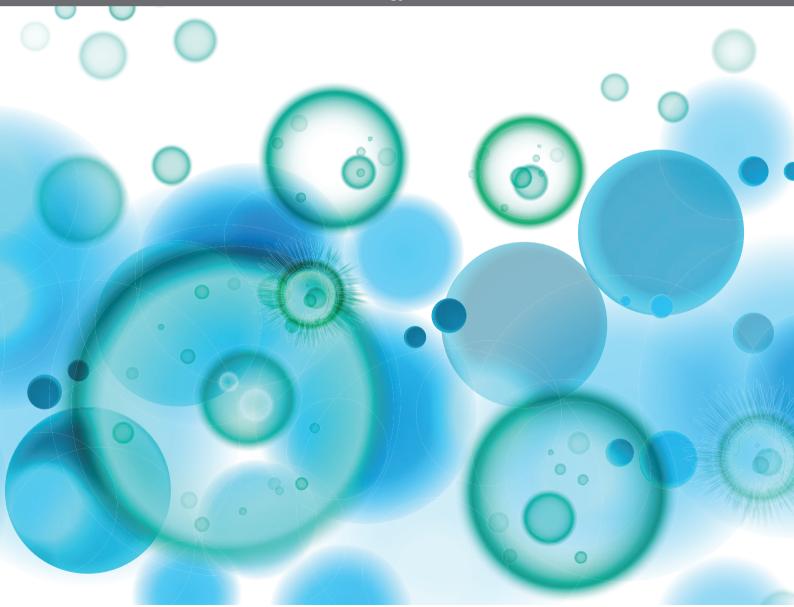
THE IMMUNOTHERAPEUTIC POTENTIAL OF ALLOGENEIC HEMATOPOIETIC STEM CELL TRANSPLANTATION (HSCT)

EDITED BY: Andrea Bacigalupo, Charles Craddock and Nicolaus Martin Kröger

PUBLISHED IN: Frontiers in Immunology







Frontiers eBook Copyright Statement

The copyright in the text of individual articles in this eBook is the property of their respective authors or their respective institutions or funders. The copyright in graphics and images within each article may be subject to copyright of other parties. In both cases this is subject to a license granted to Frontiers.

The compilation of articles constituting this eBook is the property of Frontiers.

Each article within this eBook, and the eBook itself, are published under the most recent version of the Creative Commons CC-BY licence. The version current at the date of publication of this eBook is CC-BY 4.0. If the CC-BY licence is updated, the licence granted by Frontiers is automatically updated to the new version.

When exercising any right under the CC-BY licence, Frontiers must be attributed as the original publisher of the article or eBook, as applicable.

Authors have the responsibility of ensuring that any graphics or other materials which are the property of others may be included in the CC-BY licence, but this should be checked before relying on the CC-BY licence to reproduce those materials. Any copyright notices relating to those materials must be complied with.

Copyright and source acknowledgement notices may not be removed and must be displayed in any copy, derivative work or partial copy which includes the elements in question.

All copyright, and all rights therein, are protected by national and international copyright laws. The above represents a summary only. For further information please read Frontiers' Conditions for Website Use and Copyright Statement, and the applicable CC-BY licence.

ISSN 1664-8714 ISBN 978-2-88974-537-1 DOI 10.3389/978-2-88974-537-1

About Frontiers

Frontiers is more than just an open-access publisher of scholarly articles: it is a pioneering approach to the world of academia, radically improving the way scholarly research is managed. The grand vision of Frontiers is a world where all people have an equal opportunity to seek, share and generate knowledge. Frontiers provides immediate and permanent online open access to all its publications, but this alone is not enough to realize our grand goals.

Frontiers Journal Series

The Frontiers Journal Series is a multi-tier and interdisciplinary set of open-access, online journals, promising a paradigm shift from the current review, selection and dissemination processes in academic publishing. All Frontiers journals are driven by researchers for researchers; therefore, they constitute a service to the scholarly community. At the same time, the Frontiers Journal Series operates on a revolutionary invention, the tiered publishing system, initially addressing specific communities of scholars, and gradually climbing up to broader public understanding, thus serving the interests of the lay society, too.

Dedication to Quality

Each Frontiers article is a landmark of the highest quality, thanks to genuinely collaborative interactions between authors and review editors, who include some of the world's best academicians. Research must be certified by peers before entering a stream of knowledge that may eventually reach the public - and shape society; therefore, Frontiers only applies the most rigorous and unbiased reviews. Frontiers revolutionizes research publishing by freely delivering the most outstanding

research, evaluated with no bias from both the academic and social point of view. By applying the most advanced information technologies, Frontiers is catapulting scholarly publishing into a new generation.

What are Frontiers Research Topics?

Frontiers Research Topics are very popular trademarks of the Frontiers Journals Series: they are collections of at least ten articles, all centered on a particular subject. With their unique mix of varied contributions from Original Research to Review Articles, Frontiers Research Topics unify the most influential researchers, the latest key findings and historical advances in a hot research area! Find out more on how to host your own Frontiers Research Topic or contribute to one as an author by contacting the Frontiers Editorial Office: frontiersin.org/about/contact

THE IMMUNOTHERAPEUTIC POTENTIAL OF ALLOGENEIC HEMATOPOIETIC STEM CELL TRANSPLANTATION (HSCT)

Topic Editors:

Andrea Bacigalupo, Agostino Gemelli University Polyclinic (IRCCS), Italy **Charles Craddock**, University of Birmingham, United Kingdom **Nicolaus Martin Kröger**, University of Hamburg, Germany

Citation: Bacigalupo, A., Craddock, C., Kröger, N. M., eds. (2022). The Immunotherapeutic Potential of Allogeneic Hematopoietic Stem Cell Transplantation (HSCT). Lausanne: Frontiers Media SA.

doi: 10.3389/978-2-88974-537-1

Table of Contents

04 Influence of KIR and NK Cell Reconstitution in the Outcomes of Hematopoietic Stem Cell Transplantation

Fei Gao, Yishan Ye, Yang Gao, He Huang and Yanmin Zhao

21 Efficacy and Safety of Eculizumab in the Treatment of Transplant-Associated Thrombotic Microangiopathy: A Systematic Review and Meta-Analysis

Rui Zhang, Meng Zhou, Jiaqian Qi, Wenjing Miao, Ziyan Zhang, Depei Wu and Yue Han

31 TKI Maintenance After Stem-Cell Transplantation for FLT3-ITD Positive Acute Myeloid Leukemia: A Systematic Review and Meta-Analysis

Nico Gagelmann, Christine Wolschke, Evgeny Klyuchnikov, Maximilian Christopeit, Francis Ayuk and Nicolaus Kröger

Interplay Between the Intestinal Microbiota and Acute Graft-Versus-Host Disease: Experimental Evidence and Clinical Significance

Tao Hong, Rui Wang, Xiaoqi Wang, Shijie Yang, Weihao Wang, Qiangguo Gao and Xi Zhang

57 GVHD Prophylaxis 2020

Mahasweta Gooptu and Joseph Harry Antin

70 Revisit of Optimal Donor Number Estimation in the Hong Kong Bone Marrow Donor Registry

Jenny Chung Yee Ho, Stephen Kwok Fan Cheung, Zhongyi Lui, Ivan Wing Hong Tang, Wanling Yang, Patrick Ip, Cheuk Kwong Lee, Derek Middleton and Janette Siu Yin Kwok

- 76 Chimerism, the Microenvironment and Control of Leukemia
 H. Joachim Deeg
- 83 Allogeneic Stem Cell Transplantation for Acute Myeloid Leukemia: Who, When, and How?

Justin Loke, Richard Buka and Charles Craddock

Allogeneic Hemopoietic Stem Cell Transplantation for Myelofibrosis: 2021

Andrea Bacigalupo, Idanna Innocenti, Elena Rossi, Federica Sora,
Eugenio Galli, Francesco Autore, Elisabetta Metafuni, Patrizia Chiusolo,
Sabrina Giammarco, Luca Laurenti, Giulia Benintende, Simona Sica and

Valerio De Stefano

110 Integrating CAR T-Cell Therapy and Transplantation: Comparisons of Safety and Long-Term Efficacy of Allogeneic Hematopoietic Stem Cell Transplantation After CAR T-Cell or Chemotherapy-Based Complete Remission in B-Cell Acute Lymphoblastic Leukemia

Yan-Li Zhao, De-Yan Liu, Rui-Juan Sun, Jian-Ping Zhang, Jia-Rui Zhou, Zhi-Jie Wei, Min Xiong, Xing-Yu Cao, Yue Lu, Jun-fang Yang, Xian Zhang, Dao-Pei Lu and Peihua Lu





Influence of KIR and NK Cell Reconstitution in the Outcomes of Hematopoietic Stem Cell Transplantation

Fei Gao 1,2,3, Yishan Ye 1,2,3, Yang Gao 1,2,3, He Huang 1,2,3* and Yanmin Zhao 1,2,3*

¹ Bone Marrow Transplantation Center, The First Affiliated Hospital, School of Medicine, Zhejiang University, Hangzhou, China, ² Institute of Hematology, Zhejiang University, Hangzhou, China, ³ Zhejiang Engineering Laboratory for Stem Cell and Immunotherapy, Hangzhou, China

OPEN ACCESS

Edited by:

Nicolaus Martin Kröger, Medizinische Fakultät, Universität Hamburg, Germany

Reviewed by:

Elizabeth Oaks Krieger, Virginia Commonwealth University, United States Ismael Buño, Instituto de Investigación Sanitaria Gregorio Marañón, Spain

*Correspondence:

He Huang huanghe@zju.edu.cn Yanmin Zhao yanminzhao@zju.edu.cn

Specialty section:

This article was submitted to Alloimmunity and Transplantation, a section of the journal Frontiers in Immunology

Received: 27 May 2020 Accepted: 27 July 2020 Published: 02 September 2020

Citation

Gao F, Ye Y, Gao Y, Huang H and Zhao Y (2020) Influence of KIR and NK Cell Reconstitution in the Outcomes of Hematopoietic Stem Cell Transplantation. Front. Immunol. 11:2022. doi: 10.3389/fimmu.2020.02022 Natural killer (NK) cells play a significant role in immune tolerance and immune surveillance. Killer immunoglobin-like receptors (KIRs), which recognize human leukocyte antigen (HLA) class I molecules, are particularly important for NK cell functions. Previous studies have suggested that, in the setting of hematopoietic stem cell transplantation (HSCT), alloreactive NK cells from the donor could efficiently eliminate recipient tumor cells and the residual immune cells. Subsequently, several clinical models were established to determine the optimal donors who would exhibit a graft-vs. -leukemia (GVL) effect without developing graft-vs. -host disease (GVHD). In addition, hypotheses about specific beneficial receptor-ligand pairs and KIR genes have been raised and the favorable effects of alloreactive NK cells are being investigated. Moreover, with a deeper understanding of the process of NK cell reconstitution post-HSCT, new factors involved in this process and the defects of previous models have been observed. In this review, we summarize the most relevant literatures about the impact of NK cell alloreactivity on transplant outcomes and the factors affecting NK cell reconstitution.

Keywords: KIR, NK cell reconstitution, hematopoietic stem cell transplantation, GVHD, infection, relapse

INTRODUCTION

Allogeneic hematopoietic stem cell transplantation (allo-HSCT) is an effective therapy for patients with hematological malignancies. However, relapse, graft-vs. -host disease (GVHD), and infections remain the main causes of treatment failure (1–4). Potential strategies to prevent GVHD and even infections while sparing the graft-vs. -leukemia (GVL) effect have attracted extensive attention. Natural killer (NK) cells, which are a major type of innate lymphocytes, are being researched in this context.

NK cells constitute 5–15% of human peripheral blood lymphocytes (5,6) and possess the abilities of cytotoxic lysis and rapid cytokine secretion without prior antigen presentation (7,8). These functions are regulated by various types of receptors expressed on NK cells manifesting multiple functions either activating or inhibitory (9-11) (Table 1). Among the NK cell receptors, the killer immunoglobin-like receptor (KIR) is one of the major factors that mediate self-tolerance and anti-tumor/infection responses.

TABLE 1 | NK cell receptors and their ligands.

Inhibitory and their	receptors ligands	Activating and their	g receptors ligands	Coreceptors and their ligands		
KIR2DL1	HLA-C2	KIR2DS1	HLA-C2	2B4	CD48	
KIR2DL2	HLA-C1	KIR2DS2	HLA-C1	NTB-A	NTB-A	
KIR2DL3	HLA-C1	KIR2DS3	Unknown	CS1	CS1	
KIR2DL4	HLA-G	KIR2DS4	HLA-A11	NKp80	AICL	
KIR2DL5	Unknown	KIR2DS5	Unknown	TLR	TLRL	
KIR3DL1	HLA-Bw4	KIR3DS1	HLA-F	DNM-1	PVR, Netcin-2	
KIR3DL2	HLA-A3/A11	NKG2C	HLA-E	CD96	PVR	
KIR3DL3	Unknown	NKG2D	MICA, MICB, ULBP1-4			
NKG2A	HLA-E	NKp30	B7-H6, BAT3, CMV pp65			
LIR-1	HLA class I	NKp44	Viral hemagglutinins			
		NKp46	Viral hemagglutinins			
		CD16	IgG-1, 3, 4			

It is well established that KIR genes are located on chromosome 19q13.4 (12). Based on their various structures (the number of extracellular immunoglobulin domains (D) and the long (L) or short (S) tails), 16 KIR genes (including two pseudogenes (P), KIR2DP1 and KIR3DP1) have been classified into four groups (KIR2DL1-5, KIR3DL1-3, KIR2DS1-5, and KIR3DS1). Six genes with short tails are activating KIR genes that encode activating receptors, while the eight genes with long tails are inhibitory KIR genes encoding inhibitory receptors. KIRs could be divided into haplotype A and B according to the activating genes on them. Haplotype A has only one activating gene, KIR2DS4, whereas haplotype B possesses up to five activating KIR genes, including KIR2DS1, 2, 3, 5, and 3DS1 (Figure 1). Thus, the A/A genotype is defined as homozygous for A haplotypes, and the B/x genotype consists of at least one B haplotype. Finally, according to the specific KIR gene locus on the chromosome, a centromeric (Cen) and telomeric (Tel) KIR haplotype and genotype are further determined (13-15). Five inhibitory and three activating KIRs recognize specific class I HLA (A, B, or C) ligands, with the inhibitory KIR2DL1 recognizes group 2 HLA-C alleles, KIR2DL2 and KIR2DL3 recognize group 1 HLA-C alleles, KIR3DL1 recognizes HLA-Bw4 alleles, and KIR3DL2 recognizes HLA-A3/-A11 alleles. Moreover, activating KIR2DS1, KIR2DS2, and KIR2DS4 recognize HLA-C2, C1, A11, respectively (15). The ligands of the remaining KIRs remain unknown.

As KIR genes and human leukocyte antigen (HLA) genes are located on different chromosomes, autologous KIR receptor-ligand mismatch may exist (16). Normally, NK cells acquire self-tolerance and functional competence through the education process, in which inhibitory KIRs could be inhibited by self-HLA ligands and activated in a non-self HLA environment. Besides, the decreased responsiveness of activating KIRs in the presence of their cognate ligands also prevents autoimmunity

(17–23) (**Figure 2A**). Importantly, infected and/or tumor cells may express inhibitory KIR ligands insufficiently or express activating ligands that may activate NK cells (24–31).

As the first reconstituted lymphocyte subset after transplantation (32, 33), NK cells play a critical role in controlling early relapse and infections. They also possess the ability to eliminate recipient T cells and antigen-presenting cells (APCs), to prevent graft failure and GVHD (34-38) (Figure 2B). Three models were established historically in an attempt to optimize donor selection for HSCT based on KIR (Figure 2A). The Perugia group in Italy firstly proposed the donor-recipient KIR ligand-ligand model (also known as KIR ligand model) solely based on the HLA phenotype of the donor and recipient. The KIR ligand incompatibility in the GVH direction was defined as the absence in recipients of donor class I allele group(s) recognized by KIRs. Those authors observed that the HLA haplotype-mismatched transplants reduced the rejection and relapse rate and prevented GVHD in patients with acute myeloid leukemia (AML) (36). Subsequently, the second model (named receptor-ligand model or missing ligand model) was raised by Leung et al. based on the compatibilities between the recipient HLA and donor inhibitory KIR. This model focused on donor KIR instead of donor HLA and could, therefore, be used in both HLA-matched and HLA-mismatched transplants. The results of that study suggested that the receptor-ligand model better predicted the risk of primary disease relapse, especially for lymphoid malignancies, compared with the ligand-ligand model (39). Subsequently, with a deeper understanding of KIR haplotypes, the third model analyzed and compared the KIR genotypes of different donors. Cooley et al. showed that unrelated donors with KIR-B haplotypes conferred a significant relapsefree survival (RFS) benefit to patients with AML undergoing T cell-replete HSCT (40). Based on the three models described above, numerous studies have been carried out to explore the impact of NK cell alloreactivity. Clinical results obtained from KIR ligand model, receptor ligand model and KIR haplotype and gene model were summarized in Tables 2-4, respectively. Nevertheless, the results were controversial, and several key questions remained regarding NK cell biology post-HSCT. What are the exact effects of NK cell alloreactivity on patients after HSCT? How do NK cells reconstitute post-HSCT and which factors may interfere with the reconstitution process? This review summarizes the latest literature on this important topic and offer some instructive hypothesis.

KIR AND TRANSPLANT OUTCOMES

NK Cell Alloreactivity and GVHD

GVHD is an important complication of HSCT with high morbidity and mortality in which allogeneic donor immune cells are activated by APCs and then recognize and attack the host tissue (105). Removing donor T cells from grafts reduces the occurrence of GVHD, while it also elevates the risk of graft failure and disease relapse (106–108).

As another component of immune cells, previous murine studies suggested that adoptive transfer of interleukin-2 (IL-2)-activated SCID NK cells with donor bone marrow cells

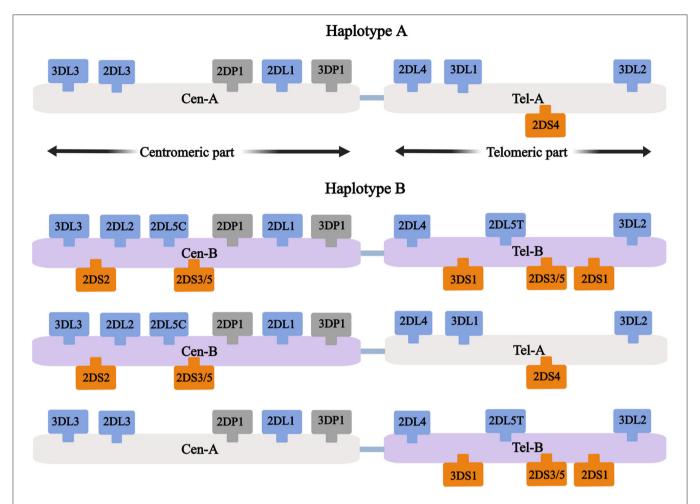


FIGURE 1 | Simplified genomic maps of KIR. Inhibitory KIR genes are color-coded in blue, activating KIR genes in orange, and pseudogenes in gray. KIR haplotype A has only one activating KIR gene: KIR2DS4, KIR B haplotype has fixable content of activating KIR genes. KIR haplotype could be further determined as Cen haplotype and Tel haplotype.

promoted engraftment in allogenic hosts with no signs of GVHD (109). Later, Asai et al. reported that hosts receiving MHCincompatible bone marrow and spleen cells (as a source of T cells) rapidly succumbed to acute GVHD, while hosts who additionally received IL-2-activated donor NK cells on day 0 experienced a significant improvement in survival because of the lower incidence of severe GVHD. They further demonstrated that that the protective effect on GVHD was dependent on the transforming growth factor-beta (TGF-β) and could be abrogated by an anti-TGF-β antibody (35). Moreover, Ruggeri et al. showed that pre-transplant alloreactive Ly49 (Ly49 receptors recognize major histocompatibility complex (MHC) class I molecules in mice, which is analogous to KIR in humans) ligand-mismatched donor NK cell transfusion successfully eliminated host tumor cells and protected against GVHD by depleting host APCs. In contrast, hosts receiving bone marrow grafts without NK cell infusion died of GVHD, and non-alloreactive Ly49 ligand matched NK cell infusion did not provide protection against GVHD (36). Consistently, subsequent studies also found that donor alloreactive NK cells suppressed GVHD by inhibiting T cell proliferation and activation (37, 110). However, the protective role of NK cells in GVHD pathogenesis has also been challenged. Pre-clinical evidence from a xenogeneic model showed that an *in vitro* IL-2-activated human NK cell infusion promoted GVHD in SCID mice via the production of cytokines such as IFN- γ and tumor necrosis factor- α (TNF- α) (111, 112). Accordingly, GVHD was inhibited after the administration of anti-IFN- γ and depletion of Poly I:C-activated NK cells in murine studies (113, 114).

In patients with hematological malignancies, a purified (115, 116) or cytokine-induced (117–121) donor NK cell transfusion was also well tolerated and seldom induced severe GVHD (grade III-IV acute GVHD or moderate-to-severe chronic GVHD). More recently, a pilot study suggested that, after haplo-HSCT, patients with refractory AML who received a donor NK cell infusion experienced a significantly lower grade II-IV GVHD than did those without NK cell infusion (122). In contrast, Shah et al. observed that patients who received a donor

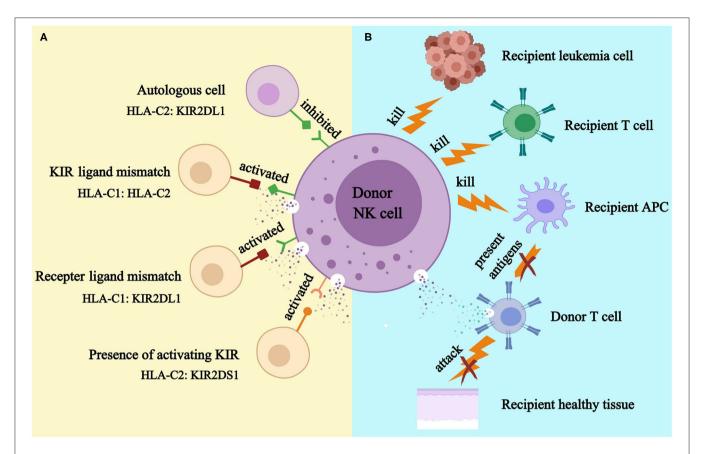


FIGURE 2 | KIR models (A) and NK cell-mediated killing (B). APC, antigen presenting cell. (A). Donor NK cell is tolerant to self because donor inhibitory KIR is inhibited by its cognate HLA ligand; donor NK cell might kill recipient cell because HLA ligand for donor inhibitory KIR presents in donor but absents in recipient (KIR ligand model); donor NK cell could kill recipient cell because recipient HLA ligand does not inhibit donor inhibitory KIR (receptor ligand model); donor NK cell could kill recipient cell because donor activating KIR is activated by recipient (KIR B haplotypes and KIR B genes). (B). Alloreactive donor NK cell could kill recipient leukemia cell to prevent relapse; it could kill recipient T cell to prevent graft rejection; and it could kill recipient APC to prevent GVHD.

IL-15/4-1BBL-activated NK cell infusion after T cell-depleted (TCD) stem cell transplantation experienced a high risk of GVHD (123).

In addition to the technique of adoptive transfer, many studies have analyzed the effects of innate donor-recipient NK cell alloreactivity on GVHD in a clinical setting. The majority of studies did not report a significant association between these parameters (41–44, 46, 47, 50, 51, 54–56, 59, 65, 66, 79, 81, 83, 87–89, 91–93, 97, 98, 102, 104), while some reported a protective effect (70, 74, 76). Moreover, several studies found that KIR ligand mismatch or receptor-ligand mismatch increased the risk of GVHD (45, 57, 60, 64, 68, 80). Accordingly, two studies performed in China that applied the 'Peking protocol' for HSCT using the granulocyte-colony stimulating factor (G-CSF)-mobilized graft containing a high dose of T cells observed promotive effects of NK cell alloreactivity on GVHD (48, 49).

It is not entirely clear why the reconstituted alloreactive NK cells were unable to prevent GVHD as the adoptively transferred NK cells. Studies have indicated that this discrepancy was probably attributable to the impaired function of early reconstituted NK cells. Shilling et al. first observed that a period

of several months or even years was required for the recipient to reconstitute an NK cell repertoire resembling that of the donor (124). Vago et al. also suggested that the NK cells that were reconstituted early after transplantation were immature and exhibited compromised cytotoxicity (125). In addition, NK cell reconstitution is affected by graft composition. Patients receiving more T cells in grafts experience a faster T cell reconstitution (126, 127), while the absolute number of reconstituted NK cells and KIR expression are impaired by the co-grafted T cells (127-130). Other than NK cells, nearly 5% of CD8⁺ T cells, 0.2% of CD4⁺ T cells, and 10% of $\gamma\delta$ T cells in the peripheral blood also express KIRs (131–133). Therefore, it is possible that the potential beneficial effects of alloreactive NK cells are overwhelmed by the strong alloreactive T cell response. In addition, it was observed that NK cells generated more IFN-y in the presence of T cells in grafts, leading to a higher occurrence of acute GVHD (aGVHD) (130). Moreover, post-transplant immune suppression also exerted negative effects on NK cell reconstitution (134, 135).

Regarding specific genotypes, some studies have reported that KIR haplotype B donors afforded a significantly reduced risk of GVHD (60, 63, 86, 96). Consistent with these findings,

TABLE 2 | Impact of KIR on clinical outcomes in KIR ligand model.

References N		Disease Donor		Graft manipulation	Clinical outcomes
Ruggeri et al. (36)	92	AML, ALL	HRD	TCD*	KIR ligand mismatch: higher EFS and OS, lower relapse (AML) KIR ligand mismatch: lower aGVHD ²⁻⁴
Davies et al. (41)	175	Mixed	URD	TCD*, TCR	KIR ligand mismatch: lower OS (myeloid cohort)
Giebel et al. (42)	130	Mixed	URD	TCD#	KIR ligand mismatch: higher OS and DFS, lower TRM
Schaffer et al. (43)	190	Mixed	URD	TCD*, TCD#	KIR ligand mismatch: higher IRM and TRM, and lower OS
Elmaagacli et al. (44)	236	CML	MSD, URD	TCR	KIR ligand mismatch: lower molecular relapse
Yabe et al. (45)	1489	Mixed	URD	TCD#, TCR	KIR ligand mismatch: higher aGVHD $^{2/3-4}$ and lower OS (HLA-C mismatched transplants)
Verneris et al. (46)	716	Pediatric AL	URD	TCD#, TCR	KIR ligand mismatch: no significant impact on OS, DFS, relapse, TRM, or aGVHD.
Ruggeri et al. (47)	112	AML	HRD	TCD*	KIR ligand mismatch: lower relapse (CR group), higher EFS, and lower risk of relapse or death
Huang et al. (48)	116	Mixed	HRD	TCD#	KIR ligand mismatch: higher aGVHD2-4 and relapse, lower OS
Zhao et al. (49)	64	Mixed	HRD	TCD#	KIR ligand mismatch: higher aGVHD;
Michaelis et al. (50)	57	Mixed	HRD	TCD*	KIR ligand mismatch: lower EFS (AML)
Mancusi et al. (51)	161	AML, ALL	HRD	TCD* TCD*+Treg/Tcon	NK-alloreactive donors: lower relapse and higher EFS (AML)
Yahng et al. (52)	100	AML	HRD	TCD#	KIR ligand mismatch (HVG): higher relapse and CMV reactivation, lower DFS
Zhao et al. (53)	180	Mixed	HRD	TCD#	KIR ligand match: lower CMV reactivation rate and higher IFN- $\!\gamma$ expression
Wanquet et al. (54)	144	Mixed	HRD	TCD#	KIR ligand mismatch: lower relapse and higher PFS (no CR group)
Shimoni et al. (55)	444	AML, ALL	HRD	TCD#	KIR ligand mismatch: a trend of higher relapse (AML), lower OS

MSD, matched sibling donor; URD, unrelated donor; HRD, haploidentical related donor; AML, acute myeloid leukemia; ALL, acute lymphoid leukemia; CML, chronic myeloid leukemia; TCD, T cell depleted; TCR: T cell replete; Treg, regulatory T cells; Tcon, conventional T cells; aGVHD: acute graft vs. host disease; cGVHD: chronic graft vs. host disease; OS, overall survival; RFS, relapse free survival; DFS, disease free survival; EFS, event free survival; IRM: infection related mortality; TRM: transplant related mortality; CMV, cytomegalovirus. TCD*: ex-vivo TCD.

TCD#: in-vivo TCD.

Sivori et al. suggested that donor NK cells expressing KIR2DS1 were efficient in killing allogenic dendric cells in the setting of haplo-HSCT, thus leading to a better GVHD control (136). However, several studies also found that donors with KIR-B/x led to higher GVHD occurrence in recipients compared with donors with A/A, probably because of the more potent production of IFN- γ by alloreactive NK cells (40, 45, 76, 77, 94, 95).

Other factors, such as HLA mismatch, disease type, patient age, GVHD prophylaxis, and graft source, were also reported to interfere with GVHD occurrence in these studies (44, 45, 63, 66, 87, 92, 93, 104). Collectively, the manner in which the reconstituted NK cells affect the risk of GVHD remains largely unknown, and the relationships between NK and T cells during the initiation and process of GVHD warrant further investigation.

NK Cell Alloreactivity and Infection

Infections are especially challenging for patients after HSCT because of the immunological derangement caused by multiple factors, including an intensive conditioning regimen, immunosuppressive agents, and other complications, such as GVHD (137, 138).

Several studies have reported that patients receiving KIR ligand-mismatched transplants are more vulnerable to infections. Schaffer et al. first reported that KIR ligand mismatch was

associated with an increased infection-related mortality (43). Similarly, results from Zhao et al. showed that recipients from the KIR ligand-mismatched group experienced a significantly higher cytomegalovirus (CMV) reactivation rate. Moreover, the percentage of interferon-gamma (IFN-γ)-expressing NK cells in the peripheral blood was significantly higher in the KIR ligand matched group 30 and 100 days post-HSCT compared with the KIR ligand-mismatched group (53). The higher level of IFN-γ secretion from the NK cells might trigger Th1 immune responses, antigen presentation cell activation, and macrophage killing (7, 8), leading to lower infection rate. While, KIR ligand mismatch may increase the risk of infection by eliminating recipient APCs by donor alloreactive NK cells (36).

Many studies have found that KIR-B genes protect patients with HSCT against infections and most of them were predominantly T cell replete (TCR) transplants (81, 84, 87, 96, 139, 140). Cook et al. first observed that KIR haplotype B donors exhibited a significant reduction in the rate of CMV reactivation in sibling allo-HSCT (139). Wu et al. and Zaia et al. reported that donors expressing higher numbers of activating KIRs were associated with a lower CMV reactivation rate (62, 84). Specifically, activating KIR2DS2 and KIR2DS4 may play a major protective role (84, 140). Importantly, transplantations from donors with KIR2DS1 correlated with better infectious control (51, 96). Mancusi et al. further demonstrated that the binding

TABLE 3 | Impact of KIR on clinical outcomes in receptor ligand model.

References N		Disease	Donor	Graft manipulation	Clinical outcomes
Leung et al. (39)	36	Mixed	HRD	TCD*	Receptor ligand mismatch: lower relapse
Cook et al. (56)	220	Mixed	MSD	/	HLA-C2C2 patients vs. HLA-C1/x patients: lower OS (myeloid cohort)
Verheyden et al. (57)	65	Mixed	MSD	TCD*, TCR	HLA-C1C2 patients vs. HLA-C1C1 or C2C2 patients: lower aGVHD
Hsu et al. (58)	1770	Mixed	URD	TCR	Missing ligand for donor iKIR: lower relapse (HLA mismatched transplants)
Clausen et al. (59)	43	Mixed	MSD	TCR	Ligand missing to KIR3DL2 plus one other iKIR vs. others: lower relapse and higher OS
Ludajic et al. (60)	124	Mixed	URD	TCD#, TCR	Missing ligand for donor KIR2DL1: higher aGVHD ²⁻⁴ ;
Linn et al. (61)	151	Mixed	MSD	TCR	Missing ligand for donor iKIR: no impact on OS and RFS
Wu et al. (62)	48	Mixed	URD	TCD#	HLA group C1 vs. C2: higher CMV reactivation rate
Gagne et al. (63)	264	Mixed	URD	TCR	Missing HLA-C1 ligand: lower OS (myeloid cohort)
Clausen et al. (64)	100	Mixed	MSD	TCR	HLA-C1C2 patients vs. HLA-C1C1 or C2C2 patients: lower relapse and aGVHD $^{2-4}$, higher RFS
Björklund et al. (65)	105	AML, MDS	MSD	TCD#, TCR	Receptor ligand mismatch: no significant impact on OS, relapse and GVHD
Wu et al. (66)	116	Mixed	URD	TCD#, TCR	Missing ligand for donor iKIR: lower relapse, higher OS and DFS (myeloid cohort);
Zhou et al. (67)	219	Mixed	MSD	/	HLA-C1C1 patients vs. HLA-C2/x patients: lower aGVHD ²⁻⁴
Sobecks et al. (68)	909	AML, MDS	URD	TCD#, TCR	Missing ligand for donor iKIR: higher aGVHD ³⁻⁴ and TRM (AML); Missing HLA-C2 for donor KIR2DL1: higher aGVHD ^{2/3-4} (AML)
Park et al. (69)	59	Mixed	MSD, URD	TCD#, TCR	Receptor ligand mismatch: higher OS, DFS and lower relapse
Cardozo et al. (70)	50	Mixed	MSD	TCR	Patients with all ligands present vs. missing ligand for donor iKIR: higher aGVHD; Missing ligand for donor iKIR: higher OS (myeloid cohort)
Faridi et al. (71)	281	Mixed	MSD, URD	TCD#	Missing ligand for donor iKIR: lower relapse and better RFS (URD)
Neuchel et al. (72)	1446	Mixed	URD	TCR	HLA-C2C2 vs. HLA-C1/x patients: lower OS, DFS, higher relapse (myeloid cohort)
Arima et al. (73)	10638	Mixed	MSD, URD	TCD*, TCD# TCR	HLA-C1C1 patients vs. HLA-C1C2 patients: lower relapse and higher RFS (AML and CML); HLA-C1C1 patients vs. HLA-C1C2 patients: higher relapse (ALL)
Gaafar et al. (74)	87	Mixed	MSD	TCR	KIR2DL1: HLA-C2 match: higher aGVHD ²⁻⁴ (AML)
Arima et al. (75)	2884	ALL	MSD, URD	TCD, TCR	HLA-C1C1 patients vs. HLA-C1C2 patients: higher relapse
Chen et al. (76)	84	Mixed	HRD	TCD#	Missing HLA-C2 ligand for donor KIR2DL1: higher OS and lower RRM (myeloid cohort); Missing HLA-C for donor iKIR: lower aGVHD ²⁻⁴ (lymphoid cohort);
Zhao et al. (77)	97	CML	HRD	TCD#	Receptor ligand match: lower relapse
Zhao et al. (78)	188	Mixed	HRD	TCD#	Receptor ligand match: lower relapse and higher LFS
Solomon et al. (79)	208	Mixed	HRD	TCD#	Receptor ligand mismatch: higher OS and DFS, lower relapse
Willem et al. (80)	51	Mixed	HRD	TCD#	KIR2DL/HLA mismatch: higher GVHD and lower relapse

AL, acute leukemia; MDS, myelodysplastic syndromes; iKIR, inhibitory KIR; LFS, leukemia-free survival.

of KIR2DS1 to HLA-C2 triggered pro-inflammatory cytokine production by alloreactive NK cells (51). Moreover, without a cognate ligand (HLA-C1) in recipients, donor KIR2DS2 was associated with a higher CMV reactivation rate after HLA-identical sibling HSCTs (81). Apart from CMV reactivation, the incidence of bacterial infections was also reduced when patients had KIR-B/x donors (87). In contrast with previous results, KIR2DS2 gene and Cen-B/x donors related to a higher incidence of CMV reactivation and infection-related mortality in TCD transplants (53, 100). The reasons for these differing results may be due to the different graft composition. As previously described, NK cells generate more IFN-γ in TCR transplants,

which may benefit the infection control (130). Of notice, the activating KIR targets outside of HLA are largely unknown, and these clinical observations still need to be confirmed by definitive functional analysis in the future.

NK Cell Alloreactivity and Relapse/Survival

Primary disease relapse remains the main obstacle that hampers the long-term survival of patients with hematological malignancies. Previous experience showed that adoptive transfer of autologous NK cell for patients with tumors was safe but inefficient (141–145), probably because autologous NK cells could not overcome the inhibition mediated by

TABLE 4 | Impact of KIR on clinical outcomes in KIR haplotype and gene model.

References	N	Disease	Donor	Graft manipulation	Clinical outcomes	
Cooley et al. (40)	448	AML	URD	TCR	KIR B/x donor: higher RFS and cGVHD	
Cook et al. (56)	220	Mixed	MSD	/	KIR2DS2: lower OS (HLA-C2C2 patients with myeloid diseases)	
Verheyden et al. (57)	65	Mixed	MSD	TCD*, TCR	Donor co-presenting KIR2DS1 and 2DS2: lower relapse	
Chen et al. (81)	131	Mixed	MSD	TCR	KIR2DS2: higher CMV reactivation (HLA-C2C2 patients); Additional activating KIR genes in donor: higher OS and lower CMV reactivation	
Yabe et al. (45)	1489	Mixed	URD	TCD#, TCR	KIR2DS2: higher aGVHD3-4 (HLA-C mismatched transplants)	
Schellekens et al. (82)	83	Mixed	MSD	TCR	KIR2DS1: higher OS (HLA-C1C1 patients); More activating KIRs in donor or patients: higher relapse; KIR2DS5 in patients or both in donor and patients: higher relapse	
van der Meer et al. (83)	70	Mixed	MSD	TCD*	KIR2DS5: higher LFS and lower relapse (HLA-C1C1 or HLA-C2C2 patients); KIR2DS5: lower LFS and higher relapse (HLA-C1C2 patients)	
Ludajic et al. (60)	124	Mixed	URD	TCD#, TCR	KIR2DS2: lower aGVHD ²⁻⁴ (HLA-C1C2 patients)	
Zaia et al. (84)	211	Mixed	MSD, URD	TCR	Donor co-presenting KIR 2DS2 and 2DS4: lower CMV reactivation; Donor aKIR gene content ≥5: lower CMV reactivation	
Wu et al. (62)	48	Mixed	URD	TCD#	High aKIRs group: lower CMV reactivation rate	
Gagne et al. (63)	264	Mixed	URD	TCR	KIR B/x donor: lower aGVHD $^{3-4}$ (HLA identical pairs with myeloid disease)	
Bao et al. (85)	75	Mixed	URD	TCD#	KIR B/x donor: higher OS	
Venstrom et al. (86)	1087	Mixed	URD	TCD*, TCR	KIR3DS1: lower aGVHD $^{2-4}$; KIR3DS1: lower aGVHD $^{2-4}$, TRM and mortality (AML, CML and ALL)	
Wu et al. (66)	116	Mixed	URD	TCD#, TCR	KIR2DS3: higher relapse, lower OS and DFS (myeloid cohort); More numbers of activating KIR genes in donor: higher relapse	
Tomblyn et al. (87)	116	Mixed	URD	TCD*, TCR	KIR B/x donor: lower bacterial infections by day 180	
Cooley et al. (88)	1409	AML, ALL	URD	TCR	KIR B/x donor: lower relapse and higher DFS (AML); Cen-BB vs. Cen-BA or AA: lower relapse and higher DFS (AML); Tel-B/x vs. Tel-AA: lower relapse (AML); B content ≥ 2: lower relapse (AML)	
Venstrom et al. (89)	1277	AML	URD	TCD*, TCR	Donor KIR2DS1 with HLA-C1/x patients vs. with HLA-C2C2 patients: lower relapse; KIR3DS1: higher OS	
Zhou et al. (67)	219	Mixed	MSD	/	Cen-B/x donor: higher OS, RFS and lower relapse	
Impola et al. (90)	134	Mixed	MSD	/	KIR 2DL2 or KIR 2DS2: better RFS (AML)	
Bao et al. (91)	210	Mixed	URD	TCD#	KIR B/x donor: higher OS, RFS and lower NRM (AML and MDS); Cen-B/x donor: higher OS, RFS (AML and MDS at standard risk)	
Cardozo et al. (70)	50	Mixed	MSD	TCR	KIR2DS2: lower OS and EFS	
Bachanova et al. (92)	614	NHL	URD	TCD#, TCR	KIR B/x donor: lower relapse and better PFS (HLA matched transplants	
Kamenaric et al. (93)	111	Mixed	MSD, URD	TCD#	KIR2DS4 (neg vs. pos): no impact on GVHD (MSD)	
Hosokai et al. (94)	106	Mixed	MSD, URD	TCR	KIR B/x donor: higher aGVHD $^{3-4}$ (more evdient in HLA mismatched transplants)	
Neuchel et al. (72)	1446	Mixed	URD	TCR	KIR2DS2: higher OS and DFS (HLA-C2C2 patients); KIR2DS1: lower relapse but higher TRM (HLA-C2C2 patients); KIR2DS5: lower relapse (HLA-C2C2 patients)	
Gaafar et al. (74)	87	Mixed	MSD	TCR	KIR2DS2: HLA-C1 match: higher aGVHD ²⁻⁴ (AML); KIR2DS1: HLA-C2 match: higher cGVHD (AML); Donor presenting KIR2DL1 or 2DS2: higher cGVHD (AML)	
Sahin et al. (95)	96	AML, CML	MSD	TCR	KIR B/x donor: higher cGVHD	
Heatley et al. (96).	152	Mixed	MSD	TCR	KIR2DS2: higher OS (AML); Cen-B/x donor: higher OS (AML) and lower aGVHD ²⁻⁴ (AML); Tel B/x donor: lower CMV reactivation	
Babor et al. (97)	317	Pediatric ALL	MSD, URD	TCD#, TCR	Higher ct-KIR score: lower relapse	
Tordai et al. (98)	314	Mixed	MSD, URD	/	The combination of KIR2DS1 donor with HLA-C2 pos patients: higher OS	

(Continued)

TABLE 4 | Continued

References	N	Disease	Donor	Graft manipulation	Clinical outcomes
Nakamura et al. (99)	288	AML	MSD, URD	TCD*, TCD#	CMV reactivation: lower relapse and higher NRM (more evident in KIR B/x donor or when donor presenting KIR2DS1)
Bultitude et al. (100)	119	AML	URD	TCD, TCR	Cen-B/x donor: lower OS and NRM, higher IRM
Weisdorf et al. (101)	2662	AML	URD	TCD#, TCR	KIR B/x donor: lower relapse and higher LFS (RIC)
Verneris et al. (46)	716	Pediatric AL	URD	TCD#, TCR	KIR gene content: no significant impact on OS, DFS, relapse, TRM, or aGVHD
Zhao et al. (49)	64	Mixed	HRD	TCD#	KIR2DS3: higher aGVHD and cGVHD; KIR2DS5: higher aGVHD
Symons et al. (102)	86	Mixed	HRD	TCD#	KIR B/x donor: lower NRM and higher OS, EFS (KIR AA patients)
Chen et al. (76)	84	Mixed	HRD	TCD#	KIR2DS2: higher OS (lymphoid cohort); KIR2DS1: higher GVHD (lymphoid cohort)
Michaelis et al. (50)	57	Mixed	HRD	TCD*	KIR B/x donor: lower relapse
Zhao et al. (77)	97	CML	HRD	TCD#	KIR2DS3: lower EFS and OS, higher TRM; KIR2DS5: higher EFS and OS, lower TRM; KIR B/x donor: higher aGVHD ³⁻⁴
Oevermann et al. (103)	85	Pediatric ALL	HRD	TCD*	KIR B/x donor: lower relapse and better EFS; High donor KIR-B content: lower relapse and better EFS
Mancusi et al. (51)	cusi et al. (51) 161 AML, ALL HRD TCD* Tel B/x vs. Tel AA: lower NRM and TCD*+Treg/Tcon KIR2DS1/3DS1: lower NRM and h		Tel B/x vs. Tel AA: lower NRM and higher EFS (NK-alloreactive donors) KIR2DS1/3DS1: lower NRM and higher EFS (NK-alloreactive donors) KIR 2DS1 binding to HLA C2: increased inflammatory cytokine		
Zhao et al. (53)	180	Mixed	HRD	TCD#	KIR2DS2: higher CMV reactivation
Solomon et al. (79)	208	Mixed	HRD	TCD#	KIR B/x donor with 2DS2 vs. KIR B/x donor without 2DS2: higher OS and DFS, lower relapse and NRM; KIR B/x donor with 2DS2 vs. KIR A/A donor: higher OS and DFS, lower NRM
Perez-Martinez et al. (104)	192	Pediatric mixed	HRD	TCD*, TCD#	KIR AA donor: higher relapse and lower DFS

pos: positive; neg: negative; NHL, non-Hodgkin lymphoma; PFS, progression-free survival; NRM: non-relapse mortality. TCD*: ex-vivo TCD; TCD#: in-vivo TCD.

tumor cells expressing self-HLA. In contrast, allogenic (117), especially haploidentical, donor NK cell infusion demonstrated wide prospects in the salvage treatment (115, 120, 121) and prophylactic treatment (118, 119) of patients with hematological malignancies. In allo-HSCT, whether the reconstituted alloreactive NK cells prevent the disease relapse remains controversial.

In HLA-mismatched transplants, the Perugia group first observed that, in the context of T cell depletion, high stem cell dose, and absence of post-transplant immune suppression, KIR ligand mismatch reduced the risk of relapse and markedly improved survival in patients with AML, but not in those with acute lymphoblast leukemia (ALL) (36). This protective effect on relapse or survival was supported by many clinical studies (42, 44, 47, 51, 54), especially in myeloid disease (44, 47, 51) and transplants with TCD grafts (42, 47, 51, 54). However, conflicting results stemmed from many studies that failed to replicate these results (39, 46, 58, 102), and some even reached the opposite conclusions (41, 43, 45, 48, 50, 55).

Studies using the receptor-ligand model including HLA-matched donor-recipient pairs also reported conflicting results. Leung et al. first reported that the receptor-ligand model was more accurate than the KIR ligand model when predicting the risk of relapse, especially for lymphoid malignancies. Moreover,

the potency of the relapse protection positively correlated with the number of receptor-ligand mismatch pairs (39). Subsequently, the protective effect of receptor-ligand mismatch has been confirmed by many investigations (58, 59, 66, 69, 71, 73, 76, 79, 80). Moreover, a survival advantage was also observed in patients with receptor-ligand mismatch compared with receptor-ligand matched pairs (59, 66, 69–71, 73, 76, 79). However, several other studies described opposite results (63, 64, 75, 77, 78). Of notice, two studies from Japan observed that the lack of the HLA-C2 ligand for donor inhibitory KIR afforded relapse protection in patients with AML and chronic myeloid leukemia, but increased the relapse rate in patients with ALL (73, 75). To date, no plausible explanation has been put forward for this disparity in relapse.

In contrast to the controversial results described above, transplantations from KIR haplotype B donors achieved greater agreement. Cooley et al. observed that patients with AML with KIR-B/x donors experienced a 30% improvement in RFS compared with those with A/A donors (40). Subsequently, many further investigations confirmed this beneficial effect of the KIR-B haplotype on relapse and survival in patients with hematological malignancies (50, 51, 57, 67, 72, 76, 79, 81, 85, 88–92, 96, 98, 101–104). Five of these studies reported that the protection effects mainly existed in the KIR Cen-B locus (67,

88, 91, 92, 96). Babor et al. further suggested that the presence of Cen-B with absence of Tel-B improved leukemia control in pediatric patients with ALL (97). At the genetic level, the KIR2DS2 gene, which is located on the Cen-B motif (72, 76, 79, 90, 92, 96), and the KIR2DS1 gene, located on the Tel-B motif (51, 72, 82, 98), were found to be related to a decreased relapse rate or an improved survival. However, several studies found that Cen-B donors indicated a lower OS (56, 70, 100). Meanwhile, Verneris et al. did not find any association between transplant outcomes and NK cell alloreactivity or KIR gene content in pediatric patients with acute leukemia (46).

Recently, Krieger et al. developed a scoring system, in which interactions of multiple KIR genes and HLA ligands were quantitatively analyzed. This comprehensive method raised an improved strategy to select a donor and exhibited great potential in the future (146).

Collectively, it is still controversial to determine an optimal donor who exhibits the best NK cell function using the three established KIR models. A better knowledge of NK cell reconstitution after HSCT may promote a better understanding of how NK cells affect the transplant outcomes in these patients. More in-depth studies focusing on "functional changes in NK cells" rather than "match or mismatch" may help us get closer to an optimal donor.

NK CELL RECONSTITUTION AFTER TRANSPLANTATION

Maturation and Differentiation of NK Cells

NK cells are derived from the CD34⁺ hematopoietic stem and precursor cells in the bone marrow, which then migrate to the periphery (147). Recent evidence suggested that not only the bone marrow, but also secondary lymphoid tissues contribute to the development of NK cells (148). According to the surface expression of CD56, NK cells could be divided in two main subtypes: CD56^{bright} and CD56^{dim} NK cells. CD56^{bright} NK cells exist mainly in lymph nodes and tonsils, while CD56^{dim} NK cells, the more mature subset transformed from CD56^{bright} NK cells, are dominant in the peripheral blood (7, 147, 149, 150). CD56^{bright} and CD56^{dim} NK cells are equipped with distinct functions. The former population responds rapidly to interleukin-mediated stimulation with proliferation and cytokine secretion, while the latter population displays higher cytolytic capacity and lower proliferation (7, 8, 149). During the process of maturation, CD94/NKG2A is the first receptor that is expressed on immature NK cells. Together with the downregulation of CD56 expression, NK cells upregulate CD16 expression, lose NKG2A, and acquire KIR receptors. Finally, a subset of CD56^{dim} cells continue to differentiate and express CD57, together with an increased KIR expression and a completely abolished proliferative ability (150, 151).

In HSCTs with post-transplant cyclophosphamide (PT-Cy) as GVHD prophylaxis, NK cells experience two waves of expansion. After graft infusion, peripheral NK cells and T cells (mainly mature cells from the donor) were detectable at very low levels. PT-Cy administration results in a further decrease in T cells and

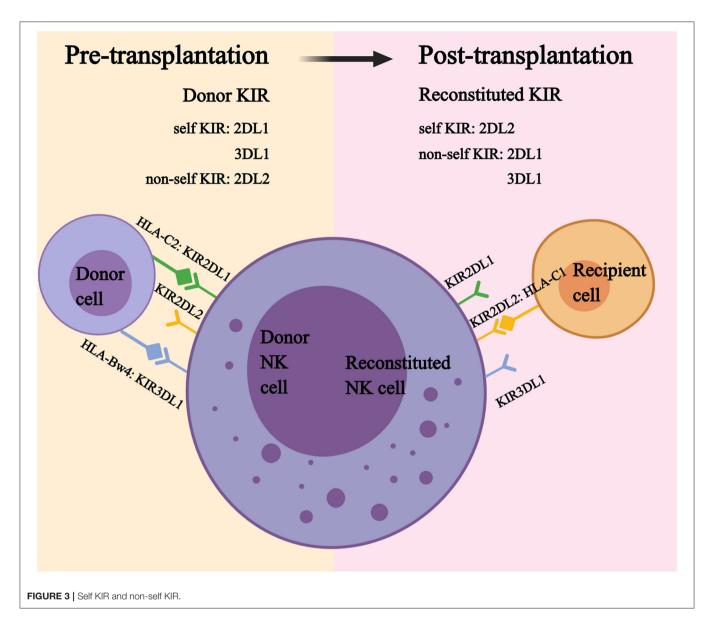
NK cells, and NK cells are barely detectable in the peripheral blood. Subsequently, the reconstituted NK cells gradually recover and express high levels of CD56 and NKG2A. Around 60 days after transplantation, the KIR expression returns to normal. The expression of CD56 and NKG2A gradually decreases and becomes stable at 9–12 months post-transplantation. Other receptors expressed on NK cells, such as DNAM-1and 2B4, also require several months to return to normal (152). In summary, post-transplantation NK cell reconstitution is a long-term process (124, 125, 152).

KIR Education: From Anergic to Responsive

As described earlier, the random combination of KIR receptor and HLA ligand can exist in healthy individuals. However, the autoimmune attack is inhibited because each NK cell expresses at least one self-inhibitory receptor. To avoid autoreactivity, NK cells must undergo an education process: NK cells expressing inhibitory KIR for self-HLA ligand (self-KIR) are educated, which means that these cells can be inhibited by self-inhibitory signals and become alloreactive against self-HLA-deficient targets. In contrast, NK cells expressing an inhibitory KIR that lacks a self-HLA ligand (non-self KIR) are uneducated, which means that they are tolerant to the self but also to infected or malignant cells (19, 21).

In the last decades, studies on KIR education have much extended our knowledge of NK cell function. After transplantation, most reconstituted NK cells express a donorlike KIR repertoire that is significantly different from that of recipient NK cells prior to transplantation (124, 151). Therefore, reconstituted NK cells expressing donor KIR may exert alloreactivity in recipients, or become anergic, as recipients may not present the cognate HLA (Figure 3). Foley et al. and Björklund et al. observed that reconstituted NK cells with nonself KIR remained tolerant, while those with self KIR acquired better functions after transplantation (65, 153). However, Yu et al. reached the opposite conclusion that alloreactive NK cells broke the self-tolerance and displayed functional capacities in the first 3 months, then gradually acquired self-tolerance by day 100 post-transplantation (154). Rathmann et al. also suggested that alloreactive NK cells were increased in the peripheral blood and exhibited a GVL effect in the early period after transplantation (155). One possible explanation for this observation is that the infusion of a megadose of donor CD34+ cells may create a transient donor dominant HLA environment in recipient bone marrow, and the early reconstituted NK cells expressing nonself KIR for the recipient may become educated by donor HLA and acquire functions (156). After migration to a recipientdominant environment, reconstituted NK cells may gradually lose their responsiveness.

In murine studies, it was observed that mature NK cells from major histocompatibility complex (MHC) class I-sufficient mice become hyporesponsive after transfusion into MHC class I-deficient mice. Conversely, anergic NK cells from MHC class I-deficient mice acquired functions after exposure to the MHC class I-sufficient environment (157, 158). Using a murine



transgenic model of HLA-B*27:05 exhibiting the Bw4 ligand for KIR3DL1, Boudreau et al. observed similar results in stem cell transplantation. CD34+ cells from KIR3DL1+ donors were transfused to B27 Tg+ and Tg- mice, respectively. A functional analysis suggested that the most cytotoxic responsive cells were KIR3DL1+ NK cells from Bw4+ donors and developed in B27 Tg+ mice (Bw4+ donors and Tg+ mice), while the least-responsive cells were KIR3DL1+ NK cells from Bw4- donors and developed in Tg- mice (Bw4- donors and Tg- mice). Recipients with the other two combinations (Bw4+ donors and Tg- mice and Bw4- donors and Tg+ mice) displayed a medium level of responsiveness. The stepwise escalation of NK cell responsiveness suggested that both the donor and recipient MHC environments are critical for the maintenance and adjustment of NK cell education (159).

Recently, the Nowak team proposed that inhibitory KIR (iKIR)-HLA pairs could predict the post-HSCT NK cell

education status, i.e., donors presenting cognate HLA for donor iKIR and recipients lacking it predict a downward education level; in contrast, recipients presenting cognate HLA for donor iKIR and donors lacking it predict an upward education level. Those authors found that the decrease in iKIR–HLA pairs post-transplantation is associated with a higher relapse and poorer survival (160–162), indicating that reconstituted NK cells acquire better functions after interaction with more cognate HLA class I ligands in recipients. Zhao et al. also observed that, when the donors and recipients expressed three major HLA ligands (HLA-C1, C2, Bw4), patients with AML and myelodysplastic syndrome (MDS) experienced the lowest relapse rate, and NK cells expressing three inhibitory receptors exhibited the greatest cytotoxicity and cytokine responsiveness against K562 targets (163).

Based on the findings described above, it is likely that three factors (donor KIR, donor HLA, and recipient HLA) all

contribute to the variation in NK cell function. Therefore, the KIR ligand and receptor-ligand models, which only take two factors into account, may not accurately predict donors that exhibit the greatest NK cell function post-transplantation.

Factors That Affect NK Cell Reconstitution

Although CMV reactivation suggests an immune-compromised state, patients who experienced CMV reactivation had a lower relapse rate or better survival (70, 98, 99, 164). This protective effect might be attributed to the rapid maturation of NK cells. During CMV reactivation, NK cells that express NKG2C rapidly expand and continue to increase for 1 year (165). The number of CD56^{dim} NK cells in the peripheral blood, their KIR expression, and IFN-y production in response to K562 cells were also elevated in patients who developed CMV reactivation (165-173). Furthermore, nearly 60% of NKG2C+ NK cells achieved complete differentiation and expressed CD57 after CMV reactivation. These cells were termed memory-like NK cells and could be detected long after the primary CMV infection, offering a long-lasting protection (147, 166). In contrast, for non-CMV-infected patients, a higher proportion of NKG2A+ NKG2C- KIR- NK cells in the peripheral blood indicates a slow NK cell maturation. Interestingly, CMV antigen exposure to recipients also leads to an increased frequency of NKG2C+ NK cells, accompanied by increased KIR expression and decreased NKG2A expression (174).

As mentioned above, T cells in the graft impair the recovery of NK cells and KIR reconstitution (127-130). A possible explanation for this observation is that T cells compete with NK cells for IL-15, a cytokine that regulates immune cell survival and development (175, 176). Unlike ex-vivo TCD grafts, pretransplant anti-thymocyte globulin (ATG) administration results in partial T cell depletion. Two recent studies found that ATG administration promoted NK cell recovery and delayed the reconstitution of CD4+ and CD8+ T cells, while sparing the effector memory T and regulatory T cells (Tregs) (177, 178). Compared with ATG, PT-Cy is more efficient in eliminating NK cells, with a higher residual ratio of CD4⁺ T cells and Tregs (179). Of note, several studies showed that T cells in the graft may contribute to a better NK cell function (153, 180). Several studies reported that CD56^{bright} NK cells in lymph nodes could be stimulated by IL-2-producing T cells, resulting in NK cell maturation with higher IFN-γ secretion and cytotoxic functions (181, 182).

The relationship between GVHD and NK cell reconstitution remains controversial. Previous studies demonstrated that GVHD correlated with an impaired NK cell reconstitution and KIR expression (183–185). Ullrich et al. found that CD56^{bright} NK cells were dramatically decreased in patients with GVHD, while CD56^{dim} NK cells, the more mature subtype, did not show significant changes (185). In addition, Hu et al. found that the NKG2A subset of CD56^{dim} NK cells was significantly decreased in patients with GVHD. Remarkably, a functional analysis showed that NKG2A⁺ NK cells from GVHD and non-GVHD patients exhibited a comparable GVL effect. Furthermore, the co-culture of donor T cells with NKG2A⁺ NK cells from non-GVHD patients suggested that NKG2A⁺ NK cells inhibit

T cell proliferation and activation, indicating that the decreased number of NKG2A⁺ NK cells might be a cause, rather than a consequence, of GVHD (186). In addition, the administration of immunosuppressive agents could also affect immune recovery. Both Ullrich et al. and Giebel et al. suggested that steroid treatment, rather than GVHD, was related to the delayed NK cell reconstitution (184, 187).

FUTURE DIRECTIONS

Numerous studies have found that alloreactive NK cells affect treatment outcomes. Although great progress has been made through both pre-clinical and clinical investigations based on the three KIR models, the controversy remains, especially regarding the benefits of KIR alloreactivity on relapse control. Recent findings showed that donor KIR, donor HLA, and recipient HLA environment all contribute to the variation of NK cell function. The newly proposed iKIR-HLA pair model needs to be further examined in the future.

NK cells, the lymphocytes that are reconstituted first after transplantation, could be negatively affected by the T cells in the graft. However, NK cell function could also be promoted through T-cell-mediated activation. The exact interactions between NK and T cells, as well as the strategy to trigger a potential synergistic NK and T cell effect remains to be investigated.

It is noteworthy that the protective role of NK cell alloreactivity in relapse protection mostly exists in myeloid disease; in fact, some studies even found that NK cell alloreactivity increased the risk of relapse for patients with lymphoid disease. The discrepancy between expressing ligands among different diseases and their binding affinity to KIR should raise more attention. In this way, we might identify which patients would benefit from the KIR-based donor selection.

CONCLUSION

In the early period after transplantation, reconstituted alloreactive NK cell may not directly influence GVHD occurrence, as it is immature and it could be affected by T cells and immunosuppressive agents. The compatibility between donor KIR and the recipient HLA ligand may protect patients from infection. In the late period after transplantation, the iKIR-HLA pair model may reflect the variation in NK cell function, and quantitative analysis of KIR-HLA interactions may provide more convincing results regarding relapse and survival.

AUTHOR CONTRIBUTIONS

YZ and HH designed. FG and YY wrote this paper. All authors revised and approved the final manuscript.

FUNDING

This work was supported by the National Natural Science Foundation of China (81670148 and 81730008) and Medical and Health Research Project of Zhejiang Province (2012KYB709).

REFERENCES

- Gratwohl A, Brand R, Frassoni F, Rocha V, Niederwieser D, Reusser P, et al. Cause of death after allogeneic haematopoietic stem cell transplantation (HSCT) in early leukaemias: an EBMT analysis of lethal infectious complications and changes over calendar time. *Bone Marrow Trans.* (2005) 36:757–69. doi: 10.1038/sj.bmt.1705140
- Wingard JR, Majhail NS, Brazauskas R, Wang Z, Sobocinski KA, Jacobsohn D, et al. Long-term survival and late deaths after allogeneic hematopoietic cell transplantation. J Clin Oncol. (2011) 29:2230–9. doi: 10.1200/JCO.2010.33.7212
- Holmqvist AS, Chen Y, Wu J, Battles K, Bhatia R, Francisco L, et al. Assessment of late mortality risk after allogeneic blood or marrow transplantation performed in childhood. *JAMA Oncol.* (2018) 4:e182453. doi: 10.1001/jamaoncol.2018.2453
- Styczynski J, Tridello G, Koster L, Iacobelli S, van Biezen A, van der Werf S, et al. Death after hematopoietic stem cell transplantation: changes over calendar year time, infections and associated factors. *Bone Marrow Trans*. (2020) 55:126–36. doi: 10.1038/s41409-019-0624-z
- Lanier LL, Le AM, Civin CI, Loken MR, Phillips JH. The relationship of CD16 (Leu-11) and Leu-19 (NKH-1) antigen expression on human peripheral blood NK cells and cytotoxic T lymphocytes. *J Immunol*. (1986) 136:4480-6.
- Almeida-Oliveira A, Smith-Carvalho M, Porto LC, Cardoso-Oliveira J, Ribeiro Ados S, Falcao RR, et al. Age-related changes in natural killer cell receptors from childhood through old age. *Hum Immunol.* (2011) 72:319– 29. doi: 10.1016/j.humimm.2011.01.009
- 7. Vivier E, Tomasello E, Baratin M, Walzer T, Ugolini S. Functions of natural killer cells. *Nat Immunol.* (2008) 9:503–10. doi: 10.1038/ni1582
- Caligiuri MA. Human natural killer cells. Blood. (2008) 112:461– 9. doi: 10.1182/blood-2007-09-077438
- Lanier LL. NK cell receptors. Annu Rev Immunol. (1998) 16:359– 93. doi: 10.1146/annurev.immunol.16.1.359
- Moretta A, Bottino C, Vitale M, Pende D, Cantoni C, Mingari MC, et al. Activating receptors and coreceptors involved in human natural killer cell-mediated cytolysis. *Annu Rev Immunol.* (2001) 19:197–223. doi: 10.1146/annurev.immunol.19.1.197
- Pegram HJ, Andrews DM, Smyth MJ, Darcy PK, Kershaw MH. Activating and inhibitory receptors of natural killer cells. *Immunol Cell Biol.* (2011) 89:216–24. doi: 10.1038/icb.2010.78
- Wilson MJ, Torkar M, Trowsdale J. Genomic organization of a human killer cell inhibitory receptor gene. *Tissue Antigens*. (1997) 49:574– 9. doi: 10.1111/j.1399-0039.1997.tb02804.x
- 13. Hsu KC, Chida S, Geraghty DE, Dupont B. The killer cell immunoglobulin-like receptor (KIR) genomic region: gene-order, haplotypes and allelic polymorphism. *Immunol Rev.* (2002) 190:40–52. doi: 10.1034/j.1600-065X.2002.19004.x
- Parham P, Moffett A. Variable NK cell receptors and their MHC class I ligands in immunity, reproduction and human evolution. *Nat Rev Immunol*. (2013) 13:133–44. doi: 10.1038/nri3370
- Manser AR, Weinhold S, Uhrberg M. Human KIR repertoires: shaped by genetic diversity and evolution. *Immunol Rev.* (2015) 267:178– 96. doi: 10.1111/imr.12316
- 16. Leung W. Use of NK cell activity in cure by transplant. *Br J Haematol.* (2011) 155:14–29. doi: 10.1111/j.1365-2141.2011.08823.x
- Kim S, Poursine-Laurent J, Truscott SM, Lybarger L, Song YJ, Yang L, et al. Licensing of natural killer cells by host major histocompatibility complex class I molecules. *Nature*. (2005) 436:709–13. doi: 10.1038/nature 03847
- Fernandez NC, Treiner E, Vance RE, Jamieson AM, Lemieux S, Raulet DH. A subset of natural killer cells achieves self-tolerance without expressing inhibitory receptors specific for self-MHC molecules. *Blood*. (2005) 105:4416–23. doi: 10.1182/blood-2004-08-3156
- Anfossi N, Andre P, Guia S, Falk CS, Roetynck S, Stewart CA, et al. Human NK cell education by inhibitory receptors for MHC class I. *Immunity*. (2006) 25:331–42. doi: 10.1016/j.immuni.2006.06.013
- Fauriat C, Ivarsson MA, Ljunggren HG, Malmberg KJ, Michaelsson
 J. Education of human natural killer cells by activating killer

- cell immunoglobulin-like receptors. *Blood.* (2010) 115:1166–74. doi: 10.1182/blood-2009-09-245746
- Schonberg K, Fischer JC, Kogler G, Uhrberg M. Neonatal NK-cell repertoires are functionally, but not structurally, biased toward recognition of self HLA class I. *Blood*. (2011) 117:5152–6. doi: 10.1182/blood-2011-02-334441
- Vivier E, Nunes JA, Vely F. Natural killer cell signaling pathways. Science. (2004) 306:1517–9. doi: 10.1126/science.1103478
- Valiante NM, Uhrberg M, Shilling HG, Lienert-Weidenbach K, Arnett KL, D'Andrea A, et al. Functionally and structurally distinct NK cell receptor repertoires in the peripheral blood of two human donors. *Immunity*. (1997) 7:739–51. doi: 10.1016/S1074-7613(00)80393-3
- Demanet C, Mulder A, Deneys V, Worsham MJ, Maes P, Claas FH, et al. Down-regulation of HLA-A and HLA-Bw6, but not HLA-Bw4, allospecificities in leukemic cells: an escape mechanism from CTL and NK attack? *Blood*. (2004) 103:3122–30. doi: 10.1182/blood-2003-07-2500
- Masuda K, Hiraki A, Fujii N, Watanabe T, Tanaka M, Matsue K, et al. Loss or down-regulation of HLA class I expression at the allelic level in freshly isolated leukemic blasts. *Cancer Sci.* (2007) 98:102–8. doi: 10.1111/j.1349-7006.2006.00356.x
- Verheyden S, Ferrone S, Mulder A, Claas FH, Schots R, De Moerloose B, et al. Role of the inhibitory KIR ligand HLA-Bw4 and HLA-C expression levels in the recognition of leukemic cells by Natural Killer cells. *Cancer Immunol Immunother*. (2009) 58:855–65. doi: 10.1007/s00262-008-0601-7
- Reusing SB, Manser AR, Enczmann J, Mulder A, Claas FH, Carrington M, et al. Selective downregulation of HLA-C and HLA-E in childhood acute lymphoblastic leukaemia. Br J Haematol. (2016) 174:477–80. doi: 10.1111/bjh.13777
- Cerwenka A, Lanier LL. Natural killer cells, viruses and cancer. Nat Rev Immunol. (2001) 1:41–9. doi: 10.1038/35095564
- Seliger B, Ritz U, Ferrone S. Molecular mechanisms of HLA class I antigen abnormalities following viral infection and transformation. *Int J Cancer*. (2006) 118:129–38. doi: 10.1002/ijc.21312
- Pende D, Marcenaro S, Falco M, Martini S, Bernardo ME, Montagna D, et al. Anti-leukemia activity of alloreactive NK cells in KIR ligand-mismatched haploidentical HSCT for pediatric patients: evaluation of the functional role of activating KIR and redefinition of inhibitory KIR specificity. *Blood.* (2009) 113:3119–29. doi: 10.1182/blood-2008-06-164103
- Thiruchelvam-Kyle L, Hoelsbrekken SE, Saether PC, Bjornsen EG, Pende D, Fossum S, et al. The activating human NK Cell receptor KIR2DS2 recognizes a beta2-microglobulin-independent ligand on cancer cells. *J Immunol*. (2017) 198:2556–67. doi: 10.4049/jimmunol.1600930
- Ogonek J, Kralj Juric M, Ghimire S, Varanasi PR, Holler E, Greinix H, et al. Immune reconstitution after allogeneic hematopoietic stem cell transplantation. Front Immunol. (2016) 7:507. doi: 10.3389/fimmu.2016.00507
- de Witte MA, Sarhan D, Davis Z, Felices M, Vallera DA, Hinderlie P, et al. Early reconstitution of NK and gammadelta T cells and its implication for the design of post-transplant immunotherapy. *Biol Blood Marrow Transplant*. (2018) 24:1152–62. doi: 10.1016/j.bbmt.2018.02.023
- Alvarez M, Sun K, Murphy WJ. Mouse host unlicensed NK cells promote donor allogeneic bone marrow engraftment. *Blood.* (2016) 127:1202– 5. doi: 10.1182/blood-2015-08-665570
- Asai O, Longo DL, Tian ZG, Hornung RL, Taub DD, Ruscetti FW, et al. Suppression of graft-versus-host disease and amplification of graft-versustumor effects by activated natural killer cells after allogeneic bone marrow transplantation. J Clin Invest. (1998) 101:1835–42. doi: 10.1172/JCI1268
- Ruggeri L, Capanni M, Urbani E, Perruccio K, Shlomchik WD, Tosti A, et al. Effectiveness of donor natural killer cell alloreactivity in mismatched hematopoietic transplants. Science. (2002) 295:2097–100. doi: 10.1126/science.1068440
- Olson JA, Leveson-Gower DB, Gill S, Baker J, Beilhack A, Negrin RS. NK cells mediate reduction of GVHD by inhibiting activated, alloreactive T cells while retaining GVT effects. *Blood.* (2010) 115:4293– 301. doi: 10.1182/blood-2009-05-222190
- 38. Hu B, Bao G, Zhang Y, Lin D, Wu Y, Wu D, et al. Donor NK Cells and IL-15 promoted engraftment in nonmyeloablative allogeneic bone marrow transplantation. *J Immunol.* (2012) 189:1661–70. doi: 10.4049/jimmunol.1103199

 Leung W, Iyengar R, Turner V, Lang P, Bader P, Conn P, et al. Determinants of antileukemia effects of allogeneic NK cells. *J Immunol.* (2004) 172:644– 50. doi: 10.4049/jimmunol.172.1.644

- Cooley S, Trachtenberg E, Bergemann TL, Saeteurn K, Klein J, Le CT, et al. Donors with group B KIR haplotypes improve relapse-free survival after unrelated hematopoietic cell transplantation for acute myelogenous leukemia. *Blood.* (2009) 113:726–32. doi: 10.1182/blood-2008-07-171926
- Davies SM, Ruggieri L, DeFor T, Wagner JE, Weisdorf DJ, Miller JS, et al. Evaluation of KIR ligand incompatibility in mismatched unrelated donor hematopoietic transplants. Killer immunoglobulin-like receptor. *Blood*. (2002) 100:3825–7. doi: 10.1182/blood-2002-04-1197
- Giebel S, Locatelli F, Lamparelli T, Velardi A, Davies S, Frumento G, et al. Survival advantage with KIR ligand incompatibility in hematopoietic stem cell transplantation from unrelated donors. *Blood.* (2003) 102:814– 9. doi: 10.1182/blood-2003-01-0091
- Schaffer M, Malmberg KJ, Ringden O, Ljunggren HG, Remberger M. Increased infection-related mortality in KIR-ligand-mismatched unrelated allogeneic hematopoietic stem-cell transplantation. *Transplantation*. (2004) 78:1081–5. doi: 10.1097/01.TP.0000137103.19717.86
- 44. Elmaagacli AH, Ottinger H, Koldehoff M, Peceny R, Steckel NK, Trenschel R, et al. Reduced risk for molecular disease in patients with chronic myeloid leukemia after transplantation from a KIR-mismatched donor. *Transplantation*. (2005) 79:1741–7. doi: 10.1097/01.TP.0000164500.16052.3C
- 45. Yabe T, Matsuo K, Hirayasu K, Kashiwase K, Kawamura-Ishii S, Tanaka H, et al. Donor killer immunoglobulin-like receptor (KIR) genotype-patient cognate KIR ligand combination and antithymocyte globulin preadministration are critical factors in outcome of HLA-C-KIR ligand-mismatched T cell-replete unrelated bone marrow transplantation. Biol Blood Marrow Transplant. (2008) 14:75–87. doi: 10.1016/j.bbmt.2007.09.012
- Verneris MR, Miller JS, Hsu KC, Wang T, Sees JA, Paczesny S, et al. Investigation of donor KIR content and matching in children undergoing hematopoietic cell transplantation for acute leukemia. *Blood Adv.* (2020) 4:1350–6. doi: 10.1182/bloodadvances.2019001284
- 47. Ruggeri L, Mancusi A, Capanni M, Urbani E, Carotti A, Aloisi T, et al. Donor natural killer cell allorecognition of missing self in haploidentical hematopoietic transplantation for acute myeloid leukemia: challenging its predictive value. *Blood.* (2007) 110:433–40. doi: 10.1182/blood-2006-07-038687
- Huang XJ, Zhao XY, Liu DH, Liu KY, Xu LP. Deleterious effects of KIR ligand incompatibility on clinical outcomes in haploidentical hematopoietic stem cell transplantation without *in vitro* T-cell depletion. *Leukemia*. (2007) 21:848–51. doi: 10.1038/sj.leu.2404566
- Zhao XY, Huang XJ, Liu KY, Xu LP, Liu DH. Prognosis after unmanipulated HLA-haploidentical blood and marrow transplantation is correlated to the numbers of KIR ligands in recipients. *Eur J Haematol.* (2007) 78:338– 46. doi: 10.1111/j.1600-0609.2007.00822.x
- Michaelis SU, Mezger M, Bornhauser M, Trenschel R, Stuhler G, Federmann B, et al. KIR haplotype B donors but not KIR-ligand mismatch result in a reduced incidence of relapse after haploidentical transplantation using reduced intensity conditioning and CD3/CD19-depleted grafts. *Ann Hematol.* (2014) 93:1579–86. doi: 10.1007/s00277-014-2084-2
- Mancusi A, Ruggeri L, Urbani E, Pierini A, Massei MS, Carotti A, et al. Haploidentical hematopoietic transplantation from KIR ligand-mismatched donors with activating KIRs reduces nonrelapse mortality. *Blood.* (2015) 125:3173–82. doi: 10.1182/blood-2014-09-599993
- 52. Yahng SA, Jeon YW, Yoon JH, Shin SH, Lee SE, Cho BS, et al. Negative impact of unidirectional host-versus-graft killer cell immunoglobulin-like receptor ligand mismatch on transplantation outcomes after unmanipulated haploidentical peripheral blood stem cell transplantation for acute myeloid leukemia. *Biol Blood Marrow Transplant.* (2016) 22:316–23. doi: 10.1016/j.bbmt.2015.09.018
- Zhao XY, Luo XY, Yu XX, Zhao XS, Han TT, Chang YJ, et al. Recipient-donor KIR ligand matching prevents CMV reactivation post-haploidentical T cell-replete transplantation. Br J Haematol. (2017) 177:766–81. doi: 10.1111/bjh.14622
- 54. Wanquet A, Bramanti S, Harbi S, Furst S, Legrand F, Faucher C, et al. Killer cell immunoglobulin-like receptor-ligand mismatch in donor versus

- recipient direction provides better graft-versus-tumor effect in patients with hematologic malignancies undergoing allogeneic t cell-replete haploidentical transplantation followed by post-transplant cyclophosphamide. *Biol Blood Marrow Transplant*. (2018) 24:549–54. doi: 10.1016/j.bbmt.2017.11.042
- Shimoni A, Labopin M, Lorentino F, Van Lint MT, Koc Y, Gulbas Z, et al. Killer cell immunoglobulin-like receptor ligand mismatching and outcome after haploidentical transplantation with post-transplant cyclophosphamide. *Leukemia*. (2019) 33:230–9. doi: 10.1038/s41375-018-0170-5
- Cook MA, Milligan DW, Fegan CD, Darbyshire PJ, Mahendra P, Craddock CF, et al. The impact of donor KIR and patient HLA-C genotypes on outcome following HLA-identical sibling hematopoietic stem cell transplantation for myeloid leukemia. *Blood.* (2004) 103:1521– 6. doi: 10.1182/blood-2003-02-0438
- Verheyden S, Schots R, Duquet W, Demanet C. A defined donor activating natural killer cell receptor genotype protects against leukemic relapse after related HLA-identical hematopoietic stem cell transplantation. *Leukemia*. (2005) 19:1446–51. doi: 10.1038/sj.leu.2403839
- Hsu KC, Gooley T, Malkki M, Pinto-Agnello C, Dupont B, Bignon JD, et al. KIR ligands and prediction of relapse after unrelated donor hematopoietic cell transplantation for hematologic malignancy. *Biol Blood Marrow Transplant*. (2006) 12:828–36. doi: 10.1016/j.bbmt.2006.04.008
- 59. Clausen J, Wolf D, Petzer AL, Gunsilius E, Schumacher P, Kircher B, et al. Impact of natural killer cell dose and donor killer-cell immunoglobulin-like receptor (KIR) genotype on outcome following human leucocyte antigenidentical haematopoietic stem cell transplantation. *Clin Exp Immunol.* (2007) 148:520–8. doi: 10.1111/j.1365-2249.2007.03360.x
- Ludajic K, Balavarca Y, Bickeboller H, Rosenmayr A, Fae I, Fischer GF, et al. KIR genes and KIR ligands affect occurrence of acute GVHD after unrelated, 12/12 HLA matched, hematopoietic stem cell transplantation. *Bone Marrow Transplant*. (2009) 44:97–103. doi: 10.1038/bmt.2008.432
- Linn YC, Phang CY, Lim TJ, Chong SF, Heng KK, Lee JJ, et al. Effect of missing killer-immunoglobulin-like receptor ligand in recipients undergoing HLA full matched, non-T-depleted sibling donor transplantation: a single institution experience of 151 Asian patients. *Bone Marrow Transplant*. (2010) 45:1031–7. doi: 10.1038/bmt.2009.303
- 62. Wu X, He J, Wu D, Bao X, Qiu Q, Yuan X, et al. KIR and HLA-Cw genotypes of donor-recipient pairs influence the rate of CMV reactivation following non-T-cell deleted unrelated donor hematopoietic cell transplantation. *Am J Hematol.* (2009) 84:776–7. doi: 10.1002/ajh.21527
- 63. Gagne K, Busson M, Bignon JD, Balere-Appert ML, Loiseau P, Dormoy A, et al. Donor KIR3DL1/3DS1 gene and recipient Bw4 KIR ligand as prognostic markers for outcome in unrelated hematopoietic stem cell transplantation. Biol Blood Marrow Transplant. (2009) 15:1366–75. doi: 10.1016/j.bbmt.2009.06.015
- 64. Clausen J, Kircher B, Auberger J, Schumacher P, Ulmer H, Hetzenauer G, et al. The role of missing killer cell immunoglobulin-like receptor ligands in T cell replete peripheral blood stem cell transplantation from HLA-identical siblings. *Biol Blood Marrow Transplant.* (2010) 16:273–80. doi: 10.1016/j.bbmt.2009.10.021
- 65. Bjorklund AT, Schaffer M, Fauriat C, Ringden O, Remberger M, Hammarstedt C, et al. NK cells expressing inhibitory KIR for non-self-ligands remain tolerant in HLA-matched sibling stem cell transplantation. *Blood.* (2010) 115:2686–94. doi: 10.1182/blood-2009-07-229740
- 66. Wu GQ, Zhao YM, Lai XY, Luo Y, Tan YM, Shi JM, et al. The beneficial impact of missing KIR ligands and absence of donor KIR2DS3 gene on outcome following unrelated hematopoietic SCT for myeloid leukemia in the Chinese population. *Bone Marrow Transplant.* (2010) 45:1514–21. doi: 10.1038/bmt.2010.3
- 67. Zhou H, Bao X, Wu X, Tang X, Wang M, Wu D, et al. Donor selection for KIR B haplotype of the centromeric motifs can improve the outcome after HLAidentical sibling hematopoietic stem cell transplantation. *Biol Blood Marrow Transplant*. (2013) doi: 10.1016/j.bbmt.2013.10.017
- Sobecks RM, Wang T, Askar M, Gallagher MM, Haagenson M, Spellman S, et al. Impact of KIR and HLA genotypes on outcomes after reducedintensity conditioning hematopoietic cell transplantation. *Biol Blood Marrow Transplant*. (2015) 21:1589–96. doi: 10.1016/j.bbmt.2015.05.002
- 69. Park S, Kim K, Jang JH, Kim SJ, Kim WS, Kang ES, et al. KIR alloreactivity based on the receptor-ligand model is associated with improved

clinical outcomes of allogeneic hematopoietic stem cell transplantation: Result of single center prospective study. *Hum Immunol.* (2015) 76:636–43. doi: 10.1016/j.humimm.2015.09.009

- Cardozo DM, Marangon AV, da Silva RF, Aranha FJP, Visentainer JEL, Bonon SHA, et al. Synergistic effect of KIR ligands missing and cytomegalovirus reactivation in improving outcomes of haematopoietic stem cell transplantation from HLA-matched sibling donor for treatment of myeloid malignancies. *Hum Immunol.* (2016) 77:861–8. doi: 10.1016/j.humimm.2016.07.003
- Faridi RM, Kemp TJ, Dharmani-Khan P, Lewis V, Tripathi G, Rajalingam R, et al. Donor-recipient matching for KIR genotypes reduces chronic GVHD and missing inhibitory KIR ligands protect against relapse after myeloablative, HLA matched hematopoietic cell transplantation. *PLoS ONE*. (2016) 11:e0158242. doi: 10.1371/journal.pone.0158242
- Neuchel C, Furst D, Niederwieser D, Bunjes D, Tsamadou C, Wulf G, et al. Impact of donor activating KIR genes on HSCT outcome in C1-ligand negative myeloid disease patients transplanted with unrelated donors-a retrospective study. *PLoS ONE*. (2017) 12:e0169512. doi: 10.1371/journal.pone.0169512
- Arima N, Kanda J, Tanaka J, Yabe T, Morishima Y, Kim SW, et al. Homozygous HLA-C1 is associated with reduced risk of relapse after HLA-matched transplantation in patients with myeloid leukemia. *Biol Blood Marrow Transplant*. (2018) 24:717–25. doi: 10.1016/j.bbmt.2017.11.029
- Gaafar A, Sheereen A, Almohareb F, Eldali A, Chaudhri N, Mohamed SY, et al. Prognostic role of KIR genes and HLA-C after hematopoietic stem cell transplantation in a patient cohort with acute myeloid leukemia from a consanguineous community. *Bone Marrow Transplant.* (2018) 53:1170–9. doi: 10.1038/s41409-018-0123-7
- Arima N, Kanda J, Yabe T, Morishima Y, Tanaka J, Kako S, et al. Increased relapse risk of acute lymphoid leukemia in homozygous HLA-C1 patients after HLA-matched allogeneic transplantation: a japanese national registry study. *Biol Blood Marrow Transplant*. (2020) 26:431–7. doi: 10.1016/j.bbmt.2019.10.032
- 76. Chen DF, Prasad VK, Broadwater G, Reinsmoen NL, DeOliveira A, Clark A, et al. Differential impact of inhibitory and activating Killer Ig-Like Receptors (KIR) on high-risk patients with myeloid and lymphoid malignancies undergoing reduced intensity transplantation from haploidentical related donors. Bone Marrow Transplant. (2012) 47:817–23. doi: 10.1038/bmt.2011.181
- 77. Zhao XY, Chang YJ, Xu LP, Zhang XH, Liu KY, Li D, et al. HLA and KIR genotyping correlates with relapse after T-cell-replete haploidentical transplantation in chronic myeloid leukaemia patients. *Br J Cancer*. (2014) 111:1080–8. doi: 10.1038/bjc.2014.423
- Zhao XY, Chang YJ, Zhao XS, Xu LP, Zhang XH, Liu KY, et al. Recipient expression of ligands for donor inhibitory KIRs enhances NK-cell function to control leukemic relapse after haploidentical transplantation. *Eur J Immunol*. (2015) 45:2396–408. doi: 10.1002/eji.201445057
- Solomon SR, Aubrey MT, Zhang X, Piluso A, Freed BM, Brown S, et al. Selecting the best donor for haploidentical transplant: impact of HLA, killer cell immunoglobulin-like receptor genotyping, and other clinical variables. *Biol Blood Marrow Transplant*. (2018) 24:789–98. doi: 10.1016/j.bbmt.2018.01.013
- 80. Willem C, Makanga DR, Guillaume T, Maniangou B, Legrand N, Gagne K, et al. Impact of KIR/HLA incompatibilities on NK cell reconstitution and clinical outcome after T cell-replete haploidentical hematopoietic stem cell transplantation with posttransplant cyclophosphamide. *J Immunol.* (2019) 202:2141–52. doi: 10.4049/jimmunol.1801489
- 81. Chen C, Busson M, Rocha V, Appert ML, Lepage V, Dulphy N, et al. Activating KIR genes are associated with CMV reactivation and survival after non-T-cell depleted HLA-identical sibling bone marrow transplantation for malignant disorders. *Bone Marrow Transplant.* (2006) 38:437–44. doi: 10.1038/sj.bmt.1705468
- 82. Schellekens J, Rozemuller EH, Petersen EJ, van den Tweel JG, Verdonck LF, Tilanus MG. Activating KIRs exert a crucial role on relapse and overall survival after HLA-identical sibling transplantation. *Mol Immunol.* (2008) 45:2255–61. doi: 10.1016/j.molimm.2007.11.014
- 83. van der Meer A, Schaap NP, Schattenberg AV, van Cranenbroek B, Tijssen HJ, Joosten I. KIR2DS5 is associated with leukemia free survival after HLA

- identical stem cell transplantation in chronic myeloid leukemia patients. *Mol Immunol.* (2008) 45:3631–8. doi: 10.1016/j.molimm.2008.04.016
- 84. Zaia JA, Sun JY, Gallez-Hawkins GM, Thao L, Oki A, Lacey SF, et al. The effect of single and combined activating killer immunoglobulin-like receptor genotypes on cytomegalovirus infection and immunity after hematopoietic cell transplantation. *Biol Blood Marrow Transplant.* (2009) 15:315–25. doi: 10.1016/j.bbmt.2008.11.030
- Bao XJ, Hou LH, Sun AN, Qiu QC, Yuan XN, Chen MH, et al. The impact of KIR2DS4 alleles and the expression of KIR in the development of acute GVHD after unrelated allogeneic hematopoietic SCT. Bone Marrow Transplant. (2010) 45:1435–41. doi: 10.1038/bmt.2009.357
- 86. Venstrom JM, Gooley TA, Spellman S, Pring J, Malkki M, Dupont B, et al. Donor activating KIR3DS1 is associated with decreased acute GVHD in unrelated allogeneic hematopoietic stem cell transplantation. *Blood.* (2010) 115:3162–5. doi: 10.1182/blood-2009-08-236943
- Tomblyn M, Young JA, Haagenson MD, Klein JP, Trachtenberg EA, Storek J, et al. Decreased infections in recipients of unrelated donor hematopoietic cell transplantation from donors with an activating KIR genotype. *Biol Blood Marrow Transplant*. (2010) 16:1155–61. doi: 10.1016/j.bbmt.2010.02.024
- Cooley S, Weisdorf DJ, Guethlein LA, Klein JP, Wang T, Le CT, et al. Donor selection for natural killer cell receptor genes leads to superior survival after unrelated transplantation for acute myelogenous leukemia. *Blood.* (2010) 116:2411–9. doi: 10.1182/blood-2010-05-283051
- Venstrom JM, Pittari G, Gooley TA, Chewning JH, Spellman S, Haagenson M, et al. HLA-C-dependent prevention of leukemia relapse by donor activating KIR2DS1. N Engl J Med. (2012) 367:805–16. doi: 10.1056/NEJMoa1200503
- Impola U, Turpeinen H, Alakulppi N, Linjama T, Volin L, Niittyvuopio R, et al. Donor haplotype B of NK KIR receptor reduces the relapse risk in HLAidentical sibling hematopoietic stem cell transplantation of AML patients. Front Immunol. (2014) 5:405. doi: 10.3389/fimmu.2014.00405
- Bao X, Wang M, Zhou H, Zhang H, Wu X, Yuan X, et al. Donor killer immunoglobulin-like receptor profile bx1 imparts a negative effect and centromeric b-specific gene motifs render a positive effect on standardrisk acute myeloid leukemia/myelodysplastic syndrome patient survival after unrelated donor hematopoietic stem cell transplantation. *Biol Blood Marrow Transplant.* (2016) 22:232–9. doi: 10.1016/j.bbmt.2015.09.007
- Bachanova V, Weisdorf DJ, Wang T, Marsh SGE, Trachtenberg E, Haagenson MD, et al. Donor KIR B genotype improves progressionfree survival of non-hodgkin lymphoma patients receiving unrelated donor transplantation. *Biol Blood Marrow Transplant*. (2016) 22:1602– 7. doi: 10.1016/j.bbmt.2016.05.016
- Burek Kamenaric M, Stingl Jankovic K, Grubic Z, Serventi Seiwerth R, Maskalan M, Nemet D, et al. The impact of KIR2DS4 gene on clinical outcome after hematopoietic stem cell transplantation. *Hum Immunol*. (2017) 78:95–102. doi: 10.1016/j.humimm.2016.11.010
- 94. Hosokai R, Masuko M, Shibasaki Y, Saitoh A, Furukawa T, Imai C. Donor killer immunoglobulin-like receptor haplotype B/x induces severe acute graft-versus-host disease in the presence of human leukocyte antigen mismatch in T cell-replete hematopoietic cell transplantation. Biol Blood Marrow Transplant. (2017) 23:606–11. doi: 10.1016/j.bbmt.2016.12.638
- Sahin U, Dalva K, Gungor F, Ustun C, Beksac M. Donor-recipient killer immunoglobulin like receptor (KIR) genotype matching has a protective effect on chronic graft versus host disease and relapse incidence following HLA-identical sibling hematopoietic stem cell transplantation. *Ann Hematol.* (2018) 97:1027–39. doi: 10.1007/s00277-018-3274-0
- Heatley SL, Mullighan CG, Doherty K, Danner S, O'Connor GM, Hahn U, et al. Activating KIR haplotype influences clinical outcome following HLA-matched sibling hematopoietic stem cell transplantation. *HLA*. (2018) 92:74–82. doi: 10.1111/tan.13327
- 97. Babor F, Peters C, Manser AR, Glogova E, Sauer M, Potschger U, et al. Presence of centromeric but absence of telomeric group B KIR haplotypes in stem cell donors improve leukaemia control after HSCT for childhood ALL. Bone Marrow Transplant. (2019) 54:1847–58. doi: 10.1038/s41409-019-0543-z
- 98. Tordai A, Bors A, Kiss KP, Balassa K, Andrikovics H, Batai A, et al. Donor KIR2DS1 reduces the risk of transplant related mortality in HLA-C2 positive young recipients with hematological

malignancies treated by myeloablative conditioning. PLoS ONE. (2019) 14:e0218945. doi: 10.1371/journal.pone.0218945

- Nakamura R, Gendzekhadze K, Palmer J, Tsai NC, Mokhtari S, Forman SJ, et al. Influence of donor KIR genotypes on reduced relapse risk in acute myelogenous leukemia after hematopoietic stem cell transplantation in patients with CMV reactivation. *Leuk Res.* (2019) 87:106230. doi: 10.1016/j.leukres.2019.106230
- 100. Bultitude WP, Schellekens J, Szydlo RM, Anthias C, Cooley SA, Miller JS, et al. Presence of donor-encoded centromeric KIR B content increases the risk of infectious mortality in recipients of myeloablative, T-cell deplete, HLA-matched HCT to treat AML. Bone Marrow Transplant. (2020) doi: 10.1038/s41409-020-0858-9. [Epub ahead of print].
- 101. Weisdorf D, Cooley S, Wang T, Trachtenberg E, Vierra-Green C, Spellman S, et al. KIR B donors improve the outcome for AML patients given reduced intensity conditioning and unrelated donor transplantation. *Blood Adv.* (2020) 4:740–54. doi: 10.1182/bloodadvances.2019001053
- 102. Symons HJ, Leffell MS, Rossiter ND, Zahurak M, Jones RJ, Fuchs EJ. Improved survival with inhibitory killer immunoglobulin receptor (KIR) gene mismatches and KIR haplotype B donors after nonmyeloablative, HLA-haploidentical bone marrow transplantation. *Biol Blood Marrow Transplant*. (2010) 16:533–42. doi: 10.1016/j.bbmt.2009.11.022
- 103. Oevermann L, Michaelis SU, Mezger M, Lang P, Toporski J, Bertaina A, et al. KIR B haplotype donors confer a reduced risk for relapse after haploidentical transplantation in children with ALL. *Blood.* (2014) 124:2744–7. doi: 10.1182/blood-2014-03-565069
- 104. Perez-Martinez A, Ferreras C, Pascual A, Gonzalez-Vicent M, Alonso L, Badell I, et al. Haploidentical transplantation in high-risk pediatric leukemia: A retrospective comparative analysis on behalf of the Spanish working Group for bone marrow transplantation in children (GETMON) and the Spanish Grupo for hematopoietic transplantation (GETH). Am J Hematol. (2020) 95:28–37. doi: 10.1002/ajh.25661
- 105. Yu H, Tian Y, Wang Y, Mineishi S, Zhang Y. Dendritic cell regulation of graft-vs.-host disease: immunostimulation and tolerance. Front Immunol. (2019) 10:93. doi: 10.3389/fimmu.2019.00093
- Horowitz MM, Gale RP, Sondel PM, Goldman JM, Kersey J, Kolb HJ, et al. Graft-versus-leukemia reactions after bone marrow transplantation. *Blood.* (1990) 75:555-62. doi: 10.1182/blood.V75.3.555.bloodjournal7 53555
- 107. Marmont AM, Horowitz MM, Gale RP, Sobocinski K, Ash RC, van Bekkum DW, et al. T-cell depletion of HLA-identical transplants in leukemia. *Blood.* (1991) 78:2120–30. doi: 10.1182/blood.V78.8.2120.bloodjournal7882120
- 108. Champlin RE, Passweg JR, Zhang MJ, Rowlings PA, Pelz CJ, Atkinson KA, et al. T-cell depletion of bone marrow transplants for leukemia from donors other than HLA-identical siblings: advantage of T-cell antibodies with narrow specificities. *Blood.* (2000) 95:3996–4003.
- Murphy WJ, Bennett M, Kumar V, Longo DL. Donor-type activated natural killer cells promote marrow engraftment and B cell development during allogeneic bone marrow transplantation. *J Immunol.* (1992) 148:2953–60.
- 110. Huber CM, Doisne JM, Colucci F. IL-12/15/18-preactivated NK cells suppress GvHD in a mouse model of mismatched hematopoietic cell transplantation. Eur J Immunol. (2015) 45:1727–35. doi: 10.1002/eji.201445200
- 111. Xun C, Brown SA, Jennings CD, Henslee-Downey PJ, Thompson JS. Acute graft-versus-host-like disease induced by transplantation of human activated natural killer cells into SCID mice. *Transplantation*. (1993) 56:409– 17. doi: 10.1097/00007890-199308000-00031
- 112. Xun CQ, Thompson JS, Jennings CD, Brown SA. The effect of human IL-2-activated natural killer and T cells on graft-versus-host disease and graft-versus-leukemia in SCID mice bearing human leukemic cells. *Transplantation*. (1995) 60:821–7. doi: 10.1097/00007890-199510270-00011
- Mowat AM. Antibodies to IFN-gamma prevent immunologically mediated intestinal damage in murine graft-versus-host reaction. *Immunology*. (1989) 68:18–23.
- 114. MacDonald GC, Gartner JG. Prevention of acute lethal graft-versus-host disease in F1 hybrid mice by pretreatment of the graft with anti-NK-1.1 and complement. *Transplantation*. (1992) 54:147–51. doi: 10.1097/00007890-199207000-00026

- Passweg JR, Tichelli A, Meyer-Monard S, Heim D, Stern M, Kuhne T, et al. Purified donor NK-lymphocyte infusion to consolidate engraftment after haploidentical stem cell transplantation. *Leukemia*. (2004) 18:1835– 8. doi: 10.1038/sj.leu.2403524
- 116. Rizzieri DA, Storms R, Chen DF, Long G, Yang Y, Nikcevich DA, et al. Natural killer cell-enriched donor lymphocyte infusions from A 3-6/6 HLA matched family member following nonmyeloablative allogeneic stem cell transplantation. *Biol Blood Marrow Transplant.* (2010) 16:1107–14. doi: 10.1016/j.bbmt.2010.02.018
- 117. Introna M, Borleri G, Conti E, Franceschetti M, Barbui AM, Broady R, et al. Repeated infusions of donor-derived cytokine-induced killer cells in patients relapsing after allogeneic stem cell transplantation: a phase I study. *Haematologica*. (2007) 92:952–9. doi: 10.3324/haematol.11132
- 118. Yoon SR, Lee YS, Yang SH, Ahn KH, Lee JH, Lee JH, et al. Generation of donor natural killer cells from CD34(+) progenitor cells and subsequent infusion after HLA-mismatched allogeneic hematopoietic cell transplantation: a feasibility study. *Bone Marrow Transplant.* (2010) 45:1038–46. doi: 10.1038/bmt.2009.304
- 119. Ciurea SO, Schafer JR, Bassett R, Denman CJ, Cao K, Willis D, et al. Phase 1 clinical trial using mbIL21 ex vivo-expanded donor-derived NK cells after haploidentical transplantation. Blood. (2017) 130:1857– 68. doi: 10.1182/blood-2017-05-785659
- Bjorklund AT, Carlsten M, Sohlberg E, Liu LL, Clancy T, Karimi M, et al. Complete Remission with Reduction of High-Risk Clones following Haploidentical NK-Cell Therapy against MDS and AML. Clin Cancer Res. (2018) 24:1834–44. doi: 10.1158/1078-0432.CCR-17-3196
- 121. Vela M, Corral D, Carrasco P, Fernandez L, Valentin J, Gonzalez B, et al. Haploidentical IL-15/41BBL activated and expanded natural killer cell infusion therapy after salvage chemotherapy in children with relapsed and refractory leukemia. Cancer Lett. (2018) 422:107–17. doi: 10.1016/j.canlet.2018.02.033
- 122. Jaiswal SR, Zaman S, Nedunchezhian M, Chakrabarti A, Bhakuni P, Ahmed M, et al. CD56-enriched donor cell infusion after post-transplantation cyclophosphamide for haploidentical transplantation of advanced myeloid malignancies is associated with prompt reconstitution of mature natural killer cells and regulatory T cells with reduced incidence of acute graft versus host disease: A pilot study. Cytotherapy. (2017) 19:531–42. doi: 10.1016/j.jcyt.2016.12.006
- 123. Shah NN, Baird K, Delbrook CP, Fleisher TA, Kohler ME, Rampertaap S, et al. Acute GVHD in patients receiving IL-15/4-1BBL activated NK cells following T-cell-depleted stem cell transplantation. *Blood.* (2015) 125:784–92. doi: 10.1182/blood-2014-07-592881
- 124. Shilling HG, McQueen KL, Cheng NW, Shizuru JA, Negrin RS, Parham P. Reconstitution of NK cell receptor repertoire following HLAmatched hematopoietic cell transplantation. *Blood.* (2003) 101:3730– 40. doi: 10.1182/blood-2002-08-2568
- 125. Vago L, Forno B, Sormani MP, Crocchiolo R, Zino E, Di Terlizzi S, et al. Temporal, quantitative, and functional characteristics of single-KIR-positive alloreactive natural killer cell recovery account for impaired graft-versus-leukemia activity after haploidentical hematopoietic stem cell transplantation. *Blood.* (2008) 112:3488–99. doi: 10.1182/blood-2007-07-103325
- 126. Fallen PR, McGreavey L, Madrigal JA, Potter M, Ethell M, Prentice HG, et al. Factors affecting reconstitution of the T cell compartment in allogeneic haematopoietic cell transplant recipients. *Bone Marrow Transplant.* (2003) 32:1001–14. doi: 10.1038/sj.bmt.1704235
- 127. Ciurea SO, Mulanovich V, Saliba RM, Bayraktar UD, Jiang Y, Bassett R, et al. Improved early outcomes using a T cell replete graft compared with T cell depleted haploidentical hematopoietic stem cell transplantation. *Biol Blood Marrow Transplant*. (2012) 18:1835–44. doi: 10.1016/j.bbmt.2012.07.003
- 128. Eissens DN, Schaap NP, Preijers FW, Dolstra H, van Cranenbroek B, Schattenberg AV, et al. CD3+/CD19+-depleted grafts in HLA-matched allogeneic peripheral blood stem cell transplantation lead to early NK cell cytolytic responses and reduced inhibitory activity of NKG2A. *Leukemia*. (2010) 24:583–91. doi: 10.1038/leu.2009.269
- 129. Pfeiffer MM, Feuchtinger T, Teltschik HM, Schumm M, Muller I, Handgretinger R, et al. Reconstitution of natural killer cell receptors influences natural killer activity and relapse rate after haploidentical

transplantation of T- and B-cell depleted grafts in children. *Haematologica*. (2010) 95:1381–8. doi: 10.3324/haematol.2009.021121

- 130. Cooley S, McCullar V, Wangen R, Bergemann TL, Spellman S, Weisdorf DJ, et al. KIR reconstitution is altered by T cells in the graft and correlates with clinical outcomes after unrelated donor transplantation. *Blood.* (2005) 106:4370–6. doi: 10.1182/blood-2005-04-1644
- 131. Bjorkstrom NK, Beziat V, Cichocki F, Liu LL, Levine J, Larsson S, et al. CD8 T cells express randomly selected KIRs with distinct specificities compared with NK cells. *Blood*. (2012) 120:3455–65. doi: 10.1182/blood-2012-03-416867
- van Bergen J, Thompson A, van der Slik A, Ottenhoff TH, Gussekloo J, Koning F. Phenotypic and functional characterization of CD4T cells expressing killer Ig-like receptors. *J Immunol.* (2004) 173:6719–26. doi: 10.4049/iimmunol.173.11.6719
- 133. Lafarge X, Pitard V, Ravet S, Roumanes D, Halary F, Dromer C, et al. Expression of MHC class I receptors confers functional intraclonal heterogeneity to a reactive expansion of gammadelta T cells. Eur J Immunol. (2005) 35:1896–905. doi: 10.1002/eji.200425837
- 134. Pradier A, Papaserafeim M, Li N, Rietveld A, Kaestel C, Gruaz L, et al. Small-molecule immunosuppressive drugs and therapeutic immunoglobulins differentially inhibit NK cell effector functions in vitro. Front Immunol. (2019) 10:556. doi: 10.3389/fimmu.2019.00556
- Schmidt S, Schubert R, Demir A, Lehrnbecher T. Distinct effects of immunosuppressive drugs on the anti-aspergillus activity of human natural killer cells. *Pathogens*. (2019) 8:4. doi: 10.3390/pathogens8040246
- 136. Sivori S, Carlomagno S, Falco M, Romeo E, Moretta L, Moretta A. Natural killer cells expressing the KIR2DS1-activating receptor efficiently kill T-cell blasts and dendritic cells: implications in haploidentical HSCT. *Blood.* (2011) 117:4284–92. doi: 10.1182/blood-2010-10-316125
- Wojtowicz A, Bochud PY. Risk stratification and immunogenetic risk for infections following stem cell transplantation. *Virulence*. (2016) 7:917– 29. doi: 10.1080/21505594.2016.1234566
- Kao RL, Holtan SG. Host and graft factors impacting infection risk in hematopoietic cell transplantation. *Infect Dis Clin North Am.* (2019) 33:311– 29. doi: 10.1016/j.idc.2019.02.001
- 139. Cook M, Briggs D, Craddock C, Mahendra P, Milligan D, Fegan C, et al. Donor KIR genotype has a major influence on the rate of cytomegalovirus reactivation following T-cell replete stem cell transplantation. *Blood.* (2006) 107:1230–2. doi: 10.1182/blood-2005-03-1039
- 140. Gallez-Hawkins GM, Franck AE, Li X, Thao L, Oki A, Gendzekhadze K, et al. Expression of activating KIR2DS2 and KIR2DS4 genes after hematopoietic cell transplantation: relevance to cytomegalovirus infection. *Biol Blood Marrow Transplant*. (2011) 17:1662–72. doi: 10.1016/j.bbmt.2011.04.008
- 141. Lister J, Rybka WB, Donnenberg AD, deMagalhaes-Silverman M, Pincus SM, Bloom EJ, et al. Autologous peripheral blood stem cell transplantation and adoptive immunotherapy with activated natural killer cells in the immediate posttransplant period. Clin Cancer Res. (1995) 1:607–14. PubMed PMID: 9816022.
- 142. Burns LJ, Weisdorf DJ, DeFor TE, Repka TL, Ogle KM, Hummer C, et al. Enhancement of the anti-tumor activity of a peripheral blood progenitor cell graft by mobilization with interleukin 2 plus granulocyte colony-stimulating factor in patients with advanced breast cancer. *Exp Hematol.* (2000) 28:96– 103. doi: 10.1016/S0301-472X(99)00129-0
- 143. Burns LJ, Weisdorf DJ, DeFor TE, Vesole DH, Repka TL, Blazar BR, et al. IL-2-based immunotherapy after autologous transplantation for lymphoma and breast cancer induces immune activation and cytokine release: a phase I/II trial. Bone Marrow Transplant. (2003) 32:177–86. doi: 10.1038/sj.bmt.1704086
- 144. Ishikawa E, Tsuboi K, Saijo K, Harada H, Takano S, Nose T, et al. Autologous natural killer cell therapy for human recurrent malignant glioma. *Anticancer Res.* (2004) 24:1861–71.
- 145. Krause SW, Gastpar R, Andreesen R, Gross C, Ullrich H, Thonigs G, et al. Treatment of colon and lung cancer patients with *ex vivo* heat shock protein 70-peptide-activated, autologous natural killer cells: a clinical phase i trial. *Clin Cancer Res.* (2004) 10:3699–707. doi: 10.1158/1078-0432.CCR-0 3-0683
- 146. Krieger E, Sabo R, Moezzi S, Cain C, Roberts C, Kimball P, et al. Killer immunoglobulin-like receptor-ligand interactions predict clinical outcomes

- following unrelated donor transplantations. *Biol Blood Marrow Transplant*. (2020) 26:672–82. doi: 10.1016/j.bbmt.2019.10.016
- 147. Montaldo E, Del Zotto G, Della Chiesa M, Mingari MC, Moretta A, De Maria A, et al. Human NK cell receptors/markers: a tool to analyze NK cell development, subsets and function. Cytometry A. (2013) 83:702–13. doi: 10.1002/cyto.a.22302
- Scoville SD, Freud AG, Caligiuri MA. Modeling human natural killer cell development in the era of innate lymphoid cells. Front Immunol. (2017) 8:360. doi: 10.3389/fimmu.2017.00360
- 149. Cooper MA, Fehniger TA, Caligiuri MA. The biology of human natural killer-cell subsets. *Trends Immunol.* (2001) 22:633–40. doi: 10.1016/S1471-4906(01)02060-9
- 150. Bjorkstrom NK, Riese P, Heuts F, Andersson S, Fauriat C, Ivarsson MA, et al. Expression patterns of NKG2A, KIR, and CD57 define a process of CD56dim NK-cell differentiation uncoupled from NK-cell education. *Blood.* (2010) 116:3853–64. doi: 10.1182/blood-2010-04-281675
- Abel AM, Yang C, Thakar MS, Malarkannan S. Natural killer cells: development, maturation, and clinical utilization. Front Immunol. (2018) 9:1869. doi: 10.3389/fimmu.2018.01869
- Russo A, Oliveira G, Berglund S, Greco R, Gambacorta V, Cieri N, et al. NK cell recovery after haploidentical HSCT with posttransplant cyclophosphamide: dynamics and clinical implications. *Blood.* (2018) 131:247–62. doi: 10.1182/blood-2017-05-780668
- 153. Foley B, Cooley S, Verneris MR, Curtsinger J, Luo X, Waller EK, et al. NK cell education after allogeneic transplantation: dissociation between recovery of cytokine-producing and cytotoxic functions. *Blood.* (2011) 118:2784–92. doi: 10.1182/blood-2011-04-347070
- 154. Yu J, Venstrom JM, Liu XR, Pring J, Hasan RS, O'Reilly RJ, et al. Breaking tolerance to self, circulating natural killer cells expressing inhibitory KIR for non-self HLA exhibit effector function after T cell-depleted allogeneic hematopoietic cell transplantation. *Blood.* (2009) 113:3875– 84. doi: 10.1182/blood-2008-09-177055
- 155. Rathmann S, Glatzel S, Schonberg K, Uhrberg M, Follo M, Schulz-Huotari C, et al. Expansion of NKG2A-LIR1- natural killer cells in HLA-matched, killer cell immunoglobulin-like receptors/HLA-ligand mismatched patients following hematopoietic cell transplantation. *Biol Blood Marrow Transplant*. (2010) 16:469–81. doi: 10.1016/j.bbmt.2009.12.008
- Moretta L, Locatelli F, Pende D, Marcenaro E, Mingari MC, Moretta A. Killer Ig-like receptor-mediated control of natural killer cell alloreactivity in haploidentical hematopoietic stem cell transplantation. *Blood.* (2011) 117:764–71. doi: 10.1182/blood-2010-08-264085
- Joncker NT, Shifrin N, Delebecque F, Raulet DH. Mature natural killer cells reset their responsiveness when exposed to an altered MHC environment. J Exp Med. (2010) 207:2065–72. doi: 10.1084/jem.20100570
- 158. Elliott JM, Wahle JA, Yokoyama WM. MHC class I-deficient natural killer cells acquire a licensed phenotype after transfer into an MHC class I-sufficient environment. J Exp Med. (2010) 207:2073–9. doi: 10.1084/jem.20100986
- 159. Boudreau JE, Liu XR, Zhao Z, Zhang A, Shultz LD, Greiner DL, et al. Cell-Extrinsic MHC Class I molecule engagement augments human NK cell education programmed by cell-intrinsic MHC class I. *Immunity*. (2016) 45:280–91. doi: 10.1016/j.immuni.2016.07.005
- 160. Rogatko-Koros M, Mika-Witkowska R, Bogunia-Kubik K, Wysoczanska B, Jaskula E, Koscinska K, et al. Prediction of NK cell licensing level in selection of hematopoietic stem cell donor, initial results. Arch Immunol Ther Exp (Warsz). (2016) 64(Suppl 1):63–71. doi: 10.1007/s00005-016-0438-2
- 161. Graczyk-Pol E, Rogatko-Koros M, Nestorowicz K, Gwozdowicz S, Mika-Witkowska R, Pawliczak D, et al. Role of donor HLA class I mismatch, KIR-ligand mismatch and HLA:KIR pairings in hematological malignancy relapse after unrelated hematopoietic stem cell transplantation. HLA. (2018) 92 Suppl 2:42–6. doi: 10.1111/tan.13386
- 162. Nowak J, Gwozdowicz S, Graczyk-Pol E, Mika-Witkowska R, Rogatko-Koros M, Nestorowicz K, et al. Epstein-Barr virus infections are strongly dependent on activating and inhibitory KIR-HLA pairs after T-cell replate unrelated hematopoietic stem cell transplantation, the principles, and method of pairing analysis. HLA. (2019) 94 Suppl 2:40–8. doi: 10.1111/tan.13770
- 163. Zhao XY, Yu XX, Xu ZL, Cao XH, Huo MR, Zhao XS, et al. Donor and host coexpressing KIR ligands promote NK education after allogeneic

hematopoietic stem cell transplantation. *Blood Adv.* (2019) 3:4312–25. doi: 10.1182/bloodadvances.2019000242

- 164. Elmaagacli AH, Steckel NK, Koldehoff M, Hegerfeldt Y, Trenschel R, Ditschkowski M, et al. Early human cytomegalovirus replication after transplantation is associated with a decreased relapse risk: evidence for a putative virus-versus-leukemia effect in acute myeloid leukemia patients. *Blood.* (2011) 118:1402–12. doi: 10.1182/blood-2010-08-304121
- 165. Della Chiesa M, Falco M, Podesta M, Locatelli F, Moretta L, Frassoni F, et al. Phenotypic and functional heterogeneity of human NK cells developing after umbilical cord blood transplantation: a role for human cytomegalovirus? *Blood.* (2012) 119:399–410. doi: 10.1182/blood-2011-08-372003
- 166. Foley B, Cooley S, Verneris MR, Pitt M, Curtsinger J, Luo X, et al. Cytomegalovirus reactivation after allogeneic transplantation promotes a lasting increase in educated NKG2C+ natural killer cells with potent function. *Blood*. (2012) 119:2665–74. doi: 10.1182/blood-2011-10-386995
- 167. Horowitz A, Guethlein LA, Nemat-Gorgani N, Norman PJ, Cooley S, Miller JS, et al. Regulation of adaptive NK cells and CD8T cells by HLA-C correlates with allogeneic hematopoietic cell transplantation and with cytomegalovirus reactivation. *J Immunol.* (2015) 195:4524–36. doi: 10.4049/jimmunol.1401990
- 168. Jin F, Lin H, Gao S, Wang H, Yan H, Guo J, et al. Characterization of IFNgamma-producing natural killer cells induced by cytomegalovirus reactivation after haploidentical hematopoietic stem cell transplantation. Oncotarget. (2017) 8:51–63. doi: 10.18632/oncotarget.13916
- 169. Beziat V, Liu LL, Malmberg JA, Ivarsson MA, Sohlberg E, Bjorklund AT, et al. NK cell responses to cytomegalovirus infection lead to stable imprints in the human KIR repertoire and involve activating KIRs. *Blood*. (2013) 121:2678–88. doi: 10.1182/blood-2012-10-459545
- 170. Djaoud Z, David G, Bressollette C, Willem C, Rettman P, Gagne K, et al. Amplified NKG2C+ NK cells in cytomegalovirus (CMV) infection preferentially express killer cell Ig-like receptor 2DL: functional impact in controlling CMV-infected dendritic cells. *J Immunol.* (2013) 191:2708–16. doi: 10.4049/jimmunol.1301138
- Charoudeh HN, Terszowski G, Czaja K, Gonzalez A, Schmitter K, Stern M. Modulation of the natural killer cell KIR repertoire by cytomegalovirus infection. Eur J Immunol. (2013) 43:480–7. doi: 10.1002/eji.201242389
- 172. Della Chiesa M, Falco M, Bertaina A, Muccio L, Alicata C, Frassoni F, et al. Human cytomegalovirus infection promotes rapid maturation of NK cells expressing activating killer Ig-like receptor in patients transplanted with NKG2C-/- umbilical cord blood. *J Immunol.* (2014) 192:1471–9. doi: 10.4049/jimmunol.1302053
- 173. Muccio L, Bertaina A, Falco M, Pende D, Meazza R, Lopez-Botet M, et al. Analysis of memory-like natural killer cells in human cytomegalovirus-infected children undergoing alphabeta+T and B cell-depleted hematopoietic stem cell transplantation for hematological malignancies. Haematologica. (2016) 101:371-81. doi: 10.3324/haematol.2015.1 34155
- 174. Davis ZB, Cooley SA, Cichocki F, Felices M, Wangen R, Luo X, et al. Adaptive natural killer cell and killer cell immunoglobulin-like receptor-expressing T cell responses are induced by cytomegalovirus and are associated with protection against cytomegalovirus reactivation after allogeneic donor hematopoietic cell transplantation. *Biol Blood Marrow Transplant*. (2015) 21:1653–62. doi: 10.1016/j.bbmt.2015.05.025
- 175. Lodolce JP, Boone DL, Chai S, Swain RE, Dassopoulos T, Trettin S, et al. IL-15 receptor maintains lymphoid homeostasis by supporting lymphocyte homing and proliferation. *Immunity*. (1998) 9:669–76. doi: 10.1016/S1074-7613(00)80664-0
- Cooper MA, Bush JE, Fehniger TA, VanDeusen JB, Waite RE, Liu Y, et al. *In vivo* evidence for a dependence on interleukin 15 for survival of natural killer cells. *Blood*. (2002) 100:3633–8. doi: 10.1182/blood-2001-12-0293

- 177. Bosch M, Dhadda M, Hoegh-Petersen M, Liu Y, Hagel LM, Podgorny P, et al. Immune reconstitution after anti-thymocyte globulin-conditioned hematopoietic cell transplantation. *Cytotherapy*. (2012) 14:1258–75. doi: 10.3109/14653249.2012.715243
- 178. Servais S, Menten-Dedoyart C, Beguin Y, Seidel L, Gothot A, Daulne C, et al. Impact of pre-transplant anti-T cell globulin (ATG) on immune recovery after myeloablative allogeneic peripheral blood stem cell transplantation. PLoS ONE. (2015) 10:e0130026. doi: 10.1371/journal.pone.0130026
- 179. Retiere C, Willem C, Guillaume T, Vie H, Gautreau-Rolland L, Scotet E, et al. Impact on early outcomes and immune reconstitution of high-dose post-transplant cyclophosphamide vs. anti-thymocyte globulin after reduced intensity conditioning peripheral blood stem cell allogeneic transplantation. *Oncotarget.* (2018) 9:11451–64. doi: 10.18632/oncotarget.24328
- 180. Nguyen S, Kuentz M, Vernant JP, Dhedin N, Bories D, Debre P, et al. Involvement of mature donor T cells in the NK cell reconstitution after haploidentical hematopoietic stem-cell transplantation. *Leukemia*. (2008) 22:344–52. doi: 10.1038/sj.leu.2405041
- 181. Fehniger TA, Cooper MA, Nuovo GJ, Cella M, Facchetti F, Colonna M, et al. CD56bright natural killer cells are present in human lymph nodes and are activated by T cell-derived IL-2: a potential new link between adaptive and innate immunity. *Blood.* (2003) 101:3052–7. doi: 10.1182/blood-2002-09-2876
- 182. Ferlazzo G, Thomas D, Lin SL, Goodman K, Morandi B, Muller WA, et al. The abundant NK cells in human secondary lymphoid tissues require activation to express killer cell Ig-like receptors and become cytolytic. *J Immunol.* (2004) 172:1455–62. doi: 10.4049/jimmunol.172.3.1455
- 183. Zhao XY, Huang XJ, Liu KY, Xu LP, Liu DH. Reconstitution of natural killer cell receptor repertoires after unmanipulated HLAmismatched/haploidentical blood and marrow transplantation: analyses of CD94:NKG2A and killer immunoglobulin-like receptor expression and their associations with clinical outcome. Biol Blood Marrow Transplant. (2007) 13:734–44. doi: 10.1016/j.bbmt.2007.02.010
- 184. Bunting MD, Varelias A, Souza-Fonseca-Guimaraes F, Schuster IS, Lineburg KE, Kuns RD, et al. GVHD prevents NK-cell-dependent leukemia and virus-specific innate immunity. *Blood.* (2017) 129:630– 42. doi: 10.1182/blood-2016-08-734020
- 185. Ullrich E, Salzmann-Manrique E, Bakhtiar S, Bremm M, Gerstner S, Herrmann E, et al. Relation between Acute GVHD and NK cell subset reconstitution following allogeneic stem cell transplantation. Front Immunol. (2016) 7:595. doi: 10.3389/fimmu.2016.00595
- 186. Hu LJ, Zhao XY, Yu XX, Lv M, Han TT, Han W, et al. Quantity and quality reconstitution of NKG2A(+) natural killer cells are associated with graftversus-host disease after allogeneic hematopoietic cell transplantation. *Biol Blood Marrow Transplant.* (2019) 25:1–11. doi: 10.1016/j.bbmt.2018.08.008
- 187. Giebel S, Dziaczkowska J, Czerw T, Wojnar J, Krawczyk-Kulis M, Nowak I, et al. Sequential recovery of NK cell receptor repertoire after allogeneic hematopoietic SCT. Bone Marrow Transplant. (2010) 45:1022– 30. doi: 10.1038/bmt.2009.384

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2020 Gao, Ye, Gao, Huang and Zhao. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.





Efficacy and Safety of Eculizumab in the Treatment of Transplant-Associated Thrombotic Microangiopathy: A Systematic Review and Meta-Analysis

Rui Zhang ^{1,2,3,4†}, Meng Zhou ^{1,2,3,4,5†}, Jiaqian Qi ^{1,2,3,4,5†}, Wenjing Miao ^{1,2,3,4}, Zivan Zhang ^{1,3,5}. Depei Wu ^{1,2,3,4,5,6*} and Yue Han ^{1,2,3,4,5,6*}

OPEN ACCESS

Edited by:

Nicolaus Martin Kröger, Universität Hamburg, Germany

Reviewed by:

Michael Medinger, University Hospital of Basel, Switzerland Jun Peng, Shandong University, China

*Correspondence:

Depei Wu wudepeisz@163.com Yue Han hanyue@suda.edu.cn

[†]These authors have contributed equally to this work and share first authorship

Specialty section:

This article was submitted to Alloimmunity and Transplantation, a section of the journal Frontiers in Immunology

Received: 22 May 2020 Accepted: 30 November 2020 Published: 20 January 2021

Citation:

Zhang R, Zhou M, Qi J, Miao W, Zhang Z, Wu D and Han Y (2021) Efficacy and Safety of Eculizumab in the Treatment of Transplant-Associated Thrombotic Microangiopathy: A Systematic Review and Meta-Analysis. Front. Immunol. 11:564647. doi: 10.3389/fimmu.2020.564647 ¹ Jiangsu Institute of Hematology, The First Affiliated Hospital of Soochow University, Suzhou, China, ² Collaborative Innovation Center of Hematology, Soochow University, Suzhou, China, ³ Institute of Blood and Marrow Transplantation, Suzhou, China, ⁴ Key Laboratory of Thrombosis and Hemostasis of Ministry of Health, Suzhou, China, ⁵ National Clinical Research Center for Hematologic Diseases, Suzhou, China, ⁶ State Key Laboratory of Radiation Medicine and Protection, Soochow University, Suzhou, China

Background: Transplant-associated thrombotic microangiopathy (TA-TMA) is a dangerous and life-threatening complication in patients undergoing hematopoietic stem cell transplantation (HSCT). Eculizumab has been used in the treatment of TA-TMA, and several studies have confirmed the benefit of Eculizumab in patients with TA-TMA. However, the results remain controversial. We conducted a systematic review and meta-analysis to evaluate the efficacy and safety of Eculizumab for TA-TMA.

Materials and Methods: We searched PubMed and Embase for studies on the efficacy and safety of Eculizumab in TA-TMA patients. Efficacy outcomes consisted of overall response rate (ORR), complete response rate (CRR), and survival rate at the last follow-up (SR). Safety outcomes were adverse events (AEs), including infection, sepsis, impaired liver function, infusion reactions, and death.

Results: A total of 116 patients from six studies were subjected to meta-analysis. The pooled estimates of ORR, CRR, and SR for TA-TMA patients were 71% (95% CI: 58–82%), 32% (95% CI: 11–56%), and 52% (95% CI: 40–65%), respectively. Only one patient presented with a severe rash, and infection was the most common AEs. The main causes of death were infection and GvHD.

Conclusion: Current evidence suggests that Eculizumab improves SR and ORR in patients with TA-TMA and that Eculizumab is well tolerated. However, the number of studies is limited, and the findings are based mainly on data from observational studies. Higher quality randomized controlled trials and more extensive prospective cohort studies are needed.

Keywords: Eculizumab, terminal complement inhibitor, transplant-associated thrombotic microangiopathy, hematopoietic stem cell transplantation, efficacy, safety, meta-analysis

INTRODUCTION

Hematopoietic stem cell transplantation (HSCT) is a recognized treatment for both malignant and non-malignant diseases. While this treatment has increased cure rates and reduced disease mortality, its complications remain life-threatening and of concern. Transplant-associated thrombotic microangiopathy (TA-TMA) is one of the most devastating complications of hematopoietic stem cell transplantation. A recent study reported a 3-year cumulative incidence rate of 3% for TA-TMA, and TA-TMA was associated with high mortality (HR = 3.1, 95% CI: 2.8-16.3%) (1). Treatment intensity, use of calcitonin inhibitors (CNIs), graft-versus-host disease (GvHD), and viral infection are risk factors for TA-TMA (2, 3). Patients with TA-TMA are characterized by microangiopathic hemolytic anemia, unexplained thrombocytopenia, elevated lactate dehydrogenase (LDH), and endothelial injury-related organ failure, such as hypertension, chronic kidney disease (CKD), pulmonary hypertension, gastrointestinal or central nervous system disease (4). TA-TMA is mainly defined with two standard diagnostic criteria. One is the International Working Group (IWG) (thrombocytopenia in the blood; new-onset, prolonged or progressive thrombocytopenia; sudden and persistent elevation of LDH; decreased hemoglobin or increased transfusion requirements, decreased serum hemoglobin) (5), and the other is overall-TMA (O-TMA) as demonstrated by Cho et al. (Elevated LDH; new-onset thrombocytopenia with a platelet count $<50 \times 10^9/L$ or thrombocytopenia >50%; new-onset anemia with hemoglobin below the lower limit of normal or requiring transfusion support; the presence of typed cells in peripheral blood or histological evidence of microangiopathy in tissue specimens; no coagulation dysfunction, negative Coombs test) (6). The mechanism of how systemic microvascular endothelial injury leads to TA-TMA remains unclear. Due to its similar histomorphology to atypical hemolytic uremic syndrome (aHUS), most studies suggest that it is dysregulation of the complement system that causes TA-TMA to occur (7-9). Subsequently, C5 is cleaved to C5a and C5b, forming a cell membrane attack complex (MAC, C5b-9) on the surface of the endothelial cells, resulting in further endothelial cell damage (8). Significantly elevated plasma C3b, sC5b-9 levels were observed in TA-TMA patients (10).

There is no consensus on care strategies for TA-TMA. Conventional treatments, including supportive care measures, withdrawal of CNIs, therapeutic plasma exchange (TPE) and pharmacological treatments such as rituximab, defibrotide, and daclizumab have been used in the treatment of TA-TMA (8). Recently, the use of Eculizumab for the treatment of TA-TMA has raised concerns. With more and more case reports reaching remission (11–13), Eculizumab has shown its benefits in the treatment of patients with TA-TMA. Eculizumab is a terminal complement inhibitor that works by inhibiting the cleavage of C5 to C5a and C5b. C5b-9 is then blocked from forming on the surface of endothelial cells (14). Since the FDA approved Eculizumab for the treatment of atypical hemolytic uremic syndrome (aHUS) (15), TA-TMA patients treated with Eculizumab were treated according to the standard regimen of

aHUS (16). Patients received induction therapy with Eculizumab 900mg once a week for four weeks. When hematological signs of TA-TMA resolved, maintenance therapy was continued with 1,200 mg given every two weeks (17, 18). Some researchers have paid attention to evaluate the therapeutic benefits of Eculizumab in patients with TA-TMA. A study by Joslyn et al. showed a hematological response rate of 70% in patients with TA-TMA after Eculizumab treatment (19), similar to the 67% reported by Michelle et al. (20), but much worse compared to the 93% reported by Stephan et al. (21). In addition, the survival rate of 60% (19) studied by Joslyn et al. was similar to that of Michelle et al.'s 67% (20) and more favorable than that of Stephan et al. (33%) (21). The efficacy results vary from study to study.

To determine which factors may contribute to diversity in response rates and survival, we systematically reviewed relevant studies of Eculizumab in patients with TA-TMA and performed a meta-analysis to better understand the efficacy and safety of Eculizumab.

MATERIALS AND METHODS

Search Strategy

PubMed and Embase databases were searched from their inception up to February 15, 2020, for relevant studies, and publication language was restricted as English. The search strategy was based on the following combined MeSH terms: (((("Transplantation"[Mesh]) OR Transplantations[Title/Abstract])) AND (("Thrombotic Microangiopathies"[Mesh]) OR (((Microangiopathies, Thrombotic[Title/Abstract])) OR Microangiopathy, Thrombotic[Title/Abstract]) OR Thrombotic Microangiopathy[Title/Abstract]))) AND ((((((Eculizumab) OR Alexion)))) OR Soliris)) OR 5G1.1) OR H5G1.1VHC +H5G1.1VLC) OR H5G1.1) OR H5G1-1) OR H5G11). The systematic review and meta-analysis were conducted and reported in compliance with the PRISMA statement (22).

Selection Criteria

Studies eligible in the meta-analysis met the following criteria: (1) Patients developed TMA after hematopoietic stem cell transplantation; (2) Eculizumab was regarded as first-line therapy or second-line therapy; (3) studies are cohort studies and data from case, letter, review, conference abstract were not taken into consideration. (4) Outcomes of this meta-analysis will include complete response, overall response, survival rate, and adverse events (AEs). To minimize bias in the selected pieces of literature, each paper with a title and a general meeting, our inclusion criteria were checked by two reviewers independently. Then full texts were identified and reevaluated carefully. Any disagreements were further discussed and resolved by consulting a senior investigator to reach a consensus.

Outcome Measures

The diagnosis of TA-TMA was identified according to IWG (5) or O-TMA (6). Hematological response (HR) was defined as disappearance of schistocytes, normalization of LDH and

haptoglobin, and dependence of transfusion. Complete response (CR) was defined as hematological response with resolution of organ dysregulation caused by TMA. Among outcomes of patients undergoing Eculizumab therapy, which included hematological response (HR), complete response (CR) and no response (NR), an effective overall response (OR) was composed of CR and HR. The survival rate (SR) was evaluated at the last follow-up of each study. Adverse events were reported at baseline and a follow-up visit with a focus on meningococcal infections, serious infections, sepsis, hepatic impairment, infusion reaction, and death.

Data Extraction

We extracted general characteristics, including the surname of the first author, year of publication, setting, sample size from each included study. Pretreatment patient data collected included age, gender, primary disease, type of transplant, diagnostic criteria, the level of serum sC5b-9, time from HSCT to TA-TMA diagnosis, time from TA-TMA diagnosis to Eculizumab use. Treatment variables included median days of Eculizumab therapy, median Eculizumab dose, outcomes and prognosis.

Quality Assessment

The Newcastle-Ottawa Quality Assessment Scale for Cohort Studies was applied to evaluate the quality and risk of bias of included studies (23).

Statistical Analysis

Efficacy was evaluated by the overall response rate (ORR), complete response rate (CRR), survival rate (SR). Safety of Eculizumab was evaluated by adverse events (AEs) including treatment-emergent adverse events (TEAEs), treatment-related adverse events (TRAEs), serious adverse events (SAEs), and cause of death. All the raw data extracted from the studies were transformed with the Freeman-Tukey double arcsine method. Estimated proportions (ES) with 95% confidence intervals (CIs) were calculated for ratio outcomes. The presence of heterogeneity was assessed by using the chi-square test of heterogeneity and the I² measure of inconsistency. Higher I² value and lower P-value indicate a greater degree of heterogeneity, and I² values ≤25%, between 25 and 50%, and ≥50% were equal to low, median, and substantial heterogeneity, respectively. A random-effects model was used regardless of heterogeneity. Considering some significant factors might affect clinical response, survival and prognosis, subgroup analyses and meta-regression for the overall response rate (ORR) and survival rate (SR) were performed based on publication year, setting, sample size, age, primary disease, median days between transplant to TA-TMA, Eculizumab as first-line therapy, overall median therapy duration, the median number of Eculizumab doses if relevant data were available. The p-value of meta-regression of publication <0.05 accounted for the existence of heterogeneity. Funnel plots were inappropriate to perform as the total number of included studies were six (<10). Sensitivity analyses were conducted further to decide the stability and reliability of the results we performed by deletion of every single investigation. All statistical analyses were conducted using R (version 3.6.2). A two-tailed P value of less than 0.05 was considered statistically significant.

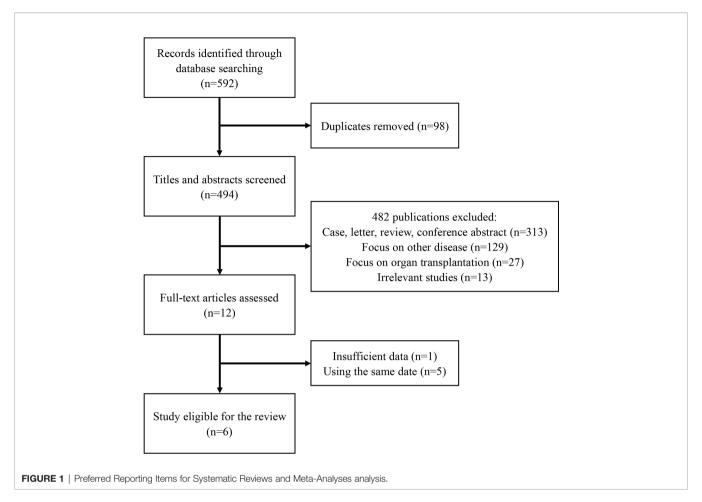
RESULTS

Data Sources

In all, 592 publications were initially identified based on literature search parameters (Figure 1). A total of 98 were discarded for duplicates, and 482 records were removed by inspecting the titles and abstracts based on prospective search criteria. After full-text evaluations of the remaining 12 articles, six were considered to be eligible for the systematic review and meta-analysis according to the selection criteria (19–21, 24–26). The necessary information of the included six articles was summarized in Table 1. These articles were published, ranging from 2015 to 2020. TA-TMA was diagnosed mainly by adopting O-TMA proposed by Cho et al. Treatment with Eculizumab was primarily administered following the recommended dose for aHUS. And for pediatric patients, the dose of Eculizumab followed the protocol of Jodele et al. Baseline characteristics of patients including age, gender, primary disease, type of transplant and Eculizumab treatment, and outcomes of efficacy and safety endpoints were described in Table 2. TA-TMA was diagnosed at a median age of 23 years (range1.2-66) posttransplantation. Most patients had transplant performed for hematological malignancy, neuroblastoma, as well as immune deficiency. Of patients with available information, 82.8% received allogeneic hematopoietic cell transplantation, 69.9% received CNI treatment at the time of diagnosis, aGvHD occurred in 39.7%, and viral infection was 26.7%.

Efficacy Outcomes

A total of six articles, including 116 patients were eligible for the analysis of overall response rate (ORR) (19-21, 24-26). It showed that the heterogeneity among the included studies was median ($I^2 = 30\%$, P = 0.21). Pooled result of ORR in TA-TMA patients treated with Eculizumab was 71% (95%CI: 58-82%) (Figure 2). Subgroup analysis and meta-regression were conducted to evaluate the potential effects of setting, sample size, age, primary disease, median days between transplant to TA-TMA, Eculizumab as first-line or second-line, therapy duration, the median number of Eculizumab doses (Table S1). Subgroup analysis of setting showed that the pooled ORR of single-center (ORR = 74%, 95%CI:57-88%) was numerically higher than that of multicenter (ORR = 63%, 95%CI:35-87%). Subgroup analysis of the number of Eculizumab dose showed that the pooled ORR of dose ≥8 was 75% (95%CI:58-89%), which is higher than that <8 (ORR = 59%, 95%CI:37-80%). The therapy duration <60 days achieved a higher survival rate (ORR = 84%, 95%CI:57-100%) than that ≥ 60 days (ORR =64%, 95%CI:54-75%). However, the p-value of meta-regression of variables were all >0.05, which did not account for the existence of heterogeneity. The application of sensitivity analysis showed that the study by Stephan et al. (21) impacted



the overall results (Figure S1), which was a historically controlled, single-center study as opposed to the other observational studies.

Among these included studies, five studies reported the complete response rate (CRR) for TA-TMA patients receiving Eculizumab treatment (19, 20, 24-26). The heterogeneity among included studies was substantial ($I^2 = 73\%$, P < 0.01) and the pooled estimate of CRR was 32% (95%CI: 11-56%), which was much lower than overall response (Figure 3). As the number of studies was limited, the source of heterogeneity could not be analyzed by meta-regression. Sensitivity analysis of CRR in TA-TMA patients treated with Eculizumab informed that Sonata et al. (26) might be the source of heterogeneity (Figure S2). The CRR of the study was 88% which was much higher than other studies, and it is an observational study consisting of 64 pediatric patients diagnosed as high-risk TA-TMA. All the patients were offered Eculizumab as first-line therapy, and the median number of Eculizumab doses given was 11, which is more than that of other studies.

The survival rate (SR) of TA-TMA patients treated with Eculizumab was analyzed in six articles (19–21, 24–26). The heterogeneity among the six included studies was low ($I^2 = 24\%$, P = 0.25). Pooled estimate of SR was 52% (95%CI: 40–65%) (**Figure 4**). Subgroup analysis and meta-regression were

conducted to evaluate the potential effects of publication year, setting, sample size, age, primary disease, median days between transplant to TA-TMA, therapy duration, the median number of Eculizumab doses (Table S2). Subgroup analysis of sample size showed that the pooled SR of the size ≤ 10 (SR = 68%, 95%CI:47– 86%) was numerically higher than that >10 (SR = 45%, 95% CI:29-61%). Subgroup analysis of age showed that the pooled SR of pediatric TA-TMA patients (SR = 59%, 95%CI:47-70%) was numerically higher than that of adult patients (SR = 40%, 95% CI:24-57%). Subgroup analysis of primary disease indicated that the pooled SR of hematological disease was 40% (95%CI:24-57%), which is lower than that of primary disease containing the hematological disease and others (SR = 59%, 95%CI:47-70%). The pooled SR of TA-TMA diagnosed during first 100 days after transplant was 59% (95%CI:48-70%), which was significantly higher than that of TA-TMA diagnosed more than 100 days after transplant (SR = 33%, 95%CI:16-53%). The therapy duration of more than 65 days achieved a higher survival rate (SR = 59%, 95%CI:47-70%) than that less than 65 days (SR = 40%, 95% CI:24-57%). P-value of median days between transplant and TA-TMA was 0.0266, which can explain the source of heterogeneity. The result of sensitivity analysis indicated that omitting the study of Prajwal et al. (25) and Stephan et al. (21) may influence the pooled results (Figure S3).

TABLE 1 | Study from which patient-level data were provided and included in meta-analysis.

Study	Year	setting	diagnostic criteria	Patients with TMA after HSCT in the study	TA-TMA patients with Eculizumab treatment in the study	Eculizumab regime
Flore et al. (24)	2015	Multicenter	O-TMA criteria	12	12	Induction therapy: 900 mg weekly for 4 weeks Maintenance therapy: 1,200 mg every 2 weeks
Prajwal et al. (25)	2016	Multicenter	O-TMA criteria	9	9	Adult patients: induction therapy was 900 mg weekly for 4 weeks, followed by 1,200 mg every 2 weeks for maintenance therapy. Pediatric patients: the first dose was based on weight, subsequent dose according to CH50.
Stephan et al. (21)	2017	Single- center	O-TMA criteria	39	15	Induction therapy: 900 mg weekly for 4 weeks Maintenance therapy: 1,200 mg every 2 weeks
Joslyn et al. (19)	2018	Single- center	O-TMA criteria	10	10	Induction therapy: 900 mg weekly for 4 weeks Maintenance therapy: 1,200 mg every 2 weeks
Michelle et al. (20)	2019	Single- center	Before 2010, IWG criteria After 2010, O-TMA criteria	9	6	Adult patients: induction therapy was 900 mg weekly for 4 weeks, followed by 1,200 mg every 2 weeks for maintenance therapy. Pediatric patients: the first dose was based on weight, subsequent dose according to CH50.
Sonata et al. (26)	2020	Single- center	O-TMA criteria	177	64	The first dose was based on weight, subsequent dose according to CH50.

CH50, a total hemolytic complement activity level; HSCT, Hematopoietic stem cell transplantation; IWG, International Working Group; O-TMA, overall-TMA; TA-TMA, Transplant associated thrombotic microangiopathy.

Safety Outcomes

All six studies were reported that treatment of TA-TMA with Eculizumab was well tolerated (19–21, 24–26). Among 116 patients, only one case was reported to get severe skin rash leading to drug discontinuation during Eculizumab therapy (19). Three studies were reported that some patients developed the infection after starting Eculizumab therapy for TA-TMA (21, 24, 26), and no meningococcal infections were reported. After the Eculizumab therapy, many survivors suffered from CKD and HTN, and much of them were still depend on dialysis (**Table 2**). As the data from the studies were limited, further studies need to analyze the prognosis.

From a total of 55 subjects who died, cause of death can be divided into four risk factors: GvHD, infection, TA-TMA related organ failure, relapse of disease, which were presented in **Figure 5**. Among these risk factors, the proportion of infection was 31% (95%CI:6–61%), which is much higher than the other three factors. And the proportion of GvHD was 26% (95%CI:2–59%). TA-TMA related death was occupied 23% (95%CI:10–38%). Death related to relapse of primary disease was the least. No study was reported that death is related to the use of Eculizumab.

Risk of Bias

The Newcastle-Ottawa scale was used to assess the risk of bias. The NOS scores of every study ranged from 7 to 9, with an average of 7.7. The detailed information of NOS scores is shown in **Table S3**.

DISCUSSION

In all six observational studies, including 116 patients, were included in a systematic review and meta-analysis to

investigate the efficacy and safety of Eculizumab in patients with TA-TMA (19-21, 24-26). After Eculizumab treatment, almost 71% of patients responded to the therapy. However, the number of patients who reached full response was significantly lower (32%). Patients with TA-TMA treated with Eculizumab had a 52% survival rate at a median follow-up of 13.5 months after HSCT. Previous treatment strategies for TA-TMA after HSCT have focused on conventional therapies, including withdrawal of CNIs, plasmapheresis, defibrillation, rituximab, and combinations of several therapies (8). A retrospective study showed that 24% of TA-TMA patients (n = 33) underwent plasmapheresis and achieved a clinical response with an SR of 45% at 100 days after diagnosis (27). Corti et al. reported a total of 12 TA-TMA patients undergoing defibrillation in two centers with an ORR and SR of 67 and 50%, respectively (28). Another study by Au et al. showed an ORR of 80% (n = 4/5) for rituximab treatment and a study SR of 60% at a median follow-up of 305 (250-440) d (29). Due to the small sample size reported above, it is difficult to perform a systematic and comprehensive comparison between conventional therapy and Eculizumab. Nonetheless, the summary ORR and SR of Eculizumab from our meta-analysis appeared higher than traditional treatment.

A retrospective study by Prajwal et al. provided an evaluation of efficacy for TA-TMA patients treated with Eculizumab (25). Although they also demonstrated higher response rates and survival in TA-TMA patients treated with Eculizumab, the articles they included were mostly cases (11–13). Cases usually report successful treatment rather than unsuccessful treatment. The benefit of Eculizumab may be significantly overestimated, and it is difficult to explain the heterogeneity between cases. In addition, this retrospective analysis focused on patients with

TABLE 2 | Baseline characteristics for patients included in meta-analysis.

Variable	Flore et al. (n = 12)	Prajwal et al. (n = 9)	Stephan et al. (n = 15)	Joslyn et al. (n = 10)	Michelle et al. (n = 6)	Sonata et al. (n = 64)
Age, median (range)	39(1.2–66)	7(2–61)	48(23–66)	44(17–59)	5.2(2.5–25)	5.5(2.7–11.7)
Gender, male (%)	7(58)	6(67)	7(47)	4(40)	4(67)	40(63)
Primary disease	Hematological disease	Hematological disease/others	Hematological disease	Hematological disease	Hematological disease/others	Hematological disease/others
Type of transplant	Allo/UCB	Allo/Auto	Allo	Allo/UCB	Auto	Allo/Auto/UCB
Conditioning regimen	MAC/RIC	MAC/RIC	MAC/RIC	MAC/RIC	MAC	MAC/RIC
Other risk factors at diagnosis, number (%)						
CNI used	8(67)	7(78)	9(60)	10(100)	NA	49(77)
aGvHD	8(67)	5(56)	12(80)	7(70)	O(O)	14(22)
Affection	6(50)	2(22)	8(67)	8(80)	1(17)	6(9)
Interval between HSCT and diagnosis,	121	68	264	93	35	<100 ^a
median days						
sC5b-9	NA	NA	456(127-810)	NA	151.5(100-460)	398(282-544)
Interval between diagnosis and Eculizumab	31	24	10	4	18	NA
therapy, median days						
Eculizumab therapy, median days	65	178	52.5	48.5	110	66
First-line therapy, number (%)/second-line	5(42)/7	2(22)/7	11(73)/4	7(70)/3	6(100)/0	64(100)/0
therapy, number						
Eculizumab dose, median dose	6	8	9	6	9.5	11
Overall response, number (%)	6(50)	7(78)	13(93) b	7(70)	4(67)	41(64)
Complete response, number (%)	2(17)	5(56)	NA	1(10)	1(17)	36(56)
Survivals, number (%)	4(33)	7(78)	5(33)	6(60)	4(67)	35(55)
Median follow-up months	14	12	8	13	30	15
AEs during Eculizumab therapy	Infection	No	Infection	Skin rash	NA	Infection
Cause of death, numbers (%)						
TA-TMA related	4(50)	0	2(20)	0	1(50)	8(28)
Infection	2(25)	0	8(80)	2(50)	0	6(21)
GvHD	2(25)	2(100)	0	1(25)	0	14(48)
Relapse of the primary disease	0	0	0	1(25)	1(50)	1(3)
Prognosis	CKD	CKD	CKD	CKD	CKD/HTN	CKD/HTN

Allo,; Auto, Autologous HSCT; CNI, calcineurin inhibitors; HTN hypertension; MAC, myeloablative regimen; RIC, reduced intensity regimen; UCB, umbilical cord blood.

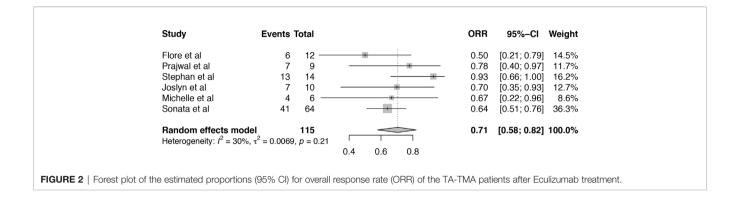
refractory discontinuation of calcineurin inhibitors and plasma exchange in patients with TA-TMA. Eculizumab was considered second-line therapy, whereas patients receiving Eculizumab as first-line therapy were not considered. What's more, adverse events during Eculizumab treatment were not analyzed. Our current meta-analysis incorporates the largest number of observational studies to date. Among these included studies, we joined the largest cohort to date, which included the terminal complement blocker Eculizumab as first-line treatment in patients with TA-TMA, which provides a higher weighting in the meta-data. Subgroup analysis and meta-regression were performed to detect heterogeneous sources. A sensitivity analysis was performed to demonstrate the stability and reliability of the findings. More importantly, our study provides not only a pooled assessment of efficacy but also of AEs and cause of death. Therefore, the current meta-analysis is a more comprehensive and credible analysis of the effectiveness and safety of Eculizumab for TA-TMA.

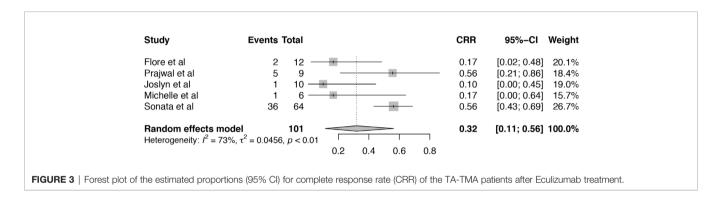
Because efficacy outcomes vary widely across centers, our current study analyzed which factors may contribute to differences in response rates and survival rates. Since the p-values of meta-regression were all greater than 0.05, the subgroup analysis of ORR could not explain the source of the

difference. Flore et al. (24) studied the lowest ORR compared to four other studies (19-21, 26). Due to the multicenter and retrospective nature of the study, there was heterogeneity in patient inclusion criteria, primary disease, stem cell transplantation characteristics, and Eculizumab regimens. It seems that this setup may be the source of the ORR discrepancy. More importantly, the overall response in the study results was good, but the complete response rate was meagre. This indicates that the kidney damage is advanced by the time of Eculizumab treatment. As a result, most patients do not get organ recovery. From this, it appears that the delay in Eculizumab treatment may prevent patients from achieving optimal response and maximum recovery of organ function. However, in the study by Joslyn et al., the median duration between diagnosis and Eculizumab therapy was shorter, at four days, compared to other studies. Only one patient achieved a complete response and organ function was restored (19). This suggests that earlier initiation of Eculizumab may have no significant effect on restoring organ function. How to improve the full response rate remains a big question. Our subgroup analysis of TA-TMA patient survival revealed that the time between HSCT and TA-TMA diagnosis is a potential source of SR heterogeneity, as the p value for meta-regression for days

^aAs 92% patients were diagnosed TA-TMA at a median of 23 days (IQR 3-48), and five had TA-TMA between 118 and 221 days after transplant, we regarded that the median days of interval between HSCT and diagnosis was less than 100 days.

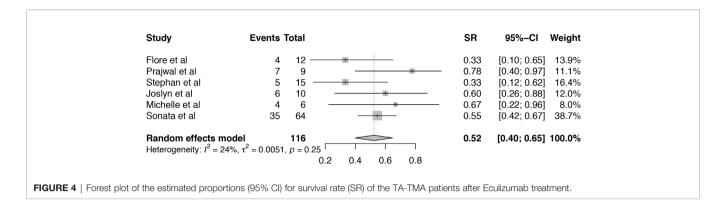
^bone unknown response due to early death.

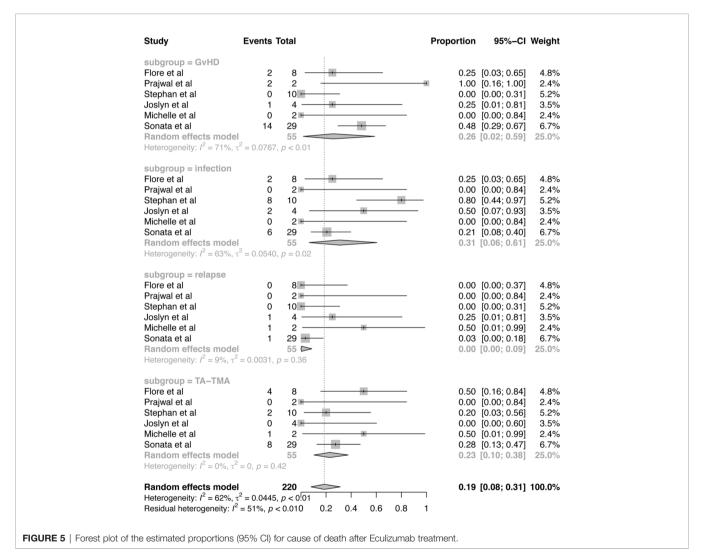




between transplantation and TA-TMA is 0.0266. TA-TMA diagnosed within the first 100 days after transplantation is more susceptible to publication bias than TA-TMA more than 100 days after transplantation. Therefore, early diagnosis of TA-TMA is the key to successful treatment of TA-TMA. However, the early diagnosis of TA-TMA faces challenges due to overlapping clinical features and the lack of standard diagnostic criteria as most studies support the idea that unregulated complement activation leads to the development of TA-TMA occurrence (7-9). The search for sensitivity and specificity of complement activation monitoring biomarkers should be of interest. Recently, Orsolya et al. showed that early elevation of sC5b-9 is a predictor of late development of TA-TMA (30). In this study, sC5b-9 levels increased from baseline levels to day 28 in patients with TA-TMA (n = 10), while the same trend was observed in only nine patients (p = 0.031) without TA-TMA (n = 23). In our meta-analysis, sC5b-9 levels were documented in three articles, and elevated sC5b-9 was observed in TA-TMA patients in two studies. However, the timing of the detection of sC5b-9 levels was not elaborated. Further studies are needed to determine whether terminal pathway activation is an independent predictor of TA-TMA after HSCT. Median age and primary disease may be another two factors contributing to significant differences between studies though their p-values of meta-regression are both 0.0827. Children seem to achieve higher SR than adults. In addition, SR of TA-TMA patients under treatment of Eculizumab reported in the studies which focused on hematological disease is lower than that of other studies. Sensitivity analysis of ORR and SR showed that the investigation by Stephan et al. (21) was a source of heterogeneity in SR of this meta-analysis. The SR (33%) in this single-center analysis was significantly lower compared to other studies. Patients in this study were diagnosed as hematological disease and the median age was 48 years, which was older than that of other five reports included. Additionally, the median days from hematopoietic stem cell transplantation to TA-TMA diagnosis were 264 days, the longest in all the studies.

The safety profile seems to indicate that Eculizumab is well tolerated. A more substantial observational data set covering a 5year registry of patients with aHUS reported that no new safety issues were identified in patients treated with adult or pediatric Eculizumab (31). In our meta-analysis, only one case of TRAEs, i.e. one patient with a severe rash, was reported, resulting in discontinuation of Eculizumab therapy. The most commonly reported AEs are infections. Eculizumab is a monoclonal antibody that inhibits C5 cleavage and prevents terminal complement activation (4). Patients treated with Eculizumab have an increased risk of infection, especially meningococcal infections, due to the lack of adequate functional complement (17). Whereas a study by Sonata et al. reported no cases of meningococcal infections in patients who had not received the meningococcal vaccine (32). And in our present study, no meningococcal infections were reported, which corresponds to the findings of Sonata et al. However, among patients treated with Eculizumab, the highest number of deaths due to infection was seen in the study by Stephan et al. Based on their report, an





increase in mortality due to infection in the group treated with Eculizumab was found compared to the conventional treatment group (21). Therefore, precaution and treatment of infection are equally urgent during the treatment of TA-TMA. GvHD is a risk factor that leads not only to TA-TMA but also to death during treatment. It has been shown that GvHD almost always precedes the diagnosis of TA-TMA, and there may be a mechanical link

between TA-TMA, GvHD, and endothelial injury (33). Another study reported that the occurrence of TA-TMA was associated with risk factors such as aGvHD grade ≥2, steroid-refractory aGvHD, and CMV reactivation/end-organ disease, but not with conditioning regimen (RIC or MAC), TBI use or TBI dose, primary condition, donor type, age, or gender. More importantly, patients diagnosed with TA-TMA combined with

aGvHD had significantly lower overall survival compared to patients with TA-TMA alone or GvHD (median 5.6 vs. 7.6 vs. 55.4 months; p < 0.0001) (34). The relationship between TA-TMA and GvHD is unclear. Future studies should provide information on the relationship between GvHD and Evidence of the TA-TMA link. TA-TMA itself is another major cause of death because of endothelial injury-related organ failure. It is necessary to explore how endothelial cells are damaged. One study from our center has reported that heme oxygenase-1 (HO-1) was significantly decreased in patients with TA-TMA and suppressed oxidative stress could attenuate complement deposition in TMA plasmachallenged HUVECs (35). The nuclear factor erythroid 2-related factor 2 (Nrf2), a transcription factor, initiates transcription of the HO-1 gene to protect cells from oxidative stress (36). Further experimental study about Nrf2 and endothelial injury is undertaken in our center.

Another area that needs to be discussed is how much Eculizumab needs to be given to achieve a hematological response and the duration of Eculizumab treatment for TA-TMA. For adult patients, the dose of induction therapy was 900 mg per week for four weeks. If the patient responds to induction therapy, the treatment is maintained at 1,200 mg administered every two weeks. For pediatric patients, the initial dose is based on body weight, and subsequent dose adjustments are based on maintaining total hemolytic complement activity (CH50) levels. Patients weighing less than 40 kg started at 600 mg and others at 900 mg. Induction therapy is also administered weekly for four weeks, and CH50 should be maintained at complement activator enzyme (CAE) levels of 0 to 3 (18, 37). The treatment then transitions to maintenance therapy, which adequately suppresses CH50 to a scale of three CAE (4) and then to maintenance therapy. Regarding the question of when Eculizumab can be safely discontinued, the study by Prajwal et al. proposed that ECU can be suspended after determining clinical symptoms and laboratory manifestations (38).

There are still some potential limitations to our study. First, there is a complete lack of randomized controlled trials and a limited study population size, and investigators have conducted limited studies on the efficacy of Eculizumab for TA-TMA. Second, although there is a great deal of heterogeneity among the included studies, the limited number of included studies prevents us from analyzing the sources of heterogeneity. Third, AEs are generalized in the article, so we do not have access to security data for AEs. Despite these limitations, our review is the first comprehensive meta-analysis of all eligible studies that analyzed the efficacy and safety of Eculizumab in patients with TA-TMA.

REFERENCES

- Epperla N, Li A, Logan B, Fretham C, Chhabra S, Aljurf M, et al. Incidence, Risk Factors for and Outcomes of Transplant-Associated Thrombotic Microangiopathy. Br J Haematol (2020) 189(6):1171–81. doi: 10.1111/bjh.16457
- Daly AS, Hasegawa WS, Lipton JH, Messner HA, Kiss TL. Transplantationassociated thrombotic microangiopathy is associated with transplantation from unrelated donors, acute graft-versus-host disease and venoocclusive disease of the liver. *Transfus Apher Sci* (2002) 27(1):3–12. doi: 10.1016/s1473-0502(02)00020-4

CONCLUSION

This systematic review and meta-analysis suggest that Eculizumab improves SR and ORR in patients with TA-TMA. Furthermore, patients with TA-TMA diagnosed within the first 100 days after HSCT are more likely to achieve better outcomes with Eculizumab compared to patients with TA-TMA diagnosed more than 100 days after HSCT. In addition, Eculizumab is well-tolerated, but the prevention and treatment of infection still require attention. Further RCTs and extensive prospective cohort studies are needed to evaluate efficacy and safety, particularly for Eculizumab for TA-TMA.

DATA AVAILABILITY STATEMENT

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found in the article/**Supplementary Material**.

AUTHOR CONTRIBUTIONS

RZ, MZ, and JQ contributed to the study conception and design, and writing the manuscript. RZ, WM, and ZZ performed data collection and analysis. DW and YH commented on the research design, data analysis, writing the manuscript, and supervision of the study. All authors contributed to the article and approved the submitted version.

FUNDING

This work was supported by the National Natural Science Foundation of China (81873432 and 81670132), grants from the Jiangsu Province of China (BE2016665, SBE2016740635 and ZDRCA2016047), The Natural Science Foundation of the Jiangsu Higher Education Institution of China (18KJA320006), and the Priority Academic Program Development of Jiangsu Higher Education Institutions (PAPD).

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fimmu.2020. 564647/full#supplementary-material

- Jodele S, Davies SM, Lane A, Khoury J, Dandoy C, Goebel J, et al. Diagnostic and risk criteria for HSCT-associated thrombotic microangiopathy: a study in children and young adults. *Blood* (2014) 124(4):645–53. doi: 10.1182/blood-2014-03-564997
- Jodele S, Laskin BL, Dandoy CE, Myers KC, El-Bietar J, Davies SM, et al. A new paradigm: Diagnosis and management of HSCT-associated thrombotic microangiopathy as multi-system endothelial injury. *Blood Rev* (2015) 29 (3):191–204. doi: 10.1016/j.blre.2014.11.001
- Ruutu T, Barosi G, Benjamin RJ, Clark RE, George JN, Gratwohl A, et al. Diagnostic criteria for hematopoietic stem cell transplant-associated

- microangiopathy: results of a consensus process by an International Working Group. *Haematologica* (2007) 92(1):95–100. doi: 10.3324/haematol.10699
- Cho BS, Yahng SA, Lee SE, Eom KS, Kim YJ, Kim HJ, et al. Validation of recently proposed consensus criteria for thrombotic microangiopathy after allogeneic hematopoietic stem-cell transplantation. *Transplantation* (2010) 90(8):918–26. doi: 10.1097/TP.0b013e3181f24e8d
- Laskin BL, Goebel J, Davies SM, Jodele S. Small vessels, big trouble in the kidneys and beyond: hematopoietic stem cell transplantation-associated thrombotic microangiopathy. Blood (2011) 118(6):1452–62. doi: 10.1182/blood-2011-02-321315
- Jodele S. Complement in Pathophysiology and Treatment of Transplant-Associated Thrombotic Microangiopathies. Semin Hematol (2018) 55(3):159– 66. doi: 10.1053/j.seminhematol.2018.04.003
- Gloude NJ, Khandelwal P, Luebbering N, Lounder DT, Jodele S, Alder MN, et al. Circulating dsDNA, endothelial injury, and complement activation in thrombotic microangiopathy and GvHD. *Blood* (2017) 130(10):1259–66. doi: 10.1182/blood-2017-05-782870
- Qi J, Wang J, Chen J, Su J, Tang Y, Wu X, et al. Plasma levels of complement activation fragments C3b and sC5b-9 significantly increased in patients with thrombotic microangiopathy after allogeneic stem cell transplantation. *Ann Hematol* (2017) 96(11):1849–55. doi: 10.1007/s00277-017-3092-9
- Peffault de Latour R, Xhaard A, Fremeaux-Bacchi V, Coppo P, Fischer AM, Helley D, et al. Successful use of eculizumab in a patient with post-transplant thrombotic microangiopathy. Br J Haematol (2013) 161(2):279–80. doi: 10.1111/bjh.12202
- Okano M, Sakata N, Ueda S, Takemura T. Recovery from life-threatening transplantation-associated thrombotic microangiopathy using eculizumab in a patient with very severe aplastic anemia. *Bone Marrow Transplant* (2014) 49 (8):1116–8. doi: 10.1038/bmt.2014.97
- Chapin J, Shore T, Forsberg P, Desman G, Van Besien K, Laurence J, et al. Hematopoietic transplant-associated thrombotic microangiopathy: case report and review of diagnosis and treatments. Clin Adv Hematol Oncol (2014) 12(9):565–73.
- Zuber J, Fakhouri F, Roumenina LT, Loirat C, Fremeaux-Bacchi V. French Study Group for a HCG. Use of eculizumab for atypical haemolytic uraemic syndrome and C3 glomerulopathies. *Nat Rev Nephrol* (2012) 8(11):643–57. doi: 10.1038/nrneph.2012.214
- US Food and Drug Administration. Soliris (eculizumab) [prescribing information]. Boston, MA: Alexion Pharmaceuticals (2018).
- Greenbaum LA, Fila M, Ardissino G, Al-Akash SI, Evans J, Henning P, et al. Eculizumab is a safe and effective treatment in pediatric patients with atypical hemolytic uremic syndrome. *Kidney Int* (2016) 89(3):701–11. doi: 10.1016/j.kint.2015.11.026
- 17. Soliris[®]. (2014).
- Legendre CM, Licht C, Muus P, Greenbaum LA, Babu S, Bedrosian C, et al. Terminal complement inhibitor eculizumab in atypical hemolytic-uremic syndrome. N. Engl J Med (2013) 368(23):2169–81. doi: 10.1056/NEJMoa1208981
- Rudoni J, Jan A, Hosing C, Aung F, Yeh J. Eculizumab for transplant-associated thrombotic microangiopathy in adult allogeneic stem cell transplant recipients. *Eur J Haematol* (2018) 101(3):389–98. doi: 10.1111/ejh.13127
- Schoettler M, Lehmann L, Li A, Ma C, Duncan C. Thrombotic Microangiopathy Following Pediatric Autologous Hematopoietic Cell Transplantation: A Report of Significant End-Organ Dysfunction in Eculizumab-Treated Survivors. Biol Blood Marrow Transplant (2019) 25(5): e163–8. doi: 10.1016/j.bbmt.2018.12.840
- Bohl SR, Kuchenbauer F, von Harsdorf S, Kloevekorn N, Schonsteiner SS, Rouhi A, et al. Thrombotic Microangiopathy after Allogeneic Stem Cell Transplantation: A Comparison of Eculizumab Therapy and Conventional Therapy. Biol Blood Marrow Transplant (2017) 23(12):2172–7. doi: 10.1016/j.bbmt.2017.08.019
- Moher D, Liberati A, Tetzlaff J, Altman DG. PRISMA Group. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. *Ann Intern. Med* (2009) 151(04):264–269, W64. doi: 10.7326/0003-4819-151-4-200908180-00135
- Wells GA, Shea B, O'Connell D, et al. The Newcastle–Ottawa Scale (NOS) for assessing the quality of nonrandomised studies in meta–analysis. Available at: http://www.ohri.ca/programs/clin- ical_epidemiology/oxford.asp (Accessed December 12, 2018).
- de Fontbrune FS, Galambrun C, Sirvent A, Huynh A, Faguer S, Nguyen S, et al. Use of Eculizumab in Patients With Allogeneic Stem Cell Transplant-

- Associated Thrombotic Microangiopathy: A Study From the SFGM-TC. Transplantation (2015) 99(9):1953–9. doi: 10.1097/TP.00000000000000001
- Dhakal P, Giri S, Pathak R, Bhatt VR. Eculizumab in Transplant-Associated Thrombotic Microangiopathy. Clin Appl Thromb Hemost (2017) 23(2):175– 80. doi: 10.1177/1076029615599439
- Jodele S, Dandoy CE, Lane A, Laskin BL, Teusink-Cross A, Myers KC, et al. Complement blockade for TA-TMA: lessons learned from a large pediatric cohort treated with eculizumab. *Blood* (2020) 135(13):1049–57. doi: 10.1182/ blood.2019004218
- Mulay S, Kreuter JD, Bryant SC, Elliott MA, Hogan WJ, Winters JL, et al. Outcomes of plasma exchange in patients with transplant-associated thrombotic microangiopathy based on time of presentation since transplant. *J Clin Apher* (2015) 30(3):147–53. doi: 10.1002/jca.21352
- Corti P, Uderzo C, Tagliabue A, Della Volpenull A, Annaloronull C, Tagliaferrinull C, et al. Defibrotide as a promising treatment for thrombotic thrombocytopenic purpura in patients undergoing bone marrow transplantation. *Bone Marrow Transplant* (2002) 29(6):542–3. doi: 10.1038/sj.bmt.1703414
- Au WY, Ma ES, Lee TL, Ha SY, Fung AT, Lie AK, et al. Successful treatment of thrombotic microangiopathy after haematopoietic stem cell transplantation with rituximab. Br J Haematol (2007) 137(5):475–8. doi: 10.1111/j.1365-2141.2007.06588.x
- Horvath O, Kallay K, Csuka D, Mezo B, Sinkovits G, Kassa C, et al. Early Increase in Complement Terminal Pathway Activation Marker sC5b-9 Is Predictive for the Development of Thrombotic Microangiopathy after Stem Cell Transplantation. Biol Blood Marrow Transplant (2018) 24(5):989–96. doi: 10.1016/j.bbmt.2018.01.009
- Rondeau E, Cataland SR, Al-Dakkak I, Miller B, Webb NJA, Landau D. Eculizumab Safety: Five-Year Experience From the Global Atypical Hemolytic Uremic Syndrome Registry. *Kidney Int Rep* (2019) 4(11):1568–76. doi: 10.1016/j.ekir.2019.07.016
- Jodele S, Dandoy CE, Danziger-Isakov L, Myers KC, El-Bietar J, Nelson A, et al. Terminal Complement Blockade after Hematopoietic Stem Cell Transplantation Is Safe without Meningococcal Vaccination. *Biol Blood Marrow Transplant* (2016) 22(7):1337–40. doi: 10.1016/j.bbmt.2016.03.032
- 33. Wall SA, Zhao Q, Yearsley M, Blower L, Agyeman A, Ranganathan P, et al. Complement-mediated thrombotic microangiopathy as a link between endothelial damage and steroid-refractory GvHD. *Blood Adv* (2018) 2 (20):2619–28. doi: 10.1182/bloodadvances.2018020321
- Kraft S, Bollinger N, Bodenmann B, Heim D, Bucher C, Lengerke C, et al. High mortality in hematopoietic stem cell transplant-associated thrombotic microangiopathy with and without concomitant acute graft-versus-host disease. Bone Marrow Transplant (2019) 54(4):540–8. doi: 10.1038/s41409-018-0293-3
- Pan T, Qi J, You T, Han S, Yang L, Miao W, et al. Circulating Heme Oxygenase-1 and Complement Activation in Transplant-Associated Thrombotic Microangiopathy. Biol Blood Marrow Transplant (2019) 25 (8):1486-91. doi: 10.1016/j.bbmt.2019.03.002
- Kensler TW, Wakabayashi N, Biswal S. Cell survival responses to environmental stresses via the Keap1-Nrf2-ARE pathway. Annu Rev Pharmacol Toxicol (2007) 47:89–116. doi: 10.1146/annurev.pharmtox.46.120604.141046
- Jodele S, Fukuda T, Vinks A, Mizuno K, Laskin BL, Goebel J, et al. Eculizumab therapy in children with severe hematopoietic stem cell transplantationassociated thrombotic microangiopathy. *Biol Blood Marrow Transplant* (2014) 20(4):518–25. doi: 10.1016/j.bbmt.2013.12.565
- Dhakal P, Bhatt VR. Is complement blockade an acceptable therapeutic strategy for hematopoietic cell transplant-associated thrombotic microangiopathy? Bone Marrow Transplant (2017) 52(3):352–6. doi: 10.1038/bmt.2016.253

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2021 Zhang, Zhou, Qi, Miao, Zhang, Wu and Han. This is an openaccess article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.





TKI Maintenance After Stem-Cell Transplantation for *FLT3*-ITD Positive Acute Myeloid Leukemia: A Systematic Review and Meta-Analysis

Nico Gagelmann, Christine Wolschke, Evgeny Klyuchnikov, Maximilian Christopeit, Francis Ayuk and Nicolaus Kröger*

Department of Stem Cell Transplantation, University Medical Center Hamburg-Eppendorf, Hamburg, Germany

OPEN ACCESS

Edited by:

Charles Craddock, University of Birmingham, United Kingdom

Reviewed by:

Federico Simonetta, Geneva University Hospitals (HUG), Switzerland Laurent Garderet, Assistance Publique Hopitaux de Paris, France

*Correspondence:

Nicolaus Kröger nkroeger@uke.uni-hamburg.de

Specialty section:

This article was submitted to Alloimmunity and Transplantation, a section of the journal Frontiers in Immunology

Received: 17 November 2020 Accepted: 22 February 2021 Published: 12 March 2021

Citation:

Gagelmann N, Wolschke C, Klyuchnikov E, Christopeit M, Ayuk F and Kröger N (2021) TKI Maintenance After Stem-Cell Transplantation for FLT3-ITD Positive Acute Myeloid Leukemia: A Systematic Review and Meta-Analysis. Front. Immunol. 12:630429. doi: 10.3389/fimmu.2021.630429 This analysis aimed to systematically review and synthesize the existing evidence regarding the outcome of tyrosine kinase inhibitor (TKI) maintenance therapy after allogeneic stem-cell transplantation for patients with FLT3-ITD-mutated acute myeloid leukemia (AML). We searched publicly available databases, references lists of relevant reviews, registered trials, and relevant conference proceedings. A total of 7 studies comprising 680 patients were included. Five studies evaluated sorafenib and 2 studies evaluated midostaurin, compared with control. The incidence of relapse was significantly reduced after TKI therapy, showing an overall pooled risk ratio (RR) of 0.35 (95% confidence interval [CI], 0.23-0.51; P < 0.001), with a marked 65% reduced risk for relapse. The overall pooled RR for relapse-free survival and overall survival showed significantly improved outcome after TKI maintenance therapy, being 0.48 (95% CI, 0.37-0.61; P < 0.001) and 0.48 (95% CI, 0.36-0.64; P < 0.001). The risk for relapse or death from any cause was reduced by 52% using TKI. No difference in outcome was seen for non-relapse mortality, and the risk for chronic or acute graft-vs. -host disease appeared to be increased, at least for sorafenib. In conclusion, post-transplant maintenance therapy with TKI was associated with significantly improved outcome in relapse and survival in patients with FLT3-ITD positive AML.

Keywords: sorafenib, midostaurin, maintenance, allogeneic stem cell transplantation, FLT3-internal tandem duplication, acute myeloid leukemia, graft-vs.-host disease

INTRODUCTION

Acute myeloid leukemia (AML) is a heterogeneous hematologic malignancy derived from hematopoietic stem cells with a series of abnormalities on the level of cytogenetics, genetics, and epigenetics (1, 2). Prognosis of this disease varies widely according to mutation profile, patient age, and comorbidities (2, 3). The duplication in Fms-like tyrosine kinase 3-internal tandem (*FLT3*-ITD) occurs in about 25% of adult AML patients (4–7). Patients harboring *FLT3*-ITD, particularly those with a high allelic ratio, show increased relapse rates and inferior survival, despite undergoing allogeneic stem-cell transplantation (6, 8).

TKIs After Transplant for FLT3-ITD AML

In the front-line setting of FLT3-mutated AML, combining conventional chemotherapy with a multi-targeted tyrosine kinase inhibitor (TKI), namely midostaurin, resulted in improved overall survival (9). Another multi-targeted TKI, sorafenib, has been approved for solid tumors such as hepatocellular and renal cell cancer (10, 11), but it has also shown efficacy in terms of prolonged progression-free survival in younger AML patients in combination with upfront chemotherapy (12), but not in the elderly population (13). In the relapsed/refractory setting, patients with FLT3-ITD-positive AML receiving TKI monotherapy showed promising outcomes (14-16), while this approach may remain a palliative strategy which is furthermore limited by emerging TKI resistance (17, 18). In contrast, when patients with FLT3-ITD-mutated AML relapsing after allogeneic stem-cell transplantation received sorafenib, the outcome may differ profoundly, as suggested by long-term remissions in selected patients (19, 20).

To reflect the increasing interest within clinical and basic research, we aimed to systematically review the current body of literature and to synthesize the existing evidence regarding the outcome of TKI maintenance therapy after allogeneic stem-cell transplantation for patients with *FLT3*-ITD-mutated AML.

METHODS

The methodology of this systematic review with meta-analysis was undergone in accordance with the Cochrane handbook. Further, dimensions of reporting were assessed with the preferred reporting items for systematic reviews and meta-analyses (PRISMA) guidelines and the meta-analysis of observational studies in epidemiology (MOOSE) checklist and adhered accordingly (21, 22). The research question was defined using the PICOS framework: population, FLT3-ITD mutated AML; intervention, stem-cell transplantation with TKI maintenance; comparator, placebo, or no maintenance; outcome, survival and relapse; study design, retrospective and prospective comparative studies.

Search Strategy

Medline and the Cochrane Library were searched (until August 11, 2020, respectively). Additionally, meeting abstracts archived between 2017 and 2020 from hematology/oncology meetings were screened. Review of clinicaltrials.gov was performed until August 11, 2020. The search strategy consisted of keywords specific to each database and considered all trial designs of human subjects and was not restricted by language. Search terms included all subject headings and associated keywords for "sorafenib or midostaurin or gilteritinib" and "leukemia or leukemia." Reference lists of relevant reports were reviewed in addition.

Study Selection, Data Extraction, and End Points

Two reviewers (NG and NK) independently screened titles, abstracts, and the full text of relevant articles. Disagreements were resolved by consensus. Studies were included if they fulfilled the following criteria: adult patients with *FLT3*-ITD

AML; prospective or retrospective studies reporting on patients receiving TKI therapy after stem-cell transplantation; evaluating a comparison with a control; reporting at least on relapse-free survival and/or cumulative incidence of relapse.

The following information was extracted from the included studies: the name of the first author, year of publication, study design, TKI treatment, control, number of participants, conditioning intensity for stem-cell transplantation, frequency of high-risk cytogenetics within the studied population, length of follow-up, and primary, and secondary outcomes. Primary end points for data synthesis were relapse-free survival and cumulative incidence of relapse. Secondary end points were overall survival, non-relapse mortality, chronic and acute graft-vs. -host disease (GVHD). Relapse-free survival was defined as time from randomization to first event of either AML relapse or death from any cause in prospective studies or as defined in retrospective studies. Definition of relapse was used in accordance with the included studies.

Risk of Bias and Quality Assessment

Risk of bias for prospective trials was addressed in accordance with tools developed by the Cochrane Collaboration, and the risk of bias for retrospective comparisons was assessed using the ROBINS-I tool (23). The certainty of the evidence for each outcome was assessed using the grading of recommendations assessment, development, and evaluation (GRADE) approach (24), including considerations of risk of bias, inconsistency, indirectness, imprecision, and publication bias. Retrospective studies were judged a priori as having serious risk of bias, in accordance with the GRADE approach. The resulting overall certainty of the evidence was assessed as high, moderate, low, or very low. All end points within the quality assessment were considered as being of critical importance.

Data Synthesis and Analysis

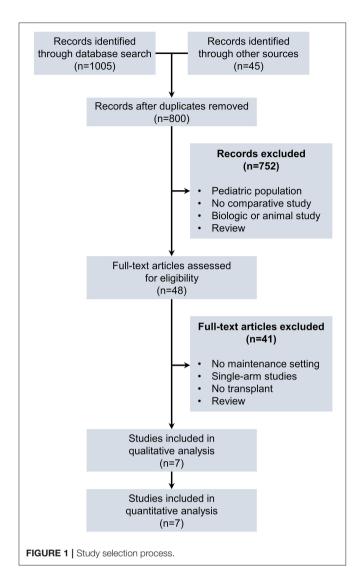
Risk ratios (RRs) and 95% confidence intervals (CIs) were calculated for primary and secondary end points by pooling the results from studies using the Mantel-Haenszel method and the random-effects model. Heterogeneity was assessed using I^2 and was categorized from moderate to high (25). Prespecified subgroups were different TKIs (midostaurin and sorafenib). All values with P < 0.05 were considered statistically significant. Means were calculated for the end point of safety. Analyses were performed using R statistical software version 3.6.1 using the meta and metafor packages (R Core Team. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. https://www.R-project.org/)(26).

RESULTS

Search Results

A total of 1050 citations were identified from the electronic database search and from other sources including meeting abstracts. After duplicates were removed, 800 unique citations remained. Based on title and abstract screening, 752 citations were excluded. Forty-one citations were excluded on the basis

TKIs After Transplant for FLT3-ITD AMI



of screening full-text articles. Reasons for exclusion were: studies with no maintenance setting; lack of direct comparison results; no patients undergoing allogeneic stem-cell transplantation; and review articles. Seven studies (27–33) were included in qualitative and quantitative analyses (**Figure 1**).

Study Characteristics and Risk of Bias

A total of 680 patients were included in the 7 studies. Three studies (28, 31, 32) were prospective randomized studies and 1 study (29) was a prospective study that compared TKI intervention with historical controls using propensity score matching. One prospective study was an abstract, and fully published data were not accessible during finalization of the present manuscript (32). The remaining 3 studies (27, 30, 33) were of retrospective design. Five studies evaluated the efficacy of sorafenib comprising 504 patients while the remaining 2 studies evaluated the TKI midostaurin and comprised 176 patients. Median age in the TKI group ranged from 24 to 55 years and frequency of patients having complete remission at time of

transplantation in the TKI group ranged from 61 to 100%. Four studies only used myeloablative conditioning transplantation. Median time of follow-up ranged from 18 to 59 months. The remaining characteristics are summarized in **Table 1**.

The duration of maintenance treatment differed between the studies. Maintenance was administered for 24 months or until occurrence of relapse, or limiting toxicity in Burchert et al. (31) In both studies from Xuan et al. (27, 28) TKI was given until day 180 after transplantation or until intolerable adverse events occurred. Maziarz et al. (32) applied TKI for twelve 4-week cycles. Patients in the study from Shi et al. (33) received TKI maintenance at a median of 238 days (range, 21–385 days). In Brunner et al. (30) TKI therapy was planned for 12–24 months, leaving continuation or early withdrawal to the discretion of the treating physician. Schlenk et al. (29) gave TKI therapy for 365 days.

Low risk of bias was assessed in 2 prospective randomized studies (28, 31), 4 studies showed moderate risk of bias (16, 27, 30, 33), and 2 studies conferred high risk of bias (29, 32). Overall, the risk of bias of the included studies according to each end point was judged to be serious. Publication bias could not be assessed due to the number of <10 studies included in the analysis, which is in accordance with the Cochrane handbook recommendations. **Supplementary Tables 1**, **2** depict the summary of the risk of bias profile for each dimension within each study and **Supplementary Table 3** summarizes the quality of evidence for each end point.

Relapse-Free Survival and Incidence of Relapse

The primary end point of relapse-free survival was assessed in all 7 studies at 18–59 months follow-up. The overall pooled RR showed significantly better relapse-free survival after TKI therapy, being 0.48 (95% CI, 0.37–0.61; P < 0.001) with no relevant heterogeneity ($I^2 = 0\%$, **Figure 2A**). The quality of the evidence was high. Subgroup analyses showed no significant difference in outcome between midostaurin and sorafenib (P = 0.21). However, the pooled RR for midostaurin was 0.60 (95% CI, 0.39–0.94; $I^2 = 0\%$) while a larger effect was seen for sorafenib, being 0.43 (95% CI, 0.32–0.58; $I^2 = 0\%$), compared with control.

Incidence of relapse was assessed in six studies. The overall pooled RR showed significantly reduced incidence of relapse, being 0.35 (95% CI, 0.23–0.51; P < 0.001) in favor of the TKI therapy with no relevant heterogeneity ($I^2 = 0\%$, **Figure 3A**). The quality of the evidence was high. Subgroup analyses showed no significant difference in outcome between midostaurin and sorafenib (P = 0.72). One study evaluated midostaurin, with a pooled RR of 0.43 (95% CI, 0.12–1.50). Sorafenib showed significantly reduced incidence of relapse showing a RR 0.34 (95% CI, 0.22–0.51; $I^2 = 0\%$), compared with control.

Overall Survival and Non-relapse Mortality

Significantly improved outcome for TKI therapy was also seen in overall survival, which was assessed in 6 studies. The overall pooled RR was 0.48 (95% CI, 0.36-0.64; P < 0.001) in favor of the TKI therapy with no relevant heterogeneity ($I^2 = 0\%$, **Figure 2B**). The quality of the evidence was high. Subgroup analyses showed no significant difference in outcome between midostaurin and

TABLE 1 | Study characteristics.

Study	Design	N	Age in TKI group (range)	TKI	Comparator	Myeloablative conditioning	CR at transplant ^b	High-risk cytogenetics	Length of follow-up
Burchert et al. (31)	Randomized phase 2	83	54 (23–74)	Sorafenib	Placebo	TKI: 42%, Placebo: 47%	TKI: 63%, placebo: 48%	TKI: 2%, Placebo: 8%	42 months
Brunner et al. (30)	Retrospective	81	55 (20–74)	Sorafenib	No TKI	TKI: 54%, No: 49%	100% (CR1)	8%	27 months
Schlenk et al. (29)	Prospective phase 2, propensity score matching with historical controls	116ª	54 (18–70)	Midostaurin	Historical control	NR	TKI: 61%, control: 43%	NR	24 months
Xuan et al. (28)	Randomized phase 3	202	35 (26–42)	Sorafenib	No TKI	100%	TKI: 73%, no: 77%	TKI: 7%, no: 5%	21 months
Xuan et al. (27)	Retrospective	82	37 (15–55)	Sorafenib	No TKI	100%	77%	TKI: 6%, no: 1%	59 months
Maziarz et al. (32)	Randomized phase 2	60	18–70 ^c	Midostaurin	No TKI	100%	NR	NR	18 months
Shi et al. (33)	Retrospective	56	24 (14–62)	Sorafenib	No TKI	100%	100%	17%	24 months

N, number; TKI, tyrosine kinase inhibitor; HSCT, hematopoietic stem-cell transplantation; MAC, myeloablative conditioning; RIC, reduced intensity conditioning; CR, complete remission; NR, not reported.

sorafenib (P=0.30). The pooled RR for midostaurin, which was evaluated only in 1 study, was 0.60 (95% CI, 0.36–1.00). A larger effect was seen for sorafenib after synthesis of the remaining 7 studies, with a RR 0.48 (95% CI, 0.36–0.64; $I^2=0$ %), compared with control.

Non-relapse mortality was assessed in 5 studies, which evaluated the efficacy of sorafenib. No significant difference between sorafenib and the control was seen, showing an overall pooled RR of 0.87 (95% CI, 0.51–1.47; P=0.60) with no relevant heterogeneity ($I^2=0\%$, **Figure 3B**). The quality of the evidence was low.

Graft-vs.-Host Disease and Safety

Chronic GVHD was assessed in 6 studies. No significant difference in the incidence was seen, with a trend toward higher incidence after TKI therapy showing an overall pooled RR of 1.14 (95% CI, 0.93–1.41; P=0.21) with no relevant heterogeneity ($I^2=0\%$, **Figure 4A**). The quality of the evidence was low. Subgroup analyses showed no significant difference in outcome between midostaurin and sorafenib (P=0.19). However, the pooled RR for midostaurin was 0.79 (95% CI, 0.43–1.44) while results for sorafenib suggested higher risk for chronic GVHD showing a RR of 0.43 (95% CI, 0.32–0.57; $I^2=0\%$), compared with control.

Similar results were yielded for acute GVHD, which was assessed in six studies. The overall pooled RR was 1.22 (95% CI, 0.96–1.55; P=0.10) with no relevant heterogeneity ($I^2=0\%$, **Figure 4B**). The quality of the evidence was high. No difference was seen between the TKIs (P=0.48). One study which evaluated midostaurin showed a RR of 1.06 (95% CI, 0.67–1.68), while risk for acute GVHD appeared to be increased after sorafenib

therapy showing a RR of 1.29 (95% CI, 0.98–1.70; $I^2 = 0$ %), when compared with control.

The safety profile could be assessed in the two randomized controlled trials on sorafenib (28, 31), for which means were calculated (**Table 2**). Frequency of adverse events were mostly comparable while skin toxicity was seen more frequently in the sorafenib group (19.5%) in comparison with the control group (6.3%), and hematologic toxicities such as neutropenia and thrombocytopenia, albeit in low absolute numbers, were more frequently observed in the sorafenib group (8.7 and 8.9%) compared with the control group (4.8 and 4.3%).

DISCUSSION

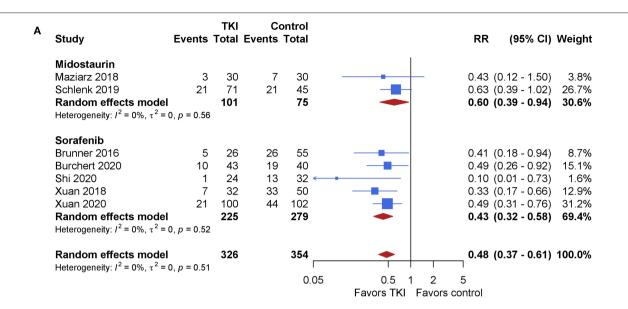
Patients with *FLT3*-ITD mutated AML undergoing allogeneic stem-cell transplantation have a high risk of relapse (34). Because oncogenic addiction is caused by *FLT3*-ITD (35), it was reasonable to hypothesize that it could be a potential therapeutic target in *FLT3*-ITD mutated patients (36). While evidence accumulated that the multi-targeted TKI midostaurin can improve outcome in the front-line setting (9), whether specifically targeting *FLT3*-ITD using TKI therapy after allogeneic stem-cell transplantation can improve outcome was long unknown (6, 37, 38).

This first evidence synthesis for TKI therapy after allogeneic stem-cell transplantation in *FLT3*-ITD mutated AML found TKI therapy using midostaurin or sorafenib in comparison with control was significantly associated with better outcome in relapse and relapse-free survival. The risk for relapse was reduced by marked 65% and the risk for relapse or death from any cause

^a The original number of patients in the study was 284, here we report on the subgroup analyses of patients that actually underwent midostaurin maintenance after stem-cell transplantation or not.

^bAs reported in the patient characteristics of the trials.

c Inclusion criteria, age distribution not given.



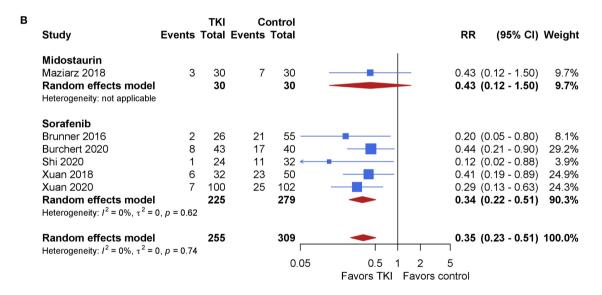
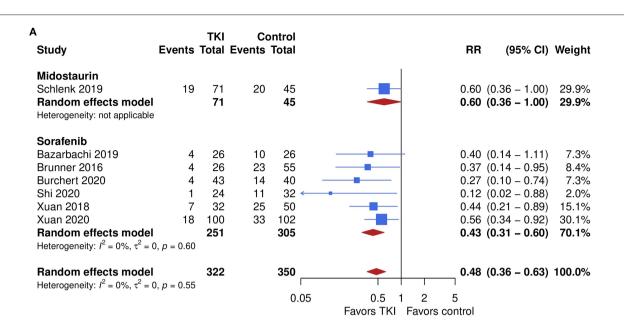


FIGURE 2 The impact of TKI therapy on primary end points of relapse-free survival and cumulative incidence of relapse. Relapse-free survival **(A)** was assessed in all 7 studies at 18–59 months follow-up. The overall pooled RR showed significantly better relapse-free survival after TKI therapy, being 0.48 (95% CI, 0.37–0.61; P < 0.001) with no relevant heterogeneity (P = 0%). Subgroup analyses showed no significant difference in outcome between midostaurin and sorafenib (P = 0.21). Incidence of relapse **(B)** was assessed in six studies. The overall pooled RR showed significantly reduced incidence of relapse, being 0.35 (95% CI, 0.23–0.51; P < 0.001) in favor of the TKI therapy with no relevant heterogeneity (P = 0%). Subgroup analyses showed no significant difference in outcome between midostaurin and sorafenib (P = 0.72).

was reduced by 53% using TKI. Furthermore, overall survival was significantly improved after TKIs with a risk reduction for death from any cause by 52%. No significant difference for non-relapse mortality was noted, which was only assessed in studies on sorafenib. The risk for GVHD appeared to be increased for TKI therapy.

Although the results of this analysis did not seem to be influenced by different TKIs, more studies evaluated the role of sorafenib (6). Two studies used midostaurin, of which 1 is a still ongoing phase 2 randomized study and 1 a priori studied the effects of midostaurin throughout the therapeutic course, with a subgroup analysis of



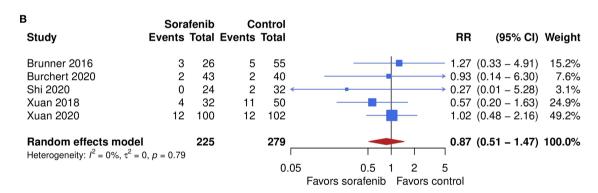
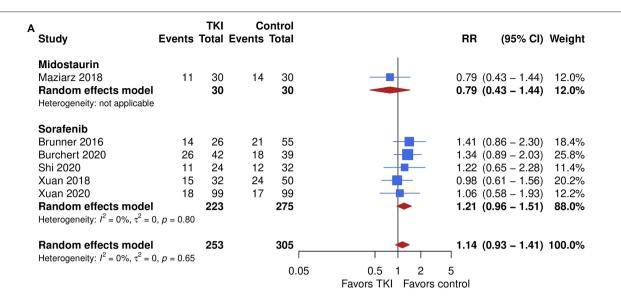


FIGURE 3 | The impact of TKI therapy on secondary end points of overall survival and non-relapse mortality. Significantly improved outcome for TKI therapy was also seen in overall survival **(A)**, which was assessed in 6 studies. The overall pooled RR was 0.48 (95% CI, 0.36–0.64; P < 0.001) in favor of the TKI therapy with no relevant heterogeneity ($I^2 = 0.001$). Subgroup analyses showed no significant difference in outcome between midostaurin and sorafenib ($I^2 = 0.001$). Non-relapse mortality **(B)** was assessed in 5 studies, which evaluated the efficacy of sorafenib. No significant difference between sorafenib and the control was seen, showing an overall pooled RR of 0.87 (95% CI, 0.51–1.47; $I^2 = 0.001$) with no relevant heterogeneity ($I^2 = 0.001$).

post-transplant therapy compared with no post-transplant therapy. Other TKIs, for example, quizartinib and gilteritinib, which inhibit FLT3 more specifically and potently in comparison with midostaurin (39), showed improvement in overall survival in relapsed/refractory patients (18, 40). Gilteritinib is also being investigated for post-transplantation maintenance in AML patients with *FLT3*-ITD in a phase 3 randomized study (NCT02997202). Further research is needed to ascertain the comparative efficacy and safety of

different TKIs post-transplantation therapy in *FLT3*-ITD mutated AML.

Given the well-described impact of minimal residual disease (MRD) on the outcomes after allogeneic stem-cell transplantation for AML (41, 42), and with the availability of a commercially available, next-generation sequencing-based MRD test for such patients, demonstration of a benefit of TKI therapy (or control) is critical to develop and incorporate TKIs into risk-based maintenance approaches (43). Both



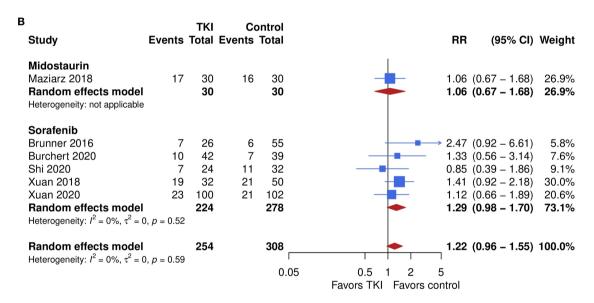


FIGURE 4 | The impact of TKI therapy on secondary end points of acute and chronic GVHD. Chronic GVHD **(A)** was assessed in six studies. No significant difference in the incidence was seen, with a trend toward higher incidence after TKI therapy showing an overall pooled RR of 1.14 (95% CI, 0.93–1.41; P=0.21) with no relevant heterogeneity ($l^2=0$ %). Subgroup analyses showed no significant difference in outcome between midostaurin and sorafenib (P=0.19). However, the pooled RR for midostaurin was 0.79 (95% CI, 0.43–1.44) while results for sorafenib suggested higher risk for chronic GVHD showing a RR of 0.43 (95% CI, 0.32–0.57; $l^2=0$ %), compared with control. Similar results were yielded for acute GVHD **(B)**, which was assessed in six studies. The overall pooled RR was 1.22 (95% CI, 0.96–1.55; P=0.10) with no relevant heterogeneity ($l^2=0$ %). No difference was seen between the TKIs (P=0.48). One study which evaluated midostaurin showed a RR of 1.06 (95% CI, 0.67–1.68), while risk for acute GVHD appeared to be increased after sorafenib therapy showing a RR of 1.29 (95% CI, 0.98–1.70; $l^2=0$ %), when compared with control.

prospective randomized studies on sorafenib showed subgroup results according to the MRD status at time of randomization (28, 31). While the Chinese study group showed significantly reduced incidence of relapse after sorafenib with hazard ratios of 0.28 for patients with undetectable MRD and 0.25 for detectable MRD (28), patients with undetectable MRD appeared to have better relapse-free survival in the

German study group, but this comparison was not statistically significant (31). In the German study group, patients with detectable MRD had significantly improved relapse-free survival, while the results need to be interpreted with caution owing to the relatively low numbers of patients in each group. The ongoing BMT CTN 1506 study on gilteritinib includes the critical objective to better understand the

TABLE 2 | Safety of sorafenib in 2 randomized controlled trials.

	Sorafenib	Control	
	(n = 143)	(n = 142)	
Neutropenia	8.7%	4.8%	
Thrombocytopenia	8.9%	4.3%	
Skin toxicity	19.5%	6.3%	
Infections	29.6%	28.0%	
Gastrointestinal toxicity	25.2%	21.7%	
Cardiac and renal insufficiency	11.8%	7.8%	

impact of MRD on outcomes with post-transplantation TKI maintenance.

Recent basic research findings indicate that the synergism of T-cells and sorafenib may metabolically reprogram AML-reactive T-cells, providing potential to contribute to immune-mediated curative treatment of *FLT3*-ITD mutated AML relapse (44). Furthermore and in general, a graft-vs.-leukemia effect is considered to be associated with the occurrence of GVHD (45). The findings of the present data synthesis suggest that at least sorafenib might increase the incidence of GVHD. Whether other mechanisms are involved in this effect requires further investigation.

In terms of safety, multi-targeted TKIs such as midostaurin and sorafenib are relatively non-specific and exert off-target activities. The prospective study on front-line midostaurin showed no unexpected adverse events (9). Higher grade 3-4 adverse events were seen for anemia (92.7 vs. 87.8%), rash (14.1 vs. 7.6%), and nausea (9.6 vs. 5.6%) in comparison with placebo, with no necessary dose modification for hematologic toxicity. With respect to sorafenib, small-sample studies have shown that the most common adverse events were related to hematological, skin, and gastrointestinal toxicities. In the present analysis, safety of post-transplantation TKI therapy could only be assessed for both prospective studies on sorafenib which showed no unexpected and comparable rates of adverse events when compared with control (Table 2). Only skin toxicity appeared to be slightly increased, but the overlap in skin rashes between an adverse event caused by sorafenib and graft-vs.host disease of the skin represents a difficulty for the differential diagnosis (27, 46). Furthermore, 60 and 50% of patients in the Chinese and German study needed a dose modification (interruption or reduction) because of adverse events. Dose reductions did not seem to limit sorafenib efficacy but more attention in view of TKI-specific toxicities and dose intensities is needed.

As with any meta-analysis, the present evidence synthesis regarding TKIs after stem-cell transplantation has several limitations. The conditioning intensity for transplantation was not homogenous. Four studies only used myeloablative conditioning transplantation (27, 28, 32, 33). Comparative

analyses on the superiority of one conditioning over another are inconclusive and may be interpreted on the subgroup level (42, 47-50), and the evidence on the impact of conditioning on outcome after TKIs is immature (51). Furthermore, the time of initiation of TKI was not homogeneous between studies and this meta-analysis could not account for differences in dosage schemes nor duration of treatment or treatment interruptions. Additionally, the present analysis may not provide any evidence for favoring one TKI over another. Further, RRs had to be calculated at different time of follow-up in the included studies, ranging from 18 to 59 months. This issue can be controlled for only when patient-level data are available. The risk of selection bias in meta-analyses of different donor stem-cell transplantation studies or due to the incorporation of findings from retrospective and prospective studies cannot be completely ruled out (52, 53). One prospective study on midostaurin was not adequately powered to identify a statistical difference between the groups (32), and on prospective study on sorafenib was prematurely terminated owing to slow patient recruitment (31). However, upfront exclusion of certain studies may even increase heterogeneity. And last, associations of allele ratios or TKD mutations cannot be addressed by analyses as presented here and further prospective evaluations are warranted.

In sum, this analysis identified a significant improvement in relapse-free survival, overall survival, and relapse incidence after post-transplant TKI therapy in *FLT3*-ITD mutated AML. These effects are irrespective of the TKI, while there is more consistent evidence for sorafenib so far. Ongoing studies could further help to better dissect patient subgroups that may benefit the most and identify refined relation of FLT3 selectivity vs. immune-stimulatory off-target activities governing TKI therapy after stem-cell transplantation in *FLT3*-ITD mutated AML.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/**Supplementary Material**, further inquiries can be directed to the corresponding author/s.

AUTHOR CONTRIBUTIONS

NG and NK had full access to all the data in the study, designed the study, retrieved, analyzed and interpreted the data, and wrote the first draft of the manuscript. All authors interpreted the data, wrote the manuscript, and approved the final version.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fimmu. 2021.630429/full#supplementary-material

REFERENCES

- Grimwade D, Ivey A, Huntly BJ. Molecular landscape of acute myeloid leukemia in younger adults and its clinical relevance. *Blood*. (2016) 127:29–41. doi: 10.1182/blood-2015-07-604496
- Papaemmanuil E, Gerstung M, Bullinger L, Gaidzik VI, Paschka P, Roberts ND, et al. Genomic Classification and Prognosis in Acute Myeloid Leukemia. N Engl J Med. (2016) 374:2209–21. doi: 10.1056/NEJMoa1516192
- 3. Döhner H, Estey E, Grimwade D, Amadori S, Appelbaum FR, Büchner T, et al. Diagnosis and management of AML in adults: 2017 ELN recommendations from an international expert panel. *Blood*. (2017) 129:424–47. doi: 10.1182/blood-2016-08-733196
- Gilliland DG, Griffin JD. The roles of FLT3 in hematopoiesis and leukemia. Blood. (2002) 100:1532-42. doi: 10.1182/blood-2002-02-0492
- Nakao M, Yokota S, Iwai T, Kaneko H, Horiike S, Kashima K, et al. Internal tandem duplication of the flt3 gene found in acute myeloid leukemia. Leukemia. (1996) 10:1911–8.
- Antar AI, Otrock ZK, Jabbour E, Mohty M, Bazarbachi A. FLT3 inhibitors in acute myeloid leukemia: ten frequently asked questions. *Leukemia*. (2020) 34:682–96. doi: 10.1038/s41375-019-0694-3
- Small D, Levenstein M, Kim E, Carow C, Amin S, Rockwell P, et al. STK-1, the human homolog of Flk-2/Flt-3, is selectively expressed in CD34⁺ human bone marrow cells and is involved in the proliferation of early progenitor/stem cells. *Proc Natl Acad Sci USA*. (1994) 91:459–63. doi: 10.1073/pnas.91.2.459
- Bazarbachi A, Bug G, Baron F, Brissot E, Ciceri F, Dalle IA, et al. Clinical practice recommendation on hematopoietic stem cell transplantation for acute myeloid leukemia patients with FLT3-internal tandem duplication: a position statement from the Acute Leukemia Working Party of the European Society for Blood and Marrow Transplantation. *Haematologica*. (2020) 105:1507–16. doi: 10.3324/haematol.2019.243410
- Stone RM, Mandrekar SJ, Sanford BL, Laumann K, Geyer S, Bloomfield CD, et al. Midostaurin plus chemotherapy for acute myeloid leukemia with a FLT3 mutation. N Engl J Med. (2017) 377:454–64. doi: 10.1056/NEJMoa1614359
- Escudier B, Eisen T, Stadler WM, Szczylik C, Oudard S, Siebels M, et al. Sorafenib in advanced clear-cell renal-cell carcinoma. N Engl J Med. (2007) 356:125–34. doi: 10.1056/NEJMoa060655
- Llovet JM, Ricci S, Mazzaferro V, Hilgard P, Gane E, Blanc J-F, et al. Sorafenib in advanced hepatocellular carcinoma. N Engl J Med. (2008) 359:378–90. doi: 10.1056/NEJMoa0708857
- Röllig C, Serve H, Hüttmann A, Noppeney R, Müller-Tidow C, Krug U, et al. Addition of sorafenib versus placebo to standard therapy in patients aged 60 years or younger with newly diagnosed acute myeloid leukaemia (SORAML): a multicentre, phase 2, randomised controlled trial. *Lancet Oncol.* (2015) 16:1691–9. doi: 10.1016/S1470-2045(15)00362-9
- Serve H, Krug U, Wagner R, Sauerland MC, Heinecke A, Brunnberg U, et al. Sorafenib in combination with intensive chemotherapy in elderly patients with acute myeloid leukemia: results from a randomized, placebo-controlled trial. J Clin Oncol. (2013) 31:3110–8. doi: 10.1200/JCO.2012.46.4990
- Zhang W, Konopleva M, Shi Y, McQueen T, Harris D, Ling X, et al. Mutant FLT3: a direct target of sorafenib in acute myelogenous leukemia. *J Natl Cancer Inst.* (2008) 100:184–98. doi: 10.1093/jnci/djm328
- Metzelder S, Wang Y, Wollmer E, Wanzel M, Teichler S, Chaturvedi A, et al. Compassionate use of sorafenib in FLT3-ITD-positive acute myeloid leukemia: sustained regression before and after allogeneic stem cell transplantation. *Blood.* (2009) 113:6567–71. doi: 10.1182/blood-2009-03-208298
- 16. Bazarbachi A, Labopin M, Battipaglia G, Djabali A, Passweg J, Socié G, et al. Sorafenib improves survival of FLT3-mutated acute myeloid leukemia in relapse after allogeneic stem cell transplantation: a report of the EBMT Acute Leukemia Working Party. Haematologica. (2019) 104:e398–401. doi: 10.3324/haematol.2018.211615
- Metzelder SK, Schroeder T, Finck A, Scholl S, Fey M, Götze K, et al. High activity of sorafenib in FLT3-ITD-positive acute myeloid leukemia synergizes with allo-immune effects to induce sustained responses. *Leukemia*. (2012) 26:2353–9. doi: 10.1038/leu.2012.105
- 18. Perl AE, Altman JK, Cortes J, Smith C, Litzow M, Baer MR, et al. Selective inhibition of FLT3 by gilteritinib in relapsed or refractory acute myeloid

- leukaemia: a multicentre, first-in-human, open-label, phase 1-2 study. *Lancet Oncol.* (2017) 18:1061–75. doi: 10.1016/S1470-2045(17)30416-3
- Metzelder SK, Schroeder T, Lübbert M, Ditschkowski M, Götze K, Scholl S, et al. Long-term survival of sorafenib-treated FLT3-ITD-positive acute myeloid leukaemia patients relapsing after allogeneic stem cell transplantation. Eur J Cancer. (2017) 86:233–9. doi: 10.1016/j.ejca.2017.09.016
- Battipaglia G, Massoud R, Ahmed SO, Legrand O, El Cheikh J, Youniss R, et al.
 Efficacy and feasibility of sorafenib as a maintenance agent after allogeneic
 hematopoietic stem cell transplantation for Fms-like tyrosine kinase 3
 mutated acute myeloid leukemia: an update. Clin Lymphoma Myeloma Leuk.
 (2019) 19:506–8. doi: 10.1016/j.clml.2019.04.004
- Moher D, Liberati A, Tetzlaff J, Altman DG. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. *BMJ*. (2009) 339:b2535. doi: 10.1136/bmj.b2535
- Stroup DF, Berlin JA, Morton SC, Olkin I, Williamson GD, Rennie D, et al. Meta-analysis of observational studies in epidemiology: a proposal for reporting. Meta-analysis Of Observational Studies in Epidemiology (MOOSE) group. *JAMA*. (2000) 283:2008–12. doi: 10.1001/jama.283.15.2008
- Sterne JA, Hernán MA, Reeves BC, Savović J, Berkman ND, Viswanathan M, et al. ROBINS-I: a tool for assessing risk of bias in non-randomised studies of interventions. BMJ. (2016) 355:i4919. doi: 10.1136/bmj.i4919
- Guyatt GH, Oxman AD, Vist GE, Kunz R, Falck-Ytter Y, Alonso-Coello P, et al. GRADE: an emerging consensus on rating quality of evidence and strength of recommendations. *BMJ*. (2008) 336:924–6. doi: 10.1136/bmj.39489.470347.AD
- Gagelmann N, Ayuk F, Wolschke C, Kröger N. Comparison of different rabbit anti-thymocyte globulin formulations in allogeneic stem cell transplantation: systematic literature review and network meta-analysis. *Biol Blood Marrow Transplant*. (2017) 23:2184–91. doi: 10.1016/j.bbmt.2017.08.027
- Balduzzi S, Rücker G, Schwarzer G. How to perform a meta-analysis with R: a practical tutorial. Evid Based Ment Health. (2019) 22:153–60. doi: 10.1136/ebmental-2019-300117
- Xuan L, Wang Y, Huang F, Jiang E, Deng L, Wu B, et al. Effect of sorafenib on the outcomes of patients with FLT3-ITD acute myeloid leukemia undergoing allogeneic hematopoietic stem cell transplantation. *Cancer*. (2018) 124:1954– 63. doi: 10.1002/cncr.31295
- 28. Xuan L, Wang Y, Huang F, Fan Z, Xu Y, Sun J, et al. Sorafenib maintenance in patients with FLT3-ITD acute myeloid leukaemia undergoing allogeneic haematopoietic stem-cell transplantation: an open-label, multicentre, randomised phase 3 trial. *Lancet Oncol.* (2020). doi: 10.1016/S1470-2045(20)30455-1
- Schlenk RF, Weber D, Fiedler W, Salih HR, Wulf G, Salwender H, et al. Midostaurin added to chemotherapy and continued single-agent maintenance therapy in acute myeloid leukemia with FLT3-ITD. *Blood*. (2019) 133:840–51. doi: 10.1182/blood-2018-08-869453
- Brunner AM, Li S, Fathi AT, Wadleigh M, Ho VT, Collier K, et al. Haematopoietic cell transplantation with and without sorafenib maintenance for patients with FLT3-ITD acute myeloid leukaemia in first complete remission. Br J Haematol. (2016) 175:496–504. doi: 10.1111/bjh.14260
- Burchert A, Bug G, Fritz LV, Finke J, Stelljes M, Röllig C, et al. Sorafenib maintenance after allogeneic hematopoietic stem cell transplantation for acute myeloid leukemia with FLT3-internal tandem duplication mutation (SORMAIN). J Clin Oncol. (2020) 38:2993–3002. doi: 10.1200/JCO.19.03345
- 32. Maziarz RT, Patnaik MM, Scott BL, Deol A, Rowley SD. Radius: a phase 2 randomized trial investigating standard of care \pm midostaurin after allogeneic stem cell transplant in FLT3-ITD-mutated AML. *Blood.* (2018) 132:662. doi: 10.1182/blood-2018-99-113582
- 33. Shi J, Cao L, Luo Y, Zhao Y, Tan Y, Yu J, et al. Maintenance sorafenib is superior to prophylactic donor lymphocyte infusion at improving the prognosis of acute myeloid leukemia with FMS-like tyrosine kinase 3 internal tandem duplication after allogeneic hematopoietic stem cell transplantation. Bone Marrow Transplant. (2020) 56:293–6. doi: 10.1038/s41409-020-01015-w
- 34. Brunet S, Labopin M, Esteve J, Cornelissen J, Socié G, Iori AP, et al. Impact of FLT3 internal tandem duplication on the outcome of related and unrelated hematopoietic transplantation for adult acute myeloid leukemia in first remission: a retrospective analysis. *J Clin Oncol.* (2012) 30:735–41. doi: 10.1200/JCO.2011.36.9868

- Smith CC, Wang Q, Chin C-S, Salerno S, Damon LE, Levis MJ, et al. Validation of ITD mutations in FLT3 as a therapeutic target in human acute myeloid leukaemia. Nature. (2012) 485:260–3. doi: 10.1038/nature11016
- Daver N, Kantarjian H. FLT3 inhibition in acute myeloid leukaemia. Lancet Oncol. (2017) 18:988–9. doi: 10.1016/S1470-2045(17)30509-0
- 37. Chen Y-B, Li S, Lane AA, Connolly C, Del Rio C, Valles B, et al. Phase I trial of maintenance sorafenib after allogeneic hematopoietic stem cell transplantation for Fms-like tyrosine kinase 3 internal tandem duplication acute myeloid leukemia. *Biol Blood Marrow Transplant*. (2014) 20:2042–8. doi: 10.1016/j.bbmt.2014.09.007
- Kindler T, Lipka DB, Fischer T. FLT3 as a therapeutic target in AML: still challenging after all these years. Blood. (2010) 116:5089–102. doi: 10.1182/blood-2010-04-261867
- Karaman MW, Herrgard S, Treiber DK, Gallant P, Atteridge CE, Campbell BT, et al. A quantitative analysis of kinase inhibitor selectivity. *Nat Biotechnol*. (2008) 26:127–32. doi: 10.1038/nbt1358
- Cortes JE, Khaled SK, Martinelli G. Efficacy and safety of single-agent quizartinib (Q), a potent and selective FLT3 inhibitor (FLT3i), in patients (pts) with FLT3-internal tandem duplication (FLT3-ITD)-mutated relapsed/refractory (R/R) acute myeloid leukemia (AML) enrolled in the global, phase 3, randomized controlled Quantum-R trial. Blood. (2018) 132:563.
- Jongen-Lavrencic M, Grob T, Hanekamp D, Kavelaars FG, Al Hinai A, Zeilemaker A, et al. Molecular minimal residual disease in acute myeloid leukemia. N Engl J Med. (2018) 378:1189–99. doi: 10.1056/NEJMoa17 16863
- 42. Hourigan CS, Dillon LW, Gui G, Logan BR, Fei M, Ghannam J, et al. impact of conditioning intensity of allogeneic transplantation for acute myeloid leukemia with genomic evidence of residual disease. *J Clin Oncol.* (2020) 38:1273–83. doi: 10.1200/JCO.19.03011
- Levis MJ, Chen Y-B, Hamadani M, Horowitz MM, Jones RJ. FLT3 inhibitor maintenance after allogeneic transplantation: is a placebocontrolled, randomized trial ethical? *J Clin Oncol.* (2019) 37:1604–7. doi: 10.1200/JCO.19.00321
- Mathew NR, Baumgartner F, Braun L, O'Sullivan D, Thomas S, Waterhouse M, et al. Sorafenib promotes graft-versus-leukemia activity in mice and humans through IL-15 production in FLT3-ITD-mutant leukemia cells. *Nat Med.* (2018) 24:282–91. doi: 10.1038/nm.4484
- Horowitz MM, Gale RP, Sondel PM, Goldman JM, Kersey J, Kolb HJ, et al. Graft-versus-leukemia reactions after bone marrow transplantation. *Blood*. (1990) 75:555–62.
- Escudier B, Worden F, Kudo M. Sorafenib: key lessons from over 10 years of experience. Expert Rev Anticancer Ther. (2019) 19:177–89. doi: 10.1080/14737140.2019.1559058
- Kröger N, Iacobelli S, Franke G-N, Platzbecker U, Uddin R, Hübel K, et al. Dose-reduced versus standard conditioning followed by allogeneic stem-cell

- transplantation for patients with myelodysplastic syndrome: a prospective randomized phase III study of the EBMT (RICMAC trial). *J Clin Oncol.* (2017) 35:2157–64. doi: 10.1200/ICO.2016.70.7349
- Ringdén O, Erkers T, Aschan J, Garming-Legert K, Le Blanc K, Hägglund H, et al. A prospective randomized toxicity study to compare reducedintensity and myeloablative conditioning in patients with myeloid leukaemia undergoing allogeneic haematopoietic stem cell transplantation. *J Intern Med*. (2013) 274:153–62. doi: 10.1111/joim.12056
- Scott BL, Pasquini MC, Logan BR, Wu J, Devine SM, Porter DL, et al. Myeloablative versus reduced-intensity hematopoietic cell transplantation for acute myeloid leukemia and myelodysplastic syndromes. *J Clin Oncol.* (2017) 35:1154–61. doi: 10.1200/JCO.2016.70.7091
- 50. Bornhäuser M, Kienast J, Trenschel R, Burchert A, Hegenbart U, Stadler M, et al. Reduced-intensity conditioning versus standard conditioning before allogeneic haemopoietic cell transplantation in patients with acute myeloid leukaemia in first complete remission: a prospective, open-label randomised phase 3 trial. *Lancet Oncol.* (2012) 13:1035–44. doi: 10.1016/S1470-2045(12)70349-2
- Appelbaum FR. Maintenance therapy after allogeneic hematopoietic cell transplantation for acute myeloid leukemia. Best Pract Res Clin Haematol. (2019) 32:101109. doi: 10.1016/j.beha.2019.101109
- 52. Gagelmann N, Bacigalupo A, Rambaldi A, Hoelzer D, Halter J, Sanz J, et al. Haploidentical stem cell transplantation with posttransplant cyclophosphamide therapy vs other donor transplantations in adults with hematologic cancers: a systematic review and meta-analysis. *JAMA Oncol.* (2019) 5:1739–48. doi: 10.1001/jamaoncol.2019. 3541
- Gagelmann N, Ljungman P, Styczynski J, Kröger N. Comparative efficacy and safety of different antiviral agents for cytomegalovirus prophylaxis in allogeneic hematopoietic cell transplantation: a systematic review and meta-analysis. Biol Blood Marrow Transplant. (2018) 24:2101–9. doi: 10.1016/j.bbmt.2018.05.017

Conflict of Interest: NK and CW were co-authors to one included study.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2021 Gagelmann, Wolschke, Klyuchnikov, Christopeit, Ayuk and Kröger. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.





Interplay Between the Intestinal Microbiota and Acute Graft-Versus-Host Disease: Experimental Evidence and Clinical Significance

Tao Hong 1† , Rui Wang 1† , Xiaoqi Wang 1 , Shijie Yang 1 , Weihao Wang 1 , Qiangguo Gao 3 and Xi Zhang 1,2*

¹ Medical Center of Hematology, Xinqiao Hospital, Third Military Medical University (Army Medical University), Chongqing, China, ² State Key Laboratory of Trauma, Burns and Combined Injury, Third Military Medical University (Army Medical University), Chongqing, China, ³ Department of Cell Biology, College of Basic Medicine, Third Military Medical University (Army Medical University), Chongqing, China

OPEN ACCESS

Edited by:

Nicolaus Martin Kröger, Universität Hamburg, Germany

Reviewed by:

Marit Inngjerdingen, University of Oslo, Norway Olle Thor, Hans Ringden, Karolinska Institutet (KI), Sweden Yuho Najima, Tokyo Metropolitan Komagome Hospital, Japan

*Correspondence:

Xi Zhang zhangxxi@sina.com

[†]These authors have contributed equally to this work

Specialty section:

This article was submitted to Alloimmunity and Transplantation, a section of the journal Frontiers in Immunology

Received: 22 December 2020 Accepted: 26 February 2021 Published: 16 March 2021

Citation:

Hong T, Wang R, Wang X, Yang S, Wang W, Gao Q and Zhang X (2021) Interplay Between the Intestinal Microbiota and Acute Graft-Versus-Host Disease: Experimental Evidence and Clinical Significance. Front. Immunol. 12:644982 doi: 10.3389/fimmu.2021.644982 Allogeneic hematopoietic stem cell transplantation (allo-HSCT) is a potentially curative therapy for many hematological disorders and autoimmune diseases, but acute graftversus-host disease (aGVHD) has remained a major obstacle that limits allo-HSCT and exhibits a daunting mortality rate. The gastrointestinal system is among the most common sites affected by aGVHD. Experimental advances in the field of intestinal microbiota research enhanced our understanding - not only of the quantity and diversity of intestinal microbiota - but also their association with homeostasis of the immune system and disease pathogenesis, including that of aGVHD. Meanwhile, ever-growing clinical evidence suggest that the intestinal microbiota is dysregulated in patients who develop aGVHD and that the imbalance may affect clinical outcomes, indicating a potential predictive role for microbiota dysregulation in aGVHD severity and prognosis. The current animal and human studies investigating the intestinal microbiota in aGVHD and the understanding of the influence and management of the microbiota in the clinic are reviewed herein. Taken together, monitoring and remodeling the intestinal microecology following allo-HSCT may provide us with promising avenues for diagnosing, preventing or treating aGVHD in the clinic.

Keywords: hematopoietic stem cell transplantation, acute graft-versus-host disease, intestinal microbiota, diversity, strategies

INTRODUCTION

Malignancies of the hematopoietic system and therapy-refractory autoimmune diseases are frequently associated with high mortality and hence represent the most common indications to perform allogeneic hematopoietic stem cell transplantation (allo-HSCT) (1, 2). However, graft-versus-host disease (GVHD), in which donor-derived T cells recognize host tissues as foreign, causing inappropriate and aberrant immune attacks, remains one of the major limitations to HSCT. Approximately 40-60% of patients receiving allo-HSCT may suffer from GVHD (3–5) with a

mortality rate of 15% to 20% (3, 4, 6–9). Clinically, prophylaxis of acute GVHD (aGVHD) involves immunosuppression of donor cells, but there is no standard approach, and it often varies by institution (3). Treatment protocols can also be challenging because therapeutic options are limited, response rates for corticosteroids are only approximately 50%, and response durations are typically brief (6, 7). In addition, several drugs are reported to be effective in patients not responding to corticosteroids, but most data are unconvincing, and combination therapies tried to date have yielded modest or no benefit over corticosteroids alone (10–12). Because of the small number of results from well-designed, large-scale, clinical studies, there is considerable variability in dealing with aGVHD worldwide, which leads to updated consensus recommendations that still have problems (13).

Much work has been done to research the biological mechanisms participating in the pathogenesis of aGVHD, but the specific nature of these interactions has not been fully elucidated, especially the relationship between aGVHD and the intestinal microbiota. The intestinal microbiota has been proven to be critical for maintaining healthy tissues and stimulating immunity (14), and increasing evidence has revealed that dysbiosis in intestinal microbial populations is linked to human disease and defects in immunity (15–18). Recent studies have notably widened our understanding of the interactions between the loss of intestinal bacterial diversity and aGVHD following allo-HSCT. This review provides an update of current knowledge on the cross-talk between them, with the purpose of determining improved prophylactics and therapies for aGVHD based on the role of the intestinal microbiota.

THE INTESTINAL TRACT AND INTESTINAL MICROBIOTA

The gastrointestinal tract consists of the mucous layer, submucous layer, muscular layer and serosa from the inner to outer layers. The intestinal mucosa, the innermost layer of the gastrointestinal tract, can be further divided into the epithelium, lamina propria and muscularis mucosae (19). The epithelium is a single-cell layer that contains unique secretory cells and stem cells, and many immune cells in the lamina propria help to monitor pathogens and maintain immune tolerance to food and commensal antigens (20). The mucosal surface maintains an intact biological barrier that prevents substantial bacterial and other detrimental invasion into the host tissue and blood circulation under steady-state homeostasis; this function is implemented by epithelial tissues, gut-associated lymphoid tissue (GALT) and important secretory components (21). The commensal intestinal microbiota also contributes to the maintenance of intestinal ecological balance. An overview of homeostasis between the microbiota and the host intestinal mucosa is shown in Figure 1A.

The human intestinal tract hosts 10¹³ to 10¹⁴ microbial organisms of approximately 1000 species (**Table 1**) (22, 23).

Although viruses and fungi are also present in considerable amounts and diversity, the vast majority of these organisms are bacteria collectively termed the gut microbiota (24-26), which play an important role in the synthesis of a variety of vitamins and amino acids, participating in the metabolism of carbohydrates and proteins and promoting the absorption of various mineral elements (27). The balance and diversity of the gut microbiota is of great importance for the human body, as researchers have found close connections between changes in the gut microbiota and human diseases, such as obesity (28), diabetes (29, 30), functional bowel syndrome (31), autism (32), and autoimmune diseases (e.g., rheumatoid arthritis (33)). For decades, the analysis of the intestinal microbiota has been largely dependent on ex vivo cultivation of bacteria, which vields only 10-30% of the population, limiting knowledge of the bacterial composition (34). In recent years, the development of nextgeneration sequencing technologies, such as 16S rDNA sequencing and metagenomics, has allowed for further identification of microorganisms, which clarifies the detailed and specific role of the intestinal microbiota in aGVHD, leading to a new era of research (35, 36).

INTESTINAL MICROBIOTA IN THE MECHANISM OF AGVHD

Pathophysiology

Development of aGVHD is considered a three-step process. The microbiota-linked pathogenesis of gastrointestinal aGVHD is summarized herein.

In the first step, when the conditioning regimen of allo-HSCT damages the intestinal epithelium, homeostasis between the host and intestinal commensals is disturbed (Figure 1B). Totalbody irradiation (TBI) induces dose-dependent damage to the gut lining including killing intestinal stem cells (ISCs), depleting or inhibiting non-epithelial cells, injuring intestinal crypts and causing gastrointestinal tract syndrome (37-39), by mechanisms such as increasing p53-mediated epithelial apoptosis (40) and plasminogen activator inhibitor-type 1 (PAI-1)-mediated enteritis (41). The intestinal mucosa is the major target tissue, and histological evidence has shown villous shortening, increased lymphocytic cell infiltration, crypt destruction and epithelial apoptosis. Crypt cell degeneration has been suggested to be the initial lesion of gastrointestinal aGVHD (42-44), and loss of goblet cells and Paneth cells has been shown to lead to translocation of dominant luminal pathogens and pathogen-associated molecular patterns (PAMPs) as well as intestinal dysbiosis, which further accelerates gastrointestinal aGVHD and infections (45). Moreover, some studies have provided evidence that these toxic effects are partially mediated by the intestinal microbiota. Lai et al. found that mice treated with antibiotics or deficient in myeloid differentiation primary response gene 88 (MyD88), a crucial adaptor for recognition of microbial molecules, showed less crypt loss and less damage to progenitor and stem

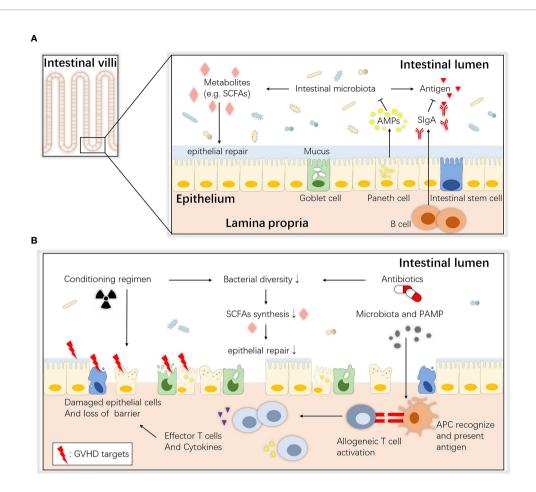


FIGURE 1 | Overview of intestinal ecology and gastrointestinal aGVHD. (A) Homeostasis between the commensals and host intestinal epithelium. At steady state, host intestinal epithelial cells live with commensals, and their interaction maintains immune and biological homeostasis. ISCs maintain the regeneration of the epithelium, Paneth cells secrete AMPs that create a sterility gradient, goblet cells produce mucus to separate the microbiota from host epithelial tissue, and immune cells such as B lymphocytes secrete SIgA to neutralize biologically active microbial antigens. Together, they maintain an intact barrier on the mucosa surface. SCFAs (e.g., butyrate) are bacterial fermentation products that can be used as an energy source and regulate the differentiation, recruitment and activation of immune cells. (B) Pathogenesis of gastrointestinal aGVHD. During allo-HSCT, the antibiotics and altered diet and cell damage caused by the conditioning regimen all lead to dysbiosis and metabolic disorders. Then, the depletion of SCFAs may also contribute to epithelial defects, allowing translocation of pathogenic bacteria and PAMPs. APCs (e.g., DCs) recognize them and elicit Th1 and Th17 responses and the release of proinflammatory factors that enhance tissue damage. ISCs, intestinal stem cells; AMPs, antimicrobial peptides; SCFAs, short-chain fatty acids; SIgA, secretory immunoglobulin A; APC, antigen-presenting cell; PAMP, pathogen-associated molecular pattern.

TABLE 1 | Bacterial taxonomy of some important microbiota constituents in the literature.

Phylum	Class	Order	Family	Genus	Species
Verrucomicrobia	Verrucomicrobiae	Verrucomicrobiales		Akkermansia	
Proteobacteria	γ-proteobacteria	Enterobacteriales	Enterobacteriaceae	Escherichia	E. coli
Bacteroidetes	Bacteroidia	Bacteroidales	Bacteroidaceae	Bacteroide	
Firmicutes	Clostridia	Clostridiales	Lachnospiraceae	Blautia	
				Eubacterium	
			Ruminococcaceae		
	Erysipelotrichia	Erysipelotrichiales		Erysipelatoclostridium	
	Bacilli	Lactobacillales	Lactobacteriaceae	Enterococcus Lactobacillus	Enterococc

cells after radiation (46). Seth et al. showed that the microbiota protects against dextran sodium sulfate-induced intestinal damage after radiation (47). It is possible that the microbiota affects the initiation of damage.

In terms of the alloreactive cells at the second step, it is well known that potential pathogens and their antigen molecules can activate T cells and affect differentiation. A study showed that cohousing laboratory mice with feral mice produced mice with

immune systems closer to those of adult humans, with a preference toward effector and memory T cell populations, suggesting that 'dirtier' mice have more intrinsic activation leading to more GVHD (48). After observing their existence, studies have shown that activated T cells access the epithelium via gut-specific homing molecules on the T cells and adhesion ligands on the vasculature, such as MAdCAM-1; the intestinal crypts are the primary location invaded by T cells, wherein they directly interact with ISCs (49–54). For T cell differentiation, the induction of both T helper (Th) and regulatory T (Treg) cells is influenced by the microbiota. Breaching the intestinal barrier or penetration by the microbiota in aGVHD activates the IL-23 pathway by JAK-STAT, stimulating epithelial cells to produce serum amyloid A proteins, which leads to Th17 differentiation, an important subset in aggravating aGVHD (55-57). Interestingly, some species, such as Bacteroides fragilis, prevent IL-17 production by releasing polysaccharide A (PSA) and promoting Foxp3+ Tregs, and Tregs maintain gastrointestinal homeostasis via the release of IL-10 (58), which could alleviate aGVHD. Other species, i.e., Clostridiales, produce SCFAs (e.g., butyrate and propionate), which block histone deacetylases (HDACs) through the G-protein receptor (GPR) to promote acetylation of histone H3 in Tregs at the Foxp3 locus, which also induces Treg differentiation (59). In addition, innate lymphoid cells 3 (ILC3s), an activated population with expression of the natural cytotoxicity receptor (NCR, such as NKp44) and nuclear hormone receptor RORyt were found mainly in mucosal tissues and emerged as modulators of conditioning-induced tissue damage in the context of aGVHD, secreting IL-22 and IL-17, which are necessary in defense against bacterial pathogens (60, 61). NCR+ ILC3-derived IL-22 has already been found to be

crucial in epithelial recovery and protect ISCs from damage by activating STAT3 and downstream regulators of cellular proliferation and survival which finally attenuates aGVHD (62, 63). But there is no direct evidence that NCR ILC3s-derived IL-17 is involved in the pathology of aGVHD (64). Granulocyte macrophage colony-stimulating factor (GM-CSF) produced by ILC3s is also essential for the normal development of intestinal dendritic cells (DCs) involved in Treg induction (65). These interactions are summarized in **Figure 2**.

The third step links cytokine storms and inflammatory amplification, which induce direct damage and establish typical aGVHD injury. Damage to the intestine plays a central role in amplifying systemic GVHD by propagating a proinflammatory cytokine milieu (66). Moreover, the many combinations of cytokines (e.g., TNF, INF-γ, IL-1, IL-2, and IL-17) and costimulatory networks at the T cell surface are definitely complex, in addition to numerous products produced by the intestinal microbiota. The role of the intestinal microbiota in regulating cytokines has been elucidated in some previous studies. Atarashi et al. showed that on the basis of high potency in enhancing Treg abundance and inducing anti-inflammatory molecules, 17 rationally selected strains of Clostridia result the increase of IL-10 in the gut (67). Another study reported that increased abundance of Enterobacteriaceae is positively correlated with IL-17A aggravating aGVHD (68).

In addition to the abovementioned mechanisms, the pathogenesis of aGVHD involves many other specific mechanisms that require deeper investigation. However, it is increasingly clear that the intestinal microbiota indeed participates in the initiation and development of aGVHD.

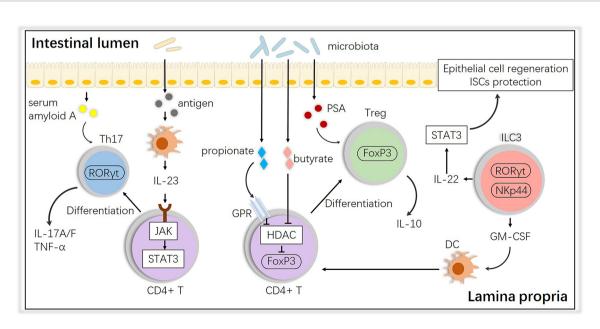


FIGURE 2 | Impact of the intestinal microbiota on T cell subsets and interactions with ILC3s. The intestinal microbiota influences the differentiation of T cells into anti-inflammatory Tregs or proinflammatory Th17 cells, and ILCs play an important role in this process. PSA, polysaccharide A; GM-CSF, granulocyte macrophage colony-stimulating factor; HDAC, histone deacetylase; GPR, G protein receptor.

Metabolites

The intestinal microbiota generates a wide range of bioactive metabolites serving as mediators and has pervasive consequences in aGVHD; modifications in bacteria-derived metabolites may be a new perspective regarding this disease (69).

Short-chain fatty acids (SCFAs) are one of the major microbial-derived metabolites found exclusively in the intestinal tract, which function in maintaining the epithelial barrier and colonocyte survival as well as play a diverse array of immune regulatory roles (70, 71). It has been reported that butyrate, one of the three main SCFAs, has a protective effect against aGVHD in murine models; butyrate restoration improved histone acetylation and IEC junctional integrity and decreased IEC apoptosis, ultimately mitigating aGVHD (72–74).

3-Indoxyl sulfate (3-IS), another promising metabolite analyzed in aGVHD, is a tryptophan metabolite of commensal colonic bacteria that has been identified as an indirect marker of a balanced microbiota and predicts the outcome of allo-HSCT (75, 76). Moreover, studies have also shown that gut tryptophan-produced indole metabolites reduce GVHD severity *via* type I interferon (IFN I) (77).

Respiratory metabolites may hold potential as surrogate markers for aGVHD (78–80). Various microbiota constituents are known to produce volatile metabolites, and volatile organic compounds (VOCs) generated during pathologic processes have been reported monitored in diseases such as obesity (81), hepatitis (82) and IBD (83). More recently, Hamilton et al. analyzed the VOCs of patients with and without aGVHD and correctly classified 89% (17 of 19) and 90% (9 of 10) of them, respectively, showing that breath analysis is a feasible and promising noninvasive method to detect and potentially monitor aGVHD (84).

Choline, phosphatidylcholine and carnitine-containing dietary ingredients can be metabolized into trimethylamine (TMA) and subsequently converted into trimethylamine N-oxide (TMAO), which could induce vascular inflammation and endothelial dysfunction (85). Dietary habit such as high-choline diet producing high level of TMAO alters distinct quality and quantity of gut microbiota which might affect microbial metabolites and GVHD severity. The latest research further explored TMAO in aGVHD and found that TMAO enhanced the allogenic GVH reaction. In an animal model, the group stimulated with TMAO showed a worse survival rate, higher GVHD scores and more damage to target tissues, which resulted from Th1 and Th17 differentiation (86).

These studies innovatively provide the link between microbiota-derived metabolites and aGVHD, which sheds light on alleviating aGVHD by controlling metabolism.

LOSS OF MICROBIOTA DIVERSITY IN AGVHD

Views about the role of the microbiota in aGVHD are separated into two roles: a direct role and an indirect role. The former considers that the intestinal microbiota directly promotes tissue injury or inflammation after allo-HSCT, and animals possess normal immunocompetence but abnormal microbiota community compositions, so eliminating this proinflammatory factor by antimicrobial treatment should protect the recipient from aGVHD (87–89). The latter considers the intestinal microbiota to play an indirect role, as animals raised in germfree conditions have abnormal development with highly aberrant immunity, and this kind of animal does not possess a prerequisite immune ability to develop aGVHD (90–92). Some researchers even support that both direct and indirect effects may occur simultaneously (93). Furthermore, we can identify some common alterations or differences from animal experiments and clinical studies.

Animal Studies

Earlier transplantation studies on animals in the 1970s focused on manifestations such as better survival and lower mortality with the management of the intestinal microbiota (94, 95). Subsequently, protection from GVHD in germ-free animals was confirmed in the 1980s (87), including xenogeneic transplantation that typically abrogates severe GVHD (88, 89). Antibiotic-mediated gut decontamination in mice (96) and dogs (97) showed that both microbes and aberrant immune development most proximately affect the development of GVHD. However, few animal studies have measured the impact of antibiotic treatment on microbiota composition or linked specific bacterial species to GVHD at this time.

In the early 21st century, advances in technology advanced research on the alterations (Table 2). Shono et al. found that piperacillin-tazobactam and imipenem-cilastatin therapy led to distinct patterns of gut microbiota composition in GVHD mice, with an increasing abundance of an Akkermansia strain, a bacterium with mucus-degrading capabilities, which raises the possibility that mucus degradation may contribute to murine GVHD (102). At the phylum level, the proportions of Firmicutes and Bacteroidetes, two of the major enteric commensals, were found to be decreased in GVHD mice, while the abundance of E. coli at the genus level was higher, which could be due to the development of systemic infection (100, 101). Furthermore, for Clostridiales and Lactobacillales, at the order level of Firmicutes, diversity analysis showed loss of the former but expansion of the latter, while eliminating Lactobacillales from mice before transplantation aggravated GVHD, and reintroducing the predominant Lactobacillus species showed a significant protective effect (99). Similar to these results, a report showed that the acute phase of GVHD was characterized by a shift toward the Enterobacteriaceae family. Bacteroides and Enterococcus abundances increased during GVHD, whereas Clostridia, Bifidobacteria and Bacillus were less abundant, but in this report, Lactobacillus abundance was decreased in GVHD (98), Moreover, subsequent studies also presented contrary results that the abundances of Lactobacillus and another uncultured bacterium from Firmicutes increased while the abundances of E. coli and another uncultured bacterium from Bacteroidetes decreased in allogeneic immunoglobulin yolk (IgY)-treated GVHD animals (103). However, the exact mechanisms of these alterations remain unclear, although several mechanisms have

TABLE 2 | Summary of nearly 10 years of experimental studies investigating the association between alterations in the intestinal microbiota and aGVHD.

Year	Mouse model	Relative abundance alteration of microbiota	Relationship with aGVHD	Ref
2010	(Age- and sex-matched) Balb/c→C57B6(TLR ^{-/-} , MyD88 ^{-/-} , TRIF ^{-/-} and WT)	Bacteroides, Enterococcus ↑ Clostridia, Bifidobacteria, Bacillus, Lactobacilli,	GVHD development is accompanied by shift towards proinflammatory bacterial species(enterobacteria, enterococci and <i>Bacteroides</i> /Prevotella).	(98)
2012	B10.BR→B6 Balb/c→B10.BR B6→BM12	Clostridiales↓ Lactobacillales↑	Increased microbial chaos early after allo-HSCT is a potential risk factor for subsequent GVHD.	(99)
2012	(F) B6→B6D2F1 (F) C3H.Sw→B6 (F) B6-Ly5.1→B6D2F1	 Firmicutes, Bacteroidetes↓ E. coli↑	Diversity of the microbial community was significantly reduced in mice with GVHD.	(100)
2015	(F) B6→BALB.B (F) B6/SJL→BALB.B	● E. coli↑	Diversity of the microbial community was significantly reduced in mice with GVHD.	(101)
2016	(F) C57BL/6→129S1	 Erysipelotrichia, Enterococcus, Akkermansia† Clostridiales↓ 	Aggravated GVHD mortality was associated with imipenem-cilastatin or piperacillin-tazobactam treatment mice which lead to an increase in Akkermansia muciniphila.	(102)
2017	C57BL/6→B6D2F1 B6D2F1→B6D2F1	Lactobacillus and another uncultured bacterium from Firmicutes↑ E. coli and another uncultured bacterium from Bacteroidetes.	Improvement of aGVHD in animals treated with immunoglobulin may be mediated by reducing pathogenic bacteria such as <i>E. coli</i> and increasing probiotic bacteria such as <i>Lactobacilius</i> .	(103)
2017	BALB/c(WT, IL-17A ^{-/-} , IL- 17RA ^{-/-})→B6(WT, IL-17A ^{-/-} , IL-17RA ^{-/-}) BALB/c(WT)→B6(WT, IL- 17RA/B/C ^{-/-})	Microbiome of WT mice shifted toward that of the IL-17RA/C-deficient mice during cohousing prior to transplant	IL-17–sensitive microbiota controls susceptibility to aGVHD with increased susceptibility to aGVHD transferred to WT mice <i>via</i> cohousing with IL-17RA or IL-17RC–deficient mice.	(57)
2019	(M) C57BL/6→BALB/c (M) C57BL/6→129S1/Sv (M) LP/J→C57BL/6	Enterococcus†	Enterococcus expands in mouse after allo-HSCT and exacerbates GVHD severity which is dependent on the lactose.	(104)
2019	(F) BALB/c→B6(WT and Nirp6- ^{-/-}) (F) C3H.sw→B6(WT and Nirp6- ^{-/-})	Verrucomicrobia, Proteobacteria,Bacteroidetes†Firmicutes↓	Host NLRP6 play a pathogenic role in aggravating GVHD which was independent of indigenous microbiota changes.	(105)

been proposed in earlier publications, including agglutination, opsonization, and toxin neutralization (106, 107).

Collectively, it is evident that not all bacteria have the same effect on GVHD, the composition of bacterial species and their function matter. The perturbations in the diversity and function of bacterial species may be caused by animal strains or research environment. However, one special bacterial species seems to be adverse in GVHD. Enterococci, gram-positive, facultative, anaerobic bacteria, occur in low abundance in the healthy host gut, with reports showing that Enterococcus expansion is associated with increased bloodstream infection and mortality in HSCT (108, 109). The data mentioned above (98) also revealed an increase in enterococci in the development of GVHD. In a recent study (104), in line with previous suspicion, enterococci similarly expanded and dominated the microbiota after allo-HSCT in GVHD mice; lactose drove this growth, and a lactose-free diet attenuated enterococcal expansion and T cell-driven inflammation in GVHD. Indeed, fecal domination by specific translocation of pathogenic bacteria, such as Enterococcus, is a significant risk factor for the development of aGVHD and increased overall GVHD-related mortality. Some easily overlooked factors, such as dietary elements, may play important roles in the progression, which should be closely managed.

The development of biogenetics has led to a focus on specific gene sites. For example, host NOD-like receptor family pyrin

domain-containing 6 (NLRP6) regulates microbiota-dependent protection in intestinal colitis and tumorigenesis (110–113), but Tomomi et al. found an inverse effect in GVHD, in which host NLRP6 played a pathogenic role in aggravating intestinal damage (105). Interestingly, this influence was independent of indigenous microbiota changes. Toll-like receptors (TLRs), which sense bacterial lipoproteins and LPS and DNA, are suspected to modulate innate immune responses, and a major role of TLR9-mediated sensing of bacterial DNA in the aggravation of GVHD has been reported (114–116). With gene knockout, Markus et al. similarly proved that bacterial innate immune receptor TLR-'- mice showed significantly reduced GVHD mortality, which was further confirmed by less pronounced GVHD scores over time (98).

Furthermore, some groups have demonstrated substantial differences between the gut microbiota of mice purchased from different commercial vendors or repositories (117). There are clear differences in microbiota composition and diversity between sexes in previous studies (118, 119). In addition, Floris et al. showed that BALB/c mice had higher abundance and diversity of immunoglobulin A (IgAs) than C57BL/6 mice which is correlated with increased microbiota diversity (120). And aging alters the gut microbiota in mice, in which aged microbiome leads to an exaggerated systemic inflammatory response and reduced levels of SCFAs in young mice (121). In conclusion, gut microbiota composition can be influenced by housing, gender, host genetic,

age, et al, and further studies are needed to determine the impact of different murine models and strains on aGVHD.

Human Studies

Early in the 1980s, in a clinical report of 130 patients with aplastic anemia undergoing allo-HSCT, gut decontamination and laminar airflow isolation were shown to lower the incidence of aGVHD (122). However, subsequent clinical research of human aGVHD has demonstrated that loss of intestinal microbiota diversity is associated with aGVHD, as microbiota disruption characterized by expansions of potentially pathogenic bacteria and reduction in alpha diversity (a variable that reflects the number of unique bacterial taxa present and their relative frequencies) have been reported. Some advanced clinical studies performed in recent years may further provide us with a better understanding of these alterations (**Table 3**).

In 2012, Jenq et al. studied changes in the microbiota of patients undergoing allo-HSCT and found that only the GVHD group had decreased microbial diversity with increased

Lactobacillales abundance and loss of Clostridiales, suggesting that shifts in diversity were a result of GVHD rather than allo-HSCT or antibiotic exposure (99). In 2014, Taur Y et al. found that loss of bacterial diversity in stool specimens was associated with increased mortality from GVHD, and survival at 3 years after allo-HSCT was 36%, 60%, and 67% for patients with low, intermediate, and high microbiota diversity, respectively (123). However, patients in this study received T cell-depleted grafts, which could also reduce GVHD. Jeng et al. further investigated 64 allo-HSCT recipients of T cell-replete grafts and found that a higher abundance of Blautia was associated with a reduced risk of GVHD-related mortality and increased overall survival (124). Blautia is a genus that belongs to the class Clostridia. As reported in previous studies, Clostridiales rescue intestinal epithelial cell damage by upregulating Treg cells through the production of the SCFA butyrate (59, 67, 130), and alteration of the indigenous microbiota with 17 rationally selected strains of high butyrateproducing Clostridia led to decreased GVHD (74). Consistent with these findings, in 2017, Simms et al. reported a significant

TABLE 3 | Summary of human studies investigating the association between the microbiota and GVHD in the past 10 years.

Year	Patients	Relative abundance alteration of microbiota	Outcomes	Ref
2012	18 adult patients (8 GVHD vs. 10 non-GVHD)	● In GVHD patients: Lactobacillales↑	GVHD group had decreased stool microbial diversity, microbial chaos early after transplantation is a potential risk factor for subsequent GVHD.	(99)
	,	Clostridiales↓		
2014	80 adult patients	 In GVHD patients: Enterococcus, Streptococcus, Lactobacillus↑ 	Increased mortality from GVHD was associated with lower diversity of microbiota at engraftment, which showed a strong predictive effect on mortality.	(123)
2015	115 adult patients	In GVHD patients:Blautia.	Increased abundance of commensal bacteria belonging to the Blautia genus is associated with reduced lethal GVHD and improved OS.	(124)
2017	29 pediatric patients	 In GVHD patients: Enterobacteriaceae, Enterococcus↑ In non-GVHD patients: anti-inflammatory Clostridia(AIC), Bacteroidetes, Bifidobacterium ↑ 	Exposure to antianaerobic antibiotics clindamycin lead to depletion of Clostridia species which is associated with GVHD in pediatric HSCT patients.	(125)
2017	66 adult patients (52 GVHD vs. 14 non-GVHD)	● In GVHD patients: oral Actinobacteria, oral Firmicutes↑ Lachnospiraceae↓	The stool microbiota at neutrophil recovery post-HSCT is predictive of subsequent development of aGVHD.	(126)
2018	81 adult patients (32 GVHD vs. 49 non-GVHD)	● In GVHD patients: Enterobacteriaceae↑ Lachnospiraceae, Ruminococcaceae↓	Intestinal microbiota might induce aGVHD by influencing the Treg/Th17 balance.	(68)
2019	141 adult patients (83 grade 0-I aGVHD vs. 58 grade II-IV aGVHD)	 ■ In GVHD patients: Proteobacteria, Gammaproteobacteria, Enterobacteriaceae† Firmicutes, Clostridia, Lachnospiraceae, Peptostreptococcaceae, Erysipelotrichaceae, Blautia, Lachnoclostridium, 	GVHD group had lower diversity of microbiota. The AIM score defined as microbiota diversity of 4 bacterials (Lachnospiraceae, Peptostreptococcaceae, Erysipelotrichaceae, Enterobacteriaceae) was positively correlated with aGVHD grade and could be predictive of the development of aGVHD.	(127)
2019	1325 adult male	Erysipelatoclostridium, Eubacterium↓ In GVHD patients: enterococcal↑	Expansion of enterococcal was associated with GVHD and mortality which can be driven by lactose.	(104)
2020	70 patients (35GVHD vs. 35 non-GVHD)	● In GVHD patients: Lachnospiraceae, Blautia, Ruminococcaceae↓	Microbiota alterations were highly specific of GI aGVHD severity with lower bacterial biomass, a-diversity and decreased butyrate.	(128)
2020	1362 adult patients from 4 centers	■ In GVHD patients: Enterococcus, Klebsiella, Escherichia, Staphylococcus, Streptococcus↑	Patterns of microbiota disruption during allo-HSCT were similar across transplantation centers and geographic locations which were characterized by loss of diversity and domination by single taxa, lower diversity was associated with higher risks of TRM and death attributable to GVHD.	(129)

OS, overall survival; AIM score, accumulated intestinal microbiota score; TRM, transplant-related mortality.

decline in anti-inflammatory Clostridia in pediatric patients with aGVHD (125). Therefore, it can be speculated that some microbial taxa, such as Blautia, are beneficial for the outcomes of HSCT and mitigation of aGVHD, as they behave as drivers in this process, which should be protected and used in a probiotic approach. By contrast, the Enterococcus genus from Lactobacillales contributes to inflammation, whose role in human aGVHD is the same as that in animals, as expansion of Enterococcus association with increased GVHD in humans has been reported (104, 131). Lactobacillus, another genus of Lactobacillales, showed a possible protective effect in human GVHD (99). In 2018, Lijie Han et al. found that GVHD patients showed a higher abundance of Proteobacteria and a lower abundance of Clostridia, which was correlated with the Treg/ Th17 ratio and H3 acetylation, indicating an interaction among alterations in the microbiota, allogenic T cell activation and histone acetylation (68). One year later, this team (127) proved again that in aGVHD, the diversity of the microbiota was significantly lower, with decreases in Clostridia, Lachnospiraceae, Blautia, Eubacterium, and Erysipelatoclostridium abundance and increases in Enterobacteriaceae abundance. This finding was consistent with a study in 2017 showing a persistent lack of Lachnospiraceae and Bacteroidaceae species in GVHD patients, whereas Lachnospiraceae was negatively correlated with neutrophil recovery (126). In addition, the specific actors in the intestinal ecosystem involved in the pathologic process of aGVHD have been explored more recently (128). Shown in stool samples, microbiota alterations were highly specific to gastrointestinal aGVHD severity, and a negative correlation was observed with the Lachnospiraceae, especially the Blautia genus, and Ruminococcaceae families. On the other hand, geographic variations matter, while a recent study analyzing 8767 fecal samples from 1362 patients with allo-HSCT at 4 different centers likewise showed a similar association between lower intestinal diversity and higher risks of transplantationrelated death and death attributable to GVHD (129).

From these data, we conclude that restoring intestinal microbiota diversity after allo-HSCT is beneficial in the clinic, protective indigenous probiotics should be preserved to balance the alteration of intestinal microbiota community for patients.

CLINICAL INTERVENTIONS AND VALUE OF THE INTESTINAL MICROBIOTA IN AGVHD

Diet

Certain diets may contribute to the development of GVHD given that food is one of the most important factors affecting the composition of the intestinal microbiota (132). One example is a choline diet, and a murine model has shown that a high-choline diet enhances the allogenic GVH reaction, which leads to more aGVHD (86). The effect of parenteral or enteral nutrition on the intestinal ecosystem during HSCT has also been evaluated, with preference toward the latter choice, as enteral feeding has been shown to protect against GVHD in several studies (133–135),

while parenteral nutrition was associated with poor outcomes and other complications (136, 137). The type of oral nutrition may be another important factor. Recently, elaborate foods known as a neutropenic diet for allo-HSCT patients have been reexamined, and advanced evidence has shown limited benefit and even potentially harm of supplying aGVHD patients with neutropenic diets (138, 139).

Prebiotics and Probiotics

Intervention in the intestinal microbiota with a nutritional approach including prebiotics and probiotics may be another promising treatment option for aGVHD.

Prebiotics are indigestible compounds, usually indigestible carbohydrates, that bacteria have an advantage in metabolizing, resulting in the production of SCFAs and metabolites with a potential immunomodulatory role (140, 141). Strategies have been studied in the setting of aGVHD to modulate the intestinal microbiota by supplementation with inulin, oligosaccharides, galacto-oligosaccharides, and potato starch, showing beneficial results. In a recent study, Yoshifuji et al. found that intake of resistant starch and GFO (glutamine, fiber, and oligosaccharide) shortened the duration of oral mucositis and diarrhea and reduced the incidence and severity of aGVHD (142). Other clinical trials focused on fructooligosaccharide, potato-based starch, and gluten-free diets are currently being studied for potential benefit (143).

Probiotics are ingestible formulations of live bacteria that can modulate intestinal homeostasis. A probiotic strategy achieved by FMT consists of introducing one strain or selected strains of microorganisms that confer a benefit. An early report showed that a probiotic-rich diet prior to HSCT is associated with earlier neutrophil engraftment and a shorter duration of febrile neutropenia (144). Some probiotics were found to be safe to administer during aGVHD, such as *Lactobacillus plantarum* reported in pediatric patients (145) and *Lactobacillus rhamnosus* GG in murine models (146). However, controversies exist. Recently, there have been some concerns regarding the safety of administering living microorganisms to immunocompromised patients with altered gut permeability, as some clinical cases have demonstrated sepsis (147), bacteremia (148) and meningitis (149) after treatment of pediatric patients.

Antibiotics

Given that the intestinal microbiota critically affects transplant outcomes, correctly managing the influence of the microbiota in GVHD—antibiotics has been encouraged. The advent of techniques to generate and maintain germ-free rodents since the 1940s made it possible to examine the microbiota in animals (123), and subsequent studies demonstrated the benefits of using antibiotics. Bekkum and Jones et al. found that germ-free mice housed in sterile conditions or mice treated with antibiotics developed less severe aGVHD (94, 95). Vossen et al. showed that in a cohort of 112 pediatric patients, recipients treated with total gastrointestinal decontamination (GID) using high doses of nonabsorbable antibiotics prevented moderate-to-severe aGVHD, suggesting that the translocation of luminal bacteria and their cell wall-derived compounds might be inhibited during

total GID (150). Weber et al. analyzed 394 patients receiving allo-HSCT and found that the treatment of rifaximin correlated with lower enterococcal positivity and higher urinary 3-indoxyl sulfate concentrations. Patients on rifaximin showed lower 1-year transplant-related mortality and higher overall survival. Infectious complications with systemic antibiotic use did not abrogate the beneficial effects of rifaximin on intestinal microbiota composition in the early course of allo-HSCT or outcome (151).

However, the use of gut-decontamination prophylactic antibiotics seems to be a doubled-edged sword. A retrospective analysis in 2018 mentioned above (68) proved that β-lactam antibiotic administration was an independent risk factor for aGVHD according to the Cox regression model for multivariate analysis of aGVHD. Although showing no significant adverse effect, in an allo-BMT murine model, treatment of both donor mice with broad-spectrum antibiotics and control SPF donor mice induced GVHD mortality at a similar rate, and reducing and altering the microbial flora in the donors had no effect on their T cell alloreactivity and induction of GVHD after allogeneic BMT (152). In human studies, the concept that gut decontamination prevents aGVHD is controversial given that some subsequent clinical trials have failed to demonstrate consistent benefits (68, 153-155). Prophylactic use of antibiotics is reported to be associated with more severe aGVHD that involves the intestinal tract and liver, impacting 1-y and 2-y overall survival (OS) in patients receiving myeloablative regimens (155). Earlier antibiotic treatment in patients prior to allo-HSCT was further associated with higher transplant-related mortality than no antibiotics (76). A potential role for anaerobic bacteria, in particular Clostridiales, was supported that Blautia abundance was inversely correlated with the risk of developing gastrointestinal GVHD (124), while clindamycin, an anti-anaerobic bacterial agent, increased the risk of GVHD by depleting anti-inflammatory clostridia (125). Reported previously, piperacillin-tazobactam and imipenemcilastatin increased the incidence, severity, and mortality of GVHD, and piperacillin-tazobactam reduced Bacteroidetes and Lactobacillus abundance (102). Subsequent evidence further supports the adverse role of anti-anaerobic bacterial penicillin derivatives and carbapenems, as both are associated with a higher incidence of GVHD (156). Although seemingly negative, these findings may enlighten us that antibiotics preserving anaerobic bacteria may potentially reduce the risk of developing gut GVHD.

In conclusion, it is obvious that there remain many controversies for GVHD patients to undergo antibiotic therapy, which needs further exploration. Thus, it is of great necessity to identify new strategies to maintain the diversity and richness of the intestinal microbiota. Different attempts have been made in clinical practice involving narrow-spectrum antibiotics and modulating the timing and duration of treatment.

FMT

Loss of microbiota diversity creates an opportunity to intervene in aGVHD by reestablishing diversity using microbes that modulate inflammation. FMT has been investigated as a potential therapeutic strategy for gut GVHD in both preventive and therapeutic strategies in recent years. The recent insight of FMT considers it the 'ultimate probiotic' for GVHD in allo-HSCT because it directly modifies the host's intestinal microbiome composition to restore eubiosis and gut homeostasis (157). Indeed, according to some previous reports, FMT significantly improves the outcome of GVHD. Kakihana et al. reported that 4 patients received FMT 92 days after HSCT, all patients responded to FMT within several days, with 3 complete responses and 1 partial response, and an increase in peripheral effector regulatory T cells during the response to FMT was also observed (158). Similarly, another study reported that two of three patients achieved a complete response with multiple FMTs, while the other obtained a partial response; the FMT response was correlated with increased microbial diversity and richness (159). More recent reports further illustrate the role of FMT in GVHD, with promising results (160-162). Currently, a number of ongoing clinical trials to investigate the role of FMT in preventing intestinal GVHD following allo-HSCT, summarized in Table 4, are being carried out to try to find the best treatment protocol.

Although many reports have shown that FMT is a safe and effective strategy used in different situations to modulate the microbiota and cure aGVHD, it should be noted that patients are usually immunocompromised with altered intestinal permeability, as infectious complications after FMT have been reported in other settings at the same time (163, 164). Moreover, the evidence mentioned above suggests a beneficial role of FMT in GVHD and needs to be confirmed in larger studies. The optimization of all practical aspects of FMT still needs to be addressed in the future.

Predictive Marker

As reported in the current literature, a reduction in microbiota diversity and alteration of metabolites can predict transplantation outcomes and aGVHD, shedding light on the value of the microbiota. In 2015, Weber et al. detected 3-indoxyl sulfate (3-IS) in urine specimens within the first 28 days after allo-HSCT in 131 patients and found that a low level of 3-IS within the first 10 days was associated with significantly higher transplant-related mortality and worse overall survival 1 year after allo-HSCT. Furthermore, the class Bacilli was proven to be associated with low 3-IS levels (75). In 2017, Golob et al. showed that a gradient of 20 types of bacterial species could predict severe aGVHD by calculating a gradient of the sum of the relative abundance of positively correlated bacteria minus the sum of the relative abundance of negative correlates (126). Similarly, a study in 2019 by Lijie Han et al. showed that microbiota diversity combined with the gradients of 4 bacteria (Peptostreptococcaceae, Erysipelotrichaceae, Enterobacteriaceae, and Lachnospiraceae) can predict the development and severity of aGVHD (127). Geographic variations in the composition of human microbial communities and differences in clinical practices across institutions raise the question of whether relationships between microbiota composition and clinical outcomes after allo-HSCT are generalizable. In 2020, Peled et al. conducted a study with data from four clinical research institutions by comparing risk scores from regularized Cox regression with cross-validation; they

TABLE 4 | Summary of ongoing microbiota-linked (not)recruiting clinical trials for GVHD.

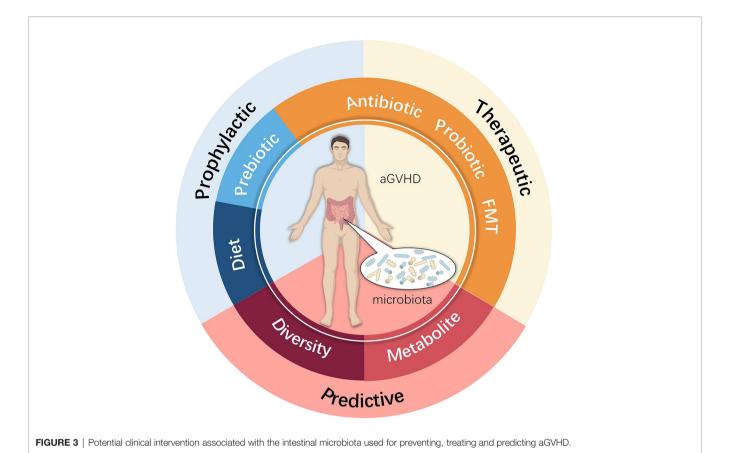
Number	Trail title	Interventions	Aims	Study design	Phase	Patients	Time
NCT03819803	Fecal microbiota transplantation in aGVHD after ASCT	FMT	To explore the employment of FMT in GI-aGVHD.	Single group assignment Open label	III	15	2017/ 3/1- 2020/
NCT04285424	FMT for steroid resistant gut acute GVHD	FMT	To evaluate safety and efficacy of FMT for the treatment of steroid resistant GVHD of the gut.	Single group assignment Open label	Early I	30	12/31 2020/ 3/1- 2022/ 3/1
NCT03812705	Fecal microbiota transplantation for steroid resistant/dependent acute GI GVHD	FMT	To evaluate the efficacy and safety of fecal microbiome transplantation in patients with steroid resistant/dependent acute gastrointestinal GVHD.	Single group assignment Open label	II	30	2018/ 12/ 13- 2022/ 12/31
NCT04269850	Fecal microbiota transplantation with Ruxolitinib and Steroids as an upfront treatment of severe acute intestinal GVHD	FMT; Ruxolitinib; Methylprednisone	To evaluate this combination treatment in the first line with FMT.	Single group assignment Open label	I/II	20	2019/ 9/1- 2023/ 9/1
NCT03371667	To compare the efficacy of the addition of Methotrexate (MTX) to current standard acute GVHD first-line treatment with corticosteroids	Methotrexate	To compare the efficacy of the addition of MTX to current standard acute GVHD first-line treatment with corticosteroids.	Randomized Multicenter Double blinded Parallel assignment	III	102	2018/ 8/16- 2021/ 9
NCT03727113	Optimization of antibiotic treatment in hematopoietic stem cell receptors	Antibiotics	To demonstrate that in ASCT receptors a predefined protocol of optimization of the antibacterial treatment will preserve the intestinal microbiota diversity which will correlate with decrease incidence of acute GVHD.	Observational Case-Control Prospective	Unknown	180	2018/ 1/16- 2020/ 5/31
NCT03727113	Choosing the Best Antibiotic to Protect Friendly Gut Bacteria During the Course of Stem Cell Transplant	Piperacillin- tazobactam	To compare the effects of different antibiotics on the community of friendly bacteria in the gut	Randomized Parallel Assignment Open Label	II	144	2017/ 2/10- 2021/ 2
(not R) NCT04059757	Fecal microbiota transplantation for the treatment of gastrointestinal acute GVHD	FMT	To see the efficacy and what side effects are seen with FMT as a treatment for GVHD.	Single group assignment Open label	II	17	2020/ 7/1- 2021/ 12
NCT04280471	Fecal microbiota transplantation for the treatment of severe acute gut Graft-Versus-Host Disease	FMT capsule	To explore side effects of using an investigational procedure (FMT) in treating patients with severe acute gut GVHD.	Single group assignment Open label	I	10	2020/ 6/30- 2022/ 7/1
NCT04139577	FMT In high-risk acute GVHD after allo-HCT	FMT	To evaluate the effectiveness of Fecal Microbiota Transplant (FMT) treatment in high-risk acute GVHD.	Single group assignment Open label	I	11	2019/ 11- 2023/
NCT03549676	Fecal microbiota Transplantation for Treatment of Refractory Graft Versus Host Disease-a Pilot Study	FMT	To evaluate safety and efficacy of FMT for the treatment of refractory GVHD of the gut.	Single group assignment Open label	I	15	5/1 2019/ 7/1- 2020/
NCT03359980	Treatment of steroid refractory gastro-intestinal acute GVHD after allogeneic HSCT With fecal microbiota transfer	FMT	To explore the employment of FMT in GlaGVHD.	Single group assignment Open label	II	32	12/31 2018/ 8/13- 2020/ 12

FMT, fecal microbiota transplantation; GI-aGVHD, gastrointestinal acute graft-versus-host disease.

showed that not only a diversity metric but also a signature of specific bacterial abundances was informative about the risk of death after allo-HSCT across institutions (129). Similarly, another study analyzing data from stool and blood samples of 150 patients from two centers who underwent allo-HSCT also showed that gut microbiota score (GMS) from a LASSO (least absolute shrinkage

and selection operator) model at neutrophil engraftment could predict aGVHD (165).

Thus, studies on the microbiota as a predictive marker for aGVHD are worth further exploration to provide assistance with the currently available tools for predicting the development of aGVHD.



CONCLUSION AND PROSPECTS

Alteration of the intestinal microbiota is a corollary given that the gut is an aGVHD target organ. Although we could further explore the intestinal microbiota by better dissecting compositional and functional microbiota structures, most of the research has concentrated on characterization, with data analyzed only *via* correlation. In this regard, determining which specific bacterial taxa are the main taxa affecting aGVHD is of great urgency and importance. In addition to descriptive investigations, mechanistic investigations are needed, which will help translate these associations into therapies for aGVHD if the microbiota is causal.

New approaches to prevent and cure aGVHD remain an unmet need that can be best addressed by understanding the complex pathophysiology, with increasing evidence indicating that the intestinal microbiota indeed participates in this process. Future studies are essential to explore further the role of the intestinal microbiota in aGVHD to elucidate the real impact of microbiota ecology. A promising approach may involve altering certain microbiota species by more precise and safer methods, which may consist of diet, prebiotics, probiotics, advanced antibiotic therapies, FMT, and microbiota metabolism analyses (Figure 3). Fully understanding the mechanism by which loss of microbiota diversity influences specific molecules and pathways and regulates the pathogenesis and development of aGVHD

remains a top concern. Only by attempting to find better prophylactic and therapeutic schemes for allo-HSCT complications can we focus on what truly matters, which is also the appeal and value of the Human Microbiome Project and precision medicine.

AUTHOR CONTRIBUTIONS

The manuscript was conceptualized by XZ and QG. TH wrote the majority of the manuscript. RW and XW co-wrote the manuscript. The figures were designed by TH and drawn by XW and SY. WW, RW, and TH summarized the tables. All authors contributed to the article and approved the submitted version.

FUNDING

This work was supported by The 2020 Open Project (Key Project) of the National Center for Clinical Medicine Research on Hematological System Diseases (Grant No. 2020ZKZC02), National Key Research and Development Plan "Stem Cell and Transformation Research" Key Special Project (grant no. 2017YFA0105502), and Science and Health Joint Project of Chongqing (grant no. 2018QNXM015).

REFERENCES

- Ferrara JLM, Levine JE, Reddy P, Holler E. Graft-versus-host disease. Lancet (2009) 373(9674):1550–61. doi: 10.1016/S0140-6736(09)60237-3
- Blazar BR, Murphy WJ, Abedi M. Advances in graft-versus-host disease biology and therapy. Nat Rev Immunol (2012) 12(6):443–58. doi: 10.1038/ nri3212
- Ramachandran V, Kolli SS, Strowd LC. Review of Graft-Versus-Host Disease. Dermatol Clin (2019) 37(4):569–82. doi: 10.1016/j.det.2019.05.014
- Jagasia M, Arora M, Flowers ME, Chao NJ, McCarthy PL, Cutler CS, et al. Risk factors for acute GVHD and survival after hematopoietic cell transplantation. *Blood* (2012) 119(1):296–307. doi: 10.1182/blood-2011-06-364265
- Atkinson K, Horowitz MM, Gale RP, van Bekkum DW, Gluckman E, Good RA, et al. Risk factors for chronic graft-versus-host disease after HLAidentical sibling bone marrow transplantation. *Blood* (1990) 75(12):2459– 64. doi: 10.1182/blood.V75.12.2459.bloodjournal75122459
- Zahid MF, Lazarus HM, Ringden O, Barrett JA, Gale RP, Hashmi SK. Can we prevent or treat graft-versus-host disease with cellular-therapy? *Blood Rev* (2020) 43:100669. doi: 10.1016/j.blre.2020.100669
- Sung AD, Chao NJ. Concise review: acute graft-versus-host disease: immunobiology, prevention, and treatment. Stem Cells Transl Med (2013) 2(1):25–32. doi: 10.5966/sctm.2012-0115
- Barton-Burke M, Dwinell DM, Kafkas L, Lavalley C, Sands H, Proctor C, et al. Graft-versus-host disease: a complex long-term side effect of hematopoietic stem cell transplant. Oncol (Williston Park) (2008) 22(11 Suppl Nurse Ed):31–45.
- Hashmi S, Ahmed M, Murad MH, Litzow MR, Adams RH, Ball LM, et al. Survival after mesenchymal stromal cell therapy in steroid-refractory acute graft-versus-host disease: systematic review and meta-analysis. *Lancet Haematol* (2016) 3(1):e45–52. doi: 10.1016/S2352-3026(15)00224-0
- Martin PJ, Inamoto Y, Flowers ME, Carpenter PA. Secondary treatment of acute graft-versus-host disease: a critical review. *Biol Blood Marrow Transpl* (2012) 18(7):982–8. doi: 10.1016/j.bbmt.2012.04.006
- 11. Schroeder MA, Khoury HJ, Jagasia M, Ali H, Schiller GJ, Staser K, et al. A phase 1 trial of itacitinib, a selective JAK1 inhibitor, in patients with acute graft-versus-host disease. *Blood Adv* (2020) 4(8):1656–69. doi: 10.1182/bloodadvances.2019001043
- Malard F, Huang XJ, Sim JPY. Treatment and unmet needs in steroidrefractory acute graft-versus-host disease. *Leukemia* (2020) 34(5):1229–40. doi: 10.1038/s41375-020-0804-2
- Penack O, Marchetti M, Ruutu T, Aljurf M, Bacigalupo A, Bonifazi F, et al. Prophylaxis and management of graft versus host disease after stem-cell transplantation for haematological malignancies: updated consensus recommendations of the European Society for Blood and Marrow Transplantation. *Lancet Haematol* (2020) 7(2):e157–e67. doi: 10.1016/ S2352-3026(19)30256-X
- Maynard CL, Elson CO, Hatton RD, Weaver CT. Reciprocal interactions of the intestinal microbiota and immune system. *Nature* (2012) 489 (7415):231–41. doi: 10.1038/nature11551
- Turnbaugh PJ, Ley RE, Mahowald MA, Magrini V, Mardis ER, Gordon JI. An obesity-associated gut microbiome with increased capacity for energy harvest. Nature (2006) 444(7122):1027–31. doi: 10.1038/nature05414
- Suez J, Korem T, Zeevi D, Zilberman-Schapira G, Thaiss CA, Maza O, et al. Artificial sweeteners induce glucose intolerance by altering the gut microbiota. Nature (2014) 514(7521):181–6. doi: 10.1038/nature13793
- Collins SM. A role for the gut microbiota in IBS. Nat Rev Gastroenterol Hepatol (2014) 11(8):497–505. doi: 10.1038/nrgastro.2014.40
- Hsiao EY, McBride SW, Hsien S, Sharon G, Hyde ER, McCue T, et al. Microbiota modulate behavioral and physiological abnormalities associated with neurodevelopmental disorders. *Cell* (2013) 155(7):1451–63. doi: 10.1016/j.cell.2013.11.024
- Luissint AC, Parkos CA, Nusrat A. Inflammation and the Intestinal Barrier: Leukocyte-Epithelial Cell Interactions, Cell Junction Remodeling, and Mucosal Repair. Gastroenterology (2016) 151(4):616–32. doi: 10.1053/ j.gastro.2016.07.008
- Peled JU, Hanash AM, Jenq RR. Role of the intestinal mucosa in acute gastrointestinal GVHD. *Blood* (2016) 128(20):2395–402. doi: 10.1182/blood-2016-06-716738

- Shono Y, van den Brink MRM. Gut microbiota injury in allogeneic haematopoietic stem cell transplantation. Nat Rev Cancer (2018) 18 (5):283–95. doi: 10.1038/nrc.2018.10
- Turnbaugh PJ, Ley RE, Hamady M, Fraser-Liggett CM, Knight R, Gordon JI. The human microbiome project. *Nature* (2007) 449(7164):804–10. doi: 10.1038/nature06244
- Murphy S, Nguyen VH. Role of gut microbiota in graft-versus-host disease.
 Leuk Lymphoma (2011) 52(10):1844–56. doi: 10.3109/10428194.2011.580476
- Ott SJ, Kuhbacher T, Musfeldt M, Rosenstiel P, Hellmig S, Rehman A, et al. Fungi and inflammatory bowel diseases: Alterations of composition and diversity. Scand J Gastroenterol (2008) 43(7):831–41. doi: 10.1080/ 00365520801935434
- Qin J, Li R, Raes J, Arumugam M, Burgdorf KS, Manichanh C, et al. A human gut microbial gene catalogue established by metagenomic sequencing. *Nature* (2010) 464(7285):59–65. doi: 10.1038/nature08821
- Reyes A, Haynes M, Hanson N, Angly FE, Heath AC, Rohwer F, et al. Viruses in the faecal microbiota of monozygotic twins and their mothers. Nature (2010) 466(7304):334–8. doi: 10.1038/nature09199
- Shono Y, Docampo MD, Peled JU, Perobelli SM, Jenq RR. Intestinal microbiota-related effects on graft-versus-host disease. *Int J Hematol* (2015) 101(5):428–37. doi: 10.1007/s12185-015-1781-5
- Rodriguez J, Hiel S, Neyrinck AM, Le Roy T, Potgens SA, Leyrolle Q, et al. Discovery of the gut microbial signature driving the efficacy of prebiotic intervention in obese patients. *Gut* (2020) 69(11):1975–87. doi: 10.1136/ gutjnl-2019-319726
- Thingholm LB, Ruhlemann MC, Koch M, Fuqua B, Laucke G, Boehm R, et al. Obese Individuals with and without Type 2 Diabetes Show Different Gut Microbial Functional Capacity and Composition. *Cell Host Microbe* (2019) 26(2):252–64 e10. doi: 10.1016/j.chom.2019.07.004
- Leiva-Gea I, Sanchez-Alcoholado L, Martin-Tejedor B, Castellano-Castillo D, Moreno-Indias I, Urda-Cardona A, et al. Gut Microbiota Differs in Composition and Functionality Between Children With Type 1 Diabetes and MODY2 and Healthy Control Subjects: A Case-Control Study. *Diabetes Care* (2018) 41(11):2385–95. doi: 10.2337/dc18-0253
- 31. Hod K, Ringel Y. Probiotics in functional bowel disorders. *Best Pract Res Clin Gastroenterol* (2016) 30(1):89–97. doi: 10.1016/j.bpg.2016.01.003
- Strati F, Cavalieri D, Albanese D, De Felice C, Donati C, Hayek J, et al. New evidences on the altered gut microbiota in autism spectrum disorders. *Microbiome* (2017) 5(1):24. doi: 10.1186/s40168-017-0242-1
- Pan H, Guo R, Ju Y, Wang Q, Zhu J, Xie Y, et al. A single bacterium restores
 the microbiome dysbiosis to protect bones from destruction in a rat model of
 rheumatoid arthritis. *Microbiome* (2019) 7(1):107. doi: 10.1186/s40168-0190719-1
- Kumari R, Palaniyandi S, Hildebrandt GC. Microbiome: An Emerging New Frontier in Graft-Versus-Host Disease. *Dig Dis Sci* (2019) 64(3):669–77. doi: 10.1007/s10620-018-5369-9
- 35. Wang WL, Xu SY, Ren ZG, Tao L, Jiang JW, Zheng SS. Application of metagenomics in the human gut microbiome. *World J Gastroenterol* (2015) 21(3):803–14. doi: 10.3748/wjg.v21.i3.803
- Morgan XC, Huttenhower C. Meta'omic analytic techniques for studying the intestinal microbiome. *Gastroenterology* (2014) 146(6):1437–48.e1. doi: 10.1053/j.gastro.2014.01.049
- Leibowitz BJ, Wei L, Zhang L, Ping X, Epperly M, Greenberger J, et al. Ionizing irradiation induces acute haematopoietic syndrome and gastrointestinal syndrome independently in mice. *Nat Commun* (2014) 5:3494. doi: 10.1038/ncomms4494
- Ch'ang HJ, Maj JG, Paris F, Xing HR, Zhang J, Truman JP, et al. ATM regulates target switching to escalating doses of radiation in the intestines. Nat Med (2005) 11(5):484–90. doi: 10.1038/nm1237
- Powell DW, Pinchuk IV, Saada JI, Chen X, Mifflin RC. Mesenchymal cells of the intestinal lamina propria. Annu Rev Physiol (2011) 73:213–37. doi: 10.1146/annurev.physiol.70.113006.100646
- Wang X, Wei L, Cramer JM, Leibowitz BJ, Judge C, Epperly M, et al. Pharmacologically blocking p53-dependent apoptosis protects intestinal stem cells and mice from radiation. Sci Rep (2015) 5:8566. doi: 10.1038/srep08566
- Rannou E, Francois A, Toullec A, Guipaud O, Buard V, Tarlet G, et al. In vivo evidence for an endothelium-dependent mechanism in radiationinduced normal tissue injury. Sci Rep (2015) 5:15738. doi: 10.1038/srep15738

- 42. Epstein RJ, McDonald GB, Sale GE, Shulman HM, Thomas ED. The diagnostic accuracy of the rectal biopsy in acute graft-versus-host disease: a prospective study of thirteen patients. *Gastroenterology* (1980) 78(4):764–71. doi: 10.1016/0016-5085(80)90681-2
- Wall AJ, Rosenberg JL, Reilly RW. Small intestinal injury in the immunologically runted mouse. Morphologic and autoradiographic studies. J Lab Clin Med (1971) 78(5):833–4.
- Sale GE, Shulman HM, McDonald GB, Thomas ED. Gastrointestinal graftversus-host disease in man. A clinicopathologic study of the rectal biopsy. *Am J Surg Pathol* (1979) 3(4):291–9. doi: 10.1097/00000478-197908000-00001
- Penack O, Henke E, Suh D, King CG, Smith OM, Na IK, et al. Inhibition of neovascularization to simultaneously ameliorate graft-vs-host disease and decrease tumor growth. *J Natl Cancer Inst* (2010) 102(12):894–908. doi: 10.1093/jnci/djq172
- Lai XY, Egan LJ. Suppression of radiation-induced DNA double-strand break repair by MyD88 is accompanied by apoptosis and crypt loss in mouse colon. Oncogenesis (2013) 2:e62. doi: 10.1038/oncsis.2013.22
- Rakoff-Nahoum S, Paglino J, Eslami-Varzaneh F, Edberg S, Medzhitov R. Recognition of commensal microflora by toll-like receptors is required for intestinal homeostasis. *Cell* (2004) 118(2):229–41. doi: 10.1016/ i.cell.2004.07.002
- Beura LK, Hamilton SE, Bi K, Schenkel JM, Odumade OA, Casey KA, et al. Normalizing the environment recapitulates adult human immune traits in laboratory mice. *Nature* (2016) 532(7600):512–6. doi: 10.1038/nature17655
- Berlin C, Bargatze RF, Campbell JJ, von Andrian UH, Szabo MC, Hasslen SR, et al. alpha 4 integrins mediate lymphocyte attachment and rolling under physiologic flow. *Cell* (1995) 80(3):413–22. doi: 10.1016/0092-8674(95) 90491-3
- 50. Butcher EC, Picker LJ. Lymphocyte homing and homeostasis. *Science* (1996) 272(5258):60–6. doi: 10.1126/science.272.5258.60
- Johansson-Lindbom B, Agace WW. Generation of gut-homing T cells and their localization to the small intestinal mucosa. *Immunol Rev* (2007) 215:226–42. doi: 10.1111/j.1600-065X.2006.00482.x
- Masopust D, Schenkel JM. The integration of T cell migration, differentiation and function. Nat Rev Immunol (2013) 13(5):309–20. doi: 10.1038/nri3442
- Mora JR, Bono MR, Manjunath N, Weninger W, Cavanagh LL, Rosemblatt M, et al. Selective imprinting of gut-homing T cells by Peyer's patch dendritic cells. *Nature* (2003) 424(6944):88–93. doi: 10.1038/nature01726
- Fu YY, Egorova A, Sobieski C, Kuttiyara J, Calafiore M, Takashima S, et al. T Cell Recruitment to the Intestinal Stem Cell Compartment Drives Immune-Mediated Intestinal Damage after Allogeneic Transplantation. *Immunity* (2019) 51(1):90–103 e3. doi: 10.1016/j.immuni.2019.06.003
- 55. Shih VF, Cox J, Kljavin NM, Dengler HS, Reichelt M, Kumar P, et al. Homeostatic IL-23 receptor signaling limits Th17 response through IL-22-mediated containment of commensal microbiota. *Proc Natl Acad Sci U S A* (2014) 111(38):13942–7. doi: 10.1073/pnas.1323852111
- Sano T, Huang W, Hall JA, Yang Y, Chen A, Gavzy SJ, et al. An IL-23R/IL-22 Circuit Regulates Epithelial Serum Amyloid A to Promote Local Effector Th17 Responses. Cell (2015) 163(2):381–93. doi: 10.1016/j.cell.2015.08.061
- Varelias A, Ormerod KL, Bunting MD, Koyama M, Gartlan KH, Kuns RD, et al. Acute graft-versus-host disease is regulated by an IL-17-sensitive microbiome. *Blood* (2017) 129(15):2172–85. doi: 10.1182/blood-2016-08-732628
- Round JL, Mazmanian SK. Inducible Foxp3+ regulatory T-cell development by a commensal bacterium of the intestinal microbiota. *Proc Natl Acad Sci U S A* (2010) 107(27):12204–9. doi: 10.1073/pnas.0909122107
- Furusawa Y, Obata Y, Fukuda S, Endo TA, Nakato G, Takahashi D, et al. Commensal microbe-derived butyrate induces the differentiation of colonic regulatory T cells. *Nature* (2013) 504(7480):446–50. doi: 10.1038/ nature12721
- Artis D, Spits H. The biology of innate lymphoid cells. *Nature* (2015) 517 (7534):293–301. doi: 10.1038/nature14189
- Cella M, Fuchs A, Vermi W, Facchetti F, Otero K, Lennerz JK, et al. A human natural killer cell subset provides an innate source of IL-22 for mucosal immunity. *Nature* (2009) 457(7230):722–5. doi: 10.1038/ nature07537

- Pickert G, Neufert C, Leppkes M, Zheng Y, Wittkopf N, Warntjen M, et al. STAT3 links IL-22 signaling in intestinal epithelial cells to mucosal wound healing. J Exp Med (2009) 206(7):1465–72. doi: 10.1084/jem.20082683
- Hanash AM, Dudakov JA, Hua G, O'Connor MH, Young LF, Singer NV, et al. Interleukin-22 protects intestinal stem cells from immune-mediated tissue damage and regulates sensitivity to graft versus host disease. *Immunity* (2012) 37(2):339–50. doi: 10.1016/j.immuni.2012.05.028
- Konya V, Mjosberg J. Innate lymphoid cells in graft-versus-host disease. Am J Transpl (2015) 15(11):2795–801. doi: 10.1111/ajt.13394
- Mortha A, Chudnovskiy A, Hashimoto D, Bogunovic M, Spencer SP, Belkaid Y, et al. Microbiota-dependent crosstalk between macrophages and ILC3 promotes intestinal homeostasis. Science (2014) 343 (6178):1249288. doi: 10.1126/science.1249288
- Teshima T, Reddy P, Zeiser R. Acute Graft-versus-Host Disease: Novel Biological Insights. Biol Blood Marrow Transpl (2016) 22(1):11–6. doi: 10.1016/j.bbmt.2015.10.001
- Atarashi K, Tanoue T, Oshima K, Suda W, Nagano Y, Nishikawa H, et al. Treg induction by a rationally selected mixture of Clostridia strains from the human microbiota. *Nature* (2013) 500(7461):232–6. doi: 10.1038/ nature12331
- Han L, Jin H, Zhou L, Zhang X, Fan Z, Dai M, et al. Intestinal Microbiota at Engraftment Influence Acute Graft-Versus-Host Disease via the Treg/Th17 Balance in Allo-HSCT Recipients. Front Immunol (2018) 9:669. doi: 10.3389/fimmu.2018.00669
- Michonneau D, Latis E, Curis E, Dubouchet L, Ramamoorthy S, Ingram B, et al. Metabolomics analysis of human acute graft-versus-host disease reveals changes in host and microbiota-derived metabolites. *Nat Commun* (2019) 10 (1):5695. doi: 10.1038/s41467-019-13498-3
- Rios-Covian D, Ruas-Madiedo P, Margolles A, Gueimonde M, de Los Reyes-Gavilan CG, Salazar N. Intestinal Short Chain Fatty Acids and their Link with Diet and Human Health. Front Microbiol (2016) 7:185. doi: 10.3389/fmicb.2016.00185
- Koh A, De Vadder F, Kovatcheva-Datchary P, Backhed F. From Dietary Fiber to Host Physiology: Short-Chain Fatty Acids as Key Bacterial Metabolites. Cell (2016) 165(6):1332–45. doi: 10.1016/j.cell.2016.05.041
- Fujiwara H, Docampo MD, Riwes M, Peltier D, Toubai T, Henig I, et al. Microbial metabolite sensor GPR43 controls severity of experimental GVHD. Nat Commun (2018) 9(1):3674. doi: 10.1038/s41467-018-06048-w
- Scott NA, Andrusaite A, Andersen P, Lawson M, Alcon-Giner C, Leclaire C, et al. Antibiotics induce sustained dysregulation of intestinal T cell immunity by perturbing macrophage homeostasis. Sci Transl Med (2018) 10(464). doi: 10.1126/scitranslmed.aao4755
- Mathewson ND, Jenq R, Mathew AV, Koenigsknecht M, Hanash A, Toubai T, et al. Gut microbiome-derived metabolites modulate intestinal epithelial cell damage and mitigate graft-versus-host disease. *Nat Immunol* (2016) 17 (5):505–13. doi: 10.1038/ni.3400
- Weber D, Oefner PJ, Hiergeist A, Koestler J, Gessner A, Weber M, et al. Low urinary indoxyl sulfate levels early after transplantation reflect a disrupted microbiome and are associated with poor outcome. *Blood* (2015) 126 (14):1723–8. doi: 10.1182/blood-2015-04-638858
- Weber D, Jenq RR, Peled JU, Taur Y, Hiergeist A, Koestler J, et al. Microbiota Disruption Induced by Early Use of Broad-Spectrum Antibiotics Is an Independent Risk Factor of Outcome after Allogeneic Stem Cell Transplantation. *Biol Blood Marrow Transpl* (2017) 23(5):845–52. doi: 10.1016/j.bbmt.2017.02.006
- Swimm A, Giver CR, DeFilipp Z, Rangaraju S, Sharma A, Ulezko Antonova A, et al. Indoles derived from intestinal microbiota act via type I interferon signaling to limit graft-versus-host disease. *Blood* (2018) 132(23):2506–19. doi: 10.1182/blood-2018-03-838193
- De Vadder F, Kovatcheva-Datchary P, Goncalves D, Vinera J, Zitoun C, Duchampt A, et al. Microbiota-generated metabolites promote metabolic benefits via gut-brain neural circuits. *Cell* (2014) 156(1-2):84–96. doi: 10.1016/j.cell.2013.12.016
- Koeth RA, Wang Z, Levison BS, Buffa JA, Org E, Sheehy BT, et al. Intestinal microbiota metabolism of L-carnitine, a nutrient in red meat, promotes atherosclerosis. *Nat Med* (2013) 19(5):576–85. doi: 10.1038/nm.3145
- 80. Rothhammer V, Quintana FJ. The aryl hydrocarbon receptor: an environmental sensor integrating immune responses in health and

- disease. Nat Rev Immunol (2019) 19(3):184-97. doi: 10.1038/s41577-019-0125-8
- Alkhouri N, Eng K, Cikach F, Patel N, Yan C, Brindle A, et al. Breathprints of childhood obesity: changes in volatile organic compounds in obese children compared with lean controls. *Pediatr Obes* (2015) 10(1):23–9. doi: 10.1111/ j.2047-6310.2014.221.x
- Hanouneh IA, Zein NN, Cikach F, Dababneh L, Grove D, Alkhouri N, et al.
 The breathprints in patients with liver disease identify novel breath biomarkers in alcoholic hepatitis. Clin Gastroenterol Hepatol (2014) 12 (3):516–23. doi: 10.1016/j.cgh.2013.08.048
- Rieder F, Kurada S, Grove D, Cikach F, Lopez R, Patel N, et al. A Distinct Colon-Derived Breath Metabolome is Associated with Inflammatory Bowel Disease, but not its Complications. Clin Transl Gastroenterol (2016) 7(11): e201. doi: 10.1038/ctg.2016.57
- Hamilton BK, Rybicki LA, Grove D, Ferraro C, Starn J, Hodgeman B, et al. Breath analysis in gastrointestinal graft-versus-host disease after allogeneic hematopoietic cell transplantation. *Blood Adv* (2019) 3(18):2732–7. doi: 10.1182/bloodadvances.2019000345
- 85. Chen ML, Zhu XH, Ran L, Lang HD, Yi L, Mi MT. Trimethylamine-N-Oxide Induces Vascular Inflammation by Activating the NLRP3 Inflammasome Through the SIRT3-SOD2-mtROS Signaling Pathway. J Am Heart Assoc (2017) 6(9). doi: 10.1161/JAHA.117.006347
- Wu K, Yuan Y, Yu H, Dai X, Wang S, Sun Z, et al. Gut microbial metabolite trimethylamine N-oxide aggravates GVHD by inducing M1 macrophage polarization in mice. *Blood* (2020) 36(4):501–15. doi: 10.1182/blood. 2019003990
- Bealmear PM, Mirand EA, Holtermann OA. Modification of graft-vs-host disease following bone marrow transplantation in germfree mice. *Prog Clin Biol Res* (1983) 132C:409–21.
- Wade AC, Luckert PH, Tazume S, Niedbalski JL, Pollard M. Characterization of xenogeneic mouse-to-rat bone marrow chimeras. I. Examination of hematologic and immunologic function. *Transplantation* (1987) 44(1):88–92. doi: 10.1097/00007890-198707000-00019
- Heidt PJ, Vossen JM. Experimental and clinical gnotobiotics: influence of the microflora on graft-versus-host disease after allogeneic bone marrow transplantation. J Med (1992) 23(3-4):161–73.
- Bauer H, Horowitz RE, Levenson SM, Popper H. The response of the lymphatic tissue to the microbial flora. Studies on germfree mice. Am J Pathol (1963) 42:471–83.
- Hooper LV, Littman DR, Macpherson AJ. Interactions between the microbiota and the immune system. Science (2012) 336(6086):1268–73. doi: 10.1126/science.1223490
- Round JL, Mazmanian SK. The gut microbiota shapes intestinal immune responses during health and disease. Nat Rev Immunol (2009) 9(5):313–23. doi: 10.1038/nri2515
- 93. Fredricks DN. The gut microbiota and graft-versus-host disease. *J Clin Invest* (2019) 129(5):1808–17. doi: 10.1172/JCI125797
- van Bekkum DW, Roodenburg J, Heidt PJ, van der Waaij D. Mitigation of secondary disease of allogeneic mouse radiation chimeras by modification of the intestinal microflora. J Natl Cancer Inst (1974) 52(2):401–4. doi: 10.1093/ jnci/52.2.401
- 95. Jones JM, Wilson R, Bealmear PM. Mortality and gross pathology of secondary disease in germfree mouse radiation chimeras. *Radiat Res* (1971) 45(3):577–88. doi: 10.2307/3573066
- Lampert IA, Moore RH, Huby R, Cohen J. Observations on the role of endotoxin in graft-versus-host disease. Prog Clin Biol Res (1988) 272:351-9.
- Vriesendorp HM, Heidt PJ, Zurcher C. Gastrointestinal decontamination of dogs treated with total body irradiation and bone marrow transplantation. *Exp Hematol* (1981) 9(9):904–16.
- Heimesaat MM, Nogai A, Bereswill S, Plickert R, Fischer A, Loddenkemper C, et al. MyD88/TLR9 mediated immunopathology and gut microbiota dynamics in a novel murine model of intestinal graft-versus-host disease. Gut (2010) 59(8):1079–87. doi: 10.1136/gut.2009.197434
- Jenq RR, Ubeda C, Taur Y, Menezes CC, Khanin R, Dudakov JA, et al. Regulation of intestinal inflammation by microbiota following allogeneic bone marrow transplantation. J Exp Med (2012) 209(5):903–11. doi: 10.1084/jem.20112408

- 100. Eriguchi Y, Takashima S, Oka H, Shimoji S, Nakamura K, Uryu H, et al. Graft-versus-host disease disrupts intestinal microbial ecology by inhibiting Paneth cell production of alpha-defensins. *Blood* (2012) 120(1):223–31. doi: 10.1182/blood-2011-12-401166
- Eriguchi Y, Nakamura K, Hashimoto D, Shimoda S, Shimono N, Akashi K, et al. Decreased secretion of Paneth cell alpha-defensins in graft-versus-host disease. *Transpl Infect Dis* (2015) 17(5):702–6. doi: 10.1111/tid.12423
- 102. Shono Y, Docampo MD, Peled JU, Perobelli SM, Velardi E, Tsai JJ, et al. Increased GVHD-related mortality with broad-spectrum antibiotic use after allogeneic hematopoietic stem cell transplantation in human patients and mice. Sci Transl Med (2016) 8(339):339ra71. doi: 10.1126/scitranslmed. aa(7311)
- 103. Bouazzaoui A, Huber E, Dan A, Al-Allaf FA, Pfirstinger J, Sprotte G, et al. Reduction of aGVHD using chicken antibodies directed against intestinal pathogens in a murine model. *Blood* (2017) 129(8):1052–5. doi: 10.1182/ blood-2016-06-722538
- 104. Stein-Thoeringer CK, Nichols KB, Lazrak A, Docampo MD, Slingerland AE, Slingerland JB, et al. Lactose drives Enterococcus expansion to promote graft-versus-host disease. *Science* (2019) 366(6469):1143–9. doi: 10.1126/ science.aax3760
- 105. Toubai T, Fujiwara H, Rossi C, Riwes M, Tamaki H, Zajac C, et al. Host NLRP6 exacerbates graft-versus-host disease independent of gut microbial composition. Nat Microbiol (2019) 4(5):800–12. doi: 10.1038/s41564-019-0373-1
- 106. Xu Y, Li X, Jin L, Zhen Y, Lu Y, Li S, et al. Application of chicken egg yolk immunoglobulins in the control of terrestrial and aquatic animal diseases: a review. *Biotechnol Adv* (2011) 29(6):860-8. doi: 10.1016/j.biotechadv.2011.07.003
- 107. Li X, Jing K, Wang X, Li Y, Zhang M, Li Z, et al. Protective effects of chicken egg yolk antibody (IgY) against experimental Vibrio splendidus infection in the sea cucumber (Apostichopus japonicus). Fish Shellfish Immunol (2016) 48:105–11. doi: 10.1016/j.fsi.2015.11.024
- Taur Y, Xavier JB, Lipuma L, Ubeda C, Goldberg J, Gobourne A, et al. Intestinal domination and the risk of bacteremia in patients undergoing allogeneic hematopoietic stem cell transplantation. Clin Infect Dis (2012) 55 (7):905–14. doi: 10.1093/cid/cis580
- 109. Ford CD, Gazdik MA, Lopansri BK, Webb B, Mitchell B, Coombs J, et al. Vancomycin-Resistant Enterococcus Colonization and Bacteremia and Hematopoietic Stem Cell Transplantation Outcomes. *Biol Blood Marrow Transpl* (2017) 23(2):340–6. doi: 10.1016/j.bbmt.2016.11.017
- Elinav E, Strowig T, Kau AL, Henao-Mejia J, Thaiss CA, Booth CJ, et al. NLRP6 inflammasome regulates colonic microbial ecology and risk for colitis. Cell (2011) 145(5):745–57. doi: 10.1016/j.cell.2011.04.022
- Levy M, Shapiro H, Thaiss CA, Elinav E. NLRP6: A Multifaceted Innate Immune Sensor. Trends Immunol (2017) 38(4):248–60. doi: 10.1016/j.it.2017.01.001
- 112. Hu B, Elinav E, Huber S, Strowig T, Hao L, Hafemann A, et al. Microbiota-induced activation of epithelial IL-6 signaling links inflammasome-driven inflammation with transmissible cancer. *Proc Natl Acad Sci U S A* (2013) 110 (24):9862–7. doi: 10.1073/pnas.1307575110
- 113. Anand PK, Malireddi RK, Lukens JR, Vogel P, Bertin J, Lamkanfi M, et al. NLRP6 negatively regulates innate immunity and host defence against bacterial pathogens. *Nature* (2012) 488(7411):389–93. doi: 10.1038/nature11250
- 114. Durakovic N, Radojcic V, Skarica M, Bezak KB, Powell JD, Fuchs EJ, et al. Factors governing the activation of adoptively transferred donor T cells infused after allogeneic bone marrow transplantation in the mouse. *Blood* (2007) 109(10):4564–74. doi: 10.1182/blood-2006-09-048124
- 115. Taylor PA, Ehrhardt MJ, Lees CJ, Panoskaltsis-Mortari A, Krieg AM, Sharpe AH, et al. TLR agonists regulate alloresponses and uncover a critical role for donor APCs in allogeneic bone marrow rejection. *Blood* (2008) 112(8):3508–16. doi: 10.1182/blood-2007-09-113670
- 116. Calcaterra C, Sfondrini L, Rossini A, Sommariva M, Rumio C, Menard S, et al. Critical role of TLR9 in acute graft-versus-host disease. *J Immunol* (2008) 181(9):6132–9. doi: 10.4049/jimmunol.181.9.6132
- 117. Denning TL, Norris BA, Medina-Contreras O, Manicassamy S, Geem D, Madan R, et al. Functional specializations of intestinal dendritic cell and macrophage subsets that control Th17 and regulatory T cell responses are dependent on the T cell/APC ratio, source of mouse strain, and regional localization. J Immunol (2011) 187(2):733–47. doi: 10.4049/jimmunol.1002701

- Org E, Mehrabian M, Parks BW, Shipkova P, Liu X, Drake TA, et al. Sex differences and hormonal effects on gut microbiota composition in mice. *Gut Microbes* (2016) 7(4):313–22. doi: 10.1080/19490976.2016.1203502
- 119. Peng C, Xu X, Li Y, Li X, Yang X, Chen H, et al. Sex-specific association between the gut microbiome and high-fat diet-induced metabolic disorders in mice. *Biol Sex Differ* (2020) 11(1):5. doi: 10.1186/s13293-020-0281-3
- 120. Fransen F, Zagato E, Mazzini E, Fosso B, Manzari C, El Aidy S, et al. BALB/c and C57BL/6 Mice Differ in Polyreactive IgA Abundance, which Impacts the Generation of Antigen-Specific IgA and Microbiota Diversity. *Immunity* (2015) 43(3):527–40. doi: 10.1016/j.immuni.2015.08.011
- Spychala MS, Venna VR, Jandzinski M, Doran SJ, Durgan DJ, Ganesh BP, et al. Age-related changes in the gut microbiota influence systemic inflammation and stroke outcome. *Ann Neurol* (2018) 84(1):23–36. doi: 10.1002/ana.25250
- 122. Storb R, Prentice RL, Buckner CD, Clift RA, Appelbaum F, Deeg J, et al. Graft-versus-host disease and survival in patients with aplastic anemia treated by marrow grafts from HLA-identical siblings. Beneficial effect of a protective environment. N Engl J Med (1983) 308(6):302–7. doi: 10.1056/NEJM198302103080602
- 123. Taur Y, Jenq RR, Perales MA, Littmann ER, Morjaria S, Ling L, et al. The effects of intestinal tract bacterial diversity on mortality following allogeneic hematopoietic stem cell transplantation. *Blood* (2014) 124(7):1174–82. doi: 10.1182/blood-2014-02-554725
- 124. Jenq RR, Taur Y, Devlin SM, Ponce DM, Goldberg JD, Ahr KF, et al. Intestinal Blautia Is Associated with Reduced Death from Graft-versus-Host Disease. Biol Blood Marrow Transpl (2015) 21(8):1373–83. doi: 10.1016/j.bbmt.2015.04.016
- 125. Simms-Waldrip TR, Sunkersett G, Coughlin LA, Savani MR, Arana C, Kim J, et al. Antibiotic-Induced Depletion of Anti-inflammatory Clostridia Is Associated with the Development of Graft-versus-Host Disease in Pediatric Stem Cell Transplantation Patients. *Biol Blood Marrow Transpl* (2017) 23(5):820–9. doi: 10.1016/j.bbmt.2017.02.004
- 126. Golob JL, Pergam SA, Srinivasan S, Fiedler TL, Liu C, Garcia K, et al. Stool Microbiota at Neutrophil Recovery Is Predictive for Severe Acute Graft vs Host Disease After Hematopoietic Cell Transplantation. Clin Infect Dis (2017) 65(12):1984–91. doi: 10.1093/cid/cix699
- 127. Han L, Zhang H, Chen S, Zhou L, Li Y, Zhao K, et al. Intestinal Microbiota Can Predict Acute Graft-versus-Host Disease Following Allogeneic Hematopoietic Stem Cell Transplantation. *Biol Blood Marrow Transpl* (2019) 25(10):1944–55. doi: 10.1016/j.bbmt.2019.07.006
- 128. Payen M, Nicolis I, Robin M, Michonneau D, Delannoye J, Mayeur C, et al. Functional and phylogenetic alterations in gut microbiome are linked to graft-versus-host disease severity. Blood Adv (2020) 4(9):1824–32. doi: 10.1182/bloodadvances.2020001531
- 129. Peled JU, Gomes ALC, Devlin SM, Littmann ER, Taur Y, Sung AD, et al. Microbiota as Predictor of Mortality in Allogeneic Hematopoietic-Cell Transplantation. N Engl J Med (2020) 382(9):822–34. doi: 10.1056/ NEJMoa1900623
- Arpaia N, Campbell C, Fan X, Dikiy S, van der Veeken J, deRoos P, et al. Metabolites produced by commensal bacteria promote peripheral regulatory T-cell generation. *Nature* (2013) 504(7480):451–5. doi: 10.1038/nature12726
- 131. Holler E, Butzhammer P, Schmid K, Hundsrucker C, Koestler J, Peter K, et al. Metagenomic analysis of the stool microbiome in patients receiving allogeneic stem cell transplantation: loss of diversity is associated with use of systemic antibiotics and more pronounced in gastrointestinal graft-versushost disease. *Biol Blood Marrow Transpl* (2014) 20(5):640–5. doi: 10.1016/i.bbmt.2014.01.030
- David LA, Maurice CF, Carmody RN, Gootenberg DB, Button JE, Wolfe BE, et al. Diet rapidly and reproducibly alters the human gut microbiome. *Nature* (2014) 505(7484):559–63. doi: 10.1038/nature12820
- Mattsson J, Westin S, Edlund S, Remberger M. Poor oral nutrition after allogeneic stem cell transplantation correlates significantly with severe graftversus-host disease. *Bone Marrow Transpl* (2006) 38(9):629–33. doi: 10.1038/si.bmt.1705493
- 134. Gonzales F, Bruno B, Alarcon Fuentes M, De Berranger E, Guimber D, Behal H, et al. Better early outcome with enteral rather than parenteral nutrition in children undergoing MAC allo-SCT. Clin Nutr (2018) 37(6 Pt A):2113–21. doi: 10.1016/j.clnu.2017.10.005

- 135. Guieze R, Lemal R, Cabrespine A, Hermet E, Tournilhac O, Combal C, et al. Enteral versus parenteral nutritional support in allogeneic haematopoietic stem-cell transplantation. Clin Nutr (2014) 33(3):533–8. doi: 10.1016/j.clnu.2013.07.012
- Pierre JF. Gastrointestinal immune and microbiome changes during parenteral nutrition. Am J Physiol Gastrointest Liver Physiol (2017) 312(3): G246–G56. doi: 10.1152/ajpgi.00321.2016
- 137. Seguy D, Berthon C, Micol JB, Darre S, Dalle JH, Neuville S, et al. Enteral feeding and early outcomes of patients undergoing allogeneic stem cell transplantation following myeloablative conditioning. *Transplantation* (2006) 82(6):835–9. doi: 10.1097/01.tp.0000229419.73428.ff
- 138. Moody KM, Baker RA, Santizo RO, Olmez I, Spies JM, Buthmann A, et al. A randomized trial of the effectiveness of the neutropenic diet versus food safety guidelines on infection rate in pediatric oncology patients. *Pediatr Blood Cancer* (2018) 65(1). doi: 10.1002/pbc.26711
- 139. Trifilio S, Helenowski I, Giel M, Gobel B, Pi J, Greenberg D, et al. Questioning the role of a neutropenic diet following hematopoetic stem cell transplantation. *Biol Blood Marrow Transpl* (2012) 18(9):1385–90. doi: 10.1016/j.bbmt.2012.02.015
- 140. Smith PM, Howitt MR, Panikov N, Michaud M, Gallini CA, Bohlooly YM, et al. The microbial metabolites, short-chain fatty acids, regulate colonic Treg cell homeostasis. *Science* (2013) 341(6145):569–73. doi: 10.1126/science.1241165
- 141. Zama D, Bossu G, Leardini D, Muratore E, Biagi E, Prete A, et al. Insights into the role of intestinal microbiota in hematopoietic stem-cell transplantation. *Ther Adv Hematol* (2020) 11. doi: 10.1177/2040620719896961
- 142. Yoshifuji K, Inamoto K, Kiridoshi Y, Takeshita K, Sasajima S, Shiraishi Y, et al. Prebiotics protect against acute graft-versus-host disease and preserve the gut microbiota in stem cell transplantation. *Blood Adv* (2020) 4 (19):4607–17. doi: 10.1182/bloodadvances.2020002604
- 143. Schwabkey ZI, Jenq RR. Microbiome Anomalies in Allogeneic Hematopoietic Cell Transplantation. Annu Rev Med (2020) 71:137–48. doi: 10.1146/annurev-med-052918-122440
- 144. Tavil B, Koksal E, Yalcin SS, Uckan D. Pretransplant nutritional habits and clinical outcome in children undergoing hematopoietic stem cell transplant. *Exp Clin Transpl* (2012) 10(1):55–61. doi: 10.6002/ect.2011.0082
- 145. Ladas EJ, Bhatia M, Chen L, Sandler E, Petrovic A, Berman DM, et al. The safety and feasibility of probiotics in children and adolescents undergoing hematopoietic cell transplantation. *Bone Marrow Transpl* (2016) 51(2):262– 6. doi: 10.1038/bmt.2015.275
- 146. Gerbitz A, Schultz M, Wilke A, Linde HJ, Scholmerich J, Andreesen R, et al. Probiotic effects on experimental graft-versus-host disease: let them eat yogurt. *Blood* (2004) 103(11):4365–7. doi: 10.1182/blood-2003-11-3769
- 147. Mehta A, Rangarajan S, Borate U. A cautionary tale for probiotic use in hematopoietic SCT patients-Lactobacillus acidophilus sepsis in a patient with mantle cell lymphoma undergoing hematopoietic SCT. Bone Marrow Transpl (2013) 48(3):461–2. doi: 10.1038/bmt.2012.153
- 148. Vahabnezhad E, Mochon AB, Wozniak LJ, Ziring DA. Lactobacillus bacteremia associated with probiotic use in a pediatric patient with ulcerative colitis. J Clin Gastroenterol (2013) 47(5):437–9. doi: 10.1097/ MCG.0b013e318279abf0
- 149. Robin F, Paillard C, Marchandin H, Demeocq F, Bonnet R, Hennequin C. Lactobacillus rhamnosus meningitis following recurrent episodes of bacteremia in a child undergoing allogeneic hematopoietic stem cell transplantation. J Clin Microbiol (2010) 48(11):4317–9. doi: 10.1128/JCM.00250-10
- 150. Vossen JM, Guiot HF, Lankester AC, Vossen AC, Bredius RG, Wolterbeek R, et al. Complete suppression of the gut microbiome prevents acute graft-versus-host disease following allogeneic bone marrow transplantation. *PloS One* (2014) 9(9):e105706. doi: 10.1371/journal.pone.0105706
- 151. Weber D, Oefner PJ, Dettmer K, Hiergeist A, Koestler J, Gessner A, et al. Rifaximin preserves intestinal microbiota balance in patients undergoing allogeneic stem cell transplantation. *Bone Marrow Transpl* (2016) 51 (8):1087–92. doi: 10.1038/bmt.2016.66
- Tawara I, Liu C, Tamaki H, Toubai T, Sun Y, Evers R, et al. Influence of donor microbiota on the severity of experimental graft-versus-host-disease. *Biol Blood Marrow Transpl* (2013) 19(1):164–8. doi: 10.1016/j.bbmt.2012.09.001
- 153. Whangbo J, Ritz J, Bhatt A. Antibiotic-mediated modification of the intestinal microbiome in allogeneic hematopoietic stem cell

- transplantation. Bone Marrow Transpl (2017) 52(2):183–90. doi: 10.1038/bmt.2016.206
- 154. Kimura S, Akahoshi Y, Nakano H, Ugai T, Wada H, Yamasaki R, et al. Antibiotic prophylaxis in hematopoietic stem cell transplantation. A metaanalysis of randomized controlled trials. J Infect (2014) 69(1):13–25. doi: 10.1016/j.jinf.2014.02.013
- 155. Routy B, Letendre C, Enot D, Chenard-Poirier M, Mehraj V, Seguin NC, et al. The influence of gut-decontamination prophylactic antibiotics on acute graft-versus-host disease and survival following allogeneic hematopoietic stem cell transplantation. *Oncoimmunology* (2017) 6(1):e1258506. doi: 10.1080/2162402X.2016.1258506
- 156. Farowski F, Bucker V, Vehreschild JJ, Biehl L, Cruz-Aguilar R, Scheid C, et al. Impact of choice, timing, sequence and combination of broad-spectrum antibiotics on the outcome of allogeneic haematopoietic stem cell transplantation. Bone Marrow Transpl (2018) 53(1):52–7. doi: 10.1038/ bmt.2017.203
- 157. DeFilipp Z, Hohmann E, Jenq RR, Chen YB. Fecal Microbiota Transplantation: Restoring the Injured Microbiome after Allogeneic Hematopoietic Cell Transplantation. Biol Blood Marrow Transpl (2019) 25(1):e17-22. doi: 10.1016/j.bbmt.2018.10.022
- 158. Kakihana K, Fujioka Y, Suda W, Najima Y, Kuwata G, Sasajima S, et al. Fecal microbiota transplantation for patients with steroid-resistant acute graft-versus-host disease of the gut. *Blood* (2016) 128(16):2083–8. doi: 10.1182/blood-2016-05-717652
- 159. Spindelboeck W, Schulz E, Uhl B, Kashofer K, Aigelsreiter A, Zinke-Cerwenka W, et al. Repeated fecal microbiota transplantations attenuate diarrhea and lead to sustained changes in the fecal microbiota in acute, refractory gastrointestinal graft-versus-host-disease. *Haematologica* (2017) 102(5):e210–e3. doi: 10.3324/haematol.2016.154351
- 160. Qi X, Li X, Zhao Y, Wu X, Chen F, Ma X, et al. Treating Steroid Refractory Intestinal Acute Graft-vs.-Host Disease With Fecal Microbiota Transplantation: A Pilot Study. Front Immunol (2018) 9:2195. doi: 10.3389/fimmu.2018.02195

- 161. Kaito S, Toya T, Yoshifuji K, Kurosawa S, Inamoto K, Takeshita K, et al. Fecal microbiota transplantation with frozen capsules for a patient with refractory acute gut graft-versus-host disease. Blood Adv (2018) 2(22):3097–101. doi: 10.1182/bloodadvances.2018024968
- 162. Zhang J, Ren G, Li M, Lu P, Yi S. The Effects of Fecal Donors with Different Feeding Patterns on Diarrhea in a Patient Undergoing Hematopoietic Stem Cell Transplantation. Case Rep Hematol (2019) 2019:4505238. doi: 10.1155/ 2019/4505238
- 163. Quera R, Espinoza R, Estay C, Rivera D. Bacteremia as an adverse event of fecal microbiota transplantation in a patient with Crohn's disease and recurrent Clostridium difficile infection. *J Crohns Colitis* (2014) 8(3):252–3. doi: 10.1016/j.crohns.2013.10.002
- 164. Kuijper EJ, Allegretii J, Hawkey P, Sokol H, Goldenberg S, Ianiro G, et al. A necessary discussion after transmission of multidrug-resistant organisms through faecal microbiota transplantations. *Lancet Infect Dis* (2019) 19 (11):1161–2. doi: 10.1016/S1473-3099(19)30545-6
- 165. Han L, Zhao K, Li Y, Han H, Zhou L, Ma P, et al. A gut microbiota score predicting acute graft-versus-host disease following myeloablative allogeneic hematopoietic stem cell transplantation. *Am J Transpl* (2020) 20(4):1014–27. doi: 10.1111/ajt.15654

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2021 Hong, Wang, Wang, Yang, Wang, Gao and Zhang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with





GVHD Prophylaxis 2020

Mahasweta Gooptu* and Joseph Harry Antin

Dana-Farber Cancer Institute, Boston, MA, United States

Graft-vs. host disease (GVHD), both acute and chronic are among the chief non-relapse complications of allogeneic transplantation which still cause substantial morbidity and mortality despite significant advances in supportive care over the last few decades. The prevention of GVHD therefore remains critical to the success of allogeneic transplantation. In this review we briefly discuss the pathophysiology and immunobiology of GVHD and the current standards in the field which remain centered around calcineurin inhibitors. We then discuss important translational advances in GVHD prophylaxis, approaching these various platforms from a mechanistic standpoint based on the pathophysiology of GVHD including in-vivo and ex-vivo T-cell depletion alongwith methods of selective T-cell depletion, modulation of T-cell co-stimulatory pathways (checkpoints), enhancing regulatory T-cells (Tregs), targeting T-cell trafficking as well as cytokine pathways. Finally we highlight exciting novel pre-clinical research that has the potential to translate to the clinic successfully. We approach these methods from a pathophysiology based perspective as well and touch upon strategies targeting the interaction between tissue damage induced antigens and T-cells, regimen related endothelial toxicity, T-cell co-stimulatory pathways and other T-cell modulatory approaches, T-cell trafficking, and cytokine pathways. We end this review with a critical discussion of existing data and novel therapies that may be transformative in the field in the near future as a comprehensive picture of GVHD prophylaxis in 2020. While calcineurin inhibitors remain the standard, post-transplant eparinsphamide originally developed to facilitate haploidentical transplantation is becoming an attractive alternative to traditional calcinuerin inhibitor based prophylaxis due to its ability to reduce severe forms of acute and chronic GVHD without compromising other outcomes, even in the HLA-matched setting. In addition T-cell modulation, particularly targeting some important T-cell co-stimulatory pathways have resulted in promising outcomes and may be a part of GVHD prophylaxis in the future. Novel approaches including targeting early events in GVHD pathogenesis such as interactions byetween tissue damage associated antigens and T-cells, endothelial toxicity, and T-cell trafficking are also promising and discussed in this review. GVHD prophylaxis in 2020 continues to evolve with novel exiciting therapies on the horizon based on a more sophisticated understanding of the immunobiology

Keywords: GvHD prophylaxis, calcineurin inhibitors, post-transplant cyclophosphamide, T-cell depletion, T-cell modulation, T-cell trafficking

OPEN ACCESS

Edited by:

Charles Craddock, University of Birmingham, United Kingdom

Reviewed by:

Philippe Saas, INSERM U1098 Interactions Hôte-Greffon-Tumeur and Ingénierie Cellulaire et Génique, France Ying-Jun Chang, Peking University People's Hospital. China

*Correspondence:

Mahasweta Gooptu mahasweta_gooptu@dfci.harvard.edu

Specialty section:

This article was submitted to Alloimmunity and Transplantation, a section of the journal Frontiers in Immunology

Received: 13 September 2020 Accepted: 22 February 2021 Published: 07 April 2021

Citation:

of GVHD.

Gooptu M and Antin JH (2021) GVHD Prophylaxis 2020. Front. Immunol. 12:605726. doi: 10.3389/fimmu.2021.605726

INTRODUCTION

GVHD prophylaxis has come a long way since the initial days of allogeneic transplantation. The improvement in GVHD outcomes has been one of the primary reasons for the reduction in non-relapse mortality over time (1) that has enhanced the success of allogeneic transplantation and allowed us to perform transplants in older patients as well as those with co-morbidities. GVHD comprises two distinct entities-acute GVHD (aGVHD) which typically presents in the first 3-6 months following transplant and manifests as a characteristic rash, secretory diarrhea, or cholestatic liver function abnormalities and chronic GVHD (cGVHD) which presents usually after the first 3 months and can affect virtually any organ system (ocular, oral, skin, musculo-skeletal, gastro-intestinal, pulmonary etc.,). Overlap syndromes are well-recognized, although relatively rare. These two entities have distinct pathophysiologies as well, however, prophylactic strategies generally try to prevent both acute and chronic varieties albeit with varying success depending on the strategy.

In HLA-matched transplantation, while the backbone of most widely used prophylactic platforms remains calcineurin-inhibitor (CNI)-based, a variety of new drugs have been added to CNI's in an attempt to improve efficacy and reduce toxicity. In this review, we discuss current standards and their evolution over time and highlight some of these translational advances. Further we touch upon novel pre-clinical advances developed on the foundation of a deeper understanding of transplant immunology and promising for translation to the clinic. We begin with a description of the immunobiology of GVHD, to better understand potential targets which have been exploited over the last few decades and currently to develop effective prophylactic therapies for GVHD.

The Immunobiology of Graft-vs.-Host Disease

Acute GVHD

One of the first models describing the biology of GVHD was proposed by Antin and Ferrara where they described a sequential cascade initiated by conditioning regimen mediated host tissue injury with the production of inflammatory cytokines (phase 1). This is followed by activation and proliferation of effector T-lymphocytes (phase II) which eventually lead to recruitment and activation of additional mononuclear effectors and amplification of a "cytokine storm" (Phase III) (2). Further extensive research has refined these concepts and identified targets for development of novel prophylactic strategies to prevent acute and chronic GVHD.

Phase 1

Both neutrophils and monocytes are involved in the initial inflammatory response in the pathogenesis of GVHD. Monocytes are activated by molecules called damage-associated molecular patterns (DAMPs) such as uric acid, ATP, Heparan sulfate, HMGB-1 or IL-33 which can initiate and perpetuate a non-infectious inflammatory response involving the innate immune system. In contrast pathogen-associated molecular patterns

(PAMPs) such as bacterial lipopolysaccharides cause infectionassociated inflammation. The DAMP and PAMP mediated inflammatory responses result in activation of the innate immune system (monocytes and neutrophils) which then cause local tissue damage mediated by reactive oxygen species. This eventually culminates in interaction of antigen-presenting cells (APCs) in the innate and adaptive immune and activation of cytokine cascades (IL-1, IL-6, TNF- α , etc.,) leading to the "cytokine storm (3)." Prophylactic strategies targeting these events have focused on arresting the cytokine storm through inhibition of particular cytokines or interrupting the interaction between APCs and PAMP.

Phase II

Primed by this cytokine storm, effector T-lymphocytes now migrate to lymphoid organs and host tissues mediated by L-selectin, CCR7. This culminates in APC mediated T-cell activation and engagement of the T-cell receptor complex and modulation by anti and co-stimulatory pathways. In particular, T-lymphocytes trafficking to the gut express high levels of integrin $\beta 7~(\alpha 4\beta 7)$ which bind corresponding host tissue ligands presenting a potential target for intervention. The T-cell activation process and proliferation process is a crucial target for GVHD prevention.

Phase III

These events lead to a self-potentiating T-lymphocyte activation causing tissue damage via direct cellular cytotoxicity and indirectly via release of soluble mediators (TNF- α , IFN- γ , IL-1, and nitric oxide) (4).

A number of additional pathways have since been implicated in GVHD pathogenesis including the canonical NOTCH pathway (5). It has been shown via monoclonal antibodies that Notch-deprived T-cells proliferate normally but produce less inflammatory cytokines with a preferential increase in Tregs (6) and all the effects were dependent on NOTCH1/2 receptors on T-cells and Dll1/4 ligands in the recipient with dominant roles for NOTCH1 and Dll4 (6).

While the pro-inflammatory signals described above potentiate GVHD, there are also anti-inflammatory components of the immune system that try to dampen these inflammatory responses. Regulatory T-cells (Tregs) are important in immunologic tolerance, partly via release of anti-inflammatory cytokines such as IL-10 and TGF-β (7). Cytokine responses are often classified as effector T helper (Th) type 1 (IL-2, INF-γ) and type 2 (IL-4, IL-10) responses where type 2 cytokines can inhibit potent proinflammatory type 1cytokines, and a Th1 to Th2 shift could be beneficial in aGVHD (8). In addition a particular subset of CD4+ cells called Th17 cells have been identified which are characterized by the production characterized by production of IL-17A and F, IL-21, and IL-22 and which in murine models migrate to GVHD target organs causing severe pulmonary and GI lesions and GVHD deaths (9). These are postulated to be anatagonistic to Tregs (10) making them an interesting target. Invariant natural killer T (iNKT) cells are another cellular subset with putative immunoregulatory functions, in part via an increase Treg numbers and IL-4 secretion, that may be important in GVHD pathophysiology.

Chronic GVHD

Chronic GVHD remains the most common late toxicity of allogeneic transplantation with significant morbidity and quality of life implications. cGVHD has its own distinctive immunobiology. Briefly we can conceptualize the pathophysiology of cGVHD in three phases: (1) Inflammation leading to tissue damage (2) chronic inflammation, thymic injury, dysregulated B- and T-cell immunity (3) tissue repair with fibrosis (11, 12). Although a more detailed discussion of these phases is beyond the scope of this review, we will focus on some of the known interventions that can prevent or reduce the incidence of cGVHD as well as some novel therapies being tested, particularly those targeting the B-cell axis.

Potential targets for developing novel prophylactic platforms have been identified based on our current and more comprehensive understanding of the biology of GVHD. In this review we discuss both current standards and important translational advances as well as exciting new potential therapies which may be translated to the clinic in the future.

Current Standards in GVHD Prophylaxis

The effective prevention of GVHD is critical to the success of allogeneic transplantation. Based on the understanding that aGVHD is primarily mediated by effector T-lymphocytes, prophylactic strategies have focused on T-cell suppression in the recipient. Calcineurin inhibitors (tacrolimus/Tac and cyclosporine/CyA) inhibit the proliferation and activation of Tcells and have been used in combination with either methotrexate (MTX) or mycophenolate mofetil (MMF) as standard prophylaxis in HLA-matched HSCT. In two randomized controlled trials (RCT) in the 1990s, the combination of Tac/MTX was found to be significantly superior to CyA/MTX is the prevention of grade II-IV aGVHD and extensive chronic GVHD in HLA-matched sibling and unrelated donors, although a benefit in overall survival (OS) was not shown (13, 14). Furthermore, a single-center phase II RCT compared Tac/MTX with Tac/MMF and found that Tac/MTX was more effective in preventing severe aGVHD, particularly in matched unrelated donor (MUD) transplantation (15). CNI based prophylaxis remains the standard in HLA-matched transplantation. However, the recent advent of post-transplant cyclophosphamide (PTCy), has been revolutionary, not only allowing related donor haploidentical transplants to be performed but also making some inroads in the field of HLA-matched transplantation.

Translational Advances in GVHD Prophylaxis

In-vivo T-Cell Depletion/Modulation

T-cell depletion or modulation *in-vivo* has been the basis for the development of a number of novel GVHD prophylaxis strategies. These have typically been incorporated into regimens where the backbone comprises CNIs. We summarize some of these approaches below.

Post-transplant Cyclophosphamide

Transplantation across HLA barriers historically has been difficult due to high rates of graft rejection and severe GVHD secondary to strong bidirectional alloreactive responses between

donor and recipient. The introduction of post-transplant cyclophosphamide (PTCy) in the context of haploidentical transplantation has been a gamechanger and allowed us to perform such mismatched transplants safely and effectively, typically from related donors.

First pioneered at Johns Hopkins, cyclophosphamide is given at doses of 50 mg/kg on days +3 and +4 following the infusion of haploidentical stem-cells. In initial studies reported by the Hopkins group, reduced intensity HSCT with PTCy along with Tac and MMF as GVHD prophylaxis resulted in engraftment in 87% of patients with acceptable rates of grade II-IV (34%) and III-IV aGVHD (6%). Further, rates of chronic severe GVHD were found to be particularly low. Relapse rates were in the 50% range (16). Numerous subsequent studies have shown similar numbers and more recently myeloablative haploidentical transplantation with PTCy based prophylaxis has also been widely adopted (17-19). It is interesting to note that the rates of grade II-IV aGVHD are not in fact significantly lower with PTCy based prophylaxis in haplotransplants compared with those seen in HLA-matched transplant with CNI based prophylaxis and the actual mechanistic implications of PTCy are being investigated.

The earlier more simplistic hypotheses postulated that PTCy results in the deletion of alloreactive T-cells with some debate regarding the effect it may have on the T-cell mediated graft-vs. leukemia (GVL) effect. However, more recently it has been shown in mice that Tregs are preserved and that T-effector cell exhaustion may play an important role (20). A comprehensive model has however not been defined yet.

The beneficial effect of PTCy on severe aGVHD and cGVHD has led to its adoption into the HLA-matched setting more recently. In a Phase II RCT (BMT CTN 1203) which compared three different GVHD prophylaxis regimens in RIC MUD HSCT (PTCy/Tac/MMF, Tac/MTX/bortezomib, and Tac/MTX/maraviroc) with standard Tac/MTX prophylaxis, the PTCy arm fared the best with comparable grade II-IV aGVHD (27%) but lower rates of grade III-IV aGVHD (2%) and cGVHD requiring immunosuppression (22%) outcomes. Relapse rates in the PTCy arm were 28%. Overall GVHD and relapse free survival (GRFS) was superior in the PTCy arm meeting the primary end-point of the trial (21). Again in an RCT from Europe, PTCy/Tac/MMF based prophylaxis fared better than CyA/MMF in the prevention of acute and chronic GVHD in HLA_matched RIC HSCT although the standard of care arm did not include Tac or MTX and numbers were limited (22). Some centers have been using low-dose ATG along with low-dose PTCy to try and improve grade II-IV aGVHD rates (23), however, this is not an universally accepted practice at this time due to limited data.

Bolstered by this data, PTCy/Tac/MMF is now being compared to standard Tac/MTX prophylaxis in RIC HLA-matched HSCT in a large phase III RCT (BMT CTN 1703). PTCy does have the potential to eliminate the effect of donor mismatch on GVHD outcomes, however, CNI based prophylaxis remains the standard until phase III data is available.

Anti-thymocyte Globulin

The polyclonal immunoglobulin product obtained from the sera of rabbits and horses immunized with human thymocytes or Tcell lines is called anti-thymocyte globulin or ATG. ATG has

Advances in GVHD Prophylaxis 2020

been used as part of transplant conditioning to effect *in-vivo* TCD with the aim to reduce both acute and chronic GVHD with varying success.

Of the four RCTs that have evaluated ATG in combination with standard CNI/MTX prophylaxis, the first used horse ATG and showed a reduction in aGVHD; however, there were higher rates of infection with resultant no difference in NRM or OS. Importantly, there was a reduction in chronic severe GVHD (24, 25). The second RCT used rabbit ATG while the third mainly used PBSC grafts. In both of these trials, although there was no effect on aGVHD, a reduction in chronic GVHD was seen once again (26). Therefore, it seems that the use of ATG can reduce severe chronic GVHD with no deleterious effect on OS, however, aGVHD is not consistently reduced.

Rabbit ATG is generally considered to deplete T-cells more effectively as well as allow greater expansion of regulatory T-cells (Tregs) (27). The beneficial effect of ATG on extensive chronic GVHD was suggested in an retrospective analysis which compared ATG to no ATG containing GVHD prophylaxis regimens in matched unrelated donor transplantation (28). Subsequently, in a recent RCT which evaluated Tac/MTX \pm anti T-lymphocyte globulin (ATLG), a form of rabbit ATG, in myeloablative unrelated donor transplantation, a significant reduction in grade II-IV aGVHD and moderate/severe chronic GVHD was seen. However, NRM and OS was impaired in the ATLG arm (29). It was suggested that a higher dose of ATLG in the trial may have contributed to increased infections and mortality.

In this context, there is evidence that increased doses or prolonged dosage schedules of ATG may have immunosuppressive toxicity with increased NRM and relapse (30). Individualized ATG dosing, based not just on weight but on absolute lymphocyte count has been proposed as a way to tailor doses of ATG for maximal benefit (31). Different doses of ATG have also been explored in the context of haploidentical transplantation (32).

In the realm of matched related donor transplantation, a recent multi-center randomized study from China demonstrated improved acute and chronic GVHD rates without compromising relapse or treatment-related mortality (33) and merits further study.

Sirolimus

Sirolimus is a mTOR inhibitor which inhibits effector T-lymphocytes and in *in-vitro* studies appeared to spare regulatory T-lymphocytes. A favorable ratio of Tregs:Teff has been shown to be associated with better GVHD outcomes and hence sirolimus has an immunologic profile that was thought to be potentially beneficial for GVHD prevention. In addition, it has a distinct toxicity profile compared to tacrolimus and is not nephrotoxic. In a large RCT in myeloablative transplants with HLA-matched donors, sirolimus in combination with tacrolimus was compared with the standard Tac/MTX platform. There was no difference in grades II-IV aGVHD and cGVHD, but better grade III-IV aGVHD outcomes with sirolimus/Tac were seen. Non-relapse mortality (NRM) and OS were similar as well (34). Hence sirolimus appears to be an acceptable alternative to MTX when

used with CNIs. In subsequent studies, sirolimus has been associated with higher rates of veno-occlusive disease (VOD) particularly when ablative busulfan is used (35) or when there are additional risk factors for VOD. In RIC transplantation, the addition of sirolimus to Tac/MTX resulted in better grade II-IV aGVHD outcomes without survival benefit in a phase II RCT (36). More recently, a phase II RCT found that the combination of sirolimus with CyA and MMF was superior to CyA/MMF; however, the comparator arm is generally considered inferior to Tac/MTX (37).

Sirolimus has been found to be particularly helpful in situations where nephrotoxicity is a concern such as in transplantation for sickle-cell disease. It is also being used with PTCy in patients with borderline renal function, with rates of engraftment and GVHD comparable to PTCy based regimens with CNI and may be a way to safely perform HSCT in patients with renal dysfunction (38).

Given its Treg sparing effects, novel combinations such as that with OX40L blockade are being investigated (39). This is discussed in greater detail in the section on OX40L blockade later in this review.

Ex-vivo T-Cell Depletion/Modulation

Ex-vivo TCD has been used for decades in allogeneic transplantation as a prophylactic strategy to prevent GVHD. Methods of T-cell depletion have included (1) negative selection (removal of T-lymphocytes) through the use of monoclonal antibodies with or without complement (40, 41), counter flow elutriation (42), and immunotoxins (43) or (2) positive selection of CD34+ hematopoietic stem cells from the graft which (currently the preferred method) usually via immunomagnetic beads with the CliniMACS CD34 Reagent System (Miltenyi Biotech, Gladbach, Germany) (44). The two methods do differ in efficacy with greater TCD being achieved by positive selection.

Pan T-Cell Depletion

An early concern with TCD was that it could affect the powerful GVL effect in HSCT which is also believed to be T-cell mediated. In a RCT in patients transplanted with marrow grafts, TCD was compared to conventional prophylaxis with CyA/MTX; the 3year disease free survival (DFS) was similar in both groups with lower rates of grade III-IV aGVHD with TCD. Relapse rates were however, higher with TCD, particularly in patients with chronic myelogenous leukemia (CML) (45). A large registry analysis also showed higher relapse rates with TCD (46). Subsequently, in a phase II trial with peripheral blood stem-cell (PBSC) transplantation (BMT CTN 0303), immunomagnetic beads were used for CD34 selection of the graft and TCD and relapse rates appeared to be comparable to historic controls while rates of acute and extensive chronic GVHD were favorable (47). Another trial comparing CD34 selected HLA-matched sibling HSCT with conventional prophylaxis showed comparable rates of GVHD, relapse and overall survival (48). Whether these results will hold up in the setting of an RCT has been tested in the recently completed multi-center RCT (NCT02345850) comparing ex-vivo CD34 selection to PTCy + MMF and conventional Tac/MTX prophylaxis, the results of which are eagerly awaited.

The other notable issue with pan-TCD has been a higher incidence of graft failure and slower immune reconstitution (IR) leading to higher rates of infectious complications, particularly viral infections (CMV, EBV) (49). The use of additional *in-vivo* TCD in the form of anti-thymocyte globulin (ATG), particularly in the haploidentical setting can potentially be helpful in achieving better engraftment rates (50). Other strategies to improve IR have incorporated direct T-cell add back strategies post stem-cell infusion (51) or the use of megadoses of CD34 selected cells (Perugia group) which seem to have a tolerizing effect (52).

Given these issues with pan-TCD, more selective methods of T-cell depletion aimed at preventing GVHD while preserving GVL are being explored with the availability of sophisticated clinical grade cell separation techniques.

Selective T-Cell Depletion Strategies

While a number of strategies have been attempted for selective TCD, they did not meet with lasting success. Depletion of CD5+T-cells and CD8+T-cells were tried in the 1990s; while rates of GVHD were encouraging, rates of relapse were high leading to the abandonment of these strategies. CD6 depletion is separately discussed in the section on T-cell modulation.

More recently Bleakley et al. have looked at CD45RA (naïve) TCD with the understanding that it is primarily the naïve T-cells in an allograft that are alloreactive. Unfortunately, this strategy has not produced encouraging results thus far. Bleakley et al. reported a first in human trial where they performed CD45RA (naïve) TCD via a two-step immunomagnetic bead-based procedure in 35 adult patients. Although 34/35 patients engrafted, rates of aGVHD were high in the 66% range. This was not improved when naïve TCD was combined with a/b TCD (described below) either and this approach is not widely used at this time.

α/β TCD

While a majority of T-cells express α/β receptors T-cell receptors (TCR), 2–10% of T-cells express γ/δ TCR. These γ/δ T-cells are believed to have important innate immune effects characterized by rapid cytokine release and killing of viral infected and tumor cells (53). This makes them an attractive candidate to potentially mediate GVL without inducing GVHD by the selective depletion of α/β T-cells. In a prospective single-arm pediatric trial in patients with acute leukemia, an encouraging GRFS of 70% was seen (54). The median follow-up for surviving patients in this study was 46 months. This approach is being tested in a number of other trials in pediatric and adult patients; in one of these a CNI-free GVHD prophylaxis strategy for acute leukemia patients undergoing 1–2 locus MMUD MAC HSCT (NCT03717480) is being looked at.

Modulating T-Cell Co-stimulatory/Co-inhibitory Pathways

During T-cell activation, following initial engagement of antigen by the TCR, a number of co-stimulatory and co-inhibitory signals come into play mediated by receptors on T-cells and APCs. This is true in acute GVHD as well. Hence the modulation of these co-stimulatory and co-inhibitory interactions is one of the new frontiers in the prophylaxis of GVHD.

CD28/CTLA-4 Blockade: Abatacept

The most promising of these has been blockade of the CD28/CTLA-4 axis. CD28 and CTLA-4 are both receptors on the T-cell which bind to B7-1/CD80 and B7-2/CD86 ligands on the APC; however, while CD28 is co-stimulatory, CTLA-4 is coinhibitory. Abatacept (CTLA4-Ig) is the soluble extra-cellular portion of CTLA-4 complexed with immunoglobulin heavy chain which blocks CD28 and CTLA-4, with more of an effect on CD28 leading ultimately to an inhibitory signal. Murine models from Blazar et al. showed that CD28/CTLA-4 blockade could reduce aGVHD lethality (55). Kean et al. performed a promising feasibility study in humans and reported results from a phase II RCT comparing standard of care (SOC) +abatacept to abatacept only in pediatric and adult patients. Grades III-IV aGVHD were significantly decreased and OS was improved in the abatacept arm (56). These impressive results have led to FDA breakthrough designation for this drug.

Since this approach blocks both stimulatory and inhibitory pathways, the concern for unwanted T-cell activation has been raised; hence more selective approaches to blocking CD28 are also being investigated. As an example, FR104 (CD28-specific pegylated-Fab') with and without sirolimus are being investigated in non-human primate models (57).

Enhancing Regulatory T-Cells

Tregs (CD4+CD25+Foxp3+) comprise a unique subset of T-lymphocytes that can be derived from the thymus or converted from CD4+CD25- cells (inducible or iTregs). Tregs play an important role in immune homeostasis and a favorable balance between Tregs and effector T-cells may be important to prevent GVHD. While *ex-vivo* expansion of Tregs is possible (58), there are concerns about their stability *ex-vivo*. Some preliminary data in alternative donor transplantation has shown that infusion of such expanded Tregs can be beneficial (59).

Hence, rather than the direct infusion of Tregs, other approaches have been attempted which can upregulate Tregs or enhance their functionality in the post-transplant immune milieu. One such approach involves invariant natural killer T (iNKT) cells.

Invariant Natural Killer T Cells

iNKT cells are unique in that they co-express both T and NK cell markers and therefore straddle both the innate and adaptive immune system with a semi-invariant TCR that recognizes glycolipid antigens presented by the major histocompatibility complex (MHC) class I-like molecule CD1d. They modulate the immune system via IL-4 and IL-10. In murine models, iNKT cells reduced GVHD both by a switch to a Th2 cytokine profile and/or IL-4-dependent Treg expansion. These mice were conditioned with a regimen incorporating total lymphoid irradiation plus ATG (TLI-ATG) (60, 61). This was then translated in a proof of concept study in humans with promising GVHD outcomes (62). An analysis of post-transplant immune reconstitution showed that low iNKT/T cell ratios were independently associated with

rates of acute GVHD (63) while another provocative study suggested that the larger numbers of iNKT cells in the donor graft correlated with improved GVHD free relapse free survival (64). Direct infusion of *ex-vivo* expanded iNKT cells is also an area of investigation.

In this contect, REGIMMUNE is a compound in which KRN7000, a synthetic alpha-galactosylceramide derivative and a CD1d ligand, is embedded in a lipid bilayer. REGIMMUNE has been shown to reduce aGVHD mortality by expanding Tregs via iNKT cells in murine models (65). In a Phase IIa trial REGIMMUNE in combination with sirolimus did reduce overall and acute GVHD although more mature data is awaited (66).

Targeting T-Cell Trafficking

Vedolizumab

Alloreactive CD8+ T-cells bound for the intestines express high levels of integrin $\beta 7$ ($\alpha 4\beta 7$) that binds to its ligand mucosal addressin cell adhesion molecule 1 (MAdCAM 1) in Peyer's patches and gut-associated lymphoid tissue (GALT) in the intestinal mucosa. Vedolizumab is a humanized moAb which prevents T-cell trafficking to the gut by targeting $\alpha 4\beta 7$ integrins on the T-cells. Early proof-of-concept and restrospective analyses have shown promising efficacy with vedolizumab in steroid refractory aGVHD (67). Given its effect on T-cell trafficking, an early critical event in GVHD pathogenesis, it was then tested in the context of prophylaxis in a phase 1b study where it was moderately safe with low rates of acute and chronic GVHD (68). A phase III RCT comparing vedolizumab + SOC prophylaxis to SOC is currently underway (NCT03657160).

Maraviroc

Maraviroc is an antagonist of CCR5, a chemokine receptor that has been implicated in T-cell trafficking during GVHD pathogenesis. Maraviroc appears to block lymphocyte chemotaxis without actually affecting T-cell function which made it an attractive candidate as a prophylactic agent. However, in a prospective non-randomized study from the BMT CTN, maraviroc in combination with standard Tac/MTX was not superior to standard of care and in this trial the PTCy/Tac/MMF arm fared the best (21).

Targeting Cytokine Pathways

Tocilizumab

Interleukin-6 (IL-6), an inflammatory cytokine has been shown to be one of the chief mediators of aGVHD in murine models (69). Therefore, IL-6 blocking agents could prevent aGVHD. In a phase II trial, tocilizumab, a humanized monoclonal antibody against the IL-6 receptor (IL-6R) was found to be promising (70); however, in a placebo-controlled phase III study from Australia, there was no significant difference in grades II-IV or III-IV aGVHD (71). This is a salient reminder that given the complex pathophysiology of aGVHD, with crosstalk between myriad cytokines and immune effector cells, targeting multiple cytokine pathways will be required for efficacy. These translational advances are summarized in **Table 1**.

Novel GVHD Prophylactic Strategies on the Horizon

There are novel therapies which have not yet been successfully translated to clinical practice but hold great promise. These therapies are based on innovative targets based on a more intricate understanding of the pathophysiology of GVHD.

Targeting Tissue Damage/Endothelial Injury Siglecs/CD24 Fc

As mentioned above in the section on the immunobiology of GVHD, conditioning regimen associated tissue damage exposes antigens which comprise pathogens or Pathogen-Associated Molecular Patterns (PAMPs) and components of damaged cells (Danger-Associated Molecular Patterns or DAMPs) which trigger activation of the innate immune system. Conversely Sialic-acid-binding-immunoglobulin-like lectins (Siglecs) are a particular class of pattern recognition receptors that downregulate innate immune responses (72). A number of Siglec homologs have been identified in mice and humans and are all characterized by immunoreceptor tyrosine-based inhibitory motifs (ITIMs) or ITIM-like regions in their intracellular domains.

A role for Siglecs in modulating adaptive T-cell mediated immune responses has also been proposed. Reddy et al. have shown that Siglec-G interacts with CD24c in murine models and this interaction CD24 suppresses TNF- α , IL-1 β , and IL-6 via NFkB and therefore is promising in the domain of GVHD prophylaxis (73).

Defibrotide

Defibrotide is a polydisperse mixture of predominantly single-stranded polydeoxyribonucleotides which in preclinical and human studies has demonstrated profibrinolytic, antithrombotic, anti-inflammatory and angio-protective effects ultimately resulting in stabilization of endothelial cells (74). Defibrotide is used in the treatment of veno-occlusive disease/sinusoidal obstruction syndrome (VOD/SOS), another morbid complication of allogeneic transplantation. Endothelial activation is also associated with transplant conditioning regimens and prime the host for GVHD. In a randomized phase II pediatric trial of defibrotide in VOD/SOS, the incidence and severity of aGVHD at days +30 and +100 were significantly better in the defibrotide-treated arm in patients who underwent HSCT (75). This signal is being further explored in a phase II randomized, open-label study comparing defibrotide + SOC vs. SOC alone in pediatric and adult patients for the prevention of aGVHD (NCT03339297).

T-Cell Modulation

Notch Pathway

The canonical NOTCH pathway has been shown to play a critical role in T-cell activation, differentiation, and function in aGVHD pathogenesis (5). Using humanized monoclonal antibodies, it has now been shown that Notch-deprived T-cells produce less inflammatory cytokines but proliferate normally, with a preferential increase in Tregs, without compromising GVL, mediated chiefly by NOTCH1, and Dll-4 (6). Selective

TABLE 1 | Translational advances and experimental strategies in GVHD prophylaxis.

	Translational advances in GVHD Prophylaxis		Experimental GVHD prophylaxis strategies	
1.	In vivo T-cell depletion/modulation		Targeting tissue damage/endothelial damage	
A.	Post-transplant cyclophosphamide	Phase II/III	Siglecs/CD24 Fc	Murine models
В.	Anti-thymocyte Globulin	Phase III	Defibrotide	Ongoing Phase II
C.	Sirolimus	Phase III		
2.	Ex-vivo T-cell depletion/modulation		T-cell modulation	
A.	Pan T-cell depletion	Phase II/III	Notch pathway	Pre-clinical
В.	Selective T-cell depletion		Mesenchymal stem-cells	Exploratory
	a/b T-cell depletion	Phase I/II		
	CD45RA (naïve) T-cell depletion	Phase I/II		
3.	Modulating T-cell co-stimulatory/inhibitory pathways		Modulating T-cell co-stimulatory/inhibitory pathways	
A.	CD28/CTLA-4 blockade/Abatacept	Phase III	OX40L blockade	Murine/non-human primate models
			ALPN101	Murine models, Phase I/II ongoing
			CD6 blockade: Itolizumab	Xenograft models, Phase I/II ongoing
4.	Enhancing Tregs (regulatory T-cells)		Targeting T-cell trafficking	
	iNKT-cells	Murine models, proof of concept in humans	PSGL-1	Murine models
	REGimmune	Phase IIa		
5.	Targeting T-cell trafficking		Subset T-cell depletion	
	Vedolizumab	Phase 1b, Phase III ongoing	Xenikos/T-guard	Exploratory
	Maraviroc	Phase III (negative study)		
6.	Targeting cytokine pathways		Targeting cytokine pathways	
	Tocilizumab	Phase III (negative study)	AAT (Alpha-1 antitrypsin)	Phase II/III ongoing
			JAK-STAT	Phase I ongoing

NOTCH blockade is an exciting new frontier and offers potential for clinical translation.

Mesenchymal Stem-Cells

Attempts have been made to utilize the immunomodulatory properties of MSCs for GVHD prophylaxis based on murine models of HLA-mismatched transplantation with co-transplantation of hematopoietic stem-cells and MSCs (76). Ning et al. showed in a small randomized trial, the incidence of grade II-IV aGVHD was 11.1% in the MSC group compared to 53.3% in the control group (77). However, the sample size in this trial was very small (n=25) and larger studies are needed to further study the effect of MSCs on preventing GVHD. A randomized phase II trial has also shown some beneficial effect on cGVHD and need to be further studied (78).

Targeting T-Cell Co-stimulatory Pathways (CD24 Fc) OX40L Blockade

OX40 (CD134) is a co-stimulatory receptor found on T-cells while its ligand OX40L is expressed on dendritic cells, B-cells, and endothelial cells. In 2003 Blazar et al. investigated the OX40 regulation of GVHD in murine models and found that antagonistic anti OX40L moAb or the use of OX40^{-/-}donor or recipient mice resulted in similar reduction in GVHD (79). Further, although OX40 was expressed on CD4 and CD8 cells, the effect of OX40 appeared to be mediated chiefly by CD4+ cells.

OX40 is also a strong negative regulator of Foxp3(+) Tregs (80) and therefore blockage could enhance Treg reconstitution which could be beneficial in GVHD. Tkachev et al. (39) then have shown that in non-human primate models, the combination of KY1005 (OX40L blocking antibody) and sirolimus has synergistic activity in reduction of GVHD mortality associated with control of both Th/Tc1 and Th/Tc17 activity. In addition, there was a Treg sparing effect with the combination. This exciting approach is now being translated to the clinic.

ALPN101

Inducible Costimulator (ICOS) is a member of the CD28/CTLA-4 family expressed on activated T-cells while the ICOS ligand (ICOSL), a B7 family member is constitutively expressed on B-cells, macrophages and dendritic cells and upregulated on APCs *via* TNF-alpha and lipopolysaccharides (LPS). In murine models of GVHD, blockade of the ICOS:ICOSL interactions via moAb or ICOS^{-/-} mice resulted in significant decrease in GVHD and GVHD related mortality both mediated by CD4+ and CD8+cells (81).

ALPN101 is a novel Fc fusion protein of a human ICOSL variant immunoglobulin domain (single domain vIgDTM) binding both ICOS and CD28 at higher affinity than wild-type molecules, designed to inhibit both the CD28 and ICOS pathways to dampen co-stimulatory responses during alloreactive T-cell activation. In a murine dose-ranging study, ALPN101 inhibited

GVHD responses at all doses and more significantly than the comparator belatacept, an approved CTLA-4-Fc protein CD28 pathway inhibitor (82).

A phase 1/2 dose finding study (BALANCE) is ongoing in patients with steroid sensitive and steroid refractory aGVHD (NCT04227938) investigating this potentially transformative effect and this is another promising target for GVHD prophylaxis.

CD6 Blockade: Itolizumab

CD6 is a co-stimulatory receptor on T-cells that binds to activated leukocyte cell adhesion molecule (ALCAM), a ligand on APCs and is involved in T-cell activation and trafficking. Historically CD6 T-cell depletion using a monoclonal antibody (moAb) was evaluated in a single-arm trial (n=112) with aGVHD rates in the 18% range using bone marrow grafts (41). More recently, itolizumab, a humanized anti-CD6 moAb was tested in xenograft models with some evidence that it could modulate T-cell activity (83). This molecule has also been fast-tracked by the FDA and is being tested in a phase I/II study for first-line treatment (with steroids) of severe aGVHD (NCT03763318) and may have a role in prophylaxis.

Targeting T-Cell Trafficking

PSGL-

P-selectin is one of a family of three glycosylated lectins (E, L, and P-selectin) which is constitutively expressed on the vascular endothelium of skin and bone marrow and inducibly expressed on other cells during inflammation. P-selectin is a receptor for PSGL-1, a glycoprotein strongly expressed on all leukocytes (84). PSGL-1 mRNA has been shown to be upregulated during GVHD in experimental models (85). P-selectin deficient mice were shown to have less GVHD morbidity and mortality; in addition T-cells were redirected from Peyer's patches and GALT to spleen and lymph nodes indicating that disruption of P-selectin interactions during GVHD pathogenesis can affect T-cell trafficking to target organs (84). Although it is likely that disruption of this pathway alone may not fully abrogate selectin interactions in GVHD, it is a promising new target.

Targeting Cytokine Pathways

Alpha-1 Antitrypsin

AAT is a liver derived serine protease inhibitor which can inhibit proinflammatory plasma cytokines and induce anti-inflammatory IL10 among other somewhat protean immunologic functions. It has also been shown to be involved in the *in-vivo* induction of Treg. In preclinical aGVHD models, AAT reduced inflammatory cytokines, altered the ratio of effector and regulatory T-cells and reduced levels of DAMPs (86). AAT has shown promise in early phase trials for SR aGVHD (87) and is being tested in phase III trials. This drug is also being tested in the prophylactic setting in a phase II/III randomized, multi-center, placebo-controlled study for prevention of aGVHD (NCT03805789).

IAK-STAT

The Janus kinase family comprises intra-cellular signaling proteins (JAK-1, 2, 3, and tyrosine kinase 2) involved in downstream transduction of various cytokine pathways (88). They are fundamentally involved in all three phases of GVHD pathogenesis by regulating the activity of APCs, T- and Blymphocytes (89). Pre-clinical studies showed that JAK-1/JAK-2 could reduce GVHD without affecting GVL (90, 91) including an effect on T-cell trafficking and enhancement of Tregs. JAK-1/JAK-3 inhibition also appears to reduce GVHD in murine models (92). Following on encouraging early phase studies with JAK-1/JAK-2 inhibitor ruxolitinib (Rux) in SR aGVHD, phase III data has now been reported (Rux vs. investigator's choice for SR aGVHD, REACH-2) which shows better overall response rates with Rux although a benefit in NRM could not be shown. JAK inhibition could be an exciting new frontier for GVHD prophylaxis as well. Choi et al. (93) showed that baricitinib (JAK1/JAK2 inhibitor) completely prevented GVHD in murine models without hampering GVL by multiple mechanisms including expansion of Tregs by preserving JAK3-STAT5 signaling; downregulation of CXCR3 and helper T cells 1 and 2.

So far clinical data is limited to a pilot study in myelofibrosis patients where a combination of ATG, ruxolitinib, and PTCy was used as GVHD prophylaxis with acceptable engraftment rates (94) and a single-arm study where Rux was used to replace CNIs in patients with CNI intolerance (95). Itacitinib, a selective JAK-1 inhibitor is being investigated in combination with CNI for primary prophylaxis of GVHD (GRAVITAS-119 trial, NCT03320642).

Subset TCD

Xenikos/T-Guard

Monoclonal antibodies conjugated with immunotoxins is a method of selective TCD that has been attempted in the past with CD5 as a target among others as described above. T-guard is a immunotoxin combination comprised of a mixture of anti-CD3 and anti-CD7 antibodies separately conjugated to recombinant ricin A (CD3/CD7-IT), which induces *in vivo* depletion of T cells and natural killer (NK) cells and suppresses T cell receptor activation.

This was first evaluated in a phase I/II trial in humans in SR aGVHD with a 50% response rate and manageable toxicities albeit with evidence of capillary leak and thrombotic microangiopathy (96). This may be a potential target for prophylaxis in the future. These experimental methods are summarized in **Table 1**.

Prevention of cGVHD

Although cGVHD is a distinct clinical and immunologic entity from aGVHD, there are limited interventions that specifically target the prevention of cGVHD. In general we know that patients who have less aGVHD will likely get less cGVHD and so the prevention of aGVHD is important in the prevention of cGVHD. In terms of donor and transplant related interventions, younger same-sex donors and the use of bone marrow product rather than PBSC has been shown to reduce rates of cGVHD (97).

T-cell directed approaches that have been quite successful include ATG and more recently PTCy as outlined above. B-cell directed approaches are an area of interest given our current understanding of the important role that B-cells play in the pathophysiology of cGVHD. Rituximab, a monoclonal antibody targeting CD20 was evaluated in a phase II trial with cGVHD rates in the 48% range cGVHD requiring immunosuppression in the 31% range (98). This was promising at the time and led to an ongoing randomized trial where obinutuzumab, another monoclonal B-cell directed antibody is being tested for cGVHD prevention (NCT02867384). It will be interesting to see how it will fare in comparison to PTCy.

Another area of interest is the augmentation of tolerance by the use of low-dose IL-2 (aldeskeukin) to enhance Treg reconstitution creating a favorable immunologic milieu for the prevention of cGVHD. This strategy has had success in the therapy of steroid-refractory cGVHD (99) and may have a role in prevention as well although remains investigational at this time.

DISCUSSION

GVHD prophylaxis has evolved over the last few decades from direct *in-vivo* and *ex-vivo* pan T-cell depletion strategies to more directed immunomodulatory strategies based on a more comprehensive understanding of GVHD immunobiology. Nevertheless, the basic backbone of CNI based prophylaxis has survived the test of time. We have touched upon this current standard in this review and further discussed important translational advances and exciting pre-clinical strategies which may be a part of future prophylactic regimens.

Of these translational advances, PTCy is arguably the most exciting and a potential replacement for standard CNI/MTX based prophylaxis in multiple transplant settings. Sometimes considered to be an elegant method of in vivo T-cell depletion, mechanistic studies have indicated that PTCy has a far more complex impact on the post-transplant immune system including a Treg sparing role which is of great interest in the community (20). In haploidentical transplants, PTCy based GVHD prophylaxis is the standard, typically incorporating a CNI and MMF in the most widely used regimens. Although rates of grades II-IV aGVHD are comparable in haploidentical transplants with PTCy based prophylaxis compared to CNI/MTX prophylaxis in MUD transplants, the rates of severe acute and chronic GVHD are far lower without significantly compromising relapse rates which makes it a very attractive strategy (100). In fact, the results in the haploidentical setting with PTCy based prophylaxis have been so impressive that this platform is now being tested in the HLA_matched setting where it is being compared to standard CNI/MTX based prophylaxis. Data from a small RCT (22) as well as a larger prospective trial (BMT CTN 1203) (21) in the reduced intensity setting have already generated encouraging signals where the PTCy arm performed better than the standard CNI arm as well as other potential novel strategies. Based on this data, some centers have already migrated to using PTCy based prophylaxis for matched unrelated donor and in some cases even in matched related donor transplantation. However, from a purist's viewpoint, data from a well-powered RCT is still not sufficient to change practice standards and the results of BMT CTN 1703 comparing PTCy based prophylaxis to CNI/MTX are eagerly awaited. In the myeloablative setting, the standard remains CNI/MTX although PTCy based prophylaxis will likely be tested in this setting as well. Other *in-vivo* TCD strategies such as ATG are still widely used, although the most profound effect of ATG appears to be on severe chronic GVHD and comes at the cost of poorer T-cell immune reconstitution and therefore more infectious complications. Sirolimus is another drug that has had promising results in reducing severe acute GVHD without much impact on chronic GVHD and is a reasonable alternative to CNI/MTX (34).

In the domain of *ex-vivo* TCD, pan TCD is still performed routinely in certain centers; once again with gains in the realm of severe chronic GVHD at the cost of more infectious complications. There have been some concerns about higher rates of relapse with *ex-vivo* TCD as well. These three important methods of T-cell depletion for GVHD prophylaxis, namely CNI/MTX, PTCy and *ex-vivo* pan-TCD have been compared in a multi-center RCT (BMT CTN 1301), the results of which are eagerly awaited as well. In the last decade the spotlight has shifted to methods of selective *ex-vivo* TCD with limited success in the clinical setting. a/b TCD which attempts to reduce GVHD without affecting GVL and can be performed without the use of post-transplant immunosuppression is promising and may be an important modality in the future.

Separate from direct TCD (in-vivo or ex-vivo), a new frontier in GVHD prophylaxis is targeting immune checkpoints which regulate T-cell activation. Given the dramatic success of checkpoint inhibitors in the world of solid tumor oncology, there has been tremendous interest and a much better understanding of these checkpoints in recent years. In the case of GVHD of course, researchers have tried to downregulate rather than upregulate T-cell activation following initial antigen engagement by the Tcell receptor complex. Although there are a number of molecules being tested at the bench and detailed in this review, the most promising of these has been blockade of the CD28/CTLA-4 axis with Abatacept (CTLA-Ig) with an eventual inhibitory signal downstream to the T-cell. Results from a RCT with pediatric and adult patients has shown a dramatic reduction in severe acute and chronic GVHD including in mismatched unrelated donors (56). With FDA breakthrough status, this molecule has the potential to be an integral part of GVHD prophylaxis in the future although it is unclear if it is more effective than PTCy based prophylaxis. Certainly cyclophosphamide, a drug that has been used for decades, is far more affordable and therefore a platform easily generalizable in more resource poor settings.

Targeting T-cell trafficking, an early event in GVHD prophylaxis, is being tried with integrin blockers such as vedolizumab. It is logical that inhibiting the very movement of effector T-lymphocytes to target organs should better prevent GVHD rather than trying to arrest the widespread inflammation and cytokine cascades which characterize the final common pathway in GVHD pathogenesis. As a testimony to that, when tocilizumab an IL-6 blocker was evaluated, despite promising phase II single-arm data, was not more effective than standard

prophylaxis in a phase III RCT (71). This has in fact been the case with numerous promising prophylactic therapies which perform well in single-arm studies but have not been a home run in well-designed phase III studies.

Within the limitations of this review, we have highlighted some of the exciting pre-clinical science that has the potential to translate into effective prophylactic therapies which target GVHD pathogenesis beyond direct T-cell depletion. Targeting the interaction between DAMPs/ PAMPs on APCs and Siglecs with a downstream inhibitory effect on cytokine cascades as well as investigating a role for Siglecs in modulating adaptive T-cell mediated immunity are areas of interest (73). Endothelial damage, another inciting pro-inflammatory event in GVHD pathogenesis is being targeted by drugs like defibrotide which have enjoyed tremendous success in the therapy of VOD. Targeting selectin interactions such as PSGL-1 (84) is another developing area in the field of GVHD therapeutics and prophylaxis.

Pathways critical in T-cell modulation during activation and proliferation such as the Notch pathway has been an area of

great interest although not ready for translation at this time (6). Bolstered by the success of Abatacept, other molecules targeting checkpoints such as OX40L blockade (including combination with sirolimus) (39), blockade of the ICOS: ICOSL interaction with ALPN101 (81) and CD6 blockade (Itolizumab) (83) are extremely exciting. The role of AAT in prophylaxis both as an immunomodulator as well as in opposing inflammatory cytokines is being looked at.

In conclusion, while GVHD prophylaxis in 2020 still incorporates the traditional paradigms of CNI based prophylaxis, PTCy is knocking on the door and a number of exciting new translational therapies and pre-clinical advances are on the horizon which promise to challenge the established paradigms.

AUTHOR CONTRIBUTIONS

JHA supervised and edited manuscript along with MG. All authors contributed to the article and approved the submitted version.

REFERENCES

- Khoury HJ, Wang T, Hemmer MT, Couriel D, Alousi A, Cutler C, et al. Improved survival after acute graft-versus-host disease diagnosis in the modern era. *Haematologica*. (2017) 102:958–66. doi: 10.3324/haematol.2016.156356
- Antin JH, Ferrara JL. Cytokine dysregulation and acute graft-versus-host disease. Blood. (1992) 80:2964–8. doi: 10.1182/blood.V80.12.2964.2964
- Zeiser R, Blazar BR. Acute graft-versus-host disease biologic process, prevention, and therapy. N Engl J Med. (2017) 377:2167–79. doi: 10.1056/NEJMra1609337
- 4. Ferrara JLM, Levine JE, Reddy P, Holler E. Graft-versus-host disease. *Lancet*. (2009) 373:1550–61. doi: 10.1016/S0140-6736(09)60237-3
- Zhang Y, Sandy AR, Wang J, Radojcic V, Shan GT, Tran IT, et al. Notch signaling is a critical regulator of allogeneic CD4+ T-cell responses mediating graft-versus-host disease. *Blood.* (2011) 117:299– 308. doi: 10.1182/blood-2010-03-271940
- Tran IT, Sandy AR, Carulli AJ, Ebens C, Chung J, Shan GT, et al. Blockade of individual Notch ligands and receptors controls graft-versus-host disease. J Clin Invest. (2013) 123:1590–604. doi: 10.1172/JCI65477
- 7. Sakaguchi S, Yamaguchi T, Nomura T, Ono M. Regulatory T cells and immune tolerance. Cell. (2008) 133:775–87. doi: 10.1016/j.cell.2008.05.009
- Krenger W, Ferrara JLM. Graft-versus-host disease and the Th1/Th2 paradigm. *Immunol Res.* (1996) 15:50–73. doi: 10.1007/BF02918284
- Carlson MJ, West ML, Coghill JM, Panoskaltsis-Mortari A, Blazar BR, Serody JS. In vitro-differentiated TH17 cells mediate lethal acute graftversus-host disease with severe cutaneous and pulmonary pathologic manifestations. Blood. (2009) 113:1365–74. doi: 10.1182/blood-2008-06-162420
- Yu Y, Wang D, Liu C, Kaosaard K, Semple K, Anasetti C, et al. Prevention of GVHD while sparing GVL effect by targeting Th1 and Th17 transcription factor T-bet and RORγt in mice. Blood. (2011) 118:5011– 20. doi: 10.1182/blood-2011-03-340315
- 11. Zeiser R, Blazar BR. Pathophysiology of chronic graft-versushost disease and therapeutic targets. N Engl J Med. (2017) 377:2565–79. doi: 10.1056/NEJMra1703472
- Cooke KR, Luznik L, Sarantopoulos S, Hakim FT, Jagasia M, Fowler DH, et al.
 The biology of chronic graft-versus-host disease: a task force report from the national institutes of health consensus development project on criteria for clinical trials in chronic graft-versus-host disease. *Biol Blood Marrow Transplant*. (2017) 23:211–34. doi: 10.1016/j.bbmt.2016.09.023

- 13. Nash RA, Antin JH, Karanes C, Fay JW, Avalos BR, Yeager AM, et al. Phase 3 study comparing methotrexate and tacrolimus with methotrexate and cyclosporine for prophylaxis of acute graft-versus-host disease after marrow transplantation from unrelated donors. *Blood.* (2000) 96:2062–8.
- Ratanatharathorn V, Nash RA, Przepiorka D, Devine SM, Klein JL, Weisdorf D, et al. Phase III study comparing methotrexate and tacrolimus (prograf, FK506) with methotrexate and cyclosporine for graft-versus-host disease prophylaxis after HLA-identical sibling bone marrow transplantation. *Blood*. (1998) 92:2303–14.
- Perkins J, Field T, Kim J, Kharfan-Dabaja MA, Fernandez H, Ayala E, et al. A randomized phase II trial comparing tacrolimus and mycophenolate mofetil to tacrolimus and methotrexate for acute graft-versus-host disease prophylaxis. Biol Blood Marrow Transplant. (2010) 16:937–47. doi: 10.1016/j.bbmt.2010.01.010
- Luznik L, O'Donnell PV, Symons HJ, Chen AR, Leffell MS, Zahurak M, et al. HLA-haploidentical bone marrow transplantation for hematologic malignancies using non-myeloablative conditioning and high-dose, posttransplantation cyclophosphamide. *Biol Blood Marrow Transplant*. (2008) 14:641–50. doi: 10.1016/j.bbmt.2008.03.005
- Solomon SR, Sizemore C, Zhang X, Brown S, Connor K, Morris LE, et al. TBI-based myeloablative haploidentical stem cell transplantation is a safe and effective alternative to unrelated donor transplantation in patients without matched sibling donors. *Blood.* (2014) 124:426. doi: 10.1182/blood.V124.21.426.426
- Raiola AM, Dominietto A, Grazia C di, Lamparelli T, Gualandi F, Ibatici A, et al. Unmanipulated haploidentical transplants compared with other alternative donors and matched sibling grafts. *Biol Blood Marrow Transplant*. (2014) 20:1573–9. doi: 10.1016/j.bbmt.2014.05.029
- Symons HJ, Chen AR, Luznik L, Kasamon YL, Meade JB, Jones RJ, et al. Myeloablative haploidentical bone marrow transplantation with T cell replete grafts and post-transplant cyclophosphamide: results of a phase II clinical trial. *Blood*. (2011) 118:4151. doi: 10.1182/blood.V118.21.4151.4151
- Wachsmuth LP, Patterson MT, Eckhaus MA, Venzon DJ, Gress RE, Kanakry CG. Posttransplantation cyclophosphamide prevents graft-versushost disease by inducing alloreactive T cell dysfunction and suppression. J Clin Invest. (2019) 129:2357–73. doi: 10.1172/JCI124218
- 21. Bolaños-Meade J, Reshef R, Fraser R, Fei M, Abhyankar S, Al-Kadhimi Z, et al. Three prophylaxis regimens (tacrolimus, mycophenolate mofetil, and cyclophosphamide; tacrolimus, methotrexate, and bortezomib; or tacrolimus, methotrexate, and maraviroc) versus tacrolimus and methotrexate for prevention of graft-versus-host

- disease with haemopoietic cell transplantation with reduced-intensity conditioning: a randomised phase 2 trial with a non-randomised contemporaneous control group (BMT CTN 1203). *Lancet Haematol.* (2019) 6:e132–43. doi: 10.1016/S2352-3026(18)30221-7
- De Jong CN, Meijer E, Bakunina K, Nur E, van Marwijk Kooij M, de Groot MR, et al. Post-transplantation cyclophosphamide after allogeneic hematopoietic stem cell transplantation: results of the prospective randomized HOVON-96 trial in recipients of matched related and unrelated donors. *Blood.* (2019) 134:1. doi: 10.1182/blood-2019-124659
- Wang Y, Wu D-P, Liu Q-F, Xu L-P, Liu K-Y, Zhang X-H, et al. Low-dose posttransplant cyclophosphamide and anti-thymocyte globulin as an effective strategy for GVHD prevention in haploidentical patients. *J Hematol Oncol*. (2019) 12:88. doi: 10.1186/s13045-019-0781-y
- 24. Bacigalupo A, Lamparelli T, Bruzzi P, Guidi S, Alessandrino PE, di Bartolomeo P, et al. Antithymocyte globulin for graft-versus-host disease prophylaxis in transplants from unrelated donors: 2 randomized studies from Gruppo Italiano Trapianti Midollo Osseo (GITMO). Blood. (2001) 98:2942–7. doi: 10.1182/blood.V98.10.2942
- Bacigalupo A, Lamparelli T, Barisione G, Bruzzi P, Guidi S, Alessandrino PE, et al. Thymoglobulin prevents chronic graft-versus-host disease, chronic lung dysfunction, and late transplant-related mortality: longterm follow-up of a randomized trial in patients undergoing unrelated donor transplantation. *Biol Blood Marrow Transplant*. (2006) 12:560– 5. doi: 10.1016/j.bbmt.2005.12.034
- Finke J, Bethge WA, Schmoor C, Ottinger HD, Stelljes M, Zander AR, et al. Standard graft-versus-host disease prophylaxis with or without anti-T-cell globulin in haematopoietic cell transplantation from matched unrelated donors: a randomised, open-label, multicentre phase 3 trial. *Lancet Oncol.* (2009) 10:855–64. doi: 10.1016/S1470-2045(09)70225-6
- 27. Feng X, Kajigaya S, Solomou EE, Keyvanfar K, Xu X, Raghavachari N, et al. Rabbit ATG but not horse ATG promotes expansion of functional CD4+CD25highFOXP3+ regulatory T cells in vitro. Blood. (2008) 111:3675–83. doi: 10.1182/blood-2008-01-130146
- 28. Mohty M, Labopin M, Balère ML, Socié G, Milpied N, Tabrizi R, et al. Antithymocyte globulins and chronic graft-vs.-host disease after myeloablative allogeneic stem cell transplantation from HLA-matched unrelated donors: a report from the Sociéte Française de Greffe de Moelle et de Thérapie Cellulaire. Leukemia. (2010) 24:1867–74. doi: 10.1038/leu.2010.200
- Soiffer RJ, Kim HT, McGuirk J, Horwitz ME, Johnston L, Patnaik MM, et al. Prospective, randomized, double-blind, phase III clinical trial of anti–T-lymphocyte globulin to assess impact on chronic graft-versushost disease–free survival in patients undergoing HLA-matched unrelated myeloablative hematopoietic cell transplantation. *J Clin Oncol.* (2017) 35:4003–4011. doi: 10.1200/JCO.2017.75.8177
- Podgorny PJ, Ugarte-Torres A, Liu Y, Williamson TS, Russell JA, Storek J. High rabbit-antihuman thymocyte globulin levels are associated with low likelihood of graft-vs.-host disease and high likelihood of posttransplant lymphoproliferative disorder. *Biol Blood Marrow Transplant*. (2010) 16:915– 26. doi: 10.1016/j.bbmt.2010.02.027
- 31. Admiraal R, Nierkens S, de Witte MA, Petersen EJ, Fleurke G-J, Verrest L, et al. Association between anti-thymocyte globulin exposure and survival outcomes in adult unrelated haemopoietic cell transplantation: a multicentre, retrospective, pharmacodynamic cohort analysis. *Lancet Haematol*. (2017) 4:e183–91. doi: 10.1016/S2352-3026(17)30029-7
- Chang Y-J, Wang Y, Mo X-D, Zhang X-H, Xu L-P, Yan C-H, et al. Optimal dose of rabbit thymoglobulin in conditioning regimens for unmanipulated, haploidentical, hematopoietic stem cell transplantation: Long-term outcomes of a prospective randomized trial. *Cancer*. (2017) 123:2881–92. doi: 10.1002/cncr.30540
- 33. Chang Y-J, Wu D-P, Lai Y-R, Liu Q-F, Sun Y-Q, Hu J, et al. Antithymocyte globulin for matched sibling donor transplantation in patients with hematologic malignancies: a multicenter, open-label, randomized controlled study. *J Clin Oncol.* (2020) 38:3367–76. doi: 10.1200/JCO.20.00150
- Cutler C, Logan B, Nakamura R, Johnston L, Choi S, Porter D, et al. Tacrolimus/sirolimus vs. tacrolimus/methotrexate as GVHD prophylaxis after matched, related donor allogeneic HCT. *Blood.* (2014) 124:1372– 7. doi: 10.1182/blood-2014-04-567164

- Cutler C, Stevenson K, Kim HT, Richardson P, Ho VT, Linden E, et al. Sirolimus is associated with veno-occlusive disease of the liver after myeloablative allogeneic stem cell transplantation. *Blood*. (2008) 112:4425– 31. doi: 10.1182/blood-2008-07-169342
- Armand P, Kim HT, Sainvil M-M, Lange PB, Giardino AA, Bachanova V, et al. The addition of sirolimus to the graft-versus-host disease prophylaxis regimen in reduced intensity allogeneic stem cell transplantation for lymphoma: a multicentre randomized trial. *Br J Haematol*. (2016) 173:96– 104. doi: 10.1111/bjh.13931
- 37. Sandmaier BM, Kornblit B, Storer BE, Olesen G, Maris MB, Langston AA, et al. Addition of sirolimus to standard cyclosporine plus mycophenolate mofetil-based graft-versus-host disease prophylaxis for patients after unrelated non-myeloablative haemopoietic stem cell transplantation: a multicentre, randomised, phase 3 trial. *Lancet Haematol.* (2019) 6:e409–18. doi: 10.1016/S2352-3026(19)30088-2
- Solomon SR, Sanacore M, Zhang X, Brown S, Holland K, Morris LE, et al. Calcineurin inhibitor–free graft-versus-host disease prophylaxis with post-transplantation cyclophosphamide and brief-course sirolimus following reduced-intensity peripheral blood stem cell transplantation. *Biol Blood Marrow Transplant*. (2014) 20:1828–34. doi: 10.1016/j.bbmt.2014.07.020
- Tkachev V, Furlan SN, Watkins B, Hunt DJ, Zheng HB, Panoskaltsis-Mortari A, et al. Combined OX40L and mTOR blockade controls effector T cell activation while preserving Treg reconstitution after transplant. Sci Transl Med. (2017) 9:eaan3085 doi: 10.1126/scitranslmed.aan3085
- 40. Reinherz EL, Geha R, Rappeport JM, Wilson M, Penta AC, Hussey RE, et al. Reconstitution after transplantation with T-lymphocyte-depleted HLA haplotype-mismatched bone marrow for severe combined immunodeficiency. *Proc Natl Acad Sci USA*. (1982) 79:6047–51. doi: 10.1073/pnas.79.19.6047
- 41. Soiffer RJ, Murray C, Mauch P, Anderson KC, Freedman AS, Rabinowe SN, et al. Prevention of graft-versus-host disease by selective depletion of CD6-positive T lymphocytes from donor bone marrow. *J Clin Oncol.* (1992) 10:1191–200. doi: 10.1200/JCO.1992.10.7.1191
- Wagner JE, Donnenberg AD, Noga SJ, Cremo CA, Gao IK, Yin HJ, et al. Lymphocyte depletion of donor bone marrow by counterflow centrifugal elutriation: results of a phase I clinical trial. *Blood.* (1988) 72:1168– 76. doi: 10.1182/blood.V72.4.1168.1168
- 43. Antin JH, Bierer BE, Smith BR, Ferrara J, Guinan EC, Sieff C, et al. Selective depletion of bone marrow T lymphocytes with anti-CD5 monoclonal antibodies: effective prophylaxis for graft-versus-host disease in patients with hematologic malignancies. *Blood.* (1991) 78:2139–49. doi: 10.1182/blood.V78.8.2139.bloodjournal7882139
- Hobbs GS, Perales M-A. Effects of T-cell depletion on allogeneic hematopoietic stem cell transplantation outcomes in AML patients. J Clin Med. (2015) 4:488–503. doi: 10.3390/jcm4030488
- 45. Wagner JE, Thompson JS, Carter SL, Kernan NA, Unrelated donor marrow transplantation trial. Effect of graft-versus-host disease prophylaxis on 3-year disease-free survival in recipients of unrelated donor bone marrow (T-cell Depletion Trial): a multi-centre, randomised phase II-III trial. *Lancet*. (2005) 366:733–41. doi: 10.1016/S0140-6736(05)66996-6
- Horowitz MM, Gale RP, Sondel PM, Goldman JM, Kersey J, Kolb HJ, et al. Graft-versus-leukemia reactions after bone marrow transplantation. *Blood*. (1990) 75:555–62. doi: 10.1182/blood.V75.3.555.555
- 47. Devine SM, Carter S, Soiffer RJ, Pasquini MC, Hari PN, Stein A, et al. Low risk of chronic graft versus host disease and relapse associated with T-cell depleted peripheral blood stem cell transplantation for acute myeloid leukemia in first remission: results of the blood and marrow transplant clinical trials network (BMT CTN) Protocol 0303. Biol Blood Marrow Transplant. (2011) 17:1343–51. doi: 10.1016/j.bbmt.2011. 02.002
- 48. Pasquini MC, Devine S, Mendizabal A, Baden LR, Wingard JR, Lazarus HM, et al. Comparative outcomes of donor graft CD34+ selection and immune suppressive therapy as graft-versus-host disease prophylaxis for patients with acute myeloid leukemia in complete remission undergoing HLA-matched sibling allogeneic hematopoietic cell transplantation. *J Clin Oncol.* (2012) 30:3194–201. doi: 10.1200/JCO.2012.41.7071
- Small TN, Avigan D, Dupont B, Smith K, Black P, Heller G, et al. Immune reconstitution following T-cell depleted bone marrow transplantation: effect

- of age and posttransplant graft rejection prophylaxis. Biol Blood Marrow Transplant. (1997) 3:65–75.
- Aversa F, Tabilio A, Velardi A, Cunningham I, Terenzi A, Falzetti F, et al. Treatment of high-risk acute leukemia with T-cell-depleted stem cells from related donors with one fully mismatched HLA haplotype. N Engl J Med. (1998) 339:1186–93. doi: 10.1056/NEJM199810223391702
- 51. Geyer MB, Ricci AM, Jacobson JS, Majzner R, Duffy D, Van de Ven C, et al. T cell depletion utilizing CD34(+)stem cell selection and CD3(+) addback from unrelated adult donors in paediatric allogeneic stem cell transplantation recipients. Br J Haematol. (2012) 157:205–19. doi: 10.1111/j.1365-2141.2012.09048.x
- 52. Rachamim N, Gan J, Segall H, Krauthgamer R, Marcus H, Berrebi A, et al. Tolerance induction by "megadose" hematopoietic transplants: donor-type human CD34 stem cells induce potent specific reduction of host anti-donor cytotoxic T lymphocyte precursors in mixed lymphocyte culture. *Transplantation*. (1998) 65:1386–93. doi: 10.1097/00007890-199805270-00017
- 53. Daniele N, Scerpa MC, Caniglia M, Bernardo ME, Rossi C, Ciammetti C, et al. Transplantation in the onco-hematology field: focus on the manipulation of $\alpha\beta$ and $\gamma\delta$ T cells. *Pathol Res Pract.* (2012) 208:67–73. doi: 10.1016/j.prp.2011.10.006
- 54. Locatelli F, Merli P, Pagliara D, Li Pira G, Falco M, Pende D, et al. Outcome of children with acute leukemia given HLA-haploidentical HSCT after $\alpha\beta$ T-cell and B-cell depletion. *Blood.* (2017) 130:677–85. doi: 10.1182/blood-2017-04-779769
- Blazar BR, Taylor PA, Linsley PS, Vallera DA. In vivo blockade of CD28/CTLA4: B7/BB1 interaction with CTLA4-Ig reduces lethal murine graft-versus-host disease across the major histocompatibility complex barrier in mice. Blood. (1994) 83:3815–25. doi: 10.1182/blood.V83.12.3815.3815
- 56. Watkins B, Qayed M, Bratrude B, Betz K, Brown M, Rhodes J, et al. T cell costimulation blockade with abatacept nearly eliminates early severe acute graft versus host disease after HLA-mismatched (7/8 HLA matched) unrelated donor transplant, with a favorable impact on disease-free and overall survival. *Blood*. (2017) 130:212. doi: 10.1182/blood.V130.Suppl_1.212.212
- Watkins BK, Tkachev V, Furlan SN, Hunt DJ, Betz K, Yu A, et al. CD28 blockade controls T cell activation to prevent graft-versus-host disease in primates. J Clin Invest. (2018) 128:3991–4007. doi: 10.1172/JCI98793
- 58. Taylor PA, Lees CJ, Blazar BR. The infusion of *ex vivo* activated and expanded CD4(+)CD25(+) immune regulatory cells inhibits graft-versus-host disease lethality. *Blood.* (2002) 99:3493–9. doi: 10.1182/blood.V99.10.3493
- Brunstein CG, Miller JS, Cao Q, McKenna DH, Hippen KL, Curtsinger J, et al. Infusion of *ex vivo* expanded T regulatory cells in adults transplanted with umbilical cord blood: safety profile and detection kinetics. *Blood*. (2011) 117:1061–70. doi: 10.1182/blood-2010-07-293795
- 60. Pillai AB, George TI, Dutt S, Teo P, Strober S. Host NKT cells can prevent graft-versus-host disease and permit graft antitumor activity after bone marrow transplantation. *J Immunol*. (2007) 178:6242–51. doi: 10.4049/jimmunol.178.10.6242
- 61. Pillai AB, George TI, Dutt S, Strober S. Host natural killer T cells induce an interleukin-4-dependent expansion of donor CD4+CD25+Foxp3+ T regulatory cells that protects against graft-versus-host disease. *Blood.* (2009) 113:4458–67. doi: 10.1182/blood-2008-06-165506
- Lowsky R, Takahashi T, Liu YP, Dejbakhsh-Jones S, Grumet FC, Shizuru JA, et al. Protective conditioning for acute graft-versus-host disease. N Engl J Med. (2005) 353:1321–31. doi: 10.1056/NEJMoa050642
- 63. Rubio M-T, Moreira-Teixeira L, Bachy E, Bouillié M, Milpied P, Coman T, et al. Early posttransplantation donor-derived invariant natural killer T-cell recovery predicts the occurrence of acute graft-versus-host disease and overall survival. *Blood.* (2012) 120:2144–54. doi: 10.1182/blood-2012-01-404673
- Malard F, Labopin M, Chevallier P, Guillaume T, Duquesne A, Rialland F, et al. Larger number of invariant natural killer T cells in PBSC allografts correlates with improved GVHD-free and progression-free survival. *Blood*. (2016) 127:1828–35. doi: 10.1182/blood-2015-12-688739
- Duramad O, Laysang A, Li J, Nguyen N, Ishii Y, Namikawa R. A liposomal formulation of KRN7000 (RGI-2001) potently reduces GvHD lethality

- through the expansion of CD4+Foxp3+ regulatory T cells in murine models. Blood. (2008) 112:3500. doi: 10.1182/blood.V112.11.3500.3500
- 66. Chen Y-B, Efebera YA, Johnston L, Ball ED, Avigan D, Lekakis LJ, et al. Increased Foxp3+Helios+ regulatory T cells and decreased acute graft-versus-host disease after allogeneic bone marrow transplantation in patients receiving sirolimus and RGI-2001, an activator of invariant natural killer T cells. Biol Blood Marrow Transplant. (2017) 23:625–34. doi: 10.1016/j.bbmt.2017.01.069
- 67. Fløisand Y, Lazarevic VL, Maertens J, Mattsson J, Shah NN, Zachée P, et al. Safety and effectiveness of vedolizumab in patients with steroid-refractory gastrointestinal acute graft-versus-host disease: a retrospective record review. *Biol Blood Marrow Transplant*. (2019) 25:720–7. doi: 10.1016/j.bbmt.2018.11.013
- 68. Chen Y-B, Shah NN, Renteria AS, Cutler C, Jansson J, Akbari M, et al. Vedolizumab for prevention of graft-versus-host disease after allogeneic hematopoietic stem cell transplantation. *Blood Adv.* (2019) 3:4136–46. doi: 10.1182/bloodadvances.2019000893
- Tawara I, Koyama M, Liu C, Toubai T, Thomas D, Evers R, et al. Interleukin-6 modulates graft-versus-host responses after experimental allogeneic bone marrow transplantation. Clin Cancer Res. (2011) 17:77– 88. doi: 10.1158/1078-0432.CCR-10-1198
- Drobyski WR, Szabo A, Zhu F, Keever-Taylor C, Hebert KM, Dunn R, et al. Tocilizumab, tacrolimus and methotrexate for the prevention of acute graft-versus-host disease: low incidence of lower gastrointestinal tract disease. Haematologica. (2018) 103:717–27. doi: 10.3324/haematol.2017.183434
- Kennedy GA, Tey S-K, Curley C, Butler JP, Misra A, Subramoniapillai E, et al. Results of a phase III double-blind study of the addition of tocilizumab vs. placebo to cyclosporin/methotrexate Gvhd prophylaxis after HLA-matched allogeneic stem cell transplantation. *Blood.* (2019) 134:368. doi: 10.1182/blood-2019-126285
- Crocker PR, Paulson JC, Varki A. Siglecs and their roles in the immune system. Nat Rev Immunol. (2007) 7:255–66. doi: 10.1038/nri2056
- Toubai T, Hou G, Mathewson N, Liu C, Wang Y, Oravecz-Wilson K, et al. Siglec-G-CD24 axis controls the severity of graft-versus-host disease in mice. *Blood*. (2014) 123:3512–23. doi: 10.1182/blood-2013-12-545335
- Richardson PG, Carreras E, Iacobelli M, Nejadnik B. The use of defibrotide in blood and marrow transplantation. *Blood Adv.* (2018) 2:1495– 509. doi: 10.1182/bloodadvances.2017008375
- Richardson PG, Soiffer RJ, Antin JH, Uno H, Jin Z, Kurtzberg J, et al. Defibrotide for the treatment of severe hepatic veno-occlusive disease and multiorgan failure after stem cell transplantation: a multicenter, randomized, dose-finding trial. *Biol Blood Marrow Transplant*. (2010) 16:1005–17. doi: 10.1016/j.bbmt.2010.02.009
- Chung NG, Jeong DC, Park SJ, Choi BO, Cho B, Kim HK, et al. Cotransplantation of marrow stromal cells may prevent lethal graft-versus-host disease in major histocompatibility complex mismatched murine hematopoietic stem cell transplantation. *Int J Hematol.* (2004) 80:370–6. doi: 10.1532/IJH97.A30409
- 77. Ning H, Yang F, Jiang M, Hu L, Feng K, Zhang J, et al. The correlation between cotransplantation of mesenchymal stem cells and higher recurrence rate in hematologic malignancy patients: outcome of a pilot clinical study. Leukemia. (2008) 22:593–9. doi: 10.1038/sj.leu.2405090
- 78. Gao L, Zhang Y, Hu B, Liu J, Kong P, Lou S, et al. Phase II multicenter, randomized, double-blind controlled study of efficacy and safety of umbilical cord-derived mesenchymal stromal cells in the prophylaxis of chronic graft-versus-host disease after HLA-haploidentical stem-cell transplantation. *J Clin Oncol.* (2016) 34:2843–50. doi: 10.1200/JCO.2015.65.3642
- Blazar BR, Sharpe AH, Chen AI, Panoskaltsis-mortari A, Lees C, Akiba H, et al. Ligation of OX40 (CD134) regulates graft-versus-host disease (GVHD) and graft rejection in allogeneic bone marrow transplant recipients. *Blood*. 101:3741–8. doi: 10.1182/blood-2002-10-3048
- 80. Vu MD, Xiao X, Gao W, Degauque N, Chen M, Kroemer A, et al. OX40 costimulation turns off Foxp3+ Tregs. *Blood*. (2007) 110:2501–10. doi: 10.1182/blood-2007-01-070748
- 81. Taylor PA, Panoskaltsis-Mortari A, Freeman GJ, Sharpe AH, Noelle RJ, Rudensky AY, et al. Targeting of inducible costimulator (ICOS) expressed on alloreactive T cells down-regulates graft-versus-host disease (GVHD)

- and facilitates engraftment of allogeneic bone marrow (BM). *Blood.* (2005) 105:3372–80. doi: 10.1182/blood-2004-10-3869
- 82. Dillon SR, Yang J, Lewis KE, Evans LS, Mudri S, Wu R, et al. Alpn-101, a dual ICOS/CD28 antagonist, demonstrates potent and dose-dependent suppression of graft vs. host disease (GvHD) in a human/NSG mouse xenograft model, with activity superior to CD28 or ICOS single pathway antagonists. *Biol Blood Marrow Transplant*. (2019) 25:S290–1. doi: 10.1016/j.bbmt.2018.12.666
- 83. Ng CT, Ampudia J, Soiffer RJ, Ritz J, Connelly S. Itolizumab as a potential therapeutic for the prevention and treatment of graft vs. host disease. *Blood.* (2019) 134:5603. doi: 10.1182/blood-2019-122787
- 84. Lu SX, Holland AM, Na I-K, Terwey TH, Alpdogan O, Bautista JL, et al. Absence of P-selectin in recipients of allogeneic bone marrow transplantation ameliorates experimental graft-versus-host-disease. *J Immunol.* (2010) 185:1912–9. doi: 10.4049/jimmunol.0903148
- Ichiba T, Teshima T, Kuick R, Misek DE, Liu C, Takada Y, et al. Early changes in gene expression profiles of hepatic GVHD uncovered by oligonucleotide microarrays. *Blood*. (2003) 102:763–71. doi: 10.1182/blood-2002-09-2748
- 86. Tawara I, Sun Y, Lewis EC, Toubai T, Evers R, Nieves E, et al. Alpha-1-antitrypsin monotherapy reduces graft-versus-host disease after experimental allogeneic bone marrow transplantation. *Proc Natl Acad Sci* USA. (2012) 109:564–9. doi: 10.1073/pnas.1117665109
- Magenau JM, Goldstein SC, Peltier D, Soiffer RJ, Braun T, Pawarode A, et al. α1-Antitrypsin infusion for treatment of steroid-resistant acute graft-versus-host disease. *Blood*. (2018) 131:1372–9. doi: 10.1182/blood-2017-11-815746
- Schwartz DM, Bonelli M, Gadina M, O'Shea JJ. Type I/II cytokines, JAKs, and new strategies for treating autoimmune diseases. *Nat Rev Rheumatol*. (2016) 12:25–36. doi: 10.1038/nrrheum.2015.167
- Schroeder MA, Choi J, Staser K, DiPersio JF. The role of janus kinase signaling in graft-versus-host disease and graft versus leukemia. *Biol Blood Marrow Transplant*. (2018) 24:1125–34. doi: 10.1016/j.bbmt.2017.12.797
- Choi J, Ziga ED, Ritchey J, Collins L, Prior JL, Cooper ML, et al. IFNγR signaling mediates alloreactive T-cell trafficking and GVHD. *Blood*. (2012) 120:4093–103. doi: 10.1182/blood-2012-01-403196
- 91. Choi J, Cooper ML, Alahmari B, Ritchey J, Collins L, Holt M, et al. Pharmacologic blockade of JAK1/JAK2 reduces GvHD and preserves the graft-versus-leukemia effect. *PLoS ONE*. (2014) 9:e109799. doi: 10.1371/journal.pone.0109799
- Zeiser R, von Bubnoff N, Butler J, Mohty M, Niederwieser D, Or R, et al. Ruxolitinib for glucocorticoid-refractory acute graft-versus-host disease. N Engl J Med. (2020) 382:1800–10. doi: 10.1056/NEJMoa19 17635

- Choi J, Cooper ML, Staser K, Ashami K, Vij KR, Wang B, et al. Baricitinibinduced blockade of interferon gamma receptor and interleukin-6 receptor for the prevention and treatment of graft-versus-host disease. *Leukemia*. (2018) 32:2483–94. doi: 10.1038/s41375-018-0123-z
- Morozova EV, Moiseev IS, Barabanshikova MV, Darskaya EI, Bondarenko SN, Zubarovskaya LS, et al. Graft-versus-host disease prophylaxis with posttransplantation cyclophosphamide and ruxolitinib in patients with myelofibrosis. *Blood*. (2017) 130:4492. doi: 10.1182/blood.V130.Suppl 1.4492.4492
- Zhao Y, Shi J, Luo Y, Gao F, Tan Y, Lai X, et al. Calcineurin inhibitors replacement by ruxolitinib as graft-versus-host disease prophylaxis for patients after allogeneic stem cell transplantation. *Biol Blood Marrow Transplant*. (2020) 26:e128–33. doi: 10.1016/j.bbmt.2020.01.012
- Groth C, Groningen LFJ van, Matos TR, Bremmers ME, Preijers FWMB, Dolstra H, et al. Phase I/II trial of a combination of anti-CD3/CD7 immunotoxins for steroid-refractory acute graft-versus-host disease. *Biol Blood Marrow Transplant*. (2019) 25:712–9. doi: 10.1016/j.bbmt.2018.10.020
- 97. Lee JW, Joachim Deeg H. Prevention of chronic GVHD. Best Pract Res Clin Haematol. (2008) 21:259–70. doi: 10.1016/j.beha.2008.02.010
- Cutler C, Kim HT, Bindra B, Sarantopoulos S, Ho VT, Chen Y-B, et al. Rituximab prophylaxis prevents corticosteroid-requiring chronic GVHD after allogeneic peripheral blood stem cell transplantation: results of a phase 2 trial. *Blood*. (2013) 122:1510–7. doi: 10.1182/blood-2013-04-495895
- Koreth J, Matsuoka K, Kim HT, McDonough SM, Bindra B, Alyea EP, et al. Interleukin-2 and regulatory T cells in graft-versus-host disease. N Engl J Med. (2011) 365:2055–66. doi: 10.1056/NEJMoa1108188
- 100. Ciurea SO, Zhang M-J, Bacigalupo AA, Bashey A, Appelbaum FR, Aljitawi OS, et al. Haploidentical transplant with posttransplant cyclophosphamide vs. matched unrelated donor transplant for acute myeloid leukemia. *Blood.* (2015) 126:1033–40. doi: 10.1182/blood-2015-04-639831

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2021 Gooptu and Antin. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.





Revisit of Optimal Donor Number Estimation in the Hong Kong Bone Marrow Donor Registry

Jenny Chung Yee Ho¹, Stephen Kwok Fan Cheung¹, Zhongyi Lui², Ivan Wing Hong Tang¹, Wanling Yang², Patrick Ip², Cheuk Kwong Lee³, Derek Middleton⁴ and Janette Siu Yin Kwok^{1*}

¹ Division of Transplantation and Immunogenetics, Queen Mary Hospital, Hong Kong, Hong Kong, ² Department of Paediatrics and Adolescent Medicine, The University of Hong Kong, Hong Kong, Hong Kong, ³ Hong Kong Red Cross Blood Transfusion Services, Hong Kong, Hong Kong, ⁴ Transplant Immunology, Royal Liverpool Hospital, Liverpool, United Kingdom

OPEN ACCESS

Edited by:

Nicolaus Martin Kröger, Universität Hamburg, Germany

Reviewed by:

Jason Dehn, Central Bone Marrow Donor Register Germany, Germany Ying-Jun Chang, Peking University People's Hospital, China

*Correspondence:

Janette Siu Yin Kwok kwoksy@ha.org.hk

Specialty section:

This article was submitted to Alloimmunity and Transplantation, a section of the journal Frontiers in Immunology

Received: 05 December 2020 Accepted: 11 March 2021 Published: 16 April 2021

Citation:

Ho JCY, Cheung SKF, Lui Z, Tang IWH, Yang W, Ip P, Lee CK, Middleton D and Kwok JSY (2021) Revisit of Optimal Donor Number Estimation in the Hong Kong Bone Marrow Donor Registry. Front. Immunol. 12:638253. doi: 10.3389/fimmu.2021.638253 High resolution typing of the HLA-DPB1 locus for patient who requested for hematopoietic stem cell transplantation (HSCT) workup has recently become mandatory by the National Marrow Donor Program (NMDP) in order to facilitate matching between donors and recipients for better outcomes. The likelihood of identifying HLA matched donors in Hong Kong, on top of the existing HLA-A, -B, -C, and -DRB1 loci, is revisited in this study. HLA-A, -B, -C, -DRB1 and -DPB1 genotypes of 5,266 volunteer unrelated Chinese donors from the Hong Kong Bone Marrow Donor Registry (HKBMDR), were included in this study. Matching models were employed to determine the matching probabilities for 10/10(DPB1) and 9/10(DPB1) HLA match. The matching probabilities are 20% at 10/10(DPB1) HLA match and 55% at 9/10(DPB1) match, based on the existing 130,000 donors in the HKBMDR. The likelihoods of match become 27% and 65% respectively, by increasing the registry to 250,000. However, if DPB T-cell-epitope (TCE) model is considered in the matching, the probability will increase to 46% at 10/10 DPB1 permissive mismatching. Our findings provide vital information about the future planning on the targeted recruitment size, HLA typing and search strategies of the donor registry and arose the transplant physicians' acceptability to 9/ 10(DBP1) or 10/10(DBP1) HLA match. Nevertheless, the marrow donor registry has planned for increasing the registry size and bringing down the age of recruited donors which will ultimately enhance patient outcome.

Keywords: alleles, frequency haplotypes, HLA antigens, Chinese, matching probability

INTRODUCTION

The detrimental graft-versus-host disease (GVHD) remains a major challenge after curative hematopoietic stem cell transplantation (HSCT). Systemic outcome analysis has shown that HLA-DPB1 mismatch had resulted in increased risk of acute GVHD. Transplantation with non-permissive DPB1 mismatch was shown to be associated with higher transplant-related mortality (1). Starting from 27 February 2021, HLA typing of DBP1 loci has become mandatory for patients

requesting for HSCT workup from the National Marrow Donor Program (NMDP). In light of better outcome for HSCT, optimal matching between donors and recipients are recommended at high resolution in the HLA-A, -B, -C, -DRB1 and -DPB1 loci. Due to the population-specific allelic variation and the extremely high level of HLA gene polymorphism, the availability of optimal HLA-matched unrelated donors and cord-blood units has always been a concern (2, 3). As a result, donors with mismatched HLA antigens may also be considered in many situations. However, these HLA mismatches may lead to an 8% reduction per loci in the 5-year overall survival rate after HSCT (4). The additional information on DPB1 loci may help clinician on final donor selection by reviewing the matching at DPB1 to enhance the patient outcome when more than one potential donors are available for HSCT.

Volunteer unrelated donor database has been managed by the Hong Kong Bone Marrow Donor Registry (HKBMDR). At present, there are close to 130,000 stem cell donors in HKBMDR and 38 million donors in the Bone Marrow Donors Worldwide (BMDW) (5). Continual growth on the number of donors has been achieved globally. However, it accompanied with significant resource implication in donor recruitment and HLA typing. Therefore, strategic donor recruitment becomes very important account of the donor registry planning. Many crucial factors, including recruitment on more young male donors (6) or focus on the recruitment of donors with rare human leukocyte antigen (HLA) phenotypes (7), donors from ethnic minority (8–11), and recruitment activities based on HLA frequency differences at regional priority setting (12–15).

Estimation of matching probability, including mixed patient population, provides vital information for donor recruitment strategy planning and framework for international stem cell donor exchange (16). We have used the calculations based on HLA-A, -B, -C and -DRB1 loci high-resolution haplotype frequencies (HF) of our own population to estimate the donor pool size earlier (17).

The linkage disequilibrium between HLA-DPB1 and other loci are weak due to a hot-spot of recombination between HLA-DPB1 and HLA-DQB1 loci (18). A big proportion of unrelated donor HSCTs were performed across HLA-DPB1 mismatches (19, 20). HLA-DPB1 alloantigens are target of graft-versusleukemia (GVL) or graft-versus-host (GVH) disease mediated by alloreactive T cells (21-24). However, only 3-57% of HLA-DPB1 were typed in the HLA DNA typed unrelated donors from varies registries (25). Since it was well known that racial and ethnic background play a profound role in adult-donor availability and match probabilities (26), the same phenomenon was proven in our previous study (17). We estimated the donor pool and matching probability on HLA 10/10(DBP1) matching with reference to our recent publication on the gene and HF of the Hong Kong population (27). To our knowledge, this is the first study to revisit the calculation of matching probabilities of our population and the estimation of donor size based on the additional DPB1 requirement.

MATERIALS AND METHODS

Sample Collection and Genotyping

The gene frequency and HF as reported previously were used in the analysis (27). In brief, Next generation sequencing supplemented with sequence-specific primer was used to define allele combinations and some specific alleles with 5,266 donors. HF was calculated from these results using Markov Chain Monte Carlo (MCMC) algorithm PHASE (28). Matching model was then utilized by using the calculated HF and effective adult-donor registry size for each group, with the assumption of genotypes in Hardy-Weinberg equilibrium (HWE) (29, 30).

HLA-DPB1 typing was assigned based on T-cell Epitope algorithms version 2.0 assignment (https://raw.githubusercontent.com/ANHIG/IMGTHLA/Latest/tce/dpb_tce.csv) and also the online tool at https://www.ebi.ac.uk/ipd/imgt/hla/dpb_v2.html (31). The TCE group assignment was reported for all HLA-DPB1 alleles according to the Release 3.38.0 of the IPD-IMGT/HLA Database, released 2019-10. The predicted immunogenicity of the HLA-DPB1 matching will be presented as Permissive, Non-Permissive GvH or Non-Permissive host-versus-graft (HvG).

Statistics Analysis

The frequencies of HLA-A, -B, -C, -DRB1 and -DPB1 alleles were calculated from the number of observed genotypes. MCMC simulation from Guo and Thompson was utilized to assess the Hardy-Weinberg equilibrium for each loci *via* PHASE (32), and the deviance of genotype frequency within each loci was detected by PHASE invoking Arlequin (33). *P* value of <0.01 was considered to be statistically significant.

Formulae described by Schmidt et al. has been utilized in this study with modification (16). In brief, the probability p(n) for any patient from their own population to identify at least one matched donor in a registry including n individuals of a donor population is given $p(n) = \sum_i f_i [1-(1-f_i)^n]$ with p(n) being the matching probability in "n" sample size, f_i being the frequencies of the i-th genotype and i-th is any genotype from the rank of genotype frequencies in the order from the highest to the lowest in a donor population. The estimated HF was used to derive the genotype frequencies under the assumption of HWE.

RESULTS AND DISCUSSION

Data from the recently published HLA genotype and haplotype frequencies of the HKBMDR (27) was applied in this study. Characteristics of these HLA haplotypes in Hong Kong were summarized in **Table 1**.

In concordance with our previous study (17), it was found that the number of haplotypes was significantly increased with number of donor samples. This increase is exclusive for our local population, as a plateau of number of haplotypes with increase in sample size was not observed in other ethnic groups, e.g. Caucasians and European populations (34). Mori et al.

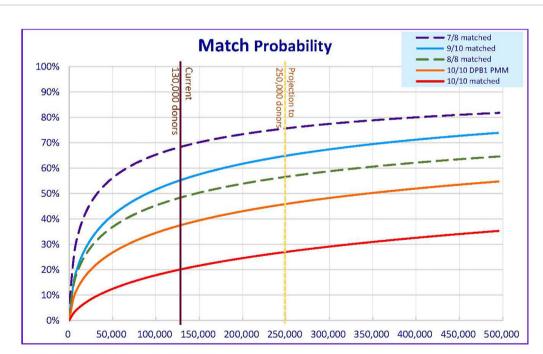
TABLE 1 | Characteristics of the haplotypes of Hong Kong

HLA loci available		A-C-B-DRB1-DPB1
Sample size (N)		5,266
Sample size (2N)		10,532
Number of haplotypes (>0.006%)		3,326
Sum (%) of haplotype frequencies within the top	10	10.7
	25	16.0
	50	21.1
	100	28.1
	250	40.2
	500	51.7
	1,000	65.6
Number of haplotypes with frequency	≥0.01	3
	≥0.005	9
	≥0.001	126
	≥0.0005	323
	≥0.0001	1,918

reported that a significant higher level of the occurrence of common haplotypes (0.01%) was observed in Asian Americans than in Caucasian Americans in the NMDP database. This suggested that the Caucasian Americans had a smaller degree of genetic diversity than Asian Americans (35). Similar findings from a large sample database that the occurrence of common haplotypes in Asian or Pacific Islanders (API) was also higher than Caucasians (34). However, whether the same phenomenon will be observed when HLA-DPB1 is considered requires further elucidation.

A similar methodology was applied in calculating the likelihood of finding a "matched" donor in US (26), likelihood of finding an 8/8 HLA match or ≥ 7/8 HLA match by different donor registry size in Hong Kong was reported in previous study for matching A, B, C, DRB1 loci only (17). With the increase in the number of donors in the HKBMDR to 130,000 as of December 2020, the likelihood of finding an available 8/8 HLA matched donor is 49% and 69% for finding 7/8 HLA matched donor (Figure 1). The results were comparable to those figures found among Asians, Pacific Islanders, and Native Americans (26). However, when taking into account of matching for HLA-DPB1 loci, the likelihood of finding an available 10/10(DBP1) HLA matched donor is 20% while 55% for finding 9/10(DBP1) HLA matched donor. Similar finding was observed in a Finnish retrospective study in which only 32.6% of local donors or 19.3% of both local and foreign donors were HLA-DPB1 matched with HSCT patients (36). In our data, the matching probability increases to 38% when taking into account of the DPB1 T-cellepitope (TCE) permissive mismatching model.

TCE Groups has been utilized in classifying HLA-DPB1 mismatches that might be tolerated (permissive) or would increase risks (non-permissive) after unrelated HSCT. If HLA-DPB1 matching with TCE Groups is considered, beneficial effect during donor selection has been shown in various studies (37, 38). Donors with a permissive HLA-DPB1 group are preferred over those showing a non-permissive HLA-DPB1 group, among those 9/10(DPB1) and 10/10(DBP1) potential donors. "DPB1 TCE3 grading" has been implanted in OptiMatch with the



The brown line is current registry size

FIGURE 1 | Matching Probability (MP) of varies level of HLA match against different donor registry size in HKBMDR. DPB1 PMM - with HLA-DPB1 permissive mismatch.

evident published by Zino et al. (39, 40). The new score was assigned based on the 3 TCE Groups algorithm according to the T cell cross-reactivity patterns (31).

IPD-IMGT/HLA website provided the original feature of DPB1 TCE3 algorithm and have been used in the BMDW Search & Match Service. The discrimination of permissive or non-permissive HLA-DPB1 mismatches is determined based on whether the donor and patient alleles belong to the same (permissive) or different (non-permissive) TCE Groups. There are total of 81 combinations of the HLA-DPB1 typing resulting for TCE version 2 assignment (Supplementary Table 1).

Greater heterogeneity in HLA typing of the Hong Kong Chinese was found where compared with other populations (34, 41, 42). Therefore, to enhance the chance of successful donor search, a larger donor pool is warranted. In concordance with the findings by Dehn and Buck, the likelihood of matching in HLA-A, -B, -C, -DRB1 and -DQB1 10 alleles for Asian Americans was also inferior than Caucasian Americans for 7/8 or 9/10(DQB1) matched unrelated donor search was also lower (98% vs 88%) (43–45).

In addition to the matching issue, attrition of donors due to age and contact unavailability may pose another negative impact on the likelihood of finding a donor. Based on the previous registry size of 100,000, the attrition rate was 2% or 2,000 per year. As shown in the projection (Figure 1), increasing the registry size to 250,000 in five-year time, 26,600 new recruitments per year is required to achieve matching likelihood at 46% for 10/10(DBP1) HLA-DPB1 permissible Match or 65% for ≥ 9/10(DBP1) HLA Match. An annual recruitment of 26,600 is a big rise compared to the current of 8,000 per year. Extra resources should be sought to cover the cost in donor recruitment and HLA typing. A survey was conducted to identify the crucial factors that affect the motivation of stem cell donation in Hong Kong (46). To enhance the recruitment ratio of the younger age group, recruitment program targeting a specific age group, especially for student at higher education may facilitate better recruitment rate and longer maintenance for donation to maximize the cost-effectiveness. Targeted educational activities such as Stem Cell Donation campaign, including educational talks to students and parents, promotion video on social media and social networking platforms and roadshows may help to enhance the recruitment of youngsters.

Racial and ethnic background in a donor registry has been reported to affect the adult-donor availability (26). The current analysis has not taken into the account of adult-donor availability which may have substantially lower match likelihoods. In addition, donors from the patient's own racial and ethnic group has shown to have the highest matching probability (47), this probability may also be enhanced if donors from other racial and ethnic groups could be available. Registry with donors that have a relatively low occurrence of inter-racial or inter-ethnic marriage might have less chance to have donors identified from other groups. The overall donor available rate is less than 30% (27) and it will expect to be lowered when additional loci is considered.

In the above estimation, the matching probability from around 3 million Chinese donors registered in China and

Taiwan registries has not taken into account, which may provide extra donor matching. Furthermore, the matching probability of the cord blood units which are readily available and require less stringent HLA matching was not included in this calculation. Cord blood would be used as an alternative when adult donor is not readily available in many transplant centers. The issue of relatively low stem cell dose for adult size recipient has been resolved by the application of double cord blood units, and has been proven success clinically (48, 49). Whether cord blood can eventually substitute the need of a large registry is still debatable.

Although only 5,266 donors HLA haplotype frequencies have been included in the current study, some rare alleles may not be covered in the presence analysis and affect the accuracy of the estimation. Nonetheless, common haplotype for those with frequencies above 0.2% should be covered. The information provided in this study provided an overview of the matching probability for the local population and facilitate the formulation of donor recruitment target and planning for extra resources in order to support the cost in donor recruitment and HLA typing. Establishment of a cost-effective bone marrow donor registry with an expanded donor pool is utmost important to enhance the likelihood of matching, shorten donor search time in the same ethnicity as domestic donors are more likely to donate stem cells (47). Moreover, it circumvents the shipment restriction or border control especially during the COVID-19 pandemic. This will facilitate timely HSCT in order to catch the best timing during patient remission period, and thus enhance the success rate of HSCT and patient outcome. A more comprehensive model of analysis for inclusion of availability of donor, incomplete or discrepant donor typing and loss of contact would be desired. With the continuation of donor HLA typing by the NGS technology, a revisit of the analysis with a larger sample size would be warranted in the future in order to obtain a more accurate estimation to cover the rare HLA alleles.

DATA AVAILABILITY STATEMENT

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

ETHICS STATEMENT

Ethical review and approval was not required for the study on human participants in accordance with the local legislation and institutional requirements. The donors have provided their written informed consent to perform HLA typing for HKBMDR.

AUTHOR CONTRIBUTIONS

The study was designed by WY and JK. Data was collected by JH, SC, IT, CKL and JK. The computation and statistical analyses were performed by IT, ZL, WY and JK. The samples were

provided by CKL and JK. The manuscript was written by JH, SC, IT, PI, CKL, DM and JK. All authors contributed to the article and approved the submitted version.

ACKNOWLEDGMENTS

The authors would like to acknowledge the Hong Kong Bone Marrow Foundation Limited for the equipment donation.

REFERENCES

- Pidala J, Lee SJ, Ahn KW, Spellman S, Wang HL, Aljurf M, et al. Nonpermissive HLA-DPB1 mismatch increases mortality after myeloablative unrelated allogeneic hematopoietic cell transplantation. *Blood* (2014) 124(16):2596–606. doi: 10.1182/blood-2014-05-576041
- Beatty PG, Mori M, Milford E. Impact of racial genetic polymorphism on the probability of finding an HLA-matched donor. *Transplantation* (1995) 60 (8):778–83. doi: 10.1097/00007890-199510270-00003
- Kollman C, Maiers M, Gragert L, Muller C, Setterholm M, Oudshoorn M, et al. Estimation of HLA-A, -B, -DRB1 haplotype frequencies using mixed resolution data from a National Registry with selective retyping of volunteers. Hum Immunol (2007) 68(12):950–8. doi: 10.1016/j.humimm.2007.10.009
- Lee SJ, Klein J, Haagenson M, Baxter-Lowe LA, Confer DL, Eapen M, et al. High-resolution donor-recipient HLA matching contributes to the success of unrelated donor marrow transplantation. *Blood* (2007) 110(13):4576–83. doi: 10.1182/blood-2007-06-097386
- Burlingham WJ, Grailer AP, Heisey DM, Claas FH, Norman D, Mohanakumar T, et al. The effect of tolerance to noninherited maternal HLA antigens on the survival of renal transplants from sibling donors. N Engl J Med (1998) 339(23):1657–64. doi: 10.1056/NEJM199812033392302
- Schmidt AH, Biesinger L, Baier D, Harf P, Rutt C. Aging of registered stem cell donors: implications for donor recruitment. *Bone Marrow Transplant* (2008) 41(7):605–12. doi: 10.1038/sj.bmt.1705950
- Schmidt AH, Stahr A, Baier D, Schumacher S, Ehninger G, Rutt C. Selective recruitment of stem cell donors with rare human leukocyte antigen phenotypes. Bone Marrow Transplant (2007) 40(9):823–30. doi: 10.1038/sj.bmt.1705832
- Johansen KA, Schneider JF, McCaffree MA, Woods GL, Council on S, and Public Health, AMA. Efforts of the United States' National Marrow Donor Program and Registry to improve utilization and representation of minority donors. *Transfus Med* (2008) 18(4):250–9. doi: 10.1111/j.1365-3148.2008.00865.x
- Laver JH, Hulsey TC, Jones JP, Gautreaux M, Barredo JC, Abboud MR. Assessment of barriers to bone marrow donation by unrelated African-American potential donors. *Biol Blood Marrow Transplant* (2001) 7(1):45– 8. doi: 10.1053/bbmt.2001.v7.pm11215698
- Pingel J, Solloch UV, Hofmann JA, Lange V, Ehninger G, Schmidt AH. Highresolution HLA haplotype frequencies of stem cell donors in Germany with foreign parentage: how can they be used to improve unrelated donor searches? Hum Immunol (2013) 74(3):330–40. doi: 10.1016/j.humimm.2012.10.029
- Schmidt AH, Solloch UV, Baier D, Yazici B, Ozcan M, Stahr A, et al. Criteria for initiation and evaluation of minority donor programs and application to the example of donors of Turkish descent in Germany. *Bone Marrow Transplant* (2009) 44(7):405–12. doi: 10.1038/bmt.2009.55
- Buhler S, Nunes JM, Nicoloso G, Tiercy JM, Sanchez-Mazas A. The heterogeneous HLA genetic makeup of the Swiss population. *PloS One* (2012) 7(7):e41400. doi: 10.1371/journal.pone.0041400
- Marroni F, Curcio M, Fornaciari S, Lapi S, Mariotti ML, Scatena F, et al. Microgeographic variation of HLA-A, -B, and -DR haplotype frequencies in Tuscany, Italy: implications for recruitment of bone marrow donors. *Tissue Antigens* (2004) 64(4):478–85. doi: 10.1111/j.1399-0039.2004.00292.x
- Rendine S, Borelli I, Barbanti M, Sacchi N, Roggero S, Curtoni ES. HLA polymorphisms in Italian bone marrow donors: a regional analysis. *Tissue Antigens* (1998) 52(2):135–46. doi: 10.1111/j.1399-0039.1998.tb02277.x
- 15. Schmidt AH, Solloch UV, Baier D, Stahr A, Wassmuth R, Ehninger G, et al. Regional differences in HLA antigen and haplotype frequency distributions in Germany and their relevance to the optimization of hematopoietic stem cell

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fimmu.2021. 638253/full#supplementary-material

Supplementary Table 1 | 81 Combinations of DPB1 TCE v2 assignment and matching outcome.

- donor recruitment. Tissue Antigens (2010) 76(5):362–79. doi: 10.1111/j.1399-0039.2010.01520.x
- Schmidt AH, Sauter J, Pingel J, Ehninger G. Toward an optimal global stem cell donor recruitment strategy. PloS One (2014) 9(1):e86605. doi: 10.1371/journal.pone.0086605
- Kwok J, Guo M, Yang W, Ip P, Chan GCF, Ho J, et al. Estimation of optimal donor number in Bone Marrow Donor Registry: Hong Kong's experience. Hum Immunol (2017) 78(10):610–3. doi: 10.1016/j.humimm.2017.08.007
- Cullen M, Erlich H, Klitz W, Carrington M. Molecular mapping of a recombination hotspot located in the second intron of the human TAP2 locus. Am J Hum Genet (1995) 56(6):1350-8.
- Petersdorf EW, Kollman C, Hurley CK, Dupont B, Nademanee A, Begovich AB, et al. Effect of HLA class II gene disparity on clinical outcome in unrelated donor hematopoietic cell transplantation for chronic myeloid leukemia: the US National Marrow Donor Program Experience. *Blood* (2001) 98(10):2922– 9. doi: 10.1182/blood.V98.10.2922
- Fleischhauer K, Shaw BE, Gooley T, Malkki M, Bardy P, Bignon JD, et al. Effect of T-cell-epitope matching at HLA-DPB1 in recipients of unrelateddonor haemopoietic-cell transplantation: a retrospective study. *Lancet Oncol* (2012) 13(4):366–74. doi: 10.1016/S1470-2045(12)70004-9
- Rutten CE, van Luxemburg-Heijs SA, Griffioen M, Marijt EW, Jedema I, Heemskerk MH, et al. HLA-DP as specific target for cellular immunotherapy in HLA class II-expressing B-cell leukemia. *Leukemia* (2008) 22(7):1387–94. doi: 10.1038/leu.2008.90
- Rutten CE, van Luxemburg-Heijs SA, Halkes CJ, van Bergen CA, Marijt EW, Oudshoorn M, et al. Patient HLA-DP-specific CD4+ T cells from HLA-DPB1mismatched donor lymphocyte infusion can induce graft-versus-leukemia reactivity in the presence or absence of graft-versus-host disease. *Biol Blood Marrow Transplant* (2013) 19(1):40–8. doi: 10.1016/j.bbmt.2012.07.020
- Gaschet J, Lim A, Liem L, Vivien R, Hallet MM, Harousseau JL, et al. Acute graft versus host disease due to T lymphocytes recognizing a single HLA-DPB1*0501 mismatch. J Clin Invest (1996) 98(1):100-7. doi: 10.1172/ JCI118753
- Gaschet J, Gallot G, Ibisch C, Lim A, Even J, Vivien R, et al. Acute graftversus-host disease after bone marrow transplantation with a single HLA-DPB1*1001 mismatch: involvement of different TCRBV subsets. *Bone Marrow Transplant* (1998) 22(4):385–92. doi: 10.1038/sj.bmt.1701336
- WMDA Global Trend Report. (2019) Available at: https://wmda.info/wp-content/uploads/2020/07/20201307-GTR-2019-Summary-slides.pdf.
- Gragert L, Eapen M, Williams E, Freeman J, Spellman S, Baitty R, et al. HLA match likelihoods for hematopoietic stem-cell grafts in the U.S. registry. N Engl J Med (2014) 371(4):339–48. doi: 10.1056/NEJMsa1311707
- Kwok J, Tang WH, Chu WK, Chan YS, Liu Z, Yang W, et al. High resolution allele genotyping and haplotype frequencies for NGS based HLA 11 loci of 5266 Hong Kong Chinese bone marrow donors. *Hum Immunol* (2020). doi: 10.1016/j.humimm.2020.08.005
- Stephens M, Smith NJ, Donnelly P. A new statistical method for haplotype reconstruction from population data. Am J Hum Genet (2001) 68(4):978–89. doi: 10.1086/319501
- Beatty PG, Boucher KM, Mori M, Milford EL. Probability of finding HLAmismatched related or unrelated marrow or cord blood donors. Hum Immunol (2000) 61(8):834–40. doi: 10.1016/S0198-8859(00)00138-5
- Mori M, Graves M, Milford EL, Beatty PG. Computer program to predict likelihood of finding and HLA-matched donor: methodology, validation, and application. *Biol Blood Marrow Transplant* (1996) 2(3):134–44.
- 31. Crivello P, Zito L, Sizzano F, Zino E, Maiers M, Mulder A, et al. The impact of amino acid variability on alloreactivity defines a functional distance predictive

- of permissive HLA-DPB1 mismatches in hematopoietic stem cell transplantation. *Biol Blood Marrow Transplant* (2015) 21(2):233–41. doi: 10.1016/i.bbmt.2014.10.017
- Guo SW, Thompson EA. Performing the exact test of Hardy-Weinberg proportion for multiple alleles. *Biometrics* (1992) 48(2):361–72. doi: 10.2307/2532296
- Excoffier L, Lischer HEL. Arlequin suite ver 3.5: a new series of programs to perform population genetics analyses under Linux and Windows. Mol Ecol Resour (2010) 10(3):564–7. doi: 10.1111/j.1755-0998.2010.02847.x
- Gragert L, Madbouly A, Freeman J, Maiers M. Six-locus high resolution HLA haplotype frequencies derived from mixed-resolution DNA typing for the entire US donor registry. *Hum Immunol* (2013) 74(10):1313–20. doi: 10.1016/j.humimm.2013.06.025
- Mori M, Beatty PG, Graves M, Boucher KM, Milford EL. HLA gene and haplotype frequencies in the North American population: the National Marrow Donor Program Donor Registry. *Transplantation* (1997) 64 (7):1017–27. doi: 10.1097/00007890-199710150-00014
- Linjama T, Rather C, Ritari J, Perasaari J, Eberhard HP, Korhonen M, et al. Extended HLA Haplotypes and Their Impact on DPB1 Matching of Unrelated Hematologic Stem Cell Transplant Donors. *Biol Blood Marrow Transplant* (2019) 25(10):1956–64. doi: 10.1016/j.bbmt.2019.07.008
- Arrieta-Bolanos E, Crivello P, Shaw BE, Ahn KW, Wang HL, Verneris MR, et al. In silico prediction of nonpermissive HLA-DPB1 mismatches in unrelated HCT by functional distance. *Blood Adv* (2018) 2(14):1773–83. doi: 10.1182/bloodadvances.2018019620
- 38. Tram K, Stritesky G, Wadsworth K, Ng J, Anasetti C, Dehn J. Identification of DPB1 Permissive Unrelated Donors Is Highly Likely. *Biol Blood Marrow Transplant* (2017) 23(1):81–6. doi: 10.1016/j.bbmt.2016.10.021
- Zino E, Frumento G, Marktel S, Sormani MP, Ficara F, Di Terlizzi S, et al. A T-cell epitope encoded by a subset of HLA-DPB1 alleles determines nonpermissive mismatches for hematologic stem cell transplantation. *Blood* (2004) 103(4):1417–24. doi: 10.1182/blood-2003-04-1279
- Zino E, Vago L, Di Terlizzi S, Mazzi B, Zito L, Sironi E, et al. Frequency and targeted detection of HLA-DPB1 T cell epitope disparities relevant in unrelated hematopoietic stem cell transplantation. *Biol Blood Marrow Transplant* (2007) 13(9):1031–40. doi: 10.1016/j.bbmt.2007.05.010
- Gourraud PA, Lamiraux P, El-Kadhi N, Raffoux C, Cambon-Thomsen A. Inferred HLA haplotype information for donors from hematopoietic stem cells donor registries. *Hum Immunol* (2005) 66(5):563–70. doi: 10.1016/j.humimm.2005.01.011

- Maiers M, Gragert L, Klitz W. High-resolution HLA alleles and haplotypes in the United States population. *Hum Immunol* (2007) 68(9):779–88. doi: 10.1016/j.humimm.2007.04.005
- Petersdorf EW. Role of major histocompatibility complex variation in graftversus-host disease after hematopoietic cell transplantation. F1000Res (2017) 6:617. doi: 10.12688/f1000research.10990.1
- Buck K, Wadsworth K, Setterholm M, Maiers M, Confer D, Hartzman R, et al. High-Resolution Match Rate of 7/8 and 9/10 or Better for the Be The Match Unrelated Donor Registry. *Biol Blood Marrow Transplant* (2016) 22(4):759–63. doi: 10.1016/j.bbmt.2015.12.012
- Dehn J, Buck K, Maiers M, Confer D, Hartzman R, Kollman C, et al. 8/8 and 10/10 high-resolution match rate for the be the match unrelated donor registry. Biol Blood Marrow Transplant (2015) 21(1):137–41. doi: 10.1016/j.bbmt.2014.10.002
- Kwok J, Leung E, Wong W, Leung K, Lee CK, Lam W, et al. Factors Influencing Hematopoietic Stem Cell Donation Intention in Hong Kong: A Web-Based Survey. Ann Transplant (2015) 20:604–13. doi: 10.12659/AOT.894165
- Linjama T, Eberhard HP, Perasaari J, Muller C, Korhonen M. A European HLA Isolate and Its Implications for Hematopoietic Stem Cell Transplant Donor Procurement. *Biol Blood Marrow Transplant* (2018) 24(3):587–93. doi: 10.1016/i.bbmt.2017.10.010
- Yamamoto H. Single cord blood transplantation in Japan; expanding the possibilities of CBT. Int J Hematol (2019) 110(1):39–49. doi: 10.1007/s12185-019-02672-4
- Izumi K, Kanda J, Nishikori M, Arai Y, Ishikawa T, Yoshioka S, et al. Outcomes of allogeneic stem cell transplantation for DLBCL: a multi-center study from the Kyoto Stem Cell Transplantation Group. *Ann Hematol* (2019) 98(12):2815–23. doi: 10.1007/s00277-019-03835-3

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2021 Ho, Cheung, Lui, Tang, Yang, Ip, Lee, Middleton and Kwok. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.





Chimerism, the Microenvironment and Control of Leukemia

H. Joachim Deeg*

Fred Hutchinson Cancer Research Center and the University of Washington, Seattle, WA, United States

Transplantation of allogeneic hematopoietic cells faces two barriers: failure of engraftment due to a host versus graft reaction, and the attack of donor cells against the patient, the graft versus host (GVH) reaction. This reaction may lead to GVH disease (GVHD), but in patients transplanted due to leukemia or other malignant disorders, this may also convey the benefit of a graft versus leukemia (GVL) effect. The interplay of transplant conditioning with donor and host cells and the environment in the patient is complex. The microbiome, particularly in the intestinal tract, profoundly affects these interactions, directly and via soluble mediators, which also reach other host organs. The microenvironment is further altered by the modifying effect of malignant cells on marrow niches, favoring the propagation of the malignant cells. The development of stable mixed donor/host chimerism has the potential of GVHD prevention without necessarily increasing the risk of relapse. There has been remarkable progress with novel conditioning regimens and selective T-cell manipulation aimed at securing engraftment while preventing GVHD without ablating the GVL effect. Interventions to alter the microenvironment and change the composition of the microbiome and its metabolic products may modify graft/host interactions, thereby further reducing GVHD, while enhancing the GVL effect. The result should be improved transplant outcome.

Keywords: chimerism, microenviroment, microbiome, GVHD prophylaxis regimens, allogeneic transplant, Graft vs. Leukemia Effect

"... he first commanded Bellerophon to kill that savage monster, the Chimaera, who was not a human being, but a goddess, ..." (Homer, *The Iliad*)

jdeeg@fredhutch.org

*Correspondence:

H. Joachim Deea

OPEN ACCESS

Catholic University of the Sacred

Virginia Commonwealth University,

Humanitas Research Hospital, Italy

Edited by: Andrea Bacigalupo,

Heart, Italy

Reviewed by:

United States

Luca Castagna.

Amir Ahmed Toor,

Specialty section:
This article was submitted to
Alloimmunity and Transplantation,
a section of the journal
Frontiers in Immunology

Received: 11 January 2021 Accepted: 17 February 2021 Published: 22 April 2021

Citation

Deeg HJ (2021) Chimerism, the Microenvironment and Control of Leukemia. Front, Immunol, 12:652105.

Front. Immunol. 12:652105. doi: 10.3389/fimmu.2021.652105

INTRODUCTION

In modern medical terminology, particularly in transplantation, the term "chimera" is applied to the result of transplantation, specifically the transplantation of *cells* from one individual into another. This cell transfer will change the recipient composition (1) and may lead to adverse events by inducing a syndrome, termed graft versus host (GVH) disease (GVHD). While GVHD is undesirable, the transferred cells also aid in eliminating the disease for which the patient is being transplanted, via a graft versus leukemia (GVL) effect. In fact, conditioning with cytotoxic therapy alone generally will not eradicate the last malignant cells, as shown in early murine models (2). The donor cell-mediated GVL effect is an essential part of the curative potential of hematopoietic cell transplantation (HCT).

GRAFT VERSUS HOST DISEASE AND GRAFT VERSUS LEUKEMIA EFFECT

A GVL effect was first reported by Barnes and colleagues in murine models in 1956 (3) and 1957 (4), just as Don Thomas et al. reported the successful transfer of normal blood-forming hematopoietic stem cells from healthy donors into human patients with leukemia (5). These reports were followed by publications considering immunotherapeutic approaches to treat leukemia (6, 7). Weiden et al. presented the first comprehensive analysis of clinical transplant results, which showed that patients with acute leukemia who were transplanted with marrow cells from human leukocyte antigen (HLA)-matched sibling donors and who developed GVHD, particularly in its chronic form, had a reduced incidence of relapse and superior survival (8, 9). The GVH reaction is triggered by the encounter of cells from two individuals, the transplant donor and the recipient, with prominent manifestations at the patient's boundaries, in particular the intestinal tract (1, 10). Since a patient's leukemic cells have the same basic genetic makeup as the patient's healthy organs and tissues, this GVL effect may not be surprising. However, the question that arises immediately is whether this effect could be achieved and exploited without the development of GVHD. It has been challenging to separate the GVL effect from GVHD, but animal models indicate that the post-transplant interaction of donor and host cells—conventional and regulatory T cells, donor and host dendritic cells of various lineages, and iNKT cells, along with components of the microenvironment can be shifted such that GVHD is largely prevented while the GVL effect is maintained (11).

The probability of post-HCT relapse depends upon numerous factors, including disease characteristics, treatment received before transplantation, remission status, including measurable residual disease (MRD) at the time of transplantation, the transplant conditioning regimen, the source of donor cells, HLA mismatch between donor and patient, and the development of (chronic) GVHD. MRD, in particular, is currently an area of extensive research. The level of detection of MRD depends upon the methodology used (e.g., deep sequencing for DNA mutations vs. multi-color flow cytometric analysis) (12-14). While flow cytometry identifies immunophenotypic abnormalities that may serve as targets for the GVH reaction and the GVL effect, this is less likely to be the case for most mutations, unless they result in changes in protein expression. A head-to-head comparison of flow and mutation data in regard to their impact on posttransplant relapse is currently not available. Further, there has been a keen interest in the role of DNA polymorphism (and the respective differences between donor and patient) and the occurrence of GVHD and GVL reactivity. While some singlenucleotide polymorphisms associated with a limited number of genes and their possible role for GVHD have been described, no firm conclusions can be drawn (15). Considering an impact of cytogenetic risk and GVHD, we carried out an analysis (Radich and Deeg, unpublished) in patients transplanted for MDS, selecting cohorts, which by conventional criteria could be considered the two extremes for relapse risk: patients who had high risk cytogenetics (16) and did not develop GVHD (acute or chronic) and patients with good risk karyotype who did develop GVHD. Remission status at the time of transplantation, donor selection, conditioning regimen, and GVHD prophylaxis were comparable. Contrary to our hypothesis that there would be a high incidence of relapse in the first cohort and a low incidence in the second, we failed to observe a significant difference. While the analysis may have had limited statistical power, the lack of any difference was striking. Clearly, risk parameters such as DNA mutations (17) (not available for our analysis) and factors that have not been incorporated into currently used risk schemes are relevant for relapse or sustained remission. It is of interest in this context that a recent report suggests a higher incidence of chronic GVHD and possibly a reduction in relapse incidence in patients transplanted from donors with clonal hematopoiesis (18).

T-CELL DEPLETION

Early data on global T-cell depletion of the donor cell inoculum before infusion into the patient showed substantial reduction of the incidence of GVHD but also resulted in a high rate of graft failure and disease relapse (19). More recent data using selective T-cell depletion appear to be more promising.

One strategy is the administration of post-transplant cyclophosphamide (CY), originally for HLA haplo-identical transplants but then extended to other donor/host combinations (20). The reduction of the incidence of GVHD, especially chronic GVHD, with this approach was interpreted as a result of the elimination of host-alloreactive donor T cells. However, more recent data from murine models show that treatment with CY favors the development of CD4+CD25+Foxp3+ regulatory T cells. In addition, some conventional alloreactive T cells persist, albeit with impaired function (21). It is this conjunction of an expansion of regulatory T cells, including those with alloantigen specificity, and altered immuno-competence of conventional T cells that is responsible for the observed prevention of GVHD (21). This mechanism was also functional in thymectomized mice, indicating that it does not require the generation or central selection of T cells. Whether the use of post-transplant CY is associated with an increase in relapse, particularly of myeloid malignancies, remains a matter of debate. Apparently, the modified donor-derived alloreactive T cells maintain GVL activity.

Another concept with similar aims, the prevention of GVHD without increasing the risk of relapse, is the depletion of CD45A+CD62L+ naïve T cells (22, 23). In murine models, the infusion of naïve T cells induced severe GVHD, while central memory T cells resulted in milder GVHD, and effector-memory T cells did not cause significant GVHD (24, 25). Memory T cells, however, conveyed anti-pathogen immunity and GVL reactivity (26). Naïve CD45+CD62L+ T cells appear to be "uncommitted" and, thus, are able to get activated by new (patient) antigens that they encounter, thereby triggering a cascade of signals that initiate GVHD. In the clinic, patients with myeloid or lymphoid malignancies conditioned with regimens

of various intensities and infused with hematopoietic cells from HLA-identical sibling donors that were in vitro depleted of CD45RA+ T cells achieved sustained engraftment, had a very low incidence of severe acute and chronic GVHD, and were not at a higher risk of relapse than patients transplanted with T cell-replete grafts (23). Further, in patients who did develop acute GVHD, generally grade II, corticosteroid treatment could be discontinued much earlier, at a median of 85 days, compared with 853 days in patients given T cell-replete grafts. No case of steroid-refractory GVHD has been observed so far after naïve T-cell depletion. This pattern of rapid response of acute GVHD to steroid therapy and the rare occurrence of chronic GVHD suggests a modified immune environment and a different biology of acute GVHD related to the removal of non-committed naïve T cells. The fact that regulatory T cells that express CD45RA are also eliminated suggests that those cells are not required for the establishment or maintenance of tolerance in this clinical model. In fact, one can speculate that elimination of those regulatory T cells might lead to a more potent GVL effect.

MIXED DONOR/RECIPIENT CHIMERISM

What is the impact of incomplete donor cell engraftment? Available data indicate that the development of *mixed* chimerism, the concurrent presence of recipient and donor lymphohematopoietic cells in the patient after transplantation, may attenuate or prevent the development of GVHD. Mixed chimerism was originally described in patients with nonmalignant disorders, in particular immune deficiencies (27) but also in aplastic anemia (28). This mixed chimerism can persist for years. Studies in a canine model indicated that administration of sublethal doses of total body irradiation before and pharmacological immunosuppression after donor cell infusion resulted in stable mixed hematopoietic donor/recipient chimerism (29). These data underscore the importance of the intensity of the transplant conditioning regimens, which for nonmalignant disorders tend to be less intensive, for the development of mixed chimerism.

Would mixed chimerism also be possible and consistent with transplant success in patients with malignant disorders? Stated differently, would the establishment of "tolerance" between patient and recipient cells include tolerance to the malignant cells and, thereby, eliminated the GVL effect? In fact, several reports have shown persistent antitumor responses even after a loss of donor cell chimerism (30, 31). What is the mechanism? The answer will at least in part depend upon which donor and patient cell sub-populations in the patient's marrow and immune system account for the mix and how the mix alters cell functions. We showed recently that in patients transplanted for myeloproliferative disorders, mixed CD33+ chimerism was associated with subsequent relapse, whereas mixed CD3+ chimerism was not and, in fact, did result in less GVHD without an increased incidence of relapse (32). We observed similar outcomes in two trials enrolling patients with acute myeloid leukemia (AML) or MDS who had been conditioned with busulfan/fludarabine and thymoglobulin (Yeh et al., unpublished observations, February 2021). The factors controlling this balance between patient and donor cells without leading to disease recurrence remain to be determined.

GRAFT VERSUS LEUKEMIA EFFECTS WITHOUT CLASSICAL HEMATOPOIETIC CELL TRANSPLANTATION

If cells from healthy donors are able to induce a GVL effect after transplantation, can such an effect be achieved with the infusion of donor cells (DLI), without carrying out an actual transplant, as has been shown for patients who relapsed after transplantation (60)? Several investigators used leukocyte infusions from HLAmismatched donors in an attempt to provide a direct GVL effect (33-35). In one study, DLI was given to patients with various malignancies to induce a GVL or GV tumor effect (33). These patients were pre-treated with interferon 2ß and given DLI, and 4 weeks later, donor chimerism (determined by PCR for marker analysis) was detected in four of 11 evaluable patients. Of note, four patients who had previously received an autologous transplant developed acute GVHD, and the three patients who could be assessed did show anti-tumor responses. GVHD is a risk associated with DLI. However, the occurrence of GVHD in patients who had previously undergone a transplant is consistent with a modified microenvironment and a role of host cells in the GVHD pathophysiology (36). However, many patients given DLI for relapse after transplantation do experience tumor responses without developing GVHD, illustrating that clinical GVHD is not required for a GVL effect to occur. The GVL effect may be mediated by a subclinical reaction or, alternatively, might involve activity against antigens with limited expression, restricted to the tumor (37). Ongoing research is exploiting this possibility, for example, by generating effector cells against minor histocompatibility antigens (HA-1) primarily expressed on lympho-hematopoietic cells and for which patient and donor differ (61).

Guo et al. reported results with the infusion of granulocyte colony-stimulating factor (G-CSF)-mobilized peripheral blood progenitor cells (including CD34+, CD3+, and NK cells) at various doses, from HLA-mismatched donors in patients with AML in first remission who were not given any GVHD prophylaxis (38). Donor chimerism as determined by the identification of cells containing the Y chromosome (from the male donor) was present in 20 of 23 female recipients as late as 1,000 days after infusion. Leukemia-free survival was significantly prolonged in patients who received higher doses of CD3+ donor cells, while no GVHD was observed. The investigators also showed, further, that in addition to a GVL effect, these patients also experienced a recipient vs. leukemia effect, suggesting activation of the patient's immune system by the infused HLA non-identical donor cells (38). The same authors subsequently reported similar results in another 185 patients with de novo AML (35), but confirmation from other centers is currently not available.

THE MICROENVIRONMENT

The marrow microenvironment is essential for the support of normal and malignant hematopoiesis. We have presented *in vitro* data from patients with MDS, which show a two-way signaling path between the clonal disease cells and non-clonal mesenchymal/stroma cells (39). Stroma cells exhibited altered gene expression and favored the survival of clonal MDS cells rather than healthy hematopoietic precursors. Exposure to the hypomethylating agent, 5-aza-citidine, normalized gene expression in stroma cells and restored their functional competence in support of normal hematopoiesis (39). It is intriguing to speculate that altered gene expression in the marrow microenvironment is a contributor to the frequently observed myelosuppression following DLI.

Data on the role of stroma in disease persistence or recurrence have also been presented for patients with AML (40, 41). Those studies show that malignant (clonal) myeloid cells trigger remodeling events in bone marrow niches, and this remodeled environment then favors the expansion of the malignant clone (39, 42). Other broad-acting contributors to the altered post-transplant milieu in the patient are the effects of endothelial cell activation (43).

Further, solid cancer models show that propagation of clonal tumor cells in the form of metastases was dependent upon the co-migration of stromal cells with those tumor cells (44). Consistent with that observation, we were not able to establish sustained engraftment of clonal MDS cells in a xenotransplant model of human MDS cells in immunodeficient mice, if MDS cells were injected by themselves. However, we did achieve long-term engraftment and expansion when MDS cells were injected along with the (transformed) human stroma cell line HS27a (45). The role of the microenvironment for effective hematopoiesis is undisputed, but what is of note in these models is the support of the *clonal disease* that is mediated at least in part by a quasi auto-feedback loop that leads to "preferential treatment" of the clone.

Possibly related to these data are observations on the development of donor-derived leukemia, i.e., the transformation of polyclonal, healthy donor cells into originator cells of a clonal myeloid disorder (assuming the absence of preexisting clonal abnormalities in donor cells). Several reports have postulated a "leukemogenic effect" of the marrow microenvironment (46, 47). Is the underlying mechanism related to signals provided by donor cells, viz., the chimeric status associated with a successful transplant?

THE MICROBIOME

Exciting research has established that the microbiome plays a central role in the development of GVHD (48–50). We recently summarized data from several laboratories on the profound effects of donor/host interactions at the boundaries of the transplant recipient and the role of the patient's microbiome, particularly in the intestinal tract, in modifying those interactions (1). Shifts in the composition of the intestinal microbiome are associated with GVHD. While some bacteria, such as *Blautia*, appear to have a beneficial effect, others, for example,

Veillonella or enterococcal species such as Enterococcus faecium or Enterococcus faecalis, favor the development or propagation of GVHD, leading to inferior transplant survival (51). These intestinal bacteria interact directly with patient cells, including GALT, L cells, and dendritic cells and thereby modify either tolerogenic or allo-reactive signals (52, 53). Various species, such as E. faecalis, can cross the intestinal barrier and migrate to intestinal lymph nodes, priming resident T and B lymphocytes. Bacterial metabolites, specifically the short-chain fatty acids butyrate or propionate, released into the bloodstream, have a protective effect against chronic GVHD (54, 55). One mechanism involves enhanced development of regulatory T cells. Conversely, a loss of species that produce high levels of butyrate would be associated with a higher incidence of GVHD. So far, there is no evidence that a shift in the composition of the intestinal microbiome impacted progression of the malignancy for which the patient underwent transplantation (51), although there is a profound impact of the mix of the gut microbiome on the response to immunotherapy in other models (56). Intriguing are some very recent observations (Chris Johnston PhD, personal communication, November 2020) indicating that bacteria can alter the methylation pattern of human DNA, thereby modifying gene expression. Conceivably, this may lead to alterations of potential targets for a GVL effect by donor cells.

Viral organisms such as picobirna viruses have also been shown to participate in these donor/host interactions (57), and the role of the cytomegalovirus (CMV) in GVHD development has been investigated extensively (58). Sellar et al. (59) studied patients with various lymphohematopoietic malignancies who were CMV+ and received transplants from CMV negative donors. The conditioning regimens were of reduced intensity and included in vivo T-cell depletion with alemtuzumab. The investigators showed that CMV-specific T cells were exclusively of host origin and protected the patients against recurrent CMV infections, indicating that the status of mixed donor/host chimerism in these patients was associated with increased immune protection. DLI to induced full donor chimerism did not trigger the development of symptomatic CMV infection, and in some patients, donor-derived CMVspecific CD8+ T lymphocytes further expanded. This conversion (from host to donor) occurred without clinical evidence of GVHD, suggesting the possibility that the presence of mixed chimerism, albeit temporary, facilitated the establishment of tolerance.

SUMMARY AND CONCLUSIONS

The interactions between donor and recipient cells following allogeneic HCT are complex, and the cast of characters of this drama is not limited to donor and recipient immune cells. Additional actors include cellular and non-cellular components of the microenvironment and, importantly, the microbiome. Nature had not envisioned *Homo sapiens* trying to break down barriers that have evolved over millions of years. Doing so upsets the balance that we observe in healthy individuals. Of course, these therapeutic interventions are directed at the

eradication of a malignant disease, which has already changed the internal milieu. A better understanding of signals that trigger the development of malignant disorders such as leukemia would allow for earlier interventions and might permit their exploitation to restrict the reactions of donor cells to the GVL effect, while preventing GVHD. Can we direct the divine ability of the chimera against the malignancy and sever the ugly head of GVHD? Current research using state of the art tools, including systems biology and machine learning, may be able to pave the way.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/supplementary material, further inquiries can be directed to the corresponding author/s.

REFERENCES

- Deeg HJ. Individuals, boundaries, graft-versus-host disease. Biol Blood Marrow Transplant. (2020) 26:e309–12. doi: 10.1016/j.bbmt.2020.09.001
- Burchenal JH, Oettgen HF, Holmberg EA, Hemphill SC, Reppert JA. Effect of total body irradiation on the transplantability of mouse leukemias. *Cancer Res.* (1960) 20:425
- Barnes DWH, Corp MJ, Loutit JF, Neal FE. Treatment of murine leukaemia with x-rays and homologous bone marrow. preliminary communication. Br Med J. (1956) 2:626–7. doi: 10.1136/bmj.2.4993.626
- Barnes DWH, Loutit JF. Treatment of murine leukaemia with x-rays and homologous bone marrow: II. Br J Haematol. (1957) 3:241–52. doi: 10.1111/j.1365-2141.1957.tb05793.x
- Thomas ED, Lochte HL Jr, Lu WC, Ferrebee JW. Intravenous infusion of bone marrow in patients receiving radiation and chemotherapy. N Engl J Med. (1957) 257:491–6. doi: 10.1056/NEJM195709122571102
- Mathe G, Amiel JL, Schwarzenberg L, Cattan A, Schneider M. Adoptive immunotherapy of acute leukemia: experimental and clinical results. *Cancer Res.* (1965) 25:1525–31.
- 7. Mathe G, Amiel JL, Schwarzenberg L, Cattan A, Schneider M, Devries MJ, et al. Successful allogeneic bone marrow transplantation in man: chimerism, induced specific tolerance and possible anti-leukemia effects. *Blood.* (1965) 25:179–96. doi: 10.1182/blood.V25.2.179.179
- Weiden PL, Flournoy N, Thomas ED, Prentice R, Fefer A, Buckner CD, et al. Antileukemic effect of graft-versus-host disease in human recipients of allogeneic-marrow grafts. N Engl J Med. (1979) 300:1068–73. doi: 10.1056/NEJM197905103001902
- 9. Weiden PL, Sullivan KM, Flournoy N, Storb R, Thomas ED, And the Seattle Marrow Transplant T. Antileukemic effect of chronic graft-versus-host disease. contribution to improved survival after allogeneic marrow transplantation. N Engl J Med. (1981) 304:1529–33. doi: 10.1056/NEJM198106183042507
- Hill GR, Ferrara JL. The primacy of the gastrointestinal tract as a target organ of acute graft-versus-host disease: rationale for the use of cytokine shields in allogeneic bone marrow transplantation. *Blood.* (2000) 95:2754– 9. doi: 10.1182/blood.V95.9.2754.009k25_2754_2759
- Morris ES, Macdonald KP, Hill GR. Stem cell mobilization with G-CSF analogs: a rational approach to separate GVHD and GVL? *Blood.* (2006) 107:3430-5. doi: 10.1182/blood-2005-10-4299
- Schuurhuis GJ, Heuser M, Freeman S, Bene MC, Buccisano F, Cloos J, et al. Minimal/measurable residual disease in AML: a consensus document from the European LeukemiaNet MRD Working Party. *Blood.* (2018) 131:1275– 91. doi: 10.1182/blood-2017-09-801498
- 13. Hourigan CS, Dillon LW, Gui G, Logan BR, Fei M, Ghannam J, et al. Impact of conditioning intensity of allogeneic transplantation for acute myeloid

AUTHOR CONTRIBUTIONS

The author confirms being the sole contributor of this work and has approved it for publication.

FUNDING

This work was supported in part by the Seattle Translational Tumor Research (STTR) program and the Miklos Kohary and Natalia Zimonyi Kohary Endowed Chair.

ACKNOWLEDGMENTS

HD want to thank Helen Crawford and Joan Vermeulen for help with manuscript preparation and for maintaining our electronic literature database.

- leukemia with genomic evidence of residual disease. J Clin Oncol. (2020) 38:1273-83. doi: 10.1200/JCO.19.03011
- Dillon LW, Gui G, Logan BR, Fei M, Ghannam J, Li Y, et al. Impact of conditioning intensity and genomics on relapse after allogeneic transplantation for patients with myelodysplastic syndrome. *JCO Precis Oncol.* (2021) 5:265–74.
- Martin PJ, Fan W, Storer BE, Levine DM, Zhao LP, Warren EH, et al. Replication of associations between genetic polymorphisms and chronic graft-versus-host disease. *Blood.* (2016) 128:2450– 6. doi: 10.1182/blood-2016-07-728063
- Schanz J, Steidl C, Fonatsch C, Pfeilstocker M, Nosslinger T, Tuechler H, et al. Coalesced multicentric analysis of 2,351 patients with myelodysplastic syndromes indicates an underestimation of poor-risk cytogenetics of myelodysplastic syndromes in the international prognostic scoring system. J Clin Oncol. (2011) 29:1963–70. doi: 10.1200/JCO.2010.28.3978
- Bejar R. Implications of molecular genetic diversity in myelodysplastic syndromes. Curr Opin Hematol. (2017) 24:73– 8. doi: 10.1097/MOH.000000000000313
- Frick M, Chan W, Arends CM, Hablesreiter R, Halik A, Heuser M, et al. Role of donor clonal hematopoiesis in allogeneic hematopoietic stem-cell transplantation. *J Clin Oncol.* (2019) 37:375–85. doi: 10.1200/JCO.2018.79.2184
- Kernan NA, Flomenberg N, Dupont B, O'reilly RJ. Graft rejection in recipients of T-cell-depleted HLA-nonidentical marrow transplants for leukemia. identification of host-derived antidonor allocytotoxic T lymphocytes. *Transplantation*. (1987) 43:842–7. doi: 10.1097/00007890-198743060-00014
- Fuchs EJ, Luznik L. HLA-haploidentical hematopoietic cell transplantation. UpToDate [Electronic Resource] (2019). Available online at: https://www.uptodate.com/contents/hla-haploidentical-hematopoietic-cell-transplantation
- Wachsmuth LP, Patterson MT, Eckhaus MA, Venzon DJ, Gress RE, Kanakry CG. Post-transplantation cyclophosphamide prevents graft-versushost disease by inducing alloreactive T cell dysfunction and suppression. J Clin Invest. (2019) 129:2357–73. doi: 10.1172/JCI124218
- Bleakley M, Heimfeld S, Jones LA, Turtle C, Krause D, Riddell SR, et al. Engineering human peripheral blood stem cell grafts that are depleted of naive T cells and retain functional pathogen-specific memory T cells. *Biol Blood Marrow Transp.* (2014) 20:705–16. doi: 10.1016/j.bbmt.2014.01.032
- Bleakley M, Heimfeld S, Loeb KR, Jones LA, Chaney C, Seropian S, et al. Outcomes of acute leukemia patients transplanted with naive T cell-depleted stem cell grafts. J Clin Invest. (2015) 125:2677–89. doi: 10.1172/JCI81229
- 24. Dutt S, Tseng D, Ermann J, George TI, Liu YP, Davis CR, et al. Naive and memory T cells induce different types of graft-versus-host disease. *J Immunol.* (2007) 179:6547–54. doi: 10.4049/jimmunol.179.10.6547

- Zheng H, Matte-Martone C, Jain D, Mcniff J, Shlomchik WD. Central memory CD8+ T cells induce graft-versus-host disease and mediate graft-versusleukemia. J Immunol. (2009) 182:5938–48. doi: 10.4049/jimmunol.0802212
- Zheng H, Matte-Martone C, Li H, Anderson BE, Venketesan S, Sheng TH, et al. Effector memory CD4+ T cells mediate graft-versus-leukemia without inducing graft-versus-host disease. *Blood.* (2008) 111:2476–84. doi: 10.1182/blood-2007-08-109678
- Heimall J, Logan BR, Cowan MJ, Notarangelo LD, Griffith LM, Puck JM, et al. Immune reconstitution and survival of 100 SCID patients post-hematopoietic cell transplant: a PIDTC natural history study. *Blood.* (2017) 130:2718– 27. doi: 10.1182/blood-2017-05-781849
- Spitzer TR, Himoe E, Cottler-Fox M, Cahill R, Deeg HJ. Long-term stable mixed chimaerism following allogeneic marrow transplantation for severe aplastic anaemia. Br J Haematol. (1990) 76:146–54. doi: 10.1111/j.1365-2141.1990.tb07850.x
- Storb R, Yu C, Wagner JL, Deeg HJ, Nash RA, Kiem HP, et al. Stable mixed hematopoietic chimerism in DLA-identical littermate dogs given sublethal total body irradiation before and pharmacological immunosuppression after marrow transplantation. *Blood.* (1997) 89:3048–54. doi: 10.1182/blood.V89.8.3048
- Rubio MT, Kim YM, Sachs T, Mapara M, Zhao G, Sykes M. Antitumor effect of donor marrow graft rejection induced by recipient leukocyte infusions in mixed chimeras prepared with nonmyeloablative conditioning: critical role for recipient-derived IFN-gamma. *Blood.* (2003) 102:2300– 7. doi: 10.1182/blood-2002-12-3949
- Dey BR, Mcafee S, Colby C, Cieply K, Caron M, Saidman S, et al. Anti-tumour response despite loss of donor chimaerism in patients treated with non-myeloablative conditioning and allogeneic stem cell transplantation.
 Br J Haematol. (2005) 128:351–9. doi: 10.1111/j.1365-2141.2004. 05328.x
- 32. Deeg HJ, Salit RB, Monahan T, Schoch G, Mcfarland C, Scott BL, et al. Early mixed lymphoid donor/host chimerism is associated with improved transplant outcome in patients with primary or secondary myelofibrosis. *Biol Blood Marrow Transplant*. (2020) 26:2197–203. doi: 10.1016/j.bbmt.2020.07.013
- Porter DL, Connors JM, Van DV, Duffy KM, Mcgarigle C, Saidman SL, et al. Graft-versus-tumor induction with donor leukocyte infusions as primary therapy for patients with malignancies. *J Clin Oncol.* (1999) 17:1234. doi: 10.1200/JCO.1999.17.4.1234
- Dickinson AM, Norden J, Li S, Hromadnikova I, Schmid C, Schmetzer H, et al. Graft-versus-leukemia effect following hematopoietic stem cell transplantation for leukemia. Front Immunol. (2017) 8:496. doi: 10.3389/fimmu.2017.00496
- Guo M, Chao NJ, Li JY, Rizzieri DA, Sun QY, Mohrbacher A, et al. HLA-mismatched microtransplant in older patients newly diagnosed with acute myeloid leukemia: results from the microtransplantation interest group. *JAMA Oncol.* (2018) 4:54–62. doi: 10.1001/jamaoncol.2017. 2656
- Shlomchik WD. Graft-versus-host disease (Review). Nat Rev Immunol. (2007) 7:340–52. doi: 10.1038/nri2000
- 37. Kolb HJ. Graft-versus-leukemia effects of transplantation and donor lymphocytes (Review). *Blood.* (2008) 112:4371–83 doi: 10.1182/blood-2008-03-077974
- Guo M, Hu KX, Liu GX, Yu CL, Qiao JH, Sun QY, et al. HLAmismatched stem-cell microtransplantation as postremission therapy for acute myeloid leukemia: long-term follow-up. *J Clin Oncol.* (2012) 30:4084– 90. doi: 10.1200/JCO.2012.42.0281
- Bhagat TD, Chen S, Bartenstein M, Barlowe AT, Von Ahrens D, Choudhary GS, et al. Epigenetically aberrant stroma in MDS propagates disease via Wnt/beta-catenin activation. *Cancer Res.* (2017) 77:4846– 57. doi: 10.1158/0008-5472.CAN-17-0282
- Konopleva M, Konoplev S, Hu W, Zaritskey AY, Afanasiev BV, Andreeff M. Stromal cells prevent apoptosis of AML cells by up-regulation of antiapoptotic proteins. *Leukemia*. (2002) 16:1713–24. doi: 10.1038/sj.leu.2402608
- Tabe Y, Konopleva M. Role of microenvironment in resistance to therapy in AML. Curr Hematol Malig Rep. (2015) 10:96– 103. doi: 10.1007/s11899-015-0253-6

- 42. Chen Y, Hoffmeister LM, Zaun Y, Arnold L, Schmid KW, Giebel B, et al. Acute myeloid leukemia-induced remodeling of the human bone marrow niche predicts clinical outcome. *Blood Adv.* (2020) 4:5257–68. doi: 10.1182/bloodadvances.2020001808
- Pagliuca S, Michonneau D, Sicre De Fontbrune F, Sutra Del Galy A, Xhaard A, Robin M, et al. Allogeneic reactivity-mediated endothelial cell complications after HSCT: a plea for consensual definitions. *Blood Adv.* (2019) 3:2424– 35. doi: 10.1182/bloodadvances.2019000143
- Duda DG, Duyverman AM, Kohno M, Snuderl M, Steller EJ, Fukumura D, et al. Malignant cells facilitate lung metastasis by bringing their own soil. *Proc Natl Acad Sci USA*. (2010) 107:21677–82. doi: 10.1073/pnas.1016234107
- Li X, Marcondes AM, Ragoczy T, Telling A, Deeg HJ. Effect of intravenous coadministration of human stroma cell lines on engraftment of long-term repopulating clonal myelodysplastic syndrome cells in immunodeficient mice. *Blood Cancer J.* (2013) 3:e113. doi: 10.1038/bcj.2013.11
- Nafa K, Bessler M, Deeg HJ, Luzzatto L. New somatic mutation in the PIG-A gene emerges at relapse of paroxysmal nocturnal hemoglobinuria. *Blood*. (1998) 92:3422–7. doi: 10.1182/blood.V92.9.3422
- Aldoss I, Song JY, Curtin PT, Forman SJ. Multiple donorderived leukemias in a recipient of allogeneic hematopoietic cell transplantation for myeloid malignancy. *Blood Adv.* (2020) 4:4798–801. doi: 10.1182/bloodadvances.2020002803
- Penack O, Holler E, Van Den Brink MR. Graft-versus-host disease: regulation by microbe-associated molecules and innate immune receptors (Review). Blood. (2010) 115:1865–72. doi: 10.1182/blood-2009-09-242784
- Jenq RR, Taur Y, Devlin SM, Ponce DM, Goldberg JD, Ahr KF, et al. Intestinal blautia is associated with reduced death from graftversus-host disease. *Biol Blood Marrow Transplant*. (2015) 21:1373– 83. doi: 10.1016/j.bbmt.2015.04.016
- Haak BW, Littmann ER, Chaubard JL, Pickard AJ, Fontana E, Adhi F, et al. Impact of gut colonization with butyrate-producing microbiota on respiratory viral infection following allo-HCT. *Blood.* (2018) 131:2978– 86. doi: 10.1182/blood-2018-01-828996
- Stein-Thoeringer CK, Nichols KB, Lazrak A, Docampo MD, Slingerland AE, Slingerland JB, et al. Lactose drives Enterococcus expansion to promote graftversus-host disease. Science. (2019) 366:1143–9. doi: 10.1126/science.aax3760
- Rescigno M, Urbano M, Valzasina B, Francolini M, Rotta G, Bonasio R, et al. Dendritic cells express tight junction proteins and penetrate gut epithelial monolayers to sample bacteria. *Nat Immunol.* (2001) 2:361– 7. doi: 10.1038/86373
- Chistiakov DA, Bobryshev YV, Kozarov E, Sobenin IA, Orekhov AN. Intestinal mucosal tolerance and impact of gut microbiota to mucosal tolerance. Front Microbiol. (2014) 5:781. doi: 10.3389/fmicb.2014. 00781
- 54. Zitvogel L, Kroemer G. Immunostimulatory gut bacteria. *Science*. (2019) 366:1077–1078. doi: 10.1126/science.aaz7595
- Markey KA, Schluter J, Gomes ALC, Littmann ER, Pickard AJ, Taylor BP, et al. The microbe-derived short-chain fatty acids butyrate and propionate are associated with protection from chronic GVHD. *Blood.* (2020) 136:130– 6. doi: 10.1182/blood.2019003369
- Gopalakrishnan V, Helmink BA, Spencer CN, Reuben A, Wargo JA. The influence of the gut microbiome on cancer, immunity, cancer immunotherapy. Cancer Cell. (2018) 33:570– 80. doi: 10.1016/j.ccell.2018.03.015
- Legoff J, Resche-Rigon M, Bouquet J, Robin M, Naccache SN, Mercier-Delarue S, et al. The eukaryotic gut virome in hematopoietic stem cell transplantation: new clues in enteric graft-versus-host disease. *Nat Med.* (2017) 23:1080– 5. doi: 10.1038/nm.4380
- Nichols WG, Corey L, Gooley T, Davis C, Boeckh M. High risk of death due to bacterial and fungal infection among cytomegalovirus (CMV)-seronegative recipients of stem cell transplants from seropositive donors: evidence for indirect effects of primary CMV infection. *J Infect Dis.* (2002) 185:273– 82. doi: 10.1086/338624
- Sellar RS, Vargas FA, Henry JY, Verfuerth S, Charrot S, Beaton B, et al. CMV promotes recipient T-cell immunity following reduced-intensity T-cell-depleted HSCT, significantly modulating chimerism status. *Blood.* (2015) 125:731–9. doi: 10.1182/blood-2014-07-589150

- Kolb HJ, Mittermüller J, Clemm C, Holler G, Ledderose G, Brehm G, et al. Donor leukocyte transfusions for treatment of recurrent chronic myelogenous leukemia in marrow transplant patients. *Blood.* (1990) 76:2462– 2465. doi: 10.1182/blood.V76.12.2462.2462
- 61. Krakow EF, Summers C, Dahlberg A, Bar M, Biernacki MA, Cunningham T, et al. Phase I study of adoptive immunotherapy with HA-1-specific CD8+ and CD4+ memory T cells for children and adults with relapsed acute leukemia after allogeneic Hematopoietic Stem Cell Transplantation (HCT): trial in progress. *Blood.* (2020) 136:45–6. doi: 10.1182/blood-2020-1 37726

Conflict of Interest: The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2021 Deeg. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.





Allogeneic Stem Cell Transplantation for Acute Myeloid Leukemia: Who, When, and How?

Justin Loke 1,2, Richard Buka 1,2 and Charles Craddock 1,2*

¹ Centre for Clinical Haematology, Queen Elizabeth Hospital, Birmingham, United Kingdom, ² CRUK Clinical Trials Unit, University of Birmingham, Birmingham, United Kingdom

Although the majority of patients with acute myeloid leukemia (AML) treated with intensive chemotherapy achieve a complete remission (CR), many are destined to relapse if treated with intensive chemotherapy alone. Allogeneic stem cell transplant (allo-SCT) represents a pivotally important treatment strategy in fit adults with AML because of its augmented antileukemic activity consequent upon dose intensification and the genesis of a potent graftversus-leukemia effect. Increased donor availability coupled with the advent of reduced intensity conditioning (RIC) regimens has dramatically increased transplant access and consequently allo-SCT is now a key component of the treatment algorithm in both patients with AML in first CR (CR1) and advanced disease. Although transplant related mortality has fallen steadily over recent decades there has been no real progress in reducing the risk of disease relapse which remains the major cause of transplant failure and represents a major area of unmet need. A number of therapeutic approaches with the potential to reduce disease relapse, including advances in induction chemotherapy, the development of novel conditioning regimens and the emergence of the concept of post-transplant maintenance, are currently under development. Furthermore, the use of genetics and measurable residual disease technology in disease assessment has improved the identification of patients who are likely to benefit from an allo-SCT which now represents an increasingly personalized therapy. Future progress in optimizing transplant outcome will be dependent on the successful delivery by the international transplant community of randomized prospective clinical trials which permit examination of current and future transplant therapies with the same degree of rigor as is routinely adopted for non-transplant therapies.

OPEN ACCESS

Edited by:

Antoine Toubert, Université Paris Diderot, France

Reviewed by:

Benedetto Bruno,
NYU Langone Health, United States
Luca Castagna,
Humanitas Research Hospital, Italy
Christoph Schmid,
Augsburg University Hospital,
Germany

*Correspondence:

Charles Craddock
Charles.Craddock@uhb.nhs.uk

Specialty section:

This article was submitted to Alloimmunity and Transplantation, a section of the journal Frontiers in Immunology

> Received: 27 January 2021 Accepted: 23 March 2021 Published: 03 May 2021

Citation:

Loke J, Buka R and Craddock C (2021) Allogeneic Stem Cell Transplantation for Acute Myeloid Leukemia: Who, When, and How? Front. Immunol. 12.659595. doi: 10.3389/fimmu.2021.659595 Keywords: acute myeloid leukemia, allogeneic stem cell transplantation, graft-versus-leukemia, chemotherapy, minimal residual disease, measurable residual disease (MRD)

INTRODUCTION

It is more than sixty years since allogeneic stem cell transplantation (allo-SCT) was pioneered as a novel and potentially curative therapeutic modality in patients with chemotherapy-resistant acute myeloid leukemia (AML) (1, 2). Subsequent analyses have confirmed the role of allo-SCT as the optimal treatment strategy in adults with AML in first complete remission (CR1) consequent upon

its ability to reduce the risk of disease relapse by more than 60% compared with intensive chemotherapy alone. Remarkably the magnitude of the augmented anti-leukemic activity of allo-SCT, result from both dose intensification and the genesis of a potent graft-versus-leukemia (GVL) effect, is similar in all biological subtypes of AML (3).

The survival benefit of the augmented anti-leukemic activity of allo-SCT is blunted by its attendant transplant related mortality (TRM). It is therefore essential to a) identify patients whose outcome with intensive chemotherapy (IC) is such that the enhanced anti-leukemic activity of allo-SCT is otiose b) identify patients whose outcome with IC is such that deployment of the enhanced anti-leukemic activity of an allograft should be considered and c) define as precisely as possible the patient population in which allo-SCT can be delivered with an acceptable morbidity and mortality. Thus the identification of patients likely to benefit from allo-SCT requires a dynamic assessment which incorporates both the predicted risk of disease relapse if the patient were to receive IC alone coupled with a prediction of the TRM were the patient to proceed to transplant (4). Accurate prediction of these parameters has been refined by progress in both risk stratification utilizing clinical, cytogenetic and molecular genetic data as well as advances in prediction of the risk of allo-SCT (5-9). Increasingly, randomized controlled trials are informing critical questions concerning relapse risk in patients treated with IC alone (10) and informing the personalization of transplant strategies (11-14). Cooperative transplant trials networks such as the US BMT CTN and the UK transplant cooperative IMPACT will play an increasingly important role in optimizing outcomes after allo-SCT in AML (15).

Who and When Should Patients With AML Be Transplanted?

The focus of therapeutic endeavor in newly diagnosed AML in recent years has primarily been on improving induction chemotherapy (16, 17). However, the increasing availability of allo-SCT coupled with the recognition that a substantial proportion of patients treated with IC alone are destined to relapse has prioritized the development of algorithms designed to identify patients likely to benefit from allo-SCT in CR1. The advent of more accurate risk stratification, utilizing genetic and measurable residual disease (MRD) analysis, coupled with increased sophistication in predicting and reducing TRM has improved decision making concerning the delivery of optimal consolidation therapy in adult AML (18).

The importance of correctly identifying patients in first CR1 who are likely to relapse is predicated by the poor, incomplete rates of remission salvage, such that a significant proportion of patient who relapse do not reach a second CR (CR2) (19). Furthermore, the use of additional intensive chemotherapy and concomitant infections often result in patients with impaired fitness prior to an allo-SCT in CR2. Studies recurrently show that patients with active disease have poorer outcomes as compared to those patients transplanted in CR, thus this should be a critical goal prior to proceeding to transplant (20, 21). Whilst patients transplanted with CR with incomplete count recovery (CRi) have inferior outcomes to patients with AML in CR, this is as a result

of increased non-relapse mortality (NRM) but not necessarily relapse risk (22). Other studies have shown the number of courses of consolidation chemotherapy delivered prior to transplant do not improve patient outcome (23).

Who Should Be Transplanted With Refractory or Relapsed Disease?

The aim of therapy in fit adults with relapsed with AML is to proceed to allo-SCT once a 2nd CR has been achieved (24). This is based on studies demonstrating very poor outcomes in patients who are not allografted in CR2 (19, 25, 26). However, there may be a subset of patients with core-binding factor translocated AML who may achieve long term remission with autologous transplantation, or in a minority, salvage chemotherapy (19, 27). A number of prognostic systems exist for patients with relapsed/refractory AML (28, 29) which may help to identify subgroups of patients with AML who are likely to have long-term survival following an allo-SCT. Important factors identified in these prognostic systems include, duration of CR1, age at relapse and cytogenetic risk at diagnosis.

Retrospective analyses of allo-SCT for AML in CR2 have demonstrated overall survival (OS) of 30-60%, with acceptable rates of TRM despite intensive pre-treatment in this cohort of patients (30–32). Results have also been encouraging in the use of alternative donors in transplantation at CR2 (32). A formal comparison of myeloablative (MAC) versus RIC regimens in this setting is not possible, but registry studies show no significant differences in OS between patients treated with the differing conditioning intensities (32). Despite this, in fit younger patients who might tolerate a MAC regimen, this is probably the preferred treatment strategy to reduce further disease relapse which remains the major risk facing this patient cohort.

A particularly challenging group of patients with AML are those with primary refractory disease, defined as failure to achieve remission following two cycles of induction chemotherapy (33). Numerous studies have shown that patients transplanted with active disease have poorer outcomes than those in remission (20, 31, 34). However, studies have demonstrated approximately 20-30% of patients with primary refractory disease may have long term survival after an allo-SCT (35) and recent work has identified risk factors that may identify patients who are likely to have primary refractory disease at an earlier stage (36). In the evolving landscape of genetic stratification, these scoring systems are likely to be refined, and the long term impact of novel salvage options from targeted therapies remains to be seen (37, 38). One recent study underlined the particularly poor outcome of patients with TP53 mutant AML, when they were transplanted with active disease (39). A challenge in assessments of such genetic risk factors will be the clonal evolution which occurs in patients with AML following treatment (40).

Finally, for patients who relapse following an allo-SCT, the outcome is very poor (41). However, for some patients, especially ones with a durable remission since transplant, and with disease control at the time of second allo-SCT, this procedure may provide an OS at 2 years of 25% (42). In patients who received an

unrelated donor transplant, no advantage for change in donor in this setting could be demonstrated.

Who Should Be Transplanted in First Complete Remission?

Donor versus no donor studies were the first to demonstrate the ability of allo-SCT to increase disease free survival (DFS) and OS in patients transplanted using a myeloablative HLA matched sibling allo-SCT (43). A selection strategy to identify patients who should be transplanted in CR1 was articulated by Cornelissen and colleagues with the European LeukaemiaNet (ELN) AML working party (4) and is based on the competing risks of relapse with chemotherapy alone versus risk of relapse after an allo-SCT and the concomitant TRM (Figure 1). Underpinning this treatment algorithm is the observation that the risk of relapse following allo-SCT is more than halved as compared to that observed in patients treated with IC alone (3)-regardless of cytogenetic risk group. At the same time recent reductions in transplant toxicity permit delivery of an allo-SCT with an NRM of 15% or less in fit adults with a well matched

sibling or volunteer unrelated donor. On this basis the ELN group recommend consideration of allo-SCT in fit adults with AML in CR1 who have a predicted relapse risk of 35-40% and a suitable donor (33). Thus adults with AML in CR1 who fulfill ELN criteria for good risk disease on the basis of cytogenetics or the presence of an NPM1 mutation without FLT3-ITD mutation, and demonstrate a good response to induction chemotherapy by MRD criteria are not routinely deemed eligible for an allo-SCT in CR1. Conversely, all other adults in CR1 in whom the predicted risk of relapse of >40% if they are treated with IC alone should, in principle, be considered transplant candidates providing a suitable stem cell donor is available (44).

Risk stratification in patients with AML in CR1 is based on clinical (5) factors, such as age and gender, as well as cytogenetic risk based on karyotyping results (6) (**Table 1**). This has been refined in recent years by the discovery of further mutations of prognostic significance in genes such as *FLT3* (45), *NPM1* (46), *ASXL1* (47), *RUNX1* (48) and *TP53* (49) as reflected in the 2017 ELN classification (33). Increasingly mutational information is available for patients as a result of next generation sequencing (NGS)

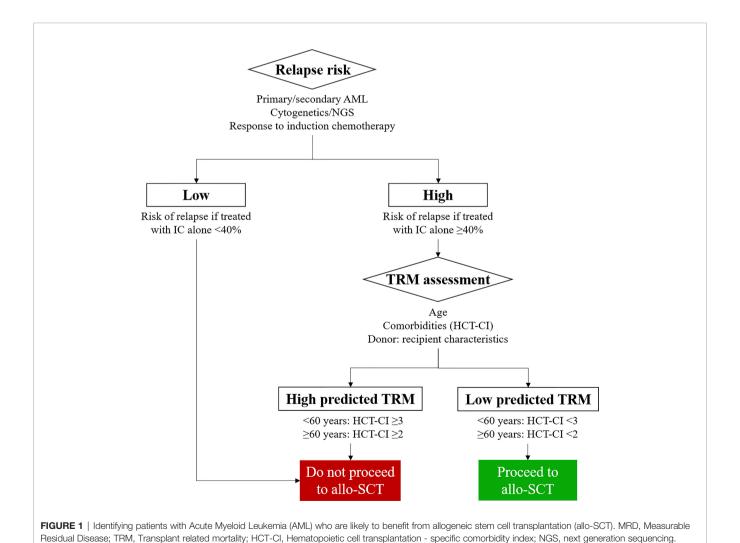


TABLE 1 | Factors determining disease risk in AML.

Clinical Variables	Molecular variables	Dynamic variables
• Age	Cytogenetic	Response to course 1 by morphology
Gender	Next generation sequencing of genes e.g. FLT3, NPM1, RUNX1, ASXL1, TP53	Response to treatment by MRD
 Presenting white cell count 		
Primary versus secondary diseasePerformance status		

technology assaying panels of commonly affected myeloid genes (33). This is of further importance as these genetic markers are now commonly used as both therapeutic targets (50, 51) and as prognostic markers of response to therapies (52). The results of these large scale sequencing efforts of AML samples at diagnosis, in combination with data relating to treatment use and clinical outcome will likely refine these risk categories. This will provide a "personalized" risk score for individuals patients based on a number of these clinical factors and allow for incorporation of combinations of genetic mutations, such as that seen recently in the study of myeloproliferative neoplasms (53, 54). It is increasingly becoming apparent that both clinical and mutational characteristics determine the kinetics of disease relapse. Importantly patients with a *FLT3* mutation are amongst those likely to relapse early in whom the timing of transplant should not be delayed (55).

Incorporation of MRD Risk Stratification

An important development in risk stratification has been the incorporation of MRD monitoring to routinely assess patients' response to chemotherapy (56) (**Table 2** and **Figure 2**). The kinetics and depth of response has been identified as being critical in re-assessing the risk of relapse in patients with otherwise favorable and intermediate risk disease. The impact of MRD monitoring appears to be the most important, independent prognostic factor in many scenarios (57, 58). The selection of the optimal MRD monitoring modality depends on the presence of leukemia specific molecular, cytogenetic or immuno-phenotypic dependent on the AML subtype. Each MRD monitoring technique has its own advantages and

disadvantages, and all require expertise in the delivery of reliable results (Table 2).

Examples of Different Uses of MRD Risk Stratification

Real-time quantitative polymerase chain reaction (RQ-PCR) monitoring of disease specific transcripts provides a sensitive and disease specific assay of MRD for patients with AML expressing a detectable fusion gene transcript (e.g. Corebinding factor (CBF) fusion gene, KMT2A fusion genes, mutant NPM1). In the case of AML with CBF translocation, although age can influence prognosis (59), the depth of response to course 1 and 2 of IC (57) are critical determinants of relapse risk. In cases with residual levels of CBF fusion transcripts at the end of treatment (60), relapse risk depends on level of transcripts, but low levels of CBF fusion gene transcripts may persist after end of treatment without affecting long-term survival. Failure to achieve a 3-log reduction in CBF fusion transcript after two cycles of chemotherapy is associated with an over 50% relapse risk in the monitoring studies of two large cooperative groups, suggesting possible benefit from an allo-SCT in these patients (57, 61).

In younger adults with NPM1 mutant AML, RQ-PCR positivity in the peripheral blood after two cycles of chemotherapy is an important predictor of relapse, identifying a population of patients who should be considered allo-SCT mandatory (58). This is supported by data which points to the beneficial effect of allo-SCT in patients with mutant NPM1 residual disease post induction chemotherapy (62). Recent studies have confirmed that, in younger adults at least, NPM1 is also a predictive biomarker. Patients with NPM1 mutant AML who have a less than 4-log reduction in peripheral blood NPM1 MRD levels demonstrated improved survival after allo-SCT compared with patients who received chemotherapy alone (62). The low relapse risk for patients who are negative for mutant NPM1 transcripts in the peripheral blood after two cycles of intensive chemotherapy outweighs other poor prognostic factors such as concomitant FLT3-ITD mutation or poor risk genotypes (7). The degree to which NPM1 mutations are a prognostic or predictive biomarker in older patients (over the age of 60 years) remains unclear (63). In part this may be due to the increased association of other poor risk cytogenetic features in more elderly patients with NPM1 mutant AML (64). Of note in patients with adverse risk cytogenetics, the presence of NPM1 mutation has no impact on survival outcomes.

TABLE 2 | Relative merits of different MRD monitoring methodologies.

Method	Multi-parameter Flow Cytometry (MFC MRD)	Quantitative PCR (RQ-PCR)	Next generation sequencing (NGS)
Advantages	Rapid results	Sensitive	Applicable to many
	Widely applicable to many patents	Easily compared with sequential results due to	patients
		quantitative range	Error correction increases
		Widely accepted standardisation	sensitivity
Disadvantages	Reliant on expertise of reporting lab	Restricted molecular targets (e.g. Core binding factor	Ongoing development of
	Phenotype of AML cells may change over time	translocations, NPM1c mutant)	technology
			Expense
Examples of	Risk stratification in younger adults, post induction	Risk stratification post chemotherapy to determine	Pre-transplant MRD
use	chemotherapy, with NPM1 negative AML.	relapse risk in NPM1 mutant AML.	monitoring.

MRD timepoint Diagnosis	Pre-transplant	Peri-transplant	Post-transplant
Treatment stage	Induction and consolidation	Conditioning	Maintenance & pre-emptive therapy
Role of MRD monitoring	Relapse risk stratification	Select conditioning intensity and GVHD prophylaxis	Identify need for Maintenance and / or pre-emptive intervention
Novel agents	Midostaurin CPX-351 Venetoclax Gemtuzumab-ozogamicin	Treosulfan	Non-targeted therapy: e.g. Azacitidine, Lenalidomide, Panobinostat, DLI Targeted agents: e.g. targeting: broad spectrum tyrosine kinases, FLT3, BCL-2, IDH-1, IDH-2, Hedgehog

FIGURE 2 | Role of measurable residual disease (MRD) and novel agents at different stages of the treatment pathway in acute myeloid leukemia (AML). GVHD, graft versus host disease; DLI, donor lymphocyte infusion.

A number of large prospective studies have confirmed the prognostic significance of multi-parametric flow cytometry (MFC) determined MRD in adults with newly diagnosed AML treated with IC. In younger adults, MFC MRD+ positive patients with standard risk, NPM1- mutated AML appeared to benefit from allo-SCT in CR1 (10) and data on this group of patients continues to be accrued, including the benefits of intensifying chemotherapy in patients with a suboptimal MRD response after first course of intensive chemotherapy. In older patients, a higher level of MRD after induction treatment is also prognostic of a worse outcome (65). However, in this age group, although MFC MRD negativity, offered improved overall survival, relapse rates remained high.

Early studies suggest a promise for NGS technology for MRD assessment (66), which has the advantage that it may be applicable for many forms of AML. Error correction methodology has become incorporated in this technology to enable higher levels of sensitivity (67), but is currently limited to research settings due to the costs. Furthermore, there has not yet been an upfront comparison of these different MRD technologies independently, or in combination, to compare technical specifications. A recent large study suggested there was an additive prognostic value of NGS MRD over MFC MRD, but interestingly the persistence of age related clonal hematopoiesis after treatment did not result in an increased relapsed rate (66)

Improving Assessment of Transplant Related Mortality in Patients With AML

A critical factor to understand whether a patient with AML is suitable for an allo-SCT is estimation of the TRM associated with

the procedure and whether it is outweighed by the improvement in relapse risk delivered by the transplant process (68-70) (Figure 1). Furthermore, these considerations are central to any discussion with patient and family as to whether the increased risk of an allograft is justifiable. The European Society for Blood and Marrow Transplantation (EBMT) risk score, originally developed in patients allografted for chronic myeloid leukemia (CML) (71), was subsequently shown to be applicable in other disease settings (72), and provided the first attempt to provide a quantifiable estimate of TRM and transplant outcome which could be routinely applied in clinic. However in patients allografted for AML more emphasis is now placed on the Hematopoietic cell transplantation-specific comorbidity index (HCT-CI) score which incorporates a weighted score based on the presence of pre-transplant comorbidities (8). This has been shown to be valid in patients undergoing an allo-SCT for myelodysplastic syndrome (MDS) or AML (9) and more recently combined with age (73), to demonstrate the varying effects of these comorbidities based on a patients' age. Of note, this analysis showed that younger patients with comorbidities were at a significant disadvantage to older fit individuals with no other significant comorbidities.

Unfortunately no scoring system for TRM can include the importance of a clinical assessment of patients based on the "end of bed" assessment and knowledge of how patients have tolerated recent intensive treatment. Thus despite improvements in mathematical modeling techniques to predict treatment related risk on a personalized basis to account for the dynamic interactions between different variables (74, 75), there remains a considerable limitation in the ability of these scoring systems to

predict TRM. Finally, the majority of these scoring systems were developed in the era of sibling or matched unrelated donor transplantation, thereby limiting their use for those with alternative donor sources, which are now of increasing use; such as for recipients of haploidentical donor or umbilical cord stem cells transplants.

What Is the Impact of Patients' Age in Considering Transplant Eligibility?

It is commonly recognized that an important challenge in the management of patients with AML is the increased frequency of this disease with age. Furthermore, the older patient faces a combined challenge of increased frequency of comorbidities and higher risk genetic features (76). Nevertheless patients over the age of 70 years with AML are routinely transplanted with acceptable results (77) but careful assessment of transplant suitability is required. The widely used, updated HCT-CI score allows some adjustments due to age (73), and this analysis showed that younger patients with comorbidities were at a significant disadvantage to older fit individuals with no other significant comorbidities. Nonetheless the HCT-CI score is still of importance in this population, as it has been shown that in patients above 60 years of age a HCT-CI score of 2 or greater results in substantially higher TRM than otherwise expected (78). Future developments to improve assessment of transplant eligibility in this cohort should involve geriatric assessments that encompass an assessment of the functional status of the patient (79).

HOW SHOULD PATIENTS WITH AML IN CR1 BE TRANSPLANTED?

The major causes of treatment failure in adults allografted for AML are transplant toxicity and disease relapse. Whilst significant progress has been made over recent decades in reducing TRM the risk of disease relapse remains stubbornly high. The key considerations in patients with allo-SCT-mandatory AML include identifying which patients should receive RIC as opposed to a MAC allo-SCT and, in patients lacking a well-matched sibling or unrelated donor, what is the preferential alternative donor stem cell source? The development of strategies with the ability to reduce the risk of disease relapse post-transplant also represents a major unmet need.

Strategies to Improve Outcomes Pre-Transplant

The design of novel treatment strategies with the potential to reduce the risk of disease relapse post allo-SCT remains a priority if we are to increase the number of patients with AML who benefit from transplant. A number of questions remain regarding the optimal management of patients' pathway before, during and after an allo-SCT (**Figure 2**). This debate has been reinvigorated in recent years by two key innovations: the widespread use of MRD technologies in patients with AML (80) and the increasing availability of novel pharmacological agents that may be applied

at different treatment stages (81) (**Figure 2**). The adverse impact of pre-transplant MRD on post-transplant outcomes has been increasingly widely recognized (14, 82) and this may inform pre-transplant treatment strategies. Furthermore, emerging data suggest that conditioning intensity and potentially graft-versus-host disease prophylaxis strategies may influence the poor prognostic impact of pre-transplant MRD (83). Finally, post-transplant monitoring of MRD may become important in identifying patients who should receive pre-emptive treatment (84) and is likely to be important in future maintenance strategies in patients post allo-SCT.

How Important Is Pre-Transplant MRD?

A number of retrospective studies have demonstrated the adverse prognostic significance of patients with MFC MRD positivity prior to transplant (82), with some likening the outcomes of these patients post allo-SCT to those with active disease (85). This draws comparison to the outcomes of younger adults with a partial response to the first cycle of induction chemotherapy who have a similar overall outcome as compared to patients who have a CR or CRi but have MFC MRD positivity (10). Two prospective studies have demonstrated the importance of pre-transplant MRD (14, 83) in patients with AML or high risk MDS. The FIGARO study investigated the impact of pretransplant MFC MRD in 244 patients entered into a randomized comparison between FLAMSA-Bu-RIC regimen and a control RIC arm. This identified a poor prognostic impact of a 0.2% threshold of residual disease. However, even in the MRD positive arm, only approximately 50% of patients relapsed: not only suggesting further strategies to identify patients at risk of relapse are required (14), but contrary to previously held opinions, this sizeable proportion of patients with high risk AML may be salvageable with an allo-SCT.

The importance of pre-transplant MRD persists regardless of the technique used to monitor MRD. RT-PCR monitoring of CBF fusion transcripts prior to allo-SCT for patients in CR2, show that those with MRD negativity have a reduced risk of relapse as compared to those with MRD positive disease pre-transplant (86).

Can We Improve Transplant Outcomes in Patients With Evidence of Pre-Transplant MRD?

It remains unknown whether additional courses of chemotherapy or whether further alterations to transplant management in patients with pre-transplant MRD would be of benefit. However, in recent years a number of provocative results have provided impetus to design clinical trials to tackle the poor prognostic impact of pre-transplant MRD.

Pre-Transplant Strategies to Alter Impact of Pre-Transplant MRD?

Studies of novel agents in recent years such as midostaurin and the liposomal cytarabine-daunorubicin preparation CPX-351, suggest that the benefits of these drugs may extend to patients who receive an allo-SCT (16, 50) (**Figure 2**). This provides interesting preliminary data that this may be through improving quality of remissions pre-transplant which may in

future studies be measured as pre-transplant MRD. In the case of the FLT3 inhibitor midostaurin which was added to intensive induction and consolidation, the overall survival benefit of the addition of midostaurin appeared to persist in the majority of patients who were allografted in first remission. Notably midostaurin was not administered as post-transplant maintenance in this study. Likewise, CPX-351 demonstrated improved remission rates and OS in patients receiving this drug over standard remission induction therapy in patients with secondary AML. In patients who subsequently received an allo-SCT, those who had received CPX-351 had improved survival as compared to those in the control arm, but the numbers in the study were small, and a smaller proportion were in a remission at time of transplant in the control arm (16). Definitive studies including the incorporation of pretransplant MRD will be important in validating or refuting the role of pre-transplant therapy in influencing pre-transplant MRD status.

In patients with comorbidities and a high chance of induction related death following intensive chemotherapy, in whom a curative pathway is still intended (87, 88), a less intensive approach may be valid prior to transplant. With the increasing availability of venetoclax based regimens, data will likely emerge as to the transplant outcomes of patients who have a remission following these lower intensity approaches as compared to conventional intensive induction regimens. At present, data on this cohort remains limited, as these regimens have been developed in cohorts of less fit individuals in which the overall transplant rates have been low (89). Certainly, it is well established that patients with AML who have non-proliferative disease, or transformed MDS can have durable remissions with azacitidine alone (90), and patients who proceed to transplant in remission may have long term outcomes which is comparable to those who have remissions from IC (91-93). Although, remission rates for patients receiving non-intensive treatment such as Azacitidine are likely to be inferior as compared to conventional induction chemotherapy alone (94-96), it is unclear whether for patients who do remit, pre-transplant MRD levels are affected by treatment intensity, and whether this has subsequent impact on post-transplant outcomes.

Can Changes in Conditioning and GVHD Prophylaxis Alter the Impact of Pre-Transplant MRD?

MRD as measured by error corrected NGS was performed in patients with AML who were enrolled onto the BMT CTN 0901 study which performed a randomized comparison of RIC versus MAC regimens (15). In a comparison of patients who were NGS MRD positive pre-transplant, patients who received a RIC regimen had an inferior outcome to those who were MRD negative at the same timepoint (83). In contrast, in patients transplanted with a MAC regimen, levels of MRD pre-transplant did not appear to affect outcomes post-transplant. This suggested that it was possible to alter transplant conditioning to improve outcomes of patients with MRD pre-transplant, but in practice would be limited to younger patients who would be eligible to receive a MAC regimen regardless (see below).

For those with NPM1 mutant transcripts pre-transplant, the risk of relapse post-transplant is increased. However, this is also dependent on the concomitant FLT3-ITD mutation status (97). The identification of T-cell depletion as an adverse risk factor in the whole cohort, and in those with positive NPM1 MRD pre-transplant, suggest a possible transplant strategy that may improve outcomes for this subset of patients.

Improving Conditioning Regimens for Patients With AML

Transplant conditioning regimens have evolved since the establishment of allo-SCT as a pivotal tool in reducing relapse risk in patients with AML. MAC regimens established the benefits of an allo-SCT in patients with AML (43, 98) but patients over the age of 40 experienced excess toxicity historically. In the last two decades the increased use of RIC regimens has allowed the routine delivery of an allo-SCT to patients over the age of 70 (77). In recent years the efforts of a number transplant cooperative groups have delivered important randomized controlled trials to optimize transplant conditioning regimens to further inform choice of conditioning regimens (12, 15, 99, 100).

What Is the Optimal Conditioning Intensity?

A MAC regimen by definition requires the infusion of donor stem cells to rescue recipients from permanent bone marrow aplasia. The original studies in allo-SCT used conditioning regimens based on radiotherapy (1). This established the basic principles required of any conditioning regimen in acute leukemia, which is to allow durable engraftment of donor hematopoiesis as well as the delivery of an anti-leukemic effect, which is in turn related to the intensity of conditioning (101).

Cyclophosphamide (Cy) based conditioning combined with total body irradiation (TBI) or busulphan are acceptable MAC regimens. The development of intravenous preparations of busulphan has improved the pharmacokinetics of this agent (102) and has practical advantages over TBI based regimens. Measuring busulphan pharmacokinetics may help predict optimal doses in conditioning (103). Cy/TBI regimens are still commonly used and may be better for patients with either central nervous system (CNS) disease or myeloid sarcoma. Nevertheless, a pivotal randomized controlled trial that demonstrated the superior tolerability of a Fludarabine/Busulphan (Flu/Bu4: 12.8 mg/kg over 4 days of IV busulfan) combination over a standard Cyclophosphamide/Busulphan combination, with acceptable tolerability in patients up to the age of 65 (12). This has resulted in the Flu/Bu4 regimen being accepted as a standard of care for fit patients where a MAC regimen is desired.

RIC regimens result in varying duration of cytopenias and are defined as containing less than ≤ 8 Gy Total Body Irradiation (TBI) or ≤ 8 mg/kg busulfan (104). The optimal RIC regimen has not been established. A number of RIC regimens have been developed over the last twenty years to enable a tolerable conditioning regimen to be delivered in patients due to either comorbidities or increased age, with varying levels of toxicity and anti-leukemic potency (e.g. Flu/Bu2: 6.4mg/kg, 2 days of IV

busulphan) (105), and Flu/melphalan (140 mg/m² of IV melphalan on 1 day) (106). The variability in the effectiveness of these regimens are exemplified by two randomized controlled trials (RCT) of RIC regimens. One study which compared the outcomes of a Flu/2Gy TBI regimen with a Flu/Bu2 regimen demonstrated increased TRM but notable decrease in relapse rates with the Flu/Bu2 regimen (107). In contrast, a recent Flu/Treosulfan study showed superior toxicity incidence to a Flu/Bu2 comparison, but is notable for a TRM in the Flu/Bu2 arm that is far in excess of historical expectations (108).

Given the improved tolerability of novel MAC regimens (12) alongside widespread experience with RIC regimens an important question arose as to whether a MAC or RIC regimen should be selected when either is available in high risk MDS and AML (109, 110). Despite this interest it was surprising that two RCTs comparing RIC and MAC regimens closed early to recruitment but did not demonstrate significant differences in relapse free or overall survival (99, 100, 111). In contrast, a Blood and Marrow Transplant Clinical Trials Network (BMT CTN) study (15) which studied a randomized comparison of RIC versus MAC regimens demonstrated a lower rate of TRM, but higher relapse risk resulting in an inferior relapse free survival (RFS) in patients receiving in the RIC arm as compared to those who received a MAC regimen. However, this study is notable for the higher than expected relapse risk in patients who received a RIC regimen.

The high relapse rates associated with RIC regimens, for patients with high risk AML resulted in the development of the FIGARO study, which compared the outcomes of a standard RIC arm with an augmented RIC schedule with sequential chemotherapy (FLAMSA-Bu) which had shown promising results in early studies in patients with primary refractory disease (112). However, this randomized controlled study demonstrated no improvement in relapse risk from the FLAMSA-Bu regimen as compared to a standard control arm (14).

GVHD Prophylaxis Strategies

The introduction of Ciclosporin was critical in establishing the deliverability of allo-SCT in patients with acute leukemia (113, 114) reducing the risk of graft-versus-host disease (GVHD). However, studies that demonstrated an inverse relationship between GVHD and relapse risk form the basis of the evidence underlying the GVL (115, 116). Commensurate with this observation, further studies demonstrated a relationship between ciclosporin exposure and risk of relapse, in the context of T-cell depleted allo-SCT (21, 117). Tacrolimus (FK506) has also been compared with Ciclosporin in a number of randomized trials with varying results (118-120), suggesting a reduction in acute GVHD with the use of Tacrolimus but no significant effect on OS or RFS. Other agents such as Sirolimus (121, 122) and Mycophenolate mofetil (123, 124) have also been used either as an addition or substitute for historical Ciclosporin/ Methotrexate combination without a definitive improvement in overall outcomes.

In vivo T-cell depletion can be achieved by either Antithymocyte globulin (ATG) or Alemtuzumab. Studies demonstrate an improvement in risk of acute GVHD without significant changes in OS (125, 126). However a US retrospective study suggested that ATG compromised relapse risk in patients undergoing a RIC allo-SCT (127) which has led to a discrepancy in the uptake of ATG on the two continents (128). More recent data suggest that variations in vivo levels of ATG may result in differences in relapse risk as well as NRM (129). It is also important to note that there appear to be different immunosuppressive properties dependent on the source of ATG, which is critical when different studies are compared (130). The humanized anti-CD52 antibody, Alemtuzumab has also been used extensively as a method of *in vivo* T-cell depletion (131, 132), with control of GVHD particularly notable in the HLA-mismatch setting (133). In more recent years, the use of post-transplant Cyclophosphamide which was pioneered for use in the haploidentical donor allo-SCT setting (134) has been used in the volunteer unrelated donor setting (135) but formal assessment in the clinical trial setting is awaited.

The variation in relapse rate from study to study for these different GVHD prophylaxis studies suggest the need to perform adequately powered studies with suitable endpoints, in order to determine the optimal GVHD prophylaxis strategies in AML.

How to Improve Outcomes of Patients With AML Post-Transplant

Improving Monitoring of Disease Post-Transplant

Whilst the cornerstone of post-transplant care remains careful clinical assessment and review, post-transplant disease monitoring to identify patients at risk of relapse, and timely intervention is becoming more important. This is particularly important with the increased use of RIC allo-SCT which is associated with a higher risk of relapse (15). Furthermore, the use of pre-emptive treatment before fulminant hematological relapse may increase the efficacy of interventions such as donor lymphocyte infusion (DLI) or Azacitidine (136–139).

MRD Monitoring Post-Transplant

Prior to hematological relapse, the prognosis of which is usually very poor, early disease re-emergence can be detected by several techniques. The ELN guidelines formally recommend monitoring for MRD post-transplant (33). Similar to pretransplant, the optimal method for monitoring MRD will be dependent on disease characteristics, and availability of technology, and expertise in the treating center. Posttransplant MRD monitoring has prognostic value. For example, the (8, 21) fusion transcript RUNX1/RUNX1T1 is suitable for MRD monitoring and has been investigated posttransplant (60, 140, 141). Similar to pre-transplant, detectable RUNX1/RUNX1T1 transcripts at 3 months after transplant was a more potent predictor of relapse than presence of c-KIT mutations (141). The most prognostic threshold of MRD may be different after transplant, as compared to that of the pretransplant setting. For example, one study determined the prognostic impact of NPM1 MRD pre- and post-transplant and found that 1% increase in transcripts pre-transplant and a 10% increase post-transplant were predictive of outcome (142). A combination of multiple methods to detect MRD may be

required to provide the most accurate prognostic information. For example combining NGS MRD for NPM1 with multicolor flow cytometry may improve relapse prediction over either modality alone (143).

Discrepancies between the most discriminatory MRD thresholds at different treatment stages illustrate how the preand post-transplant bone marrow environment is different; post-transplant, there is a complex immunological milieu of developing tolerance and GVL. As not all patients with MRD relapse, it is postulated that the GVL effect may eradicate residual disease without the need for further intervention. Although it is also logical that early intervention for patients with molecular MRD would be beneficial, there is limited evidence to support this strategy. In a sub-analysis of patients included in the UK AML17 trial, the provision of post-transplant MRD information to clinicians did not affect outcomes – although this was not a randomized comparison, and not a main aim of the study (97).

Chimerism

Post-transplant monitoring of host-donor hematopoietic chimerism is a widely used post-transplant monitoring strategy, particularly after RIC allo-SCT. Chimerism can be measured in the whole blood, or specifically in T cells (CD3+ selected) or myeloid cells (CD33+). It is known that patients with mixed chimerism post-RIC allo-SCT do have an increased risk of relapse (144), although it should be noted that chimerism and residual disease are conceptually different. Mixed chimerism does not necessarily mean the presence of residual disease, nor does complete chimerism confirm its absence. In haploidentical allo-SCT disease relapse can occur due to acquired uniparental disomy of chromosome 6p leading to loss of the mismatched HLA-haplotype on leukemia cells and subsequent immune escape (145, 146). In this context, chimerism measurement by disparate methodologies can yield different results: recipient non-HLA marker based chimerism shows an increase during relapse, whilst HLA marker based chimerism remains low in disease relapse driven by a loss of HLA (147). Nevertheless chimerism monitoring, post RIC allo-SCT is an important way of identifying patients at high risk of relapse in whom intervention with pre-emptive DLI may be beneficial. Patients who achieve full donor chimerism (FDC) with DLI have a comparable outcome to those who reach FDC spontaneously (148, 149).

There may be ways to improve the performance of chimerism monitoring, including earlier use post-transplant (150), in CD34+ cells (151–154), and, in combination with monitoring for MRD. Waterhouse et al. compared the utility of chimerism and molecular monitoring including WT1 over-expression. Of 15/70 patients in whom increasing mixed chimerism was detected, all had a positive MRD marker and/or increased WT1 expression. They found that in half, detectable MRD and mixed chimerism occurred at the same time but in the other half, mixed chimerism preceded MRD positivity (155). The FIGARO study demonstrated that the risk of relapse following pre-transplant MRD positivity, is reduced by the achievement of full donor chimerism (14), and is a key finding that should direct future treatment strategies to identify methods of increasing the rate of achieving full donor chimerism.

Post-Transplant Maintenance Strategies to Reduce Relapse

Post-transplant pharmacological interventions may have direct activity on malignant cells, and there is improving understanding that modulation of the complex immunological environment may provide additional benefit. There is improving interest in assessing the impact of routine, maintenance treatments, which do not significantly add to the burden of toxicity which includes infection, organ toxicity, and GVHD (**Table 3** and **Figure 3**).

Non-Targeted Agents

Non-targeted agents which modulate the immune system and tumor microenvironment have the advantage that they are generalizable, are not dependent on specific mutations and may maintain efficacy across the patchwork of clonally heterogeneous disease which is rapidly changing in the post-transplant bone marrow (165, 166).

Azacitidine is an epigenetic modulator that has efficacy in AML both as sole therapy and in combination with other treatments. Post-transplant, in the RICAZA study, Azacitadine was shown to be well tolerated and may both reduce the risk of GVHD through regulatory T-cell expansion and augment the GVL through upregulation of cancer associated antigens on leukemia cells (139, 156, 167). Azacitadine has also been studied in the RELAZA (157) and RELAZA2 (84) studies whereby patients were with mixed CD34+ chimerism and MRD positivity respectively were offered single-agent Azacitadine. In RELAZA, 80% patients responded and Azacitadine delayed relapse. In RELAZA2, relapse free survival at 12 months was 46% in those who had MRD detected and received Azacitadine, suggesting a delaying of haematological relapse. Despite this, a phase 3 RCT of azacitadine versus observation did not show evidence of survival benefit when used as post-transplant maintenance for patients with high risk AML, although this study was limited by the short duration of time that patients remained on treatment (158). The oral formulation of Azacitidine (CC-486) and Panobinostat, another epigenetic modulator have also shown promise in early phase studies and are both the subject of on-going RCTs (NCT04173533 and NCT04326764 respectively) (168, 169). Lenalidomide, an immunomodulator, in combination with Azacitidine is also active in post-transplant relapse (170) but is associated with GVHD when used as monotherapy in the maintenance setting (159) thus indicating the importance of studying the effects of drugs in this specific treatment stage.

DLI can induce remission in patients with hematological relapse, eradicate MRD and promote reversion to full donor chimerism. Alternatively, prophylactic DLI can be delivered to patients at high risk of relapse regardless of detectable disease. A recent observational, matched-pair study found that prophylactic DLI in patients with high-risk AML increased OS at five years by 30% (164). The on-going prospective, 2-arm, phase II PRO-DLI randomized trial will add valuable further information in this area (171). There are also developing technology to manipulate DLI to improve efficacy and limit toxicity. These are reviewed elsewhere, and studies are ongoing (172).

TABLE 3 | Examples of post-transplant maintenance strategies.

		Mechanism	Examples of use
Non-targeted	Azacitidine	Epigenetic modulator	RICAZA (2016)
agents			Phase II trial, azacitidine single agent, n=37. Reduced GvHD (156).
			RELAZA (2012)
			Phase II trial, azacitadine single agent for mixed CD34+ chimerism, n=20.
			80% responded (157).
			RELAZA2 (2018)
			Phase II trial, azacitadine single agent for MRD+ patients, n=55. Relapse free surviva
			at 12 months 46% (84).
			Oran et al. (2020)
			Phase III trial, n=187. No difference in relapse free survival or overall survival (158).
	Oral azacitidine	Epigenetic modulator	On-going phase III trial
			NCT04173533 (oral azacitidine versus placebo).
	Panobinostat	Epigenetic modulator	On-going phase II trial NCT04326764
	Lenalidomide	Immunomodulator	LENAMAINT (2012)
			Phase II trial, n=10. Stopped early due to high incidence of severe acute GVHD (159
Targeted	Sorafenib	Broad-spectrum tyrosine	SORMAIN study (2020)
agents		kinase inhibitor	Randomised phase II, n=83, FLT3-ITD. Improved relapse free survival at 2 years (85)
			versus 53%) (160).
			Xuan et al. (13)
			Randomised phase III, n=202, FLT3-ITD. Reduced relapse at 1 year (7% versus 24%) (13
	Midostaurin	Broad-spectrum tyrosine	RADIUS study (2020)
		kinase inhibitor	Phase II, n=60 (161).
	Gilteritinib	FLT-3 inhibitor	On-going phase III trial
			NCT02997202 (gilteritinib versus placebo).
	Venetoclax	BCL-2 inhibitor	Kent et al. (2020) (abstract)
			Phase II, n=23. 6 month leukemia free survival: 87% (162).
			On-going trials
			Venetoclax + azacitidine. NCT04161885 (phase III) and NCT04128501 (phase II).
	Glasdegib	Hedgehog inhibitor	Kent et al. (2020)
			Phase II, n=31, high risk patients. No apparent benefit (163).
	Ivosidenib	IDH-1 inhibitor	On-going phase I trial NCT03728335
	Enasidenib	IDH-2 inhibitor	On-going phase I trial NCT03564821
Cellular	Prophylactic donor lymphocyte	Graft-versus-leukemia	Schmid et al. (2019)
herapy	infusion (DLI)	effect	Retrospective matched-pair study of prophylactic DLI for high-risk disease. Overall
			survival benefit (69.8% vs. 40.2%) (164).
			On-going phase II trial NCT02856464

Targeted Agents and Future Areas of Development

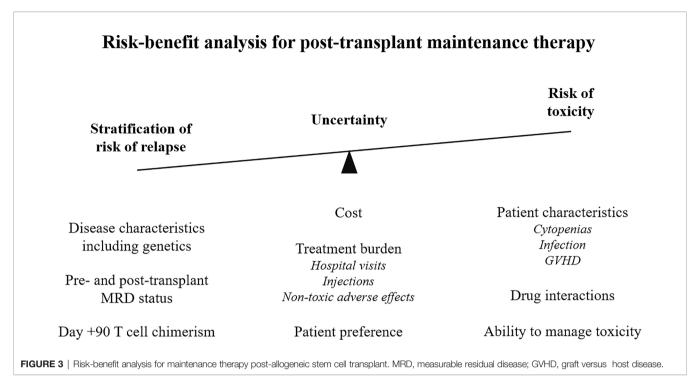
Routine application of NGS for DNA mutations have allowed for the identification of dysregulated, druggable pathways in AML. Many are only applicable to a subset of patients, but may also offer the first rung on the ladder of personalized medicine. The major challenges include identification of suitable, druggable targets in the context of clonal heterogeneity (165), and proving clinical efficacy when patient subgroups are relatively small.

An ever-expanding list of targeted treatments directed against key pathways in AML have received Food and Drug Administration FDA approval in recent years. FLT3, as described above is a tyrosine kinase, mutations in which are known to be associated with poor outcomes. In patients with FLT3 mutations, the use of post-transplant sorafenib, a broad-spectrum tyrosine kinase inhibitor (including FLT3), was associated with improved survival compared with placebo (13), findings that were consistent with the phase II SORMAIN study (160). As discussed earlier, the use of another broad-spectrum FLT3 inhibitor, midostaurin along with induction chemotherapy improves outcomes in FLT3-mutated AML (50). In the post-transplant setting, evidence of benefit from midostaurin is limited to a randomized phase II study (RADIUS) which showed a

reduction in relapse with midostaurin treatment post-transplant albeit compared with historical controls (173).

Despite some evidence of benefit, there remain concerns about the off-target toxicity and adverse events associated with the broadspectrum tyrosine kinase inhibitors. The aforementioned SORMAIN study found that the patients most likely to benefit from sorafenib post-transplant were those in whom MRD was detectable (160). For treatments where there are concerns over toxicity, especially in patients with more comorbidities, it is clear that post-transplant disease monitoring can add vital information for assessment of the risk-benefit equation. Second generation drugs which are potent, more specific FLT3 inhibitors are now available and have efficacy as monotherapy in relapsed AML (37). Clinical evaluation of Gilteritinib for post-transplant maintenance is underway (174).

Other targets of small molecule inhibitors include the antiapoptotic protein BCL2, the Hedgehog signaling pathway, and isocitrate dehydrogenase 1 and 2 (IDH1 & 2). Venetoclax is a selective BCL2 inhibitor which is currently licensed in combination with Azacitidine for the treatment of older patients who are not suitable for intensive treatment and was found to have a substantial survival benefit in this cohort when



compared with Azacitidine monotherapy (89). In a small study in the post-transplant maintenance setting, Venetoclax was reported to be safe and well tolerated but further studies are required to demonstrate benefit (162). Venetoclax is also being assessed in combination with Azacitadine as maintenance therapy post-transplant (175, 176) but its application may be limited by concerns over myelosuppression.

Glasdegib is an inhibitor of the Hedgehog signaling pathway which has evidence of modest benefit in combination with low dose Cytarabine for patients unfit for intensive treatment (38). It has been recently evaluated in a small single arm study in unselected high-risk patients in the post-transplant maintenance setting. However, there was no clear evidence of benefit either measured by MRD elimination, change in chimerism status, or clinical outcomes. Additionally, treatment was complicated by adverse events requiring pausing or cessation of treatment (163). Further studies in patients who are most likely to benefit as identified by genetic pre-stratification are required.

IDH1 and 2 are proteins which mediate the conversion of isocitrate to alpha-ketoglutarate. Gain in function mutations result in DNA and histone hypermethylation and altered downstream gene expression contributing to oncogenesis. Ivosidenib and Enasidenib, IDH1 and IDH2 inhibitors respectively both have evidence of efficacy in single-arm studies in AML (177–179) and are currently being evaluated for post-transplant maintenance (180, 181).

In summary, there is emerging, encouraging evidence that post-transplant maintenance therapies can reduce the risk of relapse, modulate the risk of GVHD, and improve survival. However, their use must be balanced in order to weigh up the additional toxicity and financial burden against the magnitude of the clinical effect. Detailed molecular analysis of a patient's

disease and post-transplant disease monitoring will allow further stratification and potentially identify the patients who are most likely to benefit from treatment (summarized in **Figure 3**).

CONCLUSION

The establishment of large transplant trial networks has improved the scientific rationale behind transplant practice at every stage of the treatment pathway. This has improved the identification of which patients who are most likely to benefit from an allo-SCT, and also provides a rigorous assessment of novel agents that may benefit patients. Finally, by embedding correlative translational science in these studies, this further improves our knowledge and understanding of the scientific basis of clinical practice. This is of direct benefit to patients, and subsequently provides a vital starting place for future studies.

AUTHOR CONTRIBUTIONS

All authors contributed to the writing of this review article. All authors contributed to the article and approved the submitted version.

FUNDING

Research support and clinical trials funding from CRUK, Bloodwise and Cure Leukaemia acknowledged. Core funding to the Birmingham ECMC Centre program is gratefully acknowledged.

REFERENCES

- Thomas ED, Lochte HLJr, Lu WC, Ferrebee JW. Intravenous infusion of bone marrow in patients receiving radiation and chemotherapy. N Engl J Med (1957) 257(11):491–6. doi: 10.1056/NEJM195709122571102
- Appelbaum FR. Hematopoietic-Cell Transplantation at 50. N Engl J Med (2007) 357(15):1472–5. doi: 10.1056/NEJMp078166
- Cornelissen JJ, Breems D, Putten WLJV, Gratwohl AA, Passweg JR, Pabst T, et al. Comparative Analysis of the Value of Allogeneic Hematopoietic Stem-Cell Transplantation in Acute Myeloid Leukemia With Monosomal Karyotype Versus Other Cytogenetic Risk Categories. *J Clin Oncol* (2012) 30(17):2140–6. doi: 10.1200/JCO.2011.39.6499
- Cornelissen J. The European LeukemiaNet AML Working Party consensus statement on allogeneic HSCT for patients with AML in remission: an integrated-risk adapted approach. Nat Rev Clin Oncol (2012) 9:579–90. doi: 10.1038/nrclinonc.2012.150
- 5. Wheatley K, Burnett AK, Goldstone AH, Gray RG, Hann IM, Harrison CJ, et al. A simple, robust, validated and highly predictive index for the determination of risk-directed therapy in acute myeloid leukaemia derived from the MRC AML 10 trial. United Kingdom Medical Research Council's Adult and Childhood Leukaemia Working Parties. Br J Haematol (1999) 107 (1):69–79. doi: 10.1046/j.1365-2141.1999.01684.x
- Grimwade D. Refinement of cytogenetic classification in acute myeloid leukemia: determination of prognostic significance of rare recurring chromosomal abnormalities among 5876 younger adult patients treated in the United Kingdom Medical Research Council trials. *Blood* (2010) 116:354–65. doi: 10.1182/blood-2009-11-254441
- Papaemmanuil E, Gerstung M, Bullinger L, Gaidzik VI, Paschka P, Roberts ND, et al. Genomic Classification and Prognosis in Acute Myeloid Leukemia. N Engl J Med (2016) 374(23):2209–21. doi: 10.1056/NEJMoa1516192
- Sorror ML, Maris MB, Storb R, Baron F, Sandmaier BM, Maloney DG, et al. Hematopoietic cell transplantation (HCT)-specific comorbidity index: a new tool for risk assessment before allogeneic HCT. *Blood* (2005) 106 (8):2912–9. doi: 10.1182/blood-2005-05-2004
- Sorror ML, Giralt S, Sandmaier BM, De Lima M, Shahjahan M, Maloney DG, et al. Hematopoietic cell transplantation specific comorbidity index as an outcome predictor for patients with acute myeloid leukemia in first remission: combined FHCRC and MDACC experiences. *Blood* (2007) 110 (13):4606–13. doi: 10.1182/blood-2007-06-096966
- Freeman SD, Hills RK, Virgo P, Khan N, Couzens S, Dillon R, et al. Measurable Residual Disease at Induction Redefines Partial Response in Acute Myeloid Leukemia and Stratifies Outcomes in Patients at Standard Risk Without NPM1 Mutations. J Clin Oncol (2018) 36(15):1486–97. doi: 10.1200/JCO.2017.76.3425
- Fuchs EJ, O'Donnell PV, Eapen M, Logan BR, Antin JH, Dawson P, et al. Double unrelated umbilical cord blood versus HLA-haploidentical bone marrow transplantation (BMT CTN 1101). Blood (2021) 137(3):420–8. doi: 10.1182/blood.2020007535
- Rambaldi A, Grassi A, Masciulli A, Boschini C, Mico MC, Busca A, et al. Busulfan plus cyclophosphamide versus busulfan plus fludarabine as a preparative regimen for allogeneic haemopoietic stem-cell transplantation in patients with acute myeloid leukaemia: an open-label, multicentre, randomised, phase 3 trial. *Lancet Oncol* (2015) 16(15):1525–36. doi: 10.1016/S1470-2045(15)00200-4
- Xuan L, Wang Y, Huang F, Fan Z, Xu Y, Sun J, et al. Sorafenib maintenance in patients with FLT3-ITD acute myeloid leukaemia undergoing allogeneic haematopoietic stem-cell transplantation: an open-label, multicentre, randomised phase 3 trial. *Lancet Oncol* (2020) 21(9):1201–12. doi: 10.1016/S1470-2045(20)30455-1
- Craddock C, Jackson A, Loke J, Siddique S, Hodgkinson A, Mason J, et al. Augmented Reduced-Intensity Regimen Does Not Improve Postallogeneic Transplant Outcomes in Acute Myeloid Leukemia. *J Clin Oncol* (2020) 0(0): JCO.20.02308. doi: 10.1200/JCO.20.02308
- Scott BL, Pasquini MC, Logan BR, Wu J, Devine SM, Porter DL, et al. Myeloablative Versus Reduced-Intensity Hematopoietic Cell Transplantation for Acute Myeloid Leukemia and Myelodysplastic Syndromes. J Clin Oncol (2017) 35(11):1154-61. doi: 10.1200/ JCO.2016.70.7091

- Lancet JE, Uy GL, Cortes JE, Newell LF, Lin TL, Ritchie EK, et al. CPX-351 (cytarabine and daunorubicin) Liposome for Injection Versus Conventional Cytarabine Plus Daunorubicin in Older Patients With Newly Diagnosed Secondary Acute Myeloid Leukemia. J Clin Oncol (2018) 36(26):2684–92. doi: 10.1200/ICO.2017.77.6112
- Löwenberg B, Pabst T, Vellenga E, van Putten W, Schouten HC, Graux C, et al. Cytarabine Dose for Acute Myeloid Leukemia. N Engl J Med (2011) 364 (11):1027–36. doi: 10.1056/NEJMoa1010222
- Gooley TA, Chien JW, Pergam SA, Hingorani S, Sorror ML, Boeckh M, et al. Reduced Mortality after Allogeneic Hematopoietic-Cell Transplantation. N Engl J Med (2010) 363(22):2091–101. doi: 10.1056/NEJMoa1004383
- Burnett AK, Goldstone A, Hills RK, Milligan D, Prentice A, Yin J, et al. Curability of Patients With Acute Myeloid Leukemia Who Did Not Undergo Transplantation in First Remission. J Clin Oncol (2013) 31(10):1293–301. doi: 10.1200/JCO.2011.40.5977
- Michallet M, Bilger K, Garban F, Attal M, Huyn A, Blaise D, et al. Allogeneic Hematopoietic Stem-Cell Transplantation After Nonmyeloablative Preparative Regimens: Impact of Pretransplantation and Posttransplantation Factors on Outcome. J Clin Oncol (2001) 19 (14):3340-9. doi: 10.1200/JCO.2001.19.14.3340
- Charles C, Sandeep N, Andrew P, Cassandra B, Laura B, Emmanouil N, et al. Factors predicting long-term survival after T-cell depleted reduced intensity allogeneic stem cell transplantation for acute myeloid leukemia. Haematologica (2010) 95(6):989–95. doi: 10.3324/haematol.2009.013920
- Innes AJ, Woolley P, Szydlo RM, Lozano S, Fernando F, Bansal D, et al. Complete remission with incomplete count recovery (CRi) prior to allogeneic HCT for acute myeloid leukaemia is associated with a high non-relapse mortality. *Leukemia* (2020) 34(2):667–70. doi: 10.1038/ s41375-019-0572-z
- Tallman MS, Rowlings PA, Milone G, Zhang MJ, Perez WS, Weisdorf D, et al. Effect of postremission chemotherapy before human leukocyte antigenidentical sibling transplantation for acute myelogenous leukemia in first complete remission. *Blood* (2000) 96(4):1254–8. doi: 10.1182/ blood.V96.4.1254
- DeWolf S, Tallman MS. How I treat relapsed or refractory AML. Blood (2020) 136(9):1023–32. doi: 10.1182/blood.2019001982
- Gale RP, Horowitz MM, Rees JK, Gray RG, Oken MM, Estey EH, et al. Chemotherapy versus transplants for acute myelogenous leukemia in second remission. *Leukemia* (1996) 10(1):13–9.
- Ganzel C, Sun Z, Cripe LD, Fernandez HF, Douer D, Rowe JM, et al. Very poor long-term survival in past and more recent studies for relapsed AML patients: The ECOG-ACRIN experience. Am J Hematol (2018) 93(8):1074– 81. doi: 10.1002/ajh.25162
- 27. Gorin NC, Labopin M, Frassoni F, Milpied N, Attal M, Blaise D, et al. Identical outcome after autologous or allogeneic genoidentical hematopoietic stem-cell transplantation in first remission of acute myelocytic leukemia carrying inversion 16 or t(8;21): a retrospective study from the European Cooperative Group for Blood and Marrow Transplantation. J Clin Oncol (2008) 26(19):3183-8. doi: 10.1200/ICO.2007.15.3106
- Breems DA, Van Putten WL, Huijgens PC, Ossenkoppele GJ, Verhoef GE, Verdonck LF, et al. Prognostic index for adult patients with acute myeloid leukemia in first relapse. *J Clin Oncol* (2005) 23(9):1969–78. doi: 10.1200/ JCO.2005.06.027
- Duval M, Klein JP, He W, Cahn JY, Cairo M, Camitta BM, et al. Hematopoietic stem-cell transplantation for acute leukemia in relapse or primary induction failure. J Clin Oncol (2010) 28(23):3730–8. doi: 10.1200/ JCO.2010.28.8852
- Tauro S, Craddock C, Peggs K, Begum G, Mahendra P, Cook G, et al. Allogeneic stem-cell transplantation using a reduced-intensity conditioning regimen has the capacity to produce durable remissions and long-term disease-free survival in patients with high-risk acute myeloid leukemia and myelodysplasia. J Clin Oncol (2005) 23(36):9387–93. doi: 10.1200/ JCO.2005.02.0057
- Weisdorf DJ, Millard HR, Horowitz MM, Hyare PS, Champlin R, Ho V, et al. Allogeneic transplantation for advanced acute myeloid leukemia: The value of complete remission. *Cancer* (2017) 123(11):2025–34. doi: 10.1002/ cncr.30536

- Gilleece MH, Labopin M, Savani BN, Yakoub-Agha I, Socie G, Gedde-Dahl T, et al. Allogeneic haemopoietic transplantation for acute myeloid leukaemia in second complete remission: a registry report by the Acute Leukaemia Working Party of the EBMT. *Leukemia* (2019) 34(1):87–99. doi: 10.1038/s41375-019-0527-4
- Döhner H, Estey E, Grimwade D, Amadori S, Appelbaum FR, Büchner T, et al. Diagnosis and management of AML in adults: 2017 ELN recommendations from an international expert panel. *Blood* (2017) 129 (4):424–47. doi: 10.1182/blood-2016-08-733196
- 34. Craddock C, Nagra S, Peniket A, Brookes C, Buckley L, Nikolousis E, et al. Factors predicting long-term survival after T-cell depleted reduced intensity allogeneic stem cell transplantation for acute myeloid leukemia. Haematologica (2010) 95(6):989–95. doi: 10.3324/haematol.2009.013920
- Craddock C, Labopin M, Pillai S, Finke J, Bunjes D, Greinix H, et al. Factors predicting outcome after unrelated donor stem cell transplantation in primary refractory acute myeloid leukaemia. *Leukemia* (2011) 25(5):808– 13. doi: 10.1038/leu.2011.13
- Ferguson P, Hills RK, Grech A, Betteridge S, Kjeldsen L, Dennis M, et al. An
 operational definition of primary refractory acute myeloid leukaemia
 allowing early identification of patients who may benefit from allogeneic
 stem cell transplantation. *Haematologica* (2016) 101(11):1351–8.
 doi: 10.3324/haematol.2016.148825
- 37. Perl AE, Altman JK, Cortes J, Smith C, Litzow M, Baer MR, et al. Selective inhibition of FLT3 by gilteritinib in relapsed or refractory acute myeloid leukaemia: a multicentre, first-in-human, open-label, phase 1-2 study. Lancet Oncol (2017) 18(8):1061–75. doi: 10.1016/S1470-2045(17)30416-3
- Cortes JE, Khaled S, Martinelli G, Perl AE, Ganguly S, Russell N, et al. Quizartinib versus salvage chemotherapy in relapsed or refractory FLT3-ITD acute myeloid leukaemia (QuANTUM-R): a multicentre, randomised, controlled, open-label, phase 3 trial. *Lancet Oncol* (2019) 20(7):984–97. doi: 10.1016/S1470-2045(19)30150-0
- Najima Y, Sadato D, Harada Y, Oboki K, Hirama C, Toya T, et al. Prognostic impact of TP53 mutation, monosomal karyotype, and prior myeloid disorder in nonremission acute myeloid leukemia at allo-HSCT. Bone Marrow Transplant (2021) 56(2):334–46. doi: 10.1038/s41409-020-01016-9
- Ding L, Ley TJ, Larson DE, Miller CA, Koboldt DC, Welch JS, et al. Clonal evolution in relapsed acute myeloid leukaemia revealed by whole-genome sequencing. *Nature* (2012) 481(7382):506–10. doi: 10.1038/nature10738
- Schmid C, Labopin M, Nagler A, Niederwieser D, Castagna L, Tabrizi R, et al. Treatment, risk factors, and outcome of adults with relapsed AML after reduced intensity conditioning for allogeneic stem cell transplantation. *Blood* (2012) 119(6):1599–606. doi: 10.1182/blood-2011-08-375840
- Christopeit M, Kuss O, Finke J, Bacher U, Beelen DW, Bornhauser M, et al. Second allograft for hematologic relapse of acute leukemia after first allogeneic stem-cell transplantation from related and unrelated donors: the role of donor change. *J Clin Oncol* (2013) 31(26):3259–71. doi: 10.1200/JCO.2012.44.7961
- Koreth J, Schlenk R, Kopecky KJ, Honda S, Sierra J, Djulbegovic BJ, et al. Allogeneic stem cell transplantation for acute myeloid leukemia in first complete remission: systematic review and meta-analysis of prospective clinical trials. *JAMA* (2009) 301(22):2349–61. doi: 10.1001/jama.2009.813
- Schlenk RF, Dohner K, Krauter J, Frohling S, Corbacioglu A, Bullinger L, et al. Mutations and treatment outcome in cytogenetically normal acute myeloid leukemia. N Engl J Med (2008) 358(18):1909–18. doi: 10.1056/ NEJMoa074306
- 45. Kottaridis PD, Gale RE, Frew ME, Harrison G, Langabeer SE, Belton AA, et al. The presence of a FLT3 internal tandem duplication in patients with acute myeloid leukemia (AML) adds important prognostic information to cytogenetic risk group and response to the first cycle of chemotherapy: analysis of 854 patients from the United Kingdom Medical Research Council AML 10 and 12 trials. Blood (2001) 98(6):1752–9. doi: 10.1182/blood.V98.6.1752
- Falini B, Mecucci C, Tiacci E, Alcalay M, Rosati R, Pasqualucci L, et al. Cytoplasmic nucleophosmin in acute myelogenous leukemia with a normal karyotype. N Engl J Med (2005) 352(3):254–66. doi: 10.1056/NEJMoa041974
- 47. Metzeler KH, Becker H, Maharry K, Radmacher MD, Kohlschmidt J, Mrozek K, et al. ASXL1 mutations identify a high-risk subgroup of older patients with primary cytogenetically normal AML within the ELN

- Favorable genetic category. *Blood* (2011) 118(26):6920–9. doi: 10.1182/blood-2011-08-368225
- Mendler JH, Maharry K, Radmacher MD, Mrozek K, Becker H, Metzeler KH, et al. RUNX1 mutations are associated with poor outcome in younger and older patients with cytogenetically normal acute myeloid leukemia and with distinct gene and MicroRNA expression signatures. *J Clin Oncol* (2012) 30(25):3109–18. doi: 10.1200/JCO.2011.40.6652
- Ortmann CA, Kent DG, Nangalia J, Silber Y, Wedge DC, Grinfeld J, et al. Effect of Mutation Order on Myeloproliferative Neoplasms. N Engl J Med (2015) 372(7):601–12. doi: 10.1056/NEJMoa1412098
- Stone RM, Mandrekar SJ, Sanford BL, Laumann K, Geyer S, Bloomfield CD, et al. Midostaurin plus Chemotherapy for Acute Myeloid Leukemia with a FLT3 Mutation. N Engl J Med (2017) 377(5):454–64. doi: 10.1056/ NEIMoa1614359
- Burd A, Levine RL, Ruppert AS, Mims AS, Borate U, Stein EM, et al. Precision medicine treatment in acute myeloid leukemia using prospective genomic profiling: feasibility and preliminary efficacy of the Beat AML Master Trial. *Nat Med* (2020) 26(12):1852–8. doi: 10.1038/s41591-020-1089-8
- DiNardo CD, Tiong IS, Quaglieri A, MacRaild S, Loghavi S, Brown FC, et al. Molecular patterns of response and treatment failure after frontline venetoclax combinations in older patients with AML. *Blood* (2020) 135 (11):791–803. doi: 10.1182/blood.2019003988
- Grinfeld J, Nangalia J, Baxter EJ, Wedge DC, Angelopoulos N, Cantrill R, et al. Classification and Personalized Prognosis in Myeloproliferative Neoplasms. N Engl J Med (2018) 379(15):1416–30. doi: 10.1056/ NEJMoa1716614
- 54. Fenwarth L, Thomas X, de Botton S, Duployez N, Bourhis JH, Lesieur A, et al. A personalized approach to guide allogeneic stem cell transplantation in younger adults with acute myeloid leukemia. *Blood* (2021) 137(4):524–32. doi: 10.1182/blood.2020005524
- Craddock C, Versluis J, Labopin M, Socie G, Huynh A, Deconinck E, et al. Distinct factors determine the kinetics of disease relapse in adults transplanted for acute myeloid leukaemia. *J Internal Med* (2018) 283 (4):371–9. doi: 10.1111/joim.12720
- Schuurhuis GJ, Heuser M, Freeman S, Bene MC, Buccisano F, Cloos J, et al. Minimal/measurable residual disease in AML: a consensus document from the European LeukemiaNet MRD Working Party. *Blood* (2018) 131 (12):1275–91. doi: 10.1182/blood-2017-09-801498
- 57. Jourdan E, Boissel N, Chevret S, Delabesse E, Renneville A, Cornillet P, et al. Prospective evaluation of gene mutations and minimal residual disease in patients with core binding factor acute myeloid leukemia. *Blood* (2013) 121 (12):2213–23. doi: 10.1182/blood-2012-10-462879
- Ivey A, Hills RK, Simpson MA, Jovanovic JV, Gilkes A, Grech A, et al. Assessment of Minimal Residual Disease in Standard-Risk AML. N Engl J Med (2016) 374(5):422–33. doi: 10.1056/NEJMoa1507471
- Prébet T, Boissel N, Reutenauer S, Thomas X, Delaunay J, Cahn J-Y, et al. Acute Myeloid Leukemia With Translocation (8;21) or Inversion (16) in Elderly Patients Treated With Conventional Chemotherapy: A Collaborative Study of the French CBF-AML Intergroup. *J Clin Oncol* (2009) 27(28):4747– 53. doi: 10.1200/JCO.2008.21.0674
- 60. Yin JA, O'Brien MA, Hills RK, Daly SB, Wheatley K, Burnett AK. Minimal residual disease monitoring by quantitative RT-PCR in core binding factor AML allows risk stratification and predicts relapse: results of the United Kingdom MRC AML-15 trial. *Blood* (2012) 120(14):2826–35. doi: 10.1182/ blood-2012-06-435669
- 61. Rücker FG, Agrawal M, Corbacioglu A, Weber D, Kapp-Schwoerer S, Gaidzik VI, et al. Measurable residual disease monitoring in acute myeloid leukemia with t(8;21)(q22;q22.1): results from the AML Study Group. *Blood* (2019) 134(19):1608–18. doi: 10.1182/blood.2019001425
- 62. Balsat M, Renneville A, Thomas X, Botton SD, Caillot D, Marceau A, et al. Postinduction Minimal Residual Disease Predicts Outcome and Benefit From Allogeneic Stem Cell Transplantation in Acute Myeloid Leukemia With NPM1 Mutation: A Study by the Acute Leukemia French Association Group. J Clin Oncol (2017) 35(2):185–93. doi: 10.1200/JCO.2016.67.1875
- 63. Ostronoff F, Othus M, Lazenby M, Estey E, Appelbaum FR, Evans A, et al. Prognostic significance of NPM1 mutations in the absence of FLT3-internal tandem duplication in older patients with acute myeloid leukemia: a SWOG

- and UK National Cancer Research Institute/Medical Research Council report. J Clin Oncol (2015) 33(10):1157-64. doi: 10.1200/JCO.2014.58.0571
- 64. Angenendt L, Röllig C, Montesinos P, Martínez-Cuadrón D, Barragan E, García R, et al. Chromosomal Abnormalities and Prognosis in NPM1-Mutated Acute Myeloid Leukemia: A Pooled Analysis of Individual Patient Data From Nine International Cohorts. J Clin Oncol (2019) 37(29):2632–42. doi: 10.1200/JCO.19.00416
- Freeman SD, Virgo P, Couzens S, Grimwade D, Russell N, Hills RK, et al. Prognostic Relevance of Treatment Response Measured by Flow Cytometric Residual Disease Detection in Older Patients With Acute Myeloid Leukemia. J Clin Oncol (2013) 31(32):4123–31. doi: 10.1200/JCO.2013.49.1753
- Jongen-Lavrencic M, Grob T, Hanekamp D, Kavelaars FG, al Hinai A, Zeilemaker A, et al. Molecular Minimal Residual Disease in Acute Myeloid Leukemia. N Engl J Med (2018) 378(13):1189–99. doi: 10.1056/ NEIMoa1716863
- Ghannam J, Dillon LW, Hourigan CS. Next-generation sequencing for measurable residual disease detection in acute myeloid leukaemia. Br J Haematol (2020) 188(1):77–85. doi: 10.1111/bjh.16362
- Loke J, Malladi R, Moss P, Craddock C. The role of allogeneic stem cell transplantation in the management of acute myeloid leukaemia: a triumph of hope and experience. *Br J Haematol* (2020) 188(1):129–46. doi: 10.1111/ bib 16355
- Cornelissen JJ, Blaise D. Hematopoietic stem cell transplantation for patients with AML in first complete remission. *Blood* (2016) 127(1):62–70. doi: 10.1182/blood-2015-07-604546
- Brunet S, Labopin M, Esteve J, Cornelissen J, Socié G, Iori AP, et al. Impact
 of FLT3 internal tandem duplication on the outcome of related and
 unrelated hematopoietic transplantation for adult acute myeloid leukemia
 in first remission: a retrospective analysis. *J Clin Oncol* (2012) 30(7):735–41.
 doi: 10.1200/JCO.2011.36.9868
- Gratwohl A, Hermans J, Goldman JM, Arcese W, Carreras E, Devergie A, et al. Risk assessment for patients with chronic myeloid leukaemia before allogeneic blood or marrow transplantation. Chronic Leukemia Working Party of the European Group for Blood and Marrow Transplantation. *Lancet* (1998) 352(9134):1087–92. doi: 10.1016/S0140-6736(98)03030-X
- Gratwohl A, Stern M, Brand R, Apperley J, Baldomero H, de Witte T, et al. Risk score for outcome after allogeneic hematopoietic stem cell transplantation: a retrospective analysis. *Cancer* (2009) 115(20):4715–26. doi: 10.1002/cncr.24531
- Sorror ML, Storb RF, Sandmaier BM, Maziarz RT, Pulsipher MA, Maris MB, et al. Comorbidity-Age Index: A Clinical Measure of Biologic Age Before Allogeneic Hematopoietic Cell Transplantation. J Clin Oncol (2014) 32: (29):3249–56. doi: 10.1200/JCO.2013.53.8157
- Gerstung M, Papaemmanuil E, Martincorena I, Bullinger L, Gaidzik VI, Paschka P, et al. Precision oncology for acute myeloid leukemia using a knowledge bank approach. *Nat Genet* (2017) 49(3):332–40. doi: 10.1038/ ng.3756
- 75. Shouval R, Labopin M, Bondi O, Mishan-Shamay H, Shimoni A, Ciceri F, et al. Prediction of Allogeneic Hematopoietic Stem-Cell Transplantation Mortality 100 Days After Transplantation Using a Machine Learning Algorithm: A European Group for Blood and Marrow Transplantation Acute Leukemia Working Party Retrospective Data Mining Study. J Clin Oncol (2015) 33(28):3144–51. doi: 10.1200/JCO.2014.59.1339
- Schiffer C, Lee E, Tomiyasu T, Wiernik P, Testa J. Prognostic impact of cytogenetic abnormalities in patients with de novo acute nonlymphocytic leukemia. *Blood* (1989) 73: (1):263–70. doi: 10.1182/blood.V73.1.263. bloodiournal731263
- Muffly L, Pasquini MC, Martens M, Brazauskas R, Zhu X, Adekola K, et al. Increasing use of allogeneic hematopoietic cell transplantation in patients aged 70 years and older in the United States. *Blood* (2017) 130(9):1156–64. doi: 10.1182/blood-2017-03-772368
- 78. Nikolousis E, Nagra S, Pearce R, Perry J, Kirkland K, Byrne J, et al. Impact of pre-transplant co-morbidities on outcome after alemtuzumab-based reduced intensity conditioning allo-SCT in elderly patients: a British Society of Blood and Marrow Transplantation study. *Bone Marrow Transplant* (2015) 50(1):82–6. doi: 10.1038/bmt.2014.215
- Muffly LS, Boulukos M, Swanson K, Kocherginsky M, Cerro PD, Schroeder L, et al. Pilot Study of Comprehensive Geriatric Assessment (CGA) in

- Allogeneic Transplant: CGA Captures a High Prevalence of Vulnerabilities in Older Transplant Recipients. *Biol Blood Marrow Transplant* (2013) 19 (3):429–34. doi: 10.1016/j.bbmt.2012.11.006
- Freeman SD, Hourigan CS. MRD evaluation of AML in clinical practice: are we there yet? *Hematol Am Soc Hematol Educ Program* (2019) 2019(1):557–69. doi: 10.1182/hematology.2019000060
- Richard-Carpentier G, DiNardo CD. Single-agent and combination biologics in acute myeloid leukemia. Hematol Am Soc Hematol Educ Program (2019) 2019(1):548–56. doi: 10.1182/hematology.2019000059
- Buckley SA, Wood BL, Othus M, Hourigan CS, Ustun C, Linden MA, et al. Minimal residual disease prior to allogeneic hematopoietic cell transplantation in acute myeloid leukemia: a meta-analysis. *Haematologica* (2017) 102(5):865–73. doi: 10.3324/haematol.2016.159343
- 83. Hourigan CS, Dillon LW, Gui G, Logan BR, Fei M, Ghannam J, et al. Impact of Conditioning Intensity of Allogeneic Transplantation for Acute Myeloid Leukemia With Genomic Evidence of Residual Disease. *J Clin Oncol* (2020) 38(12):1273–83. doi: 10.1200/JCO.19.03011
- 84. Platzbecker U, Middeke JM, Sockel K, Herbst R, Wolf D, Baldus CD, et al. Measurable residual disease-guided treatment with azacitidine to prevent haematological relapse in patients with myelodysplastic syndrome and acute myeloid leukaemia (RELAZA2): an open-label, multicentre, phase 2 trial. *Lancet Oncol* (2018) 19(12):1668–79. doi: 10.1016/S1470-2045(18) 30580-1
- Araki D, Wood BL, Othus M, Radich JP, Halpern AB, Zhou Y, et al. Allogeneic Hematopoietic Cell Transplantation for Acute Myeloid Leukemia: Time to Move Toward a Minimal Residual Disease-Based Definition of Complete Remission? J Clin Oncol (2016) 34(4):329–36. doi: 10.1200/JCO.2015.63.3826
- 86. Halaburda K, Labopin M, Mailhol A, Socié G, Craddock C, Aljurf M, et al. Allogeneic stem cell transplantation in second complete remission for core binding factor acute myeloid leukemia: a study from the Acute Leukemia Working Party of the European Society for Blood and Marrow Transplantation. *Haematologica* (2020) 105(6):1723–30. doi: 10.3324/haematol.2019.222810
- 87. Krug U, Röllig C, Koschmieder A, Heinecke A, Sauerland MC, Schaich M, et al. Complete remission and early death after intensive chemotherapy in patients aged 60 years or older with acute myeloid leukaemia: a web-based application for prediction of outcomes. *Lancet* (2010) 376(9757):2000–8. doi: 10.1016/S0140-6736(10)62105-8
- Walter RB, Othus M, Borthakur G, Ravandi F, Cortes JE, Pierce SA, et al. Prediction of early death after induction therapy for newly diagnosed acute myeloid leukemia with pretreatment risk scores: a novel paradigm for treatment assignment. J Clin Oncol (2011) 29(33):4417–23. doi: 10.1200/ ICO.2011.35.7525
- DiNardo CD, Jonas BA, Pullarkat V, Thirman MJ, Garcia JS, Wei AH, et al. Azacitidine and Venetoclax in Previously Untreated Acute Myeloid Leukemia. N Engl J Med (2020) 383(7):617–29. doi: 10.1056/ NEIMoa2012971
- Fenaux P, Mufti GJ, Hellstrom-Lindberg E, Santini V, Finelli C, Giagounidis A, et al. Efficacy of azacitidine compared with that of conventional care regimens in the treatment of higher-risk myelodysplastic syndromes: a randomised, open-label, phase III study. *Lancet Oncol* (2009) 10(3):223–32. doi: 10.1016/S1470-2045(09)70003-8
- 91. Damaj G, Duhamel A, Robin M, Beguin Y, Michallet M, Mohty M, et al. Impact of azacitidine before allogeneic stem-cell transplantation for myelodysplastic syndromes: a study by the Societe Francaise de Greffe de Moelle et de Therapie-Cellulaire and the Groupe-Francophone des Myelodysplasies. *J Clin Oncol* (2012) 30(36):4533–40. doi: 10.1200/JCO.2012.44.3499
- Gerds AT, Gooley TA, Estey EH, Appelbaum FR, Deeg HJ, Scott BL. Pretransplantation therapy with azacitidine vs induction chemotherapy and posttransplantation outcome in patients with MDS. *Biol Blood Marrow Transplant* (2012) 18(8):1211–8. doi: 10.1016/j.bbmt.2012.01.009
- 93. Voso MT, Leone G, Piciocchi A, Fianchi L, Santarone S, Candoni A, et al. Feasibility of allogeneic stem-cell transplantation after azacitidine bridge in higher-risk myelodysplastic syndromes and low blast count acute myeloid leukemia: results of the BMT-AZA prospective study. *Ann Oncol* (2017) 28 (7):1547–53. doi: 10.1093/annonc/mdx154

- Alessandrino EP, Porta MGD, Pascutto C, Bacigalupo A, Rambaldi A. Should Cytoreductive Treatment Be Performed Before Transplantation in Patients With High-Risk Myelodysplastic Syndrome? *J Clin Oncol* (2013) 31 (21):2761–2. doi: 10.1200/JCO.2012.48.0525
- Sébert M, Komrokji RS, Sekeres MA, Prebet T, Cluzeau T, Santini V, et al. Impact of baseline cytogenetic findings and cytogenetic response on outcome of high-risk myelodysplastic syndromes and low blast count AML treated with azacitidine. *Leuk Res* (2017) 63:72–7. doi: 10.1016/ j.leukres.2017.10.013
- Burnett AK, Russell NH, Hills RK, Kell J, Freeman S, Kjeldsen L, et al. Addition of gemtuzumab ozogamicin to induction chemotherapy improves survival in older patients with acute myeloid leukemia. *J Clin Oncol* (2012) 30(32):3924–31. doi: 10.1200/JCO.2012.42.2964
- 97. Dillon R, Hills R, Freeman S, Potter N, Jovanovic J, Ivey A, et al. Molecular MRD status and outcome after transplantation in NPM1-mutated AML. *Blood* (2020) 135(9):680–8. doi: 10.1182/blood.2019002959
- Cornelissen JJ, van Putten WL, Verdonck LF, Theobald M, Jacky E, Daenen SM, et al. Results of a HOVON/SAKK donor versus no-donor analysis of myeloablative HLA-identical sibling stem cell transplantation in first remission acute myeloid leukemia in young and middle-aged adults: benefits for whom? *Blood* (2007) 109(9):3658–66. doi: 10.1182/blood-2006-06-025627
- Bornhäuser M, Kienast J, Trenschel R, Burchert A, Hegenbart U, Stadler M, et al. Reduced-intensity conditioning versus standard conditioning before allogeneic haemopoietic cell transplantation in patients with acute myeloid leukaemia in first complete remission: a prospective, open-label randomised phase 3 trial. *Lancet Oncol* (2012) 13(10):1035–44. doi: 10.1016/S1470-2045 (12)70349-2
- 100. Kroger N, Iacobelli S, Franke GN, Platzbecker U, Uddin R, Hubel K, et al. Dose-Reduced Versus Standard Conditioning Followed by Allogeneic Stem-Cell Transplantation for Patients With Myelodysplastic Syndrome: A Prospective Randomized Phase III Study of the EBMT (RICMAC Trial). J Clin Oncol (2017) 35(19):2157–64. doi: 10.1200/JCO.2016.70.7349
- 101. Clift RA, Buckner CD, Appelbaum FR, Bearman SI, Petersen FB, Fisher LD, et al. Allogeneic marrow transplantation in patients with acute myeloid leukemia in first remission: a randomized trial of two irradiation regimens. Blood (1990) 76(9):1867–71. doi: 10.1182/blood.V76.9.1867.1867
- 102. Andersson BS, Kashyap A, Gian V, Wingard JR, Fernandez H, Cagnoni PJ, et al. Conditioning therapy with intravenous busulfan and cyclophosphamide (IV BuCy2) for hematologic malignancies prior to allogeneic stem cell transplantation: a phase II study. Biol Blood Marrow Transplant (2002) 8(3):145–54. doi: 10.1053/bbmt.2002.v8.pm11939604
- 103. Esteves I, Santos FPS, Fernandes JF, Seber A, Oliveira JSR, Hamerschlak N, et al. Pharmacokinetics analysis results are similar for oral compared to intravenous busulfan in patients undergoing hematopoietic stem cell transplantation, except for the earlier onset of mucositis. A controlled clinical study. Bone Marrow Transplant (2019) 54(11):1799–804. doi: 10.1038/s41409-019-0521-5
- 104. Bacigalupo A, Ballen K, Rizzo D, Giralt S, Lazarus H, Ho V, et al. Defining the intensity of conditioning regimens: working definitions. *Biol Blood Marrow Transplant* (2009) 15(12):1628–33. doi: 10.1016/j.bbmt.2009.07.004
- 105. Slavin S, Nagler A, Naparstek E, Kapelushnik Y, Aker M, Cividalli G, et al. Nonmyeloablative stem cell transplantation and cell therapy as an alternative to conventional bone marrow transplantation with lethal cytoreduction for the treatment of malignant and nonmalignant hematologic diseases. *Blood* (1998) 91(3):756–63. doi: 10.1182/blood.V91.3.756.756_756_763
- 106. Giralt S, Estey E, Albitar M, van Besien K, Rondon G, Anderlini P, et al. Engraftment of allogeneic hematopoietic progenitor cells with purine analog-containing chemotherapy: harnessing graft-versus-leukemia without myeloablative therapy. *Blood* (1997) 89(12):4531–6. doi: 10.1182/ blood.V89.12.4531
- 107. Blaise D, Tabrizi R, Boher JM, Le Corroller-Soriano AG, Bay JO, Fegueux N, et al. Randomized study of 2 reduced-intensity conditioning strategies for human leukocyte antigen-matched, related allogeneic peripheral blood stem cell transplantation: prospective clinical and socioeconomic evaluation. *Cancer* (2013) 119(3):602–11. doi: 10.1002/cncr.27786
- 108. Beelen DW, Trenschel R, Stelljes M, Groth C, Masszi T, Reményi P, et al. Treosulfan or busulfan plus fludarabine as conditioning treatment before

- allogeneic haemopoietic stem cell transplantation for older patients with acute myeloid leukaemia or myelodysplastic syndrome (MC-FludT.14/L): a randomised, non-inferiority, phase 3 trial. *Lancet Haematol* (2020) 7(1):e28–39. doi: 10.1016/S2352-3026(19)30157-7
- 109. Martino R, Iacobelli S, Brand R, Jansen T, van Biezen A, Finke J, et al. Retrospective comparison of reduced-intensity conditioning and conventional high-dose conditioning for allogeneic hematopoietic stem cell transplantation using HLA-identical sibling donors in myelodysplastic syndromes. *Blood* (2006) 108(3):836–46. doi: 10.1182/ blood-2005-11-4503
- 110. Martino R, de Wreede L, Fiocco M, van Biezen A, von dem Borne PA, Hamladji RM, et al. Comparison of conditioning regimens of various intensities for allogeneic hematopoietic SCT using HLA-identical sibling donors in AML and MDS with <10% BM blasts: a report from EBMT. Bone Marrow Transplant (2013) 48(6):761–70. doi: 10.1038/bmt.2012.236</p>
- 111. Fasslrinner F, Schetelig J, Burchert A, Kramer M, Trenschel R, Hegenbart U, et al. Long-term efficacy of reduced-intensity versus myeloablative conditioning before allogeneic haemopoietic cell transplantation in patients with acute myeloid leukaemia in first complete remission: retrospective follow-up of an open-label, randomised phase 3 trial. *Lancet Haematol* (2018) 5(4):e161–e9. doi: 10.1016/S2352-3026(18)30022-X
- 112. Schmid C, Schleuning M, Ledderose G, Tischer J, Kolb HJ. Sequential regimen of chemotherapy, reduced-intensity conditioning for allogeneic stem-cell transplantation, and prophylactic donor lymphocyte transfusion in high-risk acute myeloid leukemia and myelodysplastic syndrome. *J Clin Oncol* (2005) 23(24):5675–87. doi: 10.1200/JCO.2005.07.061
- 113. Powles RL, Clink HM, Spence D, Morgenstern G, Watson JG, Selby PJ, et al. Cyclosporin a to prevent graft-versus-host disease in man after allogeneic bone-marrow transplantation. *Lancet* (1980) 315(8164):327–9. doi: 10.1016/ S0140-6736(80)90881-8
- 114. Storb R, Deeg HJ, Pepe M, Appelbaum F, Anasetti C, Beatty P, et al. Methotrexate and cyclosporine versus cyclosporine alone for prophylaxis of graft-versus-host disease in patients given HLA-identical marrow grafts for leukemia: long-term follow-up of a controlled trial. *Blood* (1989) 73 (6):1729–34. doi: 10.1182/blood.V73.6.1729.1729
- 115. Weiden PL, Flournoy N, Thomas ED, Prentice R, Fefer A, Buckner CD, et al. Antileukemic effect of graft-versus-host disease in human recipients of allogeneic-marrow grafts. N Engl J Med (1979) 300(19):1068–73. doi: 10.1056/NEJM197905103001902
- 116. Inamoto Y, Flowers ME, Lee SJ, Carpenter PA, Warren EH, Deeg HJ, et al. Influence of immunosuppressive treatment on risk of recurrent malignancy after allogeneic hematopoietic cell transplantation. *Blood* (2011) 118(2):456– 63. doi: 10.1182/blood-2011-01-330217
- 117. Bacigalupo A, Van Lint MT, Occhini D, Gualandi F, Lamparelli T, Sogno G, et al. Increased risk of leukemia relapse with high-dose cyclosporine A after allogeneic marrow transplantation for acute leukemia. *Blood* (1991) 77 (7):1423–8. doi: 10.1182/blood.V77.7.1423.bloodjournal7771423
- 118. Ratanatharathorn V, Nash RA, Przepiorka D, Devine SM, Klein JL, Weisdorf D, et al. Phase III study comparing methotrexate and tacrolimus (prograf, FK506) with methotrexate and cyclosporine for graft-versus-host disease prophylaxis after HLA-identical sibling bone marrow transplantation. *Blood* (1998) 92(7):2303–14.
- 119. Hiraoka A, Ohashi Y, Okamoto S, Moriyama Y, Nagao T, Kodera Y, et al. Phase III study comparing tacrolimus (FK506) with cyclosporine for graft-versus-host disease prophylaxis after allogeneic bone marrow transplantation. Bone Marrow Transplant (2001) 28(2):181–5. doi: 10.1038/sj.bmt.1703097
- Ram R, Gafter-Gvili A, Yeshurun M, Paul M, Raanani P, Shpilberg O. Prophylaxis regimens for GVHD: systematic review and meta-analysis. *Bone Marrow Transplant* (2009) 43(8):643–53. doi: 10.1038/bmt.2008.373
- 121. Pidala J, Kim J, Jim H, Kharfan-Dabaja MA, Nishihori T, Fernandez HF, et al. A randomized phase II study to evaluate tacrolimus in combination with sirolimus or methotrexate after allogeneic hematopoietic cell transplantation. *Haematologica* (2012) 97(12):1882–9. doi: 10.3324/haematol.2012.067140
- 122. Furlong T, Kiem HP, Appelbaum FR, Carpenter PA, Deeg HJ, Doney K, et al. Sirolimus in combination with cyclosporine or tacrolimus plus methotrexate for prevention of graft-versus-host disease following hematopoietic cell

- transplantation from unrelated donors. Biol Blood Marrow Transplant (2008) 14(5):531-7. doi: 10.1016/j.bbmt.2008.02.009
- 123. Bolwell B, Sobecks R, Pohlman B, Andresen S, Rybicki L, Kuczkowski E, et al. A prospective randomized trial comparing cyclosporine and short course methotrexate with cyclosporine and mycophenolate mofetil for GVHD prophylaxis in myeloablative allogeneic bone marrow transplantation. Bone Marrow Transplant (2004) 34(7):621–5. doi: 10.1038/sj.bmt.1704647
- 124. Perkins J, Field T, Kim J, Kharfan-Dabaja MA, Fernandez H, Ayala E, et al. A randomized phase II trial comparing tacrolimus and mycophenolate mofetil to tacrolimus and methotrexate for acute graft-versus-host disease prophylaxis. *Biol Blood Marrow Transplant* (2010) 16(7):937–47. doi: 10.1016/j.bbmt.2010.01.010
- 125. Finke J, Bethge WA, Schmoor C, Ottinger HD, Stelljes M, Zander AR, et al. Standard graft-versus-host disease prophylaxis with or without anti-T-cell globulin in haematopoietic cell transplantation from matched unrelated donors: a randomised, open-label, multicentre phase 3 trial. *Lancet Oncol* (2009) 10(9):855–64. doi: 10.1016/S1470-2045(09)70225-6
- 126. Socié G, Schmoor C, Bethge WA, Ottinger HD, Stelljes M, Zander AR, et al. Chronic graft-versus-host disease: long-term results from a randomized trial on graft-versus-host disease prophylaxis with or without anti-T-cell globulin ATG-Fresenius. *Blood* (2011) 117(23):6375–82. doi: 10.1182/blood-2011-01-329821
- 127. Soiffer RJ, LeRademacher J, Ho V, Kan F, Artz A, Champlin RE, et al. Impact of immune modulation with anti-T-cell antibodies on the outcome of reduced-intensity allogeneic hematopoietic stem cell transplantation for hematologic malignancies. *Blood* (2011) 117(25):6963–70. doi: 10.1182/blood-2011-01-332007
- 128. Ruutu T, van Biezen A, Hertenstein B, Henseler A, Garderet L, Passweg J, et al. Prophylaxis and treatment of GVHD after allogeneic haematopoietic SCT: a survey of centre strategies by the European Group for Blood and Marrow Transplantation. Bone Marrow Transplant (2012) 47(11):1459–64. doi: 10.1038/bmt.2012.45
- 129. Admiraal R, Nierkens S, de Witte MA, Petersen EJ, Fleurke GJ, Verrest L, et al. Association between anti-thymocyte globulin exposure and survival outcomes in adult unrelated haemopoietic cell transplantation: a multicentre, retrospective, pharmacodynamic cohort analysis. *Lancet Haematol* (2017) 4 (4):e183–e91. doi: 10.1016/S2352-3026(17)30029-7
- 130. Atta EH, de Sousa AM, Schirmer MR, Bouzas LF, Nucci M, Abdelhay E. Different outcomes between cyclophosphamide plus horse or rabbit antithymocyte globulin for HLA-identical sibling bone marrow transplant in severe aplastic anemia. *Biol Blood Marrow Transplant* (2012) 18 (12):1876–82. doi: 10.1016/j.bbmt.2012.07.004
- Chakraverty R, Peggs K, Chopra R, Milligan DW, Kottaridis PD, Verfuerth S, et al. Limiting transplantation-related mortality following unrelated donor stem cell transplantation by using a nonmyeloablative conditioning regimen. *Blood* (2002) 99(3):1071–8. doi: 10.1182/blood.V99.3.1071
- 132. Perez-Simon JA, Kottaridis PD, Martino R, Craddock C, Caballero D, Chopra R, et al. Nonmyeloablative transplantation with or without alemtuzumab: comparison between 2 prospective studies in patients with lymphoproliferative disorders. *Blood* (2002) 100(9):3121–7. doi: 10.1182/blood-2002-03-0701
- 133. Mead AJ, Thomson KJ, Morris EC, Mohamedbhai S, Denovan S, Orti G, et al. HLA-mismatched unrelated donors are a viable alternate graft source for allogeneic transplantation following alemtuzumab-based reduced-intensity conditioning. *Blood* (2010) 115(25):5147–53. doi: 10.1182/blood-2010-01-265413
- 134. Luznik L, O'Donnell PV, Symons HJ, Chen AR, Leffell MS, Zahurak M, et al. HLA-haploidentical bone marrow transplantation for hematologic malignancies using nonmyeloablative conditioning and high-dose, posttransplantation cyclophosphamide. *Biol Blood Marrow Transplant* (2008) 14(6):641–50. doi: 10.1016/j.bbmt.2008.03.005
- 135. Battipaglia G, Labopin M, Kroger N, Vitek A, Afanasyev B, Hilgendorf I, et al. Posttransplant cyclophosphamide vs antithymocyte globulin in HLA-mismatched unrelated donor transplantation. *Blood* (2019) 134(11):892–9. doi: 10.1182/blood.2019000487
- 136. Schmid C, Labopin M, Nagler A, Bornhauser M, Finke J, Fassas A, et al. Donor lymphocyte infusion in the treatment of first hematological relapse after allogeneic stem-cell transplantation in adults with acute myeloid

- leukemia: a retrospective risk factors analysis and comparison with other strategies by the EBMT Acute Leukemia Working Party. *J Clin Oncol* (2007) 25(31):4938–45. doi: 10.1200/JCO.2007.11.6053
- 137. Hofmann S, Götz M, Schneider V, Guillaume P, Bunjes D, Döhner H, et al. Donor Lymphocyte Infusion Induces Polyspecific CD8+ T-Cell Responses With Concurrent Molecular Remission in Acute Myeloid Leukemia With NPM1 Mutation. J Clin Oncol (2013) 31(3):e44-7. doi: 10.1200/JCO. 2011.41.1116
- Wermke M, Thiede C, Kiani A, Ehninger G, Bornhäuser M, Platzbecker U. Successful treatment of molecular relapse in NPM1-positive AML using 5azacytidine. *Leukemia* (2010) 24(1):236–7. doi: 10.1038/leu.2009.204
- 139. Craddock C, Labopin M, Robin M, Finke J, Chevallier P, Yakoub-Agha I, et al. Clinical activity of azacitidine in patients who relapse after allogeneic stem cell transplantation for acute myeloid leukemia. *Haematologica* (2016) 101(7):879–83. doi: 10.3324/haematol.2015.140996
- 140. Qin YZ, Wang Y, Xu LP, Zhang XH, Chen H, Han W, et al. The dynamics of RUNX1-RUNX1T1 transcript levels after allogeneic hematopoietic stem cell transplantation predict relapse in patients with t(8;21) acute myeloid leukemia. J Hematol Oncol (2017) 10(1):44-. doi: 10.1186/s13045-017-0414-2
- 141. Wang Y, Wu DP, Liu QF, Qin YZ, Wang JB, Xu LP, et al. In adults with t (8;21)AML, posttransplant RUNX1/RUNX1T1-based MRD monitoring, rather than c-KIT mutations, allows further risk stratification. *Blood* (2014) 124(12):1880–6. doi: 10.1182/blood-2014-03-563403
- 142. Shayegi N, Kramer M, Bornhäuser M, Schaich M, Schetelig J, Platzbecker U, et al. The level of residual disease based on mutant NPM1 is an independent prognostic factor for relapse and survival in AML. *Blood* (2013) 122(1):83–92. doi: 10.1182/blood-2012-10-461749
- 143. Zhou Y, Othus M, Walter RB, Estey EH, Wu D, Wood BL. Deep NPM1 Sequencing Following Allogeneic Hematopoietic Cell Transplantation Improves Risk Assessment in Adults with NPM1-Mutated AML. Biol Blood Marrow Transplant (2018) 24(8):1615–20. doi: 10.1016/j.bbmt. 2018 04 017
- 144. McSweeney PA, Niederwieser D, Shizuru JA, Sandmaier BM, Molina AJ, Maloney DG, et al. Hematopoietic cell transplantation in older patients with hematologic malignancies: replacing high-dose cytotoxic therapy with graft-versus-tumor effects. *Blood* (2001) 97(11):3390–400. doi: 10.1182/blood.V97.11.3390
- 145. Vago L, Perna SK, Zanussi M, Mazzi B, Barlassina C, Stanghellini MTL, et al. Loss of Mismatched HLA in Leukemia after Stem-Cell Transplantation. N Engl J Med (2009) 361(5):478–88. doi: 10.1056/NEJMoa0811036
- 146. Villalobos IB, Takahashi Y, Akatsuka Y, Muramatsu H, Nishio N, Hama A, et al. Relapse of leukemia with loss of mismatched HLA resulting from uniparental disomy after haploidentical hematopoietic stem cell transplantation. *Blood* (2010) 115(15):3158–61. doi: 10.1182/blood-2009-11-254284
- 147. Ahci M, Toffalori C, Bouwmans E, Crivello P, Brambati C, Pultrone C, et al. A new tool for rapid and reliable diagnosis of HLA loss relapses after HSCT. Blood (2017) 130(10):1270–3. doi: 10.1182/blood-2017-05-784306
- 148. Brierley CK, Jones FM, Hanlon K, Peniket AJ, Hatton C, Collins GP, et al. Impact of graft-versus-lymphoma effect on outcomes after reduced intensity conditioned-alemtuzumab allogeneic haematopoietic stem cell transplantation for patients with mature lymphoid malignancies. Br J Haematol (2019) 184(4):547–57. doi: 10.1111/bjh.15685
- Liesveld JL, Rothberg PG. Mixed chimerism in SCT: Conflict or peaceful coexistence? Bone Marrow Transplant (2008) 42:297–310. doi: 10.1038/ bmt.2008.212
- 150. Kinsella FAM, Inman CF, Gudger A, Chan YT, Murray DJ, Zuo J, et al. Very early lineage-specific chimerism after reduced intensity stem cell transplantation is highly predictive of clinical outcome for patients with myeloid disease. *Leuk Res* (2019) 83:106173–. doi: 10.1016/j.leukres. 2019.106173
- 151. Thiede C, Lutterbeck K, Oelschlägel U, Kiehl M, Steudel C, Platzbecker U, et al. Detection of relapse by sequential monitoring of chimerism in circulating CD34+ cells. *Ann Hematol* (2002) 81(Suppl 2):S27–8.
- 152. Bornhäuser M, Oelschlaegel U, Platzbecker U, Bug G, Lutterbeck K, Kiehl MG, et al. Monitoring of donor chimerism in sorted CD34+ peripheral blood cells allows the sensitive detection of imminent relapse after allogeneic stem

- cell transplantation. Haematologica (2009) 94(11):1613–7. doi: 10.3324/haematol.2009.007765
- 153. Rosenow F, Berkemeier A, Krug U, Müller-Tidow C, Gerss J, Silling G, et al. CD34(+) lineage specific donor cell chimerism for the diagnosis and treatment of impending relapse of AML or myelodysplastic syndrome after allo-SCT. Bone Marrow Transplant (2013) 48(8):1070-6. doi: 10.1038/bmt.2013.2
- 154. Hoffmann JC, Stabla K, Burchert A, Volkmann T, Bornhäuser M, Thiede C, et al. Monitoring of acute myeloid leukemia patients after allogeneic stem cell transplantation employing semi-automated CD34+ donor cell chimerism analysis. Ann Hematol (2014) 93(2):279–85. doi: 10.1007/s00277-013-1961-4
- 155. Waterhouse M, Pfeifer D, Duque-Afonso J, Follo M, Duyster J, Depner M, et al. Droplet digital PCR for the simultaneous analysis of minimal residual disease and hematopoietic chimerism after allogeneic cell transplantation. Clin Chem Lab Med (2019) 57(5):641–7. doi: 10.1515/cclm-2018-0827
- 156. Craddock C, Jilani N, Siddique S, Yap C, Khan J, Nagra S, et al. Tolerability and Clinical Activity of Post-Transplantation Azacitidine in Patients Allografted for Acute Myeloid Leukemia Treated on the RICAZA Trial. Biol Blood Marrow Transplant (2016) 22(2):385–90. doi: 10.1016/j.bbmt.2015.09.004
- 157. Platzbecker U, Wermke M, Radke J, Oelschlaegel U, Seltmann F, Kiani A, et al. Azacitidine for treatment of imminent relapse in MDS or AML patients after allogeneic HSCT: Results of the RELAZA trial. *Leukemia* (2012) 26 (3):381–9. doi: 10.1038/leu.2011.234
- 158. Oran B, de Lima M, Garcia-Manero G, Thall PF, Lin R, Popat U, et al. A phase 3 randomized study of 5-azacitidine maintenance vs observation after transplant in high-risk AML and MDS patients. *Blood Adv* (2020) 4 (21):5580–8. doi: 10.1182/bloodadvances.2020002544
- 159. Sockel K, Bornhaeuser M, Mischak-Weissinger E, Trenschel R, Wermke M, Unzicker C, et al. Lenalidomide maintenance after allogeneic HSCT seems to trigger acute graft-versus-host disease in patients with high-risk myelodysplastic syndromes or acute myeloid leukemia and del(5q): results of the LENAMAINT trial. *Haematologica* (2012) 97(9):e34–5. doi: 10.3324/haematol.2012.067629
- 160. Burchert A, Bug G, Fritz LV, Finke J, Stelljes M, Röllig C, et al. Sorafenib Maintenance After Allogeneic Hematopoietic Stem Cell Transplantation for Acute Myeloid Leukemia With FLT3-Internal Tandem Duplication Mutation (SORMAIN). J Clin Oncol (2020) 38(26):2993–3002. doi: 10.1200/JCO.19.03345
- 161. Maziarz RT, Levis M, Patnaik MM, Scott BL, Mohan SR, Deol A, et al. Midostaurin after allogeneic stem cell transplant in patients with FLT3internal tandem duplication-positive acute myeloid leukemia. *Bone Marrow Transplant* (2020). doi: 10.1038/s41409-020-01153-1
- 162. Kent A, Pollyea DA, Winters A, Jordan CT, Smith C, Gutman JA. Venetoclax Is Safe and Tolerable As Post-Transplant Maintenance Therapy for AML Patients at High Risk for Relapse. *Blood* (2020) 136(Supplement 1):11–2. doi: 10.1182/blood-2020-138832
- 163. Kent A, Vasu S, Schatz D, Monson N, Devine S, Smith C, et al. Glasdegib as maintenance therapy for patients with AML and MDS patients at high risk for postallogeneic stem cell transplant relapse. *Blood Adv* (2020) 4(13):3102– 8. doi: 10.1182/bloodadvances.2020001991
- 164. Schmid C, Labopin M, Schaap N, Veelken H, Schleuning M, Stadler M, et al. Prophylactic donor lymphocyte infusion after allogeneic stem cell transplantation in acute leukaemia - a matched pair analysis by the Acute Leukaemia Working Party of EBMT. Br J Haematol (2019) 184(5):782–7. doi: 10.1111/bjh.15691
- 165. Quek L, Ferguson P, Metzner M, Ahmed I, Kennedy A, Garnett C, et al. Mutational analysis of disease relapse in patients allografted for acute myeloid leukemia. *Blood Adv* (2016) 1(3):193–204. doi: 10.1182/ bloodadvances.2016000760
- 166. Jan M, Leventhal MJ, Morgan EA, Wengrod JC, Nag A, Drinan SD, et al. Recurrent genetic HLA loss in AML relapsed after matched unrelated allogeneic hematopoietic cell transplantation. Blood Adv (2019) 3 (14):2199–204. doi: 10.1182/bloodadvances.2019000445
- 167. Goodyear O, Dennis M, Jilani N, Loke J, Siddique S, Ryan G, et al. Azacitidine augments expansion of regulatory T cells after allogeneic stem cell transplantation in patients with acute myeloid leukemia (AML). Blood (2012) 119:3361–9. doi: 10.1182/blood-2011-09-377044

- 168. Clinical Trials.gov. National Library of Medicine (U.S.) Randomised Study of Oral Azacitidine vs Placebo Maintenance in AML or MDS Patients After Allo-SCT Identifier NCT04173533. Available at: https://clinicaltrials.gov/ ct2/show/NCT04173533 (Retrieved April 1, 2021).
- 169. ClinicalTrials.gov. National Library of Medicine (U.S.) Panobinostat Maintenance After HSCT for High-risk AML and MDS. Identifier: NCT04326764. Available from: https://clinicaltrials.gov/ct2/show/ NCT04326764 (Retrieved April 1, 2021).
- 170. Craddock C, Slade D, De Santo C, Wheat R, Ferguson P, Hodgkinson A, et al. Combination Lenalidomide and Azacitidine: A Novel Salvage Therapy in Patients Who Relapse After Allogeneic Stem-Cell Transplantation for Acute Myeloid Leukemia. *J Clin Oncol* (2019) 37(7):580-8. doi: 10.1200/JCO 18.00889
- 171. IMPACT Partnership (Internet). PRO-DLI: A Phase II Prospective Trial of Prophylactic Donor Lymphocyte Infusions for the Prevention of Relapse post HSCT in patients with High Risk Myeloid Malignancy. Available from: https://www.impactpartnership.org.uk/the-trials/pro-dli/ (updated November 12 2020; cited April 1 2020).
- 172. Greiner J, Götz M, Bunjes D, Hofmann S, Wais V. Immunological and Clinical Impact of Manipulated and Unmanipulated DLI after Allogeneic Stem Cell Transplantation of AML Patients. J Clin Med (2019) 9(1). doi: 10.3390/jcm9010039
- 173. Schlenk RF, Weber D, Fiedler W, Salih HR, Wulf G, Salwender H, et al. Midostaurin added to chemotherapy and continued single-agent maintenance therapy in acute myeloid leukemia with FLT3-ITD. *Blood* (2019) 133(8):840–51. doi: 10.1182/blood-2018-08-869453
- 174. ClinicalTrials.gov. National Library of Medicine (U.S.). A Trial of the FMS-like Tyrosine Kinase 3 (FLT3) Inhibitor Gilteritinib Administered as Maintenance Therapy Following Allogeneic Transplant for Patients With FLT3/Internal Tandem Duplication (ITD) Acute Myeloid Leukemia (AML). Identifier NCT02997202. Available from: https://clinicaltrials.gov/ct2/show/ NCT02997202 (Retrieved April 1, 2021).
- 175. ClinicalTrials.gov. National Library of Medicine (U.S.). Venetoclax and Azacitidine for the Treatment of Acute Myeloid Leukemia in the Post-Transplant Setting. Identifier NCT04128501. Available from: https:// clinicaltrials.gov/ct2/show/NCT04128501 (Retrieved April 1, 2021).
- 176. ClinicalTrials.gov. National Library of Medicine (U.S.). (2000, February 29 -). A Study Evaluating Safety and Efficacy of Venetoclax in Combination With Azacitidine Versus Standard of Care After Allogeneic Stem Cell Transplantation (SCT) in Participants With Acute Myeloid Leukemia (AML) (VIALE-T). Identifier NCT04161885. Retrieved April 1, 2021 from: https://clinicaltrials.gov/ct2/show/NCT04161885.
- 177. Roboz GJ, DiNardo CD, Stein EM, de Botton S, Mims AS, Prince GT, et al. Ivosidenib induces deep durable remissions in patients with newly diagnosed IDH1-mutant acute myeloid leukemia. *Blood* (2020) 135(7):463–71. doi: 10.1182/blood.2019002140
- 178. DiNardo CD, Stein EM, de Botton S, Roboz GJ, Altman JK, Mims AS, et al. Durable Remissions with Ivosidenib in IDH1-Mutated Relapsed or Refractory AML. N Engl J Med (2018) 378(25):2386–98. doi: 10.1056/ NEJMoa1716984
- 179. Stein EM, DiNardo CD, Pollyea DA, Fathi AT, Roboz GJ, Altman JK, et al. Enasidenib in mutant IDH2 relapsed or refractory acute myeloid leukemia. Blood (2017) 130(6):722–31. doi: 10.1182/blood-2017-04-779405
- 180. ClinicalTrials.gov. National Library of Medicine (U.S.). Enasidenib as Maintenance Therapy in Treating Patients With Acute Myeloid Leukemia With IDH2 Mutation After Donor Stem Cell Transplant. Identifier NCT03728335. Available from: https://clinicaltrials.gov/ct2/show/ NCT03728335 (Retrieved April 1, 2021).
- 181. ClinicalTrials.gov. National Library of Medicine (U.S.). IDH1 Inhibition Using Ivosidenib as Maintenance Therapy for IDH1-mutant Myeloid Neoplasms Following Allogeneic Stem Cell Transplantation. Identifier NCT03564821. Available from: https://clinicaltrials.gov/ct2/show/NCT03564821 (Retrieved April 1 2021).

Conflict of Interest: CC has received honoraria from Celgene, Daichi-Sankyo, Novartis and Pfizer as well as research funding from Celgene. JL has received travel funding from Novartis and Daichi-Sankyo, honoraria from Pfizer, Janssen and Amgen.

The remaining author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The reviewer CS has declared past co-authorships with one of the authors CC, to the handling editor, at the time of review.

Copyright © 2021 Loke, Buka and Craddock. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.





Allogeneic Hemopoietic Stem Cell Transplantation for Myelofibrosis: 2021

Andrea Bacigalupo ^{1,2*}, Idanna Innocenti ¹, Elena Rossi ^{1,2}, Federica Sora ^{1,2}, Eugenio Galli ¹, Francesco Autore ¹, Elisabetta Metafuni ¹, Patrizia Chiusolo ^{1,2}, Sabrina Giammarco ¹, Luca Laurenti ^{1,2}, Giulia Benintende ², Simona Sica ^{1,2} and Valerio De Stefano ^{1,2}

¹ Dipartimento di Diagnostica per Immagini, Radioterapia Oncologica ed Ematologia, Fondazione Policlinico Universitario A. Gemelli IRCCS, Roma, Italy, ² Sezione di Ematologia, Dipartimento di Scienze Radiologiche ed Ematologiche, Università Cattolica del Sacro Cuore, Roma, Italy

OPEN ACCESS

Edited by:

Hermann Einsele, Julius Maximilian University of Würzburg, Germany

Reviewed by:

Federico Simonetta, Geneva University Hospitals (HUG), Switzerland Juergen Finke, University of Freiburg Medical Center, Germany

*Correspondence:

Andrea Bacigalupo andrea.bacigalupo@unicatt.it

Specialty section:

This article was submitted to Alloimmunity and Transplantation, a section of the journal Frontiers in Immunology

Received: 03 December 2020 Accepted: 13 April 2021 Published: 04 May 2021

Citation:

Bacigalupo A, Innocenti I, Rossi E, Sora F, Galli E, Autore F, Metafuni E, Chiusolo P, Giammarco S, Laurenti L, Benintende G, Sica S and De Stefano V (2021) Allogeneic Hemopoietic Stem Cell Transplantation for Myelofibrosis: 2021. Front. Immunol. 12:637512. doi: 10.3389/fimmu.2021.637512 The aim of this review is to update the current status of allogeneic hemopoietic stem cell transplants (HSCT) for patients with myelofibrosis (MF). We have first summarized the issue of an indication for allogeneic HSCT, discussing several prognostic scoring systems, developed to predict the outcome of MF, and therefore to identify patients who will benefit of an allogeneic HSCT. Patients with low risk MF are usually not selected for a transplant, whereas patients with intermediate or high risk MF are eligible. A separate issue, is how to predict the outcome of HSCT: we will outline a clinical molecular myelofibrosis transplant scoring system (MTSS), which predicts overall survival, ranging from 90% for low risk patients, to 20% for very high risk patients. We will also discuss transfusion burden and spleen size, as predictors of transplant outcome. The choice of a transplant platform including the conditioning regimen, the stem cell source and GvHD prophylaxis, are crucial for a successful program in MF, and will be outlined. Complications such as poor graft function, graft failure, GvHD and relapse of the disease, will also be reviewed. Finally we discuss monitoring the disease after HSCT with donor chimerism, driver mutations and hematologic data. We have made an effort to make this review as comprehensive and up to date as possible, and we hope it will provide some useful data for the clinicians.

Keywords: myelofibrosis, allogeneic transplantation, busulfan, thiotepa, fludarabine chimerism, splenectomy

INDICATIONS FOR HSCT

In the era of JAK inhibitors, allogeneic hematopoietic stem cell transplantation (HSCT) remains the only curative treatment for patients with Myelofibrosis (MF) (1). The American Society for Transplantation and Cellular Therapy (ASTCT) considers an allogeneic HSCT "standard of care with clinical evidence" for patients with intermediate and high risk disease (2). In order to classify patients as intermediate or high risk several models have been developed. **Table 1** outlines some of the most commonly used scoring systems and the variables they are based on: IPSS (3), DIPSS (4), DIPSS-plus (5) and MIPSS70 (6). The first two are based exclusively on clinical data, the third incorporates cytogenetics and the fourth includes mutational analysis. Survival of patients with MF

TABLE 1 | Prognostic scoring systems for patients with myelofibrosis.

IPSS [3]	DIPSS [4]	DIPSS-plus [5]	MIPSS70 [6]	
 ✓ Age > 65 years (1 point) ✓ Constitutional symptoms (1 point) ✓ Hemoglobin < 10 g/dl (1point) ✓ WBC count > 25 × 109/l (1 point) Circulating blasts ≥ 1% (1 point) 	 ✓ Age > 65 years (1 point) ✓ Constitutional symptoms (1 point) ✓ Hemoglobin < 10 g/dl (2points) ✓ WBC count > 25 x 109/l (1 point) Circulating blasts ≥ 1% (1 point) 	 ✓ RBC transfusion (1 point) ✓ PLT count < 100 × 109/l (1 point) Unfavorable karyotype ^a (1 point) 	Genetic variables: ✓ One HMR mutation (1 point) ✓ ≥2 HMR mutations (2 points) ✓ Type1/like CALR absent (1 point) Clinical variables: ✓ Hemoglobin <10g/dl (1 point) ✓ Leukocytes >25×109/l (2 points) ✓ Platelets <100×109/l (2 points) ✓ Circulating blasts ≥2% (1 point) ✓ Constitutional symptoms (1 point) ✓ Bone marrow fibrosis grade ≥2 (1 point)	✓
 Low risk: 0 points (11.3 yrs) Intermediate-1 risk: 1 point (7.9 yrs) Intermediate-2 risk: 2 points (4 yrs) High risk: ≥ 3 points (2.3 yrs) 	 Low risk: 0 point (n.r.) Intermediate-1 risk: 1–2 point (14.2 yrs) Intermediate-2 risk: 3–4 points (4 yrs) High risk: 5-6 points (1.5 yrs) 	 Low risk: 0 point (15.4 yrs) Intermediate-1 risk: 1 point (6.5 yrs) Intermediate-2 risk: 2-3 points (2.9 yrs) High risk: 4-6 points (1.3 yrs) 	 Low risk: 0-1 points (n.r.) Intermediate risk: 2-4 points (6.3 yrs) High risk: ≥ 5 points (3.1 yrs) 	•

IPSS, international prognostic scoring system; DIPSS, dynamic international prognostic scoing system; MIPSS70, mutation-enhanced international prognostic scoring system; HMR, high molecular risk (see text).

can be predicted using one of those models, and thus eligibility for a transplant procedure. However eligibility must also include transplant related variables, such as patients age up to 70-75 years, a good performance status, low transfusion burden, absence of a massive splenomegaly and portal hypertension and donor type. Older patients also tend to have one or more comorbidities which may increase the risk of transplant related mortality (TRM) or even preclude a transplant approach. A Panel of experts recommends considering allogeneic HSCT for patients with IPSS/DIPSS/DIPSS plus high or intermediate-2 risk (7) (Figure 1). The Panel also recommends considering an allogeneic HSCT for transplant-eligible patients with IPSS/ DIPSS/DIPSS-Plus intermediate-1 risk score, who present with either refractory, transfusion-dependent anemia, a percentage of blasts in peripheral blood > 2% in at least two repeated manual measurements, adverse cytogenetics, or high-risk mutations, such as such as ASXL1, EZH2, IDH1/IDH2, SRSF2 (7)(Figure 1). In this situation, the transplant procedure should be performed in a controlled setting (registries, clinical trial) (7).

More recently a mutation-based prognostic model has been proposed to identify candidates for HSCT among low or intermediate-1 risk DIPSS, who are expected to have similar overall survival as patients with a high risk DIPSS (8). Patients who are triple negative (JAK2/CALR/MPL) or CALR wild type and ASXL1 mutated, irrespective of DIPSS risk scores, should be considered for HSCT (8). A combination of mutation-based prognosis together with clinical data has been compiled in a recent scoring system (9).

In conclusion, we are now able to identify MF patients with a different median survival: there is consensus on the eligibility to transplant for DIPSS intermediate2/high risk patients. The presence of high risk mutations in DIPSS intermediate1/low risk patients may also suggest eligibility for a transplant procedure. The clinical conditions of the patient, the degree of

HLA matching of potential donors and the patient's choice must be considered in the final decision to transplant or not.

HOW TO DEAL WITH SPLENOMEGALY

Splenomegaly is a common feature in patients with advanced myelofibrosis (MF) and it is a sign of extramedullary hematopoiesis (also known as myeloid metaplasia) (10). Patients may be severely symptomatic with abdominal pain, early satiety, weight loss, cytopenia, portal hypertension, and splenic infarction (10).

Splenectomy is effective in relieving symptoms, but is associated with a number of complications, as well as significant morbidity and mortality.

Peri-operative mortality is in the range of 5%-10%. The most common complications are infections, thrombosis and bleeding, occurring in up to 30% of patients (11). Patients with thrombocytopenia seemed to have an increased probability of post-splenectomy blast transformation, although this did not result in shortened survival. Leukemic transformation is more probably related to natural progression of the disease in advanced stage and to post-splenectomy redistribution of circulating blasts, not to true clonal evolution (12, 13). Hemopoietic stem cell transplantation (HSCT) offers the potential of cure for patients with intermediate or high risk myelofibrosis (14). Splenomegaly, characteristic of those patients, may lead to sequestration of transplanted stem cells and delayed hematologic recovery (3, 15) thus affecting the transplant outcome. Surgical removal of the spleen may be effective in reducing the time for neutrophil and platelet recovery (16) but its impact on relapse rate and survival is unclear (17, 18), calling for a prospective randomized trial. Pre-transplant splenectomy in MF patients was associated with

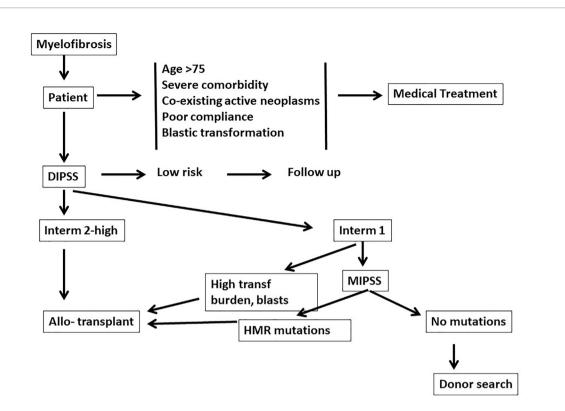


FIGURE 1 | Eligibility for a transplant procedure in patients with myelofibrosis: medical treatment should be offered for older patients (>75 years) and/or patients with comorbidities. Dynamic international prognostic scoring system (DIPSS) will then identify patients low risk patients, who should be followed. DIPSS-intermediate 2 and high risk patients are who are strong candidates for an allogeneic transplant. DIPSS-intermediate 1 patients with a high transfusion burden and blasts counts are also strong candidates for an allogeneic transplant. Patients may also be studied with a molecular international prognostic scoring system (MIPSS), and may be eligible for transplantation if high risk mutations (HMR) (see text) are identified.

a prolonged overall and event-free survival in a recently published study (19).

The advent of **Janus kinase** (**JAK**) 1/2 **inhibitors**, which decrease splenomegaly and alleviate MF-related symptoms, has had, as compared to old cytoreductive drugs, a major impact on the management of splenomegaly, removing some indications for splenectomy. However, in a proportion of patients, the splenic response is then lost. Many MF patients who proceed to allogeneic HSCT, are currently treated with JAK inhibitors, usually ruxolitinib: this should be tapered down over a 10- to 14-day period and should be discontinued just before the conditioning regimen (20). In one study, ruxolitinib was continued also during transplant in the attempt of preventing GvHD (21).

Splenic irradiation (SI) may also be used to reduce the spleen size and related symptoms; there are only few small studies on SI prior to transplant in MF patients (22, 23). It was demonstrated that SI alleviates splenic discomfort and reduces spleen size in a majority of MF patients, with a median duration of response of 6 months (24). Limitations of SI include prolonged pancytopenia with infectious complications. Comparable engraftment rate has been shown in patients receiving or not SI (25) as well as comparable acute and chronic GVHD incidence, post-transplant infectious complications and survival. The role of SI in leukemic transformation (LT) remains unclear and speculative. Radiotherapy may be indicated in patients who are

not eligible for surgery or in patients who have lost their response to JAK2 inhibitors (26–28).

PREDICTING THE OUTCOME OF HSCT

Disease based risk score. Survival of MF patients receiving medical treatment, with the exclusion of allogeneic HSCT, can be predicted by several scoring systems, reviewed in Indications for HSCT (3-7). Some studies have assessed whether these scoring systems can predict the outcome of patients after an allogeneic HSCT. DIPSS can predict post transplant survival (29), and the same has been shown for DIPSS-plus (30). In multivariate analysis, the DIPPS-plus score predicted survival, disease free survival (DFS) and TRM, together with conditioning regimen, comorbidity index (HCT-CI), patients' age and donor type (30). In 2019, a cohort of 159 patients with secondary myelofibrosis who underwent allogeneic HSCT was analyzed retrospectively to compare the predictive value of DIPSS and MYSEC (31). The four risk groups of DIPSS did not predict survival after allogeneic HSCT, whereas MYSEC maintained its predictive role also in the post-transplant setting.

Transplant based risk score (TS). Few scoring systems have been designed exclusively for allogeneic HSCT. In 2010, a study identified spleen size, transfusion history and donor type as

predictive of outcome: survival was 79% for low risk patients and 8% for high risk patients (18). In 2012 a predictive risk model including JAKV617F status, age and constitutional symptoms was proposed in the setting of 150 transplanted patients and resulted to be predictive for 5 years overall survival (OS) (32).

More recently, a scoring system has been devised, which incorporates HLA matching between donor and recipient, mutational analysis, and clinical data, at time of transplantation (MTSS), in patient with primary and secondary MF (9). This index is predictive of non-relapse mortality. In the last year we have revisited our transplant score (TS) including maximum spleen size and red blood cell transfusion burden before HSCT (18): the 5 year disease free survival (DFS) was 74% vs 36% (p=0.0001) for patients with low or high TS.

In conclusion, scoring systems designed to predict transplant outcome are available and can be used when counseling patients eligible for transplant procedures.

DONOR TYPE, STEM CELL SOURCE AND GVHD PROPHYLAXIS

Donor type is an important predictor of outcome in myelofibrosis: a study from the Center for International Blood and Marrow Transplant Research (CIBMTR) on 233 transplants for myelofibrosis (33), showed that donor type was an independent risk factor for TRM, with a relative risk of death of 3.92 for matched unrelated donor (MUD) and 9.37 for mismatched unrelated donor (MMUD), when compared to matched related donor (MRD) (33). The 5 year overall survival was 56% for MRD, 48% for MUD and 34% for MMUD. The main causes of death were GvHD, infections and organ failure, in particular among MMUD grafts (33). Similar results are reported in other studies (17, 34–37). On the other hand, contrasting data exist regarding GvHD and donor type. Some studies show no significant difference among different donor types (17, 35, 38), whereas the CIBMTR shows a higher risk of GvHD for patients receiving MUD (RR 1.98) and MMUD (RR 1.52) as compared to MRD (33). Engraftment is reported to be comparable according to donor type (33, 35, 38), whereas significant differences have been described according to the stem cell source, with faster recovery with peripheral blood grafts (37, 38). Unrelated cord blood (UCB) transplants have been rarely used in myelofibrosis, and are associated with delayed engraftment and a high TRM, probably due to the significant risk for graft failure and infectious complications (39).

The addition of ATG to conventional GvHD prophylaxis, based on calcineurin inhibitor alone or combined to methotrexate or mycophenolic acid, reduces the incidence of GvHD, as one would expect (40). However, modified regimens of GvHD prophylaxis, including the use of post-transplant cyclophosphamide (PT-CY) have reduced post-transplant complications in alternative donor grafts, especially for HLA haplo-identical donor (36). A combination of calcineurin inhibitor with ATG and PTCy after reduced intensity conditioning may further reduce the risk of GvHD, improving TRM and survival, without an increased risk of relapse (41). Very

recently, an interesting pilot study was conducted by Morozova and colleagues: GvHD prophylaxis with PTCY and ruxolitinib showed promising results in terms of GvHD control in a small cohort of patients with acceptable TRM (42).

In summary an HLA matched donor is the best option for myelofibrosis, in order to achieve optimal outcome: alternative donor grafts may be explored using modified regimens of GvHD prophylaxis.

CONDITIONING REGIMENS AND RUXOLITINIB

Conditioning regimens in myelofibrosis were historically myeloablative (MAC), predominantly busulfan plus cyclophosphamide and total body irradiation with or without cyclophosphamide (15), but transplant related mortality (TRM) and GvHD rates were high, especially in older individuals (43).

Reduced intensity conditioning (RIC) has been increasingly used in MF, in consideration of the older age of MF patients. The first prospective EBMT multicenter phase II trial of RIC SCT consisted of busulfan (10 mg/kg) orally (or equivalent IV dose) plus fludarabine (180 mg/m²) and in vivo T-cell depletion with anti-thymocyte globulin at a dose of 3 x 10 mg/kg (for related transplantation) or 3 x 20 mg/kg (for unrelated donor transplantation): this protocol resulted in low rates of primary graft failure and rapid hematologic recovery (17). Fludarabine 90 mg/m^2, combined with melphalan 140 mg/m^2 (FLU-MEL) is an alternative RIC regimen, and has been compared in a retrospective study with the BU-FLU regimen (44). Although the FLU-MEL was associated with increased early toxicity, the long-term outcome (OS and disease-free survival) was similar in the two groups. In both regimens the use of a HLA mismatched unrelated donor was associated with worse outcome, in terms of TRM, OS and progression-free survival. A randomized study comparing fludarabine in combination with busulfan 10 mg/kg i.v. or thiotepa 12 mg/kg, failed to identify significant differences in terms of clinical outcome (45): both regimens were associated with a significant degree of mixed chimerism.

In a retrospective comparisons of RIC versus MAC regimens for myelofibrosis, the latter do not appear to protect patients from relapse (46), neither there are differences within day +100 transplant-related mortality (47). A large retrospective analysis of the EBMT in 2224 patients with myelofibrosis, compared MAC regimens (781 patients) with RIC regimens (1443 patients) (48): there was no statistically significant difference in engraftment, GvHD, TRM and overall survival; there was a trend toward a higher relapse rate with RIC.

We have recently shown that a conditioning regimen including two alkylating agents (in our case busulfan and thiotepa) with fludarabine, significantly reduced the risk of relapse when compared to regimen with one alkylating agent (either busulfan or thiotepa or melphalan) in combination with fludarabine (36). Therefore, the choice of the conditioning regimen, may play a significant role in determining the control of the disease after an allogeneic HSCT.

The efficacy of the JAK1/JAK2 inhibitor ruxolitinib in reducing spleen size and systemic symptoms, in myelofibrosis, has been established (49, 50). Currently, most patients undergoing an allogeneic HSCT have been treated with this agent with the aim of reducing splenomegaly, improving the performance status and shorten time to engraftment. A phase II trial demonstrated the feasibility of ruxolitinib therapy followed by a RIC regimen for patients with myelofibrosis (51). Appropriate tapering should be scheduled (52), although recently peri-transplant ruxolitinib has been reported (42). There is no evidence, however, that the administration of ruxolitinib pre-transplant reduces the incidence of relapse after transplant.

MONITORING DISEASE CONTROL (DONOR CHIMERISM AND MUTATIONS)

Patients with myelofibrosis may have one of three driver mutations (JAK2, CALR and MPL), or lack all three (triple negative patients). Ditschkowski et al. (53) showed that survival after transplantation was not significantly different for JAK2+ (75%) versus JAK2 negative (71%) patients. More recent retrospective studies have suggested a survival advantage for CALR mutation (54, 55). A large retrospective study has investigated the role of extensive mutational profiling with a targeted 16-gene panel, and has confirmed the favorable role of a CALR mutation (56). In the same study IDH2 and ASXL1 mutations confirmed their adverse prognostic role after allogenic HSCT, whereas a triple negative status (JAK2, MPL, CALR) did not appear to modify the outcome after transplant.

Minimal residual disease (MRD) should be used to identify patients achieving a complete remission after HSCT, as well as an early evidence of relapse. Alchalby et al. has shown that JAK2 negativity after allogeneic HSCT significantly reduces the risk of relapse (57). Similar results have been obtained with MPL and CALR mutations as MRD markers (58). A recent retrospective single-center study (59) has shown that that patients with detectable mutations on day +100 or at day +180 after allogeneic HSCT have a significant higher risk of clinical relapse at 5 years, as compared to molecular-negative patients (62% vs 10%, P<0.001 and 70% vs 10%, P<0.001, respectively): single different mutations have comparable predictive value on relapse.

However, 10% to 15% of patients are triple negative and cannot be followed after transplantation with a molecular marker: in these patients chimerism studies can be helpful to identify early signs of relapse. We have recently described 120 patients with chimerism data on day +30 (60), showing that early full donor chimerism is highly predictive of long-term disease control. The cumulative incidence of relapse at 5 years, was 14% vs 40% for patients with or without full donor chimerism (40). We found that a conditioning regimen including two alkylating agents (busulfan and thiotepa) induces a significantly higher rate of complete donor chimerism on day +30, as compared to patients prepared with one alkylating agent (either busulfan, melphalan or thiotepa) (87% vs 45%, p<0.0001).

MRD positive patients or patients with declining donor chimerism, who still are receiving immunosuppressive therapy, may discontinue immunosuppressive drugs and/or receive donor lymphocyte infusions (DLI), in order to achieve again full donor chimerism.

PRIMARY GRAFT FAILURE (PRGF) AND POOR GRAFT FUNCTION (PGF)

Lack of engraftment of donor stem cells is referred to as primary graft failure (PrGF), and is characterized by neutropenia, combined with mixed or no donor chimerism on bone marrow and/or peripheral blood cells (61). PrGF should be distinguished from poor graft function, or cytopenia with full donor chimerism (62). The latter suggests inappropriate function of engrafted donor stem cells and can be treated with the infusion of selected CD34+ cells from the same donor, without a preparative regimen (62). Predictive factors have not been determined, but several conditions have been associated with unsuccessful engraftment, such as the intensity of the conditioning regimen, donor type, stem cells source, number of CD34+ cells infused, GvHD prophylaxis, degree of fibrosis, degree of splenomegaly, pre-transplant thrombocytopenia (63).

The incidence of PrGF ranges from 2 to 24%. A lower rate was reported in a large prospective study from EBMT (48), with only in 2 out 103 patients with PrGF. However, 11% of patients experienced poor graft function and required an additional stem cell boost. In a subsequent pilot study, PrGF was not influenced by the intensity of conditioning regimen (64) and no other predictors were found in other studies (17, 53). Donor type appears to influence the incidence of PrGF, which is lower in patients transplanted from HLA identical donors, as compared to transplants from family mismatched and unrelated donors (65–67). Contrasting data are reported on other factors: splenectomy before HSCT, peripheral stem cell use as source of stem cells and the absence of pre-transplant thrombocytopenia have been suggested to promote engraftment in some studied (18, 66, 68), but not in other studies (65).

Patients with full donor engraftment, may still have transfusion dependent low blood counts for variable periods of time, and this is referred to as Poor graft function (PGF). In a large retrospective analysis, the proportion of patients with less than 20x10^9/l platelets between day +50 and +100 after an allogeneic HSCT, is 10% and has not changed in the time period before 2000, 2001-2010 and beyond 2010 (unpublished). A diagnosis of myelofibrosis is a negative predictor for hematologic recovery: a low platelet count is seen in 18% vs 8% of patients with or without a diagnosis of MF (unpublished). For this reason, when looking at patients receiving a top up of CD34 selected cells for PGF, the proportion of patients with MF (26%) is higher than the proportion of MF in the transplant indications (7%) (62). These patients may remain transfusion dependent for long periods of time, and may be treated either with an infusion of CD34 selected cells from the same donor, or, more recently with high dose eltrombopag. Time to trilineage

recovery is however delayed with these approaches and longlasting supportive care must be planned.

TREATMENT OF MF RELAPSE AFTER ALLOGENEIC TRANSPLANT

Allogeneic hematopoietic stem cell transplant remains the only curative treatment for myelofibrosis (MF). A retrospective EBMT study on 1055 patients with MF transplanted between 1995 and 2014, alive and free of their disease at two years after HSCT showed that the most common cause of death (41-61%) was relapse of MF, for all time periods (2-5years, 5-10 years) (40). There is no standardized re-treatment of relapse after allogeneic transplant. Based on limited available literature, ruxolitinib, donor leukocytes infusion (DLI), and a second allogenic HSCT are three options for relapsing MF patients; obviously, the choice depends on patients age, fitness status, molecular or hematologic relapse, and the presence of GVHD.

The use of DLI and second transplant as salvage treatment for relapsed MF after allogeneic HSCT was reported in a retrospective study some years ago (69). Out of 26 relapsed patients, 39% achieved a stable response to dose-escalated DLIs. Seventeen patients, thirteen of which non-responders to DLI, underwent a second allogeneic HSCT, achieving an ORR of 80% (9 CR and 3 PR); incidence of relapse at 1-year was 24%. The 2-year overall survival and progression-free survival were 70% and 67%, respectively.

The most consistent data derive from a recent EBMT real-life retrospective study focusing on the treatment of 251/1371 (18%) MF patients, who relapsed after an allogeneic HSCT (70). DLIs were used in 23% of patients, whereas 20% underwent DLI combined with chemotherapy and 11% had chemotherapy alone. Fifty-one patients (25%) underwent second allogeneic HSCT alone and 26 (13%) underwent DLI and a second allogeneic HSCT. The median OS from the time of relapse for patients receiving DLI alone, DLI followed by a second allogeneic HSCT or second allogeneic HSCT were 76 months, 54 months, and 27 months respectively.

Recently Chabra et al. published a small number of MF patients, mostly treated with ruxolitinb pre-transplant (71): after a median follow up of >3 years, two patients out of 37 had relapsed after HSCT (5.4%), but the study lacked a strong control group of untreated ruxolitinib patients. Indeed other recently published data in the ruxolitinib era (72), have shown no improvement in survival nor in the incidence of relapse for MF. The use of ruxolitinib after allogenic HSCT is primarily attributable to the treatment of GVHD, and only in few cases for the treatment of the relapse, mostly in combination with DLIs. One study has reported peritransplant use of ruxolitinib (21).

In conclusion, although based on a small number of studies, the best therapeutic strategy for MF patients relapsing after an

REFERENCES

 Tefferi A, Partain DK, Palmer JM, Slack JL, Roy V, Hogan WJ, et al. Allogeneic hematopoietic stem cell transplant overcomes the adverse survival effect of very high risk and unfavorable karyotype allogeneic HSCT, seems to be dose -escalated DLI, or otherwise, for non-responders, a second allogeneic HSCT. The question remains whether DLI should be infused after a lympho-depleting treatment, as currently is being done for CAR-T cells.

DESIGNING A TRANSPLANT STRATEGY FOR MYELOFIBROSIS

Patients with myelofibrosis need to be discussed to identify eligibility for transplant procedures (**Figure 1**). Patients over the age of 75 years, with severe comorbidities, coexisting active neoplasms, or poor compliance, should be addressed by medical treatment. Patients less than 75 years of age and fit, should be assessed for risk factors (DIPSS or other scoring systems): low risk patients should be followed regularly. DIPSS intermediate 2 or high risk patients are eligible for a transplant procedure (**Figure 1**). DIPSS int 1 patients should be studied with next generation sequencing (NGS): if no additional adverse mutations are found (ASXL1, EZH2, SRSF2, IDH1/2) then the search for a donor can be initiated, but the transplant may be postponed. If, on the contrary, additional adverse mutations are identified the donor search may be initiated and the transplant also programmed.

Once a transplant is programmed several facts need to be considered: in addition to patient factors such as age, comorbidities and disease phase (DIPSS), other facts need to be taken in to account, including transplant variables (donor type, stem cell source, conditioning regimen, GvHD prophylaxis), the psychological status of the patient, the presence of care givers, especially for the post-transplant discharge and logistics (transplant centers may be located at a distance from the patients' home). The combination of all these factors will then lead to a tailored strategy in terms of optimal timing and choice of a transplant platform.

AUTHOR CONTRIBUTIONS

AB, SS and VS designed the study and overviewed the manuscript. Sections and authors: indications (II), splenectomy (ER), predicting outcome (FS, EG), monitoring disease (PC), Graft Failure (SG), Relapse (LL), Conditioning regimens (FA), donor type (EM), reviewed MS (GB). All authors contributed to the article and approved the submitted version.

FUNDING

This study was partly funded by AIRC, Associazione Italiana Ricerca contro il Cancro; grant to AB.

- in myelofibrosis. *Am J Hematol* (2018) 93(5):649–54. doi: 10.1002/ajh.25053
- 2. Kanate AS, Majhail NS, Savani BN, Bredeson C, Champlin RE, Crawford S, et al. Indications for Hematopoietic Cell Transplantation and Immune Effector Cell Therapy: Guidelines from the American Society for

Transplantation and Cellular Therapy. Biol Blood Marrow Transplant (2020) 26(7):1247–56. doi: 10.1016/j.bbmt.2020.03.002

- Cervantes F, Dupriez B, Pereira A, Passamonti F, Reilly JT, Morra E, et al. New prognostic scoring system for primary myelofibrosis based on a study of the International Working Group for Myelofibrosis Research and Treatment. Blood (2009) 113(13):2895–901. doi: 10.1182/blood-2008-07-170449
- 4. Passamonti F, Cervantes F, Vannucchi AM, Morra E, Rumi E, Pereira A, et al. A dynamic prognostic model to predict survival in primary myelofibrosis: a study by the IWG-MRT (International Working Group for Myeloproliferative Neoplasms Research and Treatment). Blood (2010) 115 (9):1703–8. doi: 10.1182/blood-2009-09-245837
- Gangat N, Caramazza D, Vaidya R, George G, Begna K, Schwager S, et al. DIPSS plus: a refined Dynamic International Prognostic Scoring System for primary myelofibrosis that incorporates prognostic information from karyotype, platelet count, and transfusion status. *J Clin Oncol* (2011) 29 (4):392–7. doi: 10.1200/JCO.2010.32.2446
- Guglielmelli P, Lasho TL, Rotunno G, Mudireddy M, Mannarelli C, Nicolosi M, et al. MIPSS70: Mutation-Enhanced International Prognostic Score System for Transplantation-Age Patients With Primary Myelofibrosis. J Clin Oncol (2018) 36(4):310–8. doi: 10.1200/JCO.2017.76.4886
- Barbui T, Tefferi A, Vannucchi AM, Passamonti F, Silver RT, Hoffman R, et al. Philadelphia chromosome-negative classical myeloproliferative neoplasms: revised management recommendations from European LeukemiaNet. Leukemia (2018) 32(5):1057–69. doi: 10.1038/s41375-018-0077-1
- Tefferi A, Nicolosi M, Mudireddy M, Szuber N, Finke CM, Lasho TL, et al. Driver mutations and prognosis in primary myelofibrosis: Mayo-Careggi MPN alliance study of 1,095 patients. Am J Hematol (2018) 93(3):348–55. doi: 10.1002/ajh.24978
- Gagelmann N, Ditschkowski M, Bogdanov R, Bredin S, Robin M, Cassinat B, et al. Comprehensive clinical-molecular transplant scoring system for myelofibrosis undergoing stem cell transplantation. *Blood* (2019) 133 (20):2233–42. doi: 10.1182/blood-2018-12-890889
- Santos FP, Tam CS, Kantarjian H, Cortes J, Thomas D, Pollock R, et al. Splenectomy in patients with myeloproliferative neoplasms: efficacy, complications and impact on survival and transformation. *Leuk Lymphoma* (2014) 55(1):121-7. doi: 10.3109/10428194.2013.794269
- Cervantes F. How I treat splenomegaly in myelofibrosis. Blood Cancer J (2011) 1(10):e37. doi: 10.1038/bcj.2011.36
- Tefferi A, Mesa RA, Nagorney DM, Schroeder G, Silverstein MN. Splenectomy in myelofibrosis with myeloid metaplasia: a single-institution experience with 223 patients. *Blood* (2000) 95(7):2226–33. doi: 10.1182/ blood.V95.7.2226.007k19_2226_2233
- Tefferi A, Mudireddy M, Gangat N, Hanson CA, Ketterling RP, Pardanani A, et al. Risk factors and a prognostic model for postsplenectomy survival in myelofibrosis. Am J Hematol (2017) 92(11):1187–92. doi: 10.1002/ aih.24881
- 14. Guardiola P, Anderson JE, Bandini G, Cervantes F, Runde V, Arcese W, et al. Allogeneic stem cell transplantation for agnogenic myeloid metaplasia: a European Group for Blood and Marrow Transplantation, Société Française de Greffe de Moelle, Gruppo Italiano per il Trapianto del Midollo Osseo, and Fred Hutchinson Cancer Research Center Collaborative Study. Blood (1999) 93(9):2831–8. doi: 10.1182/blood.V93.9.2831
- Martino R, Altés A, Muñiz-Díaz E, Brunet S, Sureda A, Domingo-Albós A, et al. Reduced transfusion requirements in a splenectomized patient undergoing bone marrow transplantation. Acta Haematol (1994) 92(3):167– 8. doi: 10.1159/000204213
- von Bueltzingsloewen A, Bordigoni P, Dorvaux Y, Witz F, Schmitt C, Chastagner P, et al. Splenectomy may reverse pancytopenia occurring after allogeneic bone marrow transplantation. *Bone Marrow Transpl* (1994) 14 (2):339–40.
- Kröger N, Holler E, Kobbe G, Bornhäuser M, Schwerdtfeger R, Baurmann H, et al. Allogeneic stem cell transplantation after reduced-intensity conditioning in patients with myelofibrosis: a prospective, multicenter study of the Chronic Leukemia Working Party of the European Group for Blood and Marrow Transplantation. *Blood* (2009) 114(26):5264–70. doi: 10.1182/blood-2009-07-234880
- Bacigalupo A, Soraru M, Dominietto A, Pozzi S, Geroldi S, Van Lint MT, et al. Allogeneic hemopoietic SCT for patients with primary myelofibrosis: a predictive

- transplant score based on transfusion requirement, spleen size and donor type. Bone Marrow Transplant (2010) 45(3):458–63. doi: 10.1038/bmt.2009.188
- Robin M, Zine M, Chevret S, Meignin V, Munoz-Bongrand N, Moatti H, et al. The Impact of Splenectomy in Myelofibrosis Patients before Allogeneic Hematopoietic Stem Cell Transplantation. *Biol Blood Marrow Transpl* (2017) 23(6):958-64. doi: 10.1016/j.bbmt.2017.03.002
- McLornan DP, Yakoub-Agha I, Robin M, Chalandon Y, Harrison CN, Kroger N. State-of-the-art review: allogeneic stem cell transplantation for myelofibrosis in 2019. *Haematologica* (2019) 104(4):659–68. doi: 10.3324/ haematol.2018.206151
- Kröger N, Shahnaz Syed Abd Kadir S, Zabelina T, Badbaran A, Christopeit M, Ayuk F, et al. Peritransplantation Ruxolitinib Prevents Acute Graft-versus-Host Disease in Patients with Myelofibrosis Undergoing Allogenic Stem Cell Transplantation. *Biol Blood Marrow Transpl* (2018) 24(10):2152-6. doi: 10.1016/j.bbmt.2018.05.023
- Kalman NS, Mukhopadhyay ND, Roberts CH, Chung HM, Clark WB, McCarty JM, et al. Low-dose splenic irradiation prior to hematopoietic cell transplantation in hypersplenic patients with myelofibrosis. *Leuk Lymphoma* (2017) 58(12):2983–4. doi: 10.1080/10428194.2017.1321747
- Vyas OH, Kaul E, Rosenberg AS, Gunjan L, Shah UA, Comenzo RL, et al. Splenic Irradiation and a Reduced-Intensity Conditioning Regimen Prior to Allogeneic Stem-Cell Transplantation for Myelofibrosis. *Blood* (2014) 124 (21):3170. doi: 10.1182/blood.V124.21.3170.3170
- Elliott MA, Chen MG, Silverstein MN, Tefferi A. Splenic irradiation for symptomatic splenomegaly associated with myelofibrosis with myeloid metaplasia. Br J Haematol (1998) 103(2):505–11. doi: 10.1046/j.1365-2141.1998.00998.x
- Akpek G, Pasquini MC, Logan B, Agovi MA, Lazarus HM, Marks DI, et al. Effects of spleen status on early outcomes after hematopoietic cell transplantation. Bone Marrow Transpl (2013) 48(6):825–31. doi: 10.1038/ bmt.2012.249
- Bouabdallah R, Coso D, Gonzague-Casabianca L, Alzieu C, Resbeut M, Gastaut JA. Safety and efficacy of splenic irradiation in the treatment of patients with idiopathic myelofibrosis: a report on 15 patients. *Leuk Res* (2000) 24(6):491–5. doi: 10.1016/s0145-2126(00)00018-7
- Wagner H, McKeough PG, Desforges J, Madoc-Jones H. Splenic irradiation in the treatment of patients with chronic myelogenous leukemia or myelofibrosis with myeloid metaplasia. Results of daily and intermittent fractionation with and without concomitant hydroxyurea. *Cancer* (1986) 58(6):1204–7. doi: 10.1002/ 1097-0142(19860915)58:6<1204::aid-cncr2820580605>3.0.co;2-g
- Malato A, Rossi E, Tiribelli M, Mendicino F, Pugliese N. Splenectomy in Myelofibrosis: Indications, Efficacy, and Complications. Clin Lymphoma Myeloma Leuk (2020) 20(9):588–95. doi: 10.1016/j.clml.2020.04.015
- Kröger N, Giorgino T, Scott BL, Ditschkowski M, Alchalby H, Cervantes F, et al. Impact of allogeneic stem cell transplantation on survival of patients less than 65 years of age with primary myelofibrosis. *Blood* (2015) 125(21):3347– 50; quiz 3364. doi: 10.1182/blood-2014-10-608315
- Samuelson Bannow BT, Salit RB, Storer BE, Stevens EA, Wu D, Yeung C, et al. Hematopoietic Cell Transplantation for Myelofibrosis: the Dynamic International Prognostic Scoring System Plus Risk Predicts Post-Transplant Outcomes. *Biol Blood Marrow Transpl* (2018) 24(2):386–92. doi: 10.1016/j.bbmt.2017.09.016
- 31. Gagelmann N, Eikema DJ, de Wreede LC, Koster L, Wolschke C, Arnold R, et al. CMWP of the European Society for Blood and Marrow Transplantation. Comparison of Dynamic International Prognostic Scoring System and MYelofibrosis SECondary to PV and ET Prognostic Model for Prediction of Outcome in Polycythemia Vera and Essential Thrombocythemia Myelofibrosis after Allogeneic Stem Cell Transplantation. Biol Blood Marrow Transplant (2019) 25(6):e204–8. doi: 10.1016/j.bbmt.2019.03.024
- 32. Alchalby H, Yunus DR, Zabelina T, Kobbe G, Holler E, Bornhäuser M, et al. Risk models predicting survival after reduced-intensity transplantation for myelofibrosis. *Br J Haematol* (2012) 157(1):75–85. doi: 10.1111/j.1365-2141.2011.09009.x
- Gupta V, Malone AK, Hari PN, Ahn KW, Hu ZH, Gale RP, et al. Reducedintensity hematopoietic cell transplantation for patients with primary myelofibrosis: a cohort analysis from the center for international blood and marrow transplant research. *Biol Blood Marrow Transpl* (2014) 20(1):89–97. doi: 10.1016/j.bbmt.2013.10.018

Bacigalupo et al. Allogenic HSCT for Myelofibrosis

34. Raj K, Eikema DJ, McLornan DP, Olavarria E, Blok HJ, Bregante S, et al. Family Mismatched Allogeneic Stem Cell Transplantation for Myelofibrosis: Report from the Chronic Malignancies Working Party of European Society for Blood and Marrow Transplantation. *Biol Blood Marrow Transplant* (2019) 25 (3):522–8. doi: 10.1016/j.bbmt.2018.10.017

- Keyzner A, Shapiro S, Moshier S, Schorr E, Petersen E, Najfeld B, et al. Outcome of Allogeneic Hematopoietic Stem Cell Transplantation for Patients with Chronic and Advanced Phase Myelofibrosis. Biol Blood Marrow Transplant J Am Soc Blood Marrow Transplant (2016) 08:029. doi: 10.1016/j.bbmt.2016.08.029
- Bregante S, Dominietto A, Ghiso A, Raiola AM, Gualandi F, Varaldo R, et al. Improved Outcome of Alternative Donor Transplantations in Patients with Myelofibrosis: From Unrelated to Haploidentical Family Donors. *Biol Blood Marrow Transpl* (2016) 22(2):324–9. doi: 10.1016/j.bbmt.2015.09.028
- Robin M, Tabrizi R, Mohty M, Furst S, Michallet M, Bay JO, et al. Allogeneic haematopoietic stem cell transplantation for myelofibrosis: a report of the Société Française de Greffe de Moelle et de Thérapie Cellulaire (SFGM-TC). Br J Haematol (2011) 152(3):331–9. doi: 10.1111/j.1365-2141.2010.08417.x
- Murata M, Takenaka K, Uchida N, Ozawa Y, Ohashi K, Kim SW, et al. Comparison of Outcomes of Allogeneic Transplantation for Primary Myelofibrosis among Hematopoietic Stem Cell Source Groups. *Biol Blood Marrow Transpl* (2019) 25(8):1536–43. doi: 10.1016/j.bbmt.2019.02.019
- Robin M, Giannotti F, Deconinck E, Mohty M, Michallet M, Sanz G, et al. Eurocord and Chronic Malignancies Working Party-European Group for Blood and Marrow Transplantation (CMWP-EBMT). Unrelated cord blood transplantation for patients with primary or secondary myelofibrosis. *Biol Blood Marrow Transpl* (2014) 20(11):1841–6. doi: 10.1016/j.bbmt.2014.06.011
- Robin M, de Wreede LC, Wolschke C, Schetelig J, Eikema DJ, Van Lint MT, et al. Long-term outcome after allogeneic hematopoietic cell transplantation for myelofibrosis. *Haematologica* (2019) 104(9):1782–8. doi: 10.3324/ haematol.2018.205211
- Salas MQ, Lam W, Law AD, Kim DDH, Michelis FV, Loach D, et al. Reducedintensity conditioning allogeneic transplant with dual T-cell depletion in myelofibrosis. Eur J Haematol (2019) 103(6):597–606. doi: 10.1111/ eih 13327
- Morozova EV, Barabanshikova MV, Moiseev IS, Shakirova AI, Barhatov IM, Ushal IE, et al. A Prospective Pilot Study of Graft-versus-Host Disease Prophylaxis with Post-Transplantation Cyclophosphamide and Ruxolitinib in Patients with Myelofibrosis. Acta Haematol (2020) 23:1–8. doi: 10.1159/ 000506758
- Kröger N, Zabelina T, Schieder H, Panse J, Ayuk F, Stute N, et al. Pilot study of reduced-intensity conditioning followed by allogeneic stem cell transplantation from related and unrelated donors in patients with myelofibrosis. *Br J Haematol* (2005) 128(5):690–7. doi: 10.1111/j.1365-2141.2005.05373.x
- 44. Robin M, Porcher R, Wolschke C, Sicre de Fontbrune F, Alchalby H, Christopeit M, et al. Outcome after Transplantation According to Reduced-Intensity Conditioning Regimen in Patients Undergoing Transplantation for Myelofibrosis. *Biol Blood Marrow Transpl* (2016) 22(7):1206–11. doi: 10.1016/ j.bbmt.2016.02.019
- Patriarca F, Masciulli A, Bacigalupo A, Bregante S, Pavoni C, Finazzi MC, et al. Busulfan- or Thiotepa-Based Conditioning in Myelofibrosis: A Phase II Multicenter Randomized Study from the GITMO Group. *Biol Blood Marrow Transpl* (2019) 25(5):932–40. doi: 10.1016/j.bbmt.2018.12.064
- 46. Patriarca F, Bacigalupo A, Sperotto A, Isola M, Soldano F, Bruno B, et al. Allogeneic hematopoietic stem cell transplantation in myelofibrosis: the 20-year experience of the Gruppo Italiano Trapianto di Midollo Osseo (GITMO). Haematologica (2008) 93(10):1514–22. doi: 10.3324/haematol.12828
- Abelsson J, Merup M, Birgegård G, WeisBjerrum O, Brinch L. Brune M et al; Nordic MPD Study Group. The outcome of allo-HSCT for 92 patients with myelofibrosis in the Nordic countries. *Bone Marrow Transpl* (2012) 47 (3):380–6. doi: 10.1038/bmt.2011.91
- 48. McLornan D, Szydlo R, Koster L, Chalandon Y, Robin M, Wolschke C, et al. Myeloablative and Reduced-Intensity Conditioned Allogeneic Hematopoietic Stem Cell Transplantation in Myelofibrosis: A Retrospective Study by the Chronic Malignancies Working Party of the European Society for Blood and Marrow Transplantation. Biol Blood Marrow Transplant (2019) 25(11):2167– 71. doi: 10.1016/j.bbmt.2019.06.034

- Harrison C, Kiladjian JJ, Al-Ali HK, Gisslinger H, Waltzman R, Stalbovskaya V, et al. JAK inhibition with ruxolitinib versus best available therapy for myelofibrosis. N Engl J Med (2012) 366(9):787–98. doi: 10.1056/ NEJMoa1110556
- Verstovsek S, Mesa RA, Gotlib J, Levy RS, Gupta V, DiPersio JF, et al. A double-blind, placebo-controlled trial of ruxolitinib for myelofibrosis. N Engl J Med (2012) 366(9):799–807. doi: 10.1056/NEJMoa1110557
- Gupta V, Kosiorek HE, Mead A, Klisovic RB, Galvin JP, Berenzon D, et al. Ruxolitinib Therapy Followed by Reduced-Intensity Conditioning for Hematopoietic Cell Transplantation for Myelofibrosis: Myeloproliferative Disorders Research Consortium 114 Study. *Biol Blood Marrow Transpl* (2019) 25(2):256–64. doi: 10.1016/j.bbmt.2018.09.001
- 52. Robin M, Francois S, Huynh A, Cassinat B, Bay JO, Cornillon J, et al. Ruxolitinib Before Allogeneic Hematopoietic Stem Cell Transplantation (HSCT) In Patients With myelofibrosis: a Preliminary Descriptive Report Of The JAK ALLO Study, a Phase II Trial Sponsored By Goelams-FIM In Collaboration With The Sfgmtc. Blood (2013) 122(21):306. doi: 10.1182/blood.V122.21.306.306
- Ditschkowski M, Elmaagacli AH, Trenschel R, Steckel NK, Koldehoff M, Beelen DW. No influence of V617F mutation in JAK2 on outcome after allogeneic hematopoietic stem cell transplantation (HSCT) for myelofibrosis. Biol Blood Marrow Transpl (2006) Dec12(12):1350-1. doi: 10.1016/j.bbmt.2006.07.010
- Alchalby H, Badbaran A, Zabelina T, Kobbe G, Hahn J, Wolff D, et al. Impact of JAK2V617F mutation status, allele burden, and clearance after allogeneic stem cell transplantation for myelofibrosis. *Blood* (2010) 116(18):3572–81. doi: 10.1182/blood-2009-12-260588
- Panagiota V, Thol F, Markus B, Fehse B, Alchalby H, Badbaran A, et al. Prognostic effect of calreticulin mutations in patients with myelofibrosis after allogeneic hematopoietic stem cell transplantation. *Leukemia* (2014) 28 (7):1552–5. doi: 10.1038/leu.2014.66
- Kröger N, Panagiota V, Badbaran A, Zabelina T, Triviai I, Araujo Cruz MM, et al. Impact of Molecular Genetics on Outcome in Myelofibrosis Patients after Allogeneic Stem Cell Transplantation. *Biol Blood Marrow Transpl* (2017) 23(7):1095–101. doi: 10.1016/j.bbmt.2017.03.034
- Alchalby H, Badbaran A, Bock O, Fehse B, Bacher U, Zander AR, et al. Screening and monitoring of MPL W515L mutation with real-time PCR in patients with myelofibrosis undergoing allogeneic-SCT. *Bone Marrow Transpl* (2010) 45(9):1404–7. doi: 10.1038/bmt.2009.367
- Rumi E, Passamonti F, Arcaini L, Bernasconi P, Elena C, Pietra D, et al. Molecular remission after allo-SCT in a patient with post-essential thrombocythemia myelofibrosis carrying the MPL (W515A) mutation. *Bone Marrow Transpl* (2010) 45(4):798–800. doi: 10.1038/bmt.2009.231
- Wolschke C, Badbaran A, Zabelina T, Christopeit M, Ayuk F, Triviai I, et al. Impact of molecular residual disease post allografting in myelofibrosis patients. *Bone Marrow Transpl* (2017) 52(11):1526–9. doi: 10.1038/ bmt.2017.157
- Chiusolo P, Bregante S, Giammarco S, Lamparelli T, Casarino L, Dominietto A, et al. Full Donor Chimerism After Allogeneic Hematopoietic Stem Cells Transplant For Myelofibrosis: The Role Of The Conditioning Regimen. Am J Hematol (2021) 96:234–40. doi: 10.1002/ajh.26042
- Olsson RF, Logan BR, Chaudhury S, Zhu X, Akpek G, Bolwell BJ, et al. Primary graft failure after myeloablative allogeneic hematopoietic cell transplantation for hematologic malignancies. *Leukemia* (2015) 29(8):1754–62. doi: 10.1038/leu.2015.75
- Stasia A, Ghiso A, Galaverna F, Raiola AM, Gualandi F, Luchetti S, et al. CD34 selected cells for the treatment of poor graft function after allogeneic stem cell transplantation. *Biol Blood Marrow Transpl* (2014) 20(9):1440–3. doi: 10.1016/j.bbmt.2014.05.016
- Slot S, Smits K, van de Donk NW, Witte BI, Raymakers R, Janssen JJ, et al. Effect of conditioning regimens on graft failure in myelofibrosis: a retrospective analysis. *Bone Marrow Transpl* (2015) 50(11):1424–31. doi: 10.1038/bmt.2015.172
- 64. Kröger N, Zabelina T, Schieder H, Panse J, Ayuk F, Stute N, et al. Pilot study of reduced-intensity conditioning followed by allogeneic stem cell transplantation from related and unrelated donors in patients with myelofibrosis. *Br J Haematol* (2005) 128(5):690–7. doi: 10.1111/j.1365-2141.2005.05373.x

Bacigalupo et al. Allogenic HSCT for Myelofibrosis

 Gupta V, Kröger N, Aschan J, Xu W, Leber B, Dalley C, et al. A retrospective comparison of conventional intensity conditioning and reduced-intensity conditioning for allogeneic hematopoietic cell transplantation in myelofibrosis. *Bone Marrow Transpl* (2009) 44(5):317–20. doi: 10.1038/ bmt.2009.10

- Rondelli D, Goldberg JD, Isola L, Price LS, Shore TB, Boyer M, et al. MPD-RC 101 prospective study of reduced-intensity allogeneic hematopoietic stem cell transplantation in patients with myelofibrosis. *Blood* (2014) 124(7):1183–91. doi: 10.1182/blood-2014-04-572545
- 67. Robin M, Tabrizi R, Mohty M, Furst S, Michallet M, Bay JO, et al. Allogeneic haematopoietic stem cell transplantation for myelofibrosis: a report of the Société Française de Greffe de Moelle et de Thérapie Cellulaire (SFGM-TC). Br J Haematol (2011) 152(3):331–9. doi: 10.1111/j.1365-2141.2010.08417.x
- Ballen KK, Shrestha S, Sobocinski KA, Zhang MJ, Bashey A, Bolwell BJ, et al. Outcome of transplantation for myelofibrosis. *Biol Blood Marrow Transpl* (2010) 16(3):358–67. doi: 10.1016/j.bbmt.2009.10.025
- Klyuchnikov E, Holler E, Bornhäuser M, Kobbe G, Nagler A, Shimoni A, et al. Donor lymphocyte infusions and second transplantation as salvage treatment for relapsed myelofibrosis after reduced-intensity allografting. *Br J Haematol* (2012) 159(2):172–81. doi: 10.1111/bjh.12013
- 70. McLornan DP, Szydlo R, Robin M, van Biezen A, Koster L, Blok HJP, et al. Outcome of patients with Myelofibrosis relapsing after allogeneic stem cell transplant: a retrospective study by the Chronic Malignancies

- Working Party of EBMT. Br J Haematol (2018) 182(3):418-22. doi: 10.1111/bjh.15407
- Chhabra S, Narra RK, Wu R, Szabo A, George G, Michaelis LC, et al. Fludarabine/Busulfan Conditioning-Based Allogeneic Hematopoietic Cell Transplantation for Myelofibrosis: Role of Ruxolitinib in Improving Survival Outcomes. Biol Blood Marrow Transpl (2020) 26(5):893–901. doi: 10.1016/j.bbmt.2020.01.010
- Shahnaz Syed Abd Kadir S, Christopeit M, Wulf G, Wagner E, Bornhauser M, Schroeder T, et al. Impact of ruxolitinib pretreatment on outcomes after allogeneic stem cell transplantation in patients with myelofibrosis. *Eur J Haematol* (2018) 101(3):305–17. doi: 10.1111/ejh.13099

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2021 Bacigalupo, Innocenti, Rossi, Sora, Galli, Autore, Metafuni, Chiusolo, Giammarco, Laurenti, Benintende, Sica and De Stefano. This is an openaccess article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.





Integrating CAR T-Cell Therapy and Transplantation: Comparisons of Safety and Long-Term Efficacy of Allogeneic Hematopoietic Stem Cell Transplantation After CAR T-Cell or Chemotherapy-Based Complete Remission in B-Cell Acute Lymphoblastic Leukemia

OPEN ACCESS

Edited by:

Andrea Bacigalupo, Catholic University of the Sacred Heart, Italy

Reviewed by:

Jan Comelissen, Erasmus Medical Center, Netherlands Xiao-Dong Mo, Peking University People's Hospital, China

*Correspondence:

Peihua Lu peihua_lu@126.com

Specialty section:

This article was submitted to Alloimmunity and Transplantation, a section of the journal Frontiers in Immunology

Received: 13 September 2020 Accepted: 22 April 2021 Published: 07 May 2021

Citation:

Zhao Y-L, Liu D-Y, Sun R-J,
Zhang J-P, Zhou J-R, Wei Z-J,
Xiong M, Cao X-Y, Lu Y, Yang J-f,
Zhang X, Lu D-P and Lu P (2021)
Integrating CAR T-Cell Therapy and
Transplantation: Comparisons of
Safety and Long-Term Efficacy of
Allogeneic Hematopoietic Stem Cell
Transplantation After CAR T-Cell or
Chemotherapy-Based Complete
Remission in B-Cell Acute
Lymphoblastic Leukemia.
Front. Immunol. 12:605766.
doi: 10.3389/fimmu.2021.605766

Yan-Li Zhao¹, De-Yan Liu¹, Rui-Juan Sun¹, Jian-Ping Zhang¹, Jia-Rui Zhou¹, Zhi-Jie Wei¹, Min Xiong¹, Xing-Yu Cao¹, Yue Lu¹, Jun-fang Yang², Xian Zhang², Dao-Pei Lu^{1,3} and Peihua Lu^{2,3*}

¹ Department of Bone Marrow Transplantation, Hebei Yanda Lu Daopei Hospital, Langfang, China, ² Department of Hematology and Immunology, Hebei Yanda Lu Daopei Hospital, Langfang, China, ³ Lu Daopei Institute of Hematology, Beijing, China

Patients often undergo consolidation allogeneic hematopoietic stem cell transplantation (allo-HSCT) to maintain long-term remission following chimeric antigen receptor (CAR) Tcell therapy. Comparisons of safety and efficacy of allo-HSCT following complete remission (CR) achieved by CAR-T therapy versus by chemotherapy for B-cell acute lymphoblastic leukemia (B-ALL) has not been reported. We performed a parallel comparison of transplant outcomes in 105 consecutive B-ALL patients who received allo-HSCT after achieving CR with CAR-T therapy (n=27) or with chemotherapy (n=78). The CAR-T-allo-HSCT group had more patients in second CR compared to the chemotherapy-allo-HSCT group (78% vs. 37%; p<0.01) and more with complex cytogenetics (44% vs. 6%; p<0.001) but the proportion of patients with pre-transplant minimal residual disease (MRD) was similar. The median follow-up time was 49 months (range: 25-54 months). The CAR-T cohort had a higher incidence of Grade II-IV acute graft-versus-host disease (aGVHD 48.1% [95% CI: 46.1-50.1%] vs. 25.6% [95%CI: 25.2-26.0%]; p=0.016). The incidence of Grade III-IV aGVHD was similar in both groups (11.1% vs.11.5%, p=0.945). The overall incidence of chronic GVHD in the CAR-T group was higher compared to the chemotherapy group (73.3% [95%Cl: 71.3-75.3%] vs. 55.0% [95%CI: 54.2-55.8%], p=0.107), but the rate of extensive chronic GVHD was similar (11.1% vs.11.9%, p=0.964). Efficacy measures 4 years following transplant were

all similar in the CAR-T vs. the chemotherapy groups: cumulative incidences of relapse (CIR; 11.1% vs.12.8%; p=0.84), cumulative incidences of non-relapse mortality (NRM; 18.7% vs. 23.1%; p=0.641) leukemia-free survival (LFS; 70.2% vs. 64.1%; p=0.63) and overall survival (OS; 70.2% vs. 65.4%; p=0.681). We found that pre-transplant MRD-negative CR predicted a lower CIR and a higher LFS compared with MRD-positive CR. In conclusion, our data indicate that, in B-ALL patients, similar clinical safety outcomes could be achieved with either CD19 CAR T-cell therapy followed by allo-HSCT or chemotherapy followed by allo-HSCT. Despite the inclusion of more patients with advanced diseases in the CAR-T group, the 4-year LFS and OS achieved with CAR T-cells followed by allo-HSCT. Further confirmation of these results requires larger, randomized clinical trials.

Keywords: CD19 CAR T-cell therapy, relapse/refractory B cell acute lymphoblastic leukemia, allogeneic hematopoietic cell transplantation, survival, relapse

INTRODUCTION

Refractory/relapsed (R/R) B-cell acute lymphoblastic leukemia (B-ALL) is a leading cause of morbidity and mortality in children and young adults (1–3). Allogeneic hematopoietic stem cell transplantation (allo-HSCT) is often undertaken for high-risk R/R B-ALL patients. However, many R/R patients are never able to achieve a complete remission (CR) following chemotherapy and are not referred for allo-HSCT. Therefore, relapse rates among these patients remain high despite the potential cure that is possible for B-ALL patients with an allo-HSCT. Achieving a CR prior to allo-HSCT has been shown to improve outcomes for these R/R B-ALL patients including improving leukemia free survival (LFS) following transplantation (4, 5).

In recent years, clinical trials with anti-CD19+ chimeric antigen receptor T-cell (CAR-T) therapy have demonstrated high CR rates of ~70% to 90% in patients with R/R B-ALL (6-12) and offer the hope of a potential cure for those patients who are otherwise refractory or relapsed following chemotherapy. However, remission following CAR-T therapy is often not durable with about half of CR patients relapsing within 1 year of therapy (6, 10, 13, 14). There is accumulating evidence from recent studies demonstrating that CAR-T therapy followed by allo-HSCT could potentially result in higher rates of durable, long-term remission for pediatric R/R B-ALL and reduce the relapse rates seen with CAR T-cell therapy alone (6, 8, 15-17). Yet there is still controversy around the safety and efficacy of allo-HSCT following CAR T-cell therapy and the ability to achieve long-term LFS with this sequential, dual immunotherapy. Some studies have reported high relapse rates and higher treatment related mortality following transplant after CAR-T therapy, resulting in no improvement of LFS and overall survival (OS) when compared to CAR T-cell therapy alone (11, 18).

In the present study, we conducted a parallel comparison of outcomes among R/R B-ALL patients who achieved remission from either CAR T-cell therapy or chemotherapy and who subsequently underwent allo-HSCT. We report safety and efficacy results in these two cohorts.

MATERIALS AND METHODS

Patients

We included 105 consecutive B-ALL patients who underwent allo-HSCT after achieving CR either from CAR-T therapy (n=27) or chemotherapy (n=78) at the Hebei Yanda Lu Daopei Hospital between November 2015 and August 2016. Details on the enrollment of the CAR-T group and chemotherapy group (including 13 B-ALL patients with *BCR/ABL* who received chemotherapy plus a tyrosine kinase inhibitor) are shown in **Figure 1**.

The study protocol was approved by the ethics committee of the Hebei Yanda Lu Daopei Hospital. Informed consent was obtained from the patients according to the Declaration of Helsinki.

CD19+ CAR T-Cell Therapy and CAR-T-Related Side Effects

CD19+ CAR T-cell therapy was performed according to previously described methods (6). Briefly, we used a second generation CD19+ lentiviral vector that also expressed the costimulatory 4-1BB molecule (CAR-T clinical trials No: ChiCTR-IIh-16008711). Before CAR T-cell infusion, patients received lymphodepleting chemotherapy consisting of fludarabine (30 mg/m²/day) and cyclophosphamide (250 mg/m²/day) (FC) on days -5, -4, and -3. Cytokine release syndrome (CRS) and neurotoxicity grading were performed according to previously described methods (19–22).

Clinical Transplant Protocol

Before transplantation, patients received intensive myeloablative conditioning regimens. Total body irradiation (TBI) plus cyclophosphamide/fludarabine-based chemotherapy or busulfan (Bu) plus cyclophosphamide/fludarabine-based were used according to each patient's status. TBI-based regimes were preferred if no contraindications such as severe pulmonary complications were observed. TBI was given using a horizontal beam in a linear accelerator. Patients in the TBI group received conditioning with fractioned TBI (200cGy Bid for 5-6 doses). Patients in the Bu group received Bu (0.8mg/kg i.v. Q6h for 16 doses) on days -8 to -6. TBI or Bu was followed by

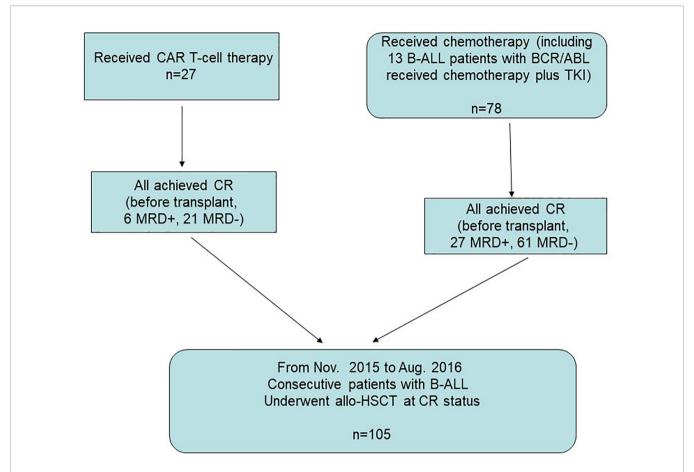


FIGURE 1 | Enrollment, and parallel comparison. Between November 2015 and August 2016, 105 consecutive B-ALL patients who underwent allo-HSCT after achieving CR either from CAR-T therapy (n=27) or chemotherapy (n=78) were enrolled for parallel comparison.

cyclophosphamide 1.8 g/m²/day for 2 days or fludarabine 30mg/ m²/day for 5 days. Rabbit anti-T Cell Globulin (ATG, Fresenius; totally 20mg/kg divided by 4 days) or thymoglobuline (ATG, Sanofi-Aventis, total dose of 7.5mg/kg divided by 4 days) were used on days -5 to -2 in mismatched unrelated transplants and haploidentical transplants (Haplo-HSCT). Cyclosporine, short-term methotrexate (15 mg/m² on day +1, then 10 mg/m² on days +3, +6, and +11 intravenously after transplantation), and mycophenolate mofetil were used for graft-versus-host disease (GVHD) prophylaxis. Grafts were granulocyte colony-stimulating factor (G-CSF) mobilized bone marrow (BM) and peripheral blood (PB) cells as described previously (23).

Acute GVHD (aGVHD) was diagnosed and graded according to modified Glucksberg criteria (24–26). Chronic GVHD (cGVHD) was evaluated using National Institutes of Health consensus criteria (27, 28) and aGVHD treatment was described previously (23, 24). Thrombotic microangiopathy (TMA) was diagnosed according to the Jodele criteria (29).

Analysis of Chimerism

Hematopoietic chimerism was evaluated by PCR amplification of short tandem repeats (STR) using both bone marrow and CD3+

cells from PB samples collected at 1, 2, 3, 6, 9, and 12 months after transplant and at 6-month intervals thereafter. Complete donor chimerism was defined as the presence of \geq 95% of the donor-type.

Statistics

Comparisons of patient characteristics between the two groups were performed using the Mann-Whitney U-test for continuous variables and $\chi 2$ for categorical data. The probabilities of survival were calculated using the Kaplan-Meier method. Cumulative incidences were estimated for aGVHD, non-relapse mortality (NRM), and relapse to accommodate for competing risks. Death and relapse were competing events for aGVHD and death was a competing event for cGVHD. NRM was the competing event for relapse and vice versa. Hazard ratios (HRs) for clinical outcomes were estimated in a multivariate analysis using Cox proportional hazards regression with a backward stepwise model selection approach. The following variables were included: gender, patient age (<14 years vs. ≥14 years), pre-HSCT treatment (CAR-T vs. chemotherapy), disease status pre-transplant (≥CR2 vs. CR1), MRD status pre-transplant (positive vs. negative), poor risk chromosomes (yes vs. no), conditioning regimens (TBI-based

vs. Bu-based), donor type (alternative donor vs. identical unrelated or sibling donor), donor-recipient gender matching (female to male vs. others), course from diagnosis to transplant, and mononuclear, CD3+ and CD34+ cell counts (using the median value as the cut-off point). Independent variables with P>0.1 were sequentially excluded from the model, and P<0.05 was considered to be statistically significant. P values were 2-sided. The SPSS 16 (SPSS Inc./IBM, Armonk, NY, USA) and the R software package (version 4.0.0; http://www.r-project.org) were used for data analyses. Surviving patients were censored on April 30th, 2020.

Definitions

CR and CR with incomplete count recovery (CRi) were defined in accordance with the National Comprehensive Cancer Network (NCCN) guideline, version 1.2018 (30). Minimal residual disease (MRD)-negative status was defined as the absence of leukemia cells in BM determined by multiparameter flow cytometry (FCM, sensitivity, 1:10,000), and the absence of leukemia-associated fusion gene in BM determined by real-time quantitative PCR (RT-PCR). Hypodiploidy, complex karyotype, t(v;11q23) or t(9;22) (q34;q11.2) determined by G band or FISH were defined as poor risk chromosomes according to the NCCN guideline, version 1.2018 (30). LFS and OS were calculated from the date of allo-HSCT to the date of relapse or death or the last follow-up time. The cumulative incidence of relapse (CIR) was calculated from date of allo-HSCT to the date of relapse.

RESULTS

Patient Characteristics

The detailed characteristics of the two groups are summarized in **Table 1**. Patients' median age was 13 years (range: 2–52 years) with a 58/42 male/female ratio. Fifty-three percent of the patients were pediatric (age <14 years) and 47% of patients were adults (age \geq 14 years). High WBC counts were observed in 31 patients (30%) at initial diagnosis.

The median time from CAR T-cell therapy to HSCT was 84 days (range: 35-293 days). CRS was observed in the majority of the patients in the CAR T-cell therapy group. Grade 1 (56%) and Grade 2 (26%) CRS made up the majority of CRS cases. Severe CRS occurred in 15% of patients—11% of patients had Grade 3, and 4% of patients had Grade 4 CRS. A total of four patients had Grade 3 neurotoxicity with seizures.

In the CAR-T group, 22 (81%) patients had R/R B-ALL, and 5 (19%) had persistent or relapsed MRD after hematological remission. Among the 21 relapsed patients in the CAR T-cell group, the median time from diagnosis to first relapse was 17 months (range: 3-47 months). Following relapse, 17 patients failed to regain CR after a median 2 courses of chemotherapy (range: 1-5 courses) and afterwards underwent CAR T-cell therapy. Four patients who had relapsed during consolidation chemotherapy received CAR-T therapy directly. In the chemotherapy group, 48 (62%) had R/R B-ALL, and 12 (15%) had persistent or recurrent MRD. The median time from

diagnosis to last relapse of the 33 relapsed patients was 31 months (range: 2-120 months). One patient relapsed 3 times within 10 years. Among 18 (17%) patients in the chemotherapy group, 9 presented with high risk ALL. As shown in **Table 1**, in the CAR-T group, prior to allo-HSCT, 22% (6/27) of the patients were in CR1 compared to 63% (49/78) of the patients in CR1 in the chemotherapy group. Compared to the chemotherapy group, the CAR-T group had more patients who were \geq CR 2 (78% ν s. 37%, respectively; p<0.001).

As assayed by FCM and RT-PCR, 22% of patients in the CAR-T group and 35% of patients in the chemotherapy group had MRD detected pre-transplant (p=0.232). The proportion of patients with extramedullary diseases at diagnosis and at relapse before transplant were not significantly different between the CAR-T group and the chemotherapy group (p=0.927). There was no significant difference in the median time from diagnosis to transplant (13.5 months [range: 4-123 months]). Complex chromosomes were present in 44% of patients in the CAR-T group and 12% of the chemotherapy group (p<0.001). There was significant difference in the presence of fusion genes (p=0.023). Poor risk BCR-ABL1 (n=13) and MLL-AF4 (n=4) were exclusively observed in the chemotherapy group (**Table 1**).

Donor Source, Graft, Conditioning Regimens and Engraftment

In the CAR-T group, 59% of patients received a transplant from haploidentical donors (Haplo-D), 30% from matched unrelated donors (MUD), and 11% from HLA-matched sibling donors (MSD). In the chemotherapy group, 64% of patients received a transplant from Haplo-Ds, 21% from MUDs and the remaining 15% of patients received a transplant from MSDs. There were no significant differences among different donor types, the donors' age and gender between the CAR-T and chemotherapy groups. In addition, there were no differences in the median mononuclear cells, CD34 and CD3 between the two groups (Table 1).

Myeloablative conditioning regimens were administered with TBI cyclophosphamide/TBI-fludarabine in 83% of patients and Bu cyclophosphamide/fludarabine in 17% of patients. There was no significant difference in conditioning regimens observed between the groups.

All patients achieved sustained neutrophil engraftment after a median of 14 days (range: 11-20 days) in the CAR-T group and 14 days (range: 10-29 days) in the chemotherapy group (p=0.97). Platelet engraftment failure occurred in 2 patients (7%) in the CAR-T group. One patient died of severe acute GVHD on day 27 after transplantation, and one died of infection at 68 days post-transplantation. All the 78 patients in the chemotherapy group achieved sustained platelet engraftment. There was a significant difference in platelet engraftment between the two groups (p=0.026). Post-transplant, the median day of platelet engraftment was significantly longer in the CAR-T group (14 days, range: 5-47 days) compared to the chemotherapy group (12 days, range: 4-32 days) (p=0.026).

No graft failure occurred (except that one patient had poor graft function) and rapid achievement of full donor chimerism

TABLE 1 | Patient characteristics.

Characteristics	Total	CAR-T group	Chemotherapy group	P value
No.	105	27	78	
Median age, years (range)	13.0(2-52)	11(3-44)	14.5(2-52)	0.441
Age group, no. (%)	, ,	, ,	, ,	0.236
≥14	49(47)	10(36)	39(50)	
<14	56(53)	17(63)	39(50)	
Median donor age, years (range)	33(8-63)	31(16-54)	33.5(8-63)	0.550
Gender, male, no. (%)	61(58%)	18(67)	43(55)	0.295
With extramedullary disease (EMD), no (%)	15(14)	4(15)	11(14)	0.230
Median duration from diagnosis to HSCT months (range)	13.5(4-123)	20(4-54)	11(4-123)	0.232
Disease risk	10.5(4-125)	20(4-04)	11(4-120)	0.232
R/R B-ALL, no (%)	70(67)	22(82)	49(60)	0.042
	70(67)	22(82)	48(62)	
From diagnosis to first relapse time, no. (%)	10(50)	44(44)	0/47)	
<18 months	19(58)	11(41)	8(17)	
18~36 months	19(44)	8(30)	11(14)	
>36 months	16(19)	2(1)	14(18)	
Primary refractory	16(19)	1(0)	15(19)	
Persistent or relapsed MRD	17(16)	5(19)	12(15)	
Others	18(17)	0	18(17)	
Disease status pre-transplant, no. (%)				< 0.001
CR1	55(52)	6(22)	49(63)	
≥CR2	50(48)	21(78)	29(37)	
Donor source, no. (%)				0.588
Haplo-d	66(63)	16(59)	50(64)	
MUD	24(23)	8(30)	16(21)	
MSD	15(14)	3(11)	12(15)	
FCM MRD-positive pre-conditioning, no. (%)	33(31)	6(22)	27(35)	0.232
Fusion genes				0.023
BCR-ABL1	13(12)	0	13(17)	
TEL-AML1	9(9)	2(7)	7(9)	
E2A-PBX1	4(4)	3(11)	1(1)	
MLL-AF4	4(4)	0	4(5)	
MLL-AF1P	1(1)	1(4)	0	
MLL-ENL	1(1)	0	1(1)	
Gene mutations	1(1)	O	1(1)	
NRAS/KRAS	12(11)	5(11)	7(9)	0.16
IKZF				0.10
TP53	9(9)	2(7)	7(9)	
Flt3 ITD/KTD	5(5)	2(7)	3(4)	0.383
	4(4)	1(4)	3(4)	0.728
High risk cytogenetics, no. (%)	48(46)	16(59)	32(41)	0.103
Complex cytogenetic, no (%)	17(16)	12(44)	5(6)	<0.001
Donor-recipient gender match, n (%)				0.263
Female to male	19(18)	7(26)	12(15)	
Others	86(82)	20(74)	66(85)	
Conditioning regimens, no. (%)				0.335
TBI-based	87(83)	24(89)	63(81)	
Bu-based	18(17)	3(11)	15(19)	
Median CD34 cells,×10 ⁶ /kg(range)	4.45(1.76-12.23)	4.6(1.76-10.18)	4.41(2.02-12.23)	0.428
Median CD3 cells,×10 ⁸ /kg(range)	1.66(0.44-4.99)	1.83(0.85-3.04)	1.59(0.44-4.99)	0.491
Graft type, no. (%)				0.41
BM+PB	80(76)	19(70)	61(78)	
PB	25(24)	8(30)	17(22)	
Neutrophil engraftment, days (range)	14(10-29)	14(11-20)	4(10-29)	0.973
Platelet engraftment, days (range)	12(4-47)	14(5-47)	12(4-32)	0.026
Median follow-up time in survivor, months (range)	49 (25-54)	49 (44-53)	49 (25-54)	0.831

was confirmed in all patients by day 30. No significant difference between the two groups was observed.

Incidence of GVHD

The cumulative incidence of Grade II-IV aGVHD was higher in the CAR T-cell group compared to the chemotherapy group (48.1% [95% CI: 46.1, 50.1%] vs. 25.6% [95% CI: 25.2, 26.0%],

respectively; p=0.016), while the incidence of Grade III-IV aGVHD were similar between the two groups (11.1% [95% CI: 10.3, 11.9%] vs. 11.5% [95% CI: 11.3, 11.7%], respectively; p=0.945) (**Figures 2A, B**). A low versus high grade CRS (Grade 0-1 vs. Grade 2-4) before transplant did not have significant effects on Grade II-IV aGVHD (47.4% [95% CI: 44.7, 50.1%] vs. 50.0% [95% CI: 42.6, 57.4%] among the CAR-

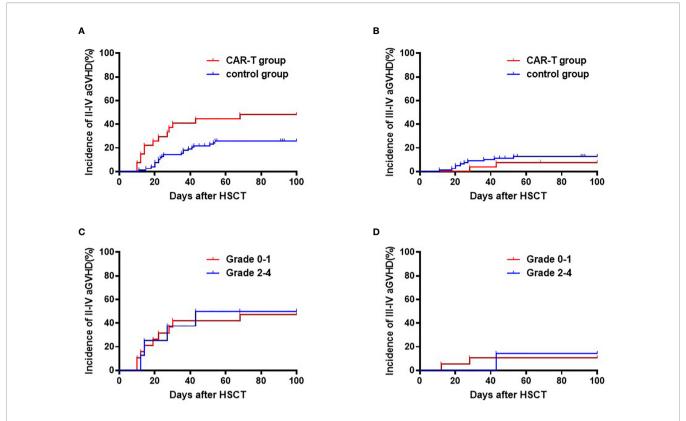


FIGURE 2 | Cumulative incidences of Grade II-IV and Grade III-IV acute GVHD. (A) Cumulative incidences of Grade II-IV acute GVHD: CAR-T group: 48.1% (95% CI:46.1, 50.1%) vs. chemotherapy group: 25.6% (95% CI: 25.2, 26.0%); p=0.016. (B) Cumulative incidences of Grade III-IV acute GVHD: CAR T-cell group: 11.1% (95% CI: 10.3, 11.9%) vs. chemotherapy group: 11.5% (95% CI: 11.3, 11.7%); p=0.945. (C) Cumulative incidences of Grade II-IV acute GVHD: CRS Grade 0-I 47.4% (95% CI: 44.7, 50.1%) vs. CRS Grade II-IV: 50.0% (95% CI: 42.6, 57.4%); p=0.95. (D) Cumulative incidences of Grade III-IV aGVHD: CRS Grade 0-I: 10.5% (95% CI: 9.5, 11.5%) vs. CRS Grade II-IV: 12.5% (95% CI: 9.4, 15.6%); p=0.92.

T and chemotherapy groups, respectively; p=0.95) or on Grade III-IV aGVHD (10.5% [95% CI: 9.5, 11.5%] *vs.* 12.5% [95% CI: 9.4, 15.6%]; p=0.92) after transplant (**Figures 2C, D**).

For patients surviving over 100 days after transplantation, cumulative incidence of cGVHD at 18 months were higher in the CAR-T group, but this difference did not reach statistical difference (71.3% [95% CI: 71.3, 75.3%] vs. 55.0% [95% CI: 54.2, 55.8%], p=0.107) (**Figure 3A**). Cumulative incidence of extensive cGVHD at 18 months was similar between the CAR-T and chemotherapy groups (11.1% [95% CI: 10.3, 11.9%] vs. 11.9% [95% CI: 11.7, 12.1%]; p=0.964) (**Figure 3B**).

CIR After HSCT

The CIRs at 4 years following transplant were 11.1% [95%CI: 10.3,11.9%] for the CAR-T group versus 12.8% [95% CI: 12.0-13.6%] for the chemotherapy group (p=0.84) (**Figure 4A**). Univariate analysis showed that disease status (HR 3.87, [95% CI 1.09-13.7], p=0.027) and MRD before transplantation (HR 2.81 [95%CI 0.961-8.24], p=0.056) were predictive factors for relapse. The multivariate analysis confirmed these predictive effects of relapse (HR 4.10, [95% CI1.13-14.84], p=0.031 and HR 3.02, [95% CI1.02-8.96], p=0.046, for disease status and MRD before transplant, respectively) (**Table 2**).

A total of 13 patients relapsed after transplant, three in CAR-T group and 10 in the chemotherapy group. All except one patient died at a median time of day 283 (range: 48-1116) after transplant. The patient that survived relapsed following a second haploidentical transplant underwent donor CD19 CAR T-cell therapy and remains in remission.

In the CAR T-cell group, 6 patients were MRD positive (MRD+CR) before transplant, including 5 patients who were CD19 negative (CD19-) and MRD+CR. Two of the CD19-MRD+CR patients relapsed with CD19- leukemia at Day 60 and at Day 275, and consequently died at Day 270 and Day 336 after transplant, respectively. Another patient died of severe GVHD on Day 27. The remaining three patients survived in remission at Month 46, 47, and 49, respectively.

Infection, TMA and NRM

No remarkable differences were observed in cytomegalovirus (CMV) reactivation (52% vs. 50%, p=0.93) between the CAR T-cell and chemotherapy groups, respectively. There were also no differences in rates of transplant-associated TMA (TA-TMA) between the CAR T-cell and chemotherapy groups, respectively (15% vs. 14%, p=0.51). In the chemotherapy group, three patients were diagnosed with viral pneumonia

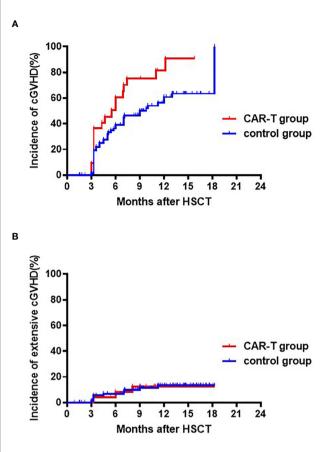


FIGURE 3 | Cumulative incidences of chronic and extensive chronic GVHD. **(A)** Cumulative incidences of chronic GVHD: CAR T-cell group: 73.3% (95% CI: 71.3, 75.3%) vs. chemotherapy group: 55.0% (95% CI: 54.2, 55.8%); p=0.107. **(B)** Cumulative incidences of extensive chronic GVHD: CAR T-cell group: 11.1% (95% CI:10.3, 11.9%) vs. the chemotherapy group: 11.9% (95% CI: 11.7, 12.1%); p=0.964.

and died. Incidences or non-relapse mortality (NRM) within 100 days were 7.4% (95% CI:6.8-8.0%) and 5.1% (95% CI: 4.9-5.3%) (p=0.64). The NRM at 1 and 4 years after transplantation was 11.1% (95% CI: 10.3-11.9%) and 18.7% (95% CI: 17.5, 19.9%), respectively, for the CAR-T group versus 16.7% (95% CI [16.3-17.1%] and 23.1% (95% CI: 22.7, 23.5%) for the chemotherapy group (p=0.64) (**Figure 4B**).

LFS, OS and Cause of Mortality

With a median follow-up of 49 months (range: 44-54 months) for surviving patients, LFS and OS at 4 years were similar in the CAR-T and chemotherapy groups (LFS: 70.2% [95% CI: 53.0, 87.4%] *vs.* 64.1% [95% CI: 53.5, 74.7%], p=0.63; OS: 70.2% [95% CI: 53.0, 87.4%] *vs.* 65.4% [95%CI:54.8, 76.0%], p=0.681) (**Figure 5A**).

Univariate and multivariate analysis showed that MRD prior to transplant was a negative prognostic factor for LFS (p=0.024 for univariate-analysis and p=0.016 (HR 2.6 [95% CI: 1.2, 5.8] for multivariate-analysis). The 4-year LFS for the MRD-CR and

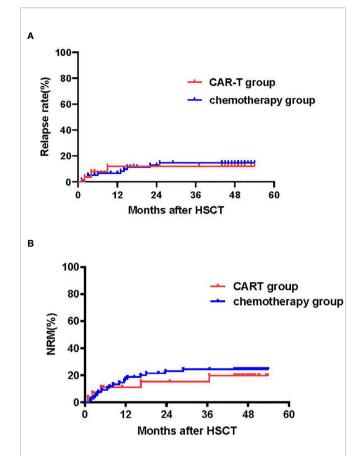


FIGURE 4 | Cumulative incidences of relapse (CIR) and NRM. **(A)** Cumulative incidence of relapse: CAR T-cell group: 4-year CIR of 11.1% (95% CI: 10.3, 11.9%) vs. chemotherapy group: 12.8% (95% CI:12.6, 13.0%) (p=0.84). **(B)** Cumulative incidence of NRM: CAR T-cell group: 4-year NRM of 18.7% (95% CI:17.5, 19.9%) vs. chemotherapy group: 4-year NRM of 23.1% (95% CI:22.7, 23.5%); p=0.64.

MRD+CR groups was 72.2% [95% CI: 61.8, 82.6%] and 51.5% [95% CI: 34.4, 68.6%], respectively (p=0.024). The 4-year OS for MRD-CR patients was 73.6% (95%CI: 63.4, 83.8%) and MRD+CR of 51.5% (95%CI: 34.4, 68.6%) (p=0.02), respectively (**Figure 5B**).

For the 97 patients who survived over 100 days after transplant, 4-year LFS for no cGVHD, and limited and extensive cGVHD was 62.2% (95% CI: 46.5, 77.9%), 85.6% (95% CI: 75.8, 95.4%) and 36.4% (95% CI: 8.0, 64.8%), respectively (no *vs.* limited cGVHD, p=0.009; limited *vs.* extensive cGVHD, p<0.001; no *vs.* extensive cGVHD, p=0.20). The 4-year OS for the no cGVHD cohort was 64.9% (95% CI:49.6, 80.2%), 85.6% (95% CI: 75.8, 95.4%) for the limited cGVHD cohort and 36.4% (95% CI: 8.0, 64.8%) for the extensive cGVHD cohort (no *vs.* limited cGVHD, p=0.019; limited *vs.* extensive cGVHD, p<0.001; no *vs.* extensive cGVHD, p=0.123) (**Figure 5C**).

At the time of the latest follow up in April 2020, 29 patients had died. The primary causes of death were relapse (8 patients), GVHD (8 patients), infection (5 patients) and TMA (5 patients) (**Table 3**).

TABLE 2 | Univariate and multivariate analysis of risk factors for clinical outcomes.

		=	II-IV GVHD	٥				=	III-IV GVHD	٩				J	CIR				Ż	MAN				5	LFS		
		Univariate		Multiv	Multivariate		Univariate	riate		Multiv	Multivariate		Univ	Univariate		Multivariate	iate		Univariate		Multivariate		Univariate			Multivariate	
	£	95% CI	p h value	HR 95%	% p	壬	95% CI		p H	HR 95%	95% CI	p H	HR 95	95% CI p	p HR value	95% CI	p value	壬	65% CI	p HR value	R 65% p CI value	壬	65% CI	p value	띂	65% CI	p value
Age(≥14 vs. <14yrs) Treatment pre-HSCT (CAR-T vs.	0.785 (0.785 0.395-1.56 0.49 2.36 1.18-4.75 0.016	0.49			0.339	0.339 0.0921-1.25 0.956 0.263-3.47		0.088 0.5	0.506 0.1081-2.37		0.39 1.	1.23 0.41	0.418-3.62 0.708	36			2.09	0.295-2.12	0.084		1.8	0.921-3.51	0.079	1.652	0.84-3.25	0.15
Disease status(≥CR2 vs. CR 1)	1.09	1.09 0.557-2.15 0.769	0.769			0.494	1 0.152-1.6		0.239			e,	3.87 1.09	1.09-13.7 0.0	0.027 4.1	1.13-	0.031	0.774	0.774 0.344-1.74 0.	0.54		1.54	0.808-2.93	3 0.192			
MRD(pos vs. neg)	1.09	1.09 0.536-2.23 0.81	0.81			2.29	0.75-7.02		0.142			2	2.81 0.96	0.961-8.24 0.056 3.02	156 3.02			1.45	0.046 1.45 0.638-3.32 0.	0.38		2.21	1.16-4.24 0.017 2.105 1.09-4.06	0.017	2.105	1.09-4.06	0.027
Poor risk chromosomes (yes vs.	1.5	0.76-2.95 0.245	0.245			2.48	0.735	-8.17 0.1	0.125			0	709 0.23	0.709 0.236-2.13 0.543	:43	0.0		0.622	0.263-1.47	0.271		0.614	0.614 0.313-1.21	0.151			
Conditioning(TBI-based vs. Bu-	0.888	0.888 0.365-2.16 0.795	0.795			0.254	1 0.0815→	8.17 0.0	0.5	0.0815-8.17 0.015 0.387 0.0929-1.61		0.19 0.6	655 0.18	0.655 0.181-2.38 0.523	523			0.937	0.314-2.79 0.903	903		0.883	0.883 0.384-2.03	3 0.765			
Donor type (haplo vs. MUD/MSD) donor-ecipients gender matched	0.894	0.894 0.45-1.78 0.753 1.16 0.464-2.89 0.761	0.753			3.17	0.715-14		0.113	0.113 0.0375 1.885 0.4163-8.54	3-8.54	1.	1.33 0.41	0.413-4.31 0.629 0.271-4.89 0.853	53			0.917	0.402-2.09	0.84		1.25	0.646-2.42 0.516 0.234-1.59 0.33	0.516			
(errate to mate vs. ouners) course from diagnosis to transplant 0.883 0.45-1.73 0.719	0.883	0.45-1.73	0.719			0.455	5 0.141-1.47		0.188			2	2.34 0.73	0.732-7.48 0.144	44			9.0	0.262-1.37 0.3	0.225		1.05	0.552-1.98	0.893			
(z median vs. <median) MNC (zmedian vs. <median) CD3 (zmedian vs. <median)< td=""><td>0.59 0</td><td>0.59 0.303-1.15 0.3</td><td>0.3</td><td></td><td></td><td>0.996</td><td>0.351-2.82</td><td></td><td>0.934</td><td></td><td></td><td>- ö</td><td>1.11 0.4</td><td>0.412-3 0.886</td><td>37</td><td></td><td></td><td>0.926</td><td>0.433-1.98</td><td>0.871</td><td></td><td>1.12</td><td>0.62-2.01</td><td>0.616</td><td></td><td></td><td></td></median)<></median) </median) 	0.59 0	0.59 0.303-1.15 0.3	0.3			0.996	0.351-2.82		0.934			- ö	1.11 0.4	0.412-3 0.886	37			0.926	0.433-1.98	0.871		1.12	0.62-2.01	0.616			
CD34 (zmedian vs. <median) (limited="" cgvhd="" extensive="" td="" vs.="" vs.<=""><td>1.01</td><td>0.511-1.98 0.986</td><td>0.986</td><td></td><td></td><td>0.682</td><td></td><td></td><td>0.51</td><td></td><td></td><td>Ö</td><td></td><td></td><td>0.128</td><td></td><td></td><td>0.916</td><td>0.408-2.06</td><td>0.834</td><td></td><td>0.576</td><td></td><td></td><td>0.644</td><td>0.34-1.22</td><td>0.18</td></median)>	1.01	0.511-1.98 0.986	0.986			0.682			0.51			Ö			0.128			0.916	0.408-2.06	0.834		0.576			0.644	0.34-1.22	0.18

DISCUSSION

Recently CAR-T therapy has shown dramatic initial responses with CR rates approaching 80-90% among R/R B-ALL patients (6–12). However, risk of relapse remains a major problem for these patients. Allo-HSCT after CAR-T therapy may have a consolidative role to further improve the durability of remission for these patients. However, whether prior CAR-T therapy can potentially increase the transplant-related mortality and toxicity remain a concern. In the present study, we compared the safety and efficacy of allo-HSCT in patients in patients after achieving CR either post CAR-T or after chemotherapy with a median follow-up of 4 years. To our knowledge, this is the first analysis comparing B-ALL patient outcomes after allo-HSCT following either prior CAR T-cell therapy or chemotherapy.

Although this is not a randomized trial, our parallel cohort study showed a similarly high LFS (70.2% vs. 64.1%) and OS (70.2% vs. 65.4%) after a median follow-up of 4 years in patients who received allo-HSCT after achieving CR from CAR-T therapy (n=27) or after achieving CR following chemotherapy (n=78), even despite having significantly more patients with advanced disease and refractory/relapsed status in the CAR-T group. There was no graft-failure in either group. The incidences of NRM, TMA and CMV reactivation within both groups were similar.

Hematopoietic reconstitution is one of the key issues in heavily pre-treated B-ALL patients after allo-HSCT. In our study, all patients achieved prompt and sustained neutrophil engraftment, at a median 14 days and achieved 100% donor chimerism in bone marrow and blood on day 28 in both the CAR-T and chemotherapy groups. Nevertheless, the engraftment of platelets was significantly slower in the CAR-T group compared to the chemotherapy group (Day 14 vs. Day 12, p=0.026). One possible reason may be due to the higher incidence of aGVHD and corresponding glucocorticoids treatment of patients in the CAR-T group. In addition, cytokine storm subsequent to CAR-T therapy might impair the endothelium system in transplant recipients (6–9), including the hematological microenvironment.

GVHD remains a major cause of morbidity and mortality following allo-HSCT. The reports of incidence and severity of GVHD after transplant post CAR T-cell therapy have been very limited. In a study by Shadman et al. from the University of Washington, it reported incidence of Grade II-IV and Grade III-IV acute GVHD of 69% and 25%, respectively (17). Jiang et al. reported no severe aGVHD except for mild skin rash and diarrhea (Grade ≤2) among 21 patients (31). In our study, Grade II-IV and Grade III-IV aGVHD were 48% and 11%, respectively, in the CAR-T group. Further analysis showed that the grade of CRS had no influence on the incidence and severity of aGVHD. Considering the limited size of the CAR-T group in our study, further clinical trials are necessary to verify the effect of CRS on aGVHD after transplantation. We found higher incidence of Grade II-IV aGVHD in the CAR T-cell group, but similar incidence of severe aGVHD compared to the chemotherapy group. Regarding the cGVHD, the overall incidence of cGVHD in the CAR-T group was higher, but the

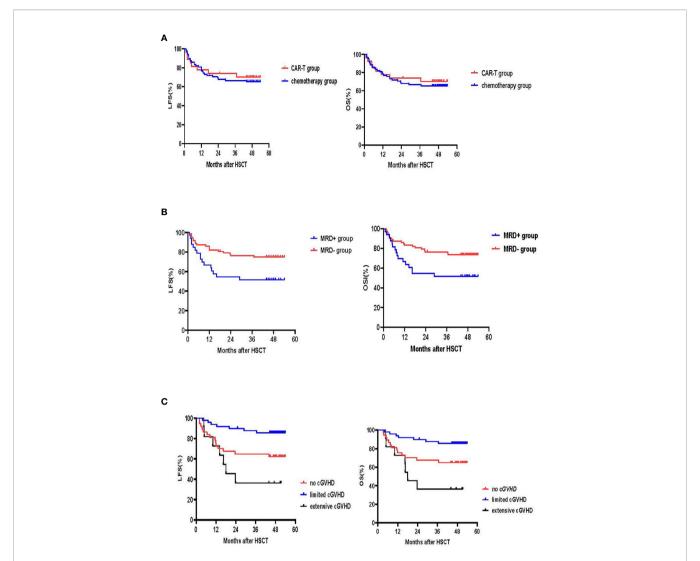


FIGURE 5 | LFS and OS. (A) LFS and OS in the CAR-T and chemotherapy groups. The 4-year LFS for the CAR-T group was 70.2% (95% CI:53.0, 87.4%) vs. 64.1% (95% CI:53.5, 74.7%) for the chemotherapy group (p=0.63). The 4-year OS for the CAR-T group was 70.2% (95% CI:53.0, 87.4%) vs. 65.4% (95% CI:54.8, 76.0%) for the chemotherapy group (p=0.681) (B) LFS and OS according to MRD. The 4-year LFS for patients who achieved MRD- CR was 72.2% (95% CI:61.8, 82.6%) and 51.5% (95% CI:34.4, 68.6%) for those that had an MRD+ CR (p=0.024). The 4-year OS for the MRD- CR group was 73.6% (95% CI:63.4, 83.8%) and 51.5% (95% CI:34.4, 68.6%) for the MRD+CR group (p=0.02). (C) LFS and OS according to cGVHD. 4-year LFS for patients without cGVHD was 62.2% (95% CI:46.5, 77.9%), 85.6% (95% CI:75.8,95.4%) for those with limited cGVHD and 36.4% (95% CI:8.0, 64.8%) for those with extensive cGVHD (no vs. limited cGVHD, p=0.009; limited vs. extensive cGVHD, p<0.001; no vs. extensive cGVHD, p=0.20). 4-year OS for the no cGVHD group was 64.9% (95% CI:49.6, 80.2%), 85.6% for the limited cGVHD group (95% CI:75.8, 95.4%) and 36.4% for the extensive cGVHD group (95% CI:8.0, 64.8%) (no vs. limited cGVHD, p=0.019; limited vs. extensive cGVHD, p<0.001; no vs. extensive cGVHD, p<0.001; no vs. extensive cGVHD, p<0.123).

TABLE 3 | Causes of death in the CAR-T and chemotherapy groups.

Causes	Total	CAR-T group	Chemotherapy group
All causes of death	34	8	26
Relapse	11	3	8
NRM	23	4	18
GVHD	8	1	7
Infection	6	2	4
TMA	5	1	4
Graft failure or rejection	1		1
Malignant arrhythmia	1		1
Acute pancreatitis	1	1	
Organic pneumonia	1		1

rate of extensive cGVHD (11.1% vs. 11.9%, in the CAR T-cell and chemotherapy groups, respectively p=0.96) was similar between the groups. There was a relative higher incidence of cGVHD in our study. It is likely because the major donor type was haplo donor (67%) in our study. In addition, considering the high risk of recurrence in this group of patients with advanced disease, immunosuppressants were withdrawn as soon as possible. And in some very high-risk patients, prophylactic DLI and interferon were applied to gain a limited chronic GVHD status in both groups. The incidence of the cGVHD was similar to our previous reports of haplo-HSCT with ATG (23, 32).

Neurotoxicity is a relatively common toxicity with CAR-T therapy (20–22). However, whether development of CAR-T-related neurotoxicity increases the neurotoxicity of fundamental immunomodulators such as cyclosporine and tacrolimus after allo-HSCT is still unclear. In our study, four patients experienced Grade III neurotoxicity with seizures after CAR-T infusion. However, none of the four patients had developed drug-induced encephalopathy or TMA after transplant. Nevertheless, patients who present with severe neurotoxicity after CAR-T should be followed up and treated with caution.

There are now more and more published studies confirming that consolidative allo-HSCT following CAR-T therapy could reduce relapse rates and improve LFS for R/R ALL patients (6, 10, 11, 14-16). However, studies comparing CIR, LFS and OS between patients post CAR T-cell therapy and post-chemotherapy have not been reported. In the present study, we found similar CIR, LFS and OS rates between our two cohorts. Among our 105 patients, there was only 11% CIR, which is lower than the CIR rate reported previously among ALL patients in CR (33-35). There are multiple potential reasons for the relatively lower CIR in our study. First, a haploidentical donor was the main donor source (67%) in our study. Mo et al. found that a haplo-donor was superior to a matched sibling donor in offsetting the detrimental effects of highrisk factors and pre-transplant MSD among ALL patients (13, 36). Second, TBI-based conditioning regimes were used in the majority of our patients (83%). TBI has demonstrated an advantage over Bu as a component of conditioning regimens for MSD, MUD, and haplo-HSCT in pediatric and adult patients with ALL (33-35, 37). Third, myeloablative conditioning regimens were used in all the patients in this study, which were more effective in eradicating residual leukemia disease.

Several studies have previously shown the prognostic relevance of disease status and MRD status among B-ALL patients (38, 39). In our multivariate analysis, we demonstrated that the CIR for MRD+ patients before transplant was 3 times as high as that of MRD- patients and a negative MRD status either after CAR-T therapy or after chemotherapy and prior to transplant predicted better results. We showed that MRD before transplant was an independent predicator for CIR after HSCT and that achieving an MRD-negative CR was crucial and equally important for optimal transplantation outcomes among both the chemotherapy and CAR-T therapy groups.

Among patients receiving CAR-T therapy, whether CR patients with an CD19-negative status before transplant will have an increased risk of relapse remains a concern and will require further investigation. The possible reason for a CD19-negative relapse could be due to a selective immune escape mechanism of the tumor cells (40). So far there is no evidence that CD19-negative leukemic clones may be more easily attained as a further escape from the graft-versus-leukemia (GVL) effect of donor cells. Allo-HSCT is an anti-HLA immunotherapy, which is independent from CD19. Excluding the one early treatment-related death within one month following transplantation, half (2/4) of the patients relapsed with CD19-negative clones in our CAR-T group. Due to the limited number of cases in our study, we cannot make any conclusions on

whether patients with a CD19-negative MRD status before transplant are more likely to relapse. Nevertheless, caution should be taken for those patients with CD19-negative MRD status who are to undergo an allo-HSCT.

In addition to CAR-T therapy, other immunotherapeutic approaches have proven successful in R/R B-ALL such as blimatumomab and inotuzomab. Currently, it remains unclear whether it is better to treat R/R B-ALL with CAR-T cell therapy vs.. blimatumomab or inotuzomab. Thus, it is important to conduct a randomized clinical trial in the future to investigate this question. A head-to-head comparison trial of blimatumomab or inotuzomab vs. CAR-T cell is undergoing (NCT03628053). Despite high CR/CRi of 67% achieved with inotuzomab for pediatric R/R ALL patients (41), 44% with blimatumomab (42), relapse remains the major problem. Without consolidative allo-HSCT, long-term disease control was limited with both blimatumomab and inotuzomab, especially for patients with high leukemic burden (43). The incidence of sinusoids obstruction syndrome (SOS), previously known as veno-occlusive disease, has been reported after allo-HSCT following inotuzomab, at an especially high rate (52%) in pediatric patients (41). In addition to efficacy and safety considerations, cost, insurance coverage, and local availability of each immunotherapy are all factors that will influence clinical decision-making. One limitation of our study is that it is not a randomized clinical trial but it is not feasible at the present time to do such randomized clinical trial as the CAR-T therapy is currently only indicated to chemotherapy refractory or relapsed patients. Nevertheless, our long-term follow-up and parallel comparison results demonstrate that pre-transplant CR induced by CAR-T therapy in R/R B-cell ALL patients carry the same prognostic significance as CR induced by conventional chemotherapy for patients without refractory disease. Although the CAR-T group had a higher incidence of Grade overall aGVHD and cGVHD, the incidence of serve aGVHD and cGVHD was comparable in both groups. Importantly, no clear increased transplant related mortality was identified in our CAR-T group. We conclude that the strategy of CAR-T therapy followed by allo-HSCT in R/R B cell-ALL was safe and effective, exhibiting similar long- term NRM, CIR, LFS and OS as those achieved among patients in the chemotherapy group.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/supplementary material. Further inquiries can be directed to the corresponding author.

AUTHOR CONTRIBUTIONS

Y-LZ did the work of data collecting, data analysis and paper writing. D-PL and PL are the director of Lu Daopei Hospital and

respond for the whole paper. Y-LZ, D-YL, R-JS, J-PZ, J-RZ, Z-JW, MX, X-YC, and YL take care of the all the HSCT patients. And J-fY and XZ took care of the patients during CAR-T therapy. All authors contributed to the article and approved the submitted version.

REFERENCES

- Fielding AK, Richards SM, Chopra R, Lazarus HM, Litzow MR, Buck G, et al. Outcome of 609 Adults After Relapse of Acute Lymphoblastic Leukemia (ALL); an MRC UKALL12/ECOG 2993 Study. Blood (2007) 109:944–50. doi: 10.1182/blood-2006-05-018192
- Nguyen K, Devidas M, Cheng SC, La M, Raetz EA, Carroll WL, et al. Factors Influencing Survival After Relapse From Acute Lymphoblastic Leukemia: A Children's Oncology Group Study. *Leukemia* (2008) 22:2142–50. doi: 10.1038/leu.2008.251
- Malempati S, Gaynon PS, Sather H, La MK, Stork LC. Outcome After Relapse Among Children With Standard-Risk Acute Lymphoblastic Leukemia: Children's Oncology Group Study CCG-1952. J Clin Oncol Off J Am Soc Clin Oncol (2007) 25:5800-7. doi: 10.1200/JCO.2007.10.7508
- Zhao Y, Tong W, Cao X-y, Xiong M, Zhang J, Wei Z, et al. Improved Outcomes of Haploidentical Blood and Marrow Transplantation in Hematologic Malignancies: A Single Center Study of 514 Cases. *Blood* (2015) 126:3224. doi: 10.1182/blood.V126.23.3224.3224
- D'Souza A, Fretham C, Lee SJ, Arora M, Brunner J, Chhabra S, et al. Current Use of and Trends in Hematopoietic Cell Transplantation in the United States. Biol Blood Marrow Transplant (2020) 26:e177–82. doi: 10.1016/ j.bbmt.2020.04.013
- Pan J, Yang JF, Deng BP, Zhao XJ, Zhang X, Lin YH, et al. High Efficacy and Safety of Low-Dose CD19-directed CAR-T Cell Therapy in 51 Refractory or Relapsed B Acute Lymphoblastic Leukemia Patients. *Leukemia* (2017) 31:2587–93. doi: 10.1038/leu.2017.145
- Maude SL, Frey N, Shaw PA, Aplenc R, Barrett DM, Bunin NJ, et al. Chimeric Antigen Receptor T Cells for Sustained Remissions in Leukemia. New Engl J Med (2014) 371:1507–17. doi: 10.1056/NEJMoa1407222
- 8. Davila ML, Riviere I, Wang X, Bartido S, Park J, Curran K, et al. Efficacy and Toxicity Management of 19-28z Car T Cell Therapy in B Cell Acute Lymphoblastic Leukemia. *Sci Trans Med* (2014) 6:224ra25. doi: 10.1126/scitranslmed.3008226
- Sommermeyer D, Hudecek M, Kosasih PL, Gogishvili T, Maloney DG, Turtle CJ, et al. Chimeric Antigen Receptor-Modified T Cells Derived From Defined CD8+ and CD4+ Subsets Confer Superior Antitumor Reactivity In Vivo. Leukemia (2016) 30:492–500. doi: 10.1038/leu.2015.247
- Zhang X, Lu X-A, Yang J, Zhang G, Li J, Song L, Su Y, et al. Efficacy and Safety of Anti-CD19 CAR T-Cell Therapy in 110 Patients With B-cell Acute Lymphoblastic Leukemia With High-Risk Features. *Blood Adv* (2020) 4:2325–38. doi: 10.1182/bloodadvances.2020001466
- Park JH, Rivière I, Gonen M, Wang X, Sénéchal B, Curran KJ, et al. Long-Term Follow-up of CD19 Car Therapy in Acute Lymphoblastic Leukemia. New Engl J Med (2018) 378:449–59. doi: 10.1056/NEJMoa1709919
- Maude SL, Laetsch TW, Buechner J, Rives S, Boyer M, Bittencourt H, et al. Tisagenlecleucel in Children and Young Adults With B-Cell Lymphoblastic Leukemia. New Engl J Med (2018) 378:439–48. doi: 10.1056/NEJMoa 1709866
- Rettinger E, Merker M, Salzmann-Manrique E, Kreyenberg H, Krenn T, Dürken M, et al. Pre-Emptive Immunotherapy for Clearance of Molecular Disease in Childhood Acute Lymphoblastic Leukemia After Transplantation. Biol Blood Marrow Transplant (2017) 23:87–95. doi: 10.1016/j.jbhmt.2016.10.006
- Hay KA, Gauthier J, Hirayama AV, Voutsinas JM, Wu Q, Li D, et al. Factors Associated With Durable EFS in Adult B-cell ALL Patients Achieving MRDnegative CR After CD19 Car T-cell Therapy. *Blood* (2019) 133:1652–63. doi: 10.1182/blood-2018-11-883710
- Lee DW, Kochenderfer JN, Stetler-Stevenson M, Cui YK, Delbrook C, Feldman SA, et al. T Cells Expressing CD19 Chimeric Antigen Receptors for Acute Lymphoblastic Leukaemia in Children and Young Adults: A Phase 1

ACKNOWLEDGMENTS

We thank Hong-xing Liu, Hui Wang, Tong Wang from Pathology & Laboratory Medicine Division in Hebei Yanda Lu Daopei Hospital for their technical assistance.

- Dose-Escalation Trial. Lancet (London England) (2015) 385:517–28. doi: 10.1016/S0140-6736(14)61403-3
- Lee M.S.-S. D.W., Yuan CM, Shah NN, Delbrook C, Yates B, Zhang H, et al. Long-Term Outcomes Following CD19 Car T Cell Therapy for B-ALL are Superior in Patients Receiving a Fludarabine/Cyclophosphamide Preparative Regimen and Post-CAR Hematopoietic Stem Cell Transplantation. *Blood* (2016) 126:218. doi: 10.1182/blood.V128.22.218.218
- Shadman M, Gauthier J, Hay KA, Voutsinas JM, Milano F, Li A, et al. Safety of Allogeneic Hematopoietic Cell Transplant in Adults After CD19-targeted Car Tcell Therapy. Blood Adv (2019) 3:3062–9. doi: 10.1182/bloodadvances.2019000593
- Bouziana S, Bouzianas D. Exploring the Dilemma of Allogeneic Hematopoietic Cell Transplantation After Chimeric Antigen Receptor T Cell Therapy: to Transplant or Not? Biol Blood Marrow Transplant J Am Soc Blood Marrow Transplant (2020) 26(8):e183–91. doi: 10.1016/j.bbmt.2020.04.003
- Lee DW, Gardner R, Porter DL, Louis CU, Ahmed N, Jensen M, et al. Current Concepts in the Diagnosis and Management of Cytokine Release Syndrome. Blood (2014) 124:188–95. doi: 10.1182/blood-2014-05-552729
- Hu Y, Sun J, Wu Z, Yu J, Cui Q, Pu C, et al. Predominant Cerebral Cytokine Release Syndrome in CD19-directed Chimeric Antigen Receptor-Modified T Cell Therapy. J Hematol Oncol (2016) 9:70. doi: 10.1186/s13045-016-0299-5
- Gust J, Hay KA, Hanafi LA, Li D, Myerson D, Gonzalez-Cuyar LF, et al. Endothelial Activation and Blood-Brain Barrier Disruption in Neurotoxicity After Adoptive Immunotherapy With CD19 Car-T Cells. Cancer Discovery (2017) 7:1404–19. doi: 10.1158/2159-8290.CD-17-0698
- Mei H, Jiang H, Wu Y, Guo T, Xia L, Jin R, et al. Neurological Toxicities and Coagulation Disorders in the Cytokine Release Syndrome During CAR-T Therapy. Br J Haematol (2018) 181:689–92. doi: 10.1111/bjh.14680
- Lu D-P, Dong L, Wu T, Huang X-J, Zhang M-J, Han W, et al. Conditioning Including Antithymocyte Globulin Followed by Unmanipulated HLAmismatched/haploidentical Blood and Marrow Transplantation can Achieve Comparable Outcomes With HLA-identical Sibling Transplantation. *Blood* (2006) 107:3065–73. doi: 10.1182/blood-2005-05-2146
- Przepiorka D, Weisdorf D, Martin P, Klingemann HG, Beatty P, Hows J, et al. 1994 Consensus Conference on Acute Gvhd Grading. *Bone Marrow Transplant* (1995) 15:825–8.
- Thomas ED, Storb R, Clift RA, Fefer A, Johnson L, Neiman PE, et al. Bone-Marrow Transplantation (Second of Two Parts). N Engl J Med (1975) 292:895–902. doi: 10.1056/NEJM197504242921706
- Deeg HJ, Storb R. Graft-Versus-Host Disease: Pathophysiological and Clinical Aspects. Annu Rev Med (1984) 35:11–24. doi: 10.1146/annurev.me. 35.020184.000303
- Filipovich AH, Weisdorf D, Pavletic S, Socie G, Wingard JR, Lee SJ, et al. National Institutes of Health Consensus Development Project on Criteria for Clinical Trials in Chronic Graft-Versus-Host Disease: I. Diag Staging Working Group Rep Biol Blood Marrow Transplant (2005) 11:945–56. doi: 10.1016/ j.bbmt.2005.09.004
- Jagasia MH, Greinix HT, Arora M, Williams KM, Wolff D, Cowen EW, et al. National Institutes of Health Consensus Development Project on Criteria for Clinical Trials in Chronic Graft-Versus-Host Disease: I. Diagnosis and Staging Working Group Report. Biol Blood Marrow Transplant J Am Soc Blood Marrow Transplant (2005) 11:945–56. doi: 10.1016/j.bbmt.2014.12.001
- Jodele S, Davies SM, Lane A, Khoury J, Dandoy C, Goebel J, et al. Diagnostic and Risk Criteria for HSCT-associated Thrombotic Microangiopathy: A Study in Children and Young Adults. *Blood* (2014) 124:645–53. doi: 10.1182/blood-2014-03-564997
- Brown PA, Shah B, Advani A, Aoun P, Barta SK, Bhatnagar B, et al. National Comprehensive Cancer Network (NCCN) Guidelines in Acute Lymphoblastic Leukemia, Version 1, Vol. 2018 (2018).
- 31. Jiang H, Li C, Yin P, Guo T, Liu L, Xia L, et al. Anti-CD19 Chimeric Antigen Receptor-Modified T-cell Therapy Bridging to Allogeneic Hematopoietic

Stem Cell Transplantation for Relapsed/Refractory B-cell Acute Lymphoblastic Leukemia: An Open-Label Pragmatic Clinical Trial. *Am J Hematol* (2019) 94:1113–22. doi: 10.1002/ajh.25582

- Huang XJ, Liu DH, Liu KY, Xu LP, Chen H, Han W, et al. Treatment of Acute Leukemia With Unmanipulated HLA-mismatched/haploidentical Blood and Bone Marrow Transplantation. *Biol Blood Marrow Transplant* (2009) 15 (2):257–65. doi: 10.1016/j.bbmt.2008.11.025
- Davies SM, Ramsay NK, Klein JP, Weisdorf DJ, Bolwell B, Cahn JY, et al. Comparison of Preparative Regimens in Transplants for Children With Acute Lymphoblastic Leukemia. J Clin Oncol Off J Am Soc Clin Oncol (2000) 18:340–7. doi: 10.1200/JCO.2000.18.2.340
- Kalaycio M, Bolwell B, Rybicki L, Absi A, Andresen S, Pohlman B, et al. BU- vs TBI-based Conditioning for Adult Patients With ALL. Bone Marrow Transplant (2011) 46:1413–7. doi: 10.1038/bmt.2010.314
- 35. Fu H, Xu L, Liu D, Liu K, Zhang X, Chen H, et al. Total Body Irradiation and Cyclophosphamide Plus Antithymocyte Globulin Regimen is Well Tolerated and Promotes Stable Engraftment as a Preparative Regimen Before T Cell-Replete Haploidentical Transplantation for Acute Leukemia. *Biol Blood Marrow Transplant* (2014) 20:1176–82. doi: 10.1016/j.bbmt.2014.04.012
- Mo XD, Xu LP, Zhang XH, Liu DH, Wang Y, Chen H, et al. Haploidentical Hematopoietic Stem Cell Transplantation in Adults With Philadelphianegative Acute Lymphoblastic Leukemia: No Difference in the High- and Low-Risk Groups. Int J Cancer (2015) 136:1697–707. doi: 10.1002/ijc.29146
- 37. Peters J-HDC, Locatelli F, Poetschger U, Pichler H, Sedlacek P, Buechner J, et al. TBI or Chemotherapy Based Conditioning for Children and AdolescentsWith ALL: A Prospective Randomized Muticenter-Study "Forum" on Behalf of the AIEOP-BFM-ALL-SG, Ibfm-Sg, Intreall-Sg AND Ebmt-Pd-Wp. EHA Library. Peters C. (2020) 294922; S102. Available at: https://library.ehaweb.org/eha/2020/eha25th/294922/c.peters.tbi.or. chemotherapy.based.conditioning.for.children.and.adolescents.html?f=listing %3D0%2Abrowseby%3D8%2Asortby%3D1%2Asearch%3Dtbi+or+chemotherapy+based+conditioning%2Bchildren%2Badolescents
- 38. Zhou Y, Slack R, Jorgensen JL, Wang SA, Rondon G, de Lima M, et al. The Effect of Peritransplant Minimal Residual Disease in Adults With Acute Lymphoblastic Leukemia Undergoing Allogeneic Hematopoietic Stem Cell

- Transplantation. Clin Lymphoma Myeloma Leuk (2014) 14:319–26. doi: 10.1016/j.clml.2014.01.002
- Gökbuget N, Kneba M, Raff T, Trautmann H, Bartram CR, Arnold R, et al. Adult Patients With Acute Lymphoblastic Leukemia and Molecular Failure Display a Poor Prognosis and are Candidates for Stem Cell Transplantation and Targeted Therapies. *Blood* (2012) 120:1868–76. doi: 10.1182/blood-2011-09.377713
- Maude SL, Rheingold SR, Aplenc R, Teachey DT, Callahan C, Baniewicz D, et al. Efficacy of Humanized Cd19-Targeted Chimeric Antigen Receptor (Car)-Modified T Cells in Children and Young Adults With Relapsed/ Refractory Acute Lymphoblastic Leukemia. *Blood* (2016) 128:217. doi: 10.1182/blood.V128.22.217.217
- Bhojwani D, Sposto R, Shah NN, Rodriguez V, Yuan C, Stetler-Stevenson M, et al. Inotuzumab Ozogamicin in Pediatric Patients With Relapsed/Refractory Acute Lymphoblastic Leukemia. *Leukemia* (2019) 33:884–92. doi: 10.1038/ s41375-018-0265-z
- Kantarjian H, Stein A, Gökbuget N, Fielding AK, Schuh AC, Ribera JM, et al. Blinatumomab Versus Chemotherapy for Advanced Acute Lymphoblastic Leukemia. New Engl J Med (2017) 376:836–47. doi: 10.1056/NEJMoa1609783
- Yurkiewicz IR, Muffly L, Liedtke M. Inotuzumab Ozogamicin: A CD22 mAbdrug Conjugate for Adult Relapsed or Refractory B-cell Precursor Acute Lymphoblastic Leukemia. *Drug Des Devel Ther* (2018) 12:2293–300. doi: 10.2147/DDDT.S150317

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2021 Zhao, Liu, Sun, Zhang, Zhou, Wei, Xiong, Cao, Lu, Yang, Zhang, Lu and Lu. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Advantages of publishing in Frontiers



OPEN ACCESS

Articles are free to reac for greatest visibility and readership



FAST PUBLICATION

Around 90 days from submission to decision



HIGH QUALITY PEER-REVIEW

Rigorous, collaborative, and constructive peer-review



TRANSPARENT PEER-REVIEW

Editors and reviewers acknowledged by name on published articles

Frontiers

Avenue du Tribunal-Fédéral 34 1005 Lausanne | Switzerland

Visit us: www.frontiersin.org

Contact us: frontiersin.org/about/contact



REPRODUCIBILITY OF RESEARCH

Support open data and methods to enhance research reproducibility



DIGITAL PUBLISHING

Articles designed for optimal readership across devices



FOLLOW US

@frontiersir



IMPACT METRICS

Advanced article metrics track visibility across digital media



EXTENSIVE PROMOTION

Marketing and promotion of impactful research



LOOP RESEARCH NETWORK

Our network increases your article's readership