

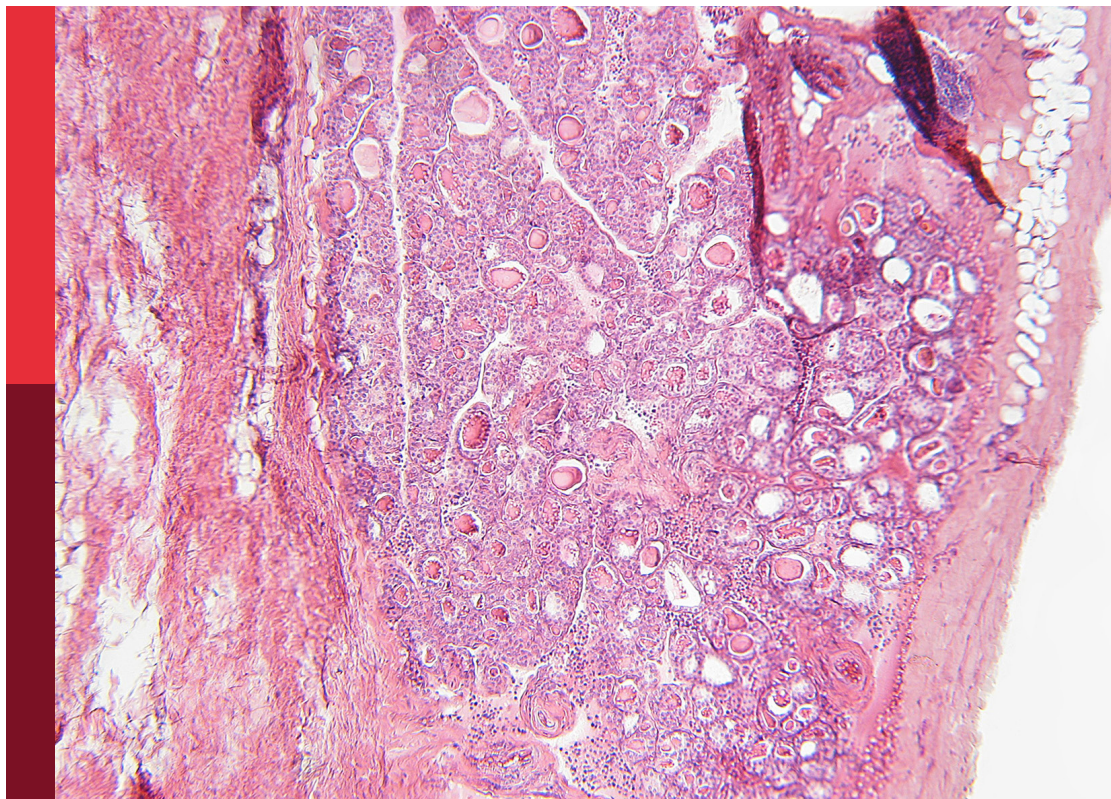
Recurrent pregnancy loss and endocrine dysfunction

Edited by

Hong Zhang, Hongbin Chi, Federico Jensen,
Lianghui Diao and Zitao Liu

Published in

Frontiers in Endocrinology



FRONTIERS EBOOK COPYRIGHT STATEMENT

The copyright in the text of individual articles in this ebook is the property of their respective authors or their respective institutions or funders. The copyright in graphics and images within each article may be subject to copyright of other parties. In both cases this is subject to a license granted to Frontiers.

The compilation of articles constituting this ebook is the property of Frontiers.

Each article within this ebook, and the ebook itself, are published under the most recent version of the Creative Commons CC-BY licence. The version current at the date of publication of this ebook is CC-BY 4.0. If the CC-BY licence is updated, the licence granted by Frontiers is automatically updated to the new version.

When exercising any right under the CC-BY licence, Frontiers must be attributed as the original publisher of the article or ebook, as applicable.

Authors have the responsibility of ensuring that any graphics or other materials which are the property of others may be included in the CC-BY licence, but this should be checked before relying on the CC-BY licence to reproduce those materials. Any copyright notices relating to those materials must be complied with.

Copyright and source acknowledgement notices may not be removed and must be displayed in any copy, derivative work or partial copy which includes the elements in question.

All copyright, and all rights therein, are protected by national and international copyright laws. The above represents a summary only. For further information please read Frontiers' Conditions for Website Use and Copyright Statement, and the applicable CC-BY licence.

ISSN 1664-8714
ISBN 978-2-8325-5190-5
DOI 10.3389/978-2-8325-5190-5

About Frontiers

Frontiers is more than just an open access publisher of scholarly articles: it is a pioneering approach to the world of academia, radically improving the way scholarly research is managed. The grand vision of Frontiers is a world where all people have an equal opportunity to seek, share and generate knowledge. Frontiers provides immediate and permanent online open access to all its publications, but this alone is not enough to realize our grand goals.

Frontiers journal series

The Frontiers journal series is a multi-tier and interdisciplinary set of open-access, online journals, promising a paradigm shift from the current review, selection and dissemination processes in academic publishing. All Frontiers journals are driven by researchers for researchers; therefore, they constitute a service to the scholarly community. At the same time, the *Frontiers journal series* operates on a revolutionary invention, the tiered publishing system, initially addressing specific communities of scholars, and gradually climbing up to broader public understanding, thus serving the interests of the lay society, too.

Dedication to quality

Each Frontiers article is a landmark of the highest quality, thanks to genuinely collaborative interactions between authors and review editors, who include some of the world's best academicians. Research must be certified by peers before entering a stream of knowledge that may eventually reach the public - and shape society; therefore, Frontiers only applies the most rigorous and unbiased reviews. Frontiers revolutionizes research publishing by freely delivering the most outstanding research, evaluated with no bias from both the academic and social point of view. By applying the most advanced information technologies, Frontiers is catapulting scholarly publishing into a new generation.

What are Frontiers Research Topics?

Frontiers Research Topics are very popular trademarks of the *Frontiers journals series*: they are collections of at least ten articles, all centered on a particular subject. With their unique mix of varied contributions from Original Research to Review Articles, Frontiers Research Topics unify the most influential researchers, the latest key findings and historical advances in a hot research area.

Find out more on how to host your own Frontiers Research Topic or contribute to one as an author by contacting the Frontiers editorial office: frontiersin.org/about/contact

Recurrent pregnancy loss and endocrine dysfunction

Topic editors

Hong Zhang — Second Affiliated Hospital of Soochow University, China

Hongbin Chi — Peking University Third Hospital, China

Federico Jensen — University of Buenos Aires, Argentina

Lianghui Diao — Shenzhen Zhongshan Obstetrics & Gynecology Hospital, China

Zitao Liu — New Hope Fertility Center, United States

Citation

Zhang, H., Chi, H., Jensen, F., Diao, L., Liu, Z., eds. (2024). *Recurrent pregnancy loss and endocrine dysfunction*. Lausanne: Frontiers Media SA.

doi: 10.3389/978-2-8325-5190-5

Table of contents

- 04 **Editorial: Recurrent pregnancy loss and endocrine dysfunction**
Hong Zhang and Lianghui Diao
- 06 **Metabonomic analysis of follicular fluid in patients with diminished ovarian reserve**
Jianan Li, Zhouhui Zhang, Yiqiu Wei, Pujia Zhu, Tailang Yin and Qiongqiong Wan
- 18 **Effects of ovarian stimulation protocols on outcomes of assisted reproductive technology in adenomyosis women: a retrospective cohort study**
Li Ge, Yexing Li, Shengnan Guan, Linlin Cui and Zi-Jiang Chen
- 28 **Association between female circulating heavy metal concentration and abortion: a systematic review and meta-analysis**
Meiqi Ren, Liantong Wang, Liqin Wen, Jinghua Chen, Song Quan and Xiao Shi
- 45 **Impact of thyroid-stimulating hormone levels after controlled ovarian hyperstimulation on *in vitro* fertilization/intracytoplasmic sperm injection outcomes in women with fresh embryo transfer: a prospective cohort study**
Ning Huang, Lixue Chen, Ying Lian, Hongbin Chi and Jie Qiao
- 52 **Sequential embryo transfer versus double cleavage-stage embryo or double blastocyst transfer in patients with recurrent implantation failure with frozen-thawed embryo transfer cycles: a cohort study**
Jiangman Gao, Yifeng Yuan, Jia Li, Tian Tian, Ying Lian, Ping Liu, Rong Li, Jie Qiao, Xiaoyu Long and Haiyan Wang
- 61 **Diagnostic workup of endocrine dysfunction in recurrent pregnancy loss: a cross-sectional study in Northeast China**
Liyang Zhang, Yushu Du, Jingshuang Zhou, Jiapo Li, Hongfei Shen, Yilin Liu, Chuanyang Liu and Chong Qiao
- 69 **Impact of the number of previous embryo implantation failures on IVF/ICSI-ET pregnancy outcomes in patients younger than 40 years: a retrospective cohort study**
Yuan Fang, Fan Jingjing, Cheng Tiantain, Xie Huanhuan and He Qiaohua
- 78 **Lupus and recurrent pregnancy loss: the role of female sex hormones and B cells**
Natalin Jimena Valeff, Maria Silvia Ventimiglia, Lianghui Diao and Federico Jensen
- 85 **Low follistatin level is a causal risk factor for spontaneous abortion: a two-sample mendelian randomization study**
Chen Gong, Wenzhi Yang, Xue Liu, Xinliang Li, Yutong Wang and Chan Tian



OPEN ACCESS

EDITED AND REVIEWED BY

Richard Ivell,
University of Nottingham, United Kingdom

*CORRESPONDENCE

Hong Zhang
✉ szzhanghong126@126.com

RECEIVED 23 June 2024

ACCEPTED 01 July 2024

PUBLISHED 09 July 2024

CITATION

Zhang H and Diao L (2024) Editorial:
Recurrent pregnancy loss and
endocrine dysfunction.
Front. Endocrinol. 15:1453336.
doi: 10.3389/fendo.2024.1453336

COPYRIGHT

© 2024 Zhang and Diao. This is an open-access article distributed under the terms of the [Creative Commons Attribution License \(CC BY\)](#). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Editorial: Recurrent pregnancy loss and endocrine dysfunction

Hong Zhang^{1*} and Lianghui Diao²

¹Department of Obstetrics and Gynecology, the Second Affiliated Hospital of Soochow University, Suzhou, China, ²Shenzhen Key Laboratory of Reproductive Immunology for Peri-implantation, Shenzhen Zhongshan Institute for Reproductive Medicine and Genetics, Shenzhen Zhongshan Obstetrics & Gynecology Hospital, Shenzhen, China

KEYWORDS

thyroid dysfunction, luteal phase deficiency, diabetes mellitus, insulin resistance, recurrent pregnancy loss, IVF/ICSI

Editorial on the Research Topic

Recurrent pregnancy loss and endocrine dysfunction

Recurrent pregnancy loss (RPL) is a distressing condition affecting a significant number of women globally, with complex etiologies that often remain elusive (1). It is estimated that approximately 8% to 12% of all cases of RPL are caused by endocrine diseases, which include thyroid dysfunction, luteal phase deficiency, hyperprolactinemia, diabetes mellitus, insulin resistance, polycystic ovarian syndrome (PCOS), and so on (2). The Research Topic “Recurrent Pregnancy Loss and Endocrine Dysfunction”, aims to deepen our understanding of how endocrine issues contribute to RPL and to identify potential therapeutic targets.

The manuscripts accepted for this Research Topic present a diverse range of studies, each providing valuable insights into different aspects of endocrine-related RPL.

The cross-sectional study by Zhang et al. highlights the high prevalence of endocrine dysfunctions, such as thyroid dysfunction, hyperprolactinemia, obesity, PCOS, and glucose abnormalities in women with RPL. They emphasize the importance of comprehensive endocrine evaluations of endocrine dysfunction in recurrent pregnancy loss, proposing that obesity may be a key endocrine factor among patients with two or more pregnancy losses, and suggesting screening of patients for endocrine-related etiology after two miscarriages.

Several studies focus on specific endocrine disorders and their impact on RPL.

Huang et al. investigates the effect of thyroid-stimulating hormone (TSH) levels post-controlled ovarian hyperstimulation on IVF/ICSI outcomes. It finds that while TSH levels do not significantly affect pregnancy rates, lower TSH levels are associated with higher preterm delivery rates.

In terms of treatment strategies, sequential embryo transfer could improve the clinical outcomes of patients with recurrent implantation failure (Gao et al.). Fang et al. found that the number of previous embryo implantation failures is an independent factor affecting implantation rate, clinical pregnancy rate, spontaneous abortion rate and live birth rate of patients underwent IVF/ICSI- ET.

Adenomyosis can induce heavy menstrual bleeding, chronic pelvic pain, infertility and RPL. Ge et al. explore the effects of ovarian stimulation protocols on assisted reproductive technology outcomes in women with adenomyosis, they recommended that an ultra-long or long protocol might be beneficial for fresh embryo transfer.

The link between lupus and RPL is explored in another study, which finds that women with lupus have a higher risk of pregnancy loss due to the autoimmune nature of the disease, adding to the complexity of managing RPL in these patients (Valeff et al.).

Ren et al. found that exposure to environmental pollutants is also a risk factor for RPL. Insulin resistance, vitamin D insufficiency, and abnormal thyroid and sex hormone concentrations might contribute to heavy metal-related abortion.

Low follistatin levels have also been identified as a potential risk factor for RPL. This study indicates that follistatin may play a protective role in maintaining pregnancy, highlighting another endocrine factor that could be crucial in managing RPL (Gong et al.).

Li et al. provide a metabolic perspective by analyzing follicular fluid in patients with diminished ovarian reserve (DOR), showing unique metabolic characteristics that could be associated with RPL.

Conclusion

In conclusion, these studies collectively enhance our understanding of the multifaceted relationship between endocrine dysfunction and RPL. They underscore the importance of comprehensive and individualized endocrine evaluations in women with RPL to improve clinical outcomes. The knowledge gained from this compilation of studies aims to reduce the burden of RPL on affected women and their families, ultimately leading to better clinical practices and improved reproductive health outcomes.

References

1. Quenby S, Gallos ID, Dhillon-Smith RK, Podsek M, Stephenson MD, Fisher J, et al. Miscarriage matters: the epidemiological, physical, psychological, and economic costs of early pregnancy loss. *Lancet*. (2021) 397:1658–67. doi: 10.1016/S0140-6736(21)00682-6
2. Pluchino N, Drakopoulos P, Wenger JM, Petignat P, Streuli I, Genazzani AR. Hormonal causes of recurrent pregnancy loss (RPL). *Hormones (Athens)*. (2014) 13:314–22. doi: 10.14310/horm.2002

We express our gratitude to all contributors for their invaluable research and to the reviewers for their critical evaluations. We hope this Research Topic will stimulate further research and enhance clinical practices in managing endocrine-related recurrent pregnancy loss.

Author contributions

HZ: Writing – review & editing. LD: Writing – original draft.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.



OPEN ACCESS

EDITED BY

Hong Zhang,
Second Affiliated Hospital of Soochow
University, China

REVIEWED BY

Lang Qin,
Sichuan University, China
Beihong Zheng,
Fujian Women and Children
Hospital, China

*CORRESPONDENCE

Tailang Yin

✉ reproductive@whu.edu.cn
Qiongqiong Wan
✉ wan.qq@whu.edu.cn

[†]These authors share first authorship

SPECIALTY SECTION

This article was submitted to
Reproduction,
a section of the journal
Frontiers in Endocrinology

RECEIVED 27 December 2022

ACCEPTED 06 February 2023

PUBLISHED 27 February 2023

CITATION

Li J, Zhang Z, Wei Y, Zhu P, Yin T and
Wan Q (2023) Metabonomic analysis of
follicular fluid in patients with diminished
ovarian reserve.
Front. Endocrinol. 14:1132621.
doi: 10.3389/fendo.2023.1132621

COPYRIGHT

© 2023 Li, Zhang, Wei, Zhu, Yin and Wan.
This is an open-access article distributed
under the terms of the [Creative Commons
Attribution License \(CC BY\)](#). The use,
distribution or reproduction in other
forums is permitted, provided the original
author(s) and the copyright owner(s) are
credited and that the original publication in
this journal is cited, in accordance with
accepted academic practice. No use,
distribution or reproduction is permitted
which does not comply with these terms.

Metabonomic analysis of follicular fluid in patients with diminished ovarian reserve

Jianan Li^{1†}, Zhouhui Zhang^{2†}, Yiqiu Wei¹, Pujia Zhu²,
Tailang Yin^{1*} and Qiongqiong Wan^{2*}

¹Reproductive Medicine Center, Renmin Hospital of Wuhan University, Wuhan, Hubei, China, ²The Institute for Advanced Studies, Wuhan University, Wuhan, Hubei, China

Background: Ovarian reserve is an important factor determining female reproductive potential. The number and quality of oocytes in patients with diminished ovarian reserve (DOR) are reduced, and even if *in vitro* fertilization-embryo transfer (IVF-ET) is used to assist their pregnancy, the clinical pregnancy rate and live birth rate are still low. Infertility caused by reduced ovarian reserve is still one of the most difficult clinical problems in the field of reproduction. Follicular fluid is the microenvironment for oocyte survival, and the metabolic characteristics of follicular fluid can be obtained by metabolomics technology. By analyzing the metabolic status of follicular fluid, we hope to find the metabolic factors that affect the quality of oocytes and find new diagnostic markers to provide clues for early detection and intervention of patients with DOR.

Methods: In this research, 26 infertile women with DOR and 28 volunteers with normal ovarian reserve receiving IVF/ET were recruited, and their follicular fluid samples were collected for a nontargeted metabonomic study. The orthogonal partial least squares discriminant analysis model was used to understand the separation trend of the two groups, KEGG was used to analyze the possible metabolic pathways involved in differential metabolites, and the random forest algorithm was used to establish the diagnostic model.

Results: 12 upregulated and 32 downregulated differential metabolites were detected by metabolic analysis, mainly including amino acids, indoles, nucleosides, organic acids, steroids, phospholipids, fatty acyls, and organic oxygen compounds. Through KEGG analysis, these metabolites were mainly involved in aminoacyl-tRNA biosynthesis, tryptophan metabolism, pantothenate and CoA biosynthesis, and purine metabolism. The AUC value of the diagnostic model based on the top 10 metabolites was 0.9936.

Conclusion: The follicular fluid of patients with DOR shows unique metabolic characteristics. These data can provide us with rich biochemical information and a research basis for exploring the pathogenesis of DOR and predicting ovarian reserve function.

KEYWORDS

diminished ovarian reserve, follicular fluid, metabonomics, oocytes, embryos, amino acids, steroids

1 Introduction

The ovarian reserve reflects the sum of follicles at different stages of development in the female ovary and the ability of these follicles to grow, develop and fertilize. With increasing age, follicles decrease gradually in number and function (1). Diminished ovarian reserve (DOR) refers to a decrease in the number and quality of oocytes, DOR is also currently diagnosed mainly by a decrease in basal sinus follicle count and antimüllerian hormone as well as elevated serum basal follicle stimulating hormone levels. DOR that occurs after the age of 40 is usually physiological, while women's early experience of DOR leads to an early decline in their reproductive function. The most common clinical manifestations of DOR are menstrual disorder, endocrine disorder, poor response to ovarian stimulation and infertility. If not treated in time, it is likely to progress to premature ovarian failure within a few years (2). The etiology of DOR is complex and mainly includes age, genetics, immunity, psychology, environment and other factors. In most patients, the main reasons are still difficult to determine. Recent studies have shown that even if IVF-ET is selected as a treatment means, the pregnancy rate of DOR patients is still reduced due to their low response to ovulation inducing drugs, and their live birth rate is significantly lower than that of women with normal ovarian reserve (3). In addition, patients with DOR have an increased incidence of hypertension during pregnancy (4) and an increased risk of recurrent abortion and aneuploid blastocysts after IVF (5, 6). Due to the unclear etiology and limited therapeutic effect of DOR, it is still one of the great challenges in the clinical treatment of infertility.

Follicular fluid (FF) is formed by the secretion of granulosa cells, membrane cells and oocytes and the diffusion of plasma components from capillaries to the follicular cavity. Its main components include hormones, growth factors, cytokines, proteins, steroids, amino acids and polysaccharides. As the microenvironment for the growth and development of follicles and oocytes, FF is the medium for the exchange of substances and energy between oocytes and the extracellular environment, and alterations in components in FF can reflect the metabolic level and developmental potential of oocytes (7). Studies also show that changes in follicular fluid metabolites, such as progesterone (8), phosphatidylcholine (9), and high-density lipoproteins (10), have an impact on fertilization and early embryonic development. Therefore, studying changes in metabolites in follicular fluid can reveal the factors that affect the development of oocytes and pregnancy outcome in DOR patients.

Metabonomics is a new omic technology developed after genomics and proteomics, and it can simultaneously qualitatively and quantitatively analyze all low molecular weight metabolites in the samples to be detected. Metabonomics reflects events downstream of gene expression and provides valuable information about cell metabolism. Common methods include nuclear magnetic resonance (NMR), gas chromatography–mass spectrometry (GC–MS), liquid chromatography–mass spectrometry (LC–MS), and capillary

electrophoresis mass spectrometry (CE–MS) (11). In recent years, the rapid development of metabonomics has also promoted the study of follicular fluid, which is used to assess the metabolic status of oocytes, study the pathogenesis of diseases and identify potential biomarkers (12). The researchers found that the FF showed different metabolic characteristics at different stages of follicular development (13), and the characteristics of FF metabolism in women of different ages were also different (14). In addition, changes in the metabolic map were also found in FF under the conditions of endometriosis (EMS), polycystic ovary syndrome (PCOS) and other diseases (15, 16). Several targeted metabonomic studies have revealed abnormalities in the metabolism of amino acids and oxidized lipids in the FF of DOR patients, and these differential metabolites are significantly related to the number of oocytes retrieved and embryo quality during IVF-ET, which indicates that the decline in oocyte quality in DOR patients may be involved in the impairment of energy utilization and the increase in oxidative stress. Differential metabolites may be used as biomarkers to predict ovarian reserve and embryonic development (17–19). At present, there are few studies on the nontargeted metabonomics of FF from patients with DOR. In this study, the researchers used nontargeted metabonomics technology (high-performance liquid chromatography–mass spectrometry) to expand the detection range of metabolites in the FF of DOR patients and screen potential biomarkers, which supplemented new data for studying the pathogenesis of DOR and the impact of metabolic changes on IVF-ET outcomes of DOR patients.

2 Materials and methods

2.1 Sample collection and preparation

This study is a prospective clinical experiment that has been approved by the Ethics Committee of Wuhan University People's Hospital (WDRY2018-K009), and all subjects signed an informed consent form. The subjects were all IVF or intracytoplasmic sperm injection (ICSI)–assisted pregnancy patients at Wuhan University People's Hospital from December 2020 to January 2022, and they were divided into an experimental group (DOR group, n=26) and a control group (CON, n=28) according to whether the ovarian reserve was normal.

The inclusion criteria of the DOR group were as follows: 1: 20–40 years old; 2. body mass index (BMI) ≤ 24 kg/m²; and 3. patients meeting the clinical diagnostic criteria of DOR (20): 1) AMH ≤ 1.1 ng/mL; 2) basic follicle stimulating hormone (bFSH) ≥ 10 IU/L, with or without FSH/luteinizing hormone (LH) ≥ 3.2 ; and 3) number of follicles in the unilateral basal sinus (bAFC) ≤ 7 . Any two of the above three items can be diagnosed as DOR. In the control group, patients with normal ovarian reserve were treated with IVF/ICSI only because of male infertility or oviduct factors (oviduct adhesion, excision, ligation, etc.). The inclusion criteria of the control group were as follows: 1, 20–35 years old; 2. BMI ≤ 24 kg/m²; 3, AMH ≥ 2 ng/mL, with or without bAFC ≥ 7 ; 4. bFSH < 10 IU/L, and estradiol (E2) < 80 pg/mL.

All subjects in this experiment should meet the following exclusion criteria: 1. History of ovarian surgery: cyst stripping, oophorectomy, etc.; 2. EMS, PCOS, ovarian cysts and other diseases that may affect the ovarian reserve function; 3. Received hormone therapy three months before the visit; 4. Any contraindication for ovulation induction therapy; 5. Other systemic abnormalities included hereditary diseases such as chromosome abnormalities, endocrine diseases such as hyperthyroidism and diabetes, infectious diseases such as hepatitis B, HIV and syphilis, and autoimmune diseases such as systemic lupus erythematosus.

2.2 Oocyte acquisition and follicular fluid collection

Subjects received an individualized ovarian stimulation protocol according to their ovarian reserve, age, weight and previous ovulation induction. In our reproductive center, the gonadotropin-releasing hormone antagonist protocol (GnRH-ant protocol) and ovarian stimulation protocol under progesterone (PPOS protocol) are used to help patients with DOR. The GnRH-ant protocol: patients begin daily gonadotropin (Gn) on days 2-3 of the menstrual cycle, and follicular development and serum hormone levels are monitored at the same time. When one follicle diameter reaches 12 mm or serum E2 > 300 pg/ml, patients should take GnRH ant 0.25/d; until three follicles have a diameter greater than 18 mm, and 60% of the follicles have a diameter of 16 mm or E2 has no significant increase or decline, they should be injected with HCG6000-10000 IU that night, and after 34-38 hours, they are punctured for oocytes under ultrasonic monitoring. The PPOS protocol is as follows: patients begin daily progesterone (8-10 mg) on days 2-5 of the menstrual cycle, and at the same time, they are injected with FSH or human menopausal gonadotropin (HMG) 150-300 IU/day. The amount of Gn was adjusted according to follicular growth. When one follicle diameter is greater than 17 mm, patients are injected with HCG 6000-10000 IU or GnRH-a 0.2 mg combined with HCG 2000 IU to induce ovulation. Oocytes were retrieved 34-38 h after ovulation induction. Patients who still have more than 2 small follicles (diameter ≤ 8 mm) after oocyte retrieval in the follicular phase, they should continue to take oral Gn, FSH or HMG (150-300 IU) in the luteal phase and adjust the Gn dose 2-3 days later according to follicular development. When at least 2-3 follicles had a diameter greater than 18 mm, and the serum E2 level reached an average of approximately 200 pg/ml for each dominant follicle, HCG (10000 IU) was injected. Oocytes were retrieved 34-38 h after injection.

Only follicular fluid from follicles larger than 18 mm in diameter was collected. The patient's oocytes were retrieved and placed in the incubator for 4-6 hours, and the follicular fluid was collected in Eppendorf tubes after merging. The collected follicular fluid was centrifuged at 4°C and 15000 rpm for 10 minutes, and the supernatant was collected and frozen at -80°C for the next step of detection.

2.3 Embryo quality assessment

The embryo quality was recorded on the third day after fertilization. The evaluation criteria for embryos are as follows: Grade I: the shape of blastomere is regular, the size is uniform, and there is no obvious DNA fragment; Grade II: the size of blastomere is slightly uneven, the shape is slightly irregular, and the fragments are not more than 20%; Grade III: blastomeres vary in size, fragments between 20% to 50%; Grade IV: blastomeres of different sizes, fragments ≥ 50%. High-quality embryos at cleavage stage are defined as follows: Day3 embryos are Class I-II, and the number of blastomeres is 6~9.

2.4 Mass spectrometry detection: sample preparation and spectrum collection

In this study, we used a timsTOF Pro mass spectrometer (Bruker Daltonics, Germany) in combination with an Ultimate 3000 liquid system (Thermo Scientific) for LC-MS analysis. MS data were obtained using a timsTOF Pro mass spectrometer with a TOP 3 data-dependent acquisition (DDA) method. The scanning range of the timsTOF Pro mass spectrometer was 20-1300 Da, and the collision energy of MS/MS was 20 eV. The ESI source conditions were set as follows: capillary voltage=3.6 kV; dry gas flow=10.0 L/min; Spray gas = 2.2 bar; drying temperature=220 °C. Before injecting each mode, we performed external calibration with sodium formate and injected 15 QC samples before FF samples to balance the instrument. During data collection, one QC sample was inserted for every 10 FF samples to monitor the consistency and stability of the whole operation process. The Ultimate 3000 liquid system was mainly used for chromatographic separation. To improve the quality and quantity of identification of hydrophilic and hydrophobic metabolites, two chromatographic separation methods, HILIC and RPLC, were both used for nontargeted metabolomic analysis. In HILIC mode, we used a Waters ACQUITY UPLC BEH amide column (2.1 mm * 100 mm, 1.7 μm), the column temperature was 40 °C, and the injection volume was 5 μL. The mobile phase consisted of A (H₂O containing 10 mM ammonium acetate and 0.1% acetic acid) and B (containing 95% ACN and 10 mM ammonium acetate and 0.1% acetic acid), with a constant flow rate of 0.5 mL/min. The chromatographic gradient program was as follows: 0 ~ 1 min, 100% B; 1 ~ 11 min, 60% B; 11~11.5 min, 40% B; 11.5 ~ 12 min, 40% B; 12 ~ 12.1 min, 100% B; 12.1 ~ 18 min, 100% B. In RPLC mode, we used Waters ACQUITY UPLC BEH C18 column (2.1mm * 100mm, 1.7 μm), the column temperature was maintained at 50°C, and the injection volume was 5μ. The mobile phase consisted of A (H₂O containing 0.1% formic acid) and B (containing 0.1% formic acid and ACN), with a constant flow rate of 0.4 mL/min. The chromatographic gradient program was as follows: 0 ~ 2 min, 2% B; 2 ~ 17 min, 100% B; 17 ~ 20 min, 100% B; 20 ~ 20.1 min, 2% B; 20.1 ~ 23 min, 2%.

2.5 Data processing and statistical analysis

MetaboScape4.0 software was used to process the original data with peak filtering (greater than 5000), calibration, feature matching and standardization to conduct comprehensive molecular feature extraction. After determining the mass to charge ratio and peak intensity of metabolites, we searched the Human metabolome Database (HMDB) and the database established by local standards to retrieve and identify the detected metabolites. Subsequently, the data of all metabolites were normalized on Metaboanalyst 5.0 (www.metaboanalyst.ca). We excluded features with CV values greater than 0.25, filled in the missing value (5.5%) with 1/5 of the positive minimum value, and normalized the data according to the sum. Benjamin Hochberg correction was applied to keep the risk of type I errors below 5%. Using a nonparametric test, we set the fold change (FC) > 1.2 and false discovery rate (FDR) < 0.05 to screen potential differential metabolites related to DOR. Unsupervised pattern recognition principal component analysis (PCA) and orthogonal partial least squares discriminant analysis (OPLS-DA) were also analyzed on SIMCA software after autoscaling. By reducing the dimensions of the obtained multidimensional data to form a matrix, OPLS-DA could show the separation trend between the two groups more clearly. In addition, the permutation test was used to verify the imitative effect and prediction ability of the OPLS-DA model. Metabolites with first principal component projection (VIP) > 1.0 and P value < 0.05 were considered to be significantly different between the two groups. Kyoto Encyclopedia of Genes and Genomes (KEGG) analysis was conducted to identify pathways that might be involved. Finally, a random forest algorithm was used to establish the diagnostic model.

The baseline characteristics of the subjects were analyzed, and the data are expressed as the mean \pm standard deviation (SD). SPSS 25.0 software was used to analyze the normality of the data. Variables with normal or near normal distributions were analyzed by Student's t test or the chi-square test, and data with nonnormal distribution were analyzed by the Kruskal–Wallis test. The Spearman correlation coefficient between differential metabolites and the ovarian function index was calculated using the psych package in R. Statistical significance was defined as $P < 0.05$.

3 Results

3.1 Clinical characteristics of patients with DOR

The patients' clinical characteristics are showed in [Table 1](#). The average age, BMI, years of infertility, fasting blood glucose (FBG) level, serum hormone level and IVF-ET outcome were compared between the DOR and CON groups. Age, bAFC, AMH, and bFSH were significantly different between the two groups ($P < 0.001$). BMI ($P=0.016$, $P<0.05$) and years of infertility ($P=0.021$, $P<0.05$) had a slight increase in the DOR group, while bLH, bE2, P and FBG had no differences ($P>0.05$). Analysis of the results of oocytes obtained and fertilization revealed that the number of oocytes retrieved, MII oocytes, 2PN fertilization and high-quality embryos on the third day in DOR patients were significantly lower than those in CON patients ($P<0.001$). This shows that the ovarian reserve of patients in DOR group is indeed reduced and our inclusion criteria are reliable. DOR is an age-related infertility, ovarian reserve and

TABLE 1 The demographic and clinical characteristics of patients with DOR and CON.

| Item | DOR group (n=26) | CON group (n=28) | P value |
|--------------------------------|-------------------|-------------------|-------------|
| Age (year) | 34.04 \pm 3.8 | 30.29 \pm 2.62 | < 0.001 *** |
| BMI (kg/m ²) | 21.57 \pm 1.43 | 20.48 \pm 1.74 | 0.016 * |
| bFSH (mIU/ml) | 11.11 \pm 4.71 | 6.60 \pm 1.75 | < 0.001 *** |
| bLH (mIU/ml) | 4.22 \pm 4.36 | 3.65 \pm 1.71 | 0.52 |
| bE2 (pg/ml) | 47.55 \pm 30.86 | 36.02 \pm 17.33 | 0.094 |
| P (ng/ml) | 1.49 \pm 2.77 | 0.78 \pm 1.03 | 0.21 |
| bAFC (n) | 3.5 \pm 1.61 | 10.18 \pm 3.35 | < 0.001 *** |
| AMH (ng/mL) | 0.72 \pm 0.36 | 4.70 \pm 2.49 | < 0.001 *** |
| FBG (mmol/L) | 4.78 \pm 0.45 | 5.02 \pm 0.31 | 0.474 |
| Duration of infertility (year) | 3.28 \pm 2.77 | 2.78 \pm 2.31 | 0.021 * |
| Oocytes retrieved (n) | 4.31 \pm 2.49 | 15.07 \pm 6.43 | < 0.001 *** |
| MI oocytes (n) | 3.12 \pm 1.77 | 10.21 \pm 4.83 | < 0.001 *** |
| 2PN Fertilizations (n) | 2.38 \pm 1.68 | 7.14 \pm 4.09 | < 0.001 *** |
| Day3 High-quality embryos (n) | 1.04 \pm 1.08 | 4.43 \pm 3.56 | < 0.001 *** |

DOR, Diminished ovarian reserve; NOR, Normal ovarian reserve; BMI, Body mass index; bFSH, basic follicle-stimulating hormone; bLH, basic luteinizing hormone; bE2, basic estrogen; P, progesterone; bAFC, basic antral follicle count; AMH, anti-Müllerian hormone; FBG, fasting blood glucose * $P \leq 0.05$ ** $P \leq 0.005$ *** $P \leq 0.001$.

oocyte quality decline with advancing age, the difference in age between the DOR group and the control group in the results may be explained by the fact that age is to some extent an etiological or synergistic factor in DOR. To make the study more rigorous, future studies should expand the sample size and distinguish between different age groups before comparing metabolic levels

3.2 Multivariate analysis of metabolites

In this experiment, we detected metabolites in follicular fluid under HILIC positive and negative ion modes and RPLC positive and negative ion modes. A total of 2897 variable features were detected under HILIC mode, and 3419 features were detected under

RPLC mode; a total of 994 metabolites were identified. The Wayne diagram in [Supplementary Figure 1 \(Figure S1\)](#) shows that the metabolites detected in the four modes are very different, which proves that detection in different modes can obtain more information about metabolites.

To compare the overall difference in the metabolic spectrum between DOR patients and the control group, PCA and OPLS-DA discriminant models were established based on the metabolite data obtained under four modes. The OPLS-DA discriminant model reveals that the DOR group and control group can be well separated, as shown in [Figures 1A, C, E, G](#), the DOR group is all clustered on the right side, the control group is all clustered on the left side, and the values of R²_Y and Q²_Y are close to 1, which indicates that the difference between the two groups is significant

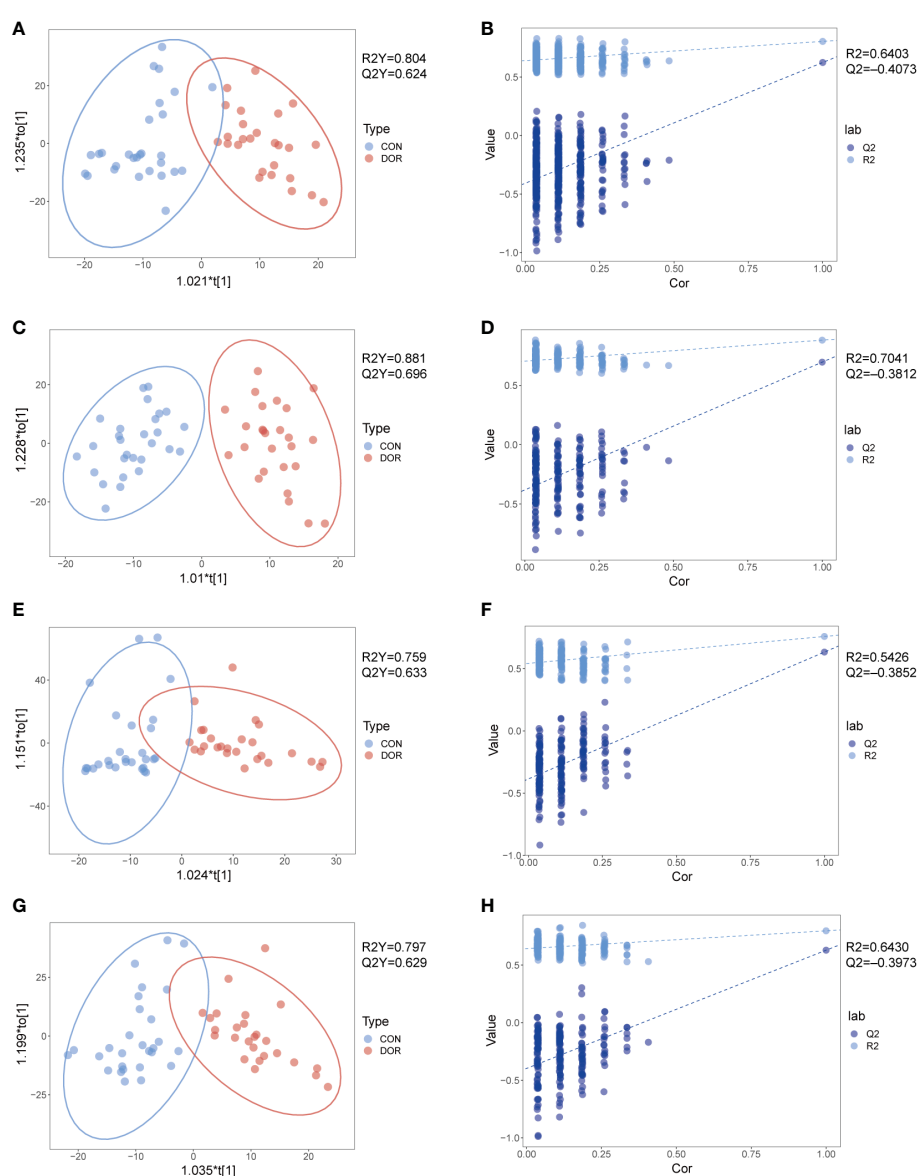


FIGURE 1

Metabolomic analysis of follicular fluid from patients with diminished ovarian reserved ($n = 26$, green dots) and healthy controls ($n = 28$, red dots) under HILIC and RPLC mode. OPLS-DA score plots for positive ionization mode (A, E) and negative ionization mode (C, G), the OPLS-DA model's permutation test for positive ionization mode (B, F) and negative ionization mode (D, H). (A-D): HILIC mode; e-h: RPLC mode.

and the prediction ability of the OPLS-DA model is good. As shown in Figures 1B, D, F, H, the permutation test shows the value of Q² and R² under the four modes, further verifying that the OPLS-DA model has a good imitative effect and prediction ability. In addition, the PCA models also show that the DOR group and control group are distributed in different regions (Figure S2), but the two groups are not completely separated in the PCA model, which indicates that the samples have individual differences. In Figure S2, the quality control samples (QC) are gathered together, which shows that the stability and repeatability of the detection system is satisfactory, and the OPLS-DA is reliable.

3.3 Identification of differential metabolites and analysis of related pathways

Univariate analysis was carried out on all features detected in follicular fluid, and a volcano plot is shown in Figures 2A–D, downregulated features in DOR samples were clustered on the left, and upregulated features were clustered on the right. A total of 12 upregulated and 32 downregulated metabolites were identified from all differential features. Differential metabolites mainly include amino acids, indoles, nucleosides, organic acids, steroids, phospholipids, fatty acyls and organic oxygen compounds. The VIP value of these differential metabolites in OPLS-DA was greater than 1, and $P < 0.05$ in univariate analysis (Supplementary Tables 1, 2), which also indicated that they were related to ovarian reserve. In addition, the heatmap also intuitively shows the change in metabolites in DOR patients (Figure 2E).

According to KEGG analysis, the differential metabolites are mainly involved in aminoacyl-tRNA biosynthesis, tryptophan metabolism, pantothenate and CoA biosynthesis, and purine metabolism (marked in Figure 3). These metabolic pathways play an important role in maintaining the normal function of cells and tissues, and the results add new data for exploring the pathogenesis and clinical treatment of DOR.

3.4 Correlation analysis between differential metabolites and clinical indicators

In this experiment, Spearman correlation analysis was used to study the relationship between differential metabolites and clinical detection indicators in FF. Ten metabolites with correlation coefficients of $R > 0.6$ and $P < 0.05$ are shown in Figure 4A. We found that pregnanediol-3-glucuronide, N-acetyl-D-tryptophan, L-aspartic acid and indole-3-carboxaldehyde were significantly positively correlated with AMH, bAFC, number of oocytes retrieved, MII oocytes, and 2PN fertilizations, while almost all metabolites did not correlate with BMI, bLH, bP, bE2, and duration of infertility ($R < 0.3$, not shown in the figure). In addition, for further study on how the metabolic profile changed in the FF of DOR patients, the correlation between metabolites is shown in Figure 4B.

3.5 Establishment of a diagnostic model based on 10 metabolites

To intervene earlier and more individually in the development of DOR, we used a random forest algorithm to distinguish the DOR groups and the CON groups and finally selected the top 10 metabolites (Figure 5A) to be included in the diagnostic model. Using a receiver operating characteristic (ROC) curve to evaluate the diagnostic performance of this model, as shown in Figure 5B, the AUC reached a very high value of 0.9936. When all the metabolites were chosen for diagnostic models, the AUC value (AUC=0.9936) was similar to that of the top 10 metabolites. Although the top 20 metabolites had the highest AUC values (AUC=0.9952) when included in the diagnostic model, it was not much higher than 0.9936, so we finally determined the top 10 metabolites for the sake of efficient clinical diagnosis. In addition, unsupervised cluster analysis showed that the top 10 metabolites could distinguish all DOR patients from normal subjects (Figure 5C). In summary, these 10 metabolites can be used as promising tools to predict ovarian function.

4 Discussion

In this study, we used nontargeted metabonomics to detect the follicular fluid of 54 subjects, and found that there were 12 upregulated and 32 downregulated metabolites in the FF of DOR patients compared with normal women. These metabolites were mainly involved in aminoacyl-tRNA biosynthesis, tryptophan metabolism, pantothenate and CoA biosynthesis, purine metabolism, which are closely related to the number of oocytes and embryo quality. In the following, we discuss in detail the possible mechanisms of differential metabolites.

As an essential amino acid, tryptophan has three metabolic pathways *in vivo*: 1) tryptophan is hydroxylated and decarboxylated to form serotonin and melatonin; 2) tryptophan generates nicotinic acid, nicotinamide adenine dinucleotide (NAD) and nicotinamide adenine dinucleotide phosphate (NADP) through karinurine metabolism; and 3) tryptophan can generate indole, indole pyruvic acid, indole lactic acid, etc., through deamination and decarboxylation. Tryptophan and its metabolites participate in many physiological processes, such as maintaining cell growth and regulating immune function. In recent years, research on tryptophan metabolism in the reproductive field has made some progress. A large number of studies have shown that serotonin, as one of the main metabolites of tryptophan, plays an important role in regulating placental function and fetal development (21). Serotonin can affect the development of oocytes by regulating progesterone secretion by granulosa cells. Animal experiments have also shown that a decrease in serotonin in mouse blood leads to damage to early embryo development (22). Serotonin is the main source of melatonin. The latest evidence showed that human ovarian granulosa cells cultured *in vitro* can express melatonin, and melatonin reduces oxidative stress by improving the mitochondrial function of oocytes (23). In addition, adding an appropriate amount of melatonin during embryo culture improves

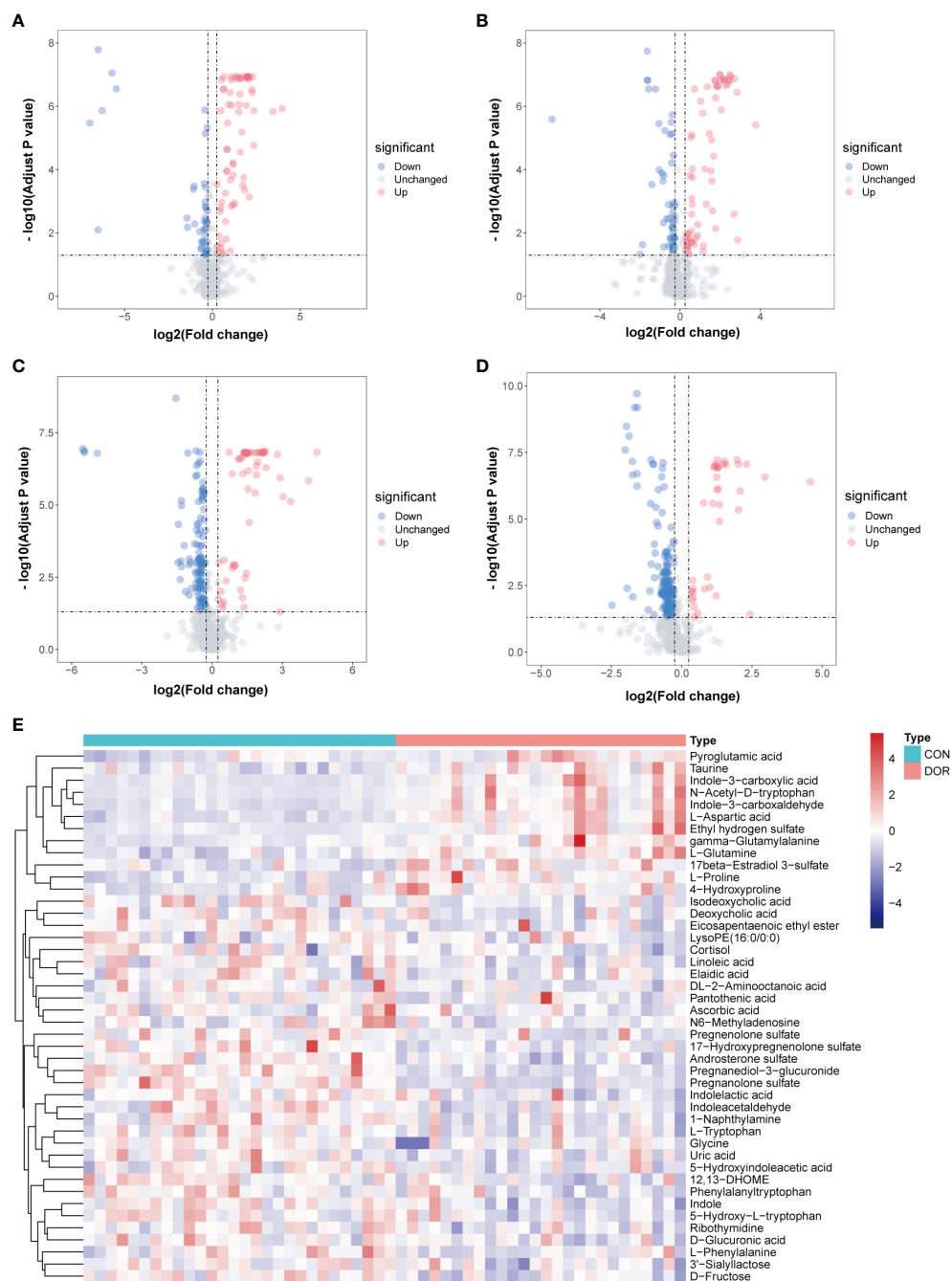
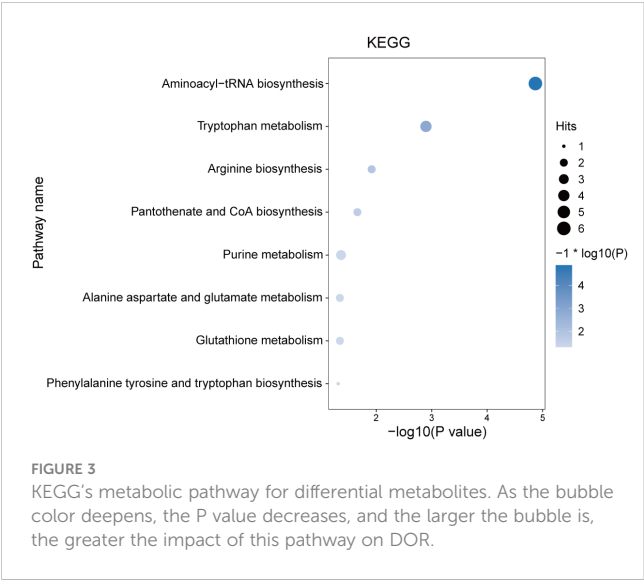


FIGURE 2

Volcanic maps under different detection modes show the distribution of different features. The blue dot represents the up regulated features while the red dot represents the down regulated. (A): positive ionization mode in HILIC; (B): negative ionization mode in HILIC; (C): positive ionization mode in RPLC; (D): negative ionization mode in RPLC; (E): Heatmap based on 44 metabolites, unsupervised cluster analysis showed that differential metabolites could distinguish DOR patients from healthy subjects.

the rate of high-quality embryos in patients with repeated low-quality embryos on the third day (24), and the quality of embryos and clinical results of patients with repeated cycles after the failure of IVF/ICSI can also be improved (25). Only a small amount of tryptophan is metabolized through the indoleacetic acid pathway, and the role of metabolites generated from this pathway in oocyte and embryo development is still unclear. In this experiment, we detected that tryptophan and its decomposition products 5-

hydroxy-L-tryptophan, 5-hydroxyindoleacetic acid (the metabolic end product of serotonin), indole, indoleacetaldehyde, and indoleacetic acid in the FF of DOR patients decreased, and were positively correlated with the number of oocytes, fertilized embryos, and high-quality embryos. The results of this study indicate that the decrease in tryptophan and its decomposition products, especially in the ovary, will affect the quantity and quality of oocytes, and may be detrimental to the early development of embryos. In addition,



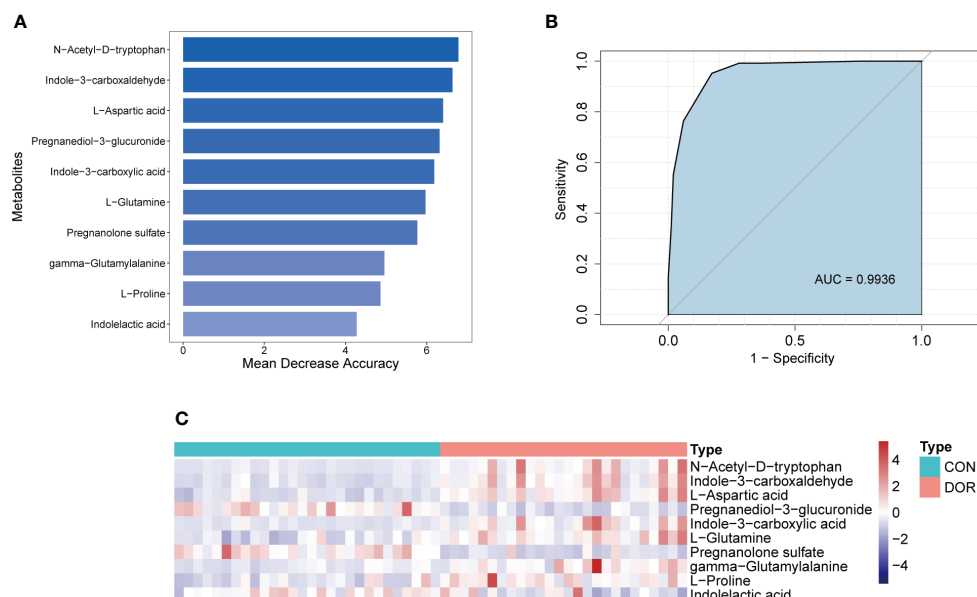


FIGURE 5

Establishment and test of diagnostic model based on 10 metabolites. (A). Top 10 metabolites selected according to random forest algorithm; (B). ROC curve and AUC value of the combined diagnostic model of the top 10 metabolites; (C). Unsupervised cluster analysis of the top 10 metabolites.

pathway (30). Maternal L-proline supplementation could also improve mouse placental development and fetal survival (31). L-glutamine is another important energy substrate that plays a role in the development of preimplantation embryos (32, 33). However, how these two amino acids affect oocyte development is still unclear. We detected changes in amino acids and their derivatives in the FF of DOR patients, and it has been shown that amino acids promote oocyte maturation and early embryonic development by reducing oxidative stress and enhancing mitochondrial function. However, whether amino acids participate in glucose metabolism to regulate cell energy utilization and what kind of synergistic or antagonistic effects they have in DOR patients need further study.

Steroids are another group of metabolites that are significantly altered in the FF of DOR patients. We found that androsterone sulfate, pregnenolone sulfate, pregnanolone sulfate, and 17-hydroxypregnenolone sulfate were decreased; taurine, glucuronic acid and pregnenol-3-glucuronic acid which are closely related to the excretion of gonadal hormones were also downregulated; while 17beta-Estradiol 3-sulfate (E2-3S) was upregulated. Pregnenolone sulfate, 17-hydroxypregnenolone sulfate and androsterone sulfate are the starting points for the synthesis of progesterone, estrogen and cortisol (34). Progesterone is mainly converted into pregnenol-3-glucuronic acid, and estrogen is mainly excreted in the form of sulfate and glucuronic acid. Our results seemed to indicate that compared with normal women, estrogen and progesterone in the FF of DOR patients are decreased. Yang et al. found that genes regulating cholesterol synthesis and transport, such as SCAP, FDFT1, CYP51A1, SRB1 and STARD1 were significantly downregulated in granulosa cells of DOR patients, and serum estradiol and progesterone were significantly lower in DOR groups (35), which complemented our findings. Dehydroepiandrosterone (DHEA) is mainly derived from 17-

hydroxypregnenolone sulfate, and study have proved that DHEA treatment improved the clinical outcome of DOR patients receiving IVF-ET (36). DHEA improves the quality of oocytes by increasing the oxidative phosphorylation of mitochondria and reducing the apoptosis of cumulus cells (37, 38). During oocyte activation, DHEA at the normal threshold could significantly improve the fertilization rate and live birth rate (39). Our study found that the downregulation of 17-hydroxypregnenolone sulfate was related to the number of oocytes retrieved, MII oocytes and high-quality embryos, which also implied that 17-hydroxypregnenolone sulfate and DHEA played an important role in the growth and development of oocytes and embryos. At present, the research on metabolites in the steroid group is not perfect, and the metabolic map obtained may not be complete, so more experiments are needed to further understand how the steroid group affects oocyte and embryo development.

The energy required for oocyte growth and meiosis mainly depends on the oxidative phosphorylation of mitochondria. However, the ability of oocytes to ingest and consume glucose is limited, the pyruvate needed for oxidative phosphorylation is mostly supplied by the glycolysis pathway of cumulus cells and granulosa cells (40), and increasing glycolysis in granulosa cells also promotes the activation of primordial follicles (41). In our study, D-fructose, pantothenic acid and 3-sialyllactose were downregulated, which seemed to point to the decreased energy metabolism of cumulus cells and oocytes in DOR patients. Pantothenic acid is necessary for coenzyme A (CoA), which is a cofactor of many enzymes. CoA participates not only in the metabolism of sugar, fat and protein but also in the antioxidation of the body. This study also detected that ascorbic acid was reduced in FF, and the bioactive form of ascorbic acid is vitamin C, which is considered an antioxidant. The results indicated that the follicular

microenvironment in patients with DOR seemed to be damaged by oxidative stress, thus adversely affecting the growth and maturation of oocytes.

Some fatty acids, such as linoleic acid, eicosapentaenoic ethyl ester, elaidic acid and 12,13-DHOME, were also downregulated, and they were significantly positively correlated with the number of oocytes retrieved, fertilized and high-quality embryos. It has been proved that defects in the synthesis of polyunsaturated fatty acids such as linoleic acid and linolenic acid lead to follicular arrest, oocyte atresia and infertility in female mice (42). Fatty acids are important energy source, mainly providing ATP for oocytes through β -Oxidation, and inhibition of β -Oxidation lead to meiosis arrest of oocytes and development failure after fertilization (43, 44). The decrease in fatty acids leads to DNA damage in granulosa cells and then accelerates cell apoptosis, which further reduces the quality of oocytes and embryos (45, 46). We know that the imbalance of oxidation/antioxidation is an important reason for DOR (47). There is evidence that arachidonic acid metabolism disorder is closely related to the reduction of ovarian reserve (18), and fatty acids and their derivatives participate in the regulation of cellular oxidative stress as antioxidants (48), which may be one of their roles in DOR.

In the FF of DOR patients, we also detected abnormal nucleotide metabolism, including decreased ribothymidine, N6-methyladenosine (m6A) and uric acid (UA). UA is the final product of human purine metabolism, and its physiological level can act as an antioxidant to reduce the damage of oxygen radicals and nitrite to cells (49). PCOS is closely involved in inflammation and oxidative stress, and some patients have obviously increased serum uric acid, which is related to the severity of the disease (50). Although no research has shown the role of UA in DOR, it can also be reasonably speculated that low UA levels in FF may cause increased oxidative stress and mitochondrial dysfunction and thus affect the growth and maturation of oocytes. N6-methyladenosine is one of the most common and abundant RNA modifications; it affects the stability, splicing and/or translation of modified RNA and therefore plays an important role in posttranscriptional regulation (51). A recent study found that in granulosa cells of aging ovaries, there was more m6A involved in the modification of mRNA, which damaged the normal process of mRNA degradation and accelerated the aging of granulosa cells (52). Mu. et al. also confirmed that m6A is indispensable in all stages of mouse follicular development, and oocytes lacking m6A cannot complete meiosis and form oosperms (53). With an increasing number of studies on m6A in granulosa cells and oocytes, m6A may become a new target for studying the pathogenesis and treatment of DOR.

5 Innovation and limitations

In the metabonomic study on the follicular fluid of patients with DOR, we used nontargeted metabonomic technology for the first time, obtained a more informative metabolic profile and established a diagnostic model based on metabolites. Our results provide data support for the study of the pathogenesis of DOR

and the search for new diagnostic markers. However, our study still has some limitations. First, the sample size of the experimental groups is less than 30, which is only one small sample study. After expanding the sample, more metabolites with apparent differences may be obtained. Second, after obtaining the differential metabolites, we did not conduct cell experiments or animal experiments for validation or further study. Third, in subsequent research, targeted metabonomic studies could be carried out according to the metabolites obtained, which may be helpful to improve the metabolic pathway of DOR. Finally, our research is limited to metabonomics, and combining our results with genomics, transcriptomics and proteomics will hopefully promote research on the mechanism of ovarian reserve reduction.

Data availability statement

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found in the article/[Supplementary Material](#).

Ethics statement

This study is a prospective clinical experiment that has been approved by the Ethics Committee of Wuhan University People's Hospital (WDRY2018-K009), and all subjects signed an informed consent form. The patients/participants provided their written informed consent to participate in this study.

Author contributions

JL, YW and TY conceived the original ideas. JL and YW collected samples and clinical data. JL, ZZ, YW and PZ conducted the metabolomics analysis. ZZ performed the statistical analysis. JL interpreted the data. JL and ZZ cowrote the manuscript. TY, SMC and QW supervised and revised the manuscript. All authors contributed to the article and approved the submitted version.

Funding

This work was supported by the following grants: the National Key Research and Development Program of China (No. 2021YFC2700700), and the Provincial Natural Science Foundation of China (No. 2022CFB200).

Acknowledgments

We would like to acknowledge all the patients included in our study and the medical staff for their contribution to this work.

Although the author Yan Zhang (YZ) does not appear in the author list of this study, she also provided some suggestions for this study and is gratefully acknowledged here.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

- Wallace WH, Kelsey TW. Human ovarian reserve from conception to the menopause. *PLoS One* (2010) 5(1):e8772. doi: 10.1371/journal.pone.0008772
- Lu Q, Shen H, Li Y, Zhang C, Wang C, Chen X, et al. Low testosterone levels in women with diminished ovarian reserve impair embryo implantation rate: a retrospective case-control study. *J Assist Reprod Genet* (2014) 31(4):485–91. doi: 10.1007/s10815-014-0186-3
- Hu S, Xu B, Jin L. Perinatal outcome in young patients with diminished ovarian reserve undergoing assisted reproductive technology. *Fertil Steril* (2020) 114(1):118–124 e1. doi: 10.1016/j.fertnstert.2020.02.112
- Han S, Zhai Y, Guo Q, Qin Y, Liu P. Maternal and neonatal complications in patients with diminished ovarian reserve in *In-vitro* Fertilization/Intracytoplasmic sperm injection cycles. *Front Endocrinol (Lausanne)* (2021) 12:648287. doi: 10.3389/fendo.2021.648287
- Bunnewell SJ, Honess ER, Karia AM, Keay SD, Al Wattar BH, Quenby S. Diminished ovarian reserve in recurrent pregnancy loss: a systematic review and meta-analysis. *Fertil Steril* (2020) 113(4):818–827 e3. doi: 10.1016/j.fertnstert.2019.11.014
- Jaswa EG, McCulloch CE, Simbulan R, Cedars MI, Rosen MP. Diminished ovarian reserve is associated with reduced euploid rates *via* preimplantation genetic testing for aneuploidy independently from age: evidence for concomitant reduction in oocyte quality with quantity. *Fertil Steril* (2021) 115(4):966–73. doi: 10.1016/j.fertnstert.2020.10.051
- Dumesic DA, Meldrum DR, Katz-Jaffe MG, Krisher RL, Schoolcraft WB. Oocyte environment: follicular fluid and cumulus cells are critical for oocyte health. *Fertil Steril* (2015) 103(2):303–16. doi: 10.1016/j.fertnstert.2014.11.015
- Nagy B, Poto L, Farkas N, Koppan M, Varnagy A, Kovacs K, et al. Follicular fluid progesterone concentration is associated with fertilization outcome after IVF: a systematic review and meta-analysis. *Reprod BioMed Online* (2019) 38(6):871–82. doi: 10.1016/j.rbmo.2018.12.045
- Wang J, Zheng W, Zhang S, Yan K, Jin M, Hu H, et al. An increase of phosphatidylcholines in follicular fluid implies attenuation of embryo quality on day 3 post-fertilization. *BMC Biol* (2021) 19(1):200. doi: 10.1186/s12915-021-01118-w
- Jia C, Nagy RA, Homminga I, Hoek A, Tietge UJF. The anti-inflammatory function of follicular fluid HDL and outcome of modified natural cycle *in vitro* fertilization. *Biol Reprod* (2020) 103(1):7–9. doi: 10.1093/biolre/i0aa061
- Wilson ID, Theodoridis G, Virgiliou C. A perspective on the standards describing mass spectrometry-based metabolic phenotyping (metabolomics/metabonomics) studies in publications. *J Chromatogr B Anal Technol BioMed Life Sci* (2021) 1164:122515. doi: 10.1016/j.jchromb.2020.122515
- Sun Z, Chang HM, Wang A, Song J, Zhang X, Guo J, et al. Identification of potential metabolic biomarkers of polycystic ovary syndrome in follicular fluid by SWATH mass spectrometry. *Reprod Biol Endocrinol* (2019) 17(1):45. doi: 10.1186/s12958-019-0490-y
- Yang J, Feng T, Li S, Zhang X, Qian Y. Human follicular fluid shows diverse metabolic profiles at different follicle developmental stages. *Reprod Biol Endocrinol* (2020) 18(1):74. doi: 10.1186/s12958-020-00631-x
- Zhang X, Wang T, Song J, Deng J, Sun Z. Study on follicular fluid metabolomics components at different ages based on lipid metabolism. *Reprod Biol Endocrinol* (2020) 18(1):42. doi: 10.1186/s12958-020-00599-8
- Liu L, Yin TL, Chen Y, Li Y, Yin L, Ding J, et al. Follicular dynamics of glycerophospholipid and sphingolipid metabolisms in polycystic ovary syndrome patients. *J Steroid Biochem Mol Biol* (2019) 185:142–9. doi: 10.1016/j.jsbmb.2018.08.008
- Karaer A, Tuncay G, Mumcu A, Dogan B. Metabolomics analysis of follicular fluid in women with ovarian endometriosis undergoing *in vitro* fertilization. *Syst Biol Reprod Med* (2019) 65(1):39–47. doi: 10.1080/19396368.2018.1478469
- de la Barca JMC, Boueillh T, Simard G, Boucret L, Ferre-L'Hottelier V, Tessier L, et al. Targeted metabolomics reveals reduced levels of polyunsaturated choline plasmalogens and a smaller dimethylarginine/arginine ratio in the follicular fluid of patients with a diminished ovarian reserve. *Hum Reprod* (2017) 32(11):2269–78. doi: 10.1093/humrep/dex303
- Liang C, Zhang X, Qi C, Hu H, Zhang Q, Zhu X, et al. UHPLC-MS-MS analysis of oxylipins metabolomics components of follicular fluid in infertile individuals with diminished ovarian reserve. *Reprod Biol Endocrinol* (2021) 19(1):143. doi: 10.1186/s12958-021-00825-x
- Al Rashid K, Taylor A, Lumsden MA, Goulding N, Lawlor DA, Nelson SM. Association of the functional ovarian reserve with serum metabolomic profiling by nuclear magnetic resonance spectroscopy: a cross-sectional study of ~ 400 women. *BMC Med* (2020) 18(1):247. doi: 10.1186/s12916-020-01700-z
- Practice Committee of the American Society for Reproductive M. Testing and interpreting measures of ovarian reserve: a committee opinion. *Fertil Steril* (2015) 103(3):e9–e17. doi: 10.1016/j.fertnstert.2014.12.093
- Bakovic P, Kesic M, Peric M, Becceheli I, Horvaticek M, George M, et al. Differential serotonin uptake mechanisms at the human maternal-fetal interface. *Int J Mol Sci* (2021) 22(15):7807. doi: 10.3390/ijms22157807
- Dube F, Amireault P. Local serotonergic signaling in mammalian follicles, oocytes and early embryos. *Life Sci* (2007) 81(25-26):1627–37. doi: 10.1016/j.lfs.2007.09.034

Supplementary material

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fendo.2023.1132621/full#supplementary-material>

SUPPLEMENTARY FIGURE 1

Number of metabolites detected in different modes. HP: HILIC positive ionization mode; HN: HILIC negative ionization mode; RP: RPLC positive ionization mode; RN: RPLC negative ionization mode.

SUPPLEMENTARY FIGURE 2

PCA analysis and evaluation of quality control samples in different modes. a-b: HILIC (+) mode; c-d: HILIC (-); e-f: RPLC (+); g-h: RPLC (-).

SUPPLEMENTARY FIGURE 3

Correlation between differential metabolites.

SUPPLEMENTARY TABLE 1

Down-regulated metabolites in FF samples from DOR groups

SUPPLEMENTARY TABLE 2

Up-regulated metabolites in FF samples from DOR groups

SUPPLEMENTARY TABLE 3

Diagnostic models based on top5, top10, top 20 and total differential metabolites

23. Liu YJ, Ji DM, Liu ZB, Wang TJ, Xie FF, Zhang ZG, et al. Melatonin maintains mitochondrial membrane potential and decreases excessive intracellular Ca(2+) levels in immature human oocytes. *Life Sci* (2019) 235:116810. doi: 10.1016/j.lfs.2019.116810
24. Bao Z, Li G, Wang R, Xue S, Zeng Y, Deng S. Melatonin improves quality of repeated-poor and frozen-thawed embryos in human, a prospective clinical trial. *Front Endocrinol (Lausanne)* (2022) 13:853999. doi: 10.3389/fendo.2022.853999
25. Zhu Q, Wang K, Zhang C, Chen B, Zou H, Zou W, et al. Effect of melatonin on the clinical outcome of patients with repeated cycles after failed cycles of *in vitro* fertilization and intracytoplasmic sperm injection. *Zygote* (2022) 30(4):471–9. doi: 10.1017/S0967199421000770
26. Broekhuizen M, Danser AHJ, Reiss IKM, Merkus D. The function of the kynurenine pathway in the placenta: A novel pharmacotherapeutic target? *Int J Environ Res Public Health* (2021) 18(21):11545. doi: 10.3390/ijerph182111545
27. Wang S, Mu L, Zhang C, Long X, Zhang Y, Li R, et al. Abnormal activation of tryptophan-kynurenine pathway in women with polycystic ovary syndrome. *Front Endocrinol (Lausanne)* (2022) 13:877807. doi: 10.3389/fendo.2022.877807
28. Li S, Guo Q, Wang YM, Li ZY, Kang JD, Yin XJ, et al. Glycine treatment enhances developmental potential of porcine oocytes and early embryos by inhibiting apoptosis. *J Anim Sci* (2018) 96(6):2427–37. doi: 10.1093/jas/sky154
29. Tscherner AK, Macaulay AD, Ortman CS, Baltz JM. Initiation of cell volume regulation and unique cell volume regulatory mechanisms in mammalian oocytes and embryos. *J Cell Physiol* (2021) 236(10):7117–33. doi: 10.1002/jcp.30352
30. Liu N, Yang Y, Si X, Jia H, Zhang Y, Jiang D, et al. L-proline activates mammalian target of rapamycin complex 1 and modulates redox environment in porcine trophectoderm cells. *Biomolecules* (2021) 11(5):742. doi: 10.3390/biom11050742
31. Liu N, Dai Z, Zhang Y, Chen J, Yang Y, Wu G, et al. Maternal l-proline supplementation enhances fetal survival, placental development, and nutrient transport in micedagger. *Biol Reprod* (2019) 100(4):1073–81. doi: 10.1093/biolre/iy240
32. Chen PR, Redel BK, Spate LD, Ji T, Salazar SR, Prather RS. Glutamine supplementation enhances development of *in vitro*-produced porcine embryos and increases leucine consumption from the medium. *Biol Reprod* (2018) 99(5):938–48. doi: 10.1093/biolre/iy129
33. Chen PR, Lucas CG, Spate LD, Prather RS. Glutaminolysis is involved in the activation of mTORC1 in *in vitro*-produced porcine embryos. *Mol Reprod Dev* (2021) 88(7):490–9. doi: 10.1002/mrd.23516
34. Hana V, Jezkova J, Kosak M, Krsek M, Hana V, Hill M. Novel GC-MS/MS technique reveals a complex steroid fingerprint of subclinical hypercortisolism in adrenal incidentalomas. *J Clin Endocrinol Metab* (2019) 104(8):3545–56. doi: 10.1210/je.2018-01926
35. Yang X, Zhao Z, Fan Q, Li H, Zhao L, Liu C, et al. Cholesterol metabolism is decreased in patients with diminished ovarian reserve. *Reprod BioMed Online* (2022) 44(1):185–92. doi: 10.1016/j.rbmo.2021.09.013
36. Li J, Yuan H, Chen Y, Wu H, Wu H, Li L. A meta-analysis of dehydroepiandrosterone supplementation among women with diminished ovarian reserve undergoing *in vitro* fertilization or intracytoplasmic sperm injection. *Int J Gynaecol Obstet* (2015) 131(3):240–5. doi: 10.1016/j.ijgo.2015.06.028
37. Li CJ, Lin LT, Tsui KH. Dehydroepiandrosterone shifts energy metabolism to increase mitochondrial biogenesis in female fertility with advancing age. *Nutrients* (2021) 13(7):2449. doi: 10.3390/nu13072449
38. Lin LT, Wang PH, Wen ZH, Li CJ, Chen SN, Tsai EM, et al. The application of dehydroepiandrosterone on improving mitochondrial function and reducing apoptosis of cumulus cells in poor ovarian responders. *Int J Med Sci* (2017) 14(6):585–94. doi: 10.7150/ijms.18706
39. Chimote BN, Chimote NM. Correction to: Dehydroepiandrosterone sulphate (DHEAS) concentrations stringently regulate fertilization, embryo development and IVF outcomes: are we looking at a potentially compelling 'oocyte-related factor' in oocyte activation? *J Assist Reprod Genet* (2021) 38(8):2223. doi: 10.1007/s10815-021-02215-z
40. Richani D, Dunning KR, Thompson JG, Gilchrist RB. Metabolic co-dependence of the oocyte and cumulus cells: essential role in determining oocyte developmental competence. *Hum Reprod Update* (2021) 27(1):27–47. doi: 10.1093/humupd/dmaa043
41. Zhang X, Zhang W, Wang Z, Zheng N, Yuan F, Li B, et al. Enhanced glycolysis in granulosa cells promotes the activation of primordial follicles through mTOR signaling. *Cell Death Dis* (2022) 13(1):87. doi: 10.1038/s41419-022-04541-1
42. Stoffel W, Schmidt-Soltan I, Binczek E, Thomas A, Thevis M, Wegner I. Dietary omega3-and omega6-polyunsaturated fatty acids reconstitute fertility of juvenile and adult Fads2-deficient mice. *Mol Metab* (2020) 36:100974. doi: 10.1016/j.molmet.2020.100974
43. Downs SM, Mosey JL, Klinger J. Fatty acid oxidation and meiotic resumption in mouse oocytes. *Mol Reprod Dev* (2009) 76(9):844–53. doi: 10.1002/mrd.21047
44. Sturmey RG, O'Toole PJ, Leese HJ. Fluorescence resonance energy transfer analysis of mitochondrial lipid association in the porcine oocyte. *Reproduction* (2006) 132(6):829–37. doi: 10.1530/REP-06-0073
45. de Barros TT, Venancio VP, Hernandez LC, Gregg Antunes LM, Hillesheim E, Salomao RG, et al. DNA Damage is inversely associated to blood levels of DHA and EPA fatty acids in Brazilian children and adolescents. *Food Funct* (2020) 11(6):5115–21. doi: 10.1039/c9fo02551k
46. Zhao Z, Fan Q, Zhu Q, He R, Li Y, Liu C, et al. Decreased fatty acids induced granulosa cell apoptosis in patients with diminished ovarian reserve. *J Assist Reprod Genet* (2022) 39(5):1105–14. doi: 10.1007/s10815-022-02462-8
47. Park SU, Walsh L, Berkowitz KM. Mechanisms of ovarian aging. *Reproduction* (2021) 162(2):R19–33. doi: 10.1530/REP-21-0022
48. Djuricic I, Calder PC. Beneficial outcomes of omega-6 and omega-3 polyunsaturated fatty acids on human health: An update for 2021. *Nutrients* (2021) 13(7):2421. doi: 10.3390/nu13072421
49. Wang Q, Wen X, Kong J. Recent progress on uric acid detection: A review. *Crit Rev Anal Chem* (2020) 50(4):359–75. doi: 10.1080/10408347.2019.1637711
50. Mu L, Pan J, Yang L, Chen Q, Chen Y, Teng Y, et al. Association between the prevalence of hyperuricemia and reproductive hormones in polycystic ovary syndrome. *Reprod Biol Endocrinol* (2018) 16(1):104. doi: 10.1186/s12958-018-0419-x
51. Huang W, Chen TQ, Fang K, Zeng ZC, Ye H, Chen YQ. N6-methyladenosine methyltransferases: functions, regulation, and clinical potential. *J Hematol Oncol* (2021) 14(1):117. doi: 10.1186/s13045-021-01129-8
52. Jiang ZX, Wang YN, Li ZY, Dai ZH, He Y, Chu K, et al. The m6A mRNA demethylase FTO in granulosa cells retards FOS-dependent ovarian aging. *Cell Death Dis* (2021) 12(8):744. doi: 10.1038/s41419-021-04016-9
53. Mu H, Zhang T, Yang Y, Zhang D, Gao J, Li J, et al. METTL3-mediated mRNA N(6)-methyladenosine is required for oocyte and follicle development in mice. *Cell Death Dis* (2021) 12(11):989. doi: 10.1038/s41419-021-04272-9



OPEN ACCESS

EDITED BY

Lianghui Diao,
Shenzhen Zhongshan Urology
Hospital, China

REVIEWED BY

Yanqiu Hu,
Nanjing Medical University, China
Mosammat Begum,
Infertility Care and Research Centre (ICRC),
Bangladesh

*CORRESPONDENCE

Linlin Cui

✉ fdclear3@126.com

†These authors have contributed
equally to this work and share
last authorship

RECEIVED 02 April 2023

ACCEPTED 26 July 2023

PUBLISHED 17 August 2023

CITATION

Ge L, Li Y, Guan S, Cui L and Chen Z-J
(2023) Effects of ovarian stimulation
protocols on outcomes of assisted
reproductive technology in adenomyosis
women: a retrospective cohort study.
Front. Endocrinol. 14:1198779.
doi: 10.3389/fendo.2023.1198779

COPYRIGHT

© 2023 Ge, Li, Guan, Cui and Chen. This is
an open-access article distributed under the
terms of the [Creative Commons Attribution
License \(CC BY\)](#). The use, distribution or
reproduction in other forums is permitted,
provided the original author(s) and the
copyright owner(s) are credited and that
the original publication in this journal is
cited, in accordance with accepted
academic practice. No use, distribution or
reproduction is permitted which does not
comply with these terms.

Effects of ovarian stimulation protocols on outcomes of assisted reproductive technology in adenomyosis women: a retrospective cohort study

Li Ge¹, Yexing Li^{2,3,4,5,6,7,8}, Shengnan Guan^{2,3,4,5,6,7,8},
Linlin Cui^{1,2,3,4,5,6,7,8*†} and Zi-Jiang Chen^{2,3,4,5,6,7,8,9,10†}

¹Center for Reproductive Medicine, the Second Hospital of Shandong University, Cheeloo College of Medicine, Shandong University, Jinan, Shandong, China, ²Center for Reproductive Medicine, Cheeloo College of Medicine, Shandong University, Jinan, Shandong, China, ³Research Unit of Gametogenesis and Health of Assisted Reproductive Technology (ART)-Offspring, Chinese Academy of Medical Sciences (No.2021RU001), Jinan, Shandong, China, ⁴Key Laboratory of Reproductive Endocrinology of Ministry of Education, Shandong University, Jinan, Shandong, China, ⁵Shandong Key Laboratory of Reproductive Medicine, Shandong Provincial Hospital Affiliated to Shandong First Medical University, Jinan, China, ⁶Shandong Provincial Clinical Research Center for Reproductive Health, Jinan, Shandong, China, ⁷Shandong Technology Innovation Center for Reproductive Health, Jinan, Shandong, China, ⁸National Research Center for Assisted Reproductive Technology and Reproductive Genetics, Shandong University, Jinan, Shandong, China, ⁹Shanghai Key Laboratory for Assisted Reproduction and Reproductive Genetics, Shanghai, China, ¹⁰Center for Reproductive Medicine, Ren Ji Hospital, School of Medicine, Shanghai Jiao Tong University, Shanghai, China

Objective: To evaluate the effects of different ovarian stimulation protocols on in vitro fertilization/intracytoplasmic sperm injection (IVF/ICSI) outcomes in infertile women with adenomyosis.

Methods: We carried out a retrospective cohort study among infertile women with adenomyosis receiving IVF/ICSI treatment, including 257 fresh embryo transfer (ET) cycles and 305 frozen embryo transfer (FET) cycles. In fresh ET cycles, ultra-long, long, short, and antagonist protocols were adopted. In FET cycles, patients received long-acting GnRH agonist (GnRHa) pretreatment or not. The primary outcome was clinical pregnancy rate (CPR), and the secondary outcomes included implantation rate (IR), miscarriage rate (MR), and live birth rate (LBR).

Results: In fresh ET cycles, compared with ultra-long and long protocols, IR (49.7%, 52.1% versus 28.2%, $P=0.001$) and CPR (64.3%, 57.4% versus 35.6%, $P=0.004$) significantly decreased in the short protocol. Similarly, compared with ultra-long and long protocols, a decreased inclination of IR (49.7%, 52.1% versus 33.3%) and CPR (57.4%, 64.3% versus 38.2%) existed in the antagonist protocol, although no statistical significance was detected because of strict P adjustment of Bonferroni method ($P_{adj}=0.008$). Compared with long protocol, LBR in short protocol decreased obviously (48.2% versus 20.3%, $P<0.001$). In FET cycles, no matter which origin of embryos, there were no statistical differences in IR, CPR, and LBR. For women ≥ 35 years receiving fresh ET, CPR was higher in ultra-long and long protocols (52.1%, 50.0% versus 20.0%, 27.5%, $P=0.031$) compared to antagonist and short protocols. For women ≥ 35 years receiving

FET, compared with ultra-long and antagonist protocols, cycles with embryos originating from long and short protocols had higher proportions of long-acting GnRHa pretreatment (30.4%, 30.00 versus 63.9%, 51.4%, $P=0.009$). IR (61.1%, 48.6% versus 32.6%, 25.0%, $P=0.020$) and CPR (58.3%, 48.6% versus 30.4%, 25.0%, $P=0.024$) in long and short protocols were higher than rates of ultra-long and antagonist protocols, but no statistical differences were supported because of strict Bonferroni method ($P_{adj}=0.008$).

Conclusion: In infertile women with adenomyosis, if a fresh embryo was planned for transfer, an ultra-long or long protocol might be beneficial. If antagonist and short protocols were used, whole embryos frozen followed by FET was recommended. In FET cycles, embryos derived from different protocols had no impact on pregnancy outcomes.

KEYWORDS

adenomyosis, infertility, *in vitro* fertilization (IVF), intracytoplasmic sperm injection (ICSI), pregnancy outcomes

Introduction

As a continuing conundrum, adenomyosis has besieged clinicians for more than one hundred years, which is manifested in the displacement of endometrial glands and stroma in the myometrium (1). Adenomyosis can induce a series of clinical problems, such as heavy menstrual bleeding, chronic pelvic pain, and infertility (2–4). As time passes, the lesions gradually exacerbate, and eventually result in infertility or other severe impacts. A cross-sectional study showed that the incidence of adenomyosis was 20%–29.7% in the infertile population (5). In infertile women receiving assisted reproductive technology (ART), the proportion could rise to 30%–40% (6).

The negative effect of adenomyosis on ART outcomes was accumulated, and persistent endeavors were made to improve the pregnancy outcomes (6–8). So far, the usage of ultra-long protocol was more extensive because of possible improvement in clinical pregnancy rate (CPR) or live birth rate (LBR) (9–12). A widely accepted mechanism was that downregulation induced by long-acting GnRHa could counter hyperestrogenism and progesterone resistance of adenomyosis (13–15). However, the ultra-long protocol had an obvious defect, that was, deep inhibition of ovarian function, which usually resulted in the increase of the duration and dosage of gonadotropin (10). More seriously, for adenomyosis patients with poorer ovarian reserve, the inhibition of long-acting GnRHa could induce poor ovarian response, manifesting in decreased oocyte retrieval and negative pregnancy outcomes. Besides ultra-long protocol, conventional protocols, such as long, antagonist, and short protocols, all could be adopted, however, systematic evaluation of these protocols was absent. In frozen embryo transfer (FET) cycles, if embryos originated from different

protocols, did any differences in pregnancy outcomes exist? There were no answers.

So, we designed this study and tried to systematically evaluate the pregnancy outcomes of different protocols in fresh embryo transfer (ET) cycles. Additionally, we also aimed to elucidate whether embryo deriving from different protocols could affect the outcomes of FET cycles.

Materials and methods

Study design and population

Patients with adenomyosis who underwent *in vitro* fertilization/ intracytoplasmic sperm injection (IVF/ICSI) at the Center for Reproductive Medicine, Shandong University from January 2016 to December 2020 were included in the retrospective cohort study. The follow-up time was up to April 2023. The study has been reviewed and approved by Ethics Committee at the Center for Reproductive Medicine, Shandong University (No. 2021-133). The inclusion criteria were as follows: (i) age below 42 years at the start of the IVF/ICSI cycles; (ii) diagnosis of adenomyosis based on a consensus opinion from the Morphological Uterus Sonographic Assessment (MUSA) group with subjective enlargement of the uterine corpus and heterogeneous myometrium, accompanying with or without asymmetrical thickening, cysts, hyperechoic islands, fan-shaped shadowing, echogenic subendometrial lines and buds, translesional vascularity, irregular junctional zone and interrupted junctional zone or not (16, 17); (iii) no history of uterine malformation, untreated hydrosalpinx or intrauterine lesions. All scans were performed by experienced imaging doctors, who had

over 8 years of experience in gynecological practice. The baseline data of scans were recorded on the electronic medical system.

Controlled ovarian stimulation protocols during IVF/ICSI

Pretreatment

Long acting GnRHa downregulation was an important pretreatment method with duration varying from 1 to 6 cycles according to the response of patients to GnRHa, which was explained in detail in the ultra-long protocol. Surgery and anti-inflammatory drugs were not used as pretreatment.

Ultra-long protocol

to receive the first injection of long-acting GnRHa (Triptorelin Acetate®; Ipsen, France; Leuprorelin Acetate®; Lizhu, China) with a dose of 3.75 mg on day 2 or 3 of the menstrual cycle. The anteroposterior diameter of the uterus was measured 28 days after each injection. If the diameter was more than 70 mm, another injection was repeated until the sixth injection. Twenty-eight days after the last dose, the effect of downregulation was evaluated by transvaginal ultrasonography (TVS) and serum hormone examination. Eligible downregulation was defined as endometrium thickness ≤ 5 mm, serum estradiol ≤ 50 pg/mL, LH ≤ 5 IU/L, and diameters of follicles < 8 mm in bilateral ovary without functional cysts.

Long protocol

A daily dose of triptorelin acetate (0.05–0.1 mg) of GnRHa (triptorelin acetate®; Ipsen, France) for 14 days. If downregulation criteria were achieved, ovarian stimulation started.

Short protocol

A daily dose (0.05–0.1 mg) of GnRHa (triptorelin acetate®; Ipsen, France) was given on days 2–4 of the menstrual cycle until HCG trigger. After 1–2 days of GnRHa, gonadotropin was used for ovarian stimulation and lasted for about 8–12 days.

Antagonist protocol

A daily dose (0.25 mg) of GnRH-ant (Orgalutran®, MSD, Netherlands) was used on the fifth or sixth day of gonadotropin until the trigger day.

Ovarian stimulation

Gonadotropin (Gonal F®, Merck Serono, Switzerland; Lishenbao®, Lizhu, China) at 150–300 IU daily was administered for COS according to age, body mass index (BMI), and ovarian reserve. The adjustment of gonadotropin and addition of recombinant LH (Luveris®, Merck Serono, Germany) were decided by follicular development. Routinely, 8000–10000 IU of urinary human chorionic gonadotropin (HCG) (hCG®; Livzon, China) was used intramuscularly for triggering when at least two

follicles measured ≥ 18 mm. If a high risk of ovarian hyperstimulation syndrome (OHSS) existed, 2000–4000 IU of HCG combined with 0.2 mg GnRHa was administrated. Oocyte retrieval was performed 34–36 h later. The choice of IVF or ICSI depended on sperm quality. Cancellation of fresh ET and whole embryo frozen was carried out in the conditions of high risk of OHSS, hydrosalpinx, or unsynchronized endometrium status. All embryos were cultured for 3 or 5 days with two high-quality cleavage-stage embryos on day 3 or one blastocyst on day 5 transferring into the uterus under the guidance of abdominal ultrasound. The high-quality embryos were defined as 2PN-derived embryos with 7–10 cells and scores ≥ 3 on day 3 or ≥ 4 BC on day 5 (18). Oral dydrogesterone tablet (Duphaston®, Abbott, Netherlands) 10 mg twice daily and vaginal progesterone gel (Crinone gel®, Merck Serono, Switzerland) 90 mg once daily or oral dydrogesterone tablet (Duphaston®, Abbott, Netherlands) 20 mg twice daily and vaginal progesterone soft capsules (Utrogestan®, Besins, Belgium) 200 mg once daily were administered as luteal phase support.

Protocols of endometrial preparation in FET cycles

GnRHa pretreatment before FET: GnRHa pretreatment was performed in patients with severer adenomyosis before FET for ≥ 1 month with 3.75 mg GnRHa per month. The uterine anteroposterior diameter was measured 28 days after each injection. If it was more than 70 mm, injection of the same dose of GnRHa was repeated until the sixth injection. Hormone replacement cycles would be administered 4 weeks after the last dose of GnRHa.

Oral estradiol valerate (Progynova®, Bayer, Germany) was administrated in a dose-escalating method, 4 mg/day for the first 5 days and subsequently 6 mg/day for the second 5 days. According to the assessment of endometrial thickness and serum hormone levels of E_2 , 8 mg/day for another 3–4 days was continued or not. When endometrial thickness ≥ 7 mm, progesterone addition was started. One frozen-thawed blastocyst was transferred into the uterus 5 days after progesterone addition. Other protocols for endometrial preparation in FET cycles had been described in previous studies (19).

Observational parameters and outcome variables

Baseline parameters included age, BMI, duration of infertility, primary infertility, antral follicle count (AFC), basal follicle-stimulating hormone (FSH), anti-Müllerian hormone (AMH), the mean diameter of the initial uterus, history of dysmenorrhea, and IVF-related parameters. The mean diameter of the uterus was calculated as the average of long and wide diameter in the longitudinal section measured by TVS on days 2–6 of the menstrual cycle. According to the verbal multidimensional scoring system (20), dysmenorrhea was classified into none, mild,

moderate, and severe. The primary pregnancy outcome was CPR, and the secondary pregnancy outcomes included implantation rate (IR), biochemical pregnancy rate (BPR), ectopic pregnancy rate (EPR), miscarriage rate (MR), early MR, late MR, and LBR, cumulative live birth rate (CLBR). Serum β -hCG level was examined 14 days after ET. Biochemical pregnancy as defined as elevated serum β -hCG level ≥ 10 mIU/mL. Clinical pregnancy was defined as the presence of intra-uterine pregnancy or visible extra-uterine pregnancy. Ectopic pregnancy was defined as the presence of a gestational sac or mass outside the uterine cavity. Miscarriage was defined as pregnancy loss before 28 gestational weeks. Clinical pregnancy loss ≤ 12 gestational weeks was an early miscarriage, vice versa, pregnancy loss >12 gestational weeks was a late miscarriage. IR was defined as the ratio of the total number of gestational sacs confirmed by TVS to the total number of transferred embryos. CLBR was defined as the number of cycles in which patients had the first live birth/number of all transfer cycles per oocyte retrieval, including all fresh and frozen ET.

Statistical analysis

All statistical analysis was conducted by SPSS 25.0 (SPSS, Chicago, USA). The normality of continuous variables was assessed by the Shapiro-Wilk test. If conforming to normality distribution, mean \pm SD and analysis of variance (ANOVA) were adopted for variables description and intra-group comparisons. Vice versa, if the non-normality distribution was ascertained, the median (25th-75th percentile) and Kruskal-Wallis test were used for variables description and intragroup comparisons. Frequencies (percentages) were utilized to describe categorical variables. The comparisons of categorical variables were evaluated by χ^2 (2) test or Fisher's exact test. Among the pairwise comparisons of four subgroups, the Bonferroni method was used to adjust p-values. In order to account for the differences among subgroups, parameters, such as age, BMI, duration of infertility, types of infertility, AFC, AMH, FSH, initial uterine diameter, and dysmenorrhea history, were included in the logistic regression model. $P < 0.05$ was considered as statistical significance.

Results

A total of 562 cycles were included with 257 cycles of fresh ET and 305 cycles of FET. In fresh ET cycles, cycles of ultra-long, long, antagonist, and short protocols were 108, 56, 34, and 59, respectively. In FET cycles, cycles of FET with embryos originating from ultra-long, long, antagonist, and short protocols were 98, 101, 54, and 52.

Pregnancy outcomes of adenomyosis in fresh ET cycles

In fresh ET cycles, except for BMI, there were significant differences in age, duration of infertility, primary infertility, AFC, FSH, AMH, mean initial diameter of the uterus, and dysmenorrhea in intragroup comparisons. Compared with ultra-long, long, and

antagonist protocols, the short protocol had older age (34.00, 32.50, 33.50 versus 37.00, $P < 0.001$), poorer AMH (1.65, 2.75, 1.32 versus 1.30, $P < 0.001$), and lower AFC (11.00, 13.00, 10.00 versus 9.0, $P < 0.001$). The proportion of severe dysmenorrhea in ultra-long protocol was highest compared with long, antagonist, and short protocols (37.96% versus 17.86%, 11.76%, 13.56%, $P < 0.001$). The data was shown in [Table 1](#).

COS-related parameters and pregnancy outcomes were shown in [Table 2](#). Compared with the long, antagonist, and short protocols, the duration of gonadotropin (9.00, 9.00, 9.00 versus 11.00, $P < 0.001$) and the dosage of gonadotropin (1800.00, 2175.00, 1975.00 versus 2775.00, $P < 0.001$) increased clearly in the ultra-long protocol. Compared with ultra-long, long and antagonist protocols, endometrial thickness on HCG trigger day was thinnest (1.1, 1.1, 1.0 versus 0.9, $P = 0.002$), and the number of retrieved oocytes was lowest (7.00, 10.00, 6.00 versus 4.00, $P < 0.001$) in the short protocol. However, no statistical significance existed among intragroup comparisons of ultra-long, long, and antagonist protocols. For high-quality embryos on day 3, long protocol had more good embryos compared with ultra-long, antagonist, and short protocols (4.00 versus 2.00, 2.00, 2.00, $P < 0.001$).

The parameters of pregnancy outcomes included IR, BPR, EPR, CPR, MR, early MR and late MR, LBR, and CLBR. Compared with ultra-long and long protocols, IR (49.7%, 52.1% versus 28.2%, $P = 0.001$) and CPR (64.3%, 57.4% versus 35.6%, $P = 0.004$) in short protocol significantly decreased. Compared with ultra-long and long protocols, the decrease of IR (49.7%, 52.1% versus 33.3%) and CPR (57.4%, 64.3% versus 38.2%) also existed in antagonist protocol, although no statistical significance was detected because of strict P adjustment of Bonferroni method ($P_{adj} = 0.008$). Compared with the long protocol, LBR in the short protocol decreased obviously (48.2% versus 20.3%, $P < 0.001$) and LBR in the antagonist protocol was also pessimistic (48.2% versus 26.5%), although no statistical significance. There were no significant differences in intragroup comparisons of BPR, MR, early MR, and late MR.

Considering the differences in baseline parameters among the four protocols, we carried out CPR and LBR-associated multinomial logistics regression analysis, which was shown in [Supplementary Tables 1, 2](#). When CPR was set as the outcome variable, we observed that long protocol was a protective factor compared with short protocol (OR 2.414, 95% CI 1.011-1.144, $P = 0.047$). AFC was also a protective factor (OR 1.073, 95% CI 1.007-1.144, $P = 0.030$), and the main risk factor was age (OR 0.87, 95% CI 0.809-0.936, $P < 0.001$). When LBR was set as the outcome variable, ultra-long (OR 2.47, 95% CI 1.005-6.07, $P = 0.049$) and long protocol (OR 2.786, 95% CI 1.055-7.353, $P = 0.039$) were both protective factors compared with short protocol. Secondary infertility (OR 2.088, 95% CI 1.018-4.283, $P = 0.045$) and AFC (OR 1.069, 95% CI 1.001-1.142, $P = 0.047$) were also protective factors. Age (OR 0.809, 95% CI 0.743-0.882, $P < 0.001$) was still a risk factor.

Pregnancy outcomes of adenomyosis in FET cycles

In FET cycles, baseline data of frozen embryo transferred was shown in [Table 3](#). Compared with ultra-long, long, and antagonist

TABLE 1 Baseline characteristics of different COS protocols in fresh ET cycles.

| | Ultra-long protocol | Long protocol | Antagonist protocol | Short protocol | P value |
|-------------------------------------|----------------------------------|------------------------------------|---------------------------------|-----------------------------------|---------|
| No. of cycles | 108 | 56 | 34 | 59 | |
| Age, years | 34.00 (31.00,37.00) ^a | 32.50 (30.00,36.00) ^b | 33.50 (32.00,41.00) | 37.00 (33.00,40.00) ^{ab} | 0.001 |
| BMI, kg/m ² | 24.40 (22.09,26.84) | 23.37 (21.37,26.63) | 25.70 (23.02,27.03) | 23.95 (21.76,26.26) | 0.249 |
| Duration of infertility, years | 3.00 (2.00,5.00) | 3.00 (2.00,5.00) | 3.50 (2.50,5.50) ^d | 2.00 (1.00,3.75) ^d | 0.007 |
| Primary infertility, n(%) | 40 (37.04) ^a | 23 (41.07) ^b | 9 (26.47) | 6 (10.17) ^{ab} | <0.001 |
| AFC | 11.00 (7.00,14.00) ^{ac} | 13.00 (10.00,16.00) ^{bce} | 10.00 (6.00,14.00) ^c | 9.00 (5.00,11.00) ^{ab} | <0.001 |
| Basal FSH, IU/L | 6.45 (5.30,7.78) ^a | 6.50 (5.88,7.23) | 6.76 (5.60,8.97) | 7.60 (6.32,8.87) ^a | 0.015 |
| AMH, ng/ml | 1.65 (1.06,3.03) ^{ac} | 2.75 (1.66,3.90) ^{bc} | 1.32 (0.92,3.37) | 1.30 (0.77,1.90) ^{ab} | <0.001 |
| Mean initial diameter of uterus, cm | 6.22 (5.64,7.30) ^{ac} | 5.43 (4.65,6.35) ^c | 6.05 (4.81,6.85) | 5.45 (4.72,6.03) ^a | <0.001 |
| History of dysmenorrhea | | | | | <0.001 |
| None, n(%) | 11 (10.19) | 19 (33.93) | 16 (47.06) | 22 (37.29) | |
| Mild, n(%) | 30 (27.78) | 19 (33.93) | 8 (23.53) | 18 (30.51) | |
| Moderate, n(%) | 26 (24.07) | 8 (14.29) | 6 (17.65) | 11 (18.64) | |
| Severe, n(%) | 41 (37.96) ^{af} | 10 (17.86) | 4 (11.76) ^f | 8 (13.56) ^a | |

Data were presented as median (25th–75th percentile) for non-normality distribution variables and frequencies (percentages) for categorical variables.

COS, controlled ovarian stimulation; BMI, body mass index; AFC, antral follicle count; FSH, follicle stimulating hormone; AMH, anti-müllerian hormone; ET, embryo transfer.

^aultra-long vs short; ^blong vs short; ^cultra-long vs long; ^dantagonist vs short; ^elong vs antagonist; ^fultra-long vs antagonist.

TABLE 2 Outcomes of ultra-long, long, antagonist, and short protocols in fresh ET cycles.

| | Ultra-long protocol | Long protocol | Antagonist protocol | Short protocol | P _{adjust} |
|--|---|--|---|---|---------------------|
| No. of cycles | 108 | 56 | 34 | 59 | |
| Total dosage of Gn, IU | 2775.00 (2250.00,3993.75) ^{acf} | 1800.00 (1500.00,2325.00) ^c | 2175.00 (1656.25,2728.12) ^f | 1975.00 (1425.00,2700.00) ^a | <0.001 |
| Duration of Gn stimulation, days | 11.00 (10.00,12.25) ^{acf} | 9.00 (9.00,11.00) ^c | 9.00 (8.00,10.00) ^f | 9.00 (7.50,10.00) ^a | <0.001 |
| LH on HCG trigger day, IU/L | 1.04 (0.56,1.59) ^{acf} | 2.30 (1.43,3.05) ^{bc} | 2.51 (1.80,5.54) ^{df} | 5.24 (3.80,7.04) ^{abd} | <0.001 |
| E ₂ on HCG trigger day, pg/ml | 2061.50 (1407.75,2884.25) | 2650.50 (1708.00,3468.25) ^{be} | 1653.50 (1081.00,2172.25) ^c | 1619.00 (1294.50,2575.50) ^b | <0.001 |
| P on HCG trigger day, ng/ml | 0.66 (0.46,0.84) | 0.58 (0.44,0.94) | 0.46 (0.35,0.80) | 0.65 (0.39,0.81) | 0.549 |
| Endometrial thickness on HCG trigger day, cm | 1.10 (0.90,1.20) ^a | 1.10 (0.90,1.25) ^b | 1.00 (0.80,1.10) | 0.90 (0.80,1.10) ^{ab} | 0.002 |
| No. of oocytes retrieved | 7.00 (4.75,12.00) ^a | 10.00 (6.00,12.00) ^b | 6.00 (3.00,9.75) | 4.00 (3.00,8.00) ^{ab} | <0.001 |
| No. of 2PN zygotes retrieved | 5.00 (3.00,7.25) ^a | 6.00 (4.00,9.00) ^b | 4.00 (2.25,6.50) | 3.00 (2.00,5.00) ^{ab} | <0.001 |
| No. of high-quality embryos retrieved on Day 3 | 2.00 (1.00,4.00) ^c | 4.00 (2.00,6.00) ^{bc} | 2.00 (2.00,4.00) | 2.00 (1.00,3.00) ^b | <0.001 |
| Pregnancy outcomes, %(n/N) | | | | | |
| IR | 49.7 (85/171) ^a | 52.1 (49/94) ^b | 33.3 (18/54) | 28.2 (24/85) ^{ab} | 0.001 |
| BPR | 10.2 (11/108) | 10.7 (6/56) | 11.8 (4/34) | 8.5 (5/59) | 0.960 |
| EPR | 0.01 (1/108) | – | – | – | – |
| CPR | 57.4 (62/108) ^a | 64.3 (36/56) ^b | 38.2 (13/34) | 35.6 (21/59) ^{ab} | 0.004 |
| MR | 30.6 (19/62) | 25.0 (9/36) | 30.8 (4/13) | 42.9 (9/21) | 0.575 |

(Continued)

TABLE 2 Continued

| | Ultra-long protocol | Long protocol | Antagonist protocol | Short protocol | P _{adjust} |
|----------|---------------------|---------------------------|---------------------|---------------------------|---------------------|
| Early MR | 16.1 (10/62) | 22.2 (8/36) | 23.1 (3/13) | 38.1 (8/21) | 0.230 ^g |
| Late MR | 14.5 (9/62) | 2.8 (1/36) | 7.7 (1/13) | 4.8 (1/21) | 0.241 ^g |
| LBR | 39.8 (43/108) | 48.2 (27/56) ^b | 26.5 (9/34) | 20.3 (12/59) ^b | 0.007 |

Data were presented as median (25th–75th percentile) for non-normality distribution variables. Gn, gonadotropin; LH, luteinizing hormone; E2, estradiol; P, progesterone; HCG, human chorionic gonadotropin IR, implantation rate; BPR, biochemical pregnancy rate; EPR, ectopic pregnancy rate; CPR, clinical pregnancy rate; MR: miscarriage rate; LBR, live birth rate.

^aultra-long vs short; ^blong vs short; ^cultra-long vs long; ^dantagonist vs short; ^elong vs antagonist; ^fultra-long vs antagonist; ^gFisher's exact test.

TABLE 3 Baseline characteristics of frozen embryo originating COS cycles.

| | Ultra-long protocol | Long protocol | Antagonist protocol | Short group | P value |
|---|----------------------------------|----------------------------------|----------------------------------|------------------------------------|---------|
| No. of cycles | 98 | 101 | 54 | 52 | |
| Age, years | 33.00 (30.00,37.00) ^a | 31.00 (30.00,35.00) ^b | 33.00 (30.25,36.00) ^d | 37.00 (34.00,40.00) ^{abd} | <0.001 |
| BMI, kg/m ² | 23.14 (20.70,26.33) ^a | 23.28 (21.13,27.18) | 24.69 (23.01,26.17) | 25.33 (23.06,26.70) ^a | 0.016 |
| Duration of infertility, years | 3.00 (2.00,4.50) | 3.50 (1.50,4.00) | 3.50 (2.50,5.50) | 2.50 (2.00,4.00) | 0.084 |
| Primary infertility, n(%) | 51 (52.04) ^a | 50 (49.5) ^b | 25 (46.3) ^d | 7 (13.46) ^{abd} | <0.001 |
| AFC | 13.50 (10.00,19.75) ^a | 15.00 (11.00,20.00) ^b | 14.00 (9.00,28.5) ^d | 9.00 (5.75,12.00) ^{abd} | <0.001 |
| Basal FSH, IU/L | 6.08 (5.06,7.57) | 6.16 (5.61,7.03) ^b | 6.31 (5.92,7.71) | 6.87 (5.57,8.35) ^b | 0.013 |
| AMH, ng/ml | 2.84 (1.57,5.90) ^a | 4.17 (2.78,5.54) ^b | 4.09 (1.73,6.90) ^d | 1.35 (0.84,2.17) ^{abd} | <0.001 |
| Mean diameter of initial uterus, cm | 6.25 (5.50,7.30) ^c | 5.70 (4.70,6.75) ^c | 5.80 (4.76,7.05) | 5.80 (4.94,6.61) | 0.010 |
| History of dysmenorrhea | | | | | <0.001 |
| None, n(%) | 15 (15.31) | 21 (20.79) | 15 (27.78) | 15 (28.85) | |
| Mild, n(%) | 21 (21.43) | 43 (42.57) | 13 (24.07) | 17 (32.69) | |
| Moderate, n(%) | 26 (26.53) | 13 (12.87) | 22 (40.74) | 9 (17.31) | |
| Severe, n(%) | 36 (36.73) ^f | 24 (23.76) | 4 (7.41) ^f | 11 (21.15) | |
| long-acting GnRHa pretreatment before FET | | | | | 0.004 |
| yes | 62(63.26%) ^c | 39(38.61%) ^c | 27(50.00%) | 22(42.31%) | |
| no | 36(36.74%) ^c | 62(61.39%) ^c | 27(50.00%) | 30(57.7%) | |

Data were presented as median (25th–75th percentile) for non-normality distribution variables and frequencies (percentages) for categorical variables.

BMI, body mass index; AFC, antral follicle count; FSH, follicle-stimulating hormone; AMH, anti-müllerian hormone; COS, controlled ovarian stimulation; FET, frozen embryo transplant.

^aultra-long vs short; ^blong vs short; ^cultra-long vs long; ^dantagonist vs short; ^elong vs antagonist; ^fultra-long vs antagonist.

protocols, the short protocol had older age (33.00, 31.00, 33.00 versus 37.00, $P<0.001$) and lower AMH (2.84, 4.17, and 4.09 versus 1.35, $P<0.001$). Age, BMI, duration of infertility, primary infertility, AFC, FSH, and AMH had no statistical difference among ultra-long, long and antagonist protocols. The related parameters of frozen-embryo-originating COS protocols and pregnancy outcomes were shown in Table 4. Compared with the long, antagonist, and short protocols, the dosage of gonadotropin (2025.00, 1875.00, and 2025.00 versus 2512.50, $P<0.001$) and the stimulation duration of gonadotropin (10.00, 9.00, 9.00 versus 11.00, $P<0.001$) significantly increased in the ultra-long protocol. Ultra-long and long protocols had higher E2 levels on HCG trigger day compared with the short protocol (3117.00, 3000.00 versus 2131.50, $P<0.001$). The number of retrieved oocytes (6.5 versus 12.00, 12.00, 11.00, $P<0.001$) and

2PN zygotes (4.00 versus 7.00, 8.00, 6.50, $P<0.001$) in short protocol were significantly lower than those in ultra-long, long and antagonist protocols. Absolutely different from fresh ET cycles, no statistical differences were detected among ultra-long, long, antagonist and short protocols on IR (45.5%, 47.2%, 41.8%, 50.0%, $P=0.851$), BPR (12.2%, 6.9%, 5.6%, 13.5%, $P=0.313$), CPR (43.9%, 49.5%, 40.7%, 50.0%, $P=0.658$), MR (41.9%, 32.0%, 45.5%, 38.5%, $P=0.791$), early MR (27.9%, 32.0%, 36.4%, 30.8%, $P=0.917$), late MR (14.0, -, 9.1%, 7.7%, $P=0.12$), and LBR (25.5%, 33.7%, 22.2%, 30.8%, $P=0.403$). The CLBR in the long protocol was significantly higher than the corresponding rate in ultra-long, antagonist, and short protocols (68.2% versus 46.2%, 36.2%, 35.9%, $p<0.001$). We also noticed that in the ultra-long protocol, even if women had severer dysmenorrhea and larger uterus, the

TABLE 4 Outcomes of FET and characteristics of frozen embryo originating COS cycles.

| | Ultra-long protocol | Long protocol | Antagonist protocol | Short protocol | P _{adjust} |
|--|---|---|--|--|---------------------|
| No. of cycles | 98 | 101 | 54 | 52 | |
| Total dosage of Gn, IU | 2512.50 (2025.00,3862.50) ^{acf} | 2025.00 (1500.00,2775.00) ^c | 1875.00 (1418.75,2531.25) ^f | 2025.00 (1556.25,2625.00) ^a | <0.001 |
| Duration of Gn stimulation, days | 11.00 (10.00,12.75) ^{acf} | 10.00 (9.00,12.00) ^c | 9.00 (9.00,11.00) ^f | 9.00 (8.00,10.25) ^a | <0.001 |
| LH on HCG trigger day, IU/L | 1.00 (0.57,1.40) ^{acf} | 1.74 (1.15,2.61) ^{bce} | 3.28 (1.98,4.92) ^{cf} | 4.79 (3.40,6.50) ^{ab} | <0.001 |
| E ₂ on HCG trigger day, pg/ml | 3117.00 (2445.00,4806.75) ^a | 3000.00 (2373.00,4300.00) ^b | 2158.00 (1741.00,4566.25) | 2131.50 (1597.75,3408.25) ^{ab} | <0.001 |
| P on HCG trigger day, ng/ml | 0.75 (0.53,0.99) | 0.74 (0.50,1.08) | 0.83 (0.43,1.18) | 0.71 (0.43,0.91) | 0.542 |
| Endometrial thickness on HCG trigger day, cm | 0.90 (0.80,1.00) | 0.90 (0.80,1.00) | 0.90 (0.80,1.00) | 0.88 (0.80,1.00) | 0.676 |
| No. of oocytes retrieved | 12.00 (9.00,17.75) ^a | 12.00 (10.00,16.00) ^b | 11.00 (5.25,15.00) ^d | 6.50 (4.00,8.25) ^{abd} | <0.001 |
| No. of 2PN zygotes retrieved | 7.00 (5.00,12.00) ^a | 8.00 (6.00,10.00) ^b | 6.50 (4.00,10.00) ^d | 4.00 (3.00,6.00) ^{abd} | <0.001 |
| No. of high-quality embryos retrieved on Day 3 | 4.00 (2.00,6.00) | 5.00 (3.00,8.00) ^b | 3.00 (1.25,6.00) | 3.00 (2.00,4.00) ^b | <0.001 |
| Pregnancy outcomes, %(n/N) | | | | | |
| IR | 45.5 (45/99) | 47.2 (51/108) | 41.8 (23/55) | 50.0 (26/52) | 0.851 |
| BPR | 56.1(55/98) | 56.4 (57/101) | 46.3 (25/54) | 63.5 (33/52) | 0.313 |
| EPR | – | – | – | 0.02 (1/52) | – |
| CPR | 43.9 (43/98) | 49.5 (50/101) | 40.7 (22/54) | 50.0 (26/52) | 0.658 |
| MR | 41.9 (18/43) | 32.0 (16/50) | 45.5 (10/22) | 38.5 (10/26) | 0.791 |
| Early MR | 27.9 (12/43) | 32.0 (16/50) | 36.4 (8/22) | 30.8 (8/26) | 0.917 |
| Late MR | 14.0 (6/43) | – | 9.1 (2/22) | 7.7 (2/26) | 0.120 ^g |
| LBR | 25.5 (25/98) | 33.7 (34/101) | 22.2 (12/54) | 30.8 (16/52) | 0.403 |
| CLBR | 46.2(67/145) ^c | 68.2(60/88) ^{bce} | 36.2(21/58) ^c | 35.9(21/58) ^b | <0.001 |
| CLBR ^{hypo} | 59.8(116/194) ^c | 76.5(91/119) ^{bce} | 50.7(38/75) ^c | 49(48/98) ^b | <0.001 |

Data were presented as median (25th–75th percentile) for non-normality distribution variables. Gn, gonadotropin; LH, luteinizing hormone; E₂, estradiol; P, progesterone; HCG, human chorionic gonadotropin; IR, implantation rate; BPR, biochemical pregnancy rate; EPR, ectopic pregnancy rate; CPR, clinical pregnancy rate; MR, miscarriage rate; LBR, live birth rate; CLBR, cumulative live birth rate, was calculated based on current finished embryo transfer cycles. CLBR^{hypo} was calculated by hypothesizing that all surplus embryos were transferred and live birth was achieved. ^aultra-long vs short; ^blong vs short; ^cultra-long vs long; ^dantagonist vs short; ^elong vs antagonist; ^fultra-long vs antagonist; ^gFisher's exact test.

CLBR was higher than those in antagonist and short protocol (46.2% versus 36.2%, 35.9%), although no statistical differences existed because of strict P adjustment of Bonferroni method ($P_{adj}=0.008$). So far, 117 blastocytes were still frozen, and we calculated the highest rate of CLBR by hypothesizing that live birth can be achieved by surplus embryo transfer. The real CLBR will fluctuate between the current data and the hypothesized data.

Pregnancy outcomes of adenomyosis with age ≥ 35 years in fresh ET and FET cycles

Considering the possible selection bias of different protocols, we carried out an analysis among women ≥ 35 years. In fresh ET cycles, cycles of ultra-long, long, antagonist, and short protocols were 48, 20, 15, and 40, respectively. There were no significant differences in BMI, basal FSH, AMH, and mean diameter of the initial uterus (Supplementary Table 3).

Antagonist protocol had significantly older age compared with ultra-long, long, and short protocols (42.00 versus 37.00, 38.00, and 39.00, $P=0.002$). The proportion of no dysmenorrhea in short protocol was significantly higher than in ultra-long protocols (45.0% versus 10.40%, $P=0.010$). Compared with antagonist and short protocols, ultra-long and long protocols had more number of oocytes (3.00, 4.00 versus 6.00, 8.00, $P=0.02$, $P=0.011$) and 2PN zygotes (3.00, 2.00 versus 4.50, 5.00, $P=0.012$). However, there was no statistical difference in high-quality embryos on day 3. As to the pregnancy outcomes, ultra-long and long protocols had higher CPR (52.1%, 50.00% versus 20.0%, 27.5%, $P=0.031$). Although there were no significant differences on IR ($P=0.183$), BPR ($P=0.87$), MR ($P=0.417$), and LBR ($P=0.071$), LBR in ultra-long and long protocols was higher than the corresponding rates in antagonist and short protocols (27.1%, 30.0% versus 6.7%, 10.0%). The data was shown in Supplementary Table 4).

In FET cycles, the FET cycles with embryos originating from ultra-long, long, antagonist, and short protocols were 46, 36, 20, and

37, respectively. The data was shown in [Supplementary Tables 5, 6](#). No significant differences existed in age between the four groups ($P=0.331$). Except for BMI, there were no statistical differences in the duration of infertility, the proportion of primary infertility, basal FSH, AMH, initial mean diameter of the uterus, the duration and dosage of gonadotropin, oocyte retrieved, number of 2PN zygotes and the final number of high-quality embryos retrieved on Day 3. The proportion of long-acting GnRHa pretreatment before FET in the long and short protocols groups was higher than in the ultra-long and antagonist groups (63.9%, 51.4% versus 30.4%, 30.00%, $P=0.009$). IR (61.1%, 48.6% versus 32.6%, 25.0%, $P=0.020$) and CPR (58.3%, 48.6% versus 30.4%, 25.0%, $P=0.024$) in long and short protocols were higher than rates of ultra-long and antagonist protocols, but no statistical differences were supported because of strict Bonferroni method ($P_{adj}=0.008$). No significant differences in BPR ($P=0.338$), MR ($P=0.634$), and LBR ($P=0.078$) were detected. The CLBR in long protocol was significantly higher than those in ultra-long, antagonist, and short protocols (68.8% versus 34.4%, 16%, 26.4%, $P<0.001$). In the ultra-long protocol, an even higher proportion of dysmenorrhea and larger uterus existed, and the CLBR was better compared with the antagonist and short protocols (34.4% versus 16%, 26.4%), although no statistical differences existed because of strict P adjustment of Bonferroni method ($P_{adj}=0.008$).

We carried out multinomial logistic regression analysis and CPR was set as the outcome variable. In fresh ET cycles, older age was an important risk factor (OR 0.787, 95% CI 0.658-0.941, $P=0.008$) and no protective factors were observed ([Supplementary Table 7](#)). In FET cycles, AFC (OR 0.903, 95% CI 0.821-0.994, $P=0.037$) and secondary infertility (OR 0.200, 95% CI 0.058-0.690, $P=0.011$) were observed as protective factors in FET cycles, which were shown in [Supplementary Table 8](#).

Discussion

Ultra-long protocol could mediate hypo-estrogen status caused by pituitary downregulation and produce positive effects on pregnancy outcomes (21). The long protocol also could mediate pituitary downregulation with weaker function. Because of endogenous inhibition of GnRHa, the duration and dosage of gonadotropin significantly increased in the ultra-long protocol. There were no statistical differences in the number of retrieved oocytes and 2PN zygotes in the intragroup comparison of ultra-long and long protocols. In our study, the ultra-long protocol had a higher proportion of severe adenomyosis, however, after long-acting GnRHa pretreatment, IR, CPR, MR, and LBR in ultra-long and long protocols were similar. Hou X's study analyzed 362 ultra-long protocol cycles and 127 long protocol cycles in fresh ET cycles among adenomyosis patients, and the dosage and duration of gonadotropin in ultra-long protocol also significantly increased. Compared with the long protocol, CPR and LBR in the ultra-long protocol were higher (10). Lan J's study ascertained that the ultra-long protocol improved pregnancy outcomes in women with adenomyosis, especially in women with diffuse adenomyosis (12). All in all, the ultra-long protocol had its advantages for the

improvement of pregnancy outcomes for severer adenomyosis, although time and economic cost was increased.

Antagonists could produce a rapid suppression of pituitary function, which could decrease the dosage of gonadotropin and the risk of ovarian hyperstimulation (22). Compared with ultra-long and long protocols, AFC in antagonist protocol was lower, however, the number of retrieved oocytes and 2PN zygotes had no statistical significance. Compare with ultra-long and long protocols, IR, CPR, and LBR in antagonist protocol were poorer, however, strict P adjustment of the Bonferroni method ($P_{adj}=0.008$) in pairwise under multiple comparisons made the statistical difference difficult to achieve. The BPR, MR, early MR, and late MR were similar among ultra-long, long, and antagonist protocols. Thalluri V. reported CPR in antagonist protocol was 23.6% in patients with adenomyosis, which was similar to our data and significantly lower than the reported CPR in ultra-long or long protocols (9–11, 23). Kolanska K compared pregnancy outcomes of GnRH-agonist versus GnRH-antagonist protocols in women with endometriosis-associated infertility and inferred GnRH-antagonist associated dysfunction of endometrial receptivity might result in decreased CPR and LBR (24).

The advantages of a short protocol could promote the secretion of FSH by flare-up effect, strengthen recruitment of early follicles and improve the response of the ovaries. Compared with ultra-long and long protocols, CPR and LBR in the short protocol were significantly decreased. Vice versa, compared with antagonist protocol, there were no statistical differences in IR, CPR, MR, and LBR. Sheng compared pregnancy outcomes of ultra-long, long, and short protocols, and CPR in the short protocol was 22.4% (25), which was similar to our data. Khan KN et al. reported short and antagonist protocols could not induce hypo-estrogenism and recover potential endometrial normalization of adenomyosis (14, 26), so when they were used, fresh ET should be prudent because of the obviously negative pregnancy outcomes.

In FET cycles, except for larger uterine diameter and a higher proportion of severe dysmenorrhea in ultra-long protocols, other baseline data of embryos originating COS cycles were similar in ultra-long, long and antagonist protocols. However, baseline differences between short and other protocols still had statistical differences, such as age, AFC, and AMH. What we should pay attention to is that no matter which was the origin of the embryo, IR, CPR, MR, and LBR had no statistical difference in FET cycles. So far, there were no other similar comparisons. As to CLBR, because patients still had frozen embryos, we calculated fluctuation ranges by hypothesizing successful live birth with embryo transfer in the future. Compared with other protocols, current data showed that long protocol had its advantages on CLBR. The baseline parameters between long and antagonist were matched, however, the CLBR was poor in antagonist protocol, therefore, antagonist protocol should be cautiously adapted. For severer adenomyosis with good ovarian reserve, the ultra-long protocol was worth considering, while more data was needed to verify a better choice between ultra-long and long protocol.

Considering the possible choosing bias, we analyzed the usage of different protocols and pregnancy outcomes among women ≥ 35 years. In fresh ET cycles, the antagonist protocol had older age than

the ultra-long protocol, while no difference in age existed in other intragroup comparisons. AMH and the final number of high-quality embryos had no statistical significance. Compared with the antagonist and short protocols, CPR in ultra-long and long protocols was higher. LBR in antagonist and short protocols were lower, although no statistical difference existed. In FET cycles, embryos from long and short protocols had a higher proportion of long-acting GnRHa pretreatment, which IR and CPR increased, indicating the possible benefit of long-acting GnRHa.

Strengths and limitations

This was the first study to make direct comparisons of different COS protocols on IVF/ICSI outcomes. Our study indicated that an ultra-long protocol might be a better choice for severe adenomyosis in fresh ET cycles. We observed that the pregnancy outcomes of antagonist and short protocols were poor in fresh ET cycles, however, the outcomes could be reversed by whole embryos being frozen and a FET strategy. The principal limitation of the study was the possible bias of a retrospective cohort study, therefore, we developed strict inclusion and exclusion criteria to minimize the bias. Prospective studies are needed to verify the conclusions.

Conclusion

If fresh ET was decided upon, an ultra-long or long protocol might be appropriate. If antagonist and short protocols were used, whole embryo frozen combined with FET is recommended. In FET cycles, embryo origin had no impact on pregnancy outcomes.

Data availability statement

Data will be made available to the editors of the journal for review or query upon request.

Ethics statement

The studies involving humans were approved by Ethics Committee at the Center for Reproductive Medicine, Shandong University (No. 2021-133). The studies were conducted in

accordance with the local legislation and institutional requirements. The participants provided their written informed consent to participate in this study.

Author contributions

LG and YL analyzed data and drafted the article. SG collected and analyzed data. LC and Z-JC designed, revised, and approved the manuscript.

Funding

This study is funded by The National Key Research and Development Program of China (2021YFC2700700), Research Unit of Gametogenesis and Health of ART-Offspring, Chinese Academy of Medical Sciences (2020RU001), China Health Promotion Foundation, Taishan Scholars Program for Young Experts of Shandong Province (tsqn201909195), Natural Science Foundation of Shandong Province (ZR2021MH390).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fendo.2023.1198779/full#supplementary-material>

References

1. Upson K, Missmer SA. Epidemiology of adenomyosis. *Semin Reprod Med* (2020) 38(2-03):89–107. doi: 10.1055/s-0040-1718920
2. Gordts S, Grimbizis G, Campo R. Symptoms and classification of uterine adenomyosis, including the place of hysteroscopy in diagnosis. *Fertil Steril* (2018) 109:380–8.e1. doi: 10.1016/j.fertnstert.2018.01.006
3. Kissler S, Hamscho N, Zangos S, Wiegatz I, Schlichter S, Menzel C, et al. Uterotubal transport disorder in adenomyosis and endometriosis—a cause for infertility. *BJOG* (2006) 113(8):902–8. doi: 10.1111/j.1471-0528.2006.00970.x
4. Guo S, Zhang D, Lu X, Zhang Q, Gu R, Sun B, et al. Hypoxia and its possible relationship with endometrial receptivity in adenomyosis: a preliminary study. *Reprod Biol Endocrinol* (2021) 19(1):7. doi: 10.1186/s12958-020-00692-y
5. Puente JM, Fabris A, Patel J, Patel A, Cerrillo M, Requena A, et al. Adenomyosis in infertile women: prevalence and the role of 3D ultrasound as a marker of severity of the disease. *Reprod Biol Endocrinol* (2016) 14(1):60. doi: 10.1186/s12958-016-0185-6

6. Younes G, Tulandi T. Effects of adenomyosis on *in vitro* fertilization treatment outcomes: a meta-analysis. *Fertil Steril* (2017) 108(3):483–490.e3. doi: 10.1016/j.fertnstert.2017.06.025
7. Vercellini P, Consonni D, Dridi D, Bracco B, Frattaruolo MP, Somigliana E. Uterine adenomyosis and *in vitro* fertilization outcome: a systematic review and meta-analysis. *Hum Reprod* (2014) 29(5):964–77. doi: 10.1093/humrep/deu041
8. Nirgianakis K, Kalaitzopoulos DR, Schwartz ASK, Spaanderman M, Kramer BW, Mueller MD, et al. Fertility, pregnancy and neonatal outcomes of patients with adenomyosis: a systematic review and meta-analysis. *Reprod BioMed Online* (2021) 42(1):185–206. doi: 10.1016/j.rbmo.2020.09.023
9. Wu Y, Huang J, Zhong G, Lan J, Lin H, Zhang Q. Long-term GnRH agonist pretreatment before frozen embryo transfer improves pregnancy outcomes in women with adenomyosis. *Reprod BioMed Online* (2022) 44(2):380–8. doi: 10.1016/j.rbmo.2021.10.014
10. Hou X, Xing J, Shan H, Mei J, Sun Y, Yan G, et al. The effect of adenomyosis on IVF after long or ultra-long GnRH agonist treatment. *Reprod BioMed Online* (2020) 41(5):845–53. doi: 10.1016/j.rbmo.2020.07.027
11. Park CW, Choi MH, Yang KM, Song IO. Pregnancy rate in women with adenomyosis undergoing fresh or frozen embryo transfer cycles following gonadotropin-releasing hormone agonist treatment. *Clin Exp Reprod Med* (2016) 43(3):169–73. doi: 10.5653/cerm.2016.43.3.169
12. Lan J, Wu Y, Wu Z, Wu Y, Yang R, Liu Y, et al. Ultra-long gnRH agonist protocol during IVF/ICSI improves pregnancy outcomes in women with adenomyosis: A retrospective cohort study. *Front Endocrinol (Lausanne)* (2021) 12:609771. doi: 10.3389/fendo.2021.609771
13. Wu HM, Chang HM, Leung PCK. Gonadotropin-releasing hormone analogs: Mechanisms of action and clinical applications in female reproduction. *Front Neuroendocrinol* (2021) 60:100876. doi: 10.1016/j.yfrne.2020.100876
14. Khan KN, Kitajima M, Hiraki K, Fujishita A, Sekine I, Ishimaru T, et al. Changes in tissue inflammation, angiogenesis and apoptosis in endometriosis, adenomyosis and uterine myoma after GnRH agonist therapy. *Hum Reprod (Oxford England)* (2010) 25:642–53. doi: 10.1093/humrep/dep437
15. Khan KN, Kitajima M, Hiraki K, Fujishita A, Nakashima M, Ishimaru T, et al. Cell proliferation effect of GnRH agonist on pathological lesions of women with endometriosis, adenomyosis and uterine myoma. *Hum Reprod (Oxford England)* (2010) 25:2878–90. doi: 10.1093/humrep/deq240
16. Van den Bosch T, Dueholm M, Leone FP, Valentin L, Rasmussen CK, Votino A, et al. Terms, definitions and measurements to describe sonographic features of myometrium and uterine masses: a consensus opinion from the Morphological Uterus Sonographic Assessment (MUSA) group. *Ultrasound obstetrics gynecology Off J Int Soc Ultrasound Obstetrics Gynecology* (2015) 46(3):284–98. doi: 10.1002/uog.14806
17. Bazot M, Darai E. Role of transvaginal sonography and magnetic resonance imaging in the diagnosis of uterine adenomyosis. *Fertil Steril* (2018) 109(3):389–97. doi: 10.1016/j.fertnstert.2018.01.024
18. Puissant F, Van Rysselberge M, Barlow P, Deweze J, Leroy F. Embryo scoring as a prognostic tool in IVF treatment. *Hum Reprod* (1987) 2(8):705–8. doi: 10.1093/oxfordjournals.humrep.a136618
19. Man Y, Bian Y, Zhao S, Zhao R, Xu X, Wei D, et al. The effect of different endometrial preparations on women with polycystic ovary syndrome undergoing initial frozen embryo transfer: A historical cohort analysis. *Acta Obstet Gynecol Scand* (2021) 100:1116–23. doi: 10.1111/aogs.14058
20. Andersch B, Milsom I. An epidemiologic study of young women with dysmenorrhea. *Am J Obstet Gynecol* (1982) 144(6):655–60. doi: 10.1016/0002-9378(82)90433-1
21. Xie M, Yu H, Zhang X, Wang W, Ren Y. Elasticity of adenomyosis is increased after GnRHa therapy and is associated with spontaneous pregnancy in infertile patents. *J Gynecol Obstet Hum Reprod* (2019) 48:849–53. doi: 10.1016/j.jogoh.2019.05.003
22. Lambalk CB, Banga FR, Huirne JA, Toftager M, Pinborg A, Homburg R, et al. GnRH antagonist versus long agonist protocols in IVF: a systematic review and meta-analysis accounting for patient type. *Hum Reprod Update* (2017) 23(5):560–79. doi: 10.1093/humupd/dmx017
23. Thalluri V, Tremellen KP. Ultrasound diagnosed adenomyosis has a negative impact on successful implantation following GnRH antagonist IVF treatment. *Hum Reprod Update* (2012) 27(12):3487–92. doi: 10.1093/humrep/des305
24. Kolanska K, Cohen J, Bendifallah S, Salleret L, Antoine JM, Chabbert-Buffet N, et al. Pregnancy outcomes after controlled ovarian hyperstimulation in women with endometriosis-associated infertility: GnRH-agonist versus GnRH-antagonist. *J Gynecol Obstet Hum Reprod* (2017) 46(9):681–6. doi: 10.1016/j.jogoh.2017.09.007
25. Sheng Y, Cai-Hong M, Yang R, Liu Z, Liu P, Qiao J. Effects of different controlled ovarian hyperstimulation for adenomyosis on the outcomes of IVF-ET. *Reprod Contracept* (2010) 30(6):375–8. doi: 10.1111/j.1479-828X.2010.01276.x
26. Tremellen K, Russell P. Adenomyosis is a potential cause of recurrent implantation failure during IVF treatment. *Aust N Z J Obstet Gynaecol* (2011) 51(3):280–3. doi: 10.1111/j.1479-828X.2010.01276.x



OPEN ACCESS

EDITED BY

Lianghui Diao,
Shenzhen Zhongshan Urology Hospital,
China

REVIEWED BY

Yihua Yang,
Guangxi Medical University, China
Ting Zhang,
Shanghai Jiao Tong University, China

*CORRESPONDENCE

Song Quan

✉ quansong@smu.edu.cn

Xiao Shi

✉ drshixiao@163.com

[†]These authors have contributed equally to this work

RECEIVED 04 May 2023

ACCEPTED 31 July 2023

PUBLISHED 29 August 2023

CITATION

Ren M, Wang L, Wen L, Chen J, Quan S and Shi X (2023) Association between female circulating heavy metal concentration and abortion: a systematic review and meta-analysis. *Front. Endocrinol.* 14:1216507. doi: 10.3389/fendo.2023.1216507

COPYRIGHT

© 2023 Ren, Wang, Wen, Chen, Quan and Shi. This is an open-access article distributed under the terms of the [Creative Commons Attribution License \(CC BY\)](#). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Association between female circulating heavy metal concentration and abortion: a systematic review and meta-analysis

Meiqi Ren^{1†}, Liantong Wang^{1†}, Liqin Wen², Jinghua Chen², Song Quan^{1*} and Xiao Shi^{1*}

¹Center for Reproductive Medicine, Department of Obstetrics and Gynaecology, NanFang Hospital, Southern Medical University, Guangzhou, China, ²The First School of Clinical Medicine, Southern Medical University, Guangzhou, China

Objective: This study aimed to evaluate the association between blood heavy metal (zinc (Zn), copper (Cu), lead (Pb), and cadmium (Cd)) concentrations and spontaneous abortion (SA) and recurrent pregnancy loss (RPL) and explore the possible endocrine dysfunction associated with it.

Methods: A literature search was performed in the PubMed, Embase, Cochrane Library, and Web of Science databases up to April 2023. The overall effects were expressed as the standard mean difference (SMD). Subgroup analysis was performed according to the type of abortion (SA or RPL). Stata 16.0 was utilized for data analysis.

Results: Based on the integrated findings, abortion women showed significantly lower Zn (SMD = -1.05, 95% CI: -1.74 to -0.36, $p = 0.003$) and Cu concentrations (SMD = -1.42, 95% CI: -1.97 to -0.87, $p < 0.001$) and higher Pb (SMD = 1.47, 95% CI: 0.89–2.05, $p < 0.001$) and Cd concentrations (SMD = 1.15, 95% CI: 0.45–1.85, $p = 0.001$) than normal pregnant women. Subgroup analysis showed that Zn and Cu deficiency and Cd and Pb exposure were significantly ($p < 0.05$) associated with RPL, whereas Cu deficiency and Cd and Pb exposure were significantly ($p < 0.05$) associated with SA.

Conclusion: Zn and Cu deficiencies and Pb and Cd exposure were associated with abortion. Endocrine dysfunction, such as insulin resistance, vitamin D insufficiency, and abnormal thyroid and sex hormone concentrations, is thought to be involved in heavy metal-related abortion.

KEYWORDS

recurrent pregnancy loss, spontaneous abortion, endocrine dysfunction, zinc, copper, lead, cadmium

1 Introduction

Spontaneous abortion (SA) is a serious reproductive health problem with various definitions. According to the World Health Organization, SA is defined as the involuntary loss of a fetus weighing ≤ 500 g before the 20th gestational week (GW) (1), whereas the Chinese Medical Association Obstetrics and Gynecology Branch defines it as the involuntary loss of a fetus weighing $\leq 1,000$ g before the 28th GW (2). SA occurs in 10%–15% of pregnancies, and approximately 80% of SA occurs before 12 weeks of pregnancy, which is known as early pregnancy loss (3). Recurrent pregnancy loss (RPL) is a special form of SA that affects 1.4% of women and causes physical and emotional challenges (4). However, the definition of RPL has been inconsistent. The European Society for Human Reproduction and Embryology (ESHRE) defines it as two or more abortions, irrespective of whether they are consecutive (5), while the American Society for Reproductive Medicine defines it as the loss of two or more consecutive pregnancies (6). There is controversy about the quantity and consecutiveness of abortions (7). The etiologies of SA and RPL, including chromosomal abnormalities, uterine malformations, and endocrine dysfunction, are complex (8–10). Exposure to environmental pollutants is also a risk factor for SA and RPL. Most pollutants are endocrine disruptors and early embryonic development is extremely sensitive to them (11, 12).

Heavy metals are among the most harmful environmental contaminants because they are not biologically degradable and can accumulate in organisms along the food chain (13). Heavy metals are mainly absorbed through air, drinking water, and contaminated food (14). They can be classified as essential (e.g., copper [Cu], zinc [Zn]) and non-essential (such as lead [Pb] and cadmium [Cd]). Essential metals play important roles in metabolism, enzymatic synthesis, and signal transduction, and their deficiency or overexposure may affect normal physiological functions of organisms (14). For instance, Zn and Cu are important components of several proteins, including antioxidant enzymes, metalloenzymes, and coenzymes, which are essential for fetal growth. Maternal Zn and Cu deficiency can reduce the fetal Zn and Cu supply through the placenta and cause fetal loss and pregnancy complications (15–17). Non-essential metals are usually toxic to humans, especially to human reproductive health, even at very low concentrations. Among all nonessential metals, Cd and Pb are endocrine-disrupting metals that can interfere with the production and secretion of sex hormones, leading to poor pregnancy outcomes (18).

Several previous studies have investigated the associations between the concentrations of Cd, Pb, Zn, and Cu in the blood and the risk of abortion (1, 19); however, the results have been inconsistent. Some studies have reported that exposure to heavy metals during early pregnancy can increase the incidence of SA and RPL (20–22), and endocrine dysfunction has been suggested as a mediator (23, 24). Other studies have reported contrasting findings (18). Given the increasing interest of clinicians and researchers, stronger evidence on the effect of heavy metal exposure on abortion and its underlying mechanisms is in demand. We performed the present meta-analysis to clarify the associations between abortion

and the concentrations of Cd, Pb, Zn, and Cu. We also systematically reviewed the previous literature to explore the relationships between endocrine dysfunction, the four metals, and RPL or SA.

2 Methods

2.1 Study selection

The systematic review was conducted according to the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) statement. PubMed, Embase, Cochrane Library, and Web of Science databases were searched for relevant studies published up to April 2023. The subject terms included 'Miscarriage,' 'Pregnancy loss,' 'Abortion, Spontaneous,' 'Zinc,' 'Copper,' 'Lead,' and 'Cadmium.' Random combinations of these subject terms and their synonyms were used for retrieval. The detailed literature search strategy is provided in the [Supplementary Material](#). We reached the corresponding authors when the data were missing.

2.2 Inclusion criteria and exclusion criteria

Studies meeting the following criteria were included in the meta-analysis. (a) Study population: Pregnant women without internal and obstetric diseases that impair the normal process of pregnancy, including infectious diseases, gestational hypertension, gestational diabetes mellitus, and infertility. (b) Measurement: Female serum, plasma, or whole blood metal concentrations. (c) Observation group: Women who had experienced abortion, including SA and RPL. SA is defined as the involuntary loss of a fetus before the 28th GW (including the 20th and 24th GW) (2). RPL is defined as two or more abortions, irrespective of whether they are consecutive (5). (d) Control group: Healthy pregnant women with normal pregnancy or delivery. (e) Study type: Observational study.

The exclusion criteria were as follows: (a) article type: review, meta-analysis, meeting, case report, letter, comment, editorial, note, trial registry record, or protocol, (b) studies that focused on non-human cases (e.g., animal studies), (c) unclear definition of SA or RPL, (d) insufficient data on metal concentration; and (e) unavailable full text.

2.3 Quality assessment

The studies that met our inclusion criteria after the initial search were case-control, nested case-control, and cross-sectional studies. Therefore, the Newcastle-Ottawa Scale (NOS) was used to assess the quality of the studies (25). Each included article was independently appraised by two authors (MR and LiqW). Based on the NOS, studies were categorized as high- (8, 9), moderate- (6, 7), or low-quality (<6). Any disagreements regarding the assessment of the studies were discussed with the third author (LiaW).

2.4 Data extraction

Two investigators independently extracted the relevant data from the included studies (MR and LiaW). All data were double-checked by the third author (LiqW). The following information was extracted from the selected studies: first author, publication year, country and continent of the study population, type of detected sample, type of article, type of heavy metal, type of abortion, follow-up endpoint, sample size, concentrations of heavy metals, and analytical method employed.

2.5 Statistical analysis

Meta-analysis was performed using Stata 16.0 (Stata Corp, College Station, TX, USA). The standard mean difference (SMD) was adopted to integrate the data on metal concentration, as it is a continuous variable with different units across various studies. The 95% confidence intervals (CIs) were computed and presented as forest plots. For each study, statistical heterogeneity was assessed using Cochran's Q-test and I^2 statistics, and a random effects model was used to estimate the relationship between metal concentrations and abortion, as there was significant heterogeneity ($p < 0.05$, $I^2 > 50\%$). To investigate the impact of metal concentration on the different types of abortions (SA and RPL), a subgroup analysis was performed. To investigate the origin of the heterogeneity, four additional subgroup analyses were performed based on the follow-up endpoints (ongoing pregnancy and live birth) of participants, continent of the study population (Africa, Asia, North America, Oceania, and Europe), type of article (case-control study, cross-sectional study, and nested case-control study), and type of detected sample (serum, plasma, and whole blood). An influence analysis (sensitivity analysis) was conducted to improve the reliability of the meta-analysis results. A funnel plot and Begg's and Egger's tests were used to detect potential

publication bias; p -values < 0.05 represented significant statistical publication bias for Begg's and Egger's tests.

3 Results

3.1 Literature search

Figure 1 illustrates the PRISMA flow diagram for the selection of studies for inclusion in the systematic review and meta-analysis. A total of 4,222 potential studies were identified through database search. Among them, 136 articles were removed for duplicates, 1,209 articles were not observational studies (including reviews, meta-analyses, meetings, case reports, letters, comments, editorials, notes, trial registry records, and protocols), and 2,829 articles were not relevant to our study based on screening of their titles and abstracts by two authors. The literature screening results were double-checked to ensure that the relevant documents were not missed and did not need to be retrieved. After an independent review of the full texts by three authors (MR, LiaW, and LiqW), 12 studies were excluded because they did not meet the inclusion criteria, and eight studies were excluded because they had insufficient data (they only reported mean values without standard deviation of the metal concentration). Twenty-eight relevant studies were subjected to a final quantitative assessment based on the exclusion and inclusion criteria. Among the 28 studies, 14 investigated Zn and Cu, 15 investigated Pb, and eight investigated Cd.

3.2 Study characteristics

Table 1 lists the baseline characteristics of the included articles. The included articles were observational studies published between 1979 and 2023 and involved 1,377 abortion cases (including 1,159 females with SA and 218 females with RPL), together with 3,289

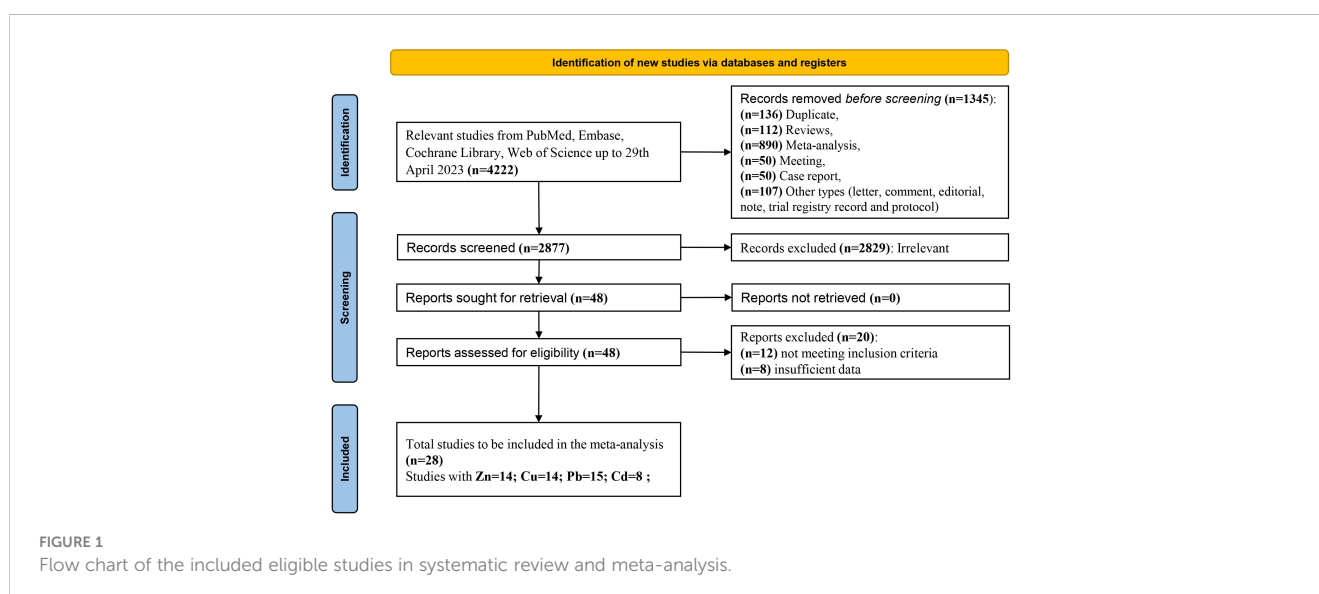


TABLE 1 Characteristics of the eligible studies.

| Sr.No | Author | Year | Country | Continent | Sample | Type of Article | Heavy Metal | Type of Abortion | Follow-up End-point | Number of subjects | | Metal concentration (mg/dL) | | p | Analytical method employed | Reference |
|-------|--------------------|------|---------|-----------|--------|-----------------|-------------|------------------|---------------------|--------------------|----------|-----------------------------|-----------------------|--------|----------------------------|-----------|
| | | | | | | | | | | Cases | Controls | Cases | Controls | | | |
| 1 | Ahmed, M.H. | 2007 | Egypt | Africa | Serum | Case-control | Zinc | SA | 12 weeks | 24 | 14 | 0.76 ± 0.06 (mg/L) | 0.86 ± 0.04 (mg/L) | 0.000 | AAS | (26) |
| | | | | | Serum | Case-control | Lead | SA | 12 weeks | 24 | 14 | 10.559 ± 1.317 (µg/dL) | 7.977 ± 3 (µg/dL) | 0.000 | AAS | |
| | | | | | Serum | Case-control | Cadmium | SA | 12 weeks | 24 | 14 | 3.2 ± 0.65 (µg/L) | 2.74 ± 0.25 (µg/L) | 0.039 | AAS | |
| 2 | Ajayi, O. O. | 2012 | Nigeria | Africa | Serum | Case-control | Zinc | RPL | 20 weeks | 35 | 34 | 99.25 ± 2.14 (µg/dL) | 99.25 ± 2.14 (µg/dL) | 0.001 | AAS | (27) |
| | | | | | Serum | Case-control | Copper | RPL | 20 weeks | 35 | 34 | 94.25 ± 3.07 (µg/dL) | 122.45 ± 2.71 (µg/dL) | 0.001 | AAS | |
| | | | | | Serum | Case-control | Lead | RPL | 20 weeks | 35 | 34 | 85.96 ± 1.09 (µg/dL) | 60.70 ± 1.40 (µg/dL) | 0.001 | AAS | |
| | | | | | Serum | Case-control | Cadmium | RPL | 20 weeks | 35 | 34 | 4.58 ± 0.77 (µg/dL) | 2.49 ± 0.09 (µg/dL) | 0.001 | AAS | |
| 3 | Alebic-Juretic, A. | 2005 | Croatia | Europe | Plasma | Case-control | Copper | SA | 14weeks | 17 | 28 | 18.2 ± 5.5 (µmol/L) | 27.1 ± 7.6 (µmol/L) | <0.001 | Photometry | (28) |
| 4 | Al-Sheikh, Y. A. | 2019 | Saudi | Asia | Plasma | Case-control | Zinc | RPL | Delivery | 28 | 28 | 2.84 ± 0.36 (µmol/l) | 3.55 ± 0.49 (µmol/l) | <0.001 | ICP-MS | (29) |
| | | | | | Plasma | Case-control | Copper | RPL | Delivery | 28 | 28 | 19.6 ± 2.75 (µmol/l) | 24.5 ± 3.41 (µmol/l) | <0.001 | ICP-MS | |
| 5 | Attalla, S.M. | 2009 | Egypt | Africa | Serum | Case-control | Zinc | RPL | 12 weeks | 40 | 24 | 77.01 ± 11.55 (µg %) | 90.01 ± 10.77 (µg %) | 0.020 | AAS | (30) |

(Continued)

TABLE 1 Continued

| Sr.No | Author | Year | Country | Continent | Sample | Type of Article | Heavy Metal | Type of Abortion | Follow-up End-point | Number of subjects | | Metal concentration (mg/dL) | | p | Analytical method employed | Reference |
|-------|---------------------|------|-----------|---------------|--------|-----------------|-------------|------------------|---------------------|--------------------|----------|-----------------------------|----------------------------|---------|----------------------------|-----------|
| | | | | | | | | | | Cases | Controls | Cases | Controls | | | |
| | | | | | Serum | Case-control | Lead | RPL | 12 weeks | 40 | 24 | 19.78 ± 3.85 (µg/dL) | 10.53 ± 1.01 (µg/dL) | <0.0001 | AAS | |
| | | | | | Serum | Case-control | Cadmium | RPL | 12 weeks | 40 | 24 | 7.51 ± 1.02 (µg/dL) | 5.06 ± 0.81 (µg/dL) | <0.0001 | AAS | |
| 6 | Bassiouni, B. A. | 1979 | Egypt | Africa | Plasma | Case-control | Copper | SA | 14 weeks | 24 | 14 | 140.12 ± 15.20 (µg/100 mL) | 188.57 ± 14.41 (µg/100 mL) | <0.01 | AAS | (31) |
| 7 | Borella, P. | 1990 | Italy | Europe | Plasma | Case-control | Zinc | SA | 16 weeks | 12 | 41 | 13.64 ± 2.35 (µmol/L) | 13.13 ± 2.37 (µmol/L) | >0.05 | AAS | (32) |
| | | | | | Plasma | Case-control | Copper | SA | 16 weeks | 12 | 41 | 28.22 ± 4.68 (µmol/L) | 26.05 ± 6.53 (µmol/L) | >0.05 | AAS | |
| 8 | Borja-Aburto, V. H. | 1999 | Mexico | North America | Serum | Case-control | Lead | SA | 20 weeks | 35 | 60 | 12 ± 6.16 (µg/dL) | 10.1 ± 5.34 (µg/dL) | 0.021 | AAS | (33) |
| 9 | Dreosti, I. E. | 1990 | Australia | Oceania | Serum | Case-control | Zinc | SA | 12 weeks | 35 | 37 | 0.75 ± 0.02 (µg/mL) | 0.69 ± 0.02 (µg/mL) | >0.05 | AAS | (34) |
| | | | | | Serum | Case-control | Copper | SA | 12 weeks | 35 | 37 | 1.36 ± 0.05 (µg/mL) | 1.48 ± 0.05 (µg/mL) | >0.05 | AAS | |
| 10 | Faikoglu, R. | 2006 | Turkey | Asia | Serum | Case-control | Lead | SA | 20 weeks | 20 | 20 | 23.2 ± 1 3.77 (µg/dL) | 18.04 ± 13.08 (µg/dL) | >0.05 | AAS | (35) |
| 11 | Ghneim, H. K. | 2016 | Saudi | Asia | Plasma | Case-control | Zinc | RPL | Delivery | 25 | 25 | 3.67 ± 0.39 (µmol/L) | 4.11 ± 0.49 (µmol/L) | <0.001 | ICP-MS | (36) |
| | | | | | Plasma | Case-control | Copper | RPL | Delivery | 25 | 25 | 25.6 ± 3.25 (µmol/L) | 28.8 ± 3.42 (µmol/L) | <0.001 | ICP-MS | |

(Continued)

TABLE 1 Continued

| Sr.No | Author | Year | Country | Continent | Sample | Type of Article | Heavy Metal | Type of Abortion | Follow-up End-point | Number of sub-jects | | Metal concentra-tion (mg/dL) | | p | Analytical method employed | Reference |
|-------|-----------------------|------|---------|---------------|-------------|-----------------|-------------|------------------|---------------------|---------------------|----------|------------------------------|--------------------------|---------|----------------------------|-----------|
| | | | | | | | | | | Cases | Controls | Cases | Controls | | | |
| 12 | Ghosh, A. | 1985 | China | Asia | Serum | Cross-section | Zinc | SA | 12 weeks | 45 | 55 | 120.18 ± 19.55 (µg/mL) | 123.03 ± 18.57 (µg/mL) | >0.05 | AAS | (37) |
| 13 | Jie, O. | 2019 | China | Asia | Whole blood | Case-control | Cadmium | SA | 12 weeks | 95 | 100 | 0.32 ± 0.28 (µg/L) | 0.22 ± 0.11 (µg/L) | 0.002 | ICP-MS | (38) |
| 14 | Lamadrid-Figueroa, H. | 2007 | Mexico | North America | Plasma | Case-control | Lead | SA | 12 weeks | 71 | 136 | 0.14 ± 0.13 (µg/L) | 0.13 ± 0.13 (µg/L) | 0.15 | ICP-MS | (39) |
| 15 | Lu, Y. | 2022 | China | Asia | Whole blood | Cross-section | Zinc | SA | 12 weeks | 92 | 103 | 5,082.32 ± 1,030.13 (µg/L) | 5,243.88 ± 960.87 (µg/L) | 0.251 | ICP-MS | (40) |
| | | | | | Whole blood | Cross-section | Copper | SA | 12 weeks | 92 | 103 | 797.36 ± 161.42 (µg/L) | 861.77 ± 188.75 (µg/L) | 0.008 | ICP-MS | |
| | | | | | Whole blood | Cross-section | Lead | SA | 12 weeks | 92 | 103 | 7.27 ± 3.01 (µg/L) | 7.61 ± 2.64 (µg/L) | 0.165 | ICP-MS | |
| 16 | Omeljaniuk, W. J. | 2015 | Poland | Europe | Serum | Case-control | Zinc | SA | Delivery | 83 | 35 | 0.5865 ± 0.1071 (mg/L) | 0.6492 ± 0.1878 (mg/L) | >0.05 | AAS | (41) |
| | | | | | Serum | Case-control | Copper | SA | Delivery | 83 | 35 | 1.1532 ± 0.2980 (mg/L) | 1.4450 ± 0.2930 (mg/L) | <0.002 | AAS | |
| 17 | Omeljaniuk, W. J. | 2018 | Poland | Europe | Whole blood | Case-control | Lead | SA | Delivery | 83 | 35 | 35.54 ± 11.0 (µg/L) | 27.11 ± 4.6 (µg/L) | <0.0001 | AAS | (19) |
| | | | | | Whole blood | Case-control | Cadmium | SA | Delivery | 83 | 35 | 2.730 ± 2.07 (µg/L) | 1.035 ± 0.59 (µg/L) | <0.0004 | AAS | |
| 18 | Ou, J. | 2020 | China | Asia | Whole blood | Case-control | Lead | SA | 12 weeks | 150 | 150 | 27.21 ± 31.43 (µg/L) | 15.96 ± 12.22 (µg/L) | 0.000 | ICP-MS | (42) |

(Continued)

TABLE 1 Continued

| Sr.No | Author | Year | Country | Continent | Sample | Type of Article | Heavy Metal | Type of Abortion | Follow-up End-point | Number of subjects | | Metal concentration (mg/dL) | | p | Analytical method employed | Reference |
|-------|-----------------|------|---------|-----------|-------------|---------------------|-------------|------------------|---------------------|--------------------|----------|-----------------------------|------------------------|--------|----------------------------|-----------|
| | | | | | | | | | | Cases | Controls | Cases | Controls | | | |
| 19 | Popovic, J. K. | 2016 | Serbia | Europe | Plasma | Case-control | Copper | SA | 12 weeks | 35 | 50 | 20.52 ± 3.76 (μmol/L) | 28.43 ± 4.45 (μmol/L) | <0.01 | Colorimetry | (43) |
| 20 | Sairoz | 2023 | India | Asia | Serum | Nested Case-control | Zinc | SA | 12 weeks | 80 | 100 | 51.7 ± 10.4 (μg/dL) | 81.6 ± 20.3 (μg/dL) | 0.0000 | Reaction with nitro-PAPS | (44) |
| | | | | | Serum | Nested Case-control | Copper | SA | 12 weeks | 80 | 100 | 222.5 ± 60.5 (μg/dL) | 302.5 ± 95.2 (μg/dL) | 0.0006 | Reaction with Di-Br-PAESA | |
| 21 | Shen, P. J. | 2015 | China | Asia | Serum | Nested Case-control | Zinc | SA | 12 weeks | 58 | 1389 | 72.67 ± 11.98 (μmol/L) | 83.25 ± 12.79 (μmol/L) | <0.05 | AAS | (45) |
| | | | | | Serum | Nested Case-control | Copper | SA | 12 weeks | 58 | 1389 | 29.96 ± 5.27 (μmol/L) | 31.24 ± 5.07 (μmol/L) | >0.05 | AAS | |
| 22 | Skalnaya, M. G. | 2019 | Russia | Europe | Serum | Case-control | Copper | SA | 28 weeks | 75 | 169 | 1.12 ± 0.29 (μg/L) | 1.60 ± 0.54 (μg/L) | <0.001 | ICP-MS | (46) |
| 23 | Tabassum, H. | 2022 | Saudi | Asia | Serum | Case-control | Lead | RPL | 12 weeks | 30 | 30 | 77.96 ± 5.51 (ppb) | 38.65 ± 0.20 (ppb) | <0.001 | ICP-MS | (47) |
| | | | | | Serum | Case-control | Cadmium | RPL | 12 weeks | 30 | 30 | 0.45 ± 0.04 (ppb) | 0.42 ± 0.01 (ppb) | <0.05 | ICP-MS | |
| 24 | Tousizadeh, S. | 2023 | Iran | Asia | Serum | Case-control | Zinc | RPL | Delivery | 60 | 60 | 5.26 ± 1.96 (mg/l) | 15.06 ± 7.17 (mg/l) | <0.001 | AAS | (48) |
| | | | | | Serum | Case-control | Lead | RPL | Delivery | 60 | 60 | 3.69 ± 2.48 (mg/l) | 0.31 ± 0.61 (mg/l) | <0.001 | AAS | |
| 25 | Vigeh, M. | 2010 | Iran | Asia | Whole blood | Case-control | Lead | SA | Delivery | 15 | 336 | 3.51 ± 1.42 (μg/dL) | 3.83 ± 1.99 (μg/dL) | 0.41 | ICP-MS | (49) |

(Continued)

TABLE 1 Continued

| Sr.No | Author | Year | Country | Continent | Sample | Type of Article | Heavy Metal | Type of Abortion | Follow-up End-point | Number of sub-jects | | Metal concentra-tion (mg/dL) | | p | Analytical method employed | Reference |
|-------|--------------|------|---------|-----------|-------------|-----------------|-------------|------------------|---------------------|---------------------|----------|------------------------------|---------------------|--------|----------------------------|-----------|
| | | | | | | | | | | Cases | Controls | Cases | Controls | | | |
| 26 | Vigeh, M. | 2021 | Iran | Asia | Whole blood | Case-control | Lead | SA | Delivery | 25 | 141 | 55.43 ± 54.3 (µg/L) | 44.97 ± 45.6 (µg/L) | 0.307 | ICP-MS | (50) |
| | | | | | Whole blood | Case-control | Cadmium | SA | Delivery | 25 | 141 | 0.51 ± 0.5 (µg/L) | 0.51 ± 0.5 (µg/L) | 0.957 | ICP-MS | |
| 27 | Wang, R. | 2020 | China | Asia | Serum | Cross-section | Zinc | SA | 12 weeks | 56 | 55 | 4.18 ± 0.26 (mg/L) | 3.24 ± 1.47 (mg/L) | >0.05 | ICP-MS | (51) |
| | | | | | Serum | Cross-section | Copper | SA | 12 weeks | 56 | 55 | 1.80 ± 0.58 (mg/L) | 1.41 ± 0.55 (mg/L) | <0.001 | ICP-MS | |
| | | | | | Serum | Cross-section | Lead | SA | 12 weeks | 56 | 55 | 0.17 ± 0.09 (mg/L) | 0.15 ± 0.10 (mg/L) | >0.05 | ICP-MS | |
| 28 | Yildirim, E. | 2019 | Turkey | Asia | Whole blood | Case-control | Lead | SA | 12 weeks | 29 | 20 | 54.11 ± 17.27 (µ/L) | 44.45 ± 12.49 (µ/L) | 0.038 | AAS | (52) |
| | | | | | Whole blood | Case-control | Cadmium | SA | 12 weeks | 29 | 20 | 0.39 ± 0.06 (µ/L) | 0.40 ± 0.05 (µ/L) | 0.704 | AAS | |

SA, Spontaneous Abortion; RPL, Recurrent Pregnancy Loss; AAS, Atomic-Absorption Spectrophotometry; ICP-MS, Inductively Coupled Plasma Mass spectrophotometry.

normal pregnant females. Of the 28 articles included in this meta-analysis, six were completed in China (37, 38, 40, 42, 45, 51), three in Egypt (26, 30, 31), three in Saudi Arabia (29, 36, 47), three in Iran (48–50), two in Mexico (33, 39), two in Turkey (35, 52), two in Poland (19, 41), one in Nigeria (27), one in Croatia (28), one in Italy (32), one in Australia (34), one in Serbia (43), one in India (44), and one in Russia (46). The sample size of the included studies ranged from 38 to 1,447.

3.3 Quality of included studies

The quality assessment results for all studies are shown in **Supplementary Table 1**. Studies with quality scores higher than 6, which is the cut-off NOS score for low quality, were considered credible. All of the included articles had quality scores above 6.

3.4 Meta-analysis for Zn

Fourteen studies investigated the association between Zn concentrations and abortion. The pooled effect size showed that the Zn concentration was negatively associated with abortion (SMD = -1.05 , 95% CI: -1.74 , -0.36 , $p = 0.003$, $I^2 = 96.9\%$; **Figure 2A**). Subgroup analysis showed that women with RPL had significantly lower Zn concentrations than healthy controls (SMD = -3.44 , 95% CI: -5.01 to -1.87 , $p < 0.001$), whereas the Zn concentrations of women with SA and healthy controls were not significantly different (SMD = -0.14 , 95% CI: -0.86 – 0.58 , $p = 0.710$). Significant heterogeneity was observed in each subgroup (SA, $p < 0.001$, $I^2 =$

96.6% ; RPL, $p < 0.001$, $I^2 = 96.6\%$; **Figure 2B**). To investigate the origin of the high heterogeneity, subgroup analyses based on follow-up endpoint, continent, type of article, and type of detected sample were performed. Subgroup analyses revealed persistently high heterogeneity (**Supplementary Figure 1**). Sensitivity analysis showed that omission of any study did not change the overall effect (**Figure 2C**). There was no evidence of publication bias among the included studies (Begg, $p = 0.511$; Egger, $p = 0.335$; **Figure 2D**).

3.5 Meta-analysis for Cu

Comparisons of Cu concentrations in women with and without abortion were reported in 14 studies. The pooled effect size of the 14 studies revealed significantly lower Cu concentrations in the abortion group than in the control group (SMD = -1.42 , 95% CI: -1.97 , -0.87 , $p < 0.001$, $I^2 = 95.4\%$) (**Figure 3A**). Subgroup analysis stratified by the type of abortion (SA and RPL) showed that patients with SA and RPL had lower Cu concentrations than the healthy controls (**Figure 3B**); the SMD was -0.97 (95% CI: -1.47 , -0.48 , $p < 0.001$) for women with SA and -3.92 (95% CI: -6.97 , -0.87 , $p = 0.012$, respectively). However, subgroup analysis based on abortion type showed obvious heterogeneity (SA, $p < 0.001$, $I^2 = 94.0\%$; RPL, $p < 0.001$, $I^2 = 97.8\%$). Subgroup analyses for the follow-up endpoint, continent, type of article, and type of detected sample showed high heterogeneity (**Supplementary Figure 2**). Sensitivity analysis showed that omission of any single study did not change the overall effect (**Figure 3C**). Publication bias was detected in the studies that included Cu (Begg: $p = 0.037$; Egger: $p = 0.012$). Visual inspection of funnel plots showed asymmetry (**Figure 3D**).

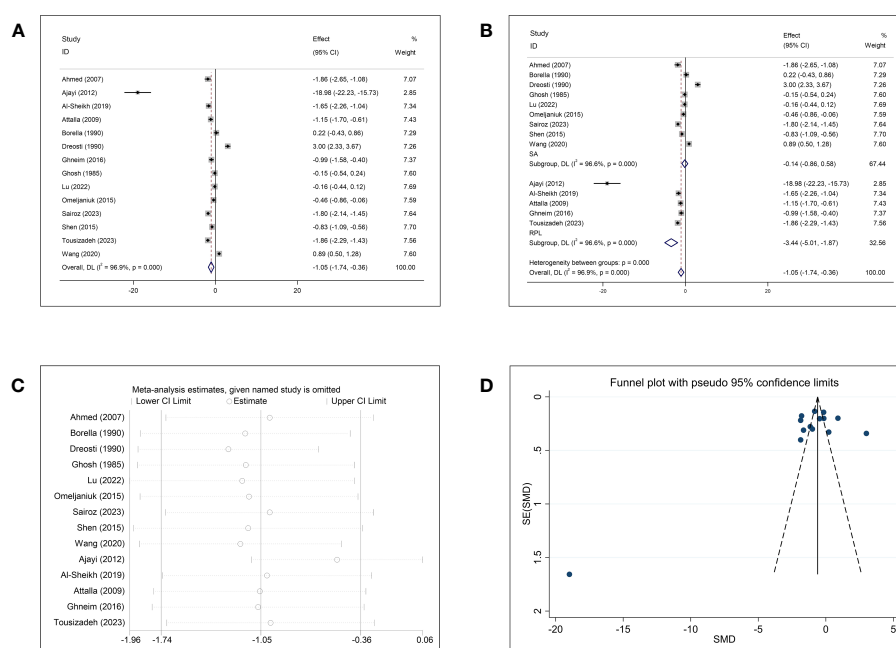


FIGURE 2

Meta-analysis outcomes of zinc. (A) Forest plot showing the meta-analysis outcomes between abortion group and normal pregnant women; (B) Subgroup analysis based on the type of abortion (SA and RPL); (C) Sensitivity analysis; and (D) Funnel plot.

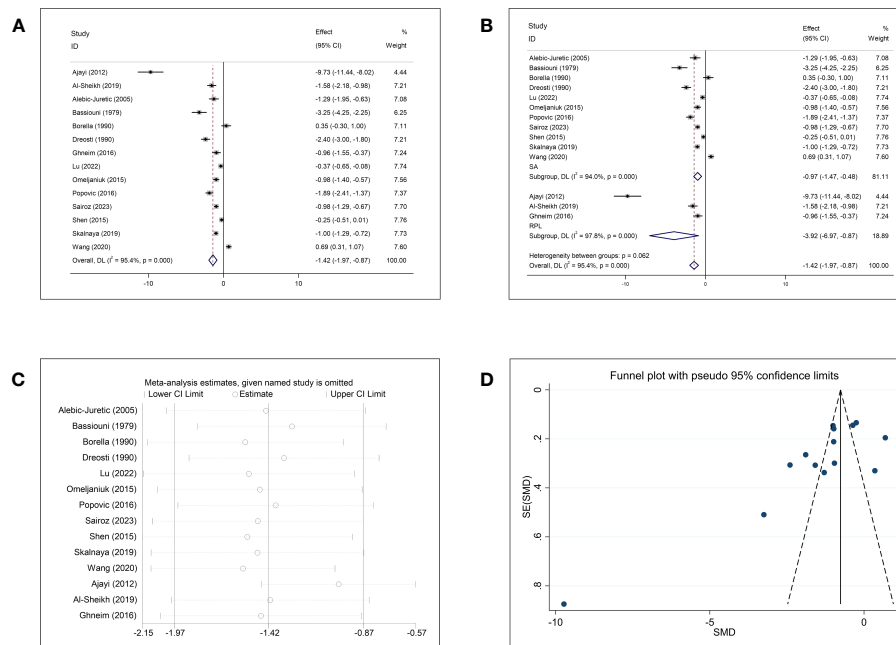


FIGURE 3

Meta-analysis outcomes of copper. (A) Forest plot showing the meta-analysis outcomes between abortion group and normal pregnant women; (B) Subgroup analysis based on the type of abortion (SA and RPL); (C) Sensitivity analysis; and (D) Funnel plot.

3.6 Meta-analysis for Pb

The meta-analysis of the association between Pb concentration and abortion included 15 studies (Figure 4A). The pooled circulating Pb concentration was significantly higher in women

who had experienced an abortion than in those with normal pregnancies (SMD = 1.47, 95% CI: 0.89–2.05, $p < 0.001$, $I^2 = 96.0\%$). Subgroup analysis for SA and RPL showed significantly higher Pb concentrations in women with SA (SMD = 0.33, 95% CI: 0.12–0.55, $p = 0.002$) and RPL (SMD = 8.19, 95% CI: 4.52–11.85,

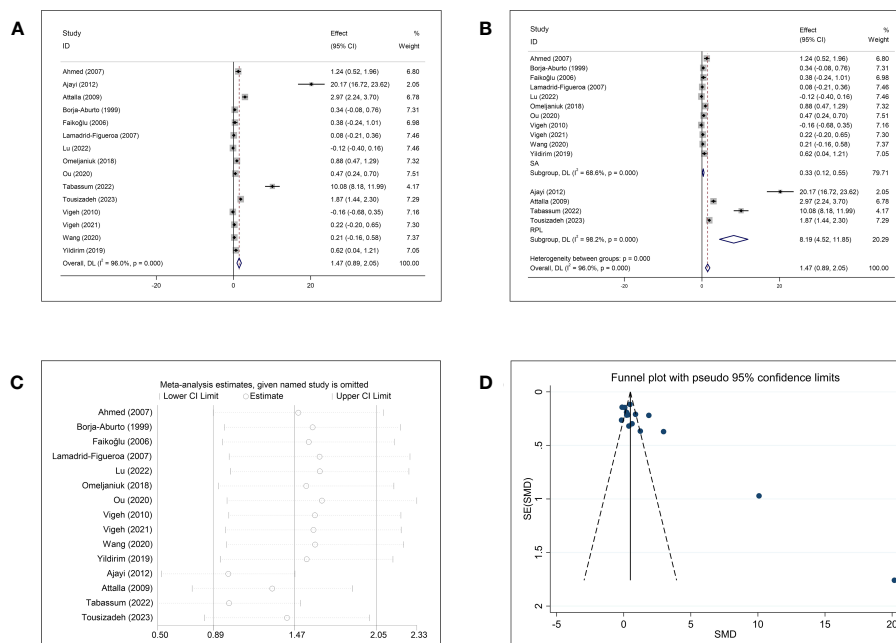


FIGURE 4

Meta-analysis outcomes of lead. (A) Forest plot showing the meta-analysis outcomes between abortion group and normal pregnant women; (B) Subgroup analysis based on the type of abortion (SA and RPL); (C) Sensitivity analysis; and (D) Funnel plot.

$p < 0.001$) than in healthy pregnant women (Figure 4B). However, significant heterogeneity was observed (SA: $p < 0.001$, $I^2 = 68.6\%$; RPL: $p < 0.001$, $I^2 = 98.2\%$). Further subgroup analyses based on the follow-up endpoint, continent, type of article, and type of detected sample also showed high heterogeneity (Supplementary Figure 3). Sensitivity analysis showed that omission of any single study did not change the overall effect (Figure 4C). Visual inspection of the funnel plots (Figure 4D) and Begg's and Egger's tests showed publication bias (Begg: $p = 0.002$; Egger: $p = 0.001$).

3.7 Meta-analysis for Cd

The pooled results of the meta-analysis of eight studies on Cd and abortion showed significantly higher Cd concentrations in women who underwent abortion than in normal pregnant women (SMD = 1.15, 95% CI: 0.45–1.85, $p = 0.001$, $I^2 = 93.7\%$) (Figure 5A). Subgroup analysis based on abortion type showed that women with SA and RPL had significantly higher Cd concentrations than the normal controls (SA: SMD = 0.42, 95% CI: 0.02–0.82, $p = 0.040$; RPL: SMD = 2.45, 95% CI: 0.85–4.04, $p = 0.003$). However, the heterogeneity was significant in each subgroup (SA: $p = 0.003$, $I^2 = 75.1\%$; RPL: $p < 0.001$, $I^2 = 94.2\%$) (Figure 5B). Further subgroup analyses based on the follow-up endpoint, continent, and type of detected sample also showed high heterogeneity (Supplementary Figure 4). All studies on Cd and abortion were case-control studies, and subgroup analysis for different article types could not be performed. The sensitivity analysis showed that the exclusion of any single study could change the overall effect (Figure 5C). Both funnel plots and

Begg's and Egger's tests showed no publication bias for Cd (Begg: $p = 0.174$; Egger: $p = 0.113$; Figure 5D).

4 Discussion

Due to widespread human exposure to (53) and bio-accumulation of heavy metals (54), there are growing concerns about the adverse effects of heavy metals on normal pregnancies. Exposure to toxic metals or deficiency of essential metals has long been suspected to lead to abortion (55). However, the results of previous studies have not been consistent (1, 19). To provide stronger evidence for this important clinical issue, we conducted the present meta-analysis, focusing on two common toxic metals (Pb and Cd) and two essential metals (Zn and Cu) (56). This study is first to investigate the overall association between blood Zn, Cu, Pb, and Cd concentrations and abortion, including RPL and SA. Zn or Cu deficiency was associated with the prevalence of abortion in women, and exposure to Pb or Cd increased the risk of abortion (SA and RPL). Only one relevant meta-analysis was carried out in 2021, showed that exposure to Cd and Pb increased the incidence of abortion (undistinguished threatened abortion, SA, and RPL) (23). Subgroup analysis based on abortion type was not performed (23). In the present study, we recruited more studies to reinforce the association between exposure to Cd and Pb and the increased risk of abortion and performed subgroup analysis based on the type of abortion to investigate the effect of Cd and Pb exposure on patients with SA and RPL. In addition, the exploration of Zn and Cu in women with RPL and SA provides a basis for clinicians who tend to intervene early against RPL in women with Zn and Cu deficiencies.

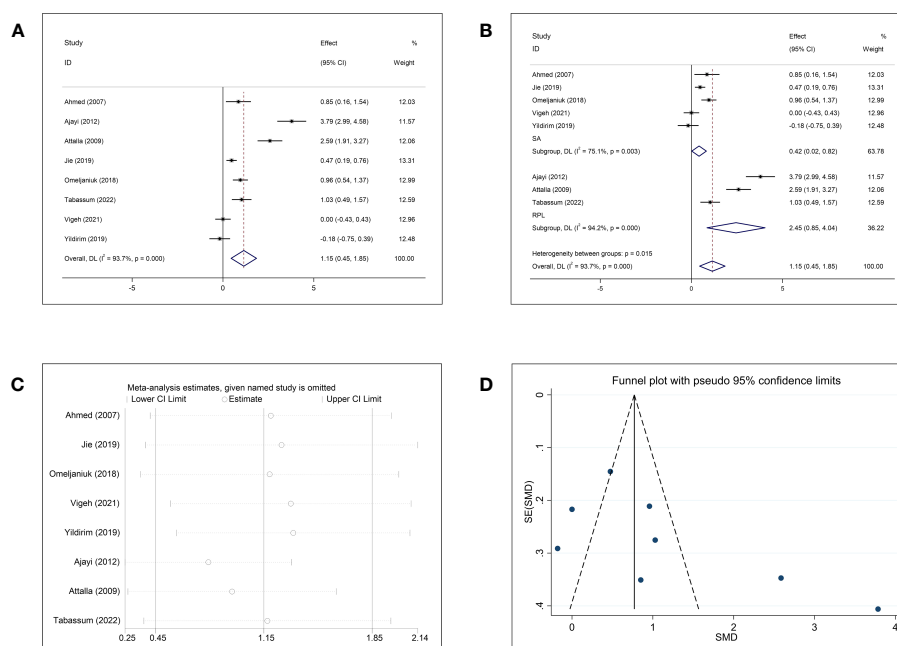


FIGURE 5

Meta-analysis outcomes of cadmium. (A) Forest plot showing the meta-analysis outcomes between abortion group and normal pregnant women; (B) Subgroup analysis based on the type of abortion (SA and RPL); (C) Sensitivity analysis; and (D) Funnel plot.

The exact mechanisms underlying the induction of SA and RPL by Pb and Cd exposure and Zn and Cu deficiency are unknown. Studies have shown that heavy metals are common environmental endocrine disruptors. Previous studies have reported that exposure to toxic metals and deficiencies of essential metals lead to abortion mainly through endocrine dysfunction, such as insulin resistance, vitamin D insufficiency, and abnormal thyroid and sex hormone concentrations, among others. The details of this process are discussed below.

4.1 Female Zn concentration and abortion

In our study, we found that Zn inadequacy tended to increase the chances of abortion, especially for RPL; however, it may not increase the incidence of SA. The underlying mechanism of Zn inadequacy-related RPL remains unknown. However, it may also be associated with endocrine dysfunction caused by Zn deficiency.

Zn deficiency has been reported to decrease insulin sensitivity and cause insulin resistance (IR) (57, 58), whereas Zn supplements can decrease IR (59, 60). IR, defined clinically as a decreased biological response to exogenous or endogenous insulin, can cause mitochondrial dysfunction in the placenta, diminished trophoblast invasion, a subclinical inflammatory state, and oxidative stress. These factors are all considered crucial in the pathophysiology of RPL (61–64). Zn can reinforce glucose transport into cells and potentiate insulin-induced glucose transport via the insulin signaling pathway (65). Zn can also act as an insulin mimetic to maintain glucose homeostasis, which may also be a mechanism underlying Zn deficiency-induced IR (66).

Apart from IR, Zn deficiency is also closely related to vitamin D deficiency, as Zn regulates the transcriptional activation of hormone-related genes via a cysteine-rich Zn-finger region in vitamin D receptors (VDRs) (67–69). Vitamin D plays a vital role in maintaining normal biological functions, such as calcium homeostasis, and cell proliferation, differentiation, and apoptosis, all of which are crucial for immunomodulation and normal pregnancy (70). Vitamin D inadequacy was reported to be associated with SA and RPL in a recent meta-analysis (71, 72). Supplementation with vitamin D can suppress inflammatory cytokine production and elevate the secretion of cathelicidin in decidual cells and trophoblasts, which can reduce the risk of abortion (73–75).

Zn deficiency appears to interfere with sex hormone synthesis and further causes RPL. Zn can affect the biosynthesis and function of sex hormones, such as progesterone and prolactin, by altering LH and FSH levels and inducing oxidative stress (17, 76–78). Zn may also promote estrogen release by forming ligand bonds with metal-binding sites on the estrogen receptor (ER) (79). Insufficient secretion of sex hormones, such as progesterone, testosterone, estrogen, and prolactin, can reduce endometrial receptivity and oocyte quality in women, which is related to RPL (55, 80, 81).

Furthermore, Zn is an essential trace element for thyroid function and homeostasis (82), and its deficiency can lead to hypothyroidism (82–84). Hypothyroidism and subclinical hypothyroidism can lead to poor pregnancy outcomes such as SA and RPL (85, 86). Zn supplements can elevate thyroxine (T4) concentrations and reduce triiodothyronine (T3) concentrations

by altering the expression of key genes (*nis*, *tpo*, *thrα*, *dio1*, *dio2*, and *ugt1ab*) in the hypothalamic–pituitary–thyroid (HPT) axis (87).

4.2 Female Cu concentration and abortion

We found that women undergoing abortion (both SA and RPL) had lower Cu concentrations, indicating that Cu deficiency may be closely related to the incidence of abortion (SA and RPL). However, the underlying mechanism remains unknown. However, previous studies have reported that Cu deficiency can induce endocrine dysfunction, such as IR, vitamin D insufficiency, and abnormal thyroid and sex hormone concentrations, which may be involved in the pathogenesis of SA and RPL. Insufficient Cu can cause IR by upregulating cytochrome c oxidase 1 (SCO1) and vascular adhesion protein-1 (VAP-1) (88–90); it can also reduce progesterone synthesis by regulating the expression of steroidogenic factor 1 (SF-1) (91). Cu deficiency can also reduce Cu/Zn superoxide dismutase (Cu/Zn-SOD) and cause oxidative stress in the ovary, ultimately leading to dysfunctional luteal formation and insufficient progesterone secretion (17). In addition, Cu deficiency can decrease the expression of estrogen synthetases such as aromatase (CYP19A1) and 17β-hydroxysteroid dehydrogenase (17β-HSD) (92). Furthermore, Cu deficiency can lead to hypothyroidism (82–84, 93) by inducing oxidative stress and decreasing thyroxine synthesis by limiting tyrosinase availability (82, 93–95).

4.3 Female Pb concentration and abortion

Our study found that women who experienced abortion (SA and RPL) had higher Pb concentrations, suggesting that Pb exposure could increase the risk of abortion (SA and RPL). Our results are consistent with those of the meta-analysis by Kaur et al. (23). Pb can substitute polyvalent cations, such as calcium (Ca²⁺), and affect various cellular processes, such as apoptosis, cell adhesion, and cell signaling (96). However, the mechanism underlying Pb-induced abortion remains unclear. Animal studies have shown that Pb exposure can downregulate IR-related genes in the PI3K and Akt signaling pathways, which are involved in hepatic gluconeogenesis and glucose production (97, 98). Low-level Pb exposure promotes the gene expression of key enzymes involved in hepatic gluconeogenesis and eventually induces hyperglycemia and impaired fasting plasma glucose, which is known as hepatic insulin resistance (99). Additionally, Pb appears to be involved in the pathology of vitamin D deficiency. Pb can diminish the activity of vitamin D by blocking the normal renal synthesis of active 1,25-dihydroxy vitamin D (1,25(OH)2D) and reduce the generation of vitamin D binding protein (DBP) (56, 100, 101). Pb can also promote degradation and block the synthesis of 1,25(OH)2D3 by upregulating the hepatic expression of Cyp24a1 enzymes and inhibiting 25-hydroxylase (CYP2R1) and 1-α-hydroxylase (CYP27B1) at the gene and protein levels (100, 102). Previous studies have suggested that Pb may be closely associated with luteal phase deficiencies. Pb can directly inhibit the expression of several key enzymes involved in progesterone synthesis, such as StAR, CYP11A1, and 3β-HSD (103, 104). Pb also appears to indirectly

interfere with progesterone synthesis by inhibiting the cAMP-PKA-dependent signaling pathway that regulates the expression of these key enzymes (104–106). Pb has adverse effects on sex hormone concentration. Pb exposure is associated with increased testosterone and prolactin concentrations and appears to reduce estrogen concentrations by decreasing the expression of estrogen synthases such as 17 β -HSD (103, 107, 108). Furthermore, Pb accumulation negatively affects thyroid function, which is also related to abortion. Excessive exposure to Pb may lead to hypo- or hyperthyroidism (109). As an oxidant, Pb can negatively impact thyroid cells by promoting oxidative stress, and it can also interact with other essential elements such as Cu, Zn, and Fe to indirectly affect thyroid function (82, 109).

4.4 Female Cd concentration and abortion

We found that the Cd concentration was significantly higher in women who experienced abortion (SA and RPL) than in normal pregnant women. Our results are in line with those of Kaur et al., who revealed that Cd exposure could increase the risk of abortion (23). Cd is a highly potent environmental pollutant that causes indirect oxidative damage to DNA, leading to the induction of cellular proliferation and inhibition of DNA repair mechanisms, causing cytotoxicity (110). However, research on the mechanisms of Cd exposure-related abortion is lacking. In recent years, an increasing number of studies have found a strong relationship between Cd and endocrine dysfunction, which is the main reason for abortions (both SA and RPL). Epidemiological surveys have shown that Cd can cause IR through perturbations in gluconeogenesis, pancreatic islet dysfunction, and metabolic and mitogen impairments in the liver and adipose tissue (111, 112). Epidemiological studies have also demonstrated that high blood Cd concentrations are negatively correlated with vitamin D concentrations (113, 114), which may be due to the interaction of Cd with renal mitochondrial hydroxylases (115). Cd is also involved in the pathogenesis of luteal phase deficiencies. It can directly or indirectly inhibit the expression of several key enzymes (StAR, CYP11A1, and 3 β -HSD) involved in progesterone synthesis by regulating the cAMP-PKA-dependent signaling pathway (24, 116, 117). Cd can also interfere with the balance of sex hormone concentrations. Cd exposure appears to decrease the expression of estrogen synthetases (CYP19A1 and 17 β -HSD), and it is also a potent xenoestrogen that can mediate the proliferation of anterior pituitary cells and prolactin secretion by mimicking estrogen (118). Cd can also negatively affect thyroid cells by promoting oxidative stress, ultimately leading to thyroid dysfunction (82, 109).

4.5 Strengths

The strength of our study is that we comprehensively investigated the relationships between blood Zn, Cu, Pb, and Cd concentrations and abortion rates (SA and RPL). In addition, we systematically reviewed previous publications on the endocrine mechanisms of metal exposure-related abortions. We propose that IR, vitamin D insufficiency, and abnormal thyroid and sex hormone concentrations

may be involved in Zn and Cu deficiencies and Pb and Cd exposure-associated abortions. Third, the large sample size of 4,666 pregnant women from 14 countries makes our estimates reliable. Fourth, most included studies were of good quality. In the included studies, the definitions of cases and controls were adequate, and the selection of controls and assessment of exposure were consistent. Fifth, a sensitivity analysis was conducted to verify the associations between the four metals and abortion.

4.6 Limitations

This study has some limitations. First, several dated documents that appeared to meet our inclusion criteria were not included because we were unable to reach the authors. Second, despite our best efforts, we were only able to find 28 related papers because of the relatively large number of animal studies and case reports. Third, most meta-analyses included in our study had high heterogeneity. To ascertain its sources, we performed a subgroup analysis based on the type of abortion, follow-up endpoint, continent, type of observational study, and type of detected sample. However, we failed to find sources of heterogeneity by subgroup analysis, as most subgroup analyses showed high heterogeneity. After carefully reviewing the included articles, we found that different diagnoses of SA or RPL may have led to clinical heterogeneity. In addition, regional variations in metal concentrations of the study participants were considered another source of heterogeneity. Local mineral deposits and their exploitation affect the metal concentrations in the environment (air, water, and soil), and different terrains can impact the diffusion of pollutants (119). Thus, participants had different risks of metal exposure. Moreover, participants of different races with different genetic backgrounds have various sensitivities to metal exposure (120). Owing to the limited information regarding the region and race of the investigated subjects in the original literature, we were only able to perform a subgroup analysis based on different continents to investigate the heterogeneity caused by regional differences. The age of the study population and the time point of blood collection in each study were also considered potential sources of heterogeneity, as heavy metals could accumulate in the human body, and older adults may have higher blood metal concentrations. Furthermore, metal concentrations can change during different trimesters and the time points of blood collection may lead to heterogeneity (121, 122). Fourth, the literature regarding Cu and Pb had obvious publication and reporting bias, although the publication and reporting the bias of literature regarding Zn and Cd were acceptable. Language and multiple publication biases were considered primary problems as only the English literature was included, and two studies had outcomes from the same study population.

4.7 Implications for treatment

The findings of the present study broaden our understanding of the effects of toxic and essential metals on the RPL and SA. Endocrine dysfunction can lead to metal exposure and abortions. It will be helpful to screen blood Zn, Cu, Pb, and Cd concentrations in females. However, well-designed prospective cohort studies are

needed to clarify the causal relationship between endocrine dysfunction and heavy-metal-induced abortion.

5 Conclusion

In the present study, we found that higher blood Pb and Cd concentrations and lower Zn and Cu concentrations in females may be associated with SA and RPL. Exposure to toxic metals, as well as deficiencies in essential metals, may cause SA and RPL through endocrine dysfunction, such as insulin resistance, vitamin D insufficiency, and abnormal thyroid and sex hormone concentrations. However, further prospective cohort and experimental studies are required to provide stronger evidence.

Data availability statement

The original contributions presented in the study are included in the article/**Supplementary Material**. Further inquiries can be directed to the corresponding authors.

Author contributions

XS and MR proposed the subject and designed the protocol for this systematic review. MR, LTW, LQW and JC conducted literature screening and data extraction. MR, LQW, and LTW assessed the quality of all studies. LTW and MR performed statistical analysis. MR, LTW, and LQW produced the tables, figures. MR, LQW and LTW drafted the manuscript. XS and SQ gave overall supervision, critical revisions, and final approval of the article. All authors listed have made a substantial, direct, and intellectual contribution to the work and approved it for publication.

Funding

This work was supported by grants from the National Natural Science Foundation for Young Scholar of China (No 82101683), the Natural Science Foundation of Guangdong Province (2022A1515010245), and the Medical Scientific Research Foundation of Guangdong Province of China (A2021150).

Acknowledgments

We are grateful to Zhekai Cui, Haoqing Liang, Yuxin Yao, Shuying Lin, Haochen Ai, Huihao Ye, and Hongjie Zou for their help on this project.

References

1. Thaker R, Oza H, Shaikh I, Kumar S. Correlation of copper and zinc in spontaneous abortion. *Int J Fertil Steril* (2019) 13(2):97–101. doi: 10.22074/ijfs.2019.5586
2. La X, Wang W, Zhang M, Liang L. Definition and multiple factors of recurrent spontaneous abortion. *Adv Exp Med Biol* (2021) 1300:231–57. doi: 10.1007/978-981-33-4187-6_11

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fendo.2023.1216507/full#supplementary-material>

SUPPLEMENTARY FIGURE 1

Subgroup analysis of circulating Zn level in abortion women or healthy pregnant women. (A). Subgroup analysis based on the follow-up endpoint (ongoing pregnancy and live birth); (B). Subgroup analysis based on the continent of study population (Africa, Asia, Europe, and Oceania); (C). Subgroup analysis based on the type of article (case-control study, cross-section study, and nested case-control study); (D). Subgroup analysis based on the type of detected sample (serum, plasma, and whole blood).

SUPPLEMENTARY FIGURE 2

Subgroup analysis of circulating Cu level in abortion women or healthy pregnant women. (A). Subgroup analysis based on the follow-up endpoint (ongoing pregnancy and live birth); (B). Subgroup analysis based on the continent of study population (Africa, Asia, Europe, and Oceania); (C). Subgroup analysis based on the type of article (case-control study, cross-section study, and nested case-control study); (D). Subgroup analysis based on the type of detected sample (serum, plasma, and whole blood).

SUPPLEMENTARY FIGURE 3

Subgroup analysis of circulating Pb level in abortion women or healthy pregnant women. (A). Subgroup analysis based on the follow-up endpoint (ongoing pregnancy and live birth); (B). Subgroup analysis based on the continent of study population (Africa, North America, Asia, and Europe); (C). Subgroup analysis based on the type of article (case-control study and cross-section study); (D). Subgroup analysis based on the type of detected sample (serum, plasma, and whole blood).

SUPPLEMENTARY FIGURE 4

Subgroup analysis of circulating Cd level in abortion women or healthy pregnant women. (A). Subgroup analysis based on the follow-up endpoint (ongoing pregnancy and live birth); (B). Subgroup analysis based on the continent of study population (Africa, Asia, and Europe); (C). Subgroup analysis based on the type of detected sample (serum and whole blood).

3. American College of Obstetricians and Gynecologists' Committee on Practice Bulletins—Gynecology. ACOG practice bulletin no. 200 summary: early pregnancy loss. *Obstet Gynecol* (2018) 132(5):1311–3. doi: 10.1097/aog.0000000000002900
4. Stray-Pedersen B, Lorentzen-Styr AM. The prevalence of toxoplasma antibodies among 11,736 pregnant women in Norway. *Scand J Infect Dis* (1979) 11(2):159–65. doi: 10.3109/inf.1979.11.issue-2.12
5. Bender Atik R, Christiansen OB, Elson J, Kolte AM, Lewis S, Middeldorp S, et al. ESHRE guideline: recurrent pregnancy loss. *Hum Reprod Open* (2018) 2018(2):hoy004. doi: 10.1093/hropen/hoy004
6. Practice Committee of the American Society for Reproductive Medicine. Definitions of infertility and recurrent pregnancy loss: a committee opinion. *Fertil Steril* (2013) 99(1):63. doi: 10.1016/j.fertnstert.2012.09.023
7. Toth B, Würfel W, Bohlmann M, Zschocke J, Rudnik-Schöneborn S, Nawroth F, et al. Recurrent miscarriage: diagnostic and therapeutic procedures. Guideline of the DGGG, OEGGG and SGGG (S2k-level, AWMF registry number 015/050). *Geburtshilfe Frauenheilkd* (2018) 78(4):364–81. doi: 10.1055/a-0586-4568
8. Krog MC, Nielsen HS, Christiansen OB, Kolte AM. Reproductive endocrinology in recurrent pregnancy loss. *Clin Obstet Gynecol* (2016) 59(3):474–86. doi: 10.1097/grf.0000000000000225
9. van den Berg MM, van Maarle MC, van Wely M, Goddijn M. Genetics of early miscarriage. *Biochim Biophys Acta* (2012) 1822(12):1951–9. doi: 10.1016/j.bbdis.2012.07.001
10. Ageron A, Bhattacharya S. Infertility and miscarriage: common pathways in manifestation and management. *Womens Health (Lond)* (2015) 11(4):527–41. doi: 10.2217/whc.15.19
11. Baser E, Kirmizi DA, Turksoy VA, Onat T, Çaltekin MD, Kara M, et al. Environmental exposures in the etiology of abortion: placental toxic and trace element levels. *Z Geburtshilfe Neonatol* (2020) 224(6):339–47. doi: 10.1055/a-1263-1698
12. Dutta S, Gorain B, Choudhury H, Roychoudhury S, Sengupta P. Environmental and occupational exposure of metals and female reproductive health. *Environ Sci Pollut Res Int* (2022) 29(41):62067–92. doi: 10.1007/s11356-021-16581-9
13. Tchounwou PB, Yedjou CG, Patlolla AK, Sutton DJ. Heavy metal toxicity and the environment. *Exp Suppl* (2012) 101:133–64. doi: 10.1007/978-3-7643-8340-4_6
14. Briffa J, Sinagra E, Blundell R. Heavy metal pollution in the environment and their toxicological effects on humans. *Heliyon* (2020) 6(9):e04691. doi: 10.1016/j.heliyon.2020.e04691
15. Grzeszczak K, Kwiatkowski S, Kosik-Bogacka D. The role of Fe, Zn, and Cu in pregnancy. *Biomolecules* (2020) 10(8):1176. doi: 10.3390/biom10081176
16. Wilson RL, Grieger JA, Bianco-Miotto T, Roberts CT. Association between maternal zinc status, dietary zinc intake and pregnancy complications: A systematic review. *Nutrients* (2016) 8(10):641. doi: 10.3390/nu8100641
17. Noda Y, Ota K, Shirasawa T, Shimizu T. Copper/zinc superoxide dismutase insufficiency impairs progesterone secretion and fertility in female mice. *Biol Reprod* (2012) 86(1):1–8. doi: 10.1095/biolreprod.111.092999
18. Rahman A, Kumarathasan P, Gomes J. Infant and mother related outcomes from exposure to metals with endocrine disrupting properties during pregnancy. *Sci Total Environ* (2016) 569–570:1022–31. doi: 10.1016/j.scitotenv.2016.06.134
19. Omeljaniuk WJ, Socha K, Soroczynska J, Charkiewicz AE, Laudanski T, Kulikowski M, et al. Cadmium and lead in women who miscarried. *Clin Lab* (2018) 64(1):59–67. doi: 10.7754/Clin.Lab.2017.170611
20. Alrashed M, Tabassum H, Almuhareb N, Almutlaq N, Alamro W, Alanazi ST, et al. Assessment of DNA damage in relation to heavy metal induced oxidative stress in females with recurrent pregnancy loss (RPL). *Saudi J Biol Sci* (2021) 28(9):5403–7. doi: 10.1016/j.sjbs.2021.05.068
21. Jahan Toma N, Anwar S, Kabir T, Hosen MJ. Lead and lead-arsenic combined exposure induces mortality and developmental impairments in zebrafish embryos: a study using wild-caught zebrafish from Bangladesh. *Drug Chem Toxicol* (2022) 45(6):2833–42. doi: 10.1080/01480545.2021.1996594
22. Amadi CN, Igweze ZN, Orisakwe OE. Heavy metals in miscarriages and stillbirths in developing nations. *Middle East Fertil Soc J* (2017) 22(2):91–100. doi: 10.1016/j.mefs.2017.03.003
23. Kaur M, Sharma P, Kaur R, Khetarpal P. Increased incidence of spontaneous abortions on exposure to cadmium and lead: a systematic review and meta-analysis. *Gynecol Endocrinol* (2022) 38(1):16–21. doi: 10.1080/09513590.2021.1942450
24. Belani M, Shah P, Banker M, Gupta S. Dual effect of insulin resistance and cadmium on human granulosa cells - *In vitro* study. *Toxicol Appl Pharmacol* (2016) 313:119–30. doi: 10.1016/j.taap.2016.10.019
25. Wells GA, Shea B, O'Connell D, Peterson J, Welch V, Losos M, et al. *The Newcastle-Ottawa Scale (NOS) for assessing the quality of nonrandomised studies in meta-analyses: The Ottawa Hospital Research Institute* (2021). Available at: https://www.ohri.ca/programs/clinical_epidemiology/oxford.asp.
26. Ahmed H, El-Desouky AI, Rashed LA. Role of antioxidants, environmental exposure to lead and cadmium in spontaneous abortion. *Egypt J Occup Med* (2007) 31:217–36.
27. Ajayi OO, Charles-Davies MA, Arinola OG. Progesterone, selected heavy metals and micronutrients in pregnant Nigerian women with a history of recurrent spontaneous abortion. *Afr Health Sci* (2012) 12(2):153–9. doi: 10.4314/ahs.v12i2.12
28. Alebic-Juretic A, Frkovic A. Plasma copper concentrations in pathological pregnancies. *J Trace Elem Med Biol* (2005) 19(2-3):191–4. doi: 10.1016/j.jtemb.2005.08.002
29. Al-Sheikh YA, Ghneim HK, Alharbi AF, Alshehly MM, Aljaser FS, Aboul-Soud MAM. Molecular and biochemical investigations of key antioxidant/oxidant molecules in Saudi patients with recurrent miscarriage. *Exp Ther Med* (2019) 18(6):4450–60. doi: 10.3892/etm.2019.8082
30. Attalla S, Eldakroory S, Mosad S, Goda H. A comparative study of lead, cadmium, zinc and selenium concentration in pregnant and aborted woman. *Mansoura J Forensic Med Clin Toxicol* (2009) 17(2):27–41.
31. Bassiouni BA, Rafei AA. 5-Hydroxytryptamine (serotonin), copper and ceruloplasmin plasma concentrations in spontaneous abortion. *Eur J Obstet Gynecol Reprod Biol* (1979) 9(2):81–8. doi: 10.1016/0028-2243(79)90003-0
32. Borella P, Szilagy A, Than G, Csaba I, Giardino A, Facchinetti F. Maternal plasma concentrations of magnesium, calcium, zinc and copper in normal and pathological pregnancies. *Sci Total Environ* (1990) 99(1-2):67–76. doi: 10.1016/0048-9697(90)90212-d
33. Borja-Aburto VH, Hertz-Picciotto I, Rojas Lopez M, Farias P, Rios C, Blanco J. Blood lead levels measured prospectively and risk of spontaneous abortion. *Am J Epidemiol* (1999) 150(6):590–7. doi: 10.1093/oxfordjournals.aje.a10057
34. Dreosti IE, MacLennan A. Maternal plasma zinc levels and first trimester abortion. *Early Hum Dev* (1990) 21(2):141–2. doi: 10.1016/0378-3782(90)90069-u
35. Faikoglu R, Savan K, Utku Ç, Takar N, Zebitay AG. Significance of maternal plasma lead level in early pregnancy loss. *J Environ Sci Health - Part A Toxic/Hazardous Substances Environ Eng* (2006) 41(3):501–6. doi: 10.1080/10934520500428435
36. Ghneim HK, Al-Sheikh YA, Alshehly MM, Aboul-Soud MA. Superoxide dismutase activity and gene expression levels in Saudi women with recurrent miscarriage. *Mol Med Rep* (2016) 13(3):2606–12. doi: 10.3892/mmr.2016.4807
37. Ghosh A, Fong LY, Wan CW, Liang ST, Woo JS, Wong V. Zinc deficiency is not a cause for abortion, congenital abnormality and small-for-gestational age infant in Chinese women. *Br J Obstet Gynaecol* (1985) 92(9):886–91. doi: 10.1111/j.1471-0528.1985.tb03067.x
38. Jie O, Peng P, Qiu L, Teng L, Li C, Han J, et al. Biomarkers of metal toxicity in embryos in the general population. *J Clin Lab Anal* (2019) 33(8):e22974. doi: 10.1002/jcla.22974
39. Lamadrid-Figueroa H, Téllez-Rojo MM, Hernández-Avila M, Trejo-Valdivia B, Solano-González M, Mercado-García A, et al. Association between the plasma/whole blood lead ratio and history of spontaneous abortion: a nested cross-sectional study. *BMC Pregnancy Childbirth* (2007) 7:22. doi: 10.1186/1471-2393-7-22
40. Lu Y, Zhang Y, Guan Q, Xu L, Zhao S, Duan J, et al. Exposure to multiple trace elements and miscarriage during early pregnancy: A mixtures approach. *Environ Int* (2022) 162:107161. doi: 10.1016/j.envint.2022.107161
41. Omeljaniuk WJ, Socha K, Borawska MH, Charkiewicz AE, Laudanski T, Kulikowski M, et al. Antioxidant status in women who have had a miscarriage. *Adv Med Sci* (2015) 60(2):329–34. doi: 10.1016/j.advms.2015.06.003
42. Ou J, Peng P, Qiu L, Teng L, Li C, Han J, et al. Effect of lead exposure on spontaneous abortion: a case-control study. *Clin Lab* (2020) 66(5):1–7. doi: 10.7754/Clin.Lab.2019.190940
43. Popovic JK, Grujic Z, Grujic I, Bogavac M, Celic D, Popovic KJ, et al. Prostaglandin E-2, trace elements and levels of oxidative processes in spontaneous miscarriages. *Eur Rev Med Pharmacol Sci* (2016) 20(22):4786–90.
44. Sairoz, Prabhu K, Poojari VG, Shetty S, Rao M, Kamath A. Maternal serum zinc, copper, magnesium, and iron in spontaneous abortions. *Indian J Clin Biochem* (2023) 38(1):128–31. doi: 10.1007/s12291-022-01043-x
45. Shen PJ, Gong B, Xu FY, Luo Y. Four trace elements in pregnant women and their relationships with adverse pregnancy outcomes. *Eur Rev Med Pharmacol Sci* (2015) 19(24):4690–7.
46. Skalnaya MG, Tinkov AA, Lobanova YN, Chang JS, Skalny AV. Serum levels of copper, iron, and manganese in women with pregnancy, miscarriage, and primary infertility. *J Trace Elem Med Biol* (2019) 56:124–30. doi: 10.1016/j.jtemb.2019.08.009
47. Tabassum H, Alrashed M, Malik A, Alanazi ST, Alenzi ND, Ali MN, et al. A unique investigation of thallium, tellurium, osmium, and other heavy metals in recurrent pregnancy loss: A novel approach. *Int J Gynecol Obstet* (2023) 160(3):790–6. doi: 10.1002/ijgo.14390
48. Tousizadeh S, Mohammadi-Moghadam F, Sadeghi R, Ahmadi A, Shakeri K. Investigation of the levels of essential and non-essential metals in women with and without abortion history: A study based on the Persian population of the Shahrekord cohort. *Chemosphere* (2023) 329:138434. doi: 10.1016/j.chemosphere.2023.138434
49. Vigeh M, Yokoyama K, Kitamura F, Afshinrokh M, Beygi A, NiroOmanesh S. Early pregnancy blood lead and spontaneous abortion. *Women Health* (2010) 50(8):756–66. doi: 10.1080/03630242.2010.532760
50. Vigeh M, Yunesian M, Matsukawa T, Shamsipour M, Jeddi MZ, Rastkari N, et al. Prenatal blood levels of some toxic metals and the risk of spontaneous abortion. *J Environ Health Sci Eng* (2021) 19(1):357–63. doi: 10.1007/s40201-020-00608-3
51. Wang R, Zhang L, Chen Y, Zhang S, Zhuang T, Wang L, et al. Elevated non-essential metals and the disordered metabolism of essential metals are associated to abnormal pregnancy with spontaneous abortion. *Environ Int* (2020) 144:106061. doi: 10.1016/j.envint.2020.106061

52. Yildirim E, Derici MK. The effect of heavy metals on miscarriage. *J Clin Obstet Gynecol* (2019) 29(1):31–8. doi: 10.5336/jcog.2018-64175
53. Zhou L, Liang K, Li M, Rong C, Zheng J, Li J. Metal elements associate with in vitro fertilization (IVF) outcomes in 195 couples. *J Trace Elem Med Biol* (2021) 68:126810. doi: 10.1016/j.jtemb.2021.126810
54. Rzymiski P, Niedzielski P, Klimaszczak P, Poniedziałek B. Bioaccumulation of selected metals in bivalves (Unionidae) and *Phragmites australis* inhabiting a municipal water reservoir. *Environ Monit Assess* (2014) 186(5):3199–212. doi: 10.1007/s10661-013-3610-8
55. Lee WL, Yeh CC, Wang PH. Risk to increase threatened abortion: deficiency of some essential trace elements and exposure of toxic heavy metals. *J Chin Med Assoc* (2019) 82(8):607–8. doi: 10.1097/jcma.000000000000133
56. Schwalfenberg GK, Genies SJ. Vitamin D, essential minerals, and toxic elements: exploring interactions between nutrients and toxicants in clinical medicine. *ScientificWorldJournal* (2015) 2015:318595. doi: 10.1155/2015/318595
57. Yang HK, Lee SH, Han K, Kang B, Lee SY, Yoon KH, et al. Lower serum zinc levels are associated with unhealthy metabolic status in normal-weight adults: The 2010 Korea National Health and Nutrition Examination Survey. *Diabetes Metab* (2015) 41(4):282–90. doi: 10.1016/j.diabet.2015.03.005
58. Kant R, Verma V, Patel S, Chandra R, Chaudhary R, Shuldiner AR, et al. Effect of serum zinc and copper levels on insulin secretion, insulin resistance and pancreatic β cell dysfunction in US adults: Findings from the National Health and Nutrition Examination Survey (NHANES) 2011–2012. *Diabetes Res Clin Pract* (2021) 172:108627. doi: 10.1016/j.diabres.2020.108627
59. Karandish M, Mozaffari-Khosravi H, Mohammadi SM, Cheraghian B, Azhdari M. The effect of curcumin and zinc co-supplementation on glycemic parameters in overweight or obese prediabetic subjects: A phase 2 randomized, placebo-controlled trial with a multi-arm, parallel-group design. *Phytother Res* (2021) 35(8):4377–87. doi: 10.1002/ptr.7136
60. Cruz KJ, Morais JB, de Oliveira AR, Severo JS, Marreiros DD. The effect of zinc supplementation on insulin resistance in obese subjects: a systematic review. *Biol Trace Elem Res* (2017) 176(2):239–43. doi: 10.1007/s12011-016-0835-8
61. Lebovitz HE. Insulin resistance: definition and consequences. *Exp Clin Endocrinol Diabetes* (2001) 109 Suppl 2:S135–48. doi: 10.1055/s-2001-18576
62. Cai WY, Luo X, Lv HY, Fu KY, Xu J. Insulin resistance in women with recurrent miscarriage: a systematic review and meta-analysis. *BMC Pregnancy Childbirth* (2022) 22(1):916. doi: 10.1186/s12884-022-05256-z
63. Zhang Y, Zhao W, Xu H, Hu M, Guo X, Jia W, et al. Hyperandrogenism and insulin resistance-induced fetal loss: evidence for placental mitochondrial abnormalities and elevated reactive oxygen species production in pregnant rats that mimic the clinical features of polycystic ovary syndrome. *J Physiol* (2019) 597(15):3927–50. doi: 10.1113/jp277879
64. Azizi R, Soltani-Zangbar MS, Sheikhsani G, Pourmoghadam Z, Mehdizadeh A, Mahdipour M, et al. Metabolic syndrome mediates inflammatory and oxidative stress responses in patients with recurrent pregnancy loss. *J Reprod Immunol* (2019) 133:18–26. doi: 10.1016/j.jri.2019.05.001
65. Tang X, Shay NF. Zinc has an insulin-like effect on glucose transport mediated by phosphoinositide-3-kinase and Akt in 3T3-L1 fibroblasts and adipocytes. *J Nutr* (2001) 131(5):1414–20. doi: 10.1093/jn/131.5.1414
66. Norouzi S, Adulcikas J, Sohal SS, Myers S. Zinc stimulates glucose oxidation and glycemic control by modulating the insulin signaling pathway in human and mouse skeletal muscle cell lines. *PLoS One* (2018) 13(1):e0191727. doi: 10.1371/journal.pone.0191727
67. Herrera-Quintana L, Vázquez-Lorente H, Molina-López J, Gamarrá-Morales Y, Martín-López JI, Planells E. Vitamin D status in critically ill patients with SIRS and its relationship with circulating zinc and related parameters during ICU stay. *Nutrients* (2022) 14(17):3580. doi: 10.3390/nu14173580
68. Potocnik FC, van Rensburg SJ, Hon D, Emsley RA, Moodie IM, Erasmus RT. Oral zinc augmentation with vitamins A and D increases plasma zinc concentration: implications for burden of disease. *Metab Brain Dis* (2006) 21(2–3):139–47. doi: 10.1007/s11011-006-9023-4
69. Amos A, Razaque MS. Zinc and its role in vitamin D function. *Curr Res Physiol* (2022) 5:203–7. doi: 10.1016/j.crphys.2022.04.001
70. Jones G, Strugnell SA, DeLuca HF. Current understanding of the molecular actions of vitamin D. *Physiol Rev* (1998) 78(4):1193–231. doi: 10.1152/physrev.1998.78.4.1193
71. Tamblin JA, Pilarski NSP, Markland AD, Marson EJ, Devall A, Hewison M, et al. Vitamin D and miscarriage: a systematic review and meta-analysis. *Fertil Steril* (2022) 118(1):111–22. doi: 10.1016/j.fertnstert.2022.04.017
72. Gonçalves DR, Braga A, Braga J, Marinho A. Recurrent pregnancy loss and vitamin D: A review of the literature. *Am J Reprod Immunol* (2018) 80(5):e13022. doi: 10.1111/ajri.13022
73. Hou H, Zhang JY, Chen D, Deng F, Morse AN, Qiu X, et al. Altered decidual and placental catabolism of vitamin D may contribute to the aetiology of spontaneous miscarriage. *Placenta* (2020) 92:1–8. doi: 10.1016/j.placenta.2020.01.013
74. Evans KN, Nguyen L, Chan J, Innes BA, Bulmer JN, Kilby MD, et al. Effects of 25-hydroxyvitamin D3 and 1,25-dihydroxyvitamin D3 on cytokine production by human decidual cells. *Biol Reprod* (2006) 75(6):816–22. doi: 10.1095/biolreprod.106.054056
75. Liu N, Kaplan AT, Low J, Nguyen L, Liu GY, Equils O, et al. Vitamin D induces innate antibacterial responses in human trophoblasts via an intracrine pathway. *Biol Reprod* (2009) 80(3):398–406. doi: 10.1095/biolreprod.108.073577
76. Pakys K, Varga B, Lázár P. Zinc protection against cadmium-induced infertility in female rats. Effect of zinc and cadmium on the progesterone production of cultured granulosa cells. *Biometals* (1997) 10(1):27–35. doi: 10.1023/a:1018362603065
77. Kochman K, Gajewska A, Kozłowski H, Masiukiewicz E, Rzeszotarska B. Increased LH and FSH release from the anterior pituitary of ovariectomized rat, in vivo, by copper-, nickel-, and zinc-LHRH complexes. *J Inorg Biochem* (1992) 48(1):41–6. doi: 10.1016/0162-0134(92)80051-v
78. Han Q, Yan X, Ye Y, Han L, Ma X, Wang T, et al. ZBTB20 regulates prolactin expression and lactotrope function in adult mice. *Endocrinology* (2022) 163(12):bqac181. doi: 10.1210/endo/bqac181
79. Humeny A, Bökenkamp D, Thole HH. The HDQVH-motif in domain E of the estradiol receptor α is responsible for zinc-binding and zinc-induced hormone release. *Mol Cell Endocrinol* (1999) 153(1–2):71–8. doi: 10.1016/s0303-7207(99)00089-1
80. Li J. [Research advances in the relationship between prolactin and spontaneous abortion]. *Zhong Xi Yi Jie He Xue Bao* (2012) 10(1):7–12. doi: 10.3736/jcim20120102
81. Semenik LM, Likhachov VK, Yuzvenko TY, Dobrovolska L, Makarov OG. Risk markers of reproductive loss in women with hyperandrogenism. *Wiad Lek* (2018) 71(8):1550–3.
82. Zhou Q, Xue S, Zhang L, Chen G. Trace elements and the thyroid. *Front Endocrinol (Lausanne)* (2022) 13:904889. doi: 10.3389/fendo.2022.904889
83. Knezevic J, Starchl C, Tmava Berisha A, Amrein K. Thyroid-gut-axis: how does the microbiota influence thyroid function? *Nutrients* (2020) 12(6):1769. doi: 10.3390/nu12061769
84. Talebi S, Ghaedi E, Sadeghi E, Mohammadi H, Hadi A, Clark CCT, et al. Trace element status and hypothyroidism: A systematic review and meta-analysis. *Biol Trace Elem Res* (2020) 197(1):1–14. doi: 10.1007/s12011-019-01963-5
85. Abalovich M, Gutierrez S, Alcaraz G, Maccallini G, Garcia A, Levalle O. Overt and subclinical hypothyroidism complicating pregnancy. *Thyroid* (2002) 12(1):63–8. doi: 10.1089/105072502753451986
86. Pluchino N, Drakopoulos P, Wenger JM, Petignat P, Streuli I, Genazzani AR. Hormonal causes of recurrent pregnancy loss (RPL). *Hormones (Athens)* (2014) 13(3):314–22. doi: 10.14310/horm.2002.1505
87. Zhong L, Zhang H, Wu L, Ru H, Wei N, Yao F, et al. Copper and zinc treatments alter the thyroid endocrine system in zebrafish embryos/larvae. *Toxics* (2022) 10(12):756. doi: 10.3390/toxics10120756
88. Wei XB, Guo L, Liu Y, Zhou SR, Liu Y, Dou X, et al. Synthesis of cytochrome c oxidase 1 (SCO1) inhibits insulin sensitivity by decreasing copper levels in adipocytes. *Biochem Biophys Res Commun* (2017) 491(3):814–20. doi: 10.1016/j.bbrc.2017.06.124
89. Bligt-Lindén E, Pihlavisto M, Szatmári I, Otwinowski Z, Smith DJ, Lázár L, et al. Novel pyridazinone inhibitors for vascular adhesion protein-1 (VAP-1): old target-new inhibition mode. *J Med Chem* (2013) 56(24):9837–48. doi: 10.1021/jm401372d
90. Karim S, Liaskou E, Fear J, Garg A, Reynolds G, Claridge L, et al. Dysregulated hepatic expression of glucose transporters in chronic disease: contribution of semicarbazide-sensitive amine oxidase to hepatic glucose uptake. *Am J Physiol Gastrointest Liver Physiol* (2014) 307(12):G1180–90. doi: 10.1152/ajpgi.00377.2013
91. Tesarik J, Conde-López C, Galán-Lázaro M, Mendoza-Tesarik R. Luteal phase in assisted reproductive technology. *Front Reprod Health* (2020) 2:595183. doi: 10.3389/frph.2020.595183
92. Nikhil Kumar Tej J, Johnson P, Krishna K, Kaushik K, Gupta PSP, Nandi S, et al. Copper and Selenium stimulates CYP19A1 expression in caprine ovarian granulosa cells: possible involvement of AKT and WNT signalling pathways. *Mol Biol Rep* (2021) 48(4):3515–27. doi: 10.1007/s11033-021-06346-5
93. Kim MJ, Kim SC, Chung S, Kim S, Yoon JW, Park YJ. Exploring the role of copper and selenium in the maintenance of normal thyroid function among healthy Koreans. *J Trace Elem Med Biol* (2020) 61:126558. doi: 10.1016/j.jtemb.2020.126558
94. Kosova F, Cetin B, Akinci M, Aslan S, Seki A, Pirhan Y, et al. Serum copper levels in benign and Malignant thyroid diseases. *Bratisl Lek Listy* (2012) 113(12):718–20. doi: 10.4149/bll_2012_162
95. Krishnamurthy HK, Reddy S, Jayaraman V, Krishna K, Song Q, Rajasekaran KE, et al. Effect of micronutrients on thyroid parameters. *J Thyroid Res* (2021) 2021:1865483. doi: 10.1155/2021/1865483
96. Garza A, Vega R, Soto E. Cellular mechanisms of lead neurotoxicity. *Med Sci Monit* (2006) 12(3):Ra57–65.
97. Nadler ST, Stoehr JP, Rabaglia ME, Schueler KL, Birnbaum MJ, Attie AD. Normal Akt/PKB with reduced PI3K activation in insulin-resistant mice. *Am J Physiol Endocrinol Metab* (2001) 281(6):E1249–54. doi: 10.1152/ajpendo.2001.281.6.E1249
98. Wang N, Sheng Z, Zhou S, Jiang F, Zhang Z. Chronic lead exposure exacerbates hepatic glucolipid metabolism disorder and gut microbiota dysbiosis in high-fat-diet mice. *Food Chem Toxicol* (2022) 170:113451. doi: 10.1016/j.fct.2022.113451
99. Wan H, Wang B, Cui Y, Wang Y, Zhang K, Chen C, et al. Low-level lead exposure promotes hepatic gluconeogenesis and contributes to the elevation of fasting

glucose level. *Chemosphere* (2021) 276:130111. doi: 10.1016/j.chemosphere.2021.130111

100. Almasmoum H, Refaat B, Ghaith MM, Almainani RA, Idris S, Ahmad J, et al. Protective effect of Vitamin D3 against lead induced hepatotoxicity, oxidative stress, immunosuppressive and calcium homeostasis disorders in rat. *Environ Toxicol Pharmacol* (2019) 72:103246. doi: 10.1016/j.etap.2019.103246
101. Uchida M, Teranishi H, Aoshima K, Katoh T, Kasuya M, Inadera H. Elevated urinary levels of vitamin D-binding protein in the inhabitants of a cadmium polluted area, Jinzu River basin, Japan. *Tohoku J Exp Med* (2007) 211(3):269–74. doi: 10.1620/tjem.211.269
102. Rahman A, Al-Awadi AA, Khan KM. Lead affects vitamin D metabolism in rats. *Nutrients* (2018) 10(3):264. doi: 10.3390/nu10030264
103. Nampoothiri LP, Gupta S. Biochemical effects of gestational coexposure to lead and cadmium on reproductive performance, placenta, and ovary. *J Biochem Mol Toxicol* (2008) 22(5):337–44. doi: 10.1002/jbt.20246
104. Wen L, Jiang X, Sun J, Li X, Li X, Tian L, et al. Cyanidin-3-O-glucoside promotes the biosynthesis of progesterone through the protection of mitochondrial function in Pb-exposed rat Leydig cells. *Food Chem Toxicol* (2018) 112:427–34. doi: 10.1016/j.fct.2017.10.008
105. Ji X, Li Z, Chen H, Li J, Tian H, Li Z, et al. Cytotoxic mechanism related to dihydrolipoamide dehydrogenase in Leydig cells exposed to heavy metals. *Toxicology* (2015) 334:22–32. doi: 10.1016/j.tox.2015.05.003
106. Kawai M, Swan KF, Green AE, Edwards DE, Anderson MB, Henson MC. Placental endocrine disruption induced by cadmium: effects on P450 cholesterol side-chain cleavage and 3beta-hydroxysteroid dehydrogenase enzymes in cultured human trophoblasts. *Biol Reprod* (2002) 67(1):178–83. doi: 10.1095/biolreprod67.1.178
107. Kim K, Pollack AZ, Nobles CJ, Sjaarda LA, Zolton JR, Radoc JG, et al. Associations between blood cadmium and endocrine features related to PCOS-phenotypes in healthy women of reproductive age: a prospective cohort study. *Environ Health* (2021) 20(1):64. doi: 10.1186/s12940-021-00749-4
108. Shen W, Chen J, Yin J, Wang SL. Selenium protects reproductive system and foetus development in a rat model of gestational lead exposure. *Eur Rev Med Pharmacol Sci* (2016) 20(4):773–80.
109. Rezaei M, Javadmoosavi SY, Mansouri B, Azadi NA, Mehrpour O, Nakhaee S. Thyroid dysfunction: how concentration of toxic and essential elements contribute to risk of hypothyroidism, hyperthyroidism, and thyroid cancer. *Environ Sci Pollut Res Int* (2019) 26(35):35787–96. doi: 10.1007/s11356-019-06632-7
110. Rani A, Kumar A, Lal A, Pant M. Cellular mechanisms of cadmium-induced toxicity: a review. *Int J Environ Health Res* (2014) 24(4):378–99. doi: 10.1080/09603123.2013.835032
111. Planchart A, Green A, Hoyo C, Mattingly CJ. Heavy metal exposure and metabolic syndrome: evidence from human and model system studies. *Curr Environ Health Rep* (2018) 5(1):110–24. doi: 10.1007/s40572-018-0182-3
112. Sarmiento-Ortega VE, Moroni-González D, Díaz A, Eduardo B, Samuel T. Oral subacute exposure to cadmium LOAEL dose induces insulin resistance and impairment of the hormonal and metabolic liver-adipose axis in wistar rats. *Biol Trace Elem Res* (2022) 200(10):4370–84. doi: 10.1007/s12011-021-03027-z
113. Fisher M, Potter B, Little J, Oulhote Y, Weiler HA, Fraser W, et al. Blood metals and vitamin D status in a pregnancy cohort: A bidirectional biomarker analysis. *Environ Res* (2022) 211:113034. doi: 10.1016/j.envres.2022.113034
114. Chen C, Zhang HJ, Zhai HL, Chen Y, Han B, Li Q, et al. Association between blood cadmium and vitamin D levels in the Yangtze Plain of China in the context of rapid urbanization. *Chin Med J (Engl)* (2020) 134(1):53–9. doi: 10.1097/cm9.0000000000001068
115. Chalkley SR, Richmond J, Barltrop D. Measurement of vitamin D3 metabolites in smelter workers exposed to lead and cadmium. *Occup Environ Med* (1998) 55(7):446–52. doi: 10.1136/oem.55.7.446
116. Zhang W, Jia H. Effect and mechanism of cadmium on the progesterone synthesis of ovaries. *Toxicology* (2007) 239(3):204–12. doi: 10.1016/j.tox.2007.07.007
117. Xiong YW, Xu XF, Zhu HL, Cao XL, Yi SJ, Shi XT, et al. Environmental exposure to cadmium impairs fetal growth and placental angiogenesis via GCN-2-mediated mitochondrial stress. *J Hazard Mater* (2021) 401:123438. doi: 10.1016/j.jhazmat.2020.123438
118. Ronchetti SA, Miler EA, Duvilanski BH, Cabilla JP. Cadmium mimics estrogen-driven cell proliferation and prolactin secretion from anterior pituitary cells. *PLoS One* (2013) 8(11):e81101. doi: 10.1371/journal.pone.0081101
119. Herath D, Pitawala A, Gunatilake J, Iqbal MCM. Using multiple methods to assess heavy metal pollution in an urban city. *Environ Monit Assess* (2018) 190(11):657. doi: 10.1007/s10661-018-7016-5
120. Theppeang K, Glass TA, Bandeen-Roche K, Todd AC, Rohde CA, Schwartz BS. Gender and race/ethnicity differences in lead dose biomarkers. *Am J Public Health* (2008) 98(7):1248–55. doi: 10.2105/ajph.2007.118505
121. Jukic AMZ, Kim SS, Meeker JD, Weiss ST, Cantonwine DE, McElrath TF, et al. A prospective study of maternal 25-hydroxyvitamin D (25OHD) in the first trimester of pregnancy and second trimester heavy metal levels. *Environ Res* (2021) 199:111351. doi: 10.1016/j.envres.2021.111351
122. Liu K, Mao X, Shi J, Lu Y, Liu C. Evaluation of lead and essential elements in whole blood during pregnancy: a cross-sectional study. *Ir J Med Sci* (2016) 185(3):677–82. doi: 10.1007/s11845-015-1339-9



OPEN ACCESS

EDITED BY

Etienne Marbaix,
Université Catholique de Louvain, Belgium

REVIEWED BY

Ariel Benor,
National Institutes of Health (NIH),
United States
Jun Zhai,
First Affiliated Hospital of Zhengzhou
University, China

*CORRESPONDENCE

Hongbin Chi
✉ chihb@163.com
Jie Qiao
✉ jie.qiao@263.net

RECEIVED 06 February 2023

ACCEPTED 11 August 2023

PUBLISHED 29 August 2023

CITATION

Huang N, Chen L, Lian Y, Chi H and Qiao J
(2023) Impact of thyroid-stimulating
hormone levels after controlled ovarian
hyperstimulation on *in vitro* fertilization/
intracytoplasmic sperm injection outcomes
in women with fresh embryo transfer: a
prospective cohort study.
Front. Endocrinol. 14:1159991.
doi: 10.3389/fendo.2023.1159991

COPYRIGHT

© 2023 Huang, Chen, Lian, Chi and Qiao.
This is an open-access article distributed
under the terms of the [Creative Commons
Attribution License \(CC BY\)](#). The use,
distribution or reproduction in other
forums is permitted, provided the original
author(s) and the copyright owner(s) are
credited and that the original publication in
this journal is cited, in accordance with
accepted academic practice. No use,
distribution or reproduction is permitted
which does not comply with these terms.

Impact of thyroid-stimulating hormone levels after controlled ovarian hyperstimulation on *in vitro* fertilization/intracytoplasmic sperm injection outcomes in women with fresh embryo transfer: a prospective cohort study

Ning Huang^{1,2,3,4}, Lixue Chen^{1,2,3,4}, Ying Lian^{1,2,3,4},
Hongbin Chi^{1,2,3,4*} and Jie Qiao^{1,2,3,4,5,6*}

¹Center for Reproductive Medicine, Department of Obstetrics and Gynecology, Peking University Third Hospital, Beijing, China, ²National Clinical Research Center for Obstetrics and Gynecology, Peking University Third Hospital, Beijing, China, ³Key Laboratory of Assisted Reproduction, Peking University, Ministry of Education, Beijing, China, ⁴Beijing Key Laboratory of Reproductive Endocrinology and Assisted Reproductive Technology, Beijing, China, ⁵Beijing Advanced Innovation Center for Genomics, Peking University, Beijing, China, ⁶Peking-Tsinghua Center for Life Sciences, Peking University, Beijing, China

Objective: Maternal hypothyroidism before and during pregnancy is associated with an increased risk of adverse pregnancy outcomes; many studies have evidenced that controlled ovarian hyperstimulation (COH) triggers a significant increase in the levels of TSH; however, no large-scale prospective studies have evaluated the impact of TSH levels after COH on assisted reproductive technology outcomes. The aim of this prospective study was to investigate whether *in vitro* fertilization/intracytoplasmic sperm injection (IVF/ICSI) outcomes are affected by TSH levels after COH in women with fresh embryo transfer (ET).

Methods: A total of 664 patients who underwent IVF/ICSI treatment and received fresh ET at the Peking University Third Hospital were included in this study. The rates of clinical pregnancy, miscarriage, live birth, and preterm delivery were analyzed.

Results: The patients were categorized into two groups based on serum TSH levels after COH ($0.55 \text{ mIU/L} < \text{TSH} < 2.5 \text{ mIU/L}$: $n = 449$, $2.5 \text{ mIU/L} \leq \text{TSH} \leq 4.78 \text{ mIU/L}$: $n = 215$). There were no significant differences in the rates of clinical pregnancy, miscarriage, and live birth between the two groups, even after adjusting for age, body mass index (BMI), thyroid antibody positivity, and COH protocols. However, the preterm delivery rate was significantly higher in women with $\text{TSH} < 2.5 \text{ mIU/L}$ than in those with $\text{TSH} \geq 2.5 \text{ mIU/L}$, even after adjusting for

relevant confounding factors. There was no significant difference in live birth weight between the two groups.

Discussion: Mildly elevated TSH levels ($\text{TSH} \geq 2.5 \text{ mIU/L}$) after COH did not affect IVF/ICSI outcomes, and strict control of TSH levels within 2.5 mIU/L after COH might not be necessary. Additionally, strictly controlled TSH levels ($\text{TSH} < 2.5 \text{ mIU/L}$) may increase preterm delivery risk.

KEYWORDS

in vitro fertilization, intracytoplasmic sperm injection, controlled ovarian hyperstimulation, clinical pregnancy, miscarriage, live birth, preterm delivery

1 Introduction

Adequate thyroid hormone levels are necessary for normal pregnancy and fetal development. Since the fetal thyroid gland is non-functional during early pregnancy, fetal growth and development are completely dependent on maternal thyroid hormone transfer. Thus, maternal hypothyroidism before and during pregnancy is associated with an increased risk of adverse pregnancy outcomes, including pregnancy loss, premature birth, and low birth weight (1, 2). Furthermore, intelligence quotient scores were reportedly lower in children of women with hypothyroidism than in children of women with normal thyrotropin concentrations, suggesting that hypothyroidism was detrimental to fetal neurocognitive development (3).

Given the potential danger of hypothyroidism, many studies suggest routine screening for thyroid function before and during pregnancy. In particular, serum thyroid-stimulating hormone (TSH) measurement is the most accurate assay for hypothyroidism evaluation because elevated levels of serum TSH are the earliest abnormal laboratory indicator of the occurrence of hypothyroidism (4). Although hypothyroidism is defined as an increase in serum TSH levels above the upper limit of normality, several guidelines suggest that the upper limit of TSH levels before pregnancy should not exceed 2.5 mIU/L ; the Endocrine Society recommends that TSH should not exceed 2.5 mIU/L before pregnancy and during the first trimester (5). Furthermore, the 2011 American Thyroid Association guidelines recommend that the upper reference limit for serum TSH concentration in the first trimester of pregnancy should be defined as 2.5 mIU/L (6). The 2021 European Thyroid Association Guideline suggests levothyroxine (LT4) treatment in subfertile women with TSH levels $>4.0 \text{ mIU/L}$ to maintain serum TSH levels $<2.5 \text{ mIU/L}$ (7). Recently, large-scale cohort studies also indicated that mildly elevated TSH levels ($\text{TSH} \geq 2.5 \text{ mIU/L}$) before pregnancy may increase the risk of adverse pregnancy outcomes (8, 9). However, whether to monitor and control TSH levels during assisted reproductive treatment remains unclear.

An increasing number of studies reported a higher incidence rate of hypothyroidism in women with infertility, a population that

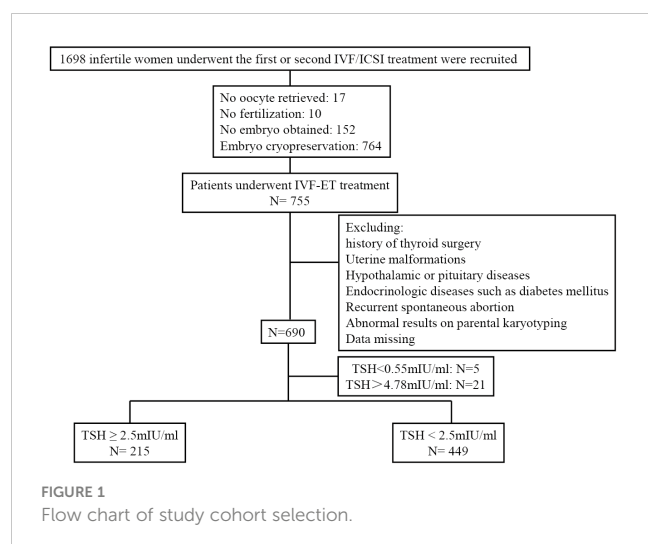
needs to achieve pregnancy using assisted reproductive technology (ART) (10). Controlled ovarian hyperstimulation (COH) is a necessary procedure in ART, and whether thyroid function should be monitored during and after COH is controversial. The most important question is whether the changes in thyroid hormone levels during COH affect ART and pregnancy outcomes. In a meta-analysis of 15 cohort studies, Li et al. found significantly increased serum TSH levels and significantly decreased free thyroxine (FT4) levels during COH (11). Another meta-analysis of 14 cohort studies, which failed to detect differences in serum FT4 levels, also showed a significant increase in serum TSH levels in patients after COH (12). Although alteration in serum FT4 levels during COH remains debatable, a significant increase in serum TSH levels after COH has been observed in many studies.

Most published studies focused on the association between TSH levels before COH on ART outcomes. However, no large-scale prospective studies have evaluated the impact of TSH levels after COH on ART outcomes, which may be very important in fresh embryo transfer (ET) cycles. To address this clinical question, we performed this prospective study to assess whether TSH levels after COH affect ART outcomes in women undergoing fresh ET.

2 Methods

2.1 Study population

We recruited 1698 infertile women who underwent the first or second *in vitro* fertilization/intracytoplasmic sperm injection (IVF/ICSI) treatment and received fresh ET at Peking University Third Hospital between June and September 2021. Patients were eligible for this study if they were 20–40 years of age and scheduled for their first or second IVF/ICSI-ET cycle. Patients were excluded if they had a history of thyroid surgery, uterine malformations, hypothalamic or pituitary diseases, diabetes mellitus or other endocrinologic or metabolic diseases, recurrent spontaneous abortion (defined as three or more previous spontaneous pregnancy losses), and abnormal results on parental karyotyping. A total of 690 women were ultimately included (Figure 1). All patients were treated using



standard COH protocols, including the ultralong gonadotropin-releasing hormone (GnRH) agonist protocol, long GnRH agonist protocol, or the GnRH antagonist protocol according to the patients' conditions. Oocytes were retrieved 34–36 hours after administration of recombinant human chorionic gonadotropin (HCG). Insemination was performed 4–6 hours after oocyte retrieval by conventional IVF or ICSI, according to sperm quality. Up to two Day 3 embryos were transferred three days after oocyte retrieval.

2.2 Laboratory tests

Serum samples for thyroid hormone testing were collected after COH. Thyroid function measurements, including TSH, FT4, thyroid peroxidase antibody (TPOAb), and thyroglobulin antibody (TGAb) levels, were performed using a fully automatic chemiluminescence immunoassay analyzer (ADVIA Centaur XP, Siemens Healthcare Diagnostics). The reference range were 0.55–4.78 mIU/L for TSH and 0.89–1.80 ng/dL for FT4. The level below 60 IU/mL indicated negative for TPOAb and TGAb. Five patients with TSH <0.55 mIU/L and 21 patients with TSH >4.78 mIU/L were excluded. The patients were divided into a low-normal TSH group (TSH <2.5 mIU/L) and a high-normal TSH group (TSH ≥2.5 mIU/L).

2.3 Study outcomes

Clinical pregnancy was defined as the presence of at least one gestational sac in the uterus, identified using ultrasonography 35 days after ET. Miscarriage was defined as the loss of clinical pregnancy before 28 weeks of gestation. Live birth was defined as the delivery of at least one live fetus. Preterm delivery was defined as delivery before 37 weeks of gestation.

2.4 Statistical analyses

Continuous variables were expressed as means (standard deviations, SDs) for normally distributed data and as medians

(interquartile ranges, IQRs) for data without a Gaussian distribution. Categorical variables were presented as numbers (percentages). Student's t-test was used to compare the differences in continuous variables with normal distribution between the two groups, and the Mann-Whitney U test was performed for continuous variables without a Gaussian distribution. Comparisons between categorical variables were performed using the Chi-squared test. Logistic regression analysis was conducted to calculate the odds ratios (ORs) with 95% confidence intervals (CIs) after adjusting for relevant factors. A two-sided $P < 0.05$ indicated statistical significance. All statistical analyses were performed using IBM SPSS Statistics for Windows version 24.0 software (Armonk, NY: IBM Corp).

3 Results

A total of 664 women who underwent IVF/ICSI treatment and fresh ET were included in this study. According to the TSH levels on HCG trigger day, 449 women with TSH <2.5 mIU/L were defined as the low-normal TSH group, and 215 women with TSH ≥2.5 mIU/L were defined as the high-normal TSH group. The baseline features of the two groups were compared, and the results were summarized in [Table 1](#). Women with TSH ≥2.5 mIU/L had significantly higher body mass index (BMI, $P = 0.034$) than women with TSH <2.5 mIU/L. No statistically significant differences were noted between the two groups in age, type of infertility, primary cause of infertility, rate of thyroid antibody positivity, or markers relevant to ovarian reserve, such as basal follicle-stimulating hormone, luteinizing hormone, estradiol, and anti-Mullerian hormone levels. We also collected data on thyroid function within six months before COH. While there was no significant difference in the basal FT4 level between two groups, a significantly higher basal TSH level was observed in the high-normal TSH group than in the low-normal TSH group ($P < 0.001$).

The group characteristics of COH and IVF are presented in [Table 2](#). We observed a significant difference in the terms of COH protocols between the two groups. The number of patients using GnRH antagonist protocols was significantly higher in the low-normal TSH group than in the high-normal TSH group. In contrast, the proportion of patients using ultralong GnRH agonists in the high-normal TSH group was higher than that in the low-normal TSH group. Although the gonadotropin dose and the days of ovarian stimulation were significantly higher in women with TSH ≥2.5 mIU/L than in those with TSH <2.5 mIU/L ($P = 0.007$ and 0.041 , respectively), no significant differences were observed between the two groups in the hormone levels on the HCG trigger day, the number of retrieved oocytes, and the number of good-quality embryos.

There were no significant differences between the low-normal TSH and high-normal TSH groups in the rates of clinical pregnancy (50.1% vs. 44.2%, $P = 0.153$), miscarriage (16.4% vs. 14.7%, $P = 0.703$), and live birth (41.9% vs. 37.7%, $P = 0.303$) ([Table 3](#)). Multiple logistic regression was performed to adjust for relevant confounding factors, and the results showed no significant differences between the two groups in the rates of clinical pregnancy (OR: 0.80, 95% CI:

TABLE 1 Baseline characteristics of patients.

| Characteristics | Low-normal TSH (n = 449) | High-normal TSH (n = 215) | P value |
|--|-----------------------------|------------------------------|----------|
| Age, mean (SD), years | 31.9 (3.7) | 32.3 (3.8) | 0.194 |
| BMI, mean (SD), kg/m ² | 22.8 (3.5) | 23.4 (3.6) | 0.034* |
| Type of infertility, no. (%) | | | |
| Primary | 281 (62.6) | 130 (60.5) | 0.599 |
| Secondary | 168 (37.4) | 85 (39.5) | |
| Primary cause of infertility, no. (%) ^a | | | |
| Male factors | 166 (37.0) | 83 (38.6) | 0.448 |
| Female factors | | | |
| Tubal factor | 142 (31.6) | 71 (33.0) | |
| Polycystic ovary syndrome | 41 (9.1) | 25 (11.6) | |
| Endometriosis | 37 (8.2) | 11 (5.1) | |
| Unknown factors | 63 (14.0) | 25 (11.6) | |
| Basal FSH, mean (IQR), mIU/mL ^b | 6.9 (5.7-8.2) | 6.8 (5.6-8.5) | 0.719 |
| Basal LH, mean (IQR), mIU/mL | 3.7 (2.5-5.0) | 3.5 (2.3-5.0) | 0.546 |
| Basal estradiol, mean (IQR), pmol/L | 146.5 (109.0-179.0) | 141.0 (102.0-169.0) | 0.136 |
| Basal FT4, mean (SD), ng/dL ^c | 1.2 (0.2) | 1.2 (0.2) | 0.628 |
| Basal TSH, mean (SD), mIU/L | 1.8 (0.8) | 2.8 (0.9) | < 0.001* |
| AMH, mean (IQR), ng/mL | 2.1 (1.2-3.4) | 1.8 (1.1-3.3) | 0.284 |
| No. of thyroid antibody positivity | 64 (14.3) | 28 (13.0) | 0.668 |

AMH, anti-Müllerian hormone; BMI, body mass index; COH, controlled ovarian hyperstimulation; FSH, follicle-stimulating hormone; FT4, free thyroxine; IQR, interquartile range; LH, luteinizing hormone; no., number; SD, standard deviation; TSH, thyroid-stimulating hormone.

^aPrimary cause of infertility indicates the most important cause for patients seeking *in vitro* fertilization/intracytoplasmic sperm injection treatment.

^bTesting for basal FSH, LH, and estradiol levels was performed between day 2 and day 4 of the menstrual cycle.

^cData of basal FT4 and TSH were collected within 6 months before COH.

*P<0.05.

0.58–1.12, P=0.193), miscarriage (OR: 0.84, 95% CI: 0.42–1.68; P=0.619), and live birth (OR: 0.87, 95% CI: 0.62–1.23, P=0.434) after adjusting for age, BMI, thyroid antibody positivity, and COH protocols (Table 4). Surprisingly, a significantly higher rate of preterm delivery was observed in women with TSH <2.5 mIU/L compared with those with TSH ≥2.5 mIU/L (22.3% vs. 11.1%, P=0.031), and this difference existed after adjusting for age, BMI, thyroid antibody positivity and COH protocols (OR: 0.39, 95% CI: 0.18–0.86, P=0.020) (Tables 3, 4). In singleton or twin pregnancies, birth weight was not significantly different between the two groups.

4 Discussion

Currently, whether screening for thyroid function after COH is needed to initiate appropriate management and intervention remains controversial. To our knowledge, this is the first large-scale prospective study to assess the impact of maternal TSH levels after COH on IVF/ICSI outcomes in women undergoing fresh ET. In this cohort of women undergoing IVF/ICSI-ET treatment, women with TSH ≥2.5 mIU/L after COH had similar pregnancy

outcomes to those of women with TSH <2.5 mIU/L. Unexpectedly, women with TSH <2.5 mIU/L had a higher preterm delivery risk, even after adjusting for relevant confounders, such as age, BMI, thyroid antibody positivity, and COH protocols. Our study suggests that TSH ≥2.5 mIU/L after COH is not a risk factor for poorer reproductive outcomes, and there is no need to strictly control TSH levels below 2.5 mIU/L after COH.

The impact of mildly elevated TSH levels on pregnancy outcomes has been debated for several years; most studies have focused on the impact of preconception TSH values on pregnancy outcomes. Two large-scale population-based cohort studies classified women into different groups according to TSH levels within 6 months before pregnancy and showed that even slightly elevated preconception TSH levels were associated with various adverse maternal outcomes, and recommended an optimal preconception TSH range between the lower reference limit and 2.50 mIU/L (8, 9). However, no large-scale study has investigated the effect of TSH levels after COH treatment on pregnancy outcomes.

COH is an important part of ART, which causes estradiol to rapidly rise to supraphysiological levels. Many studies have shown

TABLE 2 Protocols of COH and data of IVF and ET.

| Characteristics | Low-normal TSH (n = 449) | High-normal TSH (n = 215) | P value |
|---|-----------------------------|------------------------------|---------|
| Protocols of COH, no. (%) | | | |
| Ultralong GnRH agonist | 25 (5.6) | 24 (11.2) | 0.035* |
| Long GnRH agonist | 78 (17.4) | 34 (15.8) | |
| GnRH antagonist | 346 (77.1) | 157 (73.0) | |
| Gonadotropin dose, median (IQR), IU | 2625.0 (1875.0-3300.0) | 2850.0 (2025.0-3900.0) | 0.007* |
| No. of days of ovarian stimulation, median (IQR) | 10.0 (9.0-12.0) | 11.0 (9.0-13.0) | 0.041* |
| LH level on HCG trigger day, median (IQR), mIU/mL | 1.1 (0.5-2.3) | 1.2 (0.5-2.4) | 0.443 |
| Estradiol level on HCG trigger day, median (IQR), pmol/L | 6099.0 (4649.0-8449.0) | 6637.0 (4883.0-8427.0) | 0.186 |
| Progesterone level on HCG trigger day, median (IQR), nmol/L | 1.5 (1.3-1.9) | 1.6 (1.3-1.9) | 0.721 |
| No. of retrieved oocytes per cycle, median (IQR) | 9.0 (6.0-12.0) | 9.0 (6.0-11.0) | 0.632 |
| No. of good-quality embryos, median (IQR) ^a | 4.0 (2.0-5.0) | 3.0 (2.0-5.0) | 0.383 |
| No. of embryo transferred, no. (%) | | | |
| 1 | 57 (12.8) | 21 (9.9) | 0.278 |
| 2 | 389 (87.2) | 192 (90.1) | |

COH, controlled ovarian hyperstimulation; IVF, *in vitro* fertilization; ET, embryo transfer; GnRH, gonadotropin-releasing hormone; HCG, human chorionic gonadotropin; IQR, interquartile range; LH, luteinizing hormone; no., number; SD, standard deviation; TSH, thyroid-stimulating hormone.

^aThe embryos were evaluated on the third day after fertilization. Good-quality embryos were developed from two pronuclei zygotes and met the following criteria: (1) had more than five blastomeres; (2) size difference <20%; and (3) fragmentation <50%.

*P<0.05.

an association between COH and changes in thyroid function (11, 12). The underlying mechanism involves an increase in thyroid-binding globulin levels triggered by supraphysiologic estradiol levels, which reduces free thyroid hormone concentrations and, in turn, triggers serum TSH elevation. Unlike previous studies, our study analyzed TSH levels after COH instead of before, which may better reflect thyroid function before fresh ET. We did not find statistically significant differences in the rates of clinical pregnancy, miscarriage, or live birth between the low-normal and high-normal

TSH groups. Our study agrees with several previous studies that showed an association between TSH levels before COH and IVF/ICSI outcomes in infertile women. Based on a secondary data analysis of 1468 infertile women, Seungdamrong et al. showed that preconception TSH ≥ 2.5 mIU/L did not affect conception, clinical pregnancy, miscarriage, and live birth rates in infertile women without reproductive treatment (13). Two other studies, which restricted the patient populations to infertile women undergoing IVF/ICSI treatment, also found no significant

TABLE 3 Pregnancy outcomes of women with low-normal TSH levels and those with high-normal TSH levels.

| Outcomes | Low-normal TSH (n=449) | High-normal TSH (n=215) | P value |
|---|---------------------------|----------------------------|---------|
| Clinical pregnancy ^a , no. (%) | 225/449 (50.1) | 95/215 (44.2) | 0.153 |
| Miscarriage ^b , no. (%) | 37/225 (16.4) | 14/95 (14.7) | 0.703 |
| Live birth ^c , no. (%) | 188/449 (41.9) | 81/215 (37.7) | 0.303 |
| Preterm delivery ^d , no. (%) | 42/188 (22.3) | 9/81 (11.1) | 0.031* |
| Birth weight, g | | | |
| Singleton | 3300.0 (3000.0-3600.0) | 3350.0 (2990.0-3600.0) | 0.908 |
| Twin | 2550.0 (2300.0-2745.0) | 2525.0 (2150.0-2703.8) | 0.365 |

No., number; TSH, thyroid-stimulating hormone.

^aClinical pregnancy was defined as the presence of at least one gestational sac in the uterus identified using ultrasonography 35 days after embryo transfer.

^bMiscarriage was defined as the loss of a clinical pregnancy before 28 weeks of gestation. The miscarriage rate was defined as the proportion of women with miscarriage among women with clinical pregnancy.

^cLive birth was defined as the delivery of at least one living fetus.

^dPreterm delivery was defined as delivery before 37 weeks. The preterm delivery rate was defined as the proportion of women with preterm delivery among women with live birth.

*P < 0.05.

TABLE 4 Multivariate logistic regression analysis of factors associated with pregnancy outcomes.

| Factors | Clinical pregnancy | | Miscarriage | | Live birth | | Preterm delivery | |
|-------------------------------|--------------------|---------|------------------|---------|------------------|---------|------------------|---------|
| | OR (95% CI) | P value | OR (95% CI) | P value | OR (95% CI) | P value | OR (95% CI) | P value |
| Age, years | 0.95 (0.91-0.99) | 0.011* | 1.11 (1.01-1.21) | 0.023* | 0.93 (0.89-0.97) | 0.001* | 0.97 (0.88-1.06) | 0.435 |
| BMI, kg/m ² | 0.98 (0.94-1.03) | 0.455 | 1.07 (0.98-1.16) | 0.160 | 0.97 (0.93-1.02) | 0.200 | 1.07 (0.98-1.17) | 0.146 |
| TSH on HCG trigger day | | | | | | | | |
| Low-normal (reference) | NA | NA | NA | NA | NA | NA | NA | NA |
| High-normal | 0.80 (0.58-1.12) | 0.193 | 0.84 (0.42-1.68) | 0.619 | 0.87 (0.62-1.23) | 0.434 | 0.39 (0.18-0.86) | 0.020* |
| Thyroid antibody positivity | 1.00 (0.64-1.56) | 0.998 | 0.31 (0.09-1.05) | 0.060 | 1.25 (0.80-1.96) | 0.332 | 1.54 (0.68-3.46) | 0.300 |
| Protocols | | | | | | | | |
| Ultralong GnRH agonist | 1.30 (0.72-2.36) | 0.383 | 1.17 (0.40-3.42) | 0.774 | 1.17 (0.64-2.15) | 0.613 | 1.44 (0.43-4.77) | 0.554 |
| Long GnRH agonist | 1.27 (0.84-1.92) | 0.252 | 0.99 (0.44-2.21) | 0.974 | 1.25 (0.82-1.90) | 0.296 | 1.09 (0.49-2.41) | 0.834 |
| GnRH antagonist | NA | NA | NA | NA | NA | NA | NA | NA |

BMI, body mass index; CI, confidence interval; GnRH, gonadotropin-releasing hormone; HCG, human chorionic gonadotropin; NA, not available; OR, odds ratio; TSH, thyroid-stimulating hormone.

*P<0.05.

differences in the rates of clinical pregnancy, miscarriage, and live birth between the TSH <2.5 mIU/L and TSH ≥2.5 mIU/L groups (14, 15).

Most previous studies focused on the impact of the mildly elevated TSH levels on adverse ART outcomes and some guidelines recommended to maintain TSH levels below 2.5 mIU/L before pregnancy. In our study, no significant difference was found in the rates of clinical pregnancy, miscarriage and live birth between women with TSH ≥2.5 mIU/L and women with TSH <2.5 mIU/L, however, we observed a significantly higher rate of preterm delivery in women with TSH <2.5 mIU/L than in those with TSH ≥2.5 mIU/L, even after adjusting for age, BMI, thyroid antibody positivity, and COH protocols, which suggest that strictly control TSH levels below 2.5 mIU/L may trigger a detrimental effect on assisted reproductive outcomes. Our results were supported by a recent large-scale cohort study, which enrolled 175,112 women and performed a restricted cubic spline (RCS) regression model with multiple percentiles of TSH level to analyze the association between preconception TSH levels and adverse pregnancy outcomes. The study showed lower TSH was associated with a higher OR of preterm delivery (nonlinear P<0.001) (16). However, some previous studies based on TSH levels before COH showed inconsistent results. In a retrospective study, Zhang et al. reported no significant difference in the rates of preterm delivery between women with low TSH levels (TSH <2.5 mIU/L) and those with mildly elevated TSH levels (TSH ≥2.5 mIU/L) (14). Another retrospective study showed a significantly lower gestational age at delivery and lower birth weight in women with TSH ≥2.5 mIU/L than in those with TSH <2.5 mIU/L before COH (17). The controversial conclusions may result from the different timing of TSH test and screening for TSH levels after COH may be necessary in fresh ET cycles.

In our study, mildly elevated TSH levels (TSH ≥2.5 mIU/L) after COH did not affect IVF/ICSI outcomes. However, the rate of preterm delivery significantly increased in women with TSH <2.5

mIU/L. Based on our study, TSH levels after COH should not be strictly reduced to below 2.5 mIU/L. All assays for TSH and antithyroid antibodies in our study were performed in the same standardized laboratory, which reduced the potential inter-assay variability. However, our study also had some limitations. First, we only focused on TSH levels after COH and did not collect longitudinal TSH measurements throughout pregnancy. However, many previous studies have already shown the impact of TSH levels during different stages of pregnancy on reproductive outcomes (18). Second, we did not investigate the impact of the use of levothyroxine because of marked differences in the initiation time of thyroid hormone replacement. As such, further randomized controlled trials may be needed. Furthermore, we did not analyze additional pregnancy complications, such as gestational hypertension and gestational diabetes mellitus, because of the relatively small sample size. Further prospective studies with larger sample sizes are required to confirm these findings.

5 Conclusions

To our knowledge, our study is the first large-scale prospective study to demonstrate that mildly elevated TSH levels after COH may not adversely affect IVF/ICSI outcomes in women who receive fresh ET, that TSH ≥2.5 mIU/L after COH is not a risk factor for poorer reproductive outcomes, and that strict control of TSH levels to levels below 2.5 mIU/L after COH may not be necessary.

Data availability statement

The original contributions presented in the study are included in the article/supplementary material, further inquiries can be directed to the corresponding authors.

Ethics statement

The studies involving humans were approved by Peking University Third Hospital Medical Science Research Ethics Committee. The studies were conducted in accordance with the local legislation and institutional requirements. The participants provided their written informed consent to participate in this study. Written informed consent was obtained from the individual(s) for the publication of any potentially identifiable images or data included in this article.

Author contributions

NH took part in patient follow-up and wrote the initial draft of the paper. NH, LC and YL contributed to the data analysis. HC and JQ contributed to the conception and design of the study. All authors contributed to the research discussion and manuscript revision.

Funding

This work was supported by the Second Tibetan Plateau Scientific Expedition and Research (Grant No. 2019QZKK0607) and National Natural Science Foundation of China (Grant No.82171626).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

- van den Boogaard E, Vissenberg R, Land JA, van Wely M, van der Post JA, Goddijn M, et al. Significance of (sub)clinical thyroid dysfunction and thyroid autoimmunity before conception and in early pregnancy: a systematic review. *Hum Reprod Update* (2011) 17:605–19.
- Abalovich M, Gutierrez S, Alcaraz G, Maccallini G, Garcia A, Levalle O. Overt and subclinical hypothyroidism complicating pregnancy. *Thyroid* (2002) 12:63–8.
- Haddow JE, Palomaki GE, Allan WC, Williams JR, Knight GJ, Gagnon J, et al. Maternal thyroid deficiency during pregnancy and subsequent neuropsychological development of the child. *N Engl J Med* (1999) 341:549–55.
- McDermott MT. Hypothyroidism. *Ann Intern Med* (2020) 173:Itc1–itc16.
- De Groot L, Abalovich M, Alexander EK, Amino N, Barbour L, Cobin RH, et al. Management of thyroid dysfunction during pregnancy and postpartum: an Endocrine Society clinical practice guideline. *J Clin Endocrinol Metab* (2012) 97:2543–65.
- Stagnaro-Green A, Abalovich M, Alexander E, Azizi F, Mestman J, Negro R, et al. Guidelines of the American Thyroid Association for the diagnosis and management of thyroid disease during pregnancy and postpartum. *Thyroid* (2011) 21:1081–125.
- Poppe K, Bisschop P, Fugazzola L, Minziori G, Unuane D, Weghofer A. European Thyroid Association Guideline on Thyroid Disorders prior to and during Assisted Reproduction. *Eur Thyroid J* (2021) 9:281–95.
- Yang Y, Guo T, Fu J, Kuang J, Wang Y, Zhang Y, et al. Preconception thyrotropin levels and risk of adverse pregnancy outcomes in Chinese women aged 20 to 49 years. *JAMA Netw Open* (2021) 4:e215723.
- Yang Y, Guo T, Fu J, Zhao J, Wang Y, He Y, et al. Association of preconception thyrotropin levels with fecundability and risk of spontaneous abortion in China. *JAMA Netw Open* (2022) 5:e2228892.
- Dhillon-Smith RK, Tobias A, Smith PP, Middleton LJ, Sunner KK, Baker K, et al. The prevalence of thyroid dysfunction and autoimmunity in women with history of miscarriage or subfertility. *J Clin Endocrinol Metab* (2020) 105: 2667–77.
- Li D, Hu S, Meng X, Yu X. Changes in thyroid function during controlled ovarian hyperstimulation (COH) and its impact on assisted reproduction technology (ART) outcomes: a systematic review and meta-analysis. *J Assist Reprod Genet* (2021) 38:2227–35.
- Busnelli A, Cirillo F, Levi-Setti PE. Thyroid function modifications in women undergoing controlled ovarian hyperstimulation for *in vitro* fertilization: a systematic review and meta-analysis. *Fertil Steril* (2021) 116:218–31.
- Seungdamrong A, Steiner AZ, Gracia CR, Legro RS, Diamond MP, Coutifaris C, et al. Preconceptional antithyroid peroxidase antibodies, but not thyroid-stimulating hormone, are associated with decreased live birth rates in infertile women. *Fertil Steril* (2017) 108 843–50.
- Zhang Y, Wu W, Liu Y, Guan Y, Wang X, Jia L. High-normal preconception TSH levels have no adverse effects on reproductive outcomes in infertile women undergoing the first single fresh D5 blastocyst transfer. *Int J Endocrinol* (2020) 2020:1056484.
- Cai Y, Zhong L, Guan J, Guo R, Niu B, Ma Y, et al. Outcome of *in vitro* fertilization in women with subclinical hypothyroidism. *Reprod Biol Endocrinol* (2017) 15:39.
- Du H, Wu D, Zhou X, Yang H, Zhu H, Chen S, et al. and adverse pregnancy outcomes in China: A nationwide prospective cohort study. *J Clin Endocrinol Metab* (2022) 107:e2770–6.
- Baker VL, Rone HM, Pasta DJ, Nelson HP, Gvakharina M, Adamson GD. Correlation of thyroid stimulating hormone (TSH) level with pregnancy outcome in women undergoing *in vitro* fertilization. *Am J Obstet Gynecol* (2006) 194:1668–74; discussion 1674–5.
- Spencer L, Bubner T, Bain E, Middleton P. Screening and subsequent management for thyroid dysfunction pre-pregnancy and during pregnancy for improving maternal and infant health. *Cochrane Database Syst Rev* (2015) 2015: Cd011263.



OPEN ACCESS

EDITED BY

Hong Zhang,
Second Affiliated Hospital of Soochow
University, China

REVIEWED BY

Junhao Yan,
Shandong University, China
Xi Wang,
Harvard University, United States
Yong Wang,
Nanjing University, China

*CORRESPONDENCE

Haiyan Wang
✉ wangquan1991@sina.com
Xiaoyu Long
✉ long_x_y@163.com

†These authors share first authorship

RECEIVED 11 June 2023

ACCEPTED 21 August 2023

PUBLISHED 08 September 2023

CITATION

Gao J, Yuan Y, Li J, Tian T, Lian Y, Liu P,
Li R, Qiao J, Long X and Wang H (2023)
Sequential embryo transfer versus double
cleavage-stage embryo or double
blastocyst transfer in patients with
recurrent implantation failure with
frozen-thawed embryo transfer
cycles: a cohort study.
Front. Endocrinol. 14:1238251.
doi: 10.3389/fendo.2023.1238251

COPYRIGHT

© 2023 Gao, Yuan, Li, Tian, Lian, Liu, Li,
Qiao, Long and Wang. This is an open-
access article distributed under the terms of
the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/)
(CC BY). The use, distribution or
reproduction in other forums is permitted,
provided the original author(s) and the
copyright owner(s) are credited and that
the original publication in this journal is
cited, in accordance with accepted
academic practice. No use, distribution or
reproduction is permitted which does not
comply with these terms.

Sequential embryo transfer versus double cleavage-stage embryo or double blastocyst transfer in patients with recurrent implantation failure with frozen-thawed embryo transfer cycles: a cohort study

Jiangman Gao^{1,2,3,4†}, Yifeng Yuan^{1,2,3,4†}, Jia Li^{1,2,3,4}, Tian Tian^{1,2,3,4},
Ying Lian^{1,2,3,4}, Ping Liu^{1,2,3,4}, Rong Li^{1,2,3,4}, Jie Qiao^{1,2,3,4},
Xiaoyu Long^{1,2,3,4*} and Haiyan Wang^{1,2,3,4*}

¹State Key Laboratory of Female Fertility Promotion, Center for Reproductive Medicine, Department of Obstetrics and Gynecology, Peking University Third Hospital, Beijing, China, ²National Clinical Research Center for Obstetrics and Gynecology Department of Obstetrics and Gynecology, Peking University Third Hospital, Beijing, China, ³Key Laboratory of Assisted Reproduction (Peking University), Ministry of Education, Department of Obstetrics and Gynecology, Peking University Third Hospital, Beijing, China, ⁴Beijing Key Laboratory of Reproductive Endocrinology and Assisted Reproductive Technology, Department of Obstetrics and Gynecology, Peking University Third Hospital, Beijing, China

Background: Recurrent implantation failure (RIF) is more common among patients receiving assisted reproductive treatment. Many efforts have been made to increase the incidence of clinical pregnancy among patients with RIF. The effect of the sequential transfer procedure, a two-step interval transfer of a cleavage-stage embryo followed by a blastocyst in one transfer cycle, on the clinical outcomes of RIF patients remains controversial.

Methods: In total, 1774 frozen-thawed embryo transfer (FET) cycles in RIF patients were included. Of these cycles, 302 were sequential embryo transfer (ET) cycles, 979 were double day 3 cleavage-stage ET cycles, and 493 were double blastocyst ET cycles. The primary outcomes were the rates of implantation, clinical pregnancy and multiple pregnancy, and the secondary outcomes were the rates of hCG positive, early miscarriage and ectopic pregnancy.

Results: The implantation, hCG positive, and clinical pregnancy rates in the sequential ET group (32.1%, 58.9%, 50.7%) were significantly higher than those in the day 3 cleavage-stage ET group (24.9%, 46.5%, 40.4%) and were similar to those in the blastocyst transfer group (30.1%, 56.4%, 47.1%). The early miscarriage rate in the blastocyst transfer group was significantly higher than that in the cleavage-stage ET group (17.2% vs. 8.1%, $P < 0.05$), while the ectopic pregnancy rate in the blastocyst transfer group was significantly lower than that in the cleavage-stage ET group (0.4% vs. 3.0%, $P < 0.05$). The multiple pregnancy rate in the sequential ET group was significantly lower than that in the cleavage-stage ET group (17.0% vs. 25.5%, P

<0.05) and the blastocyst transfer group (17.0% vs. 27.6%, $P < 0.05$). When cycles of blastocyst culture failure were excluded, the clinical pregnancy rate was significantly higher (55.7% vs. 47.1%, $P < 0.05$), and the early miscarriage rate and multiple pregnancy rate were significantly lower (8.5% vs. 17.2%, 17.7% vs. 27.6%; $P < 0.05$, respectively) in the sequential ET group than in the double blastocyst ET group.

Conclusions: Sequential embryo transfer in FET cycles could improve the clinical outcomes of patients with RIF.

KEYWORDS

repeated implantation failure, frozen-thawed embryo transfer, sequential embryo transfer, cleavage-stage embryo transfer, blastocyst transfer

Introduction

Since 1978, many infertile couples have benefited from assisted reproductive technology (ART). As of 2019, more than 8 million children had been born after ART worldwide. Over 2.5 million *in vitro* fertilization (IVF) cycles are performed every year, resulting in over 500,000 deliveries annually (1). However, the best embryo implantation rate ranges from 25–40% (2, 3). Improving the success rate is a challenging problem associated with ART treatment programs.

Repeated implantation failure (RIF) refers to a situation when the transferred embryos fail to implant after at least three IVF-embryo transfer (IVF-ET) cycles with 1–2 high-quality embryos in each cycle (4, 5). The prevalence of RIF is 8–15% (6–8), which poses great difficulties and challenges to clinicians and embryologists. Recurrent failures of IVF-ET also bring psychological, physical, and financial distress to patients.

High-quality embryos, a receptive endometrium, and good synchrony between the embryo and endometrium are necessary conditions for successful implantation (9). The implantation process involves three phases: apposition, adhesion, and invasion. During these stages, the cross-talk between the endometrium and embryo is significant, and suboptimal endometrial receptivity is the most critical cause of RIF (10).

In recent years, scientists have proposed a sequential transfer procedure, a two-step interval transfer of a cleavage-stage embryo followed by a blastocyst in one transfer cycle to help RIF patients increase the chance of pregnancy. Several studies have suggested that sequential ET significantly improves the clinical outcomes of IVF-ET (11–13). However, the effect of sequential ET is still controversial, and its effectiveness and potential biological mechanisms have not been proven.

To further confirm the effect of sequential ET, this study analyzed the data of sequential ET at our reproductive center to evaluate the effect of sequential ET on the clinical outcomes of patients with a history of RIF in frozen-thawed embryo transfer

(FET) cycles. The primary outcome measures were implantation rate, clinical pregnancy rate and multiple pregnancy rate. The secondary outcome measures were hCG positive rate, early miscarriage rate and ectopic pregnancy rate.

Patients and methods

This retrospective observational study was performed at the Reproductive Center of Peking University Third Hospital from January 2020 to June 2022. Patients who had not conceived after three or more ET cycles and undergone sequential ET (one day 3 cleavage-stage embryo followed by a day 5/6 blastocyst in one FET cycle) were included (302 cycles). Two groups based on the ET strategy were adopted as the control groups: the double cleavage-stage embryo (day 3) transfer group (979 cycles) and the double blastocyst (day 5/6) transfer group (493 cycles). Among these participants, those employing PGT for chromosomal structural rearrangements (PGT-SR) or monogenic/single gene defects (PGT-M), those using egg donor cycles, those with a thin endometrium (thickness less than 6 mm) and those with autoimmune diseases were excluded.

Frozen-thawed embryo transfer procedure

All the included cycles were FET cycles. Endometrial preparation for FET was performed as previously described (14), and the preparation method was the artificial (hormone replacement) cycle, natural cycle, or stimulation cycle. On the transfer day, embryo grading was performed. Cleavage-stage embryos were evaluated according to the criteria of the Istanbul Embryo Evaluation Symposium (15), and blastocysts were evaluated using the Gardner grading system (16). The embryos were transferred using the Cook Sydney IVF catheter (k-jets-7019-SIVF). In the sequential ET group, one of the frozen-thawed embryos was transferred on day 3, whereas

the rest were cultured; then, one blastocyst was transferred on day 5 or day 6. In the cleavage-stage ET group, double embryos were transferred on day 3, while in the blastocyst ET group, double blastocysts were transferred on day 5.

Outcome measures

Serum β -hCG levels were measured 14–21 days after ET, with β -hCG levels ≥ 25 IU/L being defined as biochemical pregnancy, also named hCG positive. Clinical pregnancy is defined as the presence of an intrauterine gestational sac on ultrasonography. The implantation rate was defined as the number of gestational sacs divided by the total number of embryos or blastocysts transferred. Early miscarriage was defined as loss of the clinical pregnancy within 12 weeks of gestation. Ectopic pregnancy was defined as an extrauterine pregnancy. Multiple pregnancy was defined as the presence of two or more gestational sacs on ultrasound, and the rate was calculated as the number of multiple pregnancy cycles divided by the number of clinical pregnancy cycles.

Statistical analysis

Data analysis was performed using SPSS statistics software version 23 (IBM). The continuous variables are presented as the means \pm standard deviations (SDs). One-way ANOVA was used for continuous variables that had a normal distribution, while the Kruskal–Wallis test was performed for nonnormally distributed continuous data. Categorical variables are presented as counts and percentages. The chi-square test was applied to test categorical variables. In the multivariate logistic regression analysis, FET groups, parental age, infertility duration, cycles of implantation failure, insemination methods, endometrial preparation methods and endometrial thickness were included, and adjusted odds ratios (ORs) and 95% confidence intervals (95% CIs) were reported. A *P* value of < 0.05 was considered statistically significant for all tests.

Results

Characteristics of sequential embryo transfer cycles

A total of 302 sequential ET cycles were included in this study. Of these, there were 5 cycles with no embryo to culture after cleavage-stage ET, resulting in 2 cycles of cleavage-stage ET only and 3 cycles of cleavage-stage ET followed by frozen-thawed blastocyst transfer. There were 44 cycles in which the remaining frozen-thawed cleavage-stage embryos were not cultured into blastocysts after cleavage-stage ET (including 32 cycles of cleavage-stage ET only and 12 cycles of cleavage-stage ET followed by frozen-thawed blastocyst transfer). Overall, 253 cycles were completed successfully with day 3 cleavage-stage ET followed by cultured blastocyst transfer (Table 1). In total, day 3 cleavage-

TABLE 1 Cycle characteristics of sequential embryo transfer of FET.

| Sequential embryo transfer | Cycles (n) | Clinical pregnancy n (%) |
|--|------------|--------------------------|
| Total cycles | 302 | 153(50.7%) |
| No embryo to culture after a cleavage-stage embryo transfer | 5 | 1(20%) |
| A cleavage-stage embryo transfer only | 2 | 0 (0) |
| A cleavage-stage embryo transfer followed by a frozen-thawed blastocyst transfer | 3 | 1(33.3%) |
| Frozen-thawed cleavage-stage embryos did not form blastocysts after a cleavage-stage embryo transfer | 44 | 11(25%) |
| A cleavage-stage embryo transfer only | 32 | 8(25%) |
| A cleavage-stage embryo transfer followed by a frozen-thawed blastocyst transfer | 12 | 3(25%) |
| A cleavage-stage embryo transfer followed by a cultured blastocyst transfer | 253 | 141(55.7%) |

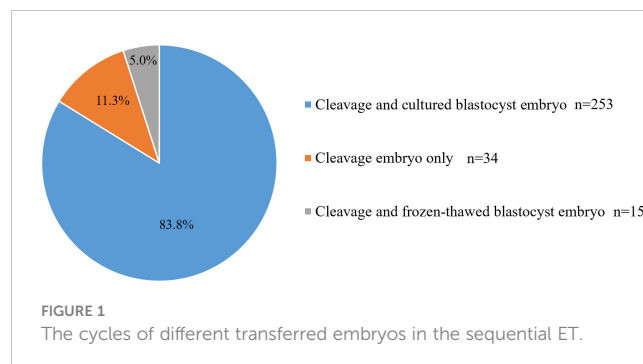
The bold values in Table 1 indicate the primary classification; non-bold values are the further subgroups.

stage embryo and cultured blastocyst transfer cycles, day 3 cleavage-stage embryo and frozen-thawed blastocyst transfer cycles, and day 3 cleavage-stage ET only cycles accounted for 83.8%, 5.0%, and 11.3% of total sequential ET cycles, respectively (Figure 1).

In summary, of the 302 sequential ET cycles performed, cleavage-stage embryos were transferred in only 34 (11.3%) cycles due to the lack of embryos for culture after ET (2 cycles) or failure of the remaining embryos to form blastocysts (32 cycles). There were fifteen (5.0%) cycles of cleavage-stage ET followed by frozen-thawed blastocyst transfer. In addition, 253 (83.8%) cycles were completed successfully with day 3 cleavage-stage ET followed by cultured blastocyst transfer.

Demographic characteristics of the three groups

We compared the baseline data, including parental age, female BMI, infertility duration, primary infertility ratio, previous failed cycles, insemination method, endometrial preparation protocols, and endometrial thickness on FET among the three groups. The infertility duration was significantly shorter (4.91 ± 3.29 years vs.



5.52 ± 3.27 years & 5.73 ± 3.46 years), and the number of cycles of previous failure were significantly lower (3.14 ± 1.69 vs. 4.83 ± 2.36 & 4.07 ± 2.00) in the cleavage-stage ET group than in the sequential ET and blastocyst transfer groups. The proportion of artificial cycles was significantly higher (65.9% vs. 53.7% & 56.2%) in the sequential ET group than in the cleavage-stage ET and blastocyst transfer groups. There were no significant differences in parental age, female BMI, primary infertility ratio, insemination method, and endometrial thickness on FET among the three groups (Table 2).

FET outcomes

Compared to those in the cleavage-stage ET group, the implantation, hCG positive, and clinical pregnancy rates were significantly higher in the sequential ET group and the blastocyst transfer group. There was no significant difference in the implantation rate, hCG positive rate, or clinical pregnancy rate between the sequential ET group and the blastocyst transfer group. The early miscarriage rate in the blastocyst transfer group was significantly higher than that in the cleavage-stage ET group, while the ectopic pregnancy rate in the blastocyst transfer group was significantly lower than that in the cleavage-stage ET group. The early miscarriage rate and ectopic pregnancy rate in the sequential ET group were not significantly different from the rates in the other two groups. The multiple pregnancy rate in the sequential ET group was significantly lower than that in the cleavage-stage ET group and the blastocyst transfer group (Figure 2).

We then compared the clinical outcomes of cycles in which sequential ET was completed successfully (a day 3 cleavage-stage ET followed by a cultured blastocyst transfer) with those of cycles in which blastocyst transfer had been completed successfully. The clinical pregnancy rate in the successfully completed sequential ET group was significantly higher than that in the blastocyst transfer group, while the early miscarriage rate and multiple pregnancy rate in the successfully completed sequential ET group were significantly lower than those in the blastocyst transfer group. There was no significant difference in the implantation rate, hCG positive rate, or ectopic pregnancy rate between the two groups (Figure 3).

Multiple logistic regression analysis with adjustments for possible confounders for clinical pregnancy, early miscarriage, and multiple pregnancy was used to evaluate the effectiveness of sequential ET (Table 3). Adjustments were made for confounding variables including FET group, parental age, infertility duration, cycles of implantation failure, insemination methods, endometrial preparation methods, and endometrial thickness. Compared to sequential ET and double blastocyst transfer, double cleavage-stage ET had a significantly lower clinical pregnancy rate (OR 0.610, 95% CI 0.432–0.861, $P=0.005$; OR 0.697, 95% CI 0.531–0.914, $P=0.009$, respectively). Endometrial thickness was associated with clinical pregnancy (OR 1.087, 95% CI 1.016–1.164, $P=0.016$). Double cleavage-stage ET had a lower early miscarriage rate than double blastocyst transfer (OR 0.439, 95% CI 0.240–0.800, $P=0.007$), female age had a significantly negative effect on early miscarriage (OR 1.162, 95% CI 1.040–1.299, $P=0.008$), and double blastocyst transfer had a significantly higher multiple pregnancy rate than sequential ET (OR 1.860, 95% CI 1.012–3.418, $P=0.046$).

TABLE 2 Demographic and cycle characteristics among the three groups according to embryo transfer of FET.

| Variable | Sequential embryo transfer (n = 302) | Cleavage-stage embryo transfer (n = 979) | Blastocyst embryo transfer (n = 493) | P value |
|-----------------------------------|--------------------------------------|--|--------------------------------------|-------------------|
| Female age (years) | 34.05 ± 4.51 | 33.63 ± 4.27 | 33.67 ± 4.01 | 0.302 |
| Male age (years) | 35.13 ± 6.82 | 34.94 ± 5.34 | 34.88 ± 5.13 | 0.291 |
| Body mass index | 22.39 ± 3.48 | 22.37 ± 3.45 | 22.36 ± 3.23 | 0.993 |
| Infertility duration (years) | 5.52 ± 3.27 | 4.91 ± 3.29 | 5.73 ± 3.46 | <0.001* |
| Primary infertility ratio (n, %) | 201 (66.6%) | 650 (66.4%) | 320 (64.9%) | 0.831 |
| Previous failed cycles (n) | 4.83 ± 2.36 | 3.14 ± 1.69 | 4.07 ± 2.00 | <0.001* |
| Insemination method (n, %) | | | | 0.246 |
| IVF | 202 (66.9%) | 603 (61.6%) | 312 (63.3%) | |
| ICSI | 100 (33.1%) | 376 (38.4%) | 181 (36.7%) | |
| Endometrial preparation (n, %) | | | | 0.006* |
| Artificial cycle | 199(65.9%) | 526 (53.7%) | 277(56.2%) | |
| Natural cycle | 88 (29.1%) | 378 (38.6%) | 185 (37.5%) | |
| Stimulation cycle | 15 (5.0%) | 75 (7.7%) | 31 (6.3%) | |
| Endometrial thickness on FET (mm) | 9.99 ± 1.73 | 9.98 ± 1.79 | 10.07 ± 1.67 | 0.757 |

IVF, in vitro fertilization; ICSI, intracytoplasmic sperm injection; FET, frozen-thawed embryo transfer; * $P<0.05$.

The bold values in Tables 2, 3 indicate significant statistical differences among groups ($P < 0.05$).

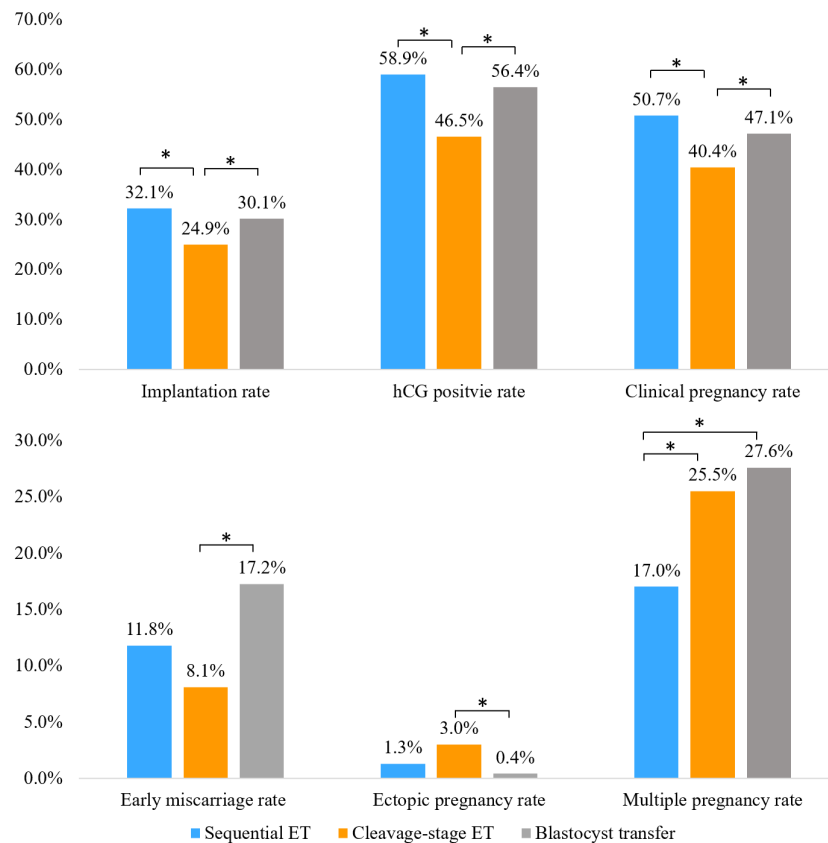


FIGURE 2

Clinical outcomes of patients in each group. *P<0.05.

Discussion

Sequential ET was first performed by Abramovici et al. in 1988 since embryo freezing was not an option in their IVF-ET program at that time (17). Since then, some clinicians have tried to use a sequential ET approach to help patients with RIF increase their chances of pregnancy. Some studies showed that sequential ET did not improve clinical outcomes for patients with RIF (18–20), while

other studies suggested that it was more effective toward increasing the clinical pregnancy and live birth rates than day 3 or day 5 ET in these patients (11–13). The study by Ji et al. included patients with a history of RIF undergoing FET cycles (18), and studies by Tehraninejad et al. (20) and Kyono et al. (19) included patients undergoing IVF fresh cycles and showed that sequential embryo transfer failed to increase the chance of pregnancy. A retrospective cohort study by Stamenov et al. (12) and a prospective and

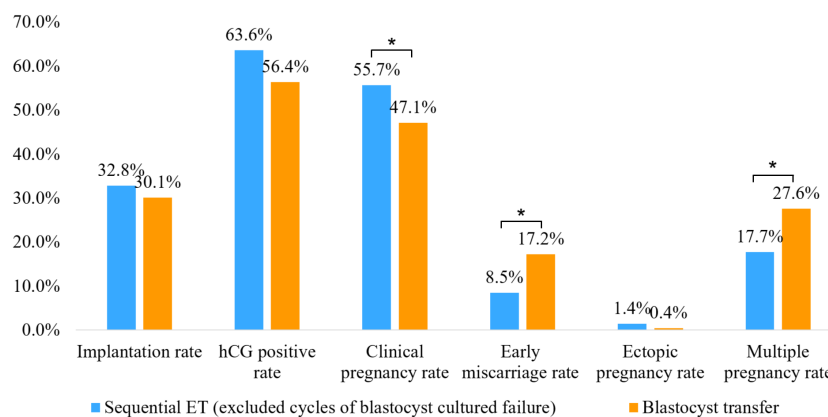


FIGURE 3

Comparison of clinical outcomes between successfully completed sequential embryo transfer group (day 3 cleavage-stage embryo transfer followed by a cultured blastocyst transfer) and double blastocyst transfer group. *P<0.05.

TABLE 3 Multivariate logistic regression analysis of clinical outcomes after adjustment.

| | Adjusted OR | 95% CI | P value |
|--|-------------|--------------|---------------|
| Clinical pregnancy | | | |
| FET group (cleavage vs. sequential) | 0.610 | 0.432-0.861 | 0.005* |
| FET group (blastocyst vs. sequential) | 0.875 | 0.612-1.251 | 0.465 |
| FET group (cleavage vs. blastocyst) | 0.697 | 0.531-0.914 | 0.009* |
| Female age | 0.965 | 0.924-1.007 | 0.104 |
| Male age | 0.983 | 0.951-1.016 | 0.317 |
| Infertility duration | 0.964 | 0.927-1.003 | 0.069 |
| Previous failed cycles | 1.005 | 0.942-1.072 | 0.880 |
| Insemination method (ICSI vs. IVF) | 0.979 | 0.766-1.252 | 0.866 |
| Endometrial preparation (natural vs. artificial) | 1.169 | 0.911-1.500 | 0.222 |
| Endometrial preparation (stimulation vs. artificial) | 0.864 | 0.528-1.415 | 0.562 |
| Endometrial preparation (natural vs. stimulation) | 1.352 | 0.815-2.243 | 0.242 |
| Endometrial thickness on FET | 1.087 | 1.016-1.164 | 0.016* |
| Early miscarriage | | | |
| Group of FET (cleavage vs. sequential) | 0.790 | 0.349-1.788 | 0.572 |
| Group of FET (blastocyst vs. sequential) | 1.802 | 0.824-3.942 | 0.140 |
| Group of FET (cleavage vs. blastocyst) | 0.439 | 0.240-0.800 | 0.007* |
| Female age | 1.162 | 1.040-1.299 | 0.008* |
| Male age | 0.929 | 0.850-1.017 | 0.110 |
| Infertility duration | 0.992 | 0.904-1.089 | 0.868 |
| Previous failed cycles | 0.862 | 0.731-1.018 | 0.079 |
| Insemination method (ICSI vs. IVF) | 1.197 | 0.694-2.063 | 0.518 |
| Endometrial preparation (natural vs. artificial) | 0.595 | 0.329-1.076 | 0.086 |
| Endometrial preparation (stimulation vs. artificial) | 1.596 | 0.599-4.322 | 0.358 |
| Endometrial preparation (natural vs. stimulation) | 0.373 | 0.129-1.073 | 0.067 |
| Endometrial thickness on FET | 0.978 | 0.842-1.137 | 0.776 |
| Multiple pregnancy | | | |
| Group of FET (cleavage vs. sequential) | 1.315 | 0.714-2.421 | 0.380 |
| Group of FET (blastocyst vs. sequential) | 1.860 | 1.012-3.418 | 0.046* |
| Group of FET (cleavage vs. blastocyst) | 0.707 | 0.448-1.115 | 0.135 |
| Female age | 0.950 | 0.872-1.034 | 0.234 |
| Male age | 0.994 | 0.929-1.065 | 0.871 |
| Infertility duration | 1.008 | 0.935-1.086 | 0.837 |
| Previous failed cycles | 1.025 | 0.916-1.147 | 0.665 |
| Insemination method (ICSI vs. IVF) | 0.786 | 0.513-1.205 | 0.270 |
| Endometrial preparation (natural vs. artificial) | 0.950 | 0.623-1.449 | 0.811 |
| Endometrial preparation (stimulation vs. artificial) | 0.319 | 0.093-1.089 | 0.068 |
| Endometrial preparation (natural vs. stimulation) | 2.980 | 0.860-10.326 | 0.085 |
| Endometrial thickness on FET | 0.951 | 0.844-1.075 | 0.434 |

IVF, in vitro fertilization; ICSI, intracytoplasmic sperm injection; FET, frozen-thawed embryo transfer; OR, odds ratio; CI, confidence interval; *P<0.05. The bold values in Tables 2, 3 indicate significant statistical differences among groups (P <0.05).

randomized trial by Torky et al. (13) showed that the implantation and clinical pregnancy rates were significantly higher in patients who underwent sequential embryo transfer than in those who underwent cleavage transfer on day 3 or blastocyst transfer on day 5 in IVF fresh cycles. Arefi et al. (11) conducted a prospective study to evaluate the improvement of pregnancy rate in sequential FET on day 3/day 5 in individuals who suffered from RIF and suggested that sequential transfer was more effective than regular day 5. A systematic review by Zhang et al. demonstrated that the clinical pregnancy rate and live birth rate were higher in the sequential ET group than in the cleavage-stage ET group for women who experienced RIF, and there were no significant differences between sequential ET and blastocyst ET (21). Even in patients with poor ovarian response, a report showed that sequential transfer had a higher live birth rate than day 3 ET and had a similar live birth rate to blastocyst transfer in the FET cycle (14). However, the sample size in each study was less than 150 subjects. In the present retrospective study, we included 302 cycles as the observation group. We also included 979 cycles of double day 3 ET and 493 cycles of double blastocyst transfer as the control groups. All of the patients had a history of three consecutive implantation failures and underwent FET cycles. We found that sequential ET had higher rates of implantation and pregnancy than conventional cleavage-stage ET. In addition, the rates of implantation and pregnancy in the sequential ET group were similar to those in the double blastocyst transfer group without increasing the risk of early miscarriage or ectopic pregnancy. This finding suggests that sequential ET may be effective in improving pregnancy outcomes in patients with RIF.

There are several advantages and potential mechanisms in sequential ET. First, successful embryo implantation requires synchronous interactions between endometrial receptivity and embryos with high developmental potential. During the implantation process, many molecular mediators, including cytokines, lipids, adhesion molecules, growth factors, and others, support the establishment of pregnancy (9). Endometrial injury by biopsy catheters during the luteal phase of the menstrual cycle has been shown to improve implantation and pregnancy rates in subsequent treatment cycles (22). During the window of implantation, endometrial preparation is guided not only by maternal factors but also by molecules secreted by the embryo, such as chorionic gonadotropin and interleukin-1 β (IL-1 β) (23). Therefore, after the first ET procedure of sequential ET, mechanical microstimulation caused by catheter insertion and cytokines produced by the embryo and endometrium may not only be a benefit for the implantation of the first transferred embryo but could also promote better implantation conditions and increase the implantation probability following blastocyst transfer (24). Second, the window of implantation is transient in humans, and implantation beyond this window results in pregnancy failure (25). Mechanical stimulation of the endometrium may slightly alter the implantation window for personalized ET, which has a beneficial effect on the receptive endometrium (26). In addition, sequential ET can probably extend the availability time for

transferred embryos to access the implantation window. Moreover, compared with double cleavage-stage day 3 ET, cleavage-stage embryos cultured *in vitro* from 3 days to 5-6 days could be screened to identify embryos with higher implantation potential, resulting in a higher pregnancy rate (27).

Compared with double blastocyst transfer, sequential ET can decrease the risk of ET cycle cancellation since prolonged culture may result in a lack of available blastocysts for transfer. In this study, the clinical pregnancy rate was approximately 25%, even when only a single day 3 embryo was transferred during sequential ET cycles. For patients with many embryos that have good developmental potential, sequential ET is likely to help them improve clinical outcomes. Our study showed that the clinical pregnancy rate was significantly higher and that the early miscarriage rate was significantly lower in the successfully completed sequential ET treatment (a day 3 cleavage-stage ET followed by transfer of a cultured blastocyst) than in the double blastocyst ET treatment. This further suggests that the first day 3 cleavage-stage ET procedure in sequential ET could probably improve the clinical outcomes of ART treatment.

The prevalence of multiple pregnancy is higher with ART than with natural pregnancy (28). This is related to the number of ETs, and there is a consequent impact on maternal and newborn outcomes (29, 30). In the last two decades, controlling the number of embryos transferred (single ET per cycle) has been advocated to reduce the risks of multiple gestations (28, 31). However, compared to the successive failure of IVF-ET, increasing the probability of pregnancy is more beneficial to patients economically and psychologically, even though it sometimes increases the risk of multiple pregnancy. In the case of multiple pregnancy, multifetal pregnancy reduction is also an option for patients to reduce the risk of maternal and fetal complications (32). Therefore, transferring two embryos is a strategy used for patients with RIF. In contrast to previous studies, this study showed that the multiple pregnancy rate in the sequential ET group was significantly lower than that in the double cleavage-stage ET group and the double blastocyst group. This seems to be another benefit of sequential ET. However, further studies are needed to investigate and define multiple pregnancy occurrence in sequential ET.

The larger population and two types of ET (double cleavage-stage embryos and double blastocysts) as controls are the strengths of our study. However, it also had some limitations, including its retrospective nature and lack of data related to live birth. Prospective studies are needed to identify this scientific issue to meet the clinical demand.

In conclusion, this study investigated the value of sequential ET in patients with RIF in FET cycles. Sequential ET was associated with a higher implantation rate, hCG positive rate, and clinical pregnancy rate than double cleavage-stage ET and comparable to those of the double blastocyst-stage ET group without increasing the risk of early miscarriage or ectopic pregnancy. Sequential ET had a lower multiple pregnancy rate than double cleavage-stage ET and double blastocyst-stage ET. In addition, the clinical pregnancy rate was significantly higher, and the early miscarriage rate was significantly lower in the

sequential ET than in the double blastocyst-stage ET when cycles of blastocyst culture failure in the sequential ET group were excluded. These findings suggested that sequential ET is an effective and beneficial option for patients with RIF.

Data availability statement

The original contributions presented in the study are included in the article. Further inquiries can be directed to the corresponding authors.

Ethics statement

The studies involving humans were approved by Peking University Third Hospital Medical Science Research Ethics Committee (M2022128). The studies were conducted in accordance with the local legislation and institutional requirements. Written informed consent for participation was not required from the participants or the participants' legal guardians/next of kin in accordance with the national legislation and institutional requirements.

Author contributions

HW and XL conceived the idea. JG and JL reviewed the literature and designed the study. YY and TT collected the data and conducted the analysis. JG designed the figures and tables and wrote the manuscript. YL, PL, RL, JQ, HW and XL coordinated the study and revised the manuscript. All authors contributed to the article and approved the submitted version.

References

1. Fauser BC. Towards the global coverage of a unified registry of IVF outcomes. *Reprod BioMed Online* (2019) 38(2):133–7. doi: 10.1016/j.rbmo.2018.12.001
2. Mitri F, Nayot D, Casper RF, Bentov Y. Current tools for the optimization of embryo transfer technique for recurrent implantation failure. *Minerva Ginecol* (2016) 68(4):431–49.
3. De Geyter C, Calhaz-Jorge C, Kupka MS, Wyns C, Mocanu E, Motrenko T, et al. ART in Europe, 2014: results generated from European registries by ESHRE: The European IVF-monitoring Consortium (EIM) for the European Society of Human Reproduction and Embryology (ESHRE). *Hum Reprod* (2018) 33(9):1586–601. doi: 10.1093/humrep/dey242
4. Coughlan C, Ledger W, Wang Q, Liu F, Demirel A, Gurgan T, et al. Recurrent implantation failure: definition and management. *Reprod BioMed Online* (2014) 28(1):14–38. doi: 10.1016/j.rbmo.2013.08.011
5. Bashiri A, Halper KI, Orvieto R. Recurrent Implantation Failure-update overview on etiology, diagnosis, treatment and future directions. *Reprod Biol Endocrinol* (2018) 16(1):121. doi: 10.1186/s12958-018-0414-2
6. Busnelli A, Reschini M, Cardellicchio L, Vegetti W, Somigliana E, Vercellini P. How common is real repeated implantation failure? An indirect estimate of the prevalence. *Reprod BioMed Online* (2020) 40(1):91–7. doi: 10.1016/j.rbmo.2019.10.014
7. Koot YE, Teklenburg G, Salker MS, Brosens JJ, Macklon NS. Molecular aspects of implantation failure. *Biochim Biophys Acta* (2012) 1822(12):1943–50. doi: 10.1016/j.bbdis.2012.05.017
8. Saxtorph MH, Hallager T, Persson G, Petersen KB, Eriksen JO, Larsen LG, et al. Assessing endometrial receptivity after recurrent implantation failure: a prospective controlled cohort study. *Reprod BioMed Online* (2020) 41(6):998–1006. doi: 10.1016/j.rbmo.2020.08.015
9. Governini L, Luongo FP, Haxhiu A, Piomboni P, Luddi A. Main actors behind the endometrial receptivity and successful implantation. *Tissue Cell* (2021) 73:101656. doi: 10.1016/j.tice.2021.101656
10. Craciunas L, Gallos I, Chu J, Bourne T, Quenby S, Brosens JJ, et al. Conventional and modern markers of endometrial receptivity: a systematic review and meta-analysis. *Hum Reprod Update* (2019) 25(2):202–23. doi: 10.1093/humupd/dmy044
11. Arefi S, Ataei M, Maleki N, Yari N, Razi S, Amirajam S. Sequential (two-step) day 3/day 5 frozen-thawed embryo transfer: does it improve the pregnancy rate of patients suffering recurrent implantation failure? *J Med Life* (2022) 15(11):1365–70. doi: 10.25122/jml-2022-0041
12. Stamenov GS, Parvanov DA, Chaushev TA. Mixed double-embryo transfer: A promising approach for patients with repeated implantation failure. *Clin Exp Reprod Med* (2017) 44(2):105–10. doi: 10.5653/cepm.2017.44.2.105
13. Torky H, Ahmad A, Hussein A, El-Desouky ES, Aly R, Ragab M, et al. Comparing sequential vs day 3 vs day 5 embryo transfers in cases with recurrent implantation failure: randomized controlled trial. *JBRA Assist Reprod* (2021) 25(2):185–92. doi: 10.5935/1518-0557.20200083
14. Hu YL, Wang Y, Geng LH, Meng XQ, Xu HJ, Adu-Gyamfi EA, et al. Effects of sequential cleavage and blastocyst embryo transfer on pregnancy outcomes in patients with poor ovarian response. *J Reprod Immunol* (2023) 155:103780. doi: 10.1016/j.jri.2022.103780
15. Medicine ASIR, Embryology ESIG. Istanbul consensus workshop on embryo assessment: proceedings of an expert meeting. *Reprod BioMed Online* (2011) 22(6):632–46. doi: 10.1016/j.rbmo.2011.02.001
16. Gardner DK, Lane M, Stevens J, Schlenker T, Schoolcraft WB. Blastocyst score affects implantation and pregnancy outcome: towards a single blastocyst transfer. *Fertil Steril* (2000) 73(6):1155–8. doi: 10.1016/S0015-0282(00)00518-5

Funding

This work was supported by the National Natural Science Foundation of China (81701407), the National Key Research and Development Program of China (2022YFC2702901, 2022YFC2703004), and the Funding from the Key Laboratory of Assisted Reproduction (Peking University), Ministry of Education (BYSYSZKF2022002).

Acknowledgments

The authors thank all the staff in our center, especially Lixue Chen for their support in collecting and collating the data.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

17. Abramovici H, Dirnfeld M, Weisman Z, Sorokin Y, Lissak A, Roife A, et al. Pregnancies following the interval double-transfer technique in an *in vitro* fertilization-embryo transfer program. *J In Vitro Fert Embryo Transf* (1988) 5(3):175–6. doi: 10.1007/BF01131183
18. Ji M, Zhang L, Fu X, Xie W, Wu X, Shu J. The outcomes of sequential embryo transfer in patients undergoing *in vitro* fertilization with frozen-thawed embryos: A retrospective study. *J Obstet Gynaecol Res* (2022) 48(10):2563–70. doi: 10.1111/jog.15369
19. Kyono K, Fukunaga N, Chiba S, Nakajo Y, Fuchinoue K, Yagi A, et al. Two-step consecutive transfer of early embryos and blastocysts. *Reprod Med Biol* (2003) 2(3):133–7. doi: 10.1046/j.1445-5781.2003.00031.x
20. Tehraninejad ES, Raisi E, Ghaleh FB, Rashidi BH, Azimineko E, Kalantari V, et al. The sequential embryo transfer compared to blastocyst embryo transfer in *in vitro* fertilization (IVF) cycle in patients with the three repeated consecutive IVF. A randomized controlled trial. *Gynecol Endocrinol* (2019) 35(11):955–9. doi: 10.1080/09513590.2019.1613639
21. Zhang J, Wang C, Zhang H, Zhou Y. Sequential cleavage and blastocyst embryo transfer and IVF outcomes: a systematic review. *Reprod Biol Endocrinol* (2021) 19(1):142. doi: 10.1186/s12958-021-00824-y
22. Gnainsky Y, Granot I, Aldo PB, Barash A, Or Y, Schechtman E, et al. Local injury of the endometrium induces an inflammatory response that promotes successful implantation. *Fertil Steril* (2010) 94(6):2030–6. doi: 10.1016/j.fertnstert.2010.02.022
23. Massimiani M, Lacconi V, La Civita F, Ticconi C, Rago R, Campagnolo L. Molecular signaling regulating endometrium-blastocyst crosstalk. *Int J Mol Sci* (2019) 21(1):23. doi: 10.3390/ijms21010023
24. Fang C, Huang R, Li TT, Jia L, Li LL, Liang XY. Day-2 and day-3 sequential transfer improves pregnancy rate in patients with repeated IVF-embryo transfer failure: a retrospective case-control study. *Reprod BioMed Online* (2013) 26(1):30–5. doi: 10.1016/j.rbmo.2012.10.004
25. Cha J, Sun X, Dey SK. Mechanisms of implantation: strategies for successful pregnancy. *Nat Med* (2012) 18(12):1754–67. doi: 10.1038/nm.3012
26. Hashimoto T, Koizumi M, Doshida M, Toya M, Sagara E, Oka N, et al. Efficacy of the endometrial receptivity array for repeated implantation failure in Japan: A retrospective, two-centers study. *Reprod Med Biol* (2017) 16(3):290–6. doi: 10.1002/rmb2.12041
27. Martins WP, Nastri CO, Rienzi L, van der Poel SZ, Gracia C, Racowsky C. Blastocyst vs cleavage-stage embryo transfer: systematic review and meta-analysis of reproductive outcomes. *Ultrasound Obstet Gynecol* (2017) 49(5):583–91. doi: 10.1002/uog.17327
28. Kim HH, Matevosian K. Are two better than one? Two sequential transfers of a single embryo may be better than a double-embryo transfer. *Fertil Steril* (2020) 114(2):267–8. doi: 10.1016/j.fertnstert.2020.04.064
29. Chambers GM, Hoang VP, Lee E, Hansen M, Sullivan EA, Bower C, et al. Hospital costs of multiple-birth and singleton-birth children during the first 5 years of life and the role of assisted reproductive technology. *JAMA Pediatr* (2014) 168(11):1045–53. doi: 10.1001/jamapediatrics.2014.1357
30. Murray SR, Norman JE. Multiple pregnancies following assisted reproductive technologies—a happy consequence or double trouble? *Semin Fetal Neonatal Med* (2014) 19(4):222–7. doi: 10.1016/j.siny.2014.03.001
31. Cutting R. Single embryo transfer for all. *Best Pract Res Clin Obstet Gynaecol* (2018) 53:30–7. doi: 10.1016/j.bpobgyn.2018.07.001
32. Sebghati M, Khalil A. Reduction of multiple pregnancy: Counselling and techniques. *Best Pract Res Clin Obstet Gynaecol* (2021) 70:112–22. doi: 10.1016/j.bpobgyn.2020.06.013



OPEN ACCESS

EDITED BY

Hong Zhang,
Second Affiliated Hospital of Soochow
University, China

REVIEWED BY

Alaa Ismail,
Women's Health Hospital, Egypt
Alan Decherney,
Clinical Center (NIH), United States

*CORRESPONDENCE

Chong Qiao
✉ qiaochong2002@163.com

[†]These authors have contributed
equally to this work and share
first authorship

RECEIVED 02 May 2023

ACCEPTED 21 August 2023

PUBLISHED 18 September 2023

CITATION

Zhang L, Du Y, Zhou J, Li J, Shen H, Liu Y,
Liu C and Qiao C (2023) Diagnostic
workup of endocrine dysfunction in
recurrent pregnancy loss: a cross-sectional
study in Northeast China.
Front. Endocrinol. 14:1215469.
doi: 10.3389/fendo.2023.1215469

COPYRIGHT

© 2023 Zhang, Du, Zhou, Li, Shen, Liu, Liu
and Qiao. This is an open-access article
distributed under the terms of the [Creative
Commons Attribution License \(CC BY\)](#). The
use, distribution or reproduction in other
forums is permitted, provided the original
author(s) and the copyright owner(s) are
credited and that the original publication in
this journal is cited, in accordance with
accepted academic practice. No use,
distribution or reproduction is permitted
which does not comply with these terms.

Diagnostic workup of endocrine dysfunction in recurrent pregnancy loss: a cross-sectional study in Northeast China

Liyang Zhang[†], Yushu Du[†], Jingshuang Zhou[†], Jiapo Li[†],
Hongfei Shen, Yilin Liu, Chuanyang Liu and Chong Qiao*

Obstetrics and Gynaecology Department, Shengjing Hospital of China Medical University,
Shenyang, China

Objective: To evaluate the prevalence of abnormal endocrine dysfunction for recurrent pregnancy loss (RPL) amongst patients with two versus three or more pregnancy losses.

Methods: This cross-sectional study retrospectively collected pre-pregnancy data of 537 women diagnosed with RPL in Shengjing Hospital of China Medical University from 2017 to 2022, including the baseline data of patients and the test results of endocrine factors. Several endocrine dysfunction included in this study were: thyroid dysfunction, obesity, hyperprolactinemia, polycystic ovary syndrome and blood glucose abnormality. Furthermore, vitamin D level were collected to study its relationship with endocrine dysfunction. Finally, we subdivided the patients according to the number of previous pregnancy loss and compared the prevalence of endocrine dysfunction between subgroups.

Results: Among 537 RPL patients, 278 (51.8%) patients had abnormal endocrine test results. The highest incidence of endocrine dysfunction was thyroid dysfunction (24.39%, 131/537), followed by hyperprolactinemia (17.34%, 85/490), obesity (10.8%, 58/537), polycystic ovary syndrome (10.50%, 56/533), and abnormal blood glucose (5.29%, 27/510). Only 2.47%(13/527) of patients have vitamin D level that reach the standard. After subdividing the population according to the number of pregnancy loss, we did not find that the incidence of endocrine dysfunction ($P=0.813$), thyroid dysfunction ($P=0.905$), hyperprolactinemia ($P=0.265$), polycystic ovary syndrome ($P=0.638$), blood glucose abnormality ($P=0.616$) and vitamin D deficiency ($P=0.908$) were different among patients with two versus three or more pregnancy losses. However, obesity ($P=0.003$) was found more frequently observed in patients with more times of pregnancy loss.

Conclusion: The prevalence of endocrine dysfunction in RPL population is high. There is no difference in the prevalence of endocrine dysfunction, except for obesity, among patients with two or more pregnancy losses, which may suggest investigations of endocrine dysfunction when patients have two pregnancy losses.

KEYWORDS

recurrent miscarriage, PCOS, hyperprolactinemia, obesity, vitamin D, thyroid dysfunction, blood glucose abnormality

Introduction

Recurrent miscarriage can be devastating for women who wish to have children, with a global prevalence ranging from 1 to 3% (1). Individuals who experience recurrent miscarriage are at increased risk of many obstetric complications, as are the emotional and psychological harms of miscarriage. This is not the end of the physical and psychological consequences of recurrent miscarriages, since complex etiologic screening and expensive systemic therapy can also be financially stressful for a family (2). Endocrinological factors are now a critical component of the screening process used by clinicians to screen patients for recurrent miscarriage. The secretion of hormones such as thyroid stimulating hormone and prolactin has an irreplaceable impact on pregnancy outcomes. Such as prolactin, it may play an important role in maintaining corpus luteum function and progesterone secretion, with potential impact on the establishment of pregnancy (3). Hyperprolactinaemia plays a critical role in infertility because it suppresses the production of GnRH and thus pituitary gonadotropins (4). The most common endocrine abnormalities seen in patients with recurrent miscarriage are luteal phase defect, polycystic ovary syndrome, thyroid dysfunction, obesity, and hyperprolactinaemia. The incidence and risks of these endocrine abnormalities in recurrent miscarriage have been the subject of many independent studies, but studies systematically describing the proportion of endocrine factor abnormalities in patients with recurrent miscarriage are still lacking.

Not only are screening factors varied and complex, but the timing of screening is also a challenge in current research on recurrent miscarriage (5). Due to regional differences in the definition of recurrent miscarriage and clinicians' own practice experience, some have begun routine etiologic screening for recurrent miscarriage following two pregnancy losses, whereas others wait for a third or more pregnancy losses before initiating etiologic screening. Based on these two points, the aims of this study include (1): To describe the proportion of endocrine dysfunction in patients with recurrent miscarriages (2). To compare whether there are differences in endocrine dysfunction between patients with recurrent miscarriages with different numbers of miscarriages.

Materials and methods

Study sample

The China Medical University Birth Cohort is an ongoing prospective cohort study that includes a sub-cohort of patients with recurrent miscarriage specifically enrolled in the Recurrent Miscarriage Clinic. In this cross-sectional study, data from 2017 to 2022 are collected from the recurrent miscarriage clinic at Shengjing Hospital, a local tertiary center, with patients being enrolled according to the following criteria: 1) Patients with two or more pregnancy losses and have completed a detailed history taking form of pregnancy losses. 2) Patients who have received a comprehensive aetiological screening prior to conception for recurrent miscarriage regarding endocrine factors (including thyroid dysfunction, obesity, hyperprolactinemia, polycystic ovary syndrome and blood glucose abnormality) at our hospital. 3) Patients did not take medicines that may affect the test results (including traditional Chinese medicine) before screening. Whereas patients with abnormal results of the endocrine factors on the initial examination will need at least a second repeat examination to make the diagnosis. Simple random sampling was adopted for the study.

Pregnancy loss as defined in this study was urine/blood β -hCG positive or ultrasound-confirmed pregnancy sacs. Pregnancy loss was defined as any spontaneous pregnancy loss or fetal weight \leq 500g before 20 weeks. Molar pregnancy, ectopic pregnancy, implantation failure and pregnancy terminations were excluded from the analysis.

Sample-size calculation

The sample size was calculated using an online sample calculation website, <http://riskcalc.org:3838/samplesize/>. The collection of data from 100 cases in the preliminary stage allowed us to roughly determine the proportion of various endocrine abnormalities in the three populations (patients with two/three/four and more pregnancy losses). Glucose abnormalities were the type of endocrine abnormality that had the lowest prevalence and had a prevalence of approximately 5%. The Type I error rate set in this study was 0.05, the degree of certainty of the study was 0.8, and the approximate ratio of

the sample size between the three populations was 1. A minimum sample size of 480 was calculated for this study.

Data collecting

A patient history collection form is used to collect baseline characteristics, which asks the patient to describe in detail her menstrual history and maternal history, including the number of miscarriages, the cause of the miscarriages, and the presence of ultrasound images to determine the occurrence of early intrauterine pregnancy. The clinician reviews this information (usually with more detailed questioning) in order to clarify the accuracy of the data collected. Whereas outcome-related data are collected primarily through the collection of laboratory test results obtained from the hospital's electronic medical record system. We do not capture data from tests conducted by patients at other hospitals, since differences in the kits can lead to inaccurate results.

The baseline characteristics collected were: age, height, weight, number of pregnancies and number of pregnancy losses. Outcome data collected included PCOS, thyroid dysfunction, hyperprolactinaemia, blood glucose abnormality, obesity, and levels of vitamin D. Premature ovarian failure was removed from the outcome events because of a small number of cases; luteal phase defect was removed from the outcome events because of diagnostic challenges. All baseline data were collected from the patients on their first visit to the recurrent miscarriage clinic. Data on all disease diagnoses were obtained in the non-pregnant state.

Diagnostic criteria

The diagnosis of PCOS is based on the revised Rotterdam diagnosis (6). Thyroid dysfunction is categorized as abnormal thyroid autoimmune antibodies alone, elevated or reduced TSH levels alone, and abnormal TSH levels in combination with abnormal levels of autoimmune antibodies; An abnormal TSH is diagnosed as a TSH of less than 0.3 μ IU/ml or more than 4.8 μ IU/ml (Specific reference values established for non-pregnant local normal individuals). Hyperprolactinaemia was defined as PRL greater than 26.72 ng/ml. abnormal blood glucose was classified as impaired glucose tolerance ($6.1 \text{ mmol/L} \leq \text{fasting glucose} < 7.0 \text{ mmol/L}$), and diabetes mellitus (fasting glucose $\geq 7.0 \text{ mmol/L}$). Diagnosis of obesity is based on the criteria for diagnosis of obesity developed by the working group on obesity in China (7). Endocrine dysfunction is determined by a combination of PCOS, abnormalities in thyroid function, hyperprolactinaemia, glucose abnormalities, and obesity; if any of these abnormalities are present, the patient is considered to have an endocrine dysfunction. Furthermore, because vitamin D may affect many endocrine factors in patients with recurrent miscarriages, vitamin D levels were also included in the present study and examined as a separate outcome; the concentration of Vitamin D was measured as 25 hydroxyvitamin D. 25-OH Vit D $\leq 20 \text{ ng/ml}$ was diagnosed as a

vitamin D deficiency; A diagnosis of vitamin D insufficiency was made if $20 < 25\text{-OH Vit D} \leq 30 \text{ ng/ml}$. Patients were asked to undergo tests related to recurrent pregnancy loss when they are nonpregnant and at least three months after their last pregnancy loss. Patients undergoing these laboratory tests are advised to avoid cold or menstrual periods, which can affect the results of laboratory tests. Screening time for hormones is 1-3 days of menstruation.

Analyzed data

For comparisons of baseline data, the information about the measure is expressed as the mean \pm the standard deviation. Counts are shown as quartiles. Comparisons of endocrine abnormality rates between those with two and three miscarriages, and those with more than three miscarriages, were made using a two-sided Pearson chi-square test. Since multiple group comparisons were performed, we used Bonferroni correction. The vitamin D comparisons were analyzed by ANOVA. All data analysis was carried out in SPSS, windows, version 25.

Results

General situation of patients and the proportion of endocrine dysfunction

The total number of RPL patients included in this study was 537, including 278 patients with a diagnosis of endocrine dysfunction, comprising 51.8% of RPL patients. Fifty-six of these patients were diagnosed with PCOS (10.50%, 56/533); 85 patients were diagnosed with hyperprolactinaemia (17.34%, 85/490); Twenty-seven patients had a diagnosis of blood glucose abnormality (5.29%, 27/510); 58 patients were diagnosed with obesity (10.80%, 58/537). A total of 537 patients had their vitamin D levels tested, and the average vitamin D value was $15.11 \pm 6.42 \text{ ng/ml}$. 83.49% of these patients, (440/527) had a diagnosis of vitamin D deficiency, 14.04% (74/537) were diagnosed with vitamin D insufficiency, and only 2.47% (13/527) of patients had vitamin D levels that were up to the standard. A total of 131 patients (24.39%) were diagnosed with thyroid function abnormalities (131/537). 101 of the patients with abnormal thyroid function were found to have abnormal autoantibodies alone, representing 77.1% of patients with abnormal thyroid function (101/131); Eighteen had an abnormal TSH level alone, representing 13.74% of patients with an abnormal thyroid function (13/131); and 12 had combined autoantibodies and TSH levels, accounting for 9.16% of patients with abnormal thyroid. Among 113 patients with positive autoimmune thyroid antibodies, 39.82% (45/113) were found to be positive for anti-Tg alone; Of these, 17.70% (20/113) were positive for anti-Tpo antibody alone; and 42.48% (48/113) were positive for both antibodies. Further details can be found in [Tables 1 and 2](#).

TABLE 1 Baseline characteristics and the proportion of endocrine disorders.

| Characteristic | n(%) |
|---------------------------|-------------|
| Age | |
| 20-29 | 133(24.77) |
| 30-34 | 272(51.03) |
| 35-45 | 132(24.58) |
| BMI | |
| <18.5 | 29(5.40%) |
| 18.5-23.9 | 321(59.78%) |
| 24-28 | 129(24.02%) |
| >28 (obesity) | 58(10.8%) |
| Times of pregnancy loss | |
| 2 | 297(55.31) |
| 3 | 184(34.52) |
| More than 3 | 56(10.51) |
| PCOS | |
| No | 477(89.50) |
| Yes | 56(10.50) |
| Thyroid Dysfunction | |
| No | 406(75.61) |
| Yes | 131(24.39) |
| HPRL | |
| No | 405(82.66) |
| Yes | 85(17.34) |
| Blood glucose abnormality | |
| No | 483(94.71) |
| IGT | 23(4.51) |
| DM | 4(0.78) |
| Vitamin D level | |
| Normal | 13(2.47) |
| Insufficient | 74(14.04) |
| Deficiency | 440(83.49) |

Multiple endocrine dysfunctions

When analyzing whether patients presented with a combination of multiple endocrine disorders, we found that 74.46% (207/278) of patients with RPL had only one endocrine disorder; 23.02% (64/278) had a combination of two endocrine disorders; 2.15% (6/278) had a combination of three endocrine disorders, and only 1 patient had a combination of 4 endocrine disorders. To further examine whether there was an interaction between the various endocrine disorders, or what types of endocrine disorders were seen more frequently together, the association between each of

the endocrine factors was analyzed. Although we did not find a significant association between any two endocrine disorders (results not shown), there was a tendency for PCOS to be associated with obesity ($p=0.059$), which is consistent with our clinical knowledge.

Endocrine dysfunction and the number of pregnancy losses

Comparisons of subgroups revealed that PCOS, thyroid dysfunction, hyperprolactinaemia, blood glucose abnormality, and vitamin D levels were not significantly different between groups. There was a significant difference in obesity between patients with different numbers of pregnancy losses ($p=0.003$). We then stratified the patients by age for different numbers of pregnancy losses and showed that all endocrine disorders were not associated with the number of miscarriages in the subgroups less than 30 years and greater than or equal to 35 years. On the other hand, among those above or equal to age 30 and below age 35, obesity was the only factor that was significantly different between patients with different numbers of pregnancy losses ($p=0.017$).

Discussion

Main findings

This cross-sectional study describes the proportion of endocrine disorders in the recurrent miscarriage population and compares whether there are differences in the distribution of endocrine disorders among those with two, three, and more pregnancy losses. Except for obesity, our results did not find a significant association between the number of pregnancy losses and the distribution of endocrine disorders. This finding may suggest that clinicians need to begin screening for endocrine factors at the beginning of two miscarriages in order to intervene early and prevent patients from experiencing further miscarriages.

Thyroid dysfunction

Currently, abnormal thyroid function is a hot topic in recurrent miscarriage research. Abnormalities in thyroid function typically include both abnormal TSH levels and abnormal autoimmune antibodies. Many clinical studies have been conducted to examine whether abnormal thyroid function can lead to pregnancy loss and other adverse pregnancy outcomes. Recent research suggests that abnormal levels of TSH alone or the presence of thyroid antibodies are associated with pregnancy loss (8). In a meta-analysis of 22 studies, high serum thyroid antibody levels have been shown to lead to recurrent miscarriage and the use of T4 replacement therapy is beneficial in pregnant patients with recurrent miscarriage (9). However, a randomized clinical trial carried in 2019 concluded that that the use of levothyroxine in euthyroid women with thyroid peroxidase antibodies did not result in a higher rate of live births than placebo in normal population (10), and the preconception use of

TABLE 2 Comparison of endocrine disorders among patients with different numbers of pregnancy loss.

| | Times of pregnancy loss | | | P value |
|---------------------------|-------------------------|------------|-----------|---------|
| | 2 | 3 | ≥4 | |
| Endocrine dysfunction | | | | 0.817 |
| No | 146(56.4%) | 88(34.0%) | 25(9.7%) | |
| Yes | 151(54.3%) | 96(34.5%) | 31(11.2%) | |
| PCOS | | | | 0.638 |
| No | 261(54.7%) | 166(34.8%) | 50(10.5%) | |
| Yes | 34(60.7%) | 16(28.6%) | 6(10.7%) | |
| Thyroid Dysfunction | | | | 0.905 |
| No | 225(55.4%) | 140(34.5%) | 41(10.1%) | |
| Yes | 72(55.0%) | 44(33.6%) | 15(11.5%) | |
| HPRL | | | | 0.265 |
| No | 220(53.9%) | 143(35.0%) | 45(11.0%) | |
| Yes | 54(63.5%) | 24(28.2%) | 7(8.2%) | |
| Blood glucose abnormality | | | | 0.616 |
| No | 272(56.3%) | 161(33.3%) | 50(10.4%) | |
| IGT | 10(43.5%) | 9(39.1%) | 4(17.4%) | |
| DM | 2(50.0%) | 2(50.0%) | 0(0.0%) | |
| Vitamin D level | | | | 0.908 |
| Normal | 7(53.8%) | 4(30.8%) | 2(15.4%) | |
| Insufficient | 38(51.4%) | 27(36.5%) | 9(12.2%) | |
| Deficiency | 248(56.4%) | 147(33.4%) | 45(10.2%) | |
| Obesity | | | | 0.003* |
| No | 277(57.8%) | 155(32.4%) | 47(9.8%) | |
| Yes | 20(34.5%) | 29(50.0%) | 9(15.5%) | |

* means P value<0.05.

levothyroxine in recurrent miscarriage population still need further research. The mechanism by which thyroid antibodies contribute to poor pregnancy outcomes may be linked to their action on immune cells in the endothelium; studies have shown that the secretion of IL-4 and IL-10 is significantly reduced in endothelial T cells and that the expression of interferon- γ is significantly increased in antibody-positive patients (11). Likewise, polyclonal B cells were overexpressed in patients with autoimmune thyroid disease, and toxic NK cell migration was significantly enhanced (12). There were no significant differences in the prevalence of thyroid dysfunction according to the number of pregnancy losses reported in this paper. These findings are also similar to previous studies.

PCOS

The second endocrine abnormality that is the focus of this paper is PCOS, whose prevalence in the recurrent miscarriage

population remains a mystery. The reported incidence of polycystic ovarian changes on ultrasound imaging in the RPL population has been reported to range from 4.8 to 82% (13–16). In a meta-analysis published in 2016, the authors included 15 articles that used the Rotterdam diagnosis as a diagnostic criterion for PCOS and concluded that the average prevalence of PCOS in the general population was 10% (17). In the RPL population, the prevalence of PCOS were reported to be 14.3% (18). Common symptoms of PCOS include insulin resistance and hyperinsulinaemia, both of which are independent risk factors for pregnancy loss (13, 19, 20). Therefore, it is important to intervene before pregnancy for PCOS patients to improve miscarriage rates. Unfortunately, there is still a lack of research on how to manage during pregnancy in PCOS patients. Researchers have found that metformin treatment might reduce the risk of miscarriage in PCOS patients (21). However, more research is needed on the safety of medication and its use in the population with recurrent miscarriage.

Obesity and glucose abnormality

Of note, about 35% of PCOS patients have a combination of obesity, which is believed to be associated with pregnancy loss (22, 23). The prevalence of obesity has been increasing in various countries in recent decades (24). The prevalence of obesity in women of reproductive age (20–39 years) in the United States rose from 28.4% to 34% between 1999 and 2008 (25). In China, the prevalence of obesity in adults rose from 3.6% in 1992 to 14.0% in 2014 (26). The hypothalamic-pituitary-ovarian axis is disrupted by obesity, and overweight women have a shorter luteal phase and lower levels of follicle-stimulating hormone, luteinizing hormone, and progesterone (27). However, in the current study, we found that obese patients were more common among women with a greater number of pregnancy losses. Obesity remained associated with the number of pregnancy losses among those aged thirty to thirty-four years even after stratifying for age. It is noteworthy that the subgroup sample sizes of the study population did not achieve the minimum sample size considered for validity following patient stratification, therefore, a larger sample size is required to confirm the association of obesity with pregnancy loss. As with obesity, blood glucose abnormality is also highly associated with pregnancy loss. In a recently published cross-sectional study, the authors suggest that women with recurrent miscarriages are more likely to have impaired β -cell function and abnormal glucose metabolism (28). Hyperglycaemia can inhibit the differentiation of trophoblasts and thereby interfere with implantation, increase oxidative stress, and affect the expression of key genes that are essential for embryogenesis (29). Hyperglycaemia promotes pregnancy loss through the promotion of premature programmed cell death of key progenitor cells within blastocysts (30). In this study, 5.29% of patients had abnormal blood glucose levels. Whereas the prevalence of diabetes among Chinese women of all ages was as high as 11%, the lower prevalence in this study may be due to the occurrence of diabetes being more prevalent in the elderly and in more economically developed regions.

Hyperprolactinaemia

Prolactin is a hormone secreted from the lactotrophic cells in the anterior pituitary. In a randomized trial, Hirahara et al. found that high levels of prolactin increased the risk of pregnancy loss in women with RPL (31). A cross-sectional study of 69 women with RPL and 31 women of reproductive age and 30 women with infertility found that the prevalence of hyperprolactinaemia was similar across groups, though it was highest in the infertility arm and not in the RPL group (32). While the deleterious effects of hyperprolactinaemia in the recurrent miscarriage population remain unclear, due to its potential risk of producing infertility and the patient's desire for children, pre-pregnancy medication is still needed.

Vitamin D

Lastly, the article also analyzed the incidence of vitamin D levels in individuals with different numbers of pregnancy losses and found no significant differences. Vitamin D concentrations are now considered essential for maintaining pregnancy. Endometrium with a greater number of vitamin D receptors is more likely to conceive, while vitamin D deficiency is more likely to result in miscarriage (33). Vitamin D deficiency can result in the development of numerous endocrine defects including PCOS, autoimmune thyroid disease, diabetes, and obesity (34). Despite such an important role of vitamin D in pregnancy, vitamin D insufficiency or deficiency is currently prevalent in the population. In this study, only 2.47% of patients had a normal vitamin D level, whereas 83.49% were diagnosed with a vitamin D deficiency level. Consistent with our results, Li et al. found that as many as 70% of women during pregnancy were deficient in vitamin D levels, and only 1.6% reached normal levels (35). Though the study population is different, the results of our study and those of Li et al. may suggest the high prevalence of insufficient and deficient vitamin D levels in both the pregnant and non-pregnant women in China. This study also suggests that vitamin D insufficiency and deficiency are not associated with the number of pregnancy losses in patients with recurrent miscarriage. Additional studies are needed to determine if vitamin D supplementation can ameliorate miscarriage.

Bias

Bias in cross-sectional studies includes many aspects and a variety of classifications of bias are summarised in the study by Wang et al (36). Because only baseline data collection involved question-based or questionnaire-based data collection (and this data was reviewed by clinicians), non-response bias, loss-to-follow-up bias, observer bias, interviewer bias, and recall bias are all relatively minor contributors to the total bias in this study. Furthermore, because data collectors were not the originators of the study, i.e. data collectors were unaware of the study objective; therefore, the study would have generated less sampling bias, as well as less allocation bias. Prevalence bias is likely to be the largest source of bias in this study, also referred to as Neyman bias, in which some patients with mild or severe diseases will be missing from the data collection process. In this study, the situation that emerged was the lack of patients with mild diseases. Because study data were collected from regional tertiary medical centers throughout the country, they were referred to patients with complex etiology and relatively severe diseases. Furthermore, because inclusion in the study required that patients undergo at least 1 complete etiologic screen for endocrine factors, these inclusion criteria also led to the loss of a proportion of patients with milder diseases. To address this bias, the study's conclusions should be similarly qualified. Tertiary care centers were more likely than local primary care to use the etiologic distribution of endocrine

factors for recurrent miscarriage derived from this study. Furthermore, a significant confounding factor between the number of pregnancy loss and endocrine dysfunction was age. In this study, we propose to use a stratified approach to remove the influence of confounding factors.

Conclusion

In summary, this study describes the proportion of endocrine factor abnormalities in patients with recurrent miscarriages and finds no significant differences in endocrine factor abnormalities other than obesity between patients with recurrent miscarriages depending on the number of pregnancy losses. The findings of this study may support the screening of patients for endocrine-related aetiology after two miscarriages.

Data availability statement

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

Ethics statement

The studies involving humans were approved by the ethics committee of China Medical University. The studies were conducted in accordance with the local legislation and institutional requirements. Written informed consent for participation was not required from the participants or the participants' legal guardians/next of kin in accordance with the national legislation and institutional requirements.

Author contributions

LZ and YD designed the study. JZ and CL were involved in the data collection. YL and HS did the data analysis. JL advised on the conduct of the study. CQ had the conception for the study. All authors listed made important intellectual contribution to the work

and approved the final version of the manuscript for publication. All authors contributed to the article and approved the submitted version.

Funding

This work was supported by National Key R&D Program of China(2016YFC1000404); The National Natural Science Foundation of China (81370735); General Program of National Natural Science Foundation of China (81771610); The Outstanding Scientific Fund of Shengjing Hospital(201706); Distinguished professor of Liaoning Province (2017); Science and Technology Project of Shenyang (20-205-4-004).

Acknowledgments

The authors thank all the patients who took part in this study.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fendo.2023.1215469/full#supplementary-material>

References

1. Bender Atik R, Christiansen OB, Elson J, Kolte AM, Lewis S, Middeldorp S, et al. ESHRE guideline: recurrent pregnancy loss. *Hum Reprod Open* (2018) 2018 (2):hoy004. doi: 10.1093/hropen/hoy004
2. Quenby S, Gallos ID, Dhillon-Smith RK, Podsek M, Stephenson MD, Fisher J, et al. Miscarriage matters: the epidemiological, physical, psychological, and economic costs of early pregnancy loss. *Lancet* (2021) 397(10285):1658–67. doi: 10.1016/S0140-6736(21)00682-6
3. Li W, Ma N, Laird SM, Ledger WL, Li TC. The relationship between serum prolactin concentration and pregnancy outcome in women with unexplained recurrent miscarriage. *J Obstet Gynaecol* (2013) 33:285–8. doi: 10.3109/01443615.2012.759916
4. Abbara A, Clarke SA, Nesbitt A, Ali S, Comninou AN, Hatfield E, Martin NM, et al. Interpretation of serum gonadotropin levels in hyperprolactinaemia. *Neuroendocrinology* (2018) 107(2):105–13. doi: 10.1159/000489264
5. Jaslow CR, Carney JL, Kutteh WH. Diagnostic factors identified in 1020 women with two versus three or more recurrent pregnancy losses. *Fertil Steril* (2010) 93 (4):1234–43. doi: 10.1016/j.fertnstert.2009.01.166
6. Rotterdam ESHRE/ASRM-Sponsored PCOS consensus workshop group. Revised 2003 consensus on diagnostic criteria and long-term health risks related to polycystic ovary syndrome (PCOS). *Hum Reprod* (2004) 19(1):41–7. doi: 10.1093/humrep/deh098

7. Zhou BF. Predictive values of body mass index and waist circumference for risk factors of certain related diseases in Chinese adults—study on optimal cut-off points of body mass index and waist circumference in Chinese adults. *BioMed Environ Sci* (2002) 15(1):83–96.
8. Dong AC, Morgan J, Kane M, Stagnaro-Green A, Stephenson MD. Subclinical hypothyroidism and thyroid autoimmunity in recurrent pregnancy loss: a systematic review and meta-analysis. *Fertil Steril* (2020) 113(3):587–600.e1. doi: 10.1016/j.fertnstert.2019.11.003
9. Xie J, Jiang L, Sadhukhan A, Yang S, Yao Q, Zhou P, et al. Effect of antithyroid antibodies on women with recurrent miscarriage: A meta-analysis. *Am J Reprod Immunol* (2020) 83(6):e13238. doi: 10.1111/aji.13238
10. Dhillon-Smith RK, Middleton LJ, Sunner KK, Cheed V, Baker K, Farrell-Carver S, et al. Levothyroxine in women with thyroid peroxidase antibodies before conception. *N Engl J Med* (2019) 380:1316–25. doi: 10.1056/NEJMoa1812537
11. Stewart-Akers AM, Krasnow JS, Brekosky J, DeLoia JA. Endometrial leukocytes are altered numerically and functionally in women with implantation defects. *Am J Reprod Immunol* (1998) 39(1):1–11. doi: 10.1111/j.1600-0897.1998.tb00326.x
12. Twig G, Shina A, Amital H, Shoenfeld Y. Pathogenesis of infertility and recurrent pregnancy loss in thyroid autoimmunity. *J Autoimmun* (2012) 38(2–3):J275–81. doi: 10.1016/j.jaut.2011.11.014
13. Cockedge KA, Saravelos SH, Metwally M, Li TC. How common is polycystic ovary syndrome in recurrent miscarriage. *Reprod BioMed Online* (2009) 19(4):572–6. doi: 10.1016/j.rbmo.2009.06.003
14. Sugiura-Ogasawara M, Sato T, Suzumori N, Kitaori T, Kumagai K, Ozaki Y. The polycystic ovary syndrome does not predict further miscarriage in Japanese couples experiencing recurrent miscarriages. *Am J Reprod Immunol* (2009) 61(1):62–7. doi: 10.1111/j.1600-0897.2008.00662.x
15. Sagle M, Bishop K, Ridley N, Alexander FM, Michel M, Bonney RC, et al. Recurrent early miscarriage and polycystic ovaries. *BMJ* (1988) 297(6655):1027–8. doi: 10.1136/bmj.297.6655.1027
16. Rai R, Backos M, Rushworth F, Regan L. Polycystic ovaries and recurrent miscarriage—a reappraisal. *Hum Reprod* (2000) 15(3):612–5. doi: 10.1093/humrep/15.3.612
17. Bozdag G, Mumusoglu S, Zengin D, Karabulut E, Yildiz BO. The prevalence and phenotypic features of polycystic ovary syndrome: a systematic review and meta-analysis. *Hum Reprod* (2016) 31(12):2841–55. doi: 10.1093/humrep/dew218
18. Mayrhofer D, Hager M, Walch K, Ghobrial S, Rogenhofer N, Marculescu R, et al. The prevalence and impact of polycystic ovary syndrome in recurrent miscarriage: A retrospective cohort study and meta-analysis. *J Clin Med* (2020) 9(9):2700. doi: 10.3390/jcm9092700
19. Pugeat M, Ducluzeau PH. Insulin resistance, polycystic ovary syndrome and metformin. *Drugs* (1999) 58(Suppl 1):41–6. doi: 10.2165/00003495-199958001-00010
20. Glueck CJ, Streicher P, Wang P. Treatment of polycystic ovary syndrome with insulin-lowering agents. *Expert Opin Pharmacother* (2002) 3(8):1177–89. doi: 10.1517/14656566.3.8.1177
21. Løvvik TS, Carlsen SM, Salvesen Ø, Steffensen B, Bixo M, Gómez-Real F, et al. Use of metformin to treat pregnant women with polycystic ovary syndrome (PregMet2): a randomised, double-blind, placebo-controlled trial. *Lancet Diabetes Endocrinol* (2019) 7:256–66. doi: 10.1016/S2213-8587(19)30002-6
22. Hamilton-Fairley D, Kiddy D, Watson H, Sagle M, Franks S. Low-dose gonadotrophin therapy for induction of ovulation in 100 women with polycystic ovary syndrome. *Hum Reprod* (1991) 6(8):1095–9. doi: 10.1093/oxfordjournals.humrep.a137491
23. Catalano PM, Shankar K. Obesity and pregnancy: mechanisms of short term and long term adverse consequences for mother and child. *BMJ* (2017) 356:j1. doi: 10.1136/bmj.j1
24. Hales CM, Carroll MD, Fryar CD, Ogden CL. Prevalence of obesity and severe obesity among adults: United States, 2017–2018. *NCHS Data Brief* (2020) 360:1–8.
25. Flegal KM, Carroll MD, Ogden CL, Curtin LR. Prevalence and trends in obesity among US adults, 1999–2008. *JAMA* (2010) 303(3):235–41. doi: 10.1001/jama.2009.2014
26. Wang L, Peng W, Zhao Z, Zhang M, Shi Z, Song Z, et al. Prevalence and treatment of diabetes in China, 2013–2018. *JAMA* (2021) 326(24):2498–506. doi: 10.1001/jama.2021.22208
27. Santoro N, Lasley B, McConnell D, Allsworth J, Crawford S, Gold EB, et al. Body size and ethnicity are associated with menstrual cycle alterations in women in the early menopausal transition: The Study of Women's Health across the Nation (SWAN) Daily Hormone Study. *J Clin Endocrinol Metab* (2004) 89(6):2622–31. doi: 10.1210/jc.2003-031578
28. Edugbe AE, James B, Akunaeziri UA, Egbodo CO, Imoh CL, Ajen AS, et al. Beta-cell dysfunction and abnormal glucose metabolism among non-diabetic women with recurrent miscarriages. *Arch Gynecol Obstet* (2020) 301(2):559–64. doi: 10.1007/s00404-019-05407-2
29. Moley KH, Chi MM, Mueckler MM. Maternal hyperglycemia alters glucose transport and utilization in mouse preimplantation embryos. *Am J Physiol* (1998) 275(1):E38–47. doi: 10.1152/ajpendo.1998.275.1.E38
30. Moley KH, Chi MM, Knudson CM, Korsmeyer SJ, Mueckler MM. Hyperglycemia induces apoptosis in pre-implantation embryos through cell death effector pathways. *Nat Med* (1998) 4(12):1421–4. doi: 10.1038/4013
31. Hirahara F, Andoh N, Sawai K, Hirabuki T, Uemura T, Minaguchi H. Hyperprolactinemic recurrent miscarriage and results of randomized bromocriptine treatment trials. *Fertil Steril* (1998) 70(2):246–52. doi: 10.1016/S0015-0282(98)00164-2
32. Triggianese P, Perricone C, Perricone R, De Carolis C. Prolactin and natural killer cells: evaluating the neuroendocrine-immune axis in women with primary infertility and recurrent spontaneous abortion. *Am J Reprod Immunol* (2015) 73(1):56–65. doi: 10.1111/aji.12335
33. Guo J, Liu S, Wang P, Ren H, Li Y. Characterization of VDR and CYP27B1 expression in the endometrium during the menstrual cycle before embryo transfer: implications for endometrial receptivity. *Reprod Biol Endocrinol* (2020) 18(1):24. doi: 10.1186/s12958-020-00579-y
34. Savastio S, Cinquatti R, Tagliaferri F, Rabbone I, Bona G. Vitamin D effects and endocrine diseases. *Minerva Pediatr* (2020) 72(4):326–39. doi: 10.23736/S0026-4946.20.05915-0
35. Li H, Ma J, Huang R, Wen Y, Liu G, Xuan M, et al. Prevalence of vitamin D deficiency in the pregnant women: an observational study in Shanghai, China. *Arch Public Health* (2020) 78:31. doi: 10.1186/s13690-020-00414-1
36. Wang X, Cheng Z. Cross-sectional studies: strengths, weaknesses, and recommendations. *Chest* (2020) 158(1S):S65–71. doi: 10.1016/j.chest.2020.03.012



OPEN ACCESS

EDITED BY

Federico Jensen,
University of Buenos Aires, Argentina

REVIEWED BY

Alicia Motta,
University of Buenos Aires, Argentina
Natalin Jimena Valeff,
CONICET Centro de Estudios
Farmacológicos y Botánicos (CEFYO),
Argentina

*CORRESPONDENCE

He Qiaohua
✉ hqhuaaaa@126.com

RECEIVED 20 June 2023

ACCEPTED 31 August 2023

PUBLISHED 29 September 2023

CITATION

Fang Y, Jingjing F, Tiantain C, Huanhuan X
and Qiaohua H (2023) Impact of the
number of previous embryo implantation
failures on IVF/ICSI-ET pregnancy
outcomes in patients younger than 40
years: a retrospective cohort study.
Front. Endocrinol. 14:1243402.
doi: 10.3389/fendo.2023.1243402

COPYRIGHT

© 2023 Fang, Jingjing, Tiantain, Huanhuan
and Qiaohua. This is an open-access article
distributed under the terms of the [Creative
Commons Attribution License \(CC BY\)](#). The
use, distribution or reproduction in other
forums is permitted, provided the original
author(s) and the copyright owner(s) are
credited and that the original publication in
this journal is cited, in accordance with
accepted academic practice. No use,
distribution or reproduction is permitted
which does not comply with these terms.

Impact of the number of previous embryo implantation failures on IVF/ICSI-ET pregnancy outcomes in patients younger than 40 years: a retrospective cohort study

Yuan Fang^{1,2}, Fan Jingjing^{1,2}, Cheng Tiantain^{1,2}, Xie Huanhuan^{1,2}
and He Qiaohua^{1,3*}

¹Reproductive Medicine Center, Henan Provincial People's Hospital, Zhengzhou, China, ²People's Hospital of Henan University, Zhengzhou, China, ³People's Hospital of Henan University, People's Hospital of Zhengzhou University, Zhengzhou, China

Objective: The objective of this study was to examine the influence of repeated embryo implantation failures on pregnancy outcomes among patients under 40 years of age undergoing *in vitro* fertilization/intracytoplasmic sperm injection embryo transfer (IVF/ICSI-ET).

Materials and methods: A retrospective analysis was conducted on the clinical data of 13,172 patients who underwent 16,975 IVF/ICSI-ET treatment cycles at Henan Reproductive Hospital between January 1, 2015, and December 31, 2018. Patients were categorized into four groups based on the number of previous embryo implantation failure cycles: Group A=no implantation failure, Group B=1 implantation failure, Group C=2 implantation failures, Group D= ≥ 3 implantation failures. Baseline characteristics and pregnancy outcomes were compared among the four groups. The impact of the number of previous embryo implantation failures on pregnancy outcomes among IVF/ICSI-ET patients was investigated using univariate and multiple regression analyses.

Results: Univariate logistic regression analysis demonstrated that factors such as the number of previous embryo implantation failures, female age, basal follicle count, endometrial thickness, total number of oocytes retrieved, type of cycle, number of high-quality embryos transferred, and stage of embryo development significantly affected implantation rate, clinical pregnancy rate, early spontaneous abortion rate, and live birth rate (all $P < 0.05$). The duration of infertility and anti-Mullerian hormone (AMH) levels were also found to influence implantation rate, clinical pregnancy rate, and live birth rate (all $P < 0.05$). Upon conducting multivariate logistic regression analysis and adjusting for confounding factors such as age, AMH levels, basal follicle count, endometrial thickness, total number of oocytes obtained, cycle type, number of high-quality embryos transferred, ovarian stimulation protocol, and stage of embryo development, it was revealed that, compared to Group A, Groups B, C, and D exhibited significantly lower implantation and live birth rates, as well as a significantly higher risk of early spontaneous abortion (all $P < 0.05$).

Conclusions: The number of previous embryo implantation failures is an independent factor affecting implantation rate, clinical pregnancy rate, spontaneous abortion rate and live birth rate of patients underwent IVF/ICSI-ET. With the increase of the number of previous embryo implantation failures, the implantation rate, clinical pregnancy rate and live birth rate of patients underwent IVF/ICSI-ET decreased significantly, and the rate of early spontaneous abortion gradually increased.

KEYWORDS

IVF/ICSI, implantation rate (IR), Live birth rate (LBR), pregnancy outcome, recurrent implantation failure (RIF)

Introduction

Infertility has increasingly become a significant global public health and sociological issue that profoundly impacts human development and health (1). The World Health Organization reports that infertility affects approximately 8–14% of women of reproductive age in western developed countries, with the prevalence rising to 25–30% in some developing regions, such as Africa and the Middle East (2). Currently, the infertile population in China exceeds 40 million, with an incidence of 12.5% among women of childbearing age (3). With the rapid advancement of assisted reproductive technology (ART), the implantation rate for a single high-quality blastocyst transfer has improved to as high as 65% (4). The *in vitro* fertilization/intracytoplasmic sperm injection-embryo transfer (IVF/ICSI-ET) technique has helped numerous infertile families achieve a healthy baby. However, approximately 10% of these individuals are unable to attain a clinical pregnancy even after three or more embryo transfers (5). Despite facing considerable emotional and financial strain, most of these patients still desire to continue their IVF/ICSI-ET treatment in hopes of increasing the “take-home-baby” rate.

Numerous factors influence the embryo implantation rate, including embryo chromosomal abnormalities, diminished endometrial receptivity, asynchrony between embryo and endometrium, maternal endocrine disorders, and thrombotic tendencies. The number of previous embryo implantation failures also plays an essential role in the embryo implantation rate (6). As the number of previous embryo implantation failures rises, the financial and psychological burden on patients grows as well. These individuals require clinicians to help predict the success probability of subsequent embryo transfer cycles based on the number of previous embryo implantation failures. However, the impact of the number of previous embryo implantation failures on the pregnancy outcome of the next IVF/ICSI-ET for infertile patients remains a contentious issue (7–11). Consequently, our study aimed to investigate the effect of the number of prior embryo implantation failures on the pregnancy outcome of the subsequent IVF/ICSI-ET treatment for patients aged <40 years.

Materials and methods

Patient and cycle characteristics

The present study analyzed the clinical data of 16,975 cycles of IVF/ICSI from 13,172 patients who were recruited from the Center of Henan Reproductive Hospital between January 2015 and December 2018. The inclusion criteria comprised the presence of indications for IVF/ICSI-ET and the absence of contraindications, in addition to an age < 40 years. The exclusion criteria included incomplete data or loss to follow-up, ultrasound-identified abnormal uterine structure, such as single or bicornuate uterine diaphragm, chromosomal abnormalities in either partner, previous pregnancy from IVF/ICSI-ET cycles, and the use of donor eggs or sperm. The IVF/ICSI-ET cycles were categorized into four groups according to the number of previous embryo implantation failures: Group A included 13172 cycles without any implantation failures; Group B included 2989 cycles with one embryo implantation failure; Group C included 658 cycles with two embryo implantation failures; and Group D included 156 cycles with three or more embryo implantation failures. The Ethics Committee of Henan Provincial People's Hospital approved the study (No. (28) Len Audit (2022)).

Controlled ovarian stimulation protocol

The appropriate protocol for controlled ovulation was selected based on the patient's age, body mass index (BMI), basal follicle count, basal sex hormone levels, anti-müllerian hormone (AMH) levels, and previous IVF/ICSI-ET protocols. Patients underwent controlled ovulation stimulation (COS) and the dosage of gonadotropin was adjusted according to follicular size and hormonal levels. When the follicular diameter was ≥ 18 mm in three or more follicles, 4000–10000 IU of human chorionic gonadotropin (hCG) (Lizhu Pharmaceutical Company, China) was administered to induce ovulation. Oocyte retrieval guided by vaginal ultrasound was performed 36–37 h later. Routine luteal

support was performed after oocyte retrieval, and one to two available embryos were transferred on either the third or fifth day after oocyte retrieval.

Preparation protocols for frozen-thawed ET

Patients with regular menstrual cycles and normal ovulation underwent natural cycle preparation of the endometrium. Transvaginal ultrasound was used to monitor follicular development and endometrial condition starting from day 11 of the menstrual cycle. Endometrial transformation occurred on the day of ovulation, and the endometrium was transformed 3 or 5 days before the transfer of cleavage-stage or blastocyst-stage embryos, respectively. For patients with irregular menstruation, hormone replacement therapy was employed using estradiol valerate (Progynova, 1 mg/tablet, Bayer, Germany). Oral administration of 4–8 mg/day was started on day 2–4 of the menstrual cycle. The dose of Progynova was maintained when the endometrial thickness was ≥ 8 mm, and the duration of administration was ≥ 11 days, while luteal support was given at the same time. After progesterone was administered, embryos were transferred on the third or fifth day. Endometrial transformation was performed on day 4 or 6 for the transfer of cleavage-stage or blastocyst-stage embryos, respectively.

Conventional luteal phase support regimen

Luteal support was provided through daily administration of vaginal progesterone gel (Crinone, 90 mg/unit, Merck & Serono) at a dosage of 90 mg, along with a once-daily oral dose of 20 mg dydrogesterone. The estrogen and progesterone doses remained unchanged after embryo transfer until blood β -hCG levels were measured on the 14th day post-embryo transfer. In case of pregnancy (β -hCG > 50 U/L), the original luteal support was maintained and the dosage was gradually reduced until the 10th week of pregnancy.

Clinical observation indicators

Clinical pregnancy was defined as the presence of an intrauterine gestational sac, confirmed by vaginal ultrasonography 4–6 weeks after embryo transfer. Spontaneous miscarriage was defined as pregnancy termination before 28 weeks of gestation with a fetal weight of less than 1000g. When spontaneous miscarriage occurs before 12 weeks of gestation, it is considered early pregnancy loss. Newborns delivered after 28 weeks of gestation and surviving are considered live births.

Other observed indicators included age, duration of infertility, BMI, type of infertility, cycle type, stage of embryo development, number(No.) of embryos transferred, endometrial thickness, No. of good-quality embryos, AMH levels, antral follicle count (AFC), infertility factors, and ovulation protocol.

Statistical analysis

Statistical analysis was conducted using Empower Stats software based on R language. Continuous variables were presented as mean \pm standard deviation (SD), and categorical variables were presented as N (%). Intergroup comparisons were performed using one-way analysis of variance (ANOVA) and χ^2 tests for categorical variables. Univariate regression analysis was used to identify potential factors affecting pregnancy outcomes. Furthermore, multivariate Logistic regression model with generalized estimating equation (GEE) was chosen to conduct univariate and multivariate analysis of the association of effect of the number of previous embryo implantation failures on pregnancy outcomes to account for the correlation between cycles from the same patient and to obtain ORs and 95% CIs for the risk of the number of implantation failures associated with pregnancy outcomes, while adjusting for confounding factors. A value of $P < 0.05$ was considered statistically significant.

Results

A total of 13,172 patients who underwent 16,957 IVF/ICSI cycles were recruited and categorized into four groups based on the number of previous embryo implantation failures. The patient demographics and characteristics are presented in Table 1. The four groups exhibited significant differences in terms of age, duration of infertility, AMH levels, AFC, endometrial thickness, number of eggs retrieved, stage of embryo development, infertility factors, No. of high-quality embryos transferred, cycle type, and ovarian stimulation protocol ($P < 0.05$). However, no significant differences were observed in BMI, type of infertility, No. of embryos transferred, or No. of quality embryos transferred among the groups ($P > 0.05$).

Comparison of clinical outcomes among the four groups

The pregnancy outcomes of the four groups are presented in Table 2. The implantation rate, clinical pregnancy rate, and live birth rate exhibited significant decreases with an increase in the number of previous embryo implantation failures ($P < 0.001$). Conversely, the rate of early spontaneous abortion increased significantly ($P < 0.001$).

Univariate analysis for clinical outcomes

The univariate logistic regression analysis revealed that the number of previous embryo implantation failures, female age, AFC, endometrial thickness, total number of eggs obtained, cycle type, No. of high-quality embryos transferred, and stage of embryo development had an impact on the implantation rate, clinical pregnancy rate, early spontaneous abortion rate, and live birth rate ($P < 0.05$). The factors affecting implantation, clinical

TABLE 1 Comparison of demographic and clinical characteristics of the four groups of patients.

| Item | group A (n = 13172) | group B (n = 2989) | group C (n = 658) | group D (n = 156) | P-value |
|--|------------------------|--------------------------------|--------------------------------|---------------------------------|---------|
| Age(y) | 30.25 ± 4.15 | 31.09 ± 4.29 ^a | 31.84 ± 4.18 ^{ab} | 33.41 ± 4.10 ^{abc} | <0.001 |
| BMI (kg/m ²) | 23.32 ± 4.69 | 23.45 ± 3.84 | 23.22 ± 3.54 | 23.37 ± 5.29 | 0.591 |
| Duration of infertility((y)) | 3.95 ± 2.77 | 2.88 ± 3.11 ^a | 3.04 ± 3.04 ^a | 3.47 ± 3.09 ^{ab} | <0.001 |
| AMH(ng/mL) | 4.48 ± 3.76 | 3.20 ± 1.77 ^a | 3.04 ± 1.90 ^a | 2.96 ± 1.92 ^a | <0.001 |
| AFC | 12.85 ± 6.24 | 8.52 ± 7.85 ^a | 8.68 ± 7.37 ^a | 7.92 ± 6.31 ^a | <0.001 |
| Endometrial thickness(mm) | 10.25 ± 2.64 | 9.83 ± 2.38 ^a | 9.69 ± 1.92 ^a | 9.59 ± 2.09 ^a | <0.001 |
| No. of Oocytes retrieved | 8.58 ± 3.89 | 8.22 ± 3.49 | 4.58 ± 5.11 ^{ab} | 5.61 ± 4.94 ^{ab} | <0.001 |
| Types of Infertility | | | | | 0.078 |
| Primary infertility | 4750(53.59%) | 697 (51.63%) | 109 (45.99%) | 25 (53.19%) | |
| Secondary Infertility | 4114(46.41%) | 653 (48.37%) | 128 (54.01%) | 22 (46.81%) | |
| Cause of infertility | | | | | 0.014 |
| PCOS | 968 (12.02%) | 197 (10.35%) | 29 (11.37%) | 3 (4.84%) | 0.117 |
| Tubal factors | 4715 (58.54%) | 1140 (61.37%) | 155 (60.78%) | 38 (61.29%) | 0.193 |
| Ovulatory dysfunction | 909 (11.29%) | 212 (11.37%) | 27 (10.59%) | 7 (11.29%) | 0.987 |
| Endometriosis | 255 (3.17%) | 51 (2.74%) | 7 (2.75%) | 1 (1.61%) | 0.686 |
| Male factors | 785(9.75%) | 191(10.25%) | 32 (12.54%) | 10 (16.12%) | 0.160 |
| Other factors | 375 (4.66%) | 63 (3.38%) | 3 (1.19%) | 1 (1.61%) | 0.001 |
| Unexplained infertility | 47 (0.57%) | 10 (0.54%) | 2 (0.78%) | 2(3.24%) | 0.282 |
| Ovarian stimulation protocol | | | | | 0.004 |
| long protocol | 7756 (60.138%) | 1776 (60.801%) | 390 (60.278%) | 87 (57.237%) | 0.877 |
| antagonist protocol | 1642 (12.732%) | 340 (11.640%) | 92 (14.219%) | 14 (9.211%) | 0.121 |
| mild ovarian stimulation protocol | 759 (5.885%) | 158 (5.409%) | 32 (4.946%) | 2 (1.316%) | 0.063 |
| PPOS | 438 (3.396%) | 91 (3.115%) | 26 (4.019%) | 12 (7.895%) | 0.012 |
| Other protocol | 2302 (17.849%) | 556 (19.035%) | 107 (16.538%) | 37 (24.342%) | 0.061 |
| stage of embryo development | | | | | <0.001 |
| Cleavage embryo | 10267(77.97%) | 1657 (55.49%) | 308 (47.17%) | 82 (52.90%) | |
| Blastocyst | 2901(22.03%) | 1329 (44.51%) | 345 (52.83%) | 73 (47.10%) | |
| No. of embryos transferred | | | | | 0.817 |
| 1 | 3417 (25.94%) | 767 (25.69%) | 175 (26.80%) | 36 (23.23%) | |
| 2 | 9754 (74.06%) | 2219 (74.31%) | 478 (73.20%) | 119 (76.77%) | |
| No. of high-quality embryos transferred | 1.68 ± 0.51 | 1.54 ± 0.50 | 1.51 ± 0.49 | 1.59 ± 0.50 | 0.213 |
| No. of high-quality embryos transferred(%) | 66.98%(15334/22921) | 59.11%(3077/5205) ^a | 57.17% (646/1131) ^a | 47.31% (130/276) ^{abc} | p<0.001 |
| Type of cycle | | | | | p<0.001 |
| Fresh cycle | 4267(50.5%) | 417(20.5%) ^a | 91(17.9%) ^a | 30 (21.3%) ^a | |
| Frozen cycle | 4184(49.5%) | 1616(79.5%) ^a | 416 (82.1%) ^a | 111(78.7%) ^a | |

overall comparison between the four groups *P < 0.05; a: compared with group A, *P < 0.05; b: compared with group B, *P < 0.05; c: compared with group C, *P < 0.05.

TABLE 2 Comparison of clinical pregnancy outcomes among the four groups of patients.

| Item | group A | group B | group C | group D | P -value |
|--------------------------------------|-------------------------|------------------------------------|-----------------------------------|-------------------------------------|----------|
| Implantation rate (n%) | 11457/22928 (49.97%) | 2085/5211 (40.01%) ^a | 436/1132 (38.52%) ^a | 72/276 (26.09%) ^{a,b,c} | <0.001 |
| Pregnancy rate (n%) | 8491/13172 (64.46%) | 1561/2989 (52.22%) ^a | 336/658 (51.06%) ^a | 57/157 (36.54%) ^{a,b,c} | <0.001 |
| Early spontaneous abortion rate (n%) | 1152/8491 (13.57%) | 265/1561 (16.97%) ^a | 74/336 (22.02%) ^{a,b} | 16/57 (28.07%) ^{a,b,c} | <0.001 |
| Live birth rate (n%) | 7261/13172 (54.78%) | 1275/2989 (42.66%) ^a | 258/658 (39.21%) ^a | 39/156 (25.00%) ^{a,b,c} | <0.001 |

overall comparison between the four groups *P < 0.05; a: compared with group A, *P < 0.05; b: compared with group B, *P < 0.05; c: compared with group C, *P < 0.05.

pregnancy, and live birth rates were duration of infertility, AMH levels, and AFC ($P < 0.05$). (Table 3)

decrease compared to group A, while early spontaneous abortion rates showed a significant increase. (Table 4)

Multiple logistic regression analysis

Multiple logistic regression was performed to analyze the impact of the number of previous embryo implantation failures on the implantation, clinical pregnancy, early spontaneous abortion, and live birth rates after IVF/ICSI-ET, while adjusting for confounding factors such as age, duration of infertility, AMH levels, AFC, endometrial thickness, total number of oocytes retrieved, infertility factors, embryonic developmental stage, and cycle type. The implantation rate, clinical pregnancy rate, and live birth rate of patients in groups B, C, and D exhibited a gradual

Discussion

The impact of the number of previous implantation failures on pregnancy outcomes remains a controversial topic that requires further investigation. Cimadomo Danilo et al (12) reported in an observational study that the embryo implantation rate and early spontaneous abortion rate in the first four cycles of IVF/ICSI-ET treatment could not be predicted based on the number of previous embryo implantation failures. However, they found that the live birth rate decreased significantly in patients with a history of ≥ 3 implantation failures compared to those who underwent their first

TABLE 3 Univariate analysis of clinical outcomes affecting the number of previous embryo implantation failures.

| Item | Implantation rate | | Pregnancy rate | | Early spontaneous abortion rate | | Live birth rate | |
|-----------------------------|-------------------|----------|-------------------|----------|---------------------------------|----------|------------------|----------|
| | OR (95% CI) | P -value | OR (95% CI) | P -value | OR (95% CI) | P -value | OR (95% CI) | P -value |
| Duration of infertility (y) | 0.98 (0.97,1.00) | 0.009* | 0.98 (0.97, 1.00) | 0.008* | 1.02 (1.00,1.04) | 0.120 | 0.98 (0.97,0.99) | 0.002* |
| AMH (ng/mL) | 1.05 (1.03, 1.06) | <0.000* | 1.05 (1.03, 1.06) | <0.000* | 0.99 (0.97,1.01) | 0.301 | 1.04 (1.03,1.05) | <0.000* |
| AFC | 1.03 (1.03, 1.04) | <0.000* | 1.03 (1.03, 1.04) | <0.000* | 0.99 (0.98,1.00) | 0.045 | 1.03 (1.02,1.03) | <0.000* |
| Endometrial thickness (mm) | 1.07 (1.05, 1.08) | <0.000* | 1.07 (1.05, 1.08) | <0.000* | 0.94 (0.92,0.97) | <0.000* | 1.07 (1.06,1.09) | <0.000* |
| No. of Oocytes retrieved | 1.04 (1.03, 1.05) | <0.000* | 1.04 (1.03, 1.05) | <0.000* | 0.95 (0.94,0.97) | <0.000* | 1.05 (1.04,1.06) | <0.000* |
| Cause of infertility (n%) | | | | | | | | |
| PCOS | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| Tubal factors | 0.79 (0.69, 0.90) | 0.000* | 0.79 (0.69, 0.90) | 0.001* | 0.77 (0.63,0.95) | 0.013* | 0.91 (0.80,1.03) | 0.139 |
| Ovulatory dysfunction | 0.77 (0.65, 0.91) | 0.002* | 0.78 (0.66, 0.93) | 0.005* | 0.78 (0.59,1.04) | 0.088 | 0.88 (0.75,1.04) | 0.128 |

(Continued)

TABLE 3 Continued

| Item | Implantation rate | | Pregnancy rate | | Early spontaneous abortion rate | | Live birth rate | |
|---|-------------------|-----------------|-------------------|-----------------|---------------------------------|-----------------|------------------|-----------------|
| | OR (95% CI) | <i>P</i> -value | OR (95% CI) | <i>P</i> -value | OR (95% CI) | <i>P</i> -value | OR (95% CI) | <i>P</i> -value |
| Endometriosis | 0.73 (0.57, 0.95) | 0.019* | 0.72 (0.56, 0.94) | 0.015* | 0.80 (0.51,1.25) | 0.333 | 0.86 (0.67,1.10) | 0.228 |
| Male factors | 1.25 (1.01, 1.55) | 0.043* | 1.23 (0.99, 1.53) | 0.056 | 0.59 (0.42,0.84) | 0.003* | 1.44 (1.18,1.76) | 0.001* |
| Other factors | 0.98 (0.78, 1.24) | 0.882 | 1.02 (0.81, 1.30) | 0.845 | 0.55 (0.36,0.84) | 0.006 | 1.19 (0.95,1.49) | 0.122* |
| Unexplained infertility | 1.04 (0.59, 1.84) | 0.882 | 1.03 (0.59, 1.82) | 0.914 | 0.86 (0.35,2.07) | 0.730 | 1.06 (0.62,1.79) | 0.836 |
| Ovarian stimulation protocol | | | | | | | | |
| long protocol | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| antagonist protocol | 1.01 (0.92, 1.11) | 0.826 | 1.01 (0.92, 1.11) | 0.870 | 0.95 (0.80,1.13) | 0.586 | 0.64 (2.92,7.38) | <0.000* |
| mild ovarian stimulation protocol | 1.07 (0.93,1.22) | 0.358 | 1.07 (0.93, 1.22) | 0.357 | 0.83 (0.64,1.06) | 0.138 | 0.58 (1.96,6.57) | <0.000* |
| PPOS | 1.07 (0.89, 1.27) | 0.458 | 1.08 (0.91, 1.29) | 0.370 | 1.13 (0.89,1.57) | 0.247 | 0.67 (1.92,1.37) | 0.001* |
| Other protocol | 1.02 (0.94, 1.11) | 0.708 | 1.03 (0.95, 1.12) | 0.529 | 0.86 (0.74,1.00) | 0.053 | 0.48 (0.32,0.44) | <0.000* |
| Type of cycle (n%) | | | | | | | | |
| Fresh cycle | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| Frozen cycle | 0.81 (0.75, 0.88) | <0.000* | 0.81 (0.75, 0.88) | <0.000* | 1.43 (1.24,1.64) | <0.000* | 0.77 (0.71,0.83) | <0.000* |
| No. of high-quality embryos transferred (%) | 1.24 (1.41,1.52) | <0.000* | 1.23 (1.31,1.65) | <0.000* | 0.95 (1.31,1.52) | <0.000* | 1.18 (1.15,1.57) | <0.000* |
| Stage of embryo development | | | | | | | | |
| Cleavage embryo | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| blastocyst | 1.25 (1.16,1.34) | <0.000* | 1.23 (1.15, 1.32) | <0.000* | 1.27 (1.13,1.43) | <0.000* | 1.11 (1.04,1.19) | 0.002* |

OR: ratio; CI is confidence interval, **P*<0.05.

IVF/ICSI cycle. A retrospective cohort study in Israel reported that the live birth rate per IVF/ICSI-ET cycle decreased significantly with the increasing number of embryo transfer cycles. The study found that the live birth rate was almost zero in patients who underwent the fourth embryo transfer cycle, suggesting that infertile patients who did not achieve a clinical pregnancy with four consecutive embryo transfer cycles should discontinue treatment or consider alternative options such as donor eggs and sperm (7). Conversely, a large multicenter study by Andrew D. A. C. Smith et al (8) showed that the success rate of IVF/ICSI-ET tended to decrease with the number of embryo transfer cycles until the ninth cycle, after which live birth was almost impossible. They support the notion that, although the live birth rate decreased after 3–4 cycles of repeated treatment, there was still a possibility of live birth. Wang et al. (10) found no significant correlation between

clinical pregnancy rate and the number of the first three transfer cycles, but observed significantly lower implantation and clinical pregnancy rates from the fourth embryo transfer cycle. The inconsistencies among these studies may be due to differences in study population characteristics, inclusion and exclusion criteria, quality of transferred embryos, level of assisted reproductive technology in each country, and sample size. Our findings demonstrate that the number of previous embryo implantation failures is an independent factor affecting implantation rate, clinical pregnancy rate, spontaneous abortion rate and live birth rate of patients underwent IVF/ICSI-ET. With the increase of the number of previous embryo implantation failures, the implantation rate, clinical pregnancy rate and live birth rate of patients underwent IVF/ICSI-ET decreased significantly, and the rate of early spontaneous abortion increased markedly.

TABLE 4 Multifactorial logistic regression analysis of the number of previous embryo implantation failures on pregnancy outcome of IVF/ICSI-ET patients.

| Item | Unadjusted | | Adjusted | |
|---------------------------------|-------------------|----------|-------------------|----------|
| | OR (95% CI) | P -value | OR (95% CI) | P- value |
| Implantation rate | | | | |
| group A | 1 | 1 | 1 | 1 |
| group B | 0.61 (0.56, 0.66) | <0.001* | 0.57 (0.51, 0.63) | <0.001* |
| group C | 0.58 (0.50, 0.68) | <0.001* | 0.53 (0.44, 0.64) | <0.001* |
| group D | 0.32 (0.23, 0.45) | <0.001* | 0.33 (0.23, 0.46) | <0.001* |
| Pregnancy rate | | | | |
| group A | 1 | 1 | 1 | 1 |
| group B | 0.60 (0.56, 0.65) | <0.001* | 0.57 (0.51, 0.63) | <0.001* |
| group C | 0.58 (0.49, 0.67) | <0.001* | 0.53 (0.44, 0.63) | <0.001* |
| group D | 0.32 (0.23, 0.44) | <0.0001* | 0.32 (0.23, 0.46) | <0.0001* |
| Early spontaneous abortion rate | | | | |
| group A | 1 | 1 | 1 | 1 |
| group B | 1.27 (1.10, 1.46) | 0.009* | 1.40 (1.16, 1.70) | < 0.001* |
| group C | 1.71 (1.32, 2.22) | <0.001* | 1.84 (1.36, 2.48) | <0.001* |
| group D | 2.61 (1.49, 4.58) | 0.008* | 2.24 (1.24, 4.02) | 0.073* |
| Live birth rate | | | | |
| group A | 1 | 1 | 1 | 1 |
| group B | 0.61 (0.57, 0.67) | <0.001* | 0.56 (0.50, 0.63) | <0.001* |
| group C | 0.53 (0.45, 0.62) | <0.001* | 0.49 (0.41, 0.58) | <0.001* |
| group D | 0.28 (0.19, 0.40) | <0.001* | 0.29 (0.20, 0.42) | <0.001* |

Group A was used as the reference group. *P < 0.05 after adjusting for confounding factors, including age, duration of infertility, AMH levels, AFC, endometrial thickness, the total number of eggs obtained, infertility factors, embryo development stage, cycle type, ovarian stimulation regimen, and the number of good-quality embryos transferred.

Effect of the number of previous embryo implantation failures on implantation, clinical pregnancy, and live birth rates in IVF/ICSI-ET patients

Embryo implantation is a crucial step in the success of IVF/ICSI-ET treatment. However, recurrent embryo implantation failure (RIF) is a challenging condition that hinders the improvement of clinical pregnancy rates in these patients. Diagnostic and therapeutic challenges arise due to the diverse etiologies of RIF. Improving embryo implantation rates has become a major challenge in improving the clinical outcomes of IVF/ICSI-ET patients. Therefore, further research on RIF patients is necessary. The definition of RIF lacks a unified international standard. Currently, most experts accept the criteria based on the patient's age, the number of failed IVF/ICSI cycles, and the number of good-quality embryos transferred (13). In China, the 2023 expert consensus defines RIF as the failure to achieve clinical pregnancy after transferring at least three good-quality embryos in three fresh or frozen cycles (14). The etiology of embryo implantation failure is complex, diverse, and partially unknown (15). Some studies have

reported that the number of previous embryo implantation failures is an independent risk factor affecting implantation rates (5, 8, 9, 12). However, other studies have suggested that patient outcomes in IVF/ICSI cycles are not significantly related to the number of embryo transfer cycles (16). In our study, we found that patients without any implantation failures had significantly higher implantation rates, clinical pregnancy rates, and live birth rates, and a lower rate of early spontaneous abortion compared to those in the other three groups. The pregnancy outcomes were comparable between one and two previous embryo implantation failure with no significant differences in implantation rate, clinical pregnancy rate, and live birth rate (see attachment for details). However, the outcomes of patients with ≥3 previous implantation failures were worse than those of the other three groups of patients.

It is well known that age is the most critical factor affecting the development of oocytes and the quality of embryos. As female age increases, the decline in ovarian function and the increased probability of aneuploidy, she is prone to embryo developmental delay and stagnation, which can lead to embryo implantation failure, and the implantation rate decreases gradually (17, 18). We selected patients less than 40 years old and adjusted age by multiple logistic

regression to avoid the impact of age on pregnancy outcomes. Impaired endometrial receptivity is also an important factor that causes embryo implantation failure (19). Therefore, patients underwent hysteroscopy after the first cycle of implantation failure to exclude interference from endometrial polyps, uterine adhesions, and chronic endometritis on implantation rates (20).

The success of ART largely depends on the quality of the embryo and the receptivity of the endometrium. In this study, the implantation rate, the clinical pregnancy rate and the live birth rate of patients underwent first embryo transfer cycle were 49.97%, 64.46% and 54.78% respectively, which was a higher rate of successful pregnancy compared to those who had previous failed cycles, which is similar to Shapiro B S study (21). Patients with one or two previous embryo implantation failures exhibited comparable pregnancy outcomes, while patients with three or more previous failures showed significantly lower rates of embryo implantation, clinical pregnancy, and live birth. Patients with three or more previous failures was RIF patients, whose infertility is often caused by multiple factors, including maternal endocrine and immune disorders and thrombophilia (22). Our cohort did not undergo etiology-related investigations and treatments for RIF patient. However, we have since put in place etiologic screening and appropriate treatment for patients with four or more embryo transfer cycles. Our findings highlight the need for further research into RIF patients, as the etiology of this condition is complex and not well understood. Therefore, patients with ≥ 3 previous implantation failures are recommended to continue subsequent cycles only after examination and treatment to improve pregnancy outcomes.

Effect of the number of previous embryo implantation failures on early spontaneous abortion rate in IVF/ICSI-ET assisted conception

Spontaneous abortion is a common complication of pregnancy in obstetrics and gynecology. It is known that spontaneous abortion after IVF/ICSI-ET-assisted clinical pregnancy reduces the live birth rate. However, it is uncertain how the number of previous embryo implantation failures affects the rate of spontaneous abortion after assisted clinical pregnancy (10, 23). Previous studies have yielded conflicting results regarding the impact of the number of embryo transfer cycles on the rate of spontaneous abortion after clinical pregnancy in IVF/ICSI-assisted conception. Some studies have shown no significant change in the spontaneous abortion rate as the number of cycles increased, while others have demonstrated an increased risk of spontaneous abortion in patients with multiple embryo transfer cycles, which partially supports the results of our study (24, 25). In our study, we found that the early spontaneous abortion rate was lowest in the first embryo transfer group. Patients with ≥ 3 embryo implantation failures had a significantly higher early spontaneous abortion rate of 28.07%, which may be due to the fact that these patients are RIF population, and the etiology of RIF

and recurrent spontaneous abortion (RSA) is similar (26). The etiology of RSA involves chromosomal or genetic abnormalities, anatomical abnormalities, autoimmune diseases, prethrombotic state (PTS), endocrine factors, infectious factors, male factors, and psychological factors (27). However, the etiological examination and correction were not carried out in our study for patients with ≥ 3 embryo implantation failures, which may explain the high risk of spontaneous abortion in these patients.

However, there are some limitations in this study. First, our study was a single-center, retrospective study and there might be some confounders that we did not control for. In addition, we prioritized some pregnancy outcomes and did not investigate neonatal outcomes. In the future, our findings need to be validated by expanding the sample size or by high-quality randomized clinical trial studies.

In conclusion, the number of previous embryo implantation failures is an independent factor affecting implantation rate, clinical pregnancy rate, spontaneous abortion rate and live birth rate of patients underwent IVF/ICSI-ET. Based on our study, patients with ≥ 3 previous implantation failures are recommended to undergo etiology-related investigations and treatments to continue subsequent cycles to improve pregnancy outcomes. Investigations for RIF includes: 1. General risk factors such as old age, poor lifestyle, smoking, etc. 2. Immune factors include autoimmunity and alloimmunity; 3. Prethrombotic states include hereditary and acquired thrombophilia; 4 Endometrial receptivity test 5. Factors of infection; 6 Reproductive anatomy; 7. Endocrine factors; 8. Male factor; 9. Chromosomes. Treatments of RIF: 1. General treatment such as weight control, healthy diet and appropriate exercise; 2.IVF-ET program optimization; 3. Regulation of immune disorders (glucocorticoids, low molecular weight heparin, immunoglobulin, etc.); 4. Low molecular weight heparin for treatment of pre-thrombotic state; 5. Doxycycline and metronidazole for chronic endometritis; 6. Treatment of submucosal myoma, polyps and hydrosalpinx and other normal anatomical structure abnormalities; 7. The man controls his weight, reduces smoking, etc. 8. Encourage both couples or those with chromosome abnormalities to perform PGT treatment.

Data availability statement

The original contributions presented in the study are included in the article/supplementary material. Further inquiries can be directed to the corresponding author.

Ethics statement

The studies involving human participants were reviewed and approved by Henan Medical Ethics Committee of Provincial People's Hospital. The patients/participants provided their written informed consent to participate in this study.

Author contributions

YF and HQ conception and design, review and final approval of the version to be published. FJ, CT, XH analyses the data. YF and HQ draft and revise the article. YF and FJ collect and analyze the data. All authors contributed to the article and approved the submitted version.

Funding

This study was supported by the Science and Technology Project of Henan Province (182102310134).

References

- Inhorn MC, Patrizio P. Infertility around the globe: new thinking on gender, reproductive technologies, and global movements in the 21st century0]. *Hum Reprod Update* (2015) 21(4):411–26. doi: 10.1093/humupd/dmv016
- Mascarenhas MN, Flaxman SR, Boerma T, Vanderpoel S, Stevens GA, et al. National, regional, and global trends in infertility prevalence since 1990: a systematic analysis of 277 health surveys. *PloS Med* (2012) 9(12):e1001356. doi: 10.1371/journal.pmed.1001356
- Sun H, Gong TT, Jiang YT, et al. *Global, regional, and national prevalence and disability-adjusted life-years for infertility in 195 countries and territories, 1990–2017: results from a global burden of disease study, 2017*. Aging (Albany NY) (2019) 11(23):10952–91. doi: 10.18632/aging.102497
- Mao X, Zhang J, Chen Q, Kuang Y, Zhang S, et al. Short-term copper intrauterine device placement improves the implantation and pregnancy rates in women with repeated implantation failure. *Fertil Steril* (2017) 108(1):55–61 e1. doi: 10.1016/j.fertnstert.2017.05.014
- Cimadomo D, Craciunas L, Vermeulen N, Vomstein K, Toth B. Definition, diagnostic and therapeutic options in recurrent implantation failure: an international survey of clinicians and embryologists. *Hum Reprod* (2021) 36(2):305–17. doi: 10.1093/humrep/deaa317
- Wang Y, Tian Y, Liu L, Li TC, Tong X, Zhu H, et al. The number of previous failed embryo transfer cycles is an independent factor affecting implantation rate in women undergoing IVF/ICSI treatment: A retrospective cohort study. *Med(Baltim)* (2021) 100(9):e25034. doi: 10.1097/MD.00000000000025034
- Simonstein F, Mashiach-Eizenberg M, Revel A, Younis JS, et al. Assisted reproduction policies in Israel: a retrospective analysis of in vitro fertilization-embryo transfer. *Fertil Steril* (2014) 102(5):1301–6. doi: 10.1016/j.fertnstert.2014.07.740
- Smith ADAC, Tilling K, Nelson SM, Lawlor DA, et al. Live-birth rate associated with repeat in vitro fertilization treatment cycles. *J Am Med Assoc* (2015) 314(24):2654–62. doi: 10.1001/jama.2015.17296
- Xu XL, Xu YY, Chen CH, Li J, Lu Y, Zh LH, et al. Impact of miscarriage in the first complete cycle of human assisted reproductive technology pregnancy on the outcome of subsequent cycles of assisted conception. *J Zhengzhou Univ (Medical Edition)* (2022) 057–001:78–82. doi: 10.13705/j.issn.1671–6825.2021.09.053
- Wang F, Sun H-X, Wang J-X, Zhang NY, Hu YL, et al. Pregnancy outcomes in repeated cycles of in vitro fertilization-embryo transfer. *Chin J Male Sci* (2010) 16(11):1007–11. doi: 10.13263/j.cnki.njia.2010.11.02
- Homburg R, Meltzer S, Robinson J, Scharf S, Anteby EY, Orvieto R, et al. Is there a limit for the number of in vitro fertilization cycles for an individual patient? *Fertil Steril* (2009) 91(4 Suppl):1329–31. doi: 10.1016/j.fertnstert.2008.03.010
- Daniilo C, Antonio C, Lisa D, Tacconi L, Soscia D, Giancani A, et al. Leave the past behind: women's reproductive history shows no association with blastocysts' euploidy and limited association with live birth rates after euploid embryo transfers. *Hum Reprod* 4(4):929–40. doi: 10.1093/humrep/deab014
- Coughlan C, Ledger W, Wang Q, Liu F, Demirolo A, Gurgan T, et al. Recurrent implantation failure: definition and management. *Reprod BioMed Online* (2014) 28(1):14–38. doi: 10.1016/j.rbmo.2013.08.011
- Chinese Physicians Association of Reproductive Medicine and Chinese Women Physicians Association of Reproductive Medicine. Chinese expert consensus on the

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

clinical management of recurrent implantation failure. *Chin Med J* (2023) 103(2):89–100. doi: 10.3760/cma.j.cn112137

15. Wang XP. New perspectives on the etiology and treatment of recurrent miscarriage and recurrent implantation failure. *Chin J Obstetrics Gynecology* (2019) 54(12):4. doi: 10.3760/cma.j.issn.0529–567x.2019.12.001

16. Silberstein T, Trimarchi JR, Gonzalez L, Keefe DL, Blazar AS, et al. Pregnancy outcome in in vitro fertilization decreases to a plateau with repeated cycles. *Fertil Sterility* (2005) 84(4):1043–5. doi: 10.1016/j.fertnstert.2005.04.026

17. Hatirnaz S, Ozer A, Hatirnaz E, Atasever M, Başaranoglu S, Kanat-Pektas M, et al. Preimplantation genetic screening among women experiencing recurrent failure of in vitro fertilization. *Int J GYNECOL OBSTET* (2017) 137(3):314–8. doi: 10.1002/ijgo.12135

18. May-Panloup P, Boucret L, Chao de la Barca JM, Desquiere-Dumas V, Ferré-L'Hottelier V, Morinière C, et al. Ovarian ageing: the role of mitochondria in oocytes and follicles. *Hum Reprod Update* (2016) 22(6):725–43. doi: 10.1093/humupd/dmw028

19. Bashiri A, Halper KI, Orvieto R. Recurrent Implantation Failure-update overview on etiology, diagnosis, treatment and future directions. *Reprod Biol Endocrinol* (2018) 16(1):121. doi: 10.1186/s12958-018-0414-2

20. Kuroda K, Horikawa T, Moriyama A, Nakao K, Juen H, Takamizawa S, et al. Impact of chronic endometritis on endometrial receptivity analysis results and pregnancy outcomes. *Immunity Inflammation Dis* (2020) 8(4):650–8. doi: 10.1002/iid3.354

21. Shapiro BS, Richter KS, Harris DC, Daneshmand ST, et al. Dramatic declines in implantation and pregnancy rates in patients who undergo repeated cycles of in vitro fertilization with blastocyst transfer after one or more failed attempts. *Fertil Steril* (2001) 76(3):538–42. doi: 10.1016/s0015-0282(01)01979-3

22. Franasik JM, Alecsandru D, Forman EJ, Gemmell LC, Goldberg JM, Llarrea N, et al. A review of the pathophysiology of recurrent implantation failure. *Fertil Steril* (2021) 116(6):1436–48. doi: 10.1016/j.fertnstert.2021.09.014

23. Zhang HZ, Xiong F, Li GG, Sun Q, Chen PL, Wan CY, et al. Clinical outcomes after repeated vitrification of frozen-thawed embryos for transfer. *J Reprod Med* (2018) 27(7):5. doi: 10.3969/j.issn.1004-3845

24. Pirtea P, De Ziegler D, Tao X, Zhan Y, Ayoubi JM, Seli E, et al. Rate of true recurrent implantation failure is low: results of three successive frozen euploid single embryo transfers. *Fertil Steril* (2021) 115(1):45–53. doi: 10.1016/j.fertnstert.2020.07.002

25. Yang R, Yang S, Li R, Chen X, Wang H, Ma C, et al. Biochemical pregnancy and spontaneous abortion in first IVF cycles are negative predictors for subsequent cycles: an over 10,000 cases cohort study. *Arch Gynecol Obstet* (2015) 292(2):453–8. doi: 10.1007/s00404-015-3639-8

26. Polanski LT, Baumgarten MN, Quenby S, Brosens J, Campbell BK, Raine-Fenning NJ, et al. What exactly do we mean by 'recurrent implantation failure'? A systematic review and opinion. *Reprod BioMed Online* (2014) 28(4):409–23. doi: 10.1016/j.rbmo.2013.12.006

27. Obstetrics and Gynecology Section of the Chinese Medical Association and Expert Consensus Group on the Diagnosis and Treatment of Recurrent Miscarriage. Expert consensus on the diagnosis and treatment of recurrent miscarriage (2022). *Chin J Obstetrics Gynecology* (2022) 57(9):653–67. doi: 10.3760/cma.j.cn112141-20220421-00259



OPEN ACCESS

EDITED BY

Richard Ivell,
University of Nottingham, United Kingdom

REVIEWED BY

Mai Shaker,
National Research Centre, Egypt
Maria Emilia Solano,
University Medical Center Regensburg,
Germany

*CORRESPONDENCE

Natalin Jimena Valeff
✉ natalivaleff@gmail.com
Maria Silvia Ventimiglia
✉ masivent@gmail.com
Federico Jensen
✉ fjensen@unaj.edu.ar

[†]These authors have contributed equally to this work

[‡]These authors have contributed equally to this work and share senior authorship

RECEIVED 02 June 2023

ACCEPTED 08 September 2023

PUBLISHED 03 October 2023

CITATION

Valeff NJ, Ventimiglia MS, Diao L and Jensen F (2023) Lupus and recurrent pregnancy loss: the role of female sex hormones and B cells.
Front. Endocrinol. 14:1233883.
doi: 10.3389/fendo.2023.1233883

COPYRIGHT

© 2023 Valeff, Ventimiglia, Diao and Jensen.
This is an open-access article distributed under the terms of the [Creative Commons Attribution License \(CC BY\)](#). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Lupus and recurrent pregnancy loss: the role of female sex hormones and B cells

Natalin Jimena Valeff^{1*†}, Maria Silvia Ventimiglia^{1*†},
Lianghui Diao^{2‡} and Federico Jensen^{1,3**}

¹Center for Pharmacological and Botanical Studies (CEFYBO-UBA-CONICET), Medical Faculty, Buenos Aires University, Buenos Aires, Argentina, ²Shenzhen Key Laboratory of Reproductive Immunology for Peri-implantation, Shenzhen Zhongshan Institute for Reproduction and Genetics, Fertility Center, Shenzhen Zhongshan Urology Hospital, Shenzhen, China, ³Centro Integrativo de Biología Y Química Aplicada. Universidad Bernardo O'Higgins, Santiago, Chile

Systemic lupus erythematosus is a debilitating autoimmune disease characterized by uncontrolled activation of adaptive immunity, particularly B cells, which predominantly affects women in a 9 to 1 ratio compared to men. This stark sex disparity strongly suggests a role for female sex hormones in the disease's onset and progression. Indeed, it is widely recognized that estradiol not only enhances the survival of autoreactive B cells but also stimulates the production of autoantibodies associated with systemic lupus erythematosus, such as anti-nuclear antibodies and anti-dsDNA antibodies. Clinical manifestations of systemic lupus erythematosus typically emerge after puberty and persist throughout reproductive life. Furthermore, symptoms often exacerbate during the premenstrual period and pregnancy, as increased levels of estradiol can contribute to disease flares. Despite being fertile, women with lupus face a heightened risk of pregnancy-related complications, including pregnancy loss and stillbirth, which significantly surpass the rates observed in the healthy population. Therefore, this review aims to summarize and discuss the existing literature on the influence of female sex hormones on B-cell activation in patients with systemic lupus erythematosus, with a particular emphasis on their impact on pregnancy loss.

KEYWORDS

recurrent pregnancy loss, lupus, B cells, hormones, pregnancy

1 Introduction

Recurrent pregnancy loss (RPL) is a distressing pregnancy disorder experienced by ~2.5% of women trying to conceive. It is defined as the spontaneous demise of two or more clinically recognized pregnancies before the fetus reaches viability; RPL includes embryonic and fetal losses from the time of conception until 24 weeks of gestation (1, 2).

Autoimmune disorders have been included along with chromosomal errors, anatomical uterine defects, and endometrial dysfunction as the most common etiologies

linked to RPL (3). Indeed, certain features commonly associated with autoimmune diseases, such as inappropriate complement activation (4–6) and the prevalence of specific autoantibodies (4, 7–11) show strong associations with RPL.

Furthermore, systemic autoimmune diseases, including systemic lupus erythematosus (SLE), have been identified as significant risk factors for RPL, similar to other autoimmune conditions (12).

SLE is a chronic autoimmune disease that predominantly affects women of reproductive age compared to men and has the potential to affect any organ in the body (13–15). The intricate clinical presentation and pathogenesis of SLE make its definition exceptionally challenging. According to the European League Against Rheumatism (EULAR) and the American College of Rheumatology (ACR), the classification criteria for SLE consist of a mandatory entry criterion of positive anti-nuclear antibodies (ANAs) at least once, followed by additive weighted criteria grouped into seven clinical domains, namely, constitutional, hematologic, neuropsychiatric, mucocutaneous, serosal, musculoskeletal, and renal, and three immunological domains: antiphospholipid antibodies (aPLs), complement proteins, and SLE-specific antibodies (16). ANAs are a group of autoantibodies that target components of the cell nucleus and can bind to proteins, nucleic acids, and protein–nucleic acid complexes (17).

From an immunological perspective, the intricate interplay of environmental, genetic, and hormonal factors results in dysregulation and abnormal activation of the innate and adaptive immune system. This leads to the generation of pathogenic autoantibodies, such as ANAs, anti-double-stranded DNA antibodies (anti-dsDNA), and aPLs, as well as the deposition of immune complexes, ultimately causing tissue damage (18, 19).

Moreover, the impact of ANAs (20) and the presence of various types of aPLs (21) significantly varies between women with RPL and autoimmune diseases, in comparison to those without autoimmunity (22). Indeed, the rate of pregnancy loss among patients with SLE is substantially higher compared to the general healthy population (3). Furthermore, the stage of SLE that the patient is in at the moment of becoming pregnant, including disease activity and renal involvement, not only impacts the health status of the mother but may also influence fetal and neonatal outcomes (23, 24). In this regard, several studies have found that increased serum levels of IL-6, IL-10, and INF- α in patients with SLE are associated with disease activity (25, 26). Regarding disease activity at the time of conception, numerous prospective studies have recently shown that women with inactive SLE generally experience minimal flares during pregnancy, while those with active SLE face an elevated risk of adverse maternal and fetal outcomes (27–29). These findings are consistent with previous reports, indicating that the rate of live births is lower in patients with clinically active SLE in the 6 months before conception compared to those with inactive disease prior to conception (30). Furthermore, RPL among women with SLE appears to be linked to a higher rate of fetal death, which is associated with the presence of aPLs (31, 32). Furthermore, it is well established that newborn babies born to mothers with SLE can develop neonatal lupus, a rare condition that is not a form of SLE, but rather a condition that affects the newborn due to the transfer of maternal autoantibodies across the placenta during pregnancy (33).

Considering the sex and age predisposition of SLE, female sex hormones are undeniably involved in the pathogenesis of the disease (34). Studies conducted on SLE-prone mice using gonadectomy/hormone deprivation and hormone supplementation have consistently confirmed this association, revealing that estrogen exacerbates the disease, while its removal ameliorates the disease in female subjects [reviewed in (35)]. In the context of pregnancy, the increase in female sex hormone levels may influence or potentiate the abnormal function of the immune cells in patients with SLE, thereby exacerbating the disease symptoms and leading to pregnancy complications, including RPL (36, 37).

Considering all the evidence, the objective of this review is to examine the current state of knowledge regarding the impact of preexisting SLE on the development of RPL, with particular focus on the role of female sex hormones in B cell activation and autoantibody production.

2 Pregnancy in patients with systemic lupus erythematosus: the impact on recurrent pregnancy loss

As mentioned earlier, SLE predominantly affects women during their reproductive age, when individuals may seek to become pregnant. However, while fertility is generally preserved in women with SLE, pregnancy in these patients can be associated with adverse maternal and fetal outcomes, including RPL (38). In a recent meta-analysis of pregnancy studies published from 2017 to 2019, it was shown that patients with SLE had markedly increased risk of stillbirth (risk ratio (RR) 16.49, 95% CI 2.95 to 92.13; $p = 0.001$) and fetal loss (RR 7.55, 95% CI 4.75 to 11.99; $p = 0.00001$) compared to healthy pregnant women (39). Despite substantial declines in rates of pregnancy loss among patients with SLE in recent years, they remain higher compared to the healthy population (40). Indeed, approximately 20% of pregnancies in patients with SLE result in miscarriages (40).

Several biomarkers have been investigated as potential predictors of pregnancy complications in women with SLE. Notably, aPLs, including anticardiolipin antibodies and lupus anticoagulants, have been associated with obstetric complications such as RPL, recurrent implantation failure, pre-eclampsia, and preterm birth (41, 42). Additionally, research has shown that low levels of complement proteins, such as C3 and C4 during the first trimester are associated with an increased risk of pregnancy loss (43) in patients with SLE.

Although the causes behind poor pregnancy outcomes in patients with SLE are diverse, there is a general consensus that active disease, characterized by the activation of autoreactive B cells and production of autoantibodies, at the time of conception and during pregnancy significantly impacts maternal and fetal outcomes (38). This is not surprising, given that a successful pregnancy relies on a precisely regulated balance between maternal immune activation and immune tolerance (44). Any disruptions or imbalances in this delicate equilibrium can lead to pregnancy loss. Conversely, during pregnancy, an increase in the levels of female sex

hormones can promote B cell autoreactivity and exacerbate the symptoms of SLE, creating a negative feedback loop. This phenomenon leads to the activation of various immune mechanisms, which can not only worsen the symptoms of SLE but also contribute to pregnancy loss. Therefore, the hormonal regulation of B cell activation during SLE and its implication in pregnancy loss will be discussed in greater detail below.

3 The impact of female sex hormones on B cell activation in patients with systemic lupus erythematosus

B cells are essential components of the adaptive immune system responsible for antibody production. They can be classified into marginal zone (MZ), B1, and B2 B cells based on their phenotype, localization, and functionality (45). While T cell activation relies on antigen presentation by antigen-presenting cells (APCs), B cells, on the other hand, can directly interact with antigens through their receptor (B cell receptor, BCR) (46). However, apart from the signal provided by antigens through BCR, B cells require a second signal for proper activation, which can be delivered by toll-like receptors (TLRs), BAFF-R, or BCR cross-linking in the case of MZ and B1 B cells (47). On the other hand, upon antigen recognition, B2 B cells migrate to the germinal center, where they receive a second signal from follicular T-helper (Tfh) cells. Subsequently, they mature into either antibody-producing plasma cells or memory B cells.

Female sex hormones play a significant role in the development and activity of the immune system (48). Both innate and adaptive immune cells bear receptors for sex hormones and respond to hormonal cues (49). Women display higher frequencies of B cells (50) along with enhanced B cell survival, maturation, and class switching. They also demonstrate greater antibody responses and higher basal levels of immunoglobulins (Igs) compared to men (51), suggesting the involvement of female sex hormones in controlling diverse B cell functions. Indeed, estrogen has been shown to reduce the production of B cell precursors, impair B cell tolerance, and increase the activation and survival of autoreactive B cells (52, 53). While B cells express both estrogen receptor (ER) α and β , it is ER α that predominantly regulates BCR signal strength (54). Elevated levels of estrogen and ER α engagement result in reduced BCR signal strength and modulation of survival regulators such as Bcl-2, CD22, and SH2-containing protein tyrosine phosphatase (SHP)-2, thereby suppressing apoptosis (52). Moreover, elevated estrogen levels result in increased serum BAFF levels, which, together with reduced BCR signal strength, promote the survival of autoreactive B cells that would otherwise be eliminated from the naive repertoire. Consequently, these autoreactive B cells gain entry into the mature B cell pool (55, 56). In such circumstances, heightened estrogen stimulation on B cells triggers a breakdown of tolerance and uncontrolled proliferation and enhances the survival of high-affinity DNA-reactive B cells, which may potentially lead to autoimmunity (54).

A significant proportion of autoreactive B cells originates from the B2 B cell pool, which requires second signals provided by

follicular T helper cells to complete their activation. The significance of these pathways in promoting autoantibody production has been demonstrated in genetically modified lupus-prone mice and using blocking antibodies against various costimulatory molecules, such as inducible costimulatory ligand (ICOS-L) and CD40 ligand. Consequently, T helper cells play a crucial role in the development and progression of SLE disease (57). Furthermore, Tfh cells not only express estrogen receptors, but it has also been demonstrated that estradiol promotes the expansion of Tfh cells and, consequently, enhances the humoral immune response (58). Therefore, in the context of SLE, estradiol appears to exert its effects on the Tfh/B2 B cell axis, promoting the development and survival of autoreactive B cells.

The fact that 90% of patients with SLE are women clearly highlights a strong sex bias in this autoimmune disease. Several hypotheses have been proposed to explain this phenomenon, with the influence of female sex hormones being the most widely accepted (59). In this regard, it is known that the clinical manifestation of the disease typically appears after puberty, affecting women between the ages of 20 to 50, a period during which levels of estradiol and progesterone significantly rise (59). The strongest evidence supporting the role of female sex hormones in SLE comes from the observation that patients with SLE experience disease exacerbation during the premenstrual period and in pregnancy (35, 59). Interestingly, a case report demonstrated that administering cross-gender hormones to a transgender female resulted in lupus nephritis, and the withdrawal of estradiol supplementation upon admission prevented the worsening of symptoms. This provides further support for the role of estradiol in driving SLE (60). Animal studies also provide support for the role of estrogen in SLE. Ovariectomized lupus-prone mice showed ameliorated disease, while estrogen supplementation in castrated male mice worsened the symptoms [reviewed in (35)]. Moreover, targeted deletion of ER α specifically in B cells has been shown to reduce the production of pathogenic autoantibodies and the development of nephritis in lupus-prone mice (61). Additionally, tamoxifen treatment significantly reduced autoantibody production and improved the course of SLE in SLE-prone mice (62).

In pregnant SLE patients, estrogen levels and ER α expression not only mediate the increase in anti-dsDNA but also alter the B-cell repertoire, leading to the expansion of autoreactive clones (63, 64). As a result, hormone levels during pregnancy have a substantial impact on the function of autoreactive B cells, intensifying SLE symptoms and contributing to adverse pregnancy outcomes, including RPL (36, 37). In fact, E2 has been demonstrated to decrease B-cell lymphopoiesis in the bone marrow at the pro-B-cell stages in mice and to alter transitional 2 (T2) B cell maturation, both during pregnancy and in patients with SLE (53, 65). Under SLE conditions, elevated BAFF levels and reduced BCR signal strength can lead to the maturation of transitional B cells into a marginal zone (MZ) B cell expansion. Under specific conditions, marginal zone (MZ) B cells can serve as precursors of unswitched memory B cells without T cell help (66). It has been previously demonstrated that during pregnancy, there is a bias toward the development of marginal zone (MZ) B cells (67). This, along with the abnormal differentiation of unswitched memory B cells

observed in patients with SLE (68) may pose a risk to the successful development of pregnancy in patients with SLE. In fact, an increase in unswitched memory B cells is observed in patients with a history of RPL and obstetric complications (69, 70).

Therefore, the presence of autoreactive B cells, along with increased B cell activation and autoantibody production in patients with SLE, poses significant challenges when it comes to achieving a full-term pregnancy.

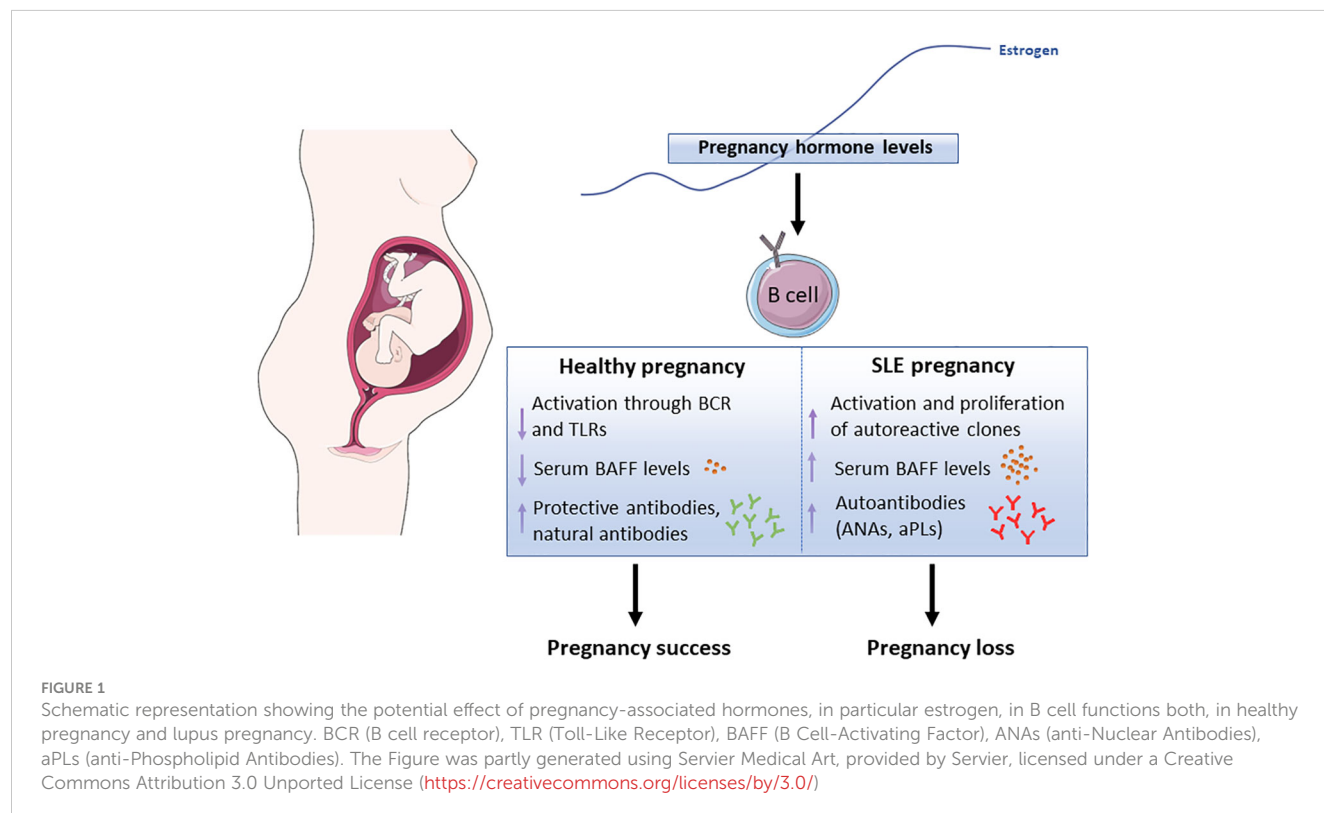
4 B-cell activation and autoantibody production in lupus: impact on pregnancy well-being

Upon activation, B cells undergo a series of tightly regulated processes that culminate in the differentiation of highly specialized cells capable of producing antibodies, as well as memory B cells (45). In addition to antibodies, activated B cells can produce a wide range of cytokines, especially when their activation goes through their BCR together with BAFF-R (71). Signaling through the BAFF-R activates several downstream pathways, including NF-KB, ERK, and MAPK, which regulate the survival functions of immature, transitional, and mature B cells (72, 73). Interestingly, it has been demonstrated that B cells from pregnant women show downregulation of transcripts associated with these pathways (74) along with reduced levels of BAFF in serum as pregnancy progresses (75), suggesting that B cells are less susceptible to being activated during pregnancy. Indeed, a transcriptomic analysis performed on B cells isolated from pregnant mice confirmed that several B cell activation pathways, including BCR,

TLR, and BAFF-R, are significantly diminished compared to B cells from non-pregnant control animals (44). Furthermore, a study by Valeff et al. (44) found that B cells isolated from pregnant women in the first trimester of pregnancy produced significantly lower levels of inflammatory cytokines when activated through their BCR and TLRs compared to B cells from non-pregnant women, reinforcing the notion of B cells being less susceptible to activation, at least during the early stages of pregnancy.

In the context of SLE, aberrant B-cell activation plays a significant role in the pathogenesis of the disease. Dysregulation of BCR and BAFF-R pathways are common and dominant factors involved in this aberrant B-cell activation (76). Furthermore, patients with SLE exhibit elevated levels of BAFF in their serum (77–79), strongly indicating the involvement of the BAFF-R pathway in B cells as a key component of SLE pathology. Indeed, mice overexpressing BAFF develop a lupus-like disease characterized by the production of ANAs and anti-dsDNA (80).

In the context of pregnancy, while the production of natural and protective antibodies is related to pregnancy success (81, 82), the presence of autoantibodies is associated with RPL (8). There is growing evidence suggesting that ANAs can play a role in both early pregnancy complications, such as embryo implantation, and pregnancy loss (83). While low titers of ANAs are common in healthy women, those with RPL often exhibit high titers of ANAs (>1:160) (83). Moreover, ANAs have been suggested to have a direct effect on the quality and development of oocytes and embryos, leading to reduced implantation rates (84). In the fetal-maternal interface, ANAs can induce the precipitation of immune complexes, attributed to elevated C3 levels, resulting in T cell activation and increased production of inflammatory cytokine (IFN- α), which in turn stimulates the humoral



immune response (85, 86). Complement activation rapidly increases the production of the pro-inflammatory cytokine TNF, which in turn recruits inflammatory cells into the placenta, ultimately contributing to pregnancy loss (87).

It is well known that imbalances toward a pro-inflammatory milieu are associated with poor pregnancy outcomes (88). Moreover, the TNF/IL-10 ratio in serum is used as an indicator or predictor of pregnancy loss (89). In line with this, the production of IL-10 by B cells is considered essential for successful pregnancies (90). Interestingly, in patients with SLE, there is a significant decrease in IL-10 production by B cells (91). Even though, the elevated serum levels of IL-10 observed in pregnant women with SLE compared to controls (25) would be an advantage in normal pregnancy conditions, the immunosuppressive and anti-inflammatory effects of this cytokine are impaired in patients with SLE compared to healthy individuals (92).

Therefore, it is reasonable to speculate that uncontrolled B cell activation in patients with SLE during gestation may lead to the production of pro-inflammatory cytokines and harmful antibodies, which could potentially compromise the well-being of the pregnancy.

In conclusion, maintaining a balanced B-cell activation is essential for a successful pregnancy. In women with preexisting SLE, hormonal changes may disrupt this balance, leading to the production of inflammatory cytokines and autoantibodies. This dysregulation can exacerbate disease symptoms and contribute to pregnancy complications, including RPL. Therefore, understanding the impact of B-cell activation and its relationship with hormonal changes during gestation is crucial for managing SLE and optimizing pregnancy outcomes (Figure 1).

Author contributions

NV designed, drafted, and revised the work. MV designed and drafted the work. LD drafted and revised the work. FJ designed,

drafted, supervised, and revised the work. All authors contributed to the article and approved the submitted version.

Funding

This work was partially supported by a grant from Agencia Nacional de Promoción Científica y Tecnológica (PICT-2020-00393) to FJ and Natural Science Foundation of Guangdong Province-General Programme, China (2022A151010650) to LD.

Acknowledgments

We especially thank the personal staff from the CEFYBO: Patricia Fernandez, María Alejandra Veron, María Cristian Lincon, Alberto Capriolo, and Alcira Mazziotti.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

- Andersen A-MN. Maternal age and fetal loss: population based register linkage study. *BMJ* (2000) 320(7251):1708–12. doi: 10.1136/bmj.320.7251.1708
- Bender Atik R, Christiansen OB, Elson J, Kolte AM, Lewis S, Middeldorp S, et al. ESHRE guideline: recurrent pregnancy loss. *Hum Reprod Open* (2018) 2018(2):hoy004. doi: 10.1093/hropen/hoy004
- Fausett MB, Branch DW. Autoimmunity and pregnancy loss. *Semin Reprod Med* (2000) 18(04):379–92. doi: 10.1055/s-2000-13728
- Ohmura K, Oku K, Kitaori T, Amengual O, Hisada R, Kanda M, et al. Pathogenic roles of anti-C1q antibodies in recurrent pregnancy loss. *Clin Immunol* (2019) 203:37–44. doi: 10.1016/j.clim.2019.04.005
- Amari Chinchilla K, Vijayan M, Taveras Garcia B, Jim B. Complement-mediated disorders in pregnancy. *Adv Chronic Kidney Dis* (2020) 27(2):155–64. doi: 10.1053/j.ackd.2020.01.002
- Girardi G, Lingo JJ, Fleming SD, Regal JF. Essential role of complement in pregnancy: from implantation to parturition and beyond. *Front Immunol* (2020) 11:1681. doi: 10.3389/fimmu.2020.01681
- Gleicher N, El-Roeiy A, Confino E, Friberg J. Reproductive failure because of autoantibodies: Unexplained infertility and pregnancy wastage. *Am J Obstetrics Gynecol* (1989) 160(6):1376–85. doi: 10.1016/0002-9378(89)90858-2
- D'Ippolito S, Ticconi C, Tersigni C, Garofalo S, Martino C, Lanzone A, et al. The pathogenic role of autoantibodies in recurrent pregnancy loss. *Am J Reprod Immunol* (2020) 83(1):0–3. doi: 10.1111/aji.13200
- Edelman PH, Rouquette AM, Verdy E, Elias A, Cabane J, Cornet D, et al. Autoimmunity, fetal losses, lupus anticoagulant: beginning of systemic lupus erythematosus or new autoimmune entity with gynaeco-obstetrical expression? *Hum Reprod* (1986) 1(5):295–7. doi: 10.1093/oxfordjournals.humrep.a136408
- Aoki K, Dudkiewicz AB, Matsuura E, Novotny M, Kaherlein G, Gleicher N. Clinical significance of β 2-glycoprotein I-dependent anticardiolipin antibodies in the reproductive autoimmune failure syndrome: Correlation with conventional antiphospholipid antibody detection systems. *Am J Obstetrics Gynecol* (1995) 172(3):926–31. doi: 10.1016/0002-9378(95)90023-3
- Harger JH, Rabin BS, Marchese SG. The prognostic value of antinuclear antibodies in women with recurrent pregnancy losses: a prospective controlled study. *Obstetrics Gynecol* (1989) 73(3 Pt 1):419–24.
- Gao R, Zeng X, Qin L. Systemic autoimmune diseases and recurrent pregnancy loss: research progress in diagnosis and treatment. *Chin Med J* (2021) 134(17):2140–2. doi: 10.1097/CM9.0000000000001691
- Tedeschi SK, Bernas B, Costenbader KH. Sexual disparities in the incidence and course of SLE and RA. *Clin Immunol* (2013) 149(2):211–8. doi: 10.1016/j.clim.2013.03.003
- Moulton VR, Suarez-Fueyo A, Meidan E, Li H, Mizui M, Tsokos GC. Pathogenesis of human systemic lupus erythematosus: A cellular perspective. *Trends Mol Med* (2017) 23(7):615–35. doi: 10.1016/j.molmed.2017.05.006
- Tsokos GC. Mechanisms of disease systemic lupus erythematosus. *N Engl J Med* (2011) 22(1):2110–21. doi: 10.1056/NEJMra1100359

16. Aringer M, Costenbader K, Daikh D, Brinks R, Mosca M, Ramsey-Goldman R, et al. 2019 european league against rheumatism/american college of rheumatology classification criteria for systemic lupus erythematosus. *Arthritis Rheumatol (Hoboken N.J.)* (2019) 71(9):1400–12. doi: 10.1002/art.40930
17. Pisetsky DS. The immunopathogenesis and immunopathology of systemic lupus erythematosus. In Schur P, Massarotti E. (eds) *Lupus Erythematosus* (New York, NY: Springer) (2012), 13–26. doi: 10.1007/978-1-4614-1189-5_2
18. Choi J, Kim ST, Craft J. The pathogenesis of systemic lupus erythematosus—an update. *Curr Opin Immunol* (2012) 24(6):651–7. doi: 10.1016/j.coi.2012.10.004
19. Mohan C, Putterman C. Genetics and pathogenesis of systemic lupus erythematosus and lupus nephritis. *Nat Rev Nephrol* (2015) 11(6):329–41. doi: 10.1038/nrneph.2015.33
20. Kwak-Kim J, Skariah A, Wu L, Salazar D, Sung N, Ota K. Humoral and cellular autoimmunity in women with recurrent pregnancy losses and repeated implantation failures: A possible role of vitamin D. *Autoimmun Rev* (2016) 15(10):943–7. doi: 10.1016/j.AUTREV.2016.07.015
21. Xu J, Chen D, Duan X, Li L, Tang Y, Peng B. The association between antiphospholipid antibodies and late fetal loss: A systematic review and meta-analysis. *Acta Obstetrica Gynecologica Scandinavica* (2019) 98(12):1523–33. doi: 10.1111/aogs.13665
22. Mekinian A, Alijotas-Reig J, Carrat F, Costedoat-Chalumeau N, Ruffatti A, Lazzaroni MG, et al. Refractory obstetrical antiphospholipid syndrome: Features, treatment, and outcome in a European multicenter retrospective study. *Autoimmun Rev* (2017) 16(7):730–4. doi: 10.1016/j.autrev.2017.05.006
23. Branch DW. Immunologic disease and fetal death. *Clin Obstetrics Gynecol* (1987) 30(2):295–311. doi: 10.1097/00003081-198706000-00009
24. Petri M. Pregnancy and systemic lupus erythematosus. *Best Pract Res Clin Obstetrics Gynaecol* (2020) 64:24–30. doi: 10.1016/j.bpobgyn.2019.09.002
25. Björkander S, Bremme K, Persson J-O, van Vollenhoven RF, Sverremark-Ekström E, Holmlund U. Pregnancy-associated inflammatory markers are elevated in pregnant women with systemic lupus erythematosus. *Cytokine* (2012) 59(2):392–9. doi: 10.1016/j.cyto.2012.04.046
26. Ruchakorn N, Ngamjanyaporn P, Suangtamai T, Kafaksom T, Polpanumas C, Petpisit V, et al. Performance of cytokine models in predicting SLE activity. *Arthritis Res Ther* (2019) 21(1):287. doi: 10.1186/s13075-019-2029-1
27. Petri M, Howard D, Repke J. Frequency of lupus flare in pregnancy. The Hopkins Lupus Pregnancy Center experience. *Arthritis Rheumatism* (1991) 34(12):1538–45. doi: 10.1002/art.1780341210
28. Clowse MEB, Magder LS, Witter F, Petri M. The impact of increased lupus activity on obstetric outcomes. *Arthritis Rheumatism* (2005) 52(2):514–21. doi: 10.1002/art.20864
29. Liu J, Zhao Y, Song Y, Zhang W, Bian X, Yang J, et al. Pregnancy in women with systemic lupus erythematosus: a retrospective study of 111 pregnancies in Chinese women. *J Maternal-Fetal Neonatal Med* (2012) 25(3):261–6. doi: 10.3109/14767058.2011.572310
30. Hayslett JP, Lynn RI. Effect of pregnancy in patients with lupus nephropathy. *Kidney Int* (1980) 18(2):207–20. doi: 10.1038/ki.1980.129
31. Lockshin MD, Druzin ML, Goei S, Qamar T, Magid MS, Jovanovic L, et al. Antibody to cardiolipin as a predictor of fetal distress or death in pregnant patients with systemic lupus erythematosus. *New Engl J Med* (1985) 313(3):152–6. doi: 10.1056/NEJM198507183130304
32. Ogasawara M, Aoki K, Hayashi Y. A prospective study on pregnancy risk of antiphospholipid antibodies in association with systemic lupus erythematosus. *J Reprod Immunol* (1995) 28(2):159–64. doi: 10.1016/0165-0378(94)00912-Q
33. Gryka-Marton M, Szukiewicz D, Teliga-Czajkowska J, Olesinska M. An overview of neonatal lupus with anti-ro characteristics. *Int J Mol Sci* (2021) 22(17):9281. doi: 10.3390/ijms22179281
34. Sachdeva R, Pal R. A pregnancy hormone-cell death link promotes enhanced lupus-specific immunological effects. *Front Immunol* (2022) 13:1051779. doi: 10.3389/fimmu.2022.1051779
35. Bose M, Jefferies C. Sex bias in systemic lupus erythematosus: a molecular insight. *Immunometabolism* (2022) 4(3):e00004. doi: 10.1097/IN.0000000000000004
36. Peart E, Clowse MEB. Systemic lupus erythematosus and pregnancy outcomes: An update and review of the literature. *Curr Opin Rheumatol* (2014) 26(2):118–23. doi: 10.1097/BOR.0000000000000030
37. Bundhun PK, Soogund MZS, Huang F. Impact of systemic lupus erythematosus on maternal and fetal outcomes following pregnancy: A meta-analysis of studies published between years 2001–2016. *J Autoimmun* (2017) 79:17–27. doi: 10.1016/j.jaut.2017.02.009
38. Singh AG, Chowdhary VR. Pregnancy-related issues in women with systemic lupus erythematosus. *Int J Rheumatic Dis* (2015) 18(2):172–81. doi: 10.1111/1756-185X.12524
39. He WR, Wei H. Maternal and fetal complications associated with systemic lupus erythematosus. *Medicine* (2020) 99(16):e19797. doi: 10.1097/MD.00000000000019797
40. Clark CA, Spitzer KA, Laskin CA. Decrease in pregnancy loss rates in patients with systemic lupus erythematosus over a 40-year period. *J Rheumatol* (2005) 32(9):1709–12.
41. Buyon JP, Kim MY, Guerra MM, Laskin CA, Petri M, Lockshin MD, et al. Predictors of pregnancy outcomes in patients with lupus. *Ann Internal Med* (2015) 163(3):153–63. doi: 10.7326/M14-2235
42. Mekinian A, Cohen J, Alijotas-Reig J, Carbillon L, Nicaise-Roland P, Kayem G, et al. Unexplained recurrent miscarriage and recurrent implantation failure: is there a place for immunomodulation? *Am J Reprod Immunol* (2016) 76(1):8–28. doi: 10.1111/aji.12493
43. Mankee A, Petri M, Magder LS. Lupus anticoagulant, disease activity and low complement in the first trimester are predictive of pregnancy loss. *Lupus Sci Med* (2015) 2(1):e000095. doi: 10.1136/lupus-2015-000095
44. Valeff N, Muzzio DO, Matzner F, Dibo M, Golchert J, Homuth G, et al. B cells acquire a unique and differential transcriptomic profile during pregnancy. *Genomics* (2021) 113(4):2614–22. doi: 10.1016/j.ygeno.2021.06.016
45. LeBien TW, Tedder TF. B lymphocytes: How they develop and function. *Blood* (2008) 112(5):1570–80. doi: 10.1182/blood-2008-02-078071
46. Rastogi I, Jeon D, Moseman JE, Muralidhar A, Potluri HK, McNeel DG. Role of B cells as antigen presenting cells. *Front Immunol* (2022) 13:954936. doi: 10.3389/fimmu.2022.954936
47. Cerutti A, Cols M, Puga I. Marginal zone B cells: virtues of innate-like antibody-producing lymphocytes. *Nat Rev Immunol* (2013) 13(2):118–32. doi: 10.1038/nri3383
48. Beagley KW, Gockel CM. Regulation of innate and adaptive immunity by the female sex hormones oestradiol and progesterone. *FEMS Immunol Med Microbiol* (2003) 38(1):13–22. doi: 10.1016/S0928-8244(03)00202-5
49. Moulton VR. Sex hormones in acquired immunity and autoimmune disease. *Front Immunol* (2018) 9:2279(OCT). doi: 10.3389/fimmu.2018.02279
50. Abdullah M, Chai P-S, Chong M-Y, Tohit ERM, Ramasamy R, Pei CP, et al. Gender effect on *in vitro* lymphocyte subset levels of healthy individuals. *Cell Immunol* (2012) 272(2):214–9. doi: 10.1016/j.cellimm.2011.10.009
51. Dodd KC, Menon M. Sex bias in lymphocytes: Implications for autoimmune diseases. *Front Immunol* (2022) 13:945762(November). doi: 10.3389/fimmu.2022.945762
52. Grimaldi CM, Cleary J, Dagtas AS, Moussai D, Diamond B. Estrogen alters thresholds for B cell apoptosis and activation. *J Clin Invest* (2002) 109(12):1625–33. doi: 10.1172/jci14873
53. Grimaldi CM, Hill L, Xu X, Peeva E, Diamond B. Hormonal modulation of B cell development and repertoire selection. *Mol Immunol* (2005) 42(7):811–20. doi: 10.1016/j.molimm.2004.05.014
54. Hill L, Jeganathan V, Chinnasamy P, Grimaldi C, Diamond B. Differential roles of estrogen receptors α and β in control of B-cell maturation and selection. *Mol Med* (2011) 17(3–4):211–20. doi: 10.2119/molmed.2010.00172
55. Bynoe MS, Grimaldi CM, Diamond B. Estrogen up-regulates Bcl-2 and blocks tolerance induction of naïve B cells. *Proc Natl Acad Sci* (2000) 97(6):2703–8. doi: 10.1073/pnas.040577497
56. Grimaldi CM, Jeganathan V, Diamond B. Hormonal regulation of B cell development: 17 β -estradiol impairs negative selection of high-affinity DNA-reactive B cells at more than one developmental checkpoint. *J Immunol* (2006) 176(5):2703–10. doi: 10.4049/jimmunol.176.5.2703
57. Tenbrock K, Rauen T. T cell dysregulation in SLE. *Clin Immunol* (2022) 239:109031. doi: 10.1016/j.clim.2022.109031
58. Monteiro C, Kasahara T, Sacramento PM, Dias A, Leite S, Silva VG, et al. Human pregnancy levels of estrogen and progesterone contribute to humoral immunity by activating TFF /B cell axis. *Eur J Immunol* (2021) 51(1):167–79. doi: 10.1002/eji.202048658
59. Tsokos GC. Systemic lupus erythematosus. *New Engl J Med* (2011) 365(22):2110–21. doi: 10.1056/NEJMra1100359
60. Hill BG, Hodge B, Misischia R. Lupus nephritis in a transgender woman on cross-sex hormone therapy: a case for the role of oestrogen in systemic lupus erythematosus. *Lupus* (2020) 29(13):1807–10. doi: 10.1177/0961203320946372
61. Tabor DE, Gould KA. Estrogen receptor alpha promotes lupus in (NZB \times NZW) F1 mice in a B cell intrinsic manner. *Clin Immunol* (2017) 174(3):41–52. doi: 10.1016/j.clim.2016.10.011
62. Stoecker ZM, Zinger H, Mozes E. Beneficial effects of the anti-oestrogen tamoxifen on systemic lupus erythematosus of (NZB \times NZW)F1 female mice are associated with specific reduction of IgG3 autoantibodies. *Ann Rheumatic Dis* (2003) 62(4):341–6. doi: 10.1136/ard.62.4.341
63. Doria A, Iaccarino L, Sarzi-Puttini P, Ghirardello A, Zampieri S, Arienti S, et al. Estrogens in pregnancy and systemic lupus erythematosus. *Ann New York Acad Sci* (2006) 1069(1):247–56. doi: 10.1196/annals.1351.022
64. Cohen-Solal JFG, Jeganathan V, Hill L, Kawabata D, Rodriguez-Pinto D, Grimaldi C, et al. Hormonal regulation of B-cell function and systemic lupus erythematosus. *Lupus* (2008) 17(6):528–32. doi: 10.1177/0961203308089402
65. Medina KL, Kincade PW. Pregnancy-related steroids are potential negative regulators of B lymphopoiesis. *Proc Natl Acad Sci USA* (1994) 91(12):5382–6. doi: 10.1073/pnas.91.12.5382
66. Sanz I, Wei C, Jenks SA, Cashman KS, Tipton C, Woodruff MC, et al. Challenges and opportunities for consistent classification of human B cell and plasma cell populations. *Front Immunol* (2019) 10:2458(OCT). doi: 10.3389/fimmu.2019.02458

67. Muzzio DO, Soldati R, Ehrhardt J, Utpatel K, Evert M, Zenclussen AC, et al. B cell development undergoes profound modifications and adaptations during pregnancy in mice. *Biol Reprod* (2014) 91(5):1–11. doi: 10.1095/biolreprod.114.122366
68. Canny SP, Jackson SW. B cells in systemic lupus erythematosus: from disease mechanisms to targeted therapies. *Rheumatic Dis Clinics North America* (2021) 47(3):395–413. doi: 10.1016/j.rdc.2021.04.006
69. Carbone J, Sarmiento E, Gallego A, Lanio N, Navarro J, García S, et al. Peripheral blood T- and B-cell immunophenotypic abnormalities in selected women with unexplained recurrent miscarriage. *J Reprod Immunol* (2016) 113:50–3. doi: 10.1016/j.jri.2015.11.003
70. Ângelo-Dias M, Martins C, Dias SS, Borrego LM, Lima J. Association of B cells with idiopathic recurrent pregnancy loss: A systematic review and meta-analysis. *Int J Mol Sci* (2022) 23(23):15200. doi: 10.3390/ijms232315200
71. Hoffman W, Lakkis FG, Chalasani G. B cells, antibodies, and more. *Clin J Am Soc Nephrol* (2016) 11(1):137–54. doi: 10.2215/CJN.09430915
72. Khan WN. B cell receptor and BAFF receptor signaling regulation of B cell homeostasis. *J Immunol* (2009) 183(6):3561–7. doi: 10.4049/jimmunol.0800933
73. Smulski CR, Eibel H. BAFF and BAFF-receptor in B cell selection and survival. *Front Immunol* (2018) 9:2285(OCT). doi: 10.3389/fimmu.2018.02285
74. Chen D, Wang W, Wu L, Liang L, Wang S, Cheng Y, et al. Single-cell atlas of peripheral blood mononuclear cells from pregnant women. *Clin Trans Med* (2022) 12(5):e821. doi: 10.1002/ctm2.821
75. Stohl HE, Stohl W. Maternal and cord blood BAFF and APRIL levels during pregnancy. *Am J Reprod Immunol* (2023) 89(3):e13654. doi: 10.1111/aji.13654
76. Kang N, Liu X, You X, Sun W, Haneef K, Sun X, et al. Aberrant B-cell activation in systemic lupus erythematosus. *Kidney Dis* (2022) 8(6):437–45. doi: 10.1159/000527213
77. Stohl W, Metyas S, Tan S-M, Cheema GS, Oamar B, Xu D, et al. B lymphocyte stimulator overexpression in patients with systemic lupus erythematosus: Longitudinal observations. *Arthritis Rheumatism* (2003) 48(12):3475–86. doi: 10.1002/art.11354
78. Petri M, Stohl W, Chatham W, McCune WJ, Chevrier M, Ryel J, et al. Association of plasma B lymphocyte stimulator levels and disease activity in systemic lupus erythematosus. *Arthritis Rheumatism* (2008) 58(8):2453–9. doi: 10.1002/art.23678
79. Vincent FB, Kandane-Rathnayake R, Koelmeyer R, Hoi AY, Harris J, Mackay F, et al. Analysis of serum B cell-activating factor from the tumor necrosis factor family (BAFF) and its soluble receptors in systemic lupus erythematosus. *Clin Trans Immunol* (2019) 8(4):e1047. doi: 10.1002/cti2.1047
80. Thorn M, Lewis RH, Mumbey-Wafula A, Kantrowitz S, Spatz LA. BAFF overexpression promotes anti-dsDNA B-cell maturation and antibody secretion. *Cell Immunol* (2010) 261(1):9–22. doi: 10.1016/j.cellimm.2009.10.004
81. Ziegler KB, Muzzio DO, Matzner F, Bommer I, Ventimiglia MS, Malinowsky K, et al. Human pregnancy is accompanied by modifications in B cell development and immunoglobulin profile. *J Reprod Immunol* (2018) 129(May):40–7. doi: 10.1016/j.jri.2018.07.003
82. Banjar S, Kadour E, Khoudja R, Ton-leclerc S, Beauchamp C, Beltempo M, et al. Intravenous immunoglobulin use in patients with unexplained recurrent pregnancy loss. *Am J Reprod Immunol* (2023) 90(2):e13737. doi: 10.1111/aji.13737
83. Liu T, Guo X, Liao Y, Liu Y, Zhu Y, Chen X. Correlation between the presence of antinuclear antibodies and recurrent pregnancy loss: A mini review. *Front Endocrinol* (2022) 13:873286. doi: 10.3389/fendo.2022.873286
84. Ying Y, Zhong Y, Zhou C, Xu Y, Wang Q, Li J, et al. Antinuclear antibodies predicts a poor IVF-ET outcome: impaired egg and embryo development and reduced pregnancy rate. *Immunol Investigations* (2012) 41(5):458–68. doi: 10.3109/08820139.2012.660266
85. Papadimitrakaki ED, Choulaki C, Koutala E, Bertsias G, Tsatsanis C, Gergianaki I, et al. Expansion of toll-like receptor 9-expressing B cells in active systemic lupus erythematosus: Implications for the induction and maintenance of the autoimmune process. *Arthritis Rheumatism* (2006) 54(11):3601–11. doi: 10.1002/art.22197
86. Zeng M, Wen P, Duan J. Association of antinuclear antibody with clinical outcome of patients undergoing *in vitro* fertilization/intracytoplasmic sperm injection treatment: A meta-analysis. *Am J Reprod Immunol* (2019) 82(3):e13158. doi: 10.1111/aji.13158
87. Girardi G. Complement inhibition keeps mothers calm and avoids fetal rejection. *Immunol Investigations* (2008) 37(5–6):645–59. doi: 10.1080/08820130802191615
88. Azizieh FY, Raghupathy RG. Tumor necrosis factor- α and pregnancy complications: A prospective study. *Med Principles Pract* (2015) 24(2):165–70. doi: 10.1159/000369363
89. Kaislasuo J, Simpson S, Petersen JF, Peng G, Aldo P, Lokkegaard E, et al. IL-10 to TNF α ratios throughout early first trimester can discriminate healthy pregnancies from pregnancy losses. *Am J Reprod Immunol* (2020) 83(1):e13195. doi: 10.1111/aji.13195
90. Danaii S, Ghorbani F, Ahmadi M, Abbaszadeh H, Koushaeian L, Soltani-Zangbar MS, et al. IL-10-producing B cells play important role in the pathogenesis of recurrent pregnancy loss. *Int Immunopharmacol* (2020) 87:106806. doi: 10.1016/j.intimp.2020.106806
91. Menon M, Blair PA, Isenberg DA, Mauri C. A regulatory feedback between plasmacytoid dendritic cells and regulatory B cells is aberrant in systemic lupus erythematosus. *Immunity* (2016) 44(3):683–97. doi: 10.1016/j.immuni.2016.02.012
92. Biswas S, Bieber K, Manz RA. IL-10 revisited in systemic lupus erythematosus. *Front Immunol* (2022) 13:970906. doi: 10.3389/fimmu.2022.970906



OPEN ACCESS

EDITED BY

Zitao Liu,
New Hope Fertility Center, United States

REVIEWED BY

Nicoletta Di Simone,
Humanitas University, Italy
Emre Pabuccu,
Ufuk University, Türkiye

*CORRESPONDENCE

Chan Tian
✉ tianchan@bjmu.edu.cn

RECEIVED 09 July 2023

ACCEPTED 30 November 2023

PUBLISHED 03 January 2024

CITATION

Gong C, Yang W, Liu X, Li X, Wang Y and
Tian C (2024) Low follistatin level is a causal
risk factor for spontaneous abortion: a two-
sample mendelian randomization study.
Front. Endocrinol. 14:1255591.
doi: 10.3389/fendo.2023.1255591

COPYRIGHT

© 2024 Gong, Yang, Liu, Li, Wang and Tian.
This is an open-access article distributed under
the terms of the [Creative Commons Attribution
License \(CC BY\)](https://creativecommons.org/licenses/by/4.0/). The use, distribution or
reproduction in other forums is permitted,
provided the original author(s) and the
copyright owner(s) are credited and that the
original publication in this journal is cited, in
accordance with accepted academic
practice. No use, distribution or reproduction
is permitted which does not comply with
these terms.

Low follistatin level is a causal risk factor for spontaneous abortion: a two-sample mendelian randomization study

Chen Gong^{1,2,3,4}, Wenzhi Yang⁵, Xue Liu⁶, Xinliang Li⁶,
Yutong Wang^{1,2,3,4} and Chan Tian^{1,2,3,4,6*}

¹State Key Laboratory of Female Fertility Promotion, Center for Reproductive Medicine, Department of Obstetrics and Gynecology, Peking University Third Hospital, Beijing, China,

²National Clinical Research Center for Obstetrics and Gynecology, Peking University Third Hospital, Beijing, China, ³Key Laboratory of Assisted Reproduction, Peking University, Ministry of Education, Beijing, China, ⁴Beijing Key Laboratory of Reproductive Endocrinology and Assisted Reproductive Technology, Beijing, China, ⁵Department of Neurology, Peking University Third Hospital, Beijing, China, ⁶Department of Medical Genetics, Center for Medical Genetics, Peking University Health Science Center, Beijing, China

Background: Recurrent pregnancy loss is a distressing event during pregnancy, and understanding its causal factors is crucial. Follistatin, a glycoprotein involved in folliculogenesis and embryogenesis, has been implicated as a potential contributor to the risk of spontaneous abortion. However, establishing a causal relationship requires rigorous investigation using robust methods.

Methods: In this study, we utilized mendelian randomization (MR), a powerful genetic epidemiological approach, to examine the causal relationship between follistatin levels and spontaneous abortion. We obtained instrumental variables strongly associated with follistatin levels from large-scale genome-wide association from the IEU database. The inverse variance weighting (IVW) method was taken as gold standard. We also performed sensitivity test to evaluate the robustness of our result.

Results: MR analysis revealed a significant causal relationship between low follistatin levels and spontaneous abortion ($p = 0.03$). Sensitivity analyses, including pleiotropy test, heterogeneity test, and leave-one-out analysis, all supported the robustness of our findings.

Conclusion: Our study provides compelling evidence supporting the causal relationship between low follistatin levels and increased risk of spontaneous abortion. These findings underscore the importance of follistatin in the etiology of spontaneous abortion and suggest potential preventive interventions. Modulating follistatin levels or relevant pathways could hold promise for reducing the incidence of spontaneous abortion and improving reproductive outcomes. The utilization of MRs strengthens the validity of our

results by mitigating confounding and reverse causality biases. Further research is needed to elucidate the underlying molecular mechanisms and explore therapeutic strategies targeting follistatin levels.

KEYWORDS

follistatin, spontaneous abortion, mendelian randomization, abortion, recurrent pregnancy loss

1 Introduction

Spontaneous abortion is a frequently encountered complication during pregnancy, characterized by the loss of pregnancy before 20 weeks of gestation (1). Approximately 9–20% of all recognized pregnancies result in spontaneous abortion. Among these, 3–5% of couples face the challenge of two or more clinically recognized pregnancies ending in failure, known as recurrent pregnancy loss (RPL) (2, 3). RPL can be devastating, bringing great trauma to both the patient and their family (4). Various factors contribute to RPL, with chromosomal abnormality being the most common, responsible for over half of RPL cases (5). Additionally, 10%–15% of women with multiple pregnancy losses exhibit uterine anomalies, such as partial or complete septum (6, 7). Hormonal causes such as luteal phase defect, pregestational diabetes mellitus, and polycystic ovary disease can lead to RPL (8). Moreover, immune disorders may contribute to RPL by dysregulating trophoblast function and endometrial angiogenesis (9). For example, primary antiphospholipid syndrome (APS), present in one-third of RPL patients, is associated with elevated serum antiphospholipid antibody (aPL) levels. Increased aPL levels reduce placental hormone production, impair trophoblast function, and result in pregnancy loss and other obstetric complications (10). Women with celiac disease often have elevated anti-transglutaminase type 2 (anti-TG2) autoantibodies, leading to reduced blood vessel formation and disrupted endometrial angiogenesis, contributing to RPL (11). Infections, exposure to environmental agents, and elevated homocysteine levels are also implicated in RPL (12). Nevertheless, the understanding of RPL remains significantly limited, as almost 50% of RPL cases are still categorized as unexplained (9).

Follistatin (FST) is a secreted protein that primarily synthesized and secreted by the liver, mainly implicated in suppressing follicle-stimulating hormone (FSH) activity through autocrine or paracrine mechanisms (13–15). Notably, FST serves as a binding protein and regulator in the transforming growth factor-beta (TGF- β) signaling pathway, selectively binding to ligands such as activins and bone morphogenetic proteins (BMP) (13, 16, 17). It restrained granulosa cell proliferation and steroidogenesis by neutralizing the action of

activin (18–20). It also enhanced basal estradiol and progesterone production (21, 22), promoting cell invasion *via* the ALK4-SMAD2/3-SMAD4 signaling pathway (23–26).

Serum FST increased significantly throughout gestation until the first day of parturition and declined afterward (27). Evidence showed its possible role as chemokine to induce trophoblast migration and invasion through the enhanced JNK signaling, contributing to maintain trophoblast function and promote placental development (28, 29). FST was upregulated in the decidua during early pregnancy, and women with RPL were observed to have a lower endometrial expression of FST during the luteal phase (30). A lower FST level in endometrium stromal cells of women with RPL was also observed (30). Conditional knockout of mice uterine *Fst* can cause severe fertility defects, reduced responsiveness to estrogen and progesterone signals, impaired artificial decidualization, and an unreceptive environment for embryo attachment. These findings suggest that *Fst* may play a crucial role in facilitating uterine receptivity (31). A decreased FST level was also observed in serum and placenta of women with preeclampsia (PE) (32–34), resulting in impaired trophoblast function through upregulating GDF11 levels in trophoblasts. The dysregulation of the FST-GDF11-Smad2/3 axis may be critical to trophoblast function, which adds more evidence to the essential role of FST on trophoblast during pregnancy (35).

Mendelian randomization (MR) is an epidemiological tool based on genetic variants related to exposure factors, helping to assess the association of these gene variations with outcomes such as disease onset or mortality. Its core relies on using genetic data as a bridge to investigate causal relationships between a particular exposure and a specific outcome (36, 37). Randomized controlled trials (RCTs) have long been recognized as the gold standard for causal inference, yet it is costly and complicated to conduct. Similar to RCTs that randomly assign participants to a trial or control group, MR studies “randomize” participants based on one or more gene alleles that influence risk factors and attempt to determine if carriers of these genetic variations have different disease onset risks compared with non-carriers (38). Traditional observational study designs rely on exposure obtained through questionnaires,

biochemical assays, or imaging, whereas genetic variations exist at birth and remain stable throughout life. Notably, information of genetic variation and diseases is easy to acquire through open-dataset, and since it leaves out the complicated implementation process and ethical restriction, it is much easier to conduct compared with RCTs (39). In view of these incomparable advantages of MR, here we performed a two-sample MR analysis of the GWAS summary data from the UK Biobank and EBI database so as to find whether there is a potential causal association between FST level and spontaneous abortion, trying to provide novel evidence in this field of research.

2 Materials and methods

2.1 Study design

Here, we conducted a two-sample MR analysis to examine the possible causal association between FST level and spontaneous abortion. Figure 1 provides an overview of the study's key factors, including the exposures, outcomes, and genetic instruments. The study was based on previously published materials and public databases and received ethical approval and participant consent from the relevant institutional review committees.

The genetic variants in this study are fully considered based on the three principles below throughout our analysis. First is relevance assumption: the genetic variant must be closely correlated with follistatin levels; second is independence assumption: the genetic variant cannot be associated with any possible confounders of follistatin levels or spontaneous abortion; and third is exclusivity assumption: the genetic variant cannot be related to the relevant outcomes.

2.2 Data sources

2.2.1 Exposure population and data

We extracted exposure data from a previous study (40), downloaded from the website of IEU OPEN GWAS PROJECT (<https://gwas.mrcieu.ac.uk/>, GWAS ID: ebi-a-GCST90012080).

2.2.2 Outcome population and data

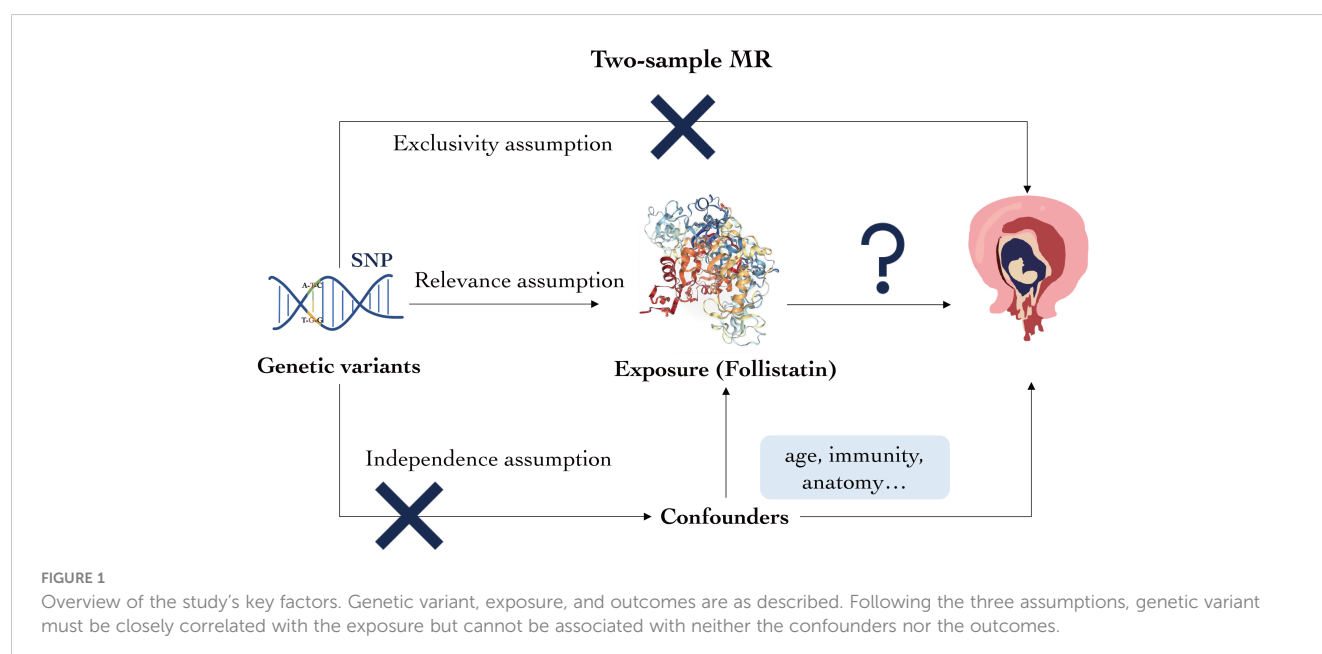
We extracted outcome data from a UK Biobank study (Dataset ID: ukb-d-O03), downloaded from the website of the IEU OPEN GWAS PROJECT (<https://gwas.mrcieu.ac.uk/>).

2.3 Statistical analysis

All the data processing and statistical analyze are performed using the R4.2.2 software (Lucent Technologies, USA). The MR, heterogeneity test and pleiotropy test were carried by the “Two Sample MR” package (41).

2.4 Extraction of instrumental variables

We performed two-step filtering to eliminate those SNPs that do not satisfy the relevance assumption and obtained the satisfying instrumental variables (IVs). We extracted SNPs that are (1) closely related to the exposure (follistatin level) at a genome-wide significance threshold of $p < 5 \times 10^{-8}$. 2) without linkage disequilibrium (LD) (linkage disequilibrium $r^2 < 0.05$), since we should make sure that no correlation LD between selected IVs and potential confounding factors exist. The R^2 value was calculated for these IVs to assess their association with the exposure (42).



2.5 Elimination of confounding factors

We used PhenoScanner (Version 2), a database of human genotype–phenotype associations to figure out whether the confounding factors such as anatomical abnormalities, hormonal imbalances (PCOS, luteal phase defect, etc.), and immune disorders (antiphospholipid syndrome, lupus erythematosus, etc.) may influence our result according to the independent assumption (43, 44). Over 65 billion associations and over 150 million unique genetic variants are recorded in PhenoScanner. We applied the “Phenoscaner” R package to investigated each of the five IVs and their corresponding phenotypes (43, 44). Any IVs exhibiting associations with confounding factors were excluded from the analysis, applying stringent criteria ($p < 1 \times 10^{-5}$, $r^2 > 0.8$).

2.6 Sensitivity test

To make sure that our MR results are robust enough for us to come up with a causal conclusion, we extensively performed three aspects of sensitivity test, namely, heterogeneity test, pleiotropy test, and leave-one-out analysis test. R software was used to visualize results with depicting scatter plot, forest plot, etc.

2.7 Two-sample MR analysis

In two-sample MR analysis, five methods are commonly used: MR-Egger, Weighted Median, Inverse Variance Weighted, Simple Mode, and Weighted Mode. Among these methods, Inverse Variance Weighted is widely accepted and considered the most effective, as it accounts for heterogeneity when assessing causality. However, the other four methods also demonstrate robustness to varying degrees. MR-Egger is particularly useful when there is potential violation of instrumental variable assumptions, such as pleiotropy. It estimates the causal effect while allowing for average pleiotropic bias (45). The Weighted Median method provides a robust estimate by considering the median of all possible instrumental variable estimates, even when up to 50% of the instruments are invalid. This method is advantageous in situations where some instrumental variables may be biased or weak. The Simple Mode and Weighted Mode methods combine the estimates from multiple instruments by either taking the mode or

using weighted averages. To evaluate the causal risk of FST levels, we performed all five methods. A positive result from any one or more of these methods would indicate a potential causal risk associated with FST levels.

3 Results

3.1 Study population

The exposure population pertains to follistatin levels, sourced from 39 cohorts of European ancestry. These data were thoroughly cleaned and summarized in a previously published study in 2020 (40), where a genome-wide meta-analysis of 90 cardiovascular-related proteins across 15 studies were identified. We extracted identified genetic variants related to follistatin levels from the recorded ebi-a dataset of IEU open GWAS project (GWAS ID: ebi-a-GCST90012080). In this dataset, 21,758 samples are involved, with 13,022,208 SNPs being reported (40). All of them are of European ancestry. The effect allele (EA), other allele (OA), beta coefficients, p value, and standard error (SE) are also included in this dataset for further investigation.

For the outcomes, summary-level data were obtained from the UK Biobank study (46). In the UK Biobank, pregnancy loss was defined as a history of self-reported spontaneous abortion or termination. We utilized the second round of Neale Lab’s genome-wide association analyses in the UK Biobank, obtaining 1,150 female patients with spontaneous abortion and 360,044 matched controls of European ancestry and 9,543,298 detected SNPs.

3.2 Five SNPs are validated as instrumental variables

MR relies on the idea of random allocation of genetic traits. If the frequency of SNPs harmonizes with the alteration of the exposure, we can tentatively deduce that the SNP is correlated with the exposure. We screen the total of 13,022,208 SNPs in the exposure dataset based on the relevance assumption and independence assumption mentioned in the “study design” and finally achieved five SNPs that satisfied for IVs. We presented some detailed information for these SNPs such as effect allele frequency,

TABLE 1 General data for five SNPs as instrumental variables.

| SNP for IV | | | | | Miscarriage (outcome) | | | | Follistatin (exposure) | | | | R ² |
|------------|----------|-------------|----|----|-----------------------|--------|--------|---------|------------------------|--------|--------|----------|----------------|
| Chr | Position | SNP ID | EA | OA | Beta | EAF | SE | p value | Beta | EAF | SE | p value | |
| 9 | 92228559 | rs10908903 | G | T | -1.43E-04 | 0.4677 | 0.0106 | 0.2850 | 0.0603 | 0.4587 | 0.0109 | 2.76E-08 | 3.43E-03 |
| 2 | 27730940 | rs1260326 | C | T | 1.05E-04 | 0.6069 | 0.0107 | 0.4381 | -0.1323 | 0.6024 | 0.0106 | 9.58E-36 | 3.21E-03 |
| 15 | 43726625 | rs150844304 | C | A | -5.26E-04 | 0.0245 | 0.0109 | 0.2187 | 0.2466 | 0.0294 | 0.0321 | 1.46E-14 | 6.98E-04 |
| 5 | 53327571 | rs31226 | C | T | 2.62E-04 | 0.6064 | 0.0123 | 0.0541 | -0.1289 | 0.5979 | 0.0107 | 1.56E-33 | 7.87E-04 |
| 12 | 57791833 | rs7974833 | C | T | 9.15E-05 | 0.2367 | 0.0321 | 0.5561 | 0.0849 | 0.2350 | 0.0123 | 5.61E-12 | 1.55E-04 |

Chr, chromosome; SNP, single-nucleotide polymorphism; EA, effect allele; OA, other allele; EAF, effect allele frequency; SE, standard error.

TABLE 2 Evaluation of instrumental variables.

| SNPs for IV | | Follistatin level (exposure) | | | |
|-------------|----------|------------------------------|--------|---------|--------|
| Chr | Position | SNP ID | MAF | beta | SE |
| 9 | 92228559 | rs10908903 | 0.4587 | 0.0603 | 0.0109 |
| 2 | 27730940 | rs1260326 | 0.6024 | -0.1323 | 0.0106 |
| 15 | 43726625 | rs150844304 | 0.0294 | 0.2466 | 0.0321 |
| 5 | 53327571 | rs31226 | 0.5979 | -0.1289 | 0.0107 |
| 12 | 57791833 | rs7974833 | 0.235 | 0.0849 | 0.0123 |

IV, instrumental variable; Chr, chromosome; SNP, single nucleotide polymorphism; MAF, minor allele frequency; Beta, the effect size; R^2 , IV explains the extent of exposure.

standard error, and effect allele (Table 1). In addition, we calculated the R^2 value for the IVs, which help to explain the extent of exposure (Table 2).

3.3 MR analysis showed that follistatin level is a causal risk for spontaneous abortion

Here, we adopted five methods to evaluate the follistatin level effect on the risk of spontaneous abortion, and the results are shown in Table 3 and Figure 2. Considering the absence of neither heterogeneity nor pleiotropy (which we would describe in detail on the next part), we selected IVW as the main method as well as the gold standard for determining the causal effect of FST level on the risk of spontaneous abortion (47). We found that the IVW method showed a p value of 0.03517563 (<0.05) and b of -0.001282787 (<0), indicating the causal relationship between low FST level in the European population.

3.4 Sensitivity analysis

In this work, sensitivity analysis is performed to (1) evaluate whether the results are robust and the conclusions are reliable; (2) assess whether the results have potential biases (such as genetic pleiotropy and data heterogeneity); and (3) evaluate whether there is a certain instrumental variable that significantly affects the outcome variable.

We detected no heterogeneity in the five IVs that we chose for the spontaneous abortion (MR-Egger Q statistics = 2.738052; Qdf = 3; Qpval = 0.4337995; IVW Q statistics = 2.875518; Qdf = 4; Qpval = 0.5788685) (Table 4). We then focused on the pleiotropy using the

MR-Egger method. The intercept with the Y-axis represents the horizontal pleiotropy. Zero horizontal pleiotropy is one of the prerequisites of applying the MR method according to the exclusive assumption. Here, we noticed no horizontal pleiotropy existed in our MR analysis results (Egger_intercept = 7.505441e-05; se = 0.0002024312; p value = 0.7354456) (Table 5). In the “leave-one-out” analysis, we sequentially removed each SNP and calculated the MR effect of the remaining SNPs. We noticed that the removal of any of these individual SNPs did not result in significant changes of the overall causal estimation effect (Figure 3). Taken together, these results suggest that our findings were robust and the exception of single IV exert no difference on the overall estimated causal effect.

The Wald ratio method was used to estimate the causal effect of each individual SNP on the risk of spontaneous abortion. The findings have been presented in a forest plot to provide a visual representation (Figure 4). The threshold of significance for the forest plot remained controversial. It can be defined as either $p < 0.05$ or $p < 0.05/n$ (n refers to the number of SNPs). Here, comprehensively regarding the p values for each single SNP on the outcome (Table 1), the leave-one-out analysis test (Figure 3), and all the SNPs combined (Figure 4), it was quite clear that a causal association existed between follistatin level and the risk of spontaneous abortion.

4 Discussion

In this study, we employed two-sample MR to investigate the causal relationship between FST levels and spontaneous abortion. The evidence from MR analysis indicates that low follistatin level was a causal risk factor for spontaneous abortion and these results

TABLE 3 Causal effect between follistatin level and spontaneous abortion by different MR analysis methods.

| Exposure | Outcome | Method | nSNP | p value | beta | R^2 |
|--------------------|----------------------|---------------------------|------|------------|--------------|----------|
| Follistatin levels | Spontaneous abortion | MR-Egger | 5 | 0.36562849 | -0.001908907 | 3.43E-03 |
| Follistatin levels | Spontaneous abortion | Weighted median | 5 | 0.03961415 | -0.001535967 | 3.21E-03 |
| Follistatin levels | Spontaneous abortion | Inverse variance weighted | 5 | 0.03517563 | -0.001282787 | 6.98E-04 |
| Follistatin levels | Spontaneous abortion | Simple mode | 5 | 0.09652113 | -0.002171829 | 7.87E-04 |
| Follistatin levels | Spontaneous abortion | Weighted mode | 5 | 0.10618687 | -0.002087265 | 1.55E-04 |

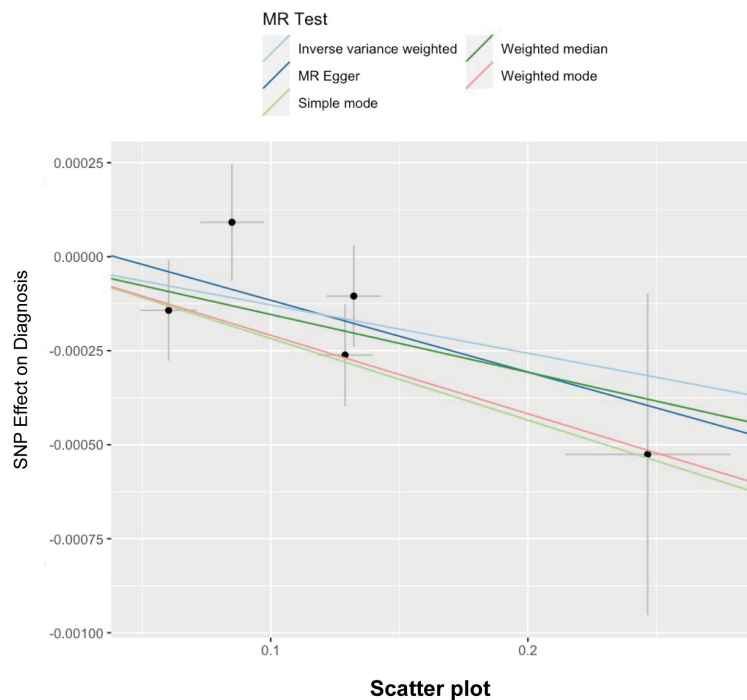


FIGURE 2

Scatter plot illustrating the distribution of individual ratio estimates of follistatin levels with spontaneous abortion as the outcome. Trend lines derived from five different 2SMR methods are also included in each scatter plot to indicate cause and effect.

were generally reliable as the sensitivity analysis strongly supports. These findings have important implications for understanding the pathogenesis of spontaneous abortion and may contribute to the development of potential preventive and therapeutic strategies.

It is widely acknowledged that establishing proper placentation involves sequential processes, notably trophoblast invasion and angiogenesis (48). The orchestrated interplay of angiogenic processes and steroid hormones induces transformative changes in the endometrium, facilitating its receptivity to the blastocyst and initiating the implantation process (49). Successful placentation and the commencement of pregnancy hinge on prerequisites such as endometrial angiogenesis, decidualization, and trophoblast invasion (48, 50, 51). Any dysregulation during the above processes may lead to pregnancy failure and RPL. High levels of anti-annexin V antibody can bind to trophoblast cells, affecting trophoblast invasiveness and causing defective placentation (52). Also, lots of proteins and related pathways have been recognized to be involved in regulating trophoblast function, such as TGF- β which governs the differentiation program of extravillous trophoblasts in the developing human placenta (53).

The underlying biological mechanisms linking low follistatin levels with spontaneous abortion warrant further investigation.

Follistatin not only has an inhibitory effect on FSH secretion from cultured anterior pituitary cells (54) but also is involved in trophoblast invasion and embryonic development. FST acts as an antagonist to the TGF- β superfamily and thereby modulates important signaling pathways such as JNK signaling, ALK4-SMAD2/3-SMAD4 signaling, and FST-GDF11-Smad2/3, further affecting trophoblast function (55). It is hypothesized that reduced follistatin levels may disrupt multiple pathways such as JNK signaling, ALK4-SMAD2/3-SMAD4 signaling, and FST-GDF11-Smad2/3, leading to trophoblast dysfunction and causing impaired implantation and defective placental development, ultimately resulting in abortion. Future research should focus on elucidating the specific molecular mechanisms through which follistatin influences pregnancy outcomes.

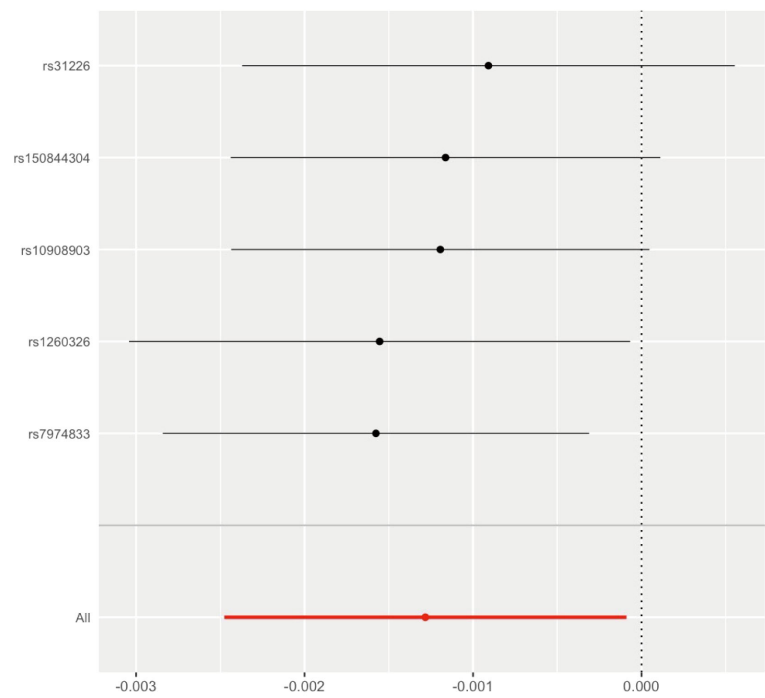
Our results are consistent with previous studies that have reported a probable association between low follistatin levels and adverse pregnancy outcomes (30, 56–58). For instance, Prakash et al. (30) observed a dramatic decrease of FST expression in endometrial stromal cells of women with spontaneous abortion. However, it should be noted that some studies reported no significant decrease of follistatin in the serum of women with spontaneous abortion (59, 60). These discrepancies may arise

TABLE 4 Heterogeneity statistics.

| Method | Q | Q_df | Q_pval |
|---------------------------|----------|------|-----------|
| MR-Egger | 2.738052 | | 0.4337995 |
| Inverse variance weighted | 2.875518 | | 0.5788685 |

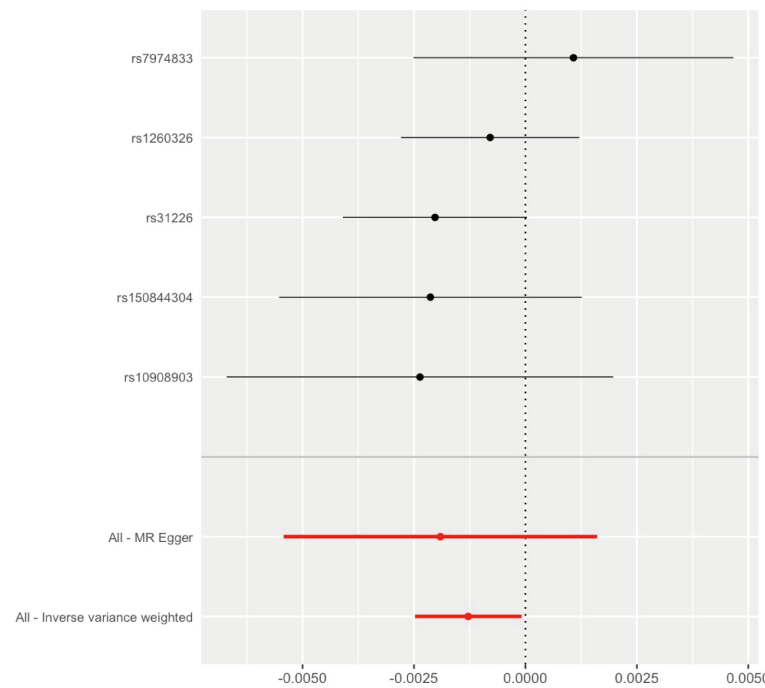
TABLE 5 Pleiotropy statistics of MR analysis.

| Method | Egger regression of intercept | SE | p value |
|----------|-------------------------------|-------------|-----------|
| MR-Egger | 7.51E-05 | 0.000202431 | 0.7354456 |



MR Leave-one-out sensitivity test

FIGURE 3
Leave-one-out analysis for follistatin levels on spontaneous abortion. The given dark dots indicate effect measures from IVW-MR analysis excluding specific SNPs. Red line indicates pooled analysis including all SNPs by the IVW-MR method (plotted for comparison).



MR effect size for follistatin level on miscarriage

FIGURE 4
Forest plot showing the causal effect of each single SNP on the risk of miscarriage.

from variations in sample size and characteristics, since it only includes around 10 abortion samples and 10 control samples.

Although previous studies have pointed out a possible association between FST and spontaneous abortion, there is no absolute evidence on a genetic aspect. Establishing a direct causal link between follistatin and spontaneous abortion presents challenges due to confounding effects from factors such as lifestyle and environmental influences. To overcome this challenge, a promising approach is the utilization of MR, which leverages genetic variants as IVs to infer causal relationships. In our study, we utilized MR to present novel evidence that demonstrates a causal relationship between low FST levels and spontaneous abortion. This finding holds significant implications for the field of reproductive health. Our results suggest that interventions targeting increased FST levels may have the potential to reduce the incidence of spontaneous abortion, which is a devastating outcome for numerous couples. However, despite the valuable insights gained from this study, several limitations should be acknowledged. We recognize that our analysis only included the European population, introducing the possibility of potential bias associated with differing ancestries.

Our study yields meaningful clinical implications. First, it indicates that FST levels might be integrated into routine antenatal assessments to evaluate the risk of pregnancy failure. This is particularly crucial for individuals with a history of spontaneous abortion, where assessing their FST levels may serve as a predictive indicator for the occurrence of RPL. Concurrently, FST may function as a prospective biomarker for targeted interventions like hormonal therapies or lifestyle adjustments and the development of personalized medical strategies. Future studies could explore interventions aimed at modulating follistatin levels to potentially prevent or mitigate the risk of spontaneous abortion. Moreover, investigations into the long-term effects of low follistatin levels on maternal and offspring health outcomes would be valuable for a comprehensive understanding of the implications.

In conclusion, our study contributes to the growing body of reliable evidence supporting the critical role of FST in successful pregnancy outcomes and highlights it as a promising therapeutic target. We remain hopeful that further research, conducted with larger sample sizes based on our observations, will provide additional insights into the underlying mechanisms that link FST and pregnancy outcomes.

Data availability statement

This study analyzed the exposure data acquired from EBI database and the outcome data sourced from the UK Biobank.

References

- Griebel CP, Halvorsen J, Golemon TB, Day AA. Management of spontaneous abortion. *Am Fam Physician*. (2005) 72(7):1243–50.
- Macklon NS, Geraedts JP, Fauser BC. Conception to ongoing pregnancy: the 'black box' of early pregnancy loss. *Hum Reprod Update*. (2002) 8(4):333–43. doi: 10.1093/humupd/8.4.333

Both datasets are publicly accessible and can be found on the IEU Open GWAS Project website (<https://gwas.mrcieu.ac.uk/>).

Author contributions

CG: Formal analysis, Investigation, Methodology, Writing – original draft, Writing – review & editing. WY: Methodology, Writing – review & editing. XL: Writing – review & editing. XLL: Writing – review & editing. YW: Validation, Writing – review & editing. CT: Conceptualization, Methodology, Supervision, Writing – review & editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This project was supported by the Natural Science Foundation of China (82171654) and the Clinical Medicine Plus X-Young Scholars Project of Peking University (Grant PKU2022LCXQ025).

Acknowledgments

We would like to express our sincere gratitude to our team members for their invaluable support and contributions to this research project. Their dedication and expertise have been instrumental in the successful completion of this work.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

- Henzel MJ, Wei Y, Mancini MA, Van Hooser A, Ranalli T, Brinkley BR, et al. Mitosis-specific phosphorylation of histone H3 initiates primarily within pericentromeric heterochromatin during G2 and spreads in an ordered fashion coincident with mitotic chromosome condensation. *Chromosoma*. (1997) 106(6):348–60. doi: 10.1007/s004120050256

4. Ruderman RS, Yilmaz BD, McQueen DB. Treating the couple: how recurrent pregnancy loss impacts the mental health of both partners. *Fertil Steril*. (2020) 114(6):1182. doi: 10.1016/j.fertnstert.2020.09.165
5. Kiwi R. Recurrent pregnancy loss: evaluation and discussion of the causes and their management. *Cleve Clin J Med* (2006) 73(10):913–21. doi: 10.3949/cjcm.73.10.913
6. Propst AM, Hill JA 3rd. Anatomic factors associated with recurrent pregnancy loss. *Semin Reprod Med* (2000) 18(4):341–50. doi: 10.1055/s-2000-13723
7. Rock JA, Murphy AA. Anatomic abnormalities. *Clin Obstet Gynecol*. (1986) 29(4):886–911. doi: 10.1097/00003081-198612000-00015
8. Amrane S, McConnell R. Endocrine causes of recurrent pregnancy loss. *Semin Perinatol*. (2019) 43(2):80–3. doi: 10.1053/j.semperi.2018.12.004
9. Ticconi C, Pietropolli A, Di Simone N, Piccione E, Fazleabas A. Endometrial immune dysfunction in recurrent pregnancy loss. *Int J Mol Sci* (2019) 20(21):5332. doi: 10.3390/ijms20215332
10. Caruso A, De Carolis S, Di Simone N. Antiphospholipid antibodies in obstetrics: new complexities and sites of action. *Hum Reprod Update*. (1999) 5(3):267–76. doi: 10.1093/humupd/5.3.267
11. Di Simone N, De Spirito M, Di Nicuolo F, Tersigni C, Castellani R, Silano M, et al. Potential new mechanisms of placental damage in celiac disease: anti-transglutaminase antibodies impair human endometrial angiogenesis. *Biol Reprod* (2013) 89(4):88. doi: 10.1095/biolreprod.113.109637
12. Di Simone N, Riccardi P, Maggiano N, Piacentani A, D'Asta M, Capelli A, et al. Effect of folic acid on homocysteine-induced trophoblast apoptosis. *Mol Hum Reprod* (2004) 10(9):665–9. doi: 10.1093/molehr/gah091
13. Lin SY, Morrison JR, Phillips DJ, de Kretser DM. Regulation of ovarian function by the TGF-beta superfamily and follistatin. *Reproduction*. (2003) 126(2):133–48. doi: 10.1530/rep.0.1260133
14. Perakakis N, Upadhyay J, Ghaly W, Chen J, Chrysafi P, Anastasilakis AD, et al. Regulation of the activins-follistatins-inhibins axis by energy status: Impact on reproductive function. *Metabolism*. (2018) 85:240–9. doi: 10.1016/j.metabol.2018.05.003
15. Rajput SK, Lee K, Zhenhua G, Di L, Folger JK, Smith GW. Embryotropic actions of follistatin: paracrine and autocrine mediators of oocyte competence and embryo developmental progression. *Reprod Fertil Dev* (2013) 26(1):37–47.
16. Thompson TB, Lerch TF, Cook RW, Woodruff TK, Jardetzky TS. The structure of the follistatin:activin complex reveals antagonism of both type I and type II receptor binding. *Dev Cell* (2005) 9(4):535–43. doi: 10.1016/j.devcel.2005.09.008
17. Nakamura T, Takio K, Eto Y, Shibai H, Titani K, Sugino H. Activin-binding protein from rat ovary is follistatin. *Science*. (1990) 247(4944):836–8. doi: 10.1126/science.2106159
18. Rabinovici J, Spencer SJ, Doldi N, Goldsmith PC, Schwall R, Jaffe RB. Activin-A as an intraovarian modulator: actions, localization, and regulation of the intact dimer in human ovarian cells. *J Clin Invest*. (1992) 89(5):1528–36. doi: 10.1172/JCI115745
19. Rabinovici J, Spencer SJ, Jaffe RB. Recombinant human activin-A promotes proliferation of human luteinized preovulatory granulosa cells in vitro. *J Clin Endocrinol Metab* (1990) 71(5):1396–8. doi: 10.1210/jcem-71-5-1396
20. Li W, Yuen BH, Leung PC. Inhibition of progesterin accumulation by activin-A in human granulosa cells. *J Clin Endocrinol Metab* (1992) 75(1):285–9. doi: 10.1210/jc.75.1.285
21. Li W, Khorasheh S, Yuen BH, Ling N, Leung PC. Stimulation of progesterone secretion by recombinant follistatin-288 in human granulosa cells. *Endocrinology*. (1993) 132(4):1750–6. doi: 10.1210/endo.132.4.8384994
22. Peng C, Ohno T, Khorasheh S, Leung PCK. Activin and follistatin as local regulators in the human ovary. *Neurosignals*. (1996) 5(2):81–9. doi: 10.1159/000109177
23. Li Y, Klausen C, Zhu H, Leung PC. Activin A increases human trophoblast invasion by inducing SNAIL-mediated MMP2 up-regulation through ALK4. *J Clin Endocrinol Metab* (2015) 100(11):E1415–27. doi: 10.1210/jc.2015-2134
24. Sun F, Cheng L, Guo L, Su S, Li Y, Yan J. Activin A promotes human trophoblast invasion by upregulating integrin beta3 via ALK4-SMAD4 signaling. *Placenta*. (2022) 129:62–9. doi: 10.1016/j.placenta.2022.10.004
25. Zhu S, Li Z, Cui L, Ban Y, Leung PCK, Li Y, et al. Activin A increases human trophoblast invasion by upregulating integrin beta1 through ALK4. *FASEB J* (2021) 35(2):e21220. doi: 10.1096/fj.202001604R
26. Li Y, Klausen C, Cheng JC, Zhu H, Leung PC. Activin A, B, and AB increase human trophoblast cell invasion by up-regulating N-cadherin. *J Clin Endocrinol Metab* (2014) 99(11):E2216–25. doi: 10.1210/jc.2014-2118
27. Koninger A, Schmidt B, Damaske D, Birdir C, Eneke A, Kimmig R, et al. Follistatin during pregnancy and its potential role as an ovarian suppressing agent. *Eur J Obstet Gynecol Reprod Biol* (2017) 212:150–4. doi: 10.1016/j.ejogrb.2017.03.001
28. Li J, Qi Y, Yang K, Zhu L, Cui X, Liu Z. Follistatin is a novel chemoattractant for migration and invasion of placental trophoblasts of mice. *Cells*. (2022) 11(23):3816. doi: 10.3390/cells11233816
29. Liu G, Qi Y, Wu J, Lin F, Liu Z, Cui X. Follistatin is a crucial chemoattractant for mouse decidualized endometrial stromal cell migration by JNK signalling. *J Cell Mol Med* (2023) 27(1):127–40. doi: 10.1111/jcmm.17648
30. Prakash A, Li TC, Tuckerman E, Laird S, Wells M, Ledger WL. A study of luteal phase expression of inhibin, activin, and follistatin subunits in the endometrium of women with recurrent miscarriage. *Fertil Steril*. (2006) 86(6):1723–30. doi: 10.1016/j.fertnstert.2006.05.040
31. Fullerton PT Jr., Monsivais D, Kommagani R, Matzuk MM. Follistatin is critical for mouse uterine receptivity and decidualization. *Proc Natl Acad Sci U S A*. (2017) 114(24):E4772–E81. doi: 10.1073/pnas.1620903114
32. Mdallase S, Moodley J, Naicker T. The role of follistatin and granulocyte-colony stimulating factor in HIV-associated pre-eclampsia. *Pregnancy Hypertens* (2020) 19:81–6. doi: 10.1016/j.preghy.2019.12.012
33. Zhao L, Shang T, Wang YL, Tang S, Li H, Liu ZH. [Study on the relationship between activin A, follistatin and preeclampsia]. *Zhonghua Fu Chan Ke Za Zhi*. (2003) 38(11):676–9.
34. Garces MF, Vallejo SA, Sanchez E, Palomino-Palomino MA, Leal LG, Angel-Muller E, et al. Longitudinal analysis of maternal serum Follistatin concentration in normal pregnancy and preeclampsia. *Clin Endocrinol (Oxf)*. (2015) 83(2):229–35. doi: 10.1111/cen.12715
35. Li H, Zhou L, Zhang C, Xi Q, Lv J, Huo W, et al. Follistatin dysregulation impaired trophoblast biological functions by GDF11-Smad2/3 axis in preeclampsia placentas. *Placenta*. (2022) 121:145–54. doi: 10.1016/j.placenta.2022.03.015
36. Smith GD, Ebrahim S. 'Mendelian randomization': can genetic epidemiology contribute to understanding environmental determinants of disease? *Int J Epidemiol* (2003) 32(1):1–22. doi: 10.1093/ije/dyg070
37. Davies NM, Holmes MV, Davey Smith G. Reading Mendelian randomisation studies: a guide, glossary, and checklist for clinicians. *BMJ* (2018) 362:k601. doi: 10.1136/bmj.k601
38. Zheng J, Baird D, Borges MC, Bowden J, Hemani G, Haycock P, et al. Recent developments in mendelian randomization studies. *Curr Epidemiol Rep* (2017) 4(4):330–45. doi: 10.1007/s40471-017-0128-6
39. Emdin CA, Khera AV, Kathiresan S. Mendelian randomization. *JAMA*. (2017) 318(19):1925–6. doi: 10.1001/jama.2017.17219
40. Folkersen L, Gustafsson S, Wang Q, Hansen DH, Hedman AK, Schork A, et al. Genomic and drug target evaluation of 90 cardiovascular proteins in 30,931 individuals. *Nat Metab* (2020) 2(10):1135–48. doi: 10.1038/s42255-020-00287-2
41. Hemani G, Zheng J, Elsworth B, Wade KH, Haberland V, Baird D, et al. The MR-Base platform supports systematic causal inference across the human phenome. *Elife* (2018) 7:e34408. doi: 10.7554/eLife.34408
42. Bowden J, Del Greco MF, Minelli C, Davey Smith G, Sheehan NA, Thompson JR. Assessing the suitability of summary data for two-sample Mendelian randomization analyses using MR-Egger regression: the role of the I² statistic. *Int J Epidemiol*. (2016) 45(6):1961–74. doi: 10.1093/ije/dyw220
43. Staley JR, Blackshaw J, Kamat MA, Ellis S, Surendran P, Sun BB, et al. PhenoScanner: a database of human genotype-phenotype associations. *Bioinformatics*. (2016) 32(20):3207–9. doi: 10.1093/bioinformatics/btw373
44. Kamat MA, Blackshaw JA, Young R, Surendran P, Burgess S, Danesh J, et al. PhenoScanner V2: an expanded tool for searching human genotype-phenotype associations. *Bioinformatics*. (2019) 35(22):4851–3. doi: 10.1093/bioinformatics/btz469
45. Hemani G, Bowden J, Davey Smith G. Evaluating the potential role of pleiotropy in Mendelian randomization studies. *Hum Mol Genet* (2018) 27(R2):R195–208. doi: 10.1093/hmg/ddy163
46. Sudlow G, Gallacher J, Allen N, Beral V, Burton P, Danesh J, et al. UK biobank: an open access resource for identifying the causes of a wide range of complex diseases of middle and old age. *PLoS Med* (2015) 12(3):e1001779. doi: 10.1371/journal.pmed.1001779
47. Sproverio W, Winchester L, Newby D, Fernandes M, Shi L, Goodday SM, et al. High blood pressure and risk of dementia: A two-sample mendelian randomization study in the UK biobank. *Biol Psychiatry* (2021) 89(8):817–24. doi: 10.1016/j.biopsych.2020.12.015
48. Huppertz B, Peeters LL. Vascular biology in implantation and placentation. *Angiogenesis*. (2005) 8(2):157–67. doi: 10.1007/s10456-005-9007-8
49. Di Simone N, D'Ippolito S, Marana R, Di Nicuolo F, Castellani R, Pierangeli SS, et al. Antiphospholipid antibodies affect human endometrial angiogenesis: protective effect of a synthetic peptide (TIF1) mimicking the phospholipid binding site of beta(2) glycoprotein I. *Am J Reprod Immunol* (2013) 70(4):299–308. doi: 10.1111/aji.12130
50. Cullinan-Bove K, Koos RD. Vascular endothelial growth factor/vascular permeability factor expression in the rat uterus: rapid stimulation by estrogen correlates with estrogen-induced increases in uterine capillary permeability and growth. *Endocrinology*. (1993) 133(2):829–37. doi: 10.1210/endo.133.2.8344219
51. Reynolds LP, Borowicz PP, Caton JS, Vonnahme KA, Luther JS, Buchanan DS, et al. Uteroplacental vascular development and placental function: an update. *Int J Dev Biol* (2010) 54(2-3):355–66. doi: 10.1387/ijdb.082799lr
52. Di Simone N, Castellani R, Caliendo D, Caruso A. Monoclonal anti-annexin V antibody inhibits trophoblast gonadotropin secretion and induces syncytiotrophoblast apoptosis. *Biol Reprod* (2001) 65(6):1766–70. doi: 10.1095/biolreprod65.6.1766
53. Haider S, Lackner AI, Dietrich B, Kunihs V, Haslinger P, Meinhardt G, et al. Transforming growth factor-beta signaling governs the differentiation program of extravillous trophoblasts in the developing human placenta. *Proc Natl Acad Sci U S A*. (2022) 119(28):e2120667119. doi: 10.1073/pnas.2120667119
54. Yokoyama Y, Nakamura T, Nakamura R, Irahara M, Aono T, Sugino H. Identification of activins and follistatin proteins in human follicular fluid and placenta. *J Clin Endocrinol Metab* (1995) 80(3):915–21. doi: 10.1210/jcem.80.3.7883850

55. Goldman-Wohl D, Yagel S. Regulation of trophoblast invasion: from normal implantation to pre-eclampsia. *Mol Cell Endocrinol* (2002) 187(1-2):233–8. doi: 10.1016/S0303-7207(01)00687-6
56. Prakash A, Laird S, Tuckerman E, Li TC, Ledger WL. Inhibin A and activin A may be used to predict pregnancy outcome in women with recurrent miscarriage. *Fertil Steril*. (2005) 83(6):1758–63. doi: 10.1016/j.fertnstert.2004.11.072
57. Daponte A, Deligeoroglou E, Garas A, Pournaras S, Hadjichristodoulou C, Messinis IE. Activin A and follistatin as biomarkers for ectopic pregnancy and missed abortion. *Dis Markers*. (2013) 35(5):497–503. doi: 10.1155/2013/969473
58. Lin SY, Craythorn RG, O'Connor AE, Matzuk MM, Girling JE, Morrison JR, et al. Female infertility and disrupted angiogenesis are actions of specific follistatin isoforms. *Mol Endocrinol* (2008) 22(2):415–29. doi: 10.1210/me.2006-0529
59. Muttukrishna S, Jauniaux E, Greenwold N, McGarrigle H, Jivraj S, Carter S, et al. Circulating levels of inhibin A, activin A and follistatin in missed and recurrent miscarriages. *Hum Reprod* (2002) 17(12):3072–8. doi: 10.1093/humrep/17.12.3072
60. Muttukrishna S, Bearfield C, Johns J, Jauniaux E. Inhibin, activin, follistatin, activin receptors and beta-glycan gene expression in the villous tissue of miscarriage patients. *Mol Hum Reprod* (2004) 10(11):793–8. doi: 10.1093/molehr/gah110

Frontiers in Endocrinology

Explores the endocrine system to find new therapies for key health issues

The second most-cited endocrinology and metabolism journal, which advances our understanding of the endocrine system. It uncovers new therapies for prevalent health issues such as obesity, diabetes, reproduction, and aging.

Discover the latest Research Topics

[See more →](#)

Frontiers

Avenue du Tribunal-Fédéral 34
1005 Lausanne, Switzerland
frontiersin.org

Contact us

+41 (0)21 510 17 00
frontiersin.org/about/contact

