. doi: 10.1007/s00213-004-1906-7
- Townshend JM, Duka T. Attentional bias associated with alcohol cues: differences between heavy and occasional social drinkers. *Psychopharmacol* (2001) 157:67–74. doi: 10.1007/s002130100764
- Field M, Cox WM. Attentional bias in addictive behaviors: a review of its development, causes, and consequences. *Drug Alcohol Depend* (2008) 97:1–20. doi: 10.1016/j.drugalcdep.2008.03.030
- Bjork JM, Hommer DW, Grant SJ, Danube C. Impulsivity in abstinent alcohol-dependent patients: Relation to control subjects and type 1-/type 2-like traits. *Alcohol* (2004) 34:133–50. doi: 10.1016/j.alcohol.2004.06.012
- Amlung M, Sweet LH, Acker J, Brown CL, MacKillop J. Dissociable brain signatures of choice conflict and immediate reward preferences in alcohol use disorders. *Addict Biol* (2014) 19:743–53. doi: 10.1111/adb.12017
- Rubio G, Jiménez M, Rodríguez-Jiménez R, Martínez I, Ávila C, Ferre F, et al. The role of behavioral impulsivity in the development of alcohol dependence: A 4-year follow-up study. *Alcohol Clin Exp Res* (2008) 32:1681–7. doi: 10.1111/j.1530-0277.2008.00746.x
- Kamarajan C, Rangaswamy M, Manz N, Chorlian DB, Pandey AK, Roopesh BN, et al. Topography, power, and current source density of theta oscillations during reward processing as markers for alcohol dependence. *Hum Brain Mapp* (2012) 33:1019–39. doi: 10.1002/hbm.21267

35. Ehrman RN, Robbins SJ, Bromwell MA, Lankford ME, Monterosso JR, O'Brien CP. Comparing attentional bias to smoking cues in current smokers, former smokers, and non-smokers using a dot-probe task. *Drug Alcohol Depend* (2002) 67:185–91. doi: 10.1016/S0376-8716(02)00065-0
36. Manchery L, Yarmush DE, Luehring-Jones P, Erlich J. Attentional bias to alcohol stimuli predicts elevated cue-induced craving in young adult social drinkers. *Addict Behav* (2017) 70:14–7. doi: 10.1016/j.addbeh.2017.01.035
37. Schacht JP, Anton RF, Myrick H. Functional neuroimaging studies of alcohol cue reactivity: a quantitative meta-analysis and systematic review. *Addict Biol* (2013) 18:121–33. doi: 10.1111/j.1369-1600.2012.00464.x
38. Knutson B, Adams CM, Fong GW, Hommer D. Anticipation of increasing monetary reward selectively recruits nucleus accumbens. *J Neurosci* (2001) 21:Rc159. doi: 10.1523/JNEUROSCI.21-16-j0002.2001
39. Dickter CL, Forestell CA, Hammett PJ, Young CM. Relationship between alcohol dependence, escape drinking, and early neural attention to alcohol-related cues. *Psychopharmacol (Berl)* (2014) 231:2031–40. doi: 10.1007/s00213-013-3348-6
40. Lang PJ, Bradley MM, Cuthbert BN. *Motivated Attention: Affect, Activation, and Action*. Lang PJ, Simons RF, Balaban MT, editors. Hillsdale, NJ: Lawrence Erlbaum Associates (1997).
41. Baler RD, Volkow ND. Drug addiction: the neurobiology of disrupted self-control. *Trends Mol Med* (2006) 12:559–66. doi: 10.1016/j.molmed.2006.10.005
42. Namkoong K, Lee E, Lee CH, Lee BO, An SK. Increased P3 amplitudes induced by alcohol-related pictures in patients with alcohol dependence. *Alcohol Clin Exp Res* (2004) 28:1317–23. doi: 10.1097/01.ALC.0000139828.78099.69
43. Martinovic J, Jones A, Christiansen P, Rose AK, Hogarth L, Field M. Electrophysiological responses to alcohol cues are not associated with Pavlovian-to-instrumental transfer in social drinkers. *PLoS One* (2014) 9:e94605. doi: 10.1016/j.jpsycho.2014.08.001
44. Krocze AM, Haeussinger FB, Hudak J, Vanes LD, Fallgatter AJ, Ehls AC. Cue reactivity essentials: Event-related potentials during identification of visual alcoholic stimuli in social drinkers. *J Stud Alcohol Drugs* (2018) 79:137–47. doi: 10.15288/jsad.2017.79.137
45. Martins JS, Bartholow BD, Cooper ML, Irvin KM, Piasecki TM. Interactive Effects of Naturalistic Drinking Context and Alcohol Sensitivity on Neural Alcohol Cue-Reactivity Responses. *Alcohol Clin Exp Res* (2019) 43:1777–89. doi: 10.1111/acer.14134
46. Bartholow BD, Lust SA, Tragesser SL. Specificity of P3 event-related potential reactivity to alcohol cues in individuals low in alcohol sensitivity. *Psychol Addict Behav* (2010) 24:220–8. doi: 10.1037/a0017705
47. Kreusch F, Quertemont E, Vilenne A, Hansenne M. Alcohol abuse and ERP components in Go/No-go tasks using alcohol-related stimuli: impact of alcohol avoidance. *Int J Psychophysiol* (2014) 94:92–9. doi: 10.1177/0269881114545663
48. Matheus-Roth C, Schenk I, Wiltfang J, Scherbaum N, Müller BW. Occipital event-related potentials to addiction-related stimuli in detoxified patients with alcohol dependence, and their association with three-month relapse. *BMC Psychiatry* (2016) 16:1–12. doi: 10.1186/s12888-016-0782-0
49. Shin E, Hopfinger JB, Lust SA, Henry EA, Bartholow BD. Electrophysiological Evidence of Alcohol-Related Attentional Bias in Social Drinkers Low in Alcohol Sensitivity. *Psychol Addict Behav* (2010) 24:508–15. doi: 10.1037/a0019663
50. Kamarajan C, Rangaswamy M, Tang Y, Chorlian DB, Pandey AK, Roopesh BN, et al. Dysfunctional reward processing in male alcoholics: An ERP study during a gambling task. *J Psychiatr Res* (2010) 44:576–90. doi: 10.1016/j.jpsychires.2009.11.019
51. Kamarajan C, Pandey AK, Chorlian DB, Manz N, Stimus AT, Bauer LO, et al. Reward processing deficits and impulsivity in high-risk offspring of alcoholics: A study of event-related potentials during a monetary gambling task. *Int J Psychophysiol* (2015) 98:182–200. doi: 10.1016/j.jpsycho.2015.09.005
52. Howse AD, Hassall CD, Williams CC, Hajcak G, Krigolson OE. Alcohol hangover impacts learning and reward processing within the medial-frontal cortex. *Psychophysiology* (2018) 55:e13081. doi: 10.1111/psyp.13081
53. Lannoy S, D'Hondt F, Dormal V, Billieux J, Muraire P. Electrophysiological correlates of performance monitoring in binge drinking: Impaired error-related but preserved feedback processing. *Clin Neurophysiol* (2017) 128:2110–21. doi: 10.1016/j.clinph.2017.08.005
54. Kann SJ, O'Rawe JF, Huang AS, Klein DN, Leung H-C. Preschool negative emotionality predicts activity and connectivity of the fusiform face area and amygdala in later childhood. *Soc Cognit Affect Neurosci* (2017) 12:1511–9. doi: 10.1093/scan/nsx079
55. Fox HC, Hong KA, Sinha R. Difficulties in emotion regulation and impulse control in recently abstinent alcoholics compared with social drinkers. *Addict Behav* (2008) 33(2):388–94. doi: 10.1016/j.addbeh.2007.10.002
56. Spada MM, Caselli G, Wells A. Metacognitions as a predictor of drinking status and level of alcohol use following CBT in problem drinkers: A prospective study. *Behav Res Ther* (2009) 47(10):882–6. doi: 10.1016/j.brat.2009.06.010
57. Koob G, Le Moal M. Drug Addiction, Dysregulation of Reward, and Allostasis. *Neuropsychopharmacology* (2001) 24:97–129. doi: 10.1016/S0893-133X(00)00195-0
58. Heilig M, Egli M, Crabbe JC, Becker HC. Acute withdrawal, protracted abstinence and negative affect in alcoholism: are they linked? *Addict Biol* (2010) 15:169–84. doi: 10.1111/j.1369-1600.2009.00194.x
59. Kornreich C. Impaired emotional facial expression recognition is associated with interpersonal problems in alcoholism. *Alcohol Alcohol* (2002) 37:394–400. doi: 10.1093/alcal/37.4.394
60. Hagan MJ, Luecken LJ, Modecki KL, Sandler IN, Wolchik SA. Childhood negative emotionality predicts biobehavioral dysregulation fifteen years later. *Emotion* (2016) 16:877–85. doi: 10.1037/emo0000161
61. Lopez-Vergara HI, Spillane NS, Merrill JE, Jackson KM. Developmental trends in alcohol use initiation and escalation from early to middle adolescence: Prediction by urgency and trait affect. *Psychol Addict Behav* (2016) 30:578–87. doi: 10.1037/adb0000173
62. Schupp HT, Flaisch T, Stockburger J, Junghofer M. Emotion and attention: event-related brain potential studies. *Prog Brain Res* (2006) 156:31–51. doi: 10.1016/s0079-6123(06)56002-9
63. Zhang J, Zhou R. Individual differences in automatic emotion regulation affect the asymmetry of the LPP component. *PLoS One* (2014) 9:e88261. doi: 10.1371/journal.pone.0088261
64. Matsuda I, Nittono H. Motivational significance and cognitive effort elicit different late positive potentials. *Clin Neurophysiol* (2015) 126:304–13. doi: 10.1016/j.clinph.2014.05.030
65. Feng C, Li W, Tian T, Luo Y, Gu R, Zhou C, et al. Arousal modulates valence effects on both early and late stages of affective picture processing in a passive viewing task. *Soc Neurosci* (2014) 9:364–77. doi: 10.1080/17470919.2014.896827
66. Yang B, Cao J, Zhou T, Dong L, Zou L, Xiang J. Exploration of Neural Activity under Cognitive Reappraisal Using Simultaneous EEG-fMRI Data and Kernel Canonical Correlation Analysis. *Comput Math Methods Med* (2018) 2018:1–11. doi: 10.1155/2018/3018356
67. Hajcak G, MacNamara A, Olvet DM. Event-Related Potentials, Emotion, and Emotion Regulation: An Integrative Review. *Dev Neuropsychol* (2010) 35:129–55. doi: 10.1080/87565640903526504
68. Hajcak G, Moser JS, Simons RF. Attending to affect: Appraisal strategies modulate the electrocortical response to arousing pictures. *Emotion* (2006) 6(3):517–22. doi: 10.1037/1528-3542.6.3.517
69. Franken IH, Nijs IM, Muris P, Van Strien JW. Alcohol selectively reduces brain activity during the affective processing of negative information. *Alcohol Clin Exp Res* (2007) 31:919–27. doi: 10.1111/j.1530-0277.2007.00424.x
70. Connell AM, Patton E, McKillop H. Binge drinking, depression, and electrocortical responses to emotional images. *Psychol Addict Behav* (2015) 29(3):673–82. doi: 10.1037/adb0000071
71. Muraire P, Campanella S, Philippot P, de Timary P, Constant E, Gauthier S, et al. Alcoholism leads to early perceptive alterations, independently of comorbid depressed state: an ERP study. *Neurophysiol Clin Neurophysiol* (2008) 38:83–7053. doi: 10.1016/j.neucli.2008.02.001
72. Foisy M, Kornreich C, Fobe A, D'Hondt L, Pelc I, Hanak C, et al. Impaired emotional facial expression recognition in alcohol dependence: do these deficits persist with mid-term abstinence? *Alcohol Clin Exp Res* (2007) 31:404–6008. doi: 10.1111/j.1530-0277.2006.00321.x
73. Bickel WK, Jarmolowicz DP, Mueller ET, Gatchalian KM, McClure SM. Are executive function and impulsivity antipodes? A conceptual reconstruction

- with special reference to addiction. *Psychopharmacol (Berl)* (2012) 221:361–87. doi: 10.1007/s00213-012-2689-x
74. Domínguez-Centeno I, Jurado-Barba R, Sion A, Martínez-Maldonado A, Castillo-Parra G, López-Muñoz F, et al. P3 Component as a Potential Endophenotype for Control Inhibition in Offspring of Alcoholics. *Alcohol Alcohol* (2018) 53:699–706. doi: 10.1093/alc/alcy051
 75. Kim H. Involvement of the dorsal and ventral attention networks in oddball stimulus processing: A meta-analysis. *Hum Brain Mapp* (2014) 35:2265–84. doi: 10.1002/hbm.22326
 76. Hada M, Porjesz B, Begleiter H, Polich J. Auditory P3a assessment of male alcoholics. *Biol Psychiatry* (2000) 48:276–86. doi: 10.1016/S0006-3223(00)00236-5
 77. Wan L, Baldridge RM, Colby AM, Stanford MS. Association of P3 amplitude to treatment completion in substance dependent individuals. *Psychiatry Res* (2010) 177:223–7. doi: 10.1016/j.psychres.2009.01.033
 78. Van Der Stelt O, Esbra-Nordmann 1998 award lecture: visual P3 as a potential vulnerability marker of alcoholism: evidence from the Amsterdam Study of Children of Alcoholics. *Alcohol Alcohol* (1999) 34:267–82. doi: 10.1093/alc/alcy34.3.267
 79. Suresh S, Porjesz B, Chorlian DB, Choi K, Jones KA, Wang K, et al. Auditory P3 in Female Alcoholics. *Alcohol Clin Exp Res* (2003) 27:1064–74. doi: 10.1097/01.ALC.0000075549.49800.AO
 80. Realmuto G, Begleiter H, Odencrantz J, Porjesz B. Event-related potential evidence of dysfunction in automatic processing in abstinent alcoholics. *Biol Psychiatry* (1993) 33:594–601. doi: 10.1016/0006-3223(93)90097-W
 81. Branchey MH, Buydens-Branchey L, Lieber CS. P3 in alcoholics with disordered regulation of aggression. *Psychiatry Res* (1988) 25:49–58. doi: 10.1016/0165-1781(88)90157-6
 82. Chen ACH, Porjesz B, Rangaswamy M, Kamarajan C, Tang Y, Jones KA, et al. Reduced Frontal Lobe Activity in Subjects With High Impulsivity and Alcoholism. *Alcohol Clin Exp Res* (2007) 31:156–65. doi: 10.1111/j.1530-0277.2006.00277.x
 83. Fein G, Cardenas VA. P3b amplitude is not reduced in abstinent alcoholics with a current MDD. *Alcohol* (2017) 63:33–42. doi: 10.1016/j.alcohol.2017.03.004
 84. Emmerson RY, Dustman RE, Shearer DE, Chamberlin HM. EEG. visually evoked and event related potentials in young abstinent alcoholics. *Alcohol* (1987) 4:241–8. doi: 10.1016/0741-8329(87)90018-8
 85. Begleiter H, Porjesz B, Rawlings R, Eckardt M. Auditory recovery function and P3 in boys at high risk for alcoholism. *Alcohol* (1987) 4:315–21. doi: 10.1016/0741-8329(87)90029-2
 86. Begleiter H, Porjesz B. What is inherited in the predisposition toward alcoholism? A proposed model. *Alcohol Clin Exp Res* (1999) 23:1125–35. doi: 10.1111/j.1530-0277.1999.tb04269.x
 87. Bauer LO, Costa L, Hesselbrock VM. Effects of alcoholism, anxiety and depression on P300 in women: a pilot study. *J Stud Alcohol* (2001) 62:571–9. doi: 10.15288/jsa.2001.62.571
 88. Malone SM, Iacono WG, McGue M. Event-related potentials and comorbidity in alcohol-dependent adult males. *Psychophysiology* (2001) 38:367–76. doi: 10.1017/S0048577201990912
 89. Cloninger C. Neurogenetic adaptive mechanisms in alcoholism. *Sci (80-)* (1987) 236:410–6. doi: 10.1126/science.2882604
 90. Lombard A, Brittain C, Wishart G, Lowe S, McCarthy A, Landschulz W, et al. Population Pharmacokinetic/ Pharmacodynamic Modelling of Auditory-Evoked Event-Related Potentials with Lorazepam. *Basic Clin Pharmacol Toxicol* (2018) 122:245–52. doi: 10.1111/bcpt.12900
 91. Brown SBRE, van der Wee NJA, van Noorden MS, Giltay EJ, Nieuwenhuis S. Noradrenergic and cholinergic modulation of late ERP responses to deviant stimuli. *Psychophysiology* (2015) 52:1620–31. doi: 10.1111/psyp.12544
 92. De Rover M, Brown SBRE, Band GP, Giltay EJ, Van Noorden MS, Van Der Wee NJA, et al. Beta receptor-mediated modulation of the oddball P3 but not error-related ERP components in humans. *Psychopharmacol (Berl)* (2015) 232:3161–72. doi: 10.1007/s00213-015-3966-2
 93. Isbel BD, Lagopoulos J, Hermens DF, Summers MJ. Mental training affects electrophysiological markers of attention resource allocation in healthy older adults. *Neurosci Lett* (2019) 698:186–91. doi: 10.1016/j.neulet.2019.01.029
 94. MacIin EL, Mathewson KE, Low KA, Boot WR, Kramer AF, Fabiani M, et al. Learning to multitask: Effects of video game practice on electrophysiological indices of attention and resource allocation. *Psychophysiology* (2011) 48:1173–83. doi: 10.1111/j.1469-8986.2011.01189.x
 95. Campanella S, Cimochowska A, Kornreich C, Hanak C, Verbanck P, Petit G. Neuropsychological correlates of response inhibition predict relapse in detoxified alcoholic patients: some preliminary evidence from event-related potentials. *Neuropsychiatr Dis Treat* (2014) 10:1025. doi: 10.2147/NDT.S61475
 96. Kamarajan C, Porjesz B, Jones KA, Choi K, Chorlian DB, Padmanabhapillai A, et al. Alcoholism is a disinhibitory disorder: Neuropsychological evidence from a Go/No-Go task. *Biol Psychol* (2005) 69:353–73. doi: 10.1016/j.biopsycho.2004.08.004
 97. Colrain IM, Sullivan EV, Ford JM, Mathalon DH, McPherson SL, Roach BJ, et al. Frontally mediated inhibitory processing and white matter microstructure: Age and alcoholism effects. *Psychopharmacol (Berl)* (2011) 213:669–79. doi: 10.1007/s00213-010-2073-7
 98. Pandey AK, Kamarajan C, Tang Y, Chorlian DB, Roopesh BN, Manz N, et al. Neurocognitive deficits in male alcoholics: An ERP/sLORETA analysis of the N2 component in an equal probability Go/NoGo task. *Biol Psychol* (2012) 89:170–82. doi: 10.1016/j.biopsycho.2011.10.009
 99. Stein M, Fey W, Koenig T, Oehy J, Moggi F. Context-Specific Inhibition is Related to Craving in Alcohol Use Disorders: A Dangerous Imbalance. *Alcohol Clin Exp Res* (2018) 42:69–80. doi: 10.1111/acer.13532
 100. Kok A. Effects of degradation of visual stimuli on components of the event-related potential (ERP) in go/nogo reaction tasks. *Biol Psychol* (1986) 23:21–38. doi: 10.1016/0301-0511(86)90087-6
 101. Smith JL, Johnstone SJ, Barry RJ. Response priming in the Go/NoGo task: The N2 reflects neither inhibition nor conflict. *Clin Neurophysiol* (2007) 118:343–55. doi: 10.1016/j.clinph.2006.09.027
 102. Herrera-Díaz A, Mendoza-Quinones R, Melie-García L, Martínez-Montes E, Sanabria-Díaz G, Romero-Quintana Y, et al. Functional Connectivity and Quantitative EEG in Women with Alcohol Use Disorders: A Resting-State Study. *Brain Topogr* (2016) 29:368–81. doi: 10.1007/s10548-015-0467-x
 103. Rangaswamy M, Jones KA, Porjesz B, Chorlian DB, Padmanabhapillai A, Kamarajan C, et al. Delta and theta oscillations as risk markers in adolescent offspring of alcoholics. *Int J Psychophysiol* (2007) 63:3–15. doi: 10.1016/j.ijpsycho.2006.10.003
 104. Jones KA, Porjesz B, Chorlian D, Rangaswamy M, Kamarajan C, Padmanabhapillai A, et al. Begleiter H. S-transform time-frequency analysis of P300 reveals deficits in individuals diagnosed with alcoholism. *Clin Neurophysiol* (2006) 117:2128–43. doi: 10.1016/j.clinph.2006.02.028
 105. Rangaswamy M, Porjesz B, Chorlian DB, Choi K, Jones KA, Wang K, et al. Theta power in the EEG of alcoholics. *Alcohol Clin Exp Res* (2003) 27(4):607–15. doi: 10.1097/01.ALC.0000060523.95470.8F
 106. Bauer L. Predicting Relapse to Alcohol and Drug Abuse via Quantitative Electroencephalography. *Neuropsychopharmacology* (2001) 25:332–40. doi: 10.1016/S0893-133X(01)00236-6
 107. Coutin-Churchman P, Moreno R, Añez Y, Vergara F. Clinical correlates of quantitative EEG alterations in alcoholic patients. *Clin Neurophysiol* (2006) 117:740–51. doi: 10.1016/j.clinph.2005.12.021
 108. Huster RJ, Enriquez-Geppert S, Lavalée CF, Falkenstein M, Herrmann CS. Electroencephalography of response inhibition tasks: Functional networks and cognitive contributions. *Int J Psychophysiol* (2013) 87(3):217–33. doi: 10.1016/j.ijpsycho.2012.08.001
 109. Tcheslavski GV, Gonen FF. Alcoholism-related alterations in spectrum, coherence, and phase synchrony of topical electroencephalogram. *Comput Biol Med* (2012) 42:394–401. doi: 10.1016/j.combiomed.2011.12.006
 110. Crespo-García M, Pinal D, Cantero JL, Díaz F, Zurrón M, Atienza M. Working memory processes are mediated by local and long-range synchronization of alpha oscillations. *J Cognit Neurosci* (2013) 27(8):1343–57. doi: 10.1162/jocn_a_00379
 111. Sadaghiani S, Scheeringa R, Lehongre K, Morillon B, Giraud A-L, Kleinschmidt A. Intrinsic connectivity networks, alpha oscillations, and tonic alertness: a simultaneous electroencephalography/functional magnetic resonance imaging study. *J Neurosci* (2010) 30(30):10243–50. doi: 10.1523/JNEUROSCI.1004-10.2010
 112. Porjesz B, Rangaswamy M. Neuropsychological endophenotypes, CNS disinhibition, and risk for alcohol dependence and related disorders. *ScientificWorldJournal* (2007) 7:131–41. doi: 10.1100/tsw.2007.203

113. Porjesz B, Begleiter H. Alcoholism and human electrophysiology. *Alcohol Res Heal* (2003) 27(2):153–60.
114. Rangaswamy M, Porjesz B, Chorlian DB, Wang K, Jones KA, Bauer LO, et al. Beta power in the EEG of alcoholics. *Biol Psychiatry* (2002) 52(8):831–42. doi: 10.1016/S0006-3223(02)01362-8
115. Huang S, Holcomb LA, Cruz SM, Marinkovic K. Altered oscillatory brain dynamics of emotional processing in young binge drinkers. *Cognit Affect Behav Neurosci* (2018) 18:43–57. doi: 10.3758/s13415-017-0551-7
116. Kovacevic S, Azma S, Irimia A, Sherfey J, Halgren E, Marinkovic K. Theta oscillations are sensitive to both early and late conflict processing stages: effects of alcohol intoxication. *PloS One* (2012) 7:e43957. doi: 10.1371/journal.pone.0043957
117. Pandey AK, Kamarajan C, Manz N, Chorlian DB, Stimus A, Porjesz B. Delta, theta, and alpha event-related oscillations in alcoholics during Go/NoGo task: Neurocognitive deficits in execution, inhibition, and attention processing. *Prog Neuropsychopharmacol Biol Psychiatry* (2016) 65:158–71. doi: 10.1016/j.pnpbp.2015.10.002
118. Kamarajan C, Porjesz B, Jones KA, Choi K, Chorlian DB, Padmanabhapillai A, et al. The role of brain oscillations as functional correlates of cognitive systems: a study of frontal inhibitory control in alcoholism. *Int J Psychophysiol* (2004) 51:155–80. doi: 10.1016/j.ijpsycho.2003.09.004
119. Harper J, Malone SM, Iacono WG. Conflict-related medial frontal theta as an endophenotype for alcohol use disorder. *Biol Psychol* (2018) 139:25–38. doi: 10.1016/j.biopsycho.2018.10.002
120. Gilmore CS, Fein G. Theta event-related synchronization is a biomarker for a morbid effect of alcoholism on the brain that may partially resolve with extended abstinence. *Brain Behav* (2012) 2:796–805. doi: 10.1002/brb3.95
121. Cavanagh JF, Frank MJ. Frontal theta as a mechanism for cognitive control. *Trends Cognit Sci* (2014) 18:414–21. doi: 10.1016/j.tics.2014.04.012
122. Aftanas LI, Varlamov AA, Pavlov SV, Makhnev VP, Reva NV. Affective picture processing: Event-related synchronization within individually defined human theta band is modulated by valence dimension. *Neurosci Lett* (2001) 303(2):115–8. doi: 10.1016/S0304-3940(01)01703-7
123. Schulte T, Muller-Oehring EM, Sullivan EV, Pfefferbaum A. Synchrony of corticostriatal-midbrain activation enables normal inhibitory control and conflict processing in recovering alcoholic men. *Biol Psychiatry* (2012) 71:269–78. doi: 10.1016/j.biopsycho.2011.10.022
124. Camchong J, Stenger A, Fein G. Resting-state synchrony during early alcohol abstinence can predict subsequent relapse. *Cereb Cortex* (2013) 23:2086–99. doi: 10.1093/cercor/bhs190
125. Muller-Oehring EM, Jung YC, Pfefferbaum A, Sullivan EV, Schulte T. The Resting Brain of Alcoholics. *Cereb Cortex* (2015) 25:4155–68. doi: 10.1093/cercor/bhu134
126. Sullivan EV, Muller-Oehring E, Pitel AL, Chanraud S, Shankaranarayanan A, Alsop DC, et al. A selective insular perfusion deficit contributes to compromised salience network connectivity in recovering alcoholic men. *Biol Psychiatry* (2013) 74:547–55. doi: 10.1016/j.biopsycho.2013.02.026
127. Fries P. Rhythms for Cognition: Communication through Coherence. *Neuron* (2015) 88:220–35. doi: 10.1016/j.neuron.2015.09.034
128. Kaplan RF, Glueck BC, Hesselbrock MN, Reed HB Jr. Power and coherence analysis of the EEG in hospitalized alcoholics and nonalcoholic controls. *J Stud Alcohol* (1985) 46:122–7. doi: 10.15288/jsa.1985.46.122
129. Coullaut-Valera R, Arbaiza I, Bajo R, Arrue R, Lopez ME, Coullaut-Valera J, et al. Drug polyconsumption is associated with increased synchronization of brain electrical-activity at rest and in a counting task. *Int J Neural Syst* (2014) 24:1450005. doi: 10.1142/s0129065714500051
130. Huang Y, Mohan A, De Ridder D, Sunaert S, Vanneste S. The neural correlates of the unified percept of alcohol-related craving: a fMRI and EEG study. *Sci Rep* (2018) 8:923. doi: 10.1038/s41598-017-18471-y
131. Michael A, Mirza KA, Mukundan CR, Channabasavanna SM. Interhemispheric electroencephalographic coherence as a biological marker in alcoholism. *Acta Psychiatr Scand* (1993) 87:213–7. doi: 10.1111/j.1600-0447.1993.tb03358.x
132. Winterer G, Enoch MA, White KV, Saylan M, Coppola R, Goldman D. EEG phenotype in alcoholism: increased coherence in the depressive subtype. *Acta Psychiatr Scand* (2003) 108:51–60. doi: 10.1034/j.1600-0447.2003.00060.x
133. Demiral SB, Golosheykin S, Anokhin AP. Genetic influences on functional connectivity associated with feedback processing and prediction error: Phase coupling of theta-band oscillations in twins. *Int J Psychophysiol* (2017) 115:133–41. doi: 10.1016/j.ijpsycho.2016.12.013
134. Dimitriadis SI, Kanatsouli K, Laskaris NA, Tsirka V, Vourkas M, Micheloyannis S. Surface EEG shows that functional segregation via phase coupling contributes to the neural substrate of mental calculations. *Brain Cognit* (2012) 80:45–52. doi: 10.1016/j.bandc.2012.04.001
135. Bu J, Ma R, Fan C, Sun S, Cheng Y, Piao Y, et al. Low-Theta Electroencephalography Coherence Predicts Cigarette Craving in Nicotine Addiction. *Front Psychiatry* (2019) 10:296. doi: 10.1016/j.jaac.2019.05.02210.3389/fpsy.2019.00296
136. Martínez-Maldonado A, Jurado-Barba R, Sion A, Domínguez-Centeno I, Castillo-Parra G, Prieto-Montalvo J, et al. Brain functional connectivity after cognitive-bias modification and behavioral changes in abstinent alcohol-use disorder patients. *Int J Psychophysiol* (2019) 154:46–58. doi: 10.1016/j.ijpsycho.2019.10.004
137. Li P, Liu H, Si Y, Li C, Li F, Zhu X, et al. EEG Based Emotion Recognition by Combining Functional Connectivity Network and Local Activations. *IEEE Trans BioMed Eng* (2019) 66:2869–81. doi: 10.1109/TBME.2019.2897651
138. Lopez-Sanz D, Garces P, Alvarez B, Delgado-Losada ML, Lopez-Higes R, Maestu F. Network Disruption in the Preclinical Stages of Alzheimer's Disease: From Subjective Cognitive Decline to Mild Cognitive Impairment. *Int J Neural Syst* (2017) 27:1750041. doi: 10.1142/s0129065717500411
139. Stam CJ, de Haan W, Daffertshofer A, Jones BF, Manshanden I, van Cappellen van Walsum AM, et al. Graph theoretical analysis of magnetoencephalographic functional connectivity in Alzheimer's disease. *Brain* (2009) 132:213–24. doi: 10.1093/brain/awn262
140. Wang Z, Suh J, Li Z, Li Y, Franklin T, O'Brien C, Childress AR. A hyper-connected but less efficient small-world network in the substance-dependent brain. *Drug Alcohol Depend* (2015) 152:102–8. doi: 10.1016/j.drugalcdep.2015.04.015
141. Ahmadi M, Ahmadi K, Rezazade M, Azad-Marzabadi E. Global organization of functional brain connectivity in methamphetamine abusers. *Clin Neurophysiol* (2013) 124:1122–31. doi: 10.1016/j.clinph.2012.12.003
142. Fornito A, Zalesky A, Bullmore ET. *Fundamentals of Brain Network Analysis*. P. A, editor. London, UK: Elsevier (2016).
143. Brandes U, Erlebach T. *Network Analysis: Methodological Foundations*. (2005). (Berlin: Heidelberg)
144. Tononi G, Sporns O, Edelman GM. A measure for brain complexity: relating functional segregation and integration in the nervous system. *Proc Natl Acad Sci U.S.A.* (1994) 91:5033–7. doi: 10.1073/pnas.91.11.5033
145. Van Dijk KA, Drzezga A. (2014). "The Default Network of the Brain." in *PET and SPECT in Neurology*. (Berlin, Heidelberg: Springer Berlin Heidelberg). 169–181. doi: 10.1007/978-3-642-54307-4_8
146. Latora V, Marchiori M. Efficient behavior of small-world networks. *Phys Rev Lett* (2001) 87:198701. doi: 10.1103/PhysRevLett.87.198701
147. Tomasi D, Volkow ND. Functional connectivity hubs in the human brain. *Neuroimage* (2011) 57:908–17. doi: 10.1016/j.biopsycho.2013.02.02610.1016/j.neuroimage.2011.05.024
148. Sjoerds Z, Stufflebeam SM, Veltman DJ, Van den Brink W, Penninx BW, Douw L. Loss of brain graph network efficiency in alcohol dependence. *Addict Biol* (2017) 22:523–34. doi: 10.1111/adb.12346
149. Cao R, Wu Z, Li H, Xiang J, Chen J. Disturbed connectivity of EEG functional networks in alcoholism: a graph-theoretic analysis. *BioMed Mater Eng* (2014) 24:2927–36. doi: 10.3233/BME-141112
150. Kopera M, Jakubczyk A, Suszek H, Glass JM, Klimkiewicz A, Wnorowska A, et al. Relationship Between Emotional Processing, Drinking Severity and Relapse in Adults Treated for Alcohol Dependence in Poland. *Alcohol Alcohol* (2017) 52:311–1. doi: 10.1093/alcac/alg010

151. Moos RH, Moos BS. Rates and predictors of relapse after natural and treated remission from alcohol use disorders. *Addiction* (2006) 101(2):212–22. doi: 10.1111/j.1360-0443.2006.01310.x
152. Polich J. Updating P300: an integrative theory of P3a and P3b. *Clin Neurophysiol* (2007) 118:2128–48. doi: 10.1016/j.clinph.2007.04.019
153. Brion M, D'Hondt F, Pitel A-L, Lecomte B, Ferauge M, de Timary P, et al. Executive functions in alcohol-dependence: A theoretically grounded and integrative exploration. *Drug Alcohol Depend* (2017) 177:39–47. doi: 10.1016/j.drugalcdep.2017.03.018
154. Wiers CE, Stelzel C, Gladwin TE, Park SQ, Pawelczack S, Gawron CK, et al. Effects of cognitivebias modification training on neural alcohol cue reactivity in alcohol dependence. *Am J Psychiatry* (2015) 172(4):335–43. doi: 10.1176/appi.ajp.2014.13111495
155. Houston RJ, Schliez NJ. Event-Related Potentials as Biomarkers of Behavior Change Mechanisms in Substance Use Disorder Treatment. *Biol Psychiatry Cognit Neurosci Neuroimaging* (2018) 3:30–40. doi: 10.1016/j.bpsc.2017.09.006
156. Campanella S, Schroder E, Kajosch H, Hanak C, Veaser J, Amiot M, et al. Neurophysiological markers of cue reactivity and inhibition subtend a three-month period of complete alcohol abstinence. *Clin Neurophysiol* (2020) 131(2):555–65. doi: 10.1016/j.clinph.2019.10.020
157. Verdejo-García A, Alcázar-Córcoles MA, Albein-Urios N. Neuropsychological Interventions for Decision-Making in Addiction: a Systematic Review. *Neuropsychol Rev* (2019) 29(1):79–92. doi: 10.1007/s11065-018-9384-6
158. de Bruin EA, Stam CJ, Bijl S, Verbaten MN, Kenemans JL. Moderate-to-heavy alcohol intake is associated with differences in synchronization of brain activity during rest and mental rehearsal. *Int J Psychophysiol* (2006) 60:304–14. doi: 10.1016/j.ijpsycho.2005.07.007

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Predicting Treatment Outcome in Major Depressive Disorder Using Serotonin 4 Receptor PET Brain Imaging, Functional MRI, Cognitive-, EEG-Based, and Peripheral Biomarkers: A NeuroPharm Open Label Clinical Trial Protocol

OPEN ACCESS

Edited by:

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Specialty section:

This article was submitted to
Molecular Psychiatry,
a section of the journal
Frontiers in Psychiatry

Received: 27 November 2019

Accepted: 19 June 2020

Published: 23 July 2020

Citation:

Köhler-Forsberg K, Jørgensen A,
Dam VH, Stenbæk DS, Fisher PM,
Ip C-T, Ganz M, Poulsen HE, Giraldi A,
Ozenne B, Jørgensen MB,
Knudsen GM and Frokjaer VG (2020)
Predicting Treatment Outcome in
Major Depressive Disorder Using
Serotonin 4 Receptor PET Brain
Imaging, Functional MRI, Cognitive-,
EEG-Based, and Peripheral
Biomarkers: A NeuroPharm Open
Label Clinical Trial Protocol.
Front. Psychiatry 11:641.
doi: 10.3389/fpsy.2020.00641

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Background: Between 30 and 50% of patients with major depressive disorder (MDD) do not respond sufficiently to antidepressant regimens. The conventional pharmacological treatments predominantly target serotonergic brain signaling but better tools to predict treatment response and identify relevant subgroups of MDD are needed to support individualized and mechanistically targeted treatment strategies. The aim of this study is to investigate antidepressant-free patients with MDD using neuroimaging, electrophysiological, molecular, cognitive, and clinical examinations and evaluate their ability to predict clinical response to SSRI treatment as individual or combined predictors.

Methods: We will include 100 untreated patients with moderate to severe depression (>17 on the Hamilton Depression Rating Scale 17) in a non-randomized open clinical trial. We will collect data from serotonin 4 receptor positron emission tomography (PET) brain scans, functional magnetic resonance imaging (fMRI), electroencephalogram (EEG), cognitive tests, psychometry, and peripheral biomarkers, before (at baseline), during, and after 12 weeks of standard antidepressant treatment. Patients will be treated with escitalopram, and in case of non-response at week 4 or intolerable side effects, offered to switch to a second line treatment with duloxetine. Our primary outcome (treatment

response) is assessed using the Hamilton depression rating subscale 6-item scores at week 8, compared to baseline. In a subset of the patients ($n = \sim 40$), we will re-assess the neurobiological response (using PET, fMRI, and EEG) 8 weeks after initiated pharmacological antidepressant treatment, to map neurobiological signatures of treatment responses. Data from matched controls will either be collected or is already available from other cohorts.

Discussion: The extensive investigational program with follow-up in this large cohort of participants provides a unique possibility to (a) uncover potential biomarkers for antidepressant treatment response, (b) apply the findings for future stratification of MDD, (c) advance the understanding of pathophysiological underpinnings of MDD, and (d) uncover how putative biomarkers change in response to 8 weeks of pharmacological antidepressant treatment. Our data can pave the way for a precision medicine approach for optimized treatment of MDD and also provides a resource for future research and data sharing.

Clinical Trial Registration: The study was registered at clinicaltrials.gov prior to initiation (NCT02869035; 08.16.2016, URL: <https://clinicaltrials.gov/ct2/results?cond=&term=NCT02869035&cntry=&state=&city=&dist=>)

Keywords: major depressive disorder, biomarker, treatment response, serotonin 4 receptor, positron emission tomography, functional magnetic resonance imaging, electroencephalogram, cognition

INTRODUCTION

Major Depressive Disorder (MDD) is one of the most severe and common brain disorders worldwide with a huge impact on life quality and socioeconomic status (1, 2). It has been linked to serotonergic dysfunction, cognitive disturbances, brain network dysfunction, vulnerability to stress, neuro-inflammation, and gene by environment factors. Still, the understanding of the pathogenesis remains limited. Guidelines for MDD treatment selection are still predominantly based on simple clinical observations about overall MDD severity, and in the case of recurrent depressive episodes, it is also based on personal patient history of treatment responses. Conventional medical treatment is mainly based on intervention of the monoaminergic system in the brain, in particular the serotonin (5-HT) system. Selective serotonin reuptake inhibitors (SSRIs) act through blockage and subsequent downregulation of the serotonin transporter (SERT) (3), which presumably induces increased extracellular 5-HT levels. However, robust evidence for a central 5-HT hypofunction in patients with MDD *in vivo* is lacking (4). Roughly one third of patients suffering from MDD do not respond sufficiently to 5-HT acting drugs (5, 6), suggesting a

diverse pathophysiology. The diagnostic criteria for MDD may cover a heterogeneous collection of various biological entities and consequently, it is not surprising that a “one size fits all” treatment strategy is suboptimal (7). Currently, the time from starting to administer a potentially efficacious drug until it can be determined if the clinical response is satisfactory is, at best, 4–6 weeks. In clinical practice, the lack of convenient and accurate tools (e.g., quantitative and/or biological) to predict treatment response prolongs the delay from diagnosis to effective treatment and constitutes a major challenge for both clinicians and patients. Therefore, stratification of subtypes and a shift toward precision medicine, e.g., through identification of predictors of treatment response, so-called biomarker(s) that can help optimize treatment choice, is of paramount importance. Candidate biomarkers could be related to neurotransmission, specific neural networks or structural alterations in specific brain regions that can be detected by brain imaging modalities such as positron emission tomography (PET), functional magnetic resonance imaging (fMRI), and electroencephalogram (EEG) or altered biophysiological or cognitive functions (4, 8). It has also been suggested that rather than a single biomarker, an algorithm involving a set of biomarkers may prove useful to subgroup patients and predict their response to certain treatment strategies in MDD (9). Several biomarkers derived from prior large studies such as iSPOT-D, EMBARC, and CANBIND for prediction of drug response in MDD have been proposed (10–12). Here, we use multimodalities (PET, fMRI, and EEG, cognitive testing, psychometrics, and peripheral biomarkers) as part of a deep phenotyping and as a unique feature to our trial, we study modes of action in the brain on a neurotransmitter level. Thus, our trial contributes with novel insights as well as

Abbreviations: MDD, Major Depressive Disorder; 5-HT: 5-hydroxytryptamine (Serotonin); SSRI, Selective serotonin reuptake inhibitor; SERT, Serotonin transporter; PET, Positron emission tomography; fMRI, Functional magnetic resonance imaging; EEG, Electroencephalogram; 5-HT₄R, Serotonin 4 receptor; rs-fMRI, Resting state functional magnetic resonance imaging; ERP, Event-related potential; LDAEP, Loudness dependence of auditory evoked potentials; CAR, Cortisol awakening response; HPA-axis, Hypothalamic-pituitary-adrenal axis; HAMD₁₇, 17-item Hamilton Depression Rating Scale; HAMD₆, 6-item Hamilton Depression Rating Scale.

provide a dataset for cross-validation of other identified predictors of psychopharmacological antidepressant treatment response. In a non-randomized, longitudinal, open clinical trial, patients with moderate to severe depression will be treated with SSRI following Danish guidelines. In order to map neurobiological signatures of treatment, we will re-examine a subset of the cohort with neuroimaging and EEG after 8 weeks of SSRI treatment and assess cognitive changes after 12 weeks. This clinical trial is part of a larger research initiative, “NeuroPharm”, which addresses pertinent and basic questions regarding human brain disease mechanisms and seeks to predict brain responses to categories of neuro-modulatory interventions as well as treatment efficacy (www.np.nru.dk). We anticipate that this study will critically advance and inform future stratification strategies, further uncover pathophysiological and treatment mechanisms and, hopefully, guide future precision medicine approaches to optimize treatment strategies for patients suffering from MDD.

Imaging techniques have vastly increased our understanding of the underpinning cerebral mechanisms involved in MDD (13). Serotonergic dysfunction is considered a central mechanism in depression, and a recent review points at the 5-HT₄ receptor (5-HT₄R) as highly implicated in MDD (14). For example, 5-HT₄R agonism has shown rapid antidepressant-like behavioral effects in rodents (15), and experimental models suggest that cerebral 5-HT₄R levels are sensitive to central 5-HT modulation in rodents (16, 17). Subsequent clinical studies from our group demonstrated that cerebral 5-HT levels can be indexed in an inverse manner through molecular brain imaging of the 5-HT₄R by using the PET-ligand 11C-SB207145 *in vivo* (18). We here aim to evaluate 5-HT₄R binding as a candidate predictor of antidepressant response to drugs targeting the 5-HT system in the hitherto largest cohort of MDD patients with PET brain imaging of serotonergic markers. We hypothesize that 1) patients with MDD differ in cerebral [¹¹C]SB207145 binding at baseline compared to healthy controls; 2) [¹¹C]SB207145 binding at baseline in patients with MDD predicts remission after 8 weeks of pharmacological serotonergic intervention; 3) After 8 weeks of serotonergic intervention, patients with remitter status have a significantly greater reduction in cerebral [¹¹C]SB207145 binding than non-responders. For an overview of primary hypotheses for other modalities, see **Appendix 1**.

fMRI can be used to assess regional activity and resting state functional networks in MDD. One systematic review found abnormal (negative bias) reactivity in amygdala responsiveness to facial expressions and emotional stimulation in patients with MDD versus healthy controls (19), and pre-treatment low amygdala reactivity has shown to be predictive for antidepressant treatment response (20). A study with 70 patients with MDD was able to predict treatment recovery with ~80%, by investigating amygdala reactivity to facial emotions and its interaction with history of early life stress (21). Another study from our group showed that amygdala reactivity was associated with brain 5-HT₄R binding and hence putatively extra synaptic 5-HT levels in healthy individuals. This established a plausible connection between 5-HT levels and amygdala activation, both involved in emotional cognitive processes (22). This exemplifies how a multimodal PET and fMRI

strategy can highlight molecular mechanisms mediating drug effects on brain function (23). Resting state fMRI (rs-fMRI) measures fluctuations in fMRI signal during the absence of an explicit task and is widely used to assess distributed intrinsic networks such as the “default mode network” (24). Alterations in rs-fMRI connectivity have been described in MDD (25) and a recent study suggested that rs-fMRI can define subtypes of MDD and predict antidepressant treatment response (26), but this has been contested by others (27). Although promising, brain imaging studies have in general been inconclusive and with small sample sizes (9, 28). In the current trial, we will use task-based and rs-fMRI in a large cohort of patients with MDD and investigate the association between 5-HT₄R levels (as a proxy for brain serotonin levels) and the clinical outcome of SSRI treatment.

EEG, a monitoring technique for direct ongoing neural activity, has been reported to be associated with treatment response in MDD [see, e.g., review (29)]. Prior studies have found that treatment responders have higher cortical alpha activity (30) and higher theta activity at rostral anterior cingulate cortex compared to treatment non-responders (31, 32). Of note, these biomarkers were derived from the resting EEG data, which is relatively easy to implement in the clinic. Furthermore, earlier evidence from event-related-potential (ERP) studies have suggested that ERP biomarkers such as auditory P300 (a positive waveform around 300 ms after stimulus onset) and loudness-dependence of auditory evoked potentials (LDAEP) can be predict drug treatment response (33, 34), and are linked to the serotonergic transmitter system (35). In the current trial, the predictive values of pretreatment EEG/ERP biomarkers will be examined.

Disturbances in cognitive processes including memory, attention, and executive functions are commonly reported in MDD (36) and contribute to psychosocial impairment and workforce disability (37). In addition, affective bias in information processing (i.e., favoring negative information over positive information at different levels of information processing) has been proposed as a central mechanism in the development and maintenance of depressive symptoms (38) which is also predictive of later treatment response to antidepressant drugs (39). Notably, cognitive disturbances do not always resolve with the remission of a depressive episode, suggesting a dissociation between core mood and cognitive symptoms in MDD (40). Combined with the low cost and relative ease of testing in a clinical setting, this distinguishes cognitive disturbances as a promising marker for stratification of depression subtypes as well as an important target for antidepressant treatment. In the present study, we therefore aim to map a broad range of cognitive disturbances in MDD, including both cold (non-emotional) and hot (emotional) cognitive processes, and explore whether they may be used to characterize clinically relevant subgroups in MDD. Based on earlier observations in healthy individuals, we expect memory performance to map onto hippocampal 5-HT₄R availability (41) and possibly affective bias in verbal memory in MDD (42).

Evidence of inflammation-associated MDD has emerged over the years (43). Patients with MDD show elevated levels of inflammatory markers in peripheral blood (44) which may

affect treatment response such that higher levels are associated with worse response (45). It has also been suggested that patients with MDD have higher levels of activated microglia, as illuminated with PET (46). Proinflammatory cytokines may influence the 5-HT homeostasis in the brain by acutely upregulate SERT through intercellular pathways (i.e., linked to p38 mitogen-activated protein kinase) and presumably thereby reduce synaptic 5-HT levels (47). Interestingly, cognitive dysfunction, a prevalent symptom in depression, also appear to be linked to an inflammatory response (48). We here aim to determine if higher levels of systemic inflammatory markers are associated with 5-HT₄R brain binding, depression status at baseline and clinical treatment response.

Another area of interest is the association between MDD and signatures of early aging. There is an increased mortality and prevalence of age-related diseases in recurrent depression (49, 50). Oxidative stress on nucleic acids is a general element of aging and has been suggested to be an underlying biological mechanism of the accelerated aging observed in depression (51). Previous research from our group has found evidence for such a link, both in studies of psychological/biological stress and oxidative stress in patients and in rodent models of depression (52–54). Earlier findings indicate alterations in levels of oxidative stress during antidepressant treatment and it is hypothesized that treatment response is related to a transient increase in oxidative stress levels, perhaps due to neurotrophic processes and/or peripheral changes in energy metabolism (55–57). Urinary 8-oxodG and 8-oxoGuo are sensitive and specific markers for systemic DNA/RNA damage from oxidation (58). We here aim to investigate urinary 8-oxodG and 8-oxoGuo as a predictive biomarker for antidepressant treatment response, its association with changes in psychopathology, structural and functional brain changes, and markers of psychological and biological stress. Additionally, we will investigate whether hormonal [estradiol, testosterone, progesterone and follicle-stimulating hormone (in females)] and metabolic status can predict antidepressant treatment response and explore whether these associations are related to genetic make-up (specified below), psychopathology and the occurrence of early life stress using self-reported childhood adverse events and parental bonding quality questionnaires, which also may interact with the 5-HT system (59, 60).

Sexual dysfunction (e.g., low sexual desire, arousal difficulties, and anorgasmia) is a prominent feature of MDD, which often leads to a decline in quality of life (61, 62). Lack of interest to what is usually pleasurable, i.e., anhedonia, is a core symptom in MDD and may also be reflected in reduced sexual desire/interest. Sexual dysfunction, in particular anorgasmia and sexual arousal difficulties may further be linked to serotonergic dysfunctions (63) as seen in MDD. As a complicating factor, impaired sexual health related to MDD may worsen with antidepressant treatments targeting the 5-HT system. For example, in a group of 704 patients with MDD treated with an antidepressant drug (SSRI) or serotonin-norepinephrine reuptake inhibitor, about half of them developed or experienced worsening in their decreased sexual desire as a side-effect, which was also

associated with reduced quality of life, lower self-esteem and adverse effects on mood and partner relations (62). We currently do not know which patient characteristics predict sexual dysfunction in response to SSRI treatment. However, differences in individual serotonergic brain architecture and/or serotonergic response to antidepressant treatment (e.g., SSRI) may play a role. In this study, we aim to map the frequency and predictors of SSRI induced sexual dysfunction and determine if serotonergic tone (measured by 5-HT₄R PET binding) pre-treatment, or changes in response to SSRI treatment, is associated with sexual desire and/or development of SSRI-induced sexual dysfunction in MDD.

Previous findings from our group have repeatedly demonstrated a coupling between key features of the 5-HT system and hypothalamus- pituitary- adrenal axis (HPA-axis), which regulates the release of the stress-hormone cortisol (64). Such HPA-axis dynamics can be measured by the cortisol awakening response. Our results support that both serotonin transporter availability (65), and serotonergic tone or direct capacity for 5-HT₄R agonism (64) support a healthy cortisol response to HPA-axis stimuli. A well-functioning and dynamic HPA-axis is critical for coping with everyday life stressors, and HPA-axis dysregulation is a prominent feature of MDD. Although heightened CAR is associated with relapse of depressive episodes in patients with a history of depression (66), in the more advanced depressed stages, i.e., chronic depression, HPA-axis dynamics are blunted as opposed to recent-onset depression (67). Notably, normalization of the HPA-axis in response to SSRI treatment appears to protect against relapse (68). Thus, the SSRI treatment response is likely to depend on restoring HPA-axis dynamics at least in a subgroup of MDD patients. In this trial, we will assess CAR in patients with MDD, the effect of SSRIs on CAR, investigate its association with baseline 5-HT₄R distribution, as well as evaluate CAR as a predictor of antidepressant treatment outcome.

MATERIALS AND METHODS

Figure 1 shows a flowchart of the scheduled data collection over the 12 weeks of pharmacological drug treatment of patients with MDD. Healthy controls (HC) will be recruited as specified below. Patients will be examined before (at baseline; week 0) and after 1, 2, 4, 8 and 12 weeks of SSRI treatment has been initiated. Depression-severity will be monitored by the Hamilton Depression Rating scale 17 items (HAMD₁₇) and its subscale of 6 items (HAMD₆) (69). A subset of patients will be offered re-examination with PET, fMRI, and EEG after 8 weeks of treatment, to assess changes from baseline and its association to treatment response. Patients from the whole spectrum of treatment responses (from poor to excellent) will be invited in a continuous fashion for this part of the study until allotted re-examinations are completed. All patients will also repeat cognitive testing at week 12. The power analysis in preparation of the study was primarily anchored to the PET modality. We estimated that we needed to include 100 patients to reach a

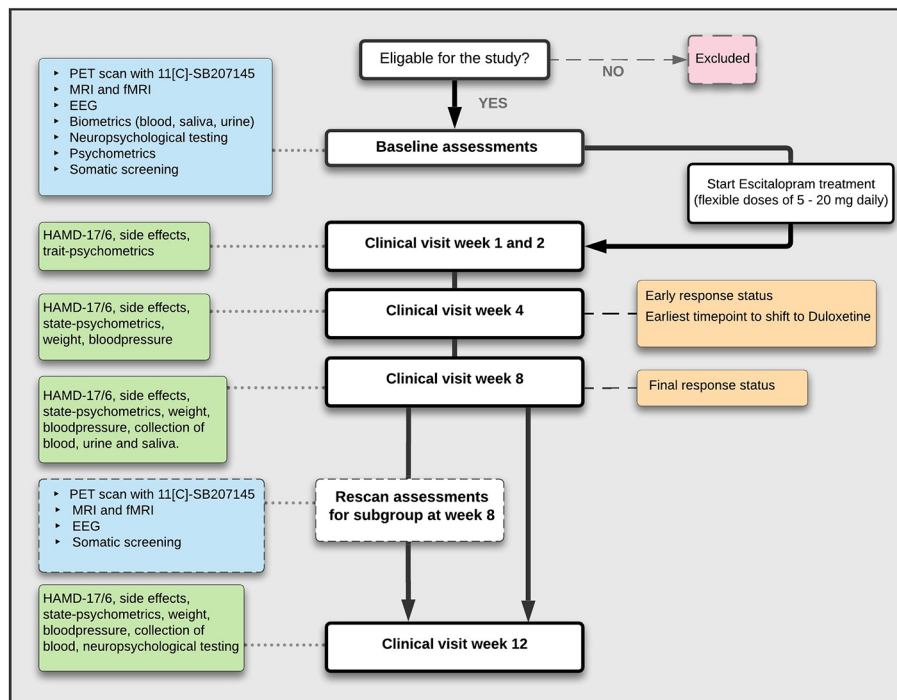


FIGURE 1 | Flowchart of study trial assessments for patients with MDD.

statistical power of 80% to detect an association between treatment response (binary classification, i.e., remitters vs non-responders, see response-definition below) and baseline 5-HT₄R non-displaceable binding potential (BP_{ND}). These calculations were based on an expected 20% maximum drop-out, ~50% remission rate after 8 weeks of treatment (5, 6) and an expected difference of 8% in 5-HT₄R binding between remitters and non-responders, corresponding to the previously found effect sizes on 5-HT₄R change in BP_{ND} after fluoxetine treatment (18). Calculations were further based on an average BP_{ND} of 0.71 and a standard deviation of 0.073 (18, 70). With a rescan subgroup of approximately 40 patients, and a Gaussian distribution of change in BP_{ND} with an SD of 0.08 (log scale), we had an expected power of 80% to identify a significant association between longitudinal changes in BP_{ND} and changes in HAMD₆ (i.e., secondary clinical outcome, continuous scale).

Participants

Patients are recruited from a central referral center within the mental health services in the Capital region of Denmark or directly referred from one of five general practitioners in collaborations with the study group (see **Figure 2** (CONSORT) for details). Data from healthy controls for the purpose of baseline comparisons to patients with MDD are available from a pre-existing database on site (71). The healthy control reference population will be supplemented with newly recruited healthy controls from a local volunteer database (www.nru.dk), as necessary.

Inclusion Criteria for Patients

Patients between 18 and 65 years of age with a moderate to severe, single, or recurrent episode of MDD consistent with the Diagnostic and Statistical Manual of Mental Disorders -5 (DSM-5) and International Statistical Classification of Diseases and Related Health Problems -10 (ICD-10) criteria will be recruited by a trained clinician. Inclusion requires a total score of >17 on HAMD₁₇ at baseline and the diagnose is confirmed by using the diagnostic tool Mini International Neuropsychiatric Interview (72). In addition, all patients are diagnostically verified by a specialist in psychiatry before final inclusion.

Exclusion Criteria for Patients

Patients with a duration of their present depressive episode exceeding two years are not included. No more than one antidepressant treatment attempt in the current episode prior to inclusion is allowed and only patients with no antidepressant medication within the last two months are eligible. Patients with known contraindications or previous non-response to an SSRI drug after an adequate trial as well as a prior or present history of other primary axis I psychiatric disorders are not included, i.e., MDD must be the primary diagnosis. Other exclusion criteria are: severe somatic illness; substance or alcohol use disorder; insufficient language skills to undergo clinical assessments; acute suicidal ideation or psychosis; patients who are deemed by a psychiatrist to require other forms of antidepressant treatments; pregnancy or breast feeding; use of any CNS drug that cannot be washed out prior

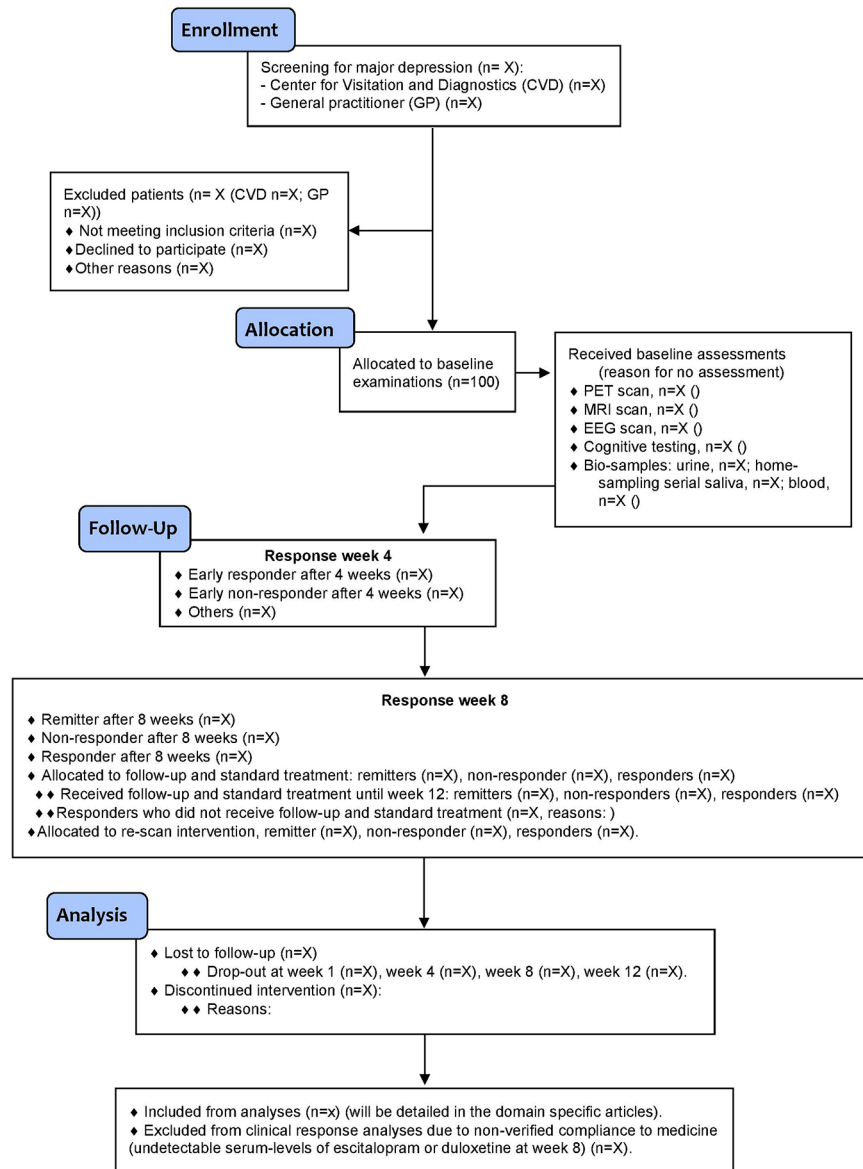


FIGURE 2 | Flow diagram (CONSORT) of the NeuroPharm trial.

to participation (e.g., metoclopramide, ondansetron, serotonergic migraine medicine, clonidine); medical conditions interfering with measurements, contraindications for PET and/or MRI scans; exposure to radioactivity >10 mSv within the last year; severe sensory or intellectual impediments interfering with comprehension of procedures or assessments and lastly any history of brain injury (i.e., loss of consciousness and amnesia or symptoms of concussion disorder).

Inclusion and Exclusion Criteria for Healthy Controls

Enrolled HC will be sought to match the patient population by gender and age distribution. All HC will be screened for MDD using a self-reporting questionnaire (major depression

inventory) (73). The HC meet the same inclusion and exclusion criteria as required for patients apart from psychiatry related issues (e.g., no current or history of mental illness or unstable somatic condition).

Treatment and Investigation Program

Baseline Assessments Before Treatment

Each patient will receive a basic physical screening including somatic status, routine blood samples, electrocardiogram including QTc interval and collection of toxicology urine tests [The Rapid Response™ Multi-Drug Test Panel (Urine)] for detection of drug abuse within the last month. Women are screened for pregnancy through self-reported use of contraceptives and a pregnancy urinary

test if relevant. All study-participants will undergo baseline assessments of brain imaging with ^{11}C -SB207145 PET and fMRI; EEG-examination; cognitive testing, collection of questionnaires and biological material (venous blood, urine, and saliva) as specified below. All HC will receive corresponding baseline assessments.

Clinical Procedure After Treatment Initiation

After completion of baseline examinations, patients will receive flexible doses of the SSRI drug escitalopram, initially 5 mg for 3–5 days depending on side effects (e.g., nausea), followed by 10 mg daily until their first follow-up visit and further adjusted individually to a maximum dose of 20 mg. Escitalopram was chosen as it binds with high selectivity to the 5-HT transporter and has minimal affinity for other receptors (74). Patients are allowed short-time treatment with cyclopyrrolone (a nonbenzodiazepine hypnotic agent) or oxazepam (a benzodiazepine) to reduce anxiety and sleep disturbances which may be prominent in the initial treatment phase and have shown not to influence treatment continuation (75), but all are requested to avoid use 3 days prior to brain scans. Clinical follow-up sessions with a study physician or trained research assistant are scheduled in an out-patient clinical setting after 1, 2, 4, 8 and 12 weeks of treatment to evaluate treatment response and side effects. Visits can deviate a maximum of one week from the original time scheduled. No cognitive behavioral therapy or other psychotherapy program is provided during clinical visits. No treatment (pharmacological or psychotherapeutically) other than the medical monotherapy provided in this study is allowed elsewhere during the trial. At week 4, early non-responders (see definition below) or patients with unacceptable side effects are offered to switch to a standard second line antidepressant treatment; duloxetine (individually adjusted doses of 30–120 mg per day), which is a serotonin-norepinephrine reuptake inhibitor. Duloxetine was chosen according to clinical guidelines for second line antidepressant treatments. Cerebral 5-HT₄R binding in humans is unaltered by injection of escitalopram (76). No prior *in vivo* studies have investigated the effect of duloxetine on 5-HT₄R binding in the human brain, but *in vitro* work has shown that duloxetine has negligible affinity for the 5-HT₄R (77). That is, none of the pharmacological compounds directly target 5-HT₄R's. The week 4 timepoint is in line with national guidelines in Denmark for switching to a second-line antidepressant treatment (4–6 weeks). Since our cohort receives frequent clinical follow-up sessions, patients can reach max dose of escitalopram (20 mg daily) already after 2 weeks. As such, switching after 4 weeks is considered appropriate for early non-responders in this trial set-up. All antidepressant medicine will be provided for free to improve compliance. Compliance will be assessed by serum escitalopram/duloxetine blood tests after eight weeks of treatment as well as tablet count at each follow-up. At each visit, depressive symptoms are rated using the HAMD₁₇ and the HAMD₆ subscale. HAMD₆ captures core symptoms of depression more directly (and disregards sleeping quality), and has been found to be sensitive to antidepressant treatment response (69). Potential side effects due to intervention will be monitored at each visit using the “Udvalg for

Kliniske Undersøgelser” scale (78). To ensure agreement and allow alignment of ratings, HAMD₁₇/HAMD₆ co-ratings between all the clinical investigators will be performed regularly during data collection. A maximum of 20% deviation from the “gold-standard” chief psychiatrist is allowed, or else a new satisfactory co-rating is needed before independent rating of study participants.

Clinical Response Status

Primary Clinical Outcome Measure. The primary outcome measure is categorical and built to capture patients with an early as well as sustained, either excellent or poor response to treatment. Patients are classified as either “remitters”, “non-responders”, or “intermediate responders” after 8 weeks of treatment. These categories are based on percentage changes of depressive symptoms from baseline, as measured by HAMD₆. Remitters must have $\geq 50\%$ reduction in HAMD₆ at 4 weeks (early responders) and a HAMD₆ score < 5 after 8 weeks of treatment. Non-responders have $< 25\%$ reduction in HAMD₆ after 4 weeks (early non-responder) and $< 50\%$ reduction in HAMD₆ after 8 weeks of treatment. Patients who do not meet the criteria above are defined as “intermediate responders” at week 8. The primary predictor analyses are directed to predict treatment response in a binary fashion (either remitter or non-responder (see **Figure 3**).

Secondary Clinical Outcome Measure. As a secondary outcome, we use a continuous response measure, i.e., HAMD₆ changes from baseline at week 8 divided by HAMD₆ at baseline, to allow analyses of the association between antidepressant treatment response and baseline characteristics or treatment-induced changes in the neurobiological modalities of interest.

Examination Modalities

PET Imaging and Quantification of 5-HT₄R Brain Binding

PET scans are conducted using a high-resolution research tomography Siemens PET scanner (CTI/Siemens, Knoxville, TN, USA) ($256 \times 256 \times 207$ voxels; $1.22 \times 1.22 \times 1.22$ mm). Participants are positioned uniformly in spine position and a specialized head holder is applied to reduce head motion during the scan. All participants undergo a 6 min transmission scan and are given an intravenous bolus of approximately 600 MBq of the PET tracer ligand [^{11}C]SB207145. The bolus is administered over 20 s followed by a 120-min dynamic PET data acquisition. The radioligand is synthesized immediately prior to injection as described elsewhere (79).

Preprocessing and PET Quantification

The 120 min dynamic PET acquisitions are reconstructed into 38 time frames (6×5 s, 10×15 s, 4×30 s, 5×2 min, 5×5 min, and 8×10 min) using a 3D-OSEM PSF algorithm (16 subsets and 10 iterations) (80) and TVTV-based attenuation correction (81). For motion correction, the AIR 5.2.5 software will be used (82), aligning each PET frame to the first 5-min frame. Structural 3-Tesla MRI scans will be used for co-registration of the PET

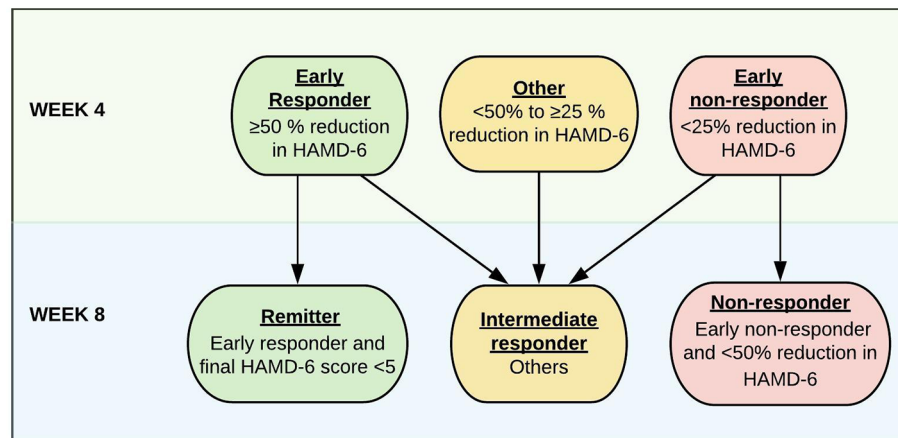


FIGURE 3 | Response categorization for patients with MDD after 4 and 8 weeks of antidepressant treatment based on changes in HAM-D₆ score.

images with SPM8 software. Automatic delineation will be carried out in a user-independent manner in PVElab software (83) and mean tissue time activity curves for grey matter volumes will be extracted for kinetic modeling. No partial volume correction will be performed because of the high resolution of the scanner. Regions of interest (ROI) have been chosen due to their known relevance in mood disorders and abundance of 5-HT₄R density (84). The selected ROIs for the primary analyses are neocortex, putamen, caudate nucleus and hippocampus. Co-registration and correct ROI placement for all subjects will be inspected in three planes by a trained investigator. PMOD version 3.0 (PMOD, Zurich, Switzerland) will be used for kinetic modeling and quantification of the 5-HT₄R binding is performed using non-displaceable binding potential (BP_{ND}) as the final outcome measure. The simplified reference tissue model will be used with cerebellum (excluding vermis) as reference region which previously has been validated in humans (76). BP_{ND} is defined as:

$$BP_{ND} = \frac{f_{ND} \times B_{avail}}{K_D}$$

where f_{ND} is the tissue free fraction of non-protein bound ¹¹C-SB207145, B_{avail} is the concentration of available 5-HT₄R and K_D is the dissociation constant for the tracer at equilibrium. Thus, BP_{ND} is proportional to the density of 5-HT₄R.

MRI and fMRI Imaging

All participants are screened for MR-compatibility and thoroughly instructed how to perform the fMRI paradigms by a trained study assistant who uses standardized instructions. All MRI scans for patients will be acquired using the same Siemens 3-Tesla Prisma scanner with a 64-channel head coil. High-resolution structural T1- and T2-weighted MR images will be acquired. Blood oxygenation level dependent fMRI scans will be obtained during a commonly used emotional faces paradigm (85, 86), reward-related guessing paradigm (87, 88) and a 10-min rs-

fMRI scan. During the rs-fMRI scan, participants are asked to close their eyes, let their mind wander and to not fall asleep. All structural scans of patients will be screened for pathological abnormalities by a medical specialist in radiology.

EEG

EEG data is recorded using a 256-channel HydroCel Sensor Net system (EGI, Inc., Eugene, OR) at 1,000 Hz with 0.1–100 Hz analog filtering where vertex electrode serve as the reference. Impedances across all electrodes are kept below 50 kΩ. EEG/ERP recording at baseline included: resting EEG (with eyes closed and open), two-tone auditory oddball and the LDAEP tasks. The same EEG/ERP recording will be re-tested in a subgroup of patients after 8 weeks of treatment.

Resting EEG

Resting EEG is recorded during four 3-min periods with a counterbalanced order of OCOC (O for eyes open, C for eyes closed) or COCO between subjects. Participants are instructed to remain still and relax, avoid eye-blinks and movements and to relax chin muscles during recording. Absolute and relative powers are computed using the following frequency bands: δ (1–4 Hz), θ (4–8 Hz), α (8–12 Hz), and beta (8–30 Hz). In addition, alpha peak frequency (APF) is identified by the frequency at maximal absolute power from the spectral range of 7–13 Hz. Frontal alpha asymmetry will be calculated using alpha power with the formula of (F4 – F3)/(F4 + F3) (Arns et al., 2015). Furthermore, theta activity will be extracted from anterior cingulate cortex with exact low-resolution electromagnetic tomography (eLoreta).

Task Elicited ERPs

The two-tone auditory oddball paradigm consists of two acoustic stimuli with different frequencies. Participants are presented with a series of standard tones (500 Hz) and deviant tones (1,000 Hz) binaurally through inserted earphones (Etymotic Research Inc., ER 3C). They are instructed to press a button when the deviant tones are presented while ignoring the standard tones. ERP components

such as N1 and P3 will be computed, both peak latency and amplitude (baseline to peak) will be extracted by the averaged trials. Participants are presented with five acoustic stimuli with different intensities (60, 70, 80, 90, and 100 dB SPL) in the same frequency of 1,000 Hz. No response is needed. The primary outcome is the slopes of peak-to-peak N1/P2 amplitudes extracted from the average trials at each intensity. A more comprehensive description of the EEG data will be presented in the subsequent reports.

Cognitive Testing

All participants undergo cognitive testing using selected tasks from the novel test battery EMOTICOM, assessing affective and social cognition including emotional face recognition, emotional threshold detection, theory of mind, and moral emotions (89). In addition, affective memory (90), working memory, reaction time and IQ will also be assessed. Testing is planned and conducted by trained neuropsychologists prior to start of drug intervention and again after 12 weeks of treatment.

Psychometrics

Apart from clinical visits including HAMD_{17/6} ratings, patients will apply self-monitoring during the study period and fill out Danish versions of online questionnaires throughout the study. All questionnaires will be imported directly to an internal database through LimeSurvey, a free and open source software. Before EEG scans and cognitive testing, all participants will report their current mood state using an in-house Likert-scale. An adjusted Likert-scale will be filled out after each MR-scan. During visits at week 4, week 8, and week 12, patients are also asked to fill out a comprehensive set of self-rating state questionnaires (see **Table 1** for a full overview). Healthy controls will be asked to fill out selected state questionnaires as part of their baseline assessments.

Biomaterials

Blood

At baseline, all participants will be screened for basic somatic status to exclude somatic conditions with possible influence on depressive symptoms. Blood samples will be collected throughout the study (see **Table 2** for a full overview) for determination of inflammatory status (high sensitivity C-reactive protein, tumor necrosis factor- α , Interleukin-6, -18, and -10) (106–109); epigenetic variations (SERT, FKBP Prolyl Isomerase 5, Catechol-O-Methyltransferase (COMT), monoamine oxidase-A, glucocorticoid-, estrogen-, oxytocin receptor and oxytocin gene-methylation); extraction of DNA for genotypes of relevance (rs41271330 (110), serotonin-transporter-linked polymorphic region (5-HTTLPR) (70), COMT, Brain-Derived Neurotrophic Factor val66met) and ABCB1, FZD7, and WNT2B (that presumably influence responsiveness to pharmacological antidepressant treatment (111)). At week 8, serum samples of the antidepressant drug (i.e., escitalopram or duloxetine) are collected as trough concentrations in steady state, with primary purpose of monitoring compliance. The samples will be stored at -20°C (or -80°C for plasma EDTA samples) until analyzation in batches at completion of the trial. Quantification of escitalopram and duloxetine in serum will be performed at the Laboratory of the

TABLE 1 | Table over questionnaires obtained throughout the study.

| Questionnaires | Time point | | | | | |
|-----------------------|------------|-------|--------|--------|--------|---------|
| | Baseline | Week1 | Week 2 | Week 4 | Week 8 | Week 12 |
| MINI | X | | | | | |
| HAMD-17/6 | X | X | X | X | X | X |
| UKU | | X | X | X | X | X |
| NEO-PIR | X | | | | | |
| CATS | X | | | | | |
| EHl | X | | | | | |
| OS-FHAM | X | | | | | |
| PBI-mother/ father | X | | | | | |
| POMS* | X | | | | X | X |
| Likert-scale* | X | | | | X | X |
| BDI-II | X | | | X | X | X |
| MDI | X | | | X | X | X |
| PSS | X | | | X | X | X |
| SHAPS | X | | | X | X | X |
| RRS | X | | | X | X | X |
| CSFQ_F_C | X | | | X | X | X |
| SUSY item 32 | X | | | X | X | X |
| Activity | X | | | X | X | X |
| GAD-10 | X | | | X | X | X |

Trait questionnaires at baseline includes personality traits with NEO-PIR (91); Child Abuse and Trauma Scale (CATS) (92) a survey about early life stress which has shown to be able to modulate the serotonin system in the brain (93); handedness with Edinburgh Handedness Inventory (EHl) (94); an in-house version of the Family History Assessment module (FHAM) questionnaire, i.e., "Online Stimulant" (OS)-FHAM; Parental Bonding Inventory (PBI) (both mother and father) (95). State conditions included a self-rating questionnaire of Profile of Mood States (POMS) (96); an in-house Likert-scale; Beck's Depression Inventory-II (BDI-II) (97); Major Depression inventory (MDI) (98); Cohen's Perceived Stress Scale (PSS) (99, 100); Snaith-Hamilton Pleasure Scale (SHAPS) (101); Rumination Response Scale (RSS) 102; Changes in Sexual Functioning Questionnaire (CSFQ) (103); "Sundhed og Sygelighed" Sex Quality Questionnaire item 32 (SUSY-item 32) (104); an in-house questionnaire about daily physical activity (71); and Generalized Anxiety Disorder-10 (GAD-10) (105). * Collected in immediate extension to EEG and MR examinations or cognitive testing.

Danish Epilepsy Centre, Filadelfia, using a routine UPLC-MS/MS method developed in-house. Standard operating procedure instructions have been established before trial initiation and will be followed during the assessment of all biomaterial.

Saliva

Saliva will be collected to determine the total cortisol output across one day as well as dynamics of the HPA-axis, as indexed by CAR. Serial saliva samples will be sampled at home and collected at baseline and at week 8 (see **Table 2**). Those visits will be placed as close to the PET-scan day or week 8 visit as possible, and patients are instructed to take samples immediately after awakening and again after 15, 30, 45, and 60 min, at 12, 6, and 11 pm. Participants are also instructed to collect saliva samples preferably during weekdays, not perform strenuous exercise <2 h and not to have any oral intake or brush their teeth <1 h prior to sampling. Cohen's Perceived Stress Scale and basic information about sleep and food intake will be filled out in conjunction with the home-sampling. All participants receive careful training in saliva collection, instructions of home-sampling procedures; cold storage of samples and fast delivery either by mail or personal delivery to the laboratory facility for preparation. When received, salivary test-tubes are centrifuged and stored at -80°C until later single-batch analysis.

TABLE 2 | Somatic status and biomaterial assessed at various timepoints throughout the study.

| Analysis | Sample | Timepoint | | |
|--------------------------------|--|-----------|--------|---------|
| | | Baseline | Week 8 | Week 12 |
| Somatic blood-sample screening | Hemoglobin, white blood cell count, metamyelo.+myelo.+promyelocytes. | X | X | X |
| | C-reactive protein. | | | |
| | Na+, K+, Creatinine | X | | |
| | ASAT, ALAT, GGT, LDH, BAP | X | | |
| | Albumin, Coagulation factors II+VI+X, thrombocytes | X | | |
| | B12, Folate | X | | |
| | 25-OH-vitamin D | X | | |
| | Blood sugar, HbA1c | X | | |
| | Triglycerides, total-cholesterol, HDL, LDL | X | | |
| | TSH, Ionized Calcium | X | | |
| | Estradiol, testosterone, progesterone, FSH (females) | X | | |
| | Electro Cardiogram (ECG) | X | | |
| Somatic examination | Neurological status | X | | |
| | Somatic status | X | | |
| Compliance to medicine control | S -escitalopram or S -duloxetine | | X | |
| Biobank | Inflammation and cytokines (hsCRP, TNF- α , IL-6, IL-18 and IL-10) | X | X | X |
| Biobank | Epigenetics (5-HTT, glucocorticoid-, FKBP5, COMT, MAO-A, estrogen-, oxytocin receptor and oxytocin gene-methylation) | X | X | X |
| Biobank | Genotypes (rs41271330, 5-HTTLPR, COMT, BDNFval66met) | X | | |
| Biobank | Gene transcription profiles (mRNA and microRNA, ABCB1, FZD7 and WNT2B) | X | X | X |
| Oxidative stress | Urine (8-oxo-dG and 8-oxo-Guo) | X | X | |
| Biobank | Saliva (Cortisol awakening response) | X | X | |

Urine

Spot-urine samples will be collected at baseline and week 8 visits for patients (see **Table 2**) in 2-ml Eppendorf tubes and will be stored at -20°C for later single-batch analysis. Apart from pregnancy and drug-screening (see *Baseline Assessments Before Treatment*), all urine samples will be analyzed for 8-oxodG and 8-oxoGuo markers for systemic DNA/RNA damage with ultra-performance liquid chromatography with tandem mass spectrometry and normalized to urinary creatinine (112).

Statistical Analyses

Evaluating Associations Between Baseline Measures, Changes From Baseline Measures, and Clinical Outcomes

Baseline data from each modality of interest, i.e., PET, EEG, fMRI, MRI, cognitive measures, peripheral molecular markers, and clinical/demographic patient profiles, will be available for

evaluating associations with the clinical outcomes for the entire group ($n = 100$ included). Changes from baseline data will be available for the subgroup (around $n = 40$ invited), who will be re-examined with brain scans and EEG for evaluating an association between changed measures and clinical outcomes. Similarly, cognitive follow-up data will be collected for all patients after 12 weeks of treatment. Primary analyses will test mean differences in baseline measures of the biomarkers from each modality between healthy controls and patients as well as response groups (remitter vs. non-responder at week 8, i.e., primary clinical outcome) using multiple linear regressions. This analysis focuses on the two extreme outcome groups. Secondary analyses will test the association between baseline measures of the biomarkers from each modality and antidepressant treatment response on a continuous scale, i.e., relative change in HAMD₆, using linear multiple regression. This analysis incorporates the full spectrum of clinical outcomes. Similar analyses will be performed to study the association between the change from baseline measures of the biomarkers and the clinical outcomes. Regression models will be adjusted for age and sex, as well as modality-specific relevant covariates. For instance, 5-HTTLPR status is predictive of 5-HT₄R binding (70) and will be adjusted for in the analyses concerning 5-HT₄R. When relevant, interactions will be evaluated, e.g., we will test if the association between the clinical outcome and 5-HT₄R is moderated by inflammatory status. Diagnostic regression tools will be used to assess model's assumptions (e.g., linearity of the effects, normality assumption for residuals). When violated, corrective procedures will be used (e.g., splines and bootstrap resampling) (113). As appropriate, adjustments for multiple comparisons will be performed within each modality. In the analysis of the PET data, we will instead use a Latent Variable Model relating the 5-HT₄R binding in several brain regions (neocortex, caudate nucleus, putamen and hippocampus) to treatment outcome *via* a latent variable (114). This allows us to assess the association between 5-HT₄R binding and clinical outcome with a single test. Patients are considered with un-verified compliance if they have taken less than 2/3 of their antidepressant medicine, missed their week 8 visit, or have undetectable serum drug levels at week 8 (i.e., <10 nM for escitalopram and <15 nM for duloxetine). Patients with un-verified compliance will not be included in primary longitudinal analyses of treatment response. Missing data will therefore be handled using complete case analysis which in regression models is valid when the probability of dropping out of the study is, conditional on the covariates, independent of the outcome. If any participants were to be excluded during the study because of their clinical outcome, a sensitivity analysis will be performed.

Evaluation of the Predictive Value of the Biomarkers Within Modality

Logistic regression models for the primary clinical outcome will be used to obtain the probability of each patient to be a remitter (vs. a non-responder) based on its clinical data and the value of a modality-specific biomarker. Given a threshold (e.g., 0.5),

patients with an estimated probability greater than the threshold will be predicted to be remitters, otherwise to be non-responders. The receiver operating characteristic (ROC) curve will be used to assess the compromise between sensitivity and specificity of this classification across thresholds. Since a 33% remission rate is expected in treatment regimen comparable to ours (5), we will focus on the ROC curve with high-specificity. The AUC (area under the curve) of the relevant part of the ROC curve will be reported as a summary of the predictive performance of each biomarker. The classification performance (accuracy, positive predictive value, negative predictive value) at the threshold optimizing the sum specificity and specificity will also be reported. To limit optimistic biases, these measures will be estimated using five-fold cross-validation (115). A permutation procedure will be used to obtain the null distribution of the predictive performance, against which the observed performance will be compared. Additional classification schemes may be considered (e.g., responder status as defined by $\geq 50\%$ reduction in HAMD₆ at week 8), with appropriate adjustment for inflated type-I error, to facilitate comparison of the current data with other relevant clinical trials. The predictor performance will be evaluated in a modality specific fashion and at a next stage, combined predictors will be evaluated.

Predictive Value of the Biomarkers Across Modalities

Two strategies will be considered to optimize prediction of treatment response using biomarkers measured at baseline across modalities. In the first strategy, we will combine the specific biomarker-candidates across all modalities (as predefined in **Appendix 1**), which will generate around 50 candidate biomarkers. A dimension reduction step will be used to define a small number of predictors (roughly 5–10) that will be used in a logistic regression model. The second strategy will use an algorithm to (i) identify, in a data-driven way, biomarkers with a predictive value among all the existing biomarkers (roughly 5,000–10,000) and (ii) predict treatment response based on the identified subset of informative biomarkers. We will investigate the use of machine learning methods (e.g., random forest, neural networks) as well as ensemble methods [e.g., Super Learner (116)]. The assessment of the predictive performance of these strategies will be carried out as described in the previous section.

Ethics Approval and Consent to Participate

The study protocol complies with the Declaration of Helsinki II and data collected during the trial will be monitored throughout the study period (for every 10th patient included) by an independent Good Clinical Practice unit in the capital region of Denmark (www.gcp-enhed.dk/en). The study has been approved by the Committees on Health Research Ethics in the Capital Region of Denmark (reference number: H-15017713), the Danish Data Protection Agency (04711/RH-2016-163), and Danish Medicines Agency (protocol number: NeuroPharm-NP1,

EudraCT-number 2016-001626-34). All potential participants will receive oral and written information about the study by the enrolling clinician, and all enrolled participants will provide written informed consent prior to inclusion. Adverse events have been scheduled to be reported annually to the Danish Medicines Agency. The study was registered at clinicaltrials.gov prior to initiation (NCT02869035), date: 08.16.2016.

Availability of Data and Materials

Data management and monitoring during the study agrees to the rules on protection of personal data. To protect confidentiality, paper-based material (e.g., cognitive test results) will be stored in a secured archive, while electronic data files that are identifiable will be stored in password secured files behind firewall in accordance to regulations. To promote data quality, the primary outcome measurement (HAMD_{17/6} scores) will be obtained during interviews on paper, manually transferred into the local database through LimeSurvey and cross-checked twice before used in analyses. Biological material will be coded with a unique identification-number and access to de-identification keys is restricted to authorized personnel only and stored in a temporary biobank located in secured areas in the laboratory facility. The biomaterial will later be analyzed in batches to reduce noise, and potential extra material after the end of the clinical trial will be transferred to the CIMBI biobank (71). All biological material will ultimately be anonymized after 15 years after the end of trial.

Progress to Date

The study opened for inclusion of patients in August 2016. To date, the remaining biological data including genetic status of healthy controls are planned to be collected. Obtained biological material is currently being analyzed and processing of imaging data is on-going. Results from the trial are planned to be communicated to the participants and public through publication in international medical journals.

DISCUSSION

The main purpose of the present study is to identify individual or combined predictors (biomarkers) of standard pharmacological antidepressant treatment outcome in MDD, by using multiple modalities such as brain imaging (PET, fMRI), EEG, cognitive tools, and clinical and molecular markers. Special emphasis in the study design has been given to evaluate the biomarker 5-HT₄R PET as a promising clinically relevant tool since the 5HT₄R availability is of interest in the pathophysiology and as a therapeutic target in MDD, and also as an index of serotonin tonus. The aim of this trial is not to investigate the specific treatment efficacy but to investigate biomarkers for response to standard treatment in a naturalistic setting, e.g., similar to the STAR*D trial (117). The study includes multiple cross-sectional and longitudinal measures in a large number of patients and controls, which

offers a unique opportunity to (a) uncover potential biomarkers or clusters of biomarkers of treatment prediction, (b) apply the findings for stratification of MDD, (c) advance the understanding of pathophysiological underpinnings of MDD, (d) map neurobiological signatures of antidepressant treatment response, and lastly (e) to ideally pave a way for a precision medicine approach for optimized treatment of MDD.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by the Committees on Health Research Ethics in the Capital Region of Denmark (reference number: H-15017713). The patients/participants provided their written informed consent to participate in this study.

AUTHOR CONTRIBUTIONS

GK, MJ, and VF conceived the concept of the study, with help and support from AJ and KK-F. KK-F, C-TI, VD, DS, PF, MG, HP, AG, and BO have supervised study design and contributed to drafting of the manuscript. All authors contributed to the article and approved the submitted version.

REFERENCES

- Global Burden of Disease Study 2013 Collaborators. Global, regional, and national incidence, prevalence, and years lived with disability for 301 acute and chronic diseases and injuries in 188 countries, 1990–2013: a systematic analysis for the Global Burden of Disease Study 2013. *Lancet* (London, England) 386(9995):743–800. doi: 10.1016/S0140-6736(15)60692-4
- Ferrari AJ, Charlson FJ, Norman RE, Patten SB, Freedman G, Murray C, et al. Burden of Depressive Disorders by Country, Sex, Age, and Year: Findings from the Global Burden of Disease Study 2010. *PLoS Med* (2013) 10(11):e1001547. doi: 10.1371/journal.pmed.1001547
- Benmansour S, Owens WA, Cecchi M, Morilak DA, Frazer A. Serotonin Clearance In Vivo Is Altered to a Greater Extent by Antidepressant-Induced Downregulation of the Serotonin Transporter than by Acute Blockade of this Transporter. *J Neurosci* (2002) 22(15):6766–72. doi: 10.1523/jneurosci.22-15-06766.20024
- Smith DF, Jakobsen S. Molecular neurobiology of depression: PET findings on the elusive correlation with symptom severity. *Front Psychiatry* (2013) 4:8. doi: 10.3389/fpsy.2013.00008
- Rush AJ, Trivedi MH, Wisniewski SR, Nierenberg AA, Stewart JW, Warden D, et al. Acute and longer-term outcomes in depressed outpatients requiring one or several treatment steps: A STAR*D report. *Am J Psychiatry* (2006) 163(11):1905–17. doi: 10.1176/ajp.2006.163.11.19056
- Nakajima S, Uchida H, Suzuki T, Watanabe K, Hirano J, Yagihashi T, et al. Is switching antidepressants following early nonresponse more beneficial in acute-phase treatment of depression?: A randomized open-label trial. *Prog Neuropsychopharmacol Biol Psychiatry* (2011) 35(8):1983–9. doi: 10.1016/j.pnpbp.2011.08.008
- Hasler G. Pathophysiology of depression: Do we have any solid evidence of interest to clinicians? *World Psychiatry* (2010) 9(3):155–61. doi: 10.1002/j.2051-5545.2010.tb00298.x
- Hellwig S, Domschke K. Update on PET imaging biomarkers in the diagnosis of neuropsychiatric disorders. *Curr Opin Neurol* (2019) 32(4):539–47. doi: 10.1097/wco.0000000000000705

FUNDING

Economic support for the study was granted from the Innovation Fund Denmark (GrantID: 5189-00087A), Research Fund of the Mental Health Services - Capital Region of Denmark, Savværksejer Jeppe Juhl og hustru Ovita Juhls Mindelegat, Augustinus Foundation (GrantID: 16-0058), Research Council of Rigshospitalet the independent research fund Denmark (GrantID: DFF-6120-00038), and the Lundbeck Foundation. H. Lundbeck A/S had no influence on study design and will not be involved in data processing or in publishing the results of the trial.

ACKNOWLEDGMENTS

We would like to thank the participating patients and volunteers, laboratory technicians, collaboration partners, study assistants, and funding supporting this study.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fpsy.2020.00641/full#supplementary-material>

- Strawbridge R, Young AH, Cleare AJ. Biomarkers for depression: Recent insights, current challenges and future prospects. *Neuropsychiatr Dis Treat* (2017) 13:1245–62. doi: 10.2147/NDT.S114542
- Williams LM, Rush AJ, Koslow SH, Wisniewski SR, Cooper NJ, Nemeroff C, et al. International Study to Predict Optimized Treatment for Depression (iSPOT-D), a randomized clinical trial: Rationale and protocol. *Trials* (2011) 12:4. doi: 10.1186/1745-6215-12-4
- Trivedi MH, McGrath PJ, Fava M, Parsey RV, Kurian BT, Phillips ML, et al. Establishing moderators and biosignatures of antidepressant response in clinical care (EMBARC): Rationale and design. *J Psychiatr Res* (2016) 78:11–23. doi: 10.1016/j.jpsychires.2016.03.001
- Lam RW, Milev R, Rotzinger S, Andreazza AC, Blier P, Brenner C, et al. Discovering biomarkers for antidepressant response: Protocol from the Canadian biomarker integration network in depression (CAN-BIND) and clinical characteristics of the first patient cohort. *BMC Psychiatry* (2016) 16:105. doi: 10.1186/s12888-016-0785-x
- Gong B, Naveed S, Hafeez DM, Afzal KI, Majeed S, Abele J, et al. Neuroimaging in Psychiatric Disorders: A Bibliometric Analysis of the 100 Most Highly Cited Articles. *J Neuroimaging* (2019) 29(1):14–33. doi: 10.1111/jon.12570
- Rebholz H, Friedman E, Castello J. Alterations of Expression of the Serotonin 5-HT₄ Receptor in Brain Disorders. *Int J Mol Sci* (2018) 19(11):3581. doi: 10.3390/ijms19113581
- Lucas G, Rymar VV, Du J, Mnie-Filali O, Bisgaard C, Manta S, et al. Serotonin(4) (5-HT₄) receptor agonists are putative antidepressants with a rapid onset of action. *Neuron* (2007) 55(5):712–25. doi: 10.1016/j.neuron.2007.07.041
- Licht CL, Marcussen AB, Wegener G, Overstreet DH, Aznar S, Knudsen GM. The brain 5-HT₄ receptor binding is down-regulated in the Flinders Sensitive Line depression model and in response to paroxetine administration. *J Neurochem* (2009) 109(5):1363–74. doi: 10.1111/j.1471-4159.2009.06050.x
- Vidal R, Valdizán EM, Mostany R, Pazos A, Castro E. Long-term treatment with fluoxetine induces desensitization of 5-HT₄ receptor-dependent

- signalling and functionality in rat brain. *J Neurochem* (2009) 110(3):1120–7. doi: 10.1111/j.1471-4159.2009.06210.x
18. Haahr ME, Fisher PM, Jensen CG, Frokjaer VG, Mc Mahon B, Madsen K, et al. Central 5-HT4 receptor binding as biomarker of serotonergic tone in humans: A [11C]SB207145 PET study. *Mol Psychiatry* (2014) 19(4):427–32. doi: 10.1038/mp.2013.147
 19. Stuhmann A, Suslow T, Dannlowski U. Facial emotion processing in major depression: A systematic review of neuroimaging findings. *Biol Mood Anxiety Disord* (2011) 1(1):10. doi: 10.1186/2045-5380-1-10
 20. Williams LM, Korgaonkar MS, Song YC, Paton R, Eagles S, Goldstein-Piekarski A, et al. Amygdala Reactivity to Emotional Faces in the Prediction of General and Medication-Specific Responses to Antidepressant Treatment in the Randomized iSPOT-D Trial. *Neuropsychopharmacology* (2015) 40(10):2398–408. doi: 10.1038/npp.2015.89
 21. Goldstein-Piekarski AN, Korgaonkar MS, Green E, Suppes T, Schatzberg AF, Hastie T, et al. Human amygdala engagement moderated by early life stress exposure is a biobehavioral target for predicting recovery on antidepressants. *Proc Natl Acad Sci* (2016) 113(42):11955–60. doi: 10.1073/pnas.1606671113
 22. Fisher PM, Haahr ME, Jensen CG, Frokjaer VG, Siebner HR, Knudsen GM. Fluctuations in [11C]SB207145 PET Binding Associated with Change in Threat-Related Amygdala Reactivity in Humans. *Neuropsychopharmacology* (2015) 40(6):1510–8. doi: 10.1038/npp.2014.339
 23. Fisher PM, Hariri AR. Linking variability in brain chemistry and circuit function through multimodal human neuroimaging. *Genes Brain Behav* (2012) 11(6):633–42. doi: 10.1111/j.1601-183X.2012.00786.x
 24. Raichle ME. The Brain's Default Mode Network. *Annu Rev Neurosci* (2015) 38:433–47. doi: 10.1146/annurev-neuro-071013-014030
 25. Kaiser RH, Andrews-Hanna JR, Wager TD, Pizzagalli DA. Large-scale network dysfunction in major depressive disorder: A meta-analysis of resting-state functional connectivity. *JAMA Psychiatry* (2015). 72(6):603–11. doi: 10.1001/jamapsychiatry.2015.0071
 26. Drysdale AT, Grosenick L, Downar J, Dunlop K, Mansouri F, Meng Y, et al. Resting-state connectivity biomarkers define neurophysiological subtypes of depression. *Nat Med* (2017) 23(1):28–38. doi: 10.1038/nm.4246
 27. Dinga R, Schmaal L, Penninx BWJH, van Tol MJ, Veltman DJ, van Velzen L, et al. Evaluating the evidence for biotypes of depression: Methodological replication and extension of. *NeuroImage Clin* (2019) 22:101796. doi: 10.1016/j.nicl.2019.101796
 28. Patel MJ, Khalaf A, Aizenstein HJ. Studying depression using imaging and machine learning methods. *NeuroImage Clin* (2016) 10:115–23. doi: 10.1016/j.nicl.2015.11.003
 29. Olbrich S, Arns M. EEG biomarkers in major depressive disorder: Discriminative power and prediction of treatment response. *Int Rev Psychiatry* (2013) 25(5):604–18. doi: 10.3109/09540261.2013.816269
 30. Arns M, Bruder G, Hegerl U, Spooner C, Palmer DM, Etkin A, et al. EEG alpha asymmetry as a gender-specific predictor of outcome to acute treatment with different antidepressant medications in the randomized iSPOT-D study. *Clin Neurophysiol* (2016) 127(1):509–19. doi: 10.1016/j.clinph.2015.05.032
 31. Pizzagalli DA. Frontocingulate dysfunction in depression: Toward biomarkers of treatment response. *Neuropsychopharmacology* (2011) 36(1):183–206. doi: 10.1038/npp.2010.166
 32. Pizzagalli DA, Webb CA, Dillon DG, et al. Pretreatment rostral anterior cingulate cortex theta activity in relation to symptom improvement in depression: A randomized clinical trial. *JAMA Psychiatry* (2018) 75(6):547–54. doi: 10.1001/jamapsychiatry.2018.0252
 33. Jaworska N, Protzner A. Electroclinical features of depression and their clinical utility in assessing antidepressant treatment outcome. *Can J Psychiatry* (2013) 58(9):509–14. doi: 10.1177/070674371305800905
 34. Mulert C, Juckel G, Brunnermeier M, Karch S, Leicht G, Mergl R, et al. Prediction of treatment response in major depression: Integration of concepts. *J Affect Disord* (2007) 98(3):215–25. doi: 10.1016/j.jad.2006.07.021
 35. Juckel G, Pogarell O, Augustin H, Mulert C, Müller-Siecheneder F, Frodl T, et al. Differential prediction of first clinical response to serotonergic and noradrenergic antidepressants using the loudness dependence of auditory evoked potentials in patients with major depressive disorder. *J Clin Psychiatry* (2007) 68(8):1206–12. doi: 10.4088/JCP.v68n0806
 36. Rock PL, Roiser JP, Riedel WJ, Blackwell AD. Cognitive impairment in depression: A systematic review and meta-analysis. *Psychol Med* (2014) 44(10):2029–40. doi: 10.1017/S003329713002535
 37. Weightman MJ, Knight MJ, Baune BT. A systematic review of the impact of social cognitive deficits on psychosocial functioning in major depressive disorder and opportunities for therapeutic intervention. *Psychiatry Res* (2019) 274:195–212. doi: 10.1016/j.psychres.2019.02.035
 38. Roiser JP, Elliott R, Sahakian BJ. Cognitive mechanisms of treatment in depression. *Neuropsychopharmacology* (2012) 37(1):117–36. doi: 10.1038/npp.2011.183
 39. Kingslake J, Dias R, Dawson GR, Simon J, Goodwin GM, Harmer CJ, et al. The effects of using the PReDiCT Test to guide the antidepressant treatment of depressed patients: Study protocol for a randomised controlled trial. *Trials* (2017) 18(1):558. doi: 10.1186/s13063-017-2247-2
 40. Zuckerman H, Pan Z, Park C, Brietzke E, Musial N, Shariq AS, et al. Recognition and Treatment of Cognitive Dysfunction in Major Depressive Disorder. *Front Psychiatry* (2018) 9:655. doi: 10.3389/fpsy.2018.00655
 41. Haahr ME, Fisher P, Holst K, Madsen K, Jensen CG, Marner L, et al. The 5-HT4 receptor levels in hippocampus correlates inversely with memory test performance in humans. *Hum Brain Mapp* (2013) 34(11):3066–74. doi: 10.1002/hbm.22123
 42. Stenbæk DS, Fisher PM, Ozenne B, Andersen E, Hjorndt LV, McMahon B, et al. Brain serotonin 4 receptor binding is inversely associated with verbal memory recall. *Brain Behav* (2017) 7(4):e00674. doi: 10.1002/brb3.674
 43. Dantzer R, O'Connor JC, Lawson MA, Kelley KW. Inflammation-associated depression: From serotonin to kynurenine. *Psychoneuroendocrinology* (2011) 36(3):426–36. doi: 10.1016/j.psyneuen.2010.09.012
 44. Lanquillon S, Krieg JC, Bening-Abu-Shach U, Vedder H. Cytokine production and treatment response in major depressive disorder. *Neuropsychopharmacology* (2000) 22(4):370–9. doi: 10.1016/S0893-133X(99)00134-7
 45. Liu JJ, Bin WY, Strawbridge R, Bao Y, Chang S, Shi L, et al. Peripheral cytokine levels and response to antidepressant treatment in depression: a systematic review and meta-analysis. *Mol Psychiatry* (2019) 25(2):339–50. doi: 10.1038/s41380-019-0474-5
 46. Richards EM, Zanotti-Fregonara P, Fujita M, Newman L, Farmer C, Ballard ED, et al. PET radioligand binding to translocator protein (TSPO) is increased in unmedicated depressed subjects. *EJNMMI Res* (2018) 8(1):57. doi: 10.1186/s13550-018-0401-9
 47. Zhu CB, Blakely RD, Hewlett WA. The proinflammatory cytokines interleukin-1beta and tumor necrosis factor-alpha activate serotonin transporters. *Neuropsychopharmacology* (2006) 31(10):2121–31. doi: 10.1038/sj.npp.1301029
 48. Allison DJ, Ditor DS. The common inflammatory etiology of depression and cognitive impairment: A therapeutic target. *J Neuroinflamm* (2014) 11:151. doi: 10.1186/s12974-014-0151-1
 49. Khan A, Faucett J, Morrison S, Brown WA. Comparative mortality risk in adult patients with schizophrenia, depression, bipolar disorder, anxiety disorders, and attention-deficit/hyperactivity disorder participating in psychopharmacology clinical trials. *JAMA Psychiatry* (2013) 70(10):1091–9. doi: 10.1001/jamapsychiatry.2013.149
 50. Laursen TM, Musliner KL, Benros ME, Vestergaard M, Munk-Olsen T. Mortality and life expectancy in persons with severe unipolar depression. *J Affect Disord* (2016) 193:203–7. doi: 10.1016/j.jad.2015.12.067
 51. Wolkowitz OW, Epel ES, Reus VI, Mellon SH. Depression gets old fast: Do stress and depression accelerate cell aging? *Depress Anxiety* (2010) 27(4):327–38. doi: 10.1002/da.20686
 52. Jørgensen A, Maigaard K, Wörtwein G, Hageman I, Henriksen T, Weimann A, et al. Chronic restraint stress in rats causes sustained increase in urinary corticosterone excretion without affecting cerebral or systemic oxidatively generated DNA/RNA damage. *Prog Neuropsychopharmacol Biol Psychiatry* (2013) 40:30–7. doi: 10.1016/j.pnpbp.2012.08.016
 53. Jørgensen A, Brødbeck K, Weimann A, Fink-Jensen A, Knorr U, Greisen Soendergaard M, Henriksen T, et al. Jørgensen: Increased Systemic Oxidatively Generated DNA and RNA Damage in Schizophrenia.

- Psychiatry Research 2013 (Epub ahead of print). *Dan Med J* (2013) 209 (3):417–23. doi: 10.1016/j.psychres.2013.01.033
54. Joergensen A, Broedbaek K, Weimann A, Semba RD, Ferrucci L, Joergensen MB, et al. Association between urinary excretion of cortisol and markers of oxidatively damaged DNA and RNA in humans. *PLoS One* (2011) 6(6): e20795. doi: 10.1371/journal.pone.0020795
 55. Jorgensen A, Krogh J, Miskowiak K, Bolwig TG, Kessing LV, Fink-Jensen A, et al. Systemic oxidatively generated DNA/RNA damage in clinical depression: Associations to symptom severity and response to electroconvulsive therapy. *J Affect Disord* (2013) 149(1–3):355–62. doi: 10.1016/j.jad.2013.02.011
 56. Walton NM, Shin R, Tajinda K, Heusner CL, Kogan JH, Miyake S, et al. Adult neurogenesis transiently generates oxidative stress. *PLoS One* (2012) 7(4):e35264. doi: 10.1371/journal.pone.0035264
 57. Chung CP, Schmidt D, Stein CM, Morrow JD, Salomon RM. Increased oxidative stress in patients with depression and its relationship to treatment. *Psychiatry Res* (2012) 206(2–3):213–6. doi: 10.1016/j.psychres.2012.10.018
 58. Jorgensen A. Oxidatively generated DNA/RNA damage in psychological stress states. *Dan Med J* (2013) 60(7):1–14.
 59. Da Cunha-Bang S, Hjortd LV, Dam VH, Stenbæk DS, Sestoft D, Knudsen GM. Anterior cingulate serotonin 1B receptor binding is positively associated with inhibitory control and amygdala reactivity to aversive faces. *Eur Neuropsychopharmacol* (2016) 92:199–204. doi: 10.1016/s0924-977x(16)31246-9
 60. Fava M, Rush AJ, Alpert JE, Balasubramani GK, Wisniewski SR, Carmin CN, et al. Difference in treatment outcome in outpatients with anxious versus nonanxious depression: A STAR*D report. *Am J Psychiatry* (2008) 165 (3):342–51. doi: 10.1176/appi.ajp.2007.06111868
 61. Montejo AL, Montejo L, Baldwin DS. The impact of severe mental disorders and psychotropic medications on sexual health and its implications for clinical management. *World Psychiatry* (2018) 17(1):3–11. doi: 10.1002/wps.20509
 62. Williams VSL, Edin HM, Hogue SL, Fehnel SE, Baldwin DS. Prevalence and impact of antidepressant-associated sexual dysfunction in three European countries: Replication in a cross-sectional patient survey. *J Psychopharmacol* (2010) 24(4):489–96. doi: 10.1177/0269881109102779
 63. Pfaus JG. Pathways of sexual desire. *J Sex Med* (2009) 6(6):1506–33. doi: 10.1111/j.1743-6109.2009.01309.x
 64. Jakobsen GR, Fisher PM, Dyssegaard A, McMahon B, Holst KK, Lehel S, et al. Brain serotonin 4 receptor binding is associated with the cortisol awakening response. *Psychoneuroendocrinology* (2016) 67:124–32. doi: 10.1016/j.psyneuen.2016.01.032
 65. Frøkjær VG, Erritzoe D, Holst KK, Jensen PS, Rasmussen PM, Fisher PM, et al. Prefrontal serotonin transporter availability is positively associated with the cortisol awakening response. *Eur Neuropsychopharmacol* (2013) 23 (4):285–94. doi: 10.1016/j.euroneuro.2012.05.013
 66. Vrshek-Schallhorn S, Doane LD, Mineka S, Zinbarg RE, Craske MG, Adam EK. The cortisol awakening response predicts major depression: Predictive stability over a 4-year follow-up and effect of depression history. *Psychol Med* (2013) 43(3):483–93. doi: 10.1017/S0033291712001213
 67. Booij SH, Bouma EMC, De Jonge P, Ormel J, Oldehinkel AJ. Chronicity of depressive problems and the cortisol response to psychosocial stress in adolescents: The TRAILS study. *Psychoneuroendocrinology* (2013) 38 (5):659–66. doi: 10.1016/j.psyneuen.2012.08.004
 68. Ruhé HG, Khoenkhoe SJ, Ottenhof KW, Koeter MW, Mocking RJT, Schene AH. Longitudinal effects of the SSRI paroxetine on salivary cortisol in Major Depressive Disorder. *Psychoneuroendocrinology* (2015) 52:261–71. doi: 10.1016/j.psyneuen.2014.10.024
 69. Timmerby N, Andersen JH, Søndergaard S, Østergaard SD, Bech P, Bech P. A Systematic Review of the Clinimetric Properties of the 6-Item Version of the Hamilton Depression Rating Scale (HAM-D6). *Psychother Psychosom* (2017) 86(3):141–9. doi: 10.1159/000457131
 70. Fisher PM, Holst KK, Mc Mahon B, Haahr ME, Madsen K, Gillings N, et al. 5-HTTLPR status predictive of neocortical 5-HT 4 binding assessed with [11C]SB207145 PET in humans. *Neuroimage* (2012) 62(1):130–6. doi: 10.1016/j.neuroimage.2012.05.013
 71. Knudsen GM, Jensen PS, Erritzoe D, Baaré W, Ettrup A, Fisher PM, et al. The Center for Integrated Molecular Brain Imaging (Cimbi) database. *Neuroimage* (2016) 124(Pt B):1213–9. doi: 10.1016/j.neuroimage.2015.04.025
 72. Sheehan DV, Lecrubier Y, Sheehan KH, Amorim P, Janavs J, Weiller E, et al. The Mini-International Neuropsychiatric Interview (M.I.N.I.): The development and validation of a structured diagnostic psychiatric interview for DSM-IV and ICD-10. *In: J Clin Psychiatry* (1998) 50(Suppl 20):22–57. doi: 10.1016/S0924-9338(99)80239-9
 73. Olsen LR, Mortensen EL, Bech P. Prevalence of major depression and stress indicators in the Danish general population. *Acta Psychiatr Scand* (2004) 109 (2):96–103. doi: 10.1046/j.0001-690X.2003.00231.x
 74. Culppepper L. Escitalopram: A New SSRI for the Treatment of Depression in Primary Care. *Prim Care Companion J Clin Psychiatry* (2002) 4(6):209–14. doi: 10.4088/PCC.v04n0601
 75. Bushnell GA, Stürmer T, Gaynes BN, Pate V, Miller M. Simultaneous antidepressant and benzodiazepine new use and subsequent long-term benzodiazepine use in adults with depression, United States, 2001–2014. *JAMA Psychiatry* (2017) 74(7):747–55. doi: 10.1001/jamapsychiatry.2017.1273
 76. Marner L, Gillings N, Madsen K, Erritzoe D, Baaré WF, Svarer C, et al. Brain imaging of serotonin 4 receptors in humans with [11C]SB207145-PET. *Neuroimage* (2010) 50(3):855–61. doi: 10.1016/j.neuroimage.2010.01.054
 77. Bymaster FP, Dreshfield-Ahmad LJ, Threlkeld PG, Shaw JL, Thompson L, Nelson DL, et al. Comparative affinity of duloxetine and venlafaxine for serotonin and norepinephrine transporters in vitro and in vivo, human serotonin receptor subtypes, and other neuronal receptors. *Neuropsychopharmacology* (2001) 25(6):871–80. doi: 10.1016/S0893-133X(01)00298-6
 78. Lingjærde O, Ahlfors UG, Bech P, Dencker SJ, Elgen K. The UKU side effect rating scale: A new comprehensive rating scale for psychotropic drugs and a cross-sectional study of side effects in neuroleptic-treated patients. *Acta Psychiatr Scand* (1987) 334:1–100. doi: 10.1111/j.1600-0447.1987.tb10566.x
 79. Marner L, Gillings N, Comley RA, Baaré WF, Rabiner EA, Wilson AA, et al. Kinetic Modeling of 11C-SB207145 Binding to 5-HT₄ Receptors in the Human Brain In Vivo. *J Nucl Med* (2009) 50(6):900–8. doi: 10.2967/jnumed.108.058552
 80. Sureau FC, Reader AJ, Comtat C, Leroy C, Ribeiro MJ, Buvat I, et al. Impact of Image-Space Resolution Modeling for Studies with the High-Resolution Research Tomograph. *J Nucl Med* (2008) 49(6):1000–8. doi: 10.2967/jnumed.107.045351
 81. Keller SH, Svarer C, Sibomana M. Attenuation correction for the HRRT PET-scanner using transmission scatter correction and total variation regularization. *IEEE Trans Med Imaging* (2013) 32(9):1611–21. doi: 10.1109/TMI.2013.2261313
 82. Woods RP, Cherry SR, Mazziotta JC. Rapid automated algorithm for aligning and reslicing pet images. *J Comput Assist Tomogr* (1992) 16 (4):620–33. doi: 10.1097/00004728-199207000-00024
 83. Svarer C, Madsen K, Hasselbalch SG, Pinborg LH, Haugbøl S, Frøkjær VG, et al. MR-based automatic delineation of volumes of interest in human brain PET images using probability maps. *Neuroimage* (2005) 24(4):969–79. doi: 10.1016/j.neuroimage.2004.10.017
 84. Madsen K, Marner L, Haahr M, Gillings N, Knudsen GM. Mass dose effects and in vivo affinity in brain PET receptor studies - a study of cerebral 5-HT 4 receptor binding with [11C]SB207145. *Nucl Med Biol* (2011) 38(8):1085–91. doi: 10.1016/j.nucmedbio.2011.04.006
 85. da Cunha-Bang S, Fisher PM, Hjortd LV, Perfalk E, Persson Skibsted A, Bock C, et al. Violent offenders respond to provocations with high amygdala and striatal reactivity. *Soc Cognit Affect Neurosci* (2017) 12(5):802–10. doi: 10.1093/scan/nsx006
 86. Nikolova YS, Iruku SP, Lin C-W, Conley E D, Puralawski R, French B, et al. FRAS1-related extracellular matrix 3 (FREM3) single-nucleotide polymorphism effects on gene expression, amygdala reactivity and perceptual processing speed: An accelerated aging pathway of depression risk. *Front Psychol* (2015) 6:1377. doi: 10.3389/fpsyg.2015.01377
 87. Forbes EE, Hariri AR, Martin SL, Silk JS, Moyses DL, Fisher PM, et al. Altered striatal activation predicting real-world positive affect in adolescent major depressive disorder. *Am J Psychiatry* (2009) 166(1):64–73. doi: 10.1176/appi.ajp.2008.07081336
 88. Nikolova YS, Ferrell RE, Manuck SB, Hariri AR. Multilocus genetic profile for dopamine signaling predicts ventral striatum reactivity. *Neuropsychopharmacology* (2011) 36(9):1940–7. doi: 10.1038/npp.2011.82

89. Bland AR, Roiser JP, Mehta MA, Schei T, Boland H, Campbell-Meiklejohn DK, et al. EMOTICOM: A Neuropsychological Test Battery to Evaluate Emotion, Motivation, Impulsivity, and Social Cognition. *Front Behav Neurosci* (2016) 10:25. doi: 10.3389/fnbeh.2016.00025
90. Jensen CG, Hjordt LV, Stenbæk DS, Andersen E, Back SK, Lansner J, et al. Development and psychometric validation of the verbal affective memory test. *Memory* (2016) 24(9):1208–23. doi: 10.1080/09658211.2015.1087573
91. Costa P, McCrae R. (2008). *The revised NEO personality inventory (NEO-PI-R)*. The SAGE Handbook of Personality Theory and Assessment. 2:179–98. doi: 10.4135/9781849200479.n9
92. Sanders B, Becker-Lausen E. The measurement of psychological maltreatment: Early data on the child abuse and trauma scale. *Child Abuse Negl* (1995) 19(3):315–23. doi: 10.1016/S0145-2134(94)00131-6
93. Harrison EL, Baune BT. Modulation of early stress-induced neurobiological changes: A review of behavioural and pharmacological interventions in animal models. *Transl Psychiatry* (2014) 4(5):e390. doi: 10.1038/tp.2014.31
94. Oldfield RC. The assessment and analysis of handedness: The Edinburgh inventory. *Neuropsychologia* (1971) 9(1):97–113. doi: 10.1016/0028-3932(71)90067-4
95. Parker G, Tupling H, Brown LB. A Parental Bonding Instrument. *Br J Med Psychol* (1979) 52:1–10. doi: 10.1111/j.2044-8341.1979.tb02487.x
96. McNair DM, Lorr M, Droppleman LF. *Revised manual for the Profile of Mood States*. (San Diego, CA: Educational and Industrial Testing Services) (1992).
97. Beck AT, Steer RA, Brown GK. Manual for the Beck depression inventory-II. *San Antonio TX Psychol Corp* (1996) doi: 10.1037/t00742-000
98. Forsell Y. The Major Depression Inventory versus schedules for clinical assessment in neuropsychiatry in a population sample. *Soc Psychiatry Psychiatr Epidemiol* (2005) 40(3):209–13. doi: 10.1007/s00127-005-0876-3
99. Cohen S, Kamarck T, Mermelstein R. A global measure of perceived stress. *J Health Soc Behav* (1983) 24(4):385–96. doi: 10.2307/2136404
100. Cohen S, Williamson G. Perceived stress in a probability sample of the United States. *Soc Psychol Heal* (1988) 31–67. doi: 10.1111/j.1559-1816.1983.tb02325.x
101. Snaith RP, Hamilton M, Morley S, Humayan A, Hargreaves D, Trigwell P. A scale for the assessment of hedonic tone. The Snaith-Hamilton Pleasure Scale. *Br J Psychiatry* (1995) 167(1):99–103. doi: 10.1192/bjp.167.1.99
102. Treynor W, Gonzalez R, Nolen-Hoeksema S. Rumination reconsidered: A psychometric analysis. *Cognit Ther Res* (2003) 27:247–59. doi: 10.1023/A:1023910315561
103. Clayton AH, McGarvey EL, Clavet GJ. The changes in sexual functioning questionnaire (CSFQ): Development, reliability, and validity. *Psychopharmacol Bull* (1997) 33(4):731–45.
104. Eplov L, Giraldi A, Davidsen M, Garde K, Kamper-Jørgensen F. Sexual desire in a nationally representative danish population. *J Sex Med* (2007) 4(1):47–56. doi: 10.1111/j.1743-6109.2006.00396.x
105. Bech P. Rating scales in depression: Limitations and pitfalls. *Dialogues Clin Neurosci* (2006) 8(2):207–15.
106. Dahl J, Ormstad H, Aass HCD, Malt UF, Bendz LT, Sandvik L, et al. The plasma levels of various cytokines are increased during ongoing depression and are reduced to normal levels after recovery. *Psychoneuroendocrinology* (2014) 45:77–86. doi: 10.1016/j.psyneuen.2014.03.019
107. Dowlati Y, Herrmann N, Swardfager W, Liu H, Sham L, Reim EK, et al. A Meta-Analysis of Cytokines in Major Depression. *Biol Psychiatry* (2010) 67(5):446–57. doi: 10.1016/j.biopsych.2009.09.033
108. Howren MB, Lamkin DM, Suls J. Associations of depression with c-reactive protein, IL-1, and IL-6: A meta-analysis. *Psychosom Med* (2009). 71(2):171–86. doi: 10.1097/PSY.0b013e3181907c1b
109. Liu Y, Ho RCM, Mak A. Interleukin (IL)-6, tumour necrosis factor alpha (TNF- α) and soluble interleukin-2 receptors (sIL-2R) are elevated in patients with major depressive disorder: A meta-analysis and meta-regression. *J Affect Disord* (2012). 139(3):230–9. doi: 10.1016/j.jad.2011.08.003
110. Tammiste A, Jiang T, Fischer K, Mägi R, Krjutskov K, Pettai K, et al. Whole-exome sequencing identifies a polymorphism in the BMP5 gene associated with SSRI treatment response in major depression. *J Psychopharmacol (Oxford, England)* (2013). 27(10):915–20. doi: 10.1177/0269881113499829
111. Uhr M, Tontsch A, Namendorf C, Ripke S, Lucae S, Ising M, et al. Polymorphisms in the Drug Transporter Gene ABCB1 Predict Antidepressant Treatment Response in Depression. *Neuron* (2008). 57(2):203–9. doi: 10.1016/j.neuron.2007.11.017
112. Rasmussen ST, Andersen JT, Nielsen TK, Cejvanovic V, Petersen KM, Henriksen T, et al. Simvastatin and oxidative stress in humans: A randomized, Double-blinded, Placebo-controlled clinical trial. *Redox Biol* (2016). 9:32–8. doi: 10.1016/j.redox.2016.05.007
113. Wood SN. *Generalized Additive Models: An Introduction with R*. 2nd ed. Taylor and Francis Inc. (2017). doi: 10.1201/9781315370279.
114. Fisher PM, Ozenne B, Svarer C, Adamsen D, Lehel S, Baaré WF, et al. BDNF val66met association with serotonin transporter binding in healthy humans. *Transl Psychiatry* (2017). 7(2):e1029. doi: 10.1038/tp.2016.295
115. Hastie TT. The Elements of Statistical Learning Second Edition. *Math Intell* (2017). 7:228–30. doi: 10.1007/b94608_7
116. Polley EC, van der Laan MJ. Super Learner In Prediction. U.C. Berkeley Division of Biostatistics Working Paper Series. (2010). Working Paper 266.
117. Rush AJ, Fava M, Wisniewski SR, Lavori PW, Trivedi MH, Sackeim HA, et al. Sequenced treatment alternatives to relieve depression (STAR*D): Rationale and design. *Control Clin Trials* (2004). 25(1):119–42. doi: 10.1016/S0197-2456(03)00112-0

Conflict of Interest: C-TI was partly employed by company H. Lundbeck A/S. VF and GK have served as consultants for SAGE therapeutics, GK has served as a speaker in a Janssen sponsored symposium. MJ and VF has given talks sponsored by Lundbeck Pharma and MJ for Boehringer Ingelheim.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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The Diagnostic Value of the Combination of Serum Brain-Derived Neurotrophic Factor and Insulin-Like Growth Factor-1 for Major Depressive Disorder Diagnosis and Treatment Efficacy

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Specialty section:

This article was submitted to
Molecular Psychiatry,
a section of the journal
Frontiers in Psychiatry

Received: 27 November 2019

Accepted: 24 July 2020

Published: 13 August 2020

Citation:

Troyan AS and Levada OA (2020) The
Diagnostic Value of the Combination
of Serum Brain-Derived Neurotrophic
Factor and Insulin-Like
Growth Factor-1 for
Major Depressive Disorder
Diagnosis and Treatment Efficacy.
Front. Psychiatry 11:800.
doi: 10.3389/fpsy.2020.00800

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Background: Last decades of psychiatric investigations have been marked by a search for biological markers that can clarify etiology and pathogenesis, confirm the diagnosis, screen individuals at risk, define the severity, and predict the course of mental disorders. In our study, we aimed to evaluate if BDNF and IGF-1 serum concentrations separately and in combination might be used as biomarkers for major depressive disorder (MDD) diagnosis and treatment efficacy and to evaluate the relationships among those proteins and clinical parameters of MDD.

Methods: Forty-one MDD patients (according to DSM-5) and 32 healthy controls (HC) were included in this study. BDNF and IGF-1 serum concentrations, psychopathological (MADRS, CGI) and neuropsychological parameters (PDQ-5, RAVLT, TMT-B, DSST), functioning according to Sheehan Disability Scale were analyzed in all subjects at admission and 30 MDD patients after 8 weeks of vortioxetine treatment. Correlational analyses were performed to explore relationships between BDNF and IGF-1 and clinical characteristics. AUC-ROCs were calculated to determine if the value of serum BDNF and IGF-1 levels could serve for MDD diagnosis.

Results: MDD patients had significantly lower serum BDNF (727.6 ± 87.9 pg/ml vs. 853.0 ± 93.9 pg/ml) and higher serum IGF-1 levels (289.15 ± 125.3 ng/ml vs. 170.2 ± 58.2 ng/ml) compared to HC. Significant correlations were obtained between BDNF levels and MDD status, depressive episode (DE) severity, precipitating factors, executive functions disruption (TMT-B, RAVLT immediate recall scores) and all subdomains of functioning. As for IGF-1, correlations were found between IGF-1 level and MDD status, DE severity, number and duration of DE, parameters of subjective and objective cognitive functioning (PDQ-5, RAVLT, TMT-B, DSST scores), and all subdomains of functioning. The associations between IGF-1 concentrations and cognitive tests' performance were stronger than those of BDNF. Separately both BDNF and IGF-1 demonstrated good

discriminating ability for MDD diagnosis with AUC of 0.840 and 0.824, respectively. However, the combination of those neurotrophins had excellent diagnostic power to discriminate MDD patients from HC, providing an AUC of 0.916. Vortioxetine treatment significantly increased BDNF and attenuated IGF-1 serum concentrations, improved all psychopathological and neuropsychological parameters and functioning.

Conclusions: The combination of IGF-1 and BDNF might be considered as a diagnostic combination for MDD.

Keywords: major depressive disorder, insulin-like growth factor-1, brain-derived neurotrophic factor, biomarkers, cognitive functions, vortioxetine

INTRODUCTION

The last decades of psychiatric investigations have been marked by a search for biological markers that can clarify etiology and pathogenesis, confirm a diagnosis, screen individuals at risk, define the severity, and predict the course of mental disorders (1–3). With respect to major depressive disorder (MDD), abnormal neuroplasticity in cerebral regions, responsible for emotional and cognitive processing, is considered to be one of the key pathogenic mechanisms (4, 5) that potentially have a biomarker value. It is associated with alterations in the expression of neurotrophic factors, such as brain-derived neurotrophic factor (BDNF), neurotrophin-3, neurotrophin-4/5, nerve growth factor, insulin-like growth factor (IGF-1), etc. (6–8). The vital role of the neurotrophins is explained by their involvement in the processes of neuronal growth, differentiation, maturation, and survival, synaptic transmission, an equilibrium between neuroregeneration and neurodegeneration, as well as memory formation (9). Therefore, there is reason to believe that two proteins—BDNF and IGF-1—can be used as marker molecules for the MDD diagnosis and therapy effectiveness (7, 8).

Recent studies have shown lower BDNF concentrations in the brains of suicidal MDD persons (10, 11). Further, postmortem studies of MDD individuals revealed an association between the diminished BDNF expression in hippocampal areas and a decrease in the volume of these anatomical structures (12). According to meta-analyses, serum and plasma BDNF reductions have been noted in antidepressant-free individuals with MDD as compared to healthy subjects (13–15). The decreased peripheral BDNF levels normalized in reprisal for several interventions [e.g., antidepressants (16, 17), electroconvulsive therapy (18, 19), aerobic exercise (20)]. Thus, we can reasonably assume that the decline in peripheral (plasma/serum) BDNF levels reflects its reduced expression in the brain and could serve as a neurobiological marker of impaired

neuroplastic processes in MDD. In addition, the increase of the neurotrophin concentration could indicate therapy effectiveness.

According to our recent systematized data, there are discrepancies in IGF-1 levels in MDD patients across the studies, although the majority demonstrate higher levels of peripheral IGF-1 compared to healthy controls (HC) (8). The elevation of peripheral total IGF-1 concentrations and its decline after antidepressants' treatment were established in several studies in MDD patients (21–23), moreover, it was reported that only patients in remission had attenuated IGF-1 concentrations following treatment (21, 22). The enhancement of the peripheral IGF-1 expression in MDD may be a compensatory mechanism in response to its brain synthesis decrease (24) or diminished neurotrophin bioavailability due to the hyposensitivity of IGF-1 receptors under the neuroinflammatory stress (25). In our recent work, we assumed that the activity of the cerebral-hepatic axis rise in response to inadequate cerebral IGF-1 levels (26). As a consequence, the production of IGF-1 in the liver increases. When cerebral IGF-1 production is restored, the liver IGF-1 production and its blood concentration decrease.

However, the clinical value, sensitivity, specificity, and predictive significance of a single biomarker for MDD remain doubtful. To solve this problem, the approach that uses the cumulative sensitivity and specificity of biomarkers' combination might be applicable (27). Therefore, in our study we aimed: 1) to evaluate if BDNF and IGF-1 serum concentrations separately and in combination might be biomarkers for MDD diagnosis and treatment response; 2) to evaluate the relationships between clinical MDD parameters and serum levels of the above-mentioned neurotrophins.

MATERIALS AND METHODS

Study Design

This was a case-control study, which included 73 participants aged 18 to 65 years. Outpatients (n=41) with MDD diagnosis according to DSM-5 criteria (28) were included through Zaporizhzhia Regional Clinical Psychiatric Hospital, Ukraine. Eligibility criteria for the study participants were described elsewhere (29). Before entering the study, all patients had received no actual antidepressant medication. Subjects were excluded if they had any other psychiatric diagnosis, high suicidal risk, substance dependence/abuse over the past year,

Abbreviations: AUC-ROC, areas under the receiver operating characteristic curves; BDNF, brain-derived neurotrophic factor; CGI-I, clinical global impression improvement; CGI-S, clinical global impression severity; DE, depressive episode; DSM-5, Diagnostic and Statistical Manual of Mental Disorders, Fifth Edition; DSST, digit symbol substitution test; IGF-1, insulin-like growth factor-1; HC, healthy controls; MADRS, Montgomery-Asberg depression rating scale; MDD, major depressive disorder; PDQ-5, perceived deficit questionnaire-5; RAVLT, Rey auditory verbal learning test; ROC, receiver-operating characteristic; SD, standard deviation; SPSS, statistical package for the social sciences; TMT-B, trail making test B.

significant neurological disorders, head trauma, unstable medical conditions, history of endocrine diseases, psychotic symptoms, the risk for the hypomanic switch (29). Thirty-two healthy controls (HC) with no current psychiatric disorder were enrolled within the same period that the MDD patients were included. HC were excluded based on the use of medications and/or illicit drugs; the intake of alcohol within 48 h of the study visit; and the presence of an unstable medical condition, which could affect cognitive function (29).

All the participants gave written informed consent to take part in the study and attended a baseline visit to undergo the evaluations. The appraisal was reiterated in 30 patients after 8 weeks of vortioxetine treatment 10–20 mg per day.

The study was approved by the local ethics committee and performed following the ethical standards laid down in the 1964 Declaration of Helsinki and its later amendments and registered at ClinicalTrials.gov (NCT03187093).

Clinical Assessments

The severity of depression at baseline and changes after treatment were assessed with MADRS (30), Clinical Global Impression Severity (CGI-S) and Improvement (CGI-I) scales (31). Perceived Deficit Questionnaire-5 (PDQ-5), which measures perceived difficulties of cognitive functioning, was used to evaluate subjective cognitive functioning (32). Functioning was rated using the Sheehan Disability Scale (SDS) (33).

Neuropsychological Assessments

To describe neurocognitive functioning, a battery of neuropsychological tests covering the most impaired cognitive domains in MDD was administered to all participants. The tests were managed using paper and pencil. The instruments included:

1. Rey Auditory Verbal Learning Test (RAVLT), to evaluate immediate verbal memory, retroactive and proactive interference effects, delayed recall, and recognition (34);
2. Trail Making Test B (TMT-B), to assess processing speed, executive function i.e., set-shifting (34);
3. Digit Symbol Substitution Test (DSST), to assess processing speed, executive function, learning, memory, attention, and concentration (34).

The procedures of the tests were described previously (29).

BDNF and IGF-1 Measurements

In MDD patients and HC, BDNF, and IGF-1 concentrations were evaluated in the morning. Blood was taken by a venous puncture between 8:00 and 11:00 a.m. within the first 2 days after clinical assessments in 73 participants at baseline and 30 patients after 8 weeks of vortioxetine (10–20 mg per day) treatment. Blood was centrifuged at 3000 g for 10 min, and the serum was stored at -20°C until further processing. IGF-1 was assessed with the chemiluminescence immunoassay Immulite 2500 (Siemens AG, Germany) and Human IGF-1 Quantikine ELISA Kit (R&D Systems Inc., Minneapolis, United States). The measurement

range for IGF-1 was from 20 to 1600 ng/ml. BDNF was measured using the chemiluminescence immunoassay «Sunrise» (TEKAN, Austria, GmbH) and BDNF Sandwich ELISA Kit («Millipore-ChemiKineTM»). The measurement range was from 15 to 1000 pg/ml.

Statistical Analysis

Data analyses were done with the SPSS for Windows, Version 20.0 (SPSS Inc., United States). The results were presented as median (interquartile range) or means (SDs) or percentages. The statistical significance of between-group comparisons was determined using non-parametric and parametric criteria when appropriate. The relationships between demographic and clinical parameters and BDNF or IGF-1 concentrations were assessed with Spearman's or Pearson's correlation coefficient. Thereafter, the areas under the receiver operating characteristic curves (AUC-ROC) were calculated to determine if the value of serum BDNF or IGF-1 level could discriminate patients with MDD from HC. A cutoff was derived from the ROC curve with empirical optimal sensitivity and specificity. In addition, a ROC curve for the combination of two biomarkers (BDNF and IGF-1) was built to assess if this combination has a higher value to discriminate patients with MDD from HC. For this purpose, we ran a binary logistic regression to get the probability and built a ROC curve using the probability as the test variable. The statistical threshold was set at $p < 0.05$.

RESULTS

Characteristics of Controls and MDD Patients

Table 1 demonstrates the main demographic, psychopathological, neuropsychological, and functional characteristics of MDD and HC groups. There were no differences in age, gender, and level of education between groups. Surveyed cohorts had a higher percentage of women than men. The mean years of education in HC and MDD patients was 15 years. Although the number of people with predisposing factors (those that put a person at risk of developing MDD, which included the presence of childhood psychotrauma, persistent stress, and MDD in relatives) was higher in MDD patients, only the presence of precipitating factors (specific events or triggers to the onset of the current DE) significantly differed MDD patients from HC.

Besides the expected statistical difference in MDD patients and HC on MADRS and CGI-S total scores, a significant distinction in neuropsychological test performance was found between the comparison groups. MDD participants were significantly worse ($p < 0.0001$) in executive functioning (DSST, TMT-B scores), processing speed (DSST, TMT-B scores), set-shifting (TMT-B), and all parameters of verbal memory (RAVLT subtests). MDD patients also had a significantly lower level of subjective cognitive functioning as compared to HC.

After 8 weeks of vortioxetine treatment in 30 MDD patients, we revealed that the intake of the antidepressant significantly improved the clinical parameters of patients (**Table 1**). Thus, we

TABLE 1 | Demographic, psychopathological, neuropsychological, functional characteristics, and serum BDNF and IGF-1 levels in healthy controls and MDD patients.

| Variables | HC (n = 32) | MDD (n = 41) | p | Treatment group(n = 30) | | p |
|--|------------------------------|-----------------------------|-------------------|------------------------------|-----------------------------------|-------------------|
| | | | | Pre-treatment | Post-treatment | |
| Demographic characteristics | | | | | | |
| Women, n (%) | 20 (62.5) ^b | 27 (65.9) | 0.77 | 20 (69) | — | |
| Age, years [§] | 38.0 (12.2) ^a | 36.4 (12.8) | 0.60 | 35.1 (12.9) | — | |
| Education, years [§] | 15.4 (1.9) ^a | 14.7 (2.0) | 0.16 | 14.6 (2.1) | — | |
| Childhood psychotrauma, n (%) | 4 (12.5) ^b | 13 (31.7) | 0.05 | 10 (34.5) | — | |
| Persistent stress, n (%) | 12 (37.5) ^b | 22 (53.7) | 0.17 | 14 (48.3) | — | |
| Precipitating factors, n (%) | 3 (9.4) ^b | 26 (63.4)** | <0.0001 | 20 (69.0)** | | |
| Number of DE | — | 1 (1-2) | — | 1 (1-2.5) | — | |
| MDD in relatives, n (%) | 4 (12.5) ^b | 12 (29.3) | 0.09 | 10 (34.5) | — | |
| History of DE, n (%) | — | 14 (34.1) | — | 11 (37.9) | — | |
| Clinical assessments | | | | | | |
| <i>Psychopathological</i> | | | | | | |
| MADRS total score | 2 (0-3.5) ^c | 28 (21.5-31.5)** | <0.0001 | 29 (24.5-33) ^d | 5 (2.5-10)^{##} | <0.0001 |
| CGI-S score | 1 (1-1) ^c | 4 (4-4.5)** | <0.0001 | 4 (4-5) ^d | 2 (1-3)^{##} | <0.0001 |
| <i>Patient-reported cognitive symptoms</i> | | | | | | |
| PDQ-5 total score | 1 (0-2) ^c | 6 (4-11.5)** | <0.0001 | 7 (4-12.5) ^d | 2 (1-4)^{##} | <0.0001 |
| <i>Neuropsychological</i> | | | | | | |
| RAVLT immediate recall total | 65.5 (58.25-69) ^c | 50.5 (45-54.5)** | <0.0001 | 51 (45-54) ^d | 69 (65-72.5)^{##} | <0.0001 |
| RAVLT proactive interference | 8 (6-9) ^c | 6 (5-7)** | <0.0001 | 6 (5-7) ^d | 8 (6-9)^{##} | <0.0001 |
| RAVLT retroactive interference | 15 (13.25-15) ^c | 11 (9-13)** | <0.0001 | 11 (9-13) ^d | 15 (14-15)^{##} | <0.0001 |
| RAVLT delayed recall | 15 (13-15) ^c | 11 (9-13)** | <0.0001 | 11 (9-13) ^d | 15 (14-15)^{##} | <0.0001 |
| RAVLT delayed recognition | 15 (15-15) ^c | 15 (14-15)* | 0.003 | 15 (13.5-15) ^d | 15 (15-15)[#] | 0.001 |
| TMT-B (s) [§] | 52.4 (12.8) ^a | 72.3 (21.2)** | <0.0001 | 72.4 (23.0) ^e | 45.4 (15.6)^{##} | <0.0001 |
| DSST | 63 (55.5-68) ^c | 50.5 (44.25-59.75)** | <0.0001 | 54 (44.5-60.5) ^d | 62 (52-73)^{##} | <0.0001 |
| Patient-rated functioning | | | | | | |
| SDS Work score | 0 (0-0) ^c | 6 (5-8)** | <0.0001 | 6 (5-8) ^d | 1 (0-3)^{##} | <0.0001 |
| SDS Social score | 0 (0-0) ^c | 7 (3.75-9.25)** | <0.0001 | 7 (4.5-10) ^d | 1 (0-3)^{##} | <0.0001 |
| SDS Family score | 0 (0-0) ^c | 6 (4-8)** | <0.0001 | 6 (4.5-8) ^d | 1 (0-2.5)^{##} | <0.0001 |
| SDS Total score | 0 (0-1.75) ^c | 19 (12-24.5)** | <0.0001 | 19.5 (13.25-25) ^d | 2.5 (0-8)^{##} | <0.0001 |
| SDS absenteeism, days | 0 (0-0) ^c | 0 (0-1)** | <0.0001 | 0 (0-1) ^d | 0 (0-0)[#] | 0.007 |
| SDS presenteism, days | 0 (0-0) ^c | 3 (2-5)** | <0.0001 | 4 (2.25-5.75) ^d | 0 (0-1.5)^{##} | <0.0001 |
| Serum protein levels | | | | | | |
| BDNF (pg/ml) [§] | 853.0 (93.9) ^a | 727.6 (87.9)** | <0.0001 | 737.3 (90.4) ^e | 905.3 (59.6)^{###} | <0.0001 |
| IGF-1 serum level (ng/ml) [§] | 170.2 (58.2) ^a | 289.2 (125.3)** | <0.0001 | 288.2 (132.6) ^e | 173.4 (71.2)^{##} | <0.0001 |

Data are presented as median (upper-lower quartile) unless otherwise stated; ^sdata are presented as means (SD).

BDNF, brain-derived neurotrophic factor; CGI-S, clinical global impression—severity of illness; DE, depressive episode; DSST, digit symbol substitution test; HC, healthy controls; IGF-1, insulin-like growth factor; MADRS, Montgomery-Asberg depression rating scale; MDD, patients with major depressive disorder; PDQ-5, perceived deficits questionnaire—5 items; RAVLT, Rey auditory verbal learning test; SDS, Sheehan disability scale; TMT-B, trail making test part B.

^aANOVA (analysis of variance) analysis, controls vs. MDD patients.

^bChi-square test, controls vs. MDD patients.

^cMann-Whitney U-test, controls vs. MDD patients.

^dWilcoxon test (paired samples), "Pre-treatment" vs. "Post-treatment".

^ePaired-samples t-test, "Pre-treatment" vs. "Post-treatment".

Compared with controls, *p < 0.05, **p < 0.01. Compared with "Pre-treatment", #p < 0.05, ##p < 0.0001.

observed a significant decrease in depression severity (MADRS, CGI-S, SGI-I scores), improvement of cognitive impairment (measured as subjectively as objectively), and functioning.

Serum Concentrations of BDNF and IGF-1

Table 1 shows serum protein levels of BDNF and IGF-1 in HC and MDD patients. It was demonstrated that serum BDNF concentrations were significantly lower in MDD persons compared to HC (p < 0.0001), whereas the concentrations of IGF-1 were significantly higher in patients than HC (p < 0.0001). Although IGF-1 concentrations were slightly higher in women than in men as in MDD [299.4 (131.5) ng/ml vs. 271.5 (116.5)] as in HC group [174.1 (53.4) vs. 164.2 (67.2)], those differences did not reach a significant level.

After 8 weeks of treatment with vortioxetine, BDNF levels were significantly higher in post-treatment than pre-treatment (p < 0.0001), moreover, they were prominently higher than in HC (F = 9.36, p = 0.003). Whereas, IGF-1 concentrations in MDD group post-treatment were significantly lower than pretreatment (p < 0.0001) and not significantly different from HC (F = 1.86, p = 0.18).

Correlations of Serum BDNF and IGF-1 Levels With Clinical Variables

Next, we performed correlational analyses to determine possible associations between serum BDNF and IGF-1 concentrations and demographic and clinical parameters in the whole sample (Table 2). We established prominent inverse relationships between BDNF concentrations and MDD status (r = -0.57, p <

TABLE 2 | Spearman's/Pearson's correlations between demographic, psychopathological, neuropsychological characteristics, and serum BDNF and IGF-1 levels in MDD patients and healthy controls.

| Variables | BDNF (pg/ml) | p | IGF-1 (ng/ml) | p |
|--------------------------------------|----------------|-------------------|----------------|-------------------|
| Demographic characteristics | | | | |
| Age, years | 0.02 | 0.9 | -0.17 | 0.16 |
| Gender | 0.15 | 0.20 | 0.10 | 0.44 |
| Depression status | -0.57** | <0.0001 | 0.50** | <0.0001 |
| Persistent stress | 0.06 | 0.59 | 0.01 | 0.93 |
| Precipitating factors | -0.33** | 0.004 | 0.20 | 0.11 |
| Recurrence of DE | -0.03 | 0.79 | 0.16 | 0.19 |
| Number of DE | -0.04 | 0.75 | 0.43** | 0.001 |
| Duration of DE, weeks | -0.02 | 0.85 | 0.37** | 0.002 |
| MDD in relatives | -0.12 | 0.32 | 0.05 | 0.68 |
| Clinical assessments | | | | |
| CGI-S score | -0.44** | <0.0001 | 0.45** | <0.0001 |
| PDQ-5 score | -0.19 | 0.16 | 0.43** | <0.0001 |
| MADRS total score | -0.43** | <0.0001 | 0.46** | <0.0001 |
| RAVLT immediate recall total score | 0.33** | 0.007 | -0.42** | 0.001 |
| RAVLT proactive interference score | 0.19 | 0.11 | -0.40** | 0.001 |
| RAVLT retroactive interference score | 0.17 | 0.16 | -0.42** | 0.001 |
| RAVLT delayed recall score | 0.23 | 0.06 | -0.37** | 0.003 |
| RAVLT delayed recognition score | 0.18 | 0.14 | -0.19 | 0.15 |
| TMT-B (s) | -0.27* | 0.03 | 0.55** | <0.0001 |
| DSST score | 0.24* | 0.048 | -0.45** | <0.0001 |
| SDS Work score | -0.37** | 0.003 | 0.40** | 0.001 |
| SDS Social score | -0.46** | <0.0001 | 0.39** | 0.002 |
| SDS Family score | -0.38** | 0.002 | 0.40** | 0.001 |
| SDS Total score | -0.43** | <0.0001 | 0.43** | <0.0001 |
| SDS absenteeism, days | -0.12 | 0.35 | 0.23 | 0.07 |
| SDS presentism, days | -0.36** | 0.003 | 0.31* | 0.01 |
| Serum proteins' levels | | | | |
| Serum BDNF level | 1 | — | -0.17 | 0.18 |
| Serum IGF-1 level | -0.17 | 0.18 | 1 | — |

BDNF, brain-derived neurotrophic factor; CGI-S, clinical global impression—severity of illness; DE, depressive episode; DSST, digit symbol substitution test; HC, healthy controls; IGF-1, insulin-like growth factor; MADRS, Montgomery-Asberg depression rating scale; MDD, patients with major depressive disorder; PDQ-5, perceived deficits questionnaire—5 items; RAVLT, Rey auditory verbal learning test; SDS, Sheehan disability scale; TMT-B, trail making test part B.

* $p < 0.05$ and ** $p < 0.01$.

0.01), precipitating factors ($r = -0.33$, $p < 0.01$), CGI-S ($r = -0.44$, $p < 0.01$), MADRS ($r = -0.43$, $p < 0.01$), TMT-B score ($r = -0.27$, $p < 0.05$), all subdomains of functioning and positive correlations between serum BDNF levels and RAVLT immediate recall level ($r = 0.33$, $p < 0.01$).

As for IGF-1, positive correlations were found between IGF-1 level and MDD status ($r = 0.50$, $p < 0.01$), number ($r = 0.43$, $p < 0.01$) and duration of DE ($r = 0.37$, $p < 0.01$), CGI-S score ($r = 0.45$, $p < 0.01$), PDQ-5 score ($r = 0.43$, $p < 0.01$), MADRS score ($r = 0.46$, $p < 0.01$), TMT-B score ($r = 0.55$, $p < 0.01$) and all subdomains of functioning and negative associations between IGF-1 and the performance of RAVLT and DSST tests. Moreover, the correlations between IGF-1 concentrations and performance of cognitive tests were higher than that of BDNF.

Serum BDNF and IGF-1 Levels for MDD Diagnosis

The discriminating ability of serum proteins' level to separate MDD participants from HC was determined with ROC analysis. The diagnostic value of BDNF and IGF-1 is shown in **Figure 1**. Separately BDNF (cutoff < 763.17 pg/ml, sensitivity = 81%, specificity = 73%) and IGF-1 (cutoff > 178.00 ng/ml, sensitivity = 84%, specificity = 64%) demonstrated good diagnostic effectiveness, with AUC of 0.840 and 0.824, respectively. However, the combination of two neurotrophins showed excellent diagnostic value for MDD diagnosis, with an AUC of 0.916.

DISCUSSION

This is the first study to investigate the diagnostic power of the combination of two biomarkers—BDNF and IGF-1—for MDD diagnosis. In our study, we found significantly lower BDNF and higher IGF-1 serum concentrations in patients compared with HC. Following 8 weeks of vortioxetine therapy, serum BDNF

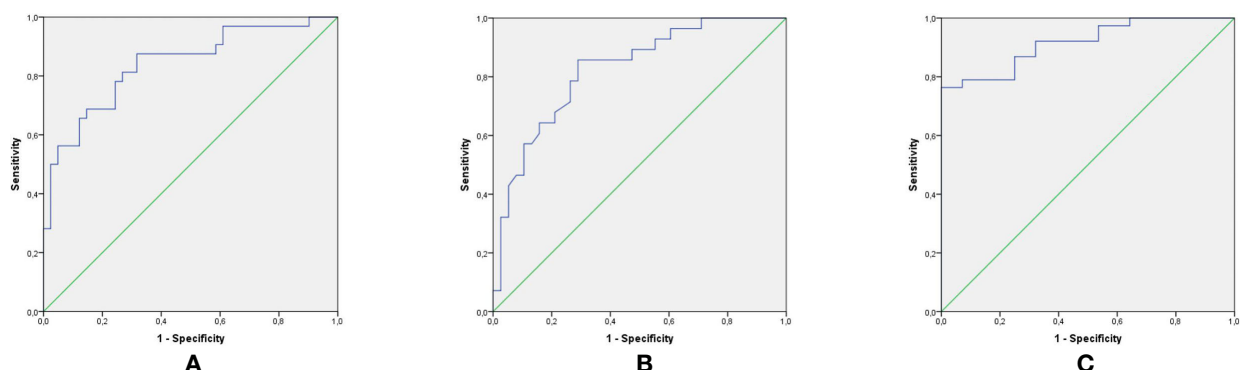


FIGURE 1 | Diagnostic value of serum BDNF and IGF-1 for MDD diagnosis. (A) ROC curve for BDNF to diagnose MDD ($p < 0.0001$, AUC: 0.840; sensitivity: 81% and specificity: 73% for a cutoff of < 763.17 pg/ml). (B) ROC curve for IGF-1 to diagnose MDD ($p < 0.0001$, AUC: 0.824; sensitivity: 84% and specificity: 64% for a cutoff of > 178.00 ng/ml). (C) ROC curve the combination of BDNF and IGF-1 to diagnose MDD ($p < 0.0001$, AUC: 0.916). AUC, Area Under the ROC Curve; BDNF, Brain-Derived Neurotrophic Factor; IGF-1, Insulin-Like Growth Factor; MDD, major depressive disorder; ROC, receiver-operating characteristic.

levels were significantly higher in post-treatment than pre-treatment and compared to the controls. IGF-1 concentrations in subjects post-treatment were significantly lower than pretreatment and not different from the controls. Significant correlations were obtained between serum BDNF levels and MDD status, the severity of the current DE, presence of precipitating factors, executive functions' disruption (TMT-B and RAVLT immediate recall scores), and all subdomains of functioning. As for IGF-1, correlations were found between IGF-1 level and MDD status, severity, number, and duration of current DE, subjective and objective cognitive functioning (PDQ-5, RAVLT subtests, TMT-B, DSST scores) and all subdomains of functioning. The associations between IGF-1 concentrations and the performance of cognitive tests were stronger than that of BDNF. Separately both BDNF and IGF-1 demonstrated good discriminating ability for MDD diagnosis; however, the combination of these two proteins had excellent diagnostic power to discriminate MDD patients from HC, providing the AUC of 0.916.

Firstly, we revealed that MDD patients had decreased BDNF concentrations and that these concentrations returned to normal after vortioxetine intake. These results are in line with previous works that found lower serum and plasma BDNF concentrations in depressed persons in comparison with healthy subjects (13, 14, 17, 27, 35–42). Several antidepressants were shown to increase serum/plasma BDNF levels in MDD persons compared to pre-treatment levels, including agomelatine (41), duloxetine (27), escitalopram (27, 43, 44), fluoxetine (41, 45), milnacipran (36), paroxetine (36, 38, 44), sertraline (44), venlafaxine (44), and vortioxetine (17, 46). However, recent works have shown that not all antidepressants increase BDNF concentrations (47), and antidepressant-induced rise in BDNF concentrations are more substantial in responders compared to non-responders (48–51). Moreover, it was demonstrated that high-intensity exercises (52), electroconvulsive therapy (15, 18, 19, 53), and deep brain stimulation (54) also increase BDNF levels. In animal studies, it has been revealed that vortioxetine enhances the behaviors of depressed rodents probably by influencing the cAMP/CREB/BDNF signal pathway and promoting the expression of BDNF in the dorsal and ventral hippocampus of the animals (55, 56). Moreover, it was shown that vortioxetine, but not fluoxetine, raised hippocampal BDNF concentrations in rats (57). Another study demonstrated that vortioxetine amplified the number of synapses and mitochondria substantially, together with increased BDNF levels, while fluoxetine showed no effect (58). It was suggested that fast changes in BDNF concentrations and synaptic/mitochondria plasticity of the hippocampus during vortioxetine treatment may be attributed to vortioxetine's modulation of 5-HT receptors (58). Longitudinal studies in humans revealed that it is more likely that BDNF is a biomarker for the state of MDD and treatment response rather than a risk factor for MDD (15, 40).

In our study, we obtained significant associations between serum BDNF levels and MDD severity, precipitating factors, executive functions' disruption (performance of TMT-B and RAVLT immediate recall), and all subdomains of functioning. Previous studies in MDD patients have found correlations between BDNF

levels and the presence of melancholic features and psychomotor retardation (59), DE severity (36, 60) and duration (60), and cognitive decline during performances of TMT-B (61) and verbal delayed recall (62). Nevertheless, one study evidenced no correlations between patients' plasma BDNF levels and cognitive functioning (38).

Regarding peripheral IGF-1 increase in MDD patients and its decrease after vortioxetine treatment, our results are consistent with some studies (21, 22, 24, 25, 63–65). Data on a significant decrease of serum IGF-1 concentrations during antidepressant treatment are supported by previous investigations, in which amitriptyline, doxepin, fluoxetine, and paroxetine led to a substantial decline of peripheral IGF-1 levels (21, 22, 66). Nevertheless, one study showed that free IGF-1, not total, concentrations did not change after antidepressant treatment (22). The authors suggested that discrepancies between total and biologically active free IGF-1 levels could be due to adaptation mechanisms to changes typically found in major depression (22).

Although in our study we found no significant differences in IGF-1 levels between women and men in both MDD and HC groups, several previous studies have shown variability across the genders in IGF-1 levels in MDD patients (67–69). These differences between genders may be explained by variations of sex hormone or fluctuations of growth hormone and IGF-1-binding protein (67).

The reported here correlations between IGF-1 level and severity, number, and duration of DE, subjective and objective cognitive functioning (PDQ-5, RAVLT subtests, TMT-B, DSST), all subdomains of functioning are in line with our previous results (23). Nevertheless, here we pointed out that the associations for IGF-1 concentrations and the level of performance of cognitive tests were stronger than those for BDNF levels.

Lastly, the results of ROC-analysis demonstrated that BDNF and IGF-1 separately provided fairly good diagnostic power to separate MDD patients from HC, meanwhile the combination of these neurotrophins showed excellent diagnostic value. Therefore, serum BDNF and IGF-1 levels might be a potential biomarker combination as a diagnostic test for MDD. To the best of our knowledge, we were the first to combine those two serum proteins for MDD diagnosis. Previously it was shown that the arrangement of tPA, BDNF, TrkB, proBDNF, and p75NTR might be a diagnostic biomarker panel for MDD (27) and the combination of BDNF, FGF-2, TNF- α , and 5-HT may predict the efficacy of escitalopram therapy (70).

Limitations

Our study was limited by its small sample size, therefore, obtained data need to be replicated in larger samples to confirm our findings.

CONCLUSIONS

Patients with MDD had significantly lower BDNF and higher IGF-1 serum concentrations compared to controls. Vortioxetine

treatment normalized those disruptions, moreover, after 8 weeks of treatment BDNF concentrations in MDD individuals were significantly higher compared to HC. In the whole sample at baseline BDNF levels correlated with the presence of precipitating factors, meanwhile, IGF-1 levels—with the number and duration of DE. Therefore, we can suggest that BDNF is related to acute stress. As for cognition, IGF-1 had stronger associations than BDNF with the disturbances in different cognitive domains, also, it correlated with the subjective cognitive level. Both factors were associated with functioning in different spheres of life. As BDNF as IGF-1 were significantly associated with MDD status and severity of a current episode and separately demonstrated good discriminating ability for MDD diagnosis. However, their combination had excellent diagnostic power to discriminate MDD patients from HC. Thus, the combination of IGF-1 and BDNF might be considered as a biomarker panel for MDD diagnosis.

DATA AVAILABILITY STATEMENT

The datasets generated for this study are available on request to the corresponding authors.

REFERENCES

- Macaluso M, Preskorn SH. How biomarkers will change psychiatry: from clinical trials to practice. Part I: introduction. *J Psychiatr Pract* (2012) 18 (2):118–21. doi: 10.1097/01.pra.0000413277.11091.25
- Kalia M, Costa E Silva J. Biomarkers of psychiatric diseases: current status and future prospects. *Metabolism* (2015) 64(3 Suppl 1):S11–5. doi: 10.1016/j.metabol.2014.10.026
- Venkatasubramanian G, Keshavan MS. Biomarkers in Psychiatry - A Critique. *Ann Neurosci* (2016) 23(1):3–5. doi: 10.1159/000443549
- Liu B, Liu J, Wang M, Zhang Y, Li L. From Serotonin to Neuroplasticity: Evolution of Theories for Major Depressive Disorder. *Front Cell Neurosci* (2017) 28(11):305. doi: 10.3389/fncel.2017.00305
- Boku S, Nakagawa S, Toda H, Hishimoto A. Neural basis of major depressive disorder: Beyond monoamine hypothesis. *Psychiatry Clin Neurosci* (2018) 72 (1):3–12. doi: 10.1111/pcn.12604
- Duman RS. Role of neurotrophic factors in the etiology and treatment of mood disorders. *Neuromol Med* (2004) 5(1):11–25. doi: 10.1385/NMM:5:1:011
- Phillips C. Brain-Derived Neurotrophic Factor, Depression, and Physical Activity: Making the Neuroplastic Connection. *Neural Plast* (2017) 2017:7260130. doi: 10.1155/2017/7260130
- Levada OA, Trojan AS. Insulin-like growth factor-1: a possible marker for emotional and cognitive disturbances, and treatment effectiveness in major depressive disorder. *Ann Gen Psychiatry* (2017) 26(16):38. doi: 10.1186/s12991-017-0161-3
- Mitre M, Mariga A, Chao MV. Neurotrophin signalling: novel insights into mechanisms and pathophysiology. *Clin Sci (Lond)* (2017) 131(1):13–23. doi: 10.1042/CS20160044
- Guilloux JP, Douillard-Guilloux G, Kota R, Wang X, Gardier AM, Martinowich K, et al. Molecular evidence for BDNF- and GABA-related dysfunctions in the amygdala of female subjects with major depression. *Mol Psychiatry* (2012) 17(11):1130–42. doi: 10.1038/mp.2011.113
- Hayley S, Du L, Litteljohn D, Palkovits M, Faludi G, Merali Z, et al. Gender and brain regions specific differences in brain derived neurotrophic factor protein levels of depressed individuals who died through suicide. *Neurosci Lett* (2015) 23(600):12–6. doi: 10.1016/j.neulet.2015.05.052

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by the local ethics committee of State Institution Zaporizhzhia Medical Academy of Postgraduate Education Ministry of Health of Ukraine. The patients/participants provided their written informed consent to participate in this study.

AUTHOR CONTRIBUTIONS

OL supervised the study. OL and AT were involved in the concept of the study and collection of the data. Both authors equally contributed to data preparation and literature search. OL and AT analyzed the data and wrote the first draft of the manuscript. All authors contributed to the article and approved the submitted version. of the manuscript.

ACKNOWLEDGMENTS

We are grateful to Viktoriya V. Levada for substantial help in data collection.

- Dunham JS, Deakin JF, Miyajima F, Payton A, Toro CT. Expression of hippocampal brain-derived neurotrophic factor and its receptors in Stanley consortium brains. *J Psychiatr Res* (2009) 43(14):1175–84. doi: 10.1016/j.jpsychires.2009.03.008
- Molendijk ML, Spinhoven P, Polak M, Bus BA, Penninx BW, Elzinga BM. Serum BDNF concentrations as peripheral manifestations of depression: evidence from a systematic review and meta-analyses on 179 associations (N=9484). *Mol Psychiatry* (2014) 19(7):791–800. doi: 10.1038/mp.2013.105
- Levada OA, Cherednichenko NV. Brain-derived neurotrophic factor (BDNF): neurobiology and marker value in neuropsychiatry. *Lik Sprava* (2015) 3-4:15–25.
- Kishi T, Yoshimura R, Ikuta T, Iwata N. Brain-Derived Neurotrophic Factor and Major Depressive Disorder: Evidence from Meta-Analyses. *Front Psychiatry* (2018) 8:308. doi: 10.3389/fpsy.2017.00308
- Matriciano F, Bonaccorso S, Ricciardi A, Scaccianoce S, Panaccione I, Wang L, et al. Changes in BDNF serum levels in patients with major depression disorder (MDD) after 6 months treatment with sertraline, escitalopram, or venlafaxine. *J Psychiatr Res* (2009) 43(3):247–54. doi: 10.1016/j.jpsychires.2008.03.014
- Sagud M, Nikolac Perkovic M, Vuksan-Cusa B, Maravic A, Svob Strac D, Mihaljevic Peles A, et al. A prospective, longitudinal study of platelet serotonin and plasma brain-derived neurotrophic factor concentrations in major depression: effects of vortioxetine treatment. *Psychopharmacol (Berl)* (2016) 233(17):3259–67. doi: 10.1007/s00213-016-4364-0
- Brunoni AR, Baeken C, Machado-Vieira R, Gattaz WF, Vanderhasselt MA. BDNF blood levels after electroconvulsive therapy in patients with mood disorders: a systematic review and meta-analysis. *World J Biol Psychiatry* (2014) 15(5):411–8. doi: 10.3109/15622975.2014.892633
- Rocha RB, Dondossola ER, Grande AJ, Colonetti T, Ceretta LB, Passos IC, et al. Increased BDNF levels after electroconvulsive therapy in patients with major depressive disorder: A meta-analysis study. *J Psychiatr Res* (2016) 83:47–53. doi: 10.1016/j.jpsychires.2016.08.004
- Kallies G, Rapp MA, Fydrich T, Fehm L, Tschorn M, Terán C, et al. Serum brain-derived neurotrophic factor (BDNF) at rest and after acute aerobic exercise in major depressive disorder. *Psychoneuroendocrinology* (2019) 102:212–5. doi: 10.1016/j.psyneuen.2018.12.015
- Deuschle M, Blum WF, Strasburger CJ, Schweiger U, Weber B, Körner A, et al. Insulin-like growth factor-I (IGF-I) plasma concentrations are increased in

- depressed patients. *Psychoneuroendocrinology* (1997) 22:493–503. doi: 10.1016/S0306-4530(97)00046-2
22. Weber-Hamann B, Blum WF, Kratzsch J, Gilles M, Heuser I, Deuschle M. Insulin-like growth factor-I (IGF-I) serum concentrations in depressed patients: relationship to saliva cortisol and changes during antidepressant treatment. *Pharmacopsychiatry* (2009) 42:23–8. doi: 10.1055/s-0028-1085442
 23. Levada OA, Troyan AS, Pinchuk IY. Serum insulin-like growth factor-1 as a potential marker for MDD diagnosis, its clinical characteristics, and treatment efficacy validation: data from an open-label vortioxetine study. *BMC Psychiatry* (2020) 20:208. doi: 10.1186/s12888-020-02636-7
 24. Bot M, Milaneschi Y, Penninx BW, Drent ML. Plasma insulin-like growth factor I levels are higher in depressive and anxiety disorders, but lower in antidepressant medication users. *Psychoneuroendocrinology* (2016) 68:148–55. doi: 10.1016/j.psyneuen.2016.02.028
 25. Tu KY, Wu MK, Chen YW, Lin PY, Wang HY, Wu CK, et al. Significantly Higher Peripheral Insulin-Like Growth Factor-1 Levels in Patients With Major Depressive Disorder or Bipolar Disorder Than in Healthy Controls: A Meta-Analysis and Review Under Guideline of PRISMA. *Med (Baltimore)* (2016) 95(4):e2411. doi: 10.1097/MD.0000000000002411
 26. Levada OA, Troyan AS. Major depressive disorder and accelerated aging from a peripheral IGF-1 overexpression perspective. *Med Hypotheses* (2020) 138:109610. doi: 10.1016/j.mehy.2020.109610
 27. Jiang H, Chen S, Li C, Lu N, Yue Y, Yin Y, et al. The serum protein levels of the tPA-BDNF pathway are implicated in depression and antidepressant treatment. *Transl Psychiatry* (2017) 7(4):e1079. doi: 10.1038/tp.2017.43
 28. American Psychiatric Association. *Diagnostic and Statistical Manual of Mental Disorders, Fifth Edition*. American Psychiatric Association: Arlington, VA (2013).
 29. Levada OA, Troyan AS. Cognitive-functional relationships in major depressive disorder: Crucial data from a Ukrainian open-label study of vortioxetine versus escitalopram. *J Affect Disord* (2019) 250:114–22. doi: 10.1016/j.jad.2019.03.040
 30. Montgomery SA, Åsberg M. A new depression scale designed to be sensitive to change. *Br J Psychiatry* (1979) 134:382–9. doi: 10.1192/bjp.134.4.382
 31. Guy W. *Clinical Global Impressions*. In: Guy W, editor. *ECDEU Assessment Manual for Psychopharmacology Revised*. Rockville: National Institute of Mental Health (1976). p. 217–22.
 32. Cha D. Perceived Deficits Questionnaire – Depression, 5-item (PDQ-D-5). In: McIntyre R, editor. *Cognitive impairment in major depressive disorder: Clinical relevance, biological substrates, and treatment opportunities*. Cambridge: Cambridge University Press (2016). p. 242–56.
 33. Sheehan DV, Harnett-Sheehan K, Raj BA. The measurement of disability. *Int Clin Psychopharmacol* (1996) 11(Suppl 3):89–95. doi: 10.1097/00004850-199606003-00015
 34. Strauss E, Sherman EMS, Spreen O. *A compendium of neuropsychological tests: Administration, norms, and commentary*. Oxford University Press: Oxford, England (2006).
 35. Karege F, Perret G, Bondol G, Schwald M, Bertschy G, Aubry JM. Decreased serum brain-derived neurotrophic factor levels in major depressed patients. *Psychiatry Res* (2002) 109(2):143–8. doi: 10.1016/S0165-1781(02)00005-7
 36. Yoshimura R, Mitoma M, Sugita A, Hori H, Okamoto T, Umene W, et al. Effects of paroxetine or milnacipran on serum brain-derived neurotrophic factor in depressed patients. *Prog Neuropsychopharmacol Biol Psychiatry* (2007) 31(5):1034–7. doi: 10.1016/j.pnpb.2007.03.001
 37. Sen S, Duman R, Sanacora G. Serum brain-derived neurotrophic factor, depression, and antidepressant medications: meta-analyses and implications. *Biol Psychiatry* (2008) 64(6):527–32. doi: 10.1016/j.biopsych.2008.05.005
 38. Munno D, Sterpone S, Fania S, Cappellin F, Mengozzi G, Saroldi M. Plasma brain derived neurotrophic factor levels and neuropsychological aspects of depressed patients treated with paroxetine. *Panminerva Med* (2013) 55(4):377–84.
 39. Polyakova M, Stuke K, Schuemberg K, Mueller K, Schoenkecht P, Schroeter ML. BDNF as a biomarker for successful treatment of mood disorders: a systematic & quantitative meta-analysis. *J Affect Disord* (2015) 174:432–40. doi: 10.1016/j.jad.2014.11.044
 40. Bus BA, Molendijk ML. The neurotrophic hypothesis of depression. *Tijdschr Psychiatr* (2016) 58(3):215–22.
 41. Gupta K, Gupta R, Bhatia MS, Tripathi AK, Gupta LK. Effect of Agomelatine and Fluoxetine on HAM-D Score, Serum Brain-Derived Neurotrophic Factor, and Tumor Necrosis Factor- α Level in Patients With Major Depressive Disorder With Severe Depression. *J Clin Pharmacol* (2017) 57(12):1519–26. doi: 10.1002/jcph.963
 42. Schröter K, Brum M, Brunkhorst-Kanaan N, Tole F, Ziegler C, Domschke K, et al. Longitudinal multi-level biomarker analysis of BDNF in major depression and bipolar disorder. *Eur Arch Psychiatry Clin Neurosci* (2019) 270(2):169–81. doi: 10.1007/s00406-019-01007-y
 43. Lee BH, Park YM, Um TH, Kim S. Lower serum brain-derived neurotrophic factor levels are associated with failure to achieve remission in patients with major depression after escitalopram treatment. *Neuropsychiatr Dis Treat* (2014) 10:1393–8. doi: 10.2147/NDT.S64913
 44. Zhou C, Zhong J, Zou B, Fang L, Chen J, Deng X, et al. Meta-analyses of comparative efficacy of antidepressant medications on peripheral BDNF concentration in patients with depression. *PLoS One* (2017) 12(2):e0172270. doi: 10.1371/journal.pone.0172270
 45. Başterzi AD, Yazici K, Aslan E, Delialioğlu N, Taşdelen B, Tot Acar S, et al. Effects of fluoxetine and venlafaxine on serum brain derived neurotrophic factor levels in depressed patients. *Prog Neuropsychopharmacol Biol Psychiatry* (2009) 33(2):281–5. doi: 10.1016/j.pnpb.2008.11.016
 46. Yan G, Zhang M, Liu Y, Yin M. Efficacy of vortioxetine combined cognitive behaviour intervention therapy on brain-derived neurotrophic factor level on depressive patients. *Psychogeriatrics* (2019) 19(5):475–81. doi: 10.1111/psyg.12426
 47. Molendijk ML, Bus BA, Spinhoven P, Penninx BW, Kenis G, Prickaerts J, et al. Serum levels of brain-derived neurotrophic factor in major depressive disorder: state-trait issues, clinical features and pharmacological treatment. *Mol Psychiatry* (2011) 16(11):1088–95. doi: 10.1038/mp.2010.98
 48. Lee HY, Kim YK. Plasma brain-derived neurotrophic factor as a peripheral marker for the action mechanism of antidepressants. *Neuropsychobiology* (2008) 57(4):194–9. doi: 10.1159/000149817
 49. Bocchio-Chiavetto L, Bagnardi V, Zanardini R, Molteni R, Nielsen MG, Placentino A, et al. Serum and plasma BDNF levels in major depression: a replication study and meta-analyses. *World J Biol Psychiatry* (2010) 11(6):763–73. doi: 10.3109/15622971003611319
 50. Kurita M, Nishino S, Kato M, Numata Y, Sato T. Plasma brain-derived neurotrophic factor levels predict the clinical outcome of depression treatment in a naturalistic study. *PLoS One* (2012) 7(6):e39212. doi: 10.1371/journal.pone.0039212
 51. Furuse K, Ukai W, Hashimoto E, Hashiguchi H, Kigawa Y, Ishii T, et al. Antidepressant activities of escitalopram and blonanserin on prenatal and adolescent combined stress-induced depression model: Possible role of neurotrophic mechanism change in serum and nucleus accumbens. *J Affect Disord* (2019) 247:97–104. doi: 10.1016/j.jad.2019.01.007
 52. Yong KJ, Chang H. The effect of exercise intensity on brain derived neurotrophic factor and memory in adolescents. *Environ Health Prev Med* (2017) 22:27. doi: 10.1186/s12199-017-0643-6
 53. van Zutphen EM, Rhebergen D, van Exel E, Oudega ML, Bouckaert F, Sienaert P, et al. Brain-derived neurotrophic factor as a possible predictor of electroconvulsive therapy outcome. *Transl Psychiatry* (2019) 9(1):155. doi: 10.1038/s41398-019-0491-9
 54. Dandekar MP, Saxena A, Saini G, Shin JH, Migut A, Giridharan VV, et al. Medial Forebrain Bundle Deep Brain Stimulation Reverses Anhedonic-Like Behavior in a Chronic Model of Depression: Importance of BDNF and Inflammatory Cytokines. *Mol Neurobiol* (2019) 56(6):4364–80. doi: 10.1007/s12035-018-1381-5
 55. Yu H, Chen JJ, Zeng BQ, Zhong QP, Xu JP, Liu YG. Role of cAMP/CREB/BDNF signaling pathway in anti-depressive effect of vortioxetine in mice. *Nan Fang Yi Ke Da Xue Xue Bao* (2017) 37(1):107–12. doi: 10.3969/j.issn.1673-4254.2017.01.20
 56. Brivio P, Corsini G, Riva MA, Calabrese F. Chronic vortioxetine treatment improves the responsiveness to an acute stress acting through the ventral hippocampus in a glucocorticoid-dependent way. *Pharmacol Res* (2019) 142:14–21. doi: 10.1016/j.phrs.2019.02.006
 57. Lu Y, Ho CS, McIntyre RS, Wang W, Ho RC. Effects of vortioxetine and fluoxetine on the level of Brain Derived Neurotrophic Factors (BDNF) in the hippocampus of chronic unpredictable mild stress-induced depressive rats. *Brain Res Bull* (2018) 142:1–7. doi: 10.1016/j.brainresbull.2018.06.007
 58. Chen F, Danladi J, Ardalani M, Elfving B, Müller HK, Wegener G, et al. A Critical Role of Mitochondria in BDNF-Associated Synaptic Plasticity After One-Week Vortioxetine Treatment. *Int J Neuropsychopharmacol* (2018) 21(6):603–15. doi: 10.1093/ijnp/pyy022

59. Primo de Carvalho Alves L, Sica da Rocha N. Lower levels of brain-derived neurotrophic factor are associated with melancholic psychomotor retardation among depressed inpatients. *Bipolar Disord* (2018) 20(8):746–52. doi: 10.1111/bdi.12636
60. Kheirouri S, Noorazar SG, Alizadeh M, Dana-Alamdari L. Elevated Brain-Derived Neurotrophic Factor Correlates Negatively with Severity and Duration of Major Depressive Episodes. *Cognit Behav Neurol* (2016) 29(1):24–31. doi: 10.1097/WNN.0000000000000089
61. Oral E, Canpolat S, Yildirim S, Gulec M, Aliyev E, Aydin N. Cognitive functions and serum levels of brain-derived neurotrophic factor in patients with major depressive disorder. *Brain Res Bull* (2012) 88(5):454–9. doi: 10.1016/j.brainresbull.2012.03.005
62. Engelmann J, Wagner S, Wollschläger D, Kaaden S, Schlicht KF, Dreimüller N, et al. Higher BDNF plasma levels are associated with a normalization of memory dysfunctions during an antidepressant treatment. *Eur Arch Psychiatry Clin Neurosci* (2019) 270: 183–93. doi: 10.1007/s00406-019-01006-z
63. Lesch KP, Rupperecht R, Müller U, Pfüller H, Beckmann H. Insulin-like growth factor I in depressed patients and controls. *Acta Psychiatr Scand* (1988) 78:684–8. doi: 10.1111/j.1600-0447.1988.tb06404.x
64. Franz B, Buysse DJ, Cherry CR, Gray NS, Grochocinski VJ, Frank E, et al. Insulin-like growth factor 1 and growth hormone binding protein in depression: a preliminary communication. *J Psychiatr Res* (1999) 33:121–7. doi: 10.1016/S0022-3956(98)00066-1
65. Kopczak A, Stalla GK, Uhr M, Lucae S, Hennings J, Ising M, et al. IGF-I in major depression and antidepressant treatment response. *Eur Neuropsychopharmacol* (2015) 25:864–72. doi: 10.1016/j.euroneuro.2014.12.013
66. Sharma AN, da Costa e Silva BF, Soares JC, Carvalho AF, Quevedo J. Role of trophic factors GDNF, IGF-1 and VEGF in major depressive disorder: A comprehensive review of human studies. *J Affect Disord* (2016) 197:9–20. doi: 10.1016/j.jad.2016.02.067
67. Sievers C, Auer MK, Klotsche J, Athanasoulia AP, Schneider HJ, Nauck M, et al. IGF-I levels and depressive disorders: results from the Study of Health in Pomerania (SHIP). *Eur Neuropsychopharmacol* (2014) 24:890–6. doi: 10.1016/j.euroneuro.2014.01.008
68. Emeny RT, Bidlingmaier M, Lacruz ME, Linkohr B, Peters A, Reincke M, et al. Mind over hormones: sex differences in associations of wellbeing with IGF-I, IGFBP-3 and physical activity in the KORA-age study. *Exp Gerontol* (2014) 59:58–64. doi: 10.1016/j.exger.2014.08.001
69. Van Varsseveld NC, Van Bunderen CC, Sohl E, Comijs HC, Penninx BW, Lips P, et al. Serum insulin-like growth factor 1 and late-life depression: a population-based study. *Psychoneuroendocrinology* (2015) 54:31–40. doi: 10.1016/j.psyneuen.2015.01.014
70. Xu Y, Wei H, Zhu Y, Zhu Y, Zhang N, Qin J, et al. Potential serum biomarkers for the prediction of the efficacy of escitalopram for treating depression. *J Affect Disord* (2019) 250:307–12. doi: 10.1016/j.jad.2019.03.008

Conflict of Interest: OL is a member of advisory and/or speaker boards of the following companies: Lundbeck, Pfizer, Acino. AT gave presentations for Lundbeck and Acino.

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Case Report: PANDAS and Persistent Lyme Disease With Neuropsychiatric Symptoms: Treatment, Resolution, and Recovery

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OPEN ACCESS

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Specialty section:

This article was submitted to
Molecular Psychiatry,
a section of the journal
Frontiers in Psychiatry

Received: 21 November 2019

Accepted: 04 January 2021

Published: 02 February 2021

Citation:

Cross A, Bouboulis D, Shimasaki C
and Jones CR (2021) Case Report:
PANDAS and Persistent Lyme Disease
With Neuropsychiatric Symptoms:
Treatment, Resolution, and Recovery.
Front. Psychiatry 12:505941.
doi: 10.3389/fpsy.2021.505941

This case report describes the diagnosis and treatment of a pre-pubertal (onset at age 7) Caucasian female with serological evidence of Lyme disease accompanied by multiple neuropsychiatric symptoms 6 months following a vacation in a tick endemic area of the United States. Prior to the diagnosis of Lyme disease, the patient also met the clinical diagnostic criteria for PANDAS (Pediatric Autoimmune Neuropsychiatric Disorder Associated with Strep), with serological evidence of three distinct episodes of streptococcal pharyngitis. All three episodes of strep occurred during the 6-months interval between suspected Lyme disease exposure and the onset of multiple neuropsychiatric symptoms. Her sometimes incapacitating symptoms followed a relapsing and remitting course that impacted her personal, family, social, and academic domains. Over a span of 31 consecutive months of treatment with various antimicrobials and three courses of intravenous immunoglobulins (IVIg) she experienced complete remission and remains symptom free at the time of this publication. Written permission was obtained from the minor patient's mother allowing the submission and publication of this case study.

Keywords: Lyme, PANDAS, Cunningham Panel, neuropsychiatric, IVIg, strep pharyngitis, basal ganglia encephalitis (BGE), autoimmune encephalitis

BACKGROUND

In current medical practice, patients with co-occurring Lyme borreliosis and autoimmune encephalitis secondary to strep infections, such as PANDAS (1–3), are met with a number of inherent challenges (4). The ability to obtain accurate serological testing results for Lyme disease and common co-infections is a challenge for patients and providers alike due to the varied reported accuracy of different Lyme tests (5). To complicate things further, Lyme disease testing is fraught with controversy regarding methodology and interpretation of test results. In 1980, the Centers for Disease Control and Prevention (CDC) began surveillance for Lyme disease, identifying only 10 states where Lyme disease was believed to occur. Currently, all 50 states have reported cases of Lyme disease (6). In 2017, the CDC received reports of a total of 42,743 confirmed and probable cases of Lyme disease, but they estimate that in the US ~300,000 patients may contract Lyme disease annually (7). One clinical sign of Lyme disease exposure from a tick bite is an erythema migrans (EM) rash, but often patients with documented Lyme disease do not present with EM (8, 9).

Medical literature supports numerous cases of neuropsychiatric symptoms in children who have a diagnosis of Lyme disease as well as other tick-borne infections. For example, a 14-year-old boy experienced a sudden onset of psychotic behavior which persisted despite multiple hospitalizations and treatment with psychotropic medications. He was subsequently diagnosed with neurobartonellosis after he developed cutaneous lesions, which has been documented as common in individuals reporting neuropsychiatric symptoms and *Bartonella* spp. infection or exposure (10). He was treated with a combination of antimicrobials and experienced a gradual progressive decrease in neuropsychiatric symptoms and was able to discontinue all psychotropic drugs (11). Another well-documented case describes a 12-year-old boy who had a compulsion to pedal a stationary bicycle, unwilling to stop long enough to eat or go to school, resulting in a 30-pound weight loss, a skeletal appearance, and multiple hospitalizations. He was found to be infected with *Borrelia* and recovered after a course of intravenous penicillin (12).

An extensive review article documents increasing evidence and recognition that Lyme borreliosis can cause psychiatric symptoms (13). Drawing from databases and using search engines along with clinical experiences, the authors concluded that *Borrelia* can “cause immune and metabolic effects that result in a gradually developing spectrum of neuropsychiatric symptoms usually presenting with significant comorbidity which may include developmental disorders, autism spectrum disorders, schizoaffective disorders, bipolar disorder, depression, anxiety disorders (panic disorder, social anxiety disorder, generalized anxiety disorder, posttraumatic stress disorder, and intrusive symptoms), eating disorder, decreased libido, sleep disorder, addiction, opioid addiction, cognitive impairments, dementia, seizure disorders, suicide, violence, anhedonia, depersonalization, dissociative episodes, derealization, and other impairments” (13).

Data from an unpublished survey of over 1,000 parents of children with PANDAS and/or PANS, conducted by Moleculera Labs in 2018, “Economic and Psychosocial Costs of PANDAS and PANS,” revealed that, on average, patients have seen up to 12 medical providers, requiring ~3 years before receiving a diagnosis of PANDAS or its broader diagnostic category, PANS (Pediatric Acute-onset Neuropsychiatric Syndrome). The survey results also revealed that at least 20% of patients with PANDAS and/or PANS experience a delay of more than 12 months before receiving appropriate treatment even after being diagnosed with this type of autoimmune encephalopathy.

INTRODUCTION

This is a case report of a previously healthy 7-year-old girl who was conceived through artificial insemination resulting in an uncomplicated, full-term pregnancy and delivery by caesarian section. She weighed 8 pounds, 10 ounces at birth and was breast fed. Her family history was unremarkable for rheumatic fever, tics, OCD, autoimmune disorders, psychiatric illness, or allergies. 6 months prior to the onset of her symptoms, the

patient and her family vacationed in a tick endemic area of the US, but the patient had no known tick attachment or erythema migrans (EM) rash. Quite abruptly, over a period of 3 weeks, the patient experienced dramatic declines in cognitive functioning, concentration, and ability to focus, a loss of math skills, the onset of dysgraphia and difficulty with social cues, decreased processing speed, word selection problems, anxiety, fatigue, nighttime awakening, chills, joint and muscle pain, moodiness, both general and separation anxiety, and panic attacks. She also experienced obsessions and compulsions and aggressive behavior which was completely out of character according to her mother. She said to her mother, “Mom, something happened to my brain.” Previously, she enjoyed music and dance lessons but her overall activity was decreased. Because the patient had consistently been identified as an academically gifted child, a call from the patient’s teacher came as a surprise when the mother was asked, “Did you drop your daughter on her head?” The patient regressed from being a year ahead of her class in math, to being unable to add beyond the number 10. She began having trouble comprehending more difficult reading. During a ride home with her mother, the patient asked, “Who are you? What’s your name again?” And “I know you are mommy but what’s your name?” Her mother began to think that her daughter may have experienced an accident or head injury, but there was no physical sign or history of any type of injury. An EEG was performed with normal findings.

CASE PRESENTATION

This previously asymptomatic, healthy 7-year-old girl experienced an abrupt onset of several physical, neurological, and psychiatric symptoms increasing in intensity over a 3-week period. The patient’s mother reports that strep pharyngitis was diagnosed by a previous physician on three separate occasions with the first episode occurring 180 days prior to the onset of neuropsychiatric symptoms. The first course of treatment was a 10-day course of amoxicillin which resulted in no change in her behavior or functioning. The strep infection recurred, and amoxicillin was again prescribed with no behavioral improvement. The third episode of strep throat (Quest Labs DNase B results of 407), (reference range <376) was treated with a course of clindamycin with a notable improvement in the patient’s symptoms (14). Neurologists at a university medical center referred the patient to the Psychiatry department. Instead, the patient’s mother chose to have her evaluated by a developmental pediatrician who arrived at a diagnosis of Pediatric Autoimmune Neuropsychiatric Disorder Associated with Strep (PANDAS) (15, 16). Due to the patient’s worsening symptoms, a second opinion was sought, and additional blood work performed including a CDC Lyme Western Blot which was positive, leading to the additional diagnosis of Lyme disease. Quest Laboratories’ Lyme disease enzyme immunoassay (EIA) with reflex to Western Blot was positive at 1.25, (0.00–0.90 negative index value). The patient was referred to the university’s infectious disease clinic where treatment to address the Lyme disease diagnosis began. A PICC line was inserted to facilitate

the administration of IV Rocephin. A lumbar puncture (LP) was performed as part of the infectious disease evaluation with normal results. At this point, because the patient had both a diagnosis of PANDAS and Lyme disease, her mother sought the opinion of a pediatric Lyme disease specialist who was also familiar with diagnosing and treating PANDAS. A graphic representation of the timeline from the initial suspected Lyme disease exposure through her complete course of treatment is shown in **Figure 1**.

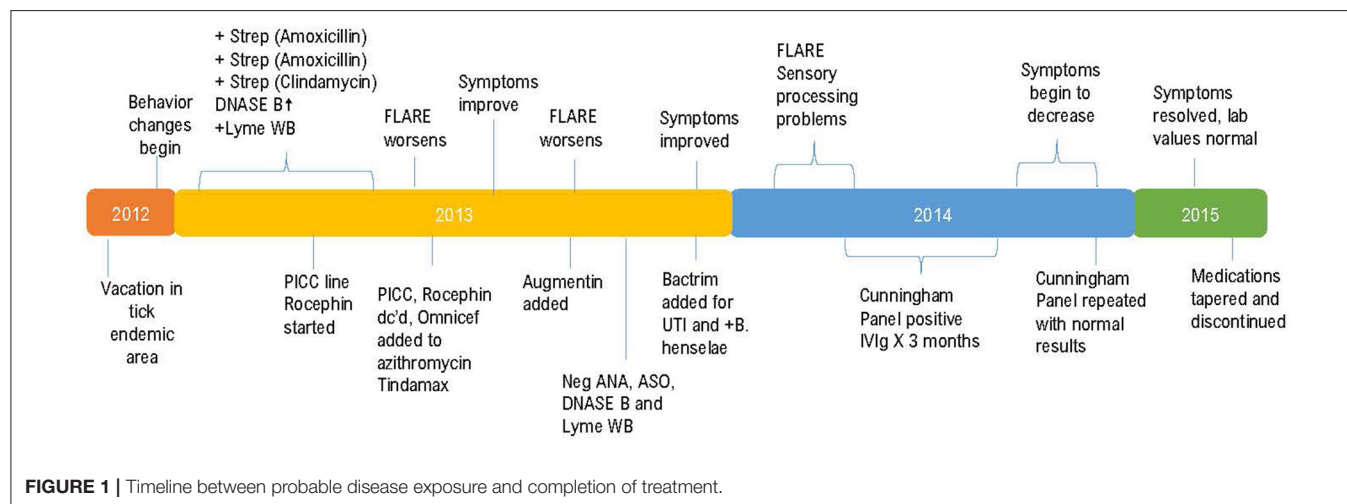
On her first visit with the pediatric Lyme disease specialist, the patient presented with crying, anxiety, headache, joint pain, decreased cognitive functioning, fatigue, nighttime awakening and an extreme fear of sleeping alone. Her ASO titer was negative but a DNase B titer was elevated at 407 (<376). Her ESR was normal. The IV Rocephin 1.5 gm QD prescribed by the previous doctor was continued, but her PICC line became occluded requiring removal. Oral Omnicef 300 mg BID was initiated to replace the Rocephin. Zithromax 250 mg BID and Tindamax 250 mg QD (Saturdays and Sundays only) were initiated.

Due to complaints of right upper quadrant pain, an abdominal ultrasound was ordered which was unremarkable ruling out concerns of possible Rocephin cholelithiasis. Ursodiol 300 mg BID was ordered to help break down cholesterol that had possibly formed in the gall bladder. A CDC Lyme WB was negative although repeated Lyme testing through IGeneX Laboratory revealed a positive Lyme WB IgM with positive double starred bands 31 and 41 and indeterminate bands 39 and 83–93. Lyme WB IgG and *Bartonella henselae* testing showed negative IgG and IgM results. Babesia FISH (RNA) was negative. Her previously elevated DNase B was now normal as was her ASO titer. An elevated *Mycoplasma pneumoniae* IgG at 2.01 (reference range ≤ 0.90 ISR) indicated a previous infection and *M. pneumoniae* IgM was negative. A basic metabolic panel, CBC, urinalysis, ESR, and ANA were completely within normal limits. Genetic testing indicated that the patient was a carrier of the *HLA-DRB1*, 2, and 4 genes which are reported to occur more frequently in patients with Lyme disease and rheumatoid arthritis (17–19). It is also been observed that *B. burgdorferi* can alter the

repertoire of self-peptides bound to MHC class II molecules and influence the likelihood of autoreactive T-cells which could lead to infection-induced autoimmune illnesses (20).

The patient's mother was given a Lyme disease checklist to complete prior to each appointment indicating symptoms and improvement or lack of improvement since the previous visit. The Lyme disease checklist asks the parent to rate the patient on 35 symptoms including (1) unexplained fevers, sweats, chills or flushing, (2) weight change (loss or gain), (3) fatigue, (4) hair loss, (5) swollen glands, (6) sore throat, (7) testicular/pelvic pain, (8) menstrual irregularity, (9) unexplained milk production/breast pain, (10) irritable bladder or dysfunction, (11) upset stomach/abdominal pain, (12) constipation/diarrhea, (13) chest pain/rib soreness, (14) shortness of breath or cough, (15) heart palpitations, (16) joint pain/swelling, (17) joint stiffness, (18) muscle pain/cramps, (19) muscle twitching, (20) headaches, (21) neck creaks/cracks/stiffness, (22) numbness/tingling/stabbing sensations, (23) facial paralysis, (24) double vision/floaters/loss of vision, (25) buzzing/ringing/ear pain/sensitivity, (26) vertigo/dizziness, (27) lightheadedness/poor balance, (28) tremors, (29) difficulty thinking, (30) difficulty concentrating, (31) forgetfulness/poor short term memory, (32) disorientation/getting lost, (33) difficulty with speech or writing, (34) mood swings/irritability/depression, and (35) too much/too little sleep. In addition, the parents were asked to rate symptom severity on a mild, moderate, or severe scale as well as symptom frequency on an occasional, often, or constant scale.

At a follow-up office visit 8 weeks later, her Lyme disease checklist indicated overall improvement with no headaches, but increased insomnia and scattered joint sensitivities. Range of motion, biometrics and vital signs were normal except for an oral temperature of 99.2° F. She had no “dark circles” under her eyes, no tremors and her balance was normal. 2 weeks later the patient experienced a sudden symptom recurrence. Omnicef was discontinued, Zithromax continued at the same dosage and Augmentin added at 500 mg BID. EMLA cream (Lidocaine 2.5% and Prilocaine 2.5%) was prescribed as a topical anesthetic to



treat her joint sensitivities. Lab values at this visit were within normal limits.

10 weeks following the regression, at her next office visit, she had no major complaints and her Lyme disease checklist indicated overall symptom improvement. She denied having headaches and reported increased energy, improved cognitive function and enjoyment of her music lessons. Her physical examination was within normal limits. ANA, CRP, ASO, Streptozyme, and *M. pneumoniae* lab results were all within normal limits. The patient was attending day camp, Monday through Friday, all day, without problems. 2 weeks following this appointment, she suddenly became argumentative, hyperactive, and combative and experienced chills and headache. Bactrim SS BID was added due to a suspected urinary tract infection and to address new IGeneX laboratory results indicating the presence of Lyme borreliosis with positive double starred bands 31 and 41 and double starred bands 39, 83–93 indeterminate. Quest laboratory testing showed an elevated *B. henselae* IgG at 1:64. *B. henselae* IgM was within normal limits at <1:20. Both Ehrlichia and Anaplasma testing had negative results.

2 months later at her next office visit, the patient presented as unhappy and experiencing some facial “twitching.” However, her appetite was good (no previous restrictive eating or food refusal was noted) and she was engaged in activities including dance and tennis.

Two additional months later, labs were repeated. Quest laboratory testing indicated negative results for *E. chaffeensis*, *A. phagocytophilum*, and *B. henselae* IgG and IgM. IGeneX testing showed an indeterminate Lyme IgM with double starred bands 31 and 41. Lyme IgG testing was interpreted as positive with double starred bands 31, 41, and 58 reactive. IGeneX interpretation guidance states the IgG WB is considered positive if two or more of the double starred bands are present from either Group 1 or Group 2. Group 1 includes bands 23–25, 31, 34, 39, 83–93. Group 2 includes bands 23–25, 34, 39, 41, 83–93. *B. henselae* IFA IgG and IgM were both reported as negative. *B. duncani* IgG was negative but IgM was reported as indicative of active infection at a level of 80 with values > 40 considered elevated. This was presumed to be a co-infection due to the original tick exposure as no additional tick exposure was identified.

CD45RA testing, which is a marker of naïve T cells, showed an elevated result of 38% (reference range 5–37%) which is an indication of the amount of time elapsed (~10 days) since the most recent antigenic stimulation (21, 22). Although the patient’s CD45RA was not significantly abnormal, as it was only slightly outside of the upper end of the normal range, it may be supportive of the confirmed presence of an additional

co-infection with *B. duncani*. The presence of *B. duncani* prompted the addition of Mepron 375 mg BID to her medication regimen. Less than 5 days following this visit, she experienced an onset of severe stomach aches and “migraines” which caused her to leave school early 4 out of 5 days each week for several weeks. These symptoms slowly resolved, and her medication regimen was continued without additional changes.

4 months later at her sixth office visit, her mother reported that she was having panic attacks and was “terrified” to sleep alone. She had some residual combativeness, lack of focus, and cognitive interference. The patient’s energy level was improved, and she was engaging in her normal activities. Physical examination and vital signs were normal. Sensory processing problems were evident with heightened sensitivity to lights and sounds. She appeared to manage well at home but struggled with these symptoms in her classroom resulting in the development and implementation of an Individual Educational Plan (IEP). Neuropsychiatric testing was ordered to assess this issue. Quantitative immunoglobulin values were all within normal limits. Quest laboratory testing at this visit revealed negative results for *B. henselae* IgG and IgM, *E. chaffeensis* IgG and IgM, and *Anaplasma Phagocytophilum* IgG and IgM. IGeneX testing showed negative results for *B. henselae* IgG and IgM, and Lyme WB IgG. Lyme WB IgM was indeterminate.

The Cunningham Panel™ of Tests (Moleculara Labs, Oklahoma City, OK) was ordered to assess the presence of antineuronal antibodies against specific neuronal receptors. The patient’s Anti-Dopamine D1 Receptor (DRD1) and Anti-Dopamine D2L Receptor (DRD2L) were elevated, her Anti-Lysoganglioside-GM1 (LYSO-GM1) was normal, her Anti-Tubulin (TUB) was elevated, and her Calcium/calmodulin-dependent protein kinase II (CaMKII) was normal (23–25). Published studies demonstrated that the elevated presence of one or more of these antineuronal antibodies and antibody mediated stimulation of CamKII was strongly associated with autoimmune neuropsychiatric symptoms such as those present in PANDAS and PANS (26, 27). Based upon the patient’s Cunningham Panel™ test results (See **Table 1**) the decision was made to prescribe intravenous immunoglobulin (IVIg) in accordance with established treatment guidelines for the patient’s level of symptom severity (28, 29). She received IVIg at 2 gm/kg given over a 2-days period for a total of three treatments at 4-weeks intervals.

A significant evidence base exists for the use of IVIg in PANDAS patients, particularly those exhibiting a relapsing and remitting course of illness with moderate to severe symptoms, although additional randomized, double blind,

TABLE 1 | Cunningham Panel results—test 1.

| | Dopamine D1 (titer) | Dopamine D2 (titer) | Lysoganglioside (titer) | Tubulin (titer) | CaM Kinase II (% of baseline) |
|----------------|---------------------|---------------------|-------------------------|-----------------|-------------------------------|
| Patient result | 8,000 | >32,000 | 320 | 4,000 | 119 |
| Normal ranges | 500–2,000 | 2,000–8,000 | 80–320 | 250–1,000 | 53–130 |
| Normal mean | 1,056 | 6,000 | 147 | 609 | 95 |
| Interpretation | Elevated | Elevated | Borderline | Elevated | Normal |

placebo control studies need to be performed (28). One well-known study, based on the hypothesis that PANDAS and Sydenham's chorea have similar group A strep etiology, proposed that immunomodulatory treatments could effectively treat neuropsychiatric symptoms. Comparing IVIg to therapeutic plasma exchange (TPE) showed that reassessment at 1-month following treatment, the TPE and IVIg groups both showed "striking improvements in obsessive-compulsive symptoms, anxiety, depression, emotional lability, and global functioning" (30).

Although controlled trials have only evaluated IVIg given as a single course, unpublished data based on the experiences of clinicians and researchers engaged in the PANS Consortium suggest that one to three repeated doses of IVIg may be helpful in children who exhibit a positive response to the initial dose but then experience a relapse as the exogenous antibodies are cleared (28).

In another randomized, controlled trial of IVIg for PANDAS, it was concluded that IVIg was safe and well-tolerated but differences between groups were smaller than anticipated and the double-blind comparison failed to demonstrate superiority of IVIg over placebo (31). It was proposed that the study design may have negated the potential observation of an improvement with IVIg by the administration of antibiotics to both groups prior to either IVIg treatment or placebo. An additional study concluded that children with PANDAS derive a favorable response to IVIg at 12-months follow up "consistent with its role in Ig replacement and immune modulation" (32).

At the conclusion of her IVIg treatments, the patient, now age 9 and in third grade, was normal in height and weight. She had no complaints and her activity level continued to improve with decreased fatigue. Her appetite was normal, and her Lyme disease checklist indicated overall improvement. She did have some residual obsessions and compulsions, intermittent hand tingling, slight facial tics, and a humming tic. Medications at this time included Mepron 375 mg BID, Omnicef 300 mg BID, Tindamax 250 mg QD (Saturdays and Sundays only), and Zithromax 250 mg QD, all of which were continued due to the presence of her residual symptoms. The protocol utilized regarding the length of time for which antibiotic treatments were administered was based upon on the patient's symptoms and laboratory results. Before discontinuance of antibiotic treatment, the patient must exhibit symptom resolution along with negative laboratory findings for a at least 2 consecutive months.

Now 21 months into her treatment, at her seventh office visit, her mother reported that she was doing "pretty well" despite a symptom flare the previous month which lasted ~3 weeks but

had completely resolved. Her Lyme disease checklist indicated that her status was stable. Vital signs were normal except for a slight elevation in her oral temperature at 99.3 F. The patient was doing well in her new school, had a normal activity level and appetite, and was enjoying her usual activities including music and dance lessons. Physical exam revealed no involuntary movements, no vocal or motor tics, clear lungs, and normal reflexes. IGeneX testing at this time showed indeterminate results for Lyme WB IgM with double starred bands 31 and 41 ++ and double starred bands 39 and 83–93 indeterminate. Lyme WB IgG was negative. *B. henselae* IFA IgG and IgM were negative. *B. duncani* IFA IgG was negative, however, *B. duncani* IgM was positive at 80 with normal values falling < 40. All findings were negative per CDC standards. A Cunningham Panel™ was repeated (Test 2) to assess the post-treatment status of antineuronal antibodies. The results of the five assays were all within normal ranges (27) (see **Table 2**).

At this point, it was determined that no further IVIg was needed. Because the family was unable to obtain insurance coverage for the IVIg treatments, they incurred the expense out-of-pocket which averaged \$12,000 per treatment (33, 34). At her follow up visit 6 months later, 31 months after her initial visit, the patient's Lyme disease checklist indicated overall improvement. Her final laboratory testing for tick-borne diseases, strep antibodies, and *M. pneumoniae* were all within normal limits.

Outcome of Treatment

Currently this patient appears to be fully recovered and has been discharged from the care of the pediatric Lyme disease specialist. She is asymptomatic and performing academically at the "top" of her class according to her mother. A summary of the serological evidence of exposure to tick borne illness and streptococcal infection is included in **Table 3**.

DISCUSSION

This patient's case may be representative of numerous other cases of autoimmune neuropsychiatric illnesses where a patient may have concomitant infections and co-morbid diagnoses. What is known with growing certainty is that post-infectious neuropsychiatric illness appears to be increasing in frequency or, at least, in frequency of diagnosis. In cases such as the one presented here, it can be challenging for a patient to deal with controversy surrounding the diagnosis of PANDAS and the legitimacy of a diagnosis of Lyme disease with a neuropsychiatric presentation. Patients with chronic neuropsychiatric symptoms

TABLE 2 | Cunningham Panel results Post-treatment—test 2.

| | Dopamine D1 (titer) | Dopamine D2 (titer) | Lysoganglioside (titer) | Tubulin (titer) | CaM Kinase II (% of baseline) |
|----------------|---------------------|---------------------|-------------------------|-----------------|-------------------------------|
| Patient result | 1,000 | 8,000 | 40 | 500 | 108 |
| Normal ranges | 500–2,000 | 2,000–8,000 | 80–320 | 250–1,000 | 53–130 |
| Normal mean | 1,056 | 6,000 | 147 | 609 | 95 |
| Interpretation | Normal | Borderline | Normal | Normal | Normal |

TABLE 3 | Serological evidence of exposure to *B. burgdorferi*, *B. henselae*, *B. duncani* and streptococcal infections.

| Serological evidence of exposure to <i>Borrelia burgdorferi</i> | |
|---|--|
| Testing 1 | Lyme ELISA +1.25 (0.00–0.94); Lyme WB IgM + 23, 41 |
| Testing 2 | Lyme WB IgM + 31, 41 |
| Testing 3 | Lyme WB IgM + 31, 41 and 58 |
| Serologic evidence of exposure to <i>B. henselae</i> | |
| <i>B. henselae</i> IgM neg; <i>B. henselae</i> IgG + 1:64 (<1:64) | |
| Serologic evidence of exposure to <i>B. duncani</i> | |
| <i>B. duncani</i> IgM 80 (IgM >40 indicates active infection) | |
| Serologic evidence of exposure to group A streptococcus | |
| DNASE B + 407 (<376 u/mL) | |

who do not respond adequately to traditional psychotropic medications may have an underlying immune-mediated condition triggered by one or more infections as evidenced in this case report. In addition, patients with genetic susceptibility to immune dysregulation, such as identified in this patient, carriers of the *HLA-DRB1*, 2, and 4 genes may increase the likelihood of an autoimmune encephalopathy indicated by the presence of antineuronal antibodies associated with neuropsychiatric symptoms. The *HLA-DRB1* gene plays a critical role in the immune system, where the HLA complex helps the immune system distinguish the body's own proteins from proteins made by foreign invaders such as viruses and bacteria. There is increasing evidence that IVIg and immunoglobulins are effective in treating autoimmune neuropsychiatric illness although the mechanism of action is uncertain.

It is often noted that patients diagnosed with PANDAS or PANS are frequently found to have multiple co-infections (35). Technically, since the initial treatment of the strep with amoxicillin did not initially result in improvement in the patient's neuropsychiatric symptoms, whereas with the addition of clindamycin the patient's symptoms began to improve, it could be concluded that Lyme may have been the pathogenic agent. However, in clinical experience it has been found that improvement in neuropsychiatric symptoms does not typically occur unless all co-infections are addressed and resolved. Another explanation may be that once the co-offending infection was also resolved, the patient's neuropsychiatric symptoms began to improve. This also brings up an issue with nomenclature; since this patient had a Lyme infection, she could be clinically diagnosed with PANS, or because of the strep diagnosis, clinically this patient could also be diagnosed with PANDAS. Because this patient had concurrent infections, we have referred to this case as

a patient with PANDAS and Lyme co-infection. We suggest that the utilization of the term “Basal Ganglia Encephalitis (BGE)” may have more clinical utility when referring to the possible pathophysiology, where the type of infectious trigger may not be essential in the nomenclature of the clinical syndrome (26, 36).

CONCLUSION

The subject of this case report had a concomitant diagnosis of Lyme borreliosis and PANDAS, both of which are consistent with the neuropsychiatric symptoms she experienced. As evidenced by her recovery and resolution of symptoms, treating both the Lyme infection and streptococcal infection, as well as treating the underlying autoimmune etiology of her neuropsychiatric symptoms resulted in a successful outcome. This case report and treatment history reiterates the complex and challenging nature of infection-triggered autoimmune neuropsychiatric disorders such as PANDAS and PANS and that multiple concomitant infectious agents can frequently be identified in patients suffering from these complex neuropsychiatric disorders. The presence of elevated antineuronal antibodies identified by the Cunningham Panel™ provided an aid in the diagnosis and in directing immunomodulatory treatment. The post-treatment resolution of these autoantibodies provided pathophysiological support for addressing both the infection(s) and the underlying immune system dysfunction which resulted in a positive medical outcome for this patient.

DATA AVAILABILITY STATEMENT

The datasets generated for this study are available on request to the corresponding author.

ETHICS STATEMENT

Written informed consent from the patient's mother was obtained allowing the submission and publication of this case study.

AUTHOR CONTRIBUTIONS

CJ: treating physician of subject of case study, provided oversight, and approval of manuscript. CS: provided oversight and technical assistance during the preparation of the manuscript. AC: provided case review and draft development of the manuscript. DB: treating physician and provided case review. All authors contributed to the article and approved the submitted version.

REFERENCES

- Swedo SE, Leonard HL, Mittleman B, Allen AJ, Perlmutter S, Lougee L, et al. Pediatric autoimmune neuropsychiatric disorders associated with streptococcal infections: clinical description of the first 50 cases. *Am J Psychiatr.* (1998). 155:264–71. doi: 10.1176/ajp.155.2.264
- Swedo SE, Seidlitz J, Kovacevic M, Elizabeth Latimer M, Hommer R, Lougee L, et al. Clinical presentation of pediatric autoimmune neuropsychiatric disorders associated with streptococcal infections in research and community settings. *J Child Adolesc Psychopharmacol.* (2015) 25:26–30. doi: 10.1089/cap.2014.0073
- Orefici G. *Pediatric Autoimmune Neuropsychiatric Disorders Associated with Streptococcal Infections (PANDAS)*. (2016). Available online at: <https://www.ncbi.nlm.nih.gov/books/NBK333433/> (accessed November 10, 2020).
- Rhee H, Cameron D. Lyme disease and pediatric autoimmune neuropsychiatric disorders associated with streptococcal

- infections (PANDAS): an overview. *Int J Gen Med.* (2012) 163:24212. doi: 10.2147/IJGM.S24212
5. Aguero-Rosenfeld ME, Wormser GP. Lyme disease: diagnostic issues and controversies. *Exp Rev Mol Diagn.* (2014) 15:1–4. doi: 10.1586/14737159.2015.989837
 6. Bacon R. *Surveillance for Lyme disease: United States, 1992–2006.* MMWR Surveillance Summary. (2006). Available online at: <https://www.ncbi.nlm.nih.gov/books/NBK390433/> (accessed November 10, 2020)
 7. Centers for Disease Control and Prevention, National Center for Emerging and Zoonotic Infectious Diseases, Division of Vector-Borne Diseases. *Lyme Disease.* (2018). Available online at: <https://www.cdc.gov/lyme/datasurveillance/recent-surveillance-data.html> (accessed November 10, 2020).
 8. Centers for Disease Control and Prevention. Centers for Disease Control and Prevention. *Recent Surveillance Data.* (2019). Available online at: <https://www.cdc.gov/lyme/datasurveillance/recent-surveillance-data.html> (accessed November 10, 2020)
 9. Campos M. *Lyme Disease: Resolving the “Lyme Wars”.* (2018). Available online at: <https://www.health.harvard.edu/blog/lyme-disease-resolving-the-lyme-wars-2018061814071> (accessed November 10, 2020).
 10. Breitschwerdt EB, Bradley JM, Maggi RG, Lashnits E, Reicherter P. *Bartonella* Associated Cutaneous Lesions (BACL) in people with neuropsychiatric symptoms. *Pathogens.* (2020) 9:1023. doi: 10.3390/pathogens9121023
 11. Breitschwerdt EB, Greenberg R, Maggi RG, Robert Mozayani B, Lewis A, Bradley JM. *Bartonella henselae* bloodstream infection in a boy with pediatric acute-onset neuropsychiatric syndrome. *J Centr Nerv Syst Dis.* (2019) 11:117957351983201. doi: 10.1177/1179573519832014
 12. Raven N. *Recovery from Lyme Induced OCD: The Bicycle Boy.* Washington, DC: The Washingtonian (1991).
 13. Bransfield R. Neuropsychiatric lyme borreliosis: an overview with a focus on a specialty psychiatrist's clinical practice. *Healthcare.* (2018) 6:104. doi: 10.3390/healthcare6030104
 14. Snider LA, Lougee L, Grant P, Swedo SE. Antibiotic prophylaxis with azithromycin or penicillin for childhood-onset neuropsychiatric disorders. *Biol Psychiatr.* (2005) 57:788–92. doi: 10.1016/j.biopsych.2004.12.035
 15. Chang K, Frankovich J, Cooperstock M, Cunningham MW, Elizabeth Latimer M, Murphy TK, et al. Clinical evaluation of youth with pediatric acute-onset neuropsychiatric syndrome (PANS): recommendations from the 2013 PANS Consensus Conference. *J Child Adolesc Psychopharmacol.* (2015) 25:3–13. doi: 10.1089/cap.2014.0084
 16. Benros ME, Waltoft BL, Nordentoft M, Østergaard SD, Eaton WW, Krogh J, et al. Autoimmune diseases and severe infections as risk factors for mood disorders. *JAMA Psychiatry.* (2013) 70:812. doi: 10.1001/jamapsychiatry.2013.1111
 17. Iliopoulou BP, Guerau-De-Arellano M, Huber BT. HLA-DR alleles determine responsiveness to *Borrelia burgdorferi* antigens in a mouse model of self-perpetuating arthritis. *Arthritis Rheumat.* (2009) 60:3831–40. doi: 10.1002/art.25005
 18. MedlinePlus Genetics. *HLA-DRB1 gene.* Available online at: <https://ghr.nlm.nih.gov/gene/HLA-DRB1> (accessed November 10, 2020).
 19. Gutierrez-Hoffmann MG, O’meally RB, Cole RN, Tiniakou E, Darrah E, Soloski MJ. *Borrelia burgdorferi*-induced changes in the class II self-immunopeptide displayed on HLA-DR molecules expressed by dendritic cells. *Front Med.* (2020) 7:568. doi: 10.3389/fmed.2020.00568
 20. Kovalchuka L, Eglite J, Lucenko I, Zalite M, Viksna L, Krumina A. Associations of HLA DR and DQ molecules with Lyme borreliosis in Latvian patients. *BMC Res Notes.* (2012) 5:438. doi: 10.1186/1756-0500-5-438
 21. Carrasco J, Danièle G, Van Pel A, Boon T, Van Der Bruggen P. CD45RA on human CD8T cells is sensitive to the time elapsed since the last antigenic stimulation. *Blood.* (2006) 108:2897–905. doi: 10.1182/blood-2005-11-07237
 22. Booth NJ, Mcquaid AJ, Sobande T, Kissane S, Agius E, Jackson SE, et al. Different proliferative potential and migratory characteristics of human CD4+ regulatory T cells that express either CD45RA or CD45RO. *J Immunol.* (2010) 184:4317–26. doi: 10.4049/jimmunol.0903781
 23. Singer HS, Mascaro-Blanco A, Alvarez K, Morris-Berry C, Kawikova I, Ben-Pazi H, et al. Neuronal antibody biomarkers for Sydenham’s chorea identify a new group of children with chronic recurrent episodic acute exacerbations of tic and obsessive-compulsive symptoms following a streptococcal infection. *PLoS ONE.* (2015) 10:120499. doi: 10.1371/journal.pone.0120499
 24. Cox CJ, Zuccolo AJ, Edwards EV, Mascaro-Blanco A, Alvarez K, Stoner J, et al. Antineuronal antibodies in a heterogeneous group of youth and young adults with tics and obsessive-compulsive disorder. *J Child Adolesc Psychopharmacol.* (2015) 25:76–85. doi: 10.1089/cap.2014.0048
 25. Fallon BA, Strobino B, Reim S, Stoner J, Cunningham MW. Antilyso ganglioside and other anti-neuronal autoantibodies in post-treatment Lyme Disease and Erythema Migrans after repeat infection. *Brain Behav Immun Health.* (2020) 2:100015. doi: 10.1016/j.bbih.2019.100015
 26. Chain JL, Alvarez K, Mascaro-Blanco A, Reim S, Bentley R, Hommer R, et al. Autoantibody biomarkers for basal ganglia encephalitis in sydenham chorea and pediatric autoimmune neuropsychiatric disorder associated with streptococcal infections. *Front Psychiatry.* (2020) 11:564. doi: 10.3389/fpsyt.2020.00564
 27. Shimasaki C, Frye RE, Trifiletti R, Cooperstock M, Kaplan G, Melamed I, et al. Evaluation of the Cunningham Panel™ in pediatric autoimmune neuropsychiatric disorder associated with streptococcal infection (PANDAS) and pediatric acute-onset neuropsychiatric syndrome (PANS): changes in antineuronal antibody titers parallel changes in patient symptoms. *J Neuroimmunol.* (2020) 339:577138. doi: 10.1016/j.jneuroim.2019.577138
 28. Frankovich J, Swedo S, Murphy T, Dale RC, Agalliu D, Williams K, et al. Clinical management of pediatric acute-onset neuropsychiatric syndrome: part II—use of immunomodulatory therapies. *J Child Adolesc Psychopharmacol.* (2017) 27:574–93. doi: 10.1089/cap.2016.0148
 29. Intravenous Immunoglobulin for the Treatment of Myasthenia Gravis. *Hayes, Inc. Medical Technology Directory. Genetics Home Reference. HLA-DRB1 Gene.* (2016). National Institutes of Health, US National Library of Medicine. Available online at: <https://ghr.nlm.nih.gov/gene/HLA-DRB1> (accessed November 10, 2020)
 30. Perlmutter SJ, Leitman SF, Garvey MA, Hamburger S, Feldman E, Leonard HL, et al. Therapeutic plasma exchange and intravenous immunoglobulin for obsessive-compulsive disorder and tic disorders in childhood. *Lancet.* (1999) 354:1153–8. doi: 10.1016/S0140-6736(98)12297-3
 31. Williams KA, Swedo SE, Farmer CA, Grantz H, Grant PJ, D’Souza P, et al. Randomized, controlled trial of intravenous immunoglobulin for pediatric autoimmune neuropsychiatric disorders associated with streptococcal infections. *J Am Acad Child Adolesc Psychiatry.* (2016) 55:17. doi: 10.1016/j.jaac.2016.06.017
 32. Younger DS, Chen X. IVIg therapy in PANDAS: analysis of the current literature. *J Neurol Neurosurg.* (2016) 3:25. doi: 10.19104/jnn.2016.25
 33. Winters JL, Brown D, Hazard E, Chainani A, Andrzejewski C Jr. Cost-minimization analysis of the direct costs of TPE and IVIg in the treatment of Guillain-Barre syndrome. *BMC Health Services Res.* (2011) 11:101. doi: 10.1186/1472-6963-11-101
 34. Jessica Katz M. *Intravenous Immunoglobulin.* (2020). Available online at: <https://emedicine.medscape.com/article/210367-overview> (accessed November 10, 2020).
 35. Cooperstock MS, Swedo SE, Pasternack MS, Murphy TK. Clinical management of pediatric acute-onset neuropsychiatric syndrome: part III—treatment and prevention of infections. *J Child Adolesc Psychopharmacol.* (2017) 27:151. doi: 10.1089/cap.2016.0151
 36. Pawela C, Brunsdon RK, Williams TA, Porter M, Dale RC, Mohammad SS. The neuropsychological profile of children with basal ganglia encephalitis. *Dev Med Child Neurol.* (2017) 59:4.

Conflict of Interest: AC and CS are employed by Moleculera Labs.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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