

Reviews in cardiovascular pharmacology 2023

Edited by

Yusof Kamisah, Mas Rizky A. A. Syamsunarno
and Ismail Laher

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Reviews in cardiovascular pharmacology: 2023

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Table of contents

| | |
|-----|---|
| 05 | Editorial: Reviews in cardiovascular pharmacology: 2023 Yusuf Kamisah, Ismail Laher and Mas Rizky A. A. Syamsunarno |
| 08 | Infection with <i>Helicobacter pylori</i> may predispose to atherosclerosis: role of inflammation and thickening of intima-media of carotid arteries Karl Aramouni, Roland K. Assaf, Maria Azar, Karen Jabbour, Abdullah Shaito, Amirhossein Sahebkar, Assaad A. Eid, Manfredi Rizzo and Ali H. Eid |
| 20 | Inclisiran: a new generation of lipid-lowering siRNA therapeutic Yanzhen Zhang, Huaigang Chen, Lang Hong, Hong Wang, Bin Li, Mengyin Zhang, Jiamei Li, Liu Yang and Fan Liu |
| 29 | Anti-atherosclerosis mechanisms associated with regulation of non-coding RNAs by active monomers of traditional Chinese medicine Guoqing Liu, Liqiang Tan, Xiaona Zhao, Minghui Wang, Zejin Zhang, Jing Zhang, Honggang Gao, Meifang Liu and Wei Qin |
| 46 | Decoding signaling mechanisms: unraveling the targets of guanylate cyclase agonists in cardiovascular and digestive diseases Qinan Yin, Xingyue Zheng, Yujie Song, Liuyun Wu, Lian Li, Rongsheng Tong, Lizhu Han and Yuan Bian |
| 70 | The pathogenesis and therapeutic strategies of heat stroke-induced myocardial injury Rui Xia, Meng Sun, Yuling Li, Jing Yin, Huan Liu, Jun Yang, Jing Liu, Yanyu He, Bing Wu, Guixiang Yang and Jianhua Li |
| 78 | Microvascular rarefaction caused by the NOTCH signaling pathway is a key cause of TKI-apatinib-induced hypertension and cardiac damage WenJuan Wang, Guodong Li, Jie Ma, Xin Fan, Jianzhong Lu, Qiyin Sun, Jiafang Yao and Qingjian He |
| 89 | Shenfu injection: a review of pharmacological effects on cardiovascular diseases Fei-Fei Xu, Xiao-Fang Xie, Hai-Yan Hu, Rong-Sheng Tong and Cheng Peng |
| 100 | The protective role of ginsenoside Rg3 in heart diseases and mental disorders Lili Shi, Jinlan Luo, Xiupan Wei, Xizhen Xu and Ling Tu |
| 116 | Non-coding RNAs: targets for Chinese herbal medicine in treating myocardial fibrosis Minghui Wang, Maocai Yan, Liqiang Tan, Xiaona Zhao, Guoqing Liu, Zejin Zhang, Jing Zhang, Honggang Gao and Wei Qin |

- 132 **Adenylyl cyclase isoforms 5 and 6 in the cardiovascular system: complex regulation and divergent roles**
Saeid Maghsoudi, Rabia Shuaib, Ben Van Bastelaere and Shyamala Dakshinamurti
- 155 **NLRP3 inflammasome and pyroptosis in cardiovascular diseases and exercise intervention**
Ping Ding, Yuanming Song, Yang Yang and Cheng Zeng
- 172 **Amentoflavone for treating cardiocerebrovascular diseases and neurological disorders**
Hang Zhang, Yin-mei Ban, De-mei Li, Gang Wang, Juan Gu and Lei Zhu



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Editorial: Reviews in cardiovascular pharmacology: 2023

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KEYWORDS

cardiac, vascular, fibrosis, atherosclerosis, inflammation, oxidative stress

Editorial on the Research Topic
[Reviews in cardiovascular pharmacology: 2023](#)

1 Introduction

Cardiovascular disease remains a leading cause of death globally, accounting for approximately 17.9 million fatalities in 2019 (World Health Organization, 2021). Cardiovascular diseases include hypertension, atherosclerosis, ischemic heart disease, stroke and heart failure, and represent a significant socioeconomic burden. The Research Topic “Reviews in Cardiovascular Pharmacology: 2023” provides an overview of both the pathogenesis and pharmacological advances in cardiovascular diseases. This Research Topic features 12 articles, each offering in-depth discussions of recent findings on the mechanisms underlying cardiovascular diseases, along with the development and application of novel cardiovascular therapies. The articles cover a wide range of topics that include recent advancements in clinical trials, emerging concepts in drug mechanisms, therapeutic strategies, and challenges related to pharmacokinetics.

Nitric oxide (NO) is a highly reactive gaseous molecule released by endothelial cells in blood vessels that plays a crucial role in mediating protective cardiovascular effects, for instance vasodilation (Siti et al., 2019). Impaired NO function often occurs before the clinical onset of cardiovascular (Abd-Elmoniem et al., 2024) disease. This endogenous vasodilator binds to soluble guanylate cyclase (sGC), stimulating the synthesis of cyclic guanosine monophosphate (cGMP). Elevated cGMP activates protein kinase G (PKG), which lowers intracellular calcium levels in vascular smooth muscle cells and induces vasodilation (Mishra et al., 2025). Yin et al. reviewed the progress of guanylate cyclase activators to stimulate the NO-sGC-cGMP signaling pathway in patients with cardiovascular disease. These include riociguat, vericiguat, praliciguat, olinciguat, cinaciguat, ataciguat, runcaciguat, mosliciguat, and BI 685509, the latter still undergoing clinical trials. Therapies such as riociguat and vericiguat show benefits such as improved quality of life in patients with heart failure and pulmonary hypertension, although definitive

conclusions for other agents are not yet available. Additionally, Yin et al. discussed the pharmacology of guanylate cyclase-C (GC-C) agonists (linaclotide and plecanatide) for the treatment of digestive conditions. These agents improve symptoms of irritable bowel syndrome with constipation (IBS-C) by activating the GC-C/cGMP pathway, reducing submucosal afferent neuron excitation and reducing abdominal pain.

While guanylate cyclase mediates its effects primarily through cGMP to cause vasodilation and smooth muscle relaxation (Siti et al., 2015), adenylyl cyclase produces cAMP to regulate diverse functions, such as metabolism, heart rate, and neurotransmitter release (Marsden and Dessauer 2019). Adenylyl cyclase isoforms 5 (AC5) and 6 (AC6) are highly expressed in cardiac tissues, where they are important in translating signals from β -adrenergic receptors into intracellular cAMP production (Marsden and Dessauer 2019). Maghsoudi et al. highlighted the distinct regulatory mechanisms and physiological roles of these isoforms, particularly in calcium handling, myocardial contractility, and adaptive responses to catecholamine stimulation. AC5 plays a key role in cardiac and vascular function, and its inhibition has cardioprotective effects. In contrast, AC6 plays a significant role in vasodilation and is particularly enriched in neonatal tissues, with its overexpression enhancing cardiac repair. The potential of targeting these isoforms for therapeutic intervention, especially in conditions such as heart failure where dysregulated β -adrenergic signaling contributes to disease progression, is also discussed.

Atherosclerosis is a significant underlying cause of cardiovascular disease. It is a chronic, progressive condition marked by the accumulation of plaques composed of lipids, cholesterol, calcium, and cellular debris within the walls of arteries (Wang et al., 2021). Inclisiran is a novel cholesterol-lowering drug with promising pharmacological properties, as described by Zhang et al. The drug utilizes small interfering RNA (siRNA) to silence the expression of the proprotein convertase subtilisin/kexin type 9 (PCSK9) gene, which encodes a protein responsible for degrading low-density lipoprotein (LDL) receptors in the liver. By inhibiting PCSK9 expression, inclisiran increases the availability of LDL receptors on hepatic cells, thereby enhancing the clearance of LDL cholesterol from the bloodstream. This leads to a sustained reduction in LDL cholesterol levels, as demonstrated in clinical trials, thereby reducing the risk of atherosclerosis progression.

Inflammation is a key contributor to the pathogenesis of many cardiovascular diseases such as atherosclerosis. The NOD-like receptor protein 3 (NLRP3) inflammasome is an inflammatory mediator activated in cardiovascular diseases. Upon activation, NLRP3 triggers pyroptosis, a form of programmed cell death characterized by inflammatory cell death and the release of pro-inflammatory cytokines (Zhang et al., 2023). Ding et al. comprehensively discussed the inhibitory effects of exercise on NLRP3 expression and pyroptosis in atherosclerosis, obesity, diabetic cardiomyopathy, myocardial infarction, hypertension, and heart failure. They demonstrated that NLRP3 plays a critical role in the pathogenesis of cardiovascular disease. In addition to these molecular mechanisms, microbial infections have also been implicated in cardiovascular disease. Aramouni et al. suggested that *Helicobacter pylori* infection may contribute to atherogenesis by promoting foam cell formation and triggering a chronic immune

response. These findings underscore the multifaceted nature of inflammation in cardiovascular disease, spanning both molecular and infectious contributions.

Traditional Chinese Medicine (TCM) has emerged as a significant source of alternative and complementary therapies, offering bioactive compounds with therapeutic potential against various diseases. Bioactive monomers extracted from TCM, such as geniposide, astragaloside IV, genkwanin, and tanshinone IIA, target non-coding RNAs to alleviate pathological processes associated with atherosclerosis. These non-coding RNAs modulate mechanisms underlying atherosclerosis, for instance inflammation, oxidative stress, adipogenesis, apoptosis, and autophagy (Liu et al.). In addition to their role in atherosclerosis, non-coding RNAs are also implicated in broader cardiovascular diseases. Active monomers such as quercetin, tripterine, notoginsenoside R1, and berberine—also derived from TCM—reduce myocardial fibrosis effects by modulating non-coding RNAs (Wang et al.). These findings emphasize the broad therapeutic potential of targeting non-coding RNAs in the treatment of various cardiovascular conditions.

In the context of cardiovascular disease, Shenfu, a TCM formulation derived from ginseng (*Panax ginseng* C.A. Mey) and aconite (*Aconitum carmichaelii* Debeaux), may have therapeutic benefits. Injecting Shenfu improves cardiac function in patients diagnosed with myocardial infarction, cardiac arrest following resuscitation, and heart failure (Xu et al.). In animal models of cardiovascular disease, it attenuates oxidative stress and inflammation by downregulating the nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B) signaling pathway. Additionally, it reduces apoptosis and fibrosis by modulating the transforming growth factor-beta (TGF- β)/Smads signaling pathway. Moreover, ginsenoside Rg3, extracted from *P. ginseng*, has shown promising effects in mitigating heart failure by reducing inflammation, oxidative stress, apoptosis, and fibrosis in various animal models of heart disease and mental illness (Shi et al.).

Zhang et al. conducted a comprehensive review on amentoflavone, a flavonoid with pharmacological potential for treating neurological disorders and cardiocerebrovascular diseases by mitigating inflammation, oxidative stress, and reducing lipid levels. They highlight the challenges in improving its pharmacokinetic profile due to its low solubility and poor oral bioavailability. The study by Wang et al. examined the challenges associated with tyrosine kinase inhibitors, a class of tumor-targeted therapies that includes apatinib. These drugs can cause microcirculatory rarefaction that impairs microvascular growth. The authors hypothesized that inhibiting the Notch signaling pathway may mitigate the microvascular changes induced by tyrosine kinase inhibitors.

Heat stroke can cause myocardial injury by disrupting electrolytes, particularly those associated with the sodium-potassium pump (Wang et al., 2019). Xia et al. explored the pathogenesis of heat stroke-induced myocardial injury, highlighting hypercytokinemia as a result of increased inflammation, metabolic abnormalities, and protein dysregulation. These changes contribute to endothelial dysfunction, circulatory shock, and ultimately cardiomyocyte death. The article also discusses several strategies for the management of heat stroke.

In conclusion, this Research Topic presents a series of articles that explore various aspects of cardiovascular health. It provides valuable insights into the molecular mechanisms underlying the pathogenesis of cardiovascular disease along with the protective effects of different cardiovascular drugs. Collectively, these articles serve as a vital resource for researchers, clinicians, and healthcare professionals striving to improve patient outcomes in cardiovascular care.

Author contributions

YK: Writing—original draft, Writing—review and editing. IL: Writing—review and editing. MS: Writing—review and editing.

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References

- Abd-Elmoniem, K. Z., Edwan, J. H., Dietsche, K. B., Villalobos-Perez, A., Shams, N., Matta, J., et al. (2024). Endothelial dysfunction in youth-onset type 2 diabetes: A clinical translational study. *Circ. Res.* 135 (6), 639–650. doi:10.1161/CIRCRESAHA.124.324272
- Laksono, S., and Kusharsamita, H. (2024). Unravelling the role of carotid atherosclerosis in predicting cardiovascular disease risk: A review. *ARYA Atheroscler* 20 (5), 52–59. doi:10.48305/arya.2024.41271.2862
- Marsden, A. N., and Dessauer, C. W. (2019). Nanometric targeting of type 9 adenylyl cyclase in heart. *Biochem. Soc. Trans.* 47 (6), 1749–1756. doi:10.1042/BST20190227
- Mishra, S., Chander, V., and Kass, D. A. (2025). Cardiac cGMP regulation and therapeutic applications. *Hypertension* 82 (2), 185–196. doi:10.1161/HYPERTENSIONAHA.124.21709
- Siti, H. N., Kamisah, Y., and Kamsiah, J. (2015). The role of oxidative stress, antioxidants, and vascular inflammation in cardiovascular disease (a review). *Vasc. Pharmacol.* 71, 40–56. doi:10.1016/j.vph.2015.03.005
- Siti, H. N., Kamisah, Y., Mohamed, S., and Jaarin, K. (2019). Effects of citrus leaf extract on aortic vascular reactivity in hypertensive rats fed repeatedly heated vegetable oil. *Appl. Physiol. Nutr. Metab.* 44 (4), 373–380. doi:10.1139/apnm-2018-0175
- Wang, J. C., Chien, W. C., Chu, P., Chung, C. H., Lin, C. Y., and Tsai, S. H. (2019). The association between heat stroke and subsequent cardiovascular diseases. *Plos One* 14, e0211386. doi:10.1371/journal.pone.0211386
- Wang, R., Wang, M., Ye, J., Sun, G., and Sun, X. (2021). Mechanism overview and target mining of atherosclerosis: Endothelial cell injury in atherosclerosis is regulated by glycolysis (Review). *Int. J. Mol. Med.* 47 (1), 65–76. doi:10.3892/ijmm.2020.4798
- World Health Organization (2021). Cardiovascular diseases (CVDs). Available at: [https://www.who.int/news-room/fact-sheets/detail/cardiovascular-diseases-\(cvds\)](https://www.who.int/news-room/fact-sheets/detail/cardiovascular-diseases-(cvds)) (Accessed Dec 24, 2024).
- Zhang, X., Wang, Z., Zheng, Y., Yu, Q., Zeng, M., Bai, L., et al. (2023). Inhibitors of the NLRP3 inflammasome pathway as promising therapeutic candidates for inflammatory diseases (Review). *Int. J. Mol. Med.* 51 (4), 35. doi:10.3892/ijmm.2023.5238



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Infection with *Helicobacter pylori* may predispose to atherosclerosis: role of inflammation and thickening of intima-media of carotid arteries

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Atherosclerosis is a major instigator of cardiovascular disease (CVD) and a main cause of global morbidity and mortality. The high prevalence of CVD calls for urgent attention to possible preventive measures in order to curb its incidence. Traditional risk factors of atherosclerosis, like age, smoking, diabetes mellitus, dyslipidemia, hypertension and chronic inflammation, are under extensive investigation. However, these only account for around 50% of the etiology of atherosclerosis, mandating a search for different or overlooked risk factors. In this regard, chronic infections, by *Helicobacter pylori* for instance, are a primary candidate. *H. pylori* colonizes the gut and contributes to several gastrointestinal diseases, but, recently, the potential involvement of this bacterium in extra-gastric diseases including CVD has been under the spotlight. Indeed, *H. pylori* infection appears to stimulate foam cell formation as well as chronic immune responses that could upregulate key inflammatory mediators including cytokines, C-reactive protein, and lipoproteins. These factors are involved in the thickening of intima-media of carotid arteries (CIMT), a hallmark of atherosclerosis. Interestingly, *H. pylori* infection was found to increase (CIMT), which along with other evidence, could implicate *H. pylori* in the pathogenesis of atherosclerosis. Nevertheless, the involvement of *H. pylori* in CVD and atherosclerosis remains controversial as several studies report no connection between *H. pylori* and atherosclerosis. This review examines and critically discusses the evidence that argues for a potential role of this bacterium in atherogenesis. However, additional basic and clinical research studies are warranted to convincingly establish the association between *H. pylori* and atherosclerosis.

KEYWORDS

cardiovascular disease, oxidative stress, extra-gastric disease, CagA, vitamin B12 deficiency

Abbreviations: Cag-A, Cytotoxin-Associated Gene; IL-6, Interleukin 6; IL-8, Interleukin 8; TNF- α , Tumor necrosis factor α ; ICAM-1, Intercellular adhesion molecule 1; CRP, C-Reactive Protein; NGAL, neutrophil gelatinase-associated lipocalin; 8-OHdG, 8-hydroxy-2'-deoxyguanosine.

1 Introduction

Atherosclerosis is precipitated by the pathological formation of plaques within blood vessels. These plaques are instigated by endothelium injury followed by infiltration of immune cells and proliferation of vascular smooth muscle cells. Thickening of the intima and hardening of the vessels then ensue, thus narrowing the vascular lumen which progresses into partial or complete obstruction of blood vessels. Atherosclerosis is a dominant cause and risk factor of cardiovascular disease (CVD), a major instigator of global mortality (Herrington et al., 2016; Libby et al., 2019). Some of the specific risk factors include age, smoking, diabetes mellitus, dyslipidemia, hypertension, and chronic inflammation (Siasos et al., 2014; Spence and Pilote, 2015; Aarabi et al., 2018; Libby et al., 2018; Yao et al., 2019; Lechner et al., 2020). These can account only for around 50% of the incidence of atherosclerosis (Herrington et al., 2016; Libby et al., 2019), making investigation into other risk factors of incidence a hot and rather attractive research area. In recent years, some attention has been given to newly identified potential atherosclerosis risk factors like chronic infections (Pothineni et al., 2017). Possible chronic infectious agents that have been reportedly linked to atherosclerosis include the infamous bacterium *Helicobacter pylori* (Karbasi-Afshar et al., 2015).

H. pylori is a gram-negative, microaerophilic, spiral-shaped bacterium. It is a key microorganism of the human microbiome and a colonizer of the gut of at least half of the world's population (Eusebi et al., 2014; Mentis et al., 2015; Burucoa and Axon, 2017; Leja et al., 2019). In 1994, this bacterium was categorized as a human class I carcinogen by the World Health Organization (Wang, 2014). Not only that, but *H. pylori* infection is the major cause of several gastric diseases including acute and chronic active gastritis, chronic atrophic gastritis, peptic ulcer, gastric adenocarcinoma, and type B low-grade mucosa-associated lymphoid tissue lymphoma (Wang et al., 2014; Wroblewski and Peek, 2016; Chang et al., 2018; de Brito et al., 2019; Sugano, 2019; Ford et al., 2020). Importantly, *H. pylori* infection has been linked to more than 50 extra-gastrointestinal manifestations in a variety of medical specializations such as dermatology, endocrinology, hematology, and cardiology, among others (Campuzano-Maya, 2014; He et al., 2022). In particular, insulin resistance (Polyzos et al., 2011), liver disorders (Waluga et al., 2015), ventilator-associated pneumonia (Dadashi and Hosseinzadeh, 2018), osteoporosis (Mizuno et al., 2015), chronic kidney disease (Hata et al., 2021), hematological manifestations like iron deficiency and vitamin B12 (cobalamin) deficiency, hypertension, and atherosclerosis have been linked to *H. pylori* infection (Tsay and Hsu, 2018; Fang et al., 2019; Franceschi et al., 2019; Mansori et al., 2020a; Mansori et al., 2020b).

Involvement of *H. pylori* in this wide array of diseases may be partly related to the fact that this bacterial infection contributes not only to local inflammation but also to systemic inflammation (Tsay and Hsu, 2018; Fang et al., 2019; Franceschi et al., 2019; Mansori et al., 2020a; Mansori et al., 2020b). In turn, the latter can instigate several extra-gastrointestinal disorders such as metabolic syndrome, diabetes mellitus, insulin resistance, and CVD (Tsay and Hsu, 2018; Fang et al., 2019; Franceschi et al., 2019; Mansori et al., 2020a; Mansori et al., 2020b; Kountouras et al., 2021). Of particular interest, recent studies have found that *H. pylori* may be responsible for the initiation, progression, and complications of atherosclerotic plaque

formation. Hence, this review was undertaken to critically examine the evidence concerning *H. pylori* chronic infection as a risk factor of atherosclerosis.

2 Current evidence

2.1 *H. pylori* infection modifies carotid intima-media thickness (CIMT)

One early indication of atherosclerosis is thickening of the carotid artery. In atherosclerosis, this blood vessel narrows, and blood flow becomes compromised thereby increasing the risk of CVD. Indeed, CIMT, which measures the thickness of the inner two layers (the intima and media) of a carotid artery, is an early diagnostic tool of atherosclerosis as this thickening appears even in asymptomatic pre-atherosclerosis patients. Relatedly, several reports support a positive correlation between *H. pylori* infection and an increase of CIMT (Table 1). Indeed, CIMT values were significantly higher in *H. pylori*-infected versus non-infected subjects, and the levels of *H. pylori*-IgG were positively correlated with the increased CIMT measurements (Shan et al., 2018). Similar results were obtained in *H. pylori*-positive men, younger than 50 years of age, who showed a higher incidence of carotid atherosclerosis than non-*H. pylori*-infected counterparts (Zhang et al., 2019). In addition, subjects without carotid atherosclerosis, but with *H. pylori* infection, were found to have higher CIMT values than those free of the infection (Zhang et al., 2019). Importantly, a follow up of subjects with persistent *H. pylori* infection after 5 years revealed a significantly higher incidence of carotid atherosclerosis compared to subjects who were *H. pylori* negative (Zhang et al., 2019). This clearly shows a potentially causative effect of this bacterial infection with atherosclerosis. Indeed, a cross-sectional study shows a positive association between *H. pylori* infection, CIMT and carotid atherosclerosis, independent of classical risk factors (Zhang et al., 2021). In agreement with this, several meta-analyses show that *H. pylori* infection can significantly increase CIMT and lead to subclinical atherosclerosis (Wang et al., 2021b; Shi et al., 2022; Simon et al., 2022).

One of the key interplayers in the potential association between atherosclerosis and *H. pylori* infection is the cytotoxin-associated gene A (CagA). Indeed, studies involving both the right and left coronary artery of patients who underwent an upper GI endoscopy show higher CIMT values among patients infected with a *H. pylori* strain positive for CagA, compared to patients infected with CagA-negative *H. pylori* (Talari et al., 2021). The CagA-positive group also exhibited higher levels of high-sensitivity C-reactive protein (hsCRP), a marker of elevated inflammatory response (Talari et al., 2021). These studies led to the conclusion that CagA+ *H. pylori* strain can induce a systemic inflammatory response that may contribute to the development of atherosclerosis (Talari et al., 2021). Notably, changes in CIMT values were found to be even more prominent when *H. pylori* infection was coupled with certain comorbidities. For instance, CIMT values were highest among subjects with an *H. pylori* infection and alcoholic liver disease (ALD), compared to other groups with only one or neither of the conditions (Bao-Ge et al., 2017). Additionally, the coexistence of *H. pylori* infection and early-stage diabetic kidney disease (DKD) causes a significant increase in CIMT measurements, and potentiates the risk of developing atherosclerosis in type

TABLE 1 Major evidence correlating *H. pylori* infection with atherosclerosis.

| Evidence | Findings | References |
|--------------------------|--|---|
| CIMT | Positive correlation between high CIMT values and <i>H. pylori</i> infection | Shan et al. (2018), Zhang et al. (2019), Wang et al. (2021b), Talari et al. (2021), Zhang et al. (2021), Shi et al. (2022), Simon et al. (2022) |
| | Correlation exacerbated when <i>H. pylori</i> infection is coupled with certain comorbidities | Bao-Ge et al. (2017), Feng et al. (2018a), Yu et al. (2019), Choi et al. (2022) |
| Dyslipidemia | <i>H. pylori</i> infected subjects have lower levels of HDL and higher levels of LDL | Lee et al. (2018), Abdu et al. (2020), Kim et al. (2016), Liang et al. (2021), Choi et al. (2019), Medrek-Socha et al. (2018), Shan et al. (2017) |
| | <i>H. pylori</i> eradication therapy increases HDL level and restores LDL/HDL ratio | Iwai et al. (2019), Park et al. (2021) |
| Systemic immune response | Chronic inflammation caused by <i>H. pylori</i> infection can generate persistent oxidative stress and modify LDL into oxidized-LDL | Hansson et al. (2006), Sies et al. (2017), Butcher et al. (2017), Gao and Liu (2017), Luo et al. (2017), Krupa et al. (2021) |
| | Serum Ox-LDL and 8-OHdG levels are higher in T2DM patients with <i>H. pylori</i> infection | Nasif et al. (2016) |
| | Potential association between <i>H. pylori</i> , the inflammatory cytokine YKL-40, and atherosclerosis | Cooke et al. (2001), Wu et al. (2004), Martinet et al. (2002), Rathcke and Vestergaard (2006), Xu et al. (2016) |
| Vitamin B12 deficiency | <i>H. pylori</i> -induced atrophic gastritis and reduction in the levels of intrinsic factor protein lead to Vitamin B12 deficiency | Supiano (1987), Kaptan et al. (2000), Carmel et al. (2001), Dierkes et al. (2003), Sipponen et al. (2003), Soyocak et al. (2021) |
| | <i>H. pylori</i> -induced vitamin B12 deficiency is suggested to play a key role in atherosclerosis via several mechanisms one of which is inhibition of the enzyme methionine synthase leading to high production of homocysteine. Hyperhomocysteinemia is implicated in atherosclerosis via several mechanisms | Zhang et al. (2000), Zhang et al. (2009), Steed and Tyagi (2011), Kozłowski et al. (2012), Zhao et al. (2017), Wu et al. (2019) |
| CagA | CagA is released systemically from CagA-injected gastric epithelial cells via exosomes. At the vascular endothelium, the virulence factor can stimulate NF- κ B and STAT3 in endothelial cells and promote the mounting of an immune reaction at the atherosclerotic plaques by interaction with anti-CagA antibodies | Ansari and Yamaoka (2020), Tegtmeyer et al. (2017), Shimoda et al. (2016), Tahmina et al. (2022), Qiang et al. (2022) |
| | CagA+ exosomes cause endothelial dysfunction | Guo et al. (2021), Xia et al. (2022) |
| | When taken up by macrophages in plaques, CagA accelerates foam cells formation | Yang et al. (2019) |
| | CagA promotes platelets aggregation | Byrne et al. (2003); Riad (2021) |

Abbreviations: CIMT, carotid intima-media thickness test; Ox-LDL, oxidized low-density lipoprotein; 8-OHdG, 8-hydroxy-2'-deoxyguanosine, T2DM, Type 2 diabetes mellitus.

2 diabetic patients (Feng et al., 2018a). In confirmation, two recent studies revealed that patients with both *H. pylori* infection and nonalcoholic fatty liver disease (NAFLD) exhibited the highest risk of carotid artery plaque formation and arterial stiffness (Yu et al., 2019; Choi et al., 2022). Taken together, these findings support the notion that *H. pylori* infection is associated with higher CIMT measurements, suggesting a correlation with the eventual incidence of atherosclerosis, in healthy individuals or ones with other co-morbidities.

2.2 *H. pylori* infection precipitates dyslipidemia

Maintaining appropriate blood lipid profiles is critical for the prevention of atherosclerotic plaque buildup. Among serum lipids, cholesterol exhibits a high tendency to accumulate on vascular walls, potentially narrowing the lumen and obstructing blood flow. Interestingly, one of the major mechanisms by which *H. pylori* could precipitate the progression of atherosclerosis is by modifying serum lipid levels and profiles (Table 1) (Vijayvergiya and Vadivelu, 2015).

Current evidence suggests that *H. pylori* is responsible for an impairment in lipid metabolism (Adachi et al., 2018). A chronic infection with *H. pylori* can modify host body lipid distribution by activating pro-inflammatory factors, decreasing lipolysis, and enhancing *de novo* synthesis of fatty acids in the liver (Wang et al., 2022). *H. pylori* can also directly act on the liver to modify body lipid profiles by inducing liver dysfunction and elevating small intestinal mucosal permeability, facilitating the invasion of bacterial endotoxins to the liver through the portal vein, causing hepatic tissue damage (Wang et al., 2022). Importantly, *H. pylori* was reported as an independent risk factor for impaired lipid profiles, manifested as reduced high-density lipoprotein (HDL) and elevated low-density lipoprotein (LDL) levels (Buzás, 2014; Zhao et al., 2019; Hashim et al., 2022; Wang et al., 2022). It is not surprising then that dyslipidemia is prevalent in *H. pylori*-suspected patients where 87.2% of *H. pylori* positive subjects had at least one abnormality in lipid profile (Abdu et al., 2020). This argument is cemented by the finding that *H. pylori*-triggered deterioration in lipid metabolism or dyslipidemia is alleviated when this bacterium is eradicated (Park et al., 2021; Wang et al., 2022).

A recent study aiming to evaluate whether current *H. pylori* infection, detected using a rapid urease test [Campylobacter-like organism test (CLO)], is correlated with subclinical

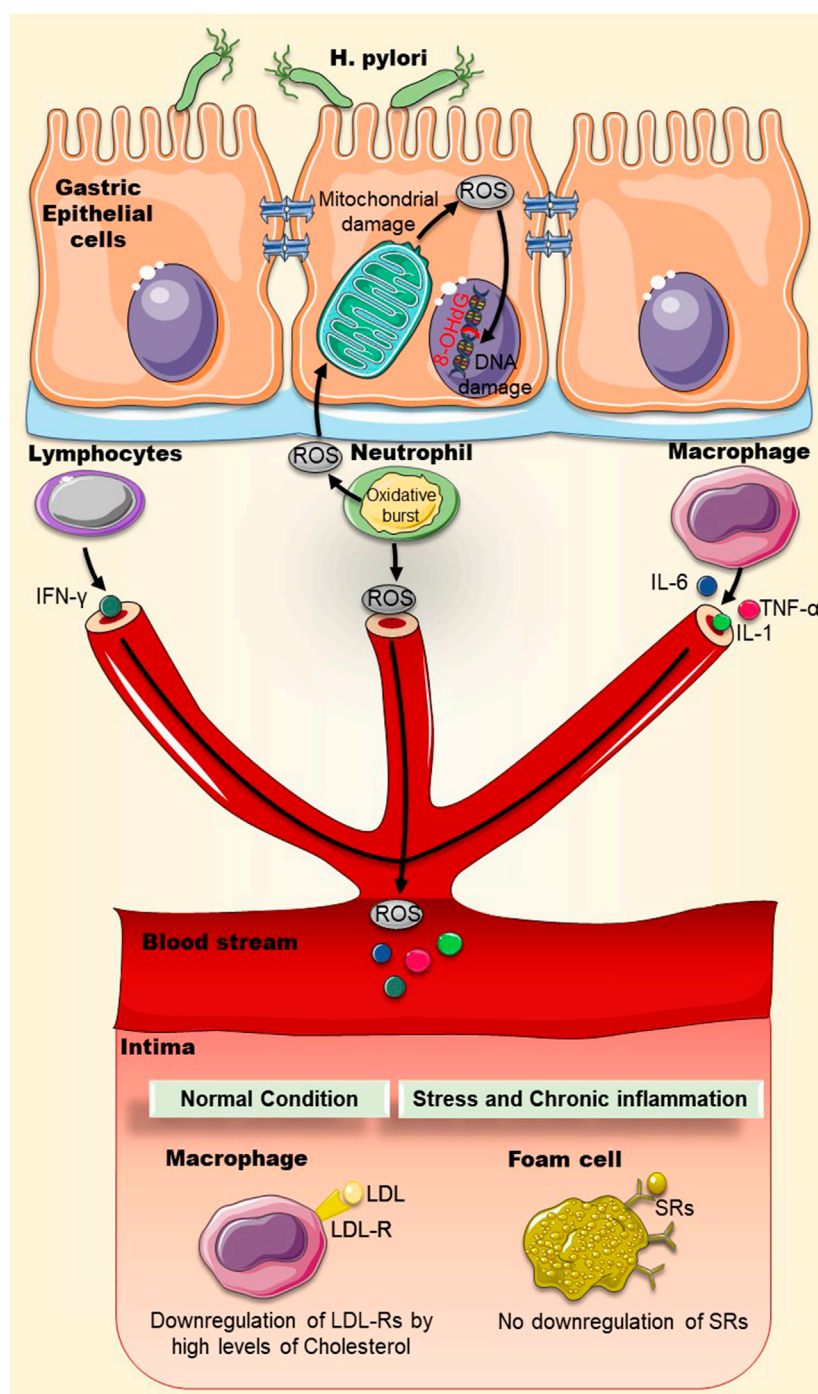


FIGURE 1

Chronic inflammation and oxidative stress may increase the uptake of oxidized-LDL particles by scavenger receptors on macrophages. Chronic *H. pylori* infection can cause local and systemic inflammation that may accelerate atherosclerotic plaque formation. Oxidative stress (ROS) and inflammatory markers (IL-1, IL-6, TNF- α , IFN- γ) generated by immune cells and mitochondrial membrane disruption can damage DNA where deoxyguanosine becomes oxidized into 8-OHdG. LDL particles are modified by oxidative stress into oxidized-LDL (Ox-LDL). Ox-LDL particles do not bind to LDL-Rs but rather bind to scavenger receptors (SRs), present on macrophages. Unlike LDL-Rs, SRs are not downregulated by high levels of cholesterol and accumulate more cholesterol and fats into macrophages, causing the eventual conversion of macrophages into foam cells. Abbreviations: LDL: Low-density lipoprotein; Ox-LDL: Oxidized low-density lipoprotein; LDL-R: low-density lipoprotein receptor; ROS: Reactive oxygen species; SRs: Scavenger receptors; 8-OHdG: 8-hydroxy-2'-deoxyguanosine.

atherosclerosis showed that CLO-positive subjects are more likely to have significant coronary artery stenosis compared to CLO-negative subjects (Lee et al., 2018). The CLO-positive

subjects also showed lower levels of HDL-cholesterol and higher mean levels of triglycerides compared to the CLO-negative subjects (Lee et al., 2018). The risk of having a

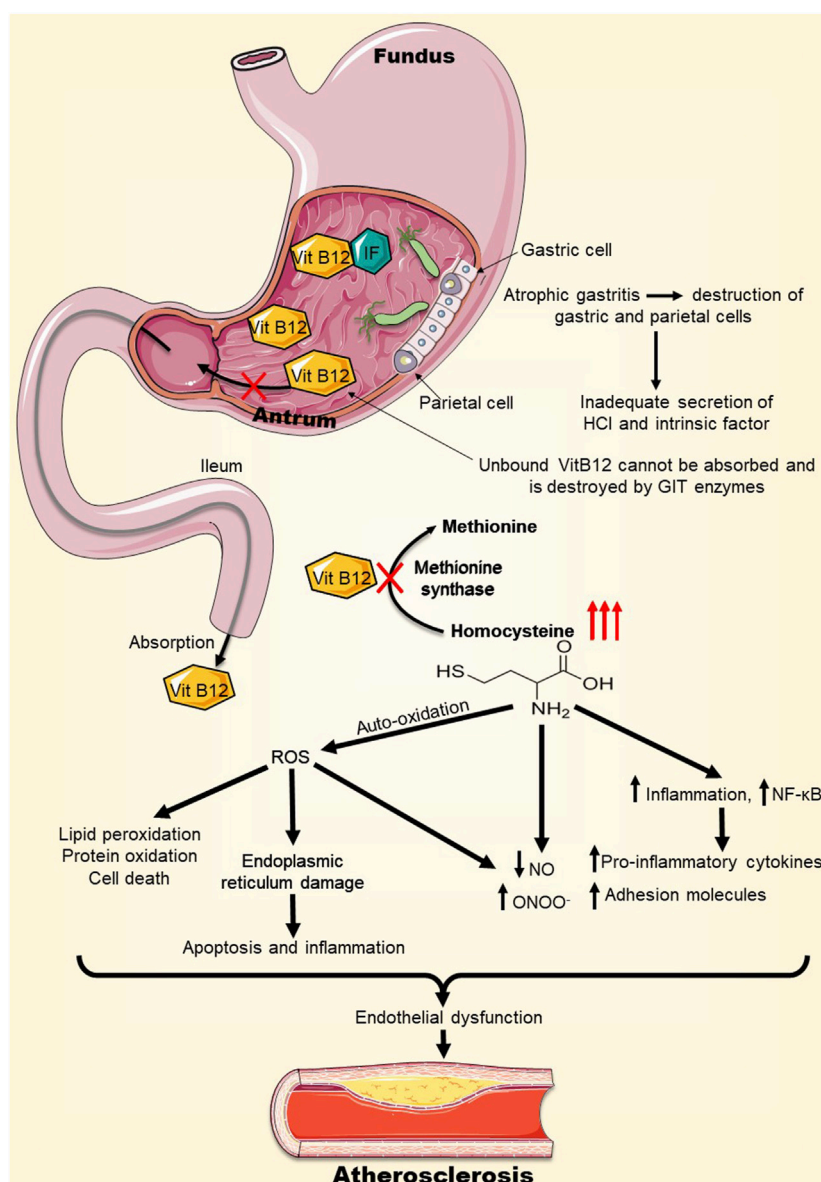


FIGURE 2

H. pylori-induced Vitamin B12 deficiency may precipitate atherosclerosis. *H. pylori* infection can lead to atrophic gastritis and destruction of parietal cells, which secrete intrinsic factor (IF), essential for vitamin B12 absorption. As a result, *H. pylori* infection can cause vitamin B12 deficiency. Vitamin B12 deficiency inhibits the enzyme methionine synthase, escalating homocysteine levels. High levels of homocysteine in the vasculature can lead to endoplasmic reticulum stress, oxidative stress, and inflammation, overall causing endothelial dysfunction which promotes atherosclerosis. Abbreviations: IF, Intrinsic Factor; ROS, Reactive oxygen species; NO, Nitric oxide; ONOO⁻, Peroxynitrite; NF-kB, Nuclear factor Kappa B.

coronary stenosis was even more prominent after adjusting for age, sex, and other factors that influence coronary artery stenosis, such as systolic blood pressure (BP), fasting glucose, HDL-cholesterol, anti-hypertension/diabetic medications, lipid-lowering agents, and antiplatelet agents (Lee et al., 2018). A recent cross-sectional study showed that *H. pylori*-infected patients exhibit higher LDL, triglycerides and cholesterol levels than control patients' (Nigatie et al., 2022). This is further supported by other findings showing that *H. pylori* seropositivity is a significant risk factor for higher levels of LDL-cholesterol, triglycerides, BMI and lower levels of

HDL-cholesterol, further establishing a role for *H. pylori* infection in dyslipidemia (Table 1) (Kim et al., 2016).

In asymptomatic healthy individuals, arterial stiffness, LDL-cholesterol levels, and the prevalence of dyslipidemia were significantly higher in the *H. pylori*-seropositive group compared to the *H. pylori*-seronegative group (Choi et al., 2019). In confirmation, findings from a recent study where participants were divided into three groups (healthy non-*H. pylori* infected, and symptomatic and asymptomatic *H. pylori* infected individuals), indicated that cholesterol levels were significantly higher in the symptomatic group than the asymptomatic and healthy groups (Medrek-Socha et al., 2018). Relevantly, both of the infected groups

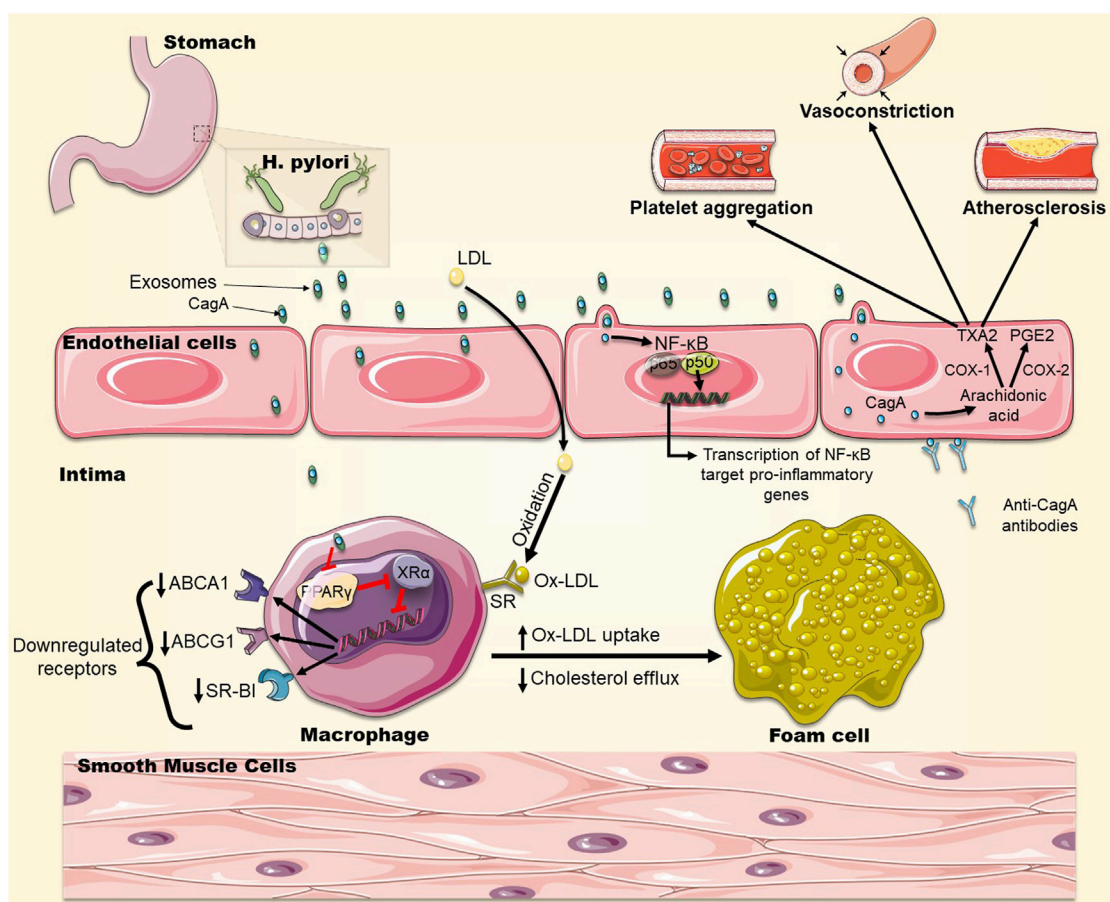


FIGURE 3

The role of CagA in foam cell formation. CagA is released in exosomes from *H. pylori*-infected gastric cells and becomes engulfed by macrophages in the intima of arteries. CagA downregulates the expression of transcription factors PPAR γ and LXRA, which control the expression of the cholesterol efflux transporters ABCA1, ABCG1 and SR-BI on the surface of macrophages leading to cholesterol accumulation in macrophages and promoting foam cell formation. In addition, OxLDL infiltrates macrophages via SRs which are not downregulated by cholesterol, exacerbating foam cell formation. CagA also stimulates NF- κ B signaling leading to the upregulation of expression of pro-inflammatory genes. CagA also elevates the levels of COX-1 and COX-2, leading to a higher production of TXA2 and PGE2, promoting platelets aggregation and vasoconstriction. CagA can be expressed on cells of the atherosclerotic plaque, to which anti-CagA antibodies will bind, exacerbating immune reactions.

showed higher LDL and lower HDL concentrations than the healthy group (Medrek-Socha et al., 2018). Other supporting data include the finding that subjects infected by *H. pylori*, specifically males between 55 and 74 years of age, had significantly higher levels of LDL-cholesterol (Shan et al., 2017). A recent observational study in chronic gastritis patients showed that *H. pylori* eradication therapy causes a significant increase in HDL levels and a significant decrease of LDL/HDL ratio, a measure of the risk of atherosclerosis (Iwai et al., 2019). Taken together, it is becoming increasingly evident that infection with *H. pylori* is associated with dyslipidemia, a key contributing factor to atherosclerotic plaque formation.

2.3 *H. pylori* infection can induce systemic immune responses

Key parameters implicated in systemic inflammation are adhesion molecules and pro-inflammatory cytokines. In this

context, it has been reported that *H. pylori* can induce upregulation of adhesion molecules on gastric epithelium as well as promote the release of several cytokines like IL-1, IL-6 and TNF- α , which then activate leukocytes and precipitate systemic inflammation (Vijayvergiya and Vadivelu, 2015). This *H. pylori*-induced chronic inflammation can precipitate persistent oxidative stress, with various noxious effects such as DNA damage, mitochondrial membrane damage and pro-inflammatory immune responses (Butcher et al., 2017; Sies et al., 2017). This contributes to a wide range of extra-gastrointestinal tract abnormalities, including atherosclerosis.

Under normal oxidative conditions, LDL particles are taken up by macrophages via receptor-mediated endocytosis by the LDL-Receptors (LDL-Rs). LDL-Rs can be downregulated by elevated levels of cholesterol, limiting the uptake of fat. However, LDL particles are extremely sensitive to oxidative damage and can be modified into oxidized-LDL (Ox-LDL) during chronic *H. pylori* infection, for instance. Ox-LDL are

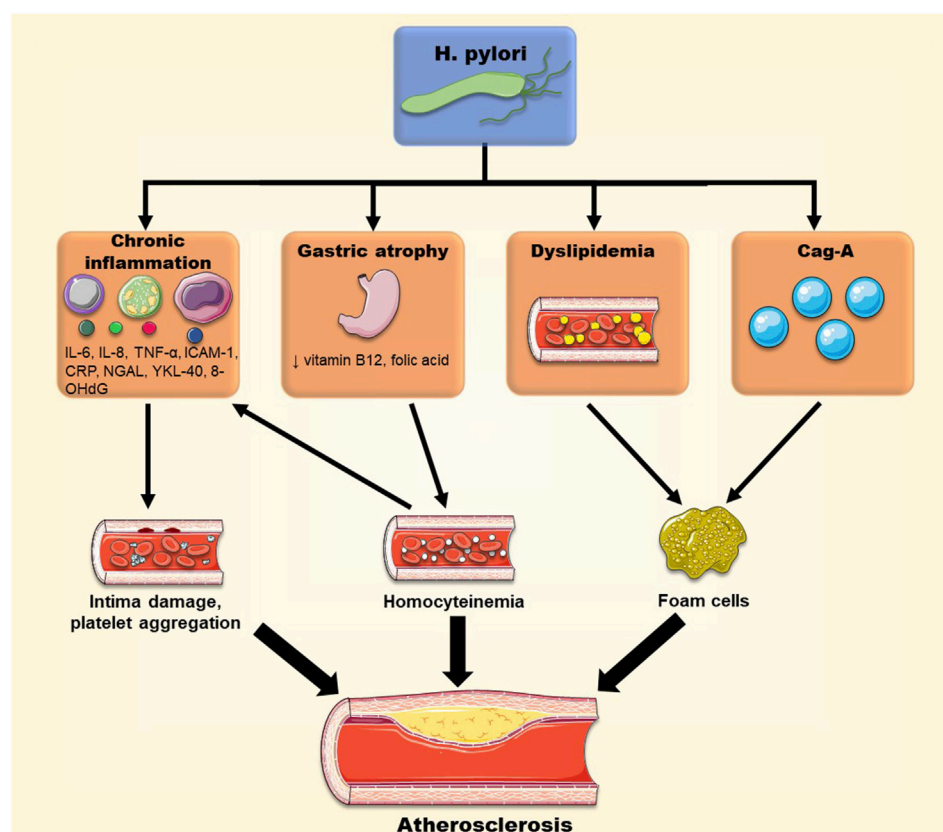


FIGURE 4

Different pathways through which *H. pylori* may promote atherosclerotic plaque formation. *H. pylori* infection can have systemic effects and cause chronic inflammation, dyslipidemia, Vitamin B12 deficiency and homocysteinemia, and promote foam cell formation, contributing to the progression of atherosclerosis.

atherogenic because they do not bind to LDL-Rs but have the capacity to bind and activate a group of receptors collectively known as scavenger receptors (SRs). These SRs are present on macrophages, but unlike LDL-Rs they are not downregulated by high levels of cholesterol, and can keep on accumulating cholesterol and fats, eventually causing the conversion of macrophages into foam cells, a hallmark of atherosclerosis (Figure 1) (Gao and Liu, 2017; Luo et al., 2017). When lipoproteins accumulate in the intima, the endothelium becomes poised to secrete a repertoire of chemokines and display several leukocyte adhesion molecules, favoring the recruitment of immune cells. An immune response is then mounted, leading to the accumulation of pro-inflammatory cytokines, proteases, and vasoactive molecules. This local inflammation can exacerbate plaque growth in blood vessels, furthering their obstruction (Hansson et al., 2006).

Ox-LDL levels were also found to be higher in serum of *H. pylori*-infected type II diabetes mellitus (T2DM) patients than in serum of diabetic patients without *H. pylori* infection or non-diabetics (Table 1) (Nasif et al., 2016). More, the role of *H. pylori* infection in accelerating foam cell formation in animals with high-fat diet has also been reported (Krupa et al., 2021). In these animals, a dramatic increase in systemic inflammatory markers fueling a pro-atherogenic endothelial cell environment was

noted. Infiltration of inflammatory cells, elevation of oxidative stress, formation of foam cells, and presence of pro-atherogenic molecules like 7-ketocholesterol and aldehydes were prominently evident (Krupa et al., 2021).

Oxidative damage also impacts both nuclear and mitochondrial DNA. For instance, deoxyguanosine becomes oxidized into 8-hydroxy-2'-deoxyguanosine (8-OHdG) (Cooke et al., 2001). Indeed, this oxidized DNA molecule is often used as a marker of oxidative damage. Moreover, it is reportedly found in rather excessive amounts in atherosclerotic plaques (Wu et al., 2004), inside macrophages, smooth muscle cells, and endothelial cells, demonstrating strong oxidative DNA damage and repair (Martinet et al., 2002). Likewise, levels of 8-OHdG and serum Ox-LDL levels in T2DM patients appear to be directly associated with *H. pylori* infection (Nasif et al., 2016). This lends support to the notion that *H. pylori* infection contributes to the pathogenesis of atherosclerosis by virtue of increasing serum Ox-LDL and 8-OHdG (Figure 1).

An interesting relation between vascular dementia, inflammation and atherosclerosis is emerging. Indeed, *H. pylori*-positive patients with vascular dementia (VD) had greater CIMT values and higher levels of YKL-40 cytokine [a biomarker of inflammation (Rathcke and Vestergaard, 2006)] than *H. pylori*-negative VD patients (Xu et al., 2016). Additionally, CIMT was

positively correlated with serum levels of YKL-40 cytokine independent of traditional atherosclerotic risk factors (Xu et al., 2016). This finding suggests a potential association between *H. pylori*, YKL-40 cytokines and atherosclerosis, warranting further investigation (Xu et al., 2016). Overall, *H. pylori* infection-induced inflammation may be able to precipitate the development of atherosclerosis as summarized in Table 1.

2.4 *H. pylori* infection can cause vitamin B12 deficiency

Atrophic gastritis is chronic inflammation of the gastric mucosa that can result in stomach atrophy along with low or absent gastric acid secretion and inadequate production of intrinsic factor (IF). This factor is a protein that binds and facilitates the transit of vitamin B12 (cobalamin) through the small intestine to be absorbed into the bloodstream (Hall and Appelman, 2019). In the absence of gastric secretions or IF, the absorption of food-bound vitamin B12 is impaired (Neumann et al., 2013). As a result, people suffering from chronic atrophic gastritis have been documented to suffer from cobalamin deficiency (Allen, 2008). Of particular interest to this review, an untreated *H. pylori* infection can eventually cause atrophic gastritis, implicating *H. pylori* in vitamin B12 deficiency (Supiano, 1987). This causality was investigated in several studies, with the conclusion that there is a probable association between cobalamin deficiency and *H. pylori* infection (Kaptan et al., 2000; Carmel et al., 2001; Dierkes et al., 2003; Sipponen et al., 2003; Raut and Chandel, 2014; Civan, 2020; Soyocak et al., 2021). These findings have potential implications to atherosclerosis because vitamin B12 malabsorption can promote the progression of atherosclerosis (Zhao et al., 2017). In fact, cobalamin is a cofactor for the enzyme methionine synthase catalyzing the conversion of homocysteine to methionine (Kozłowski et al., 2012). In the case of vitamin B12 deficiency, methionine synthase is non-functional, escalating homocysteine levels (Sipponen et al., 2003). A recent study comparing homocysteine levels of *H. pylori* positive subjects to those of *H. pylori* negative individuals found that homocysteine levels were significantly higher in the infected subjects (Huang et al., 2022).

Hyperhomocysteinemia (HHcy) is suggested to play a key role in atherosclerosis via several mechanisms (Zhao et al., 2017). The auto-oxidation of homocysteine is a source of reactive oxygen species (ROS), which at high levels in the vasculature can cause lipid peroxidation, protein oxidation, and even cell death, eventually leading to vascular injuries (Steed and Tyagi, 2011). HHcy by itself as well as HHcy-generated ROS can induce peroxynitrite (ONOO⁻) formation, effectively reducing nitric oxide (NO) bioavailability in the vasculature. Generation of ONOO⁻ and the decrease of NO levels can cause dysfunction of the vascular endothelium, instigating atherosclerosis and other diseases of the vasculature (Zhang et al., 2000). HHcy-induced oxidative stress can as well affect the endoplasmic reticulum of endothelial cells, thereby inducing apoptosis and inflammation, and dysregulation of lipid metabolism (Wu et al., 2019). Furthermore, elevated levels of homocysteine can induce the activation of nuclear factor kappa B (NF-κB) transcription factor which activates the expression of different cytokines, chemokines, and leukocyte adhesion

molecules; important mediators of vascular inflammation, pro-thrombotic state, and atherogenesis (Figure 2) (Cattaneo, 1999; Zhang et al., 2009; Riad, 2021). Taken together, it is becoming more evident that *H. pylori*-induced vitamin B12 malabsorption can lead to elevated levels of homocysteine, which may then promote the development of atherosclerosis by various mechanisms (Figure 2).

2.5 The role of CagA in atherosclerosis

CagA, a virulence factor protein expressed by certain strains of *H. pylori*, is responsible for the development of gastric diseases and is suspected to be involved in extra-gastric diseases (Ansari and Yamaoka, 2020). During infection, *H. pylori* injects this cytotoxin into host gastric cells, through a Type IV secretion system, where CagA can modify host cell signaling pathways (Tegtmeyer et al., 2017). *H. pylori*-infected gastric epithelial cells were found to release extracellular vesicles (exosomes) containing CagA. These exosomes can be delivered, through the bloodstream, to vascular endothelia distant from the primary site of infection (Table 1) (Shimoda et al., 2016). This finding may be one way to explain the systemic clinical effects thought to be mediated by *H. pylori*, including atherosclerosis. Indeed, these circulating CagA-containing exosomes may adhere to distant endothelial monolayers and deliver the CagA protein into endothelial cells, where it can stimulate NF-κB signaling. This signaling may lead to a chronic inflammatory condition, endothelial cell dysfunction, and consequently plaque formation. Similarly, Wang et al. reported that extracellular vesicle (outer membrane vesicles) derived from CagA⁺ *H. pylori*-infected cells accelerated atherosclerotic plaque formation in ApoE^{-/-} mice via NF-κB signaling (Figure 3) (Wang et al., 2021a). Exosomes-delivered CagA to endothelial cells were also found to activate the pro-inflammatory transcription factor STAT3 (Tahmina et al., 2022). In another mechanism, CagA-containing exosomes may directly fuse with cells of the atherosclerotic plaques, exposing CagA on the plaque surface to be recognized by anti-CagA antibodies, leading to immune reactions at the atherosclerotic plaque (Figure 3) (Shimoda et al., 2016).

PMSS1 is a CagA-positive *H. pylori* strain which can secrete the CagA virulence factor into host cells. PMSS1 *H. pylori*-infected mice have shown accelerated growth of atherosclerotic plaques that was dependent on dietary risk factors and genetic susceptibility (Yang et al., 2019). Immunohistochemical analysis of the vasculature of these mice revealed that the atherosclerotic plaques contained macrophage-derived foam cells. These cells significantly accumulate neutral lipid in the presence of PMSS1 infection, suggesting that the bacteria may help in the formation of foam cells (Yang et al., 2019).

This notion was reinforced by other *in vitro* and *in vivo* studies. Exosomes derived from *H. pylori*-infected gastric epithelial cells (Hp-GEC-EVs), were able to induce conversion of macrophages into foam cells when taken up by macrophages. Interestingly, CagA present in Hp-GEC-EVs would end up in the atherosclerotic plaques leading to exacerbation of the obstructive inflammatory process (Yang et al., 2019). It was also found that CagA mediated the formation of foam cells through downregulation of the expression of transcription factors PPARγ and LXRA, which control the expression of the cholesterol efflux transporters ABCA1, ABCG1 and SR-BI on the surface of macrophages, leading to cholesterol accumulation in macrophages and promoting foam cell

formation (Figure 3) (Yang et al., 2019). A recent study comparing the virulence of CagA⁺ *H. pylori* to CagA⁻ *H. pylori* in mice found that the former strain colonized gastric mucosa more effectively, causing endothelial dysfunction and enhancing atherosclerosis through ROS production, which was induced by CagA⁺ exosomes (Xia et al., 2022). Another study demonstrated that *H. pylori* infection was an independent risk factor for intracranial atherosclerosis, especially in women younger than 60 years of age, and that CagA⁺ exosomes significantly limited brain endothelial functioning *in vitro* (Guo et al., 2021). As well, a meta-analysis showed that *H. pylori* infection can significantly increase CIMT, especially in CagA⁺ individuals (Shi et al., 2022).

CagA-positive *H. pylori* strains can also promote platelet aggregation. Indeed, CagA is suspected to increase cyclooxygenase-1 (COX-1) and cyclooxygenase-2 (COX-2) production in the vascular endothelium resulting in higher thromboxane (TXA2) and prostaglandins levels, both of which are known to modify platelet aggregation and contribute to atherosclerosis (Figure 3) (Byrne et al., 2003; Riad, 2021).

Hence, CagA⁺ *H. pylori* infection may accelerate atherosclerotic plaque formation via several CagA-dependent mechanisms: CagA packaged in exosomes can modify macrophage intracellular cholesterol levels by increasing the import of cholesterol or decreasing cholesterol efflux which facilitates foam cell formation. Otherwise, CagA can promote the development of atherosclerotic plaques via the stimulation of NF- κ B, platelet aggregation, and the interaction with anti-CagA antibodies (Figure 3).

3 The counter-evidence

Despite the multitude of studies demonstrating a positive association between *H. pylori* infection and atherosclerosis, some studies report otherwise (Ahmadnia et al., 2013; Jukic et al., 2017; Feng et al., 2018b; Wernly et al., 2022). A recent cross-sectional study aiming to examine the relationship between *H. pylori* infection and the severity of coronary atherosclerosis included patients with coronary artery disease (CAD) and who underwent coronary artery bypass grafting surgery. Coronary angiograms were scored by quantitative assessment based on three angiographic parameters: the vessel score, Gensini score, and angiographic severity score. The results showed that hypertension, systolic pressure, diastolic pressure, and total cholesterol values were higher in *H. pylori*-positive subjects compared to *H. pylori*-negative subjects. HDL-cholesterol values, on the other hand, were markedly lower in the *H. pylori*-positive than in the negative subjects. However, no significant differences were found in vessel score, Gensini score and angiographic severity score, or in the use of medications that affect atherogenesis, concluding that the pathogenesis of stable chronic CAD is mainly caused by traditional risk factors rather than the influence of *H. pylori* infection (Jukic et al., 2017). Furthermore, a cross-sectional investigating the correlation between the average of CIMT values and *H. pylori* infection did not report significant differences in CIMT measurements between *H. pylori* positive and negative subjects (Feng et al., 2018b). However, traditional atherosclerosis risk factors (like gender, BMI, waistline, lipid profiles) did correlate with significant differences in CIMT scores suggesting no correlation between *H. pylori* infection and early atherosclerosis (Feng et al., 2018b). Moreover, the presence of *H. pylori* in atherosclerotic

plaques was investigated in the iliac arteries of 25 patients with end stage renal disease (ESRD) who were undergoing kidney transplantation. Although atherosclerotic plaques were present in 21 patients (84%), *H. pylori* was not detected by polymerase chain reaction and no significant relationship between atherosclerosis and gastric *H. pylori* infection could be established (Ahmadnia et al., 2013). Moreover, despite reporting an independent risk between *H. pylori* infection and CVD, the difference did not translate into CVD mortality between patients with and without the bacteria in a cohort (Wernly et al., 2022). Hence, conflicting data exist regarding the relationship between *H. pylori* and atherosclerosis and further investigations are needed in order to elucidate this relationship.

4 Conclusion

The extra-gastrointestinal manifestations of *H. pylori* infection have linked this pathogen to the progression of atherosclerosis. In addition, the bacteria may account for the development of atherosclerotic plaques through several mechanisms including dyslipidemia, systemic inflammation, oxidative stress and acceleration of foam cell formation. When different strains of the bacteria were compared, *H. pylori* strains positive for CagA showed a higher correlation with the incidence and pathogenesis of atherosclerosis, through various mechanisms involving exosomes, for example, (Figure 4). As such, interventions to eradicate *H. pylori* may help in the reduction of the prevalence of atherosclerosis. Paradoxically, some studies could not establish a role for *H. pylori* in atherosclerosis (Ahmadnia et al., 2013; Jukic et al., 2017; Feng et al., 2018b). The contradiction may be related to the presence of confounding variables, such as the methods used for diagnosis of *H. pylori* diagnostic (serology vs. rapid urease test), selection bias of geographic regions, or the study of different *H. pylori* strains, like CagA-positive or CagA-negative strains. In conclusion, *H. pylori* may exacerbate atherosclerosis, but further supporting evidence is warranted to place it alongside the traditional risk factors of atherosclerosis.

Author contributions

KA: Writing—original draft. RA: Writing—original draft. MA: Writing—original draft. KJ: Writing—original draft. AbS: Writing: review and editing. AmS: Writing: review and editing. AAE: Writing: review and editing. MR: Writing: review and editing. AHE: Conceptualization, Funding acquisition, Project administration, Supervision, Writing—original draft, Writing—review and editing.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

- Arabi, G., Heydecke, G., and Seedorf, U. (2018). Roles of oral infections in the pathomechanism of atherosclerosis. *Int. J. Mol. Sci.* 19 (7), 1978. doi:10.3390/ijms19071978
- Abdu, A., Cheneke, W., Adem, M., Belete, R., and Getachew, A. (2020). Dyslipidemia and associated factors among patients suspected to have *Helicobacter pylori* infection at jimma university medical center, jimma, Ethiopia. *Int. J. Gen. Med.* 13, 311–321. doi:10.2147/IJGM.S243848
- Adachi, K., Mishiro, T., Toda, T., Kano, N., Fujihara, H., Mishima, Y., et al. (2018). Effects of *Helicobacter pylori* eradication on serum lipid levels. *J. Clin. Biochem. Nutr.* 62 (3), 264–269. doi:10.3164/jcbn.17-88
- Ahmadnia, H., Vossoughinia, H., Mansourian, E., and Gaffarzadegan, K. (2013). No detection of *Helicobacter pylori* in atherosclerotic plaques in end stage renal disease patients undergoing kidney transplantation. *Indian J. Nephrol.* 23 (4), 259–263. doi:10.4103/0971-4065.114483
- Allen, L. H. (2008). Causes of vitamin B12 and folate deficiency. *Food Nutr. Bull.* 29 (2), S20–S34. doi:10.1177/156482650802925105
- Ansari, S., and Yamaoka, Y. (2020). *Helicobacter pylori* virulence factor cytotoxin-associated gene A (CagA)-Mediated gastric pathogenicity. *Int. J. Mol. Sci.* 21 (19), 7430. doi:10.3390/ijms21197430
- Bao-Ge, Q., Hui, W., Yi-Guo, J., Ji-Liang, S., Zhong-Dong, W., Ya-Fei, W., et al. (2017). The correlation and risk factors between carotid intima-media thickening and alcoholic liver disease coupled with *Helicobacter pylori* infection. *Sci. Rep.* 7, 43059. doi:10.1038/srep43059
- Buruco, C., and Axon, A. (2017). Epidemiology of *Helicobacter pylori* infection. *Helicobacter* 22 (1). doi:10.1111/hel.12403
- Butcher, L. D., den Hartog, G., Ernst, P. B., and Crowe, S. E. (2017). Oxidative stress resulting from *Helicobacter pylori* infection contributes to gastric carcinogenesis. *Cell Mol. Gastroenterol. Hepatol.* 3 (3), 316–322. doi:10.1016/j.jcmgh.2017.02.002
- Buzás, G. M. (2014). Metabolic consequences of *Helicobacter pylori* infection and eradication. *World J. Gastroenterol.* 20 (18), 5226–5234. doi:10.3748/wjg.v20.i18.5226
- Byrne, M. F., Murphy, J. F., Corcoran, P. A., Atherton, J. C., Sheehan, K. M., Cox, D., et al. (2003). *Helicobacter pylori* induces cyclooxygenase-1 and cyclooxygenase-2 expression in vascular endothelial cells. *Scand. J. Gastroenterol.* 38 (10), 1023–1030. doi:10.1080/00365520310005622
- Campuzano-Maya, G. (2014). Hematologic manifestations of *Helicobacter pylori* infection. *World J. Gastroenterol.* 20 (36), 12818–12838. doi:10.3748/wjg.v20.i36.12818
- Carmel, R., Aurangzeb, I., and Qian, D. (2001). Associations of food-cobalamin malabsorption with ethnic origin, age, *Helicobacter pylori* infection, and serum markers of gastritis. *Am. J. Gastroenterol.* 96 (1), 63–70. doi:10.1111/j.1572-0241.2001.03453.x
- Cattaneo, M. (1999). Hyperhomocysteinemia, atherosclerosis and thrombosis. *Thromb. Haemost.* 81 (2), 165–176. doi:10.1055/s-0037-1614438
- Chang, W. L., Yeh, Y. C., and Sheu, B. S. (2018). The impacts of *H. pylori* virulence factors on the development of gastroduodenal diseases. *J. Biomed. Sci.* 25 (1), 68. doi:10.1186/s12929-018-0466-9
- Choi, J. M., Lim, S. H., Han, Y. M., Lee, H., Seo, J. Y., Park, H. E., et al. (2019). Association between *Helicobacter pylori* infection and arterial stiffness: Results from a large cross-sectional study. *PLoS One* 14 (8), e0221643. doi:10.1371/journal.pone.0221643
- Choi, J. M., Park, H. E., Han, Y. M., Lee, J., Lee, H., Chung, S. J., et al. (2022). Non-alcoholic/Metabolic-Associated fatty liver disease and *Helicobacter pylori* additively increase the risk of arterial stiffness. *Front. Med. (Lausanne)* 9, 844954. doi:10.3389/fmed.2022.844954
- Civan, H. A. (2020). Vitamin B12 status in children with *Helicobacter pylori* gastritis. *Med. J. Bakirkoy* 16 (2). doi:10.5222/BMJ.2020.03522
- Cooke, M. S., Evans, M. D., Burd, R. M., Patel, K., Barnard, A., Lunec, J., et al. (2001). Induction and excretion of ultraviolet-induced 8-oxo-2'-deoxyguanosine and thymine dimers *in vivo*: Implications for PUVA. *J. Invest. Dermatol.* 116 (2), 281–285. doi:10.1046/j.1523-1747.2001.01251.x
- Dadashi, A., and Hosseinzadeh, N. (2018). High seroprevalence of anti-*Helicobacter pylori* antibodies in patients with ventilator-associated pneumonia. *J. Res. Med. Sci.* 23, 79. doi:10.4103/jrms.JRMS_117_18
- de Brito, B. B., da Silva, F. A. F., Soares, A. S., Pereira, V. A., Santos, M. L. C., Sampaio, M. M., et al. (2019). Pathogenesis and clinical management of *Helicobacter pylori* gastric infection. *World J. Gastroenterol.* 25 (37), 5578–5589. doi:10.3748/wjg.v25.i37.5578
- Dierkes, J., Ebert, M., Malfertheiner, P., and Luley, C. (2003). *Helicobacter pylori* infection, vitamin B12 and homocysteine. A review. *Dig. Dis.* 21 (3), 237–244. doi:10.1159/000073341
- Eusebi, L. H., Zagari, R. M., and Bazzoli, F. (2014). Epidemiology of *Helicobacter pylori* infection. *Helicobacter* 19 (1), 1–5. doi:10.1111/hel.12165
- Fang, Y., Fan, C., and Xie, H. (2019). Effect of *Helicobacter pylori* infection on the risk of acute coronary syndrome: A systematic review and meta-analysis. *Med. Baltim.* 98 (50), e18348. doi:10.1097/MD.00000000000018348
- Feng, L., Deng, C., and Li, Y. (2018a). Assessment of the relationship between carotid intima-media thickening and early-stage diabetic kidney disease coupled with *Helicobacter pylori* infection. *Dis. Markers* 2018, 3793768. doi:10.1155/2018/3793768
- Feng, Y., Zhou, W., Luo, L., and Xu, W. (2018b). *Helicobacter pylori* infection is not related to increased carotid intima-media thickness in general population. *Sci. Rep.* 8 (1), 14180. doi:10.1038/s41598-018-32465-4
- Ford, A. C., Yuan, Y., Forman, D., Hunt, R., and Moayyedi, P. (2020). *Helicobacter pylori* eradication for the prevention of gastric neoplasia. *Cochrane Database Syst. Rev.* 7, CD005583. doi:10.1002/14651858.CD005583.pub3
- Franceschi, F., Covino, M., and Roubaud Baudron, C. (2019). Review: *Helicobacter pylori* and extragastric diseases. *Helicobacter* 24 (1), e12636. doi:10.1111/hel.12636
- Gao, S., and Liu, J. (2017). Association between circulating oxidized low-density lipoprotein and atherosclerotic cardiovascular disease. *Chronic Dis. Transl. Med.* 3 (2), 89–94. doi:10.1016/j.cdtm.2017.02.008
- Guo, Y., Xu, C., Zhang, L., Chen, Z., and Xia, X. (2021). *Helicobacter pylori* infection acts as an independent risk factor for intracranial atherosclerosis in women less than 60 Years old. *Front. Cardiovasc. Med.* 8, 819315. doi:10.3389/fcvm.2021.819315
- Hall, S. N., and Appelman, H. D. (2019). Autoimmune gastritis. *Arch. Pathol. Lab. Med.* 143 (11), 1327–1331. doi:10.5858/arpa.2019-0345-RA
- Hansson, G. K., Robertson, A. K., and Soderberg-Naucler, C. (2006). Inflammation and atherosclerosis. *Annu. Rev. Pathol.* 1, 297–329. doi:10.1146/annurev.pathol.1.110304.100100
- Hashim, M., Mohammed, O., Egzeabeher, T., and Wolde, M. (2022). The association of *Helicobacter Pylori* infection with dyslipidaemia and other atherogenic factors in dyspeptic patients at St. Paul's Hospital Millennium Medical College. *Heliyon* 8 (5), e09430. doi:10.1016/j.heliyon.2022.e09430
- Hata, K., Koyama, T., Ozaki, E., Kuriyama, N., Mizuno, S., Matsui, D., et al. (2021). Assessing the relationship between *Helicobacter pylori* and chronic kidney disease. *Healthc. (Basel)* 9 (2), 162. doi:10.3390/healthcare9020162
- He, J., Liu, Y., Ouyang, Q., Li, R., Li, J., Chen, W., et al. (2022). *Helicobacter pylori* and unignorable extragastric diseases: Mechanism and implications. *Front. Microbiol.* 13, 972777. doi:10.3389/fmicb.2022.972777
- Herrington, W., Lacey, B., Sherliker, P., Armitage, J., and Lewington, S. (2016). Epidemiology of atherosclerosis and the potential to reduce the global burden of atherothrombotic disease. *Circ. Res.* 118 (4), 535–546. doi:10.1161/CIRCRESAHA.115.307611
- Huang, X., Qiu, Y., Zheng, X., Luo, M., Wang, Y., Yi, J., et al. (2022). Correlation of *Helicobacter pylori* infection with atherosclerosis and its risk factors. doi:10.21203/rs.3.rs-1929881/v1
- Iwai, N., Okuda, T., Oka, K., Hara, T., Inada, Y., Tsuji, T., et al. (2019). *Helicobacter pylori* eradication increases the serum high density lipoprotein cholesterol level in the infected patients with chronic gastritis: A single-center observational study. *PLoS One* 14 (8), e0221349. doi:10.1371/journal.pone.0221349
- Jukic, A., Bozic, D., Kardum, D., Becic, T., Luksic, B., Vrsalovic, M., et al. (2017). *Helicobacter pylori* infection and severity of coronary atherosclerosis in patients with

chronic coronary artery disease. *Ther. Clin. Risk Manag.* 13, 933–938. doi:10.2147/tcr.m.1342193

Kaplan, K., Beyan, C., Ural, A. U., Cetin, T., Ayvci, F., Gulsen, M., et al. (2000). *Helicobacter pylori* is it a novel causative agent in Vitamin B12 deficiency? *Arch. Intern. Med.* 160 (9), 1349–1353. doi:10.1001/archinte.160.9.1349

Karbasi-Afshar, R., Khedmat, H., and Izadi, M. (2015). *Helicobacter pylori* infection and atherosclerosis: a systematic review. *Acta Med. Iran.* 53 (2), 78–88.

Kim, T. J., Lee, H., Kang, M., Kim, J. E., Choi, Y. H., Min, Y. W., et al. (2016). *Helicobacter pylori* is associated with dyslipidemia but not with other risk factors of cardiovascular disease. *Sci. Rep.* 6, 38015. doi:10.1038/srep38015

Kountouras, J., Papaefthymiou, A., Polyzos, S. A., Deretzi, G., Vardaka, E., Soteriades, E. S., et al. (2021). Impact of *Helicobacter pylori*-related metabolic syndrome parameters on arterial hypertension. *Microorganisms* 9 (11), 2351. doi:10.3390/microorganisms9112351

Kozłowski, P. M., Kamachi, T., Kumar, M., and Yoshizawa, K. (2012). Reductive elimination pathway for homocysteine to methionine conversion in cobalamin-dependent methionine synthase. *J. Biol. Inorg. Chem.* 17 (4), 611–619. doi:10.1007/s00775-012-0881-4

Krupa, A., Gonciarz, W., Rusek-Wala, P., Rechciński, T., Gajewski, A., Samsel, Z., et al. (2021). *Helicobacter pylori* infection acts synergistically with a high-fat diet in the development of a proinflammatory and potentially proatherogenic endothelial cell environment in an experimental model. *Int. J. Mol. Sci.* 22 (7), 3394. doi:10.3390/ijms22073394

Lechner, K., von Schacky, C., McKenzie, A. L., Worm, N., Nixdorff, U., Lechner, B., et al. (2020). Lifestyle factors and high-risk atherosclerosis: Pathways and mechanisms beyond traditional risk factors. *Eur. J. Prev. Cardiol.* 27 (4), 394–406. doi:10.1177/2047487319869400

Lee, M., Baik, H., Park, J. S., Kim, S., Kyung, C., Baik, S. J., et al. (2018). Current *Helicobacter pylori* infection is significantly associated with subclinical coronary atherosclerosis in healthy subjects: A cross-sectional study. *PLoS One* 13 (3), e0193646. doi:10.1371/journal.pone.0193646

Leja, M., Grinberga-Derica, I., Bilgiler, C., and Steininger, C. (2019). Review: Epidemiology of *Helicobacter pylori* infection. *Helicobacter* 24 (1), e12635. doi:10.1111/hel.12635

Liang, H., Lin, S., Ji, Y., Xiao, Y., and Zheng, G. (2021). *Helicobacter pylori* increases the risk of carotid plaque formation: a clinical evidence. *Ann. Med.* 53 (1), 1448–1454. doi:10.1080/07853890.2021.1927169

Libby, P., Buring, J. E., Badimon, L., Hansson, G. K., Deanfield, J., Bittencourt, M. S., et al. (2019). Atherosclerosis. *Nat. Rev. Dis. Prim.* 5 (1), 56. doi:10.1038/s41572-019-0106-z

Libby, P., Loscalzo, J., Ridker, P. M., Farkouh, M. E., Hsue, P. Y., Fuster, V., et al. (2018). Inflammation, immunity, and infection in atherothrombosis: JACC review topic of the week. *J. Am. Coll. Cardiol.* 72 (17), 2071–2081. doi:10.1016/j.jacc.2018.08.1043

Luo, Y., Duan, H., Qian, Y., Feng, L., Wu, Z., Wang, F., et al. (2017). Macrophagic CD146 promotes foam cell formation and retention during atherosclerosis. *Cell Res.* 27 (3), 352–372. doi:10.1038/cr.2017.8

Mansori, K., Dehghanbanadaki, H., Naderpour, S., Rashti, R., Moghaddam, A. B., and Moradi, Y. (2020a). A systematic review and meta-analysis of the prevalence of *Helicobacter pylori* in patients with diabetes. *Diabetes Metab. Syndr.* 14 (4), 601–607. doi:10.1016/j.dsx.2020.05.009

Mansori, K., Moradi, Y., Naderpour, S., Rashti, R., Moghaddam, A. B., Saed, L., et al. (2020b). *Helicobacter pylori* infection as a risk factor for diabetes: a meta-analysis of case-control studies. *BMC Gastroenterol.* 20 (1), 77. doi:10.1186/s12876-020-01223-0

Martinet, W., Knaapen, M. W., De Meyer, G. R., Herman, A. G., and Kockx, M. M. (2002). Elevated levels of oxidative DNA damage and DNA repair enzymes in human atherosclerotic plaques. *Circulation* 106 (8), 927–932. doi:10.1161/01.cir.0000026393.47805.21

Medrek-Socha, M., Chojnacki, J., Smigielski, J., Konrad, P., and Chojnacki, C. (2018). Changes in the lipid profile of patients with asymptomatic and symptomatic *Helicobacter pylori* infection. *Wiad. Lek.* 71 (8), 1467–1473.

Mentis, A., Lehours, P., and Megraud, F. (2015). Epidemiology and diagnosis of *Helicobacter pylori* infection. *Helicobacter* 20 (1), 1–7. doi:10.1111/hel.12250

Mizuno, S., Matsui, D., Watanabe, I., Ozaki, E., Kuriyama, N., and Watanabe, Y. (2015). Serologically determined gastric mucosal condition is a predictive factor for osteoporosis in Japanese men. *Dig. Dis. Sci.* 60 (7), 2063–2069. doi:10.1007/s10620-015-3576-1

Nasif, W. A., Mukhtar, M. H., Nour Eldein, M. M., and Ashgar, S. S. (2016). Oxidative DNA damage and oxidized low density lipoprotein in Type II diabetes mellitus among patients with *Helicobacter pylori* infection. *Diabetol. Metab. Syndr.* 8, 34. doi:10.1186/s13098-016-0149-1

Neumann, W. L., Coss, E., Rugge, M., and Genta, R. M. (2013). Autoimmune atrophic gastritis-pathogenesis, pathology and management. *Nat. Rev. Gastroenterol. Hepatol.* 10 (9), 529–541. doi:10.1038/nrgastro.2013.101

Nigatie, M., Melak, T., Asmelash, D., and Worede, A. (2022). Dyslipidemia and its associated factors among *Helicobacter pylori*-infected patients attending at university of gondar comprehensive specialized hospital, gondar, north-west Ethiopia: A comparative cross-sectional study. *J. Multidiscip. Healthc.* 15, 1481–1491. doi:10.2147/jmdh.S368832

Park, Y., Kim, T. J., Lee, H., Yoo, H., Sohn, I., Min, Y. W., et al. (2021). Eradication of *Helicobacter pylori* infection decreases risk for dyslipidemia: A cohort study. *Helicobacter* 26 (2), e12783. doi:10.1111/hel.12783

Polyzos, S. A., Kountouras, J., Zavos, C., and Deretzi, G. (2011). The association between *Helicobacter pylori* infection and insulin resistance: a systematic review. *Helicobacter* 16 (2), 79–88. doi:10.1111/j.1523-5378.2011.00822.x

Pothineni, N. V. K., Subramany, S., Kuriakose, K., Shirazi, L. F., Romeo, F., Shah, P. K., et al. (2017). Infections, atherosclerosis, and coronary heart disease. *Eur. Heart J.* 38 (43), 3195–3201. doi:10.1093/eurheartj/ehx362

Qiang, L., Hu, J., Tian, M., Li, Y., Ren, C., Deng, Y., et al. (2022). Extracellular vesicles from *Helicobacter pylori*-infected cells and *Helicobacter pylori* outer membrane vesicles in atherosclerosis. *Helicobacter* 27 (2), e12877. doi:10.1111/hel.12877

Rathcke, C. N., and Vestergaard, H. (2006). YKL-40, a new inflammatory marker with relation to insulin resistance and with a role in endothelial dysfunction and atherosclerosis. *Inflamm. Res.* 55 (6), 221–227. doi:10.1007/s00011-006-0076-y

Raut, S. C., and Chandel, R. S. (2014). Comparison of vitamin B12 levels in gastritis with and without *H. pylori*. *Walawalkar Int. Med. J.* 1, 28–32.

Riad, M. (2021). Association of *Helicobacter pylori* infection with coronary artery disease: Is it an independent risk factor? *Egypt Heart J.* 73 (1), 61. doi:10.1186/s43044-021-00185-2

Shan, J., Bai, X., Han, L., Yuan, Y., Yang, J., and Sun, X. (2018). Association between atherosclerosis and gastric biomarkers concerning *Helicobacter pylori* infection in a Chinese healthy population. *Exp. Gerontol.* 112, 97–102. doi:10.1016/j.exger.2018.09.009

Shan, J. H., Bai, X. J., Han, L. L., Yuan, Y., and Sun, X. F. (2017). Changes with aging in gastric biomarkers levels and in biochemical factors associated with *Helicobacter pylori* infection in asymptomatic Chinese population. *World J. Gastroenterol.* 23 (32), 5945–5953. doi:10.3748/wjg.v23.i32.5945

Shi, H., Li, Y., Dong, C., Si, G., Xu, Y., Peng, M., et al. (2022). *Helicobacter pylori* infection and the progression of atherosclerosis: A systematic review and meta-analysis. *Helicobacter* 27 (1), e12865. doi:10.1111/hel.12865

Shimoda, A., Ueda, K., Nishiumi, S., Murata-Kamiya, N., Mukai, S. A., Sawada, S., et al. (2016). Exosomes as nanocarriers for systemic delivery of the *Helicobacter pylori* virulence factor CagA. *Sci. Rep.* 6, 18346. doi:10.1038/srep18346

Siasos, G., Tsigkou, V., Kokkou, E., Oikonomou, E., Vavuranakis, M., Vlachopoulos, C., et al. (2014). Smoking and atherosclerosis: mechanisms of disease and new therapeutic approaches. *Curr. Med. Chem.* 21 (34), 3936–3948. doi:10.2174/092986732134141015161539

Sies, H., Berndt, C., and Jones, D. P. (2017). Oxidative stress. *Annu. Rev. Biochem.* 86, 715–748. doi:10.1146/annurev-biochem-061516-045037

Simon, O. A., Görbe, A., Hegyi, P., Szakó, L., Oštarijaš, E., Dembrowsky, F., et al. (2022). *Helicobacter pylori* infection is associated with carotid intima and media thickening: A systematic review and meta-analysis. *J. Am. Heart Assoc.* 11 (3), e022919. doi:10.1161/jaha.121.022919

Sipponen, P., Laxen, F., Huotari, K., and Harkonen, M. (2003). Prevalence of low vitamin B12 and high homocysteine in serum in an elderly male population: association with atrophic gastritis and *Helicobacter pylori* infection. *Scand. J. Gastroenterol.* 38 (12), 1209–1216. doi:10.1080/00365520310007224

Soyocak, A., Ergun, D. D., Koc, G., Ergun, S., and Ozsobaci, N. P. (2021). Investigation of aryl hydrocarbon receptor, zinc, and vitamin B12 levels in chronic gastritis with *Helicobacter pylori* infection. *Biol. Trace Elem. Res.* 199 (7), 2431–2437. doi:10.1007/s12011-021-02667-5

Spence, J. D., and Pilote, L. (2015). Importance of sex and gender in atherosclerosis and cardiovascular disease. *Atherosclerosis* 241 (1), 208–210. doi:10.1016/j.atherosclerosis.2015.04.806

Steed, M. M., and Tyagi, S. C. (2011). Mechanisms of cardiovascular remodeling in hyperhomocysteinemia. *Antioxid. Redox Signal* 15 (7), 1927–1943. doi:10.1089/ars.2010.3721

Sugano, K. (2019). Effect of *Helicobacter pylori* eradication on the incidence of gastric cancer: a systematic review and meta-analysis. *Gastric Cancer* 22 (3), 435–445. doi:10.1007/s10120-018-0876-0

Supiano, M. A. (1987). Atrophic gastritis and vitamin B12 deficiency. *J. Am. Geriatr. Soc.* 35 (5), 478. doi:10.1111/j.1532-5415.1987.tb04684.x

Tahmina, K., Hikawa, N., Takahashi-Kanemitsu, A., Knight, C. T., Sato, K., Itoh, F., et al. (2022). Transgenically expressed *Helicobacter pylori* CagA in vascular endothelial cells accelerates arteriosclerosis in mice. *Biochem. Biophys. Res. Commun.* 618, 79–85. doi:10.1016/j.bbrc.2022.06.010

Talari, H. R., Moniri, R., Mollaghanbari, M., Haddad Kashani, H., and Jalalian, M. N. (2021). Evaluating the relationship between *Helicobacter pylori* infection and carotid intima-media thickness a cross sectional study. *Ann. Med. Surg. (Lond)* 69, 102659. doi:10.1016/j.amsu.2021.102659

Tegtmeyer, N., Wessler, S., Necchi, V., Rohde, M., Harrer, A., Rau, T. T., et al. (2017). *Helicobacter pylori* employs a unique basolateral type IV secretion mechanism for CagA delivery. *Cell Host Microbe* 22 (4), 552–560. doi:10.1016/j.chom.2017.09.005

- Tsay, F. W., and Hsu, P. I. (2018). *H. pylori* infection and extra-gastrointestinal diseases. *J. Biomed. Sci.* 25 (1), 65. doi:10.1186/s12929-018-0469-6
- Vijayvergiya, R., and Vadivelu, R. (2015). Role of *Helicobacter pylori* infection in pathogenesis of atherosclerosis. *World J. Cardiol.* 7 (3), 134–143. doi:10.4330/wjc.v7.i3.134
- Waluga, M., Kukla, M., Żorniak, M., Bacik, A., and Kotulski, R. (2015). From the stomach to other organs: *Helicobacter pylori* and the liver. *World J. Hepatol.* 7 (18), 2136–2146. doi:10.4254/wjh.v7.i18.2136
- Wang, F., Meng, W., Wang, B., and Qiao, L. (2014). *Helicobacter pylori*-induced gastric inflammation and gastric cancer. *Cancer Lett.* 345 (2), 196–202. doi:10.1016/j.canlet.2013.08.016
- Wang, N., Zhou, F., Chen, C., Luo, H., Guo, J., Wang, W., et al. (2021a). Role of outer membrane vesicles from *Helicobacter pylori* in atherosclerosis. *Front. Cell Dev. Biol.* 9, 673993. doi:10.3389/fcell.2021.673993
- Wang, X., He, Q., Jin, D., Ma, B., Yao, K., and Zou, X. (2021b). Association between *Helicobacter pylori* infection and subclinical atherosclerosis: A systematic review and meta-analysis. *Med. Baltim.* 100 (46), e27840. doi:10.1097/md.00000000000027840
- Wang, Y. C. (2014). Medicinal plant activity on *Helicobacter pylori* related diseases. *World J. Gastroenterol.* 20 (30), 10368–10382. doi:10.3748/wjg.v20.i30.10368
- Wang, Z., Wang, W., Gong, R., Yao, H., Fan, M., Zeng, J., et al. (2022). Eradication of *Helicobacter pylori* alleviates lipid metabolism deterioration: a large-cohort propensity score-matched analysis. *Lipids Health Dis.* 21 (1), 34. doi:10.1186/s12944-022-01639-5
- Wernly, S., Semmler, G., Völcker, A., Flamm, M., Aigner, E., Niederseer, D., et al. (2022). *Helicobacter pylori* and cardiovascular risk: Only a dead *Helicobacter* is a good *Helicobacter*? *Helicobacter* 27 (6), e12928. doi:10.1111/hel.12928
- Wroblewski, L. E., and Peek, R. M., Jr. (2016). *Helicobacter pylori*, cancer, and the gastric microbiota. *Adv. Exp. Med. Biol.* 908, 393–408. doi:10.1007/978-3-319-41388-4_19
- Wu, L. L., Chiou, C. C., Chang, P. Y., and Wu, J. T. (2004). Urinary 8-OHdG: a marker of oxidative stress to DNA and a risk factor for cancer, atherosclerosis and diabetes. *Clin. Chim. Acta* 339 (1–2), 1–9. doi:10.1016/j.cccn.2003.09.010
- Wu, X., Zhang, L., Miao, Y., Yang, J., Wang, X., Wang, C. C., et al. (2019). Homocysteine causes vascular endothelial dysfunction by disrupting endoplasmic reticulum redox homeostasis. *Redox Biol.* 20, 46–59. doi:10.1016/j.redox.2018.09.021
- Xia, X., Zhang, L., Wu, H., Chen, F., Liu, X., Xu, H., et al. (2022). CagA(+) *Helicobacter pylori*, not CagA(-) *Helicobacter pylori*, infection impairs endothelial function through exosome-mediated ROS formation. *Front. Cardiovasc. Med.* 9, 881372. doi:10.3389/fcvm.2022.881372
- Xu, Y., Wang, Q., Liu, Y., Cui, R., Lu, K., and Zhao, Y. (2016). Association between *Helicobacter pylori* infection and carotid atherosclerosis in patients with vascular dementia. *J. Neurol. Sci.* 362, 73–77. doi:10.1016/j.jns.2016.01.025
- Yang, S., Xia, Y. P., Luo, X. Y., Chen, S. L., Li, B. W., Ye, Z. M., et al. (2019). Exosomal CagA derived from *Helicobacter pylori*-infected gastric epithelial cells induces macrophage foam cell formation and promotes atherosclerosis. *J. Mol. Cell Cardiol.* 135, 40–51. doi:10.1016/j.jmcc.2019.07.011
- Yao, B. C., Meng, L. B., Hao, M. L., Zhang, Y. M., Gong, T., and Guo, Z. G. (2019). Chronic stress: a critical risk factor for atherosclerosis. *J. Int. Med. Res.* 47 (4), 1429–1440. doi:10.1177/0300060519826820
- Yu, L. Y., Hu, K. C., Liu, C. J., Hung, C. L., Bair, M. J., Chen, M. J., et al. (2019). *Helicobacter pylori* infection combined with non-alcoholic fatty liver disease increase the risk of atherosclerosis: Focus in carotid artery plaque. *Med. Baltim.* 98 (9), e14672. doi:10.1097/MD.00000000000014672
- Zhang, D., Jiang, X., Fang, P., Yan, Y., Song, J., Gupta, S., et al. (2009). Hyperhomocysteinemia promotes inflammatory monocyte generation and accelerates atherosclerosis in transgenic cystathionine beta-synthase-deficient mice. *Circulation* 120 (19), 1893–1902. doi:10.1161/CIRCULATIONAHA.109.866889
- Zhang, L., Chen, Z., Xia, X., Chi, J., Li, H., Liu, X., et al. (2019). *Helicobacter pylori* infection selectively increases the risk for carotid atherosclerosis in young males. *Atherosclerosis* 291, 71–77. doi:10.1016/j.atherosclerosis.2019.10.005
- Zhang, P., He, Q., Song, D., Wang, Y., Liu, X., Ding, G., et al. (2021). Association of *Helicobacter pylori* infection with carotid atherosclerosis in a northern Chinese population: A cross-sectional study. *Front. Cardiovasc. Med.* 8, 795795. doi:10.3389/fcvm.2021.795795
- Zhang, X., Li, H., Jin, H., Ebin, Z., Brodsky, S., and Goligorsky, M. S. (2000). Effects of homocysteine on endothelial nitric oxide production. *Am. J. Physiol. Ren. Physiol.* 279 (4), F671–F678. doi:10.1152/ajprenal.2000.279.4.F671
- Zhao, J., Chen, H., Liu, N., Chen, J., Gu, Y., Chen, J., et al. (2017). Role of hyperhomocysteinemia and hyperuricemia in pathogenesis of atherosclerosis. *J. Stroke Cerebrovasc. Dis.* 26 (12), 2695–2699. doi:10.1016/j.jstrokecerebrovasdis.2016.10.012
- Zhao, M. M., Krebs, J., Cao, X., Cui, J., Chen, D. N., Li, Y., et al. (2019). *Helicobacter pylori* infection as a risk factor for serum bilirubin change and less favourable lipid profiles: a hospital-based health examination survey. *BMC Infect. Dis.* 19 (1), 157. doi:10.1186/s12879-019-3787-8



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Inclisiran: a new generation of lipid-lowering siRNA therapeutic

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Atherosclerotic heart disease (AHD) is a major cause of morbidity and mortality worldwide. Lowering low-density lipoprotein cholesterol (LDL-C) levels is a key strategy to prevent and treat AHD. Inclisiran is a novel siRNA drug that targets proprotein convertase subtilisin/kexin type 9 (PCSK9) gene expression and reduces LDL-C levels with only two or three injections per year. This review summarizes the mechanism, efficacy, safety, and applications of Inclisiran in various populations and settings, based on recent literature. It also compares Inclisiran with other lipid-lowering drugs, especially other PCSK9 inhibitors. We conclude that Inclisiran is a promising lipid-lowering agent that can provide convenience and effectiveness for patients with high cardiovascular risk. However, some challenges and limitations remain for Inclisiran, such as its long-term safety and efficacy, its cost-effectiveness and accessibility, and its interactions and synergies with other drugs. These issues need further investigation and evaluation in future studies.

KEYWORDS

lipid-lowering therapies, cardiovascular risk, PCSK9, Inclisiran, siRNA

1 Introduction

Atherosclerotic heart disease (AHD) remains a global public health problem despite of significant advances in its prevention and treatment (Dong et al., 2022). It is caused by the accumulation of cholesterol and other substances in the arterial walls, which led to narrowing and hardening of the arteries and reduce blood flow to the heart and other organs. This can result in chest pain, heart attack, stroke or death. A positive correlation between circulating low-density lipoprotein cholesterol (LDL-C) levels and coronary heart disease has been demonstrated by numerous epidemiological and clinical studies (Mortensen and Nordestgaard, 2020). LDL-C is a kind of cholesterol that can damage the arteries and increase the risk of atherosclerosis (Ference et al., 2013; Liu et al., 2021). LDL-C is the main carrier of cholesterol in the blood and can be taken up by the arterial wall cells via LDL receptors, contributing to plaque formation and inflammation. This process can lead to atherosclerosis and cardiovascular disease (Robinson et al., 2018). In China, the percentage of adults with dyslipidemia, which is an abnormal level of lipids in the blood, is as high as 33.8%, while the percentage of those with increased LDL-C level, which is a major risk factor for coronary artery disease, is 4.0%. These statistics indicate a high prevalence and burden of lipid metabolic disorders in China (Lu et al., 2021). How to effectively reduce LDL-C is one of the research questions to be answered.

In recent years, PCSK9 has gained much attention as a star target for reducing LDL-C levels in patients (German and Shapiro, 2020). PCSK9 is a protein that regulates the degradation of LDL receptors on the surface of liver cells, thereby affecting the clearance of LDL-C from the blood. PCSK9 binds to LDL receptors and promotes their degradation in lysosomes, thus reducing the number of LDL receptors available for clearing LDL-C from the blood. Therefore, inhibiting PCSK9 can increase the expression of LDL receptors and lower LDL-C levels (Tibolla et al., 2011). At present, the drug research and development targeting PCSK9 is mainly focused on preventing the binding of PCSK9 to the low-density lipoprotein (LDL) receptor. There are two main types of drugs that can achieve this goal: monoclonal antibodies and small interfering RNAs (siRNA). Monoclonal antibodies are proteins that can bind to PCSK9 and block its interaction with LDL receptors (Stroes et al., 2014). siRNA are short nucleic acids that can silence the expression of PCSK9 gene in liver cells (Reyes-Soffer et al., 2017), inhibiting the expression and secretion of PCSK9 (Fitzgerald et al., 2017).

Among these drugs, monoclonal antibodies that block the binding of PCSK9 to LDL receptors are now available in China. The lipid-lowering effect of this drug is very effective, require monthly or every biweekly injection. However, the shortcoming of low compliance and the high cost of preparation of monoclonal antibody preparations are still the major obstacles to the development of these drugs (Kaddoura et al., 2020). This drug refers to the monoclonal antibodies that target PCSK9, such as evolocumab and alirocumab. These drugs can significantly reduce LDL-C levels and cardiovascular events in patients with dyslipidemia (Stroes et al., 2014; Robinson et al., 2015; Sabatine et al., 2017; Diaz et al., 2021). In the meantime, Alirocumab can reduced the risk of any stroke and ischemic stroke without increasing hemorrhagic stroke (Jukema et al., 2019). However, they also have some drawbacks, such as low patient adherence due to frequent injections, high production cost and limited accessibility. Therefore, new lipid-lowering drugs are needed urgently.

Recent studies have shown that PCSK9 is a key protein that regulates low density lipoprotein receptor (LDL-R) expression in hepatocytes, and is positively correlated with LDL-C levels (Tibolla et al., 2011). Targeting PCSK9 has brought new opportunities for lipid-lowering therapies. However, current treatments still face challenges such as poor medication adherence (Jahangir et al., 2021). However, as an emerging therapeutic approach, the long-term safety, cardiovascular risk reduction efficacy, and applicability across diverse populations of siRNA therapeutics need to be verified through more studies (Kosmas et al., 2018).

Inclisiran (trade name Leqvio[®]) is a lipid-lowering novel drug developed by Novartis Group. It is the first siRNA drug approved for the treatment of hypercholesterolemia or mixed dyslipidemia, as well as ASCVD or heterozygous familial hypercholesterolemia (HeFH) patients who need further lowering of LDL-C levels. It is a short-chain, synthetic small interfering ribonucleic acid (siRNA) (Khvorova, 2017). siRNAs are types of double-stranded RNAs that can induce gene silencing by degrading the complementary mRNAs in a sequence-specific manner. However, some issues and challenges remain for Inclisiran, such as its effect on type 2 diabetes, its long-term outcomes, its affordability and availability, and its combination

with other drugs. Moreover, the molecular mechanisms and pathways of Inclisiran-mediated LDL-C reduction and cardiovascular protection need to be clarified, and the development of novel siRNA drugs for lipid metabolism or cardiovascular disease may offer new opportunities for personalized and precision medicine. Considering the important value of Inclisiran as a novel siRNA drug in lipid-lowering therapy, we reviewed the recent literature on this drug, and outlined its mechanism of action, pharmacodynamics, safety evaluation, application in China and future application, etc., to provide a comprehensive overview of the advantages and existing problems of Inclisiran, and to offer reference for its position in lipid-lowering therapy.

2 Methods

2.1 Literature search

A literature search was conducted using PubMed, EMBASE, and SinoMed databases. Search terms pre-defined in titles, abstracts, and keywords were used to identify pertinent studies. The retrieval period spanned from the inception of the databases up to March 2023.

3 Inclisiran delivery and mechanism of action

3.1 Mechanism of action

Inclisiran specifically binds to n-acetylgalactosamine (GalNAc) and the asialoglycoprotein receptor (ASGPR) on liver cell membranes (Kosmas et al., 2018). GalNAc is a sugar molecule that enhances the affinity and specificity of Inclisiran for ASGPR, which is a receptor that mediates the endocytosis of glycoproteins in liver cells. After entering the hepatocytes, it binds to the RNA-induced silencing complex (RISC). RISC is a protein complex that recognizes and cleaves the target mRNA based on the guide strand of siRNA. At the same time, it binds to the mRNA who encoding the PCSK9 protein mediated by the antisense chain, which leads to a decrease in PCSK9 protein production. PCSK9 is a protein that regulates the expression of LDL receptors on the surface of liver cells. PCSK9 binds to LDL receptors and promotes their degradation in lysosomes, thus reducing the number of LDL receptors available for clearing LDL-C from the blood. Therefore, by reducing PCSK9 production, Inclisiran can enhance LDL receptor expression and lower LDL-C levels. To illustrate the mechanism of action of Inclisiran, we present a schematic diagram in Figure 1.

3.2 Inclisiran efficacy

After a single dose of Inclisiran, the level of LDL-C was decreased by about 50% and maintained for up to 6 months (Barale et al., 2021; Miname et al., 2021). It is noteworthy that the silencing complex remains active after mRNA degradation occurs. Therefore, the lipid-lowering effect of Inclisiran is effective in the long term.

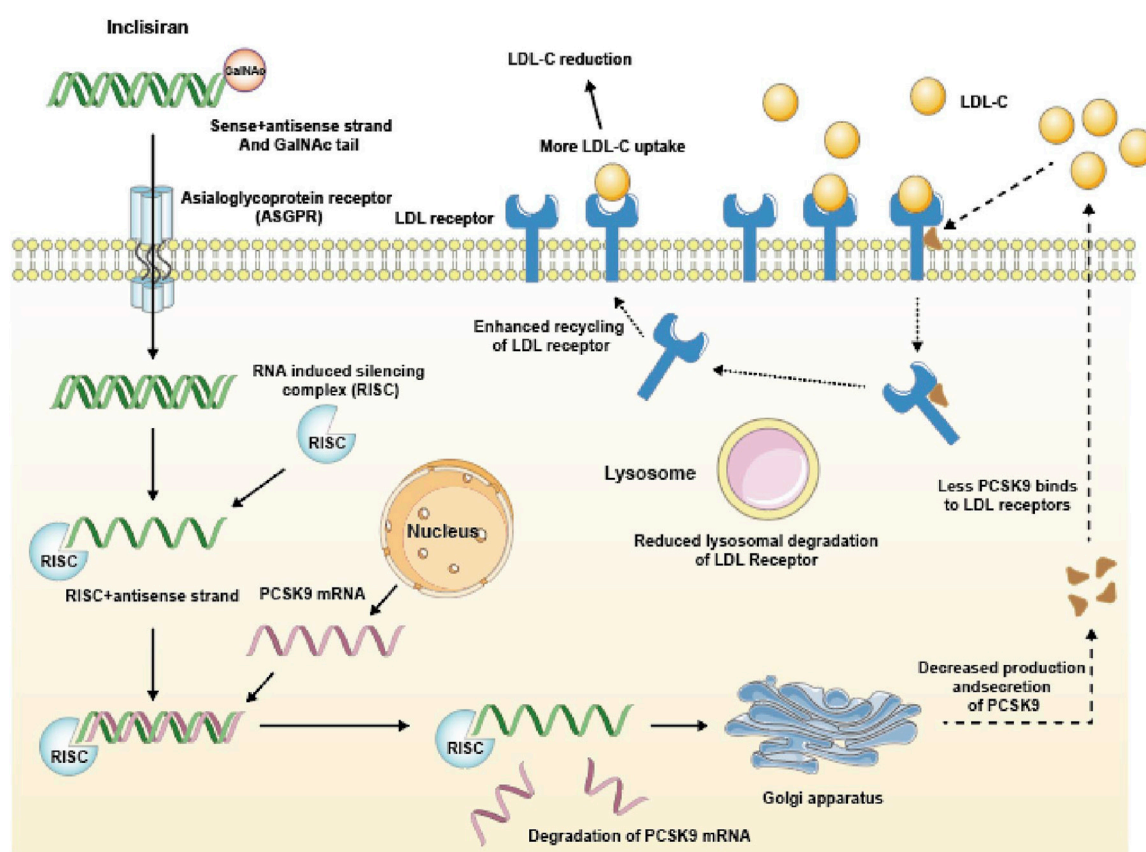


FIGURE 1

Mechanism of action of Inclisiran. GalNAc tail, N-acetylgalactosamine tail; RISC, RNA-induced silencing complex; PCSK9, Proprotein convertase subtilisin/kexin type 9.

4 Clinical efficacy, safety evaluation and exploration and application in China for different types of hypercholesterolemia and cardiovascular disease patients

4.1 Clinical trials of inclisiran for hypercholesterolemia

Inclisiran is involved in five global studies: ORION 4, 9, 10, 11, 18. These studies are phase III, double-blind, randomized, placebo-controlled trials that aim to evaluate the efficacy and safety of Inclisiran in patients with different types of hypercholesterolemia.

4.1.1 ORION 9: inclisiran for heterozygous familial hypercholesterolemia

ORION 9 enrolled patients with heterozygous familial hypercholesterolemia (HeFH), a genetic disorder that causes high LDL-C levels and increased risk of cardiovascular disease. The trial tested the hypothesis that Inclisiran, an innovative siRNA agent that silences PCSK9 gene expression in the liver and increases LDL receptor availability, would reduce LDL-C levels more than placebo in patients who were on maximally tolerated statin and ezetimibe.

4.1.2 ORION 10 and 11: inclisiran for atherosclerotic cardiovascular disease or risk equivalents

ORION 10 and 11 enrolled patients with atherosclerotic cardiovascular disease (ASCVD) or ASCVD risk equivalents, such as diabetes mellitus, chronic kidney disease or peripheral artery disease (Ray et al., 2020). These are conditions that can damage the blood vessels and impair the blood flow to the heart and other organs. The patients enrolled in the study received at least 30 days of treatment with a statin at the maximum tolerable dose. Statins are drugs that can lower LDL-C levels by inhibiting the enzyme that produces cholesterol. The dose of statins was constant throughout the study with or without ezetimibe. Ezetimibe is a cholesterol absorption inhibitor that can lower LDL-C levels by blocking the uptake of cholesterol from the intestine. Enrolled patients were randomized to the Inclisiran group or the placebo group. On the basis of the maximum tolerable dose of statin ± ezetimibe, Inclisiran 300 mg or placebo was given subcutaneously at D0, D90, D270, and D450. The primary endpoint of the studies was the percentage change in LDL-C levels from baseline to day 510. The secondary endpoints included the absolute change in LDL-C levels from baseline to day 510, the time-adjusted percentage change in LDL-C levels from day 90 to day 540, and the safety and tolerability of Inclisiran. The Inclisiran group showed a 51% decrease in LDL-C levels,

TABLE 1 Characteristics and outcomes of the clinical trials of Inclisiran for hypercholesterolemia.

| Name | Population | Inclusion criteria | Exclusion criteria | Trial design | Follow-up time | Primary endpoint | Secondary endpoint |
|----------|---|--|---|---|--|---|---|
| ORION-9 | Patients with HeFH | 18–80 years old, genetic or clinical diagnosis of HeFH, LDL-C \geq 2.6 mmol/L, maximum tolerated dose of statin \pm other lipid-lowering drugs \geq 30 days | Pregnant or lactating women, liver dysfunction, other serious diseases or conditions unsuitable for trial participation | Double-blind, randomized, placebo-controlled, multicenter, phase III trial, randomization ratio of 1:1 | Observation and measurement on day 0, day 90, day 180, day 270, day 360, day 450 and day 540, for a total of 18 months | Percentage change in LDL-C level from baseline to day 510 | Absolute change in LDL-C level from baseline to day 510; time-adjusted percentage change in LDL-C level from day 90 to day 540; safety and tolerability of Inclisiran |
| ORION-10 | Patients with ASCVD or risk equivalent (US) | 18–85 years old, with ASCVD or risk equivalent (such as diabetes, chronic kidney disease or peripheral artery disease), LDL-C \geq 1.8 mmol/L (70 mg/dL), maximum tolerated dose of statin \pm other lipid-lowering drugs \geq 30 days | Pregnant or lactating women, liver dysfunction, other serious diseases or conditions unsuitable for trial participation | Double-blind, randomized, placebo-controlled, multicenter, phase III trial, randomization ratio of 1:1 | Observation and measurement on day 0, day 90, day 180, day 270, day 360, day 450 and day 540, for a total of 18 months | Percentage change in LDL-C level from baseline to day 510 | Absolute change in LDL-C level from baseline to day 510; time-adjusted percentage change in LDL-C level from day 90 to day 540; safety and tolerability of Inclisiran |
| ORION-11 | Patients with ASCVD or risk equivalent (Europe) | 18–85 years old, with ASCVD or risk equivalent (such as diabetes, chronic kidney disease or peripheral artery disease), LDL-C \geq 1.8 mmol/L (70 mg/dL), maximum tolerated dose of statin \pm other lipid-lowering drugs \geq 30 days | Pregnant or lactating women, liver dysfunction, other serious diseases or conditions unsuitable for trial participation | Double-blind, randomized, placebo-controlled, multicenter, phase III trial, randomization ratio of 1:1 | Observation and measurement on day 0, day 90, day 180, day 270, day 360, day 450 and day 540, for a total of 18 months | Percentage change in LDL-C level from baseline to day 510 | Absolute change in LDL-C level from baseline to day 510; time-adjusted percentage change in LDL-C level from day 90 to day 540; safety and tolerability of Inclisiran |
| ORION-4 | Patients with ASCVD or high risk (global) | 55–80 years old, with ASCVD or high risk (such as diabetes, chronic kidney disease or peripheral artery disease), LDL-C \geq 1.8 mmol/L (70 mg/dL), optimized lipid-lowering therapy \geq 30 days | Pregnant or lactating women, liver dysfunction, other serious diseases or conditions unsuitable for trial participation | Randomized, placebo-controlled, multicenter phase III trial to evaluate the effect of Inclisiran on cardiovascular outcomes. Randomization ratio of 1:1 | At least 4 years of follow-up, Observe and test every 6 months | Composite endpoint of coronary heart disease death, nonfatal myocardial infarction, fatal or nonfatal ischemic stroke or coronary revascularization (major adverse cardiovascular events, MACE) | Other cardiovascular outcomes and safety and tolerability of Inclisiran |
| ORION-18 | Patients with primary hypercholesterolemia or mixed hyperlipidemia (Asia) | 18–85 years old -with primary hypercholesterolemia or mixed hyperlipidemia -LDL-C \geq 2.6 mmol/L (100 mg/dL) -maximum tolerated dose of statin \pm other lipid-lowering drugs \geq 30 days | Pregnant or lactating women -liver dysfunction -other serious diseases or conditions unsuitable for trial participation | Double-blind -randomized -placebo-controlled -multicenter phase III trial to evaluate the efficacy and safety of Inclisiran in Asian populations Randomization ratio of 1:1 | Observation and measurement on day, 0, day, 90, day, 180, day, 270, day, 360, day, 450 and day, 540, for a total of, 18 months | Percentage change in LDL-C level from baseline to day,510 | Other lipid parameters and safety and tolerability of Inclisiran |

HeFH, heterozygous familial hypercholesterolemia; LDL-C, Low-Density Lipoprotein Cholesterol; ASCVD, atherosclerotic cardiovascular disease.

compared to the placebo group, which was statistically significant and clinically meaningful (Leiter et al., 2019; Jahangir et al., 2021). The reduction was consistent and sustained across all subgroups and time points. Inclisiran also reduced other lipid parameters, such as non-

HDL-C, apolipoprotein B and lipoprotein (a). Inclisiran was well tolerated and had a similar safety profile to placebo. The most common adverse events were injection site reactions, which were mild or moderate and self-limiting.

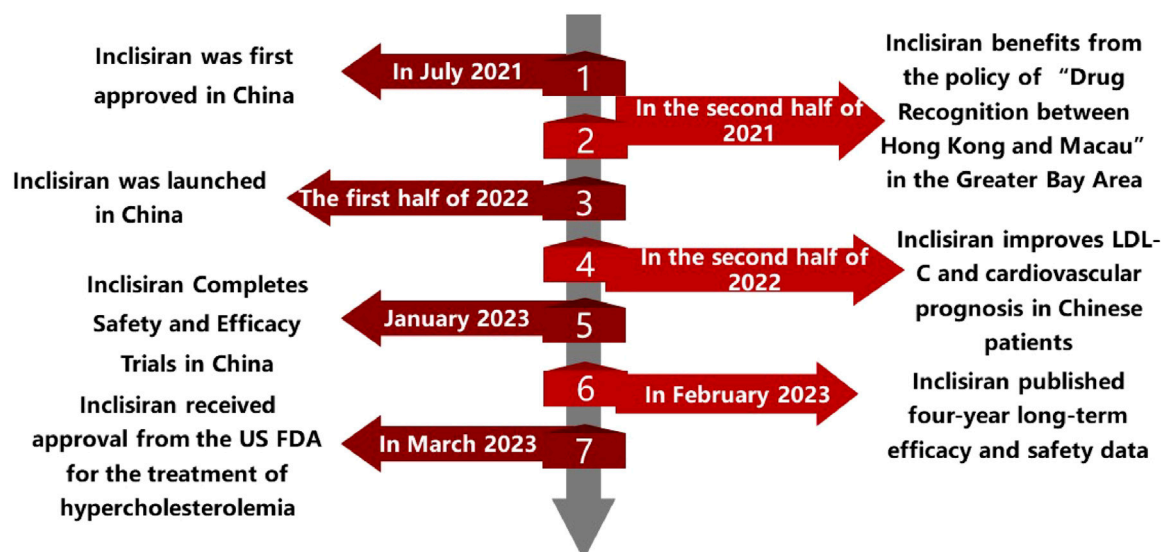


FIGURE 2
The development and use of Inclisiran in China: a chronological overview.

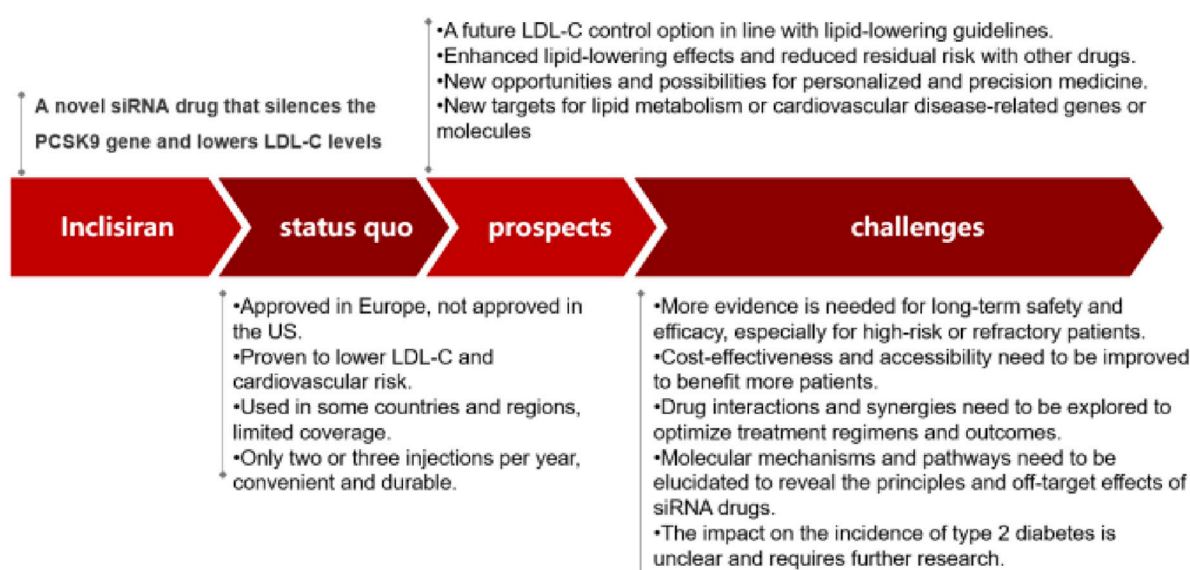


FIGURE 3
Status quo, prospects and challenges of Inclisiran.

4.1.3 ORION-18: inclisiran for asian population

Inclisiran's phase III clinical study-ORION-18, which is also taking place in China and other Asian countries for the first time. This study will enroll about 1,500 patients with primary hypercholesterolemia or mixed dyslipidemia who are not adequately controlled by statins alone or in combination with other lipid-lowering therapies. The primary endpoint is the percentage change in LDL-C levels from baseline to day 510. This study will provide more evidence for the efficacy and safety of Inclisiran in Asian populations, who may have different genetic

and environmental factors that affect their lipid metabolism and response to treatment (Jahangir et al., 2021; Ray et al., 2023).

4.1.4 ORION 4: inclisiran for cardiovascular outcomes

Additionally, ORION4 studies are being conducted to further assess the long-term effectiveness, safety, and cardiovascular benefits of Inclisiran (Brandts and Ray, 2021). This is a large-scale, multicenter, randomized trial that will enroll about 15,000 patients with ASCVD or high risk of ASCVD who have

elevated LDL-C levels despite optimal lipid-lowering therapy. It will compare the effects of Inclisiran versus placebo on cardiovascular outcomes. The primary outcome is the composite of coronary heart disease death, non-fatal myocardial infarction, fatal or non-fatal ischemic stroke or coronary revascularization. These are major adverse cardiovascular events (MACE) that can cause significant morbidity and mortality. The trial will follow the patients for at least 4 years and the results could be announced in 2025. This trial will provide important evidence for the long-term benefits and safety of Inclisiran in reducing cardiovascular risk.

A comprehensive summary of the details and findings of the clinical trials of Inclisiran for various hypercholesterolemia populations is given in the table below (See [Table 1](#)).

4.2 Safety of inclisiran

4.2.1 Advantages of inclisiran safety profile

Compared with other lipid-lowering therapies, Inclisiran has a low risk of adverse events and no major safety concerns. The most common adverse event reported in the Inclisiran group was injection site reaction, which occurred in about 5% of the patients. However, these reactions were mostly mild and transient, and none of them were severe or persistent. Injection site reactions may be related to the lipid nanoparticles used for siRNA delivery and may decrease with repeated administration. Moreover, Inclisiran did not cause any significant toxicity in liver, kidney, muscle and platelet functions. Laboratory examination showed that there was no change in the levels of alanine aminotransferase, aspartate aminotransferase, creatinine, creatine kinase or platelet count after Inclisiran treatment. Inclisiran also did not affect the cardiovascular system. There was no difference in blood pressure, heart rate or electrocardiogram parameters between the Inclisiran group and the placebo group. Therefore, Inclisiran appears to be a safe and well-tolerated drug for lowering LDL-C levels in patients with hypercholesterolemia or mixed dyslipidemia (Hardy et al., 2021; Wright et al., 2021; Ray et al., 2023).

4.2.2 Adverse reactions of inclisiran and safety comparison with siRNA drugs

One of the concerns about siRNA drugs is their possible off-target effects and immune responses (Meng and Lu, 2017; Hu et al., 2020). However, Inclisiran has been developed to reduce these risks by using a highly specific and stable siRNA sequence and a GalNAc conjugate that enhances the delivery to liver cells (Hardy et al., 2021). Inclisiran has a good overall safety profile (Wright et al., 2020). However, recently, there have been some reports that Inclisiran may have adverse reactions such as diarrhea and headache, which has aroused concern about its long-term safety. Further studies could continue monitoring its long-term safety across diverse populations.

4.3 Comparison with PCSK9 monoclonal antibody therapy

Compared with PCSK9 monoclonal antibodies, Inclisiran acts downstream by degrading mRNA to achieve sustained suppression

of PCSK9, whereas monoclonal antibodies directly block the binding of PCSK9 to LDL receptors (Leiter et al., 2019). The dosing frequency of Inclisiran is once or twice yearly, which is significantly less frequent than the 2-week or monthly injections required for monoclonal antibodies (Ray et al., 2020). This may greatly improve patient compliance. In addition, Inclisiran provides more durable lipid-lowering effects, with a single dose maintaining efficacy for 3–6 months. Overall, as an siRNA therapy, Inclisiran has unique advantages in mechanism, longer duration of action, and more convenient administration compared to monoclonal antibodies. Compared with PCSK9 monoclonal antibodies, Inclisiran demonstrated more durable lipid-lowering effects, which was confirmed by the long-term results of ORION-3 (Ray et al., 2023). The treatment course of PCSK9 monoclonal antibodies is generally 1–2 years, while Inclisiran can maintain the effect for up to 4 years. This may make Inclisiran more suitable for long-term use.

4.4 The applications of inclisiran in China nowadays

In addition to conducting clinical trials and obtaining approval in developed countries such as Europe and America, Inclisiran has also initiated a series of investigations and applications in China. However, there are still concerns about the cost and affordability and accessibility of Inclisiran. Inclisiran is priced high, costing about 20,000 yuan per injection, and about 40,000 yuan per year. The drug has not been included in the medical insurance list yet, so patients have to pay for it themselves. The high price of Inclisiran may limit its widespread use in China, which requires improving its accessibility by controlling drug costs and negotiating with medical insurance. We look forward to the real-world application data in the future, to evaluate the cost-effectiveness of Inclisiran in China.

4.4.1 Inclisiran's benefits for Chinese patients

In the near future, this drug will be more widely available in more cities and hospitals, which will have a positive effect on the quality of life of the Chinese people. Inclisiran is expected to benefit millions of Chinese patients with hypercholesterolemia or mixed dyslipidemia who are not adequately controlled by current therapies (Luo et al., 2023).

4.4.2 Inclisiran's entry into China

On July 2 2021, Novartis Pharmaceuticals held a press conference to announce that its new siRNA lipid-lowering drug, Inclisiran, had completed the first injection in China in Boao Super Hospital. The drug was subsequently used in some private hospitals in the Greater Bay Area as the 17th drug approved through the policy of "drug communication between Hong Kong and Macao". This policy allows the use of drugs that have been approved by the regulatory authorities of Hong Kong or Macao in designated medical institutions in Hainan province without additional approval from the mainland authorities. In this way, the policy can facilitate the access of innovative drugs to Chinese patients and promote the development of the healthcare industry in Hainan. The following timeline ([Figure 2](#)) summarizes the main events and dates of the introduction and utilization of Inclisiran in China.

5 Discussion

In this review, Inclisiran had been introduced as a novel siRNA lipid-lowering agent that can silence PCSK9 gene expression and lower LDL-C levels with only two or three injections per year. The characteristics and prospects of Inclisiran in terms of its clinical trials, safety and applications in China had also been summarized. In this section, some of the remaining issues and challenges for Inclisiran had been discussed, such as its impact on the risk of type 2 diabetes, its long-term safety and efficacy, its cost-effectiveness and accessibility, and its interactions and synergies with other drugs.

One of the issues that need to be further investigated is the impact of Inclisiran on the risk of type 2 diabetes. Up-regulating LDL receptors in order to reduce LDL-C levels had been shown to have mixed results in terms of its effect on type 2 diabetes incidence. Up-regulating LDL receptors in order to reduce LDL-C levels has been shown to have mixed results in terms of its effect on type 2 diabetes incidence (Ray et al., 2023). Type 2 diabetes is a metabolic disorder that can increase the risk of cardiovascular complications. Some studies suggest that it has a beneficial effect, while others find no significant effect. It is possible that the discrepancy between studies may be due to gender differences, as some studies have reported that women may have a higher risk of developing type 2 diabetes after LDL receptor upregulation than men (Shi et al., 2020). Mendelian randomization had previously proved that the reduction of LDL-C due to genetic variation at PCSK9 does increase the risk of type 2 diabetes (Schmidt et al., 2017). At present, the exact mechanism by which PCSK9 inhibitors increase the risk of type 2 diabetes is unclear. In the same way, it remained to be further discussed whether Inclisiran increased the prevalence of type 2 diabetes.

We believed that the impact of Inclisiran on the risk of type 2 diabetes is an unresolved issue, which may be influenced by gender, genotype and other drugs, and requires more research on individualized, precision and combination therapy to elucidate its mechanism and optimize its application.

Another issue that needs to be addressed is the long-term safety and efficacy of Inclisiran. The results of the clinical trial studies published so far confirmed the efficacy of Inclisiran in lowering LDL-C. However, even with lipid-lowering therapy, there are still some considerable residual risks of cardiovascular disease. These risks are considered to be caused by dysglycemia, hypertension, procoagulant state and inflammation. These are factors that can impair the endothelial function and promote the development of atherosclerosis. Clinical findings regarding lipids have emerged in recent years, and as a result, consensus lipid management guidelines have proposed lower LDL-C control goals. These guidelines are based on the evidence that lower LDL-C levels can reduce the risk of cardiovascular events and mortality in patients with dyslipidemia. Lipid management was gradually changing from “Lower is Better” to “Lower for Longer.” This means that achieving and maintaining low LDL-C levels for a long period of time is more important than short-term fluctuations (Ray et al., 2023). We believed that the long-term safety and effectiveness of Inclisiran is an important and uncertain issue, and more follow-up and observation are needed to assess its impact on the prevention and treatment of cardiovascular disease.

A third issue that needs to be considered is the cost-effectiveness and accessibility of Inclisiran. However, there were still some challenges and limitations for Inclisiran and other siRNA drugs. These drugs can effectively lower LDL-C levels by inhibiting the protein that degrades LDL receptors in the liver. The low frequency of treatment of Inclisiran could improve patients' compliance and significantly reduce the complication incidence, which may provide a future solution required for LDL-C-lowering treatment. For instance, the long-term safety and efficacy of Inclisiran need to be further evaluated in larger and more diverse populations, especially in patients with high cardiovascular risk or familial hypercholesterolemia. These are patients who may have more severe or resistant forms of dyslipidemia and may require more intensive or novel treatments. Moreover, the cost-effectiveness and accessibility of Inclisiran need to be improved to make it more affordable and available for patients who need it. The current price and supply of Inclisiran might limit its widespread use and adoption in clinical practice (Desai et al., 2022).

A final issue that needs to be explored is the potential interactions and synergies between Inclisiran and other lipid-lowering agents or cardiovascular drugs. There may be additional or synergistic effects of combining Inclisiran with other drugs that can lower LDL-C levels by different mechanisms or that can modulate other cardiovascular risk factors. The cost-effectiveness and accessibility of Inclisiran is an issue to consider, but Inclisiran and other siRNA drugs still face some challenges and limitations that need to be overcome or addressed.

Additionally, the molecular mechanisms and pathways involved in Inclisiran-mediated LDL-C reduction and cardiovascular protection need to be elucidated to reveal the underlying biology and pharmacology of siRNA drugs. This might help to understand the mode of action and the potential off-target effects of Inclisiran and other siRNA drugs. Finally, the development of novel siRNA drugs targeting other genes or molecules related to lipid metabolism or cardiovascular disease might offer new opportunities and possibilities for personalized and precision medicine. These drugs may provide more specific and tailored treatments for different subtypes or phenotypes of patients with dyslipidemia or cardiovascular disease (Scicchitano et al., 2021; Mercep et al., 2022).

6 Conclusion

In conclusion, Inclisiran was a promising siRNA lipid-lowering agent that can silence PCSK9 gene expression and lower LDL-C levels with only two or three injections per year. It had shown favorable results in various clinical trials for different types of hypercholesterolemia and cardiovascular disease patients. It had also demonstrated a good safety profile and a potential application in China. Globally, Inclisiran has been approved by the US FDA and the European EMA for the treatment of primary hypercholesterolemia or mixed dyslipidemia. In China, Inclisiran has been approved for the general evaluation of drugs in Hainan Province, and some private hospitals can use this innovative lipid-regulating drug. It is expected that Inclisiran will obtain the approval of China NDAs in the near future. However, there were still some issues and challenges that need to be addressed for Inclisiran, such as its impact on the risk of type 2 diabetes, its long-term safety and

efficacy, its cost-effectiveness and accessibility, and its interactions and synergies with other drugs. Moreover, the molecular mechanisms and pathways involved in Inclisiran-mediated LDL-C reduction and cardiovascular protection need to be elucidated, and the development of novel siRNA drugs targeting other genes or molecules related to lipid metabolism or cardiovascular disease may offer new opportunities and possibilities for personalized and precision medicine. Compared with PCSK9 antibodies, Inclisiran had the advantages of less frequent administration, lower injection volume, and potentially lower cost. However, it should be noted that Inclisiran had not yet proven its ability to reduce cardiovascular events and mortality in large-scale trials, while PCSK9 antibodies had already demonstrated such benefits in several studies. Therefore, the clinical outcomes of Inclisiran need to be further confirmed by ongoing or future trials. To summarize the main points of our review, we had provided a graphical abstract in [Figure 3](#) (See [Figure 3](#)).

7 Prospects

Recently, the indication of Inclisiran has been expanded to populations with high cardiovascular risk factors but without a history of cardiovascular events, for primary prevention. This expanded indication provides a new option for early intervention in high-risk populations. The sustained lipid-lowering effect and convenient dosing regimen of Inclisiran may help improve medication adherence and therapeutic outcomes in these patients. This opens up a promising primary prevention market for Inclisiran. We look forward to more clinical study results to verify the long-term safety and efficacy of Inclisiran in diverse populations.

Author contributions

YZ: Conceptualization, Methodology, Writing—original draft. HC: Conceptualization, Software, Visualization, Writing—review and editing. LH: Funding acquisition, Supervision, Writing—review

and editing. HW: Project administration, Supervision, Writing—review and editing. BL: Project administration, Supervision, Writing—review and editing. MZ: Project administration, Writing—review and editing. JL: Software, Writing—review and editing. LY: Conceptualization, Methodology, Writing—review and editing, Writing—original draft. FL: Conceptualization, Methodology, Writing—review and editing.

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Conflict of interest

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References

- Barale, C., Melchionda, E., Morotti, A., and Russo, I. (2021). Pcsk9 biology and its role in atherothrombosis. *Int. J. Mol. Sci.* 22. doi:10.3390/ijms22115880
- Brandts, J., and Ray, K. K. (2021). Clinical implications and outcomes of the orion phase iii trials. *Future Cardiol.* 17, 769–777. doi:10.2217/fca-2020-0150
- Desai, N. R., Campbell, C., Electricwala, B., Petrou, M., Trueman, D., Woodcock, F., et al. (2022). Cost effectiveness of inclisiran in atherosclerotic cardiovascular patients with elevated low-density lipoprotein cholesterol despite statin use: a threshold analysis. *Am. J. Cardiovasc. Drugs* 22, 545–556. doi:10.1007/s40256-022-00534-9
- Diaz, R., Li, Q. H., Bhatt, D. L., Bittner, V. A., Baccara-Dinet, M. T., Goodman, S. G., et al. (2021). Intensity of statin treatment after acute coronary syndrome, residual risk, and its modification by alirocumab: insights from the odyssey outcomes trial. *Eur. J. Prev. Cardiol.* 28, 33–43. doi:10.1177/2047487320941987
- Dong, C., Bu, X., Liu, J., Wei, L., Ma, A., and Wang, T. (2022). Cardiovascular disease burden attributable to dietary risk factors from 1990 to 2019: a systematic analysis of the global burden of disease study. *Nutr. Metab. Cardiovasc. Dis.* 32, 897–907. doi:10.1016/j.numecd.2021.11.012
- Ference, B. A., Wiklund, O., Ginsberg, H. N., Krauss, R. M., Tokgozoglu, L., Graham, I., et al. (2013). Low-density lipoproteins cause atherosclerotic cardiovascular disease. 1. Evidence from genetic, epidemiologic, and clinical studies. A consensus statement from the European Atherosclerosis Society Consensus Panel. *Eur. Heart. J.* 38, 2459–2472. doi:10.1093/eurheartj/ehx144
- Fitzgerald, K., White, S., Borodovsky, A., Bettencourt, B. R., Strahs, A., Clausen, V., et al. (2017). A highly durable rna therapeutic inhibitor of pcsk9. *N. Engl. J. Med.* 376, 41–51. doi:10.1056/NEJMoa1609243
- German, C. A., and Shapiro, M. D. (2020). Small interfering rna therapeutic inclisiran: a new approach to targeting pcsk9. *Biodrugs* 34, 1–9. doi:10.1007/s40259-019-00399-6
- Hardy, J., Niman, S., Pereira, E., Lewis, T., Reid, J., Choksi, R., et al. (2021). A critical review of the efficacy and safety of inclisiran. *Am. J. Cardiovasc. Drugs* 21, 629–642. doi:10.1007/s40256-021-00477-7
- Hu, B., Zhong, L., Weng, Y., Peng, L., Huang, Y., Zhao, Y., et al. (2020). Therapeutic sirna: state of the art. *Signal Transduct. Target Ther.* 5, 101. doi:10.1038/s41392-020-0207-x
- Jahangir, A., Sahra, S., and Krzyzak, M. (2021). Can clinicians start prescribing inclisiran for hypercholesterolemia today? A review of clinical studies for internal medicine physicians and endocrinologists. *Cureus* 13, e16664. doi:10.7759/cureus.16664
- Jukema, J. W., Zijlstra, L. E., Bhatt, D. L., Bittner, V. A., Diaz, R., Drexel, H., et al. (2019). Effect of alirocumab on stroke in odyssey outcomes. *Circulation* 140, 2054–2062. doi:10.1161/CIRCULATIONAHA.119.043826

- Kaddoura, R., Orabi, B., and Salam, A. M. (2020). Pcsk9 monoclonal antibodies: an overview. *Heart Views* 21, 97–103. doi:10.4103/HEARTVIEWS.HEARTVIEWS_20_20
- Khvorova, A. (2017). Oligonucleotide therapeutics - a new class of cholesterol-lowering drugs. *N. Engl. J. Med.* 376, 4–7. doi:10.1056/NEJMp1614154
- Kosmas, C. E., Munoz, E. A., Sourlas, A., Silverio, D., Hilario, E., Montan, P. D., et al. (2018). Inclisiran: a new promising agent in the management of hypercholesterolemia. *Diseases* 6. doi:10.3390/diseases6030063
- Leiter, L. A., Teoh, H., Kallend, D., Wright, R. S., Landmesser, U., Wijngaard, P., et al. (2019). Inclisiran lowers ldl-c and pcsk9 irrespective of diabetes status: the orion-1 randomized clinical trial. *Diabetes Care* 42, 173–176. doi:10.2337/dc18-1491
- Liu, Y., Liu, F., Zhang, L., Li, J., Kang, W., Cao, M., et al. (2021). Association between low density lipoprotein cholesterol and all-cause mortality: results from the nhanes 1999–2014. Berlin, Germany: Nature Publishing Group.
- Lu, Y., Zhang, H., Lu, J., Ding, Q., Li, X., Wang, X., et al. (2021). Prevalence of dyslipidemia and availability of lipid-lowering medications among primary health care settings in China. *Jama Netw. Open* 4, e2127573. doi:10.1001/jamanetworkopen.2021.27573
- Luo, Z., Huang, Z., Sun, F., Guo, F., Wang, Y., Kao, S., et al. (2023). The clinical effects of inclisiran, a first-in-class ldl-c lowering sirna therapy, on the ldl-c levels in Chinese patients with hypercholesterolemia. *J. Clin. Lipidol.* 2023. doi:10.1016/j.jacl.2023.04.010
- Meng, Z., and Lu, M. (2017). Rna interference-induced innate immunity, off-target effect, or immune adjuvant? *Front. Immunol.* 8, 331. doi:10.3389/fimmu.2017.00331
- Mercep, I., Friscic, N., Strikic, D., and Reiner, Z. (2022). Advantages and disadvantages of inclisiran: a small interfering ribonucleic acid molecule targeting pcsk9-a narrative review. *Cardiovasc Ther.*, 2022, 8129513. doi:10.1155/2022/8129513
- Miname, M. H., Rocha, V. Z., and Santos, R. D. (2021). The role of rna-targeted therapeutics to reduce ascvd risk: what have we learned recently? *Curr. Atheroscler. Rep.* 23, 40. doi:10.1007/s11883-021-00936-1
- Mortensen, M. B., and Nordestgaard, B. G. (2020). Elevated ldl cholesterol and increased risk of myocardial infarction and atherosclerotic cardiovascular disease in individuals aged 70–100 years: a contemporary primary prevention cohort. *Lancet* 396, 1644–1652. doi:10.1016/S0140-6736(20)32233-9
- Ray, K. K., Troquay, R., Visseren, F., Leiter, L. A., Scott, W. R., Vikarunnessa, S., et al. (2023). Long-term efficacy and safety of inclisiran in patients with high cardiovascular risk and elevated ldl cholesterol (orion-3): results from the 4-year open-label extension of the orion-1 trial. *Lancet Diabetes Endocrinol.* 11, 109–119. doi:10.1016/S2213-8587(22)00353-9
- Ray, K. K., Wright, R. S., Kallend, D., Koenig, W., Leiter, L. A., Raal, F. J., et al. (2020). Two phase 3 trials of inclisiran in patients with elevated ldl cholesterol. *N. Engl. J. Med.* 382, 1507–1519. doi:10.1056/NEJMoa1912387
- Reyes-Soffer, G., Pavlyha, M., Ngai, C., Thomas, T., Holleran, S., Ramakrishnan, R., et al. (2017). Effects of pcsk9 inhibition with alirocumab on lipoprotein metabolism in healthy humans. *Circulation* 135, 352–362. doi:10.1161/CIRCULATIONAHA.116.025253
- Robinson, J. G., Farnier, M., Krempf, M., Bergeron, J., Luc, G., Aversa, M., et al. (2015). Efficacy and safety of alirocumab in reducing lipids and cardiovascular events. *N. Engl. J. Med.* 372, 1489–1499. doi:10.1056/NEJMoa1501031
- Robinson, J. G., Williams, K. J., Gidding, S., Boren, J., Tabas, I., Fisher, E. A., et al. (2018). Eradicating the burden of atherosclerotic cardiovascular disease by lowering apolipoprotein b lipoproteins earlier in life. *J. Am. Heart Assoc.* 7, e9778. doi:10.1161/JAHA.118.009778
- Sabatine, M. S., Giugliano, R. P., Keech, A. C., Honarpour, N., Wiviott, S. D., Murphy, S. A., et al. (2017). Evolocumab and clinical outcomes in patients with cardiovascular disease. *N. Engl. J. Med.* 376, 1713–1722. doi:10.1056/NEJMoa1615664
- Schmidt, A. F., Swerdlow, D. I., Holmes, M. V., Patel, R. S., Fairhurst-Hunter, Z., Lyall, D. M., et al. (2017). Pcsk9 genetic variants and risk of type 2 diabetes: a mendelian randomisation study. *Lancet Diabetes Endocrinol.* 5, 97–105. doi:10.1016/S2213-8587(16)30396-5
- Scicchitano, P., Milo, M., Mallamaci, R., De Palo, M., Caldarola, P., Massari, F., et al. (2021). Inclisiran in lipid management: a literature overview and future perspectives. *Biomed. Pharmacother.* 143, 112227. doi:10.1016/j.biopha.2021.112227
- Shi, J., Zhang, W., Niu, Y., Lin, N., Li, X., Zhang, H., et al. (2020). Association of circulating proprotein convertase subtilisin/kexin type 9 levels and the risk of incident type 2 diabetes in subjects with prediabetes: a population-based cohort study. *Cardiovasc Diabetol.* 19, 209. doi:10.1186/s12933-020-01185-3
- Stroes, E., Colquhoun, D., Sullivan, D., Civeira, F., Rosenson, R. S., Watts, G. F., et al. (2014). Anti-pcsk9 antibody effectively lowers cholesterol in patients with statin intolerance: the gauss-2 randomized, placebo-controlled phase 3 clinical trial of evolocumab. *J. Am. Coll. Cardiol.* 63, 2541–2548. doi:10.1016/j.jacc.2014.03.019
- Tibolla, G., Norata, G. D., Artali, R., Meneghetti, F., and Catapano, A. L. (2011). Proprotein convertase subtilisin/kexin type 9 (pcsk9): from structure-function relation to therapeutic inhibition. *Nutr. Metab. Cardiovasc Dis.* 21, 835–843. doi:10.1016/j.numecd.2011.06.002
- Wright, R. S., Collins, M. G., Stoenbroek, R. M., Robson, R., Wijngaard, P., Landmesser, U., et al. (2020). Effects of renal impairment on the pharmacokinetics, efficacy, and safety of inclisiran: an analysis of the orion-7 and orion-1 studies. *Mayo Clin. Proc.* 95, 77–89. doi:10.1016/j.mayocp.2019.08.021
- Wright, R. S., Ray, K. K., Raal, F. J., Kallend, D. G., Jaros, M., Koenig, W., et al. (2021). Pooled patient-level analysis of inclisiran trials in patients with familial hypercholesterolemia or atherosclerosis. *J. Am. Coll. Cardiol.* 77, 1182–1193. doi:10.1016/j.jacc.2020.12.058



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Anti-atherosclerosis mechanisms associated with regulation of non-coding RNAs by active monomers of traditional Chinese medicine

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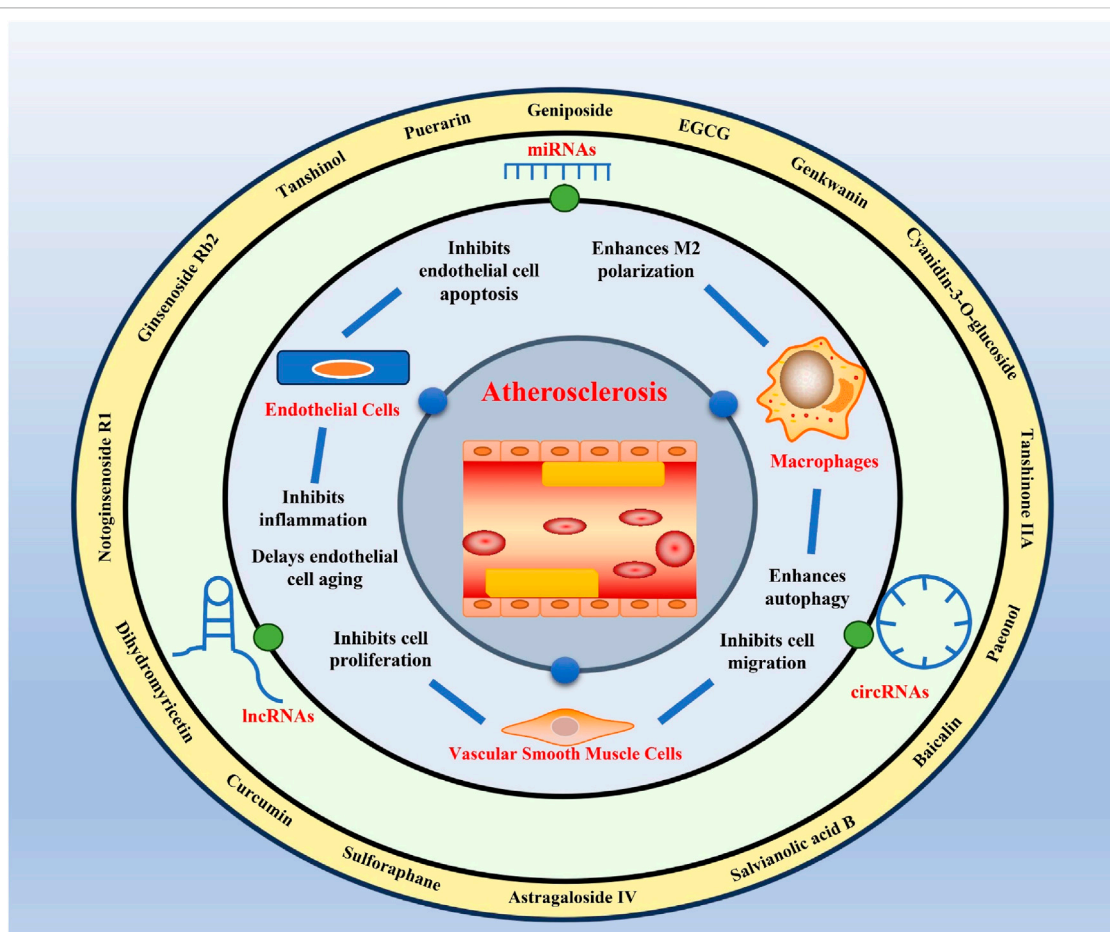
Atherosclerosis is the leading cause of numerous cardiovascular diseases with a high mortality rate. Non-coding RNAs (ncRNAs), RNA molecules that do not encode proteins in human genome transcripts, are known to play crucial roles in various physiological and pathological processes. Recently, researches on the regulation of atherosclerosis by ncRNAs, mainly including microRNAs, long non-coding RNAs, and circular RNAs, have gradually become a hot topic. Traditional Chinese medicine has been proved to be effective in treating cardiovascular diseases in China for a long time, and its active monomers have been found to target a variety of atherosclerosis-related ncRNAs. These active monomers of traditional Chinese medicine hold great potential as drugs for the treatment of atherosclerosis. Here, we summarized current advancement of the molecular pathways by which ncRNAs regulate atherosclerosis and mainly highlighted the mechanisms of traditional Chinese medicine monomers in regulating atherosclerosis through targeting ncRNAs.

KEYWORDS

non-coding RNAs, traditional Chinese medicine, atherosclerosis, cardiovascular diseases, active monomer

1 Introduction

Atherosclerosis is characterized by fibro-fat lesions on the walls of arteries with extremely high morbidity and mortality rate (Libby et al., 2019). Atherosclerosis is considered to be an important pathological basis for cerebrovascular and cardiovascular diseases such as cerebral infarction, coronary heart disease and myocardial infarction (Libby, 2021). There are many causes of atherosclerosis, such as inflammatory reactions (Zhu et al., 2018), cell death and aging (Bazioti et al., 2022), and endothelial to mesenchymal transition (Liang et al., 2022), among which chronic inflammation is the reason that has been frequently studied in the past few years. At the molecular level, telomere damage, genomic DNA damage, and mitochondrial DNA damage accumulate in vascular endothelial cells, which induce endothelial cell aging and chronic inflammation.



GRAPHICAL ABSTRACT

Persistent inflammation results in increased accumulation of lymphocytes and macrophages, leading to atherosclerosis (Wang and Bennett, 2012; Okuyama et al., 2015; Ruparelia and Choudhury, 2020). The pathogenesis and therapeutic targets of atherosclerosis have long been the focus in the field of cardiovascular researches. Statins, inhibitors of the hydroxymethylglutaryl-CoA (HMG-CoA) reductase enzyme, are powerful cholesterol lowering medications and the most commonly used clinical drugs for the prevention and treatment of atherosclerosis (Okuyama et al., 2015). Statins can reduce morbidity and mortality in patients with cardiovascular diseases. However, statins may affect the drug-drug interactions because different safety and tolerability, especially when in combination with other cardiovascular drugs, which will lead to increased risk of statin-associated hepatotoxicity and myopathy (Bellosa and Corsini, 2018). Therefore, it is urgent to discover new therapeutic targets and new drugs for atherosclerosis.

RNAs in mammalian cells are complex, some of which have the function of encoding proteins, but some of which lack this function. At present, the RNAs that lack the function of encoding proteins are named as non-coding RNAs (ncRNAs), of which the most studied are microRNAs (miRNAs), long non-coding RNAs (lncRNAs), and circular RNAs (circRNAs) (Lu and Rothenberg,

2018; Chen et al., 2021a; Bridges et al., 2021). ncRNAs have been proved to play important regulatory roles in pathogenesis of atherosclerosis through affecting inflammatory reaction, cell activation and proliferation, and lipid metabolism (Feinberg and Moore, 2016; Aryal and Suarez, 2019). Nowadays, therapeutic strategies targeting ncRNAs have entered the clinical testing phase for the treatment of cancers and have been considered as an attractive approach for the treatment of atherosclerosis.

Since ancient times, numerous herbal medicines have been used for the treatment of atherosclerosis-related diseases and decoction is the main form of traditional Chinese medicine used in clinic. With the development of separation technology, it has become possible to separate more pharmacologically active monomers from traditional Chinese medicine. This allows researchers to conduct pharmacological studies on the specific monomers rather than the whole of medicinal plants. At present, studies have found that many active monomers of traditional Chinese medicine have positive effects on atherosclerosis, such as saponins (Luo et al., 2022), flavonoids (Park et al., 2006) and alkaloids (Li et al., 2021d). For example, berberine, an active ingredient extracted from *Berberis aristata* DC., can activate PPAR- γ pathway in macrophages, resulting in decreased expressions of inflammatory factors like monocyte chemoattractant protein-1 (MCP-1) and tumor

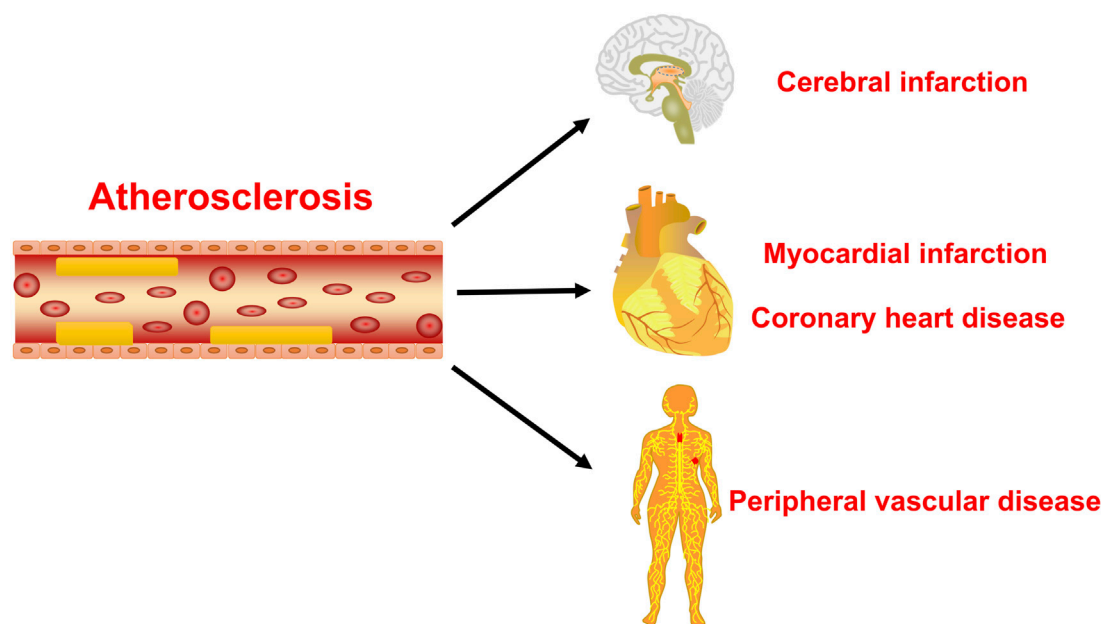


FIGURE 1

The main hazards of atherosclerosis. The atherosclerosis is the main cause of coronary heart disease, cerebral infarction, and peripheral vascular disease.

necrosis factor- α (TNF- α) (Chen et al., 2008). Another study found that hydroxysafflor yellow A, a natural compound from *Carthamus tinctorius* L., exerts protective effects on atherosclerosis by suppressing vascular endothelial cell dysfunction, vascular smooth muscle cell proliferation and migration, foam cell formation, and platelet activation (Xue et al., 2021). These active monomers of traditional Chinese medicine are predicted to have high therapeutic potential in atherosclerosis treatment. However, the specific drug targets of these active ingredient are not fully understood, which limits clinical application.

Recently, more and more studies have found that ncRNAs are the key mediators of the pharmacological effect of traditional Chinese medicine. Here, we summarized current advances of mechanisms of ncRNAs in regulating atherosclerosis. Furthermore, we highlighted the current advances in the active monomers of traditional Chinese medicine which have atheroprotective effects by regulating ncRNAs.

2 The role of ncRNAs in regulating atherosclerosis

2.1 miRNAs and atherosclerosis

MiRNAs (typically 20–25 nucleotides) are single-stranded RNA molecules that can bind to complementary sequences within the 3' untranslated region of mRNA targets. Once the miRNA binds to the mRNA, it can degrade the mRNA via cleavage or inhibit the translation of mRNAs into proteins (Winter et al., 2009; Thum and Mayr, 2012). MiRNAs are the most studied ncRNAs in atherosclerosis and have been shown to regulate the fate and

function of atherosclerosis associated cells, including endothelial cells, inflammatory cells, and vascular smooth muscle cells (VSMCs). MiRNAs can affect endothelial cell function by exacerbating senescence of endothelial cells, which is considered as a key mechanism of atherosclerosis (Menghini et al., 2009; Fiedler and Thum, 2016). There are many miRNAs involved in the regulation of endothelial cell senescence, such as miR-146a and miR-217 (Wang et al., 2021; Xiao et al., 2021). Studies have found that mesenchymal stem cell-derived extracellular vesicles attenuate endothelial cell senescence by regulating miR-146a/Src signaling (Xiao et al., 2021). MiR-217 can also promote endothelial cell senescence through the SIRT1/p53 signaling pathway (Wang et al., 2021). In addition, miRNAs can control the inflammatory state of the vasculature by affecting leukocyte activation and infiltration (Perez-Sanchez et al., 2017; Pankratz et al., 2018). In the setting of atherosclerosis, miR-126 promotes macrophage polarization to the M2 phenotype by downregulating VEGFA and krüppel-like factor 4 (KLF4) (Shou et al., 2023). MiRNAs have also been shown to affect foam cell formation and subsequent plaque formation (Eken et al., 2017; Maitrias et al., 2017). MiR-302a has been shown to promote the formation of foam cells and increase the outflow of cholesterol in macrophage by increasing ATP-binding cassette transporter A1 (ABCA1) activity (Meiler et al., 2015). In addition, the function of VSMCs can also be regulated by miRNAs. For example, miR-146b-5p reduces the expression of its target genes Bag1 and Mmp16, thereby affecting the proliferation and migration of VSMCs during atherosclerosis (Sun et al., 2020). A study also found that miR-374 may be a potential biomarker for the diagnosis of atherosclerosis, and overexpression of miR-374 promotes the proliferation and migration of VSMCs (Wang et al., 2020b). MiR-663 can target

TABLE 1 Active monomers of traditional Chinese medicine and their ncRNA targets.

| Active monomers | ncRNA | Target genes | Related hallmark | Model | References |
|------------------------|--------------|--------------|--|---|--|
| Geniposide | miR-101 | MKP-1 | Inhibits inflammation | <i>In vivo</i> : ApoE ^{-/-} mice | Cheng et al. (2019) |
| | | | | <i>In vitro</i> : RAW264.7 | |
| | miR-21 | PTEN | Inhibits inflammation and oxidative stress | <i>In vitro</i> : HUVECs | Zhou et al. (2020) |
| Astragaloside IV | circ-0000231 | miR-135a-5p | Inhibits apoptosis, inflammation, and oxidative stress; Promotes the viability and migration ability | <i>In vitro</i> : HUVECs | Shao et al. (2021) |
| | miR-33a | ABCA1 | Promotes the outflow of cholesterol | <i>In vivo</i> : ApoE ^{-/-} mice | Qin et al. (2018) |
| | | | | <i>In vitro</i> : THP-1 | |
| | miR-17-5p | PCSK9/VLDLR | Inhibits inflammation | <i>In vivo</i> : ApoE ^{-/-} mice | Qin et al. (2022) |
| | | | | <i>In vitro</i> : VSMCs | |
| | lncRNA H19 | DUSP5 | Inhibits autophagy and mineralization | <i>In vivo</i> : ApoE ^{-/-} mice C57BL/6J mice | Song et al. (2019) |
| | | | | <i>In vitro</i> : HASMCs | |
| Notoginsenoside R1 | miR-147a | MyD88 | Inhibits inflammation and oxidative stress | <i>In vitro</i> : HUVECs | Li and Huang (2021) |
| | miR-221-3p | TLR4 | Inhibits apoptosis, inflammation, and oxidative stress | <i>In vitro</i> : HUVECs | Zhu et al. (2020) |
| | miR-34a | SIRT1 | Delays aging | <i>In vitro</i> : HUVECs | Lai et al. (2018) |
| Tanshinone IIA | miR-130b | WNT5A | Inhibits inflammation and adipogenesis | <i>In vitro</i> : THP-1 | Yuan et al. (2020) |
| | miR-712-5p | ? | Inhibits inflammation and cell proliferation | <i>In vitro</i> : VSMCs | Qin et al. (2020) |
| | miR-375 | KLF4 | Enhances autophagy and M2 polarization of macrophages | <i>In vivo</i> : ApoE ^{-/-} mice | Chen et al. (2019a) |
| | | | | <i>In vitro</i> : RAW264.7 | |
| | miR-21-5p | TPM1 | Inhibits proliferation and migration | <i>In vitro</i> : HASMCs | Jia et al. (2019) |
| Salvianolic acid B | miR-146a | ? | Inhibits proliferation | <i>In vivo</i> : Carotid bifurcation ligated mice | Zhao et al. (2019) |
| | | | | <i>In vitro</i> : VSMCs | |
| Tanshinol | lncRNA TUG1 | miR-26a | Inhibits apoptosis | <i>In vivo</i> : ApoE ^{-/-} mice | Chen et al. (2016) |
| | | | | <i>In vitro</i> : HAECs, ECV304 cells | |
| Genkwanin | miR-101 | MKP-1 | Inhibits inflammation | <i>In vitro</i> : RAW264.7 | Gao et al. (2014) |
| Dihydromyricetin | miR-21 | DDAH1 | Increases NO production and weakens endothelial dysfunction | <i>In vivo</i> : ApoE ^{-/-} mice | Yang et al. (2018), Yang et al. (2020) |
| | | | | <i>In vitro</i> : HUVECs, THP-1 | |
| Sulforaphane | miR-34a | SIRT1 | Reduces oxidative stress | <i>In vitro</i> : HUVECs | Li et al. (2021c) |
| Cyanidin-3-O-glucoside | miR-204-5p | SIRT1 | Inhibits inflammation and apoptosis | <i>In vivo</i> : Rabbit model of HFD + balloon catheter injury | Wang et al. (2020c) |
| | | | | <i>In vitro</i> : HUVECs | |
| Baicalin | miR-126-5p | HMGB1 | Inhibits proliferation and migration | <i>ex vivo</i> : Blood of atherosclerosis patients and healthy people | Chen et al. (2019b) |
| | | | | <i>In vitro</i> : VSMCs | |
| Curcumin | lncRNA MIAT | EZH2 | Inhibits inflammation | <i>In vivo</i> : ApoE ^{-/-} mice | Ouyang et al. (2022) |
| | | | | <i>In vitro</i> : HUVECs | |
| | miR-125a-5p | SIRT6 | Promotes cholesterol efflux | <i>In vitro</i> : THP-1 | Tan et al. (2021) |

(Continued on following page)

TABLE 1 (Continued) Active monomers of traditional Chinese medicine and their ncRNA targets.

| Active monomers | ncRNA | Target genes | Related hallmark | Model | References |
|-----------------|------------|--------------|--|---|---------------------|
| EGCG | miR-33a | ABCA1 | Promotes cholesterol efflux | <i>In vitro</i> : THP-1 | Yang et al. (2016) |
| Ginsenoside Rb2 | miR-216a | Smad3 | Inhibits inflammation and aging | <i>In vitro</i> : HUVECs, HAECs | Chen et al. (2021b) |
| Paeonol | miR-223 | STAT3 | Inhibits inflammation | <i>In vivo</i> : ApoE ^{-/-} mice | Liu et al. (2018) |
| | | | | <i>In vitro</i> : HUVECs, THP-1 | |
| | miR-223 | ? | Inhibits inflammation | <i>In vivo</i> : SD rats | Shi et al. (2020) |
| | | | | <i>In vitro</i> : RAECs | |
| | miR-126 | VCAM-1 | Inhibits monocyte adhesion to endothelial cells | <i>In vivo</i> : SD rats | Yuan et al. (2016) |
| | | | | <i>In vitro</i> : VECs isolated from the thoracic aorta of rats | |
| | miR-21 | PTEN | Inhibits inflammation | <i>In vivo</i> : SD rats | Liu et al. (2014) |
| | | | | <i>In vitro</i> : VECs isolated from the thoracic aorta of rats | |
| Puerarin | miR-30a | Beclin-1 | Inhibits autophagy | <i>In vivo</i> : SD rats | Li et al. (2018a) |
| | | | | <i>In vitro</i> : VECs isolated from the thoracic aorta of rats | |
| | miR-338-3p | TET2 | Inhibiting apoptosis, inflammation, and oxidative stress | <i>In vitro</i> : VECs isolated from the thoracic aorta of mice | Yu et al. (2020b) |
| Puerarin | miR-29b-3p | IGF1 | Inhibits inflammation and proliferation | <i>In vivo</i> : ApoE ^{-/-} mice | Li et al. (2023) |
| | | | | <i>In vitro</i> : hVSMCs | |

MKP-1, mitogen-activated protein kinase phosphatase 1; PTEN, phosphatase and tensin homolog, ABCA1 ATP-binding cassette transporter A1, PCSK9 proprotein convertase subtilisin/kexin type 9, VLDLR, very low-density lipoprotein receptor, KLF4 krüppel-like factor 4, DUSP5 dual specificity phosphatase 5, MyD88 myeloid differentiation primary response 88, TLR4 toll-like receptor 4, SIRT1 sirtuin-1, p53 tumor protein 53, WNT5A wingless/integrated-5A, TPM1 tropomyosin 1, DDAH1 dimethylarginine dimethylaminohydrolase 1, HMGB1 high mobility group box 1 protein, EZH2 enhancer of zeste homolog 2, Smad3 sma- and mad-related protein 3, STAT3 signal transducer and activator of transcription 3, VCAM-1, Vascular cell adhesion molecule-1, IGF1 insulin-like growth factor 1, ApoE^{-/-} mice apolipoprotein e-knockout mice, RAW264.7 RAW, 264.7 mouse leukemia macrophage cell line, HUVECs, human umbilical vein endothelial cells; THP-1, human acute monocytic leukemia cell line; VSMCs, vascular smooth muscle cells; HASMCs, human aortic vascular smooth muscle cells; HAEC, human aortic endothelial cells; SD, rats sprague-dawley rats; RAECs, rat aortic endothelial cells; VECs, vascular endothelial cells; hVSMCs, human vascular smooth muscle cells, TET2 tet methylcytosine dioxygenase 2.

high mobility group AT-hook 2 (HMGA2) to inhibit the proliferation of VSMCs, thereby delaying the development of atherosclerosis (Deng and Li, 2022). In conclusion, miRNAs regulate atherosclerosis through affecting the function of endothelial cells, macrophages, and VSMCs.

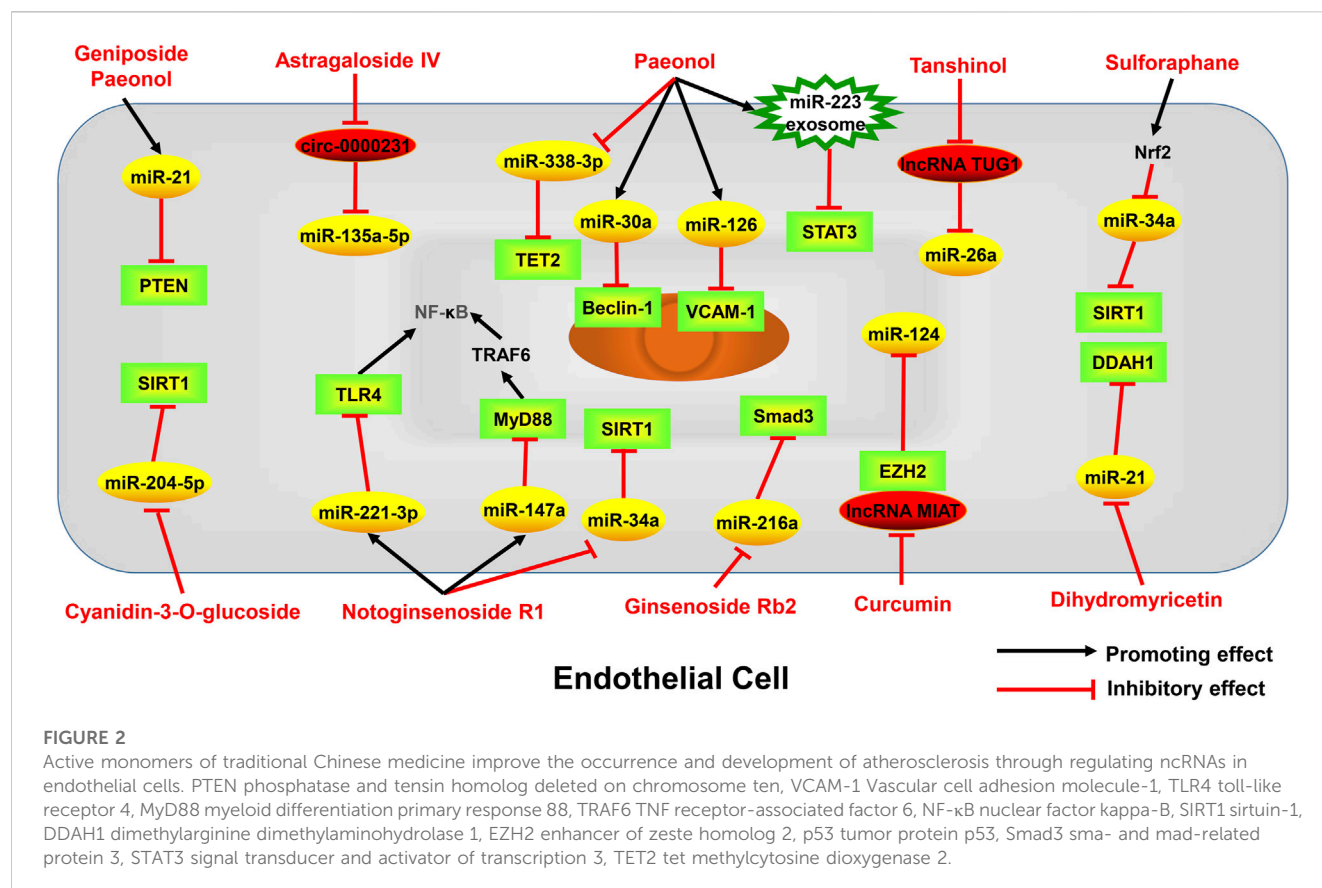
2.2 lncRNAs and atherosclerosis

lncRNAs are ncRNAs longer than 200 nucleotides (Di Mauro et al., 2018), which are abnormally expressed in many pathological tissues (Li et al., 2020; Zang et al., 2020). Unlike miRNAs, the actions of lncRNAs are relatively complex. lncRNA can be a source of miRNA. For example, miR-31 gene is embedded within an intron of the lncRNA LOC554202 and its transcription is regulated by the methylation state of the host gene promoter (Augoff et al., 2012). Moreover, lncRNAs can bind to DNA, mRNA and proteins to regulate their expressions or functions (Guttman and Rinn, 2012). The most widely known mechanism is the competitive endogenous RNA (ceRNA), in which way lncRNA acts as a negative regulator of miRNA (Salmena et al., 2011). In recent years, studies have shown that lncRNAs are dynamically expressed in developing and diseased blood vessels, suggesting

that lncRNAs have profound biological functions in atherosclerosis (Guo et al., 2019; Simion et al., 2020). lncRNAs can regulate atherosclerosis by influencing the function of vascular cells. For example, lncRNA HOXA11-AS is significantly upregulated in aortic tissue of atherosclerotic mice and oxidized low-density lipoprotein (ox-LDL)-induced endothelial cells. HOXA11-AS knockdown attenuates endothelial injuries by directly regulating the miR-515-5p/ROCK1/eNOS axis (Gao et al., 2022). In addition to endothelial cells, lncRNA also plays a role in atherosclerosis by affecting VSMCs and macrophages. For example, lncRNA TUG1 can promote the proliferation of VSMCs by regulating the miRNA-21/PTEN axis (Li et al., 2018b). lncRNA MAARS interacts with HuR to increase macrophage apoptosis in the blood vessels (Simion et al., 2020). What's more, lncRNA kcnq1ot1 can compete with miR-452-3p to promote macrophage lipid accumulation and accelerate the development of atherosclerosis (Yu et al., 2020a).

2.3 circRNAs and atherosclerosis

CircRNAs are closed circular molecules, which distinguishes them from other linear RNA molecules. CircRNAs were originally considered as by-products of mRNA cleavage, but now they are



thought to be stable and functional ncRNAs (Chen, 2016). Compared to miRNAs, circRNAs are less studied ncRNAs in atherosclerosis. Still, there are studies that have shown circRNAs can regulate the fate and function of atherosclerosis-associated cells, including endothelial cells, macrophages, and VSMCs. As with lncRNAs, circRNAs can also compete with miRNAs as ceRNAs, which is the mostly investigated mechanism (Ren et al., 2021). In endothelial cells, a study demonstrated that circ-RELL1 plays a pro-inflammatory role in endothelial cells by directly binding to miR-6873-3p and subsequently activating NF-κB signaling pathway (Huang et al., 2020). Circ_0086296 induces aberrant endothelial cell phenotypes by sponging miR-576-3p, resulting in severe atherosclerotic lesions (Zhang et al., 2022). In VSMCs, circRNA-0044073 promotes the proliferation and invasion of VSMCs by targeting miR-107 and activating the JAK/STAT signaling pathway (Shen et al., 2019). In macrophages, overexpression of circ_0004104 results in dysregulation of atherosclerosis-related genes in THP-1-derived macrophages (Wang et al., 2019). It is noticed that the role of circRNAs in atherosclerosis has rarely been studied, which may become a research hotspot in the future.

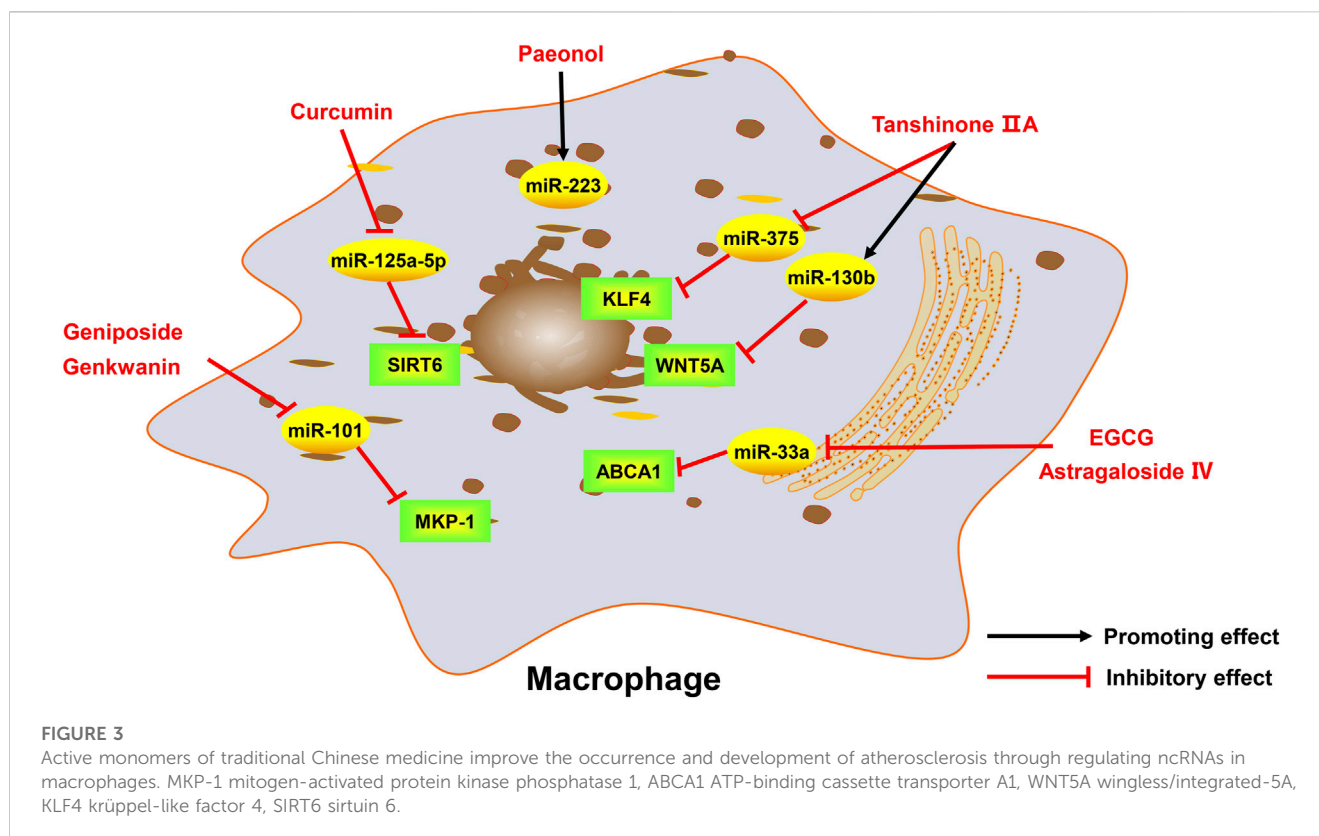
Since the role of ncRNAs in atherosclerosis is emerging, they have been considered as potential drug targets in developing therapeutic agents. As we know, traditional Chinese medicine has a long history of treating atherosclerosis in China. In particular, studies have shown that the monomers extracted from traditional Chinese medicine are the main functional components that possess anti-atherosclerotic activity, and these activities can be mediated by ncRNAs.

3 Active monomers of traditional Chinese medicine relieve atherosclerosis by regulating ncRNAs

Nowadays, the researches about the regulation of atherosclerosis by active monomers of traditional Chinese medicine are tremendous. However, the drug targets of traditional Chinese medicine remain unclear, which affects the clinical application of these medicine. It is clear that ncRNAs appear to be important players during atherosclerosis and important targets of traditional Chinese medicine. Therefore, it is particularly important to discover the mechanism by which the active monomers of traditional Chinese medicine relieve atherosclerosis through ncRNAs.

3.1 Geniposide

Geniposide, an iridoid glucoside, is the main active ingredient of *Gardenia jasminoides* J. Ellis. Geniposide exhibits a variety of anti-inflammatory and anti-oxidative functions and has good therapeutic effects on cardiovascular diseases (Fu et al., 2012b). A study has found that geniposide treatment reduces lipid levels and plaque size in the mouse model of atherosclerosis. Mechanistically, geniposide downregulates miR-101 to upregulate mitogen-activated protein kinase phosphatase-1 (MKP-1) and suppresses the production of inflammatory factors in macrophages (Cheng et al., 2019). MiR-21 has been shown to play an important role in regulating inflammatory responses by targeting phosphatase and tensin



homolog (PTEN) (Sheedy, 2015; Li et al., 2022). A study established a endothelial cell injury model by using ox-LDL and found geniposide protects endothelial cells from ox-LDL-induced injury by inhibiting oxidative stress and inflammation, and these effects are partly due to the enhancement of the miR-21/PTEN pathway (Zhou et al., 2020). Taken together, miR-101 and miR-21 are involved in the anti-inflammatory effect of geniposide in the setting of atherosclerosis.

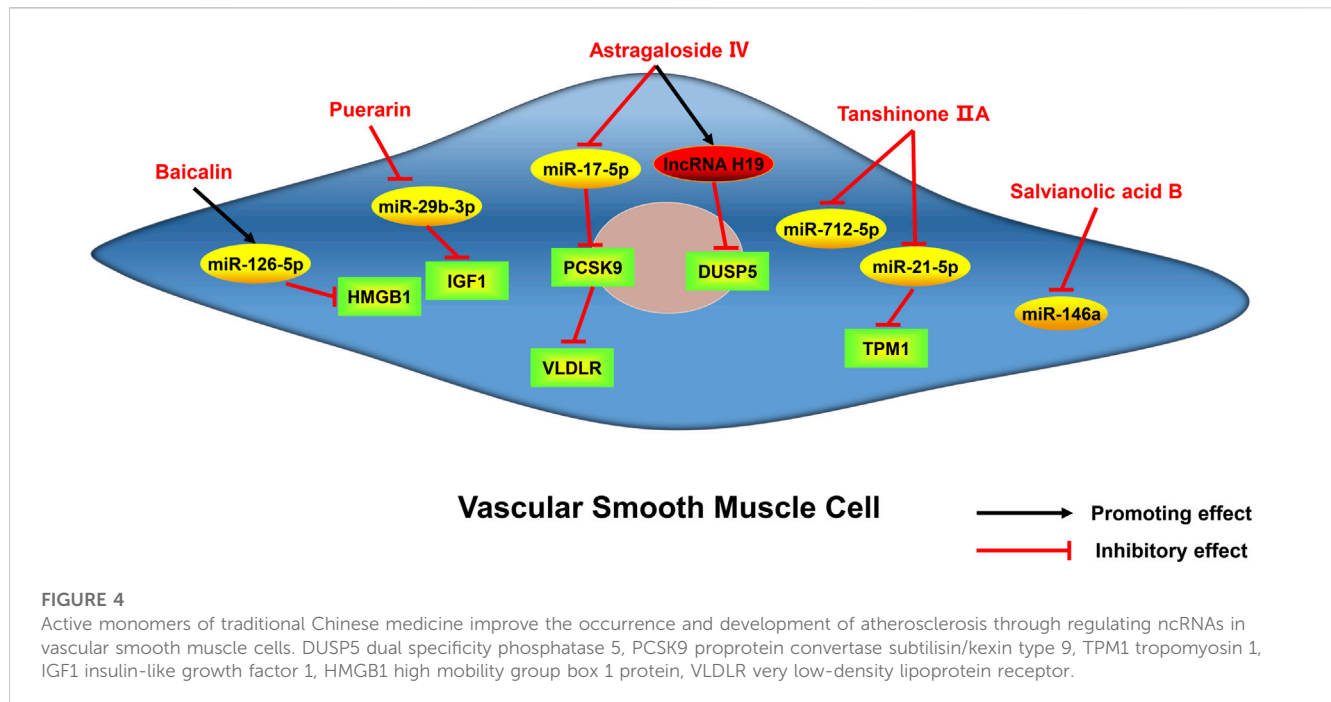
3.2 Astragaloside IV

Astragaloside IV is a saponin isolated from *Astragalus membranaceus* (Fisch.) Bunge, which has excellent cardioprotective effects (Tan et al., 2020). Astragaloside IV has been reported to protect endothelial cells from oxidative damage caused by ox-LDL through regulating the LOX-1/NLRP3 signaling pathway (Qian et al., 2019a). Recently, a study found that circ_0000231 is the key downstream target of astragaloside IV, which regulates miR-135a-5p to target chloride intracellular channel 4 (CLIC4), and contributes to the protective role of astragaloside IV in ox-LDL-induced endothelial cell injury (Shao et al., 2021). CLIC4 is also a protein associated with endothelial cell apoptosis (Zhang et al., 2020b), indicating astragaloside IV may also inhibit endothelial cell apoptosis by regulating CLIC4 through circ_0000231. Several miRNAs have been shown to be the targets of astragaloside IV. For example, astragaloside IV can protect cardiomyocytes from hypoxia-induced injury by downregulating miR-23a and miR-92a (Gong et al., 2018). ABCA1, a membrane transporter that mediates cholesterol efflux (Chen et al., 2022), has been proved to be a target of miR-33a (Gao et al.,

2018). A study has found that astragaloside IV can promote cholesterol efflux in macrophages and inhibit atherosclerosis through regulating miR-33a/ABCA1 pathway (Qin et al., 2018). The serum miR-17-5p is elevated in patients with atherosclerosis and miR-17-5p knockdown can alleviate atherosclerotic lesions and inhibit the proliferation and migration of VSMCs by directly up-regulating very low density lipoprotein receptor (VLDLR), or indirectly regulate VLDLR by affecting proprotein convertase subtilisin kexin 9 (PCSK9) (Tan et al., 2017). Astragaloside IV has been shown to downregulate miR-17-5p and further affect VLDLR expression, thus inhibiting vascular inflammation (Qin et al., 2022). In addition, lncRNA H19 has also been reported to mediate astragaloside IV's anti-atherosclerotic effect. H19 negatively regulates dual-specificity phosphatase 5 (DUSP5) expression and represses DUSP5/ERK1/2 axis (Tao et al., 2016). Astragaloside IV could attenuate autophagy and mineralization of VSMCs in atherosclerosis by up-regulating H19 and inhibiting DUSP5 (Song et al., 2019). In summary, astragaloside IV can regulate the function of endothelial cells, VSMCs, and macrophages in atherosclerosis by targeting multiple miRNAs, lncRNAs and circRNAs. Therefore, it can be expected that astragaloside IV can exert an excellent anti-atherosclerotic effect through ncRNAs in the clinic.

3.3 Notoginsenoside R1

Notoginsenoside R1, the monomer extracted from *Panax notoginseng* (Burkill) F.H.Chen, has a unique effect of promoting



blood circulation and has been used on clinical treatment of cardiovascular diseases (Lei et al., 2022). Myeloid differentiation primary response gene 88 (MyD88) is an important immunoregulatory factor, and studies have found that inhibiting MyD88 has a good effect on diabetes (Androulidaki et al., 2018). Notoginsenoside R1 was found to relieve high glucose-induced endothelial cell inflammation and oxidative stress by downregulating the MyD88 via up-regulating miR-147a (Li and Huang, 2021). The Toll-like receptor 4 (TLR4)/Nuclear factor- κ B (NF- κ B) pathway participates in oxidative stress and induces atherosclerosis in ApoE^{-/-} mice by up-regulating inflammatory cytokines (Tang et al., 2015). A study revealed that notoginsenoside R1 could upregulate the expression of miR-221-3p to target TLR4/NF- κ B pathway, thereby inhibiting ox-LDL-induced endothelial cell apoptosis, oxidative stress, and inflammation (Zhu et al., 2020). Notoginsenoside R1 may also play a role in delaying senescence of endothelial cells. Notoginsenoside R1 can decrease the expressions of miR-34a and p53, while increase the expression of SIRT1, thus enhancing the intracellular superoxide dismutase (SOD) activity and cell proliferation capacity in hydrogen peroxide-induced endothelial cell aging model (Lai et al., 2018). These studies suggest that notoginsenoside R1 has a strong and multifaceted endothelial protective effect through regulating ncRNAs.

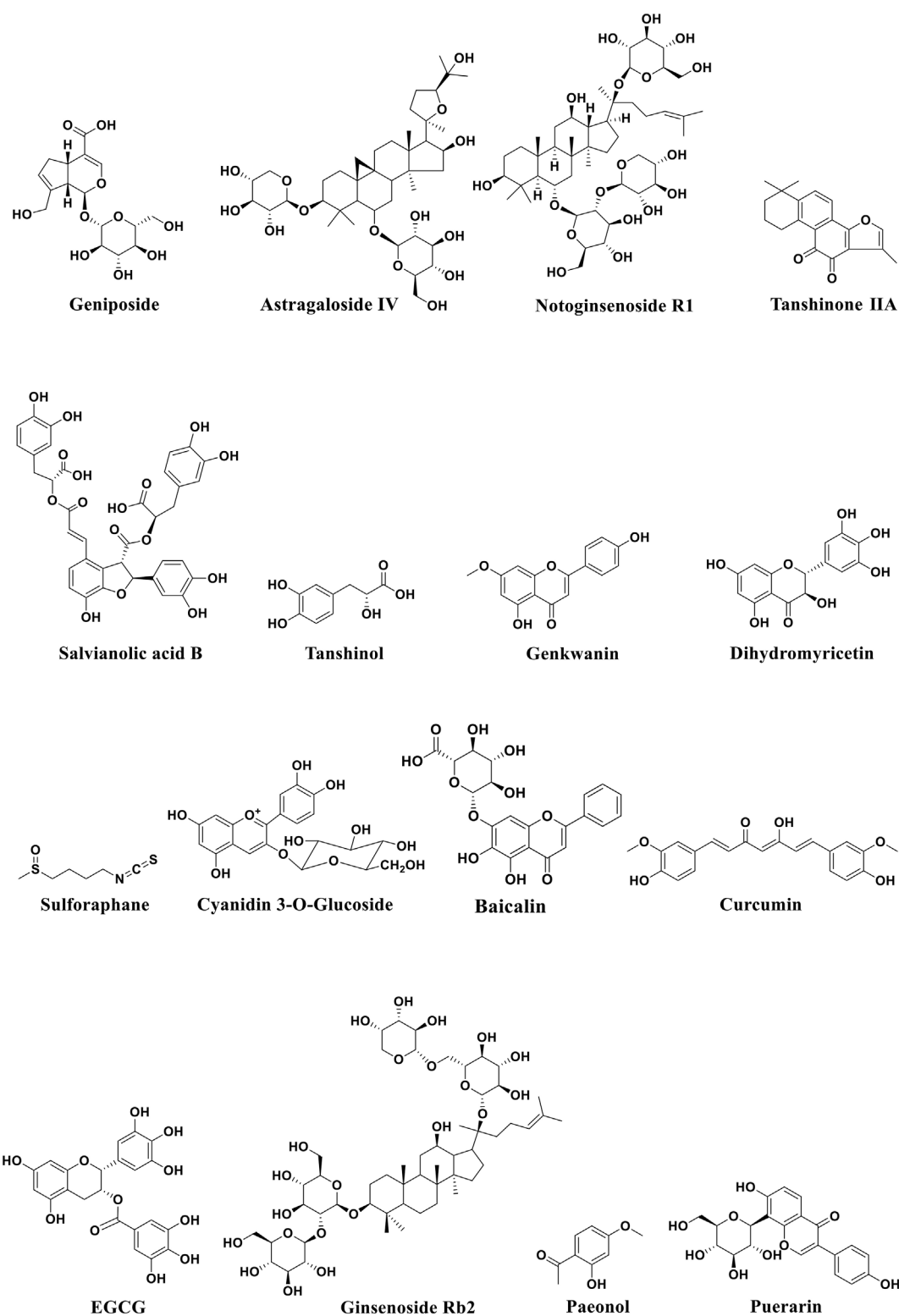
3.4 Tanshinone IIA, salvianolic acid B, tanshinol

Tanshinone, extracted from the traditional Chinese medicine *Salvia miltiorrhiza* Bunge, is a fat-soluble phenanthrene quinone compound with bacteriostatic effect (Wang et al., 2017). Among tanshinone, tanshinone IIA has been clinically proved to have a more significant effect on cardiovascular diseases, especially its

anti-inflammatory effect on macrophages. Tanshinone IIA reduces the production of inflammatory factors and adipogenesis in macrophages by up-regulating miR-130b and downregulating WNT5A, thereby relieving the development of atherosclerosis (Yuan et al., 2020). Previous studies have demonstrated that miR-712 is involved in atherosclerosis-related pathological processes, such as VSMCs calcification and endothelial cell inflammation (Son et al., 2013). Tanshinone IIA can inhibit VSMCs inflammation and proliferation by inhibiting miR-712-5p (Qin et al., 2020). KLF4, an evolutionarily conserved zinc-finger-containing transcription factor, is thought to induce M2 and inhibit M1 macrophage polarization (Liao et al., 2011). A study found that the miR-375/KLF4 pathway plays a dominant role in macrophage polarization and autophagy, and tanshinone IIA could activate KLF4 by inhibiting miR-375, leading to enhanced autophagy as well as M2 polarization of macrophages (Chen et al., 2019a). Tropomyosin 1 (TPM1), as a target gene for miR-21-5p (Baker, 2011), is involved in the formation, stabilization and regulation of cytoskeletal actin fibers (Gunning et al., 2015). It was found that tanshinone IIA could downregulate miR-21-5p and then target TPM1, which helps to inhibit the proliferation and migration of VSMCs (Jia et al., 2019).

Salvianolic acid B, a water-soluble compound extracted from *S. miltiorrhiza* Bunge, has been used to treat cardiovascular diseases for hundreds of years. MiR-146a is involved in the regulation of cell proliferation, migration, differentiation, and apoptosis (Cheng et al., 2013). A study has found that salvianolic acid B can inhibit angiotensin II-induced VSMCs proliferation and improve carotid artery ligation-induced neointimal hyperplasia by downregulating miR-146a (Zhao et al., 2019).

Tanshinol is also an active ingredient isolated from *S. miltiorrhiza* Bunge which has the effect of protecting vascular

**FIGURE 5**

Structural formula of active monomers of traditional Chinese medicine that exhibit anti-atherosclerotic activities.

endothelium and reducing atherosclerosis (Song et al., 2014). MiR-26a has been proved to have anti-apoptotic effect on endothelial cells (Zhang et al., 2015). A study found that tanshinol inhibits apoptosis

of endothelial cells and reduces atherosclerotic lesions via decreasing lncRNA TUG1 and increasing miR-26a in endothelial cells (Chen et al., 2016).

3.5 Genkwanin

Genkwanin is one of the major non-glycosylated flavonoids extracted from *Daphne genkwa* Siebold & Zucc. (Bao et al., 2019). MKP-1 is a key negative regulator of macrophage signaling in response to inflammatory stimulus and is responsible for shutting down the production of pro-inflammatory cytokines (Chen et al., 2002; Chi et al., 2006). Genkwanin was proved to potently decrease the production of proinflammatory mediators through downregulating miR-101 and increasing MKP-1 (Gao et al., 2014).

3.6 Dihydromyricetin

Dihydromyricetin, a bioactive flavonoid isolated from *Ampelopsis cantoniensis* var. *grossedentata* Hand. -Mazz. and *Ziziphus jujuba* Mill., has been found to have a wide range of pharmacological activities, such as anti-inflammatory (Sun et al., 2021), analgesic (Guan et al., 2019), anti-tumor (Chen et al., 2020) and hepatoprotective effects (Silva et al., 2020). Nitric oxide (NO), produced by endothelial nitric oxide synthase (eNOS), plays a key role in maintaining endothelial function, and impaired NO biosynthesis is a hallmark of atherosclerosis (Tousoulis et al., 2012; Cyr et al., 2020). There is evidence that overexpression of dimethylarginine dimethylaminohydrolase-1 (DDAH1) increases NO production through an asymmetric dimethylarginine (ADMA) manner (Pope et al., 2009). Studies suggested that dihydromyricetin treatment inhibits atherosclerotic lesion formation by increasing NO production by endothelial cells. MiR-21 expression can be reduced by dihydromyricetin in endothelial cells, which increases DDAH1 and reduces ADMA levels (Yang et al., 2018; Yang et al., 2020). Taken together, dihydromyricetin activates endothelial DDAH1/ADMA/eNOS/NO pathway by reducing miR-21, which relieves the pathogenesis of atherosclerosis.

3.7 Sulforaphane

Sulforaphane is an isothiocyanate, which is produced by the conversion of glucoraphanin through the myrosinase (Vanduchova et al., 2019). Sulforaphane, a potent antioxidant, is primarily found in several Brassicaceae vegetables, such as broccoli, cauliflower, cabbage, and Brussels sprouts. Sulforaphane has often been shown to protect cells from oxidative stress in cardiomyocytes and neural cells (Guerrero-Beltran et al., 2012). The nuclear factor erythroid-2-related factor 2 (Nrf2), a basic leucine zipper transcription factor that serves as a defense mechanism against oxidative stress, has been shown to be activated by sulforaphane (Bai et al., 2015; Houghton et al., 2016). SIRT1 is a potential target gene of miR-34a (Yamakuchi et al., 2008) and the role of the miR-34a/SIRT1 axis in oxidative stress-induced cellular damage has been demonstrated (Guo et al., 2017). Sulforaphane was found to protect endothelial cells from oxidative stress by regulating the miR-34a/SIRT1 axis through upregulation of Nrf2 (Li et al., 2021c). In addition, a study found that sulforaphane can reduce lipopolysaccharide-induced cell damage and oxidative stress by inhibiting miR-155 in microglia (Eren et al., 2018). MiR-155 was

proved to aggravate the carotid atherosclerotic lesion through induction of endothelial cell apoptosis and activation of inflammasome in macrophages (Yin et al., 2019b). Therefore, it is possible that sulforaphane may limit the formation of atherosclerotic lesions by inhibiting miR-155, but clearly, more studies are needed to confirm this hypothesis.

3.8 Cyanidin-3-O-glucoside

Anthocyanins are abundant natural water-soluble pigments, which are relatively rich in the skin of *Glycine max* (L.) Merr. These compounds have been shown to exert antioxidant and anti-inflammatory properties (Zhang et al., 2020a). Cyanidin-3-O-glucoside is one of the most abundant anthocyanins in nature. A study found that cyanidin-3-O-glucoside treatment not only suppresses blood lipids, but also improves endothelial cell function in a rabbit atherosclerotic model. Mechanistically, these effects are due to decreased expression of miR-204-5p, which leads to the increased expression of SIRT1 and enhanced endothelial cell function (Wang et al., 2020c).

3.9 Baicalin

Baicalin, one of the flavonoid compounds, is the main active component of traditional Chinese medicine *Scutellaria baicalensis* Georgi (Li et al., 2009). It has been shown that baicalin can alleviate the development of atherosclerosis through its anti-adipogenic, anti-inflammatory and antioxidant effects (Wu et al., 2018). The expression of miR-126 was found to be reduced in the peripheral blood of atherosclerotic patients (Jiang et al., 2014). High mobility group box 1 protein (HMGB1) is an essential facilitator of atherosclerosis by enhancing inflammation (Boteanu et al., 2017). It has been found that baicalin induces the upregulation of miR-126-5p and the downregulation of HMGB1, inhibiting ox-LDL-induced proliferation and migration of VSMCs (Chen et al., 2019b).

3.10 Curcumin

Curcumin is the main active ingredient of *Curcuma longa* L. and is mainly extracted from dried powdered turmeric. There is evidence that curcumin can modulate the inflammatory response and alleviate inflammatory diseases like atherosclerosis (Hasan et al., 2014; Chen et al., 2015). Studies found that the activated miR-126-3p from endothelial cells and VSMCs plays a key role in reducing vascular calcification (Zeng et al., 2021) and curcumin upregulates miR-126-3p expression (Li et al., 2021b). Therefore, we infer that miR-126-3p may be one of the targets of curcumin in the treatment of atherosclerosis. LncRNA MIAT has been shown to aggravate the atherosclerotic damage through the activation of PI3K/Akt signaling pathway (Sun et al., 2019). A study found that reduced expression of MIAT contributes to the protective effect of curcumin on atherosclerosis. MIAT regulates miR-124 by interacting with enhancer of zeste homolog 2 (EZH2), thereby relieving inflammation in endothelial cells (Ouyang et al., 2022). In addition, curcumin markedly suppresses miR-125a-5p and

upregulates SIRT6 in macrophages, thereby regulating the ABCA1 expression and promoting cholesterol efflux of macrophages (Tan et al., 2021).

3.11 Epigallocatechin gallate

Epigallocatechin gallate (EGCG) is the most abundant catechin in green tea. EGCG has been shown to have various pharmacological effects including the anti-atherosclerotic effect, which is primarily achieved by promoting intracellular cholesterol efflux in macrophages (Jiang et al., 2012). Recent studies showed that miR-33a is an upstream regulator of ABCA1 (Wijesekara et al., 2012) and EGCG exerts anti-atherosclerotic effect by reducing miR-33a, thereby upregulating ABCA1 and promoting the efflux of cholesterol in macrophages (Yang et al., 2016).

3.12 Ginsenoside Rb2

Ginsenoside Rb2, extracted from *Panax ginseng* C.A. Mey., is a commonly used traditional Chinese medicine with antioxidant (Huang et al., 2014), anti-inflammatory (Huang et al., 2017) and anti-apoptotic activities (Gao et al., 2015). In macrophages, ginsenoside Rb2 has been found to exert anti-inflammatory effects by upregulating the expression of an ω -3 fatty acid receptor GPR120 (Huang et al., 2017). A recent study showed that ginsenoside Rb2 can also inhibit endothelial senescence and inflammation. Mechanistically, ginsenoside Rb2 has a specific binding affinity for miR-216a and further attenuates miR-216a-induced inflammatory processes and aging states through the Smad3/I κ B α signaling pathway (Chen et al., 2021b).

3.13 Paeonol

Paeonol is one of the main active compounds in Tree Peony Bark, which has been found to have anti-inflammatory, anti-thrombotic and antioxidant properties (Fu et al., 2012a; Bao et al., 2013). Paeonol could increase the expression of miR-223 in macrophage-derived exosomes, and after the uptake of exosomes by endothelial cells, the STAT3 signaling and the related inflammatory response in endothelial cells can be attenuated (Liu et al., 2018). Another study also found similarly protective results of paeonol on endothelial cells in hyperlipidemia-induced atherosclerosis, which is also attributed to cellular uptake of exosomal miR-223 (Shi et al., 2020). Additionally, paeonol also promotes miR-126 expression to inhibit monocyte adhesion to endothelial cells and block the activation of the PI3K/Akt/NF- κ B signaling pathway (Yuan et al., 2016). Moreover, miR-21 and its target PTEN also contribute to the protective effects of paeonol on ox-LDL-induced endothelial injury (Liu et al., 2014). MiR-338-3p was proved to be increased in atherosclerotic lesions, and paeonol treatment could downregulate the expression of miR-338-3p and upregulate the expression of Tet methylcytosine dioxygenase 2 (TET2), thereby relieving ox-LDL-induced endothelial cell damage (Yin et al., 2019a; Yu et al., 2020b). Paeonol can also weaken ox-LDL-induced endothelial autophagy through regulating miR-30a/

beclin-1 signaling (Li et al., 2018a). Overall, these studies indicate that paeonol has strong endothelial protective effects, which is associated with regulation of various miRNAs and their targets.

3.14 Puerarin

Pueraria lobata is the dried roots of legumes *P. lobata* (Willd.) Ohwi and *Pueraria thunbergiana* (Siebold & Zucc.) Benth. It is clinically used in the treatment of cardiovascular and cerebrovascular diseases (Wang et al., 2020a). Puerarin, an active monomer in *Pueraria lobata*, was reported to inhibit the proliferation and inflammation of VSMCs in atherosclerosis by reducing the expression of miR-29b-3p, thereby increasing the expression of insulin-like growth factor 1 (IGF1) (Li et al., 2023). Therefore, puerarin may have a beneficial effect in the treatment of atherosclerosis by regulating miRNA.

4 Conclusion and prospects

Atherosclerosis is a major cause of coronary heart disease, cerebral infarction, and some peripheral vascular diseases (Figure 1). With the improvement of living standards, the incidence and mortality of atherosclerosis-induced cardiovascular diseases have increased rapidly in recent years. During the development of atherosclerosis, abnormal expressions of ncRNAs affect the physiological functions of endothelial cells, macrophages, and VSMCs by regulating related signaling pathways or specific proteins. China has a long history of using herbal medicine to treat cardiovascular diseases and the anti-atherosclerotic effects of several herbal medicines are also demonstrated in animal experiments and human studies. The traditional Chinese medicine monomers have recently attracted more attention in the treatment of diseases because they have certain molecular structures, predicted pharmacological effects, less drug-drug interactions, and clear mechanisms of action. Many active monomers derived from traditional Chinese medicines have been evaluated *in vivo* and *in vitro* to ameliorate the development of atherosclerosis by targeting ncRNAs. This article reviews 16 active monomers in traditional Chinese medicine that can improve the development of atherosclerosis by targeting ncRNAs in endothelial cells, macrophages, and VSMCs (Table 1; Figures 2–4). Their structures are shown in Figure 5. Besides monomeric Chinese herbal extracts, Chinese herbal formulas and decoctions have also been proved to treat atherosclerosis by targeting ncRNAs. For example, Tongxinluo Capsule inhibits vascular inflammation and neointimal hyperplasia by inhibiting the expression of miR-155, thereby blocking the feedback loop between miR-155 and TNF- α (Zhang et al., 2014). Alismatis rhizoma decoction, a classic traditional Chinese Medicinal formula used for the treatment of cardiovascular and cerebrovascular diseases, can inhibit the expression of ERK1/2 and miR-17~92a to inhibit ox-LDL-stimulated VSMCs proliferation (Shen et al., 2020). Among the ncRNAs regulated by active monomers of traditional Chinese medicine, miRNAs are the most studied. However, whether traditional Chinese medicine can exert functions via

regulating lncRNAs, circRNAs or other ncRNAs is not well studied and requires more research.

ncRNAs are the most abundant transcripts in cells. In addition to searching ncRNAs from published papers, we can identify or screen new ncRNAs in the following ways: firstly, we can utilize publicly available genomic and transcriptomic databases, such as Ensembl, NCBI, and UCSC to identify regions of the genome that are transcribed but not coding for proteins, indicating potential ncRNAs; additionally, high-throughput sequencing like RNA-Seq can be used to identify novel transcripts, including potential ncRNAs; furthermore, we can also compare the genomic sequences across various species in order to find conserved non-coding regions, which may potentially serve as ncRNAs; besides, machine learning algorithms based on sequence features and structural properties of ncRNAs can also be used to predict potential novel ncRNAs. After discovering new ncRNAs, techniques like CRISPR/Cas9, RNA interference, qRT-PCR, Northern blotting, *in situ* hybridization and other functional assays can be used to identify the specific biological functions of the ncRNAs. It can be expected that future studies will find more and more ncRNAs that related to atherosclerosis and these ncRNAs can be used as drug targets for development of anti-atherosclerotic drugs.

Over the past decades, substantial effort has been made towards the clinical application of RNA-based therapeutics, such as small interfering RNAs and antisense oligonucleotides. However, since the hurdle of immunogenicity, specificity, and delivery, some studies demonstrated limited efficacy or toxicity of ncRNAs-based therapies. Therefore, traditional Chinese medicine may become alternative drugs by targeting ncRNAs to treat atherosclerosis. It is worth noting that most studies suggest that traditional Chinese medicine treats atherosclerosis by targeting a specific ncRNA. However, the mechanism of ceRNA suggests that ncRNAs may have complex interactions in cells. What's more, a ncRNA may also have multiple targets. Therefore, we should further explore the anti-atherosclerotic mechanisms and clinical safety of these traditional Chinese medicine in more detail. It is hoped that by studying the regulation of ncRNAs by traditional Chinese medicine, it will provide theoretical support for the future research and clinical application of traditional Chinese medicine for treatment of atherosclerosis.

While many traditional Chinese medicines have therapeutic effects on atherosclerosis, some research has also identified potential side effects of certain Chinese herbs that can exacerbate atherosclerosis. For example, a moderate dosage of marijuana proves highly efficient in alleviating chronic pain (Carter et al., 2015), but marijuana can also cause cardiovascular side effects, such as endothelial dysfunction and atherosclerosis (Feng et al., 2022). Proanthocyanidin A1 can promote the production of platelets to ameliorate chemotherapy-induced thrombocytopenia (Wang et al., 2022)

and TMEA, a polyphenol in *Sanguisorba officinalis* L., can facilitate megakaryocyte differentiation and platelet production (Li et al., 2021a). However, the increased platelets can raise the risk of blood clot formation in patients with atherosclerosis (Barrett et al., 2019). Therefore, when patients have concurrent atherosclerosis, the use of these Chinese herbal medicines should be avoided. Furthermore, studying the ncRNAs that may mediate these effects is of significant importance, but this field is still lacking in research and requires further investigation.

Author contributions

GL: Conceptualization, Data curation, Methodology, Writing–original draft. LT: Writing–review and editing. XZ: Data curation, Writing–review and editing. MW: Software, Writing–review and editing. ZZ: Data curation, Writing–review and editing. JZ: Validation, Writing–review and editing. HG: Data curation, Writing–review and editing. ML: Writing–review and editing. WQ: Funding acquisition, Supervision, Writing–review and editing, Conceptualization.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

- Androulidaki, A., Wachsmuth, L., Polykratis, A., and Pasparakis, M. (2018). Differential role of MyD88 and TRIF signaling in myeloid cells in the pathogenesis of autoimmune diabetes. *PLoS ONE* 13 (3), e0194048. doi:10.1371/journal.pone.0194048
- Aryal, B., and Suarez, Y. (2019). Non-coding RNA regulation of endothelial and macrophage functions during atherosclerosis. *Vasc. Pharmacol.* 114, 64–75. doi:10.1016/j.vph.2018.03.001

- Augoff, K., McCue, B., Plow, E. F., and Sossey-Alaoui, K. (2012). miR-31 and its host gene lncRNA LOC554202 are regulated by promoter hypermethylation in triple-negative breast cancer. *Mol. Cancer* 11, 5. doi:10.1186/1476-4598-11-5
- Bai, Y., Wang, X., Zhao, S., Ma, C., Cui, J., and Zheng, Y. (2015). Sulforaphane protects against cardiovascular disease via Nrf2 activation. *Oxid. Med. Cell. Longev.* 2015, 407580. doi:10.1155/2015/407580
- Baker, A. H. (2011). MicroRNA 21 "shapes" vascular smooth muscle behavior through regulating tropomyosin 1. *Arterioscler. Thromb. Vasc. Biol.* 31 (9), 1941–1942. doi:10.1161/ATVBAHA.111.231985
- Bao, M. H., Zhang, Y. W., and Zhou, H. H. (2013). Paeonol suppresses oxidized low-density lipoprotein induced endothelial cell apoptosis via activation of LOX-1/p38MAPK/NF- κ B pathway. *J. Ethnopharmacol.* 146 (2), 543–551. doi:10.1016/j.jep.2013.01.019
- Bao, Y., Sun, Y. W., Ji, J., Gan, L., Zhang, C. F., Wang, C. Z., et al. (2019). Genkwanin ameliorates adjuvant-induced arthritis in rats through inhibiting JAK/STAT and NF- κ B signaling pathways. *Phytomedicine* 63, 153036. doi:10.1016/j.phymed.2019.153036
- Barrett, T. J., Schlegel, M., Zhou, F., Gorenchtein, M., Bolstorff, J., Moore, K. J., et al. (2019). Platelet regulation of myeloid suppressor of cytokine signaling 3 accelerates atherosclerosis. *Sci. Transl. Med.* 11 (517), eaax0481. doi:10.1126/scitranslmed.aax0481
- Bazioti, V., La Rose, A. M., Maassen, S., Bianchi, F., de Boer, R., Halmos, B., et al. (2022). T cell cholesterol efflux suppresses apoptosis and senescence and increases atherosclerosis in middle aged mice. *Nat. Commun.* 13 (1), 3799. doi:10.1038/s41467-022-31135-4
- Bellosta, S., and Corsini, A. (2018). Statin drug interactions and related adverse reactions: an update. *Expert Opin. Drug Saf.* 17 (1), 25–37. doi:10.1080/14740338.2018.1394455
- Boteanu, R. M., Suica, V. I., Uyy, E., Ivan, L., Dima, S. O., Popescu, I., et al. (2017). Alarmins in chronic noncommunicable diseases: atherosclerosis, diabetes and cancer. *J. Proteomics* 153, 21–29. doi:10.1016/j.jprot.2016.11.006
- Bridges, M. C., Daulagala, A. C., and Kourtidis, A. (2021). LNCcation: lncRNA localization and function. *J. Cell Biol.* 220 (2), e202009045. doi:10.1083/jcb.202009045
- Carter, G. T., Javaher, S. P., Nguyen, M. H. V., Garret, S., and Carlini, B. H. (2015). Re-branding cannabis: the next generation of chronic pain medicine? *Pain Manag.* 5 (1), 13–21. doi:10.2217/pmt.14.49
- Chen, C., Cheng, G., Yang, X., Li, C., Shi, R., and Zhao, N. (2016). Tanshinol suppresses endothelial cells apoptosis in mice with atherosclerosis via lncRNA TUG1 up-regulating the expression of miR-26a. *Am. J. Transl. Res.* 8 (7), 2981–2991.
- Chen, F. L., Yang, Z. H., Liu, Y., Li, L. X., Liang, W. C., Wang, X. C., et al. (2008). Berberine inhibits the expression of TNF α , MCP-1, and IL-6 in AcLDL-stimulated macrophages through PPAR γ pathway. *Endocrine* 33 (3), 331–337. doi:10.1007/s12020-008-9089-3
- Chen, F. Y., Zhou, J., Guo, N., Ma, W. G., Huang, X., Wang, H., et al. (2015). Curcumin retunes cholesterol transport homeostasis and inflammation response in M1 macrophage to prevent atherosclerosis. *Biochem. Biophys. Res. Commun.* 467 (4), 872–878. doi:10.1016/j.bbrc.2015.10.051
- Chen, L., Wang, C., Sun, H., Wang, J., Liang, Y., Wang, Y., et al. (2021a). The bioinformatics toolbox for circRNA discovery and analysis. *Brief. Bioinform.* 22 (2), 1706–1728. doi:10.1093/bib/bbaa001
- Chen, L., Yang, Z. S., Zhou, Y. Z., Deng, Y., Jiang, P., and Tan, S. L. (2020). Dihydromyricetin inhibits cell proliferation, migration, invasion and promotes apoptosis via regulating miR-21 in human cholangiocarcinoma cells. *J. Cancer* 11 (19), 5689–5699. doi:10.7150/jca.45970
- Chen, L., Zhao, Z. W., Zeng, P. H., Zhou, Y. J., and Yin, W. J. (2022). Molecular mechanisms for ABCA1-mediated cholesterol efflux. *Cell Cycle* 21 (11), 1121–1139. doi:10.1080/15384101.2022.2042777
- Chen, L. L. (2016). The biogenesis and emerging roles of circular RNAs. *Nat. Rev. Mol. Cell Biol.* 17 (4), 205–211. doi:10.1038/nrm.2015.32
- Chen, P., Li, J., Barnes, J., Kokkonen, G. C., Lee, J. C., and Liu, Y. (2002). Restraint of proinflammatory cytokine biosynthesis by mitogen-activated protein kinase phosphatase-1 in lipopolysaccharide-stimulated macrophages. *J. Immunol.* 169 (11), 6408–6416. doi:10.4049/jimmunol.169.11.6408
- Chen, W., Li, X., Guo, S., Song, N., Wang, J., Jia, L., et al. (2019a). Tanshinone IIA harmonizes the crosstalk of autophagy and polarization in macrophages via miR-375/KLF4 pathway to attenuate atherosclerosis. *Int. Immunopharmacol.* 70, 486–497. doi:10.1016/j.intimp.2019.02.054
- Chen, Y., Wang, S., Yang, S., Li, R., Yang, Y., Chen, Y., et al. (2021b). Inhibitory role of ginsenoside Rb2 in endothelial senescence and inflammation mediated by microRNA-216a. *Mol. Med. Rep.* 23 (6), 415. doi:10.3892/mmr.2021.12054
- Chen, Z., Pan, X., Sheng, Z., Yan, G., Chen, L., and Ma, G. (2019b). Baicalin suppresses the proliferation and migration of ox-LDL-VSMCs in atherosclerosis through upregulating miR-126-5p. *Biol. Pharm. Bull.* 42 (9), 1517–1523. doi:10.1248/bpb.b19-00196
- Cheng, H. S., Sivachandran, N., Lau, A., Boudreau, E., Zhao, J. L., Baltimore, D., et al. (2013). MicroRNA-146 represses endothelial activation by inhibiting pro-inflammatory pathways. *EMBO Mol. Med.* 5 (7), 1017–1034. doi:10.1002/emmm.201202318
- Cheng, S., Zhou, F., Xu, Y., Liu, X., Zhang, Y., Gu, M., et al. (2019). Geniposide regulates the miR-101/MKP-1/p38 pathway and alleviates atherosclerosis inflammatory injury in ApoE(-/-) mice. *Immunobiology* 224 (2), 296–306. doi:10.1016/j.imbio.2018.12.005
- Chi, H., Barry, S. P., Roth, R. J., Wu, J. J., Jones, E. A., Bennett, A. M., et al. (2006). Dynamic regulation of pro- and anti-inflammatory cytokines by MAPK phosphatase 1 (MKP-1) in innate immune responses. *Proc. Natl. Acad. Sci. U. S. A.* 103 (7), 2274–2279. doi:10.1073/pnas.0510965103
- Cyr, A. R., Huckaby, L. V., Shiva, S. S., and Zuckerbraun, B. S. (2020). Nitric Oxide and endothelial dysfunction. *Crit. Care Clin.* 36 (2), 307–321. doi:10.1016/j.ccc.2019.12.009
- Deng, Z., and Li, L. (2022). Effect of miR-663 on atherosclerosis by regulating the proliferation of vascular smooth muscle cells in lipid plaques. *Vascular* 23, 170853812210988. doi:10.1177/17085381221098826
- Di Mauro, V., Barandalla-Sobrados, M., and Catalucci, D. (2018). The noncoding-RNA landscape in cardiovascular health and disease. *NCRNA* 3 (1), 12–19. doi:10.1016/j.ncrna.2018.02.001
- Eken, S. M., Jin, H., Chernogubova, E., Li, Y., Simon, N., Sun, C., et al. (2017). MicroRNA-210 enhances fibrous cap stability in advanced atherosclerotic lesions. *Circ. Res.* 120 (4), 633–644. doi:10.1161/CIRCRESAHA.116.309318
- Eren, E., Tufekci, K. U., Isci, K. B., Tastan, B., Genc, K., and Genc, S. (2018). Sulforaphane inhibits lipopolysaccharide-induced inflammation, cytotoxicity, oxidative stress, and miR-155 expression and switches to mox phenotype through activating extracellular signal-regulated kinase 1/2-nuclear factor erythroid 2-related factor 2/antioxidant response element pathway in murine microglial cells. *Front. Immunol.* 9, 36. doi:10.3389/fimmu.2018.00036
- Feinberg, M. W., and Moore, K. J. (2016). MicroRNA regulation of atherosclerosis. *Circ. Res.* 118 (4), 703–720. doi:10.1161/CIRCRESAHA.115.306300
- Feng, X., Xu, S., and Weng, J. (2022). Marijuana and endothelial dysfunction: new mechanism and therapy. *Trends Mol. Med.* 28 (8), 613–615. doi:10.1016/j.molmed.2022.05.009
- Fiedler, J., and Thum, T. (2016). New insights into miR-17-92 cluster regulation and angiogenesis. *Circ. Res.* 118 (1), 9–11. doi:10.1161/CIRCRESAHA.115.307935
- Fu, P. K., Wu, C. L., Tsai, T. H., and Hsieh, C. L. (2012a). Anti-inflammatory and anticoagulative effects of paeonol on LPS-induced acute lung injury in rats. *Evid. Based Complement. Altern. Med.* 2012, 837513. doi:10.1155/2012/837513
- Fu, Y., Liu, B., Liu, J., Liu, Z., Liang, D., Li, F., et al. (2012b). Geniposide, from *Gardenia jasminoides* Ellis, inhibits the inflammatory response in the primary mouse macrophages and mouse models. *Int. Immunopharmacol.* 14 (4), 792–798. doi:10.1016/j.intimp.2012.07.006
- Gao, B., Huang, Q., Jie, Q., Zhang, H. Y., Wang, L., Guo, Y. S., et al. (2015). Ginsenoside-Rb2 inhibits dexamethasone-induced apoptosis through promotion of GPR120 induction in bone marrow-derived mesenchymal stem cells. *Stem Cells Dev.* 24 (6), 781–790. doi:10.1089/scd.2014.0367
- Gao, F., Wang, X. C., Luo, Z. D., Hu, G. Q., Ma, M. Q., Liang, Y., et al. (2022). LncRNA HOXA11-AS promotes vascular endothelial cell injury in atherosclerosis by regulating the miR-515-5p/ROCK1 axis. *Esc. heart Fail* 9 (4), 2259–2271. doi:10.1002/ehf2.13815
- Gao, J. H., Zeng, M. Y., Yu, X. H., Zeng, G. F., He, L. H., Zheng, X. L., et al. (2018). Visceral adipose tissue-derived serine protease inhibitor accelerates cholesterol efflux by up-regulating ABCA1 expression via the NF- κ B/miR-33a pathway in THP-1 macrophage-derived foam cells. *Biochem. Biophys. Res. Commun.* 500 (2), 318–324. doi:10.1016/j.bbrc.2018.04.066
- Gao, Y., Liu, F., Fang, L., Cai, R., Zong, C., and Qi, Y. (2014). Genkwanin inhibits proinflammatory mediators mainly through the regulation of miR-101/MKP-1/MAPK pathway in LPS-activated macrophages. *PLoS ONE* 9 (5), e96741. doi:10.1371/journal.pone.0096741
- Gong, L., Chang, H., Zhang, J., Guo, G., Shi, J., and Xu, H. (2018). Astragaloside IV protects rat cardiomyocytes from hypoxia-induced injury by downregulation of miR-23a and miR-92a. *Cell. Physiol. Biochem.* 49 (6), 2240–2253. doi:10.1159/000493827
- Guan, S., Shen, Y., Ge, H., Xiong, W., He, L., Liu, L., et al. (2019). Dihydromyricetin alleviates diabetic neuropathic pain and depression comorbidity symptoms by inhibiting P2X(7) receptor. *Front. Psychiatry* 10, 770. doi:10.3389/fpsy.2019.00770
- Guerrero-Beltran, C. E., Calderon-Oliver, M., Pedraza-Chaverri, J., and Chirino, Y. I. (2012). Protective effect of sulforaphane against oxidative stress: recent advances. *Exp. Toxicol. Pathol.* 64 (5), 503–508. doi:10.1016/j.etp.2010.11.005
- Gunning, P. W., Hardeman, E. C., Lappalainen, P., and Mulvihill, D. P. (2015). Tropomyosin - master regulator of actin filament function in the cytoskeleton. *J. Cell Sci.* 128 (16), 2965–2974. doi:10.1242/jcs.172502
- Guo, F. X., Wu, Q., Li, P., Zheng, L., Ye, S., Dai, X. Y., et al. (2019). The role of the lncRNA-FA2H-2-MLKL pathway in atherosclerosis by regulation of autophagy flux and inflammation through mTOR-dependent signaling. *Cell Death Differ.* 26 (9), 1670–1687. doi:10.1038/s41418-018-0235-z
- Guo, Y., Li, P., Gao, L., Zhang, J., Yang, Z., Bledsoe, G., et al. (2017). Kallistatin reduces vascular senescence and aging by regulating microRNA-34a-SIRT1 pathway. *Aging Cell* 16 (4), 837–846. doi:10.1111/acel.12615

- Guttman, M., and Rinn, J. L. (2012). Modular regulatory principles of large non-coding RNAs. *Nature* 482 (7385), 339–346. doi:10.1038/nature10887
- Hasan, S. T., Zingg, J. M., Kwan, P., Noble, T., Smith, D., and Meydani, M. (2014). Curcumin modulation of high fat diet-induced atherosclerosis and steatohepatitis in LDL receptor deficient mice. *Atherosclerosis* 232 (1), 40–51. doi:10.1016/j.atherosclerosis.2013.10.016
- Houghton, C. A., Fasset, R. G., and Coombes, J. S. (2016). Sulforaphane and other nutrigenomic Nrf2 activators: can the clinician's expectation be matched by the reality? *Oxid. Med. Cell. Longev.* 2016, 7857186. doi:10.1155/2016/7857186
- Huang, H. S., Huang, X. Y., Yu, H. Z., Xue, Y., and Zhu, P. L. (2020). Circular RNA circ-RELL1 regulates inflammatory response by miR-6873-3p/MyD88/NF- κ B axis in endothelial cells. *Biochem. Biophys. Res. Commun.* 525 (2), 512–519. doi:10.1016/j.bbrc.2020.02.109
- Huang, Q., Gao, B., Jie, Q., Wei, B. Y., Fan, J., Zhang, H. Y., et al. (2014). Ginsenoside-Rb2 displays anti-osteoporosis effects through reducing oxidative damage and bone-resorbing cytokines during osteogenesis. *Bone* 66, 306–314. doi:10.1016/j.bone.2014.06.010
- Huang, Q., Wang, T., and Wang, H. Y. (2017). Ginsenoside Rb2 enhances the anti-inflammatory effect of omega-3 fatty acid in LPS-stimulated RAW264.7 macrophages by upregulating GPR120 expression. *Acta Pharmacol. Sin.* 38 (2), 192–200. doi:10.1038/aps.2016.135
- Jia, S., Ma, W. D., Zhang, C. Y., Zhang, Y., Yao, Z. H., Quan, X. H., et al. (2019). Tanshinone IIA attenuates high glucose induced human VSMC proliferation and migration through miR-21-5p-mediated tropomyosin 1 downregulation. *Arch. Biochem. Biophys.* 677, 108154. doi:10.1016/j.abb.2019.108154
- Jiang, J., Mo, Z. C., Yin, K., Zhao, G. J., Lv, Y. C., Ouyang, X. P., et al. (2012). Epigallocatechin-3-gallate prevents TNF- α -induced NF- κ B activation thereby upregulating ABCA1 via the Nrf2/Keap1 pathway in macrophage foam cells. *Int. J. Mol. Med.* 29 (5), 946–956. doi:10.3892/ijmm.2012.924
- Jiang, Y., Wang, H. Y., Li, Y., Guo, S. H., Zhang, L., Cai, J. H., et al. (2014). Peripheral blood miRNAs as a biomarker for chronic cardiovascular diseases. *Sci. Rep.* 4, 5026. doi:10.1038/srep05026
- Lai, X. H., Lei, Y., Yang, J., and Xiu, C. K. (2018). Effect of microRNA-34a/SIRT1/p53 signal pathway on notoginsenoside R1 delaying vascular endothelial cell senescence. *China J. Chin. Mater. Med.* 43 (3), 577–584. doi:10.19540/j.cnki.cjmm.20180110.001
- Lei, W., Yan, Y., Ma, Y., Jiang, M., Zhang, B., Zhang, H., et al. (2022). Notoginsenoside R1 regulates ischemic myocardial lipid metabolism by activating the AKT/mTOR signaling pathway. *Front. Pharmacol.* 13, 905092. doi:10.3389/fphar.2022.905092
- Li, C., Yang, L., Wu, H., and Dai, M. (2018a). Paeonol inhibits oxidized low-density lipoprotein-induced vascular endothelial cells autophagy by upregulating the expression of miRNA-30a. *Front. Pharmacol.* 9, 95. doi:10.3389/fphar.2018.00095
- Li, C., Zhou, L., Lin, G., and Zuo, Z. (2009). Contents of major bioactive flavones in proprietary traditional Chinese medicine products and reference herb of radix scutellariae. *J. Pharm. Biomed. Anal.* 50 (3), 298–306. doi:10.1016/j.jpba.2009.04.028
- Li, F. P., Lin, D. Q., and Gao, L. Y. (2018b). LncRNA TUG1 promotes proliferation of vascular smooth muscle cell and atherosclerosis through regulating miRNA-21/PTEN axis. *Eur. Rev. Med. Pharmacol. Sci.* 22 (21), 7439–7447. doi:10.26355/eurrev_201811_16284
- Li, H., Jiang, X., Shen, X., Sun, Y., Jiang, N., Zeng, J., et al. (2021a). TMEA, a polyphenol in *Sanguisorba officinalis*, promotes thrombocytopoiesis by upregulating PI3K/akt signaling. *Front. Cell Dev. Biol.* 9, 708331. doi:10.3389/fcell.2021.708331
- Li, J., Li, Y., Yuan, X., Yao, D., Gao, Z., Niu, Z., et al. (2023). The effective constituent puerarin, from *Pueraria lobata*, inhibits the proliferation and inflammation of vascular smooth muscle in atherosclerosis through the miR-29b-3p/IGF1 pathway. *Pharm. Biol.* 61 (1), 1–11. doi:10.1080/13880209.2022.2099430
- Li, R., Hu, Y., and Hou, S. (2022). An exploration of oral-gut pathogens mediating immune escape of pancreatic cancer via miR-21/PTEN axis. *Front. Microbiol.* 13, 928846. doi:10.3389/fmicb.2022.928846
- Li, S., Stockl, S., Lukas, C., Herrmann, M., Brochhausen, C., König, M. A., et al. (2021b). Curcumin-primed human BMSC-derived extracellular vesicles reverse IL-1 β -induced catabolic responses of OA chondrocytes by upregulating miR-126-3p. *Stem Cell Ther.* 12 (1), 252. doi:10.1186/s13287-021-02317-6
- Li, T., Pang, Q., Liu, Y., Bai, M., Peng, Y., and Zhang, Z. (2021c). Sulforaphane protects human umbilical vein endothelial cells from oxidative stress via the miR-34a/SIRT1 axis by upregulating nuclear factor erythroid-2-related factor 2. *Exp. Ther. Med.* 21 (3), 186. doi:10.3892/etm.2021.9617
- Li, X. Q., and Huang, T. Y. (2021). Notoginsenoside R1 alleviates high glucose-induced inflammation and oxidative stress in HUVECs via upregulating miR-147a. *Kaohsiung J. Med. Sci.* 37 (12), 1101–1112. doi:10.1002/kjm2.12433
- Li, Y., Zhang, Y., Lu, J., Yin, Y., Xie, J., and Xu, B. (2021d). Anti-inflammatory mechanisms and research progress of colchicine in atherosclerotic therapy. *J. Cell. Mol. Med.* 25 (17), 8087–8094. doi:10.1111/jcmm.16798
- Li, Z., Xie, X., Fan, X., and Li, X. (2020). Long Non-coding RNA MINCR regulates miR-876-5p/GSPT1 axis to aggravate glioma progression. *Neurochem. Res.* 45 (7), 1690–1699. doi:10.1007/s11064-020-03029-8
- Liang, G., Wang, S., Shao, J., Jin, Y. J., Xu, L., Yan, Y., et al. (2022). Tenascin-X mediates flow-induced suppression of EndMT and atherosclerosis. *Circ. Res.* 130 (11), 1647–1659. doi:10.1161/CIRCRESAHA.121.320694
- Liao, X., Sharma, N., Kapadia, F., Zhou, G., Lu, Y., Hong, H., et al. (2011). Kruppel-like factor 4 regulates macrophage polarization. *J. Clin. Invest.* 121 (7), 2736–2749. doi:10.1172/JCI45444
- Libby, P. (2021). The changing landscape of atherosclerosis. *Nature* 592 (7855), 524–533. doi:10.1038/s41586-021-03392-8
- Libby, P., Buring, J. E., Badimon, L., Hansson, G. K., Deanfield, J., Bittencourt, M. S., et al. (2019). Atherosclerosis. *Nat. Rev. Dis. Prim.* 5 (1), 56. doi:10.1038/s41572-019-0106-z
- Liu, Y., Li, C., Wu, H., Xie, X., Sun, Y., and Dai, M. (2018). Paeonol attenuated inflammatory response of endothelial cells via stimulating monocytes-derived exosomal MicroRNA-223. *Front. Pharmacol.* 9, 1105. doi:10.3389/fphar.2018.01105
- Liu, Y. R., Chen, J. J., and Dai, M. (2014). Paeonol protects rat vascular endothelial cells from ox-LDL-induced injury *in vitro* via downregulating microRNA-21 expression and TNF- α release. *Acta Pharmacol. Sin.* 35 (4), 483–488. doi:10.1038/aps.2013.190
- Lu, T. X., and Rothenberg, M. E. (2018). MicroRNA. *J. Allergy Clin. Immunol.* 141 (4), 1202–1207. doi:10.1016/j.jaci.2017.08.034
- Luo, H., Chen, J., Su, C., and Zha, L. (2022). Advances in the bioactivities of phytochemical saponins in the prevention and treatment of atherosclerosis. *Nutrients* 14 (23), 4998. doi:10.3390/nu14234998
- Maitrias, P., Metzinger-Le Meuth, V., Nader, J., Reix, T., Caus, T., and Metzinger, L. (2017). The involvement of miRNA in carotid-related stroke. *Arterioscler. Thromb. Vasc. Biol.* 37 (9), 1608–1617. doi:10.1161/ATVBAHA.117.309233
- Meiler, S., Baumer, Y., Toulmin, E., Seng, K., and Boisvert, W. A. (2015). MicroRNA 302a is a novel modulator of cholesterol homeostasis and atherosclerosis. *Arterioscler. Thromb. Vasc. Biol.* 35 (2), 323–331. doi:10.1161/ATVBAHA.114.304878
- Menghini, R., Casagrande, V., Cardellini, M., Martelli, E., Terrinoni, A., Amati, F., et al. (2009). MicroRNA 217 modulates endothelial cell senescence via silent information regulator 1. *Circulation* 120 (15), 1524–1532. doi:10.1161/CIRCULATIONAHA.109.864629
- Okuyama, H., Langsjoen, P. H., Hamazaki, T., Ogushi, Y., Hama, R., Kobayashi, T., et al. (2015). Statins stimulate atherosclerosis and heart failure: pharmacological mechanisms. *Expert Rev. Clin. Pharmacol.* 8 (2), 189–199. doi:10.1586/17512433.2015.1011125
- Ouyang, S., Zhang, O., Xiang, H., Yao, Y. H., and Fang, Z. Y. (2022). Curcumin improves atherosclerosis by inhibiting the epigenetic repression of lncRNA MIAT to miR-124. *Vascular* 30, 1213–1223. doi:10.1177/17085381211040974
- Pankratz, F., Hohnloser, C., Bemtgen, X., Jaenich, C., Kreuzaler, S., Hoefer, I., et al. (2018). MicroRNA-100 suppresses chronic vascular inflammation by stimulation of endothelial autophagy. *Circ. Res.* 122 (3), 417–432. doi:10.1161/CIRCRESAHA.117.311428
- Park, K. H., Park, Y. D., Han, J. M., Im, K. R., Lee, B. W., Jeong, I. Y., et al. (2006). Anti-atherosclerotic and anti-inflammatory activities of catecholic xanthenes and flavonoids isolated from *Cudrania tricuspidata*. *Bioorg. Med. Chem. Lett.* 16 (21), 5580–5583. doi:10.1016/j.bmcl.2006.08.032
- Perez-Sanchez, C., Aguirre, M. A., Ruiz-Limon, P., Abalos-Aguilera, M. C., Jimenez-Gomez, Y., Arias-de la Rosa, I., et al. (2017). Ubiquinol effects on antiphospholipid syndrome prothrombotic profile: a randomized, placebo-controlled trial. *Arterioscler. Thromb. Vasc. Biol.* 37 (10), 1923–1932. doi:10.1161/ATVBAHA.117.309225
- Pope, A. J., Karpuriah, K., Kearns, P. N., Xia, Y., and Cardounel, A. J. (2009). Role of dimethylarginine dimethylaminohydrolases in the regulation of endothelial nitric oxide production. *J. Biol. Chem.* 284 (51), 35338–35347. doi:10.1074/jbc.M109.037036
- Qian, W., Cai, X., Qian, Q., Zhuang, Q., Yang, W., Zhang, X., et al. (2019a). Astragaloside IV protects endothelial progenitor cells from the damage of ox-LDL via the LOX-1/NLRP3 inflammasome pathway. *Drug Des. devel. Ther.* 13, 2579–2589. doi:10.2147/DDDT.S207774
- Qin, H. W., Li, Y. J., Zhang, Z. X., Ren, K., Li, S. J., and Lu, Y. B. (2018). Anti-atherosclerosis effect of astragaloside is related to miR-33a/ABCA1 signaling. *Chin. J. Pathophysiol.* 34 (12), 2120–2126. doi:10.3969/j.issn.1000-4718.2018.12.002
- Qin, H. W., Zhang, Q. S., Li, Y. J., Li, W. T., and Wang, Y. (2022). Molecular mechanism of astragaloside IV against atherosclerosis by regulating miR-17-5p and PCSK9/VLDLR signal pathway. *China J. Chin. Mater. Med.* 47 (2), 492–498. doi:10.19540/j.cnki.cjmm.20210918.701
- Qin, Y., Zheng, B., Yang, G. S., Zhou, J., Yang, H. J., Nie, Z. Y., et al. (2020). Tanshinone IIA inhibits VSMC inflammation and proliferation *in vivo* and *in vitro* by downregulating miR-712-5p expression. *Eur. J. Pharmacol.* 880, 173140. doi:10.1016/j.ejphar.2020.173140
- Ren, P., Wang, J., Li, L., Lin, X., Wu, G., Chen, J., et al. (2021). Identification of key genes involved in the recurrence of glioblastoma multiforme using weighted gene co-expression network analysis and differential expression analysis. *Bioengineered* 12 (1), 3188–3200. doi:10.1080/21655979.2021.1943986
- Ruparelia, N., and Choudhury, R. (2020). Inflammation and atherosclerosis: what is on the horizon? *Heart* 106 (1), 80–85. doi:10.1136/heartjnl-2018-314230

- Salmena, L., Poliseno, L., Tay, Y., Kats, L., and Pandolfi, P. P. (2011). A ceRNA hypothesis: the rosetta stone of a hidden RNA language? *Cell* 146 (3), 353–358. doi:10.1016/j.cell.2011.07.014
- Shao, X., Liu, Z., Liu, S., Lin, N., and Deng, Y. (2021). Astragaloside IV alleviates atherosclerosis through targeting circ_0000231/miR-135a-5p/CLIC4 axis in AS cell model *in vitro*. *Mol. Cell. Biochem.* 476 (4), 1783–1795. doi:10.1007/s11010-020-04035-8
- Sheedy, F. J. (2015). Turning 21: induction of miR-21 as a key switch in the inflammatory response. *Front. Immunol.* 6, 19. doi:10.3389/fimmu.2015.00019
- Shen, J., Wei, W., Wang, X., Yang, J., Lu, L., Lv, X., et al. (2020). Proliferation of vascular smooth muscle cells under ox-LDL is regulated by alismatis rhizoma decoction via Inhibiting ERK1/2 and miR-17-92a cluster activation. *Evid. Based Complement. Alternat. Med.* 2020, 7275246. doi:10.1155/2020/7275246
- Shen, L., Hu, Y., Lou, J., Yin, S., Wang, W., Wang, Y., et al. (2019). CircRNA-0044073 is upregulated in atherosclerosis and increases the proliferation and invasion of cells by targeting miR-107. *Mol. Med. Rep.* 19 (5), 3923–3932. doi:10.3892/mmr.2019.10011
- Shi, X., Xie, X., Sun, Y., He, H., Huang, H., Liu, Y., et al. (2020). Paeonol inhibits NLRP3 mediated inflammation in rat endothelial cells by elevating hyperlipidemic rats plasma exosomal miRNA-223. *Eur. J. Pharmacol.* 885, 173473. doi:10.1016/j.ejphar.2020.173473
- Shou, X., Wang, Y., Jiang, Q., Chen, J., and Liu, Q. (2023). miR-126 promotes M1 to M2 macrophage phenotype switching via VEGFA and KLF4. *PeerJ* 11, e15180. doi:10.7717/peerj.15180
- Silva, J., Yu, X., Moradian, R., Folk, C., Spatz, M. H., Kim, P., et al. (2020). Dihydromyricetin protects the liver via changes in lipid metabolism and enhanced ethanol metabolism. *Alcohol. Clin. Exp. Res.* 44 (5), 1046–1060. doi:10.1111/acer.14326
- Simion, V., Zhou, H., Haemmig, S., Pierce, J. B., Mendes, S., Tesmenitsky, Y., et al. (2020). A macrophage-specific lncRNA regulates apoptosis and atherosclerosis by tethering HuR in the nucleus. *Nat. Commun.* 11 (1), 6135. doi:10.1038/s41467-020-19664-2
- Son, D. J., Kumar, S., Takabe, W., Kim, C. W., Ni, C. W., Alberts-Grill, N., et al. (2013). The atypical mechanosensitive microRNA-712 derived from pre-ribosomal RNA induces endothelial inflammation and atherosclerosis. *Nat. Commun.* 4, 3000. doi:10.1038/ncomms4000
- Song, W., Pu, J., and He, B. (2014). Tanhinol protects human umbilical vein endothelial cells against hydrogen peroxide-induced apoptosis. *Mol. Med. Rep.* 10 (5), 2764–2770. doi:10.3892/mmr.2014.2541
- Song, Z., Wei, D., Chen, Y., Chen, L., Bian, Y., Shen, Y., et al. (2019). Association of astragaloside IV-inhibited autophagy and mineralization in vascular smooth muscle cells with lncRNA H19 and DUSP5-mediated ERK signaling. *Toxicol. Appl. Pharmacol.* 364, 45–54. doi:10.1016/j.taap.2018.12.002
- Sun, D., Xiang, G., Wang, J., Li, Y., Mei, S., Ding, H., et al. (2020). miRNA 146b-5p protects against atherosclerosis by inhibiting vascular smooth muscle cell proliferation and migration. *Epigenomics* 12 (24), 2189–2204. doi:10.2217/epi-2020-0155
- Sun, G., Li, Y., and Ji, Z. (2019). Up-regulation of MIAT aggravates the atherosclerotic damage in atherosclerosis mice through the activation of PI3K/Akt signaling pathway. *Drug Deliv.* 26 (1), 641–649. doi:10.1080/10717544.2019.1628116
- Sun, Y., Liu, S., Yang, S., Chen, C., Yang, Y., Lin, M., et al. (2021). Mechanism of dihydromyricetin on inflammatory diseases. *Front. Pharmacol.* 12, 794563. doi:10.3389/fphar.2021.794563
- Tan, C., Zhou, L., Wen, W., and Xiao, N. (2021). Curcumin promotes cholesterol efflux by regulating ABCA1 expression through miR-125a-5p/SIRT6 axis in THP-1 macrophage to prevent atherosclerosis. *J. Toxicol. Sci.* 46 (5), 209–222. doi:10.2131/jts.46.209
- Tan, L., Meng, L., Shi, X., and Yu, B. (2017). Knockdown of microRNA-17-5p ameliorates atherosclerotic lesions in ApoE(-/-) mice and restores the expression of very low density lipoprotein receptor. *Biotechnol. Lett.* 39 (7), 967–976. doi:10.1007/s10529-017-2337-y
- Tan, Y. Q., Chen, H. W., and Li, J. (2020). Astragaloside IV: an effective drug for the treatment of cardiovascular diseases. *Drug Des. devel. Ther.* 14, 3731–3746. doi:10.2147/DDDT.S272355
- Tang, Y. L., Jiang, J. H., Wang, S., Liu, Z., Tang, X. Q., Peng, J., et al. (2015). TLR4/NF- κ B signaling contributes to chronic unpredictable mild stress-induced atherosclerosis in ApoE^{-/-} mice. *PLoS ONE* 10 (4), e0123685. doi:10.1371/journal.pone.0123685
- Tao, H., Cao, W., Yang, J. J., Shi, K. H., Zhou, X., Liu, L. P., et al. (2016). Long noncoding RNA H19 controls DUSP5/ERK1/2 axis in cardiac fibroblast proliferation and fibrosis. *Cardiovasc. Pathol.* 25 (5), 381–389. doi:10.1016/j.carpath.2016.05.005
- Thum, T., and Mayr, M. (2012). Review focus on the role of microRNA in cardiovascular biology and disease. *Cardiovasc. Res.* 93 (4), 543–544. doi:10.1093/cvr/cvs085
- Tousoulis, D., Kampoli, A. M., Tentolouris, C., Papageorgiou, N., and Stefanadis, C. (2012). The role of nitric oxide on endothelial function. *Curr. Vasc. Pharmacol.* 10 (1), 4–18. doi:10.2174/157016112798829760
- Vanduchova, A., Anzenbacher, P., and Anzenbacherova, E. (2019). Isothiocyanate from broccoli, sulforaphane, and its properties. *J. Med. Food* 22 (2), 121–126. doi:10.1089/jmf.2018.0024
- Wang, D., Lu, C., Sun, F., Cui, M., Mu, H., Duan, J., et al. (2017). A tanshinone I derivative enhances the activities of antibiotics against *Staphylococcus aureus* *in vitro* and *in vivo*. *Res. Microbiol.* 168 (1), 46–54. doi:10.1016/j.resmic.2016.08.002
- Wang, J. C., and Bennett, M. (2012). Aging and atherosclerosis: mechanisms, functional consequences, and potential therapeutics for cellular senescence. *Circ. Res.* 111 (2), 245–259. doi:10.1161/CIRCRESAHA.111.261388
- Wang, L., Shen, C., Wang, Y., Zou, T., Zhu, H., Lu, X., et al. (2019). Identification of circular RNA Hsa_circ_0001879 and Hsa_circ_0004104 as novel biomarkers for coronary artery disease. *Atherosclerosis* 286, 88–96. doi:10.1016/j.atherosclerosis.2019.05.006
- Wang, R., Hu, X., Wang, J., Zhou, L., Hong, Y., Zhang, Y., et al. (2022). Proanthocyanidin A1 promotes the production of platelets to ameliorate chemotherapy-induced thrombocytopenia through activating JAK2/STAT3 pathway. *Phytomedicine* 95, 153880. doi:10.1016/j.phymed.2021.153880
- Wang, S., Zhang, S., Wang, S., Gao, P., and Dai, L. (2020a). A comprehensive review on pueraria: insights on its chemistry and medicinal value. *Biomed. Pharmacother.* 131, 110734. doi:10.1016/j.biopha.2020.110734
- Wang, W., Ma, F., and Zhang, H. (2020b). MicroRNA-374 is a potential diagnostic biomarker for atherosclerosis and regulates the proliferation and migration of vascular smooth muscle cells. *Cardiovasc. Diagn. Ther.* 10 (4), 687–694. doi:10.21037/cdt-20-444
- Wang, Z., Shi, D., Zhang, N., Yuan, T., and Tao, H. (2021). MiR-217 promotes endothelial cell senescence through the SIRT1/p53 signaling pathway. *J. Mol. Histol.* 52 (2), 257–267. doi:10.1007/s10735-020-09945-x
- Wang, Z., Zhang, M., Wang, Z., Guo, Z., Wang, Z., and Chen, Q. (2020c). Cyanidin-3-O-glucoside attenuates endothelial cell dysfunction by modulating miR-204-5p/SIRT1-mediated inflammation and apoptosis. *Biofactors* 46 (5), 803–812. doi:10.1002/biof.1660
- Wijesekara, N., Zhang, L. H., Kang, M. H., Abraham, T., Bhattacharjee, A., Warnock, G. L., et al. (2012). miR-33a modulates ABCA1 expression, cholesterol accumulation, and insulin secretion in pancreatic islets. *Diabetes* 61 (3), 653–658. doi:10.2337/db11-0944
- Winter, J., Jung, S., Keller, S., Gregory, R. I., and Diederichs, S. (2009). Many roads to maturity: microRNA biogenesis pathways and their regulation. *Nat. Cell Biol.* 11 (3), 228–234. doi:10.1038/ncb0309-228
- Wu, Y., Wang, F., Fan, L., Zhang, W., Wang, T., Du, Y., et al. (2018). Baicalin alleviates atherosclerosis by relieving oxidative stress and inflammatory responses via inactivating the NF- κ B and p38 MAPK signaling pathways. *Biomed. Pharmacother.* 97, 1673–1679. doi:10.1016/j.biopha.2017.12.024
- Xiao, X., Xu, M., Yu, H., Wang, L., Li, X., Rak, J., et al. (2021). Mesenchymal stem cell-derived small extracellular vesicles mitigate oxidative stress-induced senescence in endothelial cells via regulation of miR-146a/Src. *Signal Transduct. Target Ther.* 6 (1), 354. doi:10.1038/s41392-021-00765-3
- Xue, X., Deng, Y., Wang, J., Zhou, M., Liao, L., Wang, C., et al. (2021). Hydroxysafflor yellow A, a natural compound from *Carthamus tinctorius* L with good effect of alleviating atherosclerosis. *Phytomedicine* 91, 153694. doi:10.1016/j.phymed.2021.153694
- Yamakuchi, M., Ferlito, M., and Lowenstein, C. J. (2008). miR-34a repression of SIRT1 regulates apoptosis. *Proc. Natl. Acad. Sci. U. S. A.* 105 (36), 13421–13426. doi:10.1073/pnas.0801613105
- Yang, D., Tan, S., Yang, Z., Jiang, P., Qin, C., Yuan, Q., et al. (2018). Dihydromyricetin attenuates TNF- α -induced endothelial dysfunction through miR-21-mediated DDAH1/ADMA/NO signal pathway. *Biomed. Res. Int.* 2018, 1047810. doi:10.1155/2018/1047810
- Yang, D., Yang, Z., Chen, L., Kuang, D., Zou, Y., Li, J., et al. (2020). Dihydromyricetin increases endothelial nitric oxide production and inhibits atherosclerosis through microRNA-21 in apolipoprotein E-deficient mice. *J. Cell. Mol. Med.* 24 (10), 5911–5925. doi:10.1111/jcmm.15278
- Yang, H. X., Gao, Y., Jiang, H. B., and Liu, L. S. (2016). EGCG upregulated ABCA1 expression by decreasing miR-33a generation to reduce lipid accumulation of macrophage-derived foam cells. *Chin. Pharmacol. Bull.* 32, 1279–1284. doi:10.3969/j.issn.1001-1978.2016.09.018
- Yin, J., Hou, X., and Yang, S. (2019a). microRNA-338-3p promotes ox-LDL-induced endothelial cell injury through targeting BAMBI and activating TGF- β /Smad pathway. *J. Cell. Physiol.* 234 (7), 11577–11586. doi:10.1002/jcp.27814
- Yin, R., Zhu, X., Wang, J., Yang, S., Ma, A., Xiao, Q., et al. (2019b). MicroRNA-155 promotes the ox-LDL-induced activation of NLRP3 inflammasomes via the ERK1/2 pathway in THP-1 macrophages and aggravates atherosclerosis in ApoE^{-/-} mice. *Ann. Palliat. Med.* 8 (5), 676–689. doi:10.21037/apm.2019.10.11
- Yu, X. H., Deng, W. Y., Chen, J. J., Xu, X. D., Liu, X. X., Chen, L., et al. (2020a). LncRNA kcnq1ot1 promotes lipid accumulation and accelerates atherosclerosis via functioning as a ceRNA through the miR-452-3p/HDAC3/ABCA1 axis. *Cell Death Dis.* 11 (12), 1043. doi:10.1038/s41419-020-03263-6

- Yu, Y., Yan, R., Chen, X., Sun, T., and Yan, J. (2020b). Paeonol suppresses the effect of ox-LDL on mice vascular endothelial cells by regulating miR-338-3p/TET2 axis in atherosclerosis. *Mol. Cell. Biochem.* 475 (1-2), 127–135. doi:10.1007/s11010-020-03865-w
- Yuan, L., Li, Q., Zhang, Z., Liu, Q., Wang, X., and Fan, L. (2020). Tanshinone IIA inhibits the adipogenesis and inflammatory response in ox-LDL-challenged human monocyte-derived macrophages via regulating miR-130b/WNT5A. *J. Cell. Biochem.* 121 (2), 1400–1408. doi:10.1002/jcb.29375
- Yuan, X., Chen, J., and Dai, M. (2016). Paeonol promotes microRNA-126 expression to inhibit monocyte adhesion to ox-LDL-injured vascular endothelial cells and block the activation of the PI3K/Akt/NF- κ B pathway. *Int. J. Mol. Med.* 38 (6), 1871–1878. doi:10.3892/ijmm.2016.2778
- Zang, H. L., Li, Y. H., and Huang, G. M. (2020). Long-chain non-coding RNA Linc00888 promotes the proliferation and migration of esophageal cancer cells by downregulating miR-34a expression. *Eur. Rev. Med. Pharmacol. Sci.* 24 (21), 11081–11089. doi:10.26355/eurrev_202011_23594
- Zeng, P., Yang, J., Liu, L., Yang, X., Yao, Z., Ma, C., et al. (2021). ERK1/2 inhibition reduces vascular calcification by activating miR-126-3p-DKK1/LRP6 pathway. *Theranostics* 11 (3), 1129–1146. doi:10.7150/thno.49771
- Zhang, H., Xu, Z., Zhao, H., Wang, X., Pang, J., Li, Q., et al. (2020a). Anthocyanin supplementation improves anti-oxidative and anti-inflammatory capacity in a dose-response manner in subjects with dyslipidemia. *Redox Biol.* 32, 101474. doi:10.1016/j.redox.2020.101474
- Zhang, M., Zhu, Y., Zhu, J., Xie, Y., Wu, R., Zhong, J., et al. (2022). circ_0086296 induced atherosclerotic lesions via the IFIT1/STAT1 feedback loop by sponging miR-576-3p. *Cell. Mol. Biol. Lett.* 27 (1), 80. doi:10.1186/s11658-022-00372-2
- Zhang, R. N., Zheng, B., Li, L. M., Zhang, J., Zhang, X. H., and Wen, J. K. (2014). Tongxinluo inhibits vascular inflammation and neointimal hyperplasia through blockade of the positive feedback loop between miR-155 and TNF- α . *Am. J. Physiol. Heart Circ. Physiol.* 307 (4), H552–H562. doi:10.1152/ajpheart.00936.2013
- Zhang, X., Wang, Z., Li, W., Huang, R., Zheng, D., and Bi, G. (2020b). MicroRNA-217-5p ameliorates endothelial cell apoptosis induced by ox-LDL by targeting CLIC4. *Nutr. Metab. Cardiovasc. Dis.* 30 (3), 523–533. doi:10.1016/j.numecd.2019.09.027
- Zhang, Y., Qin, W., Zhang, L., Wu, X., Du, N., Hu, Y., et al. (2015). MicroRNA-26a prevents endothelial cell apoptosis by directly targeting TRPC6 in the setting of atherosclerosis. *Sci. Rep.* 5, 9401. doi:10.1038/srep09401
- Zhao, X. S., Zheng, B., Wen, Y., Sun, Y., Wen, J. K., and Zhang, X. H. (2019). Salvianolic acid B inhibits Ang II-induced VSMC proliferation *in vitro* and intimal hyperplasia *in vivo* by downregulating miR-146a expression. *Phytomedicine* 58, 152754. doi:10.1016/j.phymed.2018.11.014
- Zhou, S., Sun, Y., Zhao, K., Gao, Y., Cui, J., Qi, L., et al. (2020). miR21/PTEN pathway mediates the cardioprotection of geniposide against oxidized lowdensity lipoprotein-induced endothelial injury via suppressing oxidative stress and inflammatory response. *Int. J. Mol. Med.* 45 (5), 1305–1316. doi:10.3892/ijmm.2020.4520
- Zhu, L., Gong, X., Gong, J., Xuan, Y., Fu, T., Ni, S., et al. (2020). Notoginsenoside R1 upregulates miR-221-3p expression to alleviate ox-LDL-induced apoptosis, inflammation, and oxidative stress by inhibiting the TLR4/NF- κ B pathway in HUVECs. *Braz. J. Med. Biol. Res.* 53 (6), e9346. doi:10.1590/1414-431x20209346
- Zhu, Y., Xian, X., Wang, Z., Bi, Y., Chen, Q., Han, X., et al. (2018). Research progress on the relationship between atherosclerosis and inflammation. *Biomolecules* 8 (3), 80. doi:10.3390/biom8030080

Glossary

| | |
|--------------------------------|--|
| ABCA1 | ATP-binding cassette transporter A1 |
| ADMA | Asymmetric dimethylarginine |
| ApoE^{-/-} mice | Apolipoprotein e-knockout mice |
| ceRNA | Competitive endogenous RNA |
| circRNAs | Circular RNAs |
| CLIC4 | Chloride intracellular channel 4 |
| DDAH1 | Dimethylarginine dimethylaminohydrolase-1 |
| DUSP5 | Dual-specificity phosphatase 5 |
| EGCG | Epigallocatechin gallate |
| eNOS | Endothelial nitric oxide synthase |
| EZH2 | Enhancer of zeste homolog 2 |
| HAEC | Human aortic endothelial cells |
| HASMCs | Human airway smooth muscle cells |
| HA-VSMCs | Human aortic vascular smooth muscle cells |
| HMGA2 | High mobility group AT-hook 2 |
| HMGB1 | High mobility group box 1 protein |
| HMG-CoA | Hydroxymethylglutaryl-CoA |
| HUVECs | Human umbilical vein endothelial cells |
| hVSMCs | Human vascular smooth muscle cells |
| IGF1 | Insulin-like growth factor 1 |
| KLF4 | Krüppel-like factor 4 |
| lncRNAs | Long non-coding RNAs |
| MCP-1 | Monocyte chemoattractant protein 1 |
| miRNAs | MicroRNAs |
| MKP-1 | Mitogen-activated protein kinase phosphatase-1 |
| MyD88 | Myeloid differentiation primary response gene 88 |
| ncRNAs | Non-coding RNAs |
| NF-κB | Nuclear factor-κB |
| NO | Nitric oxide |
| Nrf2 | Nuclear factor erythroid-2-related factor 2 |
| ox-LDL | Oxidized low-density lipoprotein |
| p53 | Tumor protein 53 |
| PCSK9 | Proprotein convertase subtilisin kexin 9 |
| PTEN | Phosphatase and tensin homolog |
| RAECs | Rat aortic endothelial cells |
| RAW264.7 | RAW 264.7 mouse leukemia macrophage cell line |
| SD rats | Sprague-dawley rats |
| SIRT1 | Sirtuin 1 |
| SIRT6 | Sirtuin 6 |
| Smad3 | Sma- and mad-related protein 3 |

| | |
|---------------|--|
| SOD | Superoxide dismutase |
| STAT3 | Signal transducer and activator of transcription 3 |
| TET2 | Tet methylcytosine dioxygenase 2 |
| THP-1 | Human acute monocytic leukemia cell line |
| TLR4 | Toll-like receptor 4 |
| TNF-α | Tumor necrosis factor-α |
| TPM1 | Tropomyosin 1 |
| TRAF6 | TNF receptor-associated factor 6 |
| VCAM-1 | Vascular cell adhesion molecule-1 |
| VECs | Vascular endothelial cells |
| VLDLR | Very low-density lipoprotein receptor |
| VSMCs | Vascular smooth muscle cells |



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Decoding signaling mechanisms: unraveling the targets of guanylate cyclase agonists in cardiovascular and digestive diseases

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Soluble guanylate cyclase agonists and guanylate cyclase C agonists are two popular drugs for diseases of the cardiovascular system and digestive systems. The common denominator in these conditions is the potential therapeutic target of guanylate cyclase. Thanks to in-depth explorations of their underlying signaling mechanisms, the targets of these drugs are becoming clearer. This review explains the recent research progress regarding potential drugs in this class by introducing representative drugs and current findings on them.

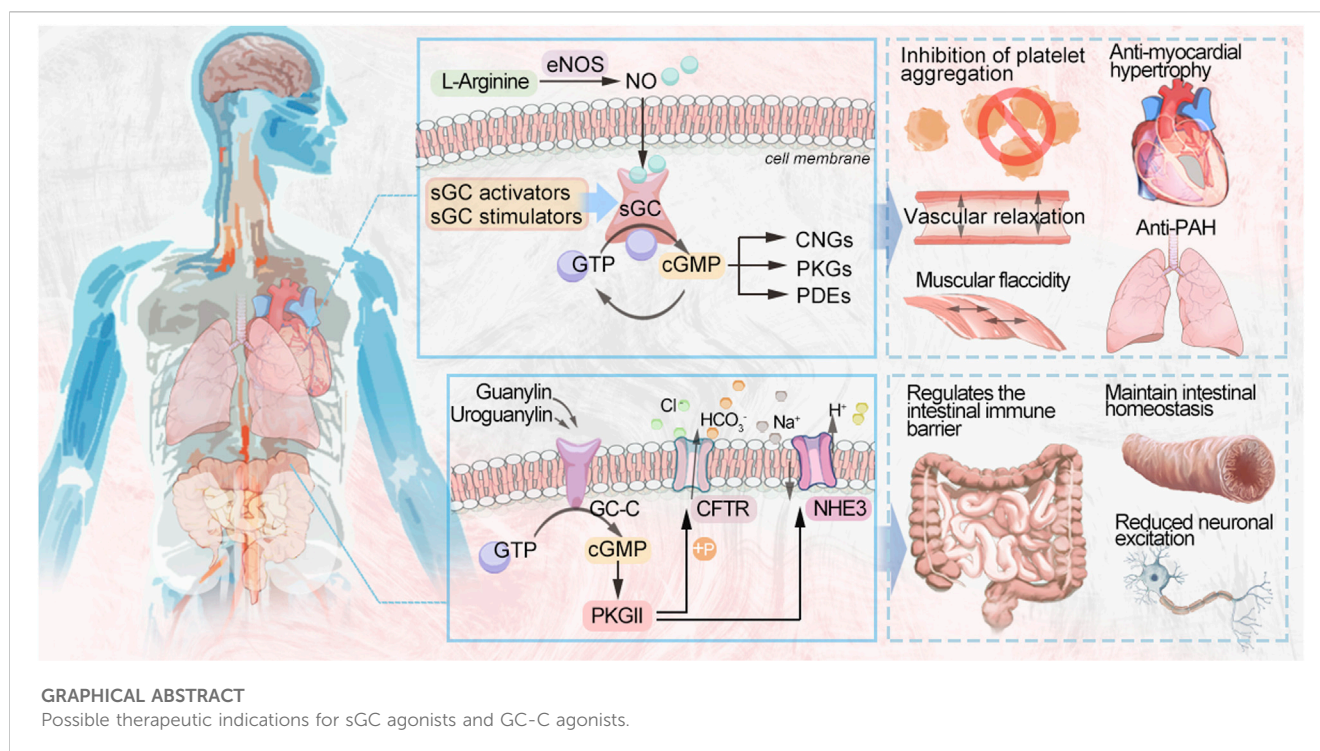
KEYWORDS

soluble guanylate cyclase stimulators, soluble guanylate cyclase activators, guanylate cyclase-c agonists, mechanism, signaling pathway

Introduction

Guanylate cyclase (GC) is a protease that catalyzes the conversion of guanylate triphosphate (GTP) to cyclic guanosine monophosphate (cGMP). Based on the properties of the distributed form of the enzyme, it can be divided into two categories: membrane-bound guanylate cyclase and soluble guanylate cyclase (sGC). Both are widely distributed in the human body, including the lung, brain, kidney, blood vessels, and other tissues (Garbers, 1990). The agonists of membrane-bound guanylate cyclase are peptides (natriuretic peptides A, B, and C), and the agonists of sGC are gaseous mediators (NO and CO) (Grzesk et al., 2023).

sGC is a heterodimer with an α -subunit and β -subunit, of which the latter contains the heme-nitric oxide/oxygen (H-NOX) domain (Argyriou et al., 2021). As a result of its affinity for nitric oxide (NO), this enzyme has also been called an NO-sensitive guanosyl cyclase. NO binding to sGC heme increases GTP cyclase activity, resulting in the production of cGMP, which regulates multiple signaling pathways in cells (Stasch et al., 2011). The cardiovascular, pulmonary, and neurological systems, as well as organs like the kidney, brain, and liver, are highly dependent on NO-sGC-cGMP regulation. Regarding its role in regulation, fibroblasts, cardiomyocytes, platelets, neurons, and immune cells are also affected by cGMP, and it controls fibrosis, the inflammatory response, and neurotransmission (Derbyshire and Marletta, 2009; Grzesk and Nowacznyk, 2021; Nowacznyk et al., 2021).



Guanylate cyclase C (GC-C) is a member of the membrane-bound GC family. It consists of an extracellular domain (ECD) and an intracellular domain, connected by a single strand across the membrane region. In the cell, it synthesizes cGMP and regulates an array of intracellular physiological functions (Hasegawa and Shimonishi, 2005). Reviewing recent progress in research on sGC agonists and GC-C agonists, along with their mechanisms, is the purpose of this paper.

Soluble guanylate cyclase agonists

sGC is a heterodimeric enzyme with a prosthetic heme group (Archer, 2013). Many physiological functions depend on NO signaling as their primary sensor. sGC binding of NO results in a significant increase in sGC enzyme activity. Additionally, cGMP is produced in a complex with its ligand, NO. The cascades of NO-driven signaling are amplified by sGC (Kang et al., 2019). Only the sGC receptor responds to gaseous NO (Schmidt et al., 2003). cGMPs produced by sGC help regulate the cardiovascular, neuronal, and digestive systems. In pharmacology and therapeutics, improving sGC activity and preventing or reversing inactivation are important goals (Stuehr et al., 2021). There are two forms of sGC in the body: those that respond to NO and those that do not. In brief, sGC contains a heme moiety, which is either ferrous (reduced sGC) or ferric (oxidized sGC). In these forms, the sGC stimulator can only target the reduced, heme-containing sGC, whereas the sGC activator binds to the oxidized or heme-free sGC, resulting in increased production of cGMPs (Dai and Stuehr, 2022). Although sGC activators stimulate heme-containing enzymes independently of NO, NO enhances their activity; even when oxidative stress occurs, cGMP is released by sGC activators (Dai and Stuehr,

2022). These novel pharmacological principles of sGC stimulation and activation appear to have very broad therapeutic potential. Signaling pathways such as NO-sGC-cGMP play an important role in cellular homeostatic maintenance and physiology (Sandner et al., 2021a). The current progress in understanding sGC agonists is summarized in Tables 1, 2.

The NO-sGC-cGMP signaling pathway

Three different nitric oxide synthases (NOS) are involved in reducing L-arginine to citrulline endogenously (Figure 1): endothelial nitric oxide synthase (eNOS), neuronal nitric oxide synthase (nNOS), and induced nitric oxide synthase (iNOS) (Sandner et al., 2018). During vascular NO-sGC-cGMP signaling, L-arginine is converted to NO in the endothelial monolayer by endothelial nitric oxide synthase (eNOS) and diffuses into the vascular lumen and the vessel wall, thereby activating sGC. Heme-dependent sGC stimulators and nonheme-dependent sGC activators increase cellular cGMP concentrations by directly activating sGC, leading to vascular relaxation and inhibiting platelet aggregation (Evora et al., 2012). NO produced through endothelial cells plays an important biological role. cGMP is synthesized by NO by binding to the active heme-containing sGC in vascular smooth muscle cells. Smooth muscle sGCs are NO signaling targets. The binding of NO to sGC leads to the conversion of GTP to cGMP. The resulting cGMP is hydrolyzed after binding to one of three types of cGMP effector proteins, including gated cation channels, the protein kinases (PKGs) that are dependent on cGMP, and the phosphodiesterases (PDEs) that are regulated by it (Feil et al., 2022).

TABLE 1 Current progress on soluble guanylate cyclase stimulators.

| Drug | Trial name | Design | Population | Dose | Trial ID | Endpoint | Safety outcome | Stage of development | Conclusion | Reference |
|------------|--------------------|---|--------------------------|------------------------------------|-------------|---|-------------------------------------|------------------------|--|--|
| Riociguat | PATENT-1 | Randomized, Double-blind, Placebo-controlled, Multicenter, Multinational Trial | Patients with PAH | 1–2.5 mg tid | NCT00810693 | Change in 6MWD | AEs, SAEs and deaths | Completed | Riociguat significantly improved exercise capacity and secondary efficacy end points in patients with PAH | Toxvig et al. (2019) |
| Riociguat | RESPITE | Open-label, International, Multicenter, Single-arm, Uncontrolled, Phase IIIb Trial | Patients with CTEPH | 1–2.5 mg tid | NCT02007629 | Change in 6MWD | AEs | Completed | Patients with PAH may benefit from switching from PDE5i to riociguat | Barnikel et al. (2022) |
| Riociguat | RioSAPH | Double Blind, Placebo Controlled Trial | Patients with SAPH | 2.5 mg tid | NCT02625558 | Time until clinical worsening | AEs | Unknown | Riociguat was effective in preventing clinical worsening and improving exercise capacity in patients with SAPH. | Baughman et al. (2022) |
| Riociguat | ISE-IIP | Randomized, Double-blind, Placebo-controlled Phase II Trial | Patients with IIP or PAH | 0.5–2.5 mg tid | NCT02138825 | Change in PVR at week 26 | AEs and SAEs | Terminated | Riociguat should not be used in patients with PH-IIP due to increased serious adverse events and mortality and an unfavorable risk-benefit profile | Nathan et al. (2019) |
| Riociguat | PATENT-CHILD | Open-label, Individual Trial | Children with PAH | 0.5–2.5 mg tid | NCT02562235 | Change in 6MWD to Week 16 | AEs and SAEs | Active, not recruiting | A suitable riociguat dosing strategy for pediatric patients with PAH have an acceptable safety profile with potential efficacy signals | Garcia et al. (2022) |
| Riociguat | RISE-SSc | Randomized, Double-Blind, Placebo-Controlled Phase II Trial | Patients with dcSSc | 0.5–2.5 mg tid | NCT02283762 | Change in mRSS | AEs and SAEs | Completed | Riociguat did not significantly benefit mRSS versus placebo | Khanna et al. (2020) |
| Vericiguat | SOCRATES-REDUCED | Randomized Parallel-group, Placebo-controlled, Double-blind, Multicenter Phase II Trial | Patients with HF | 1.25 mg, 2.5 mg, 5 mg or 10 mg, qd | NCT01951625 | Change in NT-proBNP | Changes in LVEF; LVEDV; LVESV | Completed | Vericiguat had no significant effect on NT-proBNP levels in patients with worsening chronic HF and reduced LVEF but was well-tolerated | Gheorghiad et al. (2015) |
| Vericiguat | VICTORIA | Randomized Parallel-Group, Placebo-Controlled, Double-Blind, Multi-Center Pivotal Phase III Trial | Patients with HF | 2.5, 5.0, or 10.0 mg, qd | NCT02861534 | Time to Composite Endpoint of Cardiovascular Death or Heart Failure Hospitalization | Symptomatic hypotension and syncope | Completed | Compared to placebo, vericiguat significantly reduced the incidence of the composite endpoint | Armstrong et al. (2020a) |
| Vericiguat | SOCRATES-PRESERVED | Randomized Parallel-group, Placebo-controlled, Double-blind, | Patients with HFpEF | fixed-dose (1.25 mg or 2.5 mg) and | NCT01951638 | Change in NT-proBNP and LAV From Baseline | AEs and SAEs | Completed | Vericiguat was well tolerated and did not alter NT-proBNP and LAV at | Pieske et al. (2017) |

(Continued on following page)

TABLE 1 (Continued) Current progress on soluble guanylate cyclase stimulators.

| Drug | Trial name | Design | Population | Dose | Trial ID | Endpoint | Safety outcome | Stage of development | Conclusion | Reference |
|------------|----------------|--|---|-----------------------------------|-------------|---|----------------------|----------------------|--|--|
| | | Multicenter Dose Finding Phase II Trial | | up-titrated to 5 mg or 10 mg, qd | | | | | 12 weeks compared to placebo, but was associated with improved quality of life in patients with HFpEF | |
| Vericiguat | VITALITY | Multicenter, randomized, double-blind, placebo-controlled phase IIb trial | Patients with HFpEF | up-titrated to 10 mg or 15 mg, qd | NCT03547583 | Change in KCCQ physical limitation score and 6MWD from baseline | TEAEs | Completed | No significant change in KCCQ or 6MWD compared to placebo at 24 weeks | Armstrong et al. (2020b) |
| Vericiguat | VENICE | Multicenter, Randomized, Placebo-controlled, Double-blind Group Comparison Trial | Patients with CCSs | 2.5, 5, or 10 mg, qd | NCT02617550 | Measurements of the hemodynamic profile | AEs and SAEs | Completed | The combination of Vericiguat with nitroglycerin administered to patients with CCSs was well tolerated | Boettcher et al. (2022) |
| Praliguat | CAPACITY HFpEF | Multicenter, Randomized, Double-blind, Placebo-controlled, Phase 2 Trial | Patients with HFpEF | 10, 20, or 40 mg daily | NCT03254485 | Change in peak VO2 | TEAEs | Completed | These findings do not support the use of praliguat in patients with HFpEF. | Udelson et al. (2020) |
| Praliguat | / | Phase IIA, double-blind, placebo-controlled trial | Patients with 2 diabetes and hypertension | 20 mg bid or 40 mg qd | NCT03091920 | ABPM and HOMA-IR | TEAEs | Completed | Praliguat was well tolerated and showed positive trends in metabolic and BP variables | Hanrahan et al. (2020a) |
| Praliguat | / | Randomized, Double-Blind, Placebo-Controlled, Phase 2 Trial | Patients with type 2 Diabetes Mellitus combined with Diabetic Nephropathy | 20 mg or 40 mg qd | NCT03217591 | Change in urine albumin–creatinine ratio | TEAEs | Completed | Praliguat treatment did not significantly reduce albuminuria compared with placebo | Hanrahan et al. (2020a) |
| Olinciguat | / | Multicenter, Randomized, Double-blind, Placebo-controlled, Parallel-group, Single-dose, Phase 2a Study | Patients with Achalasia | Single 5 mg dose | NCT02931565 | Change in BFT | TEAEs, SAEs and ADOs | Terminated | / | / |
| Olinciguat | STRONG SCD | Randomized, Placebo-controlled, Phase 2 Study | Patients with Sickle Cell Disease | | NCT03285178 | / | TEAEs and SAEs | Completed | / | / |

TABLE 2 Current progress in soluble guanylate cyclase activators.

| Drug | Trial name | Design | Population | Dose | Trial ID | Endpoint | Safety outcome | Stage of development | Conclusion | Reference |
|-------------|----------------|--|---|--|-------------|--|---|----------------------|--|--|
| Cinaciguat | / | Placebo Controlled, Randomized, Double-blind, Multicenter, Multinational Phase IIb Study | Patients with ADHF | Uptitration from 100–600 µg/g, over maximum 48 h | NCT00559650 | Change in PCWP | AEs , SAEs and TEAEs | Terminated | Cinaciguat unloaded the heart in patients with ADHF. High doses were associated with hypotension | Erdmann et al. (2013) |
| Cinaciguat | COMPOSE 1 | Placebo Controlled, Randomized, Double-Blind, Multicenter, Phase IIb Study | Patients with ADHF | 50 µg/h; 100 µg/h or 150 µg/h during 48 h | NCT01065077 | Change in PCWP or LOCF | Change in heart rate; SBP; frequency of SAEs and TEAEs | Terminated | It is doubtful that further studies with intravenous cinaciguat would prove beneficial in ADHF patients | Gheorghiad et al. (2012) |
| Cinaciguat | COMPOSE 2 | Placebo Controlled, Randomized, Double-Blind, Multicenter, Phase IIb Study | Patients with ADHF | 10 µg/h and 25 µg/h during 48 h | NCT01067859 | Change in PCWP or LOCF | Change in heart rate; SBP; frequency of SAEs and TEAEs | Terminated | No statistical analysis was performed | Gheorghiad et al. (2012) |
| Cinaciguat | COMPOSEE EARLY | placebo controlled, double-blind and randomized study | Patients with ADHF | 50 µg/h; 100 µg/h or 150 µg/h during 48 h | NCT01064037 | The change in dyspnea assessed using a VAS or LOCF | Change in heart rate; SBP; frequency of SAEs and TEAEs | Terminated | No significant clinical benefit of cinaciguat | Gheorghiad et al. (2012) |
| Ataciguat | / | Phase Ib Randomized, Placebo-controlled, Double-blinded Study | Patients with Moderate CAVS | 50 mg, 100 mg, or 200 mg qd | NCT02049203 | The change in blood pressure following the transition from sitting to standing | Number of patients experiencing orthostatic hypotension | Completed | / | / |
| Ataciguat | SERENEATI | Randomized, Double-blind, Placebo-controlled, Crossover Study | patients with neuropathic pain | 200 mg qd | NCT00799656 | Change in average daily pain intensity; change in NPSI | Rescue medication intake | Completed | / | / |
| Ataciguat | ACCELA | Randomized, Double-blind, Placebo-controlled, Parallel Group Trial | Patients with PAD | / | NCT00443287 | change in ICD | AEs | Completed | / | / |
| Ataciguat | CAVS | Phase II Randomized, Placebo-Controlled, Double-Blinded Study | Patients with AVC | 200 mg qd | NCT02481258 | Changes in Aortic Valve Calcium Levels | / | Completed | / | / |
| MGV354 | / | Randomized, Double-masked, Placebo-Controlled, Safety, Tolerability and Early Efficacy Study | Patients with Ocular Hypertension or Glaucoma | 3 µg per eye to 300 µg per eye | NCT02743780 | Change in Diurnal IOP | AEs and TEAEs | Completed | Human glaucomatous trabecular meshwork may have levels of oxidized sGC that are too low to benefit from MGV354 | Stacy et al. (2018) |
| Runcaciguat | CONCORD | Randomized, Double-blind, Placebo-controlled, Multicenter Study | Patients with CKD With Diabetes and/or Hypertension | / | NCT04507061 | Mean change in UACR | TEAEs; number of subjects with early discontinuations | Completed | / | / |
| Runcaciguat | NEON-NPDR | | | Oral dose | NCT04722991 | DRSS improvement | TEAEs | | / | / |

(Continued on following page)

TABLE 2 (Continued) Current progress in soluble guanylate cyclase activators.

| Drug | Trial name | Design | Population | Dose | Trial ID | Endpoint | Safety outcome | Stage of development | Conclusion | Reference |
|------------|------------|--|----------------------------|----------------------------------|-------------|--------------------------------|------------------------------|------------------------|--|---|
| | | Phase 2 Randomized, Placebo-controlled, Double-masked Proof-of-concept Study | Patients with NPDR | | | | | Active, not recruiting | | |
| Moslicigat | ATMOS | Nonrandomized Two Part Multicenter, Open-label, Single Dose Trial | Patients with PAH or CTEPH | up to a maximum dose of 4,000 µg | NCT03754660 | reduction in PVR | TEAEs | Completed | / | / |
| BI-685509 | / | Randomized, Double-blind, Placebo-controlled and Parallel Group Trial | Patients with CSPH | / | NCT05161481 | Percentage change in HVPg | Decompensation events; CTCAE | Recruiting | / | Reiberger et al. (2023) |
| BI-685509 | / | Randomized, Open-label and Parallel Group Trial | Patients with CSPH | | NCT05282121 | Percentage change in HVPg | Decompensation events; CTCAE | Recruiting | / | Reiberger et al. (2023) |
| BI-685509 | / | Phase II, Randomized, Placebo-controlled, Double-blind, Parallel Group, Study | Patients with SSc | / | NCT05559580 | Rate of decline in FVC | Time to treatment failure | Recruiting | / | / |
| BI-685509 | / | Randomized, Double-blind (Within Dose Groups), Placebo Controlled and Parallel Group Trial | Patients with CKD | / | NCT04736628 | Change in UACR | / | Active, not recruiting | / | / |
| BI-685509 | / | Randomized, Double-blind, Placebo-controlled Trial | Patients with DKD | 1 mg tid; 3 mg qd; 3 mg tid | NCT03165227 | Change in log transformed UACR | AEs , SAEs | Completed | BI 685509 was generally well tolerated | Cherney et al. (2023) |
| BI-685509 | / | Randomized, Double-blind (Within Dose Groups), Placebo Controlled and Parallel Group Trial | Patients with NPDR | / | NCT04736628 | Change in log transformed UACR | / | Active, not recruiting | / | / |
| BI-685509 | | Randomized, Double-blind (Within Dose Groups), Placebo-controlled and Parallel Group Trial | Patients with DKD | / | NCT04750577 | Change in log transformed UACR | / | Completed | / | / |

PCWP, pulmonary capillary wedge pressure; AEs, adverse events; SAEs, serious adverse events; TEAEs, treatment-emergent adverse events; ADHF, acute decompensated heart failure; SBP, systolic blood pressure; VAS, visual analog scale; LOCF, last observation carried forward; CAVS, calcific aortic valve stenosis; NPDI, neuropathic pain symptom inventory; PAD, peripheral arterial disease; ICD, initial claudication distance; AVC, aortic valve calcification; CKD, chronic kidney disease; UACR, urinary albumin-to-creatinine ratio; NPDR, nonproliferative diabetic retinopathy; DRSS, diabetic retinopathy severity scale; PAH, pulmonary arterial hypertension; CTEPH, chronic thromboembolic pulmonary hypertension; CSPH, clinically significant portal hypertension; HVPg, hepatic venous pressure gradient; CTCAE, common terminology criteria for adverse events; SSc, systemic sclerosis; FVC, forced vital capacity; DKD, diabetic kidney disease.

Disruption of the NO-sGC-cGMP signaling pathway is central to the pathogenesis of pulmonary arterial hypertension (PAH) and chronic thromboembolic pulmonary hypertension (CTEPH), in which endothelial dysfunction leads to impaired NO synthesis. The progression of PAH and CTEPH is also associated with low NO production. The sGC stimulators have a dual role in that they directly stimulate the native form of the enzyme, making it more sensitive to endogenous NO and increasing sGC activity regardless of NO and cGMP levels, leading to an increase in cGMP. (Benza et al., 2019). The vasodilation effect of cGMP in the pulmonary circulation is mediated by a variety of subcellular mechanisms, one of which is the activation of cGMP-dependent protein kinase, phosphorylating calcium-sensitive potassium channels (BKCa), leading to potential hyperpolarization of the pulmonary artery smooth muscle membrane and inhibiting calcium inflow through L-type Ca^{2+} channels (LTCCs). sGC is a redox-sensitive enzyme that is activated by hydrogen peroxide, causing pulmonary artery blood vessels to dilate. However, in cases of excessive oxidative stress, as occurs in disease states, reactive oxygen species or nitrosylation can change the oxidation state of sGC from normal reduced heme iron (Fe^{2+}) to oxidized heme (Fe^{3+}), making it less active and less responsive to NO. The oxidized sGC then loses its heme portion, after which it will eventually be degraded by the proteasome. The heme-free form of sGC is the target of sGC activators (Dasgupta et al., 2015). In the pulmonary circulation, NO acts as an endogenous pulmonary vasodilator and that synthesized by the action of eNOS on L-arginine in pulmonary vascular endothelial cells and then diffuses to adjacent vascular smooth muscle cells (VSMCs) to activate sGC (Triposkiadis et al., 2022). In VSMCs, MLCP, RhoA, RGS-2, IRAG, and BKCa are phosphorylated by PKG to promote vasodilation (Klinger and Kadowitz, 2017; Sadek et al., 2020). Many proteins are changed in their phosphorylation state when endogenous PKG is activated, including VASP and IRAG, Rap1B and Rap1Gap2, GRP2, and IP3 receptors, among others, which inhibit platelets as well (Gambaryan, 2022). Several studies have suggested vasoconstriction/vasodilation and proliferation/anti-proliferation alterations as possible pathogenic mechanisms of PAH (Biswas et al., 2020). PKG acts in pulmonary artery smooth muscle cells (PASMCs) by several mechanisms. PKG phosphorylates the BKCa channel and MLCK, leading to the relaxation of PASMCs (Christou and Khalil, 2022; Barenco-Marins et al., 2023). Vasodilator-stimulated phosphoprotein (VASP) is an actin-binding protein, and phosphorylation of VASP by PKG inhibits PASMC proliferation (Chen et al., 2004). Bone morphogenetic protein (BMP) is a signaling molecule belonging to the transforming growth factor- β (TGF- β) superfamily, which plays an important role in regulating cell proliferation, differentiation, and apoptosis. Recent studies have found that PKGI enhances the phosphorylation of the downstream signal small mothers against decapentaplegic (SMAD) 1/5 by BMP, promotes the antiproliferative and pro-differentiation effects of BMP, and keeps PASMCs in a proliferation-inhibited state (Watanabe, 2018).

The NO-sGC-cGMP signaling pathway maintains the normal function of the cardiovascular system in healthy individuals, and sGC activity falls as heart failure with reduced ejection fraction (HFrEF) progresses due to endothelial dysfunction and oxidative stress. sGC stimulation leads to increased cGMP synthesis, which can inhibit myocardial fibrosis, reduce vascular wall hardness, and

induce vasodilation (Belenkov and Kozhevnikova, 2023). A lack of sGC stimulation and a decrease in cGMP production are associated with heart failure (HF) and decreased NO bioavailability. Elevated levels of plasma inflammatory cytokines, including TNF- α and IL-6, in patients with HF are associated with endothelial dysfunction with low NO-sGC-cGMP signaling in the heart and blood vessels (Numata and Takimoto, 2022). Impairment of the NO-sGC-cGMP pathway in HF, a key second messenger pathway mediating vascular and cardiac dilation, in patients with dysfunction of the endothelium, myocardium, and blood vessels may play a role in the progression of cardiovascular disease (CVD). Notably, both HFrEF and heart failure with preserved ejection fraction (HFpEF) patients have cGMP deficiency. There are also cases where oxidative stress results in heart muscle cell loss, collagen replacement, and fibrosis due to autophagy, apoptosis, or necrosis (Hulot et al., 2021). Disruption of the NO-sGC-cGMP pathway results in the narrowing of blood vessels, clumping of platelets, inflammation, scarring, and notably, maladaptive enlargement of the heart. Hence, NO-sGC-cGMP pathway restoration is a promising pharmacological target for HF treatment (Rudebusch et al., 2022). In the heart, PKG phosphorylates phospholamban (PLB) (Eggermont et al., 1988) and RyR2 (Xiao et al., 2006), thus activating sarcoplasmic reticulum Ca^{2+} ATPase (SERCA) to promote the transport of calcium ions to the endoplasmic reticulum. In addition, PKG can phosphorylate numerous membrane channels, including L-type calcium channels (Yang et al., 2007) and transient receptor potential typical channel type 6. This ultimately reduces the inflow of extracellular calcium. Troponin I (Tsai and Kass, 2009), titin (Kruger et al., 2009), and CMYBP-C (Thoonen et al., 2015) act as structural proteins that regulate contraction of the myocardium, leading to myocardial relaxation through phosphorylation of PKG. PDE5 breaks down cGMP in cardiomyocytes into GTP, which is eventually recycled (Mihalek et al., 2022). Its activity counteracts vascular constriction and helps maintain vital organs, the highest expression of sGC being found in perfusion cardiomyocytes. cGMP produced by sGC causes ventricular diastole and decreased contractility (Hulot et al., 2021). In anti-myocardial hypertrophy, regulators of G-protein signaling (RGS), namely, RGS2 and RGS4, have a central role, leading to cGMP-mediated anti-myocardial hypertrophic effects by inactivating G-protein-coupled signaling, as RGS2 and RGS4 are targets of PKG (Klaiber et al., 2010). The Ca^{2+} -dependent mechanism of calcitonin/nFAT hypertrophy is further inhibited by the phosphorylation of the LTCC and transient receptor potential canonical channel type 6 (TRPC6) channels by cGMP/PKG (Kinoshita et al., 2010), and these mechanisms may be related to cGMP-mediated antihypertrophy, antifibrosis and alleviation of cardiac dysfunction (Sandner and Stasch, 2017).

Since the NO-cGMP signaling cascade is active in many tissues *in vivo*, the pathway is currently being explored for other roles in conditions other than cardiovascular disease, such as chronic kidney disease, fibrotic disease, neuroprotection, and dementia (Sandner, 2018). TGF- β signaling plays an important role in cellular fibrosis, and several preclinical studies have demonstrated that inhibition of TGF- β signaling exerts a potent antifibrotic effect in different organs in a variety of animal models (Distler et al., 2019; Englert et al., 2023). PKG inhibits the phosphorylation of extracellular signal-regulated kinase (ERK) by TGF- β signaling, suppresses ERK

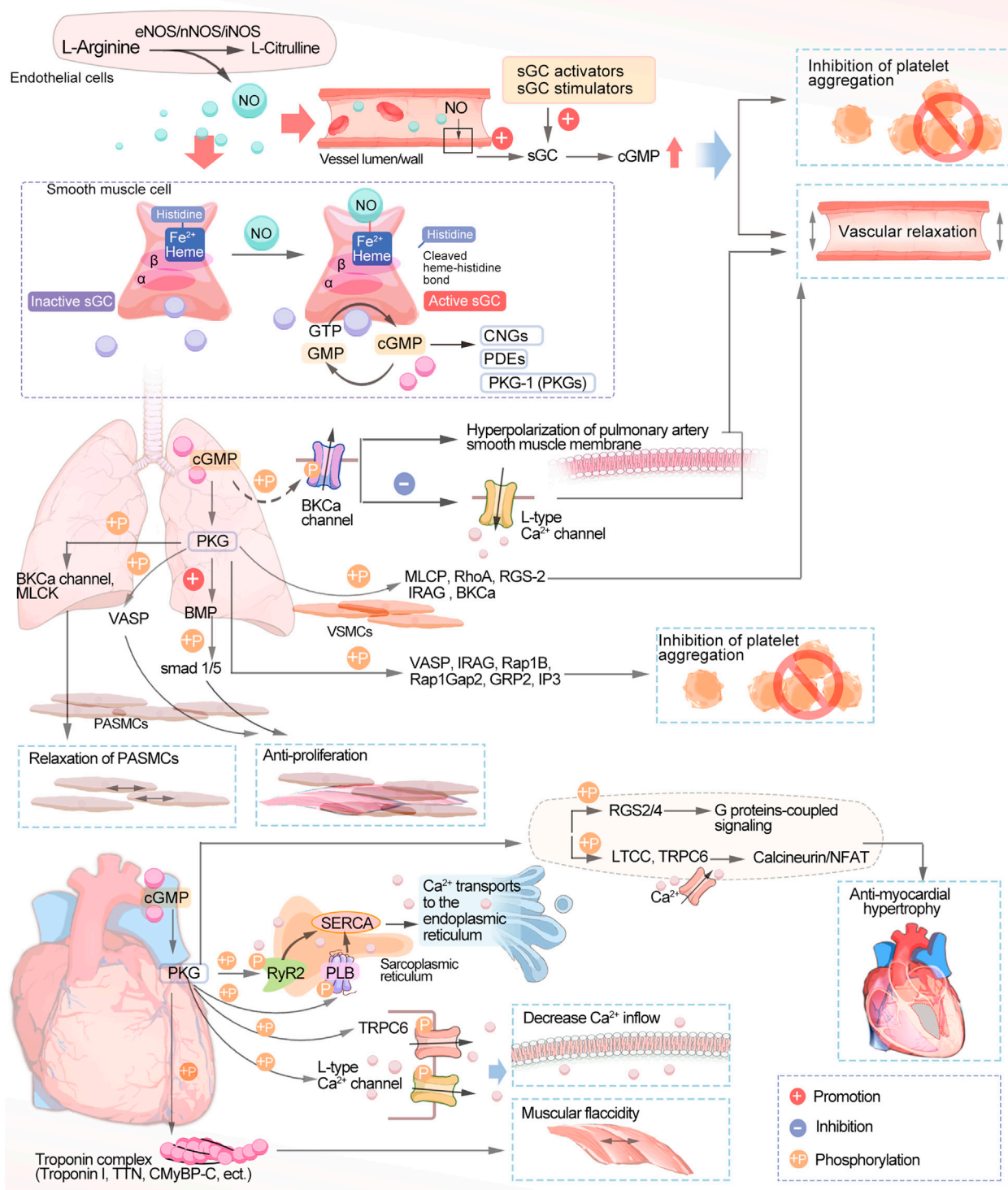


FIGURE 1

Schematic diagram of the NO-sGC-cGMP signaling pathway. NO, nitric oxide; sGC, soluble guanylate cyclase; cGMP, cyclic guanosine monophosphate; GTP, guanosine triphosphate; CNGs, cyclic nucleotide-gated ion channels; PDEs, phosphodiesterases; PKGs, protein kinases; BKCa, calcium-sensitive potassium channels; MLCK, myosin light chain kinase; VASP, vasodilator-stimulated phosphoprotein; BMP, bone morphogenetic protein; SMAD, small mothers against decapentaplegic; PSMCs, pulmonary artery smooth muscle cells; MLCP, myosin light chain phosphatase; RhoA, Ras homolog family member A; RGS-2, regulator of G-protein signaling 2; Rap1Gap2, Rap1 GTPase-activating protein 2; Rap1b, Ras-related protein 1; IRAG, IP3-induced calcium release; GRP2, guanyl-releasing protein 2; TTN, Titin; CMYBP-C, Cardiac myosin-binding protein-C; SERCA, sarco endoplasmic reticulum Ca²⁺-ATPase; PLB, phospholamban; RyR2, cardiac ryanodine receptor; TRPC6, transient receptor potential canonical channel type 6; RGS, regulator of G protein signaling; LTCCs, L-type calcium channels; NFAT, calcineurin-nuclear factor of activated T cells.

signaling, and prevents its translocation to the nucleus, blocking TGF- β -mediated extracellular matrix (ECM) generation, fibroblast differentiation to myofibroblasts, and cell proliferation (Hu et al., 2017; Sandner and Stasch, 2017).

Furthermore, cGMP modulates renal blood flow, renin secretion, glomerular function, and tubular exchange processes through its direct effects on cGMP signaling cascades. It is possible to develop renal diseases such as chronic kidney disease (CKD) when NO-sGC-cGMP signaling is downregulated. As a result, therapeutic strategies that maintain or increase cGMP activity may be effective against progressive kidney disease (Krishnan et al., 2018).

Riociguat

Currently, riociguat is the only FDA-approved sGC stimulator for treating PAH and CTEPH. It is an orally administered drug that can be rapidly absorbed, with a bioavailability of 94.3%. Riociguat's half-life varies significantly from individual to individual, at approximately 12 h for PAH/CTEPH patients and 7 h for healthy people. Riociguat promotes cGMP synthesis. It has shown good efficacy in clinical trials and is well tolerated. Riociguat has a valuable place in the treatment of pulmonary hypertension (Kenny et al., 2022).

Riociguat in PAH

The IPATENT-1 trial (NCT00810693) found that riociguat significantly improved the 6-min walking distance of PAH patients by 36 m (m) over placebo. The PHIRST-1 trial (NCT00125918) and the SUPER-1 trial (NCT00644605) trials showed that riociguat had similar results to those of tadalafil (mean difference of 33 m) and sildenafil (mean difference of 50 m), which are both rivals of this product. Pharmacokinetics (PK) and adverse events (AEs) were also similar. Riociguat was used in the RESPITE study (NCT02007629) to treat PAH patients who failed to respond adequately to either tadalafil or sildenafil. Based on these findings, riociguat may outperform PDE-5 inhibitors in efficacy (Toxvig et al., 2019). Riociguat has not shown any new safety signals since the FDA approved it in 2013, according to an evidence-based review of its safety and tolerability. The overall incidence of AEs was 93%–96%. The most common AE types are nasopharyngitis and peripheral edema. The most common severe AEs (SAEs) include syncope, right ventricular (RV) failure, hypotension, and hemoptysis. Although common side effects are reported, they are tolerated in 87%–92% of people (Donaldson et al., 2020). It is also well tolerated in older patients (Barnikel et al., 2022). Study SE-IIP (NCT02138825) of idiopathic interstitial pneumonia with pulmonary hypertension (PH) was terminated early, as patients taking riociguat experienced a higher occurrence of SAEs and mortality, as well as no efficacy signal. In the main study, 11 patients died (8 in the riociguat group and 3 in the placebo group); riociguat was associated with more SAEs among PH-IIP patients, as well as adverse risk/benefit profiles. Therefore, patients with PH-IIP should avoid using riociguat (Nathan et al., 2019). This meta-analysis encompassed the incorporation of eight randomized controlled trials with 1,606 participants of riociguat's effects on all types of PH. For PAH and CTEPH patients, riociguat treatment significantly extended the 6MWD compared with placebo and decreased N-terminal pro-B-type natriuretic peptide (NT-

proBNP), mean pulmonary artery pressure (PAP), pulmonary vascular resistance (PVR), and right atrial pressure (RAP). The cardiac index (CI) increased, cardiac output increased, and adverse events and clinical exacerbations decreased. A significant difference in efficacy outcomes and safety outcomes was not observed in other types of PH. Patients with PAH and CTEPH benefit from riociguat treatment, but patients with other types of PH only see partial hemodynamic improvements (Wang et al., 2021). The haemoDYNAMIC trial (NCT02744339) was a placebo-controlled, randomized, double-blind clinical trial that enrolled 114 patients with PH combined with HFpEF, who were randomly assigned to take riociguat or placebo. At 26 weeks, the riociguat group was significantly better in the primary efficacy measure, resting cardiac output, as determined through a right cardiac catheter (Dachs et al., 2022). In a multicenter, phase III, open-label, randomized controlled trial (NCT02634203), at week 26, riociguat therapy resulted in a more significant decrease in pulmonary vascular resistance than balloon pulmonary angioplasty (BPA) in 53 inoperable CTEPH patients, while BPA was performed on 52 patients. Treatment-related SAEs occurred more frequently in the BPA group: 22 of 52 (42%) of them experienced treatment-related SAEs vs. 5 of 53 (9%) in the riociguat group. Out of the 52 patients in the BPA group, 18 (35%) experienced lung injury, while 2 out of 53 patients (4%) reported severe hypotension leading to syncope. Deaths related to treatment were not reported (Jais et al., 2022). For patients requiring combination therapy, one review mentioned the possibility of pretreatment with riociguat plus an endothelin receptor antagonist in patients with PAH at high risk of death after 1 year (Rahaghi et al., 2023). A trial was conducted to test the combination of riociguat and ambrisentan in patients suffering from functional grade III PAH using a prospective, single-arm, open-label approach (NCT02634203) (Weatherald et al., 2022). In patients with sarcoidosis-associated pulmonary hypertension (SAPH), a double-blind placebo-controlled trial compared riociguat with placebo and examined the outcome of prolonged clinical worsening events (NCT02625558). Sixteen patients were randomized to riociguat ($n = 8$) and placebo ($n = 8$). By log-rank analysis, patients treated with riociguat stayed in the study significantly longer, their 6MWD scores tending to increase. At 1 year, riociguat was effective in preventing deterioration and in improving motor capacity in patients with clinical SAPH (Baughman et al., 2022). Riociguat is currently approved for use in adults only. It was tested for use in children in a multicenter, single-trial, 24-week, open-label phase 3 study, PATENT-CHILD (NCT02562235). The PK and safety of oral riociguat in pediatric patients with PAH were evaluated in World Health Organization Functional Classification (WHO-FC) Grade I–III patients aged 6–17 years who were being treated with stable endothelin receptor antagonists and/or prostacyclin analogs. They were given 0.5–2.5 mg of riociguat three times a day. A total of twenty-four individuals, with an average age of 12.8 years, were enrolled. Eighteen of these were WHO-FC II. Twenty patients (83%) reported primarily mild or moderate AEs. SAEs occurred in 4 cases (17%). All problems were resolved by the end of the study, and two out of the total (8%) were believed to be associated with the experimental medication. There were 3 cases of hypotension and 1 case of hemoptysis (all mild/moderate intensity). These children had similar blood concentrations of riociguat to those published in

adult patients. From baseline to week 24, the mean \pm standard deviation of the 6-min walking distance of patients increased by 23 ± 69 m ($n = 19$), and the mean NT-proBNP decreased by -66 ± 585 pg/mL ($n = 14$). The WHO-FC of the patients did not change. A clinical worsening event occurred in two patients after they were hospitalized with right heart failure. Riociguat is a safe and effective treatment for pediatric PAH, based on the PK results. The data suggest that riociguat has an acceptable safety profile and a potential efficacy signal in a pediatric cohort (Garcia et al., 2022).

Anti-cardiac-remodeling effects of riociguat

To test the hypothesis that sGC stimulation with riociguat can prevent pathological cardiac remodeling and HF caused by chronic pressure overload, an animal model of C57BL/6N mouse HF was established. After 3 weeks of transverse aortic constriction (TAC) treatment, the animals were randomized to receive riociguat or its solvent (Sol). After 5 weeks of treatment, the left ventricular rejection fraction (LVEF) improved (TAC + Rio $43.4\% \pm 6.0\%$, TAC + Sol $20.9\% \pm 4.3\%$; $p < 0.001$), and the left ventricular mass to body weight (LVM/BW) ratio, myocardial fibrosis, and the myocyte cross-sectional area decreased (5.5 ± 0.4 mg/g vs 7.7 ± 0.7 mg/g; $p < 0.001$). RNA sequencing indicated that riociguat administration reduced the expression of myocardial remodeling genes (e.g., Nppa, Nppb, Myh7, collagen genes) and downregulated signaling pathways associated with hypertrophic cardiomyopathy and heart failure. Muscle and fibroblast cultures showed a reversal of pathological stress responses when riociguat was administered (Rudebusch et al., 2022). Another bioinformatic analysis of C57BL/6N mice subjected to TAC showed that riociguat administration improved the expression of markers of pathways, metabolism, and energy production related to cardiovascular disease induced by TAC. Changes in the levels of myosin heavy chain 7 (MYH7), cardiac phosphoprotein (PLN), and ankyrin repeat domain-containing protein 1 (ANKRD1) were reversed. Riociguat also attenuates TAC-induced changes in left ventricular microRNA levels. This suggests that riociguat has beneficial effects on cardiac structure and function during stress overload, supporting the potential use of riociguat as a novel treatment for HF (Benkner et al., 2022).

Riociguat in diffuse cutaneous systemic sclerosis (dcSSc) and peripheral arterial disease

The effectiveness and safety of riociguat in patients at high risk of progressing skin fibrosis in early dcSSc were evaluated in a randomized, double-blind, placebo-controlled phase 1b trial (NCT02283762), an 18-month study of 60 participants with a modified Rodman skin score (mRSS) of 10–22 units who received riociguat orally three times a day for 18 months ($n = 60$). At week 52, the change in mRSS units from baseline was -2.09 ± 5.66 ($n = 57$) in the riociguat group and -0.77 ± 8.24 ($n = 52$) in the placebo group (mean least squares difference was -2.34 (95% CI: -4.99 – 0.30) ($p = 0.08$). Among patients with interstitial lung disease, forced lung capacity decreased by 2.7% in the riociguat group and 7.6% in the placebo group. In the riociguat group, 41.3% (19 out of 46) of patients showed an improvement of 250% in their Raynaud scores at week 14, compared to 26.0% (13 out of 50) in the placebo group. In the safety assessment, no new signals of inflammation or treatment-related death were identified. Riociguat had no significant

benefit on mRSS units. A secondary analysis and exploratory analysis revealed potential efficacy signals worth testing in further trials. Riociguat was well tolerated (Khanna et al., 2020).

Researchers have studied new blood vessel formation in mice with limb ischemia following the administration of riociguat. A 28-day treatment with 3 mg/kg/d riociguat was given intragastrically to C57BL/6 mice. Induction of posterior limb ischemia was achieved by surgically removing the femoral artery after 2 weeks of treatment. *In vitro* matrix tests showed that riociguat stimulated tubule formation in human umbilical vein endothelial cells (HUVECs) in a dose-dependent manner. Cell migration (assessed by scratch test) indices were also increased in HUVECs treated with riociguat compared to controls. At the cellular level, administration of riociguat resulted in swift initiation of the p44/p42 mitogen-activated protein kinase (MAPK) cascade in HUVECs. HUVECs treated with riociguat exhibited a reduced response to PKG inhibition. During ischemia, riociguat therapy improves blood flow recovery (as measured by laser Doppler imaging) and increases capillary density in ischemic muscle (as measured by CD31 immunostaining). The clinical results have shown that these indices reduce movement disorders and ischemic injuries significantly. The number of bone marrow-derived proangiogenic cells (PACs) was also increased by 94% when mice were treated with riociguat. Riociguat treatment was associated with significant improvements in PAC function, including migration ability, adherence to endothelial monolayers, and integration into the endodermal tubular network. GC stimulants can promote angiogenesis and improve neovascularization after ischemia, whose mechanism involves PKG-dependent p44/p42 MAPK activation, an improvement in PAC function, and an increase in their quantity. In patients with severe atherosclerosis, sGC stimulation may offer a novel approach to reducing tissue ischemia (Dhahri et al., 2023).

Vericiguat

Vericiguat was the first sGC stimulator to be marketed as of 2021, for patients with symptomatic chronic heart failure accompanied by an ejection fraction of 45%, to reduce the risk of cardiovascular death and heart failure hospitalization after heart failure hospitalization or the need for outpatient IV diuretics (Markham and Duggan, 2021). Vericiguat works synergistically with endogenous NO and enhances the affinity of sGC for low levels of NO (Singh et al., 2017). Therefore, vericiguat therapy is expected to restore the activity of an impaired NO-sGC-cGMP pathway, resulting in a variety of pharmacological effects, including improved cardiac and vascular function and reduced levels of profibrotic and inflammatory pathway markers. By increasing cGMP levels, vericiguat can also cause vascular relaxation and enhance the control of vascular tone and myocardial dysfunction (Hulot et al., 2021).

The average steady-state vericiguat distribution volume in healthy subjects is approximately 44 L, with a high bioavailability of approximately 93% when taken with food at a dose of 10 mg. Its serum albumin binding rate is approximately 98%, and its clearance rate is 1.6% L/h. It takes 30 h for vericiguat to reach half-life in patients with HF. Ninety-five percent of the vericiguat dose is metabolized primarily by glucosylation of UGT1A1 (minor) and UGT1A9 (major), resulting in inactive N-glucuronic acid

metabolites. CYP450-mediated metabolism is a minor clearance pathway, accounting for only 5% of the total clearance capacity. Fifty-three percent of the metabolized drug is excreted in urine, and 45% is excreted in feces (Campbell et al., 2022; Chiles and Al-Horani, 2022). Vericiguat, at doses up to 10.0 mg QD for 7 days, is generally well tolerated in healthy men in Europe, China, and Japan. Oral vericiguat 15.0 mg was not well tolerated, and drug-related treatment-emergent adverse events (TEAEs) were predominantly neurologic disorders such as headache and postural dizziness, which may be related to the mode of action of vericiguat (namely, vasodilation) (Boettcher et al., 2021).

Vericiguat in HFrEF

Preliminary evidence regarding vericiguat in the context of HFrEF was derived from two clinical trials, namely, the phase II study SOCRATES-REDUCED and the phase III study VICTORIA (Vannuccini et al., 2022). Four hundred fifty-six patients with clinical stability with an LVEF less than 45% within 4 weeks of worsening chronic heart failure, defined as congestion symptoms and elevated natriuretic peptide levels requiring inpatient or outpatient intravenous diuretics, were randomly assigned to an arm of the SOCRATES-REDUCED trial (NCT01951625). Overall, 351 patients (77.0%) completed investigational drug therapy with 12 weeks of effective NT-proBNP levels without major protocol bias, meeting the criteria for primary endpoint evaluation. In the preliminary analysis, the AE rates were 77.2% and 71.4% in the placebo and 10 mg vericiguat groups, respectively. The effects of vericiguat on NT-proBNP levels at 12 weeks in patients with chronic heart failure exacerbations and reduced LVEF were not significant compared with placebo, but the vericiguat was well tolerated. This drug had a dose–response relationship, indicating that further clinical trials will be needed to determine the effectiveness of the drug in treating worsening chronic heart failure patients (Gheorghade et al., 2015). In the VICTORIA III trial (NCT02861534), 5,050 patients were diagnosed with chronic heart failure (class II, III, or IV according to the New York Heart Association) with an ejection fraction of less than 45%. They were given vericiguat (10 mg once daily, target dose) or placebo (no treatment), along with guideline-based medication. The combined result of cardiovascular death and the first hospitalization for heart failure was the primary outcome. At a median of 10.8 months, 897 of 2,526 patients (35.5%) in the vericiguat group had had a primary outcome event, compared with 972 of 2,524 patients (38.5%) in the placebo group (hazard ratio, 0.90; 95% CI, 0.82–0.98; $p = 0.02$). A total of 691 (27.4%) patients were hospitalized for heart failure in the vericiguat group versus 747 (29.6%) patients in the placebo group (hazard ratio 0.90; 95% CI, 0.81–1.00). There were 414 (16.4%) deaths from cardiovascular causes in the vericiguat group and 441 (17.5%) deaths from cardiovascular causes in the placebo group (hazard ratio, 0.93; 95% CI, 0.81–1.06). A total of 957 (37.9%) patients died from any cause or were hospitalized for heart failure in the vericiguat group versus 1,032 (40.9%) patients in the placebo group (hazard ratio: 0.90; 95% CI, 0.83–0.98; $p = 0.02$). Symptomatic hypotension occurred in 9.1% and 7.9% of patients in the vericiguat and placebo groups ($p = 0.12$), respectively, and syncope occurred in 4.0% and 3.5% of patients in the vericiguat and placebo groups ($p = 0.30$), respectively. In

conclusion, among high-risk patients with heart failure, treatment with vericiguat led to a lower incidence of death from cardiovascular causes or hospitalization for heart failure than placebo (Armstrong et al., 2020a).

Vericiguat in HFpEF

Socrates-preserved (NCT01951638) was a 12-week, double-blind, placebo-controlled, phase 2b trial of 447 patients with deteriorating symptoms of chronic hypertension and an LVEF $\geq 45\%$ (Pieske et al., 2017). Patients were randomized to receive placebo or vericiguat once daily (1.25, 2.5, 2.5–5.0, or 2.5–10.0 mg). Patients tolerated vericiguat well (AEs: vericiguat 10 mg arm, 69.8%; placebo, 73.1%), but the two groups had similar changes from baseline in NT-proBNP (one-sided $p = 0.90$, two-sided $p = 0.20$) and left atrial volume (one-sided $p = 0.81$, two-sided $p = 0.37$) at 12 weeks. A larger proportion of patients treated with 10 mg vericiguat achieved clinically meaningful improvements in the Kansas City Cardiomyopathy Questionnaire Clinical Summary Score (KCCQ-CSS) and the 5-dimensional EuroQol questionnaire (EQ-5D). Given the encouraging results in terms of quality of life, there is a need for further studies on the impact of vericiguat in patients with HF. The VITALITY study (NCT03547583) was designed to understand the efficacy and safety of vericiguat on quality of life and exercise tolerance in patients with HF and HFpEF (Armstrong et al., 2020b). A total of 789 patients had chronic HF, EF $\geq 45\%$, New York Heart Association functional class II or III, decompensation in the last 6 months (no hospitalization due to HF or need for diuretics for intravenous treatment of HF), and elevated natriuretic peptides. Patients were randomized to receive up to 15 mg ($n = 264$), 10 mg ($n = 263$) or placebo ($n = 262$). No significant differences were found between groups in the physical limitation score of the KCCQ or the 6MWD after 24 weeks of treatment. The different outcomes between SOCRATES-PRESERVED and VITALITY in terms of improving quality of life in patients with HFpEF are of interest and require further investigation.

Vericiguat in chronic coronary syndromes

Vericiguat plus nitroglycerin was compared to nitroglycerin alone for safety, tolerability, and pharmacodynamic effects in patients with chronic coronary syndromes (CCSs). The VENICE (NCT02617550) randomized, double-blinded, phase I, multicenter trial randomized 36 patients with CCSs to receive either 2.5 mg vericiguat (increased doses every 2 weeks to 5 mg and 10 mg) or placebo. There were 31 patients in the study (21 receiving vericiguat plus nitroglycerin; 10 receiving placebo plus nitroglycerin). The combination of vericiguat with nitroglycerin did not increase the number of adverse events nor the risk of SAEs in patients with CCSs (Boettcher et al., 2022).

Praliguat

Praliguat (also known as IW-1973) is an sGC stimulator in the clinical stage of testing. It is being studied in clinical studies for the treatment of heart failure and diabetic nephropathy with retained ejection. Treatment with 1–10 mg/kg IW-1973 significantly reduced blood pressure in rats with normal and spontaneous hypertension in nonclinical models. IW-1973 reduced blood pressure, inflammatory

factors, and markers of kidney disease, such as proteinuria and renal fibrosis, in rat models. IW-1973 has a wide tissue distribution. It shows renoprotective, anti-inflammatory, and antifibrotic effects (Tobin et al., 2018). In a randomized, placebo-controlled phase I study, different doses of praliguat were evaluated for safety, tolerability, PK, and pharmacodynamics (PD) in healthy adults ($n = 44$). It was tolerable at various doses, and no SAEs were reported. The most common adverse reactions were headaches and decreased blood pressure. The PK was proportional to the dose, and the effective half-life was 24–37 h. The administration of praliguat resulted in a rise in plasma cGMP that was proportional to the dosage, indicating activation of sGC. Repeated daily medication can lead to a drop in blood pressure (Hanrahan et al., 2019).

Praliguat in HFpEF

There are no approved sGC stimulants for treating HFpEF, but such treatments are being investigated. CAPACITY HFpEF (NCT03254485) is a phase 2, multicenter, randomized, double-blind, placebo-controlled, parallel-group trial designed to evaluate the safety and efficacy of approximately 181 patients with HFpEF over 12 weeks. A total of 155 participants have completed the trial. The praliguat group ($n = 65$) showed a change in the peak rate of oxygen consumption (Vo_2) of -0.26 mL/kg/min (95% CI, -0.83 to 0.31), compared to 0.04 mL/kg/min (95% CI, -0.49 – 0.56) in the placebo group ($n = 78$). Changes in the 6MWD were 41.4 m (95% CI, 8.2 – 74.5) and 58.1 m (95% CI, 26.1 – 90.1), respectively. No significant benefit of praliguat was observed in HFpEF compared to placebo over a 12-week follow-up period, but it was accompanied by more hypotension and headache. These findings do not support the use of praliguat in patients with HFpEF (Udelson et al., 2020).

Praliguat's different effects in patients with T2D

Praliguat has renoprotective properties. In a rat model of obese diabetic nephropathy (DN), praliguat alone reduced proteinuria. Praliguat monotherapy had no effect on hemodynamics, whereas combined enalapril reduced proteinuria, but monotherapy reduced blood pressure and did not reduce proteinuria (Liu et al., 2020). In a phase II trial (NCT03217591) involving 156 adults with type 2 diabetes, praliguat did not significantly reduce proteinuria over 12 weeks (Hanrahan et al., 2020b). Furthermore, a mouse model of diet-induced obesity revealed some beneficial metabolic effects of praliguat (Schwartzkopf et al., 2022). Another phase II, double-blind, placebo-controlled trial of praliguat (NCT03091920) involved 26 patients with type 2 diabetes combined with hypertension. Praliguat was well tolerated and showed positive trends in metabolic and BP variables (Hanrahan et al., 2020b).

One study evaluated the effect of praliguat on hind limb ischemic (HLI) recovery in mice with type 2 diabetes. Praliguat significantly increased the diameter of their small arteries, decreased the expression of intercellular adhesion molecule 1 (ICAM1), prevented the accumulation of oxidative proangiogenic and proinflammatory muscle fibers, and significantly downregulated the expression of Myh2 and Cxcl12 mRNA in cultured myoblasts (Foussard et al., 2023).

Other medicines

BAY-747

BAY-747 is a long-acting, next-generation GC stimulator for the treatment of refractory hypertension that is administered once daily and has sustained effects on blood pressure and heart rate (up to 24 h). BAY-747 is taken orally in a single dose of 0.5–15 mg in the form of a polyethylene glycol (PEG) solution and is well tolerated in healthy volunteers. Blood concentrations of BAY-747 peaked within 2–6 h, independent of dose intensity. A single dose of 3.5 mg oral BAY-747 significantly increased heart rate and reduced blood pressure and mean arterial pressure, these effects being most pronounced within the first 4 h after taking the study drug. A single oral dose of 10 mg BAY-747 had significant effects on heart rate, cardiac output, and cardiac index, with maximum effects achieved within 4 h of administration. In the 0.5–20 mg dose range, a single oral dose of BAY-747 did not appear to affect stroke volume (Vakalopoulos et al., 2023).

Olinciguat

Olinciguat is a new oral sGC stimulator currently in phase II clinical development (NCT02931565 and NCT03285178). Olinciguat has cardioprotective effects and reduces blood pressure. In addition, it has renal-protective effects, and in a rat ZSF1 model, there is a correlation between decreased levels of glucose, cholesterol, and triglycerides (Zimmer et al., 2020). In a mouse model of TNF α -induced inflammation, olinciguat treatment was associated with reduced levels of soluble adhesion derived from endothelial cells and white blood cells (Tchernychev et al., 2021). Accordingly, it may be suitable for treating diseases characterized by vascular and extravascular lesions as well as a wide range of potential therapeutic applications.

MK-2947

MK-2947 is a novel, potent, selective sGC stimulator (Brockunier et al., 2020). A pharmacological study demonstrated that MK-2947 effectively ameliorates angiogenic performance and blunts the myofibroblast-like profibrotic phenotype of SSc dermal microvascular endothelial cells (SSc-MVECs), thus providing new evidence for the benefit of repurposing sGC stimulators for SSc (Romano et al., 2023).

CYR715

CYR715, first described by Rennie et al., is a novel carboxylic acid-containing sGC stimulator that exhibited similar dose-dependent hemopharmacology in normotensive rats. Compared to the previously described IWP-051, CYR715 had a better pharmacokinetic profile in rats and exhibits similar dose-dependent hemodynamics in normotensive rats (Rennie et al., 2021). A recent study found that preincubating red blood cells from type 2 diabetes (T2D) patients with CYR715 and administering them to isolated rat hearts enhanced left ventricular diastolic pressure recovery, reduced infarct size, and mitigated endothelial dysfunction. Therefore, CYR715 appears to be an attractive therapeutic strategy for preventing cardiovascular injury in patients with T2D (Jiao et al., 2023).

6MWD, 6-min walking distance; PAH, pulmonary arterial hypertension; CTEPH, chronic thromboembolic pulmonary hypertension; PVR, pulmonary Vascular resistance; IIP, idiopathic interstitial pneumonias; dcSSc, diffuse cutaneous systemic sclerosis; AEs, adverse events; SAEs, serious adverse events; mRSS, modified Rodnan skin score; HF, heart failure; NT-proBNP, N-terminal pro-B-type natriuretic peptide; HF, heart failure; LVEF, left ventricular ejection fraction; LVEDV, left ventricular end-diastolic volume; LVESV, left ventricular end-systolic volume; LAV, left atrial volume; KCCQ, Kansas City Cardiomyopathy Questionnaire; CCSs, chronic coronary syndromes; TEAEs, treatment-emergent adverse events; HFpEF, heart failure with preserved ejection fraction; ABPM, ambulatory BP monitoring; HOMA-IR, homeostatic model assessment of insulin resistance; SAPH, sarcoidosis associated pulmonary hypertension; ADEs, adverse events resulting in study drug discontinuation; BFT, supine bolus flow time.

Soluble guanylate cyclase activators

While sGC stimulators target reduced and heme-containing forms of sGC, sGC activators target oxidized or heme-free sGC. Since the status of these enzymes occurs mainly in the condition of diseases accompanied by oxidative stress, this binding pattern is quite attractive for the clinical use of activator drugs (Evgenov et al., 2006). Cinaciguat, a type of amino dicarboxylic acid, is the first characteristic drug of this new sGC activator class (Stasch et al., 2002). Although other sGC activators have been identified, no sGC activators are available to patients.

Cinaciguat

A 1997 ultrahigh-throughput screening (uHTS) identified cinaciguat as an sGC activator (Sandner et al., 2021b). As a direct, NO-independent activator of sGC, cGMP levels directly increase in the presence of heightened oxidative stress and impaired endothelial function, which could yield notable efficacy. However, it increases the risk of low blood pressure (Mitrovic et al., 2011).

Cinaciguat in PAH

In a randomized, double-blind, multicenter, multinational phase IIb study (NCT00559650), cinaciguat significantly reduced pulmonary capillary wedge pressure (PCWP) and mean right atrial pressure in patients with decompensated chronic congestive heart failure. There was also a decrease in both pulmonary and systemic vascular resistance, as well as a reduction in mean arterial pressure, and the cardiac index increased (Mitrovic et al., 2011). Cinaciguat was associated with 71% of adverse events, and placebo was associated with 45%. There were no adverse events associated with 30-day mortality. When the dose was increased to 200 g/h, hypotensive events increased, and the trial was terminated (Erdmann et al., 2013).

Cinaciguat in acute HF

Three phase IIb trials, including COMPOSE 1 (NCT01065077), COMPOSE 2 (NCT01067859), and the COMPOSE Early Trial (NCT01064037), were subsequently conducted to investigate the safety and efficacy of varying doses of cinaciguat 200 µg/h *versus* placebo in treating patients with acute HF initiated at different time points. However, because hypotensive events occurred and no significant benefit was observed, the clinical development of cinaciguat was discontinued (Breitenstein et al., 2017). The clinical trials led to the discontinuation of cinaciguat. Most of the research since then has been conducted on animals (Benaldo et al., 2022; Dai and Stuehr, 2023).

Other medicines

Ataciguat

Ataciguat, formerly known as HMR 1766, is an anthranilic acid derivative that is a novel sGC activator (Schafer et al., 2010). Researchers found that ataciguat normalized vasodilation and the vascular response to exogenous NO in rats with congestive heart failure and reduced platelet activation (Schafer et al., 2010). In a rat model of inflammatory chronic renal impairment, ataciguat exhibited beneficial BP-independent effects on kidney structure and urinary albumin excretion (Benz et al., 2007). Ataciguat is currently being studied in clinical trials for numerous indications, including changes in tolerated blood pressure and orthostatic tolerance (i.e., ability to stand without passing out) in patients with mild to moderate calcified aortic stenosis (NCT02049203), effectiveness in relieving patients with neuropathic pain (NCT00799656), improving claudication in patients with PAD (NCT00443287), and slowing the progression of valve calcification in patients with moderate calcific aortic valve stenosis (NCT02481258). However, the results of these clinical trials have not yet been made publicly available.

MGV354

MGV354 is a novel sGC activator that effectively lowers IOP in glaucoma models in preclinical studies (Prasanna et al., 2018). Unfortunately, in a clinical trial (NCT02743780), MGV354 did not cause a statistically significant reduction in IOP compared to placebo (Stacy et al., 2018).

Mosliciguat

Mosliciguat (BAY 1237592) is an sGC activator designed for topical application in the lung for the treatment of PAH. Inhalation of mosliciguat specifically activates apo-sGC, leading to a selective effect in the lung (Becker-Pelster et al., 2022). Mosliciguat was shown to activate heme-free NO-GC and improve cardiopulmonary circulation in minipigs and rats (Becker-Pelster et al., 2022). Based on these results, mosliciguat is currently in phase Ib clinical development (NCT03754660) as an inhaled therapy for PAH.

Runcaciguat

As a once-daily oral sGC activator, runcaciguat demonstrated good PK distribution when administered by Bayer (Hahn et al.,

2021). Preclinical studies have demonstrated that runcaciguat may be effective in preventing CKD caused by hypertension, diabetes, and obesity (Benardeau et al., 2021). The oral sGC activator runcaciguat is currently in a phase II clinical program for patients with proteinuric CKD (NCT04507061).

BI 685509

BI 685509 is an orally bioavailable, potent sGC activator that exhibits significant renal protection properties and antifibrotic activity in preclinical models of kidney disease and injury (Reinhart et al., 2023). In addition, in a preclinical rat model of thioacetamide-induced nodularity, hepatic fibrosis, portal hypertension and portosystemic shunts, BI 685509 reduced Sirius red morphometry (SRM) by 38%, alpha-smooth muscle actin (α SMA)-positive area by 55%, portal pressure by 26% and portal shunt by 10%; hence, it could be used as a potential treatment for cirrhosis-associated portal hypertension (Jones et al., 2023). BI 685509 effectively inhibited the induction effect of activated platelet-rich plasma on the SSC-related chemokine CXCL4, more strongly than riociguat did (Nabozny et al., 2023). These findings suggest that BI 685509 is a new drug to treat SSC that is superior to riociguat. In a phase Ib study (NCT03165227), BI 685509 was generally well tolerated in patients with diabetic kidney disease (DKD) (Cherney et al., 2023). BI 685509 is currently in phase II trials for CKD (NCT04736628) and DKD (NCT04750577). Studies are currently underway on the indications of PAH and CTEPH (NCT03754660) and SSC (NCT05559580).

BI 703704 and GSK2181236A

The sGC activators BI 703704 and GSK2181236A have demonstrated sustained protection against preclinical models of CKD (Costell et al., 2012; Stasch et al., 2015; Hu et al., 2022). The results of these preclinical studies need to be confirmed in humans before these agents can be considered alternatives to current recommended treatments.

Guanylate cyclase-C agonists

GC-C is a transmembrane protein receptor that has received increasing attention for its importance in digestive diseases. It plays a key role in regulating water and electrolyte balance, maintaining gastrointestinal function, relieving abdominal pain, controlling inflammation, regulating intestinal ecology, inhibiting tumor growth and regulating cell proliferation and is considered a potential therapeutic target for digestive system diseases (Waldman and Camilleri, 2018). It was discovered in 1970 as a receptor for heat-stable endotoxins of exogenous diarrhea-causing bacteria and can be detected not only in intestinal mucosal cells but also in primary and metastatic colorectal cancer, peripheral blood, lymph nodes and liver tissues (Smith and Gyles, 1970). The signaling pathway of GC-C/cGMP has a significant impact on digestive disorders, and agonists are on the market to treat these associated gastrointestinal conditions. GC-C agonists include natural and synthetic ligands; natural ligands include endogenous ligands such as uroguanylin and guanylin, exogenous ligands such as heat-resistant enterotoxin, and synthetic ligands such as linaclotide,

plecanatide and dolcanatide (Kuhn, 2016). The current progress of GC-C agonists is summarized in Table 3.

The GC-C/cGMP signaling pathway

As shown in Figure 2, intestinal guanylin and uroguanylin are effective regulators of fluid ion homeostasis. They are secreted by various cells of the intestinal mucosa, including intestinal chromaffin cells, epithelial cells, goblet cells, and Pan's cells. These peptide hormones act as receptor GC-C ligands that produce intracellular cGMP and activate PKGII. PKGII phosphorylates the cystic fibrosis transmembrane conduction regulator (CFTR) and increases the secretion of chloride ions (Cl^-) into the intestinal lumen. cGMP can also increase the content of cyclic adenosine monophosphate (cAMP) by inhibiting the activity of PDE3. cAMP activates PKA and coactivates CFTR along with PKGII. Promoting the discharge of chloride ions and bicarbonate (Vaandrager, 2002), cGMP activates HCO_3^- secretion through an unknown mechanism. In addition, cGMP can inhibit the sodium/hydrogen exchanger NHE3, thereby reducing the absorption of sodium, preventing hyponatremia and unnecessary hypovolemic shock, and maintaining the fluid balance in the intestine. By these mechanisms, cGMP can maintain the hydration state of colon mucus and ion homeostasis (Brierley, 2012; Waldman and Camilleri, 2018; Samanta and Chaudhuri, 2021). In addition, it can regulate the intestinal immune barrier and increase the levels of IL-2 and IFN- γ and paracellular permeability (Xing et al., 2021). Abdominal pain is a major symptom of inflammatory bowel disease (IBD), and therapies modulating the GC-C/cGMP pathway promote visceral analgesia in patients with these diseases in animal models and clinical trials (Waldman and Camilleri, 2018). The mechanism of its analgesia is mainly through the afferent pathway involved in the regulation of gastrointestinal pain. After receptor-ligand binding in epithelial cells, elevated intracellular cGMP can be transported to the extracellular space through the silencing of the multidrug resistance-related protein 4 cyclic nucleoglycine efflux pump located on the basolateral membrane, reducing the excitation of submucosal afferent neurons and relieving abdominal pain. Another mechanism underlying this action is the prevention of intraluminal factors from affecting pain afferents and immune mechanisms in the lamina propria and other areas to indirectly promote analgesia (Shailubhai et al., 2015; Brierley et al., 2022). It can also inhibit the proliferation of intestinal epithelial cells and maintain genomic stability, thus inhibiting the occurrence of intestinal tumors. Following the activation of GC-C, the level of cGMP increases to activate cyclic phospho-dependent PKGII, which can inhibit the protein kinase B (Akt)-related signaling pathway and thus upregulate the expression of the tumor suppressor p53. p38 MAPK is also activated, which increases the phosphorylation of the transcription factor Sp1, leading to the accumulation of p21 in cells, which promotes cell aging and reduces the risk of cancer (Basu et al., 2014). Activation of GC-C induces apoptosis by promoting the degradation of β -catenin and opposing the pro-proliferative Wnt/ β -catenin/Tcf-4 signaling pathway (Thompson et al., 2000). Because Akt enhances β -catenin activity either directly or indirectly, the suppression of Akt signaling may also be associated with the reduction of β -catenin/T-cell factor

TABLE 3 The current progress on guanylate cyclase-C agonists.

| Drug | Trial name | Design | Population | Dose | Trial ID | Endpoint | Safety outcome | Stage of development | Conclusion | Reference |
|-------------|-------------------------|---|---------------------------------|------------------------|-------------|--|----------------------|----------------------|--|------------------------|
| Linaclotide | / | Phase III, International, Multicenter, Randomized, Double-blind, Placebo-controlled, Parallel-group Trial | Patients with IBS-C | 290 µg daily | NCT01880424 | Composite Endpoint of Abdominal Pain and IBS | AEs | Completed | Linaclotide was efficacious and well-tolerated in Chinese | Liang and Liang (2023) |
| | | | | | | | | | Patients with IBS-C, with rapid onset of effect | |
| Linaclotide | / | Phase IIIb, Randomized, Double-blind, Placebo-controlled, Parallel-group Trial | Patients with IBS-C | 290 µg daily | NCT03573908 | Change in abdominal Score | TEAEs | Completed | Linaclotide significantly reduced multiple abdominal symptoms important to patients with IBS-C | Brenner et al. (2023) |
| Linaclotide | / | Phase II Multicenter, Randomized, Double-Blind, Placebo-Controlled, Parallel Group Trial | Patients with OIC | 145 µg or 290 µg daily | NCT02270983 | change in SBMs/ week | AEs and SAEs | Completed | Linaclotide significantly improved OIC symptoms and was well tolerated in patients with chronic noncancer pain | Brenner et al. (2020) |
| Linaclotide | / | Phase I, Randomized, Placebo-Controlled Trial | Patients with Colorectal Cancer | 0.87 mg daily | NCT01950403 | difference in mean cGMP levels after 7 days | AEs | Completed | Linaclotide was associated with homeostatic signaling, including phosphorylation of vasodilator-stimulated phosphoprotein and inhibition of proliferation quantified by fewer Ki67-positive epithelial cells | Weinberg et al. (2017) |
| Linaclotide | / | Phase III Randomized, Double-blind, Placebo-controlled trial | Patients with CC | 0.5 mg daily | NCT02809105 | Change from baseline in average weekly SBM frequency | AEs , SAEs | Completed | Linaclotide 0.5 mg/day is effective and safe in Japanese CC patients | Fukudo et al. (2019) |
| Plecanatide | The CIC3 Study | Randomized, Double-Blind, Placebo-Controlled trial | Patients with CIC | 3 mg or 6 mg daily | NCT01982240 | percentage of CSBM | AEs | Completed | Plecanatide significantly improved constipation and related symptoms with a low rate of adverse events | Miner et al. (2017) |
| Plecanatide | The National CIC3 Study | Randomized, 12-Week, Double-Blind, Placebo-Controlled trial | Patients with CIC | 3 mg or 6 mg daily | NCT02122471 | Number of Durable Overall CSBM Responders | AEs , SAEs and TEAEs | Completed | Plecanatide has a positive efficacy and safety profile in CIC patients | DeMicco et al. (2017) |

IBS-C, irritable bowel syndrome with constipation; AEs, adverse events; SAEs, serious adverse events; TEAEs, treatment-emergent adverse events; OIC, opioid-induced constipation; SBMs, spontaneous bowel movements; CIC, chronic idiopathic constipation; CSBM, complete spontaneous bowel movement; CC, chronic constipation.

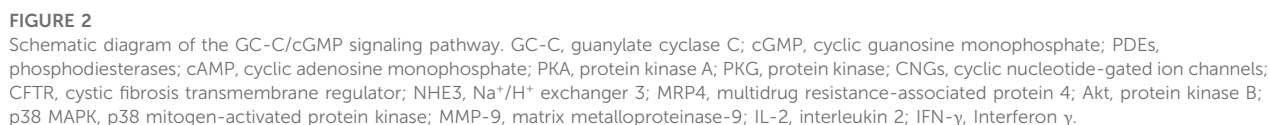


FIGURE 2
Schematic diagram of the GC-C/cGMP signaling pathway. GC-C, guanylate cyclase C; cGMP, cyclic guanosine monophosphate; PDEs, phosphodiesterases; cAMP, cyclic adenosine monophosphate; PKA, protein kinase A; PKG, protein kinase; CNGs, cyclic nucleotide-gated ion channels; CFTR, cystic fibrosis transmembrane regulator; NHE3, Na⁺/H⁺ exchanger 3; MRP4, multidrug resistance-associated protein 4; Akt, protein kinase B; p38 MAPK, p38 mitogen-activated protein kinase; MMP-9, matrix metalloproteinase-9; IL-2, interleukin 2; IFN- γ , Interferon γ .

Linaclootide

linaclotide is taken, approximately 3%–5% of the active peptide is excreted in the stool. Linaclotide and its metabolites are excreted through the kidneys (33%–45%) and biliary tract (48%–59%). Its dosage should be adjusted with particular care in patients with moderate liver impairment or mild to severe liver impairment (data are not available for patients with severe liver impairment). It is metabolized by multiple cytochrome P450 (CYP) enzymes and has no effect on major CYP subtypes, giving it a low risk of clinically relevant drug interactions (Frey et al., 2018). A therapeutic dose of linaclotide should be taken at least 30 min before meals, as effectiveness and tolerability can be affected by high-fat foods (Bassotti et al., 2018).

Linaclootide in chronic constipation syndrome

Linaclootide was evaluated at doses between 62.5 g and 600 g for efficacy and safety in patients ($n = 4,107$) with CC in a meta-analysis. More patients completed spontaneous defecation (CSBM) at different doses, which was more significant in the very low-dose group, and there were no adverse reactions in the high-dose group (Yang and Lei, 2021). Japanese researchers (NCT02809105) found that linaclootide at a dose of 0.5 mg/day was effective and safe in patients with CC, with mild and occasional adverse reactions being the most common (Fukudo et al., 2019). A prospective study from China evaluated patients taking linaclootide ($n = 97$) on bowel movements, abdominal symptoms, IBS Symptom Severity Scale (IBS-SSS), and IBS Quality of Life Questionnaire (IBS-QOL) and found a significant increase in weekly

bowel movements and a significant improvement in patients' quality of life. Diarrhea occurred in 11 cases (11.3%). IBS-C symptoms and severity were improved with linaclotide, and the drug was safe and effective (Liu et al., 2022). Another phase III clinical trial involving 659 Chinese IBS-C patients also showed that linaclotide (290 µg/day) was effective and well tolerated in Chinese IBS-C patients with rapid onset of action (NCT01880424) (Peng et al., 2022). Another study found that linaclotide reduced reaction time in patients with IBS-C (Brenner et al., 2023). In a trial in the elderly population, the most common side effect was diarrhea, but the incidence of diarrhea and constipation in elderly patients, even at reduced doses, did not differ significantly from that in nonelderly patients, and multivariate analysis showed that age, sex, and dose were not associated with diarrhea caused by linaclotide treatment. Thus, linaclotide was effective and safe in elderly patients (Chang et al., 2021; Ishigo et al., 2021). In special pediatric populations, AEs are relatively common, although studies have found that nearly half of children with FC or IBS-C benefit from linaclotide treatment, so further research is needed (Baaleman et al., 2021). In a cohort study of patients with SSc, 31 patients were treated with linaclotide. Twenty-eight of the 31 patients responded to treatment, while only three (9.7%) reported ineffective or intolerable side effects. Diarrhea, cramping, and bloating were the most commonly reported side effects (11/31, 35%). Linaclotide is a well-tolerated, effective drug that can be used to treat refractory symptoms of a low GI score on SSc (Dein et al., 2021). A multicenter phase II clinical study evaluated the efficacy and safety of linaclotide in the treatment of opioid-induced constipation (OIC) in patients with chronic noncancer pain syndrome (NCT02270983). Compared with placebo, linaclotide significantly improved stool consistency, diarrhea, bloating, and treatment satisfaction scores ($p < 0.05$). Linaclotide significantly improved OIC symptoms and was well tolerated in patients with chronic noncancerous pain (Brenner et al., 2020).

Linaclotide in visceral pain and colon cancer

Linaclotide treatment was found to reduce vaginal hyperalgesia and mechanical hyperalgesia associated with endometriosis through viscerovaginal crosstalk (Ge et al., 2019). In patients with colon cancer, a US phase I clinical trial (NCT01950403) showed that administration of linaclotide (870 µg/d) for 7 days after oral preparation of the intestine with polyethylene glycol increased the level of cGMP and reduced the proportion of Ki-67-positive colon epithelial cells (a higher proportion of Ki-67-positive cells suggested a faster rate of cell proliferation). These results suggest that linaclotide inhibits colonic epithelial cell proliferation in human colons (Weinberg et al., 2017).

Plecanatide

An oral GC-C agonist, plecanatide, consists of a 16-amino acid synthetic peptide equivalent to human uroguanylin and is used to treat gastrointestinal (GI) disorders. For the treatment of chronic idiopathic constipation (CIC) in adults, plecanatide received its first global approval in 2017 (Al-Salama and Syed, 2017; Rao, 2018). Plecanatide metabolism occurs in the gastrointestinal tract. After oral administration of 3 mg of plecanatide, blood levels of plecanatide and its active metabolites were lower than detectable levels. Standard pharmacokinetic parameters cannot be calculated, and the amount of plecanatide or its metabolites in the tissue is negligible due to the small amount of drug absorbed (Miner, 2020).

In two large randomized, double-blind, placebo-controlled studies evaluating the efficacy and safety of plecanatide (3 mg, 6 mg) vs placebo in patients with CIC, plecanatide treatment also significantly reduced the severity of other CIC symptoms (tension, consistent stools, bloating). In addition, the satisfaction and quality of life of patients treated with plecanatide improved significantly. The low incidence of adverse reactions after plecanatide treatment shows that it is safe and effective for CIC. In addition, plecanatide combined with acid suppressants is safe and effective in patients with CIC (DeMicco et al., 2017; Miner et al., 2017).

A meta-analysis evaluating the efficacy and tolerability of GC-C agonists included eight randomized controlled trials with 10,369 patients. Both drugs were effective in treating CIC, and the incidence of diarrhea was higher than that in the placebo group. Linaclotide and plecanatide were similar in efficacy and tolerability to IBS-C and CIC. There was no difference in the incidence of adverse reactions (diarrhea) between linaclotide and plecanatide (Shah et al., 2018).

Other medicines

Dolcanatide

Dolcanatide (SP-333), an oral uroguanylinoid, is replaced by selected D-amino acids to enhance stability and extend persistence, activating GUCY2C in the small and large intestines. A phase I double-blind, placebo-controlled trial (NCT03300570) of 27 mg dolcanatide administered orally daily for 7 days in healthy volunteers did not show activation of GUCY2C in distal rectal epithelial cells, as quantified by the accumulation of its product cGMP. These data suggest that the high stability of dolcanatide and its persistence along the rostral-caudal axis of the small and large intestines are insufficient to regulate GUCY2C throughout the colorectal region to prevent tumorigenesis. These results highlight the importance of developing GUCY2C anticancer agonists that target colorectal release and activity (Weinberg et al., 2021). Current research on dolcanatide focuses on its potential use for the treatment of colon cancer.

Summary

GC is widely distributed throughout the human body, mostly as sGC and GC-C, which catalyze GTP and thus increase the intracellular content of the second messenger cGMP, leading to the modulation of various intracellular physiological regulatory processes. The development of drugs on the basis of functional modulation of the GC pathway has been a fast-moving area of research over the past few years. A series of clinical studies have validated the therapeutic potential of sGC stimulators in patients with HF and PAH. Due to promising clinical trial results, riociguat and vericiguat have been approved by the US FDA for the treatment of chronic heart failure and pulmonary hypertension. In addition, the role of sGC stimulants in improving cGMP signaling might enable them to play an active role in a wide range of clinical indications, such as metabolic diseases, fibrotic diseases, urinary diseases, and neurodegenerative diseases. However, clinical studies have

shown that the benefit for dcSSc patients and patients with diabetic nephropathy is debatable, and more clinical studies are needed to support its use or nonuse in them. In contrast to sGC stimulants, the treatment potency of sGC activators is not fully clarified, although they can bind to sGC when the body is in a disease condition involving oxidative stress. The development of this class of drugs is still at the clinical study stage. In intestinal epithelial cells, GC-C is the dominant supplier of cGMP. The GC-C molecule is important for maintaining fluid and ion homeostasis in the intestinal tract, and linaclotide and plecanatide are the main drugs targeting it. Preclinical and clinical evidence shows that modulation of GC-C may improve symptoms and be tolerated by patients with IBS-C and CIC. Recent studies have shown that GC-C is also relevant to intestinal inflammation, dysbiosis and cancer, the specific mechanisms of which remain to be explored. In conclusion, cGMP has a broad spectrum of physiological effects, which gives it considerable development prospects. We systematically reviewed the transduction procedures of the NO-sGC-cGMP signaling pathway and the potential use of sGC stimulators and GC-C stimulators. Novel compounds are being developed based on the structures of sGC and GC-C, some of which are being studied in clinical trials. These clinical results may open up new therapeutic approaches for cardiovascular, renal and other diseases. Various questions remain regarding the mechanisms of their effects and the therapeutic potential of these treatments for diseases besides PAH and HF, and much more research is needed in the future.

Author contributions

QY: Writing–original draft, Writing–review and editing. XZ: Writing–original draft, Writing–review and editing. YS:

Writing–original draft. LW: Drew the figures and tables, Writing–original draft. LL: Drew the figures and tables, Writing–original draft. RT: Writing–review and editing. LH: Writing–review and editing. YB: Supervision, Writing–review and editing.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

- Ahsan, M. K., Tchernychev, B., Kessler, M. M., Solinga, R. M., Arthur, D., Linde, C. I., et al. (2017). Linaclotide activates guanylate cyclase-C/cGMP/protein kinase-II-dependent trafficking of CFTR in the intestine. *Physiol. Rep.* 5, e13299. doi:10.14814/phy2.13299
- Al-Salama, Z. T., and Syed, Y. Y. (2017). Plecanatide: first global approval. *Drugs* 77, 593–598. doi:10.1007/s40265-017-0718-0
- Archer, S. L. (2013). Riociguat for pulmonary hypertension--a glass half full. *N. Engl. J. Med.* 369, 386–388. doi:10.1056/NEJMe1306684
- Argyriou, A. I., Makrynitsa, G. I., Dalkas, G., Georgopoulou, D. A., Salagiannis, K., Vazoura, V., et al. (2021). Replacement of heme by soluble guanylate cyclase (sGC) activators abolishes heme-nitric oxide/oxygen (H-NOX) domain structural plasticity. *Curr. Res. Struct. Biol.* 3, 324–336. doi:10.1016/j.crstbi.2021.11.003
- Armstrong, P. W., Lam, C., Anstrom, K. J., Ezekowitz, J., Hernandez, A. F., O'Connor, C. M., et al. (2020a). Effect of vericiguat vs placebo on quality of life in patients with heart failure and preserved ejection fraction: the VITALITY-HFpEF randomized clinical trial. *JAMA* 324, 1512–1521. doi:10.1001/jama.2020.15922
- Armstrong, P. W., Pieske, B., Anstrom, K. J., Ezekowitz, J., Hernandez, A. F., Butler, J., et al. (2020b). Vericiguat in patients with heart failure and reduced ejection fraction. *N. Engl. J. Med.* 382, 1883–1893. doi:10.1056/NEJMoa1915928
- Baaleman, D. F., Gupta, S., Benninga, M. A., Bali, N., Vaz, K. H., Yacob, D., et al. (2021). The use of linaclotide in children with functional constipation or irritable bowel syndrome: a retrospective chart review. *Paediatr. Drugs* 23, 307–314. doi:10.1007/s40272-021-00444-4
- Barenco-Marins, T. S., Seara, F., Ponte, C. G., and Nascimento, J. (2023). Pulmonary circulation under pressure: pathophysiological and therapeutic implications of BK channel. *Cardiovasc. Drugs Ther.* 2023, 07503. doi:10.1007/s10557-023-07503-7
- Barnikel, M., Kneidinger, N., Arnold, P., Waelde, A., Behr, J., and Milger, K. (2022). Riociguat in patients with CTEPH and advanced age and/or comorbidities. *J. Clin. Med.* 11, 1084. doi:10.3390/jcm11041084
- Bassotti, G., Usai-Satta, P., and Bellini, M. (2018). Linaclotide for the treatment of chronic constipation. *Expert Opin. Pharmacother.* 19, 1261–1266. doi:10.1080/14656566.2018.1494728
- Basu, N., Saha, S., Khan, I., Ramachandra, S. G., and Visweswariah, S. S. (2014). Intestinal cell proliferation and senescence are regulated by receptor guanylyl cyclase C and p21. *J. Biol. Chem.* 289, 581–593. doi:10.1074/jbc.M113.511311
- Baughman, R. P., Shlobin, O. A., Gupta, R., Engel, P. J., Stewart, J. I., Lower, E. E., et al. (2022). Riociguat for sarcoidosis-associated pulmonary hypertension: results of a 1-year double-blind, placebo-controlled trial. *Chest* 161, 448–457. doi:10.1016/j.chest.2021.07.2162
- Becker-Pelster, E. M., Hahn, M. G., Delbeck, M., Dietz, L., Huser, J., Kopf, J., et al. (2022). Inhaled mosliciguat (BAY 1237592): targeting pulmonary vasculature via activating apo-sGC. *Respir. Res.* 23, 272. doi:10.1186/s12931-022-02189-1
- Belenkov, Y. N., and Kozhevnikova, M. V. (2023). Soluble guanylate cyclase: restoration of the NO-sGC-cGMP signaling pathway activity. A new opportunity in the treatment of heart failure. *Kardiologiia* 63, 68–76. doi:10.18087/cardio.2023.5.n2422
- Benaldo, F. A., Araya-Quijada, C., Ebersperger, G., Herrera, E. A., Reyes, R. V., Moraga, F. A., et al. (2022). Cinaciguat (BAY-582667) modifies cardiopulmonary and systemic circulation in chronically hypoxic and pulmonary hypertensive neonatal lambs in the alto andino. *Front. Physiol.* 13, 864010. doi:10.3389/fphys.2022.864010
- Benardeau, A., Kahnert, A., Schomber, T., Meyer, J., Pavkovic, M., Kretschmer, A., et al. (2021). Runcaciguat, a novel soluble guanylate cyclase activator, shows

renoprotection in hypertensive, diabetic, and metabolic preclinical models of chronic kidney disease. *Naunyn Schmiedeb. Arch. Pharmacol.* 394, 2363–2379. doi:10.1007/s00210-021-02149-4

Benkner, A., Rudebusch, J., Nath, N., Hammer, E., Grube, K., Gross, S., et al. (2022). Riociguat attenuates the changes in left ventricular proteome and microRNA profile after experimental aortic stenosis in mice. *Br. J. Pharmacol.* 179, 4575–4592. doi:10.1111/bph.15910

Benz, K., Orth, S. R., Simonaviciene, A., Linz, W., Schindler, U., Rutten, H., et al. (2007). Blood pressure-independent effect of long-term treatment with the soluble heme-independent guanylyl cyclase activator HMR1766 on progression in a model of noninflammatory chronic renal damage. *Kidney Blood Press Res.* 30, 224–233. doi:10.1159/000104091

Benza, R., Corris, P., Ghofrani, A., Kanwar, M. K., McLaughlin, V. V., Raina, A., et al. (2019). EXPRESS: switching to riociguat: a potential treatment strategy for the management of CTEPH and PAH. *Pulm. Circ.* 10, 2045894019837849. doi:10.1177/2045894019837849

Biswas, S., Kojonazarov, B., Hadzic, S., Majer, M., Bajraktari, G., Novoyatleva, T., et al. (2020). IRAG1 deficient mice develop PKG1 β dependent pulmonary hypertension. *Cells* 9, 2280. doi:10.3390/cells9102280

Boettcher, M., Dungen, H. D., Donath, F., Mikus, G., Werner, N., Thuermann, P. A., et al. (2022). Vericiguat in combination with short-acting nitroglycerin in patients with chronic coronary syndromes: the randomized, phase Ib, VENICE study. *Clin. Pharmacol. Ther.* 111, 1239–1247. doi:10.1002/cpt.2574

Boettcher, M., Thomas, D., Mueck, W., Loewen, S., Arens, E., Yoshikawa, K., et al. (2021). Safety, pharmacodynamic, and pharmacokinetic characterization of vericiguat: results from six phase I studies in healthy subjects. *Eur. J. Clin. Pharmacol.* 77, 527–537. doi:10.1007/s00228-020-03023-7

Breitenstein, S., Roessig, L., Sandner, P., and Lewis, K. S. (2017). Novel sGC stimulators and sGC activators for the treatment of heart failure. *Handb. Exp. Pharmacol.* 243, 225–247. doi:10.1007/164_2016_100

Brenner, D. M., Argoff, C. E., Fox, S. M., Bochenek, W., D'Astoli, P., Blakesley, R. E., et al. (2020). Efficacy and safety of linacotide for opioid-induced constipation in patients with chronic noncancer pain syndromes from a phase 2 randomized study. *Pain* 161, 1027–1036. doi:10.1097/j.pain.0000000000001754

Brenner, D. M., Lacy, B. E., Ford, A. C., Bartolini, W., Wu, J., Shea, E. P., et al. (2023). Linacotide reduced response time for irritable bowel syndrome with constipation symptoms: analysis of 4 randomized controlled trials. *Am. J. Gastroenterol.* 118, 872–879. doi:10.14309/ajg.00000000000002064

Brierley, S. M. (2012). Guanylate cyclase-C receptor activation: unexpected biology. *Curr. Opin. Pharmacol.* 12, 632–640. doi:10.1016/j.coph.2012.10.005

Brierley, S. M., Grundy, L., Castro, J., Harrington, A. M., Hannig, G., and Camilleri, M. (2022). Guanylate cyclase-C agonists as peripherally acting treatments of chronic visceral pain. *Trends Pharmacol. Sci.* 43, 110–122. doi:10.1016/j.tips.2021.11.002

Brockunier, L., Stelmach, J., Guo, J., Spencer, T., Rosauer, K., Bansal, A., et al. (2020). Soluble guanylate cyclase stimulators for the treatment of hypertension: discovery of MK-2947. *Bioorg Med. Chem. Lett.* 30, 127574. doi:10.1016/j.bmcl.2020.127574

Campbell, N., Kalabak-Hoganson, J., and Frey, K. (2022). Vericiguat: a novel oral soluble guanylate cyclase stimulator for the treatment of heart failure. *Ann. Pharmacother.* 56, 600–608. doi:10.1177/10600280211041384

Chang, L., Lacy, B. E., Moshiree, B., Kassebaum, A., Abel, J. L., Hanlon, J., et al. (2021). Efficacy of linacotide in reducing abdominal symptoms of bloating, discomfort, and pain: a phase 3B trial using a novel abdominal scoring system. *Am. J. Gastroenterol.* 116, 1929–1937. doi:10.14309/ajg.0000000000001334

Chen, L., Daum, G., Chitaley, K., Coats, S. A., Bowen-Pope, D. F., Eigenthaler, M., et al. (2004). Vasodilator-stimulated phosphoprotein regulates proliferation and growth inhibition by nitric oxide in vascular smooth muscle cells. *Arterioscler. Thromb. Vasc. Biol.* 24, 1403–1408. doi:10.1161/01.ATV.0000134705.39654.53

Cherney, D., de Zeeuw, D., Heerspink, H., Cardona, J., Desch, M., Wenz, A., et al. (2023). Safety, tolerability, pharmacodynamics and pharmacokinetics of the soluble guanylyl cyclase activator BI 685509 in patients with diabetic kidney disease: a randomized trial. *Diabetes Obes. Metab.* 25, 2218–2226. doi:10.1111/dom.15099

Chiles, R., and Al-Horani, R. A. (2022). Vericiguat: a new hope for heart failure patients. *Cardiovasc Ther.* 2022, 1554875. doi:10.1155/2022/1554875

Christou, H., and Khalil, R. A. (2022). Mechanisms of pulmonary vascular dysfunction in pulmonary hypertension and implications for novel therapies. *Am. J. Physiol. Heart Circ. Physiol.* 322, H702–H724. doi:10.1152/ajpheart.00021.2022

Costell, M. H., Ancellin, N., Bernard, R. E., Zhao, S., Upson, J. J., Morgan, L. A., et al. (2012). Comparison of soluble guanylate cyclase stimulators and activators in models of cardiovascular disease associated with oxidative stress. *Front. Pharmacol.* 3, 128. doi:10.3389/fphar.2012.00128

Dachs, T. M., Duca, F., Rettl, R., Binder-Rodriguez, C., Dalos, D., Ligios, L. C., et al. (2022). Riociguat in pulmonary hypertension and heart failure with preserved ejection fraction: the haemoDYNAMIC trial. *Eur. Heart J.* 43, 3402–3413. doi:10.1093/eurheartj/ehac389

Dai, Y., and Stuehr, D. J. (2022). Inactivation of soluble guanylyl cyclase in living cells proceeds without loss of haem and involves heterodimer dissociation as a common step. *Br. J. Pharmacol.* 179, 2505–2518. doi:10.1111/bph.15527

Dai, Y., and Stuehr, D. J. (2023). BAY58-2667 activates different soluble guanylyl cyclase species by distinct mechanisms that indicate its principal target in cells is the heme-free soluble guanylyl cyclase-heat shock protein 90 complex. *Mol. Pharmacol.* 103, 286–296. doi:10.1124/molpharm.122.000624

Dasgupta, A., Bowman, L., D'Arigney, C. L., and Archer, S. L. (2015). Soluble guanylate cyclase: a new therapeutic target for pulmonary arterial hypertension and chronic thromboembolic pulmonary hypertension. *Clin. Pharmacol. Ther.* 97, 88–102. doi:10.1002/cpt.10

Dein, E. J., Wigley, F. M., and McMahan, Z. H. (2021). Linacotide for the treatment of refractory lower bowel manifestations of systemic sclerosis. *BMC Gastroenterol.* 21, 174. doi:10.1186/s12876-021-01738-0

DeMicco, M., Barrow, L., Hickey, B., Shailubhai, K., and Griffin, P. (2017). Randomized clinical trial: efficacy and safety of plectanatin in the treatment of chronic idiopathic constipation. *Ther. Adv. Gastroenterol.* 10, 837–851. doi:10.1177/1756283X17734697

Derbyshire, E. R., and Marletta, M. A. (2009). Biochemistry of soluble guanylate cyclase. *Handb. Exp. Pharmacol.* 2009, 17–31. doi:10.1007/978-3-540-68964-5_2

Dhahri, W., Dussault, S., Raguema, N., Desjarlais, M., and Rivard, A. (2023). Stimulation of soluble guanylate cyclase activity with riociguat promotes angiogenesis and improves neovascularization after limb ischemia. *Atherosclerosis* 372, 32–40. doi:10.1016/j.atherosclerosis.2023.03.017

Distler, J., Gyorfi, A. H., Ramanujam, M., Whitfield, M. L., Konigshoff, M., and Lafyatis, R. (2019). Shared and distinct mechanisms of fibrosis. *Nat. Rev. Rheumatol.* 15, 705–730. doi:10.1038/s41584-019-0322-7

Donaldson, S., Ogunti, R., Kibreab, A., and Mehari, A. (2020). Riociguat in the treatment of chronic thromboembolic pulmonary hypertension: an evidence-based review of its place in therapy. *Core Evid.* 15, 31–40. doi:10.2147/CE.S172791

Eggermont, J. A., Vrolix, M., Wuytack, F., Raeymaekers, L., and Casteels, R. (1988). The (Ca²⁺-Mg²⁺)-ATPases of the plasma membrane and of the endoplasmic reticulum in smooth muscle cells and their regulation. *J. Cardiovasc Pharmacol.* 12 (5), S51–S55. doi:10.1097/00005344-198800125-00010

Englert, N., Burkard, P., Aue, A., Rosenwald, A., Nieswandt, B., and Friebe, A. (2023). Anti-fibrotic and anti-inflammatory role of NO-sensitive guanylyl cyclase in murine lung. *Int. J. Mol. Sci.* 24, 11661. doi:10.3390/ijms241411661

Erdmann, E., Semigran, M. J., Nieminen, M. S., Gheorghiad, M., Agrawal, R., Mitrovic, V., et al. (2013). Cinaciguat, a soluble guanylate cyclase activator, unloads the heart but also causes hypotension in acute decompensated heart failure. *Eur. Heart J.* 34, 57–67. doi:10.1093/eurheartj/ehs196

Evgenov, O. V., Pacher, P., Schmidt, P. M., Hasko, G., Schmidt, H. H., and Stasch, J. P. (2006). NO-independent stimulators and activators of soluble guanylate cyclase: discovery and therapeutic potential. *Nat. Rev. Drug Discov.* 5, 755–768. doi:10.1038/nrd2038

Evora, P. R., Evora, P. M., Celotto, A. C., Rodrigues, A. J., and Joviliano, E. E. (2012). Cardiovascular therapeutics targets on the NO-sGC-cGMP signaling pathway: a critical overview. *Curr. Drug Targets* 13, 1207–1214. doi:10.2174/138945012802002348

Fang, D., Hawke, D., Zheng, Y., Xia, Y., Meisenhelder, J., Nika, H., et al. (2007). Phosphorylation of beta-catenin by AKT promotes beta-catenin transcriptional activity. *J. Biol. Chem.* 282, 11221–11229. doi:10.1074/jbc.M611871200

Feil, R., Lehnert, M., Stehle, D., and Feil, S. (2022). Visualising and understanding cGMP signals in the cardiovascular system. *Br. J. Pharmacol.* 179, 2394–2412. doi:10.1111/bph.15500

Foussard, N., Rouault, P., Cornuault, L., Reynaud, A., Buys, E. S., Chapouly, C., et al. (2023). Praliciguat promotes ischemic leg reperfusion in leptin receptor-deficient mice. *Circ. Res.* 132, 34–48. doi:10.1161/CIRCRESAHA.122.322033

Frey, R., Becker, C., Saleh, S., Unger, S., van der Mey, D., and Muck, W. (2018). Clinical pharmacokinetic and pharmacodynamic profile of riociguat. *Clin. Pharmacokinet.* 57, 647–661. doi:10.1007/s40262-017-0604-7

Fukudo, S., Miwa, H., Nakajima, A., Kinoshita, Y., Kosako, M., Hayashi, K., et al. (2019). High-dose linacotide is effective and safe in patients with chronic constipation: a phase III randomized, double-blind, placebo-controlled study with a long-term open-label extension study in Japan. *Neurogastroenterol. Motil.* 31, e13487. doi:10.1111/nmo.13487

Gambaryan, S. (2022). The role of NO/sGC/cGMP/PKG signaling pathway in regulation of platelet function. *Cells* 11, 3704. doi:10.3390/cells11223704

Garbers, D. L. (1990). Guanylate cyclase receptor family. *Recent Prog. Horm. Res.* 46, 85–96. doi:10.1016/b978-0-12-571146-3.50008-0

Garcia, A. H., Gorenflo, M., Ivy, D. D., Moledina, S., Castaldi, B., Ishida, H., et al. (2022). Riociguat in children with pulmonary arterial hypertension: the PATENT-CHILD study. *Pulm. Circ.* 12, e12133. doi:10.1002/pul2.12133

Ge, P., Ren, J., Harrington, A. M., Grundy, L., Castro, J., Brierley, S. M., et al. (2019). Linacotide treatment reduces endometriosis-associated vaginal hyperalgesia and mechanical allodynia through viscerovisceral cross-talk. *Pain* 160, 2566–2579. doi:10.1097/j.pain.0000000000001657

Gheorghiad, M., Greene, S. J., Butler, J., Filippatos, G., Lam, C. S., Maggioni, A. P., et al. (2015). Effect of vericiguat, a soluble guanylate cyclase stimulator, on natriuretic

peptide levels in patients with worsening chronic heart failure and reduced ejection fraction: the SOCRATES-REDUCED randomized trial. *JAMA* 314, 2251–2262. doi:10.1001/jama.2015.15734

Gheorghiadu, M., Greene, S. J., Filippatos, G., Erdmann, E., Ferrari, R., Levy, P. D., et al. (2012). Cinaciguat, a soluble guanylate cyclase activator: results from the randomized, controlled, phase IIb COMPOSE programme in acute heart failure syndromes. *Eur. J. Heart Fail* 14, 1056–1066. doi:10.1093/eurjhf/hfs093

Grzesek, G., and Nowaczky, A. (2021). Current modulation of guanylate cyclase pathway activity-mechanism and clinical implications. *Molecules* 26, 3418. doi:10.3390/molecules26113418

Grzesek, G., Witczynska, A., Weglarz, M., Wolowicz, L., Nowaczky, J., Grzesek, E., et al. (2023). Soluble guanylyl cyclase activators-promising therapeutic option in the pharmacotherapy of heart failure and pulmonary hypertension. *Molecules* 28, 861. doi:10.3390/molecules28020861

Hahn, M. G., Lampe, T., El, S. S., Griebenow, N., Woltering, E., Schlemmer, K. H., et al. (2021). Discovery of the soluble guanylate cyclase activator runcaciguat (BAY 1101042). *J. Med. Chem.* 64, 5323–5344. doi:10.1021/acs.jmedchem.0c02154

Hanrahan, J. P., de Boer, I. H., Bakris, G. L., Wilson, P. J., Wakefield, J. D., Seferovic, J. P., et al. (2020a). Effects of the soluble guanylate cyclase stimulator pralicigat in diabetic kidney disease: a randomized placebo-controlled clinical trial. *Clin. J. Am. Soc. Nephrol.* 16, 59–69. doi:10.2215/CJN.08410520

Hanrahan, J. P., Seferovic, J. P., Wakefield, J. D., Wilson, P. J., Chickering, J. G., Jung, J., et al. (2020b). An exploratory, randomised, placebo-controlled, 14 day trial of the soluble guanylate cyclase stimulator pralicigat in participants with type 2 diabetes and hypertension. *Diabetologia* 63, 733–743. doi:10.1007/s00125-019-05062-x

Hanrahan, J. P., Wakefield, J. D., Wilson, P. J., Mihova, M., Chickering, J. G., Ruff, D., et al. (2019). A randomized, placebo-controlled, multiple-ascending-dose study to assess the safety, tolerability, pharmacokinetics, and pharmacodynamics of the soluble guanylate cyclase stimulator pralicigat in healthy subjects. *Clin. Pharmacol. Drug Dev.* 8, 564–575. doi:10.1002/cpdd.627

Hasegawa, M., and Shimonishi, Y. (2005). Recognition and signal transduction mechanism of *Escherichia coli* heat-stable enterotoxin and its receptor, guanylate cyclase C. *J. Pept. Res.* 65, 261–271. doi:10.1111/j.1399-3011.2005.00218.x

Hu, L., Chen, Y., Zhou, X., Hoek, M., Cox, J., Lin, K., et al. (2022). Effects of soluble guanylate cyclase stimulator on renal function in ZSF-1 model of diabetic nephropathy. *PLoS One* 17, e0261000. doi:10.1371/journal.pone.0261000

Hu, L., Wang, Z., Yi, R., Yi, H., Xiao, S., Chen, Z., et al. (2017). Soluble guanylate cyclase: a new therapeutic target for fibrotic diseases. *Curr. Med. Chem.* 24, 3203–3215. doi:10.2174/0929867324666170509115433

Hulot, J. S., Trochu, J. N., Donal, E., Galinier, M., Logeart, D., De Groote, P., et al. (2021). Vericiguat for the treatment of heart failure: mechanism of action and pharmacological properties compared with other emerging therapeutic options. *Expert Opin. Pharmacother.* 22, 1847–1855. doi:10.1080/14656566.2021.1937121

Isigoh, T., Shimotsubo, T., Takada, R., Nakano, K., Fujii, S., Kitagawa, M., et al. (2021). Efficacy and safety of linacotide in elderly patients. *Yakugaku Zasshi* 141, 255–262. doi:10.1248/yakushi.20-00176

Jais, X., Brenot, P., Bouvaist, H., Jevnikar, M., Canuet, M., Chabanne, C., et al. (2022). Balloon pulmonary angioplasty versus riociguat for the treatment of inoperable chronic thromboembolic pulmonary hypertension (RACE): a multicentre, phase 3, open-label, randomised controlled trial and ancillary follow-up study. *Lancet Respir. Med.* 10, 961–971. doi:10.1016/S2213-2600(22)00214-4

Jiao, T., Collado, A., Mahdi, A., Tengbom, J., Tratsiakovich, Y., Milne, G. T., et al. (2023). Stimulation of erythrocyte soluble guanylyl cyclase induces cGMP export and cardioprotection in type 2 diabetes. *JACC Basic Transl. Sci.* 8, 907–918. doi:10.1016/j.jacmts.2023.02.017

Jones, A. K., Chen, H., Ng, K. J., Villalona, J., McHugh, M., Zeveleva, S., et al. (2023). Soluble guanylyl cyclase activator BI 685509 reduces portal hypertension and portosystemic shunting in a rat thioacetamide-induced cirrhosis model. *J. Pharmacol. Exp. Ther.* 386, 70–79. doi:10.1124/jpet.122.001532

Kang, Y., Liu, R., Wu, J. X., and Chen, L. (2019). Structural insights into the mechanism of human soluble guanylate cyclase. *Nature* 574, 206–210. doi:10.1038/s41586-019-1584-6

Kazerounian, S., Pitari, G. M., Shah, F. J., Frick, G. S., Madesh, M., Ruiz-Stewart, I., et al. (2005). Proliferative signaling by store-operated calcium channels opposes colon cancer cell cytoskeleton induced by bacterial enterotoxins. *J. Pharmacol. Exp. Ther.* 314, 1013–1022. doi:10.1124/jpet.105.089052

Kenny, M., Clarke, M. M., and Pogue, K. T. (2022). Overview of riociguat and its role in the treatment of pulmonary hypertension. *J. Pharm. Pract.* 35, 437–444. doi:10.1177/0897190020961291

Khanna, D., Allanore, Y., Denton, C. P., Kuwana, M., Matucci-Cerinic, M., Pope, J. E., et al. (2020). Riociguat in patients with early diffuse cutaneous systemic sclerosis (RISE-SSc): randomised, double-blind, placebo-controlled multicentre trial. *Ann. Rheum. Dis.* 79, 618–625. doi:10.1136/annrheumdis-2019-216823

Kinoshita, H., Kuwahara, K., Nishida, M., Jian, Z., Rong, X., Kiyonaka, S., et al. (2010). Inhibition of TRPC6 channel activity contributes to the antihypertrophic effects of

natriuretic peptides-guanylyl cyclase-A signaling in the heart. *Circ. Res.* 106, 1849–1860. doi:10.1161/CIRCRESAHA.109.208314

Klaiber, M., Kruse, M., Volker, K., Schroter, J., Feil, R., Freichel, M., et al. (2010). Novel insights into the mechanisms mediating the local antihypertrophic effects of cardiac atrial natriuretic peptide: role of cGMP-dependent protein kinase and RGS2. *Basic Res. Cardiol.* 105, 583–595. doi:10.1007/s00395-010-0098-z

Klinger, J. R., and Kadowitz, P. J. (2017). The nitric oxide pathway in pulmonary vascular disease. *Am. J. Cardiol.* 120, S71–9. doi:10.1016/j.amjcard.2017.06.012

Krishnan, S. M., Kraehling, J. R., Eitner, F., Benardeau, A., and Sandner, P. (2018). The impact of the nitric oxide (NO)/Soluble guanylyl cyclase (sGC) signaling cascade on kidney Health and disease: a preclinical perspective. *Int. J. Mol. Sci.* 19, 1712. doi:10.3390/ijms19061712

Kruger, M., Kotter, S., Grutzner, A., Lang, P., Andresen, C., Redfield, M. M., et al. (2009). Protein kinase G modulates human myocardial passive stiffness by phosphorylation of the titin springs. *Circ. Res.* 104, 87–94. doi:10.1161/CIRCRESAHA.108.184408

Kuhn, M. (2016). Molecular physiology of membrane guanylyl cyclase receptors. *Physiol. Rev.* 96, 751–804. doi:10.1152/physrev.00022.2015

Liang, W. L., and Liang, B. (2023). Soluble guanylate cyclase activators and stimulators in patients with heart failure. *Curr. Cardiol. Rep.* 25, 607–613. doi:10.1007/s11886-023-01884-9

Liu, G., Shea, C. M., Jones, J. E., Price, G. M., Warren, W., Lonie, E., et al. (2020). Pralicigat inhibits progression of diabetic nephropathy in ZSF1 rats and suppresses inflammation and apoptosis in human renal proximal tubular cells. *Am. J. Physiol. Ren. Physiol.* 319, F697–F711. doi:10.1152/ajprenal.00003.2020

Liu, L., Zhang, W., Zhao, W., Guo, S., Wang, Y., Lv, X., et al. (2022). Linacotide for treating patients with irritable bowel syndrome with predominant constipation: a multicentre study of real-world data in China. *Ther. Adv. Gastroenterol.* 15, 17562848221092596. doi:10.1177/17562848221092596

Loretah, C., Baktiar, K., Hong, Z., and Stephen, N. J. (2021). Mdm2 phosphorylation by Akt regulates the p53 response to oxidative stress to promote cell proliferation and tumorigenesis. *Proc. Natl. Acad. Sci.* 118, 118. doi:10.1073/pnas.2003193118

Love, B. L., Johnson, A., and Smith, L. S. (2014). Linacotide: a novel agent for chronic constipation and irritable bowel syndrome. *Am. J. Health Syst. Pharm.* 71, 1081–1091. doi:10.2146/ajhp130575

Lubbe, W. J., Zuzga, D. S., Zhou, Z., Fu, W., Pelta-Heller, J., Muschel, R. J., et al. (2009). Guanylyl cyclase C prevents colon cancer metastasis by regulating tumor epithelial cell matrix metalloproteinase-9. *Cancer Res.* 69, 3529–3536. doi:10.1158/0008-5472.CAN-09-0067

Markham, A., and Duggan, S. (2021). Vericiguat: first approval. *Drugs* 81, 721–726. doi:10.1007/s40265-021-01496-z

Mihalek, A. D., Scott, C. D., and Mazimba, S. (2022). Evaluating riociguat in the treatment of pulmonary arterial hypertension: a real-world perspective. *Vasc. Health Risk Manag.* 18, 823–832. doi:10.2147/VHRM.S383572

Miner, P. B. (2020). Plecanatide for the treatment of constipation-predominant irritable bowel syndrome. *Expert Rev. Gastroenterol. Hepatol.* 14, 71–84. doi:10.1080/17474124.2020.1722101

Miner, P. J., Koltun, W. D., Wiener, G. J., De La Portilla, M., Prieto, B., Shailubhai, K., et al. (2017). A randomized phase III clinical trial of plecanatide, a uroguanylin analog, in patients with chronic idiopathic constipation. *Am. J. Gastroenterol.* 112, 613–621. doi:10.1038/ajg.2016.611

Mitrovic, V., Jovanovic, A., and Lehinant, S. (2011). Soluble guanylate cyclase modulators in heart failure. *Curr. Heart Fail Rep.* 8, 38–44. doi:10.1007/s11897-010-0045-1

Nabozny, G., Wang, C., Daley, L., Ebenezer, D., Delic, D., Bretschneider, T., et al. (2023). POS0620 BI 685509: a potent activator of soluble guanylate cyclase (sGC) as a novel treatment of vasculopathy and fibrosis in systemic sclerosis (SSc). *Ann. Rheumatic Dis.* 82 (1), S583–S584. doi:10.1136/annrheumdis-2023-eular.1599

Nathan, S. D., Behr, J., Collard, H. R., Cottin, V., Hoepfer, M. M., Martinez, F. J., et al. (2019). Riociguat for idiopathic interstitial pneumonia-associated pulmonary hypertension (RISE-IIP): a randomised, placebo-controlled phase 2b study. *Lancet Respir. Med.* 7, 780–790. doi:10.1016/S2213-2600(19)30250-4

Nowaczky, A., Kowalska, M., Nowaczky, J., and Grzesek, G. (2021). Carbon monoxide and nitric oxide as examples of the youngest class of transmitters. *Int. J. Mol. Sci.* 22, 6029. doi:10.3390/ijms22116029

Numata, G., and Takimoto, E. (2022). Cyclic GMP and PKG signaling in heart failure. *Front. Pharmacol.* 13, 792798. doi:10.3389/fphar.2022.792798

Peng, L. H., Fang, J. Y., Dai, N., Shen, X. Z., Yang, Y. L., Sun, J., et al. (2022). Efficacy and safety of linacotide in patients with irritable bowel syndrome with constipation: Chinese sub-cohort analysis of a phase III, randomised, double-blind, placebo-controlled trial. *J. Dig. Dis.* 23, 99–110. doi:10.1111/1751-2980.13081

Pieske, B., Maggioni, A. P., Lam, C., Pieske-Kraigher, E., Filippatos, G., Butler, J., et al. (2017). Vericiguat in patients with worsening chronic heart failure and preserved ejection fraction: results of the SOLuble guanylate Cyclase stimulator in heart failure

- patientS with PRESERVED EF (SOCRATES-PRESERVED) study. *Eur. Heart J.* 38, 1119–1127. doi:10.1093/eurheartj/ehw593
- Prasanna, G., Ferrara, L., Adams, C., Ehara, T., Li, B., Yang, L., et al. (2018). A novel selective soluble guanylate cyclase activator, MGV354, lowers intraocular pressure in preclinical models, following topical ocular dosing. *Invest. Ophthalmol. Vis. Sci.* 59, 1704–1716. doi:10.1167/jovs.18-23772
- Rahaghi, F. F., Trivieri, M. G., and Sahay, S. (2023). The role of riociguat in combination therapies for pulmonary arterial hypertension. *Respir. Med.* 211, 107196. doi:10.1016/j.rmed.2023.107196
- Rao, S. (2018). Plecanatide: a new guanylate cyclase agonist for the treatment of chronic idiopathic constipation. *Ther. Adv. Gastroenterol.* 11, 1756284818777945. doi:10.1177/1756284818777945
- Rappaport, J. A., and Waldman, S. A. (2020). An update on guanylyl cyclase C in the diagnosis, chemoprevention, and treatment of colorectal cancer. *Expert Rev. Clin. Pharmacol.* 13, 1125–1137. doi:10.1080/17512433.2020.1826304
- Reiberger, T., Berzigotti, A., Trebicka, J., Ertle, J., Gashaw, I., Swallow, R., et al. (2023). The rationale and study design of two phase II trials examining the effects of BI 685509, a soluble guanylyl cyclase activator, on clinically significant portal hypertension in patients with compensated cirrhosis. *Trials* 24, 293. doi:10.1186/s13063-023-07291-3
- Reinhart, G. A., Harrison, P. C., Lincoln, K., Chen, H., Sun, P., Hill, J., et al. (2023). The novel, clinical-stage soluble guanylate cyclase activator BI 685509 protects from disease progression in models of renal injury and disease. *J. Pharmacol. Exp. Ther.* 384, 382–392. doi:10.1124/jpet.122.001423
- Rennie, G. R., Barden, T. C., Bernier, S. G., Carvalho, A., Deming, R., Germano, P., et al. (2021). Discovery of CYR715: a novel carboxylic acid-containing soluble guanylate cyclase stimulator. *Bioorg Med. Chem. Lett.* 40, 127886. doi:10.1016/j.bmcl.2021.127886
- Romano, E., Rosa, I., Fioretto, B. S., Giuggioli, D., Manetti, M., and Matucci-Cerinic, M. (2023). Soluble guanylate cyclase stimulation fosters angiogenesis and blunts myofibroblast-like features of systemic sclerosis endothelial cells. *Rheumatol. Oxf.* 62, 1125–1137. doi:10.1093/rheumatology/keac433
- Rudebusch, J., Benkner, A., Nath, N., Fleuch, L., Kaderali, L., Grube, K., et al. (2022). Stimulation of soluble guanylyl cyclase (sGC) by riociguat attenuates heart failure and pathological cardiac remodelling. *Br. J. Pharmacol.* 179, 2430–2442. doi:10.1111/bph.15333
- Sadek, M. S., Cachorro, E., El-Armouche, A., and Kammerer, S. (2020). Therapeutic implications for PDE2 and cGMP/cAMP mediated crosstalk in cardiovascular diseases. *Int. J. Mol. Sci.* 21, 7462. doi:10.3390/ijms21207462
- Samanta, S., and Chaudhuri, A. G. (2021). Guanylin and uroguanylin: a promising nexus in intestinal electrolyte and fluid homeostasis. *J. Physiol. Pharmacol.* 72, 72. doi:10.26402/jpp.2021.5.02
- Sandner, P. (2018). From molecules to patients: exploring the therapeutic role of soluble guanylate cyclase stimulators. *Biol. Chem.* 399, 679–690. doi:10.1515/hsz-2018-0155
- Sandner, P., Becker-Pelster, E. M., and Stasch, J. P. (2018). Discovery and development of sGC stimulators for the treatment of pulmonary hypertension and rare diseases. *Nitric Oxide* 77, 88–95. doi:10.1016/j.niox.2018.05.001
- Sandner, P., Follmann, M., Becker-Pelster, E., Hahn, M. G., Meier, C., Freitas, C., et al. (2021a). Soluble GC stimulators and activators: past, present and future. *Br. J. Pharmacol.* 2021. doi:10.1111/bph.15698
- Sandner, P., and Stasch, J. P. (2017). Anti-fibrotic effects of soluble guanylate cyclase stimulators and activators: a review of the preclinical evidence. *Respir. Med.* 122 (1), S1–S9. doi:10.1016/j.rmed.2016.08.022
- Sandner, P., Zimmer, D. P., Milne, G. T., Follmann, M., Hobbs, A., and Stasch, J. P. (2021b). Soluble guanylate cyclase stimulators and activators. *Handb. Exp. Pharmacol.* 264, 355–394. doi:10.1007/164_2018_197
- Sarathi, J. B., Trumbull, A. M., Abazari, S. M., van Unen, V., Chan, J. E., Joo, N. S., et al. (2023). Critical role of down-regulated in adenoma bicarbonate transporter in linacotide stimulated intestinal bicarbonate secretion. *bioRxiv*. Available at: <https://doi.org/10.1101/2023.05.05.539132>.
- Schafer, A., Fraccarollo, D., Werner, L., and Bauersachs, J. (2010). Guanylyl cyclase activator ataciguat improves vascular function and reduces platelet activation in heart failure. *Pharmacol. Res.* 62, 432–438. doi:10.1016/j.phrs.2010.06.008
- Schmidt, P., Schramm, M., Schroder, H., and Stasch, J. P. (2003). Mechanisms of nitric oxide independent activation of soluble guanylyl cyclase. *Eur. J. Pharmacol.* 468, 167–174. doi:10.1016/s0014-2999(03)01674-1
- Schwartzkopf, C. D., Hadcock, J. R., Liu, G., Germano, P., Roux, J., Shea, C. M., et al. (2022). Beneficial metabolic effects of praliciguat, a soluble guanylate cyclase stimulator, in a mouse diet-induced obesity model. *Front. Pharmacol.* 13, 852080. doi:10.3389/fphar.2022.852080
- Shah, E. D., Kim, H. M., and Schoenfeld, P. (2018). Efficacy and tolerability of guanylate cyclase-C agonists for irritable bowel syndrome with constipation and chronic idiopathic constipation: a systematic review and meta-analysis. *Am. J. Gastroenterol.* 113, 329–338. doi:10.1038/ajg.2017.495
- Shailubhai, K., Palejwala, V., Arjunan, K. P., Saykhedkar, S., Nefsky, B., Foss, J. A., et al. (2015). Plecanatide and dolcanatide, novel guanylate cyclase-C agonists, ameliorate gastrointestinal inflammation in experimental models of murine colitis. *World J. Gastrointest. Pharmacol. Ther.* 6, 213–222. doi:10.4292/wjgpt.v6.i4.213
- Singh, A., Laribi, S., Teerlink, J. R., and Mebazaa, A. (2017). Agents with vasodilator properties in acute heart failure. *Eur. Heart J.* 38, 317–325. doi:10.1093/eurheartj/ehv755
- Smith, H. W., and Gyles, C. L. (1970). The relationship between two apparently different enterotoxins produced by enteropathogenic strains of *Escherichia coli* of porcine origin. *J. Med. Microbiol.* 3, 387–401. doi:10.1099/00222615-3-3-387
- Stacy, R., Huttner, K., Watts, J., Peace, J., Wirta, D., Walters, T., et al. (2018). A randomized, controlled phase I/II study to evaluate the safety and efficacy of MGV354 for ocular hypertension or glaucoma. *Am. J. Ophthalmol.* 192, 113–123. doi:10.1016/j.ajo.2018.05.015
- Stasch, J. P., Pacher, P., and Evgenov, O. V. (2011). Soluble guanylate cyclase as an emerging therapeutic target in cardiopulmonary disease. *Circulation* 123, 2263–2273. doi:10.1161/CIRCULATIONAHA.110.981738
- Stasch, J. P., Schlossmann, J., and Hoher, B. (2015). Renal effects of soluble guanylate cyclase stimulators and activators: a review of the preclinical evidence. *Curr. Opin. Pharmacol.* 21, 95–104. doi:10.1016/j.coph.2014.12.014
- Stasch, J. P., Schmidt, P., Alonso-Alija, C., Apeler, H., Dembowski, K., Haerter, M., et al. (2002). NO- and haem-independent activation of soluble guanylyl cyclase: molecular basis and cardiovascular implications of a new pharmacological principle. *Br. J. Pharmacol.* 136, 773–783. doi:10.1038/sj.bjp.0704778
- Stuehr, D. J., Misra, S., Dai, Y., and Ghosh, A. (2021). Maturation, inactivation, and recovery mechanisms of soluble guanylyl cyclase. *J. Biol. Chem.* 296, 100336. doi:10.1016/j.jbc.2021.100336
- Chernychev, B., Li, H., Lee, S. K., Gao, X., Ramanarasimhaiah, R., Liu, G., et al. (2021). Olinciguat, a stimulator of soluble guanylyl cyclase, attenuates inflammation, vaso-occlusion and nephropathy in mouse models of sickle cell disease. *Br. J. Pharmacol.* 178, 3463–3475. doi:10.1111/bph.15492
- Thompson, W. J., Piazza, G. A., Li, H., Liu, L., Fetter, J., Zhu, B., et al. (2000). Exisulind induction of apoptosis involves guanosine 3',5'-cyclic monophosphate phosphodiesterase inhibition, protein kinase G activation, and attenuated beta-catenin. *Cancer Res.* 60, 3338–3342.
- Thoonen, R., Giovannini, S., Govindan, S., Lee, D. I., Wang, G. R., Calamaras, T. D., et al. (2015). Molecular screen identifies cardiac myosin-binding protein-C as a protein kinase G- α substrate. *Circ. Heart Fail.* 8, 1115–1122. doi:10.1161/CIRCHEARTFAILURE.115.002308
- Tobin, J. V., Zimmer, D. P., Shea, C., Germano, P., Bernier, S. G., Liu, G., et al. (2018). Pharmacological characterization of IW-1973, a novel soluble guanylate cyclase stimulator with extensive tissue distribution, antihypertensive, anti-inflammatory, and antifibrotic effects in preclinical models of disease. *J. Pharmacol. Exp. Ther.* 365, 664–675. doi:10.1124/jpet.117.247429
- Toxvig, A. K., Wehland, M., Grimm, D., Infanger, M., and Kruger, M. (2019). A focus on riociguat in the treatment of pulmonary arterial hypertension. *Basic Clin. Pharmacol. Toxicol.* 125, 202–214. doi:10.1111/bcpt.13272
- Tripskiadis, F., Xanthopoulos, A., Skoularigis, J., and Starling, R. C. (2022). Therapeutic augmentation of NO-sGC-cGMP signalling: lessons learned from pulmonary arterial hypertension and heart failure. *Heart Fail Rev.* 27, 1991–2003. doi:10.1007/s10741-022-10239-5
- Tsai, E. J., and Kass, D. A. (2009). Cyclic GMP signaling in cardiovascular pathophysiology and therapeutics. *Pharmacol. Ther.* 122, 216–238. doi:10.1016/j.pharmthera.2009.02.009
- Udelson, J. E., Lewis, G. D., Shah, S. J., Zile, M. R., Redfield, M. M., Burnett, J. J., et al. (2020). Effect of praliciguat on peak rate of oxygen consumption in patients with heart failure with preserved ejection fraction: the CAPACITY HFpEF randomized clinical trial. *JAMA* 324, 1522–1531. doi:10.1001/jama.2020.16641
- Vaandrager, A. B. (2002). Structure and function of the heat-stable enterotoxin receptor/guanylyl cyclase C. *Mol. Cell Biochem.* 230, 73–83. doi:10.1023/A:1014231722696
- Vakalopoulos, A., Wunder, F., Hartung, I. V., Redlich, G., Jautelat, R., Buchgraber, P., et al. (2023). New generation of sGC stimulators: discovery of imidazo[1,2-a]pyridine carboxamide BAY 1165747 (BAY-747), a long-acting soluble guanylate cyclase stimulator for the treatment of resistant hypertension. *J. Med. Chem.* 66, 7280–7303. doi:10.1021/acs.jmedchem.2c02082
- Vannuccini, F., Campora, A., Barilli, M., and Palazzuoli, A. (2022). Vericiguat in heart failure: characteristics, scientific evidence and potential clinical applications. *Biomedicines* 10, 2471. doi:10.3390/biomedicines10102471
- Waldman, S. A., and Camilleri, M. (2018). Guanylate cyclase-C as a therapeutic target in gastrointestinal disorders. *Gut* 67, 1543–1552. doi:10.1136/gutjnl-2018-316029
- Wang, L., Zhu, L., Wu, Y., Li, Q., and Liu, H. (2021). Riociguat therapy for pulmonary hypertension: a systematic review and meta-analysis. *Ann. Palliat. Med.* 10, 1117–11128. doi:10.21037/apm-21-2656
- Watanabe, H. (2018). Treatment selection in pulmonary arterial hypertension: phosphodiesterase type 5 inhibitors versus soluble guanylate cyclase stimulator. *Eur. Cardiol.* 13, 35–37. doi:10.15420/ecr.2017.22:2

- Weatherald, J., Thakrar, M. V., Varughese, R. A., Kularatne, M., Liu, J., Harper, L., et al. (2022). Upfront riociguat and ambrisentan combination therapy for newly diagnosed pulmonary arterial hypertension: a prospective open-label trial. *J. Heart Lung Transpl.* 41, 563–567. doi:10.1016/j.healun.2022.01.002
- Weinberg, D. S., Foster, N. R., Della’Zanna, G., McMurray, R. P., Kraft, W. K., Pallotto, A., et al. (2021). Phase I double-blind, placebo-controlled trial of dolcanatide (SP-333) 27 mg to explore colorectal bioactivity in healthy volunteers. *Cancer Biol. Ther.* 22, 544–553. doi:10.1080/15384047.2021.1967036
- Weinberg, D. S., Lin, J. E., Foster, N. R., Della’Zanna, G., Umar, A., Seisler, D., et al. (2017). Bioactivity of oral linaclotide in human colorectum for cancer chemoprevention. *Cancer Prev. Res. (Phila)* 10, 345–354. doi:10.1158/1940-6207.CAPR-16-0286
- Xiao, B., Zhong, G., Obayashi, M., Yang, D., Chen, K., Walsh, M. P., et al. (2006). Ser-2030, but not Ser-2808, is the major phosphorylation site in cardiac ryanodine receptors responding to protein kinase A activation upon beta-adrenergic stimulation in normal and failing hearts. *Biochem. J.* 396, 7–16. doi:10.1042/BJ20060116
- Xing, C., Zhang, T., Liu, X., Li, C., Yang, G., Zhang, H., et al. (2021). Sleep disturbance induces depressive behaviors and neuroinflammation by altering the circadian oscillations of clock genes in rats. *Acta histochem.* 171, 124–132. doi:10.1016/j.neures.2021.03.006
- Yang, J., and Lei, Y. (2021). Comparison of the efficacy and safety of different doses of linaclotide for patients with chronic constipation: a meta-analysis and bayesian analysis. *Evid. Based Complement. Altern. Med.* 2021, 9923879. doi:10.1155/2021/9923879
- Yang, L., Liu, G., Zakharov, S. I., Bellinger, A. M., Mongillo, M., and Marx, S. O. (2007). Protein kinase G phosphorylates Cav1.2 alpha1c and beta2 subunits. *Circ. Res.* 101, 465–474. doi:10.1161/CIRCRESAHA.107.156976
- Zimmer, D. P., Shea, C. M., Tobin, J. V., Tchernychev, B., Germano, P., Sykes, K., et al. (2020). Olinciguat, an oral sGC stimulator, exhibits diverse pharmacology across preclinical models of cardiovascular, metabolic, renal, and inflammatory disease. *Front. Pharmacol.* 11, 419. doi:10.3389/fphar.2020.00419

Glossary

| | |
|-----------------|---|
| GTP | guanosine triphosphate |
| cGMP | cyclic guanosine monophosphate |
| sGC | soluble guanylate cyclase |
| NO | nitric oxide |
| CO | carbon monoxide |
| H-NOX | N-hemoglobin-nitric oxide |
| GC-C | guanylate cyclase C |
| ECD | extracellular domain |
| NOS | nitric oxide synthases |
| eNOS | endothelial nitric oxide synthase |
| nNOS | neuronal nitric oxide synthase |
| PKGs | protein kinases |
| PDEs | phosphodiesterases |
| PLB | phospholamban |
| RyR2 | Cardiac ryanodine receptor |
| SERCA | sarco endoplasmic reticulum Ca ²⁺ -ATPase |
| ER | endoplasmic reticulum |
| LTCCs | L-type calcium channels |
| TRPC6 | transient receptor potential canonical channel type 6 |
| TnI | troponin I |
| TTN | titin |
| CMyBP-C | cardiac myosin-binding protein-C |
| VSMC | venous smooth muscle cell |
| MLCP | myosin light chain phosphatase |
| RhoA | Ras homolog family member A |
| RGS-2 | regulator of G-protein signaling 2 |
| IP3 | inositol 1,4,5-trisphosphate |
| Rap1Gap2 | Rap1 GTPase-activating protein 2 |
| Rap1b | Ras-related protein 1 |
| IRAG | IP3-induced calcium release |
| GRP2 | guanyl-releasing protein 2 |
| BKCa | calcium-sensitive potassium channels |
| PASMCs | pulmonary artery smooth muscle cells |
| MLCK | myosin light chain kinase |
| VASP | vasodilator-stimulated phosphoprotein |
| BMP | bone morphogenetic protein |
| SMAD | small mothers against decapentaplegic |
| PDE5 | phosphodiesterase 5 |
| NFAT | calcineurin-nuclear factor of activated T cells |
| HF | heart failure |
| CVD | cardiovascular disease |

| | |
|---------------|--|
| HFrEF | heart failure with reduced ejection fraction |
| HFpEF | heart failure with preserved ejection fraction |
| ERK | extracellular signal-regulated kinase |
| ECM | extracellular matrix |
| CKD | chronic kidney disease |
| PAH | pulmonary arterial hypertension |
| CTEPH | chronic thromboembolic pulmonary hypertension |
| AE | adverse event |
| SAE | serious adverse event |
| RV | right ventricular |
| PH-IIP | pulmonary hypertension associated with idiopathic interstitial pneumonia |
| 6MWD | 6-min walk distance |
| PAP | pulmonary artery pressure |
| PVR | pulmonary vascular resistance |
| RAP | right atrial pressure |
| CI | cardiac index |
| CO | cardiac output |
| BPA | balloon pulmonary angioplasty |
| SAPH | sarcoidosis associated pulmonary hypertension |
| PK | pharmacokinetics |
| WHO-FC | World Health Organization Functional Classification |
| TAC | transverse aortic constriction |
| LVEF | left ventricular rejection fraction |
| LVM/BW | left ventricular mass to body weight ratio |
| MYH7 | myosin heavy chain 7 |
| PLN | cardiac phosphoprotein |
| ANKRD1 | ankyrin repeat domain-containing protein 1 |
| dcSSc | diffuse cutaneous systemic sclerosis |
| mRSS | modified Rodnan skin score |
| HUVECs | human umbilical vein endothelial cells |
| PACs | pro-angiogenic cells |
| CCSs | chronic coronary syndromes |
| BP | blood pressure |
| SBP | systolic blood pressure |
| DBP | diastolic blood pressure |
| PD | pharmacodynamics |
| ECG | electrocardiogram |
| DN | diabetic nephropathy |
| RPTC | renal proximal tubular epithelial cells |
| T2D | type 2 diabetes |
| RHI | reactive hyperemia index |

| | | | |
|-------------------------|---|------------------|---|
| HLI | hind limb ischemia | CIC | chronic idiopathic constipation |
| ICAM1 | intercellular adhesion molecule 1 | AUC | area under the concentration-time curve |
| Myh2 | MyHCLa | Cmax | maximum concentration |
| Cxcl12 | C-X-C motif chemokine ligand 12 | T1/2 | drug half-life |
| PEG | polyethylene glycol | GUCY2C | Guanylate cyclase C |
| SSc-MVECs | SSc dermal microvascular endothelial cells | TEAEs | treatment-emergent adverse events |
| RBCs | red blood cells | IIP | Idiopathic Interstitial Pneumonias |
| TGFβ | transforming growth factors β | NT-proBNP | N-terminal pro-B-type natriuretic peptide |
| CYP450 | cytochrome P450 | LVEDV | left ventricular end-diastolic volume |
| cGKI | cGMP-dependent protein kinase type I | LVESV | left ventricular end-systolic volume |
| CLb | blood clearance | ABPM | Ambulatory BP monitoring |
| uHTS | ultrahigh-throughput screening | HOMA-IR | Homeostatic Model Assessment of Insulin Resistance |
| PCWP | pulmonary capillary wedge pressure | ADOs | adverse events resulting in study drug discontinuation |
| ADHF | acute decompensated heart failure | BFT | Supine Bolus Flow Time |
| IOP | intraocular pressure | VAS | visual analog scale |
| CAVS | calcific aortic valve stenosis | LOCF | last observation carried forward |
| SMA | smooth muscle actin | NPSI | Neuropathic Pain Symptom Inventory |
| PRP | platelet rich plasma | PAD | Peripheral Arterial Disease |
| SRM | sirius red morphometry | ICD | initial claudication distance |
| DKD | diabetic kidney disease | AVC | Aortic Valve Calcification |
| CFTR | cystic fibrosis transmembrane regulator | UACR | urinary albumin-to-creatinine ratio |
| Cl⁻ | chloride ions | NPDR | nonproliferative diabetic Retinopathy |
| cAMP | cyclic adenosine monophosphate | DRSS | Diabetic Retinopathy Severity Scale |
| PKA | protein kinase A | CSPH | clinically significant portal hypertension |
| HCO3⁻ | bicarbonate ion | HVPG | hepatic venous pressure gradient |
| NHE3 | Na ⁺ /H ⁺ exchanger 3 | CTCAE | Common Terminology Criteria for Adverse Events |
| IBDs | inflammatory bowel diseases | FVC | forced vital capacity |
| MRP4 | multidrug resistance-associated protein 4 | KCCQ-CSS | Kansas City Cardiomyopathy Questionnaire Clinical Summary Score |
| Akt | protein kinase B | EQ-5D | 5-dimension EuroQol questionnaire |
| p38 MAPK | p38 mitogen-activated protein kinase | LAV | left atrial volume |
| CNGs | cyclic nucleotide-gated ion channels | | |
| MMP-9 | matrix metalloproteinase-9 | | |
| IBS-C | irritable bowel syndrome with constipation | | |
| CC | chronic constipation | | |
| CSBM | complete spontaneous defecation | | |
| IBS-SSS | IBS Symptom Severity Scale | | |
| IBS-QOL | IBS Quality of Life Questionnaire | | |
| CI | confidence interval | | |
| FC | functional constipation | | |
| GI | gastrointestinal | | |
| OIC | opioid-induced constipation | | |
| SBMs | spontaneous bowel movements | | |



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The pathogenesis and therapeutic strategies of heat stroke-induced myocardial injury

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Heat stroke (HS) is a febrile illness characterized by an elevation in the core body temperature to over 40°C, accompanied by central nervous system impairment and subsequent multi-organ dysfunction syndrome. In recent years, the mortality rate from HS has been increasing as ambient temperatures continue to rise each year. The cardiovascular system plays an important role in the pathogenesis process of HS, as it functions as one of the key system for thermoregulation and its stability is associated with the severity of HS. Systemic inflammatory response and endothelial cell damage constitute pivotal attributes of HS, other factors such as ferroptosis, disturbances in myocardial metabolism and heat shock protein dysregulation are also involved in the damage to myocardial tissue in HS. In this review, a comprehensively detailed description of the pathogenesis of HS-induced myocardial injury is provided. The current treatment strategies and the promising therapeutic targets for HS are also discussed.

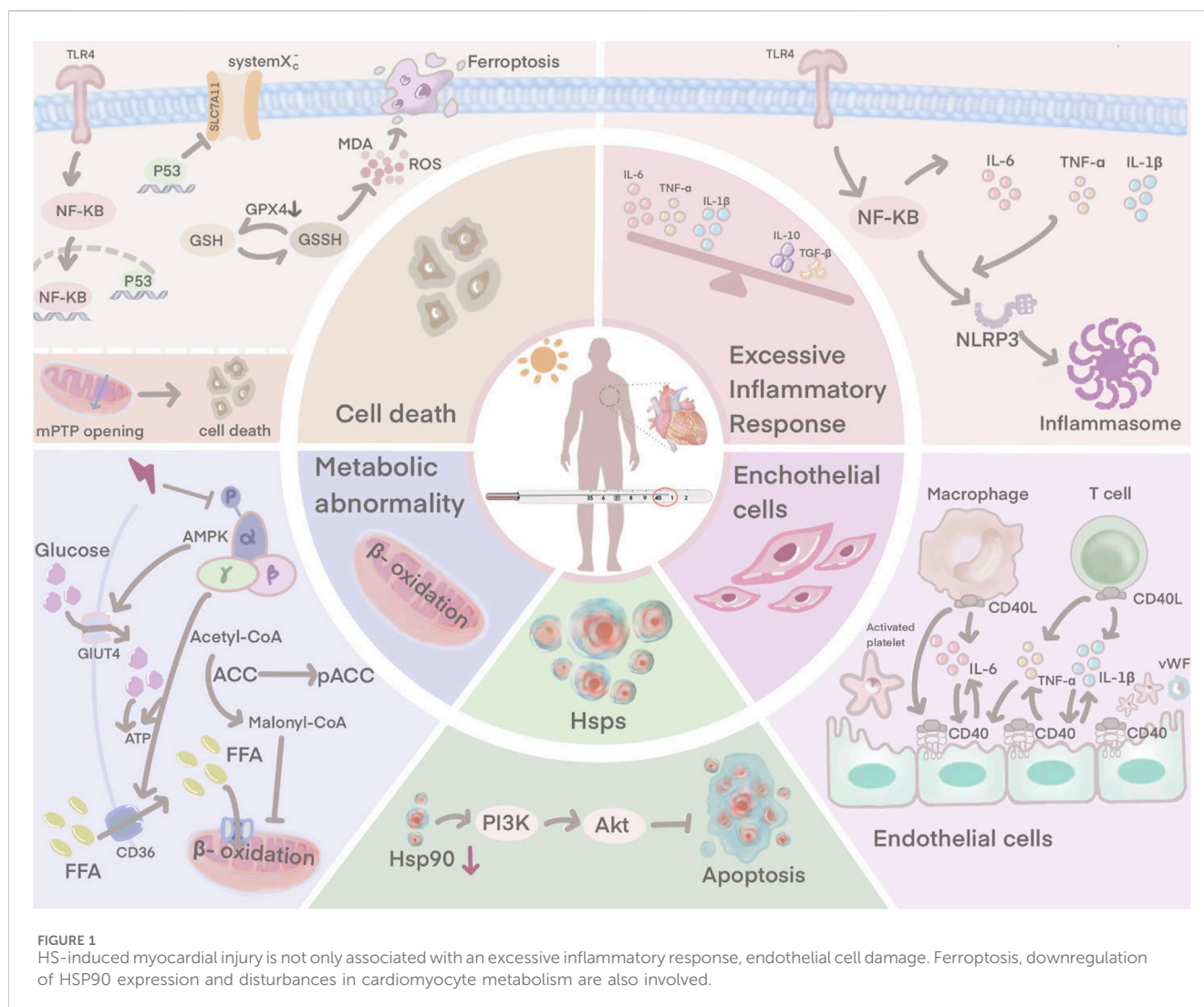
KEYWORDS

heat stroke, myocardial injury, pathogenesis, therapeutic strategy, inflammation

1 Introduction

Heat stroke (HS) is an illness characterized by a rapid rise of core temperature over 40°C with the complication of systemic inflammatory responses and central nervous system dysfunction (Bouchama and Knochel, 2002; Leon and Helwig, 2010; Peiris et al., 2017). In recent years, heat-related deaths have increased significantly due to anthropogenic climate change (Toutant et al., 2011). Frequency of severe heat waves is threatening human health worldwide and poses huge challenges to public health, attracting widespread attention in various research fields (M. Zheng et al., 2020). HS can be divided into classic heat stroke (CHS) and exertional heat stroke (EHS) depending on the involvement of skeletal muscle contraction (Bouchama et al., 2022). CHS often occurs in older people having pre-existing illnesses, while EHS typically occurs in healthy younger individuals during strenuous

Abbreviations: HS, Heat stroke; CHS, Classic heat stroke; EHS, Exertional heat stroke; SIRS, Systemic inflammatory response syndrome; TNF- α , Tumor necrosis factor- α ; IL-1 β , Interleukin-1 β ; NLRP3, NOD-like receptor family pyrin domain containing3; vWF, von Willebrand factor; DIC, Disseminated intravascular coagulation; ROS, Reactive oxygen species; GSH, Glutathione; SLC7A11, Solute carrier family 7 member 11; MDA, Malondialdehyde; ACC, Acyl CoA-carboxylase; FAs, Fatty acids; GLUT4, Glucose transporter 4; HSP, Heat shock protein.



exercise in hot environments (Peiris et al., 2017; Bouchama et al., 2022). HS, regardless of the type, is associated with extensive multi-organ tissue damage as a result of the interaction of cytotoxic, inflammatory, and clotting reactions (Piver et al., 1999). The heart, being a vulnerable organ in heat injury (Low et al., 2011; Lou et al., 2019; Ko et al., 2020), is susceptible to arrhythmia, function failure and focal myocardial necrosis (Argaud et al., 2007; Desai et al., 2023).

Abnormalities in temperature regulation, cardiovascular function and tissue perfusion are among the factors involved in multiple organ dysfunction syndrome (Low et al., 2011; Cramer et al., 2022). In an effort to dissipate heat, the body increases blood flow to the skin, redistributes blood and eventually develops hypotension and perfusion disorders (S. H. Chen et al., 2006). Thus, the regulation of the cardiovascular system plays a key role in the pathogenesis of HS. Elucidation of the mechanism of HS-induced myocardial injury can help in establishing the treatment to improve circulatory function and reduce mortality rates of HS. However, the pathogenesis of HS is still to be known and prevention strategies of myocardial injury during HS is lacking. This article provides a systematic review to further the

understanding of HS-induced myocardial injury and to provide a reference for future research (Figure 1).

2 Heat stroke and myocardial injury

Under normal conditions, a 0.3°C increase in core temperature triggers a cardiovascular regulatory response to protect the body from heat damage. This regulation increases heat dissipation by speeding up the heart rate, enhancing cardiac contractility, raising cardiac output and reducing blood flow and volume in non-skin areas (Crandall et al., 2008; Crandall and González-Alonso, 2010).

During HS, when the surrounding hot environment persists, the above regulation continues to function actively. A substantial volume of blood is pumped from the heart towards the peripheral blood vessels to dissipate heat through sweat, but this also results in hyperthermic dehydration of the body, reduced circulating blood volume, inadequate tissue perfusion, hypoxia and necrosis of myocardial cells (Crandall and González-Alonso, 2010; G. D; Chen et al., 2019). At the same time, the loss of body fluids disturbs electrolytes and interrupts the sodium-potassium

pump, which alters the heart's pacing rhythm, signal conduction and systolic-diastolic functional state, ultimately leading to myocardial ischemia, necrosis, arrhythmia and heart failure (Hausfater et al., 2010; Chen, et al., 2019; Tseng et al., 2019; Wang et al., 2019).

3 The related mechanisms of HS-induced myocardial injury

3.1 Dysregulation of the pro-inflammatory and anti-inflammatory balance

In HS, the systemic pro- and anti-inflammatory balance is disturbed, triggering a systemic inflammatory response syndrome (SIRS) that is thought to be characteristic (Epstein and Yanovich, 2019). The pathogenesis of heat stroke is closely similar to that of sepsis (Roberts et al., 2008). In a hot environment, the dilatation of blood vessels on the body surface due to heat dissipation leads to reduced blood flow to internal organs, especially intestinal mucosa, which causes increased intestinal epithelial permeability and bacterial translocation in the intestine, inducing leakage of intestinal endotoxins through the intestine into the circulation and triggering SIRS, ultimately leading to multi-organ dysfunction and death (Yang et al., 2007; Lambert, 2008; Leon and Helwig, 2010). The systemic inflammation associated with heat stroke plays a key role in myocardial injury. Currently, it is thought that the myocardial inflammatory response may be the primary cause of progressive systolic dysfunction (Dörge et al., 2000; Dörge et al., 2002). A large infiltration of inflammatory cells is usually found within the foci of myocardial infarction. Previous studies have shown that suppression of the inflammatory response is an important tool in the treatment of HS-induced myocardial injury (Lin et al., 2017; Lin et al., 2018; Lin et al., 2020).

During HS, the body undergoes a state of hypercytokinemia, releasing many cytokines such as tumor necrosis factor- α (TNF- α) and interleukin-1 β (IL-1 β) (Leon and Helwig, 2010; Z. T; Zhang et al., 2021). TNF- α , a key factor in the inflammatory response, plays an important role in neutrophil recruitment and the inflammatory cascade reaction (Yu et al., 2010). In addition, TNF- α induces the production of other inflammatory cytokines and also stimulates the migration and adhesion of neutrophils, leading to dysregulation of pro- and anti-inflammatory factors and inducing an inflammatory cascade reaction, which results in tissue damage (Yu et al., 2010). At the same time, the injured myocardial tissue also releases pro-inflammatory cytokines, including TNF- α and IL-6, which further exacerbate the systemic inflammatory response (Shen et al., 2019).

The TLR4/NF- κ B signaling pathway has a major contribution to HS-induced inflammation. TLR4 is an essential member of the TLR family and plays a central role in the recognition and response to microbial pathogens and in maintaining the integrity of the intestinal epithelial barrier (D. Yao et al., 2019). Rats subjected to heat stress have significantly elevated levels of TLR4 (D. Chen et al., 2023). When rats are affected by heat stress, NF- κ B is activated by the induced TLR4, leading to the release of pro-inflammatory factors. The production and release of pro-inflammatory factors further activates NF- κ B, which induces the NLRP3 inflammasome, leading to a sustained amplification of the initial inflammatory

signal, thus causing the so-called inflammatory cascade effect (Z. Huang et al., 2016; X; Zhang et al., 2017). TLR4 exhibits its highest expression in cardiac myocytes, and during HS, TLR4/NF- κ B signaling controls the production of pro-inflammatory factors to induce myocardial tissue damage (X. Liu et al., 2016). Inhibition of the TLR4 signaling pathway may reduce HS-induced inflammatory responses and improve abnormal cardiac function in rats (Chen et al., 2023).

3.2 Endothelial cell damage and dysfunction

Cardiac ultrastructure in HS patients exhibits severe endothelial cell damage (Sohal et al., 1968). Vascular endothelial cells cover the surface of the lumen and maintain the structural integrity and microcirculatory function of the coronary microvasculature (Chang et al., 2021). It also acts as a defensive barrier against the penetration of microorganisms, immune cells and coagulation components, which reduces the risk of thrombosis (Chang et al., 2021). Activated *in vivo* crosstalk exists between vascular endothelium, inflammation and coagulation during HS (Bouchama et al., 1991; al-Mashhadani et al., 1994; Roberts et al., 2008). Endothelial cell dysfunction plays a key role in the initiation and progression of HS (W. Huang et al., 2022).

Endothelial cells possess an anti-inflammatory effect under normal physiological conditions, repelling circulating neutrophils from adhesion (Chang et al., 2021). However, when rat myocardial tissue is damaged by heat stress, endothelial cells upregulate a variety of adhesion molecules that attract pro-inflammatory cells (neutrophils and macrophages) to secrete pro-inflammatory cytokines (Harlan et al., 1991; Wihastuti et al., 2018; Chang et al., 2021). Large amounts of pro-inflammatory factors such as IL-6 and TNF- α can trigger endothelial dysfunction and microvascular damage (F. Chen et al., 2017). Damaged endothelial cells express CD40, and in the presence of CD40 interacting with CD40 ligand (CD40L), endothelial cells actively secrete von Willebrand factor (vWF), which promotes platelet adhesion to endothelial cells and contributes to thrombosis (Keuren et al., 2004; Han et al., 2018). The interaction between CD40 and CD40L also stimulates platelets and endothelial cells to activate macrophages and T cells, which further amplifies the inflammatory response (Urbich et al., 2002). Damage to the endothelium, a natural barrier against thrombosis, upregulates procoagulant factors and downregulates anticoagulant factors, thereby disturbing the dynamic balance between pro- and anti-thrombotic activities and inducing microthrombosis (Koupenova et al., 2017). Obstruction of small vessels contributes to infarction and necrosis of myocardial tissue. The damaged tissue releases plasminogen activator which induces the development of disseminated intravascular coagulation (DIC) (Sohal et al., 1968). Hearts of patients with HS show evidence of extensive visual and microscopic haemorrhage (Sohal et al., 1968).

Aspirin, a non-steroidal anti-inflammatory drug, that not only inhibits platelet aggregation but also maintains the integrity of endothelial gap junctions (Zhou et al., 2019). Animal study has shown that the treatment with aspirin significantly improves the morphological damage and related enzyme activity of chicken cardiomyocytes induced by heat stress (Wu et al., 2016).

3.3 Abnormal cardiomyocyte death

HS instigates multiple toxic effects on the cardiovascular system, including abnormal cardiomyocyte death (Chen et al., 2017; Chen et al., 2023). The damaged myocardial cells exhibit vacuolar changes and partial necrosis (Fan et al., 2015; Chen et al., 2019). Ferroptosis is an essential form of abnormal cardiomyocyte death caused by HS, resulting from the excessive accumulation of iron-dependent lipid reactive oxygen species (ROS) in cells, where lipid peroxidation is a key component in triggering ferroptosis (Del Re et al., 2019; Stockwell et al., 2020). HS disrupts the oxidation-antioxidant balance, as evidenced by a decrease in glutathione (GSH) and solute carrier family 7 member 11 (SLC7A11), an increase in malondialdehyde (MDA), ROS, and Fe^{2+} . HS also induces shrinkage of mitochondria and an increase in the membrane density, which are key features of ferroptosis (Jiang et al., 2021; Chen et al., 2023). This suggests that ferroptosis is actively involved in HS-induced myocardial injury and causes abnormal cardiomyocyte death. Chen et al. further explored the mechanism of ferroptosis in the HS myocardial injury model (D. Chen et al., 2023). P53 expression levels were closely associated with the triggering of ferroptosis (Lei et al., 2021), and its involvement as a transcriptional repressor of SLC7A11 to ferroptosis significantly reduced the expression of SLC7A11, which in turn inhibited the activity of system X_c^- , a component of SLC7A11 (Koppula et al., 2018), thereby inhibiting cysteine uptake and reducing GPX4 activity leading to depletion of GSH biosynthesis (Xu et al., 2021; Zhang et al., 2022). Consequently, lipid peroxide accumulation ensued, ultimately culminating in cellular ferroptosis (Ma et al., 2022). P53, one of the molecules downstream of TLR4, is activated by the TLR4/NF- κ B signaling pathway, which plays an active role in the systemic inflammatory response induced by HS (Zhu et al., 2011). In view of this, Chen et al. suggested that HS may induce ferroptosis through the TLR4/NF- κ B/P53 signaling pathway (Chen et al., 2023). Inhibition of TLR4 and NF- κ B under HS conditions downregulated P53 expression, upregulated SLC7A11 and GPX4 levels, improved ferroptosis-related indicators and attenuated myocardial injury, respectively (Chen et al., 2023).

Disruption of mitochondrial structure and function can lead to severe cellular damage and death (Zamzami et al., 1997; D'Orsi et al., 2017). Mitochondria plays a crucial role in maintaining intracellular calcium homeostasis (D'Orsi et al., 2017). From rat cardiomyocytes, we know that heat stress causes mitochondrial changes in cardiac myocytes including mitochondrial swelling, rupture of cristae and disruption of the surrounding membrane (Petit et al., 1998; Qian et al., 2004). Ca^{2+} -ATPase on the mitochondrial membrane serves as critical factor in the regulation of calcium homeostasis. However, disruption by heat stress leads to a decrease in Ca^{2+} -ATPase activity, which results in reduced mitochondrial uptake of calcium ions from the cytoplasm and intracellular calcium overload (McCormack and Denton, 1989; Walkon et al., 2022). Intracellular calcium overload further activates calcium-dependent protein kinases, which promote membrane phospholipid hydrolysis, disrupting the cytoskeleton and damaging the integrity of the nucleus, causing severe damage (Vassalle and Lin, 2004). HS directly induces the opening of mitochondrial mPTP, a pivotal event in triggering the cell death

pathway (Halestrap, 2009; Bauer and Murphy, 2020). mPTP opening results in a series of cytological effects that lead to the release of cytochrome c, activation of caspase family proteases and apoptosis of cardiomyocytes (H. Yao et al., 2022). The mechanism by which HS induces mPTP opening is not yet clear, and the Fas pathway is an important signaling pathway to consider. It induces caspase-8 activation, which subsequently directly activates caspase-3 and leads to the opening of mPTP (Nakamura et al., 2000). However, whether the Fas pathway is involved in HS-induced mPTP opening remains to be explored.

3.4 Metabolic abnormalities

The link between metabolic dysregulation and cardiotoxicity has been well established (Russo et al., 2021). Mitochondrial damage caused by HS not only results in abnormal death of cardiomyocytes but also leads to disturbances in energy metabolism (Azevedo et al., 2013). Energy abnormalities in the heart are associated with the development of many heart diseases (X. Wang et al., 2023). Heat stress disrupts the integrity of the mitochondria, which is the basis for normal mitochondrial function, resulting in a suppression of energy production from the oxidative metabolism of cardiomyocytes (Patra and Hay, 2014; Laitano et al., 2020; Deng et al., 2022). However, in response to the high temperatures of the external environment, the heart requires a greater supply of energy to enhance cardiac function, which leads to a significant decrease in the ATP content of the cardiomyocytes and eventual death due to energy deficiency (Qian et al., 2004).

Glucose and fatty acids are essential substrates for oxidative phosphorylation. Glucose and lipid metabolism plays an important role in cardiac myocytes by providing energy and maintaining cellular function (H. Tian et al., 2023). However, studies in murine models of EHS have revealed that HS alters cardiomyocyte metabolic pathways, disrupts the glycolytic and oxidative phosphorylation pathways by upregulating glycolysis-related enzymes, thereby enhancing lactate production to impair cardiomyocyte function (Laitano et al., 2020). The perturbation of glucose and lipid metabolism by HS may be related to the inhibition of the AMPK signaling pathway (Rodríguez et al., 2021). AMPK increases ATP production in cardiomyocytes through stimulation of glucose metabolism and fatty acid oxidation. AMPK phosphorylation at Thr¹⁷² induces acyl CoA-carboxylase (ACC) phosphorylation to inhibit the conversion of acetyl-CoA to malonyl-CoA during fatty acids (FAs) synthesis (Carling et al., 2008). Beyond the inhibition of lipid anabolism, p-AMPK also promotes FAs uptake by inducing the activity of the FAs transporter CD36, enhancing β -oxidation (Habets et al., 2009). Glucose metabolism is also regulated by AMPK. p-AMPK increases glucose transporter 4 (GLUT4), which promotes glucose uptake and thus provides a source of energy (D. Zheng et al., 2001). Under HS conditions, phosphorylation of AMPK is inhibited, leading to dysregulation of glucolipid metabolism and disruption of energy metabolism (Roths et al., 2023). This ultimately leads to cell death and impaired cardiac function. Therefore, targeting glucose and lipid metabolism may be an effective way to counteract HS-induced myocardial injury.

3.5 Heat shock protein dysregulation

Cells from a murine model of myocardial tissue turn on their intrinsic defense mechanisms in the face of heat injury, with a dramatic increase in heat shock protein (HSP) expression being a key part of the heat shock response (Tang et al., 2013; Tang et al., 2016). It can interlock with apoptosis, inflammation and autophagy to regulate cellular homeostasis and prevent tissue damage (Hsu et al., 2013; Shen et al., 2019). It was mentioned earlier that patients with HS can develop severe vascular endothelial cell damage. After heat exposure, strong positive signals for HSP90 and HSP70 are detected in rat cardiac microvascular endothelial cells, helping the vascular endothelium to resist heat injury (X. Zhang et al., 2020). An increase in HSP90 activates the PI3K/Akt signaling pathway. Phosphorylated Akt negatively regulates the expression of pro-apoptotic proteins and contributes to cell survival (Zhang et al., 2020). It is known from rat-related experiments that HSP levels vary with the duration of heat stress. In the early stages of HS, HSP rises sharply, and as time progresses, HSP is heavily depleted, resulting in abnormally low HSP levels in the later stages (H. B. Chen et al., 2015; Lin et al., 2020). When HSP90 is crushed, the interaction of HSP90 with Akt is reduced, weakening the protective effect. This results in the vascular endothelium exhibiting a more sensitive state to heat stress and more severe damage (Zhang et al., 2020).

4 The treatment strategy for HS

The prognosis of patients with heat stroke is directly related to the degree and duration of the increase in core temperature (Hadaad et al., 2004). Therefore, whole-body cooling is the current treatment of choice for HS. Following the onset of HS, hypotension and altered cardiac protein profiles are demonstrated, which can be reversed by whole-body cooling (Ko et al., 2020). Temperature reduction is achieved mainly by conduction, evaporation and convection (Hadaad et al., 2004). In addition, symptomatic support therapy is an integral part of the treatment. When hypotension occurs in patients, aggressive fluid resuscitation and vasoactive medication should be administered with the avoidance of alpha-adrenergic drugs as they exacerbate peripheral vasoconstriction and inhibit core body temperature reduction (Atha, 2013; Asmara, 2020). Excessive inflammation and coagulation disorders are important pathogenic mechanisms of HS, therefore anti-inflammatory and anticoagulant therapies are also available as treatment options (Y. F. Tian et al., 2013; Kobayashi et al., 2018). For patients who progress to multi-organ dysfunction despite hypothermia treatment, continuous blood purification and plasma exchange are often selected, aiming not only to alleviate the body's catabolic state but also to eliminate inflammatory mediators from the bloodstream to facilitate the recovery of HS patients (Wakino et al., 2005; K. J. Chen et al., 2014). Given the danger and intractability of HS, prevention strategies are far more beneficial than any present treatment strategies. People at risk of heat exposure should be thermally acclimatized in advance, with consumption of sufficient fluids and adequate nutrition (Asmara, 2020).

5 Discussion

HS involves a complex biochemical cascade of reactions and is caused by a combination of factors. The cardiovascular system is considered to be the first system affected by HS. Circulatory shock occurs in approximately 20%–65% of patients, and an even higher 85% of patients will develop ECG abnormalities (Austin and Berry, 1956; Asmara, 2020). However, a dearth of clinical directives exists regarding the efficacious management of cardiovascular ailments amidst elevated temperatures. A precise comprehension of the fundamental mechanisms whereby heightened temperatures inflict harm upon myocardial tissue is imperative to judiciously formulate preventative and therapeutic strategies. This review summarizes the possible pathogenesis of HS-induced myocardial injury, which can help provide new targets for the treatment of HS.

The predominant body of research scrutinizing myocardial impairment due to hyperthermia predominantly comprises animal studies, with a paucity of involvement from clinical cohorts. The acquisition of clinical data assumes heightened significance. A comprehensive database analysis encompassing 27 countries spanning the years 1979–2019 revealed a 7% escalation in mortality among patients with ischemic heart disease during episodes of soaring temperatures (Alahmad et al., 2023). Moreover, a meta-analysis delineated a 2.8% augmentation in the risk of developing coronary heart disease for each 1°C ascent in temperature (J. Liu et al., 2022). Endothelial cell damage within cardiac vasculature due to pyrexia precipitates thrombosis, culminating in acute coronary incidents. Clinical investigations have documented a substantial surge in hospitalizations linked to coronary artery disease following exposure to elevated temperatures (Fuhrmann et al., 2016). Long-term monitoring of hyperthermia-stricken patients corroborates the critical role of intact myocardial tissue, with a meager 1-year survival rate of merely 24% observed in cases with markedly elevated troponin levels (Marchand and Gin, 2022). This underscores the profound impact of myocardial impairment on the prognosis of hyperthermia-afflicted individuals. Consequently, the primary focus of research should pivot towards averting myocardial damage induced by HS.

The incomplete comprehension of HS pathogenesis, coupled with the absence of evidence-based medical guidance for clinical interventions, has resulted in the inadequacies of current treatment modalities. Predominantly, supportive therapies such as whole-body cooling and fluid resuscitation constitute the primary approach. Regrettably, a lack of standardized endpoint objectives for whole-body cooling persists to date. Furthermore, despite numerous animal studies affirming the favorable efficacy of anti-inflammatory and anti-endotoxin agents for HS, their translation into clinical success remains limited. Aspirin, despite demonstrating effectiveness against heat-induced injury in avian cardiac tissues, fails to manifest any clinical benefit and may potentially exacerbate coagulation disorders and hepatic dysfunction (Tek and Olshaker, 1992). Individuals with cardiovascular ailments not only contend with the vulnerability of their cardiac systems in the face of HS but also grapple with an elevated risk due to commonly prescribed cardiac medications. β -blockers impede the capacity to augment cardiac output in response to HS, while diuretics exacerbate hypovolemia and elevate the risk of electrolyte imbalances (Marchand and Gin, 2022). This begs the question of which

medications can be initiated or stopped during extreme heat conditions.

In cases where the heart receives heat damage, significant changes in cardiac metabolism occur. These changes are not only passive bystanders, but are actual participants in causing heat stress damage to the myocardium. Targeting myocardial metabolism could be the tool for our effective treatment. Interventions in cardiac metabolic processes have been successfully used to reduce infarct size in animal models of myocardial ischaemia-reperfusion injury (Valls-Lacalle et al., 2018; Zuurbier et al., 2020). However, suitable drug targets for conversion in patients with acute myocardial infarction are still awaited. Cardiometabolic therapies are challenging, but fortunately, recent methodological advances in detecting metabolic changes within the heart will make our efforts more achievable.

In conclusion, HS-induced myocardial injury arises from a combination of excessive inflammation, endothelial cell damage, abnormal cardiac metabolism, and heat shock protein dysregulation. In the treatment, in addition to systemic supportive therapy it should also focus on precise targeting of myocardial tissue. Only with a deeper and clearer understanding of the mechanisms underlying the development of HS will there be an opportunity to establish more effective treatment.

6 Future perspective

Given the increasing mortality rate associated with HS, extensive research has been conducted to explore this condition. A meticulous examination of the literature has revealed potential molecular targets for HS treatment, encompassing TLR4, P53, AMPK, and HSP. Additionally, the Fas signaling pathway presents a novel avenue for HS management. However, the majority of these investigations have been confined to the realm of animal studies, and the therapeutic strategies delineated await clinical validation. Consequently, we should focus more on clinical trials to find relevant drug targets that can serve clinical HS patients. Furthermore, the absence of targeted therapy for HS-induced myocardial injury underscores the need for advancements in this

area. Fortunately, the rapid development of modern bioinformatics technologies offers us valuable tools to deepen our understanding of the pathogenesis of HS-induced myocardial injury and implement precise treatments.

Author contributions

RX: Writing–original draft, Writing–review and editing. MS: Writing–original draft. YL: Writing–review and editing. JY: Writing–review and editing. HL: Software, Visualization, Writing–review and editing. JY: Software, Visualization, Writing–review and editing. JL: Software, Writing–review and editing. YH: Visualization, Writing–review and editing. BW: Validation, Writing–review and editing. GY: Validation, Writing–review and editing. JL: Supervision, Writing–review and editing.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

- Alahmad, B., Khraishah, H., Royé, D., Vicedo-Cabrera, A. M., Guo, Y., Papatheodorou, S. I., et al. (2023). Associations between extreme temperatures and cardiovascular cause-specific mortality: results from 27 countries. *Circulation* 147, 35–46. doi:10.1161/circulationaha.122.061832
- al-Mashhadani, S. A., Gader, A. G., al Harthi, S. S., Kangav, D., Shaheen, F. A., and Bogus, F. (1994). The coagulopathy of heat stroke: alterations in coagulation and fibrinolysis in heat stroke patients during the pilgrimage (Haj) to Makkah. *Blood Coagul. Fibrinolysis* 5, 731–736. doi:10.1097/00001721-199410000-00009
- Argaud, L., Ferry, T., Le, Q. H., Marfisi, A., Ciorba, D., Achache, P., et al. (2007). Short- and long-term outcomes of heatstroke following the 2003 heat wave in Lyon, France. *Arch. Intern. Med.* 167, 2177–2183. doi:10.1001/archinte.167.20.1017
- Asmara, I. G. Y. (2020). Diagnosis and management of heatstroke. *Acta Med. Indones.* 52, 90–97.
- Atha, W. F. (2013). Heat-related illness. *Emerg. Med. Clin. North Am.* 31, 1097–1108. doi:10.1016/j.emc.2013.07.012
- Austin, M. G., and Berry, J. W. (1956). Observations on one hundred cases of heatstroke. *J. Am. Med. Assoc.* 161, 1525–1529. doi:10.1001/jama.1956.02970160005002
- Azevedo, P. S., Minicucci, M. F., Santos, P. P., Paiva, S. A., and Zornoff, L. A. (2013). Energy metabolism in cardiac remodeling and heart failure. *Cardiol. Rev.* 21, 135–140. doi:10.1097/CRD.0b013e318274956d
- Bauer, T. M., and Murphy, E. (2020). Role of mitochondrial calcium and the permeability transition pore in regulating cell death. *Circ. Res.* 126, 280–293. doi:10.1161/circresaha.119.316306
- Bouchama, A., Abuyassin, B., Lehe, C., Laitano, O., Jay, O., O'Connor, F. G., et al. (2022). Classic and exertional heatstroke. *Nat. Rev. Dis. Prim.* 8, 8. doi:10.1038/s41572-021-00334-6
- Bouchama, A., and Knochel, J. P. (2002). Heat stroke. *N. Engl. J. Med.* 346, 1978–1988. doi:10.1056/NEJMra011089
- Bouchama, A., Parhar, R. S., el-Yazigi, A., Sheth, K., and al-Sedairy, S. (1991). Endotoxemia and release of tumor necrosis factor and interleukin 1 alpha in acute heatstroke. *J. Appl. Physiol.* 70, 2640–2644. doi:10.1152/jappl.1991.70.6.2640
- Carling, D., Sanders, M. J., and Woods, A. (2008). The regulation of AMP-activated protein kinase by upstream kinases. *Int. J. Obes. (Lond)* 32 (Suppl. 4), S55–S59. doi:10.1038/ijo.2008.124

- Chang, X., Lochner, A., Wang, H. H., Wang, S., Zhu, H., Ren, J., et al. (2021). Coronary microvascular injury in myocardial infarction: perception and knowledge for mitochondrial quality control. *Theranostics* 11, 6766–6785. doi:10.7150/thno.60143
- Chen, D., Geng, Y., Deng, Z., Li, P., Xue, S., Xu, T., et al. (2023). Inhibition of TLR4 alleviates heat stroke-induced cardiomyocyte injury by down-regulating inflammation and ferroptosis. *Molecules* 28. doi:10.3390/molecules28052297
- Chen, F., Li, H., Zhu, G., Chen, X., and Tang, Z. (2017). Sodium tanshinone IIA sulfonate improves inflammation, aortic endothelial cell apoptosis, disseminated intravascular coagulation and multiple organ damage in a rat heat stroke model. *Mol. Med. Rep.* 16, 87–94. doi:10.3892/mmr.2017.6573
- Chen, G. D., Fan, H., and Zhu, J. H. (2019). Salidroside pretreatment protects against myocardial injury induced by heat stroke in mice. *J. Int. Med. Res.* 47, 5229–5238. doi:10.1177/0300060519868645
- Chen, H. B., Zhang, X. C., Cheng, Y. F., Abdelnasir, A., Tang, S., Kemper, N., et al. (2015). Association of heat shock protein 70 expression with rat myocardial cell damage during heat stress *in vitro* and *in vivo*. *Genet. Mol. Res.* 14, 1994–2005. doi:10.4238/2015.March.20.9
- Chen, K. J., Chen, T. H., Sue, Y. M., Chen, T. J., and Cheng, C. Y. (2014). High-volume plasma exchange in a patient with acute liver failure due to non-exertional heat stroke in a sauna. *J. Clin. Apher.* 29, 281–283. doi:10.1002/jca.21315
- Chen, S. H., Niu, K. C., and Lin, M. T. (2006). Cerebrovascular dysfunction is an attractive target for therapy in heat stroke. *Clin. Exp. Pharmacol. Physiol.* 33, 663–672. doi:10.1111/j.1440-1681.2006.04429.x
- Cramer, M. N., Gagnon, D., Laitano, O., and Crandall, C. G. (2022). Human temperature regulation under heat stress in health, disease, and injury. *Physiol. Rev.* 102, 1907–1989. doi:10.1152/physrev.00047.2021
- Crandall, C. G., and González-Alonso, J. (2010). Cardiovascular function in the heat-stressed human. *Acta Physiol. (Oxf.)* 199, 407–423. doi:10.1111/j.1748-1716.2010.02119.x
- Crandall, C. G., Wilson, T. E., Marving, J., Vogelsang, T. W., Kjaer, A., Hesse, B., et al. (2008). Effects of passive heating on central blood volume and ventricular dimensions in humans. *J. Physiol.* 586, 293–301. doi:10.1113/jphysiol.2007.143057
- Del Re, D. P., Amgalan, D., Linkermann, A., Liu, Q., and Kitsis, R. N. (2019). Fundamental mechanisms of regulated cell death and implications for heart disease. *Physiol. Rev.* 99, 1765–1817. doi:10.1152/physrev.00022.2018
- Deng, C. C., Zhang, J. P., Huo, Y. N., Xue, H. Y., Wang, W., Zhang, J. J., et al. (2022). Melatonin alleviates the heat stress-induced impairment of Sertoli cells by reprogramming glucose metabolism. *J. Pineal Res.* 73, e12819. doi:10.1111/jpi.12819
- Desai, Y., Khraishah, H., and Alahmad, B. (2023). Heat and the heart. *Yale J. Biol. Med.* 96, 197–203. doi:10.59249/hgal4894
- Dörge, H., Neumann, T., Behrends, M., Skyschally, A., Schulz, R., Kasper, C., et al. (2000). Perfusion-contraction mismatch with coronary microvascular obstruction: role of inflammation. *Am. J. Physiol. Heart Circ. Physiol.* 279, H2587–H2592. doi:10.1152/ajpheart.2000.279.6.H2587
- Dörge, H., Schulz, R., Belosjorow, S., Post, H., van de Sand, A., Konietzka, I., et al. (2002). Coronary microembolization: the role of TNF- α in contractile dysfunction. *J. Mol. Cell Cardiol.* 34, 51–62. doi:10.1006/jmcc.2001.1489
- D'Orsi, B., Mateyka, J., and Prehn, J. H. M. (2017). Control of mitochondrial physiology and cell death by the Bcl-2 family proteins Bax and Bok. *Neurochem. Int.* 109, 162–170. doi:10.1016/j.neuint.2017.03.010
- Epstein, Y., and Yanovich, R. (2019). *Heatstroke*. *N. Engl. J. Med.* 380, 2449–2459. doi:10.1056/NEJMr1810762
- Fan, H., Zhao, Y., Zhu, J. H., Song, F. C., Ye, J. H., Wang, Z. Y., et al. (2015). Thrombocytopenia as a predictor of severe acute kidney injury in patients with heat stroke. *Ren. Fail.* 37, 877–881. doi:10.3109/0886022x.2015.1022851
- Fuhrmann, C. M., Sugg, M. M., Konrad, C. E., and Waller, A. (2016). Impact of extreme heat events on emergency department visits in North Carolina (2007–2011). *J. Community Health* 41, 146–156. doi:10.1007/s10900-015-0080-7
- Habets, D. D., Coumans, W. A., El Hasnaoui, M., Zarrinpashneh, E., Bertrand, L., Viollet, B., et al. (2009). Crucial role for LKB1 to AMPK α 2 axis in the regulation of CD36-mediated long-chain fatty acid uptake into cardiomyocytes. *Biochim. Biophys. Acta* 1791, 212–219. doi:10.1016/j.bbalip.2008.12.009
- Hadad, E., Rav-Acha, M., Heled, Y., Epstein, Y., and Moran, D. S. (2004). Heat stroke: a review of cooling methods. *Sports Med.* 34, 501–511. doi:10.2165/00007256-200434080-00002
- Halestrap, A. P. (2009). What is the mitochondrial permeability transition pore? *J. Mol. Cell Cardiol.* 46, 821–831. doi:10.1016/j.jmcc.2009.02.021
- Han, L., Dai, L., Zhao, Y. F., Li, H. Y., Liu, O., Lan, F., et al. (2018). CD40L promotes development of acute aortic dissection via induction of inflammation and impairment of endothelial cell function. *Aging (Albany NY)* 10, 371–385. doi:10.18632/aging.101394
- Harlan, J. M., Vedder, N. B., Winn, R. K., and Rice, C. L. (1991). Mechanisms and consequences of leukocyte-endothelial interaction. *West J. Med.* 155, 365–369.
- Hausfater, P., Doumenc, B., Chopin, S., Le Manach, Y., Santin, A., Dautheville, S., et al. (2010). Elevation of cardiac troponin I during non-exertional heat-related illnesses in the context of a heatwave. *Crit. Care* 14, R99. doi:10.1186/cc9034
- Hsu, S. F., Chao, C. M., Huang, W. T., Lin, M. T., and Cheng, B. C. (2013). Attenuating heat-induced cellular autophagy, apoptosis and damage in H9c2 cardiomyocytes by pre-inducing HSP70 with heat shock preconditioning. *Int. J. Hyperth.* 29, 239–247. doi:10.3109/02656736.2013.777853
- Huang, W., Mao, L., Xie, W., Cai, S., Huang, Q., Liu, Y., et al. (2022). Impact of UCP2 depletion on heat stroke-induced mitochondrial function in human umbilical vein endothelial cells. *Int. J. Hyperth.* 39, 287–296. doi:10.1080/02656736.2022.2032846
- Huang, Z., Zhuang, X., Xie, C., Hu, X., Dong, X., Guo, Y., et al. (2016). Exogenous hydrogen sulfide attenuates high glucose-induced cardiotoxicity by inhibiting NLRP3 inflammasome activation by suppressing TLR4/NF- κ B pathway in H9c2 cells. *Cell Physiol. Biochem.* 40, 1578–1590. doi:10.1159/000453208
- Jiang, X., Stockwell, B. R., and Conrad, M. (2021). Ferroptosis: mechanisms, biology and role in disease. *Nat. Rev. Mol. Cell Biol.* 22, 266–282. doi:10.1038/s41580-020-00324-8
- Keuren, J. F., Baruch, D., Legendre, P., Denis, C. V., Lenting, P. J., Girma, J. P., et al. (2004). von Willebrand factor C1C2 domain is involved in platelet adhesion to polymerized fibrin at high shear rate. *Blood* 103, 1741–1746. doi:10.1182/blood-2003-07-2267
- Ko, W. C., Lin, C. H., Lee, J. J., Chang, C. P., and Chao, C. M. (2020). Therapeutic hypothermia protects against heat stroke-induced arterial hypotension via promoting left ventricular performance in rats. *Int. J. Med. Sci.* 17, 525–535. doi:10.7150/ijms.39745
- Kobayashi, K., Mimuro, S., Sato, T., Kobayashi, A., Kawashima, S., Makino, H., et al. (2018). Dexmedetomidine preserves the endothelial glycocalyx and improves survival in a rat heatstroke model. *J. Anesth.* 32, 880–885. doi:10.1007/s00540-018-2568-7
- Koppula, P., Zhang, Y., Zhuang, L., and Gan, B. (2018). Amino acid transporter SLC7A11/xCT at the crossroads of regulating redox homeostasis and nutrient dependency of cancer. *Cancer Commun. (Lond)* 38, 12. doi:10.1186/s40880-018-0288-x
- Koupenova, M., Kehrel, B. E., Corkrey, H. A., and Freedman, J. E. (2017). Thrombosis and platelets: an update. *Eur. Heart J.* 38, 785–791. doi:10.1093/eurheartj/ehw550
- Laitano, O., Garcia, C. K., Mattingly, A. J., Robinson, G. P., Murray, K. O., King, M. A., et al. (2020). Delayed metabolic dysfunction in myocardium following exertional heat stroke in mice. *J. Physiol.* 598, 967–985. doi:10.1113/jp279310
- Lambert, G. P. (2008). Intestinal barrier dysfunction, endotoxemia, and gastrointestinal symptoms: the 'canary in the coal mine' during exercise-heat stress? *Med. Sport Sci.* 53, 61–73. doi:10.1159/000151550
- Lei, G., Zhang, Y., Hong, T., Zhang, X., Liu, X., Mao, C., et al. (2021). Ferroptosis as a mechanism to mediate p53 function in tumor radiosensitivity. *Oncogene* 40, 3533–3547. doi:10.1038/s41388-021-01790-w
- Leon, L. R., and Helwig, B. G. (2010). Heat stroke: role of the systemic inflammatory response. *J. Appl. Physiol.* 109, 1980–1988. doi:10.1152/japplphysiol.00301.2010
- Lin, X., Lin, C. H., Liu, R., Li, C., Jiao, S., Yi, X., et al. (2020). Myricetin against myocardial injury in rat heat stroke model. *Biomed. Pharmacother.* 127, 110194. doi:10.1016/j.biopha.2020.110194
- Lin, X., Lin, C. H., Zhao, T., Zuo, D., Ye, Z., Liu, L., et al. (2017). Quercetin protects against heat stroke-induced myocardial injury in male rats: antioxidative and antiinflammatory mechanisms. *Chem. Biol. Interact.* 265, 47–54. doi:10.1016/j.cbi.2017.01.006
- Lin, X., Zhao, T., Lin, C. H., Zuo, D., Ye, Z., Lin, S., et al. (2018). Melatonin provides protection against heat stroke-induced myocardial injury in male rats. *J. Pharm. Pharmacol.* 70, 760–767. doi:10.1111/jphp.12895
- Liu, J., Varghese, B. M., Hansen, A., Zhang, Y., Driscoll, T., Morgan, G., et al. (2022). Heat exposure and cardiovascular health outcomes: a systematic review and meta-analysis. *Lancet Planet Health* 6, e484–e495. doi:10.1016/s2542-5196(22)00117-6
- Liu, X., Wang, N., Fan, S., Zheng, X., Yang, Y., Zhu, Y., et al. (2016). The citrus flavonoid naringenin confers protection in a murine endotoxaemia model through AMPK-ATF3-dependent negative regulation of the TLR4 signalling pathway. *Sci. Rep.* 6, 39735. doi:10.1038/srep39735
- Lou, Y., Lin, H., Wang, H., Chen, W., Wu, Y., and Li, H. (2019). Research progress in the heatstroke-induced myocardial injury. *Zhonghua Wei Zhong Bing Ji Jiu Yi Xue* 31, 1304–1306. doi:10.3760/cma.j.issn.2095-4352.2019.10.025
- Low, D. A., Keller, D. M., Wingo, J. E., Brothers, R. M., and Crandall, C. G. (2011). Sympathetic nerve activity and whole body heat stress in humans. *J. Appl. Physiol.* 111, 1329–1334. doi:10.1152/japplphysiol.00498.2011
- Ma, X. H., Liu, J. H., Liu, C. Y., Sun, W. Y., Duan, W. J., Wang, G., et al. (2022). ALOX15-launched PUFA-phospholipids peroxidation increases the susceptibility of ferroptosis in ischemia-induced myocardial damage. *Signal Transduct. Target Ther.* 7, 288. doi:10.1038/s41392-022-01090-z
- Marchand, M., and Gin, K. (2022). The cardiovascular system in heat stroke. *CJC Open* 4, 158–163. doi:10.1016/j.cjco.2021.10.002

- McCormack, J. G., and Denton, R. M. (1989). The role of Ca²⁺ ions in the regulation of intramitochondrial metabolism and energy production in rat heart. *Mol. Cell Biochem.* 89, 121–125.
- Nakamura, T., Ueda, Y., Juan, Y., Katsuda, S., Takahashi, H., and Koh, E. (2000). Fas-mediated apoptosis in adriamycin-induced cardiomyopathy in rats: *in vivo* study. *Circulation* 102, 572–578. doi:10.1161/01.cir.102.5.572
- Patra, K. C., and Hay, N. (2014). The pentose phosphate pathway and cancer. *Trends Biochem. Sci.* 39, 347–354. doi:10.1016/j.tibs.2014.06.005
- Peiris, A. N., Jaroudi, S., and Noor, R. (2017). Heat stroke. *Jama* 318, 2503. doi:10.1001/jama.2017.18780
- Petit, P. X., Goubern, M., Diolet, P., Susin, S. A., Zamzami, N., and Kroemer, G. (1998). Disruption of the outer mitochondrial membrane as a result of large amplitude swelling: the impact of irreversible permeability transition. *FEBS Lett.* 426, 111–116. doi:10.1016/s0014-5793(98)00318-4
- Piver, W. T., Ando, M., Ye, F., and Portier, C. J. (1999). Temperature and air pollution as risk factors for heat stroke in Tokyo. *Environ. Health Perspect.* 107, 911–916. doi:10.1289/ehp.99107911
- Qian, L., Song, X., Ren, H., Gong, J., and Cheng, S. (2004). Mitochondrial mechanism of heat stress-induced injury in rat cardiomyocyte. *Cell Stress Chaperones* 9, 281–293. doi:10.1379/csc-20r.1
- Roberts, G. T., Ghebeh, H., Chishti, M. A., Al-Mohanna, F., El-Sayed, R., Al-Mohanna, F., et al. (2008). Microvascular injury, thrombosis, inflammation, and apoptosis in the pathogenesis of heatstroke: a study in baboon model. *Arterioscler. Thromb. Vasc. Biol.* 28, 1130–1136. doi:10.1161/atvbaha.107.158709
- Rodríguez, C., Muñoz, M., Contreras, C., and Prieto, D. (2021). AMPK, metabolism, and vascular function. *Febs J.* 288, 3746–3771. doi:10.1111/febs.15863
- Roths, M., Freestone, A. D., Rudolph, T. E., Michael, A., Baumgard, L. H., and Selsby, J. T. (2023). Environment-induced heat stress causes structural and biochemical changes in the heart. *J. Therm. Biol.* 113, 103492. doi:10.1016/j.jtherbio.2023.103492
- Russo, M., Della Sala, A., Tocchetti, C. G., Porporato, P. E., and Ghigo, A. (2021). Metabolic aspects of anthracycline cardiotoxicity. *Curr. Treat. Options Oncol.* 22, 18. doi:10.1007/s11864-020-00812-1
- Shen, H. H., Tseng, Y. S., Kuo, N. C., Kung, C. W., Amin, S., Lam, K. K., et al. (2019). Alpha-lipoic acid protects cardiomyocytes against heat stroke-induced apoptosis and inflammatory responses associated with the induction of Hsp70 and activation of autophagy. *Mediat. Inflamm.* 2019, 8187529. doi:10.1155/2019/8187529
- Sohal, R. S., Sun, S. C., Colcolough, H. L., and Burch, G. E. (1968). Heat stroke. An electron microscopic study of endothelial cell damage and disseminated intravascular coagulation. *Arch. Intern. Med.* 122, 43–47. doi:10.1001/archinte.122.1.43
- Stockwell, B. R., Jiang, X., and Gu, W. (2020). Emerging mechanisms and disease relevance of ferroptosis. *Trends Cell Biol.* 30, 478–490. doi:10.1016/j.tcb.2020.02.009
- Tang, S., Buriro, R., Liu, Z., Zhang, M., Ali, I., Adam, A., et al. (2013). Localization and expression of Hsp27 and α B-crystallin in rat primary myocardial cells during heat stress *in vitro*. *PLOS ONE* 8, e69066. doi:10.1371/journal.pone.0069066
- Tang, S., Chen, H., Cheng, Y., Nasir, M. A., Kemper, N., and Bao, E. (2016). The interactive association between heat shock factor 1 and heat shock proteins in primary myocardial cells subjected to heat stress. *Int. J. Mol. Med.* 37, 56–62. doi:10.3892/ijmm.2015.2414
- Tek, D., and Olshaker, J. S. (1992). Heat illness. *Emerg. Med. Clin. North Am.* 10, 299–310.
- Tian, H., Zhao, X., Zhang, Y., and Xia, Z. (2023). Abnormalities of glucose and lipid metabolism in myocardial ischemia-reperfusion injury. *Biomed. Pharmacother.* 163, 114827. doi:10.1016/j.biopha.2023.114827
- Tian, Y. F., Lin, C. H., Hsu, S. F., and Lin, M. T. (2013). Melatonin improves outcomes of heatstroke in mice by reducing brain inflammation and oxidative damage and multiple organ dysfunction. *Mediat. Inflamm.* 2013, 349280. doi:10.1155/2013/349280
- Toutant, S., Gosselin, P., Bélanger, D., Bustanza, R., and Rivest, S. (2011). An open source web application for the surveillance and prevention of the impacts on public health of extreme meteorological events: the SUPREME system. *Int. J. Health Geogr.* 10, 39. doi:10.1186/1476-072x-10-39
- Tseng, M. F., Chou, C. L., Chung, C. H., Chien, W. C., Chen, Y. K., Yang, H. C., et al. (2019). Association between heat stroke and ischemic heart disease: a national longitudinal cohort study in Taiwan. *Eur. J. Intern. Med.* 59, 97–103. doi:10.1016/j.ejim.2018.09.019
- Urbich, C., Dernbach, E., Aicher, A., Zeiher, A. M., and Dimmeler, S. (2002). CD40 ligand inhibits endothelial cell migration by increasing production of endothelial reactive oxygen species. *Circulation* 106, 981–986. doi:10.1161/01.cir.0000027107.54614.1a
- Valls-Lacalle, L., Barba, I., Miró-Casas, E., Ruiz-Meana, M., Rodríguez-Sinovas, A., and García-Dorado, D. (2018). Selective inhibition of succinate dehydrogenase in reperfused myocardium with intracoronary malonate reduces infarct size. *Sci. Rep.* 8, 2442. doi:10.1038/s41598-018-20866-4
- Vassalle, M., and Lin, C. I. (2004). Calcium overload and cardiac function. *J. Biomed. Sci.* 11, 542–565. doi:10.1007/bf02256119
- Wakino, S., Hori, S., Mimura, T., Fujishima, S., Hayashi, K., Inamoto, H., et al. (2005). Heat stroke with multiple organ failure treated with cold hemodialysis and cold continuous hemodiafiltration: a case report. *Ther. Apher. Dial.* 9, 423–428. doi:10.1111/j.1744-9987.2005.00321.x
- Walkon, L. L., Strubbe-Rivera, J. O., and Bazil, J. N. (2022). Calcium overload and mitochondrial metabolism. *Biomolecules* 12. doi:10.3390/biom12121891
- Wang, J. C., Chien, W. C., Chu, P., Chung, C. H., Lin, C. Y., and Tsai, S. H. (2019). The association between heat stroke and subsequent cardiovascular diseases. *PLOS ONE* 14, e0211386. doi:10.1371/journal.pone.0211386
- Wang, X., Huang, Y., Zhang, K., Chen, F., Nie, T., Zhao, Y., et al. (2023). Changes of energy metabolism in failing heart and its regulation by SIRT3. *Heart Fail Rev.* 28, 977–992. doi:10.1007/s10741-023-10295-5
- Wihastuti, T. A., Aini, F. N., Lutfiana, N. C., Heriansyah, T., and Zamrudah, N. (2018). Exploration of adhesion molecule expression in cardiac muscle of early atherosclerosis dyslipidemic sprague dawley rats. *Open Med. Chem. J.* 12, 124–129. doi:10.2174/1874104501812010124
- Wu, D., Zhang, M., Xu, J., Song, E., Lv, Y., Tang, S., et al. (2016). *In vitro* evaluation of aspirin-induced HspB1 against heat stress damage in chicken myocardial cells. *Cell Stress Chaperones* 21, 405–413. doi:10.1007/s12192-016-0666-8
- Xu, C., Sun, S., Johnson, T., Qi, R., Zhang, S., Zhang, J., et al. (2021). The glutathione peroxidase Gpx4 prevents lipid peroxidation and ferroptosis to sustain Treg cell activation and suppression of antitumor immunity. *Cell Rep.* 35, 109235. doi:10.1016/j.celrep.2021.109235
- Yang, P. C., He, S. H., and Zheng, P. Y. (2007). Investigation into the signal transduction pathway via which heat stress impairs intestinal epithelial barrier function. *J. Gastroenterol. Hepatol.* 22, 1823–1831. doi:10.1111/j.1440-1746.2006.04710.x
- Yao, D., Dong, M., Dai, C., and Wu, S. (2019). Inflammation and inflammatory cytokine contribute to the initiation and development of ulcerative colitis and its associated cancer. *Inflamm. Bowel Dis.* 25, 1595–1602. doi:10.1093/ibd/izz149
- Yao, H., Xie, Q., He, Q., Zeng, L., Long, J., Gong, Y., et al. (2022). Pretreatment with panaxatriol saponin attenuates mitochondrial apoptosis and oxidative stress to facilitate treatment of myocardial ischemia-reperfusion injury via the regulation of keap1/nrf2 activity. *Oxid. Med. Cell Longev.* 2022, 9626703. doi:10.1155/2022/9626703
- Yu, M., Wen, N., Wenzhong, Z., Yuanchang, X., Xiaoming, D., and Yongjin, L. (2010). Effect of repeated ischaemic preconditioning on TLR4 and proinflammatory cytokines TNF- α and IL-1 β in myocardial ischaemia-reperfusion injury in a rat model. *Arch. Med. Sci.* 6, 843–847. doi:10.5114/aoms.2010.19289
- Zamzami, N., Hirsch, T., Dallaporta, B., Petit, P. X., and Kroemer, G. (1997). Mitochondrial implication in accidental and programmed cell death: apoptosis and necrosis. *J. Bioenerg. Biomembr.* 29, 185–193. doi:10.1023/a:1022694131572
- Zhang, W., Qiao, W., and Zuo, L. (2022). A(1) and A(2b) adenosine receptors regulate GPX4 against ferroptosis of cardiomyocytes in myocardial infarction rat model and *in vitro*. *Tissue Cell* 77, 101828. doi:10.1016/j.tice.2022.101828
- Zhang, X., Chen, B., Wu, J., Sha, J., Yang, B., Zhu, J., et al. (2020). Aspirin enhances the protection of Hsp90 from heat-stressed injury in cardiac microvascular endothelial cells through PI3K-Akt and PKM2 pathways. *Cells* 9. doi:10.3390/cells9010243
- Zhang, X., Du, Q., Yang, Y., Wang, J., Dou, S., Liu, C., et al. (2017). The protective effect of Luteolin on myocardial ischemia/reperfusion (I/R) injury through TLR4/NF- κ B/NLRP3 inflammasome pathway. *Biomed. Pharmacother.* 91, 1042–1052. doi:10.1016/j.biopha.2017.05.033
- Zhang, Z. T., Gu, X. L., Zhao, X., He, X., Shi, H. W., Zhang, K., et al. (2021). NLRP3 ablation enhances tolerance in heat stroke pathology by inhibiting IL-1 β -mediated neuroinflammation. *J. Neuroinflammation* 18, 128. doi:10.1186/s12974-021-02179-y
- Zheng, D., MacLean, P. S., Pohnert, S. C., Knight, J. B., Olson, A. L., Winder, W. W., et al. (2001). Regulation of muscle GLUT-4 transcription by AMP-activated protein kinase. *J. Appl. Physiol.* 91, 1073–1083. doi:10.1152/jappl.2001.91.3.1073
- Zheng, M., Zhang, J., Shi, L., Zhang, D., Pangali Sharma, T. P., and Prodhan, F. A. (2020). Mapping heat-related risks in northern Jiangxi province of China based on two spatial assessment frameworks approaches. *Int. J. Environ. Res. Public Health* 17. doi:10.3390/ijerph17186584
- Zhou, X., Wu, Y., Ye, L., Wang, Y., Zhang, K., Wang, L., et al. (2019). Aspirin alleviates endothelial gap junction dysfunction through inhibition of NLRP3 inflammasome activation in LPS-induced vascular injury. *Acta Pharm. Sin. B* 9, 711–723. doi:10.1016/j.apsb.2019.02.008
- Zhu, B. S., Xing, C. G., Lin, F., Fan, X. Q., Zhao, K., and Qin, Z. H. (2011). Blocking NF- κ B nuclear translocation leads to p53-related autophagy activation and cell apoptosis. *World J. Gastroenterol.* 17, 478–487. doi:10.3748/wjg.v17.i4.478
- Zuurbier, C. J., Bertrand, L., Beauloye, C. R., Andreadou, I., Ruiz-Meana, M., Jespersen, N. R., et al. (2020). Cardiac metabolism as a driver and therapeutic target of myocardial infarction. *J. Cell Mol. Med.* 24, 5937–5954. doi:10.1111/jcmm.15180



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Microvascular rarefaction caused by the NOTCH signaling pathway is a key cause of TKI-apatinib-induced hypertension and cardiac damage

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With the advancement of tumour-targeted therapy technology, the survival of cancer patients has continued to increase, and cardiovascular events have gradually become an important cause of death in cancer patients. This phenomenon occurs due to adverse cardiovascular reactions caused by the cardiovascular toxicity of antitumour therapy. Moreover, the increase in the proportion of elderly patients with cancer and cardiovascular diseases is due to the extension of life expectancy. Hypertension is the most common cardiovascular side effect of small molecule tyrosine kinase inhibitors (TKIs). The increase in blood pressure induced by TKIs and subsequent cardiovascular complications and events affect the survival and quality of life of patients and partly offset the benefits of antitumour therapy. Many studies have confirmed that in the pathogenesis of hypertension, arterioles and capillary thinness are involved in its occurrence and development. Our previous findings showing that apatinib causes microcirculation rarefaction of the superior mesenteric artery and impaired microvascular growth may inspire new therapeutic strategies for treating hypertension. Thus, by restoring microvascular development and branching patterns, total peripheral resistance and blood pressure are reduced. Therefore, exploring the key molecular targets of TKIs that inhibit the expression of angiogenic factors and elucidating the specific molecular mechanism involved are key scientific avenues for effectively promoting endothelial cell angiogenesis and achieving accurate repair of microcirculation injury in hypertension patients.

KEYWORDS

hypertension, microcirculation, rarefaction, TKIs, apatinib

1 Introduction

Hypertension has a high prevalence in China and accounts for a large proportion of cardiovascular diseases. Hypertension is also a major risk factor for cardiovascular-related diseases. In recent years, a number of studies have reported a relationship between tumours and hypertension. Tumours and hypertension have common risk factors and overlapping pathophysiological mechanisms (Milan et al., 2014; Bray et al., 2021; Dolmatova et al.,

TABLE 1 Incidence of hypertension caused by TKIs.

| Drug | Category | Target spot | Hypertension (%) |
|--------------|----------|---|------------------|
| axitinib | TKIs | VEGFR1-3,c-KIT,PDGFR | 22–84 |
| cabozantinib | TKIs | MET,VEGFR2,RET,AXL,FLT3 | 28–61 |
| Lenvatinib | TKIs | VEGFR1-3, FGFR 1–4,PDGFR, c-KIT,RET | 42–73 |
| Pazopanib | TKIs | VEGFR1-3,PDGFR, FGFR,c-KIT | 40–42 |
| Ponatinib | TKIs | BCR-ABL,VEGFR, PDGFR,FGFR,EPH,c-KIT,RET,TIE2,FLT3 | 53–74 |
| regorafenib | TKIs | VEGFR1-3,PDGFR,c-KIT,RET,RAF-1 | 28–67 |
| Sorafenib | TKIs | VEGFR1-3,PDGFR,c-KIT,RET,RAF-1 | 4–31 |
| sunitinib | TKIs | VEGFR2,PDGFR,c-KIT | 20–27 |
| Wandtaneb | TKIs | VEGFR2,PDGFR,c-KIT | 4–40 |

2023). Therefore, several experts have gradually formed the theory of onco-hypertension. Previous studies by our group have shown that hypertension and antihypertensive drug use are closely related to breast cancer. With increasing age, the incidence of hypertension in breast cancer patients increases, and the mechanism is related to inflammatory mediators and angiogenesis (Zhao et al., 2018; Wang et al., 2021). Another reason is that women's estrogen levels decline as they age. Estrogen has a protective effect on blood vessels, so such patients are more likely to induce hypertension. And estrogen for breast cancer patients, with the increase of age, although estrogen in the body decreases, but some patients may be converted to androgen, leading to the increase in the incidence of breast cancer. In recent years, new antineoplastic drugs have prolonged the survival of cancer patients, but the increase in blood pressure caused by new antineoplastic drugs and the subsequent cardiovascular complications and events affect the survival and quality of life of patients, which partly offset the benefits of antineoplastic therapy. It is worth exploring this topic further.

2 Incidence of hypertension caused by TKIs

VEGF signalling pathway inhibitors include monoclonal antibodies against VEGF A factor, VEGF traps, monoclonal antibodies against VEGF receptors, and TKIs (Pucci et al., 2019; Le et al., 2021; Lawler, 2022). Studies have shown that the probability of hypertension caused by VEGF signalling pathway inhibitors during antitumour therapy is approximately 11%–45%, among which the incidence of hypertension above grade 3 (referring to CTCAE grade 3–4, that is, SBP \geq 160 and/or DBP \geq 100 mmHg) or life-threatening hypertension, even requiring emergency treatment, ranges from 2 to 20% (Xu et al., 2021). According to the type and dose of TKIs, the incidence of hypertension can reach 20%–90%, and the incidence of severe hypertension can reach 6%–43% in patients using TKIs alone or in combination. The incidence of hypertension induced by TKIs is shown in Table 1.

Apatinib mesylate is an antitumour drug that was independently developed in China and belongs to the class of TKIs. In the past, it was mainly used for the treatment of solid tumours, such as

advanced gastric cancer or gastroesophageal junction adenocarcinoma (Li et al., 2016). An international multicentre phase III study (SHR-1210-III.-310 study) demonstrated that the approved PD-1 inhibitor camrelizumab in combination with apatinib had a significant survival benefit and a tolerable safety profile in the first-line treatment of advanced liver cancer, with a median overall survival (OS) of 22.1 months. It is the combination therapy with the longest OS benefit in the first-line treatment of advanced liver cancer (Qin et al., 2023). Therefore, the National Medical Products Administration (NMPA) proposed the combination of “Shuang'ai” for the first-line treatment of advanced hepatocellular carcinoma, which is the first approval in China to use a combination of PD-1 inhibitors and TKIs for the treatment of advanced hepatocellular carcinoma. The application of this treatment regimen has led to new drug options for patients with advanced liver cancer. The resulting cardiovascular side effects of nitrogen are also very noteworthy. Therefore, studying the cardiovascular toxicity and side effects caused by TKIs is highly important for guiding antitumour therapy in clinical cancer patients.

3 Causal relationship between microcirculation damage and hypertension

3.1 Microcirculation and related definitions of microcirculation damage

Microcirculation typically includes small resistance arteries (300–100 μ m in diameter), precapillary arterioles (100–10 μ m), capillaries (5–15 μ m) and venules (10–100 μ m) (le Noble et al., 2023). Various studies have shown that damage to microcirculatory tissue may involve multiple mechanisms. The main categories of such damage include impaired endothelial cell function, oxidative stress, decreased angiogenesis (most commonly in patients after the use of targeted drugs), increased endothelial permeability, enhanced leukocyte adhesion, immune cell activation, lymphatic dysfunction, impaired autoregulation, microvascular constriction, and microcirculation obstruction (Sorop et al., 2020; Sabe et al., 2022; De Ciuceis et al., 2023; Mengozzi et al., 2023).

3.2 Microcirculation and types of microcirculation damage

Studies have shown a relationship between hypertension and changes in the microvascular network in spontaneously hypertensive rats (SHRs), with reduced arteriolar density and increased postcapillary venule density (Martens and Gelband, 1998). It lead to increased postvascular resistance, which may further contribute to the development of hypertension. In addition, it has been shown that a sparse microvascular network structure increases total peripheral resistance, which eventually leads to increased blood pressure (le Noble et al., 2023). Many previous studies have confirmed the importance of arteriolar and capillary scarcity in the pathogenesis of hypertension (Levy et al., 2001; Levy et al., 2008). That is, the rarefaction of capillaries may contribute to the development of hypertension.

Microcirculation injury can occur through two main forms of microvascular rarefaction (Agabiti-Rosei, 2003; Ungvari et al., 2021). Functional rarefaction means that the total number of anatomically present vessels is not reduced but rather that perfusion of this part of the microvascular network is absent. However, as the vascular tone continues to increase, the lumen area increases, resulting in a decrease in the number of blood vessels perfused. Structural sparseness also occurs, which reduces the number of blood vessels that can be found during tissue dissection. This reduction in vascular mesh may be due to altered anatomical changes in the vascular segments or other impairments in the vascular network during the growth and development process during early tissue development. Several experts have also noted that functional rarefaction can progress to structural rarefaction in a rat model of SHR. However, there are also studies showing that patients with essential hypertension, recruitment of perfused capillaries is impaired, which can be explained by both functional and structural rarefaction (Serné et al., 2001). Therefore, whether the microcirculation dilution in hypertension is structurally sparse or functional, or even both, will be our further research plan. This part of the study is crucial for them to identify the specific microcirculation sparse so that we can improve and treat it according to the cause, especially in patients with hypertension caused by targeted drugs.

3.3 Mechanisms of microcirculation damage leading to hypertension

Microvascular injury plays an indispensable role in the occurrence and development of hypertension, and some studies have shown that it plays an important role in the progression of hypertension, hypertension-mediated organ damage and related cardiovascular events (Jonk et al., 2007; Karaca et al., 2014; Rizzoni et al., 2023). However, the causal relationship between thinning of microvessels and hypertension is difficult to explain, and some experts speculate that diffuse generalized thinness of microvessels may be one of the main causes of hypertension (Marinescu et al., 2015; Horton and Barrett, 2021). In study, after 5 weeks of treatment with telatinib, the capillary density decreased from 20.8 at baseline to 16.7 (Steehgs et al., 2008). Mourad et al. reported a significant reduction in kinetic and structural capillary density in a group of patients with metastatic colon

cancer treated with bevacizumab for 6 months (Mourad et al., 2008). However, this rarefaction is most likely functional because blood pressure increases rapidly after the start of treatment and returns to normal immediately after the discontinuation of VEGF inhibitors (van Dorst et al., 2021). Therefore, further studies to determine whether microvascular thinning is the cause or effect of the actions of VEGF inhibitors on hypertension are highly clinically important.

Animal experiments revealed that a rise in blood pressure leads to an increase in the production of reactive oxygen species in mice. Therefore, researchers have speculated that elevated blood pressure may be responsible for microvascular function and structural alterations, further contributing to the manifestation of vascular thinning (Ungvari et al., 2004; Jacobson et al., 2007). However, there is considerable evidence that microvascular changes may also be a cause rather than a consequence of hypertension. In animal models of hypertension, increased reactive oxygen species production and sparse arterioles occur even in the vasculature not exposed to hypertension (Boegehold et al., 1991). Our previous results showed that blood pressure was significantly increased in an apatinib-treated mouse model of gastric cancer, and sparsity of arteriolar vessels was detected in the mesenteric arteries of experimental model mice. Therefore, we speculate that the microcirculation damage caused by apatinib during antitumour treatment may be one of the main causes of hypertension. However, this study did not further confirm whether functional or structural vascular alterations are the cause, and it is very important to clarify this classification for the treatment of hypertension induced by TKIs. Because we did not further lower the blood pressure after the elevation of blood pressure, we investigated whether the vascular density of the superior mesenteric artery in the mice was further restored.

In addition, microvascular rarefaction is observed in the early stages of hypertension development, and microvascular rarefaction can be detected in individuals with a family history of hypertension, even if their blood pressure is normal. Similarly, in animal experiments, microvascular abnormalities occur early in the development of hypertension in SHRs (Antonios et al., 2003). In addition, the latest results from our research indicated that the administration of the ROCK inhibitor Y27632 can prevent microvascular rarefaction caused by apatinib and improve blood pressure (Wang et al., 2022a). Thus, microvascular growth disorders may have given rise to a new paradigm of hypertension treatment aimed at restoring microvascular development and branching, thereby further reducing peripheral resistance and improving the structural reduction of arterial blood pressure.

4 Relationship between microcirculation-induced hypertension and heart failure

Hypertension is an important risk factor for heart failure (Garg et al., 2021). Long-term uncontrolled hypertension can exert excessive pressure on the heart and gradually impair heart function, leading to cardiac hypertrophy and decreased myocardial contractility. Over time, this overload can impede the ability of the heart to pump blood effectively, triggering heart failure (Slivnick and Lampert, 2019;

Jackson et al., 2021; Redfield and Borlaug, 2023). The most common type of heart failure is heart failure with preserved ejection fraction (HFpEF). There are multiple mechanisms involved in the development of hypertension from HFpEF, and these mechanisms can be divided into several categories. The most common is microcirculatory dysfunction, which mainly includes an increase in inflammatory factors, the occurrence of oxidative stress, impaired endothelial function, and the occurrence of vascular endothelial fibrosis (Gryglewska et al., 2011; Paulus and Tschöpe, 2013; Stupin et al., 2021; Marra et al., 2022).

Firstly, left atrial pressure increases in hypertensive patients, with a consequent increase in left ventricular end-diastolic pressure, and left atrial remodeling occurs. This process is an indication of left ventricular diastolic dysfunction. Left atrial enlargement and dysfunction, including reduced left atrial systolic reserve, can eventually lead to the development of HFpEF (Nagai et al., 2023; Li et al., 2024). Secondly, in patients with chronic systemic inflammation such as hyperemia, systemic secreted inflammatory cytokines can cause the accumulation and inflammatory response of epicardial adipose tissue, which can promote the migration and transformation of mesenchymal stem cells and local secretion of inflammatory cytokines, leading to deep myocardial cell inflammation, increased myocardial stiffness, deep myocardial fibrosis, and finally HFpEF (Obokata et al., 2017; Gevaert et al., 2022; Gao et al., 2023; Rossi et al., 2023). In addition, microcirculation disorders also play an important role in the occurrence and development of HFpEF (Camici et al., 2020). Systemic inflammation can lead to microvascular inflammation and endothelial activation, resulting in significant structural and functional changes in cardiomyocytes and extracellular matrix, including the decrease of NO and vasodilator peptide levels, which leads to the enhancement of vasoconstriction, left ventricular stiffness and myocardial collagen-ization, and ultimately the formation of HFpEF (Vancheri et al., 2020; Agrawal et al., 2023). Overall, in most patients with hypertension, left ventricular diastolic dysfunction is the first apparent manifestation of heart failure. Our previous study (Wang et al., 2022a) showed that apatinib increased blood pressure in gastric cancer model mice, which further led to left ventricular hypertrophy and fibrosis. Therefore, exploring the mechanism through which apatinib-induced hypertension leads to HFpEF in gastric cancer mice is highly important, as is exploring drugs that can delay the progression of HFpEF.

Abnormal structure and function of miniscule arteries and capillaries results in hypoperfusion of blood flow (Mourad and Levy, 2011; Struijker-Boudier and Heijnen, 2011; Wu et al., 2023). Studies have shown that sparse microcirculation may be one of the main causes of hypertension (Ciuffetti et al., 2003). Chronic high blood pressure damages the heart and blood vessels. Pathological changes in miniscule arteries and capillaries, such as intimal thickening, fibrosis, and luminal narrowing, further inhibit blood flow perfusion (Triantafyllou et al., 2015). This microcirculatory pathology affects normal oxygen availability and nutrient delivery to the heart and other tissues (De Backer et al., 2014; Cusack et al., 2022). Ischaemia and hypoxia lead to cardiac cell damage and death, which eventually leads to weakened cardiac systolic function and the occurrence of heart failure (Premont et al., 2020). Therefore, microcirculatory impairment plays a key role in hypertension and cardiac dysfunction. Therefore, protecting

microcirculation function is highly important for the prevention and treatment of hypertensive heart failure.

5 Mechanisms related to hypertension caused by TKIs

5.1 Relationship between VEGF signaling pathway and TKI-induced hypertension

The most common pathways are the vascular endothelial growth factor (VEGF), platelet-derived growth factor (PDGF), angiotensin-1, and Notch signalling pathways (Viallard and Larrivée, 2017; Vimalraj, 2022; Huang et al., 2023) and so on. VEGF is one of the most important proangiogenic mediators. Recently, studies have shown that VEGF is the core factor affecting endothelial cell angiogenesis and is closely related to hypertensive microcirculation damage (Chade and Kelsen, 2010). However, the foundation of TKI antitumor therapy is the inhibition of vascular endothelial growth factor, which brings certain challenges to antitumor therapy. The latest research from our team revealed that in a mouse model of gastric cancer, the mechanism of apatinib-induced hypertension in mice may be related to the sparse vascular density of the superior mesenteric artery (Wang et al., 2022a).

VEGF signaling pathway inhibitors include monoclonal antibodies against VEGFA, vascular endothelial growth factor inhibitors (VEGF trap), monoclonal antibodies against VEGF receptors, and TKIs (Saif, 2013; Shughoury et al., 2023). TKIs are effective signaling cascade inhibitors that inhibit tumor blood vessel growth by inhibiting vascular endothelial growth factor receptor (VEGFR) (Vano et al., 2022). VEGFR-TKIs have become the main treatment for many solid malignant tumors. However, TKIs can induce vascular endothelial damage, hypertension and myocardial injury by targeting VEGFR, platelet-derived growth factor receptor (PDGFR) and stem cell factor receptor (SCFR) (van Crujisen et al., 2009; Lennartsson and Rönnstrand, 2012). It can also damage mitochondria and affect myocardial energy metabolism through the "off-target effect", eventually leading to cardiovascular complications (Tullemans et al., 2018; Rodríguez-Agustín et al., 2023). Therefore, the incidence of cardiovascular toxicity related to VEGFR-TKIs is high, which can cause the occurrence and development of cardiovascular complications such as hypertension, left ventricular systolic dysfunction/heart failure, and atherosclerosis.

Anti-angiogenesis targeting drugs mainly act on VEGF and VEGFR. VEGFR-1, VEGFR-2 and VEGFR-3 are the major VEGF receptors (Sallinen et al., 2009; Bernatz et al., 2021). VEGF inhibitors can increase the risk of heart failure, coronary heart disease, hypertension and thromboembolic diseases through endothelial injury, vasoconstriction and remodeling, inflammatory response and platelet activation.

5.2 Relationship between RhoA/ROCK signaling pathway and TKIs-induced hypertension

In addition, the latest study from our team revealed that apatinib has a considerable therapeutic effect on a mouse model of gastric

cancer (Wang et al., 2022a). However, apatinib can also lead to an increase in blood pressure, accompanied by activation of the RhoA/ROCK signalling pathway. And the ratio between vessel thickness and lumen diameter was significantly increased in the apatinib group. We also observed that apatinib in combination with the ROCK inhibitor Y27632 did not affect the antitumour therapeutic effect of apatinib. Finally, our combined administration of ROCK inhibitors significantly reduced the increase in blood pressure in an apatinib-induced gastric cancer mouse model (Wang et al., 2022a). Therefore, Y27632 has wide application prospects for the treatment of tumour-induced hypertension.

Further studies at the cellular level have shown that knocking down the key gene LARG in the RhoA/ROCK signalling pathway can improve the abnormal proliferation and impaired cell function of vascular smooth muscle cells caused by apatinib (Wang et al., 2022b). This is mainly because apatinib can increase blood pressure by activating the RhoA/ROCK signaling pathway. Therefore, exploring the key molecular targets of TKIs that inhibit the expression of angiogenic factors and elucidating the specific molecular mechanism involved are key for effectively promoting endothelial cell angiogenesis and achieving accurate repair of microcirculation injury in hypertension patients.

5.3 Relationship between notch signaling pathway and TKIs-induced hypertension

Notch and DLL4 are specifically expressed on vascular endothelial cells (EC). JAG has been shown to promote cell survival and proliferation, interacting with NOTCH and hematopoietic stem and progenitor cells (HSPCs) (Kangsamaksin et al., 2015). In addition, high expression of JAG promotes cancer development. We hypothesized that inhibition of Notch signaling activation would not only inhibit tumor development, but also help angiogenesis. Therefore, exploring the mechanism of angiogenesis dysfunction induced during antitumour therapy is highly clinically important.

Notch signalling pathway, as a classical signaling pathway, is closely related to cardiovascular and tumor microcirculation. Combined with our previous results, Notch signaling pathway may also be involved in TKI-induced blood pressure elevation. The regulation of the Notch signalling pathway is closely related to the proliferation, migration and tube formation of vascular endothelial cells during tumour angiogenesis. Further research on the relationship between the Notch signalling pathway and tumour-related cardiovascular disease is expected to lead to new therapeutic strategies and targets for the prevention and treatment of tumours.

5.4 Other mechanisms associated with TKIs-induced hypertension

At present, there are many studies on hypertension caused by TKIs, and the relevant mechanisms involved may be related to the following aspects: 1) decreased bioavailability of nitric oxide (Robinson et al., 2010); 2) enhanced oxidative stress (Neves et al., 2018); 3) increased secretion of endothelin-1 (Mirabito Colafella et al., 2020); 4) decreased bioavailability of prostacyclin (Wheeler-Jones et al., 1997); 5) increased bioavailability of endothelial

microparticles (Neves et al., 2019); 6) sparse microvessels (Steeghs et al., 2008); 7) vascular sclerosis (Catino et al., 2018); 8) activation of renin angiotensin (Li et al., 2022); and 9) salt-sensitive hypertension (Tsai et al., 2017).

6 Role of the notch signalling pathway in the cardiovascular system

The Notch signalling pathway is a common signalling pathway that plays a certain role in cell development. The Notch protein is a transmembrane receptor that is located on the cell surface and mediates important cellular functions (Kwak et al., 2022; Takahashi et al., 2023). The interaction between the Notch protein and its ligand initiates a signalling cascade associated with cell fate that plays a key role in differentiation, proliferation, and apoptosis in many tissue types (Zhao et al., 2019; Li et al., 2020). The Notch protein, as well as its ligand, contains extracellular EGF-like repeats that interact with the DSL domain of the ligand. Activation of the Notch signalling pathway is accompanied by proteolysis, which releases the intracellular domain of Notch (NICD) (Sprinzak and Blacklow, 2021; Fang et al., 2022). The NICD is a fragment containing a RAM23 domain (RAM) that can enhance interactions with the Notch and CSL proteins, thereby facilitating the transmission of Notch to the nuclear localization signal (NLS) (Lubman et al., 2007; Johnson and Barrick, 2012). The NICD fragment can further mediate interactions with other proteins in the Notch signalling pathway (Figure 1).

The VEGF signalling pathway interacts with the Notch signalling pathway, coordinates the differentiation of arteries and veins, and is also involved in changes in vascular budding and vascular branching (Laham et al., 2003; Li et al., 2017; Souilhol et al., 2022; Li et al., 2023a). In addition, the VEGF signalling pathway can regulate key processes such as lumen remodelling of blood vessels.

Microvascular rarefaction leading to hypertension involves multiple signalling pathways, such as vascular endothelial development, angiogenesis, inflammation, oxidative stress, and endogenous short peptides (Feihl et al., 2008; Bruno et al., 2018; Do et al., 2023; Zdravkovic et al., 2023). Vascular endothelial cells play a key role in maintaining normal vascular biological function and regulating vascular tone. The abnormal regulation of vascular endothelial growth factor and platelet-derived growth factor during angiogenesis may also be involved in the occurrence of microvascular rarefaction (Cohen et al., 2023). Studies have shown that the Notch signalling pathway plays an important role in angiogenesis (Maynard et al., 2003).

The Notch signalling pathway plays a key role in the regulation of angiogenesis, especially in balancing the proangiogenic effects of the VEGF signalling pathway. The Notch signalling pathway antagonizes angiogenesis *via* VEGF signalling. It also stimulates quiescent endothelial cells and promotes apical cell formation. Thus, this pathway mediates the formation of vascular buds and the further growth of new vascular buds (Marinescu et al., 2015). Under conditions of stress, the cascade between DLL4-mediated Notch signalling and VEGFA-VEGFR2 signalling induces endothelial cells adjacent to the dominant TCS to maintain high levels of Notch signalling, thereby inhibiting their differentiation into TCShh cells. Furthermore, it can inhibit angiogenesis (Gerhardt

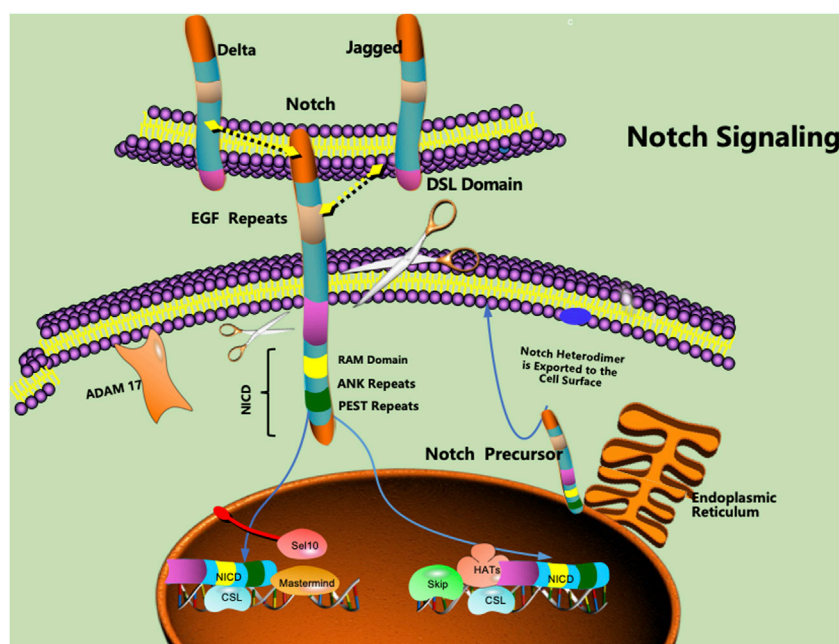


FIGURE 1

Notch proteins are cell surface transmembrane-spanning receptors that mediate critically important cellular functions through direct cell–cell contact. EGF: epidermal growth factor, NICD: intracellular domain of Notch, NLS: nuclear localization signal, CSL: CBF1/Su(H)/Lag-1.

et al., 2003; Jakobsson et al., 2010; Herbert and Stainier, 2011). Angiogenesis inhibition is thought to be controlled by Notch 1-mediated downregulation of Flt 4 and upregulation of soluble Flt 1 (sVEGFR-1/sFlt 1) (Chappell et al., 2009; Trindade et al., 2012). In particular, upregulation of sFlt1 reduces local VEGF bioavailability and inhibits angiogenesis. The Notch ligand Jag 1 is expressed mainly in stem cells and is thought to specifically block Dll 4-Notch 1 signalling in apical cells (Benedito et al., 2009). In the cardiovascular system, excessive activation of the Notch signalling pathway can lead to vascular endothelial cell proliferation and congestion and an increase in vessel wall thickness, which leads to vascular stenosis and hypertension (Niessen and Karsan, 2007; MacGrogan et al., 2018; Gomez et al., 2021).

The Notch signalling pathway is also involved in the occurrence and development of heart failure. Activation of Notch signalling can lead to cardiomyocyte proliferation and hypertrophy and can also affect angiogenesis and repair processes in the cardiovascular system (Fortini et al., 2014; Matsushita et al., 2023). These changes may adversely affect myocardial function and lead to the development of heart failure. Notch1 is a transmembrane receptor found in a variety of cells, including smooth muscle cells and endothelial cells in the cardiovascular system. During heart wall formation, Notch signalling regulates the ratio of cardiomyocytes to noncardiomyocytes by inhibiting myogenesis, thereby further promoting atrioventricular canal remodelling and maturation and heart valve formation (Niessen et al., 2011; Peng et al., 2023). Activation of the Notch1 signalling pathway is achieved mainly through the binding of the Notch1 receptor to its ligand. When the Notch1 receptor binds to its ligand, secondary cleavage occurs, and the active region of the Notch1 receptor is released. Then the receptor enters the nucleus and interacts with transcription factors to promote gene transcription and

expression (Wang S. et al., 2022). Studies have also shown that activation of the Notch1 signalling pathway is closely related to cardiac function and can affect the proliferation and differentiation of cardiomyocytes, thus affecting the development and function of the heart (Pahlavani, 2022). In addition, activation of the Notch1 signalling pathway can also regulate the expression of the actin gene in cardiac cells, affecting cardiac contractility and cardiac contraction rhythm (Hrstka et al., 2017). Therefore, the Notch signalling pathway plays an integral and critical role in maintaining and regulating blood pressure and cardiac function.

7 Role of the notch1 signalling pathway in tumour systems

The Notch signalling pathway is involved in cell-to-cell interactions and communication and plays an important role in embryonic development and the maintenance of tissue homeostasis in adult organisms. Moreover, aberrant Notch signalling pathway activity has also been found to be closely related to the occurrence and development of a variety of tumour types.

In terms of tumours, studies have shown that abnormal activation of the Notch1 signalling pathway is related to the occurrence and development of a variety of diseases, such as tumour proliferation and metastasis and abnormal responses of the immune system (Yang et al., 2018; Peng et al., 2023). Therefore, we hypothesized that inhibition of the Notch1 signalling pathway may ameliorate the adverse effects on blood pressure and cardiac function. Therefore, what is the underlying mechanism involved in reducing blood pressure and cardiac dysfunction? This will be another important dimension that needs to be studied.

In hepatocellular carcinoma (HCC), Notch signalling is activated by different ligands and plays a polymorphic role depending on the cell type, affecting tumour growth, invasive ability, and stem cell-like properties (Giovannini et al., 2021). Thus, interfering with Notch signalling may be a promising therapeutic approach. In tumour therapy, the downregulation of Notch1 can achieve synergistic effects and reduce chemoresistance when targeted drugs are used alone or in combination with chemotherapy. In addition, Notch mutations have been proposed to be predictive biomarkers for immune checkpoint blockade therapy in many cancers (Zhou et al., 2022; Li et al., 2023b).

8 Possible treatment of hypertension and myocardial damage caused by the notch signalling pathway

Recently, it has been observed that Epac1 can negatively regulate the Notch signalling pathway. Epac1 knockdown in endothelial cell lines resulted in a significant increase in the intracellular Notch1 protein level, and Epac1 knockdown increased the protein levels of NICD and DLL4, thereby further inhibiting angiogenesis. Therefore, overexpression of Epac1 may be an effective way to alleviate microvascular rarefaction (Fritz et al., 2015; Lan et al., 2020; Slika et al., 2023). Interestingly, studies have shown that Epac1 knockdown can inhibit pathological angiogenesis but has no significant effect on physiological angiogenesis, which is worthy of attention and discussion (Lan et al., 2020). Previous studies have shown that Epac1 is a β -secretase enzyme that inhibits the activation of the Notch signalling pathway (Fujita et al., 2017; Tan et al., 2022). In addition, Epac1 has been shown to enhance the VEGF signalling pathway and promote pathological angiogenesis (Namkoong et al., 2009; Ramos et al., 2018). Therefore, we speculate that the occurrence of hypertension caused by TKIs may lead to microcirculation disorders and microvascular thinning, and overexpressing the Epac1 gene may be a very meaningful way to improve or treat microvascular thinning caused by TKIs. NOTCH mutations can be used as predictive markers for treatment with immune checkpoint blockade in some tumors. In summary, it is necessary to comprehensively evaluate the NOTCH pathway from a new perspective, so as to apply it in the subsequent clinical diagnosis and treatment.

9 Mitochondrial dysfunction may also be involved in the regulation of notch signalling

In addition, Epac1 can play a role in cell-to-cell molecular conduction by increasing its interaction with macromolecular complexes, including voltage-dependent anion channel 1 (VDAC1), chaperone glucose-regulated protein 75 (GRP75), and inositol triphosphate receptor 1 (IP3R1). The interaction between Epac1 and macromolecular complexes can further promote the exchange of Ca^{2+} between the endoplasmic reticulum and mitochondria, which can eventually lead to mitochondrial Ca^{2+} overload and the opening of mitochondrial permeability transition pores (Fazal et al., 2017). Therefore, Epac1 is expected to be a new target for the treatment of ischaemic myocardial injury.

Mitochondria are important organelles for cellular energy production, and dysfunction of these organelles plays a key role in the emergence and progression of cardiovascular diseases (Poznyak et al., 2021; Wu et al., 2022). Normally, mitochondrial proteostasis is monitored by an extensive system: the mitochondrial unfolded protein response (UPRmt) is activated when mitochondria are stimulated by misfolded protein stress. The latter promotes the upregulation of ClpP, HSP6, HSP-60, ATFS-1 and other markers through a series of signalling cascades to alleviate mitochondrial stress (Merkwirth et al., 2016; Kumar et al., 2022; Xin et al., 2022; Guo et al., 2023). Under conditions of stress, cells can protect mitochondria by activating the UPRmt. It has been shown that the inhibition of Epac1 prevents ferroptosis-induced cell death and disruption of mitochondrial integrity, whereas the inhibition of Epac2 has a limited effect (Laudette et al., 2021; Musheshe et al., 2022). However, the potential effects of Epac on mitochondrial function are still unclear, and additional experiments are needed for further elucidation.

Several studies have shown that UPRmt activation is accompanied by the upregulation of the mitophagy markers PINK1, PARK2, BNIP3, P62, and LC3 and the mitochondrial oxidative phosphorylation (OXPHOS) markers Cox5a, Cox2, Nd1, and Sdhc. These results indicate that the UPRmt, mitophagy and OXPHOS play synergistic roles in maintaining mitochondrial protein balance and mitochondrial function (Kang et al., 2017; Martinez et al., 2017; Sorrentino et al., 2017; Poole and Macleod, 2021). The latest article published in NATURE refers to a series of mitochondrial stress responses that cause abnormal changes in the UPRmt, mitophagy, and OXPHOS under specific pathological or stimulus conditions, such as the mitochondrial stress response (MSR) (Ruan et al., 2017; Sorrentino et al., 2017; Jakobsen et al., 2019). The MSR is a key mechanism regulating mitochondrial protein balance and the mitochondrial stress response. EPAC1 can activate the UPRmt and protect mitochondrial function. Additionally, it is a key molecule that regulates the UPRmt and mitophagy (Gariani et al., 2016; Jayarajan et al., 2019; Aslam and Ladilov, 2021).

10 Summary

TKIs can induce vascular endothelial damage, hypertension and myocardial injury by acting on the targets VEGFR, platelet-derived growth factor receptor (PDGFR) and stem cell factor receptor (SCFR). It can also damage mitochondria and affect myocardial energy metabolism through "off-target effects", eventually leading to cardiovascular complications. Therefore, the goal of antitumour therapy is to maximize the antitumour effect while reducing treatment-related cardiovascular events. Therefore, studying the mechanism of hypertension during the treatment of cancer patients with TKIs and finding compounds or key factors that can reduce blood pressure without affecting antitumour efficacy are highly valuable. We hypothesize that inhibition of Notch signalling could ameliorate TKI-induced microvascular rarefaction, thereby further ameliorating the increase in blood pressure or the resulting cardiac dysfunction caused by microvascular rarefaction. Overexpression of Epac 1, a key gene in the Notch1 signalling pathway, can improve the pathological

vascular inhibition induced by TKIs. The mechanism by which apatinib, a representative TKI, induces microcirculation damage, hypertension and heart failure through the Notch1 signalling pathway will also be explored at the cellular and animal levels to provide clinical guidance for patients with hypertension induced by cancer treated with TKIs.

In summary, side effects such as cardiovascular toxicity caused by antitumour therapy with TKIs have become one of the main factors limiting antitumour therapy with TKIs. For the treatment of such hypertension, the effect of traditional antihypertensive drugs is not ideal, and traditional antihypertensive drugs are also closely related to the occurrence and development of some tumours. Exploring the underlying mechanism of cardiovascular complications caused by antitumour treatment with TKIs is crucial for ensuring the smooth clinical application of TKIs, and identifying this mechanism is also an urgent need. New directions for improving the treatment of hypertension induced by these drugs should be explored. The ultimate goal of our team will be to improve the clinical outcome as soon as possible and to provide greater benefits for cancer patients.

Author contributions

WW: Methodology, Resources, Writing—original draft, Writing—review and editing. GL: Investigation, Writing—review and editing. JM: Writing—review and editing. XF: Formal Analysis, Writing—review and editing. JL: Conceptualization, Writing—review and editing. QS: Supervision, Writing—review and editing. JY: Data curation, Writing—review and editing. QH: Conceptualization, Investigation, Supervision, Validation, Writing—review and editing.

References

- Agabiti-Rosei, E. (2003). Structural and functional changes of the microcirculation in hypertension: influence of pharmacological therapy. *Drugs* 63, 19–29.
- Agrawal, V., Kropski, J. A., Gokey, J. J., Kobeck, E., Murphy, M. B., Murray, K. T., et al. (2023). Myeloid cell derived IL1 β contributes to pulmonary hypertension in HFPcEF. *Circ. Res.* 133 (11), 885–898. doi:10.1161/CIRCRESAHA.123.323119
- Antonios, T. F., Rattray, F. M., Singer, D. R., Markandu, N. D., Mortimer, P. S., and MacGregor, G. A. (2003). Rarefaction of skin capillaries in normotensive offspring of individuals with essential hypertension. *Heart* 89 (2), 175–178. doi:10.1136/heart.89.2.175
- Aslam, M., and Ladilov, Y. (2021). Regulation of mitochondrial homeostasis by sAC-derived cAMP pool: basic and translational aspects. *Cells* 10 (2), 473. doi:10.3390/cells10020473
- Benedito, R., Roca, C., Sörensen, I., Adams, S., Gossler, A., Fruttiger, M., et al. (2009). The notch ligands Dll4 and Jagged1 have opposing effects on angiogenesis. *Cell* 137 (6), 1124–1135. doi:10.1016/j.cell.2009.03.025
- Bernatz, S., Monden, D., Gessler, F., Radic, T., Hattingen, E., Senft, C., et al. (2021). Influence of VEGF-A, VEGFR-1-3, and neuropilin 1-2 on progression-free: and overall survival in WHO grade II and III meningioma patients. *J. Mol. Histol.* 52 (2), 233–243. doi:10.1007/s10735-020-09940-2
- Boegehold, M. A., Johnson, M. D., and Overbeck, H. W. (1991). Pressure-independent arteriolar rarefaction in hypertension. *Am. J. Physiol.* 261 (1 Pt 2), H83–H87. doi:10.1152/ajpheart.1991.261.1.H83
- Bray, F., Laversanne, M., Weiderpass, E., and Soerjomataram, I. (2021). The ever-increasing importance of cancer as a leading cause of premature death worldwide. *Cancer* 127 (16), 3029–3030. doi:10.1002/cnrc.33587
- Bruno, R. M., Masi, S., Taddei, M., Taddei, S., and Virdis, A. (2018). Essential hypertension and functional microvascular ageing. *High. Blood Press Cardiovasc Prev.* 25 (1), 35–40. doi:10.1007/s40292-017-0245-9
- Camici, P. G., Tschöpe, C., Di Carli, M. F., Rimoldi, O., and Van Linthout, S. (2020). Coronary microvascular dysfunction in hypertrophy and heart failure. *Cardiovasc Res.* 116 (4), 806–816. doi:10.1093/cvr/cvaa023
- Catino, A. B., Hubbard, R. A., Chirinos, J. A., Townsend, R., Keefe, S., Haas, N. B., et al. (2018). Longitudinal assessment of vascular function with sunitinib in patients with metastatic renal cell carcinoma. *Circ. Heart Fail* 11 (3), e004408. doi:10.1161/CIRCHEARTFAILURE.117.004408
- Chade, A. R., and Kelsen, S. (2010). Renal microvascular disease determines the responses to revascularization in experimental renovascular disease. *Circ. Cardiovasc Interv.* 3 (4), 376–383. doi:10.1161/CIRCINTERVENTIONS.110.951277
- Chappell, J. C., Taylor, S. M., Ferrara, N., and Bautch, V. L. (2009). Local guidance of emerging vessel sprouts requires soluble Flt-1. *Dev. Cell* 17 (3), 377–386. doi:10.1016/j.devcel.2009.07.011
- Ciuffetti, G., Schillaci, G., Innocente, S., Lombardini, R., Pasqualini, L., Notaristefano, S., et al. (2003). Capillary rarefaction and abnormal cardiovascular reactivity in hypertension. *J. Hypertens.* 21 (12), 2297–2303. doi:10.1097/00004872-200312000-00018
- Cohen, J. B., Brown, N. J., Brown, S. A., Dent, S., van Dorst, D. C. H., Herrmann, S. M., et al. (2023). Cancer therapy-related hypertension: a scientific statement from the American heart association. *Hypertension* 80 (3), e46–e57. doi:10.1161/HYP.0000000000000224
- Cusack, R., Leone, M., Rodriguez, A. H., and Martin-Loeches, I. (2022). Endothelial damage and the microcirculation in critical illness. *Biomedicines* 10 (12), 3150. doi:10.3390/biomedicines10123150
- De Backer, D., Orbezo Cortes, D., Donadello, K., and Vincent, J. L. (2014). Pathophysiology of microcirculatory dysfunction and the pathogenesis of septic shock. *Virulence* 5 (1), 73–79. doi:10.4161/viru.26482

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Conflict of interest

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- De Ciuceis, C., Rizzoni, D., and Palatini, P. (2023). Microcirculation and physical exercise in hypertension. *Hypertension* 80 (4), 730–739. doi:10.1161/HYPERTENSIONAHA.122.19465
- Do, T., Van, A., Ataei, A., Sharma, S., and Mohandas, R. (2023). Microvascular dysfunction in obesity hypertension. *Curr. Hypertens. Rep.* 25 (12), 447–453. doi:10.1007/s11906-023-01272-2
- Dolmatova, E., Waheed, N., Olson, B. M., Patel, S. A., and Mandawat, A. (2023). The intersection of prostate cancer and hypertension: a call to action. *Curr. Treat. Options Oncol.* 24 (7), 892–905. doi:10.1007/s11864-023-01094-z
- Fang, Z. Q., Ruan, B., Liu, J. J., Duan, J. L., Yue, Z. S., Song, P., et al. (2022). Notch-triggered maladaptation of liver sinusoidal endothelium aggravates nonalcoholic steatohepatitis through endothelial nitric oxide synthase. *Hepatology* 76 (3), 742–758. doi:10.1002/hep.32332
- Fazal, L., Laudette, M., Paula-Gomes, S., Pons, S., Conte, C., Tortosa, F., et al. (2017). Multifunctional mitochondrial Epac1 controls myocardial cell death. *Circ. Res.* 120 (4), 645–657. doi:10.1161/CIRCRESAHA.116.309859
- Feihl, F., Liaudet, L., Levy, B. I., and Waeber, B. (2008). Hypertension and microvascular remodelling. *Cardiovasc. Res.* 78 (2), 274–285. doi:10.1093/cvr/cvn022
- Fortini, C., Cesselli, D., Beltrami, A. P., Bergamin, N., Caragnano, A., Moretti, L., et al. (2014). Alteration of Notch signaling and functionality of adipose tissue derived mesenchymal stem cells in heart failure. *Int. J. Cardiol.* 174 (1), 119–126. doi:10.1016/j.ijcard.2014.03.173
- Fritz, A. L., Adil, M. M., Mao, S. R., and Schaffer, D. V. (2015). cAMP and EPAC signaling functionally replace OCT4 during induced pluripotent stem cell reprogramming. *Mol. Ther.* 23 (5), 952–963. doi:10.1038/mt.2015.28
- Fujita, T., Umehura, M., Yokoyama, U., Okumura, S., and Ishikawa, Y. (2017). The role of Epac in the heart. *Cell Mol. Life Sci.* 74 (4), 591–606. doi:10.1007/s00018-016-2336-5
- Gao, Q., He, S., Peng, Y., Su, P., and Zhao, L. (2023). Proteomic profiling of epicardial fat in heart failure with preserved versus reduced and mildly reduced ejection fraction. *J. Cell Mol. Med.* 27 (5), 727–735. doi:10.1111/jcmm.17695
- Garg, P., Lewis, R. A., Johns, C. S., Swift, A. J., Capener, D., Rajaram, S., et al. (2021). Cardiovascular magnetic resonance predicts all-cause mortality in pulmonary hypertension associated with heart failure with preserved ejection fraction. *Int. J. Cardiovasc. Imaging* 37 (10), 3019–3025. doi:10.1007/s10554-021-02279-z
- Gariani, K., Menzies, K. J., Ryu, D., Wegner, C. J., Wang, X., Ropelle, E. R., et al. (2016). Eliciting the mitochondrial unfolded protein response by nicotinamide adenine dinucleotide depletion reverses fatty liver disease in mice. *Hepatology* 63 (4), 1190–1204. doi:10.1002/hep.28245
- Gerhardt, H., Golding, M., Fruttiger, M., Ruhrberg, C., Lundkvist, A., Abramsson, A., et al. (2003). VEGF guides angiogenic sprouting utilizing endothelial tip cell filopodia. *J. Cell Biol.* 161 (6), 1163–1177. doi:10.1083/jcb.200302047
- Gevaert, A. B., Kataria, R., Zannad, F., Sauer, A. J., Damman, K., Sharma, K., et al. (2022). Heart failure with preserved ejection fraction: recent concepts in diagnosis, mechanisms and management. *Heart* 108 (17), 1342–1350. doi:10.1136/heartjnl-2021-319605
- Giovannini, C., Fornari, F., Piscaglia, F., and Gramantieri, L. (2021). Notch signaling regulation in HCC: from hepatitis virus to non-coding RNAs. *Cells* 10 (3), 521. doi:10.3390/cells10030521
- Gomez, A. H., Joshi, S., Yang, Y., Tune, J. D., Zhao, M. T., and Yang, H. (2021). Bioengineering systems for modulating notch signaling in cardiovascular development, disease, and regeneration. *J. Cardiovasc. Dev. Dis.* 8 (10), 125. doi:10.3390/jcdd8100125
- Gryglewska, B., Necki, M., Zelawski, M., Cwynar, M., Baron, T., Mrozek, M., et al. (2011). Fractal dimensions of skin microcirculation flow in subjects with familial predisposition or newly diagnosed hypertension. *Cardiol. J.* 18 (1), 26–32.
- Guo, Y., Guan, T., Shafiq, K., Yu, Q., Jiao, X., Na, D., et al. (2023). Mitochondrial dysfunction in aging. *Ageing Res. Rev.* 88, 101955. doi:10.1016/j.arr.2023.101955
- Herbert, S. P., and Stainier, D. Y. (2011). Molecular control of endothelial cell behaviour during blood vessel morphogenesis. *Nat. Rev. Mol. Cell Biol.* 12 (9), 551–564. doi:10.1038/nrm3176
- Horton, W. B., and Barrett, E. J. (2021). Microvascular dysfunction in diabetes mellitus and cardiometabolic disease. *Endocr. Rev.* 42 (1), 29–55. doi:10.1210/endo/rev/bnaa025
- Hrstka, S. C., Li, X., Nelson, T. J., and Wanek Program Genetics Pipeline Group (2017). NOTCH1-Dependent nitric oxide signaling deficiency in hypoplastic left heart syndrome revealed through patient-specific phenotypes detected in bioengineered cardiogenesis. *Stem Cells* 35 (4), 1106–1119. doi:10.1002/stem.2582
- Huang, C., Li, H., Xu, Y., Xu, C., Sun, H., Li, Z., et al. (2023). BICC1 drives pancreatic cancer progression by inducing VEGF-independent angiogenesis. *Signal Transduct. Target Ther.* 8 (1), 271. doi:10.1038/s41392-023-01478-5
- Jackson, A. M., Jhund, P. S., Anand, I. S., Düngen, H. D., Lam, C. S. P., Lefkowitz, M. P., et al. (2021). Sacubitril-valsartan as a treatment for apparent resistant hypertension in patients with heart failure and preserved ejection fraction. *Eur. Heart J.* 42 (36), 3741–3752. doi:10.1093/eurheartj/ehab499
- Jacobson, A., Yan, C., Gao, Q., Rincon-Skinner, T., Rivera, A., Edwards, J., et al. (2007). Aging enhances pressure-induced arterial superoxide formation. *Am. J. Physiol. Heart Circ. Physiol.* 293 (3), H1344–H1350. doi:10.1152/ajpheart.00413.2007
- Jakobsen, E., Lange, S. C., and Bak, L. K. (2019). Soluble adenylyl cyclase-mediated cAMP signaling and the putative role of PKA and EPAC in cerebral mitochondrial function. *J. Neurosci. Res.* 97 (8), 1018–1038. doi:10.1002/jnr.24477
- Jakobsson, L., Franco, C. A., Bentley, K., Collins, R. T., Ponsioen, B., Aspö, I. M., et al. (2010). Endothelial cells dynamically compete for the tip cell position during angiogenic sprouting. *Nat. Cell Biol.* 12 (10), 943–953. doi:10.1038/ncb2103
- Jayarajan, V., Appukuttan, A., Aslam, M., Reusch, P., Regitz-Zagrosek, V., and Ladilov, Y. (2019). Regulation of AMPK activity by type 10 adenylyl cyclase: contribution to the mitochondrial biology, cellular redox and energy homeostasis. *Cell Mol. Life Sci.* 76 (24), 4945–4959. doi:10.1007/s00018-019-0152-y
- Johnson, S. E., and Barrick, D. (2012). Dissecting and circumventing the requirement for RAM in CSL-dependent Notch signaling. *PLoS One* 7 (8), e39093. doi:10.1371/journal.pone.0039093
- Jonk, A. M., Houben, A. J., de Jongh, R. T., Serné, E. H., Schaper, N. C., and Stehouwer, C. D. (2007). Microvascular dysfunction in obesity: a potential mechanism in the pathogenesis of obesity-associated insulin resistance and hypertension. *Physiol. (Bethesda)* 22, 252–260. doi:10.1152/physiol.00012.2007
- Kang, M. H., Das, J., Gurunathan, S., Park, H. W., Song, H., Park, C., et al. (2017). The cytotoxic effects of dimethyl sulfoxide in mouse preimplantation embryos: a mechanistic study. *Theranostics* 7 (19), 4735–4752. doi:10.7150/thno.21662
- Kangsamaksin, T., Murtomaki, A., Kofler, N. M., Cuervo, H., Chaudhri, R. A., Tattersall, I. W., et al. (2015). NOTCH decoys that selectively block DLL/NOTCH or JAG/NOTCH disrupt angiogenesis by unique mechanisms to inhibit tumor growth. *Cancer Discov.* 5 (2), 182–197. doi:10.1158/2159-8290.CD-14-0650
- Karaca, Ü., Schram, M. T., Houben, A. J., Muris, D. M., and Stehouwer, C. D. (2014). Microvascular dysfunction as a link between obesity, insulin resistance and hypertension. *Diabetes Res. Clin. Pract.* 103 (3), 382–387. doi:10.1016/j.diabres.2013.12.012
- Kumar, R., Chaudhary, A. K., Woytash, J., Inigo, J. R., Gokhale, A. A., Bshara, W., et al. (2022). A mitochondrial unfolded protein response inhibitor suppresses prostate cancer growth in mice via HSP60. *J. Clin. Invest.* 132 (13), e149906. doi:10.1172/JCI49906
- Kwak, M., Southard, K. M., Kim, W. R., Lin, A., Kim, N. H., Gopalappa, R., et al. (2022). Adherens junctions organize size-selective proteolytic hotspots critical for Notch signalling. *Nat. Cell Biol.* 24 (12), 1739–1753. doi:10.1038/s41556-022-01031-6
- Laham, R. J., Li, J., Tofukujji, M., Post, M., Simons, M., and Sellke, F. W. (2003). Spatial heterogeneity in VEGF-induced vasodilation: VEGF dilates microvessels but not epicardial and systemic arteries and veins. *Ann. Vasc. Surg.* 17 (3), 245–252. doi:10.1007/s10016-001-0299-x
- Lan, C., Shen, J., Wang, Y., Li, J., Liu, Z., He, M., et al. (2020). Camrelizumab plus apatinib in patients with advanced cervical cancer (clap): a multicenter, open-label, single-arm, phase II trial. *J. Clin. Oncol.* 38 (34), 4095–4106. doi:10.1200/JCO.20.01920
- Laudette, M., Sainte-Marie, Y., Cousin, G., Bergonnier, D., Belhabib, I., Brun, S., et al. (2021). Cyclic AMP-binding protein Epac1 acts as a metabolic sensor to promote cardiomyocyte lipotoxicity. *Cell Death Dis.* 12 (9), 824. doi:10.1038/s41419-021-04113-9
- Lawler, J. (2022). Counter regulation of tumor angiogenesis by vascular endothelial growth factor and thrombospondin-1. *Semin. Cancer Biol.* 86 (Pt 2), 126–135. doi:10.1016/j.semcancer.2022.09.006
- Le, X., Nilsson, M., Goldman, J., Reck, M., Nakagawa, K., Kato, T., et al. (2021). Dual EGFR-VEGF pathway inhibition: a promising strategy for patients with EGFR-mutant nscl. *J. Thorac. Oncol.* 16 (2), 205–215. doi:10.1016/j.jtho.2020.10.006
- Lennartsson, J., and Rönnstrand, L. (2012). Stem cell factor receptor/c-Kit: from basic science to clinical implications. *Physiol. Rev.* 92 (4), 1619–1649. doi:10.1152/physrev.00046.2011
- le Noble, F. A. C., Mourad, J. J., Levy, B. I., and Struijker-Boudier, H. A. J. (2023). VEGF (vascular endothelial growth factor) inhibition and hypertension: does microvascular rarefaction play a role? *Hypertension* 80 (5), 901–911. doi:10.1161/HYPERTENSIONAHA.122.19427
- Levy, B. I., Ambrosio, G., Pries, A. R., and Struijker-Boudier, H. A. (2001). Microcirculation in hypertension: a new target for treatment? *Circulation* 104 (6), 735–740. doi:10.1161/hc3101.091158
- Levy, B. I., Schiffrin, E. L., Mourad, J. J., Agostini, D., Vicaute, E., Safar, M. E., et al. (2008). Impaired tissue perfusion: a pathology common to hypertension, obesity, and diabetes mellitus. *Circulation* 118 (9), 968–976. doi:10.1161/CIRCULATIONAHA.107.763730
- Li, C., Ma, L., Wang, Q., Shao, X., Guo, L., Chen, J., et al. (2022). Rho kinase inhibition ameliorates vascular remodeling and blood pressure elevations in a rat model of apatinib-induced hypertension. *J. Hypertens.* 40 (4), 675–684. doi:10.1097/HJH.0000000000003060
- Li, J., Qin, S., Xu, J., Xiong, J., Wu, C., Bai, Y., et al. (2016). Randomized, double-blind, placebo-controlled phase III trial of apatinib in patients with chemotherapy-refractory

- advanced or metastatic adenocarcinoma of the stomach or gastroesophageal junction. *J. Clin. Oncol.* 34 (13), 1448–1454. doi:10.1200/JCO.2015.63.5995
- Li, L., Liu, H., Xu, C., Deng, M., Song, M., Yu, X., et al. (2017). VEGF promotes endothelial progenitor cell differentiation and vascular repair through connexin 43. *Stem Cell Res. Ther.* 8 (1), 237. doi:10.1186/s13287-017-0684-1
- Li, S., Shi, Y., Yuan, S., Ruan, J., Pan, H., Ma, M., et al. (2024). Inhibiting the MAPK pathway improves heart failure with preserved ejection fraction induced by salt-sensitive hypertension. *Biomed. Pharmacother.* 170, 115987. doi:10.1016/j.biopha.2023.115987
- Li, T., Xu, X. H., Guo, X., Yuan, T., Tang, Z. H., Jiang, X. M., et al. (2020). Activation of notch 3/c-MYC/CHOP axis regulates apoptosis and promotes sensitivity of lung cancer cells to mTOR inhibitor everolimus. *Biochem. Pharmacol.* 175, 113921. doi:10.1016/j.bcp.2020.113921
- Li, X., Souilhol, C., Canham, L., Jia, X., Diabougua, M., Ayllon, B. T., et al. (2023a). DLL4 promotes partial endothelial-to-mesenchymal transition at atherosclerosis-prone regions of arteries. *Vasc. Pharmacol.* 150, 107178. doi:10.1016/j.vph.2023.107178
- Li, X., Yan, X., Wang, Y., Kaur, B., Han, H., and Yu, J. (2023b). The Notch signaling pathway: a potential target for cancer immunotherapy. *J. Hematol. Oncol.* 16 (1), 45. doi:10.1186/s13045-023-01439-z
- Lubman, O. Y., Ilagan, M. X., Kopan, R., and Barrick, D. (2007). Quantitative dissection of the Notch:CSL interaction: insights into the Notch-mediated transcriptional switch. *J. Mol. Biol.* 365 (3), 577–589. doi:10.1016/j.jmb.2006.09.071
- MacGrogan, D., Münch, J., and de la Pompa, J. L. (2018). Notch and interacting signalling pathways in cardiac development, disease, and regeneration. *Nat. Rev. Cardiol.* 15 (11), 685–704. doi:10.1038/s41569-018-0100-2
- Marinescu, M. A., Löffler, A. I., Ouellette, M., Smith, L., Kramer, C. M., and Bourque, J. M. (2015). Coronary microvascular dysfunction, microvascular angina, and treatment strategies. *JACC Cardiovasc. Imaging* 8 (2), 210–220. doi:10.1016/j.jcmg.2014.12.008
- Marra, A. M., Sherman, A. E., Salzano, A., Guazzi, M., Saggari, R., Squire, I. B., et al. (2022). Right side of the heart pulmonary circulation unit involvement in left-sided heart failure: diagnostic, prognostic, and therapeutic implications. *Chest* 161 (2), 535–551. doi:10.1016/j.chest.2021.09.023
- Martens, J. R., and Gelband, C. H. (1998). Ion channels in vascular smooth muscle: alterations in essential hypertension. *Proc. Soc. Exp. Biol. Med.* 218 (3), 192–203. doi:10.3181/00379727-218-44286
- Martinez, B. A., Petersen, D. A., Gaeta, A. L., Stanley, S. P., Caldwell, G. A., and Caldwell, K. A. (2017). Dysregulation of the mitochondrial unfolded protein response induces non-apoptotic dopaminergic neurodegeneration in *C. elegans* models of Parkinson's disease. *J. Neurosci.* 37 (46), 11085–11100. doi:10.1523/JNEUROSCI.1294-17.2017
- Matsushita, K., Marchandot, B., Trimaille, A., Hmadeh, S., Kibler, M., Heger, J., et al. (2023). Determinants and treatments of heart failure after transcatheter aortic valve implantation: moving up a notch. *Esc. Heart Fail* 10 (4), 2183–2199. doi:10.1002/ehf2.14435
- Maynard, S. E., Min, J. Y., Merchan, J., Lim, K. H., Li, J., Mondal, S., et al. (2003). Excess placental soluble fms-like tyrosine kinase 1 (sFlt1) may contribute to endothelial dysfunction, hypertension, and proteinuria in preeclampsia. *J. Clin. Invest.* 111 (5), 649–658. doi:10.1172/JCI17189
- Mengozzi, A., de Ciuceis, C., Dell'oro, R., Georgiopoulos, G., Lazaridis, A., Nosalski, R., et al. (2023). The importance of microvascular inflammation in ageing and age-related diseases: a position paper from the ESH working group on small arteries, section of microvascular inflammation. *J. Hypertens.* 41 (10), 1521–1543. doi:10.1097/HJH.0000000000003503
- Merkwirth, C., Jovaisaite, V., Durieux, J., Matilainen, O., Jordan, S. D., Quiros, P. M., et al. (2016). Two conserved histone demethylases regulate mitochondrial stress-induced longevity. *Cell* 165 (5), 1209–1223. doi:10.1016/j.cell.2016.04.012
- Milan, A., Puglisi, E., Ferrari, L., Bruno, G., Losano, I., and Veglio, F. (2014). Arterial hypertension and cancer. *Int. J. Cancer* 134 (10), 2269–2277. doi:10.1002/ijc.28334
- Mirabito Colafella, K. M., Neves, K. B., Montezano, A. C., Garrelds, I. M., van Veghel, R., de Vries, R., et al. (2020). Selective ETA vs dual ETA/B receptor blockade for the prevention of sunitinib-induced hypertension and albuminuria in WKY rats. *Cardiovasc. Res.* 116 (10), 1779–1790. doi:10.1093/cvr/cvz260
- Mourad, J. J., des Guetz, G., Debbabi, H., and Levy, B. I. (2008). Blood pressure rise following angiogenesis inhibition by bevacizumab. A crucial role for microcirculation. *Ann. Oncol.* 19 (5), 927–934. doi:10.1093/annonc/mdm550
- Mourad, J. J., and Levy, B. I. (2011). Mechanisms of antiangiogenic-induced arterial hypertension. *Curr. Hypertens. Rep.* 13 (4), 289–293. doi:10.1007/s11906-011-0206-y
- Musheshe, N., Oun, A., Sabogal-Guáqueta, A. M., Trombetta-Lima, M., Mitchel, S. C., Adzemovic, A., et al. (2022). Pharmacological inhibition of Epac1 averts ferroptosis cell death by preserving mitochondrial integrity. *Antioxidants (Basel)* 11 (2), 314. doi:10.3390/antiox11020314
- Nagai, M., Dote, K., and Förster, C. Y. (2023). Denervation or stimulation? Role of sympatho-vagal imbalance in HFpEF with hypertension. *Hypertens. Res.* 46 (7), 1727–1737. doi:10.1038/s41440-023-01272-4
- Namkoong, S., Kim, C. K., Cho, Y. L., Kim, J. H., Lee, H., Ha, K. S., et al. (2009). Forskolin increases angiogenesis through the coordinated cross-talk of PKA-dependent VEGF expression and Epac-mediated PI3K/Akt/eNOS signaling. *Cell Signal* 21 (6), 906–915. doi:10.1016/j.cellsig.2009.01.038
- Neves, K. B., Rios, F. J., Jones, R., Evans, T. R. J., Montezano, A. C., and Touyz, R. M. (2019). Microparticles from vascular endothelial growth factor pathway inhibitor-treated cancer patients mediate endothelial cell injury. *Cardiovasc. Res.* 115 (5), 978–988. doi:10.1093/cvr/cvz021
- Neves, K. B., Rios, F. J., van der Mey, L., Alves-Lopes, R., Cameron, A. C., Volpe, M., et al. (2018). VEGFR (vascular endothelial growth factor receptor) inhibition induces cardiovascular damage via redox-sensitive processes. *Hypertension* 71 (4), 638–647. doi:10.1161/HYPERTENSIONAHA.117.10490
- Niessen, K., and Karsan, A. (2007). Notch signaling in the developing cardiovascular system. *Am. J. Physiol. Cell Physiol.* 293 (1), C1–C11. doi:10.1152/ajpcell.00415.2006
- Niessen, K., Zhang, G., Ridgway, J. B., Chen, H., Kolumam, G., Siebel, C. W., et al. (2011). The Notch1-Dll4 signaling pathway regulates mouse postnatal lymphatic development. *Blood* 118 (7), 1989–1997. doi:10.1182/blood-2010-11-319129
- Obokata, M., Reddy, Y. N. V., Pislaru, S. V., Melenovsky, V., and Borlaug, B. A. (2017). Evidence supporting the existence of a distinct obese phenotype of heart failure with preserved ejection fraction. *Circulation* 136 (1), 6–19. doi:10.1161/CIRCULATIONAHA.116.026807
- Pahlavani, H. A. (2022). Exercise-induced signaling pathways to counteracting cardiac apoptotic processes. *Front. Cell Dev. Biol.* 10, 950927. doi:10.3389/fcell.2022.950927
- Paulus, W. J., and Tschöpe, C. (2013). A novel paradigm for heart failure with preserved ejection fraction: comorbidities drive myocardial dysfunction and remodeling through coronary microvascular endothelial inflammation. *J. Am. Coll. Cardiol.* 62 (4), 263–271. doi:10.1016/j.jacc.2013.02.092
- Peng, X., Wang, S., Chen, H., and Chen, M. (2023). Role of the Notch1 signaling pathway in ischemic heart disease (Review). *Int. J. Mol. Med.* (3), 51. doi:10.3892/ijmm.2023.5230
- Poole, L. P., and Macleod, K. F. (2021). Mitophagy in tumorigenesis and metastasis. *Cell Mol. Life Sci.* 78 (8), 3817–3851. doi:10.1007/s00018-021-03774-1
- Poznyak, A. V., Nikiforov, N. G., Wu, W. K., Kirichenko, T. V., and Orekhov, A. N. (2021). Autophagy and mitophagy as essential components of atherosclerosis. *Cells* 10 (2), 443. doi:10.3390/cells10020443
- Premont, R. T., Reynolds, J. D., Zhang, R., and Stamler, J. S. (2020). Role of nitric oxide carried by hemoglobin in cardiovascular physiology: developments on a three-gas respiratory cycle. *Circ. Res.* 126 (1), 129–158. doi:10.1161/CIRCRESAHA.119.315626
- Pucci, G., Milan, A., Paini, A., Salvetti, M., Cerasari, A., and Vaudo, G. (2019). Acute blood pressure elevation associated with biological therapies for cancer: a focus on VEGF signaling pathway inhibitors. *Expert Opin. Biol. Ther.* 19 (5), 433–442. doi:10.1080/14712598.2019.1594770
- Qin, S., Chan, S. L., Gu, S. B., Bai, Y., Ren, Z., Lin, X., et al. (2023). Camrelizumab plus rivoceranib versus sorafenib as first-line therapy for unresectable hepatocellular carcinoma (CARES-310): a randomised, open-label, international phase 3 study. *Lancet* 402 (10408), 1133–1146. doi:10.1016/S0140-6736(23)00961-3
- Ramos, C. J., Lin, C., Liu, X., and Antonetti, D. A. (2018). The EPAC-Rap1 pathway prevents and reverses cytokine-induced retinal vascular permeability. *J. Biol. Chem.* 293 (2), 717–730. doi:10.1074/jbc.M117.815381
- Redfield, M. M., and Borlaug, B. A. (2023). Heart failure with preserved ejection fraction: a review. *Jama* 329 (10), 827–838. doi:10.1001/jama.2023.2020
- Rizzoni, D., Agabiti-Rosei, C., Boari, G. E. M., Muiresan, M. L., and De Ciuceis, C. (2023). Microcirculation in hypertension: a therapeutic target to prevent cardiovascular disease? *J. Clin. Med.* 12 (15), 4892. doi:10.3390/jcm12154892
- Robinson, E. S., Khankin, E. V., Choueiri, T. K., Dhawan, M. S., Rogers, M. J., Karumanchi, S. A., et al. (2010). Suppression of the nitric oxide pathway in metastatic renal cell carcinoma patients receiving vascular endothelial growth factor-signaling inhibitors. *Hypertension* 56 (6), 1131–1136. doi:10.1161/HYPERTENSIONAHA.110.160481
- Rodríguez-Agustín, A., Casanova, V., Grau-Expósito, J., Sánchez-Palomino, S., Alcamí, J., and Climent, N. (2023). Immunomodulatory activity of the tyrosine kinase inhibitor dasatinib to elicit NK cytotoxicity against cancer, HIV infection and aging. *Pharmaceutics* 15 (3), 917. doi:10.3390/pharmaceutics15030917
- Rossi, V. A., Nebunu, D., Haider, T., Laptseva, N., Naegele, M. P., Ruschitzka, F., et al. (2023). Diverging role of epicardial adipose tissue across the entire heart failure spectrum. *Esc. Heart Fail* 10 (6), 3419–3429. doi:10.1002/ehf2.14483
- Ruan, L., Zhou, C., Jin, E., Kucharavy, A., Zhang, Y., Wen, Z., et al. (2017). Cytosolic proteostasis through importing of misfolded proteins into mitochondria. *Nature* 543 (7645), 443–446. doi:10.1038/nature21695
- Sabe, S. A., Feng, J., Sellke, F. W., and Abid, M. R. (2022). Mechanisms and clinical implications of endothelium-dependent vasomotor dysfunction in coronary microvasculature. *Am. J. Physiol. Heart Circ. Physiol.* 322 (5), H819–h841. doi:10.1152/ajpheart.00603.2021

- Saif, M. W. (2013). Anti-VEGF agents in metastatic colorectal cancer (mCRC): are they all alike? *Cancer Manag. Res.* 5, 103–115. doi:10.2147/CMAR.S45193
- Sallinen, H., Anttila, M., Narvainen, J., Koponen, J., Hamalainen, K., Kholova, I., et al. (2009). Antiangiogenic gene therapy with soluble VEGFR-1, -2, and -3 reduces the growth of solid human ovarian carcinoma in mice. *Mol. Ther.* 17 (2), 278–284. doi:10.1038/mt.2008.258
- Serné, E. H., Gans, R. O., ter Maaten, J. C., Tangelde, G. J., Donker, A. J., and Stehouwer, C. D. (2001). Impaired skin capillary recruitment in essential hypertension is caused by both functional and structural capillary rarefaction. *Hypertension* 38 (2), 238–242. doi:10.1161/01.hyp.38.2.238
- Shughoury, A., Bhatwadekar, A., Jusufbegovic, D., Hajrasouliha, A., and Ciulla, T. A. (2023). The evolving therapeutic landscape of diabetic retinopathy. *Expert Opin. Biol. Ther.* 23 (10), 969–985. doi:10.1080/14712598.2023.2247987
- Slika, H., Mansour, H., Nasser, S. A., Shaito, A., Kobeissy, F., Orekhov, A. N., et al. (2023). Epac as a tractable therapeutic target. *Eur. J. Pharmacol.* 945, 175645. doi:10.1016/j.ejphar.2023.175645
- Slivnick, J., and Lampert, B. C. (2019). Hypertension and heart failure. *Heart Fail Clin.* 15 (4), 531–541. doi:10.1016/j.hfc.2019.06.007
- Sorop, O., van de Wouw, J., Chandler, S., Ohanyan, V., Tune, J. D., Chilian, W. M., et al. (2020). Experimental animal models of coronary microvascular dysfunction. *Cardiovasc Res.* 116 (4), 756–770. doi:10.1093/cvr/cvaa002
- Sorrentino, V., Romani, M., Mouchiroud, L., Beck, J. S., Zhang, H., D'Amico, D., et al. (2017). Enhancing mitochondrial proteostasis reduces amyloid- β proteotoxicity. *Nature* 552 (7684), 187–193. doi:10.1038/nature25143
- Souilhol, C., Tardajos Ayllon, B., Li, X., Diabougba, M. R., Zhou, Z., Canham, L., et al. (2022). JAG1-NOTCH4 mechanosensing drives atherosclerosis. *Sci. Adv.* 8 (35), eabo7958. doi:10.1126/sciadv.abo7958
- Sprinzak, D., and Blacklow, S. C. (2021). Biophysics of notch signaling. *Annu. Rev. Biophys.* 50, 157–189. doi:10.1146/annurev-biophys-101920-082204
- Steehgs, N., Gelderblom, H., Roodt, J. O., Christensen, O., Rajagopalan, P., Hovens, M., et al. (2008). Hypertension and rarefaction during treatment with telatinib, a small molecule angiogenesis inhibitor. *Clin. Cancer Res.* 14 (11), 3470–3476. doi:10.1158/1078-0432.CCR-07-5050
- Struijker-Boudier, H. A., and Heijnen, B. F. (2011). Early life microcirculation and the development of hypertension. *Hypertension* 58 (5), 768–769. doi:10.1161/HYPERTENSIONAHA.111.181107
- Stupin, A., Drenjančević, I., Šušnjara, P., Debeljak, Ž., Kolobarić, N., Jukić, I., et al. (2021). Is there association between altered adrenergic system activity and microvascular endothelial dysfunction induced by a 7-day high salt intake in young healthy individuals. *Nutrients* 13 (5), 1731. doi:10.3390/nu13051731
- Takahashi, H., Sakakibara-Konishi, J., Furuta, M., Shoji, T., Tsuji, K., Morinaga, D., et al. (2023). Notch pathway regulates osimertinib drug-tolerant persistence in EGFR-mutated non-small-cell lung cancer. *Cancer Sci.* 114 (4), 1635–1650. doi:10.1111/cas.15674
- Tan, Y. Q., Li, J., and Chen, H. W. (2022). Epac, a positive or negative signaling molecule in cardiovascular diseases. *Biomed. Pharmacother.* 148, 112726. doi:10.1016/j.biopha.2022.112726
- Triantafyllou, A., Anyfanti, P., Pyrasopoulou, A., Triantafyllou, G., Aslanidis, S., and Douma, S. (2015). Capillary rarefaction as an index for the microvascular assessment of hypertensive patients. *Curr. Hypertens. Rep.* 17 (5), 33. doi:10.1007/s11906-015-0543-3
- Trindade, A., Djokovic, D., Gigante, J., Badenes, M., Pedrosa, A. R., Fernandes, A. C., et al. (2012). Low-dosage inhibition of Dll4 signaling promotes wound healing by inducing functional neo-angiogenesis. *PLoS One* 7 (1), e29863. doi:10.1371/journal.pone.0029863
- Tsai, S. H., Lu, G., Xu, X., Ren, Y., Hein, T. W., and Kuo, L. (2017). Enhanced endothelin-1/Rho-kinase signalling and coronary microvascular dysfunction in hypertensive myocardial hypertrophy. *Cardiovasc Res.* 113 (11), 1329–1337. doi:10.1093/cvr/cvx103
- Tullemans, B. M. E., Heemskerck, J. W. M., and Kuijpers, M. J. E. (2018). Acquired platelet antagonism: off-target antiplatelet effects of malignancy treatment with tyrosine kinase inhibitors. *J. Thromb. Haemost.* 16 (9), 1686–1699. doi:10.1111/jth.14225
- Ungvari, Z., Csizsar, A., Kaminski, P. M., Wolin, M. S., and Koller, A. (2004). Chronic high pressure-induced arterial oxidative stress: involvement of protein kinase C-dependent NAD(P)H oxidase and local renin-angiotensin system. *Am. J. Pathol.* 165 (1), 219–226. doi:10.1016/S0002-9440(10)63290-7
- Ungvari, Z., Toth, P., Tarantini, S., Prodan, C. I., Sorond, F., Merkely, B., et al. (2021). Hypertension-induced cognitive impairment: from pathophysiology to public health. *Nat. Rev. Nephrol.* 17 (10), 639–654. doi:10.1038/s41581-021-00430-6
- Vancheri, F., Longo, G., Vancheri, S., and Henein, M. (2020). Coronary microvascular dysfunction. *J. Clin. Med.* 9 (9), 2880. doi:10.3390/jcm9092880
- van Cruysen, H., van der Veldt, A., and Hoekman, K. (2009). Tyrosine kinase inhibitors of VEGF receptors: clinical issues and remaining questions. *Front. Biosci. Landmark Ed.* 14 (6), 2248–2268. doi:10.2741/3377
- van Dorst, D. C. H., Dobbin, S. J. H., Neves, K. B., Herrmann, J., Herrmann, S. M., Versmissen, J., et al. (2021). Hypertension and prohypertensive antineoplastic therapies in cancer patients. *Circ. Res.* 128 (7), 1040–1061. doi:10.1161/CIRCRESAHA.121.318051
- Vano, Y. A., Elaidi, R., Bennamoun, M., Chevreau, C., Borchellini, D., Pannier, D., et al. (2022). Nivolumab, nivolumab-ipilimumab, and VEGFR-tyrosine kinase inhibitors as first-line treatment for metastatic clear-cell renal cell carcinoma (BIONIKK): a biomarker-driven, open-label, non-comparative, randomised, phase 2 trial. *Lancet Oncol.* 23 (5), 612–624. doi:10.1016/S1470-2045(22)00128-0
- Viallard, C., and Larrivée, B. (2017). Tumor angiogenesis and vascular normalization: alternative therapeutic targets. *Angiogenesis* 20 (4), 409–426. doi:10.1007/s10456-017-9562-9
- Vimalraj, S. (2022). A concise review of VEGF, PDGF, FGF, Notch, angiopoietin, and HGF signalling in tumor angiogenesis with a focus on alternative approaches and future directions. *Int. J. Biol. Macromol.* 221, 1428–1438. doi:10.1016/j.ijbiomac.2022.09.129
- Wang, S., Zhu, G., Jiang, D., Rhen, J., Li, X., Liu, H., et al. (2022c). Reduced Notch1 cleavage promotes the development of pulmonary hypertension. *Hypertension* 79 (1), 79–92. doi:10.1161/HYPERTENSIONAHA.120.16065
- Wang, W., He, Q., Li, C., Zhuang, C., Zhang, H., Wang, Q., et al. (2022a). Research on the mechanism and prevention of hypertension caused by apatinib through the RhoA/ROCK signaling pathway in a mouse model of gastric cancer. *Front. Cardiovasc Med.* 9, 873829. doi:10.3389/fcvm.2022.873829
- Wang, W., He, Q., Zhang, H., Zhuang, C., Wang, Q., Li, C., et al. (2021). A narrative review on the interaction between genes and the treatment of hypertension and breast cancer. *Ann. Transl. Med.* 9 (10), 894. doi:10.21037/atm-21-2133
- Wang, W., He, Q., Zhuang, C., Zhang, H., Fan, X., Wang, Q., et al. (2022b). Apatinib through activating the RhoA/ROCK signaling pathway to cause dysfunction of vascular smooth muscle cells. *Appl. Biochem. Biotechnol.* 194 (11), 5367–5385. doi:10.1007/s12010-022-04020-5
- Wheeler-Jones, C., Abu-Ghazaleh, R., Cospedal, R., Houliston, R. A., Martin, J., and Zachary, I. (1997). Vascular endothelial growth factor stimulates prostacyclin production and activation of cytosolic phospholipase A2 in endothelial cells via p42/p44 mitogen-activated protein kinase. *FEBS Lett.* 420 (1), 28–32. doi:10.1016/s0014-5793(97)01481-6
- Wu, Y., Fu, J., Huang, Y., Duan, R., Zhang, W., Wang, C., et al. (2023). Biology and function of pericytes in the vascular microcirculation. *Anim. Model Exp. Med.* 6 (4), 337–345. doi:10.1002/ame2.12334
- Wu, Y., Jiang, T., Hua, J., Xiong, Z., Dai, K., Chen, H., et al. (2022). PINK1/Parkin-mediated mitophagy in cardiovascular disease: from pathogenesis to novel therapy. *Int. J. Cardiol.* 361, 61–69. doi:10.1016/j.ijcard.2022.05.025
- Xin, N., Durieux, J., Yang, C., Wolff, S., Kim, H. E., and Dillin, A. (2022). The UPRmt preserves mitochondrial import to extend lifespan. *J. Cell Biol.* 221 (7), 221. doi:10.1083/jcb.202201071
- Xu, J., Shen, J., Gu, S., Zhang, Y., Wu, L., Wu, J., et al. (2021). Camrelizumab in combination with apatinib in patients with advanced hepatocellular carcinoma (rescue): a nonrandomized, open-label, phase II trial. *Clin. Cancer Res.* 27 (4), 1003–1011. doi:10.1158/1078-0432.CCR-20-2571
- Yang, Z., Qi, Y., Lai, N., Zhang, J., Chen, Z., Liu, M., et al. (2018). Notch1 signaling in melanoma cells promoted tumor-induced immunosuppression via upregulation of TGF- β 1. *J. Exp. Clin. Cancer Res.* 37 (1), 1. doi:10.1186/s13046-017-0664-4
- Zdravkovic, M., Popadic, V., Klasnja, S., Klasnja, A., Ivankovic, T., Lasica, R., et al. (2023). Coronary microvascular dysfunction and hypertension: a bond more important than we think. *Med. Kaunas.* 59 (12), 2149. doi:10.3390/medicina59122149
- Zhao, L., Ben-Yair, R., Burns, C. E., and Burns, C. G. (2019). Endocardial notch signaling promotes cardiomyocyte proliferation in the regenerating zebrafish heart through wnt pathway antagonism. *Cell Rep.* 26 (3), 546–554. doi:10.1016/j.celrep.2018.12.048
- Zhao, Y., Wang, Q., Zhao, X., Meng, H., and Yu, J. (2018). Effect of antihypertensive drugs on breast cancer risk in female hypertensive patients: evidence from observational studies. *Clin. Exp. Hypertens.* 40 (1), 22–27. doi:10.1080/10641963.2017.1288736
- Zhou, B., Lin, W., Long, Y., Yang, Y., Zhang, H., Wu, K., et al. (2022). Notch signaling pathway: architecture, disease, and therapeutics. *Signal Transduct. Target Ther.* 7 (1), 95. doi:10.1038/s41392-022-00934-y



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Shenfu injection: a review of pharmacological effects on cardiovascular diseases

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Shenfu injection (SFI), composed of ginseng and aconite, is a Chinese patent developed from the classic traditional prescription Shenfu Decoction created more than 700 years ago. SFI has been widely used in China for over 30 years for treating cardiovascular diseases. The main components in it include ginsenosides and aconitum alkaloids. In recent years, the role of SFI in the treatment of cardiovascular diseases has attracted much attention. The pharmacological effects and therapeutic applications of SFI in cardiovascular diseases are summarized here, highlighting pharmacological features and potential mechanisms developments, confirming that SFI can play a role in multiple ways and is a promising drug for treating cardiovascular diseases.

KEYWORDS

Shenfu injection, cardiovascular disease, heart failure, pharmacology, TCM (traditional Chinese medicine), clinical trial

1 Introduction

Cardiovascular diseases (CVDs) remain the predominant cause of mortality and morbidity worldwide over the past 20 years, including atherosclerosis, coronary heart disease, arrhythmia, hypertension, cardiomyopathy, stroke and heart failure (Parikh et al., 2018; Benjamin et al., 2019; Feng et al., 2019; Makhmudova et al., 2021). According to the World Health Organization (WHO) Report 2021, noncommunicable diseases (NCDs) kill more than 40 million people every year, and CVDs are the world's leading cause of death, accounting for almost one in three of all reported deaths globally. Data from the World Heart Report 2023 shows that 20.5 million people died from CVDs in 2021 (Mariappan et al., 2023). CVDs are caused by a variety of pathological factors, such as atherosclerosis, hypertension, hyperlipidemia, diabetes mellitus and so on, associated with energy metabolism disorder, mitochondrial structure abnormality, oxidative stress injury, cardiomyocyte apoptosis, inflammatory reaction, but the specific pathogenesis has not yet been fully elucidated (Parikh et al., 2018; Benjamin et al., 2019; Feng et al., 2019; Makhmudova et al., 2021). Based on the complex pathophysiological mechanisms, there are numerous drugs recommended for the treatment of CVDs, including angiotensin-converting enzyme inhibitors, angiotensin receptor antagonists, β -receptor antagonists, vasodilators, diuretics, α -receptor antagonists, positive inotropes, lipid-lowering drugs, antiarrhythmics, calcium channel blockers, etc. However, their potential serious adverse effects caused by these drug, such as hyperkalemia, cardiac depression, and electrolyte disturbance, cannot be ignored



FIGURE 1
Shenfu injection.

(Alhawassi et al., 2018; Núñez-Acevedo et al., 2018; Pall et al., 2021). Therefore, folk medicine is widely used to treat CVDs, among which traditional Chinese medicine (TCM) is well known in the world. Along with the long history of development for TCM, some classic recipes for the treatment of CVDs have been used in the clinic since then. Zhigancao Decoction, originated from *Treatise on Febrile Diseases* in the Eastern Han Dynasty (25–280 AD), is composed of *Glycyrrhiza uralensis* Fisch [Leguminosae; Glycyrrhizae Radix et Rhizoma], *Zingiber officinale* Rosc [Zingiberaceae; Zingiberis Rhizoma Recens], *Cinnamomum cassia* Presl [Lauraceae; Cinnamomi Ramulus], *Panax ginseng* C.A.Mey [Araliaceae; Ginseng Radix et Rhizoma Rubra], *Rehmannia glutinosa* Libosch [Scrophulariaceae; Rehmanniae Radix], *Equus asinus* L [Equidae; Asini Corii Colla], *Ophiopogon japonicus* (L.f) Ker-Gawl [Liliaceae; Ophiopogonis Radix], *Cannabis sativa* L [Moraceae; Cannabis Fructus], *Ziziphus jujuba* Mill [Rhamnaceae; Jujubae Fructus], and used to treat arrhythmia and heart failure (Xiong, 2019; Zhang N. et al., 2021; Wu et al., 2021; Yang Y. et al., 2022). Xuefu Zhuyu Decoction, recorded in the classic *Yi Lin Gai Cuo* in the Qing dynasty (1830 AD), composed of eleven commonly used herbs, including *Prunus persica* (L.) Batsch [Rosaceae; Persicae Semen], *Carthamus tinctorius* L [Compositae; Carthami Flos], *Angelica sinensis* (Oliv.) Diels [Umbelliferae; Angelicae Sinensis Radix], *Rehmannia glutinosa* Libosch [Scrophulariaceae; Rehmanniae Radix], *Achyranthes bidentata* Bl [Amaranthaceae; Achyranthis Bidentatae Radix], *Ligusticum chuanxiong* Hort [Umbelliferae; Chuanxiong Rhizoma], *Platycodon grandiflorum* (Jacq.) A. DC [Campanulaceae; Platycodonis Radix], *Paeonia lactiflora* Pall [Ranunculaceae; Paeoniae Radix Rubra], *Citrus aurantium* L [Rutaceae; Aurantii Fructus], *Glycyrrhiza uralensis* Fisch [Leguminosae; Glycyrrhizae Radix et Rhizoma], *Bupleurum chinense* DC [Umbelliferae; Bupleuri Radix], is used to treat hyperlipidemia and coronary heart disease (Wang and Qiu, 2019; Zhang S. et al., 2021; Yang et al., 2023). Zhenwu Decoction, was firstly recorded in *Treatise on Febrile Diseases*. It includes five herbs: *Poria cocos* (Schw.) Wolf [Polyporaceae; Poria], *Paeonia lactiflora* Pall [Ranunculaceae; Paeoniae Radix Alba], *Zingiber officinale* Rosc [Zingiberaceae; Zingiberis Rhizoma Recens], *Aconitum carmichaelii* Debx



FIGURE 2
Ginseng Radix et Rhizoma Rubra.



FIGURE 3
Aconiti lateralis radix praeparata.

[Ranunculaceae; Aconiti Lateralis Radix Praeparata], *Atractylodes macrocephala* Koidz [Compositae; Atractylodis Macrocephalae Rhizoma], which is applied to treat chronic heart failure (Tang et al., 2018; Han et al., 2022). It is believed in TCM that CVDs is related to the imbalance of Qi, Xue, Yin and Yang in the human body. When Qi and Yang is insufficient, CVDs are prone to occur.

SFI (Figure 1) is widely used in China to treat numerous ailments, including shock (Zhang X. et al., 2020; Wang et al., 2022; Zhang and Li, 2023), pulmonary fibrosis (Liu et al., 2021), sepsis (Luo et al., 2021; Li X. et al., 2022; Xu et al., 2022), pneumonia (Niu et al., 2021; Shi et al., 2022), cancer (Gao and Zhang, 2023; Wen et al., 2023), cerebral infarction (Zhou et al., 2020), CVDs, and has shown promising results. With development of pharmacological research, SFI has been identified as an effective drug for the treatment of CVDs. This paper reviews the latest reports in the past 20 years (2003–2022) from PubMed, Web of Science, and National Knowledge Infrastructure (CNKI) using the keywords “Shenfu injection” and “cardiovascular diseases”. The pharmacological action and therapeutic application of SFI in treating CVDs were discussed, and its pharmacological characteristics and potential mechanism was emphasized.

2 SFI -basic characteristics and history of use

SFI is a commonly used traditional Chinese medicine injection that has been used in clinical for over 30 years (Liu et al., 2021). It originated from the traditional Chinese classical formula “Shenfu Decoction”, which was first recorded in *Yan’s Prescriptions for Rescuing Lives* in the Song Dynasty (1253 AD). SFI is composed of *Panax ginseng* C.A.Mey [Araliaceae; *Ginseng radix et rhizoma rubra*] (RG) (Figure 2) and *Aconitum carmichaelii* Debx [Ranunculaceae; *Aconiti lateralis radix praeparata*] (RA) (Figure 3), which has the function of restoring Yang and invigorating Qi (Pei et al., 2021; Zhou et al., 2022). The existing studies reported that RG can be used to treat coronary heart disease and atherosclerosis by reducing blood lipid levels and improving inflammation (Hernández-García et al., 2019; Lu et al., 2019; Im, 2020). Additionally, it can inhibit arrhythmia by affecting the ion channels, such as activating potassium channel while blocking calcium channel and sodium current (Liu Z. et al., 2019; Gou et al., 2020). Furthermore, by ameliorating mitochondrial function and reducing oxidative damage in cardiomyocytes, it can prevent ventricular remodeling and heart failure. Moreover, it has the potential to improve the function of vascular endothelial cells, thereby lowering blood pressure (Yang F. et al., 2022; Liu et al., 2022). Meanwhile, RA has cardiotonic effects by accelerating β -adrenergic receptor synthesis (Tong et al., 2021), has anti-inflammatory effects through the Toll-like receptor4/Nuclear factor κ B (TLR4/NF- κ B) pathway (Yan et al., 2020), and has anti-arrhythmic effects (Wang et al., 2023). SFI, composed of RA and RG, is a common drug for the treatment of CVDs.

Modern chemical studies have shown that SFI mainly contains ginsenosides, aconite alkaloids, organic acids, nucleosides, amino acids and other components (Song et al., 2015). Ginsenosides and aconite alkaloids are the main active components of SFI. The content of ginsenosides is 676–742 μ g/mL, and the content of aconite alkaloids is 3–7 μ g/mL (Yang et al., 2014; Ge et al., 2015; Song et al., 2015). It is known that aconite has certain toxicity, and the use of RG and RA in combination can achieve the effect of potentiation and detoxification. Ginsenosides can promote the metabolism of the toxic component aconitine, prolong the elimination half-life of active ingredients such as hypaconitine, benzoylmesaconine and songorine, and significantly increase the *in vivo* exposure of active ingredients. At the same time, some studies have found that ginseng can inhibit the ion disorders, toxicity in calcineurin-nuclear factor of activated T cells (CaN-NFAT3) pathway and inhibition of the cytochrome P450 2J3 (CYP2J3) expression caused by aconitine, and enhance the antioxidant effect of myocardial cells (Liu et al., 2020; Yang et al., 2021; Chen Z. Y. et al., 2022; Bao et al., 2023). Therefore, the compatibility of aconite and ginseng has the effect of ‘reducing toxicity and increasing efficiency’.

3 Bioavailability and metabolism of SFI

Pharmacokinetic data of rodents show that aconitum alkaloids can be rapidly eliminated after intravenous injection of SFI. Protopanaxatriol (PPT) ginsenosides such as ginsenoside Re (Figure 4), Rg1 (Figure 5) and Rg2 (Figure 6) can be rapidly excreted into bile when ginsenosides was given to rats (Cai et al., 2022). The elimination rate of protopanaxadiol ginsenosides such as

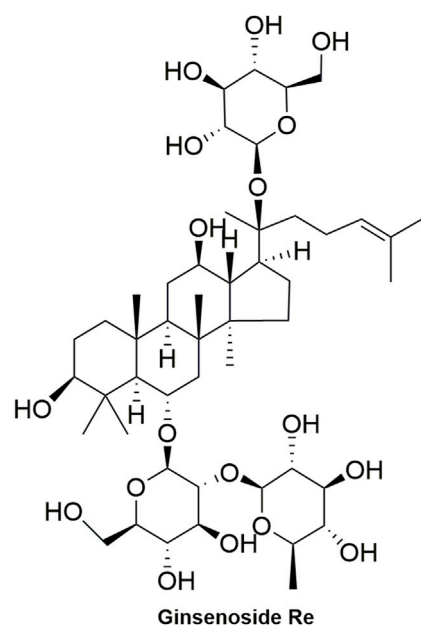


FIGURE 4
Structural formula of ginsenoside Re.

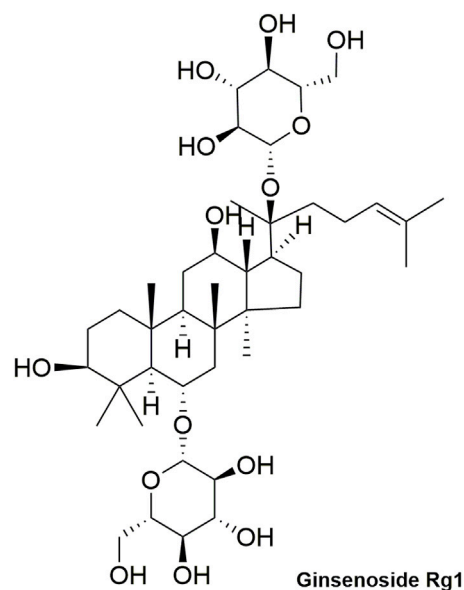
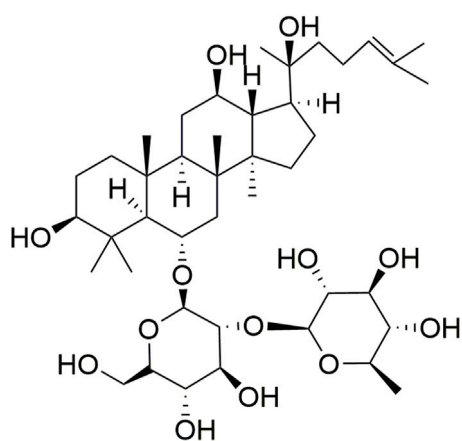


FIGURE 5
Structural formula of ginsenoside Rg1.

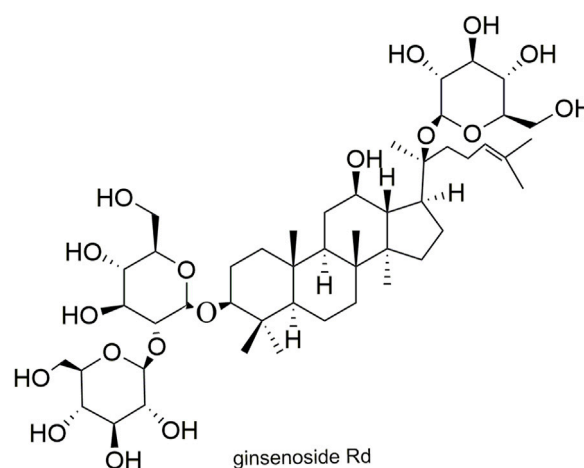
ginsenoside Rb1 (Figure 7), Rd (Figure 8) and Rh2 (Figure 9) is slower than that of PPT ginsenosides (Li et al., 2015; Zhang et al., 2016; Shen et al., 2021). The pharmacokinetic properties of ginsenosides (ginsenoside Rg1, ginsenoside Rb1, ginsenoside Rc (Figure 10)) and aconitine alkaloids (benzoylmesaconine (Figure 11), aconitine (Figure 12)) in SFI showed a linear relationship in the dose range of 2–8 mL/kg (Zhang et al., 2016; Li S. et al., 2022).

Modern studies have shown that SFI is almost safe at conventional therapeutic doses, and the incidence of adverse reactions is relatively low



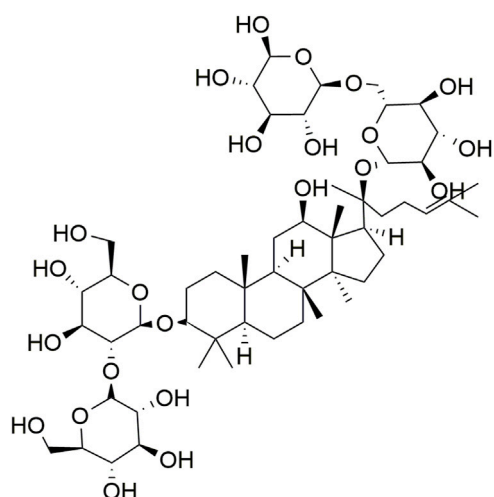
Ginsenoside Rg2

FIGURE 6
Structural formula of ginsenoside Rg2.



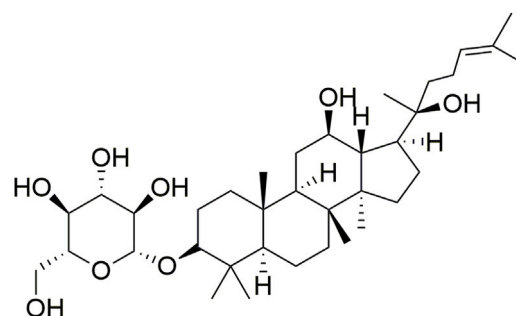
ginsenoside Rd

FIGURE 8
Structural formula of ginsenoside Rd.



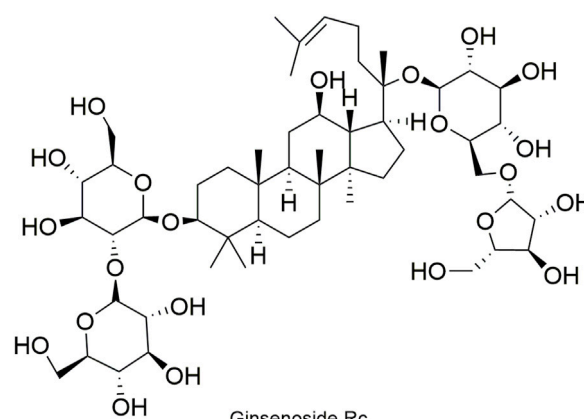
Ginsenoside b1

FIGURE 7
Structural formula of ginsenoside Rb1.



Ginsenoside Rh2

FIGURE 9
Structural formula of ginsenoside Rh2.



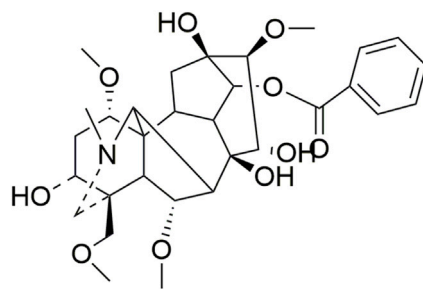
Ginsenoside Rc

FIGURE 10
Structural formula of ginsenoside Rc.

(0.076%), such as rash, itching, nausea, vomiting, dizziness, abdominal pain, and palpitation (Wang Z. F. et al., 2017).

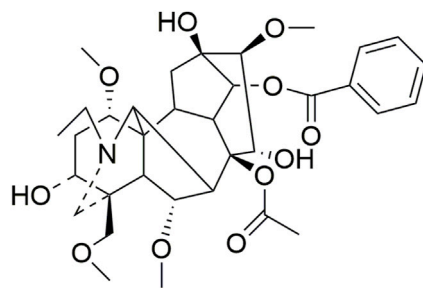
4 Pharmacological activities of SFI on CVDs

Many studies have confirmed that SFI has therapeutic effects on a variety of CVDs, such as myocardial hypertrophy, heart failure, ischemia-reperfusion injury, cardiac arrest, and arrhythmia. Its mechanism of action is mainly related to reducing inflammation



Benzoylmesaconine

FIGURE 11
Structural formula of benzoylmesaconine.



Aconitine

FIGURE 12
Structural formula of aconitine.

through NF- κ B signaling pathway, oxidative stress by reducing free radical damage, dilating blood vessels by increasing nitric oxide (NO) content, decreasing fibrosis through TGF- β /Smads signaling pathway and reducing apoptosis by increasing the expression of apoptosis proteins (Figure 13).

4.1 Cardiac hypertrophy and heart failure

Cardiac hypertrophy is mainly manifested as thickening of ventricular walls and an increase in cardiomyocyte size, closely related to cardiac fibrosis and heart failure (Feng et al., 2019). As time progresses and in settings of sustained stress, cardiac hypertrophy and fibrosis will eventually lead to heart failure (Gallo et al., 2019; Zhao D. et al., 2021; Methatham et al., 2021). Inhibiting cardiac hypertrophy and fibrosis is an effective way to treat heart failure.

TGF- β /Smads plays a key role in the pathogenesis of myocardial fibrosis. The previous research suggested that TGF- β 1 bind to receptor, recruited and phosphorylated type I receptor, induced phosphorylation of Smad2 and Smad3. The phosphorylated Smad2 and Smad3 formed a trimer complex with Smad4. Then the complex transferred into the nucleus and regulated the transcription of target genes, regulating the synthesis of collagen fibers and the activation of fibroblasts. Smad7 can competitively bind to the type I receptor of TGF- β 1 and inhibit the signal transduction of TGF- β 1/Smads pathway (Stewart et al., 2018; Wang L. et al., 2021). Ni et al.

(2017) found that SFI can effectively improve cardiac function in the rat model of congestive heart failure (CHF) and attenuate ventricular remodeling and myocardial fibrosis by regulating TGF- β /Smads signaling pathway, upregulating Smad7 and downregulating TGF- β 1, Smad2 and Smad3 gene expression.

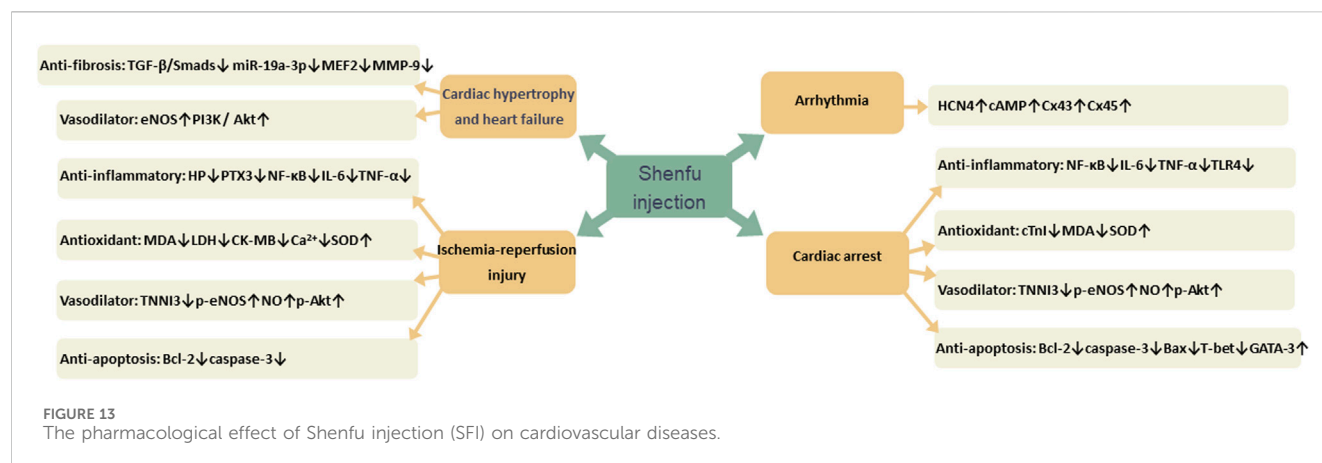
Inflammatory response is triggered in the myocardial infarction area which can result in cardiac remodeling and heart failure, accumulating high levels of monocytes and neutrophils (Halade and Lee, 2022). Inflammatory cells can also stimulate repair pathways, with increasing the content of extracellular matrix in the myocardium, including matrix metalloproteinase-9 (MMP-9) and collagen (Valiente-Alandi et al., 2018). SFI has anti-inflammatory effect, which can reduce the content of inflammatory factors such as tumor necrosis factor- α (TNF- α), interleukin-1 β (IL-1 β) and interleukin-6 (IL-6) in serum of rats, and also reduce the expression of fibronectin, collagen I, collagen III and MMP-9 protein (Ni et al., 2017; Guo et al., 2022).

Endothelial function is essential for maintaining normal vasomotor function, and disruption of this function can result in vasomotor dysfunction (Sabe et al., 2022). When heart failure occurs, endothelial function is impaired, with vasoconstrictor substances increasing, and vasodilator substances decreasing (Monteiro et al., 2019). NO is the most famous vasodilator, while endothelin-1 (ET-1) is the most widely recognized vasoconstrictor (Miyauchi and Sakai, 2019). The production of NO mainly depends on the activity and quantity of endothelial nitric oxide synthase (eNOS). When the expression of eNOS mRNA increases, the number of eNOS synthesis increases. When eNOS binds with calmodulin (CaM), the activity of eNOS enhance, adversely, when binds with caveolin-1 (Cav-1), decrease. The phosphorylation of eNOS depends on the phosphatidylinositol 3-kinase/protein kinase B (PI3K-Akt) signaling pathway, which produces phosphorylated tyrosine residues, thereby provides an anchor site for the recruitment of PI3K to the membrane (Garcia and Sessa, 2019; Suvorava et al., 2022). Zhu et al. (2020) found that SFI can increase the expression of eNOS mRNA and CaM and promote NO synthesis, decrease the expression of Cav-1 and ET-1 content, promote eNOS phosphorylation via the PI3K/Akt signaling pathway.

MicroRNAs (miRNAs), encoded by myosin heavy chain (MHC) genes, are important regulatory factors of CVDs (Omidkhoda et al., 2019). MiR19a-3p plays an important role in cardiac hypertrophy. Myocytes specific enhancer factor 2A (MEF2A) is a target gene of miR-19a-3p and is highly expressed in cardiac hypertrophy while miR-19a-3p has low expression (Mao et al., 2018). SFI can upregulate the expression of miR-19a-3p, and downregulate the expression of MEF2A and β -myosin heavy chain (β -MHC), so attenuate cardiac hypertrophy (Mao et al., 2018).

4.2 Ischemia-reperfusion injury

Myocardial ischemia-reperfusion injury is a significant factor that has a negative impact on the prognosis of myocardial infarction patients, causing myocardial stunning, no-reflow phenomena, reperfusion arrhythmia, and even permanent cardiomyocyte death. Therefore, it is critical to understand the mechanism of myocardial ischemia reperfusion and develop efficient treatments (Mokhtari-Zaer et al., 2018; Deng, 2021).



Apoptosis is involved in the pathogenesis of various CVDs and plays an important role in myocardial ischemia-reperfusion injury. Bcl-2 family proteins are important regulators of the process, prevent apoptosis by acting upstream of apoptosis proteins, such as caspase-3 and caspase-9 (King et al., 2023; Sahoo et al., 2023). Previous research found that SFI can upregulate the anti-apoptosis protein Bcl-2 and inhibit the consecutive activation of caspase-3 and caspase-9, both of which are intimately associated to apoptosis (Cao et al., 2005; Wang et al., 2009; Guo et al., 2016).

Oxidative stress is a risk factor for CVDs, and abnormally increased reactive oxygen species (ROS) is the main cause of oxidative stress. ROS combined with proteins and lipids damage cardiomyocytes (Peng et al., 2022). The superoxide dismutase (SOD) is the major antioxidant enzyme that degrade superoxide (Eleutherio et al., 2021). The glutathione system is widely recognized as one of the most potent endogenous antioxidant systems within cardiovascular system. Glutathione, one of the endogenous antioxidant molecules, can directly scavenge ROS caused by myocardial ischemia (Panday et al., 2020; Tan et al., 2023). Taurine is a common endogenous sulfur-containing amino acid with antioxidant activity and can inhibit the abnormal increase of ROS (Li et al., 2020). SFI dramatically decreased glutathione and taurine, increased SOD activity, and inhibited the rise in malondialdehyde (MDA), which is closely related to oxidative stress (Zheng et al., 2004; Cao et al., 2005; Wu et al., 2019).

Numerous studies have revealed that NO is a vasodilator with the ability to operate on cardiomyocytes and vascular endothelium via a variety of signaling pathways (Boycott et al., 2020; Cyr et al., 2020). The action of eNOS is primarily responsible for NO generation. NO generated by eNOS phosphorylation induces soluble guanylate cyclase (sGC) to create cyclic guanosine monophosphate (cGMP), a second messenger with cardiovascular protective properties (Mount et al., 2007; Zhang Q. et al., 2020; Lee et al., 2021). SFI activated eNOS phosphorylation via Akt, thereby promoting the production of NO (Wu et al., 2011; Wang et al., 2018).

4.3 Cardiac arrest

Cardiac arrest (CA), one of the leading causes of death, has a significant impact on the public health, particularly due to its persistent increase worldwide (Vazquez and Sudhir, 2023). Post-cardiac arrest

syndrome (PCAS) is a group of diseases characterized by systemic ischemia/reperfusion injury, hypoxic brain injury and myocardial dysfunction after cardiac arrest (Jou et al., 2020). It is associated with cardiovascular ischemia/reperfusion injury and cardiovascular toxicity, including factors such as excessive activation of inflammatory cytokines and catecholamines (Lazzarin et al., 2022). Matrix metalloproteinases, tumor necrosis factor, and interleukins each have a special prognostic function in PCAS. High inflammatory cytokine levels have been linked to poor neurologic and/or death outcomes (Jou et al., 2020).

NF-κB signaling pathway is one of the important pathways regulating inflammation and plays a key regulatory role in the occurrence and development of various CVDs (Cheng et al., 2023). Study have reported that in cardiac arrest swine, SFI remarkably decreased levels of many inflammatory cytokines, such as TNF-α, IL-6, mRNA and protein levels of myocardial TLR4 and NF-κB (Gu et al., 2021). TLR4, as a 'portal' protein, regulates the initiation of the inflammatory chain reaction of the body's immunity and mediates the inflammatory response (Fitzgerald and Kagan, 2020).

Na⁺-K⁺-ATPase enzyme and Ca²⁺-ATPase enzyme, ubiquitous enzymes in the heart, play a crucial role in process of CVDs (Fedosova et al., 2021). Na⁺-K⁺-ATPase transports two Na⁺ ions extrude out of the cell in exchange for one K⁺ ions, thereby maintaining the concentration gradients across the cell membrane (Fedosova et al., 2021; Obradovic et al., 2023). Ca²⁺-ATPase, a crucial role for cellular Ca²⁺ homeostasis, maintains normal intracellular calcium concentration and prevents calcium overload (Nguyen et al., 2023; Ye et al., 2023). It is reported that SFI increased Na⁺-K-ATPase and Ca²⁺-ATPase activity (Ji et al., 2011).

4.4 Arrhythmia

The normal electrical activity of the heart is initiated by special pacemaker cells located in the sinoatrial node (Liang et al., 2021). Dysfunction or loss of pacemaker cells can cause arrhythmia (Liu and Yuan, 2021b). The transplantation of stem cells is regarded as a kind of feasible treatment for arrhythmia (Sattayaprasert et al., 2020). It has been reported that bone marrow mesenchymal stem cells (BMSCs) with specific phenotypes can be transformed into

TABLE 1 Clinical trial of SFI in cardiovascular disease.

| Disease | Number of patients | Dose of SFI (mL) | Duration | Route of administration | Outcome measures | References |
|-----------------------|--------------------|------------------|----------|-------------------------|--|---------------------|
| acute heart failure | 80 patients | 50 | 7 days | i.v | cardiac function, clinical symptoms and quality of life | Wang et al. (2019b) |
| | 55 patients | 100 | 24 h | i.v | CI, cardiac output and stroke volume index | Li et al. (2022a) |
| Chronic heart failure | 80 patients | 50 | 7 days | i.v | LVEF, LVED, BNP,Fas, TNF- α , IL-6, mortality, readmission rate | Liu et al. (2015) |
| | 171 patients | 60 | 2 weeks | i.v | the all-cause mortality,6MWT | Wang et al. (2017a) |
| | 55 patients | 50 | 7 days | i.v | Cardiacfunction, LVEF, NT-proBNP, TNF- α , IL-6 | Gao et al. (2021) |
| | 91 patients | 40 | 7 days | i.v | Cardiac function, LVEF, NT-proBNP, BNP | Guo et al. (2021) |
| coronary syndrome | 74 patients | 40 | 4–6 h | i.v | the level of NGAL in urine | Guo et al. (2017) |
| myocardial infarction | 20 patients | 80 | 5 days | i.v | the area of myocardial infarction | Wang et al. (2021b) |
| cardiac arrest | 492 patients | 100 | 28 days | i.v | 28-day and 90-day survival rates, the mechanical ventilation time and hospitalization time | Zhang et al. (2017) |

pacemaker-like cells after special treatment (Chauveau et al., 2014). Moreover, HMSCs possess the capability to regulate arrhythmia substrates by altering their secretory groups in diseases (Sattayaprasert et al., 2020).

In vitro, SFI can activate inward pacemaker current of BMSCs in a concentration-dependent manner, increase HCN4 expression and cAMP content in BMSCs, induce BMSCs proliferation, promote their differentiation into pacemaker-like cells (Zhao X. et al., 2021). The HCN4 gene serves as the molecular basis for the pacemaker current, contributing significantly to inward current during depolarization and playing a crucial role in the generation and autonomous regulation of heart rate (Bucchi et al., 2012; D’Souza et al., 2021; Hoekstra et al., 2021). Bone marrow mesenchymal stem cells treated with SFI retained the function of sinoatrial node in rabbits with sinoatrial node syndrome, improved the expression of HCN4 gene and gap junction proteins (Cx43 and Cx45), and significantly upregulated the expression of cAMP in sinoatrial node (Chen Q. et al., 2022).

In addition, SFI has certain pharmacological effects on nervous system, respiratory system and digestive system. For example, SFI has a protective effect on lipopolysaccharide-induced septic shock in rabbits (Liu X. et al., 2019). It can reduce bile duct injury in rats with acute obstructive cholangitis (Tan et al., 2019) and increase the level of acetylcholine in acute liver injury in septic young rats (Wu et al., 2022). It also has a protective effect on lung and intestinal epithelial injury in mice with acute gastrointestinal injury (Zheng et al., 2022).

5 Clinical trial of SFI in CVDs

There are many clinical trials related to SFI, and nine clinical trials have been conducted to study its role in the treatment of CVDs. These clinical trials have demonstrated that SFI can improve cardiac function and corresponding indicators in patients with

CVDs, including heart failure, myocardial infarction, cardiac arrest after resuscitation, coronary syndrome, coronary heart disease and other diseases (Table 1).

5.1 The effect of SFI in patients with acute heart failure

Infusion of SFI in 80 patients with acute heart failure can improve cardiac function, clinical symptoms and quality of life (Wang et al., 2019b). Fifty patients with acute decompensated heart failure were treated with combination therapy. Compared with simple infusion of levosimendan, the improvement of hemodynamic parameters including CI, cardiac output and stroke volume index was more significant, especially in patients with acute decompensated heart failure with hypotension (Li M. et al., 2022).

5.2 The effect of SFI in patients with chronic heart failure

SFI was used to treat 80 patients with acute exacerbation of chronic heart failure, which could improve the symptoms, quality of life, exercise tolerance, improve left ventricular ejective fraction (LVEF), reduce left ventricular end diastolic diameter (LVED), plasma brain natriuretic peptide (BNP) and cytokine Fas, TNF- α , IL-6 levels, reduce mortality and readmission rate (Liu et al., 2015). SFI was administered to 171 patients suffering from chronic heart failure on the basis of Western medicine. Compared with Western medicine alone, it could reduce the all-cause mortality by 30.99%, increase the 6-min walking distance (6MWT) and improve the quality of life (Wang X. et al., 2017). Patients with coronary heart disease complicated with chronic heart failure were treated

with SFI and furosemide injection for 7 days. The effect was better than that of furosemide injection alone in improving cardiac function, LVEF, N-terminal B-type natriuretic peptide (NT-proBNP), TNF- α , IL-6 (Gao et al., 2021). For 7 days, SFI and sodium nitroprusside were administered intravenously to 91 patients who had coronary heart disease and chronic heart failure. The effect was better than that of sodium nitroprusside injection alone in improving cardiac function, LVEF, NT-proBNP and BNP (Guo et al., 2021).

5.3 The effect of SFI in patients with other CVDs

Infusion of SFI 1h before coronary angiography in 74 patients with coronary syndrome undergoing percutaneous coronary intervention (PCI) significantly reduced the level of neutrophil gelatinase-associated lipocalin (NGAL) in urine and effectively prevent contrast-induced acute kidney injury (Guo et al., 2017). SFI was used to treat patients with ST-segment elevation myocardial infarction before PCI and maintained for 5 days after PCI. Compared with patients treated with placebo, SFI reduced the area of myocardial infarction (Wang X. et al., 2021). A total of 492 cardiac arrest patients received bi-daily intravenous SFI infusions over a span of 28 days. The 28-day and 90-day survival rates were improved, the mechanical ventilation time and hospitalization time were shortened, and the recovery of spontaneous circulation after cardiac arrest was effectively improved (Zhang et al., 2017).

6 Concluding remarks and future perspectives

The results of numerous research studies in the past have demonstrated that SFI exerts varying degrees of therapeutic effects on various types of CVDs, such as heart failure, myocardial hypertrophy, myocardial ischemia, cardiac arrest, arrhythmia, and so forth. SFI plays a therapeutic role through multiple different targets, such as TGF-Smads, PI3K-Akt, eNOS-Akt pathway, and so on. SFI has been used in China for more than

30 years. It is a commonly used drug for clinical treatment of CVDs. No serious adverse reactions have been found so far.

In general, SFI is a promising drug for the treatment of CVDs. However, SFI has the characteristics of multi-component, multi-target and multi-pathway, which increases the difficulty of research. There is still a lack of in-depth study on the mechanism of SFI. In addition, large-scale, high-quality, multi-center clinical trials are needed to determine the comparison of SFI with traditional CVDs treatment regimens.

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F-FX: Writing—original draft. X-FX: Writing—original draft. H-YH: Writing—original draft. R-ST: Writing—review and editing. CP: Writing—review and editing.

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Conflict of interest

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References

- Alhawassi, T. M., Krass, I., and Pont, L. G. (2018). Antihypertensive-related adverse drug reactions among older hospitalized adults. *Int. J. Clin. Pharm.* 40, 428–435. doi:10.1007/s11096-017-0583-7
- Bao, Y., Zhang, R., Jiang, X., Liu, F., He, Y., Hu, H., et al. (2023). Detoxification mechanisms of ginseng to aconite: a review. *J. Ethnopharmacol.* 304, 116009. doi:10.1016/j.jep.2022.116009
- Benjamin, E. J., Muntner, P., Alonso, A., Bittencourt, M. S., Callaway, C. W., Carson, A. P., et al. (2019). Heart disease and stroke statistics-2019 update: a report from the American heart association. *Circulation* 139, e56–e528. doi:10.1161/cir.0000000000000659
- Boycott, H. E., Nguyen, M. N., Vrellaku, B., Gehmlich, K., and Robinson, P. (2020). Nitric oxide and mechano-electrical transduction in cardiomyocytes. *Front. Physiol.* 11, 606740. doi:10.3389/fphys.2020.606740
- Bucchi, A., Barbuti, A., Difrancesco, D., and Baruscotti, M. (2012). Funny current and cardiac rhythm: insights from HCN knockout and transgenic mouse models. *Front. Physiol.* 3, 240. doi:10.3389/fphys.2012.00240
- Cai, J., Huang, K., Han, S., Chen, R., Li, Z., Chen, Y., et al. (2022). A comprehensive system review of pharmacological effects and relative mechanisms of Ginsenoside re: recent advances and future perspectives. *Phytomedicine* 102, 154119. doi:10.1016/j.phymed.2022.154119
- Cao, J., Zheng, C. D., Zhang, G. X., Zhang, Y. J., and Min, S. (2005). Protective effect of Shenfu injection on myocardial mitochondria injured by ischemia-reperfusion in rabbits. *Chin. Med. J. Engl.* 118, 505–507.
- Chauveau, S., Brink, P. R., and Cohen, I. S. (2014). Stem cell-based biological pacemakers from proof of principle to therapy: a review. *Cytotherapy* 16, 873–880. doi:10.1016/j.jcyt.2014.02.014
- Chen, Q., Kang, L., Li, Y., Lin, Z., Chu, Q., Cai, Y., et al. (2022a). Effect of shenfu injection on differentiation of bone marrow mesenchymal stem cells into pacemaker-like cells and improvement of pacing function of sinoatrial node. *Oxid. Med. Cell. Longev.* 2022, 4299892. doi:10.1155/2022/4299892
- Chen, Z. Y., Wei, X. Y., Qiu, Z. D., Huang, Y., Tan, T., Feng, Y. L., et al. (2022b). Compatibility of fuzi and ginseng significantly increase the exposure of aconitines. *Front. Pharmacol.* 13, 883898. doi:10.3389/fphar.2022.883898
- Cheng, W., Cui, C., Liu, G., Ye, C., Shao, F., Bagchi, A. K., et al. (2023). NF- κ B, A potential therapeutic target in cardiovascular diseases. *Cardiovasc. Drugs Ther.* 37, 571–584. doi:10.1007/s10557-022-07362-8

- Cyr, A. R., Huckaby, L. V., Shiva, S. S., and Zuckerbraun, B. S. (2020). Nitric oxide and endothelial dysfunction. *Crit. Care Clin.* 36, 307–321. doi:10.1016/j.ccc.2019.12.009
- Deng, J. (2021). Advanced research on the regulated necrosis mechanism in myocardial ischemia-reperfusion injury. *Int. J. Cardiol.* 334, 97–101. doi:10.1016/j.ijcard.2021.04.042
- D'Souza, A., Wang, Y., Anderson, C., Bucchi, A., Baruscotti, M., Olieslagers, S., et al. (2021). A circadian clock in the sinus node mediates day-night rhythms in Hcn4 and heart rate. *Heart rhythm.* 18, 801–810. doi:10.1016/j.hrthm.2020.11.026
- Eleutherio, E. C. A., Silva Magalhães, R. S., de Araújo Brasil, A., Monteiro Neto, J. R., and de Holanda Paranhos, L. (2021). SOD1, more than just an antioxidant. *Arch. Biochem. Biophys.* 697, 108701. doi:10.1016/j.abb.2020.108701
- Fedosova, N. U., Habeck, M., and Nissen, P. (2021). Structure and function of Na,K-ATPase-The sodium-potassium pump. *Compr. Physiol.* 12, 2659–2679. doi:10.1002/cphy.c200018
- Feng, X., Sureda, A., Jafari, S., Memariani, Z., Tewari, D., Annunziata, G., et al. (2019). Berberine in cardiovascular and metabolic diseases: from mechanisms to therapeutics. *Theranostics* 9, 1923–1951. doi:10.7150/thno.30787
- Fitzgerald, K. A., and Kagan, J. C. (2020). Toll-like receptors and the control of immunity. *Cell* 180, 1044–1066. doi:10.1016/j.cell.2020.02.041
- Gallo, S., Vitacolonna, A., Bonzano, A., Comoglio, P., and Crepaldi, T. (2019). ERK: a key player in the pathophysiology of cardiac hypertrophy. *Int. J. Mol. Sci.* 20, 2164. doi:10.3390/ijms20092164
- Gao, W., and Zhang, K. (2023). Network meta-analysis of 8 types of traditional Chinese medicine injection combined with chemotherapy in colorectal cancer treatment. *J. Cancer Res. Clin. Oncol.* 149, 9823–9838. doi:10.1007/s00432-023-04892-y
- Gao, Y., Gao, Y., Zhu, R., and Tan, X. (2021). Shenfu injection combined with furosemide in the treatment of chronic heart failure in patients with coronary heart disease: a protocol of randomized controlled trial. *Med. Baltim.* 100, e24113. doi:10.1097/md.00000000000024113
- Garcia, V., and Sessa, W. C. (2019). Endothelial NOS: perspective and recent developments. *Br. J. Pharmacol.* 176, 189–196. doi:10.1111/bph.14522
- Ge, A. H., Li, J., Donnapée, S., Bai, Y., Liu, J., He, J., et al. (2015). Simultaneous determination of 2 aconitum alkaloids and 12 ginsenosides in Shenfu injection by ultraperformance liquid chromatography coupled with a photodiode array detector with few markers to determine multicomponents. *J. Food Drug Anal.* 23, 267–278. doi:10.1016/j.jfda.2014.10.013
- Gou, D., Pei, X., Wang, J., Wang, Y., Hu, C., Song, C., et al. (2020). Antiarrhythmic effects of ginsenoside Rg2 on calcium chloride-induced arrhythmias without oral toxicity. *J. Ginseng Res.* 44, 717–724. doi:10.1016/j.jgr.2019.06.005
- Gu, W., Hou, X. M., and Li, C. S. (2021). Effects of shenfu injection on inflammatory response during post-resuscitation myocardial dysfunction after cardiac arrest in swine. *Chin. J. Integr. Med.* 27, 417–423. doi:10.1007/s11655-021-2855-2
- Guo, B., Yang, T., Nan, J., Huang, Q., Wang, C., and Xu, W. (2021). Efficacy and safety of Shenfu injection combined with sodium nitroprusside in the treatment of chronic heart failure in patients with coronary heart disease: a protocol of randomized controlled trial. *Med. Baltim.* 100, e24414. doi:10.1097/md.00000000000024414
- Guo, F., Wang, X., Guo, Y., Wan, W., Cui, Y., Wang, J., et al. (2022). Shenfu administration improves cardiac fibrosis in rats with myocardial ischemia-reperfusion through adenosine A(2a) receptor activation. *Hum. Exp. Toxicol.* 41, 9603271221077684. doi:10.1177/09603271221077684
- Guo, Z., Niu, D., Yu, Y., Zhen, D., and Li, W. (2017). Effects of hydration combined with Shenfu injection on contrast-induced acute kidney injury in acute coronary syndrome patients undergoing percutaneous coronary intervention. *Biomed. Rep.* 7, 477–481. doi:10.3892/br.2017.986
- Guo, Z. J., Wu, C. J., and Li, C. S. (2016). Shen-Fu Injection alleviates post-resuscitation myocardial dysfunction by up-regulating expression of sarcoplasmic reticulum Ca(2+)-ATPase. *Chin. J. Integr. Med.* 22, 503–509. doi:10.1007/s11655-015-2156-8
- Halade, G. V., and Lee, D. H. (2022). Inflammation and resolution signaling in cardiac repair and heart failure. *EBioMedicine* 79, 103992. doi:10.1016/j.ebiom.2022.103992
- Han, Y., Huang, L., Zhong, G., Chang, X., Zhu, Q., Xu, M., et al. (2022). Evaluation of the safety and efficacy of Zhenwu decoction as adjuvant therapy for the treatment of heart failure with reduced ejection fraction: a protocol for systematic review and meta-analysis. *Med. Baltim.* 101, e28672. doi:10.1097/md.00000000000028672
- Hernández-García, D., Granado-Serrano, A. B., Martín-Gari, M., Naudí, A., and Serrano, J. C. (2019). Efficacy of Panax ginseng supplementation on blood lipid profile. A meta-analysis and systematic review of clinical randomized trials. *J. Ethnopharmacol.* 243, 112090. doi:10.1016/j.jep.2019.112090
- Hoekstra, M., van Ginneken, A. C. G., Wilders, R., and Verkerk, A. O. (2021). HCN4 current during human sinoatrial node-like action potentials. *Prog. Biophys. Mol. Biol.* 166, 105–118. doi:10.1016/j.biombio.2021.05.006
- Im, D. S. (2020). Pro-resolving effect of ginsenosides as an anti-inflammatory mechanism of Panax ginseng. *Biomolecules* 10, 444. doi:10.3390/biom10030444
- Ji, X. F., Yang, L., Zhang, M. Y., Li, C. S., Wang, S., and Cong, L. H. (2011). Shen-fu injection attenuates postresuscitation myocardial dysfunction in a porcine model of cardiac arrest. *Shock* 35, 530–536. doi:10.1097/SHK.0b013e31820e2058
- Jou, C., Shah, R., Figueroa, A., and Patel, J. K. (2020). The role of inflammatory cytokines in cardiac arrest. *J. Intensive Care Med.* 35, 219–224. doi:10.1177/0885066618817518
- King, L. E., Hohorst, L., and García-Sáez, A. J. (2023). Expanding roles of BCL-2 proteins in apoptosis execution and beyond. *J. Cell Sci.* 136, jcs260790. doi:10.1242/jcs.260790
- Lazzarin, T., Tonon, C. R., Martins, D., Fávero, E. L., Jr., Baumgratz, T. D., Pereira, F. W. L., et al. (2022). Post-cardiac arrest: mechanisms, management, and future perspectives. *J. Clin. Med.* 12, 259. doi:10.3390/jcm12010259
- Lee, G. H., Kim, C. Y., Zheng, C., Jin, S. W., Kim, J. Y., Lee, S. Y., et al. (2021). Rutacarpine increases nitric oxide synthesis via eNOS phosphorylation by TRPV1-dependent CaMKII and CaMKK β /AMPK signaling pathway in human endothelial cells. *Int. J. Mol. Sci.* 22, 9407. doi:10.3390/ijms22179407
- Li, M., Zhang, Y., Wan, Q., Li, Y., Qu, T., and Yuan, F. (2022a). Use of levosimendan combined with Shenfu injection to treat acute heart failure patients with hypotension: a prospective randomized controlled single-blind study. *BMC Cardiovasc Disord.* 22, 130. doi:10.1186/s12872-022-02572-2
- Li, S., Yu, L., Shi, Q., Liu, Y., Zhang, Y., Wang, S., et al. (2022b). An insight into current advances on pharmacology, pharmacokinetics, toxicity and detoxification of aconitine. *Biomed. Pharmacother.* 151, 113115. doi:10.1016/j.biopha.2022.113115
- Li, W., Yang, J., Lyu, Q., Wu, G., Lin, S., Yang, Q., et al. (2020). Taurine attenuates isoproterenol-induced H9c2 cardiomyocytes hypertrophy by improving antioxidative ability and inhibiting calpain-1-mediated apoptosis. *Mol. Cell. Biochem.* 469, 119–132. doi:10.1007/s11010-020-03733-7
- Li, X., Huang, F., Zhu, L., Luo, T., Zhang, Y., Gu, H., et al. (2022c). Effects of combination therapy with Shenfu Injection in critically ill patients with septic shock receiving mechanical ventilation: a multicentric, real-world study. *Front. Pharmacol.* 13, 1041326. doi:10.3389/fphar.2022.1041326
- Li, Z., Zhang, R., Wang, X., Hu, X., Chen, Y., and Liu, Q. (2015). Simultaneous determination of seven ginsenosides in rat plasma by high-performance liquid chromatography coupled to time-of-flight mass spectrometry: application to pharmacokinetics of Shenfu injection. *Biomed. Chromatogr.* 29, 167–175. doi:10.1002/bmc.3272
- Liang, D., Xue, Z., Xue, J., Xie, D., Xiong, K., Zhou, H., et al. (2021). Sinoatrial node pacemaker cells share dominant biological properties with glutamatergic neurons. *Protein Cell* 12, 545–556. doi:10.1007/s13238-020-00820-9
- Liu, C., Hou, Y., Wang, X., Zhao, Z., Liu, Z., Zhai, J., et al. (2015). Clinical assessment of Shenfu injection loading in the treatment of patients with exacerbation of chronic heart failure due to coronary heart disease: study protocol for a randomized controlled trial. *Trials* 16, 222. doi:10.1186/s13063-015-0729-7
- Liu, L., Hu, J., Mao, Q., Liu, C., He, H., Hui, X., et al. (2022). Functional compounds of ginseng and ginseng-containing medicine for treating cardiovascular diseases. *Front. Pharmacol.* 13, 1034870. doi:10.3389/fphar.2022.1034870
- Liu, M., Li, Y., Tang, Y., Zheng, L., and Peng, C. (2020). Synergistic effect of Aconiti Lateralis Radix Praeparata water-soluble alkaloids and Ginseng Radix et Rhizoma total ginsenosides compatibility on acute heart failure rats. *J. Chromatogr. B. Anal. Technol. Biomed. Life Sci.* 1137, 121935. doi:10.1016/j.jchromb.2019.121935
- Liu, P., Yang, S., Wang, Z., Dai, H., and Wang, C. (2021). Feasibility and mechanism analysis of shenfu injection in the treatment of idiopathic pulmonary fibrosis. *Front. Pharmacol.* 12, 670146. doi:10.3389/fphar.2021.670146
- Liu, X., Liu, R., Dai, Z., Wu, H., Lin, M., Tian, F., et al. (2019a). Effect of Shenfu injection on lipopolysaccharide (LPS)-induced septic shock in rabbits. *J. Ethnopharmacol.* 234, 36–43. doi:10.1016/j.jep.2019.01.008
- Liu, Y., and Yuan, X. (2021). Logic analysis of arrhythmia triggered by pacemaker special functions - an educational presentation. *Braz. J. Cardiovasc. Surg.* 36, 412–415. doi:10.21470/1678-9741-2020-0630
- Liu, Z., Song, L., Zhang, P., Cao, Z., Hao, J., Tian, Y., et al. (2019b). Ginsenoside Rb1 exerts antiarrhythmic effects by inhibiting I(Na) and I(CaL) in rabbit ventricular myocytes. *Sci. Rep.* 9, 20425. doi:10.1038/s41598-019-57010-9
- Lu, S., Luo, Y., Zhou, P., Yang, K., Sun, G., and Sun, X. (2019). Ginsenoside compound K protects human umbilical vein endothelial cells against oxidized low-density lipoprotein-induced injury via inhibition of nuclear factor- κ B, p38, and JNK MAPK pathways. *J. Ginseng Res.* 43, 95–104. doi:10.1016/j.jgr.2017.09.004
- Luo, S., Gou, L., Liu, S., and Cao, X. (2021). Efficacy and safety of Shenfu injection in the treatment of sepsis: a protocol for systematic review and meta-analysis. *Med. Baltim.* 100, e27196. doi:10.1097/md.00000000000027196
- Makhmudova, U., Schulze, P. C., Lütjohann, D., and Weingärtner, O. (2021). Phytosterols and cardiovascular disease. *Curr. Atheroscler. Rep.* 23, 68. doi:10.1007/s11883-021-00964-x
- Mao, Z. J., Zhang, Q. L., Shang, J., Gao, T., Yuan, W. J., and Qin, L. P. (2018). Shenfu Injection attenuates rat myocardial hypertrophy by up-regulating miR-19a-3p expression. *Sci. Rep.* 8, 4660. doi:10.1038/s41598-018-23137-4

- Mariappan, V., Srinivasan, R., Pratheesh, R., Jujjuvarapu, M. R., and Pillai, A. B. (2023). Predictive biomarkers for the early detection and management of heart failure. *Heart fail. Rev.* doi:10.1007/s10741-023-10347-w
- Methatham, T., Tomida, S., Kimura, N., Imai, Y., and Aizawa, K. (2021). Inhibition of the canonical Wnt signaling pathway by a β -catenin/CBP inhibitor prevents heart failure by ameliorating cardiac hypertrophy and fibrosis. *Sci. Rep.* 11, 14886. doi:10.1038/s41598-021-94169-6
- Miyauchi, T., and Sakai, S. (2019). Endothelin and the heart in health and diseases. *Peptides* 111, 77–88. doi:10.1016/j.peptides.2018.10.002
- Mokhtari-Zaer, A., Marefati, N., Atkin, S. L., Butler, A. E., and Sahebkar, A. (2018). The protective role of curcumin in myocardial ischemia-reperfusion injury. *J. Cell Physiol.* 234, 214–222. doi:10.1002/jcp.26848
- Monteiro, J. P., Bennett, M., Rodor, J., Caudrillier, A., Ulitsky, I., and Baker, A. H. (2019). Endothelial function and dysfunction in the cardiovascular system: the long non-coding road. *Cardiovasc. Res.* 115, 1692–1704. doi:10.1093/cvr/cvz154
- Mount, P. F., Kemp, B. E., and Power, D. A. (2007). Regulation of endothelial and myocardial NO synthesis by multi-site eNOS phosphorylation. *J. Mol. Cell Cardiol.* 42, 271–279. doi:10.1016/j.yjmcc.2006.05.023
- Nguyen, H. T., Noriega Polo, C., Wiederkehr, A., Wollheim, C. B., and Park, K. S. (2023). CDN1163, an activator of sarco/endoplasmic reticulum Ca(2+) ATPase, up-regulates mitochondrial functions and protects against lipotoxicity in pancreatic β -cells. *Br. J. Pharmacol.* 180, 2762–2776. doi:10.1111/bph.16160
- Ni, J., Shi, Y., Li, L., Chen, J., Li, L., Li, M., et al. (2017). Cardioprotection against heart failure by shenfu injection via TGF- β /smads signaling pathway. *Evid. Based Complement. Altern. Med.* 2017, 7083016. doi:10.1155/2017/7083016
- Niu, L., Xiao, L., Zhang, X., Liu, X., Liu, X., Huang, X., et al. (2021). Comparative efficacy of Chinese herbal injections for treating severe pneumonia: a systematic review and bayesian network meta-analysis of randomized controlled trials. *Front. Pharmacol.* 12, 743486. doi:10.3389/fphar.2021.743486
- Núñez-Acevedo, B., Domínguez-Ortega, J., Rodríguez-Jiménez, B., Kindelan-Recarte, C., and Pérez-Fernández, M. A. (2018). Severe and rare adverse reaction to hydrochlorothiazide. *Rev. Alerg. Mex.* 65, 442–445. doi:10.29262/ram.v65i4.363
- Obadovic, M., Sudar-Milovanovic, E., Gluvic, Z., Banjac, K., Rizzo, M., and Isenovic, E. R. (2023). The Na(+)/K(+) ATPase: a potential therapeutic target in cardiometabolic diseases. *Front. Endocrinol. (Lausanne)* 14, 1150171. doi:10.3389/fendo.2023.1150171
- Omidkhoda, N., Wallace Hayes, A., Reiter, R. J., and Karimi, G. (2019). The role of MicroRNAs on endoplasmic reticulum stress in myocardial ischemia and cardiac hypertrophy. *Pharmacol. Res.* 150, 104516. doi:10.1016/j.phrs.2019.104516
- Pall, A. H., Rasmussen, E. R., and Wadelius, M. (2021). Pharmacogenetics of angiotensin-converting enzyme inhibitor-induced angioedema. *Pharmacogenomics* 22, 319–321. doi:10.2217/pgs-2021-0036
- Panday, S., Talreja, R., and Kavdia, M. (2020). The role of glutathione and glutathione peroxidase in regulating cellular level of reactive oxygen and nitrogen species. *Microvasc. Res.* 131, 104010. doi:10.1016/j.mvr.2020.104010
- Parikh, M., Netticadan, T., and Pierce, G. N. (2018). Flaxseed: its bioactive components and their cardiovascular benefits. *Am. J. Physiol. Heart Circ. Physiol.* 314, H146–H159. doi:10.1152/ajpheart.00400.2017
- Pei, H., Ma, Y., Wang, L., Wang, L., Xu, L., and Wang, R. (2021). Effects of Shenfu injection on inflammatory factors and immune function in children with Mycoplasma pneumoniae: a protocol for a double-blind, randomized controlled trial. *Med. Baltim.* 100, e27585. doi:10.1097/md.00000000000027585
- Peng, M. L., Fu, Y., Wu, C. W., Zhang, Y., Ren, H., and Zhou, S. S. (2022). Signaling pathways related to oxidative stress in diabetic cardiomyopathy. *Front. Endocrinol. (Lausanne)* 13, 907757. doi:10.3389/fendo.2022.907757
- Sabe, S. A., Feng, J., Sellke, F. W., and Abid, M. R. (2022). Mechanisms and clinical implications of endothelium-dependent vasomotor dysfunction in coronary microvasculature. *Am. J. Physiol. Heart Circ. Physiol.* 322, H819–H841. doi:10.1152/ajpheart.00603.2021
- Sahoo, G., Samal, D., Khandayataray, P., and Murthy, M. K. (2023). A review on caspases: key regulators of biological activities and apoptosis. *Mol. Neurobiol.* 60, 5805–5837. doi:10.1007/s12035-023-03433-5
- Sattayaprasert, P., Vasireddi, S. K., Bektik, E., Jeon, O., Hajjiri, M., Mackall, J. A., et al. (2020). Human cardiac mesenchymal stem cells remodel in disease and can regulate arrhythmia substrates. *Circ. Arrhythm. Electrophysiol.* 13, e008740. doi:10.1161/circep.120.008740
- Shen, B. Q., Qu, C., Mi, L., Wang, H. Y., and Yang, H. (2021). Simultaneous quantification of twenty-eight components of Shenfu Injection in rat plasma by UHPLC-QQQ MS and its application to a pharmacokinetic study. *J. Pharm. Biomed. Anal.* 203, 114211. doi:10.1016/j.jpba.2021.114211
- Shi, S., Wang, F., Chen, B., Pan, J., Luo, D., Pei, C., et al. (2022). Efficacy and safety of shenfu injection for severe pneumonia in the elderly: a systematic review and meta-analysis based on western and eastern medicine. *Front. Pharmacol.* 13, 779942. doi:10.3389/fphar.2022.779942
- Song, Y., Zhang, N., Shi, S., Li, J., Zhang, Q., Zhao, Y., et al. (2015). Large-scale qualitative and quantitative characterization of components in Shenfu injection by integrating hydrophilic interaction chromatography, reversed phase liquid chromatography, and tandem mass spectrometry. *J. Chromatogr. A* 1407, 106–118. doi:10.1016/j.chroma.2015.06.041
- Stewart, A. G., Thomas, B., and Koff, J. (2018). TGF- β : master regulator of inflammation and fibrosis. *Respirology* 23, 1096–1097. doi:10.1111/resp.13415
- Suvorava, T., Metry, S., Pick, S., and Kojda, G. (2022). Alterations in endothelial nitric oxide synthase activity and their relevance to blood pressure. *Biochem. Pharmacol.* 205, 115256. doi:10.1016/j.bcp.2022.115256
- Tan, H. Y., Li, P. Z., Gong, J. P., and Yang, K. (2019). Shenfu injection attenuates bile duct injury in rats with acute obstructive cholangitis. *Surg. Infect. (Larchmt)* 20, 424–430. doi:10.1089/sur.2018.304
- Tan, M., Yin, Y., Ma, X., Zhang, J., Pan, W., Tan, M., et al. (2023). Glutathione system enhancement for cardiac protection: pharmacological options against oxidative stress and ferroptosis. *Cell Death Dis.* 14, 131. doi:10.1038/s41419-023-05645-y
- Tang, Q., Wang, Y., and Li, K. (2018). Zhenwu decoction for chronic heart failure: protocol for a systematic review and meta-analysis. *Med. Baltim.* 97, e11559. doi:10.1097/md.00000000000011559
- Tong, H., Zhu, J., Gong, F., Zhong, L., and Xu, T. (2021). Relationship between cardiotonic activity of Fuzi (Radix Aconiti Lateralis Preparata) and its fingerprint determined by liquid chromatography-mass spectrometry. *J. Tradit. Chin. Med.* 41, 140–149. doi:10.19852/j.cnki.jtcm.2021.01.016
- Valiente-Alandi, I., Potter, S. J., Salvador, A. M., Schafer, A. E., Schips, T., Carrillo-Salinas, F., et al. (2018). Inhibiting fibronectin attenuates fibrosis and improves cardiac function in a model of heart failure. *Circulation* 138, 1236–1252. doi:10.1161/circulationaha.118.034609
- Vazquez, A. R., and Sudhir, A. (2023). Cardiac arrest as a public health issue. *Emerg. Med. Clin. North Am.* 41, 405–411. doi:10.1016/j.emc.2023.05.003
- Wang, L., Wang, H. L., Liu, T. T., and Lan, H. Y. (2021a). TGF-beta as a master regulator of diabetic nephropathy. *Int. J. Mol. Sci.* 22, 7881. doi:10.3390/ijms22157881
- Wang, M., Hu, W. J., Zhou, X., Yu, K., Wang, Y., Yang, B. Y., et al. (2023). Ethnopharmacological use, pharmacology, toxicology, phytochemistry, and progress in Chinese crude drug processing of the lateral root of Aconitum carmichaelii Debeaux. (Fuzi): a review. *J. Ethnopharmacol.* 301, 115838. doi:10.1016/j.jep.2022.115838
- Wang, S., Liu, G., Chen, L., Xu, X., Jia, T., Zhu, C., et al. (2022). EFFECTS OF SHENFU INJECTION ON SUBLINGUAL MICROCIRCULATION IN SEPTIC SHOCK PATIENTS: A RANDOMIZED CONTROLLED TRIAL. *Shock* 58, 196–203. doi:10.1097/shk.0000000000001975
- Wang, S., and Qiu, X. J. (2019). The efficacy of Xue Fu Zhu Yu prescription for hyperlipidemia: a meta-analysis of randomized controlled trials. *Complement. Ther. Med.* 43, 218–226. doi:10.1016/j.ctim.2019.02.008
- Wang, X., Hou, Y., Mao, J., Zhang, Y., Li, Z., Zhao, Y., et al. (2017a). Western medication plus Traditional Chinese Medicine preparations in patients with chronic heart failure: a prospective, single-blind, randomized, controlled, and multicenter clinical trial. *J. Tradit. Chin. Med.* 37, 756–766. doi:10.1016/s0254-6272(18)30038-4
- Wang, X., Miao, H., Yan, Y., Guo, R., Gong, W., He, Y., et al. (2021b). Effect of shenfu injection on reperfusion injury in patients undergoing primary percutaneous coronary intervention for st segment elevation myocardial infarction: a pilot randomized clinical trial. *Front. Cardiovasc. Med.* 8, 736526. doi:10.3389/fcvm.2021.736526
- Wang, X., Zhao, Z., Mao, J., Du, T., Chen, Y., Xu, H., et al. (2019b). Randomized, double-blinded, multicenter, placebo-controlled trial of shenfu injection for treatment of patients with chronic heart failure during the acute phase of symptom aggravation (Yang and Qi deficiency syndrome). *Evid. Based Complement. Altern. Med.* 2019, 9297163. doi:10.1155/2019/9297163
- Wang, Y. L., Wang, C. Y., Zhang, B. J., and Zhang, Z. Z. (2009). Shenfu injection suppresses apoptosis by regulation of Bcl-2 and caspase-3 during hypoxia/reoxygenation in neonatal rat cardiomyocytes in vitro. *Mol. Biol. Rep.* 36, 365–370. doi:10.1007/s11033-007-9188-x
- Wang, Y. Y., Li, Y. Y., Li, L., Yang, D. L., Zhou, K., and Li, Y. H. (2018). Protective effects of shenfu injection against myocardial ischemia-reperfusion injury via activation of eNOS in rats. *Biol. Pharm. Bull.* 41, 1406–1413. doi:10.1248/bpb.b18-00212
- Wang, Z. F., Yu, J. Y., and Xie, Y. M. (2017b). Clinical safety intensive hospital monitoring on Shenfu injection with 30 106 cases. *Zhongguo Zhong Yao Za Zhi* 42, 2871–2876. doi:10.19540/j.cnki.cjcmm.20170705.008
- Wen, L., Xie, L., Gong, F., Zhang, S., and Xi, T. (2023). Efficacy and safety of Chinese medicine injections in combination with docetaxel and cisplatin for non-small cell lung cancer: a network meta-analysis. *Front. Pharmacol.* 14, 1277284. doi:10.3389/fphar.2023.1277284
- Wu, H., Dai, Z., Liu, X., Lin, M., Gao, Z., Tian, F., et al. (2019). Pharmacodynamic evaluation of shenfu injection in rats with ischemic heart failure and its effect on small molecules using matrix-assisted laser desorption/ionization-mass spectrometry imaging. *Front. Pharmacol.* 10, 1424. doi:10.3389/fphar.2019.01424
- Wu, J., Yu, W., Zhu, J., Liu, Y., Shen, Y., Sun, M., et al. (2022). Effect of Shenfu injection on the level of acetylcholine in acute liver injury of young rats with sepsis. *Minerva Gastroenterol. (Torino)* 68, 367–369. doi:10.23736/s2724-5985.21.03064-3
- Wu, Q., Zhang, Q., Li, Y., Yu, L., Zhang, Y., and Ao, M. (2021). A systematic review and meta-analysis of high-frequency prescription of zhigancao decoction combined

with conventional western medicine in the treatment of chronic heart failure. *Evid. Based Complement. Altern. Med.* 2021, 7140044. doi:10.1155/2021/7140044

Wu, Y., Xia, Z. Y., Meng, Q. T., Zhu, J., Lei, S., Xu, J., et al. (2011). Shen-Fu injection preconditioning inhibits myocardial ischemia-reperfusion injury in diabetic rats: activation of eNOS via the PI3K/Akt pathway. *J. Biomed. Biotechnol.* 2011, 384627. doi:10.1155/2011/384627

Xiong, X. J. (2019). Exploration on connotation of Zhigancao Decoction formula syndrome from the perspective of modern pathophysiology and severe cases of critical care and its clinical efficacy on cardioversion, maintenance of sinus rhythm, hemostasis, increasing platelets count, and tonifying deficiency. *Zhongguo Zhong Yao Za Zhi* 44, 3842–3860. doi:10.19540/j.cnki.cjcm.20190416.501

Xu, C., Xia, Y., Jia, Z., Wang, S., Zhao, T., and Wu, L. (2022). The curative effect of Shenfu-injection in the treatment of burn sepsis and its effect on the patient's immune function, HMGB, and vWF. *Am. J. Transl. Res.* 14, 2428–2435.

Yan, P., Mao, W., Jin, L., Fang, M., Liu, X., Lang, J., et al. (2020). Crude radix Aconiti lateralis preparata (fuzi) with Glycyrrhiza reduces inflammation and ventricular remodeling in mice through the TLR4/NF- κ B pathway. *Mediat. Inflamm.* 2020, 5270508. doi:10.1155/2020/5270508

Yang, F., Yang, M. Y., Le, J. Q., Luo, B. Y., Yin, M. D., Chao, L., et al. (2022a). Protective effects and therapeutics of ginsenosides for improving endothelial dysfunction: from therapeutic potentials, pharmaceutical developments to clinical trials. *Am. J. Chin. Med.* 50, 749–772. doi:10.1142/s0192415x22500318

Yang, H., Liu, L., Gao, W., Liu, K., Qi, L. W., and Li, P. (2014). Direct and comprehensive analysis of ginsenosides and diterpene alkaloids in Shenfu injection by combinatory liquid chromatography-mass spectrometric techniques. *J. Pharm. Biomed. Anal.* 92, 13–21. doi:10.1016/j.jpba.2013.12.041

Yang, L., Huang, G. Y., Wang, Y. G., Han, B. Q., Zheng, B., Zhu, J. M., et al. (2021). Efficacy of renshen (radix ginseng) plus fuzi (radix Aconiti lateralis preparata) on myocardial infarction by enhancing autophagy in rat. *J. Tradit. Chin. Med.* 41, 909–918. doi:10.19852/j.cnki.jtcm.2021.06.009

Yang, Y., Ge, F. L., Huang, Q., Zeng, R., Zhang, X. Y., Liu, P., et al. (2022b). Randomized controlled trials of zhigancao decoction combined with metoprolol in the treatment of arrhythmia: a systematic review and meta-analysis. *Front. Cardiovasc. Med.* 9, 795903. doi:10.3389/fcvm.2022.795903

Yang, Y., Su, C., Zhang, X. Z., Li, J., Huang, S. C., Kuang, H. F., et al. (2023). Mechanisms of Xuefu Zhuyu Decoction in the treatment of coronary heart disease based on integrated metabolomics and network pharmacology approach. *J. Chromatogr. B Anal. Technol. Biomed. Life Sci.* 1223, 123712. doi:10.1016/j.jchromb.2023.123712

Ye, B., Zhou, H., Chen, Y., Luo, W., Lin, W., Zhao, Y., et al. (2023). USP25 ameliorates pathological cardiac hypertrophy by stabilizing SERCA2a in cardiomyocytes. *Circ. Res.* 132, 465–480. doi:10.1161/circresaha.122.321849

Zhang, M. Q., and Li, C. S. (2023). Therapeutic effects of shenfu injection in shock. *Chin. J. Integr. Med.* 29, 1142–1146. doi:10.1007/s11655-023-3631-2

Zhang, N., Zhao, Y., Liu, Y., Tang, N., Zheng, W., Mao, M., et al. (2021a). A double-blinded, placebo-controlled randomized trial evaluating the efficacy and safety of

Zhigancao Tang granules for treating HFpEF: study protocol for a randomized controlled trial. *Trials* 22, 293. doi:10.1186/s13063-021-05232-6

Zhang, Q., Li, C., Shao, F., Zhao, L., Wang, M., and Fang, Y. (2017). Efficacy and safety of combination therapy of shenfu injection and postresuscitation bundle in patients with return of spontaneous circulation after in-hospital cardiac arrest: a randomized, assessor-blinded, controlled trial. *Crit. Care Med.* 45, 1587–1595. doi:10.1097/ccm.0000000000002570

Zhang, Q., Lyu, W., Yu, M., and Niu, Y. (2020a). Sulfur dioxide induces vascular relaxation through PI3K/Akt/eNOS and NO/cGMP signaling pathways in rats. *Hum. Exp. Toxicol.* 39, 1108–1117. doi:10.1177/0960327120911428

Zhang, S., Chen, Z. L., Tang, Y. P., Duan, J. L., and Yao, K. W. (2021b). Efficacy and safety of xue-fu-zhu-yu decoction for patients with coronary heart disease: a systematic review and meta-analysis. *Evid. Based Complement. Altern. Med.* 2021, 9931826. doi:10.1155/2021/9931826

Zhang, X., Guo, T., Zhang, K., Guo, W., An, X., and Gao, P. (2020b). Effect of shenfu injection on microcirculation in shock patients: a protocol for systematic review and meta-analysis. *Med. Baltim.* 99, e22872. doi:10.1097/md.00000000000022872

Zhang, Y., Tian, D., Huang, Y., Li, L., Mao, J., Tian, J., et al. (2016). Pharmacokinetic evaluation of Shenfu Injection in beagle dogs after intravenous drip administration. *Acta. Pharm. Sin. B* 6, 584–592. doi:10.1016/j.apsb.2016.05.006

Zhao, D., Zhong, G., Li, J., Pan, J., Zhao, Y., Song, H., et al. (2021a). Targeting E3 ubiquitin ligase WWP1 prevents cardiac hypertrophy through destabilizing DVL2 via inhibition of K27-linked ubiquitination. *Circulation* 144, 694–711. doi:10.1161/circulationaha.121.054827

Zhao, X., Chu, Q., Wu, W., Wu, H., Wang, S., Qing, L., et al. (2021b). Shenfu injection: a famous Chinese prescription that promotes cardiac hypertrophy through destabilizing mesenchymal stem cells. *Evid. Based Complement. Altern. Med.* 2021, 9912844. doi:10.1155/2021/9912844

Zheng, S. Y., Sun, J., Zhao, X., and Xu, J. G. (2004). Protective effect of shen-fu on myocardial ischemia-reperfusion injury in rats. *Am. J. Chin. Med.* 32, 209–220. doi:10.1142/s0192415x04001874

Zheng, Y., Zheng, M., Shao, J., Jiang, C., Shen, J., Tao, R., et al. (2022). Upregulation of claudin-4 by Chinese traditional medicine Shenfu attenuates lung tissue damage by acute lung injury aggravated by acute gastrointestinal injury. *Pharm. Biol.* 60, 1981–1993. doi:10.1080/13880209.2022.2128824

Zhou, D., Xie, L., Wang, Y., Wu, S., Liu, F., Zhang, S., et al. (2020). Clinical efficacy of tonic traditional Chinese medicine injection on acute cerebral infarction: a bayesian network meta-analysis. *Evid. Based Complement. Altern. Med.* 2020, 8318792. doi:10.1155/2020/8318792

Zhou, W., Chen, Z., Fang, Z., and Xu, D. (2022). Network analysis for elucidating the mechanisms of Shenfu injection in preventing and treating COVID-19 combined with heart failure. *Comput. Biol. Med.* 148, 105845. doi:10.1016/j.combiomed.2022.105845

Zhu, J., Song, W., Xu, S., Ma, Y., Wei, B., Wang, H., et al. (2020). Shenfu injection promotes vasodilation by enhancing eNOS activity through the PI3K/Akt signaling pathway *in vitro*. *Front. Pharmacol.* 11, 121. doi:10.3389/fphar.2020.00121



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The protective role of ginsenoside Rg3 in heart diseases and mental disorders

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Ginsenoside Rg3, a compound derived from *Panax ginseng* C. A. Mey., is increasingly recognized for its wide range of pharmacological effects. Under the worldwide healthcare challenges posed by heart diseases, Rg3 stands out as a key subject in modern research on Chinese herbal medicine, offering a novel approach to therapy. Mental illnesses are significant contributors to global disease mortality, and there is a well-established correlation between cardiac and psychiatric conditions. This connection is primarily due to dysfunctions in the sympathetic-adrenomedullary system (SAM), the hypothalamic-pituitary-adrenal axis, inflammation, oxidative stress, and brain-derived neurotrophic factor impairment. This review provides an in-depth analysis of Rg3's therapeutic benefits and its pharmacological actions in treating cardiac and mental health disorders respectively. Highlighting its potential for the management of these conditions, Rg3 emerges as a promising, multifunctional therapeutic agent.

KEYWORDS

ginsenoside Rg3, heart diseases, mental disorders, heart failure, depression

1 Introduction

Despite extensive research over the years, both heart diseases and mental disorders continue to exert significant pressure on global healthcare systems. Heart diseases maintain their status as the leading cause of death globally (Liu and Miao, 2022; Lupisella et al., 2022). The pharmacological landscape for heart diseases is diverse, encompassing a range of drugs. However, these treatments largely fail to restore cardiac function fundamentally and are frequently associated with side effects.

At the same time, mental illness significantly contributes to the global disease burden. A recent meta-analysis reveals that 14.3% of global deaths annually, equivalent to roughly eight million fatalities, are linked to mental disorders (Walker et al., 2015). Individuals with severe mental illnesses, such as schizophrenia, bipolar disorder, and major depressive disorder (MDD), face a mortality rate of two to three times higher than the average population. The elevated rate corresponds to a reduced life expectancy of 10–25 years (Correll et al., 2015). However, issues associated with treatment discontinuation and ineffectiveness are prevalent.

Up to now, numerous studies have established a strong link between cardiac diseases and psychiatric conditions. It is frequently observed that individuals with cardiac ailments often experience psychiatric disturbances (Piepenburg et al., 2019). Inversely, those with mental disorders appear to have a higher risk of developing heart diseases (Hagi et al., 2021).

Several biological mechanisms are suggested to clarify the association between mental disorders and cardiac events. Mental disorders are linked to dysfunctions in the sympathetic-adrenomedullary system (SAM), and the hypothalamic-pituitary-adrenal (HPA) axis, as well as to inflammation, oxidative stress, and impairments in the brain-derived neurotrophic factor (BDNF) system. All these physiological processes play significant roles in the onset and progression of cardiac diseases.

Recently, the field of Chinese herbal medicine has captured the interest of the scientific community, owing to its extensive range of pharmacological properties and a lower incidence of adverse side effects. Particularly, the effects of ginsenoside Rg3 (Rg3) in cardiac and mental diseases have gained more and more attention.

Consequently, in this review, we provide a detailed investigation of the therapeutic effects and pharmacological action of Rg3 in addressing heart and mental disorders (Table 1). Based on the association between heart and mental disorders, we aim to provide prospects on the potential effects and mechanisms of Rg3 in the comorbid conditions.

2 Origin and structure of Rg3

Rg3 is a constituent of ginsenosides extracted from *Panax ginseng* C. A. Mey. (Nakhjavani et al., 2020). Key ginsenoside constituents, specifically Rb1, Rb2, and Rd, possess the capacity for enzymatic transformation into Rg3 (Lee H. et al., 2020). Rg3 is categorized into two distinct stereoisomers based on its unique spatial configurations at the C20 position: 20(R)-Rg3, as illustrated in Figure 1, and 20(S)-Rg3, depicted in Figure 2. Research findings have underscored the pivotal role of Rg3 across diverse domains, encompassing its involvement in anti-aging mechanisms, anticancer properties, bone development, cellular differentiation, neuroprotection, and cardiac function (Lee et al., 2019; Zarneshan et al., 2022).

3 The relationships between heart diseases and mental disorders

3.1 Mental disorders induce the occurrence of heart diseases

Recent meta-analytic studies have shown that mental illness is a high risk for cardiac events (Zarneshan et al., 2022). In addition, a Mendelian randomization analysis revealed a significant genetic association between MDD and coronary artery disease (CAD). Specifically, the analysis indicated that for each one-unit increase in the natural logarithm of odds of MDD, the odds ratio for CAD was 1.16 (95% confidence interval: 1.05 to 1.29; $p = 0.0047$) (Tang et al., 2020). Recent observational research has indicated a correlation between genetic predispositions for schizophrenia and notable changes in cardiac structure. These alterations have been found to potentially aggravate cardiac health outcomes (Pillinger et al., 2023). Mental health disorders, such as MDD, anxiety, and stress-related conditions, may contribute to behaviors including smoking, inactivity, and drinking adversely affecting cardiac function.

3.2 Heart diseases promote the development of mental disorders

Clinically, a subset of patients with heart diseases commonly experience co-occurring mental disorders, specifically anxiety and depression. Multiple meta-analyses have demonstrated that individuals diagnosed with heart failure (HF) are at an elevated risk of experiencing depression (Hagi et al., 2021). Certain drugs used in managing heart diseases, including beta-blockers, have the potential to initiate or aggravate symptoms associated with anxiety and depression (Bornand et al., 2022; Carnovale et al., 2023). Additionally, managing heart conditions, including HF, entails a prolonged treatment course that can result in significant financial strain for patients and create a disparity between hospitalization needs and employment stability. This significantly heightens the risk of developing mental disorders in individuals with heart diseases.

3.3 The major mechanisms linking heart diseases and mental disorders

3.3.1 Inflammation

Inflammatory processes are associated with the initiation and progression of mental disorders (Ben-Azu et al., 2022a; Ben-Azu et al., 2023). Mental disorders may enhance the expression of pro-inflammatory cytokines, such as interleukin-6 (IL-6) and tumor necrosis factor-alpha (TNF- α), which are associated with endothelial dysfunction and metabolic alterations (Vaccarino et al., 2018; Misiak et al., 2022; Tsioufis et al., 2022; Chen et al., 2023). Inflammation, vascular dysfunction, and metabolic abnormalities frequently contribute to the pathogenesis of heart diseases. Furthermore, inflammatory processes within heart tissue can result in elevated blood levels of pro-inflammatory cytokines and other acute-phase reactants (Hartupee and Mann, 2017). Inflammatory factors may lead to the upregulation of indoleamine 2,3-dioxygenase (IDO), diverting tryptophan (TRP) into the kynurenine (KYN) pathway, potentially reducing serotonin (5-HT) synthesis and contributing to the onset of mental disorders, including depression. 5-HT functions as a neurotransmitter in the central nervous system, a blood factor, and a neurohormone that regulates the function of various peripheral organs (De Deurwaerdere and Di Giovanni, 2020). A deficiency in 5-HT increases susceptibility to social defeat stress and impairs responses to antidepressants (Sachs et al., 2015). Additionally, pro-inflammatory cytokines are associated with a marked decrease in both BDNF gene and protein expression (Zhang et al., 2014). BDNF is crucial for the plasticity of glutamatergic and gamma aminobutyric acid (GABA)ergic synapses and is intimately linked to severe mental illnesses (Colucci-D'Amato et al., 2020). Interestingly, animal studies have demonstrated that in post-myocardial infarction, BDNF expression is upregulated through neuronal signaling originating from the heart. This upregulation serves to shield the myocardium from ischemic damage, thereby exerting a protective effect against cardiac remodeling (Cannavo et al., 2023). Furthermore, studies have noted lower BDNF levels in patients with HF when compared to healthy controls (Xie et al., 2023; Wang et al., 2024). BDNF plays a crucial role in supporting the survival of endothelial cells during the development of the cardiovascular system (Kermani and Hempstead, 2019).

TABLE 1 Summary of effects of Rg3 on heart and mental conditions.

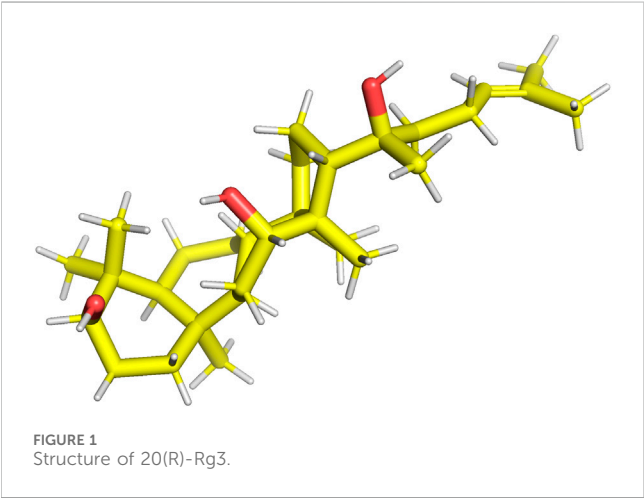
| Disease | Model | Type | Treatment (dose, duration) | | Described effects and mechanisms | References (PMID) | |
|----------------|---------------|-----------|-------------------------------|-----------------|---|----------------------|--|
| | | | Animal | Cell | | | |
| HF | TAC-mice | 20(R)-Rg3 | 20 mg/kg | 10 μM | Improving cardiac function, inhibiting cardiomyocyte hypertrophy and ER stress via enhancing SUMOylation of SERCA2a | 34,428,586 | |
| | ISO-HL-1 cell | | 4 weeks | 24 h | | | |
| | TAC-mice | — | 10 or 20 mg/kg | 10 μM | Modulating glucose metabolism and insulin resistance through activation of the AMPK pathway | 35,509,823 | |
| | Insulin-H9C2 | | 4 weeks | 24 h | | | |
| | TAC-mice | — | 10 or 20 mg/kg | — | Regulating pyruvate metabolism and sustaining glucose homeostasis in cardiac tissue through maintaining the PDHc activity | 36,572,672 | |
| | | | 4 weeks | | | | |
| | CAL-mice | 20(S)-Rg3 | 7.5, 15 or 30 mg/kg | 1, 5 or 25 μM | Inhibiting myocardial fibrosis via the ACY1/TGFβ1/Smad3 signaling pathway | 34916608 | |
| | Ang II-CFs | | 2 weeks | 24 h | | | |
| | LAD-rat | — | 10, 20 or 30 mg/kg | 10 μm | Activating mitophagy via the ULK1/FUNDC1 pathway | 37,659,296 | |
| H9C2 | 4 weeks | | 20 h | | | | |
| MI | CAL-rat | — | 30 mg/kg/day | — | Inhibiting the inflammatory response via the NF-κB pathway | 33193843 | |
| | | | 1 week | | | | |
| | ISO-mice | — | 5 mg/kg | — | Upregulating autophagy process through the AMPK signaling pathway | 32420095 | |
| | | | 2 days | | | | |
| | LAD-mice | 20(S)-Rg3 | 20 or 40 mg/kg | 5, 10 or 20 μM | Alleviating myocardial fibrosis through the TGFBR1 signaling pathway | 38107395 | |
| TGFβ1-CFs | 4 weeks | | | | | | |
| MIRI | Rat | — | 5 or 20 mg/kg | — | Antiapoptosis and anti-inflammation | 28105061 | |
| | | | 7 days | | | | |
| | Rat | — | 0.5 mg | 10 nM | Antifibrosis, inhibiting oxidative stress and inflammation via SIRT1/PGC1-α/Nrf and IκBα/NF-κB signal pathway | 31,783,047 | |
| | H9C2 | | | 24 h | | | |
| | Rat | — | 60 mg/kg | 10 mM | Inhibiting apoptosis and oxidative stress via Akt/eNOS signaling and the Bcl2/Bax pathway | 25,672,441 | |
| NRCs | 24 h | | | | | | |
| Cardiotoxicity | ADM-rats | — | 10, 20 or 40 mg/kg | 1, 10 or 100 μM | Inhibiting oxidative stress via activation of the Nrf2-ARE pathway through the activation of Akt | 26,321,736 | |
| | CMEC | | 2 weeks | 24 h | | | |
| | Mice | — | 10, 40 or 80 mg/kg | 60 μM | Inhabeting apoptosis and oxidative stress through the miR-128-3p/MDM4 axis | 37,990,515 | |
| | H9C2 | | 4 weeks | 25 h | | | |
| DCM | Mice | — | 25, 50 or 100 mg/kg | 5, 10 or 20 μM | Modulating glucose and lipid metabolism by directly binding to PPAR-γ and activation of the adiponectin pathway | 38,069,059 | |
| | H9C2 | | 12 weeks | 48 h | | | |
| | 3T3-L1 | | | | | | |

(Continued on following page)

TABLE 1 (Continued) Summary of effects of Rg3 on heart and mental conditions.

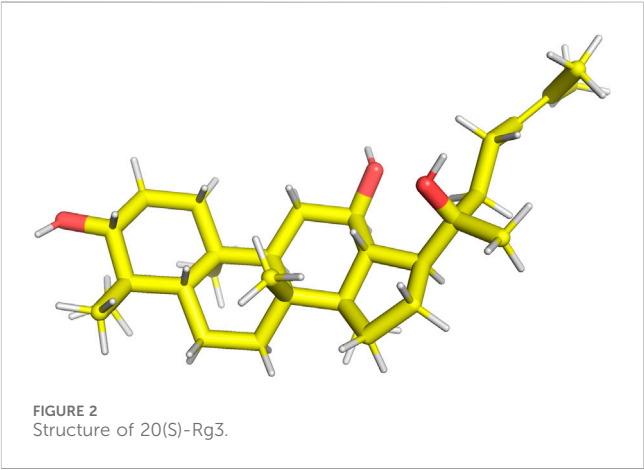
| Disease | Model | Type | Treatment (dose, duration) | | Described effects and mechanisms | References (PMID) |
|---------|------------------------|--|----------------------------|---------------|---|-------------------|
| | | | Animal | Cell | | |
| MDD | LPS-mice | — | 20 or 40 mg/kg | — | Inhibition of neuroinflammatory disturbance and the regulation of TRP-KYN metabolism | 28,762,741 |
| | | | 3 days | | | |
| | Mice | — | 10 or 20 mg/kg | — | Promotion of the BDNF signaling pathway | 28,013,484 |
| | | | 2 weeks | | | |
| | Mice | — | 50, 100 or 150 mg/kg | 1, 5 or 10 μM | Recovering proliferation and inhibiting apoptosis via CREB and BDNF signaling pathway | 28,461,003 |
| | NMDA-HT22 | | 4 weeks | | | |
| Anxiety | <i>Xenopus</i> oocytes | 20(S)-Rg3 | — | 100 μM | Regulating GABA _A receptor channel activity | 23,499,684 |
| PTSD | Rat | — | 25 or 50 mg/kg | — | Regulating the HPA axis and activating the BDNF-TrkB pathway | 35,982,366 |
| | | | 2 weeks | | | |
| ADHD | Mice | YY162(including 20(R)-Rg3 and 20(S)-Rg3) | 200 mg/kg | 100 μg/mL | Inhibiting oxidative stress | 24,394,491 |
| | SH-SY5Y | | 2 weeks | 50 h | | |

TAC, transverse aortic constriction; ISO, isoproterenol; ER, endoplasmic reticulum; SERCA2a, sarcoplasmic/endoplasmic reticulum Ca²⁺-ATPase; AMPK, AMP-activated protein kinase; PDHc, pyruvate dehydrogenase complex; CAL, coronary artery ligation; AngII, angiotensin II; ACY1, aminoacylase-1; TGFβ1, transforming growth factor-β 1; LAD, left anterior descending coronary artery ligation; ULK1, Unc51-like-kinase 1; FUNDC1, FUN14 domain-containing protein 1; NFκB, nuclear factor κB; TGFBR1, transforming growth factor beta receptor 1; PGC1-α, peroxisome proliferators-activated receptor γ coactivator 1 alpha; IκBα, inhibitor of kappa B alpha; Nrf2, nuclear factor erythroid 2-related Factor 2; Akt, protein kinase B; eNOS, endothelial nitric oxide synthase; Bcl-2, B cell lymphoma-2; Bax, Bcl2-associated X protein; ARE, antioxidant response element; MDM4, double minute 4 protein; PPAR-γ, peroxisome proliferator-activated receptor γ; TRP, tryptophan; KYN, kynurenine; BDNF, brain-derived neurotrophic factor; CREB, cyclic adenosine monophosphate response element binding protein; HPA, hypothalamic-pituitary-adrenal; TrkB, tropomyosin-related kinase B; LPS, lipopolysaccharide; HT22, murine hippocampal neuronal; NMDA, n-methyl-d-aspartate; HF, heart failure; MI, myocardial infarction; MIRI, myocardial ischemia-reperfusion injury; MDD, major depressive disorder; DCM, diabetic cardiomyopathy; PTSD, post-traumatic stress disorder; ADHD, attention deficit hyperactivity disorder.



3.3.2 HPA axis

The HPA axis serves as a critical component of the neuroendocrine system, orchestrating responses to both internal and external stressors. Various mental disorders have been shown to trigger the activation of the HPA axis (Sur and Lee, 2022b; Emudainohwo et al., 2023; Menke, 2024). This activation leads to an upsurge in cortisol synthesis within the adrenal cortex, along with enhanced production of adrenaline and noradrenaline in the adrenal medulla (Ugwu et al., 2022). Hypercortisolemia, commonly observed in mental disorders, can



lead to escalated steroid production, elevated blood pressure, and an increase in visceral fat (García-Eguren et al., 2019; Favero et al., 2021; Teng et al., 2021). These changes significantly heighten the risk of heart diseases. Elevated cortisol levels following stress are directly linked to the hypertrophy of cardiomyocytes and cardiac remodeling (Opinion et al., 2023). Additionally, plasma cortisol concentrations have been identified as an independent risk factor for cardiac events and mortality (Crawford et al., 2019; Kim et al., 2022). Furthermore, the stimulation of the HPA axis results in increased aldosterone levels. Studies have shown that stress-related aldosterone activity has been linked to

hypertension, myocardial necrosis, and fibrosis. Additionally, a rise in aldosterone has been associated with increased insulin resistance, oxidative stress, and pro-inflammatory responses (Bothou et al., 2020; Tsai et al., 2021; Campana et al., 2022). In chronic HF, elevated serum cortisol levels have been identified as an independent predictor of increased mortality risk (Güder et al., 2007). Moreover, increased cortisol levels may exacerbate the brain's vulnerability to oxidative stress, potentially leading to detrimental effects on neurobehavioral health. Stress-induced cortisol secretion may lower brain 5-HT and BDNF function, potentially leading to the onset of depressive symptoms and anxiety (Bhagwagar et al., 2002; Motta et al., 2021).

3.3.3 SAM

Negative psychological states can activate the SAM system, resulting in elevated catecholamine levels. Catecholamines, including epinephrine, norepinephrine, and dopamine, are tyrosine-derived hormones and neurotransmitters primarily synthesized in the adrenal medulla, sympathetic nerves, and brain. Elevated catecholamine levels can induce vasoconstriction, leading to cardiac injury, HF, myocardial ischemia, and necrosis (Szatko et al., 2023). Persistent catecholamine elevation may cause myocardial calcium overload in cytosolic and mitochondrial compartments, trigger oxidative stress, increase mitochondrial permeability, and cell death (Szatko et al., 2023). In patients with HF, there is an activation of the neuroendocrine systems, particularly the sympathetic nervous system and the renin-angiotensin-aldosterone system, leading to elevated levels of neurohormones such as catecholamines (Manolis et al., 2023). Dopamine, an important catecholamine, plays a crucial role in regulating various mental and physical functions, including anxiety, fear, attention deficit hyperactivity disorder (ADHD), and schizophrenia (Jayanti et al., 2023). Additionally, norepinephrine, another key catecholamine, acts as a neurotransmitter and is essential in mediating physiological and behavioral responses to stress (Schmidt et al., 2019).

4 Pharmacological action of Rg3 in heart diseases

4.1 HF

HF is a complicated condition that leads to aggressive hazards to human health. The common therapeutic regimens for HF patients predominantly involve diuretics, vasoactive drugs, and other pharmaceutical approaches (Truby and Rogers, 2020). Medications such as angiotensin-converting enzyme inhibitors (ACEIs), beta-blockers, mineralocorticoid receptor antagonists, and angiotensin receptor-neprilysin inhibitors can partially alleviate HF symptoms. However, these treatments are often insufficient in significantly reducing rehospitalization rates and mortality, even when patients adhere to established guidelines (Lai et al., 2022). Moreover, traditional medication can lead to adverse effects, including hypotension, hypokalemia, and renal function impairment (Hubers and Brown, 2016; Marciniak and Serebruan, 2019; Rossello et al., 2022). Consequently, there is a pressing need to explore new therapeutic agents to enhance survival rates and improve the quality of life for HF patients. Rg3 has

emerged as a promising treatment option in HF, offering protective benefits such as promoting cardiomyocyte relaxation, enhancing mitochondrial structure and function, regulating metabolism, reducing cardiac fibrosis, and preventing cell apoptosis.

Cardiac sarcoplasmic/endoplasmic reticulum Ca^{2+} -ATPase (SERCA2a), a crucial protein in the Ca^{2+} cycle of cardiomyocytes, is involved in Ca^{2+} reuptake into the cytoplasm and subsequent transport to the endoplasmic reticulum (ER), ultimately promoting cardiomyocytes relaxation (Liu et al., 2021; Chen et al., 2022). SUMO binds to certain lysine sites on SERCA2a, forming the SERCA2a/SUMO complex. Notably, HF patients and mice exhibited significantly reduced levels of SERCA2a SUMOylation (Mendler et al., 2016). However, Rg3 treatment could increase the SUMOylation of SERCA2a, further increase intracellular Ca^{2+} cycle protein levels, suppress ER stress and prevent reactive oxygen species (ROS) generation, thus improving cardiac function and inhibiting cardiomyocyte hypertrophy in transverse aortic constriction (TAC) induced HF mice (Liu et al., 2021).

Moreover, Rg3 could improve disordered mitochondrial ultrastructure, functions such as ATP production and spare respiratory capacity, and regulate glucose uptake, and myocardial insulin resistance (Ni et al., 2022a). Rg3 regulated glucose uptake and myocardial insulin resistance through the activation of insulin receptor substrate (IRS)-phosphoinositide 3 kinase (PI3K)-protein kinase B (Akt) signaling pathway (Ni et al., 2022a). Several pieces of experimental evidence suggested that the decoupling of glucose oxidation to glycolysis may be the cause of unaltered or reduced pyruvate oxidation in mitochondria in HF (Pound et al., 2009; Bertero and Maack, 2018; Fillmore et al., 2018). Pyruvate dehydrogenase complex (PDHc) plays a pivotal role in regulating mitochondrial pyruvate metabolism with dihydrolipoamide dehydrogenase (DLD), serving as a crucial part of PDHc (Staretz-Chacham et al., 2021; Kim et al., 2023). P300 and tat-interacting protein 60 (TIP60) are recognized as 2-hydroxyisobutyryltransferases that could regulate the activity of PDHc (Sabari et al., 2017; Huang et al., 2018). In HF, the 2-hydroxyisobutyrylation of DLD was significantly upregulated, resulting from the downregulation of PDHc activity. However, Rg3 can lower the 2-hydroxyisobutyrylation levels of DLD and maintain the PDHc activity by suppressing the acyltransferase activity of P300, further regulating pyruvate metabolism and sustaining glucose homeostasis in cardiac tissue, consequently, improving cardiac function (Ni et al., 2022b).

Another study demonstrated that in HF mice, Rg3 administration increased the expression of aminoacylase-1 (ACY1) and inhibited cardiac fibrosis, thereby, ameliorating heart function through the ACY1-mediated transforming growth factor- β 1 (TGF- β 1)/Smad3 pathway. In this study, metoprolol served as the positive control. Both Rg3 and metoprolol significantly enhanced cardiac function. Notably, the impact of Rg3 at high dosage on HF was found to be comparable to that of metoprolol. Furthermore, in murine cardiac fibroblasts, the intervention of angiotensin II (AngII) resulted in an upregulation of collagen 1, collagen 3, α -smooth muscle actin, tissue inhibitor of metalloproteinases 1, and the TGF- β 1/Smad3 signaling pathway, which could be reversed in case of overexpressing ACY1 and Rg3 administration (Lai et al., 2022). Additionally, Baoyuan decoction, a mixture of several Chinese herbs, consists of *Astragalus membranaceus* (Fisch.) Bunge,

Glycyrrhiza uralensis Fisch., *Cinnamomum cassia* Presl and *P. ginseng* C. A. Mey. (Wang et al., 2020). A study demonstrated that its active component Rg3 effectively suppressed cardiomyocyte apoptosis via angiotensin type 1 receptor (AT1)-cardiac ankyrin repeat protein (CARP) signaling pathway (Wang et al., 2020). CARP, a downstream protein of AT1 can accelerate apoptosis by activating the P53-mitochondrial apoptotic pathway (Shen et al., 2015). In another study, Rg3 could be an Unc51-like-kinase 1 (ULK1) regulator to facilitate FUN14 domain-containing protein 1 (FUNDC1)-mediated mitophagy, thus restoring mitochondria homeostasis and energy metabolism in HF (Wang et al., 2023). Additionally, trimetazidine was selected as the positive control. The findings indicated that the efficacy of trimetazidine was equivalent to that of a medium dose of Rg3 in rats with HF. Furthermore, a high dose of Rg3 exhibited the most pronounced therapeutic effectiveness (Wang et al., 2023).

4.2 Myocardial infarction (MI) and myocardial ischemia-reperfusion injury (MIRI)

MI and MIRI are global health issues characterized by high incidence and mortality rates across different countries. The pathophysiological basis of MI is closely related to mitochondria dysfunction, the depletion of endogenous antioxidants, and lipid peroxidation (Wang and Kang, 2021; Li D. et al., 2023; Cai et al., 2023). Following an acute MI event, clinical intervention typically involves thrombolysis, percutaneous coronary intervention (PCI), and coronary artery bypass grafting (Doenst et al., 2019; Mackman et al., 2020; Sabatine and Braunwald, 2021). These therapeutic strategies are effective in restoring blood flow, alleviating pain, and minimizing myocardial damage. Although these treatments have contributed to a substantial decrease in mortality rates, a subset of patients may experience complications, including hemorrhage and MIRI (Reed et al., 2017; McCarthy et al., 2018; Doenst et al., 2019). It has been observed that an incidence of 10%–25% of recurrent acute MI and a hospital death rate of 6%–14% among MIRI patients following PCI therapy (Wu et al., 2019). In MIRI, several medications, including antiplatelet drugs may extend survival times. However, a significant proportion of these treatments are associated with adverse effects such as bleeding and suboptimal targeting efficiency (Zhang et al., 2022). Additionally, some studies suggest that conventional antiplatelet medications do not markedly improve clinical symptoms (Li et al., 2021). Thus, combining with traditional Chinese medicine may be a promising therapeutic method under virtue of fewer adverse reactions (Tu et al., 2020).

A few studies in animals have proved that Rg3 exerted protective action on MI by promoting mitophagy, and inhibiting apoptosis, myocardial fibrosis as well as inflammation. A study showed that in isoproterenol (ISO)-induced MI mice, Rg3 pretreatment could decrease ROS content in the myocardium, promote autophagy via the AMP-activated protein kinase (AMPK)/acetyl CoA carboxylase (ACC) signal pathway, and inhibit apoptosis (Sun et al., 2020). Another study displayed that in MI rats, Rg3 downregulated the levels of pro-inflammatory cytokines in serum and cardiac tissue such as TNF- α , interleukin-1 β (IL-1 β), and IL-6 and increased the levels of anti-inflammatory cytokine

interleukin-10 (IL-10). The anti-inflammation mechanism of Rg3 is related to the increased expression levels of sirtuin 1 (SIRT1) and decreased expression of p-P65 (Tu et al., 2020). P65 is a crucial part of the nuclear factor κ B (NF- κ B). SIRT1-deacetylated P65 inhibits NF- κ B activation by impeding its nuclear translocation and further restrains the transcription of TNF- α , IL-6, and other inflammatory genes downstream of NF- κ B, thereby mitigating the inflammatory response (Chen et al., 2016; Tu et al., 2020). A study showed in MI mice and TGF β 1-stimulated primary cardiac fibroblasts (CFs), Rg3 suppressed CF proliferation along with collagen deposition by inactivation of transforming growth factor beta receptor 1 (TGFBR1)/Smads signaling dose-dependently (Xu et al., 2023). TGFBR1 overexpression partially abolished Rg3's inhibition on Smad2/Smad3 activation, CFs growth, together with collagen production. In this study, captopril was employed as the positive control. The results indicated that Rg3 improved cardiac function in a dose-dependent manner, with the high dose of Rg3 demonstrating effects comparable to those of captopril (Xu et al., 2023).

In addition, Rg3 exerted a beneficial role in MIRI via inhibiting inflammation, apoptosis, oxidative stress, and attenuating cardiac fibrosis. In MIRI rat models, the use of Rg3 downregulated significantly the levels of inflammatory cytokines in plasma by inhibiting inhibitor of kappa B α (I κ B α)/NF- κ B signal pathway and further improved cardiac function (Zhang et al., 2016; Li et al., 2020). Moreover, Rg3 ameliorated myocardial collagen deposition via the blockage of the TGF- β /Smad signaling pathway and exerted an anti-apoptotic effect via the Akt/endothelial nitric oxide synthase (eNOS) signaling pathway and the B cell lymphoma-2 (Bcl-2)/Bcl2-associated X protein (Bax) pathway (Wang Y. et al., 2015; Li et al., 2020). Hypoxia/reoxygenation (H/R) is a typical approach to generate MIRI injury in cardiocytes (Du et al., 2023). Rg3 administration strongly targeted FoxO3a to decrease the ROS content via the SIRT1/peroxisome proliferators-activated receptor γ coactivator-1 α (PGC1- α)/nuclear factor erythroid 2-related factor (Nrf) pathway, and inhibit inflammation through I κ B α /NF- κ B signal pathway in H9C2 cells induced by H/R (Li et al., 2020).

4.3 Cardiotoxicity

Cardiotoxicity is an essential consideration in evaluating whether drugs can be marketed during preclinical trials and is a major reason for treatment withdrawal even after approval (Coelho et al., 2017). Even though some drugs have been used, cardiotoxicity confines their use in clinical practice. Nowadays, a good chunk of drugs, mostly anti-cancer medicine, usually lead to cardiotoxicity. Cancer therapy-induced cardiotoxicity significantly impacts patients mortality, long-term prognosis, and overall quality of life. Anthracyclines play an essential role in cancer therapy, including doxorubicin (DOX), epirubicin, daunorubicin, actinomycin, and valrubicin, have been widely applied to treat multiple types of cancer (Sawicki et al., 2021; Qu et al., 2022). However, their clinical application has been constrained by diverse adverse actions, including cardiotoxicity (Qu et al., 2022). DOX-induced cardiotoxicity involves a complex mechanism, including excessive ROS production, alterations in cell membrane integrity, and apoptosis (Upadhyay et al., 2020; Rawat et al., 2021; Tai et al.,

2023). Standard treatments for cardiotoxicity encompass ACEIs, angiotensin receptor blockers (ARBs), beta-blockers, and calcium channel blockers. Research has indicated that ACEI/ARB and beta-blockers may have a positive impact on reducing the long-term health risks associated with anthracycline-induced cardiac dysfunction (Livi et al., 2021). However, previous studies have yielded inconsistent conclusions, possibly related to tumor subtypes and staging. Consequently, there is a need for further investigation into novel medications and complementary therapeutic approaches. A substantial array of natural medicines plays a significant role in mitigating drug-induced cardiotoxicity (Qu et al., 2022).

Traditional Chinese medicine has gained increasing attention for the treatment of various diseases. The protective effects of Rg3 on drug-induced cardiotoxicity have been proved in experiments. Rg3 demonstrated a remarkable ability to inhibit the upregulation of ROS and malondialdehyde, in the meantime, promote superoxide dismutase (SOD), and restore the balance of SOD/glutathione peroxidase in DOX-treated cardiac microvascular endothelial cells and rats through activating the Nrf2/antioxidant response element (ARE) and PI3K/Akt pathway (Wang X. et al., 2015). Furthermore, Rg3 could enhance endothelial function, thus playing a protective impact on cardiac function (Geng et al., 2020).

Another study showed the administration of Rg3 could enhance cardiac function in a dose-dependent manner. Specifically, in cardiotoxicity induced by microcystin, Rg3 acted by downregulating miR-128-3p and upregulating the expression of double minute 4 protein (MDM4), thereby mitigating oxidative stress and reducing cell apoptosis (Zhou and Xia, 2023).

4.4 Diabetic cardiomyopathy (DCM)

DCM is a pathophysiological condition induced by diabetes, potentially leading to HF. The diminished performance of the diabetic heart is attributed to multiple factors, including hyperglycemia, elevated fatty acids, and inflammatory cytokines (Dillmann, 2019). Current therapeutic options for DCM primarily comprise sodium-glucose cotransporter 2 inhibitors, glucagon-like peptide-1 receptor agonists, metformin, thiazolidinediones, and dipeptidyl peptidase 4 inhibitors. While clinical trials have confirmed their effectiveness in ameliorating cardiac dysfunction, their ability to fully cure or substantially improve the prognosis of DCM remains limited. Consequently, there is an urgent need to discover novel treatments specifically targeting DCM. A recent research highlights the potential of Rg3 in DCM management (Zhang et al., 2023). Rg3 appears to protect against DCM by modulating glucose and lipid metabolism, achieved through direct binding to peroxisome proliferator-activated receptor γ (PPAR- γ) and stimulating the adiponectin pathway. Additionally, Rg3 reduces proinflammatory cytokines and mitigates mitochondrial dysfunction. In this study, metformin served as a positive control, and varying doses of Rg3 were evaluated in DCM mice models. The findings indicated that high doses of Rg3, similar to metformin, effectively improved body weight, blood glucose, body fat, and serum lipid levels. However, lower doses of Rg3 did not exhibit these benefits.

Moreover, high-dose Rg3 significantly decreased serum creatine kinase (CK), creatine kinase-MB (CK-MB), and lactate dehydrogenase (LDH) levels in diabetic mice, indicative of reduced cardiac dysfunction. In contrast, metformin and low-dose Rg3 only partially improved CK and LDH levels. Therefore, high-dose Rg3 demonstrated more comprehensive systemic effects than metformin.

5 Pharmacological action of Rg3 in mental disorders

5.1 MDD

MDD, commonly referred to as depression, is a long-lasting, recurring, and possibly life-threatening psychiatric illness that impacts up to 20% of the global population (Nabavi et al., 2017). It is usually characterized by diminished self-esteem, cognitive and emotional impairments, reduced energy levels, as well as unexplained pain. Depression has been recognized as one of the primary contributors to the global burden of disease (Akpinar and Karadağ, 2022). The management of depression has become a critical issue for humanity. Primary drugs include serotonin reuptake inhibitors, serotonin-norepinephrine reuptake inhibitors, serotonin modulators, and atypical antidepressants (Chin et al., 2022). Meta-analyses indicate that the efficacy of existing antidepressants is observed in only about half to one-third of patients with depression (Kennedy et al., 2009; Jensen et al., 2023). Moreover, a significant number of patients experience recurrent episodes of depression. Consequently, there is an urgent need to explore and develop new antidepressant medications. Several investigations have demonstrated that Rg3 exhibits a therapeutic effect on depression.

Rg3 could alleviate depression-like behaviors such as anorexia, anhedonia, and decreased social exploration (Kang et al., 2017; You et al., 2017; Zhang et al., 2017). The exact mechanism refers to inhibiting inflammation cytokines such as IL-1 β and IL-6, restoring the balance of TRP-KYN metabolism, enhancing cell proliferation, and suppressing apoptosis. In MDD patients and animal models of depression, the TRP-KYN metabolic pathway is over-activated (Parrott et al., 2016). The metabolic processing of neurotoxic KYN initiates depressive-like behavior after peripheral immune activation. However, Rg3 administration could decrease the level of KYN (Kang et al., 2017). Moreover, one extensively supported hypothesis regarding depression is the neurotrophic hypothesis, positing that the pathogenesis of depression is associated with impaired functioning of the BDNF system within the brain (Colucci-D'Amato et al., 2020). BDNF is crucial for neural signal transduction and the facilitation of neuronal plasticity, achieved through its specific binding and subsequent activation of tropomyosin-related kinase B (TrkB) receptors (Moya-Alvarado et al., 2022; Pahlavani, 2023). In the chronic mild stress mouse model, Rg3 markedly enhanced the expression of BDNF and the phosphorylation of cyclic adenosine monophosphate response element binding protein (CREB), thereby mitigating depressive symptoms (You et al., 2017). Meanwhile, fluoxetine was employed as a positive control, revealing that to achieve a comparable antidepressant effect to that of fluoxetine, a higher dosage of Rg3 is necessary (You et al., 2017). Additionally, it was

observed that chronic stress exposure markedly decreased body weight. While fluoxetine was effective in mitigating this weight loss, Rg3 exhibited no significant impact on body weight.

5.2 Anxiety disorders

Anxiety disorders constitute the most prevalent category of mental illness, typically originating before or during early adulthood. Key characteristics encompass excessive fear and anxiety or avoidance of perceived threats that are persistent and impairing (Penninx et al., 2021). In adults, anxiety prevention has been evaluated in a few trials of selective or indicated prevention (Deady et al., 2017). Effective treatments for anxiety disorders are available, which not only alleviate symptoms of anxiety but also enhance overall quality of life and functioning. Both pharmacotherapy and psychotherapy are regarded as primary approaches to managing anxiety disorders. Studies have demonstrated that routine medications are mildly to moderately effective in treating these disorders, although there is a noted variability in response rates (Penninx et al., 2021). Chinese herbs have shown therapeutic effects on anxiety including Rg3.

Rg3 has shown a beneficial role in GABA_A receptor-related anxiety (Lee et al., 2013). GABA_A receptors consist of three subunits ($\alpha 1$, $\beta 1$, $\gamma 2$). The GABA_A receptor is responsible for fast inhibitory synaptic transmission. The principal physiological and pharmacological functions of GABA_A receptors encompass the mitigation of anxiety symptoms in patients. Additionally, the $\gamma 2$ subunit of the GABA_A receptor is pivotal in the control and management of human epilepsy (Lee et al., 2013). In a study, Rg3 exhibited enhancing effects on the GABA-induced inward current (I_{GABA}) with $\gamma 2$ subunit in a dependent manner. When the $\gamma 2$ subunit expression ratio was raised, the degree of the Rg3-induced activation of the GABA_A receptor increased (Lee et al., 2013).

5.3 Post-traumatic stress disorder (PTSD)

PTSD is a severe psychiatric condition linked to substantial distress and impaired functional capacity (Bryant et al., 2023). Psychological therapies are established as the primary treatment modality for PTSD (Bisson and Olff, 2021). Numerous systematic reviews have consistently affirmed the efficacy of these therapies in addressing PTSD symptoms (Bisson et al., 2013). Nevertheless, the effectiveness of psychological interventions may fluctuate based on individual factors, including the intensity of PTSD. Consequently, pharmacotherapy plays a crucial role in the symptom management of PTSD patients. While common pharmacological treatments have shown encouraging outcomes in diminishing the severity of PTSD symptoms, their overall efficacy remains moderate, and they may precipitate adverse reactions. Therefore, there is an imperative need for the exploration and development of innovative therapeutic strategies.

Rg3 has been reported to have a role in improving fear memory and spatial memory (Sur and Lee, 2022a). The HPA axis and monoamine imbalance in the medial prefrontal cortex and

hippocampus contribute to the pathogenesis of PTSD. However, Rg3 administration could be involved in regulating the HPA axis and BDNF-TrkB pathway (Sur and Lee, 2022a). In rats administered with Rg3, a significant decrease in serum corticosterone and adrenocorticotrophic hormone levels was observed, alongside a marked increase in BDNF, TrkB, catecholamine, and 5-HT concentrations. The study utilized paroxetine hydrochloride as a positive control. Compared to paroxetine hydrochloride, Rg3 demonstrated superior effects in behavioral tests. Notably, a dosage of 50 mg/kg of Rg3 was effective in elevating 5-HT levels, while significant differences were observed in the response to paroxetine hydrochloride treatment.

5.4 ADHD

ADHD is a mental disorder that typically emerges in childhood and is characterized by developmentally inappropriate and impairing inattention, motor hyperactivity, and impulsivity, with difficulties often continuing into adulthood. Medication and behavioral therapies are widely recognized and frequently utilized treatments for ADHD, demonstrating substantial efficacy and notable rates of symptom remission. Furthermore, a recent meta-analysis has revealed a significant preference for herbal medicine as an effective treatment option for ADHD (Dutta et al., 2022). An open-label pilot study demonstrated that the combination of omega-3 and Korean red ginseng may improve ADHD symptoms and cognitive functions including attention, memory, and executive function in children with ADHD (Lee J. et al., 2020). Additionally, YY162, consisting of Rg3 extracted from *P. ginseng* C. A. Mey. and *Ginkgo biloba* L. (Ben-Azu et al., 2022b; Asiwe et al., 2023), exerted protective activity on ADHD through modulating oxidative metabolism and the BDNF/TrkB pathway (Nam et al., 2014). Yet, the mechanism of Rg3 on ADHD requires further research and exploration.

6 Rg3 in the coexistence of cardiac diseases and psychiatric disorders

Current investigations into the therapeutic effects and mechanisms of Rg3, especially concerning the comorbidity of heart and mental disorders, are still in their early stages. Nowadays, there is substantial basic research supporting Rg3's efficacy in treating heart or mental disorders separately, based on their relationship, Rg3 may show potential effects in treating their comorbidity (Figure 3).

Rg3 might play a dual preventive role, potentially arresting the progression or aiding the recovery of mental disorders when used in cardiac disease treatments. Its potential mechanisms might involve modulating inflammation and oxidative stress. In cardiac disease management, Rg3 chiefly functions by reducing inflammation and curtailing oxidative stress. Anti-inflammatory action is linked to the regulation of TRP/KYN metabolism, enhancing 5-HT and BDNF levels, and thereby alleviating mental disorders (Kuo et al., 2021; Fellendorf et al., 2022; Gong et al., 2023). MDD is characterized by reduced antioxidant concentrations in plasma (Bhatt et al., 2020). Oxidative stress, characterized by the overproduction of ROS and

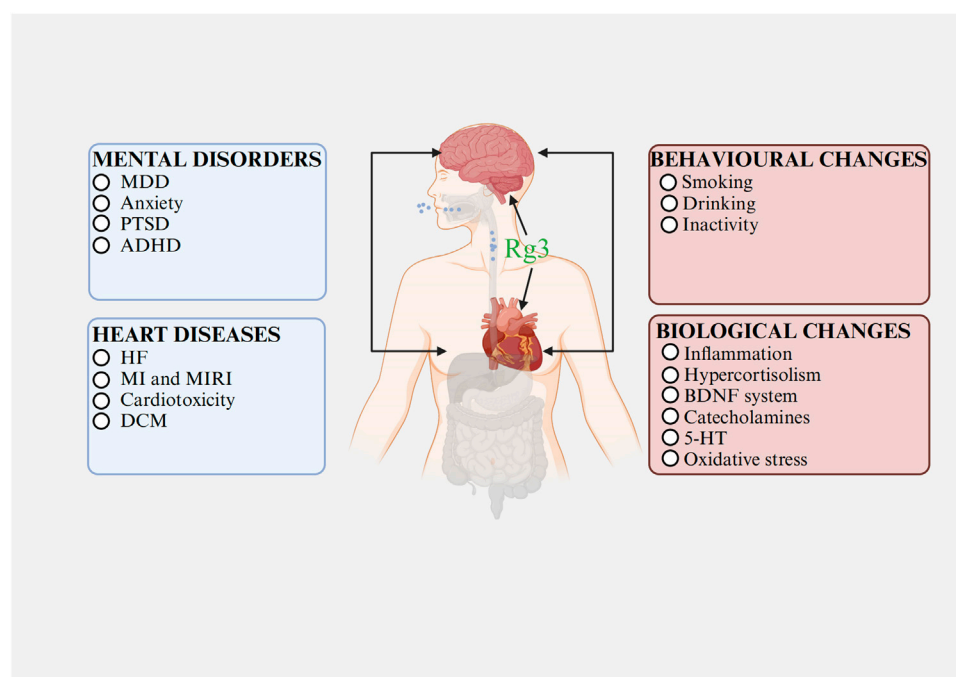


FIGURE 3

The relationships between heart diseases and mental disorders Abbreviations: MDD, major depressive disorder; PTSD, post-traumatic stress disorder; ADHD, attention deficit hyperactivity disorder; HF, heart failure; MI, myocardial infarction; MIRI, myocardial ischemia-reperfusion injury; DCM, diabetic cardiomyopathy; BDNF, brain-derived neurotrophic factor; 5-HT, serotonin. Image created with [BioRender.com](https://www.biorender.com), with permission.

the depletion of antioxidative defenses, leads to pro-inflammatory signaling and induces cellular apoptosis (Bhatt et al., 2020). Counteracting oxidative stress can therefore diminish inflammation, reduce cellular death, and improve mental health conditions.

Furthermore, in the treatment of mental disorders, Rg3 may also enhance cardiac function. For mental conditions, Rg3 works by suppressing inflammation and HPA axis activity, reducing oxidative stress, boosting BDNF levels, and lowering catecholamine levels. Inflammation and oxidative stress are known to worsen myocardial fibrosis and impair cardiac function. Elevated cortisol levels can directly lead to hypertrophy of cardiomyocytes and cardiac remodeling (Braukyliene et al., 2022). A normal BDNF system is essential for cardiovascular development, while high catecholamine levels can cause vasoconstriction, contributing to cardiac injury, pathological remodeling, heart failure, myocardial ischemia, and necrosis (Du et al., 2021; Agorrody et al., 2022; Tsai et al., 2022). Thus, the mechanism through which Rg3 enhances cardiac function while ameliorating mental disorders may involve modulating inflammation, catecholamine, the BDNF system, the HPA axis, and oxidative stress.

7 Adverse reactions of Rg3

Few studies have shown the side effects of Rg3. Existing clinical investigations of Rg3 at various dosages have not revealed any harmful reaction. The sole adverse observation demonstrated

increased but reversible kidney weight in dogs that received 60 mg/kg 20(S)-Rg3 (Gao et al., 2020; Nakhjavani et al., 2020). However, given that Rg3 is extracted from ginseng, high doses of ginseng (15 g/d) will lead to ginseng abuse syndrome. Most symptoms encompass headache, dizziness, breast pain, nausea, asthma, and so on (Siegel, 1979; Deng et al., 2023). Although Rg3 offers a broad spectrum of clinical applications in cancer patients, it should not be abused. Taking the recommended dose of Rg3 will not cause serious adverse reactions. Due to the multiple ingredients of ginseng, the adverse effects of Rg3 are not completely equal to ginseng and need to be explored.

8 Conclusion and perspectives

Recent studies have provided increasing evidence of the broad pharmacological effects of Rg3 in treating heart diseases and mental disorders through a variety of signaling pathways and influencing changes at the transcriptional level (Figure 4; Figure 5).

However, the majority of studies primarily concentrate on *in vitro* cell or animal models, with less emphasis on clinical research. The shenyi capsule, comprising Rg3 monomer, is currently employed in the clinical treatment of cancer patients. Clinical studies have demonstrated that shenyi capsules mitigate the toxicity of platinum-based chemotherapy in non-small-cell lung cancer (NSCLC), including a reduction in platelet toxicity (Xu et al., 2020). Additionally, in a double-blind, randomized, crossover study involving 23 participants, oral administration of Rg3-enriched Korean red ginseng at a dose of 400 mg/day for 7 days

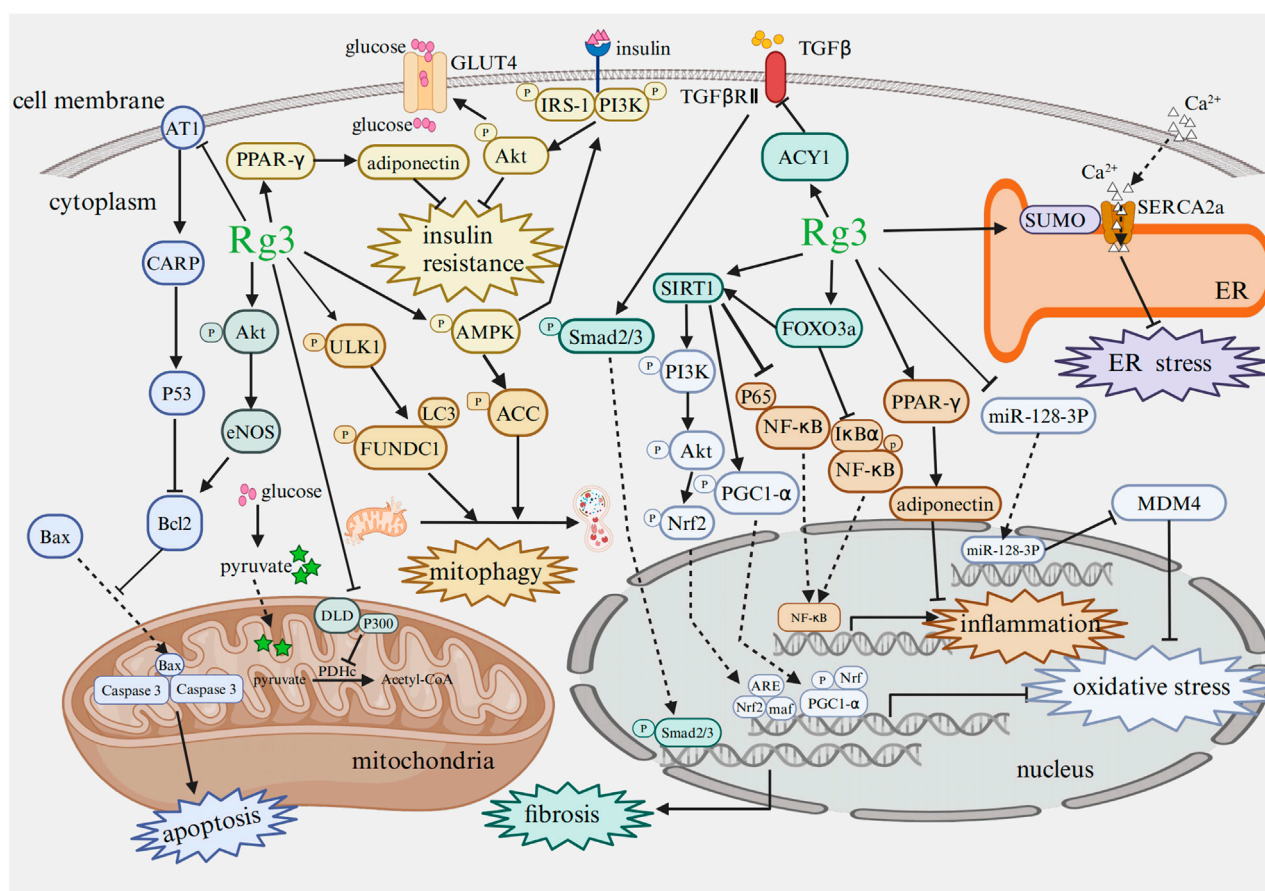


FIGURE 4

Mechanism of Rg3 on cardiac protection Abbreviations: AT1, angiotensin type 1 receptor; CARP, cardiac ankyrin repeat protein; Bcl2, B cell lymphoma-2; Bax, Bcl2-associated X protein; Akt, protein kinase B; eNOS, endothelial nitric oxide synthase; DLD, dihydrolipoamide dehydrogenase; GLUT4, glucose transporter 4; PDHc, pyruvate dehydrogenase complex; ULK1, Unc51-like-kinase 1; FUNDC1, FUN14 domain-containing protein 1; LC3, microtubule-associated protein 1 light chain 3; PI3K, phosphoinositide 3 kinase; IRS, insulin receptor substrate; AMPK, AMP-activated protein kinase; ACC, acetyl CoA carboxylase; Nrf, nuclear factor erythroid 2-related factor; PGC1-α, peroxisome proliferators-activated receptor γ coactivator-1α; PPAR-γ, peroxisome proliferator-activated receptor γ; NF-κB, nuclear factor κB; IκBα, inhibitor of kappa B alpha; ARE, antioxidant response element; maf, musculoaponeurotic fibrosarcoma oncogene homolog; SIRT1, sirtuin 1; FOXO3a, forkhead box O3a; ER, endoplasmic reticulum; SERCA2a, sarcoplasmic/endoplasmic reticulum Ca²⁺-ATPase; MDM4, double minute 4 protein. Image created with BioRender.com, with permission.

demonstrated a decrease in aortic stiffness and central blood pressure (Jovanovski et al., 2014). These findings suggest that Rg3 supplementation may help reduce risk factors for heart diseases such as platelet abnormality, vascular stiffness, and hypertension. However, a clinical study was conducted to recruit NSCLC patients with normal cardiac function. The study showed the treatment group with exclusive administration of Rg3 and the control group with placebo treatment exhibited comparable cardiac functions before and after the therapy (Li D.-R. et al., 2023). Furthermore, an open-label pilot study indicated that a combination of omega-3 and Korean red ginseng, which is rich in Rg3, could potentially alleviate symptoms of ADHD (Lee J. et al., 2020).

However, to date, there is no research exploring the exclusive or combined clinical use of Rg3 for treating heart diseases or mental disorders or the comorbidity of both in humans. The limited clinical research on Rg3 may be attributed to several factors. First, existing studies indicate that a higher concentration of Rg3 is necessary to

attain similar therapeutic outcomes as routine drugs (You et al., 2017). Then, the utilization of Rg3 in the clinic is restrictive due to low bioavailability and poor intestinal absorption capability with a percentage of approximately 10% (Xiong et al., 2023). Moreover, common methods for preparing Rg3, such as heat treatment, acid-base treatment, and biological transformation (Fu, 2019; Siddiqi et al., 2020), are hindered by the low natural concentration of Rg3 in ginseng plants (Yan et al., 2019). This results in the inability of most methods to achieve large-scale production of Rg3. Finally, there is a necessity to increase awareness and understanding of Rg3's therapeutic potential in mental disorders and its possible synergistic use in cases of mental disorders co-occurring with heart diseases.

Given these potential reasons, clinical practitioners and researchers are urged to pursue further studies on Rg3. Currently, Rg3 is primarily administered orally. However, novel administration methods, such as catheter-based endocardial or intramyocardial injections, should be explored to enhance its

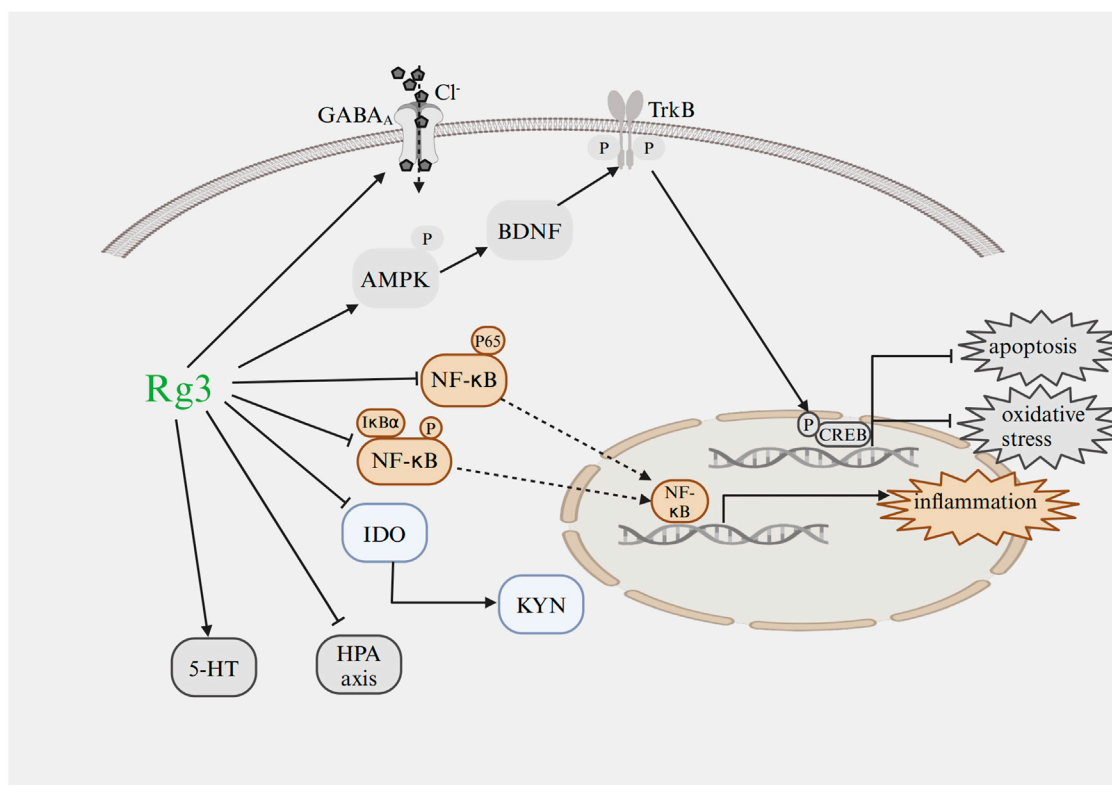


FIGURE 5

Mechanism of Rg3 on mental disorders Abbreviations: GABA, gamma aminobutyric acid; IDO, indoleamine 2,3-dioxygenase; NF-κB, nuclear factor κB; IκBα, inhibitor of kappa B alpha; KYN, kynurenine; HPA, hypothalamic-pituitary-adrenal; CREB, cyclic adenosine monophosphate response element binding protein; TrkB, tropomyosin-related kinase B; BDNF, brain-derived neurotrophic factor; 5-HT, serotonin; AMPK, AMP-activated protein kinase. Image created with BioRender.com, with permission.

clinical utility. Polyethylene glycol (PEG) is commonly utilized as a hydrophilic block in drug delivery systems for its resistance to protein adsorption and minimal toxicity (Li et al., 2020). Conversely, polypropylene sulfide (PPS) is favored as the hydrophobic block due to its pronounced hydrophobic properties. This PEG-b-PPS amphiphilic block copolymer demonstrates potential as a ROS-responsive nanovesicle, specifically designed for efficient drug delivery (Li et al., 2020). The combination of PEG and PPS offers a promising platform for enhancing the therapeutic effects of Rg3 (Li et al., 2020). Additionally, the development of Rg3-loaded nanoparticles with specific targeting abilities could enable non-invasive, tissue-specific drug delivery (Li et al., 2020). Furthermore, there is also a pressing need for extensive, well-controlled clinical trials to more accurately determine Rg3's benefits on cardiac and mental health. Regarding Rg3, there exist two variants: 20(R)-Rg3 and 20(S)-Rg3. The distinct impacts of these variants on cardiac and mental health remain to be elucidated. Given that these two epimers have already shown varied anticancer effects, their potential differential influence on heart diseases and mental disorders presents a research opportunity.

In summary, we have reviewed recent findings that illustrate the important role of Rg3 as a novel therapeutic agent in heart diseases and

mental disorders. More clinical trials are motivated to be investigated for Rg3 in heart and mental diseases, especially in patients with the coexistence of cardiac diseases and psychiatric disorders.

Author contributions

LS: Writing–original draft. JL: Writing–review and editing. XW: Writing–review and editing. XX: Writing–review and editing. LT: Writing–review and editing.

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Conflict of interest

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References

- Agorrody, G., Peclat, T. R., Peluso, G., Gonano, L. A., Santos, L., van Schooten, W., et al. (2022). Benefits in cardiac function by CD38 suppression: improvement in NAD(+) levels, exercise capacity, heart rate variability and protection against catecholamine-induced ventricular arrhythmias. *J. Mol. Cell. Cardiol.* 166, 11–22. doi:10.1016/j.jmcc.2022.01.008
- Akpınar, Ş., and Karadağ, M. G. (2022). Is vitamin D important in anxiety or depression? What is the truth? *Curr. Nutr. Rep.* 11 (4), 675–681. doi:10.1007/s13668-022-00441-0
- Asiwe, J. N., Ekene, E. N., Agbugba, L. C., Moke, E. G., Akintade, A. V., Ben-Azu, B., et al. (2023). Ginkgo biloba supplement abates lead-induced endothelial and testicular dysfunction in Wistar rats via up-regulation of Bcl-2 protein expression, pituitary-testicular hormones and down-regulation of oxido-inflammatory reactions. *J. Trace. Elem. Med. Biol.* 79, 127216. doi:10.1016/j.jtemb.2023.127216
- Ben-Azu, B., Adebayo, O. G., Jarikre, T. A., Oyoewi, M. O., Edje, K. E., Omogbiya, I. A., et al. (2022a). Taurine, an essential β -amino acid insulates against ketamine-induced experimental psychosis by enhancement of cholinergic neurotransmission, inhibition of oxidative/nitrogenic imbalances, and suppression of COX-2/iNOS immunoreactions in mice. *Metab. Brain. Dis.* 37 (8), 2807–2826. doi:10.1007/s11011-022-01075-5
- Ben-Azu, B., Adebayo, O. G., Wopara, I., Aduema, W., Onyeleonu, I., Umoren, E. B., et al. (2022b). Lead acetate induces hippocampal pyramidal neuron degeneration in mice via up-regulation of executioner caspase-3, oxido-inflammatory stress expression and decreased BDNF and cholinergic activity: reversal effects of Ginkgo biloba supplement. *J. Trace. Elem. Med. Biol.* 71, 126919. doi:10.1016/j.jtemb.2021.126919
- Ben-Azu, B., Uruaka, C. I., Ajayi, A. M., Jarikre, T. A., Nwangwa, K. E., Chilaka, K. C., et al. (2023). Reversal and preventive pleiotropic mechanisms involved in the antipsychotic-like effect of taurine, an essential β -amino acid in ketamine-induced experimental schizophrenia in mice. *Neurochem. Res.* 48 (3), 816–829. doi:10.1007/s11064-022-03808-5
- Bertero, E., and Maack, C. (2018). Metabolic remodelling in heart failure. *Nat. Rev. Cardiol.* 15 (8), 457–470. doi:10.1038/s41569-018-0044-6
- Bhagwagar, Z., Hafizi, S., and Cowen, P. J. (2002). Cortisol modulation of 5-HT-mediated growth hormone release in recovered depressed patients. *J. Affect. Disord.* 72 (3), 249–255. doi:10.1016/s0165-0327(01)00467-0
- Bhatt, S., Nagappa, A. N., and Patil, C. R. (2020). Role of oxidative stress in depression. *Drug Discov. Today* 25 (7), 1270–1276. doi:10.1016/j.drudis.2020.05.001
- Bisson, J. I., and Olff, M. (2021). Prevention and treatment of PTSD: the current evidence base. *Eur. J. Psychotraumatol.* 12 (1), 1824381. doi:10.1080/20008198.2020.1824381
- Bisson, J. I., Roberts, N. P., Andrew, M., Cooper, R., and Lewis, C. (2013). Psychological therapies for chronic post-traumatic stress disorder (PTSD) in adults. *Cochrane Database Syst. Rev.* 2013 (12), CD003388. doi:10.1002/14651858.CD003388.pub4
- Bornand, D., Reinau, D., Jick, S. S., and Meier, C. R. (2022). β -blockers and the risk of depression: a matched case-control study. *Drug Saf.* 45 (2), 181–189. doi:10.1007/s40264-021-01140-5
- Bothou, C., Beuschlein, F., and Spyroglou, A. (2020). Links between aldosterone excess and metabolic complications: a comprehensive review. *Diabetes Metab.* 46 (1), 1–7. doi:10.1016/j.diabet.2019.02.003
- Braukyliene, R., Aldujeli, A., Haq, A., Maciulevicius, L., Jankauskaite, D., Jurenas, M., et al. (2022). Impact of mineralocorticoid receptor gene NR3C2 on the prediction of functional classification of left ventricular remodeling and arrhythmia after acute myocardial infarction. *Int. J. Environ. Res. Public Health* 20 (1), 12. doi:10.3390/ijerph20010012
- Bryant, R. A., McFarlane, A. C., Silove, D., O'Donnell, M. L., Forbes, D., and Creamer, M. (2023). The lingering impact of resolved PTSD on subsequent functioning. *Focus (Am. Psychiatr. Publ.)* 21 (3), 290–295. doi:10.1176/appi.focus.23021016
- Cai, S., Zhao, M., Zhou, B., Yoshii, A., Bugg, D., Villet, O., et al. (2023). Mitochondrial dysfunction in macrophages promotes inflammation and suppresses repair after myocardial infarction. *J. Clin. Invest.* 133 (4), e159498. doi:10.1172/jci159498
- Campana, P., Palaia, M. E., Conte, M., Cante, T., Petraglia, L., Femminella, G. D., et al. (2022). The elderly at risk: aldosterone as modulator of the immune response to SARS-CoV-2 infection. *Geroscience* 44 (2), 567–572. doi:10.1007/s11357-021-00481-4
- Cannavo, A., Jun, S., Rengo, G., Marzano, F., Agrimi, J., Liccardo, D., et al. (2023). β 3AR-dependent brain-derived neurotrophic factor (BDNF) generation limits chronic postischemic heart failure. *Circ. Res.* 132 (7), 867–881. doi:10.1161/circresaha.122.321583
- Carnovale, C., Perrotta, C., Baldelli, S., Cattaneo, D., Montrasio, C., Barbieri, S. S., et al. (2023). Antihypertensive drugs and brain function: mechanisms underlying therapeutically beneficial and harmful neuropsychiatric effects. *Cardiovasc. Res.* 119 (3), 647–667. doi:10.1093/cvr/cvac110
- Chen, G. D., Yu, W. D., and Chen, X. P. (2016). Sirt1 activator represses the transcription of TNF- α in THP-1 cells of a sepsis model via deacetylation of H4K16. *Mol. Med. Rep.* 14 (6), 5544–5550. doi:10.3892/mmr.2016.5942
- Chen, J., Liu, Y., Pan, D., Xu, T., Luo, Y., Wu, W., et al. (2022). Estrogen inhibits endoplasmic reticulum stress and ameliorates myocardial ischemia/reperfusion injury in rats by upregulating SERCA2a. *Cell Commun. Signal.* 20 (1), 38–16. doi:10.1186/s12964-022-00842-2
- Chen, M. H., Hsu, J. W., Huang, K. L., Tsai, S. J., Tu, P. C., and Bai, Y. M. (2023). Inflammatory cytokines in and cognitive function of adolescents with first-episode schizophrenia, bipolar disorder, or major depressive disorder. *CNS Spectr.* 28 (1), 70–77. doi:10.1017/S1092852921000857
- Chin, T., Huyghebaert, T., Svrcek, C., and Oluboka, O. (2022). Individualized antidepressant therapy in patients with major depressive disorder: novel evidence-informed decision support tool. *Can. Fam. Physician* 68 (11), 807–814. doi:10.4674/cfp.6811807
- Coelho, A. R., Martins, T. R., Couto, R., Deus, C., Pereira, C. V., Simoes, R. F., et al. (2017). Berberine-induced cardioprotection and sirt3 modulation in doxorubicin-treated H9c2 cardiomyoblasts. *Biochim. Biophys. Acta Mol. Basis Dis.* 1863 (11), 2904–2923. doi:10.1016/j.bbdis.2017.07.030
- Colucci-D'Amato, L., Speranza, L., and Volpicelli, F. (2020). Neurotrophic factor BDNF, physiological functions and therapeutic potential in depression, neurodegeneration and brain cancer. *Int. J. Mol. Sci.* 21 (20), 7777. doi:10.3390/ijms21020777
- Correll, C. U., Detraux, J., De Lepeleire, J., and De Hert, M. (2015). Effects of antipsychotics, antidepressants and mood stabilizers on risk for physical diseases in people with schizophrenia, depression and bipolar disorder. *World Psychiatry* 14 (2), 119–136. doi:10.1002/wps.20204
- Crawford, A. A., Soderberg, S., Kirschbaum, C., Murphy, L., Eliasson, M., Ebrahim, S., et al. (2019). Morning plasma cortisol as a cardiovascular risk factor: findings from prospective cohort and mendelian randomization studies. *Eur. J. Endocrinol.* 181 (4), 429–438. doi:10.1530/eje-19-0161
- Deady, M., Choi, I., Calvo, R. A., Glozier, N., Christensen, H., and Harvey, S. B. (2017). eHealth interventions for the prevention of depression and anxiety in the general population: a systematic review and meta-analysis. *BMC Psychiatry* 17 (1), 310–314. doi:10.1186/s12888-017-1473-1
- Deurwaerdere, P., and Di Giovanni, G. (2020). Serotonin in health and disease. *Int. J. Mol. Sci.* 21 (10), 3500. doi:10.3390/ijms21103500
- Deng, W., Liu, H., Guo, L., Liu, Y., and Ma, Z. (2023). Panax ginseng abuse exhibits a pro-inflammatory effect by activating the NF- κ B pathway. *Food Sci. Nutr.* 11 (5), 2130–2140. doi:10.1002/fsn3.3011
- Dillmann, W. H. (2019). Diabetic cardiomyopathy. *Circ. Res.* 124 (8), 1160–1162. doi:10.1161/circresaha.118.314665
- Doenst, T., Haverich, A., Serruys, P., Bonow, R. O., Kappetein, P., Falk, V., et al. (2019). PCI and CABG for treating stable coronary artery disease: JACC review topic of the week. *J. Am. Coll. Cardiol.* 73 (8), 964–976. doi:10.1016/j.jacc.2018.11.053
- Du, Y., Demillard, L. J., and Ren, J. (2021). Catecholamine-induced cardiotoxicity: a critical element in the pathophysiology of stroke-induced heart injury. *Life Sci.* 287, 120106. doi:10.1016/j.lfs.2021.120106
- Du, Y., Li, J., Cai, C., Gong, F., Zhou, G., Liu, F., et al. (2023). Plantamajoside alleviates hypoxia-reoxygenation injury through integrin-linked kinase/c-Src/Akt and the mitochondrial apoptosis signaling pathways in H9c2 myocardial cells. *BMC Complement. Med. Ther.* 23 (1), 64. doi:10.1186/s12906-023-03880-6
- Dutta, T., Anand, U., Mitra, S. S., Ghorai, M., Jha, N. K., Shaikh, N. K., et al. (2022). Phytotherapy for attention deficit hyperactivity disorder (ADHD): a systematic review and meta-analysis. *Front. Pharmacol.* 13, 827411. doi:10.3389/fphar.2022.827411
- Emudainohwo, J. O. T., Ben-Azu, B., Adebayo, O. G., Aduema, W., Uruaka, C., Ajayi, A. M., et al. (2023). Normalization of HPA axis, cholinergic neurotransmission, and inhibiting brain oxidative and inflammatory dynamics Are Associated with the adaptogenic-like effect of rutin against psychosocial defeat stress. *J. Mol. Neurosci.* 73 (1), 60–75. doi:10.1007/s12031-022-02084-w

- Favero, V., Cremaschi, A., Falchetti, A., Gaudio, A., Gennari, L., Scillitani, A., et al. (2021). Management and medical therapy of mild hypercortisolism. *Int. J. Mol. Sci.* 22 (21), 11521. doi:10.3390/ijms222111521
- Fellendorf, F. T., Bonkat, N., Dalkner, N., Schönthaler, E. M. D., Manchia, M., Fuchs, D., et al. (2022). Indoleamine 2,3-dioxygenase (Ido)-activity in severe psychiatric disorders: a systematic review. *Curr. Top. Med. Chem.* 22 (25), 2107–2118. doi:10.2174/1568026622666220718155616
- Fillmore, N., Levasseur, J. L., Fukushima, A., Wagg, C. S., Wang, W., Dyck, J. R. B., et al. (2018). Uncoupling of glycolysis from glucose oxidation accompanies the development of heart failure with preserved ejection fraction. *Mol. Med.* 24 (1), 3. doi:10.1186/s10020-018-0005-x
- Fu, Y. (2019). Biotransformation of ginsenoside Rb1 to Gyp-XVII and minor ginsenoside Rg3 by endophytic bacterium *Flavobacterium* sp. GE 32 isolated from *Panax ginseng*. *Lett. Appl. Microbiol.* 68 (2), 134–141. doi:10.1111/lam.13090
- Gao, Y., Wang, G., Wang, T., Li, G., Lin, J., Sun, L., et al. (2020). A 26-week 20(S)-ginsenoside Rg3 oral toxicity study in Beagle dogs. *Regul. Toxicol. Pharmacol.* 110, 104522. doi:10.1016/j.yrtph.2019.104522
- García-Eguren, G., Giró, O., Romero, M. D. M., Grasa, M., and Hanzu, F. A. (2019). Chronic hypercortisolism causes more persistent visceral adiposity than HFD-induced obesity. *J. Endocrinol.* 242 (2), 65–77. doi:10.1530/joe-19-0168
- Geng, J., Fu, W., Yu, X., Lu, Z., Liu, Y., Sun, M., et al. (2020). Ginsenoside Rg3 alleviates ox-LDL induced endothelial dysfunction and prevents atherosclerosis in ApoE(-/-) mice by regulating PPAR γ /FAK signaling pathway. *Front. Pharmacol.* 11, 500. doi:10.3389/fphar.2020.00500
- Gong, X., Chang, R., Zou, J., Tan, S., and Huang, Z. (2023). The role and mechanism of tryptophan - kynurenine metabolic pathway in depression. *Rev. Neurosci.* 34 (3), 313–324. doi:10.1515/revneuro-2022-0047
- Güder, G., Bauersachs, J., Frantz, S., Weismann, D., Alolio, B., Ertl, G., et al. (2007). Complementary and incremental mortality risk prediction by cortisol and aldosterone in chronic heart failure. *Circulation* 115 (13), 1754–1761. doi:10.1161/circulationaha.106.653964
- Hagi, K., Nosaka, T., Dickinson, D., Lindenmayer, J. P., Lee, J., Friedman, J., et al. (2021). Association between cardiovascular risk factors and cognitive impairment in people with schizophrenia: a systematic review and meta-analysis. *JAMA Psychiatry* 78 (5), 510–518. doi:10.1001/jamapsychiatry.2021.0015
- Hartup, J., and Mann, D. L. (2017). Neurohormonal activation in heart failure with reduced ejection fraction. *Nat. Rev. Cardiol.* 14 (1), 30–38. doi:10.1038/nrcardio.2016.163
- Huang, H., Luo, Z., Qi, S., Huang, J., Xu, P., Wang, X., et al. (2018). Landscape of the regulatory elements for lysine 2-hydroxyisobutyrylation pathway. *Cell Res.* 28 (1), 111–125. doi:10.1038/cr.2017.149
- Hubers, S. A., and Brown, N. J. (2016). Combined angiotensin receptor antagonism and neprilysin inhibition. *Circulation* 133 (11), 1115–1124. doi:10.1161/circulationaha.115.018622
- Jayanti, S., Dalla Verde, C., Tiribelli, C., and Gazzin, S. (2023). Inflammation, dopaminergic brain and bilirubin. *Int. J. Mol. Sci.* 24 (14), 11478. doi:10.3390/ijms241411478
- Jensen, K. H. R., Dam, V. H., Ganz, M., Fisher, P. M., Ip, C. T., Sankar, A., et al. (2023). Deep phenotyping towards precision psychiatry of first-episode depression-the brain drugs-depression cohort. *BMC Psychiatry* 23 (1), 151. doi:10.1186/s12888-023-04618-x
- Jovanovski, E., Bateman, E. A., Bhardwaj, J., Fairgrieve, C., Mucalo, I., Jenkins, A. L., et al. (2014). Effect of Rg3-enriched Korean red ginseng (*Panax ginseng*) on arterial stiffness and blood pressure in healthy individuals: a randomized controlled trial. *J. Am. Soc. Hypertens.* 8 (8), 537–541. doi:10.1016/j.jash.2014.04.004
- Kang, A., Xie, T., Zhu, D., Shan, J., Di, L., and Zheng, X. (2017). Suppressive effect of ginsenoside Rg3 against lipopolysaccharide-induced depression-like behavior and neuroinflammation in mice. *J. Agric. Food Chem.* 65 (32), 6861–6869. doi:10.1021/acs.jafc.7b02386
- Kennedy, S. H., Andersen, H. F., and Thase, M. E. (2009). Escitalopram in the treatment of major depressive disorder: a meta-analysis. *Curr. Med. Res. Opin.* 25 (1), 161–175. doi:10.1185/03007990802622726
- Kermani, P., and Hempstead, B. (2019). BDNF actions in the cardiovascular system: roles in development, adulthood and response to injury. *Front. Physiol.* 10, 455. doi:10.3389/fphys.2019.00455
- Kim, J., Yun, K. S., Cho, A., Kim, D. H., Lee, Y. K., Choi, M. J., et al. (2022). High cortisol levels are associated with oxidative stress and mortality in maintenance hemodialysis patients. *BMC Nephrol.* 23 (1), 98. doi:10.1186/s12882-022-02722-w
- Kim, M. J., Lee, H., Chanda, D., Thoudam, T., Kang, H. J., Harris, R. A., et al. (2023). The role of pyruvate metabolism in mitochondrial quality control and inflammation. *Mol. Cells* 46 (5), 259–267. doi:10.14348/molcells.2023.2128
- Kuo, C. Y., Lin, C. H., and Lane, H. Y. (2021). Molecular basis of late-life depression. *Int. J. Mol. Sci.* 22 (14), 7421. doi:10.3390/ijms22147421
- Lai, Q., Liu, F. M., Rao, W. L., Yuan, G. Y., Fan, Z. Y., Zhang, L., et al. (2022). Aminoacylase-1 plays a key role in myocardial fibrosis and the therapeutic effects of 20(S)-ginsenoside Rg3 in mouse heart failure. *Acta Pharmacol. Sin.* 43 (8), 2003–2015. doi:10.1038/s41401-021-00830-1
- Lee, B. H., Kim, H. J., Chung, L., and Nah, S. Y. (2013). Ginsenoside Rg3 regulates GABAA receptor channel activity: involvement of interaction with the γ_2 subunit. *Eur. J. Pharmacol.* 705 (1–3), 119–125. doi:10.1016/j.ejphar.2013.02.040
- Lee, H., Hong, Y., Tran, Q., Cho, H., Kim, M., Kim, C., et al. (2019). A new role for the ginsenoside Rg3 in antiaging via mitochondria function in ultraviolet-irradiated human dermal fibroblasts. *J. Ginseng Res.* 43 (3), 431–441. doi:10.1016/j.jgr.2018.07.003
- Lee, H., Kong, G., Tran, Q., Kim, C., Park, J., and Park, J. (2020a). Relationship between ginsenoside Rg3 and metabolic syndrome. *Front. Pharmacol.* 11, 130. doi:10.3389/fphar.2020.00130
- Lee, J., Lee, A., Kim, J. H., Shin, Y. M., Kim, S. J., Cho, W. D., et al. (2020b). Effect of omega-3 and Korean red ginseng on children with attention deficit hyperactivity disorder: an open-label pilot study. *Clin. Psychopharmacol. Neurosci.* 18 (1), 75–80. doi:10.9758/cpn.2020.18.1.75
- Li, D., Zhang, G., Wang, Z., Guo, J., Liu, Y., Lu, Y., et al. (2023b). Idebenone attenuates ferroptosis by inhibiting excessive autophagy via the ROS-AMPK-mTOR pathway to preserve cardiac function after myocardial infarction. *Eur. J. Pharmacol.* 943, 175569. doi:10.1016/j.ejphar.2023.175569
- Li, D.-R., Hou, W., Hua, B.-J., Zhang, P.-T., Xiong, L., Liu, H., et al. (2023a). Shenyi capsule prolongs postoperative survival of patients with nonsmall cell lung cancer: a multicenter, randomized, controlled trial. *World J. Traditional Chin. Med.* 9 (3), 314–321. doi:10.4103/2311-8571.382023
- Li, L., Wang, Y., Guo, R., Li, S., Ni, J., Gao, S., et al. (2020). Ginsenoside Rg3-loaded, reactive oxygen species-responsive polymeric nanoparticles for alleviating myocardial ischemia-reperfusion injury. *J. Control. Release.* 317, 259–272. doi:10.1016/j.jconrel.2019.11.032
- Li, Y., Li, Y., Li, B., Liu, Y., Zhang, J., Kuang, W., et al. (2021). Antiplatelet therapy with integrated traditional Chinese and western medicine for use in myocardial ischemia-reperfusion injury: a review of clinical applications and mechanisms. *Evid. Based Complement. Altern. Med.* 2021, 7409094. doi:10.1155/2021/7409094
- Liu, Y., and Miao, J. (2022). An emerging role of defective copper metabolism in heart disease. *Nutrients* 14 (3), 700. doi:10.3390/nu14030700
- Liu, Z., Bian, X., Gao, W., Su, J., Ma, C., Xiao, X., et al. (2021). Rg3 promotes the SUMOylation of SERCA2a and corrects cardiac dysfunction in heart failure. *Pharmacol. Res.* 172, 105843. doi:10.1016/j.phrs.2021.105843
- Livi, L., Barletta, G., Martella, F., Saieva, C., Desideri, I., Bacci, C., et al. (2021). Cardioprotective strategy for patients with nonmetastatic breast cancer who are receiving an anthracycline-based chemotherapy: a randomized clinical trial. *JAMA Oncol.* 7 (10), 1544–1549. doi:10.1001/jamaoncol.2021.3395
- Lupisella, J. A., Shirude, P. S., Wurtz, N. R., and Garcia, R. A. (2022). Formyl peptide receptor 2 and heart disease. *Semin. Immunol.* 59, 101602. doi:10.1016/j.smim.2022.101602
- Mackman, N., Bergmeier, W., Stouffer, G. A., and Weitz, J. I. (2020). Therapeutic strategies for thrombosis: new targets and approaches. *Nat. Rev. Drug Discov.* 19 (5), 333–352. doi:10.1038/s41573-020-0061-0
- Manolis, A. A., Manolis, T. A., and Manolis, A. S. (2023). Neurohumoral activation in heart failure. *Int. J. Mol. Sci.* 24 (20), 15472. doi:10.3390/ijms242015472
- Marciniak, T. A., and Serebruany, V. (2019). Ranolazine, ACE inhibitors, and angiotensin receptor blockers. *Am. J. Med.* 132 (12), e844–e845. doi:10.1016/j.amjmed.2019.02.032
- McCarthy, C. P., Vaduganathan, M., McCarthy, K. J., Januzzi, J. L., Jr., Bhatt, D. L., and McEvoy, J. W. (2018). Left ventricular thrombus after acute myocardial infarction: screening, prevention, and treatment. *JAMA Cardiol.* 3 (7), 642–649. doi:10.1001/jamacardio.2018.1086
- Mendler, L., Braun, T., and Muller, S. (2016). The ubiquitin-like SUMO system and heart function: from development to disease. *Circ. Res.* 118 (1), 132–144. doi:10.1161/CIRCRESAHA.115.307730
- Menke, A. (2024). The HPA axis as target for depression. *Curr. Neuropharmacol.* 22 (5), 904–915. doi:10.2174/1570159x21666230811141557
- Misiak, B., Wójta-Kempa, M., Samochowiec, J., Schiweck, C., Aichholzer, M., Reif, A., et al. (2022). Peripheral blood inflammatory markers in patients with attention deficit/hyperactivity disorder (ADHD): a systematic review and meta-analysis. *Prog. Neuropsychopharmacol. Biol. Psychiatry* 118, 110581. doi:10.1016/j.pnpb.2022.110581
- Motta, J. R., Jung, I., Azzolin, V. F., Teixeira, C. F., Braun, L. E., De Oliveira Nerys, D. A., et al. (2021). Avocado oil (*Persea americana*) protects SH-SY5Y cells against cytotoxicity triggered by cortisol by the modulation of BDNF, oxidative stress, and apoptosis molecules. *J. Food Biochem.* 45 (2), e13596. doi:10.1111/jfbc.13596
- Moya-Alvarado, G., Guerra, M. V., Tiburcio, R., Bravo, E., and Bronfman, F. C. (2022). The Rab11-regulated endocytic pathway and BDNF/TrkB signaling: roles in plasticity changes and neurodegenerative diseases. *Neurobiol. Dis.* 171, 105796. doi:10.1016/j.nbd.2022.105796
- Nabavi, S. M., Daglia, M., Braid, N., and Nabavi, S. F. (2017). Natural products, micronutrients, and nutraceuticals for the treatment of depression: a short review. *Nutr. Neurosci.* 20 (3), 180–194. doi:10.1080/1028415X.2015.1103461

- Nakhjavani, M., Smith, E., Townsend, A. R., Price, T. J., and Hardingham, J. E. (2020). Anti-angiogenic properties of ginsenoside Rg3. *Molecules* 25 (21), 4905. doi:10.3390/molecules25214905
- Nam, Y., Shin, E. J., Shin, S. W., Lim, Y. K., Jung, J. H., Lee, J. H., et al. (2014). YY162 prevents ADHD-like behavioral side effects and cytotoxicity induced by Aroclor1254 via interactive signaling between antioxidant potential, BDNF/TrkB, DAT and NET. *Food Chem. Toxicol.* 65, 280–292. doi:10.1016/j.fct.2013.12.046
- Ni, J., Liu, Z., Jiang, M., Li, L., Deng, J., Wang, X., et al. (2022a). Ginsenoside Rg3 ameliorates myocardial glucose metabolism and insulin resistance via activating the AMPK signaling pathway. *J. Ginseng Res.* 46 (2), 235–247. doi:10.1016/j.jgr.2021.06.001
- Ni, J., Zhang, H., Wang, X., Liu, Z., Nie, T., Li, L., et al. (2022b). Rg3 regulates myocardial pyruvate metabolism via P300-mediated dihydrolipoamide dehydrogenase 2-hydroxyisobutyrylation in TAC-induced cardiac hypertrophy. *Cell Death Dis.* 13 (12), 1073. doi:10.1038/s41419-022-05516-y
- Opinion, A. G. R., Vanhomwegen, M., De Boeck, G., and Aerts, J. (2023). Long-term stress induced cortisol downregulation, growth reduction and cardiac remodeling in Atlantic salmon. *J. Exp. Biol.* 226 (22), jeb246504. doi:10.1242/jeb.246504
- Pahlavani, H. A. (2023). Exercise therapy to prevent and treat Alzheimer's disease. *Front. Aging Neurosci.* 15, 1243869. doi:10.3389/fnagi.2023.1243869
- Parrott, J. M., Redus, L., Santana-Coelho, D., Morales, J., Gao, X., and O'Connor, J. C. (2016). Neurotoxic kynurenine metabolism is increased in the dorsal hippocampus and drives distinct depressive behaviors during inflammation. *Transl. Psychiatry* 6 (10), e918. doi:10.1038/tp.2016.200
- Penninx, B. W., Pine, D. S., Holmes, E. A., and Reif, A. (2021). Anxiety disorders. *Lancet* 397 (10277), 914–927. doi:10.1016/s0140-6736(21)00359-7
- Piepenburg, S. M., Faller, H., Stork, S., Ertl, G., and Angermann, C. E. (2019). Symptom patterns and clinical outcomes in women versus men with systolic heart failure and depression. *Clin. Res. Cardiol.* 108 (3), 244–253. doi:10.1007/s00392-018-1348-6
- Pillinger, T., Osimo, E. F., de Marvao, A., Shah, M., Francis, C., Huang, J., et al. (2023). Effect of polygenic risk for schizophrenia on cardiac structure and function: a UK biobank observational study. *Lancet Psychiatry* 10 (2), 98–107. doi:10.1016/S2215-0366(22)00403-5
- Pound, K. M., Sorokina, N., Ballal, K., Berkich, D. A., Fasano, M., Lanoue, K. F., et al. (2009). Substrate-enzyme competition attenuates upregulated anaplerotic flux through malic enzyme in hypertrophied rat heart and restores triacylglyceride content: attenuating upregulated anaplerosis in hypertrophy. *Circ. Res.* 104 (6), 805–812. doi:10.1161/circresaha.108.189951
- Qu, P. R., Jiang, Z. L., Song, P. P., Liu, L. C., Xiang, M., and Wang, J. (2022). Saponins and their derivatives: potential candidates to alleviate anthracycline-induced cardiotoxicity and multidrug resistance. *Pharmacol. Res.* 182, 106352. doi:10.1016/j.phrs.2022.106352
- Rawat, P. S., Jaiswal, A., Khurana, A., Bhatti, J. S., and Navik, U. (2021). Doxorubicin-induced cardiotoxicity: an update on the molecular mechanism and novel therapeutic strategies for effective management. *Biomed. Pharmacother.* 139, 111708. doi:10.1016/j.biopha.2021.111708
- Reed, G. W., Rossi, J. E., and Cannon, C. P. (2017). Acute myocardial infarction. *Lancet* 389 (10065), 197–210. doi:10.1016/s0140-6736(16)30677-8
- Rossello, X., Menon, V., and Vranckx, P. (2022). Acetazolamide for patients with acute decompensated heart failure with volume overload. *Eur. Heart J. Acute Cardiovasc. Care* 11 (9), 712–713. doi:10.1093/ehjacc/zuac108
- Sabari, B. R., Zhang, D., Allis, C. D., and Zhao, Y. (2017). Metabolic regulation of gene expression through histone acylations. *Nat. Rev. Mol. Cell Biol.* 18 (2), 90–101. doi:10.1038/nrm.2016.140
- Sabatine, M. S., and Braunwald, E. (2021). Thrombolysis in myocardial infarction (TIMI) study group: JACC focus seminar 2/8. *J. Am. Coll. Cardiol.* 77 (22), 2822–2845. doi:10.1016/j.jacc.2021.01.060
- Sachs, B. D., Ni, J. R., and Caron, M. G. (2015). Brain 5-HT deficiency increases stress vulnerability and impairs antidepressant responses following psychosocial stress. *Proc. Natl. Acad. Sci. U. S. A.* 112 (8), 2557–2562. doi:10.1073/pnas.1416866112
- Sawicki, K. T., Sala, V., Prever, L., Hirsch, E., Ardehali, H., and Ghigo, A. (2021). Preventing and treating anthracycline cardiotoxicity: new insights. *Annu. Rev. Pharmacol. Toxicol.* 61, 309–332. doi:10.1146/annurev-pharmtox-030620-104842
- Schmidt, K. T., Makhijani, V. H., Boyt, K. M., Cogan, E. S., Pati, D., Pina, M. M., et al. (2019). Stress-induced alterations of norepinephrine release in the bed nucleus of the stria terminalis of mice. *ACS Chem. Neurosci.* 10 (4), 1908–1914. doi:10.1021/acscchemneuro.8b00265
- Shen, L., Chen, C., Wei, X., Li, X., Luo, G., Zhang, J., et al. (2015). Overexpression of ankyrin repeat domain 1 enhances cardiomyocyte apoptosis by promoting p53 activation and mitochondrial dysfunction in rodents. *Clin. Sci. (Lond)* 128 (10), 665–678. doi:10.1042/CS20140586
- Siddiqi, M. Z., Srinivasan, S., Park, H. Y., and Im, W. T. (2020). Exploration and characterization of novel glycoside hydrolases from the whole genome of lactobacillus ginsenosidimutans and enriched production of minor ginsenoside Rg3(S) by a recombinant enzymatic process. *Biomolecules* 10 (2), 288. doi:10.3390/biom10020288
- Siegel, R. K. (1979). Ginseng abuse syndrome. problems with the panacea. *Jama* 241 (15), 1614–1615. doi:10.1001/jama.241.15.1614
- Staretz-Chacham, O., Pode-Shakked, B., Kristal, E., Abraham, S. Y., Porper, K., Wormser, O., et al. (2021). The effects of a ketogenic diet on patients with dihydrolipoamide dehydrogenase deficiency. *Nutrients* 13 (10), 3523. doi:10.3390/nul13103523
- Sun, G. Z., Meng, F. J., Cai, H. Q., Diao, X. B., Zhang, B., and Bai, X. P. (2020). Ginsenoside Rg3 protects heart against isoproterenol-induced myocardial infarction by activating AMPK mediated autophagy. *Cardiovasc. Diagn. Ther.* 10 (2), 153–160. doi:10.21037/cdt.2020.01.02
- Sur, B., and Lee, B. (2022a). Ginsenoside Rg3 modulates spatial memory and fear memory extinction by the HPA axis and BDNF-TrkB pathway in a rat post-traumatic stress disorder. *J. Nat. Med.* 76 (4), 821–831. doi:10.1007/s11418-022-01636-z
- Sur, B., and Lee, B. (2022b). Myricetin inhibited fear and anxiety-like behaviors by HPA axis regulation and activation of the BDNF-ERK signaling pathway in posttraumatic stress disorder rats. *Evid. Based Complement. Altern. Med.* 2022, 8320256. doi:10.1155/2022/8320256
- Szatko, A., Glinicki, P., and Gietka-Czernel, M. (2023). Pheochromocytoma/ paraganglioma-associated cardiomyopathy. *Front. Endocrinol. (Lausanne)* 14, 1204851. doi:10.3389/fendo.2023.1204851
- Tai, P., Chen, X., Jia, G., Chen, G., Gong, L., Cheng, Y., et al. (2023). WGX50 mitigates doxorubicin-induced cardiotoxicity through inhibition of mitochondrial ROS and ferroptosis. *J. Transl. Med.* 21 (1), 823. doi:10.1186/s12967-023-04715-1
- Tang, B., Yuan, S., Xiong, Y., He, Q., and Larsson, S. C. (2020). Major depressive disorder and cardiometabolic diseases: a bidirectional mendelian randomisation study. *Diabetologia* 63 (7), 1305–1311. doi:10.1007/s00125-020-05131-6
- Teng, T., Shively, C. A., Li, X., Jiang, X., Neigh, G. N., Yin, B., et al. (2021). Chronic unpredictable mild stress produces depressive-like behavior, hypercortisolemia, and metabolic dysfunction in adolescent cynomolgus monkeys. *Transl. Psychiatry* 11 (1), 9. doi:10.1038/s41398-020-01132-6
- Truby, L. K., and Rogers, J. G. (2020). Advanced heart failure: epidemiology, diagnosis, and therapeutic approaches. *JACC Heart. Fail.* 8 (7), 523–536. doi:10.1016/j.jchf.2020.01.014
- Tsai, C. H., Pan, C. T., Chang, Y. Y., Peng, S. Y., Lee, P. C., Liao, C. W., et al. (2021). Aldosterone excess induced mitochondria decrease and dysfunction via mineralocorticoid receptor and oxidative stress *in vitro* and *in vivo*. *Biomedicines* 9 (8), 946. doi:10.3390/biomedicines9080946
- Tsai, C. K., Chen, B. H., Chen, H. H., Hsieh, R. J., Lee, J. C., Chu, Y. T., et al. (2022). Low-dose propranolol prevents functional decline in catecholamine-induced acute heart failure in rats. *Toxics* 10 (5), 238. doi:10.3390/toxics10050238
- Tsioufis, P., Theofilis, P., Tsioufis, K., and Tousoulis, D. (2022). The impact of cytokines in coronary atherosclerotic plaque: current therapeutic approaches. *Int. J. Mol. Sci.* 23 (24), 15937. doi:10.3390/ijms232415937
- Tu, C., Wan, B., and Zeng, Y. (2020). Ginsenoside Rg3 alleviates inflammation in a rat model of myocardial infarction via the SIRT1/NF- κ B pathway. *Exp. Ther. Med.* 20 (6), 238. doi:10.3892/etm.2020.9368
- Ugwu, P. I., Ben-Azu, B., Ugwu, S. U., Uruaka, C. I., Nworgu, C. C., Okorie, P. O., et al. (2022). Preventive putative mechanisms involved in the psychopathologies of mice passively coping with psychosocial defeat stress by quercetin. *Brain Res. Bull.* 183, 127–141. doi:10.1016/j.brainresbull.2022.03.004
- Upadhyay, S., Mantha, A. K., and Dhiman, M. (2020). Glycyrrhiza glabra (Licorice) root extract attenuates doxorubicin-induced cardiotoxicity via alleviating oxidative stress and stabilising the cardiac health in H9c2 cardiomyocytes. *J. Ethnopharmacol.* 258, 112690. doi:10.1016/j.jep.2020.112690
- Vaccarino, V., Sullivan, S., Hammadah, M., Wilmot, K., Al Mheid, I., Ramadan, R., et al. (2018). Mental stress-induced-myocardial ischemia in young patients with recent myocardial infarction: sex differences and mechanisms. *Circulation* 137 (8), 794–805. doi:10.1161/CIRCULATIONAHA.117.030849
- Walker, E. R., McGee, R. E., and Druss, B. G. (2015). Mortality in mental disorders and global disease burden implications: a systematic review and meta-analysis. *JAMA Psychiatry* 72 (4), 334–341. doi:10.1001/jamapsychiatry.2014.2502
- Wang, L., Lu, Z., Teng, Y., Pan, W., Li, Y., Su, S., et al. (2024). Cognitive impairment is associated with BDNF-TrkB signaling mediating synaptic damage and reduction of amino acid neurotransmitters in heart failure. *Faseb J.* 38 (1), e23351. doi:10.1096/fj.202301699RR
- Wang, X., Chen, L., Wang, T., Jiang, X., Zhang, H., Li, P., et al. (2015a). Ginsenoside Rg3 antagonizes adriamycin-induced cardiotoxicity by improving endothelial dysfunction from oxidative stress via upregulating the Nrf2-ARE pathway through the activation of akt. *Phytomedicine* 22 (10), 875–884. doi:10.1016/j.phymed.2015.06.010
- Wang, X., Ling, G., Wei, Y., Li, W., Zhang, Y., Tan, N., et al. (2023). Activation of ULK1 to trigger FUNDC1-mediated mitophagy in heart failure: effect of ginsenoside Rg3 intervention. *Phytomedicine* 120, 155042. doi:10.1016/j.phymed.2023.155042
- Wang, X., Meng, H., Wang, Q., Shao, M., Lu, W., Chen, X., et al. (2020). Baoyuan decoction ameliorates apoptosis via AT1-CARP signaling pathway in H9C2 cells and heart failure post-acute myocardial infarction rats. *J. Ethnopharmacol.* 252, 112536. doi:10.1016/j.jep.2019.112536

- Wang, X. D., and Kang, S. (2021). Ferroptosis in myocardial infarction: not a marker but a maker. *Open Biol.* 11 (4), 200367. doi:10.1098/rsob.200367
- Wang, Y., Hu, Z., Sun, B., Xu, J., Jiang, J., and Luo, M. (2015b). Ginsenoside Rg3 attenuates myocardial ischemia/reperfusion injury via Akt/endothelial nitric oxide synthase signaling and the B-cell lymphoma/B-cell lymphoma-associated X protein pathway. *Mol. Med. Rep.* 11 (6), 4518–4524. doi:10.3892/mmr.2015.3336
- Wu, J., Hall, M., Dondo, T. B., Wilkinson, C., Ludman, P., DeBelder, M., et al. (2019). Association between time of hospitalization with acute myocardial infarction and in-hospital mortality. *Eur. Heart. J.* 40 (15), 1214–1221. doi:10.1093/eurheartj/ehy835
- Xie, C., Zhan, Y., Wu, Y., Zhang, Z., Xiang, Y., Wang, L., et al. (2023). Expression and clinical significance of serum sST2, BDNF, CTnI, and BUN/Cr in patients with heart failure. *Altern. Ther. Health Med.* 29 (1), 176–181.
- Xiong, J., Yuan, H., Fei, S., Yang, S., You, M., and Liu, L. (2023). The preventive role of the red ginseng ginsenoside Rg3 in the treatment of lung tumorigenesis induced by benzo(a)pyrene. *Sci. Rep.* 13 (1), 4528. doi:10.1038/s41598-023-31710-9
- Xu, H., Miao, H., Chen, G., Zhang, G., Hua, Y., Wu, Y., et al. (2023). 20(S)-ginsenoside Rg3 exerts anti-fibrotic effect after myocardial infarction by alleviation of fibroblasts proliferation and collagen deposition through TGFBR1 signaling pathways. *J. Ginseng Res.* 47 (6), 743–754. doi:10.1016/j.jgr.2023.06.007
- Xu, Y., Peng, W., Han, D., Wang, Z., Gu, C., Feng, F., et al. (2020). Combined treatment of non-small-cell lung cancer using shenqi capsule and platinum-based chemotherapy: a meta-analysis and systematic review. *Evid. Based Complement. Altern. Med.* 2020, 3957193. doi:10.1155/2020/3957193
- Yan, H., Jin, H., Fu, Y., Yin, Z., and Yin, C. (2019). Production of rare ginsenosides Rg3 and Rh2 by endophytic bacteria from *Panax ginseng*. *J. Agric. Food Chem.* 67 (31), 8493–8499. doi:10.1021/acs.jafc.9b03159
- You, Z., Yao, Q., Shen, J., Gu, Z., Xu, H., Wu, Z., et al. (2017). Antidepressant-like effects of ginsenoside Rg3 in mice via activation of the hippocampal BDNF signaling cascade. *J. Nat. Med.* 71 (2), 367–379. doi:10.1007/s11418-016-1066-1
- Zarneshan, S. N., Fakhri, S., and Khan, H. (2022). Targeting Akt/CREB/BDNF signaling pathway by ginsenosides in neurodegenerative diseases: a mechanistic approach. *Pharmacol. Res.* 177, 106099. doi:10.1016/j.phrs.2022.106099
- Zhang, C., Yu, H., Ye, J., Tong, H., Wang, M., and Sun, G. (2023). Ginsenoside Rg3 protects against diabetic cardiomyopathy and promotes adiponectin signaling via activation of PPAR- γ . *Int. J. Mol. Sci.* 24 (23), 16736. doi:10.3390/ijms242316736
- Zhang, H., Zhou, Z., Chen, Z., Zhong, Z., and Li, Z. (2017). Ginsenoside Rg3 exerts anti-depressive effect on an NMDA-treated cell model and a chronic mild stress animal model. *J. Pharmacol. Sci.* 134 (1), 45–54. doi:10.1016/j.jphs.2017.03.007
- Zhang, J. C., Wu, J., Fujita, Y., Yao, W., Ren, Q., Yang, C., et al. (2014). Antidepressant effects of TrkB ligands on depression-like behavior and dendritic changes in mice after inflammation. *Int. J. Neuropsychopharmacol.* 18 (4), pyu077. doi:10.1093/ijnp/pyu077
- Zhang, L. P., Jiang, Y. C., Yu, X. F., Xu, H. L., Li, M., Zhao, X. Z., et al. (2016). Ginsenoside Rg3 improves cardiac function after myocardial ischemia/reperfusion via attenuating apoptosis and inflammation. *Evid. Based Complement. Altern. Med.* 2016, 6967853. doi:10.1155/2016/6967853
- Zhang, Z., Dalan, R., Hu, Z., Wang, J. W., Chew, N. W., Poh, K. K., et al. (2022). Reactive oxygen species scavenging nanomedicine for the treatment of ischemic heart disease. *Adv. Mat.* 34 (35), e2202169. doi:10.1002/adma.202202169
- Zhou, X., and Xia, X. (2023). Ginsenoside Rg3 improves microcystin-induced cardiotoxicity through the miR-128-3p/MDM4 axis. *Drug Chem. Toxicol.* 2023, 1–11. doi:10.1080/01480545.2023.2251716

Glossary

| | | | |
|----------------|---|-------------------------|---|
| Rg3 | Ginsenoside Rg3 | BDNF | Brain-derived neurotrophic factor |
| CAD | Coronary artery disease | CREB | Cyclic adenosine monophosphate response element binding protein |
| MDD | Major depressive disorder | I_{GABA} | GABA-induced inward current |
| MI | Myocardial infarction | PTSD | Post-traumatic stress disorder |
| HF | Heart failure | ADHD | Attention Deficit Hyperactivity Disorder |
| MIRI | Myocardial ischemia-reperfusion injury | HPA | Hypothalamic-pituitary-adrenal |
| SERCA2a | Sarcoplasmic/endoplasmic reticulum Ca ²⁺ -ATPase | TrkB | Tropomyosin-related kinase B |
| ER | Endoplasmic reticulum | SAM | Sympathetic-Adrenomedullary System |
| ROS | Reaction oxygen species | MDM4 | Double minute 4 protein |
| PDHc | Pyruvate dehydrogenase complex | CK | Creatine kinase |
| DLD | Dihydrolipoamide dehydrogenase | CK-MB | Creatine kinase-MB |
| ACY1 | Aminoacylase-1 | LDH | Lactate dehydrogenase |
| AngII | Angiotensin II | PPAR-γ | Peroxisome proliferator-activated receptor γ |
| TGF-β1 | Transforming growth factor-β1 | TGFBR1 | Transforming Growth Factor Beta Receptor I |
| AT1 | Angiotensin type 1 receptor | DCM | Diabetic cardiomyopathy |
| CARP | Cardiac ankyrin repeat protein | SIRT1 | Sirtuin 1 |
| ULK1 | Unc51-like-kinase 1 | GABA | Gamma aminobutyric acid |
| FUNDC1 | FUN14 domain-containing protein 1 | ISO | Isoproterenol |
| PCI | Percutaneous coronary intervention | TIP60 | Tat-interacting protein 60 |
| AMPK | AMP-activated protein kinase | Akt | Protein kinase B |
| ACC | Acetyl CoA carboxylase | | |
| TNF-α | Tumor necrosis factor-α | | |
| IL-1β | Interleukin 1 beta | | |
| NF-κB | Nuclear factor κB | | |
| eNOS | Endothelial nitric oxide synthase | | |
| Bcl-2 | B cell lymphoma-2 | | |
| Bax | Bcl2-associated X protein | | |
| H/R | Hypoxia/reoxygenation | | |
| PGC1-α | Peroxisome proliferators-activated receptor γ coactivator 1 alpha | | |
| IκBa | Inhibitor of kappa B alpha | | |
| DOX | Doxorubicin | | |
| SOD | Superoxide dismutase | | |
| Nrf | Nuclear factor erythroid 2-related factor | | |
| ARE | Antioxidant response element | | |
| PI3K | Phosphoinositide 3 kinase | | |
| PEG | Polyethylene glycol | | |
| PPS | Polypropylene sulfide | | |
| IRS | Insulin receptor substrate | | |
| TRP | Tryptophan | | |
| KYN | Kynurenine | | |



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Non-coding RNAs: targets for Chinese herbal medicine in treating myocardial fibrosis

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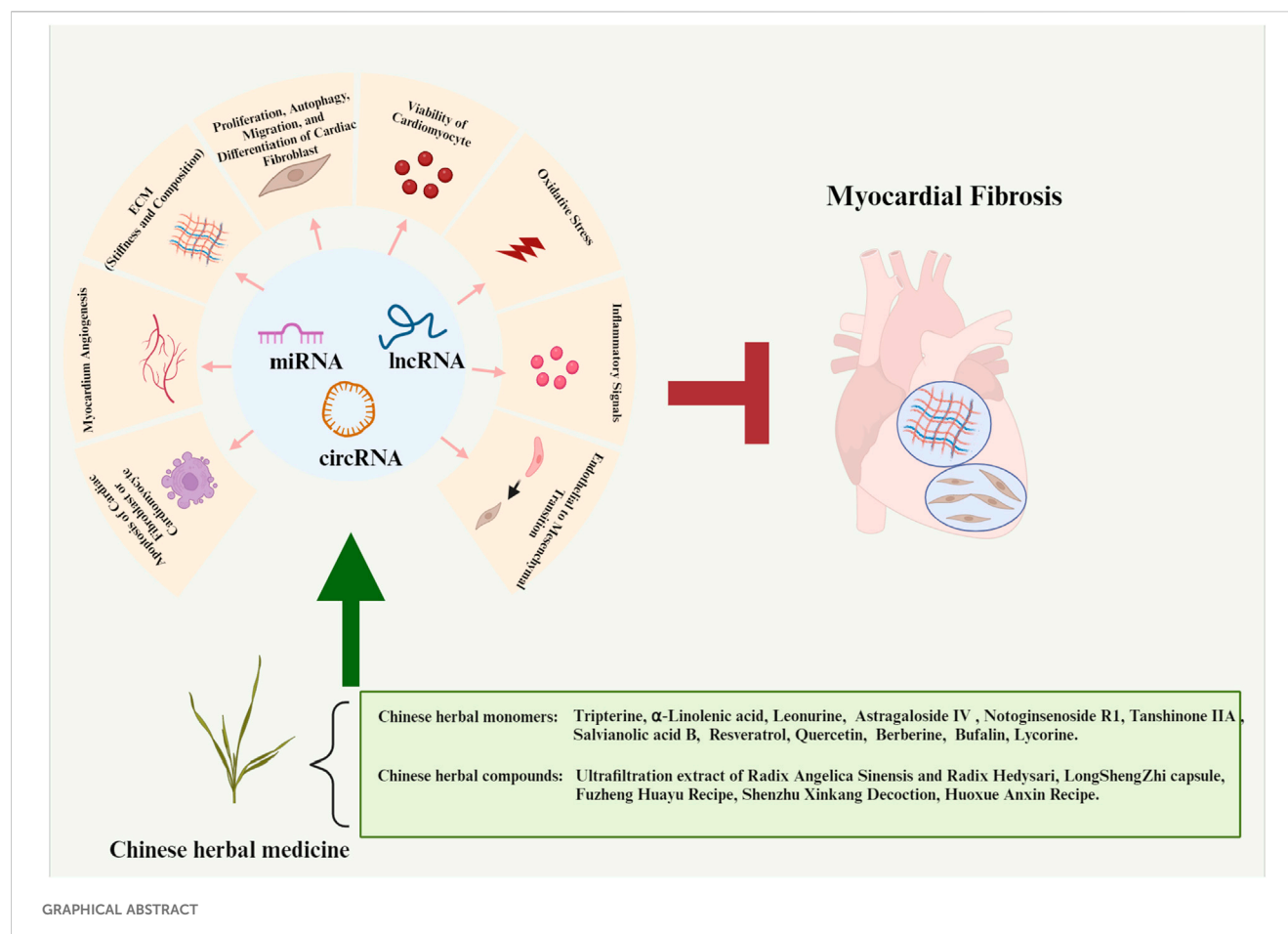
Cardiovascular diseases have become the leading cause of death in urban and rural areas. Myocardial fibrosis is a common pathological manifestation at the adaptive and repair stage of cardiovascular diseases, easily predisposing to cardiac death. Non-coding RNAs (ncRNAs), RNA molecules with no coding potential, can regulate gene expression in the occurrence and development of myocardial fibrosis. Recent studies have suggested that Chinese herbal medicine can relieve myocardial fibrosis through targeting various ncRNAs, mainly including microRNAs (miRNAs), long non-coding RNAs (lncRNAs), and circular RNAs (circRNAs). Thus, ncRNAs are novel drug targets for Chinese herbal medicine. Herein, we summarized the current understanding of ncRNAs in the pathogenesis of myocardial fibrosis, and highlighted the contribution of ncRNAs to the therapeutic effect of Chinese herbal medicine on myocardial fibrosis. Further, we discussed the future directions regarding the potential applications of ncRNA-based drug screening platform to screen drugs for myocardial fibrosis.

KEYWORDS

myocardial fibrosis, Chinese herbal medicine, ncRNAs, miRNAs, lncRNAs, circRNAs

1 Introduction

The main pathological features of myocardial fibrosis (MF) are excessive deposition of extracellular matrix in myocardial interstitium (Espeland et al., 2018). MF is a pathological manifestation of many cardiovascular diseases, like myocardial infarction (Talman and Ruskoaho, 2016), myocarditis (Tyminska et al., 2021), coronary heart disease (He et al., 2020) and hypertension (Rai et al., 2019). For example, following myocardial infarction, a large number of cardiac fibroblasts (CFs) are activated and transdifferentiate into myofibroblasts, which has stronger contraction function and extracellular matrix synthesis ability. Then increased collagen can deposit and form scar tissue in the infarcted area to provide support for the heart (González et al., 2018). Besides myocardial infarction, pressure overload due to hypertension or aortic stenosis (Verjans et al., 2018), volume overload induced by mitral valve (Dit Beaufls et al., 2021) or aortic valve insufficiency (Fragasso et al., 2022), and the release of inflammatory factors caused by oxidative stress (Lu et al., 2022) can also result in MF. MF is the end-stage pathological manifestation of most cardiovascular diseases and is closely related to cardiac death. It affects cardiac function in various ways. On one hand, perivascular fibrosis can impact



coronary blood supply, leading to myocardial ischemia, hypoxia, and even necrosis (Ytrehus et al., 2018). On the other hand, the deposition of a large number of collagen fibers results in an increase in ventricular wall hardness and a decrease in compliance, leading to reduced myocardial contraction, synchronization, and overall cardiac function (Sutton and Sharpe, 2000; Prabhu, 2005). Additionally, the deposition of collagen fibers in the matrix affects the conduction of myocardial electrical signals, easily forming reentrant rings and conduction blocks, which can result in arrhythmia (Francis Stuart et al., 2016). Therefore, studying the therapeutic strategies of MF is helpful to slow down the progression of cardiovascular diseases and reduce the mortality rate of patients.

At present, the treatment of MF mainly involves two approaches: medication and surgery. Medication methods are more commonly used at the early stage of cardiovascular diseases to prevent the development of MF. These drugs mainly include angiotensin-converting enzyme inhibitors (Francis Stuart et al., 2016), diuretics (Liu and Yu, 2022), β -blockers (Kobayashi et al., 2004), and anticoagulants (Oh et al., 2022). Surgical methods are used at the late stage of cardiovascular diseases, mainly including endocardial resection, atrioventricular valve repair/replacement, and heart transplantation. Endocardial resection can achieve the purpose of treatment by removing endocardial fibrous hyperplasia and calcification (Eckardt et al., 2000). Atrioventricular valve repair/replacement can repair/replace the fibrotic valve and restore its normal function (Mbanze et al., 2020). The heart transplantation

can be performed when other treatments for heart problems have not worked, leading to heart failure (Srivastava and Kittleson, 2024). The aforementioned Western medicine methods offer the advantages of quick and potent effects, but their associated risks and side effects cannot be ignored. For instance, some medications may cause varying degrees of side effects, such as nausea, headaches, and liver function impairment; different patients may exhibit varied responses to the same treatment methods, leading to inconsistent treatment outcomes; some treatment methods may be relatively expensive, posing an economic burden on patients.

Chinese herbal medicine is an important historical treasure. Unlike Western medicine, Chinese herbal medicine offers the advantages of having minimal side effects in clinical applications. Prescriptions are tailored to individual patients, considering factors such as time and condition. From the perspectives of “blood stasis” and “phlegm turbidity”, Chinese herbal medicine is utilized in the treatment of MF, often in combination with compounds that promote blood circulation, remove blood stasis, and address phlegm and turbidity. Clinically, widely used formulations include Qishen Yiqi pill (Lv et al., 2021), Tongxinluo Capsule (Yin et al., 2019), Baoxin Decoction (Sun et al., 2017), Ginseng Dingzhi Decoction (Wanget al., 2022a), Huoxue Anxin Recipe (Wang et al., 2016), and others.

Non-coding RNAs (ncRNAs) are identified as RNA molecules with no coding potential (Yan and Bu, 2021). With the development of RNA sequencing technology, a large number of ncRNAs have

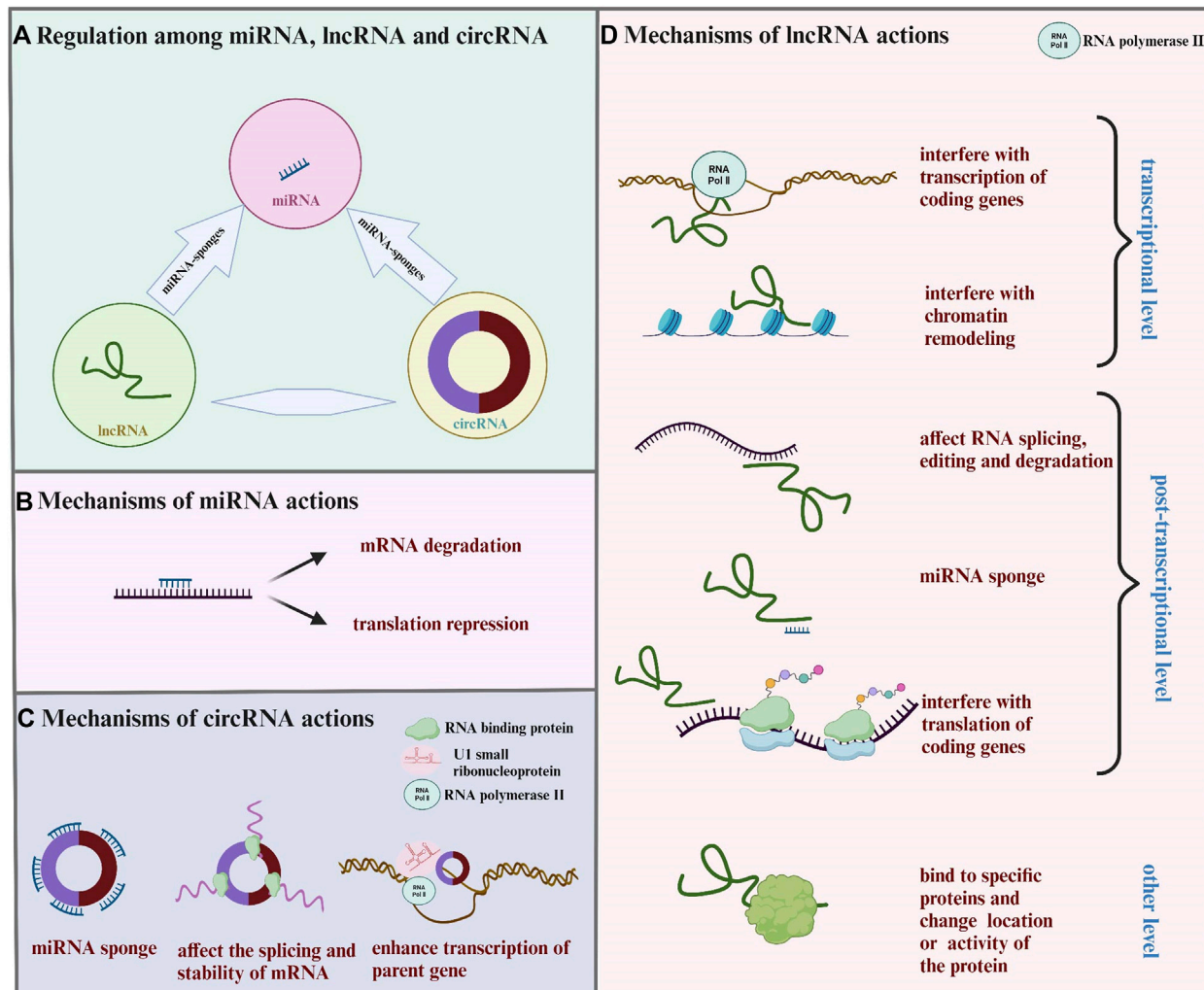


FIGURE 1

Mechanisms of controlling gene expression through ncRNAs. (A) lncRNA and circRNA act as miRNA sponges and affect the expression of miRNA and its target gene. (B) MiRNA binds to mRNA, leading to mRNA degradation or translation inhibition. (C) CirRNA combines with miRNA, RNA binding protein, U1 small ribonucleoprotein particles, and RNA polymerase II to affect the function of miRNA, interfere with the splicing or stability of mRNA, and enhance the transcription of parent genes. (D) lncRNA regulates gene expression from the transcriptional level, post-transcriptional level, and other level. Transcriptional level: lncRNA interferes with the transcription or chromatin remodeling of coding genes. Post-transcriptional level: lncRNA acts as miRNA sponges; lncRNA forms RNA double strand with mRNA, affecting mRNA splicing, editing, degradation, and translation. Other level: lncRNA binds to specific proteins, changing the location or activity of the protein.

been identified in different species and tissues. For a long time, it is generally believed that most genetic information is processed by protein-coding genes, while ncRNAs were regarded as junk nucleic acid sequences (Palazzo and Lee, 2015). In recent years, systematic analysis of cardiovascular genome and transcriptome has profoundly changed people's understanding of ncRNAs. Numerous studies have confirmed that ncRNAs are important regulatory factors in heart development and have an inseparable relationship with the occurrence and development of most cardiovascular diseases (Qu et al., 2016). By forming complexes with RNA, DNA, or proteins, ncRNAs modulate numerous pivotal signaling pathways implicated in MF (Chen et al., 2018b; Dilmaghani et al., 2021). Moreover, the expression changes of ncRNAs in plasma and tissues can be regarded as biomarkers for early warning and predicting prognosis of MF (Jiang et al., 2020;

Ghafouri-Fard et al., 2021). As such, they represent promising future clinical targets for modulating both MF and its associated cardiovascular conditions. In recent years, studies have shown that ncRNAs are the targets of Chinese herbal medicine in the treatment of MF. In this study, we have discussed recent progress in the modulation of ncRNAs through Chinese herbal medicine for managing MF, with a focus on herbal monomers and compounds.

2 Mechanisms of ncRNAs in regulating gene expression

In the past few decades, ncRNAs have been proved to regulate gene expression at multiple levels in the occurrence and development of MF, and act as new drug targets for MF

treatment. These ncRNAs mainly include microRNA (miRNA), long non-coding RNA (lncRNA), and circular RNA (circRNA), of which miRNA-mediated regulation have mostly been studied and documented (Dong et al., 2019b). Interestingly, there is also an interaction between the three ncRNAs, called competitive endogenous RNA (ceRNA) mechanism, which complicates gene regulation. The mechanisms of ncRNAs in regulating gene expression and their inter-regulations are shown in Figure 1.

2.1 MiRNA in regulating gene expression

MiRNA is a highly conserved gene family with a length of about 22–25 nucleotides (Davoodvandi et al., 2021). MiRNA binds to the untranslated region at the 3' end of mRNA to inhibit mRNA transcription or translation (Mohr and Mott, 2015). Studies have shown that pri-miRNA, the primary transcription product of miRNA, is cut into hairpin precursor miRNA (pre-miRNA) by Drosha enzyme of ribonuclease 3 family. After preliminary cutting, pre-miRNA is transported from the nucleus to the cytoplasm by transporter. Then, the pre-miRNA is further cut by the combined action of Dicer enzyme of ribonuclease 3 family and dsRNA binding protein (dsRBP). Finally, the mature miRNA binds to argonaute (AGO) protein to form RNA-inducing silencing complex (RISC), leading to the interaction between the complex and the target mRNA (Leitão and Enguita, 2022). There are two ways in which miRNA regulates gene expression: when miRNA is completely complementary to mRNA, the mRNA can be directly cut; when miRNA binds with mRNA incompletely, the translation of mRNA is prevented but the stability of mRNA is not affected (Fabian et al., 2010).

Through regulating fibrosis-related factors, miRNA plays various roles in MF. For example, mir-338-3p acts as a therapeutic target in MF through fibroblast growth factor receptor 2 (FGFR2) suppression (Huang et al., 2022a). MiR-125b is critical for induction of MF by targeting p53 and Apelin mRNA (Nagpal et al., 2016). The matrix metalloproteinases (MMPs) and tissue inhibitors of MMPs (TIMPs) play crucial roles as regulators of extracellular matrix turnover and tissue remodeling, significantly influencing MF (Serraino et al., 2023). A previous study has shown that miR-146b-5p can bind to TIMP4 mRNA, regulating the balance between TIMP4 and MMP9, which is associated with atrial fibrosis (Ye et al., 2021). Additionally, MMP2 and MMP9 have been identified as potential targets for miR-29a and miR-133a (Jones et al., 2011). Connective tissue growth factor (CTGF) exerts chemotactic and mitogenic effects on fibroblasts, closely related to the occurrence and development of fibrosis in various tissues and organs (Ramazani et al., 2018). MiR-30a attenuates MF in rats by targeting CTGF (Chen et al., 2018a). Furthermore, Galectin-3 promotes the proliferation and collagen synthesis of CFs, while miR-335 inhibits MF by directly targeting this gene Sun.

2.2 LncRNA in regulating gene expression

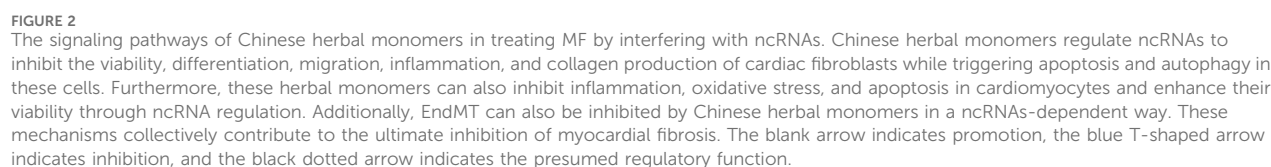
Compared with miRNA, the length of lncRNA is usually longer, generally more than 200 nt, which has the similar structure with

mRNA (Jha et al., 2023). Firstly, ceRNA mechanism is one of the important ways for lncRNA to regulate genes (Huang, 2018). As a natural miRNA sponge, lncRNA competes with mRNA to bind with miRNA, which affects gene silencing induced by miRNA. For example, lncRNA PFL acts as a ceRNA of let-7d to promote fibrogenesis (Liang et al., 2018). Besides ceRNA mechanism, lncRNA can also directly bind to DNA, mRNA and protein. This modulation can be categorized into three levels: transcription level, post-transcription level and other level (Li et al., 2023b). Transcriptional level means lncRNA can interfere with the transcription or chromatin remodeling of coding genes (Dykes and Emanueli, 2017). For example, lncTCF7 recruit switch/sucrose non-fermentable (SWI/SNF) complex to transcription factor 7 (TCF7) promoter region, leading to transcription of TCF7 gene (Wang et al., 2015b). Post-transcriptional level means lncRNA can influence mRNA splicing or translation (Ma et al., 2013). For example, lncRNA Safe can complementarily combine with secreted frizzled-related protein 2 (Sfrp2) mRNA to form a Safe-Sfrp2 RNA duplex to stabilize each other (Hao et al., 2019). Moreover, lncRNA can also bind to specific proteins, changing the location of the protein or regulating its activity (Guttman and Rinn, 2012). For example, lncRNA HOX transcript antisense RNA (HOTAIR) could bind with polypyrimidine tract-binding protein 1 (PTBP1) to increase the stability of Wnt5a (Tan et al., 2022).

The mechanism of regulating MF by lncRNA is complicated, with numerous lncRNAs involved in the ceRNA network to exert their functions. For example, lncRNA DANCER targets miR-758-3p to regulate proteoglycan 4 (PRG4) and the downstream Smad pathway, influencing the progression of cardiac dysfunction and fibrosis (Huang and Huang, 2023). lncRNA CFAR promotes MF via targeting miR-449a-5p to regulate the lysyl oxidase-like protein-3 (LOXL3)/mammalian target of rapamycin (mTOR) axis (Zhang et al., 2023). lncRNA TUG1 exacerbates MF in diabetic cardiomyopathy by modulating the miR-145a-5p/cofilin-2 (Cfl2) axis (Wang et al., 2023). Unlike the above mechanism, some lncRNAs regulate MF by directly decoying proteins. For example, the regulatory role of lncRNA Wisper in CFs proliferation, migration, and survival depends on its association with TIA1-related protein (Micheletti et al., 2017). lncRNA MetBil directly binds to methyltransferase like 3 (METTL3) protein to regulate its expression in ubiquitin-proteasome pathway, thereby regulating the expression of the methylated fibrosis-associated genes in ischemia-induced MF (Zhuang et al., 2023).

2.3 CircRNA in regulating gene expression

CircRNA has a closed loop structure without 5' cap and 3' polyA tail, and the average length of circRNA in human body is about 500 nt (Dragomir and Calin, 2018). CircRNA contains abundant miRNA binding sites and can be used as a sponge of miRNA (Salmena et al., 2011). For example, circRNA-005647 has a binding site with miR-27b-3p and inhibits the binding of miR-27b-3p with fibrosis-related genes (Yuan et al., 2019). In addition to ceRNA mechanism, circRNA can affect the splicing and stability of mRNA by binding to RNA binding protein (RBP). For example, circFndc3b enhances the expression and signal transduction of vascular endothelial growth factor-A (VEGF-A) by interacting

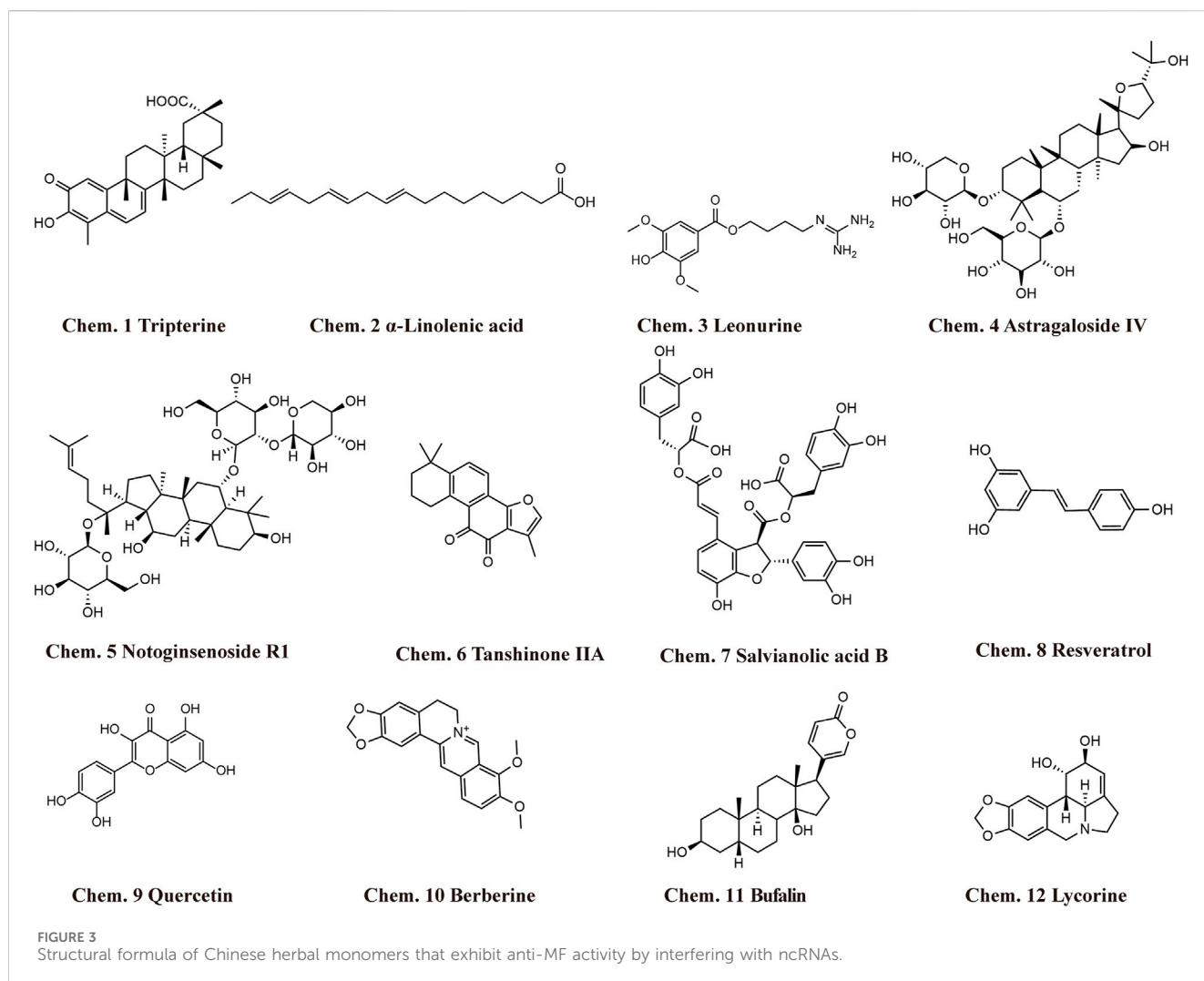


The mechanisms currently reported for circRNA in regulating MF mainly involve acting as a ceRNA, interacting with proteins, and encoding proteins. For example, circSMAD4 promotes MF by acting as a sponge against miR-671-5p (Jeong et al., 2023). CircYap directly binds to tropomyosin-4 (TMP4) and gamma-actin (ACTG) to make the interaction between the two proteins more stable, resulting in the inhibition of actin polymerization and subsequent MF (Wu et al., 2021). Circ_0036176 has the ability to encode a protein containing 208 amino acids named Myo9a-208, which mediates the inhibitory effect of circ_0036176 on the proliferation of CFs (Guo et al., 2022).

Chinese herbal medicine has the characteristics of wide sources and small side effects, thus having a unique advantage in treating human diseases including MF. Recent studies have suggested that Chinese herbal medicine can exert anti-MF effects by regulating ncRNAs. 12 kinds of Chinese herbal monomers and 5 kinds of Chinese herbal compounds have been shown to treat MF by interfering with ncRNAs. The herbal monomers include tripterine, α -linolenic acid, leonurine, astragaloside IV,

3.1 Chinese herbal monomers and their targeting ncRNAs in treating MF

Chinese herb *Tripterygium wilfordii* Hook. F. has the effects of dispelling dampness, relieving swelling and pain, and resisting inflammation (Zeng et al., 2024). Triptolide and tripterine are the two most active components in the extract of *Tripterygium wilfordii* Hook. F. In recent years, it has been found that tripterine can inhibit CFs viability and collagen production by down-regulating the expression of miR-21, the activator of ERK signaling pathway, leading to relieved MF and cardiac dysfunction (Cheng et al., 2016). In fact, miR-21 has been found to be a pro-fibrotic factor in various animal models, targeting multiple fibrotic pathways and promoting MF. For example, in a mouse model of myocardial



infarction, miR-21 promotes CF activation and MF via TGF- β /Smad7 signaling pathway (Yuan et al., 2017). In another diabetes-induced MF mouse model, silencing miR-21 inhibits high glucose-induced endothelial to mesenchymal transition (EndMT) (Li et al., 2020). These results suggest that the anti-fibrotic effect of tripterine is mediated by miR-21 and the downstream ERK, TGF- β , and EndMT signaling pathways.

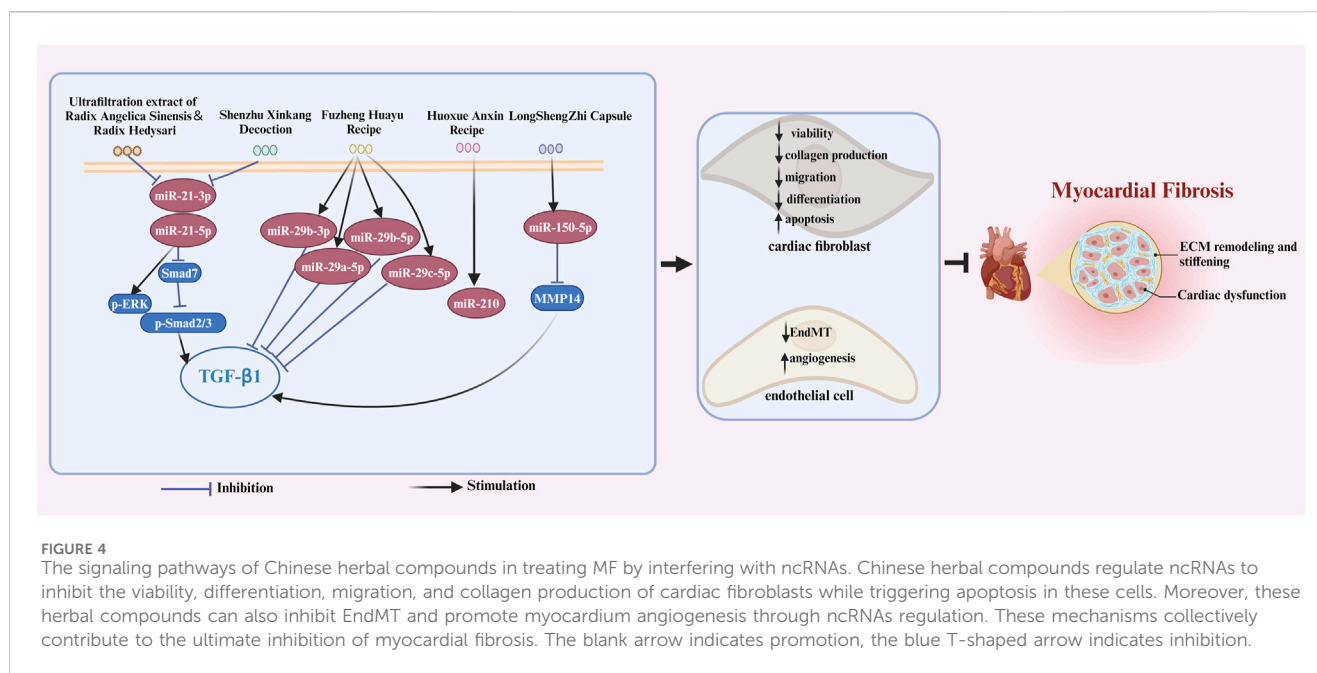
3.1.2 α -Linolenic acid (ALA)

Flaxseed oil is extracted from *Linum usitatissimum* L., and is rich in omega-3 fatty acids such as ALA. It is usually used to lower cholesterol, resist atherosclerosis, and reduce heart load (Prasad et al., 2020). A study has shown that in a rat model of myocardial infarction, flaxseed oil exerts cardioprotective effect and decreases collagen deposition via selectively up-regulating the expression of miR-133a, miR-135a, and miR-29b. The author speculated this effect may be attributed to ALA component in the flaxseed oil (Parikh et al., 2020). Previous studies have shown that the three miRNAs are all anti-fibrotic miRNAs. MiR-133a reduces MF by suppressing transforming growth factor- β 1 (TGF- β 1) signaling in an acute myocardial infarction model (Yu et al., 2019). MiR-135a could target transient receptor potential melastatin 7 (TRPM7) to

inhibit the activation of TGF- β /Smads pathway, thus relieving MF (Wei et al., 2020). MiR-29b inhibits many genes involved in extracellular matrix formation and fibrosis, such as Col1a1, Col1a2, Col3a1, fibrillin 1, Elastin, and TGF- β 1 (Rooij et al., 2008; Zhang et al., 2014).

3.1.3 Leonurine

Leonurine, an alkaloid extracted from *leonurus japonicus* Hout., has been shown to have various pharmacological effects including protecting myocardial ischemia-reperfusion injury, resisting blood platelet aggregation, reducing blood viscosity, promoting angiogenesis, lowering blood pressure, and inducing diuresis (Huang et al., 2021). Recent studies have shown that leonurine can treat MF induced by isoproterenol by upregulating miR-1, which could directly target Fibulin-2 (Fbln2) to reverse cardiac remodeling (Karakikes et al., 2013; Lu et al., 2018). In addition, evidence has shown that all mature miR-29 family members in post-myocardial infarction tissues are downregulated (Wang et al., 2021). Leonurine treatment can significantly upregulate the expression of miR-29a-3p and downregulate its target proteins including TGF- β , Col3a1, and Col1a1, to attenuate fibrosis and cardiac remodeling (Wang et al.,



2021). What's more, a study confirmed that leonurine can also promote apoptosis of CFs through regulating miR-29a-3p (Xi et al., 2023). These evidences suggest that the anti-fibrotic effect of leonurine may be mediated by miR-1 and miR-29a-3p.

3.1.4 Astragaloside IV

Astragaloside IV is one of the best bioactive components from the root of *Astragalus membranaceus* (Fisch.) Bunge (Jing et al., 2021). Studies have shown that astragaloside IV inhibits MF by up-regulating the expression of miR-135a which targets transient receptor potential melastatin 7 (TRPM7) (Wei et al., 2020). Moreover, in an experiment of high glucose-induced injury of cardiomyocytes, it was found that astragaloside IV could reduce the expression of miR-34a (Zhu et al., 2019). According to previous studies, inhibition of miR-34a can treat MF by inhibiting cardiomyocytes apoptosis (Dong F. et al., 2019a). Therefore, it can be speculated that astragaloside IV may play an anti-fibrotic role through repressing cardiomyocytes apoptosis by decreasing miR-34a.

3.1.5 Notoginsenoside R1

Panax notoginseng saponins are the main component of the roots of *Panax notoginseng* (Burk.) F. H. Chen. Panax notoginseng saponins have the effects of promoting blood circulation, removing blood stasis, and dredging collaterals, which are commonly used to treat coronary heart disease and blood stasis syndrome (Jiao et al., 2021; Yang et al., 2022). Notoginsenoside R1 is the primary active component of Panax notoginseng saponins. It was found that notoginsenoside R1 inhibits isoproterenol-induced MF through the intervention of miRNA-mRNA regulatory network, among which the expression level of miR-21 decreased, while miR-29c, miR-30c and miR-133b increased (Ning, 2016). These miRNAs have all been proven to be fibrosis-related miRNAs. It has been shown that miR-21 activates ERK signaling pathway,

EndMT and TGF- β signaling pathway to promote MF (Cheng et al., 2016; Yuan et al., 2017; Li et al., 2020). MiR-29c has been proved to target multiple fibrosis-related genes including Col1a1, Col1a2, Col3a1 and Col5a1, fibrillin 1 and TGF- β 1 (Liu et al., 2017). MiR-30c inhibits the proliferation, differentiation, migration and collagen production of CFs by targeting TGF- β R2 (Xu et al., 2018); MiR-133b has also been reported to alleviate doxorubicin-induced cardiomyocyte apoptosis and MF by targeting PTBP1 and transgelin 2 (TAGLN2) (Li et al., 2021). Therefore, notoginsenoside R1 can target multiple fibrosis-related miRNA and can be regarded as attractive anti-fibrotic candidate medicine.

3.1.6 Tanshinone IIA

Tanshinone IIA is extracted from the dried root and rhizome of *Salvia miltiorrhiza* Bunge, which exerts a wide range of cardioprotective effects in the diseases like myocardial infarction, myocardial ischemia-reperfusion injury, myocardial hypertrophy, atherosclerosis, and cardiomyopathy (Zhang et al., 2019; Yang et al., 2020). Recent studies have shown that tanshinone IIA can relieve MF through up-regulating miR-29b (Yang et al., 2015), miR-205-3p (Qiao et al., 2021), and miR-618 (Yan et al., 2022) expressions. MiR-29b and miR-205-3p could downregulate the expression of TGF- β 1, Col1a1, and Col3a1, so as to resist MF following myocardial infarction (Yang et al., 2015; Qiao et al., 2021). MiR-618 could inhibit the expression of TIMP1 and TIMP4 to mediate the anti-fibrotic effects of tanshinone IIA on CFs (Yan et al., 2022). In addition, it has been found that tanshinone IIA can inhibit fibroblast proliferation by down-regulating lncRNA human-specific regulatory loci (HSRL) in skin hypertrophic scar tissue. HSRL could promote the expression of sorting nexin 9 (SNX9) and strengthen its interaction with p-Smad3, thus activating TGF- β signaling (Shi et al., 2020). Therefore, inhibiting HSRL may also mediate the anti-fibrotic effect of tanshinone IIA.

3.1.7 Salvianolic acid B

Salvianolic acid B is also derived from the root and rhizome of *Salvia miltiorrhiza* Bunge. A study showed that lncRNA maternally expressed gene 3 (Meg3), mainly expressed in CFs, is a new promotor of MF. Silencing Meg3 prevents MMP2 production, leading to the decreased MF and improved cardiac function (Piccoli et al., 2017). In cardiomyocytes of oxygen and glucose deprivation (OGD), salvianolic acid B was reported to represses Meg3 expression, which influences murine double minute 2 (MDM2)/p53 and AMP-activated Protein Kinase (AMPK) signalling pathways, leading to incresed viability and reduced apoptosis of cardiomyocytes (Yang et al., 2019a). Therefore, it is speculated that salvianolic acid B may have the cardioprotective effect of inhibiting MF by down-regulating Meg3, but more researches are needed to test this hypothesis.

3.1.8 Resveratrol

Resveratrol is an active polyphenol, derived from many herbal medicines, such as *Morus alba* L., *Polygonum cuspidatum* Sieb. et Zucc., and *Rubus idaeus* L. Resveratrol is proved to have anti-bacterial, anti-inflammatory and immunomodulatory effects (Malaguarnera, 2019). In cardiovascular system, it exerts protective effects on atherosclerosis, myocardial infarction, and heart failure (Raj et al., 2021). Recent studies showed that resveratrol can inhibit proliferation and induce cell death of CFs (Lieben Louis et al., 2019). A study assessed the impact of resveratrol on microRNAs linked to MF, and found resveratrol inhibits the expressions of miR-17, miR-34a and miR-181a in TGF- β 1-induced CFs (Zhang et al., 2018). Overexpression of miR-17 decreases Smad7 expression level, indirectly promotes TGF- β 1 signaling (Zhang et al., 2018). MiR-34a activates TGF- β 1 signaling through increasing Smad4 expression (Huang et al., 2014). MiR-181a suppresses the expression of PH domain leucine-rich repeat protein phosphatase 2 (PHLPP2) and subsequently activates AKT signaling, leading to enhanced proliferation of keloid fibroblast cells (Rang et al., 2016). In addition, resveratrol was reported to inhibit the expression of lncRNA metastasis associated lung adenocarcinoma transcript 1 (MALAT1), which could act as a ceRNA of miR-145 and promote MF progression (Yang et al., 2019b; Huang et al., 2019). Therefore, downregulation of MALAT1 may be one of the potential mechanisms of resveratrol in treating MF. It is evident that resveratrol, with its multiple ncRNA targets, holds significant promise for the management of MF.

3.1.9 Quercetin

Quercetin is a kind of flavonol that widely exists in flowers, leaves and fruits of many plants, such as *Sophora japonica* L., *Asparagus officinalis* L., and *Acanthopanax senticosus* (Rupr. et Maxim.) Harms. Previous studies have shown that quercetin restrains the level of fibrotic proteins including TGF- β 1, α -SMA, Col1a1, and Col3a1 in heart tissue of myocardial infarction model (Albadrani et al., 2021). Studies are progressively uncovering that the promotion of CFs autophagy can effectively inhibit MF and enhance cardiac function (Wang et al., 2015a). Quercetin was found to prevent isoprenaline-induced MF by increasing autophagy of CFs via decreasing miR-223-3p and increasing forkhead Box O3 (FOXO3) (Hu et al., 2021).

3.1.10 Berberine

Berberine is a quaternary ammonium alkaloid contained in the rhizome of *Coptis chinensis* Franch. and has various cardiovascular protective effects, such as anti-heart failure, anti-arrhythmia, and lowering cholesterol effects (Pagliaro et al., 2015; Zhao et al., 2020). It was found that berberine relieves hypertension-induced MF by increasing the expression of miR-29b and decreasing its targets Col1a1 and Col3a1 (Zheng et al., 2020). Moreover, in a myocardial ischemia-reperfusion mouse model, the protective effect of berberine is exerted by inducing miR-26b-5p and inhibiting its downstream PTGS2 and MAPK members, which results in the increased viability, and decreased apoptosis, inflammatory, and oxidative stress in cardiomyocytes (Jia et al., 2022). Since the injury of the cardiomyocytes is the major causes of MF, it is speculated that miR-26b-5p may be a potential target of berberine in the treatment of MF. In addition, berberine was also proved to inhibit lncRNA myocardial infarction-associated transcript (MIAT) to improve myocardial hypertrophy (Zeng et al., 2019). Since MIAT is a pro-fibrotic lncRNA governing MF by down-regulating miR-24 and up-regulating Furin and TGF- β 1 (Qu et al., 2017), we can infer that berberine may have the effect to improve MF by inhibiting MIAT.

3.1.11 Bufalin and lycorine

Bufalin comes from dried *toad*, while lycorine is an alkaloid found in the bulb of *Lycoris radiata* (L'Hér.) Herb. Recently, high-throughput natural compound library screening identified bufalin and lycorine to be effective anti-fibrotic molecules in hypertension-induced MF mouse model (Schimmel et al., 2020). The study found the level of miR-671-5p is reduced after treatment of CFs with bufalin and lycorine, which leads to the increased expression of anti-fibrotic protein selenoprotein P1 (SEPP1) (Schimmel et al., 2020). Another study discovered that bufalin and lycorine can reduce the expression of miR-29 while increasing the expression of circRNA CDR1as. This, in turn, leads to a decrease in the infarction area and fibrotic area in a heart failure pig model (Mester-Tonczar et al., 2020). Consequently, the anti-fibrotic impact of bufalin and lycorine can be ascribed to the reduction of miR-671-5p and miR-29 levels and the elevation of CDR1as levels.

3.2 Chinese herbal compounds and their targeting ncRNAs in treating MF

3.2.1 Ultrafiltration extract of radix angelica sinensis and radix hedysari

Radix Angelica Sinensis is usually used in combination with other drugs to treat cardiovascular diseases such as radiation-induced heart disease, atherosclerosis, and ischemic heart disease (Huang et al., 2022b; Li et al., 2023a; Yuan et al., 2023), while Radix Hedysari has been proved to have significant effects in treating non-alcoholic fatty liver disease (Sun et al., 2014). In a rat model of X-irradiation-induced MF, ultrafiltration extract derived from dried root of Radix Angelica Sinensis and Radix Hedysari downregulates miR-21-3p and miR-21-5p, inducing the apoptosis of CFs and alleviating MF (Ma et al., 2019). It has been shown that miR-21 activates ERK signaling pathway, EndMT, and TGF- β signaling pathway to promote MF (Cheng et al., 2016;

TABLE 1 Mechanisms of Chinese herbal medicine in treating MF by regulating ncRNAs.

| Herbal medicine | ncRNAs | Direct ncRNAs' targets | Biological function | References |
|--------------------|---------------|------------------------|--|--|
| Tripterine | miR-21↓ | Smad7 | ↓CFs viability, differentiation, migration, and collagen production | Cheng et al. (2016), Yuan et al. (2017), Li et al. (2020) |
| | | | ↓EndMT process | |
| α-Linolenic acid | miR-29b↑ | TGF-β1 | ↓CFs differentiation and collagen production | Roosij et al. (2008), Zhang et al. (2014), Parikh et al. (2020) |
| | | Col1a1 | | |
| | | Col1a2 | | |
| | | Col3a1 fibrillin 1 | | |
| | | Elastin | | |
| | miR-133a↑ | ? | ↓CFs differentiation and collagen production | Yu et al. (2019), Parikh et al. (2020) |
| | miR-135a↑ | TRPM7 | ↓CFs viability, differentiation, and collagen production | Parikh et al. (2020), Wei et al. (2020) |
| Leonurine | miR-1↑ | Fbln2 | ↓CFs collagen production | Karakikes et al. (2013), Lu et al. (2018) |
| | miR-29a-3p↑ | TGF-β1 | ↓CFs viability, differentiation, migration, and collagen production | Wang et al. (2021), Xi et al. (2023) |
| | | Col3a1 | ↑CFs apoptosis | |
| | | Col1a1 | | |
| Astragaloside IV | miR-135a↑ | TRPM7 | ↓ CFs viability, differentiation, and collagen production | Wei et al. (2020) |
| | *miR-34a↓ | Sirt1 | ↓CMs apoptosis | Dong et al. (2019a), Zhu et al. (2019) |
| Notoginsenoside R1 | miR-21↓ | Smad7 | ↓CFs viability, differentiation, migration, and collagen production | Cheng et al. (2016); Ning (2016), Yuan et al. (2017), Li et al. (2020) |
| | | | ↓EndMT process | |
| | miR-29c↑ | ? | ↓CFs differentiation and collagen production | Ning (2016); Liu et al. (2017) |
| | miR-30c↑ | TGF-β RII | ↓CFs viability, differentiation, migration, and collagen production | Ning (2016); Xu et al. (2018) |
| | miR-133b↑ | PTBP1 | ↓CFs collagen production | Ning (2016); Li et al. (2021) |
| | | TAGLN2 | ↓CMs apoptosis | |
| Tanshinone IIA | miR-29b↑ | ? | ↓CFs differentiation and collagen production | Yang et al. (2015) |
| | miR-205-3p↑ | TGF-β1 | ↓CFs collagen production | Qiao et al. (2021) |
| | miR-618↑ | TIMP1 | ↓CFs viability, differentiation, and collagen production | Yan et al. (2022) |
| | | TIMP4 | | |
| | *lncRNA HSRL↓ | SNX9 | ↓CFs viability, differentiation, and collagen production | Shi et al. (2020) |
| Salvianolic acid B | *lncRNA Meg3↓ | p53 | ↓CFs collagen production | Piccoli et al. (2017), Yang et al. (2019a) |
| | | | ↓CMs apoptosis | |
| | | | ↑CMs viability | |
| Resveratrol | miR-17↓ | Smad7 | ↓CFs viability and collagen production | Zhang et al. (2018) |
| | miR-34↓ | Smad4 | ↓ CFs viability, differentiation, migration, and collagen production | Huang et al. (2014), Zhang et al. (2018) |
| | miR-181a↓ | PHLPP2 | ↓CFs viability | Rang et al. (2016), Zhang et al. (2018) |

(Continued on following page)

TABLE 1 (Continued) Mechanisms of Chinese herbal medicine in treating MF by regulating ncRNAs.

| Herbal medicine | ncRNAs | Direct ncRNAs' targets | Biological function | References |
|---|-----------------|------------------------|---|---|
| | *lncRNA MALAT1↓ | miR-145 | ↓CFs viability, differentiation, and collagen production | Yang et al. (2019b), Huang et al. (2019) |
| Quercetin | miR-223-3p↓ | FOXO3 | ↑CFs autophagy | Hu et al. (2021) |
| | | | ↓CFs viability and collagen production | |
| Berberine | *miR-26b-5p↑ | ? | ↑CMs viability | Jia et al. (2022) |
| | | | ↓CMs apoptosis, inflammation, and oxidative stress | |
| | miR-29b↑ | ? | ↓CFs collagen production | Zheng et al. (2020) |
| | *lncRNA MIAT↓ | miR-24 | ↓CFs viability and collagen production | Qu et al. (2017), Zeng et al. (2019) |
| Bufalin and Lycorine | miR-29↓ | ? | ↓CFs collagen production | Mester-Tonczar et al. (2020) |
| | circRNA CRD1as↑ | ? | | |
| | miR-671-5p↓ | SEPP1 | ↓CFs differentiation, collagen production, and inflammation | Schimmel et al. (2020) |
| Ultrafiltration extract of Radix Angelica Sinensis and Radix Hedysari | miR-21-3p↓ | Smad7 | ↓CFs viability, differentiation, migration, and collagen production | Cheng et al. (2016), Yuan et al. (2017), Ma et al. (2019), Li et al. (2020) |
| | miR-21-5p↓ | | ↑CFs apoptosis | |
| | | | ↓EndMT process | |
| LongShengzhi capsule | miR-150-5p↑ | MMP14 | ↓CFs collagen production | Gu et al. (2022) |
| Fuzheng Huayu Recipe | miR-29b-3p↑ | ? | ↓CFs collagen production | Qi et al. (2019) |
| | miR-29a-5p↑ | | | |
| | miR-29b-5p↑ | | | |
| | miR-29c-5p↑ | | | |
| Shenzhu Xinkang Decoction | miR-21↓ | ? | ↓EndMT process | Zhai et al. (2022) |
| Huoxue Anxin Recipe | miR-210↑ | ? | ↑Myocardium angiogenesis | Wang et al. (2016) |

(Marked with * is the speculated mechanism).
Abbreviations: CMs, cardiomyocytes; CFs, cardiac fibroblasts; circRNA, circular RNA; EndMT, endothelial to mesenchymal transition; Fbln2, Fibulin-2; FOXO3, forkhead box O3; lncRNA, long noncoding RNA; MALAT1, metastasis associated lung adenocarcinoma transcript 1; Meg3, maternally expressed gene 3; MF, myocardial fibrosis; miRNA, microRNA; MIAT, myocardial infarction-associated transcript; MMP14, matrix metalloproteinase 14; PHLPP2, PH, domain leucine-rich repeat protein phosphatase 2; PTBP1, polypyrimidine tract-binding protein 1; SEPP1, selenoprotein P1; SNX9, sorting nexin 9; TAGLN2, transgelin 2; TIMP1, tissue inhibitors of matrix metalloproteinase 1; TIMP4, tissue inhibitors of matrix metalloproteinase 4; TRPM7, transient receptor potential melastatin 7.

Yuan et al., 2017; Li et al., 2020). Therefore, the Radix Angelica Sinensis and Radix Hedysari ultrafiltration extract may be developed as a medical countermeasure for the mitigation of radiation-induced MF.

3.2.2 LongShengZhi capsule

Buyang Huanwu Decoction is a famous herbal prescription that has been used to treat stroke for centuries (Gao et al., 2021). Previous studies have shown that Buyang Huanwu Decoction can alleviate MF (Wang et al., 2022b). The compatible components of LongShengZhi capsule are similar to those of Buyang Huanwu Decoction, and this is referred to as the modern application of Buyang Huanwu Decoction. A study found that LongShengZhi capsule attenuates Angiotensin II-induced cardiac hypertrophy and fibrosis in rats. Mechanically,

Longshengzhi capsule up-regulates miR-150-5p to target MMP14 in CFs, leading to reduced cardiac remodeling (Gu et al., 2022).

3.2.3 Fuzheng Huayu Recipe

Fuzheng Huayu Recipe, a traditional Chinese herbal prescription, is often used in China to treat fibrosis (Wang et al., 2010; Sun et al., 2022). Recently, a study has suggested that Fuzheng Huayu Capsule inhibits myocardial infarction-induced MF by facilitating the expression of miR-29b-3p, miR-29a-5p, miR-29b-5p, and miR-29c-5p (Qi et al., 2019). It has been shown that miR-29 family are all anti-fibrotic factors with the effect of inhibiting TGF-β1 signaling and its downstream targets, resulting in reduced proliferation and collagen production of CFs (Rooij et al., 2008; Zhu et al., 2013; Zhang

et al., 2014). Therefore, miR-29 family are the key mediators for the anti-fibrotic effect of Fuzheng Huayu Recipe.

3.2.4 Shenzhu Xinkang Decoction

Shenzhu Xinkang Decoction is a representative prescription for the treatment of chronic heart failure and fibrosis (Zhao et al., 2023). EndMT has been shown to contribute to cardiac fibrosis (Zeisberg et al., 2007) and Shenzhu Xinkang Decoction was proved to inhibit EndMT to play an anti-fibrotic role. The possible mechanism is related to the downregulation of miR-21 level and inhibition of PTEN/PI3K/AKT pathway (Zhai et al., 2022).

3.2.5 Huoxue Anxin Recipe

Huoxue Anxin Recipe is a novel formula of Chinese herbal medicine that has a good cardioprotective effect, such as promoting myocardial angiogenesis, exhibiting anti-oxidative stresses activity, and improving cardiac function during myocardial infarction (Zhang et al., 2012; Zhang et al., 2013; Wang et al., 2016). A recent study has demonstrated that Huoxue Anxin Recipe could reduce the infarction area, alleviate fibrosis, and improve the cardiac function of myocardial infarction rats, which is mainly attributed to enhanced angiogenesis by upregulation of miR-210 and VEGF (Wang et al., 2016).

4 Conclusion and perspective

The targeted relationship between Chinese herbal medicine and ncRNAs are hot spots in current research, which opens up a new avenue for exploring the mechanism of Chinese herbal medicine in prevention and treatment of cardiovascular diseases. This review discussed 12 kinds of Chinese herbal monomers and 5 kinds of Chinese herbal compounds which have been shown to treat MF by interfering with ncRNAs (Table 1). Through targeting ncRNAs, mainly including miRNA, lncRNA and circRNA, those herbal medicine relieves MF by inhibiting the proliferation/activation/inflammation of CFs, increasing apoptosis/autophagy of CFs, inhibiting apoptosis/inflammation/oxidative stress of cardiomyocytes, increasing viability of cardiomyocytes, repressing EndMT, and promoting myocardium angiogenesis.

At present, however, several challenges persist in the investigation of ncRNAs' role in the anti-MF effect of Chinese herbal medicine: (1) The regulatory effect of Chinese herbal medicine on ncRNAs is primarily validated in animal and cell models, and it is not yet guaranteed whether these effects still exist in complex human bodies. (2) The most reported ncRNA targets of Chinese herbal medicine are miRNAs. Whether lncRNA, circRNA, and other ncRNAs act as key mediators of Chinese herbal medicine's effect has not been explored sufficiently. (3) We noted that some Chinese herbal medicine could target ncRNAs known to be associated with MF. However, there is currently no direct evidence to support the idea that these herbal medicines can relieve MF by modulating these ncRNAs. Further experiments are required to validate these scientific hypotheses.

NcRNAs have been implicated in various diseases and serve as key targets for disease treatment. However, the clinical transformation of RNA-based therapies is hindered by problems related to specificity, delivery and tolerance. Specificity problems indicate undesirable targeting effects caused by uptake of cells other than the target cells, and off-target effects caused by sequence similarity or overdose to a level much higher than endogenous expectations (Sledz and Williams, 2004; Yu et al., 2020). In addition, there is a lack of delivery vectors suitable for delivering ncRNAs to target organs and cell types (Krieg, 2011). What's more, natural RNA molecules are highly susceptible to enzymatic degradation by serum and cellular RNases. Notably, both single-stranded and double-stranded RNAs can trigger the body's viral defense system via pathogen-associated molecular pattern (PAMP) receptors (Kumar et al., 2011). Due to these reasons, RNA-based therapies are often lack of efficiency in clinical trials. Therefore, targeting specific ncRNAs with small molecules displays potential as a therapeutic approach for disease treatment. Developing effective tools to screen small molecules against particular ncRNAs is very important and urgent. Recently, a study published in Nature has introduced an innovative technique for screening small molecules that bind to ncRNAs. They devised an unbiased screen based on affinity-selection mass spectrometry to identify reversibly binding ability between lncRNA Xist and 50,000 compounds. They found 20 analogues that has similar structure with the one positive hit and finally obtained one positive compound X1 that can effectively regulate the function of Xist (Aguilar et al., 2022). This research broadens the scope of ncRNA pharmaceutical field, which will enable the development of RNA-targeting drugs by high-throughput and large-scale screening methods. Therefore, by utilizing this screening system with MF-related ncRNAs as binding targets, it holds the promise of identifying a greater number of anti-MF drugs, including Chinese herbal medicine. For example, it has been shown that miR-21 activates the ERK signaling pathway, EndMT, and the TGF- β signaling pathway to promote MF (Cheng et al., 2016; Yuan et al., 2017; Li et al., 2020). Based on the preceding statement, miR-21 has been proven to be a target for many Chinese herbal medicines in combating MF. Tripterine, Notoginsenoside R1, Shenzhu Xinkang Decoction, and the ultrafiltration extract of Radix Angelica Sinensis and Radix Hedysari can all alleviate MF by inhibiting miR-21. Therefore, miR-21 may serve as a binding target for screening anti-MF drugs.

In conclusion, it is clear in this emerging field that ncRNAs appear to be important players in mediating Chinese herbal medicine's effect in various diseases. Indeed, as we have reviewed, ncRNAs interact with fibrosis-related genes and signalling pathways, making them a pivotal bridge in mediating the therapeutic effect of Chinese herbal medicine on MF. Furthermore, ncRNAs represent promising clinical drug targets and establishing an anti-MF drug screening platform based on ncRNAs to screen drugs including Chinese herbal medicine represents a challenging but promising field for future drug development in cardiovascular diseases.

Author contributions

MW: Conceptualization, Data curation, Writing—original draft. MY: Writing—original draft. LT: Writing—original draft, Data curation. XZ: Data curation, Writing—original draft, Methodology. GL: Data curation, Writing—original draft, Visualization. ZZ: Data curation, Writing—original draft. JZ: Writing—review and editing. HG: Writing—review and editing, Methodology, Software, Validation. WQ: Writing—review and editing, Funding acquisition, Supervision, Conceptualization.

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References

- Aguilar, R., Spencer, K. B., Kesner, B., Rizvi, N. F., Badmalia, M. D., Mrozowich, T., et al. (2022). Targeting Xist with compounds that disrupt RNA structure and X inactivation. *Nature* 604 (7904), 160–166. doi:10.1038/s41586-022-04537-z
- Albadrani, G. M., BinMowyna, M. N., Bin-Jumah, M. N., El-Akabawy, G., Aldera, H., and Al-Farga, A. M. (2021). Quercetin prevents myocardial infarction adverse remodeling in rats by attenuating TGF- β 1/Smad3 signaling: different mechanisms of action. *Saudi J. Biol. Sci.* 28 (5), 2772–2782. doi:10.1016/j.sjbs.2021.02.007
- Chen, L., Ji, Q., Zhu, H., Ren, Y., Fan, Z., and Tian, N. (2018a). miR-30a attenuates cardiac fibrosis in rats with myocardial infarction by inhibiting CTGF. *Exp. Ther. Med.* 15 (5), 4318–4324. doi:10.3892/etm.2018.5952
- Chen, Z., Li, C., Lin, K., Cai, H., Ruan, W., Han, J., et al. (2018b). Non-coding RNAs in cardiac fibrosis: emerging biomarkers and therapeutic targets. *Cardiol. J.* 25 (6), 732–741. doi:10.5603/CJ.a2017.0153
- Cheng, M., Wu, G., Song, Y., Wang, L., Tu, L., Zhang, L., et al. (2016). Celastrol-induced suppression of the MiR-21/ERK signalling pathway attenuates cardiac fibrosis and dysfunction. *Cell Physiol. biochem.* 38 (5), 1928–1938. doi:10.1159/000445554
- Davoodvandi, A., Marzban, H., Goleij, P., Sahebkar, A., Morshedi, K., Rezaei, S., et al. (2021). Effects of therapeutic probiotics on modulation of microRNAs. *Cell Commun. Signal* 19 (1), 4. doi:10.1186/s12964-020-00668-w
- Dilmaghani, A. N., Shoorai, H., Sharifi, G., Mohaqiq, M., Majidpoor, J., Dinger, M. E., et al. (2021). Non-coding RNAs modulate function of extracellular matrix proteins. *Biomed. Pharmacother.* 136, 111240. doi:10.1016/j.biopha.2021.111240
- Dit Beaufils, A. L., Huttin, O., Jobbe-Duval, A., Senage, T., Filippetti, L., Piriou, N., et al. (2021). Replacement myocardial fibrosis in patients with mitral valve prolapse: relation to mitral regurgitation, ventricular remodeling, and arrhythmia. *Circulation* 143 (18), 1763–1774. doi:10.1161/circulationaha.120.050214
- Dong, F., Dong, S., Liang, Y., Wang, K., Qin, Y., and Zhao, X. (2019a). MiR-34a promotes myocardial infarction in rats by inhibiting the activity of SIRT1. *Eur. Rev. Med. Pharmacol. Sci.* 23 (16), 7059–7065. doi:10.26355/eurrev_201908_18750
- Dong, Y., Chen, H., Gao, J., Liu, Y., Li, J., and Wang, J. (2019b). Bioactive ingredients in Chinese herbal medicines that target non-coding RNAs: promising new choices for disease treatment. *Front. Pharmacol.* 10, 515. doi:10.3389/fphar.2019.00515
- Dragomir, M., and Calin, G. A. (2018). Circular RNAs in cancer - lessons learned from microRNAs. *Front. Oncol.* 8, 179. doi:10.3389/fonc.2018.00179
- Dykes, I. M., and Emanuel, C. (2017). Transcriptional and post-transcriptional gene regulation by long non-coding RNA. *Genom. Proteom. Bioinf.* 15 (3), 177–186. doi:10.1016/j.gpb.2016.12.005
- Eckardt, L., Haverkamp, W., Johna, R., Böcker, D., Deng, M. C., Breithardt, G., et al. (2000). Arrhythmias in heart failure: current concepts of mechanisms and therapy. *J. Cardiovasc. Electrophysiol.* 11 (1), 106–117. doi:10.1111/j.1540-8167.2000.tb00746.x
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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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- Hu, J., Wang, X., Cui, X., Kuang, W., Li, D., and Wang, J. (2021). Quercetin prevents isoprenaline-induced myocardial fibrosis by promoting autophagy via regulating miR-223-3p/FOXO3. *Cell Cycle* 20 (13), 1253–1269. doi:10.1080/15384101.2021.1932029
- Huang, C., Wang, R., Lu, J., He, Y., Wu, Y., Ma, W., et al. (2022a). MicroRNA-338-3p as a therapeutic target in cardiac fibrosis through FGFR2 suppression. *J. Clin. Lab. Anal.* 36 (8), e24584. doi:10.1002/jcla.24584
- Huang, L., Xu, D., Chen, Y., Yue, S., and Tang, Y. (2021). Leonurine, a potential drug for the treatment of cardiovascular system and central nervous system diseases. *Brain Behav.* 11 (2), e01995. doi:10.1002/brb3.1995
- Huang, Q., and Huang, Q. (2023). Inhibition of lncRNA DANCER prevents heart failure by ameliorating cardiac hypertrophy and fibrosis via regulation of the miR-758-3p/PRG4/smad Axis. *J. Cardiovasc. Transl. Res.* 16 (6), 1357–1372. doi:10.1007/s12265-023-10428-z
- Huang, S., Zhang, L., Song, J., Wang, Z., Huang, X., Guo, Z., et al. (2019). Long noncoding RNA MALAT1 mediates cardiac fibrosis in experimental postinfarct myocardium mice model. *J. Cell. Physiol.* 234 (3), 2997–3006. doi:10.1002/jcp.27117
- Huang, Y. (2018). The novel regulatory role of lncRNA-miRNA-mRNA axis in cardiovascular diseases. *J. Cell. Mol. Med.* 22 (12), 5768–5775. doi:10.1111/jcmm.13866
- Huang, Y., Cheng, M., Wang, X., Dong, H., and Gao, J. (2022b). Dang Gui Bu Xue Tang, a conventional Chinese herb decoction, ameliorates radiation-induced heart disease via Nrf2/HMGB1 pathway. *Front. Pharmacol.* 13, 1086206. doi:10.3389/fphar.2022.1086206
- Huang, Y., Qi, Y., Du, J., and Zhang, D. (2014). MicroRNA-34a regulates cardiac fibrosis after myocardial infarction by targeting Smad4. *Expert Opin. Ther. Targets* 18 (12), 1355–1365. doi:10.1517/14728222.2014.961424
- Jeong, A., Lim, Y., Kook, T., Kwon, D. H., Cho, Y. K., Ryu, J., et al. (2023). Circular RNA circSMAD4 regulates cardiac fibrosis by targeting miR-671-5p and FGFR2 in cardiac fibroblasts. *Mol. Ther. Nucleic Acids* 34, 102071. doi:10.1016/j.omtn.2023.102071
- Jha, S., Thasmas Loganathbabu, V. K., Kumaran, K., Krishnasamy, G., and Aruljothi, K. N. (2023). Long non-coding RNAs (lncRNAs) in heart failure: a comprehensive review. *Non-coding RNA* 10 (1), 3. doi:10.3390/ncrna10010003
- Jia, X., Shao, W., and Tian, S. (2022). Berberine alleviates myocardial ischemia-reperfusion injury by inhibiting inflammatory response and oxidative stress: the key function of miR-26b-5p-mediated PTGS2/MAPK signal transduction. *Pharm. Biol.* 60 (1), 652–663. doi:10.1080/13880209.2022.2048029
- Jiang, L., Wang, X., Zhan, X., Kang, S., Liu, H., Luo, Y., et al. (2020). Advance in circular RNA modulation effects of heart failure. *Gene X* 763s (2020), 100036. doi:10.1016/j.gene.2020.100036
- Jiao, D., Liu, Y., Hou, T., Xu, H., Wang, X., Shi, Q., et al. (2021). Notoginsenoside R1 (NG-R1) promoted lymphatic drainage function to ameliorating rheumatoid arthritis in TNF-Tg mice by suppressing NF- κ B signaling pathway. *Front. Pharmacol.* 12, 730579. doi:10.3389/fphar.2021.730579
- Jing, H., Xie, R., Bai, Y., Duan, Y., Sun, C., Wang, Y., et al. (2021). The mechanism actions of astragaloside IV prevents the progression of hypertensive heart disease based on network pharmacology and experimental pharmacology. *Front. Pharmacol.* 12, 755653. doi:10.3389/fphar.2021.755653
- Jones, J. A., Stroud, R. E., O'Quinn, E. C., Black, L. E., Barth, J. L., Eleftheriades, J. A., et al. (2011). Selective microRNA suppression in human thoracic aneurysms: relationship of miR-29a to aortic size and proteolytic induction. *Circ. Cardiovasc. Genet.* 4 (6), 605–613. doi:10.1161/circgenetics.111.960419
- Karakikes, I., Chaanine, A. H., Kang, S., Mukete, B. N., Jeong, D., Zhang, S., et al. (2013). Therapeutic cardiac-targeted delivery of miR-1 reverses pressure overload-induced cardiac hypertrophy and attenuates pathological remodeling. *J. Am. Heart Assoc.* 2 (2), e000078. doi:10.1161/jaha.113.000078
- Kobayashi, M., Machida, N., Mitsuishi, M., and Yamane, Y. (2004). Beta-blocker improves survival, left ventricular function, and myocardial remodeling in hypertensive rats with diastolic heart failure. *Am. J. Hypertens.* 17 (12), 1112–1119. doi:10.1016/j.amjhyper.2004.07.007
- Krieg, A. M. (2011). Is RNAi dead? *Mol. Ther.* 19 (6), 1001–1002. doi:10.1038/mt.2011.94
- Kumar, H., Kawai, T., and Akira, S. (2011). Pathogen recognition by the innate immune system. *Int. Rev. Immunol.* 30 (1), 16–34. doi:10.3109/08830185.2010.529976
- Leitão, A. L., and Enguita, F. J. (2022). A structural view of miRNA biogenesis and function. *Non-coding RNA* 8 (1), 10. doi:10.3390/ncrna8010010
- Li, Q., Yao, Y., Shi, S., Zhou, M., Zhou, Y., Wang, M., et al. (2020). Inhibition of miR-21 alleviated cardiac perivascular fibrosis via repressing EndMT in T1DM. *J. Cell. Mol. Med.* 24 (1), 910–920. doi:10.1111/jcmm.14800
- Li, W. Y., Long, Q. Y., Fu, X. Y., Ma, L., Tan, W., Li, Y. L., et al. (2023a). Effects of Buyang Huanwu decoction and astragali radix-angelicae sinensis radix combination on inflammatory responses in atherosclerotic mice. *Chin. J. Chin. Mater. Med.* 48 (15), 4164–4172. doi:10.19540/j.cnki.cjcm.20230418.401
- Li, X., Liu, Q., and Liu, J. (2023b). Long non-coding RNAs: discoveries, mechanisms, and research strategies in seeds. *Genes* 14 (12), 2214. doi:10.3390/genes14122214
- Li, Z., Ye, Z., Ma, J., Gu, Q., Teng, J., and Gong, X. (2021). MicroRNA-133b alleviates doxorubicin-induced cardiomyocyte apoptosis and cardiac fibrosis by targeting PTBP1 and TAGLN2. *Int. J. Mol. Med.* 48 (1), 125. doi:10.3892/ijmm.2021.4958
- Liang, H., Pan, Z., Zhao, X., Liu, L., Sun, J., Su, X., et al. (2018). lncRNA PFL contributes to cardiac fibrosis by acting as a competing endogenous RNA of let-7d. *Theranostics* 8 (4), 1180–1194. doi:10.7150/thno.20846
- Lieben Louis, X., Meikle, Z., Chan, L., DeGagne, G., Cummer, R., Meikle, S., et al. (2019). Divergent effects of resveratrol on rat cardiac fibroblasts and cardiomyocytes. *Molecules* 24 (14), 2604. doi:10.3390/molecules24142604
- Liu, L., Ning, B., Cui, J., Zhang, T., and Chen, Y. (2017). MiR-29c is implicated in the cardioprotective activity of Panax notoginseng saponins against isoproterenol-induced myocardial fibrogenesis. *J. Ethnopharmacol.* 198, 1–4. doi:10.1016/j.jep.2016.12.036
- Liu, W., and Yu, S. (2022). Research progress on the cardiorenal protection of non-steroid mineralocorticoid receptor antagonists in patients with chronic kidney disease. *Acta physiol. Sin.* 74 (6), 1023–1030. doi:10.13294/j.aps.2022.0092
- Lu, L., Yuan, L., Liang, B., Luo, J., Yang, H., Li, L., et al. (2018). Effect of leonurine on expression of miR-1 in rats with myocardial fibrosis induced by isoproterenol. *Chin. J. Pathophysiol.* 34 (11), 1928–1934. doi:10.3969/j.issn.1000-4718.2018.11.002
- Lu, T., Wu, Y., Chen, W., and Hung, Y. (2022). Targeting oxidative stress and endothelial dysfunction using Tanshinone IIA for the treatment of tissue inflammation and fibrosis. *Oxid. Med. Cell Longev.* 2022, 2811789. doi:10.1155/2022/2811789
- Lun, J., Guo, J., Yu, M., Zhang, H., and Fang, J. (2023). Circular RNAs in inflammatory bowel disease. *Front. Immunol.* 14, 1307985. doi:10.3389/fimmu.2023.1307985
- Lv, S., Yuan, P., Lu, C., Dong, J., Li, M., Qu, F., et al. (2021). QiShenYiQi pill activates autophagy to attenuate reactive myocardial fibrosis via the PI3K/AKT/mTOR pathway. *Aging* 13 (4), 5525–5538. doi:10.18632/aging.202482
- Ma, C., Fu, Z., Guo, H., Wei, H., Zhao, X., and Li, Y. (2019). The effects of Radix Angelica Sinensis and Radix Hedysari ultrafiltration extract on X-irradiation-induced myocardial fibrosis in rats. *Biomed. Pharmacother.* 112, 108596. doi:10.1016/j.biopha.2019.01.057
- Ma, L., Bajic, V. B., and Zhang, Z. (2013). On the classification of long non-coding RNAs. *RNA Biol.* 10 (6), 925–933. doi:10.4161/rna.24604
- Malaguarnera, L. (2019). Influence of resveratrol on the immune response. *Nutrients* 11 (5), 946. doi:10.3390/nu11050946
- Mbanze, J., Cumbane, B., Jive, R., and Mocumbi, A. (2020). Challenges in addressing the knowledge gap on endomyocardial fibrosis through community-based studies. *Cardiovasc. Diagn. Ther.* 10 (2), 279–288. doi:10.21037/cdt.2019.08.07
- Mester-Tonczar, J., Winkler, J., Einzinger, P., Hasimbegovic, E., Kastner, N., Lukovic, D., et al. (2020). Association between circular RNA CDR1as and post-infarction cardiac function in pig ischemic heart failure: influence of the anti-fibrotic natural compounds bufalin and lycorine. *Biomolecules* 10 (8), 1180. doi:10.3390/biom10081180
- Micheletti, R., Plaisance, I., Abraham, B. J., Sarre, A., Ting, C. C., Alexanian, M., et al. (2017). The long noncoding RNA Wisper controls cardiac fibrosis and remodeling. *Sci. Transl. Med.* 9 (395), eaai9118. doi:10.1126/scitranslmed.aai9118
- Mohr, A. M., and Mott, J. L. (2015). Overview of microRNA biology. *Semin. Liver Dis.* 35 (1), 3–11. doi:10.1055/s-0034-1397344
- Nagpal, V., Rai, R., Place, A. T., Murphy, S. B., Verma, S. K., Ghosh, A. K., et al. (2016). MiR-125b is critical for fibroblast-to-myofibroblast transition and cardiac fibrosis. *Circulation* 133 (3), 291–301. doi:10.1161/circulationaha.115.018174
- Ning, B. (2016). *Study on the inhibitory effect of Panax notoginseng saponin R1 on myocardial fibrosis induced by ISO and its mechanism*[Ph.D.]. Shanghai: Shanghai University of Traditional Chinese Medicine.
- Oh, H., Park, H. E., Song, M. S., Kim, H., and Baek, J. H. (2022). The therapeutic potential of anticoagulation in organ fibrosis. *Front. Med.* 9, 866746. doi:10.3389/fmed.2022.866746
- Pagliaro, B., Santolamazza, C., Simonelli, F., and Rubattu, S. (2015). Phytochemical compounds and protection from cardiovascular diseases: a state of the art. *Biomed. Res. Int.* 2015, 918069. doi:10.1155/2015/918069
- Palazzo, A. F., and Lee, E. S. (2015). Non-coding RNA: what is functional and what is junk? *Front. Genet.* 6, 2. doi:10.3389/fgene.2015.00002
- Parikh, M., Kura, B., O'Hara, K. A., Dibrov, E., Neticadan, T., Slezak, J., et al. (2020). Cardioprotective effects of dietary flaxseed post-infarction are associated with changes in microRNA expression. *Biomolecules* 10 (9), 1297. doi:10.3390/biom10091297
- Piccoli, M. T., Gupta, S. K., Viereck, J., Foinquinos, A., Samolovac, S., Kramer, F. L., et al. (2017). Inhibition of the cardiac fibroblast-enriched lncRNA Meg3 prevents cardiac fibrosis and diastolic dysfunction. *Circ. Res.* 121 (5), 575–583. doi:10.1161/circresaha.117.310624
- Prabhu, S. D. (2005). Post-infarction ventricular remodeling: an array of molecular events. *J. Mol. Cell. Cardiol.* 38 (4), 547–550. doi:10.1016/j.yjmc.2005.01.014
- Prasad, K., Khan, A. S., and Shoker, M. (2020). Flaxseed and its components in treatment of hyperlipidemia and cardiovascular disease. *Int. J. Angiol.* 29 (4), 216–222. doi:10.1055/s-0040-1709129

- Qi, Y., Zhu, K., Ren, X., Lou, L., Wu, A., Guo, Q., et al. (2019). Effects of Fuzheng Huayu Capsule on the expression of miR-29 family in rats with myocardial fibrosis after myocardial infarction. *Glob. Tradit. Chin. Med.* 12 (6), 839–844. doi:10.3969/j.issn.1674-1749.2019.06.005
- Qiao, P., Xu, J., Liu, X., and Li, X. (2021). Tanshinone IIA improves ventricular remodeling following cardiac infarction by regulating miR-205-3p. *Dis. Markers* 2021, 8740831. doi:10.1155/2021/8740831
- Qu, X., Du, Y., Shu, Y., Gao, M., Sun, F., Luo, S., et al. (2017). MIAT is a pro-fibrotic long non-coding RNA governing cardiac fibrosis in post-infarct myocardium. *Sci. Rep.* 7, 42657. doi:10.1038/srep42657
- Qu, X., Song, X., Yuan, W., Shu, Y., Wang, Y., Zhao, X., et al. (2016). Expression signature of lncRNAs and their potential roles in cardiac fibrosis of post-infarct mice. *Biosci. Rep.* 36 (3), e00337. doi:10.1042/bsr20150278
- Rai, R., Sun, T., Ramirez, V., Lux, E., Eren, M., Vaughan, D. E., et al. (2019). Acetyltransferase p300 inhibitor reverses hypertension-induced cardiac fibrosis. *J. Cell Mol. Med.* 23 (4), 3026–3031. doi:10.1111/jcmm.14162
- Raj, P., Thandapilly, S. J., Wigle, J., Zieroth, S., and Neticadan, T. (2021). A comprehensive analysis of the efficacy of resveratrol in atherosclerotic cardiovascular disease, myocardial infarction and heart failure. *Molecules* 26 (21), 6600. doi:10.3390/molecules26216600
- Ramazani, Y., Knops, N., Elmonem, M. A., Nguyen, T. Q., Arcolino, F. O., van den Heuvel, L., et al. (2018). Connective tissue growth factor (CTGF) from basics to clinics. *Matrix Biol.* 68–69, 44–66. doi:10.1016/j.matbio.2018.03.007
- Rang, Z., Wang, Z., Pang, Q., Wang, Y., Yang, G., and Cui, F. (2016). MiR-181a targets PHILP2 to augment Akt signaling and regulate proliferation and apoptosis in human keloid fibroblasts. *Cell Physiol. Biochem.* 40 (3–4), 796–806. doi:10.1159/000453139
- Rooij, E. V., Sutherland, L. B., Thatcher, J. E., DiMaio, J. M., Naseem, R. H., Marshall, W. S., et al. (2008). Dysregulation of microRNAs after myocardial infarction reveals a role of miR-29 in cardiac fibrosis. *Proc. Natl. Acad. Sci. U. S. A.* 105 (35), 13027–13032. doi:10.1073/pnas.0805038105
- Salmena, L., Poliseno, L., Tay, Y., Kats, L., and Pandolfi, P. P. (2011). A ceRNA hypothesis: the Rosetta Stone of a hidden RNA language? *Cell* 146 (3), 353–358. doi:10.1016/j.cell.2011.07.014
- Schimmel, K., Jung, M., Foinquinos, A., José, G. S., Beaumont, J., Bock, K., et al. (2020). Natural compound library screening identifies new molecules for the treatment of cardiac fibrosis and diastolic dysfunction. *Circulation* 141 (9), 751–767. doi:10.1161/circulationaha.119.042559
- Serraino, G. F., Jiritano, F., Costa, D., Ielapi, N., Battaglia, D., Bracale, U. M., et al. (2023). Metalloproteinases in cardiac surgery: a systematic review. *Biomolecules* 13 (1), 113. doi:10.3390/biom13010113
- Shi, J., Lai, J., Lin, Y., Xu, X., Guo, S., Wang, H., et al. (2020). Tanshinone IIA down-regulated p-Smad3 signaling to inhibit TGF- β 1-mediated fibroblast proliferation via lncRNA-HSRL/SNX9. *Int. J. Biochem. Cell Biol.* 129, 105863. doi:10.1016/j.biocel.2020.105863
- Sledz, C. A., and Williams, B. R. (2004). RNA interference and double-stranded-RNA-activated pathways. *Biochem. Soc. Trans.* 32 (6), 952–956. doi:10.1042/bst0320952
- Srivastava, P. K., and Kittleson, M. M. (2024). Modern advances in heart transplantation. *Prog. Cardiovasc. Dis.* doi:10.1016/j.pcad.2024.01.012
- Sun, R., Wang, J., Zheng, Y., Li, X., Xie, T., Li, R., et al. (2017). Traditional Chinese medicine baixin decoction improves cardiac fibrosis of rats with dilated cardiomyopathy. *Exp. Ther. Med.* 13 (5), 1900–1906. doi:10.3892/etm.2017.4223
- Sun, W. M., Wang, Y. P., Duan, Y. Q., Shang, H. X., and Cheng, W. D. (2014). Radix Hedyari polysaccharide suppresses lipid metabolism dysfunction in a rat model of non-alcoholic fatty liver disease via adenosine monophosphate-activated protein kinase pathway activation. *Mol. Med. Rep.* 10 (3), 1237–1244. doi:10.3892/mmr.2014.2327
- Sun, X., Tan, Y., Lyu, J., Liu, H. L., Zhao, Z. M., and Liu, C. H. (2022). Active components formulation developed from Fuzheng Huayu recipe for anti-liver fibrosis. *Chin. J. Integr. Med.* 28 (6), 538–544. doi:10.1007/s11655-021-3293-x
- Sutton, M. G., and Sharpe, N. (2000). Left ventricular remodeling after myocardial infarction: pathophysiology and therapy. *Circulation* 101 (25), 2981–2988. doi:10.1161/01.cir.101.25.2981
- Talman, V., and Ruskoaho, H. (2016). Cardiac fibrosis in myocardial infarction-from repair and remodeling to regeneration. *Cell Tissue Res.* 365 (3), 563–581. doi:10.1007/s00441-016-2431-9
- Tan, W., Wang, K., Yang, X., Wang, K., Wang, N., and Jiang, T. (2022). LncRNA HOTAIR promotes myocardial fibrosis in atrial fibrillation through binding with PTBP1 to increase the stability of Wnt5a. *Int. J. Cardiol.* 369, 21–28. doi:10.1016/j.ijcard.2022.06.073
- Tymińska, A., Ozierański, K., Caforio, A. L. P., Marcolongo, R., Marchel, M., Kaplon-Cieśliska, A., et al. (2021). Myocarditis and inflammatory cardiomyopathy in 2021: an update. *Pol. Arch. Intern. Med.* 131 (6), 594–606. doi:10.20452/pamw.16010
- Verjans, R., Peters, T., Beaumont, F. J., van Leeuwen, R., van Herwaarden, T., Verhesen, W., et al. (2018). MicroRNA-221/222 family counteracts myocardial fibrosis in pressure overload-induced heart failure. *Hypertension* 71 (2), 280–288. doi:10.1161/hypertensionaha.117.10094
- Wang, J., Chen, P., Cao, Q., Wang, W., and Chang, X. (2022a). Traditional Chinese medicine Ginseng Dingzhi decoction ameliorates myocardial fibrosis and high glucose-induced cardiomyocyte injury by regulating intestinal flora and mitochondrial dysfunction. *Oxid. Med. Cell. Longev.* 2022, 9205908. doi:10.1155/2022/9205908
- Wang, J., Zhang, Y., Liu, Y. M., Guo, L. L., Wu, P., Dong, Y., et al. (2016). Huoxue Anxin Recipe promotes myocardium angiogenesis of acute myocardial infarction rats by up-regulating miR-210 and vascular endothelial growth factor. *Chin. J. Integr. Med.* 22 (9), 685–690. doi:10.1007/s11655-016-2508-z
- Wang, K., Lin, Y., Shen, H., Yu, S., and Xu, J. (2023). LncRNA TUG1 exacerbates myocardial fibrosis in diabetic cardiomyopathy by modulating the microRNA-145a-5p/cf2 Axis. *J. Cardiovasc. Pharmacol.* 81 (3), 192–202. doi:10.1097/fjc.0000000000001391
- Wang, Q. L., Yuan, J. L., Tao, Y. Y., Zhang, Y., Liu, P., and Liu, C. H. (2010). Fuzheng Huayu recipe and vitamin E reverse renal interstitial fibrosis through counteracting TGF- β 1-induced epithelial-to-mesenchymal transition. *J. Ethnopharmacol.* 127 (3), 631–640. doi:10.1016/j.jep.2009.12.011
- Wang, R., Peng, L., Lv, D., Shang, F., Yan, J., Li, G., et al. (2021). Leonurine attenuates myocardial fibrosis through upregulation of miR-29a-3p in mice post-myocardial infarction. *J. Cardiovasc. Pharmacol.* 77 (2), 189–199. doi:10.1097/fjc.0000000000000957
- Wang, T., Jiang, X., Ruan, Y., Zhuang, J., and Yin, Y. (2022b). Based on network pharmacology and *in vitro* experiments to prove the effective inhibition of myocardial fibrosis by Buyang Huanwu decoction. *Bioengineered* 13 (5), 13767–13783. doi:10.1080/21655979.2022.2084253
- Wang, W., Wang, H., Geng, Q., Wang, H., Miao, W., Cheng, B., et al. (2015a). Augmentation of autophagy by atorvastatin via Akt/mTOR pathway in spontaneously hypertensive rats. *Hypertens. Res.* 38 (12), 813–820. doi:10.1038/hr.2015.85
- Wang, Y., He, L., Du, Y., Zhu, P., Huang, G., Luo, J., et al. (2015b). The long noncoding RNA lncTCF7 promotes self-renewal of human liver cancer stem cells through activation of Wnt signaling. *Cell Stem Cell* 16 (4), 413–425. doi:10.1016/j.stem.2015.03.003
- Wei, Y., Wu, Y., Feng, K., Zhao, Y., Tao, R., Xu, H., et al. (2020). Astragaloside IV inhibits cardiac fibrosis via miR-135a-TRPM7-TGF- β /Smads pathway. *J. Ethnopharmacol.* 249, 112404. doi:10.1016/j.jep.2019.112404
- Wu, N., Xu, J., Du, W. W., Li, X., Awan, F. M., Li, F., et al. (2021). YAP circular RNA, circYap, attenuates cardiac fibrosis via binding with tropomyosin-4 and gamma-actin decreasing actin polymerization. *Mol. Ther.* 29 (3), 1138–1150. doi:10.1016/j.yjmt.2020.12.004
- Xi, T., Wang, R., Pi, D., Ouyang, J., and Yang, J. (2023). The p53/miR-29a-3p axis mediates the antifibrotic effect of leonurine on angiotensin II-stimulated rat cardiac fibroblasts. *Exp. Cell Res.* 426 (1), 113556. doi:10.1016/j.yexcr.2023.113556
- Xu, J., Wu, H., Chen, S., Qi, B., Zhou, G., Cai, L., et al. (2018). MicroRNA-30c suppresses the pro-fibrogenic effects of cardiac fibroblasts induced by TGF- β 1 and prevents atrial fibrosis by targeting TGF β RII. *J. Cell Mol. Med.* 22 (6), 3045–3057. doi:10.1111/jcmm.13548
- Yan, H., and Bu, P. (2021). Non-coding RNA in cancer. *Essays Biochem.* 65 (4), 625–639. doi:10.1042/ebc20200032
- Yan, N., Xiao, C., Wang, X., Xu, Z., and Yang, J. (2022). Tanshinone IIA from *Salvia miltiorrhiza* exerts anti-fibrotic effects on cardiac fibroblasts and rat heart tissues by suppressing the levels of pro-fibrotic factors: the key role of miR-618. *J. Food Biochem.* 46 (2), e14078. doi:10.1111/jfbc.14078
- Yang, B., Zheng, C., Yu, H., Zhang, R., Zhao, C., and Cai, S. (2019a). Cardio-protective effects of salvinolic acid B on oxygen and glucose deprivation (OGD)-treated H9c2 cells. *Artif. Cells Nanomed. Biotechnol.* 47 (1), 2274–2281. doi:10.1080/21691401.2019.1621885
- Yang, F., Li, P., Li, H., Shi, Q., Li, S., and Zhao, L. (2015). MicroRNA-29b mediates the antifibrotic effect of tanshinone IIA in postinfarct cardiac remodeling. *J. Cardiovasc. Pharmacol.* 65 (5), 456–464. doi:10.1097/fjc.0000000000000214
- Yang, G., Wang, F., Wang, Y., Yu, X., Yang, S., Xu, H., et al. (2020). Protective effect of tanshinone IIA on H(2)O(2)-induced oxidative stress injury in rat cardiomyocytes by activating Nrf2 pathway. *J. Recept. Signal Transduct. Res.* 40 (3), 264–272. doi:10.1080/10799893.2020.1731535
- Yang, H., Liu, Z., Hu, X., Liu, X., Gui, L., Cai, Z., et al. (2022). Protective effect of Panax Notoginseng Saponins on apolipoprotein-E-deficient atherosclerosis-prone mice. *Curr. Pharm. Des.* 28 (6), 671–677. doi:10.2174/1381612828666220128104636
- Yang, K., Li, W., Duan, W., Jiang, Y., Huang, N., Li, Y., et al. (2019b). Resveratrol attenuates pulmonary embolism associated cardiac injury by suppressing activation of the inflammasome via the MALAT1-miR-22-3p signaling pathway. *Int. J. Mol. Med.* 44 (6), 2311–2320. doi:10.3892/ijmm.2019.4358
- Ye, Q., Liu, Q., Ma, X., Bai, S., Chen, P., Zhao, Y., et al. (2021). MicroRNA-146b-5p promotes atrial fibrosis in atrial fibrillation by repressing TIMP4. *J. Cell. Mol. Med.* 25 (22), 10543–10553. doi:10.1111/jcmm.16985
- Yin, Y., Zhang, Q., Zhao, Q., Ding, G., Wei, C., Chang, L., et al. (2019). Tongxinluo attenuates myocardial fibrosis after acute myocardial infarction in rats via inhibition of endothelial-to-mesenchymal transition. *Biomed. Res. Int.* 2019, 6595437. doi:10.1155/2019/6595437

- Ytrehus, K., Hulot, J. S., Perrino, C., Schiattarella, G. G., and Madonna, R. (2018). Perivascular fibrosis and the microvasculature of the heart. Still hidden secrets of pathophysiology? *Vasc. Pharmacol.* 107, 78–83. doi:10.1016/j.vph.2018.04.007
- Yu, A. M., Choi, Y. H., and Tu, M. J. (2020). RNA drugs and RNA targets for small molecules: principles, progress, and challenges. *Pharmacol. Rev.* 72 (4), 862–898. doi:10.1124/pr.120.019554
- Yu, B., Yu, N., Wang, Y., Zhang, H., Wan, K., Sun, X., et al. (2019). Role of miR-133a in regulating TGF- β 1 signaling pathway in myocardial fibrosis after acute myocardial infarction in rats. *Eur. Rev. Med. Pharmacol. Sci.* 23 (19), 8588–8597. doi:10.26355/eurrev_201910_19175
- Yuan, J., Chen, H., Ge, D., Xu, Y., Xu, H., Yang, Y., et al. (2017). Mir-21 promotes cardiac fibrosis after myocardial infarction via targeting Smad7. *Cell Physiol. Biochem.* 42 (6), 2207–2219. doi:10.1159/000479995
- Yuan, S., Liang, J., Zhang, M., Zhu, J., Pan, R., Li, H., et al. (2019). CircRNA_005647 inhibits expressions of fibrosis-related genes in mouse cardiac fibroblasts via sponging miR-27b-3p. *J. South. Med. Univ.* 39 (11), 1312–1319. doi:10.12122/j.jissn.1673-4254.2019.11.08
- Yuan, X., Liu, K., Dong, P., and Han, H. (2023). Protective effect and mechanism of different proportions of "Danggui-Kushen" herb pair on ischemic heart disease. *Heliyon* 9 (11), e22150. doi:10.1016/j.heliyon.2023.e22150
- Zeisberg, E. M., Tarnavski, O., Zeisberg, M., Dorfman, A. L., McMullen, J. R., Gustafsson, E., et al. (2007). Endothelial-to-mesenchymal transition contributes to cardiac fibrosis. *Nat. Med.* 13 (8), 952–961. doi:10.1038/nm1613
- Zeng, L., Yu, G., Yang, K., He, Q., Hao, W., Xiang, W., et al. (2024). Exploring the mechanism of Celastrol in the treatment of rheumatoid arthritis based on systems pharmacology and multi-omics. *Sci. Rep.* 14 (1), 1604. doi:10.1038/s41598-023-48248-5
- Zeng, Z., Pan, Y., Wu, W., Li, L., Wu, Z., Zhang, Y., et al. (2019). Myocardial hypertrophy is improved with berberine treatment via long non-coding RNA MIAT-mediated autophagy. *J. Pharm. Pharmacol.* 71 (12), 1822–1831. doi:10.1111/jphp.13170
- Zhai, S., Chen, Z., Cheng, X., Zhu, X., Zhao, Q., and Yu, Z. (2022). Experimental study on the effect of Shenzhu Xinkang decoction on myocardial fibrosis by regulating EndMT through miRNA-21. *Guid. J. Tradit. Chin. Med. Pharm.* 28 (09), 1–6+47. doi:10.13862/j.cn43-1446/r.2022.09.001
- Zhang, M., Zhang, B., Wang, X., Song, J., Tong, M., Dong, Z., et al. (2023). LncRNA CFAR promotes cardiac fibrosis via the miR-449a-5p/LOXL3/mTOR axis. *Sci. China Life Sci.* 66 (4), 783–799. doi:10.1007/s11427-021-2132-9
- Zhang, X., Wang, Q., Wang, X., Chen, X., Shao, M., Zhang, Q., et al. (2019). Tanshinone IIA protects against heart failure post-myocardial infarction via AMPKs/mTOR-dependent autophagy pathway. *Biomed. Pharmacother.* 112, 108599. doi:10.1016/j.biopha.2019.108599
- Zhang, Y., Huang, X., Wei, L., Chung, A., Yu, C., and Lan, H. (2014). miR-29b as a therapeutic agent for angiotensin II-induced cardiac fibrosis by targeting TGF- β /Smad3 signaling. *Mol. Ther.* 22 (5), 974–985. doi:10.1038/mt.2014.25
- Zhang, Y., Lu, Y., Ong'achwa, M. J., Ge, L., Qian, Y., Chen, L., et al. (2018). Resveratrol inhibits the TGF- β 1-induced proliferation of cardiac fibroblasts and collagen secretion by downregulating miR-17 in rat. *Biomed. Res. Int.* 2018, 8730593. doi:10.1155/2018/8730593
- Zhang, Y., Wang, J., Guo, L., Liu, Y., Wu, P., Dong, Y., et al. (2012). Experimental study on attenuating ischemic injury of acute myocardial infarction rats by Huoxue Anxin Recipe. *J. Chin. Med.* 32 (7), 939–943.
- Zhang, Y., Wang, J., Guo, L., and Wu, G. (2013). Huoxue anxin recipe alleviated peroxidation damage of acute myocardial infarction rats by regulating iNOS/eNOS imbalance: an experimental research. *Chin. J. Integr. Tradit. West. Med.* 33 (10), 1356–1360. doi:10.7661/CJIM.2013.10.1356
- Zhao, J., Wang, Y., Gao, J., Jing, Y., and Xin, W. (2020). Berberine mediated positive inotropic effects on rat hearts via a Ca(2+)-dependent mechanism. *Front. Pharmacol.* 11, 821. doi:10.3389/fphar.2020.00821
- Zhao, M., Yu, Z., Zhao, Q., Li, S., and Chen, J. (2023). To explore the mechanism of Shenzhu Xinkang Prescription in treating chronic heart failure based on network pharmacology. *Clin. J. Tradit. Chin. Med.* 35 (02), 265–272. doi:10.16448/j.cjctm.2023.0215
- Zheng, Y., Li, X., Li, Q., Zhang, Y., and Chen, M. (2020). Effect of Berberine on the expression of miRNA-29b in left ventricular hypertrophy and the mechanism of inhibition of myocardial fibrosis. *Chin. Med. Her.* 17 (14), 19–22. doi:10.13764/j.cnki.ncdm.2020.03.007
- Zhu, J., Chen, R., Fu, Y., Lin, Q., Huang, S., Guo, L., et al. (2013). Smad3 inactivation and MiR-29b upregulation mediate the effect of carvedilol on attenuating the acute myocardium infarction-induced myocardial fibrosis in rat. *PLoS One* 8 (9), e75557. doi:10.1371/journal.pone.0075557
- Zhu, Y., Qian, X., Li, J., Lin, X., Luo, J., Huang, J., et al. (2019). Astragaloside-IV protects H9C2(2-1) cardiomyocytes from high glucose-induced injury via miR-34a-mediated autophagy pathway. *Artif. Cells Nanomed. Biotechnol.* 47 (1), 4172–4181. doi:10.1080/21691401.2019.1687492
- Zhuang, Y., Li, T., Hu, X., Xie, Y., Pei, X., Wang, C., et al. (2023). MetBil as a novel molecular regulator in ischemia-induced cardiac fibrosis via METTL3-mediated m6A modification. *FASEB J.* 37 (3), e22797. doi:10.1096/fj.202201734R

Glossary

| | | | |
|------------------|--|----------------|---|
| AGO | Argonute | RISC | RNA-inducing silencing complex |
| ALA | α -linolenic acid | SEPP1 | selenoprotein P1 |
| AMPK | AMP-activated protein kinase | Sfrp2 | secreted frizzled related protein 2 |
| ACTG | gamma-actin | SNX9 | sorting connexin 9 |
| ceRNA | competitive endogenous RNA | SWI/SNF | switch/sucrose non-fermentable |
| Cfl2 | cofilin-2 | TAGLN2 | transgelin 2 |
| CFs | cardiac fibroblasts | TCF7 | transcription factor 7 |
| circRNA | circular RNA | TIMP | tissue inhibitors of matrix metalloproteinase |
| CTGF | connective tissue growth factor | TIMP1 | tissue inhibitors of matrix metalloproteinase 1 |
| dsRBP | dsRNA binding protein | TIMP4 | tissue inhibitors of matrix metalloproteinase 4 |
| ECM | extracellular matrix | TMP4 | tropomyosin-4 |
| EndMT | endothelial to mesenchymal transition | TRPM7 | transient receptor potential melastatin 7 |
| FGFR2 | fibroblast growth factor receptor 2 | VEGF | vascular endothelial growth factor |
| FOXO3 | forkhead box O3 | | |
| FUS | fused in sarcoma | | |
| Fbln2 | fibullin-2 | | |
| HOTAIR | HOX transcript antisense RNA | | |
| HSRL | human-specific regulatory loci | | |
| lncRNA | long non-coding RNA | | |
| LOXL3 | lysyl oxidase-like protein-3 | | |
| MALAT1 | metastasis associated lung adenocarcinoma transcript 1 | | |
| Meg3 | maternally expressed gene 3 | | |
| METTL3 | methyltransferase like 3 | | |
| MIAT | myocardial infarction-associated transcript | | |
| MF | myocardial fibrosis | | |
| miRNA | microRNA | | |
| MMP2 | matrix metalloproteinase 2 | | |
| MMP9 | matrix metalloproteinase 9 | | |
| MMP14 | matrix metalloproteinase 14 | | |
| MDM2 | murine double minute 2 | | |
| mTOR | mammalian target of rapamycin | | |
| ncRNAs | non-coding RNAs | | |
| OGD | oxygen and glucose deprivation | | |
| PAMP | pathogen-associated molecular pattern | | |
| PHLPP2 | PH domain leucine-rich repeat protein phosphatase 2 | | |
| pre-miRNA | precursor miRNA | | |
| PTBP1 | polypyrimidine tract-binding protein 1 | | |
| PRG4 | proteoglycan 4 | | |
| RBP | RNA binding protein | | |



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Adenylyl cyclase isoforms 5 and 6 in the cardiovascular system: complex regulation and divergent roles

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Adenylyl cyclases (ACs) are crucial effector enzymes that transduce divergent signals from upstream receptor pathways and are responsible for catalyzing the conversion of ATP to cAMP. The ten AC isoforms are categorized into four main groups; the class III or calcium-inhibited family of ACs comprises AC5 and AC6. These enzymes are very closely related in structure and have a paucity of selective activators or inhibitors, making it difficult to distinguish them experimentally. AC5 and AC6 are highly expressed in the heart and vasculature, as well as the spinal cord and brain; AC6 is also abundant in the lungs, kidney, and liver. However, while AC5 and AC6 have similar expression patterns with some redundant functions, they have distinct physiological roles due to differing regulation and cAMP signaling compartmentation. AC5 is critical in cardiac and vascular function; AC6 is a key effector of vasodilatory pathways in vascular myocytes and is enriched in fetal/neonatal tissues. Expression of both AC5 and AC6 decreases in heart failure; however, AC5 disruption is cardio-protective, while overexpression of AC6 rescues cardiac function in cardiac injury. This is a comprehensive review of the complex regulation of AC5 and AC6 in the cardiovascular system, highlighting overexpression and knockout studies as well as transgenic models illuminating each enzyme and focusing on post-translational modifications that regulate their cellular localization and biological functions. We also describe pharmacological challenges in the design of isoform-selective activators or inhibitors for AC5 and AC6, which may be relevant to developing new therapeutic approaches for several cardiovascular diseases.

KEYWORDS

adenylyl cyclase, G protein-coupled receptors, cyclic 3',5'-adenosine monophosphate, signal transduction, heart disease, drug discovery

Introduction

Over the course of life, heart health and aging have been correlated with diminished left ventricular (LV) function and weakened cardiac β -adrenergic receptor (β AR) responsiveness. The reasons for this dysfunctionality may include phenotype changes in the LV, reduced β AR density, upregulation of regulatory proteins Gai and G-protein coupled receptor kinases, or abnormalities in the β AR signaling system (Ferrara et al., 2014). Adenylyl cyclases (ACs), as the central effector molecules for β AR signaling, play a crucial

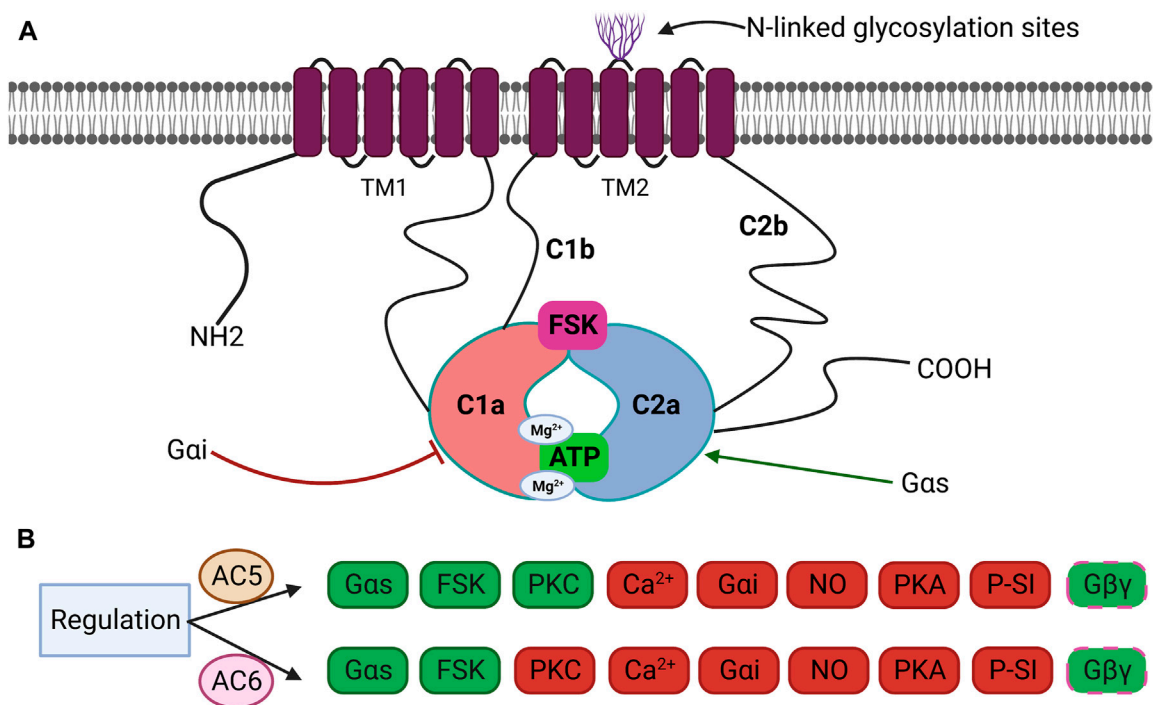


FIGURE 1

(A) Schematic structure of transmembrane adenylyl cyclases (TMACs). TMACs contain an intracellular N-terminus, two repetitions of six TM1 and TM2, and two cytoplasmic domains (C1 and C2) further divided into C1a and C2a. C1a–C2a forms the catalytic domain and FSK-binding site. Inhibitor Gai binds to C1a, whereas activator Gas binds to C2a. C1b and C2b are regulatory subdomains, and the N-terminus participates in several protein–protein interactions. Some isoforms are glycosylated on extracellular loops 5 or 6. (B) Specific regulation of AC5 and AC6 by various effectors. Both isoforms are fully activated by Gas and FSK and partially or conditionally activated by Gβγ. AC5 is activated by PKC, whereas AC6 is inhibited by PKC. Both isoforms are inhibited by Ca²⁺, Gai, NO, PKA, and P-site inhibitors (P-SI). Green: stimulation, red: inhibition, and dashed line: conditionally.

role in cardiac contractility, relaxation, and LV diastolic function (Tang et al., 2011). The failing heart has poor AC signaling and decreased LV cAMP production, leading to impaired βAR responsiveness to ligands (White et al., 1994; Roth et al., 2002). Persistent activation of the sympatho-adrenergic system in patients with congestive heart failure can also lead to unfavorable cardiac remodeling due to cardiomyocyte loss and fibrotic replacement (Zhang et al., 2013). The distribution of AC isoforms varies within cardiac tissues; ACs 2, 3, 4, 5, 6, and 7 are expressed by cardiac fibroblasts (Ostrom et al., 2003), while AC1 and AC8 are found in sinoatrial node (SAN) cells (Mattick et al., 2007; Younes et al., 2008; Robinson et al., 2021), and AC5 and AC6 are the major AC isoforms expressed in the adult ventricle (Göttle et al., 2009; Efendiev and Dessauer, 2011). AC6 also serves as the principal effector of vasodilator signaling and a regulator of membrane potential in vascular myocytes (Nelson et al., 2011). In this review, we examine the similarities and differences between ACs 5 and 6 in cardiovascular function, highlighting their divergent functional roles and opportunities for pharmacological targeting.

Overview of adenylyl cyclases

Cyclic 3',5'-adenosine monophosphate (cAMP), a ubiquitous second messenger that mediates a variety of cellular responses, is generated by more than thousand nucleotidyl cyclase proteins

classified into six groups according to the amino acid sequence of the catalytic domain (Kamenetsky et al., 2006; Seifert and Beste, 2012). The class III cyclases including eukaryotic adenylyl cyclases (ACs) are indispensable effectors of cAMP (Linder, 2006). Enumerated in order of discovery, mammalian ACs 1–9 (~120–140 kDa) have been identified as transmembrane adenylyl cyclases (TMACs), while AC10 is a soluble enzyme. ACs are canonically regulated by heterotrimeric G-proteins, Gai and Gas, upon stimulation of G protein-coupled receptors (GPCRs) (Halls and Cooper, 2017). There are significant variations in the distribution and biochemical characteristics among AC isoforms, as well as distinctive chromosomal loci of individual isoforms (Vatner et al., 2013). The TMACs share a similar framework but different regulation (Sunahara et al., 1996). Despite arising from distinct transcripts, they own a similar overall structure with highly homologous active sites, making it challenging to design selective activators or inhibitors and to differentiate their functions in the organs and tissues (Rana et al., 2017). The mammalian ACs share an intracellular N-terminus, two membrane-spanning domains (TM₁ and TM₂), and conserved catalytic domains, C1 and C2 (each ~40 kDa), together forming the catalytic core. The C1 and C2 cytoplasmic domains are further divided into the highly conserved catalytic regions, C1a and C2a, and the less conserved regulatory regions, C1b and C2b. Two active sites are shaped by their interface: the ATP catalytic site and forskolin (FSK) binding pocket (Figure 1A)

(Schmid et al., 2014; Antoni, 2020). Crystallographic studies have revealed that the hydrophobic pocket generated by C1 and C2 catalytic subunits is the allosteric site for the interaction of FSK with ACs (Pavan et al., 2009; Bhatia et al., 2023). Despite the sequence similarity of the regulatory domains in C1–C2, various signals such as G-proteins, kinases, FSK, and Ca^{2+} uniquely regulate AC isoforms (Brand et al., 2013). Indeed, the C1–C2 domain, at the cytoplasmic surface of the molecule, accommodates binding sites for Gas, Gai, FSK, ATP, Mg^{2+} , the regulator of G protein signaling (RGAS2), proteins associated with Myc (PAM), Snapin, Ric8a, A-kinase-anchoring protein (AKAP79), PH domain leucine-rich protein phosphatase 2 (PHLPP2), and phosphorylation and dephosphorylation sites for protein kinase A (PKA) and protein kinase C (PKC) (Sunahara et al., 1997). The crystal structures of the complex made up of the C1 cyclase homology domain (CHD) from AC5 and the C2 CHD from AC2 ($\text{VC}_1\text{:IIC}_2$) bound to FSK and GTP-S-activated Gas served as the fundamental model for the prototype AC catalytic core (Mou et al., 2009). Binding sites for cofactors Mg^{2+} and Mn^{2+} are further located in the active site of AC and must be occupied for catalytic activity, based on structural analyses of activated $\text{VC}_1\text{:IIC}_2$ complexes coupled to substrate analogs (Sinha and Sprang, 2006; Mou et al., 2009). Forskolin, a natural diterpene, specifically binds opposite to the ATP-binding site within the catalytic core, using a mixture of hydrophobic and hydrogen interactions to approximate the two cytoplasmic domains, resulting in increased enzyme activity regardless of Gas docking. No endogenous ligand has yet been identified for the FSK-binding site (Pavan et al., 2009; Dessauer et al., 2017; Bhatia et al., 2023).

While the C1–C2 framing the catalytic core remains highly conserved among individual AC isoforms, the N-terminus is significantly varied across isoforms (Antoni, 2020). The length and composition of the N- and C-terminal domains of ACs are significantly variable outside of the non-conserved domains (Sunahara et al., 1996). The N-terminus of AC1 to AC5 spans 60 to 240 residues in length. Pharmacologically, these are the regions where type-specific regulation of ACs can occur (Halls and Cooper, 2017). The TM domains are not required for catalytic activity since C1a and C2a can be individually shaped into active enzymes by Gas and FSK (Tang and Gilman, 1995). Removing TM domains and fusing just C1 with C2 make a small but versatile protein which can be used to replicate numerous beneficial effects of AC (Tan et al., 2019). However, the TM regions play important roles in balancing the stoichiometry of their relative catalytic domains (Sunahara et al., 1996), in regulating functional assembly and trafficking of AC (Gu et al., 2002), and may also function as receptors for extracellular signals (Seth et al., 2020).

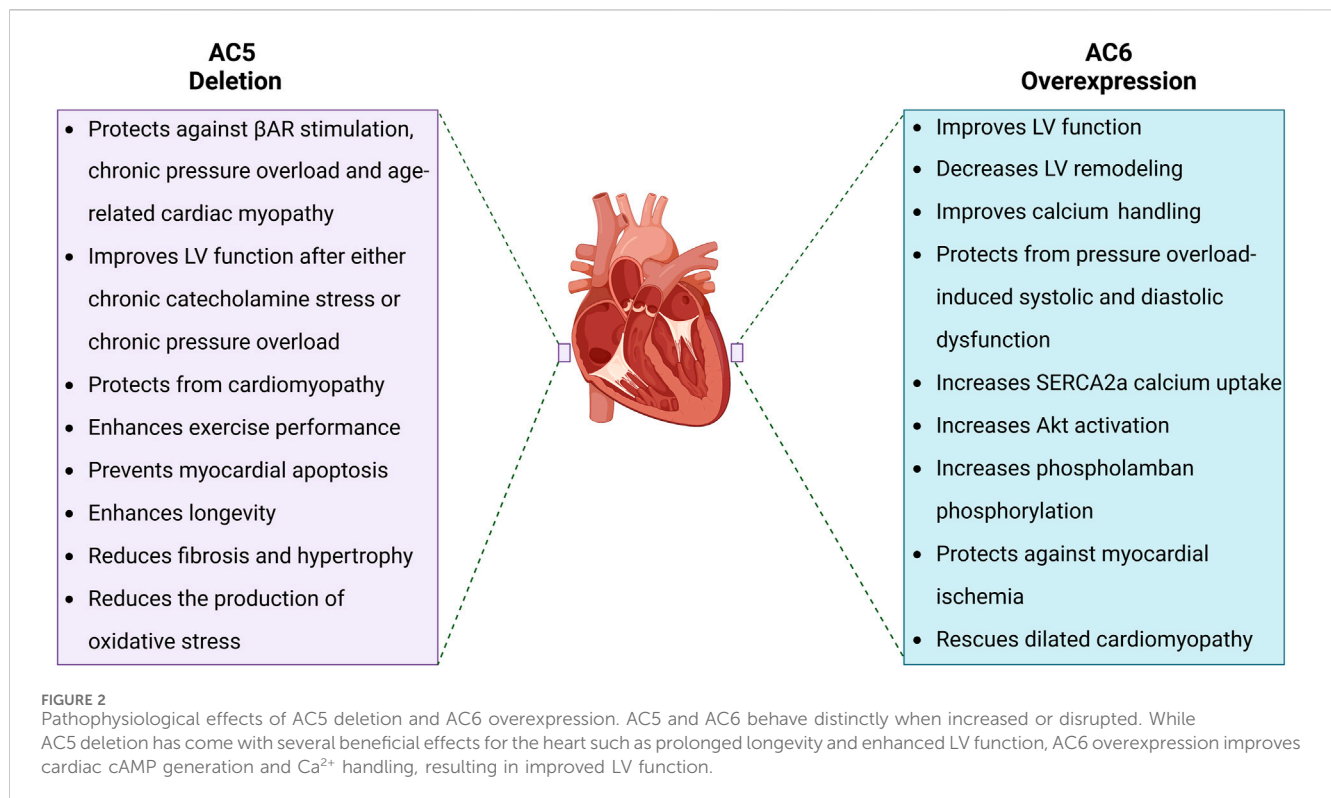
From a regulatory standpoint, the mammalian AC isoforms are classified into four groups: *Group I*, ACs 1, 3, and 8, stimulated by calcium/calmodulin ($\text{Ca}^{2+}/\text{CaM}$); *Group II*, ACs 2, 4, and 7, stimulated by $\text{G}\beta\gamma$; *Group III*, ACs 5 and 6, inhibited by Gai/ Ca^{2+} ; and *Group IV*, AC9, which is partially stimulated by FSK (Ostrom et al., 2022). All isoforms are activated by Gas while differentially regulated by Gai, $\text{G}\beta\gamma$, and protein kinases (PKA, PKC, Raf-1, and CaM kinases) (Figure 1B) (Antoni, 2020). Most cells and tissues express multiple AC isoforms; however, their

varying abundance in specific tissues is noteworthy (Halls and Cooper, 2017; Ostrom et al., 2022).

Role of ACs in cardiac tissue

Key cardiac isoforms AC5 and AC6, known as the Ca^{2+} -inhibited family of ACs, share about 91.5% sequence alignment, except for the AC6 N-terminus (aa 1–86) (Wu et al., 2017). These two isoforms have considerable similarities in their expression patterns and functions, despite their evolution at independent gene loci; the AC5 gene is on chromosome 3 at position 3q13.2–3q21, and AC6 is on chromosome 12 at position 12q12–12q13 (Haber et al., 1994). The canine (type V-a) and the rat (type V-b) forms of AC5 are in fact mRNA splice variants, co-expressed in both species (Iwami et al., 1995). Both AC5 and AC6 are non-competitively inhibited by Gai at C1a, opposite from the docking site for Gas at C2a (Taussig et al., 1994). Their tissue distribution and developmental mRNA expression patterns have been studied in several species, including humans (Tobise et al., 1994; Espinasse et al., 1995; Wang and Brown, 2004). AC6 is more expressed in the neonatal heart, while AC5 appears to be predominantly an adult isoform (Espinasse et al., 1995). The maximal amount of AC6 mRNA is expressed during fetal development, while steadily diminishing with age, and reaching its lowest amount in fully mature adults. On the contrary, a minimal amount of AC5 mRNA is expressed during fetal development, but progressively increasing with age, and reaching its highest point in the fully developed adult (Tobise et al., 1994; Okumura et al., 2009). In the mouse heart, the abundance of AC5 protein reaches its highest point before 1 week and then decreases to levels similar to those of mice studied at 3, 6, and 24 months of age. In the rat heart, however, the AC5 protein level peaks before 2 weeks of age and subsequently decreases by 3 months. In pigs, cardiac AC5 protein abundance is high in 1-day-old hearts and then diminishes with age, reaching its lowest content in the heart at 5–6 months (Hu et al., 2009). Feedback regulation of AC expression also varies as a function of age; PDE3 inhibition (PDE3i) has been found to alter AC expression levels in adult and pediatric subjects with dilated cardiomyopathy (DCM) and in pediatric subjects with a congenital single right ventricle. These groups demonstrated distinctive AC isoform mRNA expression patterns. Compared to the non-failing adult LV myocardium, the adult DCM myocardium exhibits upregulation of AC6; chronic PDE3i treatment enhances mRNA expression of AC5 and AC6, while all other isoforms are expressed at levels comparable to those of DCM patients not receiving PDE3i treatment. In contrast, the non-failing pediatric right ventricle expresses less AC5 than does the pediatric single ventricle; and in pediatric DCM, AC5 and AC6 remained unchanged by PDE3i (Nakano et al., 2017). Due to their structural homology and the paucity of type-specific antibodies, definitive levels of protein expression in the heart have been difficult to establish (Hu et al., 2009). Even following the deletion of AC5, 60% of immunodetectable AC5/6 was observed in cardiac myocytes, suggesting some degree of epitope overlap (Okumura et al., 2003b).

To distinguish their specific roles in cardiac function, AC5 and AC6 have been the subject of several transgenic overexpression and deletion studies. Both isoforms are highly expressed in the heart and



are important negative feedback responses of cardiac rhythmicity (Cosson et al., 2019). While both isoforms regulate heart rate (HR) and contractility, AC6 appears more important at baseline cardiac function (Vatner et al., 2013). AC5 and AC6 may function distinctly in the pathogenesis of cardiac stress responses. The protein abundance of AC5 and AC6 responds oppositely to pressure overload LV hypertrophy (Okumura et al., 2003b; Sugano et al., 2011), with upregulation of AC5 and downregulation of AC6, implying that it is AC5 that plays a role in chronic pressure overload cardiomyopathy (Vatner et al., 2013). It is probable that these two isoforms also play different physiological roles in regulating cardiac function during dilated cardiomyopathy. Both AC5 and AC6 mRNAs appear decreased in dogs with pacing-induced Congestive Heart Failure (CHF) (Ishikawa et al., 1994); while in a swine model of tachypacing and severe heart failure, AC6 but not AC5 is downregulated (Ping et al., 1997). While some studies suggest AC5 overexpression improves cardiac function in transgenic mice during exercise (Esposito et al., 2008), deletion of AC5 protects the heart in most models of cardiac stress including heart failure; in contrast, AC6 overexpression has been correlated with cardioprotective effects (Wu et al., 2017; Ostrom et al., 2022). The overexpression of AC6 leads to increased LV function, improved cAMP and Ca^{2+} handling, and may protect the heart from pressure overload-induced systolic and diastolic dysfunction (Phan et al., 2007; Guellich et al., 2010). AC5 deletion, on the other hand, results in protecting the heart from cardiomyopathy, chronic catecholamine stress, and chronic pressure overload (Okumura et al., 2003b; Okumura et al., 2007). Isoform-specific effects are not restricted to myocardial AC5 and AC6. Cardiac-specific overexpression of AC8 in 3-month-old transgenic mice alters several pathways, resulting in elevated AC activity and cAMP-

induced cardiac workload until up to 1 year of age, without excess mortality or onset of heart failure (Tarasov et al., 2022; Qu et al., 2024). However, other studies have shown cardiac overexpression of AC8 to cause early and accelerated cardiac remodeling, resulting in development of heart failure and a shortened life span, suggesting that alterations in cAMP/PKA signaling can hasten cardiac aging, in part via the glycogen-synthase-kinase $3\alpha/\beta$ (GSK3 α/β) phosphorylation pathway (Mougenot et al., 2019).

Although there remains some controversy regarding the specific regulation and roles of AC5 and AC6 isoforms in controlling cardiac function, their roles in the production of required cAMP to initiate cardiac chronotropic and inotropic responses are indispensable. It is thus important to differentiate the regulation and biological functions of AC5 *versus* AC6, leading the way to development of targeted treatments for different cardiac pathologies. The respective therapeutic outcomes of AC5 deletion and AC6 overexpression in the cardiovascular system are included in Figure 2.

Regulation of AC5 and AC6 by calcium

Ca^{2+} serves as a pivotal regulator of AC5 and AC6, playing a crucial role in the control of cellular homeostasis and heart function regulation. The inhibition of ACs 5 and 6 by Ca^{2+} , which is non-competitive with respect to ATP (Guillou et al., 1999), contributes to pacemaking in cardiac tissue and also sustains endothelial cell permeability. Inhibition of these cardiac AC isoforms by Ca^{2+} , which their own cAMP generation permits to enter the cardiomyocyte, serves as a form of feedback control regulating pacemaker rhythmicity as well as the force of contraction (Mou

et al., 2009). A wide range of *in vitro* inhibitory sensitivities are reported. Systolic $[Ca^{2+}]$ 1 μM is required to inhibit the activity of AC5/6, while at higher intracellular $[Ca^{2+}]$ (10–25 μM), the activity of all AC isoforms is inhibited (Guillou et al., 1999; Halls and Cooper, 2017). However, AC5 and AC6 inhibition by Ca^{2+} is biphasic, involving low- and high-affinity binding sites, so Ca^{2+} can inhibit AC5 and AC6 in sub- to supramicromolar concentration ranges (Steer and Levitzki, 1975; HANSKI et al., 1977; OLDHAM et al., 1984; Fagan et al., 1998; Guillou et al., 1999). The sub-micromolar $[Ca^{2+}]$ (0.2–0.6 μM) inhibits AC5 and AC6 in membrane preparations from different tissues, cultured cell lines, and in recombinant systems (Beazely and Watts, 2006). Mechanistically, Ca^{2+} antagonizes the activation of AC5 and AC6 by Mg^{2+} (Mou et al., 2009). Site-directed mutations of AC5 in the presence of other divalent cations indicate that the Mg^{2+} -binding loci at the AC5 catalytic site are essential for its Ca^{2+} -mediated inhibition (Hu et al., 2002; Mou et al., 2009). In addition to being a hub for GPCR AC interactions, lipid rafts are locations for capacitive Ca^{2+} entry channels; Group III AC isoforms that are localized in lipid rafts are also regulated by Ca^{2+} (Ostrom and Insel, 2004). Disruption of lipid rafts results in loss of capacitive Ca^{2+} entry and dysregulation of Ca^{2+} -regulated AC isoforms (Dessauer et al., 2017).

Subcellular localization of AC5 and AC6

GPCR signaling pathways are type-specific for either AC5 or AC6, as determined by the colocalization of receptors and AC isoforms in lipid raft or non-raft plasma membranes (Ostrom et al., 2000). ACs 4–7 are expressed in the heart and vascular system, but among them, AC5 and AC6 are dominant subtypes in adult mammalian heart (Dessauer et al., 2017). The compartmentation of AC5 and AC6 in caveolae or membrane lipid rafts provides a key biochemical process for temporal and spatial segregation of signal transduction as well as cross-talk between signaling cascades, resulting in the compartmentalization of cAMP signaling (Thangavel et al., 2009). While native AC5/6 are found in caveolar fractions, overexpressed AC6 in rat aortic smooth muscle cells is found in non-caveolin-rich fractions, where it is functional and increases cAMP (Ostrom et al., 2002). In these cells, low levels of AC6 overexpression do not much change cAMP levels at baseline nor responses to adenosine A2b receptor challenge (Ostrom et al., 2002; Thangavel et al., 2009). In contrast, in cardiomyocytes, overexpressed AC6 localizes similarly to endogenous AC6, which is targeted to caveolae, demonstrating that GPCR and AC5/6 compartmentation to caveolin-rich membranes is cell-type dependent (Ostrom et al., 2000; Ostrom et al., 2001; Ostrom et al., 2002). Overexpression of AC6 improves β AR responses without affecting signaling of other Gas-coupled receptors in a variety of cells, including airway smooth muscle, lung fibroblasts, and neonatal myocytes (Sunahara et al., 1996; Liu et al., 2008; Sadana and Dessauer, 2009; Dessauer et al., 2017). In vascular smooth muscle cells (VSMCs), AC3, AC5, and AC6 are the most abundant isoforms in β AR-mediated signaling, but during vasodilation, AC6 is the principal isoform involved in β AR-mediated cAMP/PKA signaling and activation of the ATP-sensitive potassium current, thus playing an important role in setting the membrane potential. Its counterpart AC5 does not have similar activity, (Nelson et al., 2011). In ventricular myocytes, isoforms

AC5 and AC6 have separate subcellular compartmentalization. While AC5 is mostly found deep in transverse tubules interacting with caveolin-3 (CAV3) and phosphodiesterases (PDEs), AC6 is located in the plasma membrane outside the t-tubular area (Figure 3) (Timofeyev et al., 2013). β AR receptors are also differentially distributed; β 1- and β 2AR are found within t-tubules, but β 1AR (comprising >70% of cardiac β AR) is found on the sarcolemma outside t-tubules (Brodde, 1991). β 1AR triggers cAMP generation, which increases PKA-mediated phosphorylation of L-type Ca^{2+} channels as well as other regulatory proteins, greatly increasing cardiac contractility; β 2AR couples with both Gas and Gai, resulting in a smaller increment of contractility (Zheng et al., 2005). Inactivation of Gai by pertussis toxin enhances β 1AR-mediated inotropy despite β 1AR not coupling to Gai, indicating that Gai also inhibits the AC5/6 receptor independently (Cosson et al., 2019). Due to subcellular localization, AC5 has been connected to β 2AR signaling, while AC6 is responsible for extra-tubular β 1AR signaling as well as β 1AR-mediated augmentation of the L-type Ca^{2+} channel current ($I_{Ca,L}$) in ventricular myocytes (Timofeyev et al., 2013) (Figure 3). Both β AR subtypes have obvious inotropic effects; dobutamine, a non-selective β AR agonist, increases cardiac contractility in both AC5KO and AC6KO mice (Tang et al., 2006; Tang et al., 2008). While β 2AR overexpression enhances ventricular function and activates cell survival pathways (Milano et al., 1994), β 1AR overexpression appears catastrophic, causing both cardiac hypertrophy and dilated cardiomyopathy (Engelhardt et al., 1999). The opposite effects have been reported following manipulation of their more proximal AC subtypes; AC5 disruption in t-tubules is cardioprotective (Okumura et al., 2003b), while overexpression of AC6 rescues heart function in cardiac injury (Gao et al., 2016). However, functional coupling may not only reflect localization. In AC6KO, a shift from dominant AC6 coupling to AC5 coupling occurs in the β 1AR signaling cascade. This change in the AC assignment in AC6KO results in rearranged signaling compartmentalization as well as an alteration in the PDE isoform control of the cAMP pool (Cosson et al., 2019; Ostrom et al., 2022). These findings suggest some functional redundancy of AC5 or AC6 for β 1AR-mediated inotropic responses. Additionally, in CHF models largely ascribed to excess β 1AR activity, β 2AR redistributes to the cell surface, and this loss of cAMP compartmentation correlates with heart failure (Nikolaev et al., 2010). Studies of detubulated cardiomyocytes reveal that stimulation of both β 1AR and β 2AR is more functionally effective at the sarcolemmal surface, rather than within t-tubules (Cros and Brette, 2013). Pharmacological inhibition studies confirm that β 1AR preferentially couples with AC5 at t-tubules, while β 2AR couples with AC6 when present at the cell surface, resulting in the observed AC subtype-specific effects on cardiac function and survival signaling (Tsunematsu et al., 2015).

Compartmentalization of cAMP signaling in the heart

While most tissues express a plethora of AC isoforms (Figure 4), the specific roles of each isoform may overlap within that cell type (Sadana and Dessauer, 2009). Functional redundancy or compensatory roles have been proposed within individual AC groups, as they are regulated or expressed in a comparable manner (Defer et al., 2000). Changes in AC activity may depend

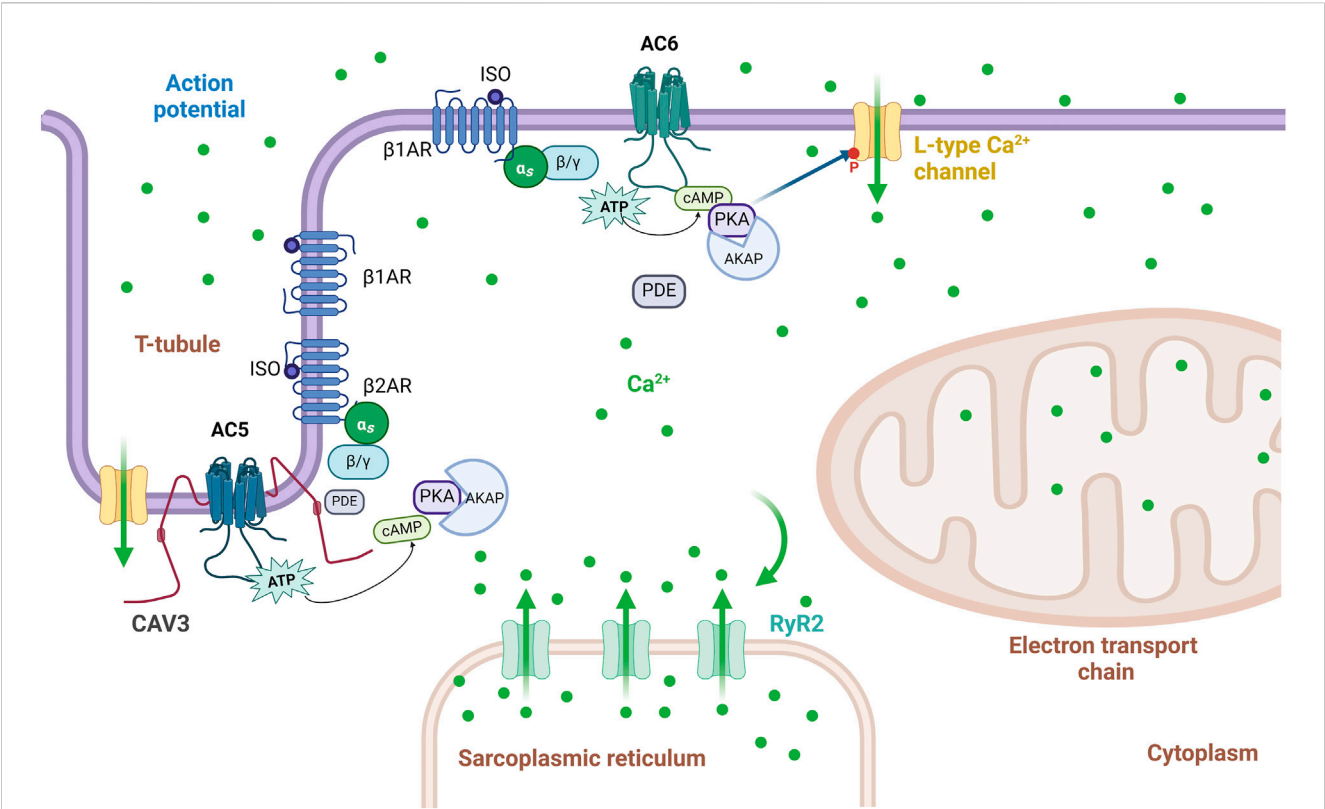


FIGURE 3 Schematic representation of the localization of AC5 and AC6 signaling within the t-tubule and sarcolemmal membrane. In ventricular myocytes, AC6 is localized to the plasma membrane outside of the t-tubular region, and it interacts only with β1AR signaling-mediated augmentation of the L-type Ca²⁺ current (ICa,L), while AC5 is localized to the membrane in t-tubular regions, and its function on ICa,L is restrained by cAMP degradation by phosphodiesterases. Adapted from (Timofeyev et al., 2013); image designed using BioRender.

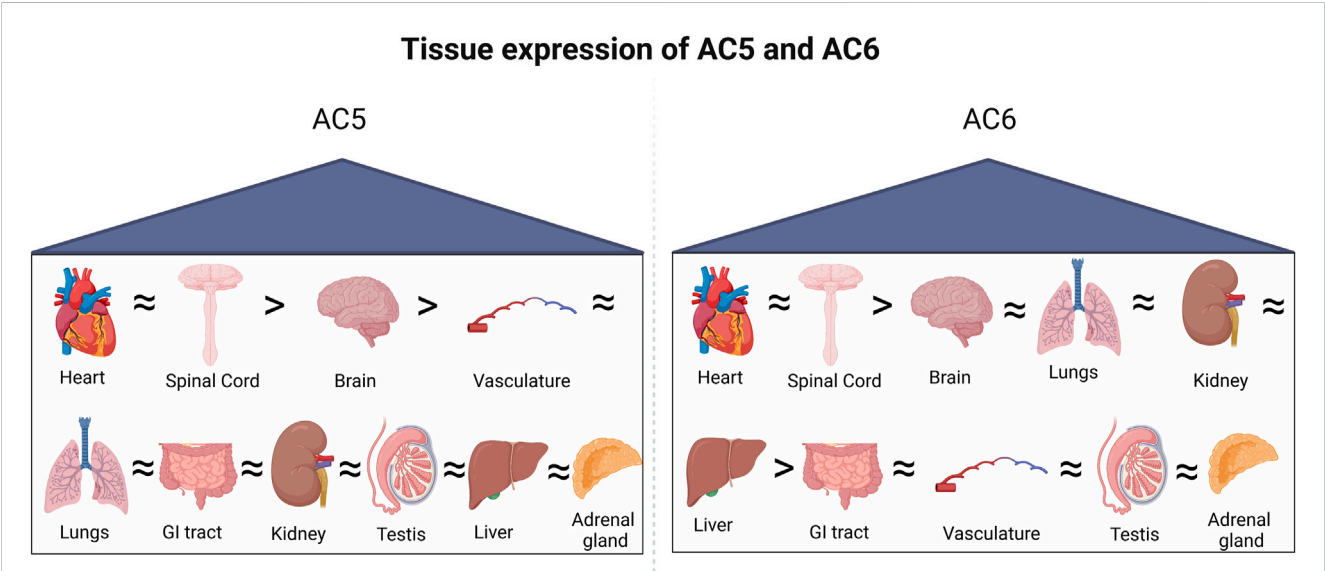


FIGURE 4 Tissue-specific expression of AC5 and AC6. Both isoforms are highly expressed in the heart and spinal cord, followed by the brain.

on the profile of isoforms present in a certain cell or tissue (Tao et al., 1998). Differing phenotypes of AC5 and AC6 activity suggest the maintenance of distinct pools of cAMP within cardiac myocytes, generated by each enzyme. Regulation and localization of AC5 and AC6 isoforms tailor the generation of cAMP by concurrent signals, assisting in dynamic control of the cAMP signal. The

compartmentation of cAMP signaling in the heart arises, in part, from specific signaling complexes which generate distinct cell responses, with AC5 and AC6 being central effectors to the formation and maintenance of these compartments.

The organization of cAMP microdomains in isolated cardiac cells has been researched extensively, using various effector proteins to sort distinct cAMP pools (Patel and Gold, 2015). Several downstream effectors can regulate cAMP production or hydrolysis, including PDEs, PKA, cyclic nucleotide-gated (CNG) channels, hyperpolarization-activated cyclic nucleotide-gated exchange proteins, exchange protein directly activated by cAMP (Epac), and the Popeye domain-containing (POPDC) protein family. At the level of cAMP signal longevity, PDE isoforms, importantly PDE3/4, play pivotal roles in the discrete regulation of localized cAMP levels in neonatal cardiac myocytes (Willoughby and Cooper, 2007). PDE4 isoforms regulate cAMP signaling resulting from β 1AR and β 2AR stimulation in cardiac myocytes; PDE4B controls β 1AR but not β 2AR responses in cardiac myocytes, suggesting a localized function to control β 1AR-dependent excitation–contraction coupling (Mika et al., 2014). PDE3 acts globally on cAMP in normal and heart failure models, modulating cAMP-mediated regulation of Ca^{2+} re-uptake in the sarcoplasmic reticulum (SR) (Calamera et al., 2022). Spatial and temporal organization of cAMP-dependent pathways is further provided by cAMP-related protein scaffolding, biasing the cAMP signal toward different end functions.

AKAP organizes AC5 and AC6 cAMP in the heart

A-kinase-anchoring proteins (AKAPs) are scaffold proteins which coordinate signaling components into multiprotein complexes, giving rise to regulation of the spatial and temporal organization and fine-tuning of cAMP signaling (Kritzer et al., 2012), ensuring correct targeting of cAMP-dependent PKA and other signaling enzymes to precise subcellular compartments (Diviani et al., 2016). AKAPs function through binding to the regulatory subunits of PKA and crucial PKA phosphorylation sites to initiate rapid and targeted coupling of the kinase to downstream effectors (Ostrom et al., 2022). AC5/6 in the heart utilizes the interactions with AKAPs to cohort with upstream and/or downstream effectors. Over 50 AKAPs are known to target PKA to various cellular locations; important cardiovascular AKAPs include AKAP15/18, AKAP79/150, Yotiao, mAKAP, AKAP-Lbc, and Gravin (Baldwin and Dessauer, 2018). The interaction of these AKAPs with AC5 or AC6 determines the localization and activity of their cAMP signaling complexes (Efendiev and Dessauer, 2011). As scaffolding effectors, AKAPs not only anchor the localization of ACs but also preserve local pools of cAMP by assembling macromolecular complexes (Baldwin and Dessauer, 2018). In cardiomyocytes, the arrangement of MAP kinase, Ca^{2+} , and cAMP-dependent pathways is coordinated through the muscle A-kinase anchoring protein β , playing a key role in cellular hypertrophy (Kapiloff et al., 2009). AC5 binds to a specific N-terminal site on mAKAP-(245–340), resulting in impairment of AC5 activity; interruption of mAKAP β –AC5 complexes could be targeted therapeutically to reduce cAMP generation and

hypertrophy in cardiac myocytes (Bauman et al., 2006; Kapiloff et al., 2009). Both AC5 and AC6 interact with AKAP79/150 along with PKA, generating a negative feedback loop where cAMP production is impaired by PKA phosphorylation of AC5/6 (Baldwin and Dessauer, 2018).

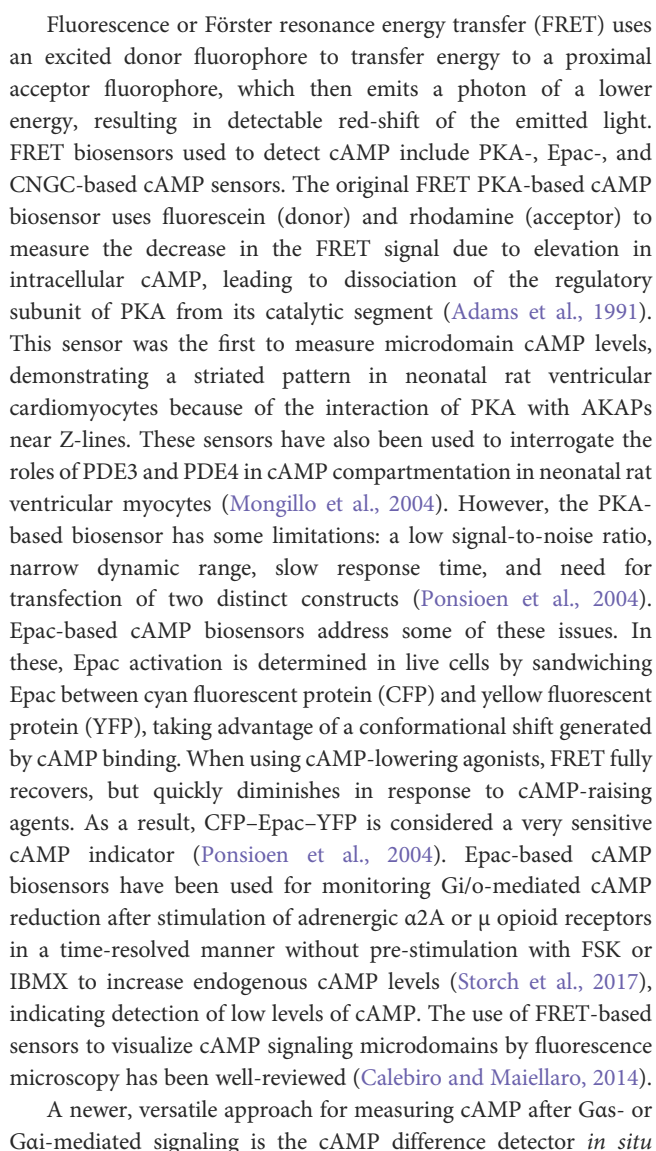
AC5 and AC6 cAMP regulation by Epac in the heart

Another downstream cAMP effector is Epac, a family of cAMP-regulated guanine nucleotide exchange factors functioning independently of PKA as mediators of cAMP signaling. While considered less important in cardiomyocytes than PKA, Epac has high affinity to bind to cAMP, resulting in activation of small GTPases Rap1 and Rap2 (Tan et al., 2022). Epac1 and Epac2 play roles in cardiovascular Ca^{2+} signaling and vascular endothelial barrier formation (Lezoualc'h et al., 2016). AC5 transgenic mice with knockout of the Epac1 gene had decreased cardiac dysfunction and were less susceptible to pacing-induced atrial fibrillation after chronic isoproterenol infusion compared to controls while evincing less cardiac apoptosis and fibrosis than the AC5 transgenics, suggesting that Epac1 mediates the deleterious effects of AC5 overexpression on cardiac function and rhythmogenesis (Cai et al., 2016). On the other hand, mice lacking Epac1 are protected against pressure overload or chronic catecholamine stress-induced cardiac dysfunction (Fujita et al., 2017), reinforcing the notion that rate- and pressure-induced cardiac failure may arise from separate cAMP pools.

Advances in cAMP biosensors in live cells

Various constructs have been developed to measure real-time cAMP levels in living cells or tissues. Previous biochemical methods or radioligand tools were not able to precisely measure the spatiotemporal resolution of intercellular cAMP fluctuations; however, several fluorescent or luminescent biosensors have been introduced that can track real-time cAMP levels in living cells. These cAMP sensors have the advantage of facilitating the study of complex, compartment-specific cAMP-dependent responses (Warrier et al., 2005).

Classical methods of determining AC activity have involved detection of radiolabeled substrate ATP conversion to AMP and detection of enzyme reaction products by chromatography (Streeto and Reddy, 1967; Krishna et al., 1968); these methods can be applied to cell or tissue lysates. Loss of ATP-bound lanthanide fluorescence has been used as an indirect but specific indicator of AC ATPase activity (Spangler et al., 2008b). The terbium–norfloxacin AC activity assay measures the substrate turnover as an assessment of AC catalytic function; varying the exogenous ATP substrate concentration permits the calculation of Michaelis–Menten kinetics (Spangler et al., 2008a). In contrast, intact cell cAMP assays are cumulative measures of product formation in the absence of exogenous substrates, reflecting real-time steady state inclusive of cAMP generation, degradation, or export. Genetically encoded cAMP sensors can also be targeted to subcellular location.



BRET biosensors (bioluminescence resonance energy transfer) use a donor luciferase to oxidize luciferin-generating bioluminescence that excites the acceptor, emitting light at an extended wavelength (Wu and Jiang, 2022). BRET enables detection of fusion protein interactions directly, without the need of an external light source to stimulate a fluorescent energy donor (Prinz et al., 2006). This technology further addresses the problems of photobleaching, autofluorescence, and signal-to-noise ratio in FRET methods; the drawbacks include difficulty of substrate delivery and stoichiometry and potential for cytotoxicity (Wu and Jiang, 2022). BRET-based Epac1-, PKA-, and nano lantern-based cAMP sensors have been developed to investigate cAMP generation (Masri et al., 2008; Hunter et al., 2017; Valkovic et al., 2018).

All plasmid-based cAMP sensors have advantages and disadvantages, with respect to ease of introduction without

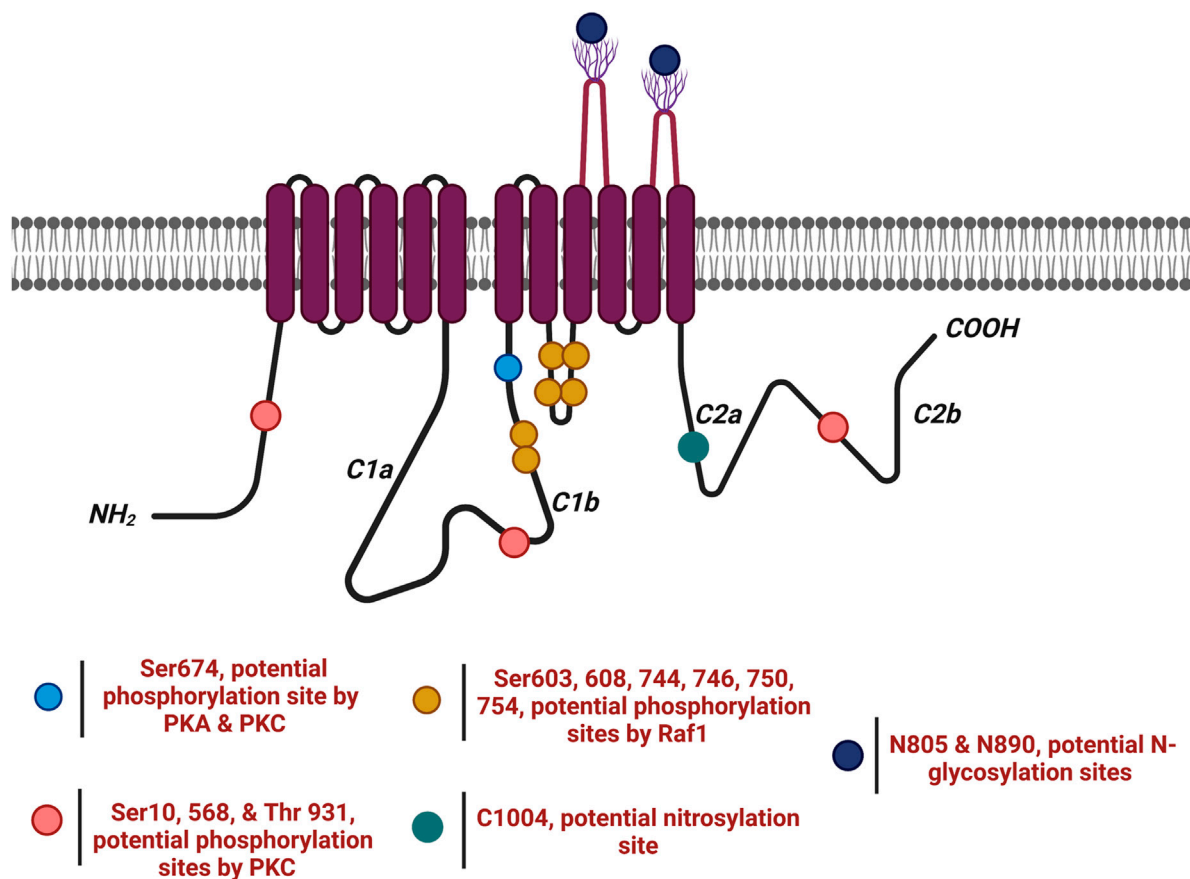


FIGURE 6

AC6 undergoes several post-translational modifications (●). AC6 is phosphorylated by PKA and PKC at Ser674 (●); phosphorylated by PKC at Ser10 and 568 and Thr931 (●); S-nitrosylated by NO at C1004 (●); Raf1 on Ser603, 608, 744, 746, 750, and 754 (●); and glycosylated at N805 and N890 on extracellular loops 5 and 6.

toxicity, targetability of expression in specific subcellular locations, temporal and spatial resolution of cAMP detection, cAMP detection ranges, and buffering from metabolic processes or post-translational modifications in living cells or tissues. Rapid cAMP binding, instant transmission of signals, and rapid reversibility of the binding event would denote an ideal cAMP sensor (Paramonov et al., 2015). Further optimization is needed for detection of low unstimulated cAMP levels in sensitive primary cells or tissues. Available cAMP biosensors have recently been reviewed in detail (Kim et al., 2021).

Regulatory post-translational modifications of AC5 and AC6

AC5/6 phosphorylation by PKA and PKC

PKA and PKC modify AC5 and AC6 activity by phosphorylating serine (Ser) or threonine (Thr) residues. In the heart, PKA regulates metabolism, gene transcription, ion fluxes, and contraction (Colombe and Pidoux, 2021) and also acts as a feedback inhibitor for both AC5 and AC6 through inhibitory phosphorylation near the end of the C1b domain, resulting in

the desensitization of AC activity (Sadana and Dessauer, 2009). Stimulation of AC6 was lost after treatment of PKA even in the presence of high concentrations of active Gas (Chen et al., 1997). Mutational analysis indicates Ser674 is the target of phosphorylation and inhibition of AC6 by PKA in intact cells (Figure 6) (Beazely and Watts, 2006; Chen et al., 1997). AC5 possesses 14 putative PKA phosphorylation sites, including a Ser 788 that corresponds to Ser674 in AC6; however, a specific PKA phosphorylation site in AC5 has not been confirmed (Iwami et al., 1995; Chen et al., 1997).

While both AC5 and AC6 isoforms are inhibited through phosphorylation by PKA, PKC inhibits only AC6 (Kamide et al., 2015; Liu et al., 2022). Previous studies have shown that AC2, AC3, and AC5 can be stimulated by PKC, while AC6 activity is inhibited (Lai et al., 1997; Defer et al., 2000). Phosphorylation of AC5 by PKCα or PKCζ enhances basal activity as well as FSK- or Gas-stimulated cAMP accumulation; PKC-ζ phosphorylation of AC5 results in a 20-fold increase in AC activity (Kawabe et al., 1996). In the heart, generation of phosphatidylinositol-3,4,5 triphosphate via hormonal or growth factor activation of phosphatidylinositol 3-kinase can activate PKC-ζ, thus directly activating AC5 production of cAMP (Hanoune and Defer, 2001). In contrast, PKC activators have either no effect or inhibitory effect

on AC6 activity (Chen and Iyengar, 1993; Jacobowitz et al., 1993; Lai et al., 1997; Lai et al., 1999). Mutational analysis of the N-terminus of AC6 as a regulatory domain showed that elimination of residues from 1 to 86 or a single mutation of Ser10 prevents phosphorylation and inhibition of AC6 by PKC (Figure 6) (Lai et al., 1999). Subsequent studies also identified AC6 C1 Ser 568 and 674 as well as AC6 C2 Thr931 as inhibitory targets of PKC, suggesting that phosphorylation of this complex of Ser and Thr might trigger a conformational change in the catalytic core, changing AC6 catalytic activity (Lin et al., 2002).

AC5/6 regulation by glycosylation

AC6 can be glycosylated on two asparagine residues, namely, N805 and N890, on extracellular loops of TM2 (Figure 6) (Wu et al., 2001). The glycosylation of AC6 results in alteration of not only its catalytic activity but also its regulation by Gai or by PKC. Inhibition of glycosylation by tunicamycin impairs FSK-stimulated AC6 activity, and mutation of glycosylation sites resulted in significantly lower FSK-, Mn^{2+} -, and Mg^{2+} -stimulated enzyme activities than did wild-type AC6, suggesting that glycosylation may be necessary for maintenance of AC6 activity (Wu et al., 2001). AC5 glycosylation has not been verified.

AC5/6 regulation by nitrosylation

Cardiac function, airway, and vascular tone, as well as regulation of immunological defense and neuronal plasticity, are regulated through nitric oxide or reactive nitrogen species (Bhatia et al., 2021). NO-mediated S-nitrosylation of AC5/AC6 plays a crucial regulatory role in physiological processes of cardiovascular function. Previous studies showed that NO, independent of its action on the guanylyl cyclase (GC) pathway, inhibits cAMP in *Dictyostelium discoideum*, indicating that NO can have a direct regulatory effect on ACs via modification of an AC regulatory domain (McVey et al., 1999). Agonist-stimulated cAMP accumulation is inhibited when N18TG2 neuroblastoma cells (Tao et al., 1998) or cardiac myocytes (Joe et al., 1998; Vila-Petroff et al., 1999) are treated with NO or NO donors (Tao et al., 1998). This inhibition is not dependent on the effect of Gai (McVey et al., 1999) nor on PDE activity (Watson et al., 2001). NO or NO donors were shown to selectively decrease FSK-stimulated AC5/6 activity but not AC1 or AC2 (Hill et al., 2000), while calmodulin stimulation of AC1 is inhibited by NO (Duhe et al., 1994). NO suppression of hormonal or FSK-stimulated AC activity in neuroblastoma plasma membranes does not require CaM (McVey et al., 1999) and directly inhibits FSK-stimulated AC5 and AC6 activity (Hill et al., 2000).

The presence of caveolae, coincidentally the location of endogenous NO generation by endothelial nitric oxide synthase (eNOS), is required for NO inhibition of AC6 activity. Inhibition of AC5/6 by NO relies upon their localization in lipid rafts with caveolin signaling complexes (Ostrom et al., 2004). While lipid raft depletion with β -cyclodextrin prevented the activation of AC activity by β AR and Gas, it has no influence on the prostanoid receptors, which are located outside of caveolin-rich microdomains and can still activate AC. Both native cardiac myocytes and

pulmonary artery endothelial cells overexpressing AC6 are inhibited by the NO donor S-nitroso-N-acetylpenicillamine (SNAP), inhibiting both basal and FSK-stimulated cAMP production (Ostrom et al., 2004). This process is subject to reversal by reducing agents, indicating the involvement of cysteine residue(s) as the target for S-nitrosylation (McVey et al., 1999; Ostrom et al., 2004). The reaction between NO and superoxide (O_2^-) results in the production of reactive nitrogen oxygen species (RNOS) capable of altering a broader variety of biomolecules than NO itself (Ridnour et al., 2004). At higher levels of O_2^- , NO inactivation is followed in turn by generation of the potent and short-lived oxidant peroxynitrite ($ONOO^-/ONOOH$), which can directly react with metal ions and thiols (Piacenza et al., 2022). $ONOO^-$ contributes more to S-nitrosylation of adjacent proteins than does NO (Maccarrone et al., 2000). It is also proposed that inhibition of AC by NO may be through S-nitrosylation caused by the reaction of another NO intermediate, nitrosonium (NO^+), with cysteine residues (Tao et al., 1998). In a quantitative mass spectrometry screening investigation of modified cysteines utilizing a bioorthogonal cleavable-linker switch technique, AC6 featured among proteins identified as S-nitrosylated (Mnatsakanyan et al., 2019). AC6 inhibition due to S-nitrosylation was also demonstrated in pulmonary arteries (Sikarwar et al., 2018). Identification of AC6 cysteines susceptible to S-nitrosylation has been explored using site-directed mutagenesis of cysteines identified by bioinformatics analysis to reside within an SNO motif (Jaggupilli et al., 2018; Bhagirath et al., 2022). Mutation of cysteine 1004, located in the conserved C2 domain of AC6 near the Gas docking position (Figure 6), decreases the basal and stimulated activity of AC6, indicating the importance of this residue in AC6 for intact catalytic activity and also susceptibility to inhibition if nitrosylated (Bhagirath et al., 2022).

AC5/6 regulation by other PTMs

AC6 may be phosphorylated through receptor tyrosine kinases (RTKs); Ser residues 603, 608, 744, 746, 750, and 754 have been implicated following RTK activation by IGF-1 or tyrosine phosphatase inhibition with sodium orthovanadate (Figure 6). Augmentation of AC6 catalytic function following RTK activity is inhibited by endogenous $p74^{raf-1}$ activity, but not by inhibitors of ERK, PKC, PKA, or PI3 kinase activity (Tan et al., 2001).

Role of AC5 in cardiovascular function

In this section, we review the studies reporting the role of AC5 in cardiomyopathies induced by catecholamine stress including chronic isoproterenol stimulation, aging, and pressure overload. Upregulated AC5 mRNA expression in spontaneously hypertensive rats suggests complex regulation of LV hypertrophy in hypertension (Vatner et al., 2013). Myocardial AC5 mRNA increases from 5 to 12 weeks in spontaneously hypertensive rats, associated with the development of LV hypertrophy (Fujino et al., 2003). On the other hand, AC5KO mice have diminished sympathetic and parasympathetic responses and disrupted Ca^{2+} -mediated cardiac regulation. Both basal and isoproterenol-stimulated AC activity

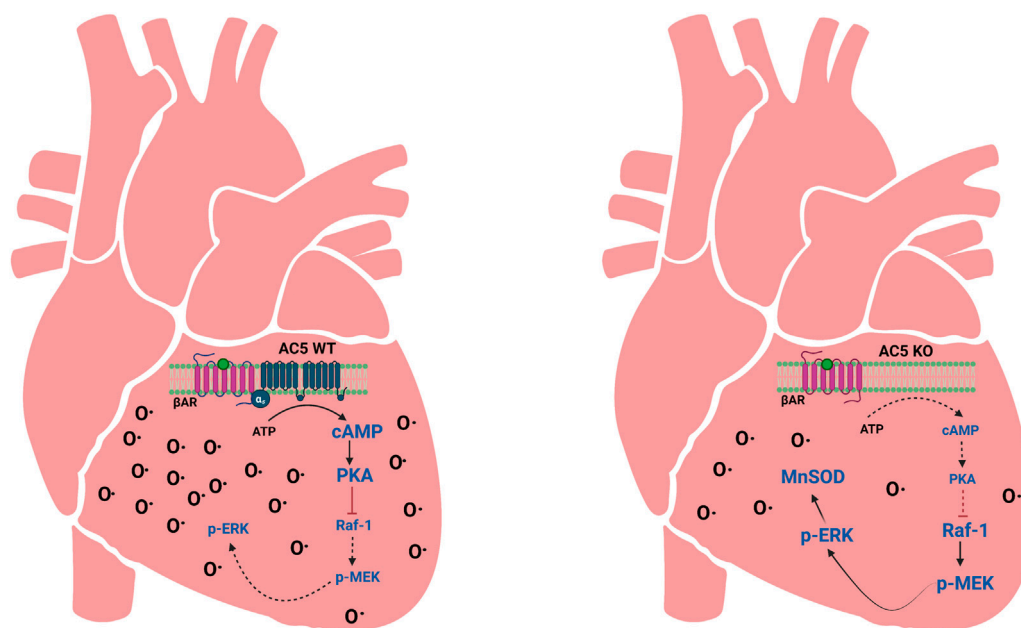


FIGURE 7

Depiction of AC5 signaling effects on cellular antioxidant imbalances in AC5WT and AC5KO hearts. When AC5 is disrupted (AC5KO), there is less β AR-stimulated cAMP, which decreases PKA activity. Lack of PKA leads to the activation of the Raf-1/MEK-ERK signaling pathway to increase the expression of MnSOD. This results in attenuation of oxidative stress. MnSOD antioxidant defense is not as present in AC5WT. $O\bullet$ indicates oxidative stress. Adapted from (Chester and Watts, 2007); image designed using BioRender.

are attenuated at 30%–40% in the cardiac membranes of AC5KO mice, with no compensatory increase in other AC isoforms; this reduction in AC activity does not alter cardiac function at baseline but compromises the LV inotropic response to adrenergic stimulation (Okumura et al., 2003a). In pressure overload, while there was no difference between AC5WT and AC5KO in cardiac muscle mass at baseline, AC5KO protected the heart from deleterious effects of pressure overload on LV ejection fraction, through restriction of myocardial apoptosis via upregulation of Bcl-2 (Okumura et al., 2003b). Others have demonstrated enhancement in basal LV function in AC5KO as well as impairment in the responsiveness of LV to β AR stimulation (Tang et al., 2006). AC5KO mice also showed better heart function following chronic catecholamine stimulation, again through reduction in cardiac apoptosis, due to increased Bcl-2 expression and Akt signaling (Okumura et al., 2007). In contrast to the outcomes after prolonged isoproterenol infusion in AC5WT and AC5KO mice, the ejection fraction response of the LV to an abrupt isoproterenol challenge was lowered in AC5KO, which is in line with the downregulation of AC5 catalytic activity. These data indicate that AC5 impairs survival signaling after long-term catecholamine infusion, while AC5 deletion improves cardiac desensitization, suggesting a novel strategy for heart failure therapy.

Zhang et al. investigated ways in which selected Gas-coupled receptors (GsPCRs) regulate cardiomyocyte viability by generating distinct signaling complexes governing pro-survival *versus* pro-death signaling. It was revealed that among the five GsPCRs studied, stimulation of β 1AR and histamine-H2-receptor (H2R) has negative effects on the survival of cardiomyocytes, mediated through cAMP production by AC5 but not AC6 and via PKA

activity stimulating pannexin-1 to release ATP into the extracellular space. On the other hand, activation of pro-survival GsPCRs adenosine-A2-receptor (A2R), calcitonin-gene-related-peptide-receptor (CGRPR), or relaxin-family peptide-receptor 1 (RXFP1) results in protective effects on cardiomyocyte survival as a function of cAMP generation by AC6, resulting in activation of cAMP efflux pumps. These findings indicate that selection of AC5 *versus* AC6 by GsPCRs determines cAMP localization which controls cAMP fate, thus altering cardiomyocyte survival (Zhong et al., 2023).

In an examination of heart rate variability during transient microgravity in parabolic flight, autonomic dysregulation became worse in AC5KO mice, while heart rate stability improved as a function of AC5 overexpression, indicating that AC5 may improve autonomic regulation (Okumura et al., 2008). It was further shown that AC5 activity is required to achieve constant responses in the low- and high-frequency ratio or normalized high frequency, two markers of sympathetic and parasympathetic activity, respectively (Bai et al., 2012). Thus, while inhibition of AC5 is beneficial for preventing cardiac myocyte myocardial apoptosis induced by excessive β AR stimulation, activation of this isoform may be advantageous in acute heart failure with low rate.

Transgenic mice overexpressing both AC5 and Gαq show efficient β AR-stimulated AC activity and cardiac contractility, but their hearts also appear pathologically fibrotic and hypertrophic (Tepe and Liggett, 1999). Microarray analysis of these hearts showed upregulation of various genes involved in pressure overload LV hypertrophy (Park et al., 2011). Examining genes relevant to ventricular hypertrophy upregulated in AC5 transgenic hearts even at baseline, transcription factor binding analysis revealed enrichment for the binding sites of nuclear factor of activated

TABLE 1 Physiological and pathophysiological effects of AC5 and AC6.

| Study of cardiomyopathy | Physiological and pathophysiological effects | | | |
|------------------------------|---|------------------------|---|------------------------|
| | AC5 | Ref | AC6 | Ref |
| Chronic pressure-overload | AC5 ^{-/-} showed better toleration, increased LV function, and less fibrosis and protected from apoptosis | Okumura et al. (2003b) | AC6-Tg (1) increased LV systolic wall stress, (2) reduced LVEF, and (3) unpreserved cardiac function | Guellich et al. (2010) |
| | AC5-Tg (1) manifested various upregulated genes related to LV hypertrophy and (2) marked enrichment of NFAT binding sites | Park et al. (2011) | Activation of AC6 expression led to better Ca ²⁺ handling due to enhanced PLN phosphorylation, decreased NCX1 and PP1 expression, and increased SR [Ca ²⁺] | Sugano et al. (2011) |
| | Expression of AC5 but not AC6 is upregulated in AMPKα2 KO-TAC mice, leading to cause hypertrophy and aggravating of cardiac function, as evidenced by a higher decrease in EF | Garnier et al. (2023) | AC6KO resulted in reduced LV hypertrophy with no sign of LV dilation and protection of LV function in female mice. This was further correlated with reduced expression of FHL1 and periostin | Tang et al. (2010) |
| Longevity | AC5KO did not change in exercise performance. Increase in exercise capacity in AC5KO mice results in enhanced skeletal muscle function, not cardiac function | Vatner et al. (2015) | Cardiac-directed expression of AC6 protects the heart from myocardial hypertrophy, improves cardiac function, increases cAMP generation, and prolongs survival in Gαq cardiomyopathy | Roth et al. (2002) |
| Chronic catecholamine stress | MnSOD expression was downregulated, and its effect was 40% lower in AC5-Tg than in WT, but 40% higher in AC5KO, leading to cause chronic catecholamine stress | Lai et al. (2009) | AC6 ^{ΔN/ΔN} and AC6 ^{-/-} mice (1) responded more to apoptotic myocytes and cardiac remodeling. (2) AC6 plays the main role on Src-dependent STAT3 activation in the sarcolemma and preserved cardiomyocytes against cardiac stress through a PKA/STAT3-dependent pathway | Wu et al. (2017) |

T-cells (NFAT), vital for the development and progress of cardiomyocyte hypertrophy (Park et al., 2011). NFAT binding determines the expression of cytoskeletal proteins (Schubert et al., 2003). So AC5 overexpression can mediate calcineurin–NFAT signaling involved in the development of LV hypertrophy (Park et al., 2011).

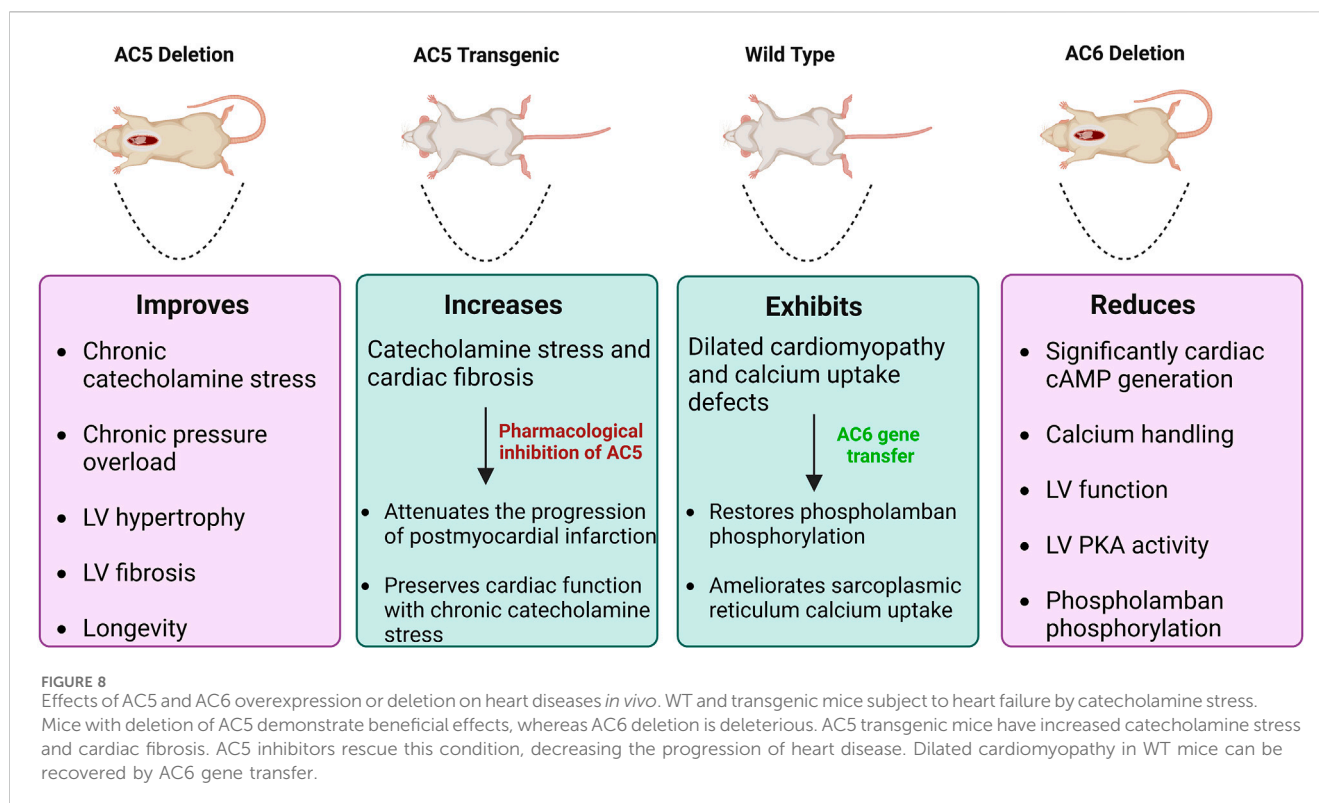
AC5KO mice also demonstrate increased physical performance, mediated through upregulation of the sirtuin-1 (SIRT1) pathway and regulating the antioxidant enzyme manganese superoxide dismutase (MnSOD) in the heart and liver of ACKO mice (Yan et al., 2012). It is proposed that SIRT1 is inhibited by AC5, leading to disruption of an interaction between SIRT1 and forkhead box O3 (FoxO3a), which eventually decreases MnSOD expression, thereby augmenting oxidative stress. The increase in MnSOD in myocytes subject to adenoviral AC5 KO is abolished by inhibition of either MEK or sirtuin, indicating MnSOD upregulation by both the mitogen-activated protein kinase kinase/extracellular signal-regulated kinases (MEK/ERK) and SIRT1/FoxO3a pathways (Chester and Watts, 2007; Lai et al., 2013). AC5KO mice are found to be resistant to cardiac stress with an enhanced median life span of roughly 30%, with protection from bone demineralization, and decreased susceptibility to fractures or aging-induced cardiomyopathy (Yan et al., 2007). The longevity, healthful aging, and stress resistance detected in AC5KO mice were correlated to diminished cAMP and PKA, resulting in activation of the Raf/MEK/ERK pathway, leading to the enhanced level of MnSOD (Yan et al., 2007). Additionally, AC5KO appears to increase NO signaling, demonstrating another mechanism by which AC5 may antagonize beneficial pathways induced by exercise (Guers et al., 2017). Two highly expressed genes encoding glutathione S-transferase (Gstk1 and Gstm) were downregulated in hypertrophic hearts, importantly because glutathione S-transferase plays the role of an antioxidant by

conjugating glutathione on various substrates to protect the heart from oxidative stress (Cho et al., 2003; Yan et al., 2012).

Overall, these data place AC5 in a crucial role regulating life span and cardiac stress resistance. Figure 7 summarizes AC5 signaling in the context of cellular antioxidant imbalances in AC5WT and AC5KO. Studies delineating the role of AC5 in cardiovascular function are highlighted in Table 1.

Role of AC6 in cardiovascular function

In contrast to AC5, several studies have shown that increase in AC6 expression is beneficial for the failing heart, preserving LV contractile function and reducing dilation and dysfunction in hearts showing pressure overload (Takahashi et al., 2006; Sugano et al., 2011). The protective effect of AC6 in the heart is postulated as cAMP pathway dependence and independence (Tang et al., 2004; Gao et al., 2008; Gao et al., 2009; Gao et al., 2017). AC6 overexpression prevents cardiac hypertrophy, fibrosis, and cardiomyopathy (Roth et al., 1999; Tang et al., 2013), while cardiac-directed expression of catalytically inactive AC6 restored the detrimental effects of sustained catecholamine infusion, through diminished myocardial cAMP production (Gao et al., 2017). In addition, in ischemic cardiomyopathy, cAMP production and systolic and diastolic LV function are enhanced after activation of AC6 expression (Lai et al., 2008). AC6KO mice exhibit reduced cAMP production and have significantly higher mortality compared to AC6WT, but remain susceptible to βAR stimulation-induced cardiomyopathy, compromised electrophysiological characteristics including diminished longitudinal conduction velocity, and impaired connexin 43 phosphorylation at Ser 368, which may be part of the mechanism of ventricular dysfunction (Tang et al., 2013). Cardiac-directed AC6 overexpression with Gαq expression triggered



improved β AR-stimulated AC activity, cAMP generation, and cardiac function *in vivo* and *ex vivo*, with no sign of hypertrophy and fibrosis (Roth et al., 1999), as well as improved Ca^{2+} handling and LV contractility manifested by ejection fraction, pressure development rate, and slope of the LV end-systolic pressure–volume relationship in aging mice (Tang et al., 2011). In 23-month-old rats, AC6 expression improved SR Ca^{2+} storage, while AC6 expression in 7-month-old mice did not show any difference in LV function and Ca^{2+} uptake (Tang et al., 2011). While cardiac troponin I (cTnI) phosphorylation diminishes with cardiac age (Jiang et al., 1993), improved LV function by AC6 is associated with phosphorylation of cTnI at Ser 23/24, which regulates thin filament function and thus contractility (Lai et al., 2008; Tang et al., 2011). While AC6 expression increases PKA activity and SR Ca^{2+} uptake (Tang et al., 2011), deletion of AC6 results in diminished PKA activity, phospholamban (PLB) phosphorylation, and decreased SR Ca^{2+} -ATPase activity 2a (SERCA2) affinity toward Ca^{+2} in failing hearts (Takahashi et al., 2006; Tang et al., 2008). Increased AC6 content increases the expression of activating transcription factor-3, which extinguishes PLB promoter activity, resulting in reduced PLB expression (Gao et al., 2004).

Enhanced AC6 expression is also correlated with increased nuclear phospho-Akt promoting phosphorylation of Akt at Ser473 and Thr308. This process appears to be independent of PKA or β AR stimulation (Gao et al., 2008). Akt activity is reversely regulated by PH domain leucine-rich repeat protein phosphatase (PHLPP), responsible for dephosphorylation of Akt at Ser473. AC6 inhibits PHLPP activity in cardiomyocytes, bringing about high levels of Akt phosphorylation. This PHLPP suppression is, however, rescued rapidly by isoproterenol and FSK stimulation, leading to significant dephosphorylation of Akt at

Ser473, but not Thr308. As PLB is an Akt target, active phospho-Akt increases PLB activity and thus improves sarcoplasmic Ca^{2+} cycling (Gao et al., 2009). Figure 8 gives an illustrated summary of the effects of AC5 and AC6 addition or deletion *in vivo*.

As AC6 shows remarkable physiological benefits for the heart, while its content and function are diminished in the failing heart (Ostrom et al., 2022), there has been increased interest in the efficacy and safety of adenoviral delivery of AC6 in patients with heart failure (Hammond et al., 2016) (AC6 Clinical Trials). In a mouse model, the AC6 C₁–C₂ construct was shown to effectively improve cardiac dysfunction caused by prolonged β AR stimulation (Roth et al., 2002). Although AC6 transgenic mice with cardiac-directed C₁–C₂ expression exhibited less cAMP production, they preserved normal cardiac function through ameliorated Ca^{2+} handling and tolerated sustained isoproterenol infusion and pressure overload without detrimental effects on LV contractility (Gao et al., 2016; Tan et al., 2019). Preclinical analysis of AC6 gene therapy recently transitioned into clinical trials. The phase 2 results for one-time AC6 gene transfer in 56 adult patients with symptomatic heart failure (ischemic or non-ischemic) and an ejection fraction of 40% resulted in safely ameliorated LV function above standard heart failure therapy (Hammond et al., 2016). The subsequent FLOURISH trial, a double-blinded placebo-controlled, multicenter phase 3 trial of 536 patients, aims to decrease heart failure hospitalization rates and improve ejection fraction, while minimizing adverse events after intracoronary injection of the human adenovirus 5 encoding human AC6 (Ad5. hAC6) gene in patients with heart failure and diminished LV ejection fraction; patient recruitment is ongoing (Penny et al., 2018).

Studies contextualizing the AC6 role in cardiovascular function are highlighted in Table 1.

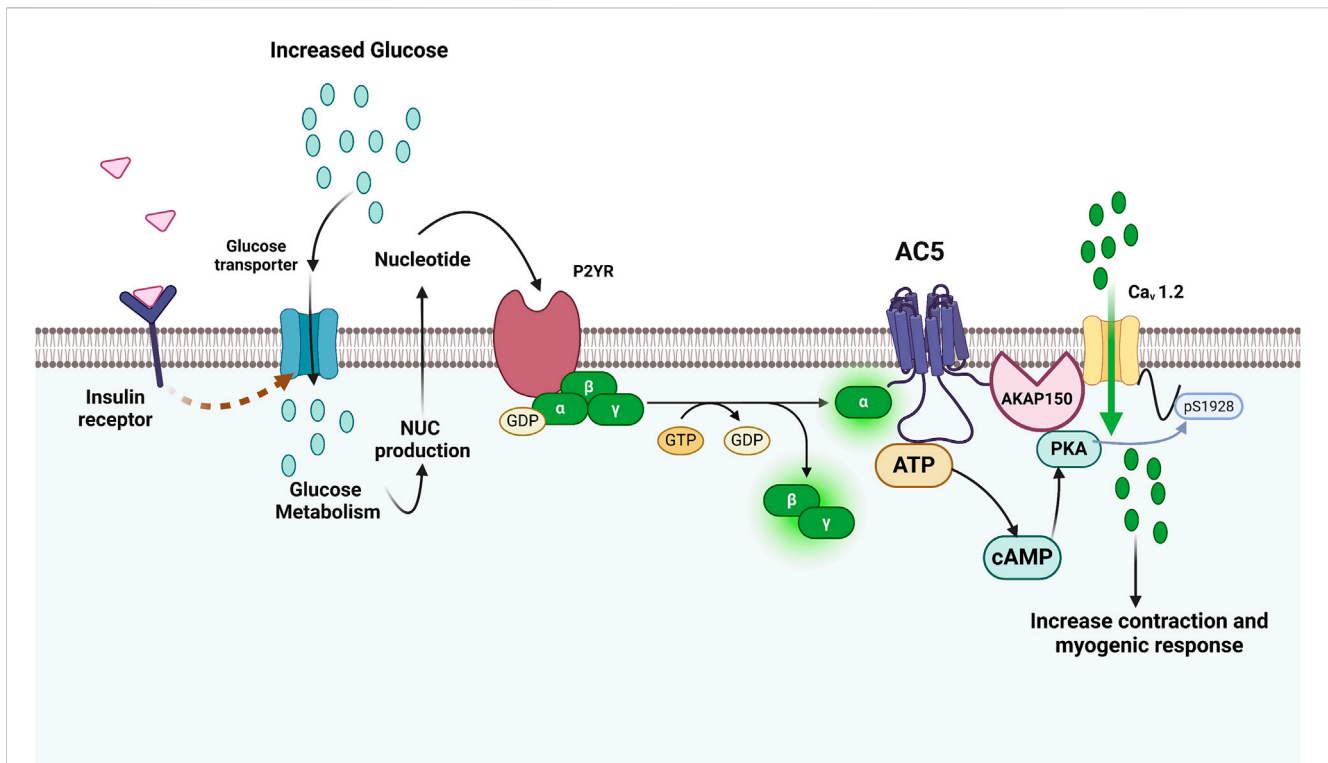


FIGURE 9

Involvement of AC5-mediated localized cAMP generation in diabetes and extracellular glucose activation of L-type Ca^{2+} channels and vasoconstriction. The illustration shows how glucose regulates L-type Ca^{2+} channel activity and vascular reactivity in an AC5-dependent manner. Increases in extracellular glucose can trigger P2YR linked to G_{as} signaling through extracellular nucleotide signaling because of transport and metabolism. Because AC5 and $\text{Ca}_v1.2$ are close in proximity, this cAMP microdomain may stimulate a pool of AKAP150-anchored PKA that is jointly linked to $\text{Ca}_v1.2$, making it to become more phosphorylated at Ser1928 and increasing channel function. Adapted from (Syed et al., 2019); image designed using BioRender.

Roles of AC5 and AC6 in vasculature

AC activity in vascular cells regulates vascular reactivity, apoptosis, hypertrophy, and proliferation (Gros et al., 2006). The distribution of AC isoforms varies in the vasculature compared to the myocardium. Based on RT-PCR studies, ACs 3, 5, and 6 are expressed in the adult rat aorta (Ostrom et al., 2002), while in the perinatal period, the ductus arteriosus expresses more AC2 and 6 than does the aorta (Yokoyama et al., 2010). The pulmonary artery expresses AC6 primarily, followed by ACs 7, 9, and 3 (Sikarwar et al., 2018). In vascular myocytes, AC6 is the main AC isoform involved in β AR-mediated cAMP/PKA signaling and activation of the K_{ATP} current, important for harmonizing the membrane potential and regulating vascular tone (Nelson et al., 2011). Subjects who carry the genetic variation *ADCY6 A674S* have increased blood pressure, with a hyperdynamic cardiac profile that is compatible with the effect of elevated AC function (Hodges et al., 2010). Vasodilation is antagonized by G_{aq} -mediated signaling, in part by direct Ca^{2+} inhibition of AC5 and AC6 (von Hayn et al., 2010).

Changes in AC activity have been related to the development of diabetes, heart failure, and hypertension (Matsumoto et al., 2005; Hodges et al., 2010). Abnormal arterial myocyte contractility, in addition to compromised endothelium-dependent vasodilation, is an important contributing factor to vascular complications including altered myogenic tone, both in diabetic mice and in

patients with diabetes (Montero et al., 2013; Sena et al., 2013; Tykocki et al., 2017; Syed et al., 2019). The contractile state of smooth muscle cells in the vessel wall determines the vascular tone, measured by a balance between the effects of vasoconstrictor and vasodilator signaling pathways, inclusive of adrenergic receptors and ACs (Shi et al., 2020). Physiological targets of ACs and downstream PKA include potassium channel phosphorylation, inducing hyperpolarization and vasodilation (Nelson et al., 2011). In resistance arteries and arterioles, the myogenic tone generated through the pulsatile stretch of the vascular wall modulates baseline smooth muscle contraction (Tykocki et al., 2017); in diabetic hyperglycemia, altered expression or function of potassium channels is linked to increased myogenic tone (Syed et al., 2019). Elevated glucose can trigger G_{as} signaling (Lemaire et al., 2004) but may drive vasoconstriction through glucose-induced cAMP production via AC5, which results in the activation of an anchored PKA pool, and in turn, phosphorylates the L-type Ca^{2+} channel pore-forming $\text{Ca}_v1.2$ subunit at Ser 1928 (Nystoriak et al., 2017; Prada et al., 2019; Syed et al., 2019). These interactions cause potentiation of L-type Ca^{2+} channel activity, enhanced $[\text{Ca}^{2+}]_i$, and vasoconstriction (Figure 9). Supporting this, interruption of AKAP5 function in arterial myocytes prevents cAMP generation in response to either increased glucose or selective purinergic P2Y11 agonist NF546; in AKAP5-null arterial myocytes or arteries, there is no clustering of P2Y11/P2Y11-like receptors,

AC5, PKA, and $\text{Ca}_v1.2$ into nanocomplexes at the plasma membrane; therefore, glucose- and NF546-induced potentiation of L-type Ca^{2+} channels and vasoconstriction does not occur (Prada et al., 2020). These data implicate AKAP5 and AC5 in the spatial confinement of cAMP signaling induced by elevated glucose via activation of P2Y11/P2Y11-like receptors in arterial myocytes (Prada et al., 2019; Prada et al., 2020).

ACs in endothelial cells play a role in vascular permeability. Prostacyclin-mediated signaling via AC6 (but not AC5) forms part of a feedback circuit that increases endothelial barrier function. Endothelial cells overexpressing AC6 have increased prostacyclin response, reducing the permeability of the endothelial barrier. In human umbilical vein endothelial cells, adenoviral-mediated gene transfer of AC6 increased prostacyclin receptor-stimulated cAMP synthesis and concurrently decreased thrombin-stimulated increases in endothelial cell barrier function (Bundey and Insel, 2006).

Role of ACs in cardiac automaticity

The sinoatrial node (SAN) is a crescent-like shaped cluster of myocytes split by connective tissue, spread over a few millimeters (Kashou et al., 2017), located at the convergence of the superior vena cava opening and the crista terminalis in the upper wall of the right atrium. The SAN comprises coordinated actions of pacemaker cells capable of generating a cyclic electrical impulse (Kashou et al., 2017). Cardiac arrhythmia due to the abnormality of the SAN can affect annually up to 1 per 1000 adults >45 years of age (John and Kumar, 2016). The SAN is the main regulator of heart rate and is regulated by β AR signaling (Irisawa et al., 1993; Tsutsui et al., 2021). While the expression of a number of AC isoforms (types 1, 2, 3, 5, 6, 8, and 9) has been identified in rabbit SAN, AC1 and AC8, two of the Ca^{2+} -activated AC isoforms, are predominantly distributed in atrial and SAN cells (Mattick et al., 2007; Younes et al., 2008; Robinson et al., 2021). In contrast, a recent study showed AC1 and AC6 but not AC8 expressed in SAN cells at the transcript and protein levels (Ren et al., 2022).

Ca^{2+} is a vital modulator of pacemaker potential via the Ca^{2+} clock (Maltsev and Lakatta, 2012), where the ryanodine receptor 2 (RyR2) facilitates the spontaneous release of Ca^{2+} from the SR, which in turn pushes Ca^{2+} to be released from the cytosol via the $\text{Na}^+-\text{Ca}^{2+}$ exchanger (Maltsev and Lakatta, 2012; Ren et al., 2022). Adrenergic control of the cardiac pacemaker current has been ascribed to AC1 (Mattick et al., 2007). AC1 mediates cAMP signaling in the SAN, in a functional microdomain with CAV3, hyperpolarization-activated cyclic nucleotide-gated 4 (HCN4), $\text{Ca}_v1.2$, and RyR2. cAMP released by AC1 leads to elevation of intracellular Ca^{2+} via Ca^{2+} channels, which triggers a positive feedback to AC1 and negative feedback to AC5/6 (Ren et al., 2022). While cardiac-specific overexpression of AC8 transgenic mice in the SAN is reported to augment the heart rate and rhythm (Moen et al., 2019), others found no differences in automaticity, basal heart rate, or isoproterenol responses in AC8-null mice compared with wild-type (Ren et al., 2022). In guinea pig atrial myocytes, sarcoplasmic reticulum type 2 inositol trisphosphate (IP3) receptors colocalize with AC8, with AC1 localized proximally. Functional AC1 and AC8 are required for the positive chronotropic effect of phenylephrine on the SAN, and activity of both AC1 and

AC8 plus PKA is required for the effect of IP3 on cellular Ca^{2+} transients (Capel et al., 2021).

Pacemaker current is generated by hyperpolarization-activated cyclic nucleotide-gated channels (HCN). To assess the role of Ca^{2+} homeostasis in autonomic regulation, AC1 and AC6 were expressed in cultures of spontaneously beating neonatal rat ventricle cells co-expressing HCN2. AC1, but not AC6 expression, increased intracellular cAMP and automaticity; AC1-mediated cAMP generation was resistant to β -adrenergic blockade, but the HCN2 response to an adrenergic agonist in the presence of AC1 (but not AC6) was sensitive to Ca^{2+} chelation, implying that the effect of Ca^{2+} homeostasis on the adrenergic regulation of the pacemaker rate could be accounted by the existence of a Ca^{2+} sensitive AC isoform (Kryukova et al., 2012; Robinson et al., 2021). However in a study of *in vivo* adenoviral gene transfer of AC6 in a porcine atrioventricular node block model, the AC6 injected group developed an escape rhythm of ~100 beats/min originating at the LV injection site, while control animals had RV escape rhythms, suggesting that biological pacemaker activity could also be triggered by AC6 (Ruhparwar et al., 2010).

AC5 and AC6 as potential drug targets for cardiovascular disease

Although β AR agonists and antagonists have therapeutic effectiveness for heart failure treatment, there are still patients who do not respond effectively; thus, heart failure is the most common cause of mortality worldwide (Capote et al., 2015). A weak cardiac response to catecholamine stimulation is a characteristic of the heart failure phenotype; hence, stimulation of the adrenergic pathway has been targeted to increase cardiac function. However, β ARs can undergo downregulation after prolonged stimulation by agonists or antagonists, which leads to a reduction in their cell surface density (tachyphylaxis), modifications in subtype composition, or increase in PKA or G-protein-coupled receptor kinase (GRK) activity, resulting in uncoupling of β AR from G proteins (desensitization) (Mahmood et al., 2022). β AR desensitization also serves as a compensatory mechanism, which may disrupt cardiac function and promote arrhythmias (Ho et al., 2010).

In contrast to GPCRs, AC5 and AC6 are less likely subject to desensitization after sustained ligand exposure (Pierre et al., 2009). The AC5 and AC6 isoforms have been indirectly targeted by GPCR agonists or antagonists and PDE inhibitors due to their action as the central relay site that assembles and amplifies a wide variety of signals (Pavan et al., 2009). AC5 and AC6 have pivotal roles in cardiac disease. Despite the promising results from knockout and transgenic mice, suggesting that AC5 and AC6 can be potential drug targets, advancements to selectively and therapeutically target these isoforms have been hindered by their structural similarity. However, many positive steps in this direction are reviewed below.

AC5 and AC6 activators for the treatment of heart failure

The most potent stimulus for increasing the cardiac output is initiated by sympathetic nervous system activation through the

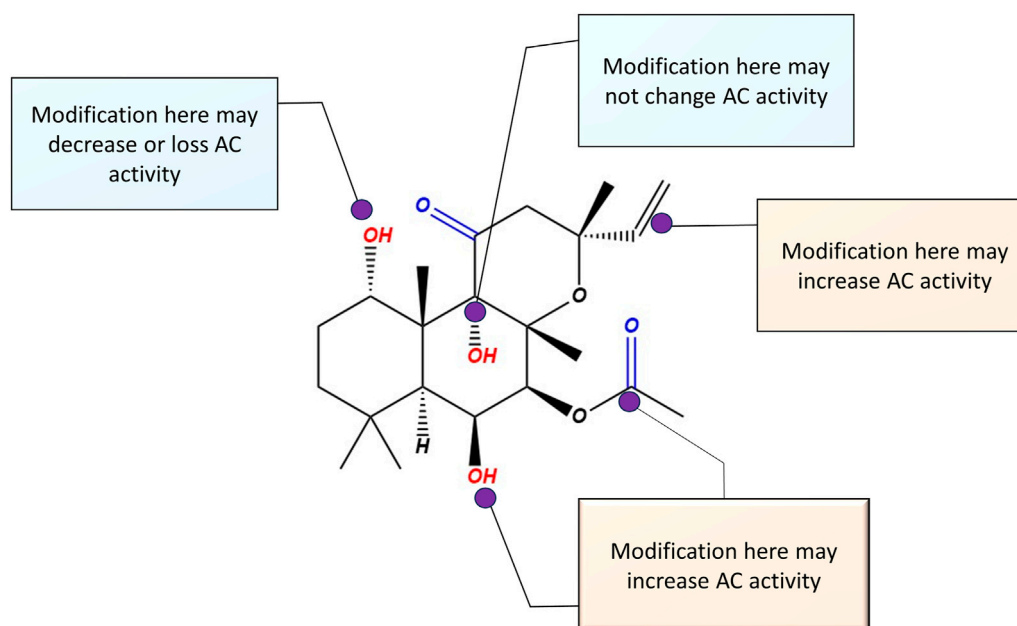


FIGURE 10
Possible modification sites on the parent compound forskolin molecule, for the design of selective AC5 or AC6 activators. Based on previous studies, any modification on C (1) reduces AC activity. Modifications on C (6) and (7) have the potential to enhance AC5 and AC6 activity.

activation of β AR and Gas, which in turn activates AC5 or AC6 to enhance cardiac contractility and rate. In the failing heart, production of basal and stimulated cAMP is diminished, but driving cardiac cAMP levels via adrenergic agents can result in higher long-term mortality (Bristow et al., 1982; Packer et al., 1991). FSK is a well-known activator of all transmembrane ACs, but its lack of selectivity impedes its clinical application; in addition to ACs, it also targets nuclear receptors, ion channels, glucose transporters, and P-glycoprotein multidrug resistance transporters (Dessauer et al., 2017). Clinically, in patients with heart failure, forskolin decreases ventricular filling pressures and vascular resistance while increasing cardiac output, ejection fraction, and stroke volume (Bristow et al., 1984). However, novel FSK-derived compounds have been synthesized that selectively target AC5 and AC6 isoforms (Pavan et al., 2009).

To design selective AC5 and AC6 activators, maintaining hydrogen bonding of FSK C1-OH, C7-acetyl, and C9-OH has been deemed crucial (Onda et al., 2001). There has also been specific focus on modification of C6 and C7 positions, due to a large open area apparent from AC crystallographic studies (Tesmer et al., 1997), suggesting that modification on these FSK positions may increase AC activity with selectivity and/or specificity. Figure 10 illustrates the possible sites of modifications for designing novel AC5 or AC6 activators. It is important to note that many of the earlier studies which established FSK positions suitable for modification evaluated only a subset of the AC isoforms. Testing of candidate compounds on each AC isoform remains necessary to determine which site is particular to each isoform.

The development of AC5 activator 6-[3-(dimethylamino)propionyl]FSK (NKH477, FD5, or colforsin daropate hydrochloride) (Toya et al., 1998) grabbed the attention of scientists working on AC drug discovery. Colforsin is a potent

water-soluble derivative of the C6 position of FSK, initially approved in Japan for the treatment of advanced congestive heart failure due to the elevation of cAMP in cardiac tissue, resulting in the enhancement of cardiac contractility. From the standpoint of AC isoform selectivity, colforsin activates AC5 > AC2~AC3 (Toya et al., 1998). Using the isolated perfused canine heart, the chronotropic, inotropic, coronary vasodilatory effects, and AC activity of colforsin were compared to those in catecholamines isoproterenol, dopamine, and dobutamine. All the drugs demonstrated positive chronotropic, inotropic, and coronary vasodilator effects. Colforsin cardiovascular actions were in the following order: coronary vasodilation >> positive inotropy > positive chronotropy, while isoproterenol, dopamine, and dobutamine evinced positive inotropy >> coronary vasodilation > positive chronotropy. While inotropic effects are desirable, chronotropic agents have potential arrhythmogenic effects and may impair cardiac filling; however, at doses with comparable inotropic effects, colforsin demonstrated a higher positive chronotropic effect than catecholamines or PDE inhibitors, with ventricular tachycardia triggered equally by colforsin and isoproterenol. Nevertheless, the coronary vasodilator activity of colforsin was more potent. The strong coronary vasodilator function of colforsin may therefore be applicable for the treatment of heart disease where coronary blood flow is limited and β AR-dependent signaling is also downregulated (Yoneyama et al., 2002). In a model of canine respiratory acidosis causing cardiac dysfunction, colforsin increased cardiac rate and decreased systemic vascular resistance to the same degree as did dobutamine, without attenuation by acidosis (Itami et al., 2019). Colforsin has also demonstrated pulmonary artery vasodilation (Yokochi et al., 2010), which is mediated largely through AC6 rather than AC5 (Sikarwar et al., 2018), indicating that colforsin may not be AC5-selective (Jaggupilli

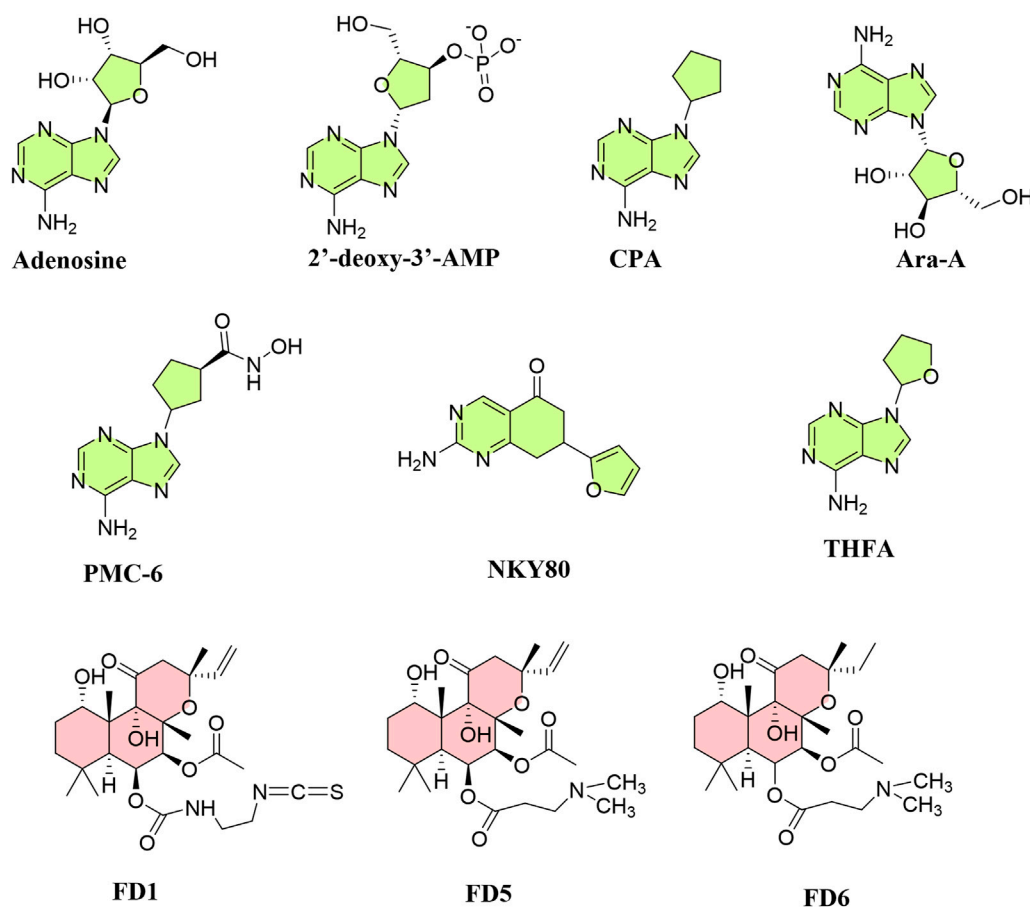


FIGURE 11
Structures of known inhibitors and activators targeting AC5 and AC6 allosteric or catalytic sites. Inhibitors (green) are mainly P-site inhibitors, and activators (red) are forskolin derivatives.

et al., 2018). 6-[3-(Dimethylamino)propionyl]-14-15-dihydro-FSK (FD6) is another FSK derivative, structurally similar to colforsin, but with reduced 14-15 alkyne bond, that showed AC5 and AC6 selectivity (Yokoyama et al., 2010). The lack of complete AC5 *versus* AC6 selectivity of colforsin and similar compounds does complicate our understanding of isoform-specific cardiac effects of AC activation, in the absence of more preclinical data. Protective or detrimental effects of individual AC manipulations on cardiac function can perhaps be best deciphered from gene overexpression or deletion studies; precise pharmacological interrogation of AC5 or AC6 activation would require further development of more selective tools.

A list of published inhibitors and FSK derivatives having AC5 and/or AC6 selectivity is depicted in Figure 11.

AC5 and AC6 inhibitors for the treatment of heart failure

As AC5 deletion appears to be protective in heart failure, there has been much research into isoform-selective AC5 inhibitors, which may be advantageous over β -blockers (Pierre et al., 2009). AC inhibitors are classified into two categories: competitive

inhibitors [(M)ANT- and TNP-nucleotides] and non-competitive P-site inhibitors. Among AC5 and AC6 inhibitors discovered, 2'-d-3'-MANT-GDP has inhibitory effects on AC6 (~8-fold) compared to AC5 (Gille et al., 2004). Among P-site inhibitors with metal chelating characteristics (PMC), PMC-6 has shown selectivity for AC5, protecting cardiac myocytes from β AR-induced apoptosis without degrading cAMP synthesis or contractility (Iwatsubo et al., 2004). P-site inhibitors that are adenosine analogs interact with ACs when PP_i is present and act by obstructing the interaction of other substrates with P-sites (Dessauer and Gilman, 1997). Unlike FSK, P-site inhibitors attach to the catalytic site for substrate ATP (Tesmer et al., 2000), generating a dead-end complex with PP_i (Bushfield et al., 1990). Ribose-substituted P-ligands such as 9-(cyclopentyl)-adenine (CPA) and 9-(tetrahydrofuryl)-adenine (THFA) have IC_{50} values in the micromolar range, selectively inhibiting AC5 more than AC3 and AC2; while 2'-deoxy-3'-AMP and 3'-AMP inhibit AC3 and AC5 more than AC2 (Onda et al., 2001; Iwatsubo et al., 2003). 2-Amino-7-(2-furanyl)-7,8-dihydro-5(6H)-quinazolinone (NKY80), derived from 9-(tetrahydro-2-furanyl)-9H-purin-6-amine (SQ22,536), like 9-(tetrahydro-2-furyl)adenine (THFA), inhibits AC5 despite not possessing an adenine ring. NKY80, though less potent, exhibits a similar AC5 selectivity to THFA in the inhibition of AC5 catalytic

activity, having a selectivity ratio of 210 between AC5 and AC2 with an IC₅₀ of 8.3 μ M for AC5, 132 μ M for AC3, and 1.7 mM for AC2 when Gas-GTP γ S-forskolin is present (Onda et al., 2001).

The antiviral drug adenine 9- β -D-arabinofuranoside (Ara-A), known as vidarabine, has been found to selectively inhibit AC5; Ara-A markedly diminished AC activity in AC5 transgenic mice, but not in AC5KO, and had a minor effect in either WT or AC6 transgenic mice (Iwatsubo et al., 2012). However, another study indicated that Ara-A is also a potent AC6 inhibitor (pIC₅₀: 5.67 and 5.34, for AC5 and AC6, respectively); indeed, SQ22,536, NKY80, and Ara-A inhibit both AC5 and AC6 without distinguishing them (Brand et al., 2013). Inhibition of AC5 by Ara-A is thought to be mediated through the MEK/ERK pathway. The Ca²⁺-binding protein annexin A4 (ANXA4) selectively inhibits AC5. Both ANXA4 and a peptide encompassing the ANXA4 N-terminal sequence (A4N₁₋₂₂) reduced cAMP generation in AC5; ANXA4 co-immunoprecipitates with AC5 but not AC6, binding to its N-terminal domain. The peptide A4N₁₋₂₂ diminishes recruitment of the L-type Ca²⁺ current (I_{CaL}) and prevents action potential prolongation after catecholamine challenge, an effect similar to the loss of the β 1AR signal in AC5KO models (Heinick et al., 2020). Another novel AC5 inhibitor, C90, inhibits cAMP production to FSK by 42% in WT but not in AC5KO, suggesting its selective AC5 inhibitory effect; it is five times less potent in inhibiting AC2 and AC6. C90 also produced the interesting effect of decreasing myocardial infarct size even when administered after coronary reperfusion (Zhang et al., 2018).

Conclusion

Although there have been a number of AC5 and AC6 activators/inhibitors tested *in vitro*, more *in vivo* studies are required for evaluation of the cardiac AC isoforms as drug targets and more precision of selectivity testing, given the similar structures and expressions patterns of cardiac ACs. Advances in detailed structural information (Qi et al., 2019) as well as computational homology modeling and mutational analysis of ligand-binding regions (Bhatia et al., 2023) will provide further direction to drug discovery for selective targeting of cardiovascular ACs, as will the selection of appropriate pharmacokinetic features and targeted drug delivery systems to decrease off-target adverse effects.

The prevalence of cardiovascular disease is on the rise, despite the rapid advancements in drug discovery for therapeutics addressing the alteration in the adrenergic signaling profile in the failing heart. Increased activity of AC6 has a beneficial effect on

cardiac contractility, cell survival, and Ca²⁺ handling, while the activation of AC5 is deleterious. Differentiating the regulation of AC5 and AC6 and their signaling pathways opens up potential avenues for selective manipulation of physiologically important cAMP pools. The development of AC5/6 selective activators or inhibitors is still in its infancy, generating novel stimulators and inhibitors or taking advantage of drug repositioning; AC6 gene transfer also holds promise for heart failure treatment. Further research in these directions is clearly warranted.

Author contributions

SM: conceptualization, data curation, supervision, and writing—original draft. RS: writing—original draft. BV: writing—original draft. SD: conceptualization, funding acquisition, project administration, resources, supervision, and writing—review and editing.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

- Adams, S. R., Harootunian, A. T., Buechler, Y. J., Taylor, S. S., and Tsien, R. Y. (1991). Fluorescence ratio imaging of cyclic AMP in single cells. *Nature* 349 (6311), 694–697. doi:10.1038/349694a0
- Antoni, F. A. (2020). The chilling of adenylyl cyclase 9 and its translational potential. *Cell. Signal.* 70, 109589. doi:10.1016/j.cellsig.2020.109589
- Bai, Y., Tsunematsu, T., Jiao, Q., Ohnuki, Y., Mototani, Y., Shiozawa, K., et al. (2012). Pharmacological stimulation of type 5 adenylyl cyclase stabilizes heart rate under both microgravity and hypergravity induced by parabolic flight. *J. Pharmacol. Sci.* 119 (4), 381–389. doi:10.1254/jphs.12102fp
- Baldwin, T. A., and Dessauer, C. W. (2018). Function of adenylyl cyclase in heart: the AKAP connection. *J. Cardiovasc. Dev. Dis.* 5 (1), 2. doi:10.3390/jcdd5010002
- Baldwin, T. A., Li, Y., Brand, C. S., Watts, V. J., and Dessauer, C. W. (2019). Insights into the regulatory properties of human adenylyl cyclase type 9. *Mol. Pharmacol.* 95 (4), 349–360. doi:10.1124/mol.118.114595
- Bauman, A. L., Soughayer, J., Nguyen, B. T., Willoughby, D., Carnegie, G. K., Wong, W., et al. (2006). Dynamic regulation of cAMP synthesis through anchored PKA-adenylyl cyclase V/VI complexes. *Mol. Cell* 23 (6), 925–931. doi:10.1016/j.molcel.2006.07.025

- Beazely, M. A., and Watts, V. J. (2006). Regulatory properties of adenylate cyclases type 5 and 6: a progress report. *Eur. J. Pharmacol.* 535 (1-3), 1–12. doi:10.1016/j.ejphar.2006.01.054
- Bhagirath, A. Y., Bhatia, V., Medapati, M. R., Singh, N., Hinton, M., Chelikani, P., et al. (2022). Critical cysteines in the functional interaction of adenylyl cyclase isoform 6 with Gas. *FASEB BioAdvances* 4 (3), 180–196. doi:10.1096/fba.2021-00073
- Bhatia, V., Elnagary, L., and Dakshinamurti, S. (2021). Tracing the path of inhaled nitric oxide: biological consequences of protein nitrosylation. *Pediatr. Pulmonol.* 56 (2), 525–538. doi:10.1002/ppul.25201
- Bhatia, V., Maghsoudi, S., Hinton, M., Bhagirath, A. Y., Singh, N., Jaggupilli, A., et al. (2023). Characterization of adenylyl cyclase isoform 6 residues interacting with forskolin. *Biology* 12 (4), 572. doi:10.3390/biology12040572
- Brand, C. S., Hocker, H. J., Gorfe, A. A., Cavaotto, C. N., and Dessauer, C. W. (2013). Isoform selectivity of adenylyl cyclase inhibitors: characterization of known and novel compounds. *J. Pharmacol. Exp. Ther.* 347 (2), 265–275. doi:10.1124/jpet.113.208157
- Bristow, M. R., Ginsburg, R., Minobe, W., Cubicciotti, R. S., Sageman, W. S., Lurie, K., et al. (1982). Decreased catecholamine sensitivity and beta-adrenergic-receptor density in failing human hearts. *N. Engl. J. Med.* 307 (4), 205–211. doi:10.1056/NEJM198207223070401
- Bristow, M. R., Ginsburg, R., Strosberg, A., Montgomery, W., and Minobe, W. (1984). Pharmacology and inotropic potential of forskolin in the human heart. *J. Clin. investigation* 74 (1), 212–223. doi:10.1172/JCI111404
- Brodde, O. E. (1991). Beta 1- and beta 2-adrenoceptors in the human heart: properties, function, and alterations in chronic heart failure. *Pharmacol. Rev.* 43 (2), 203–242.
- Bundey, R. A., and Insel, P. A. (2006). Adenylyl cyclase 6 overexpression decreases the permeability of endothelial monolayers via preferential enhancement of prostacyclin receptor function. *Mol. Pharmacol.* 70 (5), 1700–1707. doi:10.1124/mol.106.028035
- Bushfield, M., Shoshani, I., Cifuentes, M., Stübner, D., and Johnson, R. A. (1990). Inhibition of adenylate cyclase by polyadenylate. *Archives Biochem. biophysics* 278 (1), 88–98. doi:10.1016/0003-9861(90)90235-q
- Cai, W., Fujita, T., Hidaka, Y., Jin, H., Suita, K., Prajapati, R., et al. (2016). Disruption of Epac1 protects the heart from adenylyl cyclase type 5-mediated cardiac dysfunction. *Biochem. biophysical Res. Commun.* 475 (1), 1–7. doi:10.1016/j.bbrc.2016.04.123
- Calamera, G., Moltzau, L. R., Levy, F. O., and Andressen, K. W. (2022). Phosphodiesterases and compartmentation of cAMP and cGMP signaling in regulation of cardiac contractility in normal and failing hearts. *Int. J. Mol. Sci.* 23 (4), 2145. doi:10.3390/ijms23042145
- Calebiro, D., and Maiellaro, I. (2014). cAMP signaling microdomains and their observation by optical methods. *Front. Cell Neurosci.* 8, 350. doi:10.3389/fncel.2014.00350
- Capel, R. A., Bose, S. J., Collins, T. P., Rajasundaram, S., Ayagama, T., Zaccolo, M., et al. (2021). IP3-mediated Ca²⁺ release regulates atrial Ca²⁺ transients and pacemaker function by stimulation of adenylyl cyclases. *Am. J. Physiology-Heart Circulatory Physiology* 320 (1), H95–H107. doi:10.1152/ajpheart.00380.2020
- Capote, L. A., Perez, R. M., and Lymperopoulos, A. (2015). GPCR signaling and cardiac function. *Eur. J. Pharmacol.* 763, 143–148. doi:10.1016/j.ejphar.2015.05.019
- Cattani-Cavaliere, I., Li, Y., Margolis, J., Bogard, A., Roosan, M. R., and Ostrom, R. S. (2023). Quantitative phosphoproteomic analysis reveals unique cAMP signaling pools emanating from AC2 and AC6 in human airway smooth muscle cells. *Front. Physiology* 14, 1149063. doi:10.3389/fphys.2023.1149063
- Chen, J., and Iyengar, R. (1993). Inhibition of cloned adenylyl cyclases by mutant-activated Gi-alpha and specific suppression of type 2 adenylyl cyclase inhibition by phorbol ester treatment. *J. Biol. Chem.* 268 (17), 12253–12256. doi:10.1016/s0021-9258(18)31381-4
- Chen, Y., Harry, A., Li, J., Smit, M. J., Bai, X., Magnusson, R., et al. (1997). Adenylyl cyclase 6 is selectively regulated by protein kinase A phosphorylation in a region involved in Galphas stimulation. *Proc. Natl. Acad. Sci.* 94 (25), 14100–14104. doi:10.1073/pnas.94.25.14100
- Chester, J. A., and Watts, V. J. (2007). Adenylyl cyclase 5: a new clue in the search for the "fountain of youth"? *Science's STKE* 2007 (413), pe64. doi:10.1126/stke.4132007pe64
- Cho, C. G., Kim, H. J., Chung, S. W., Jung, K. J., Shim, K. H., Yu, B. P., et al. (2003). Modulation of glutathione and thioredoxin systems by calorie restriction during the aging process. *Exp. Gerontol.* 38 (5), 539–548. doi:10.1016/s0531-5565(03)00005-6
- Colombe, A.-S., and Pidoux, G. (2021). Cardiac cAMP-PKA signaling compartmentalization in myocardial infarction. *Cells* 10 (4), 922. doi:10.3390/cells10040922
- Cosson, M.-V., Hiis, H. G., Moltzau, L. R., Levy, F. O., and Krobert, K. A. (2019). Knockout of adenylyl cyclase isoform 5 or 6 differentially modifies the β 1-adrenoceptor-mediated inotropic response. *J. Mol. Cell. Cardiol.* 131, 132–145. doi:10.1016/j.jymcc.2019.04.017
- Cros, C., and Brette, F. (2013). Functional subcellular distribution of β 1- and β 2-adrenergic receptors in rat ventricular cardiac myocytes. *Physiol. Rep.* 1 (3), e00038. doi:10.1002/phy2.38
- Defer, N., Best-Belpomme, M., and Hanoune, J. (2000). Tissue specificity and physiological relevance of various isoforms of adenylyl cyclase. *Am. J. Physiology-Renal Physiology* 279 (3), F400–F416. doi:10.1152/ajprenal.2000.279.3.F400
- Dessauer, C. W., and Gilman, A. G. (1997). The catalytic mechanism of mammalian adenylyl cyclase: equilibrium binding and kinetic analysis of P-site inhibition. *J. Biol. Chem.* 272 (44), 27787–27795. doi:10.1074/jbc.272.44.27787
- Dessauer, C. W., Watts, V. J., Ostrom, R. S., Conti, M., Dove, S., and Seifert, R. (2017). International union of basic and clinical pharmacology. CI. Structures and small molecule modulators of mammalian adenylyl cyclases. *Pharmacol. Rev.* 69 (2), 93–139. doi:10.1124/pr.116.013078
- Diviani, D., Reggi, E., Arambasic, M., Caso, S., and Maric, D. (2016). Emerging roles of A-kinase anchoring proteins in cardiovascular pathophysiology. *Biochimica Biophysica Acta (BBA)-Molecular Cell Res.* 1863 (7), 1926–1936. doi:10.1016/j.bbamcr.2015.11.024
- Du, R., Nielsen, M. D., Dittman, A. H., Villacres, E. C., Choi, E.-J., and Storm, D. (1994). Oxidation of critical cysteine residues of type I adenylyl cyclase by o-iodosobenzoate or nitric oxide reversibly inhibits stimulation by calcium and calmodulin. *J. Biol. Chem.* 269 (10), 7290–7296. doi:10.1016/s0021-9258(17)37282-4
- Efendiev, R., and Dessauer, C. W. (2011). A kinase-anchoring proteins and adenylyl cyclase in cardiovascular physiology and pathology. *J. Cardiovasc. Pharmacol.* 58 (4), 339–344. doi:10.1097/FJC.0b013e31821bc3f0
- Engelhardt, S., Hein, L., Wiesmann, F., and Lohse, M. J. (1999). Progressive hypertrophy and heart failure in beta1-adrenergic receptor transgenic mice. *Proc. Natl. Acad. Sci. U. S. A.* 96 (12), 7059–7064. doi:10.1073/pnas.96.12.7059
- Espinasse, I., Iourgenko, V., Defer, N., Samson, F., Hanoune, J., and Mercadier, J.-J. (1995). Type V, but not type VI, adenylyl cyclase mRNA accumulates in the rat heart during ontogenic development. Correlation with increased global adenylyl cyclase activity. *J. Mol. Cell. Cardiol.* 27 (9), 1789–1795. doi:10.1016/0022-2828(95)90002-0
- Esposito, G., Perrino, C., Ozaki, T., Takaoka, H., Defer, N., Petretta, M. P., et al. (2008). Increased myocardial contractility and enhanced exercise function in transgenic mice overexpressing either adenylyl cyclase 5 or 8. *Basic Res. Cardiol.* 103, 22–30. doi:10.1007/s00395-007-0688-6
- Fagan, K. A., Mons, N., and Cooper, D. M. (1998). Dependence of the Ca²⁺-inhibitable adenylyl cyclase of C6-2B glioma cells on capacitative Ca²⁺ entry. *J. Biol. Chem.* 273 (15), 9297–9305. doi:10.1074/jbc.273.15.9297
- Ferrara, N., Comici, K., Corbi, G., Pagano, G., Furgi, G., Rengo, C., et al. (2014). β -adrenergic receptor responsiveness in aging heart and clinical implications. *Front. physiology* 4, 396. doi:10.3389/fphys.2013.00396
- Fujino, T., Hasebe, N., Kawabe, J.-i., Fujita, M., Fukuzawa, J., Tobise, K., et al. (2003). Effect of beta-adrenoceptor antagonist and angiotensin-converting enzyme inhibitor on hypertension-associated changes in adenylyl cyclase type V messenger RNA expression in spontaneously hypertensive rats. *J. Cardiovasc. Pharmacol.* 41 (5), 720–725. doi:10.1097/00005344-200305000-00008
- Fujita, T., Umemura, M., Yokoyama, U., Okumura, S., and Ishikawa, Y. (2017). The role of Epac in the heart. *Cell. Mol. life Sci.* 74, 591–606. doi:10.1007/s00018-016-2336-5
- Gao, M. H., Lai, N. C., Giamouridis, D., Kim, Y. C., Guo, T., and Hammond, H. K. (2017). Cardiac-directed expression of a catalytically inactive adenylyl cyclase 6 protects the heart from sustained β -adrenergic stimulation. *PLoS one* 12 (8), e0181282. doi:10.1371/journal.pone.0181282
- Gao, M. H., Lai, N. C., Giamouridis, D., Kim, Y. C., Tan, Z., Guo, T., et al. (2016). Cardiac-directed expression of adenylyl cyclase catalytic domain reverses cardiac dysfunction caused by sustained beta-adrenergic receptor stimulation. *JACC Basic Transl. Sci.* 1 (7), 617–629. doi:10.1016/j.jacbs.2016.08.004
- Gao, M. H., Miyahara, A., Feramisco, J. R., and Tang, T. (2009). Activation of PH-domain leucine-rich protein phosphatase 2 (PHLPP2) by agonist stimulation in cardiac myocytes expressing adenylyl cyclase type 6. *Biochem. biophysical Res. Commun.* 384 (2), 193–198. doi:10.1016/j.bbrc.2009.04.110
- Gao, M. H., Tang, T., Guo, T., Miyahara, A., Yajima, T., Pestonjamas, K., et al. (2008). Adenylyl cyclase type VI increases Akt activity and phospholamban phosphorylation in cardiac myocytes. *J. Biol. Chem.* 283 (48), 33527–33535. doi:10.1074/jbc.M805825200
- Gao, M. H., Tang, T., Guo, T., Sun, S. Q., Feramisco, J. R., and Hammond, H. K. (2004). Adenylyl cyclase type VI gene transfer reduces phospholamban expression in cardiac myocytes via activating transcription factor 3. *J. Biol. Chem.* 279 (37), 38797–38802. doi:10.1074/jbc.M405701200
- Garnier, A., Leroy, J., Delomé, C., Mateo, P., Viollet, B., Veksler, V., et al. (2023). Modulation of cardiac cAMP signaling by AMPK and its adjustments in pressure overload-induced myocardial dysfunction in rat and mouse. *Plos one* 18 (9), e0292015. doi:10.1371/journal.pone.0292015
- Gille, A., Lushington, G. H., Mou, T.-C., Doughty, M. B., Johnson, R. A., and Seifert, R. (2004). Differential inhibition of adenylyl cyclase isoforms and soluble guanylyl cyclase by purine and pyrimidine nucleotides. *J. Biol. Chem.* 279 (19), 19955–19969. doi:10.1074/jbc.M312560200
- Göttle, M., Geduhn, J., König, B., Gille, A., Höcherl, K., and Seifert, R. (2009). Characterization of mouse heart adenylyl cyclase. *J. Pharmacol. Exp. Ther.* 329 (3), 1156–1165. doi:10.1124/jpet.109.150953
- Gros, R., Ding, Q., Chorazyczewski, J., Pickering, J. G., Limbird, L. E., and Feldman, R. D. (2006). Adenylyl cyclase isoform-selective regulation of vascular smooth muscle

proliferation and cytoskeletal reorganization. *Circulation Res.* 99 (8), 845–852. doi:10.1161/01.RES.0000245189.21703.c0

Gu, C., Cali, J. J., and Cooper, D. M. (2002). Dimerization of mammalian adenylate cyclases: functional, biochemical and fluorescence resonance energy transfer (FRET) studies. *Eur. J. Biochem.* 269 (2), 413–421. doi:10.1046/j.0014-2956.2001.02708.x

Guellich, A., Gao, S., Hong, C., Yan, L., Wagner, T. E., Dhar, S. K., et al. (2010). Effects of cardiac overexpression of type 6 adenyl cyclase affects on the response to chronic pressure overload. *Am. J. Physiology-Heart Circulatory Physiology* 299 (3), H707–H712. doi:10.1152/ajpheart.00148.2010

Guers, J. J., Zhang, J., Campbell, S. C., Oydanich, M., Vatner, D. E., and Vatner, S. F. (2017). Disruption of adenyl cyclase type 5 mimics exercise training. *Basic Res. Cardiol.* 112, 59–12. doi:10.1007/s00395-017-0648-8

Guillou, J.-L., Nakata, H., and Cooper, D. M. (1999). Inhibition by calcium of mammalian adenyl cyclases. *J. Biol. Chem.* 274 (50), 35539–35545. doi:10.1074/jbc.274.50.35539

Haber, N., Stengel, D., Defer, N., Roeckel, N., Mattei, M. G., and Hanoune, J. (1994). Chromosomal mapping of human adenyl cyclase genes type III, type V and type VI. *Hum. Genet.* 94 (1), 69–73. doi:10.1007/BF02272844

Hackley, C. R., Mazzoni, E. O., and Blau, J. (2018). cAMP: a single-wavelength fluorescent sensor for cyclic AMP. *Sci. Signal.* 11 (520), eaah3738. doi:10.1126/scisignal.aah3738

Halls, M. L., and Cooper, D. M. (2017). Adenyl cyclase signalling complexes—Pharmacological challenges and opportunities. *Pharmacol. Ther.* 172, 171–180. doi:10.1016/j.pharmthera.2017.01.001

Hammond, H. K., Penny, W. F., Traverse, J. H., Henry, T. D., Watkins, M. W., Yancy, C. W., et al. (2016). Intracoronary gene transfer of adenyl cyclase 6 in patients with heart failure: a randomized clinical trial. *JAMA Cardiol.* 1 (2), 163–171. doi:10.1001/jamacardio.2016.0008

Hanoune, J., and Defer, N. (2001). Regulation and role of adenyl cyclase isoforms. *Annu. Rev. Pharmacol. Toxicol.* 41 (1), 145–174. doi:10.1146/annurev.pharmtox.41.1.145

Hanski, E., Sevilla, N., and Levitzki, A. (1977). The allosteric inhibition by calcium of soluble and partially purified adenylate cyclase from Turkey erythrocytes. *Eur. J. Biochem.* 76 (2), 513–520. doi:10.1111/j.1432-1033.1977.tb11621.x

Heinick, A., Pluteanu, F., Hermes, C., Klemme, A., Domnik, M., Husser, X., et al. (2020). Annexin A4 N-terminal peptide inhibits adenyl cyclase 5 and limits β -adrenoceptor-mediated prolongation of cardiac action potential. *FASEB J.* 34 (8), 10489–10504. doi:10.1096/fj.20190209ARR

Hill, J., Howlett, A., and Klein, C. (2000). Nitric oxide selectively inhibits adenyl cyclase isoforms 5 and 6. *Cell. Signal.* 12 (4), 233–237. doi:10.1016/s0898-6568(99)00082-0

Ho, D., Yan, L., Iwatsubo, K., Vatner, D. E., and Vatner, S. F. (2010). Modulation of beta-adrenergic receptor signaling in heart failure and longevity: targeting adenyl cyclase type 5. *Heart Fail. Rev.* 15, 495–512. doi:10.1007/s10741-010-9183-5

Hodges, G. J., Gros, R., Hegele, R. A., Van Uum, S., Shoemaker, J. K., and Feldman, R. D. (2010). Increased blood pressure and hyperdynamic cardiovascular responses in carriers of a common hyperfunctional variant of adenyl cyclase 6. *J. Pharmacol. Exp. Ther.* 335 (2), 451–457. doi:10.1124/jpet.110.172700

Hu, B., Nakata, H., Gu, C., De Beer, T., and Cooper, D. M. (2002). A critical interplay between Ca^{2+} inhibition and activation by Mg^{2+} of AC5 revealed by mutants and chimeric constructs. *J. Biol. Chem.* 277 (36), 33139–33147. doi:10.1074/jbc.M112373200

Hu, C.-L., Chandra, R., Ge, H., Pain, J., Yan, L., Babu, G., et al. (2009). Adenyl cyclase type 5 protein expression during cardiac development and stress. *Am. J. Physiology-Heart Circulatory Physiology* 297 (5), H1776–H1782. doi:10.1152/ajpheart.00050.2009

Hunter, M. R., Finlay, D. B., Macdonald, C. E., Cawston, E. E., Grimsey, N. L., and Glass, M. (2017). Real-time measurement of cannabinoid receptor-mediated cAMP signaling. *Methods Enzymol.* 593, 43–59. doi:10.1016/bs.mie.2017.05.001

Irisawa, H., Brown, H., and Giles, W. (1993). Cardiac pacemaking in the sinoatrial node. *Physiol. Rev.* 73 (1), 197–227. doi:10.1152/physrev.1993.73.1.197

Ishikawa, Y., Sorota, S., Kiuchi, K., Shannon, R., Komamura, K., Katsushika, S., et al. (1994). Downregulation of adenyl cyclase types V and VI mRNA levels in pacing-induced heart failure in dogs. *J. Clin. investigation* 93 (5), 2224–2229. doi:10.1172/JCI117219

Itami, T., Hanazono, K., Oyama, N., Sano, T., Makita, K., and Yamashita, K. (2019). Cardiovascular effects of intravenous colforsin in normal and acute respiratory acidosis canine models: a dose-response study. *PLoS One* 14 (7), e0213414. doi:10.1371/journal.pone.0213414

Iwami, G., Akanuma, M., Kawabe, J., Cannon, P. J., Homcy, C. J., and Ishikawa, Y. (1995). Multiplicity in type V adenyl cyclase: type Va and type Vb. *Mol. Cell. Endocrinol.* 110 (1–2), 43–47. doi:10.1016/0303-7207(95)03514-8

Iwatsubo, K., Bravo, C., Uechi, M., Baljinnayam, E., Nakamura, T., Umemura, M., et al. (2012). Prevention of heart failure in mice by an antiviral agent that inhibits type 5 cardiac adenyl cyclase. *Am. J. Physiology-Heart Circulatory Physiology* 302 (12), H2622–H2628. doi:10.1152/ajpheart.00190.2012

Iwatsubo, K., Minamisawa, S., Tsunematsu, T., Nakagome, M., Toya, Y., Tomlinson, J. E., et al. (2004). Direct inhibition of type 5 adenyl cyclase prevents myocardial apoptosis without functional deterioration. *J. Biol. Chem.* 279 (39), 40938–40945. doi:10.1074/jbc.M314238200

Iwatsubo, K., Tsunematsu, T., and Ishikawa, Y. (2003). Isoform-specific regulation of adenyl cyclase: a potential target in future pharmacotherapy. *Expert Opin. Ther. targets* 7 (3), 441–451. doi:10.1517/14728222.7.3.441

Jacobowitz, O., Chen, J., Premont, R., and Iyengar, R. (1993). Stimulation of specific types of Gs-stimulated adenyl cyclases by phorbol ester treatment. *J. Biol. Chem.* 268 (6), 3829–3832. doi:10.1016/s0021-9258(18)53547-x

Jaggupilli, A., Dhanaraj, P., Pritchard, A., Sorensen, J. L., Dakshinamurti, S., and Chelikani, P. (2018). Study of adenyl cyclase-GaS interactions and identification of novel AC ligands. *Mol. Cell Biochem.* 446 (1–2), 63–72. doi:10.1007/s11010-018-3273-4

Jiang, M. T., Moffat, M. P., and Narayanan, N. (1993). Age-related alterations in the phosphorylation of sarcoplasmic reticulum and myofibrillar proteins and diminished contractile response to isoproterenol in intact rat ventricle. *Circulation Res.* 72 (1), 102–111. doi:10.1161/01.res.72.1.102

Joe, E. K., Schussheim, A. E., Longrois, D., Mäki, T., Kelly, R. A., Smith, T. W., et al. (1998). Regulation of cardiac myocyte contractile function by inducible nitric oxide synthase (iNOS): mechanisms of contractile depression by nitric oxide. *J. Mol. Cell. Cardiol.* 30 (2), 303–315. doi:10.1006/jmcc.1997.0593

John, R. M., and Kumar, S. (2016). Sinus node and atrial arrhythmias. *Circulation* 133 (19), 1892–1900. doi:10.1161/CIRCULATIONAHA.116.018011

Kamenetsky, M., Middelhaufe, S., Bank, E. M., Levin, L. R., Buck, J., and Steegborn, C. (2006). Molecular details of cAMP generation in mammalian cells: a tale of two systems. *J. Mol. Biol.* 362 (4), 623–639. doi:10.1016/j.jmb.2006.07.045

Kamide, T., Okumura, S., Ghosh, S., Shinoda, Y., Mototani, Y., Ohnuki, Y., et al. (2015). Oscillation of cAMP and Ca^{2+} in cardiac myocytes: a systems biology approach. *J. physiological Sci.* 65 (2), 195–200. doi:10.1007/s12576-014-0354-3

Kapiloff, M. S., Piggott, L. A., Sadana, R., Li, J., Heredia, L. A., Henson, E., et al. (2009). An adenyl cyclase-mAKAPbeta signaling complex regulates cAMP levels in cardiac myocytes. *J. Biol. Chem.* 284 (35), 23540–23546. doi:10.1074/jbc.M109.030072

Kashou, A. H., Basit, H., and Chhabra, L. (2017). *Physiology, sinoatrial node*.

Kawabe, J.-i., Ebina, T., Toya, Y., Oka, N., Schwencke, C., Duzic, E., et al. (1996). Regulation of type V adenyl cyclase by PMA-sensitive and -insensitive protein kinase C isoenzymes in intact cells. *FEBS Lett.* 384 (3), 273–276. doi:10.1016/0014-5793(96)00331-6

Kim, N., Shin, S., and Bae, S. W. (2021). cAMP biosensors based on genetically encoded fluorescent/luminescent proteins. *Biosens. (Basel)* 11 (2), 39. doi:10.3390/bios11020039

Krishna, G., Weiss, B., and Brodie, B. B. (1968). A simple, sensitive method for the assay of adenyl cyclase. *J. Pharmacol. Exp. Ther.* 163 (2), 379–385.

Kritzer, M. D., Li, J., Dodge-Kafka, K., and Kapiloff, M. S. (2012). AKAPs: the architectural underpinnings of local cAMP signaling. *J. Mol. Cell Cardiol.* 52 (2), 351–358. doi:10.1016/j.jmcc.2011.05.002

Kryukova, Y. N., Protas, L., and Robinson, R. B. (2012). Ca^{2+} -activated adenyl cyclase 1 introduces Ca^{2+} -dependence to beta-adrenergic stimulation of HCN2 current. *J. Mol. Cell. Cardiol.* 52 (6), 1233–1239. doi:10.1016/j.jmcc.2012.03.010

Lai, H.-L., Lin, T.-H., Kao, Y.-Y., Lin, W.-J., Hwang, M.-J., and Chern, Y. (1999). The N terminus domain of type VI adenyl cyclase mediates its inhibition by protein kinase C. *Mol. Pharmacol.* 56 (3), 644–650. doi:10.1124/mol.56.3.644

Lai, H.-L., Yang, T.-H., Messing, R. O., Ching, Y.-H., Lin, S.-C., and Chern, Y. (1997). Protein kinase C inhibits adenyl cyclase type VI activity during desensitization of the A2a-adenosine receptor-mediated cAMP response. *J. Biol. Chem.* 272 (8), 4970–4977. doi:10.1074/jbc.272.8.4970

Lai, L., Yan, L., Gao, S., Hu, C.-L., Ge, H., Davidow, A., et al. (2013). Type 5 adenyl cyclase increases oxidative stress by transcriptional regulation of manganese superoxide dismutase via the SIRT1/FoxO3a pathway. *Circulation* 127 (16), 1692–1701. doi:10.1161/CIRCULATIONAHA.112.001212

Lai, L., Yan, L., Hu, C.-L., Gao, S., Iwatsubo, K., Ishikawa, Y., et al. (2009). Increased Type 5 adenyl cyclase expression mediates chronic catecholamine stress via increases in oxidative stress and down-regulation of MnSOD. *Am. Heart Assoc.*

Lai, N. C., Tang, T., Gao, M. H., Saito, M., Takahashi, T., Roth, D. M., et al. (2008). Activation of cardiac adenyl cyclase expression increases function of the failing ischemic heart in mice. *J. Am. Coll. Cardiol.* 51 (15), 1490–1497. doi:10.1016/j.jacc.2008.01.015

Lemaire, K., Van de Velde, S., Van Dijck, P., and Thevelein, J. M. (2004). Glucose and sucrose act as agonist and mannose as antagonist ligands of the G protein-coupled receptor Gpr1 in the yeast *Saccharomyces cerevisiae*. *Mol. Cell* 16 (2), 293–299. doi:10.1016/j.molcel.2004.10.004

Lezoualc'h, F., Fazal, L., Laudette, M., and Conte, C. (2016). Cyclic AMP sensor EPAC proteins and their role in cardiovascular function and disease. *Circulation Res.* 118 (5), 881–897. doi:10.1161/CIRCRESAHA.115.306529

Lin, T.-H., Lai, H.-L., Kao, Y.-Y., Sun, C.-N., Hwang, M.-J., and Chern, Y. (2002). Protein kinase C inhibits type VI adenyl cyclase by phosphorylating the regulatory N

- domain and two catalytic C1 and C2 domains. *J. Biol. Chem.* 277 (18), 15721–15728. doi:10.1074/jbc.M111537200
- Linder, J. U. (2006). Class III adenylyl cyclases: molecular mechanisms of catalysis and regulation. *Cell Mol. Life Sci.* 63 (15), 1736–1751. doi:10.1007/s00018-006-6072-0
- Liu, X., Thangavel, M., Sun, S. Q., Kaminsky, J., Mahautmr, P., Stitham, J., et al. (2008). Adenylyl cyclase type 6 overexpression selectively enhances beta-adrenergic and prostacyclin receptor-mediated inhibition of cardiac fibroblast function because of colocalization in lipid rafts. *Naunyn-Schmiedeberg's archives Pharmacol.* 377, 359–369. doi:10.1007/s00210-007-0196-0
- Liu, Y., Chen, J., Fontes, S. K., Bautista, E. N., and Cheng, Z. (2022). Physiological and pathological roles of protein kinase A in the heart. *Cardiovasc. Res.* 118 (2), 386–398. doi:10.1093/cvr/cvab008
- Maccarrone, M., Bari, M., Lorenzon, T., Bisogno, T., Di Marzo, V., and Finazzi-Agro, A. (2000). Anandamide uptake by human endothelial cells and its regulation by nitric oxide. *J. Biol. Chem.* 275 (18), 13484–13492. doi:10.1074/jbc.275.18.13484
- Mahmood, A., Ahmed, K., and Zhang, Y. (2022). β -Adrenergic receptor desensitization/down-regulation in heart failure: a friend or foe? *Front. Cardiovasc. Med.* 9, 925692. doi:10.3389/fcvm.2022.925692
- Maltsev, V. A., and Lakatta, E. G. (2012). The funny current in the context of the coupled-clock pacemaker cell system. *Heart rhythm.* 9 (2), 302–307. doi:10.1016/j.hrthm.2011.09.022
- Masri, B., Salahpour, A., Didriksen, M., Ghisi, V., Beaulieu, J.-M., Gainetdinov, R. R., et al. (2008). Antagonism of dopamine D2 receptor/beta-arrestin 2 interaction is a common property of clinically effective antipsychotics. *Proc. Natl. Acad. Sci.* 105 (36), 13656–13661. doi:10.1073/pnas.0803522105
- Matsumoto, T., Wakabayashi, K., Kobayashi, T., and Kamata, K. (2005). Functional changes in adenylyl cyclases and associated decreases in relaxation responses in mesenteric arteries from diabetic rats. *Am. J. Physiology-Heart Circulatory Physiology* 289 (5), H2234–H2243. doi:10.1152/ajpheart.00971.2004
- Mattick, P., Parrington, J., Oda, E., Simpson, A., Collins, T., and Terrar, D. (2007). Ca^{2+} -stimulated adenylyl cyclase isoform AC1 is preferentially expressed in Guinea-pig sino-atrial node cells and modulates the if pacemaker current. *J. physiology* 582 (3), 1195–1203. doi:10.1113/jphysiol.2007.133439
- McVey, M., Hill, J., Howlett, A., and Klein, C. (1999). Adenylyl cyclase, a coincidence detector for nitric oxide. *J. Biol. Chem.* 274 (27), 18887–18892. doi:10.1074/jbc.274.27.18887
- Mika, D., Richter, W., Westenbroek, R. E., Catterall, W. A., and Conti, M. (2014). PDE4B mediates local feedback regulation of β_1 -adrenergic cAMP signaling in a sarcolemmal compartment of cardiac myocytes. *J. Cell Sci.* 127 (5), 1033–1042. doi:10.1242/jcs.140251
- Milano, C. A., Allen, L. F., Rockman, H. A., Dolber, P. C., McMinn, T. R., Chien, K. R., et al. (1994). Enhanced myocardial function in transgenic mice overexpressing the beta 2-adrenergic receptor. *Science* 264 (5158), 582–586. doi:10.1126/science.8160017
- Mnatsakanyan, R., Markoutsas, S., Walbrunn, K., Roos, A., Verhelst, S. H., and Zahedi, R. P. (2019). Proteome-wide detection of S-nitrosylation targets and motifs using bioorthogonal cleavable-linker-based enrichment and switch technique. *Nat. Commun.* 10 (1), 2195. doi:10.1038/s41467-019-10182-4
- Moen, J. M., Matt, M. G., Ramirez, C., Tarasov, K. V., Chakir, K., Tarasova, Y. S., et al. (2019). Overexpression of a neuronal type adenylyl cyclase (type 8) in sinoatrial node markedly impacts heart rate and rhythm. *Front. Neurosci.* 13, 615. doi:10.3389/fnins.2019.00615
- Mongillo, M., McSorley, T., Evellin, S., Sood, A., Lissandron, V., Terrin, A., et al. (2004). Fluorescence resonance energy transfer-based analysis of cAMP dynamics in live neonatal rat cardiac myocytes reveals distinct functions of compartmentalized phosphodiesterases. *Circulation Res.* 95 (1), 67–75. doi:10.1161/01.RES.0000134629.84732.11
- Montero, D., Walther, G., Pérez-Martin, A., Vicente-Salar, N., Roche, E., and Vinet, A. (2013). Vascular smooth muscle function in type 2 diabetes mellitus: a systematic review and meta-analysis. *Diabetologia* 56, 2122–2133. doi:10.1007/s00125-013-2974-1
- Mou, T.-C., Masada, N., Cooper, D. M., and Sprang, S. R. (2009). Structural basis for inhibition of mammalian adenylyl cyclase by calcium. *Biochemistry* 48 (15), 3387–3397. doi:10.1021/bi802122k
- Mougenot, N., Mika, D., Czibik, G., Marcos, E., Abid, S., Houssaini, A., et al. (2019). Cardiac adenylyl cyclase overexpression precipitates and aggravates age-related myocardial dysfunction. *Cardiovasc. Res.* 115 (12), 1778–1790. doi:10.1093/cvr/cvy306
- Nakano, S. J., Sucharov, J., Van Dusen, R., Cecil, M., Nunley, K., Wickers, S., et al. (2017). Cardiac adenylyl cyclase and phosphodiesterase expression profiles vary by age, disease, and chronic phosphodiesterase inhibitor treatment. *J. cardiac Fail.* 23 (1), 72–80. doi:10.1016/j.cardfail.2016.07.429
- Nelson, C. P., Rainbow, R. D., Brignell, J. L., Perry, M. D., Willets, J. M., Davies, N. W., et al. (2011). Principal role of adenylyl cyclase 6 in K^+ channel regulation and vasodilator signalling in vascular smooth muscle cells. *Cardiovasc. Res.* 91 (4), 694–702. doi:10.1093/cvr/cvr137
- Nikolaev, V. O., Moshkov, A., Lyon, A. R., Miragoli, M., Novak, P., Paur, H., et al. (2010). Beta2-adrenergic receptor redistribution in heart failure changes cAMP compartmentation. *Science* 327 (5973), 1653–1657. doi:10.1126/science.1185988
- Nystoriak, M. A., Nieves-Cintrón, M., Patriarchi, T., Buonarati, O. R., Prada, M. P., Morotti, S., et al. (2017). Ser1928 phosphorylation by PKA stimulates the L-type Ca^{2+} channel $\text{Ca}_v1.2$ and vasoconstriction during acute hyperglycemia and diabetes. *Sci. Signal.* 10 (463), eaaf9647. doi:10.1126/scisignal.aaf9647
- Odaka, H., Arai, S., Inoue, T., and Kitaguchi, T. (2014). Genetically-encoded yellow fluorescent cAMP indicator with an expanded dynamic range for dual-color imaging. *PLoS one* 9 (6), e100252. doi:10.1371/journal.pone.0100252
- Ohta, Y., Furuta, T., Nagai, T., and Horikawa, K. (2018). Red fluorescent cAMP indicator with increased affinity and expanded dynamic range. *Sci. Rep.* 8 (1), 1866. doi:10.1038/s41598-018-20251-1
- Okumura, S., Kawabe, J.-i., Yatani, A., Takagi, G., Lee, M.-C., Hong, C., et al. (2003a). Type 5 adenylyl cyclase disruption alters not only sympathetic but also parasympathetic and calcium-mediated cardiac regulation. *Circulation Res.* 93 (4), 364–371. doi:10.1161/01.RES.0000086986.35568.63
- Okumura, S., Suzuki, S., and Ishikawa, Y. (2009). New aspects for the treatment of cardiac diseases based on the diversity of functional controls on cardiac muscles: effects of targeted disruption of the type 5 adenylyl cyclase gene. *J. Pharmacol. Sci.* 109 (3), 354–359. doi:10.1254/jphs.08r26fm
- Okumura, S., Takagi, G., Kawabe, J.-i., Yang, G., Lee, M.-C., Hong, C., et al. (2003b). Disruption of type 5 adenylyl cyclase gene preserves cardiac function against pressure overload. *Proc. Natl. Acad. Sci.* 100 (17), 9986–9990. doi:10.1073/pnas.1733772100
- Okumura, S., Tsunematsu, T., Bai, Y., Jiao, Q., Ono, S., Suzuki, S., et al. (2008). Type 5 adenylyl cyclase plays a major role in stabilizing heart rate in response to microgravity induced by parabolic flight. *J. Appl. Physiology* 105 (1), 173–179. doi:10.1152/japplphysiol.01166.2007
- Okumura, S., Vatner, D. E., Kurotani, R., Bai, Y., Gao, S., Yuan, Z., et al. (2007). Disruption of type 5 adenylyl cyclase enhances desensitization of cyclic adenosine monophosphate signal and increases Akt signal with chronic catecholamine stress. *Circulation* 116 (16), 1776–1783. doi:10.1161/CIRCULATIONAHA.107.698662
- Oldham, S. B., Rude, R. K., Molloy, C. T., Lipson, L. G., and Boggs, T. T. (1984). The effects of magnesium on calcium inhibition of parathyroid adenylate cyclase. *Endocrinology* 115 (5), 1883–1890. doi:10.1210/endo-115-5-1883
- Onda, T., Hashimoto, Y., Nagai, M., Kuramochi, H., Saito, S., Yamazaki, H., et al. (2001). Type-specific regulation of adenylyl cyclase: selective pharmacological stimulation and inhibition of adenylyl cyclase isoforms. *J. Biol. Chem.* 276 (51), 47785–47793. doi:10.1074/jbc.M107233200
- Ostrom, K. F., LaVigne, J. E., Brust, T. F., Seifert, R., Dessauer, C. W., Watts, V. J., et al. (2022). Physiological roles of mammalian transmembrane adenylyl cyclase isoforms. *Physiol. Rev.* 102 (2), 815–857. doi:10.1152/physrev.00013.2021
- Ostrom, R. S., Bunday, R. A., and Insel, P. A. (2004). Nitric oxide inhibition of adenylyl cyclase type 6 activity is dependent upon lipid rafts and caveolin signaling complexes. *J. Biol. Chem.* 279 (19), 19846–19853. doi:10.1074/jbc.M313440200
- Ostrom, R. S., Gregorian, C., Drenan, R. M., Xiang, Y., Regan, J. W., and Insel, P. A. (2001). Receptor number and caveolar co-localization determine receptor coupling efficiency to adenylyl cyclase. *J. Biol. Chem.* 276 (45), 42063–42069. doi:10.1074/jbc.M105348200
- Ostrom, R. S., and Insel, P. A. (2004). The evolving role of lipid rafts and caveolae in G protein-coupled receptor signaling: implications for molecular pharmacology. *Br. J. Pharmacol.* 143 (2), 235–245. doi:10.1038/sj.bjp.0705930
- Ostrom, R. S., Liu, X., Head, B. P., Gregorian, C., Seasholtz, T. M., and Insel, P. A. (2002). Localization of adenylyl cyclase isoforms and G protein-coupled receptors in vascular smooth muscle cells: expression in caveolin-rich and noncaveolin domains. *Mol. Pharmacol.* 62 (5), 983–992. doi:10.1124/mol.62.5.983
- Ostrom, R. S., Naugle, J. E., Hase, M., Gregorian, C., Swaney, J. S., Insel, P. A., et al. (2003). Angiotensin II enhances adenylyl cyclase signaling via Ca^{2+} /Calmodulin: gq-Gs cross-talk regulates collagen production in cardiac fibroblasts. *J. Biol. Chem.* 278 (27), 24461–24468. doi:10.1074/jbc.M212659200
- Ostrom, R. S., Violin, J. D., Coleman, S., and Insel, P. A. (2000). Selective enhancement of beta-adrenergic receptor signaling by overexpression of adenylyl cyclase type 6: colocalization of receptor and adenylyl cyclase in caveolae of cardiac myocytes. *Mol. Pharmacol.* 57 (5), 1075–1079.
- Packer, M., Carver, J. R., Rodeheffer, R. J., Ivanhoe, R. J., DiBianco, R., Zeldis, S. M., et al. (1991). Effect of oral milrinone on mortality in severe chronic heart failure. The PROMISE Study Research Group. *N. Engl. J. Med.* 325 (21), 1468–1475. doi:10.1056/NEJM199111233252103
- Paramonov, V. M., Mamaeva, V., Sahlgren, C., and Rivero-Müller, A. (2015). Genetically-encoded tools for cAMP probing and modulation in living systems. *Front. Pharmacol.* 6, 196. doi:10.3389/fphar.2015.00196
- Park, M., Park, J. Y., Lee, J. A., Tian, B., Lai, L., Iwatsubo, K., et al. (2011). Cardiac overexpression of adenylyl cyclase type 5 induces left ventricular hypertrophy potentially by activating calcineurin-NFAT signaling. Wiley Online Library.
- Patel, N., and Gold, M. G. (2015). The genetically encoded tool set for investigating cAMP: more than the sum of its parts. *Front. Pharmacol.* 6, 164. doi:10.3389/fphar.2015.00164

- Pavan, B., Biondi, C., and Dalpiaz, A. (2009). Adenylyl cyclases as innovative therapeutic goals. *Drug Discov. today* 14 (19–20), 982–991. doi:10.1016/j.drudis.2009.07.007
- Penny, W. F., Henry, T. D., Watkins, M. W., Patel, A. N., and Hammond, H. K. (2018). Design of a Phase 3 trial of intracoronary administration of human adenovirus 5 encoding human adenylyl cyclase type 6 (RT-100) gene transfer in patients with heart failure with reduced left ventricular ejection fraction: the FLOURISH Clinical Trial. *Am. Heart J.* 201, 111–116. doi:10.1016/j.ahj.2018.04.005
- Phan, H. M., Gao, M. H., Lai, N. C., Tang, T., and Hammond, H. K. (2007). New signaling pathways associated with increased cardiac adenylyl cyclase 6 expression: implications for possible congestive heart failure therapy. *Trends Cardiovasc. Med.* 17 (7), 215–221. doi:10.1016/j.tcm.2007.07.001
- Piacenza, L., Zeida, A., Trujillo, M., and Radi, R. (2022). The superoxide radical switch in the biology of nitric oxide and peroxynitrite. *Physiol. Rev.* 102 (4), 1881–1906. doi:10.1152/physrev.00005.2022
- Pierre, S., Eschenhagen, T., Geisslinger, G., and Scholich, K. (2009). Capturing adenylyl cyclases as potential drug targets. *Nat. Rev. Drug Discov.* 8 (4), 321–335. doi:10.1038/nrd2827
- Ping, P., Anzai, T., Gao, M., and Hammond, H. (1997). Adenylyl cyclase and G protein receptor kinase expression during development of heart failure. *Am. J. Physiology-Heart Circulatory Physiology* 273 (2), H707–H717. doi:10.1152/ajpheart.1997.273.2.H707
- Ponsioen, B., Zhao, J., Riedl, J., Zwartkruis, F., van der Krogt, G., Zaccolo, M., et al. (2004). Detecting cAMP-induced Epac activation by fluorescence resonance energy transfer: Epac as a novel cAMP indicator. *EMBO Rep.* 5 (12), 1176–1180. doi:10.1038/sj.embor.7400290
- Prada, M. P., Syed, A. U., Buonarati, O. R., Reddy, G. R., Nystoriak, M. A., Ghosh, D., et al. (2019). A Gs-coupled purinergic receptor boosts Ca²⁺ influx and vascular contractility during diabetic hyperglycemia. *Elife* 8, e42214. doi:10.7554/eLife.42214
- Prada, M. P., Syed, A. U., Reddy, G. R., Martín-Aragón Baudel, M., Flores-Tamez, V. A., Sasse, K. C., et al. (2020). AKAP5 complex facilitates purinergic modulation of vascular L-type Ca²⁺ channel CaV1.2. *Nat. Commun.* 11 (1), 5303. doi:10.1038/s41467-020-18947-y
- Prinz, A., Diskar, M., Erlbruch, A., and Herberg, F. W. (2006). Novel, isotype-specific sensors for protein kinase A subunit interaction based on bioluminescence resonance energy transfer (BRET). *Cell. Signal.* 18 (10), 1616–1625. doi:10.1016/j.cellsig.2006.01.013
- Qi, C., Sorrentino, S., Medalia, O., and Korkhov, V. M. (2019). The structure of a membrane adenylyl cyclase bound to an activated stimulatory G protein. *Science* 364 (6438), 389–394. doi:10.1126/science.aav0778
- Qu, J.-H., Chakir, K., Tarasov, K. V., Riordon, D. R., Perino, M. G., Silvester, A. J., et al. (2024). Reprogramming of cardiac phosphoproteome, proteome, and transcriptome confers resilience to chronic adenylyl cyclase-driven stress. *Elife* 12, RP88732. doi:10.7554/eLife.88732
- Rana, N., Conley, J. M., Soto-Velasquez, M., León, F., Cutler, S. J., Watts, V. J., et al. (2017). Molecular modeling evaluation of the enantiomers of a novel adenylyl cyclase 2 inhibitor. *J. Chem. Inf. Model.* 57 (2), 322–334. doi:10.1021/acs.jcim.6b00454
- Ren, L., Thai, P. N., Gopireddy, R. R., Timofeyev, V., Ledford, H. A., Woltz, R. L., et al. (2022). Adenylyl cyclase isoform 1 contributes to sinoatrial node automaticity via functional microdomains. *JCI insight* 7 (22), e162602. doi:10.1172/jci.insight.162602
- Ridnour, L. A., Thomas, D. D., Mancardi, D., Espey, M. G., Miranda, K. M., Paolocci, N., et al. (2004). The chemistry of nitrosative stress induced by nitric oxide and reactive nitrogen oxide species. *Putt. perspective stressful Biol. situations.* doi:10.1515/BC.2004.001
- Ripoll, L., and Von Zastrow, M. (2023). Spatial organization of adenylyl cyclase and its impact on dopamine signaling in neurons. *bioRxiv*, 2023.12.06.570478. 2023.2012.2006.570478. doi:10.1101/2023.12.06.570478
- Robinson, R. B., Dun, W., and Boyden, P. A. (2021). Autonomic modulation of sinoatrial node: role of pacemaker current and calcium sensitive adenylyl cyclase isoforms. *Prog. Biophysics Mol. Biol.* 166, 22–28. doi:10.1016/j.pbiomolbio.2020.08.004
- Roth, D. M., Bayat, H., Drumm, J. D., Gao, M. H., Swaney, J. S., Ander, A., et al. (2002). Adenylyl cyclase increases survival in cardiomyopathy. *Circulation* 105 (16), 1989–1994. doi:10.1161/01.cir.0000014968.54967.d3
- Roth, D. M., Gao, M. H., Lai, N. C., Drumm, J., Dalton, N., Zhou, J. Y., et al. (1999). Cardiac-directed adenylyl cyclase expression improves heart function in murine cardiomyopathy. *Circulation* 99 (24), 3099–3102. doi:10.1161/01.cir.99.24.3099
- Ruhparwar, A., Kallenbach, K., Klein, G., Bara, C., Ghodsizad, A., Sigg, D. C., et al. (2010). Adenylate-cyclase VI transforms ventricular cardiomyocytes into biological pacemaker cells. *Tissue Eng. Part A* 16 (6), 1867–1872. doi:10.1089/ten.TEA.2009.0537
- Sadana, R., and Dessauer, C. W. (2009). Physiological roles for G protein-regulated adenylyl cyclase isoforms: insights from knockout and overexpression studies. *Neurosignals* 17 (1), 5–22. doi:10.1159/000166277
- Schmid, A., Meili, D., and Salathe, M. (2014). Soluble adenylyl cyclase in health and disease. *Biochimica Biophysica Acta (BBA)-Molecular Basis Dis.* 1842 (12), 2584–2592. doi:10.1016/j.bbdis.2014.07.010
- Schubert, W., Yang, X. Y., Yang, T. T., Factor, S. M., Lisanti, M. P., Molkentin, J. D., et al. (2003). Requirement of transcription factor NFAT in developing atrial myocardium. *J. Cell Biol.* 161 (5), 861–874. doi:10.1083/jcb.200301058
- Seifert, R., and Beste, K. Y. (2012). Allosteric regulation of nucleotidyl cyclases: an emerging pharmacological target. *Sci. Signal.* 5 (240), pe37. doi:10.1126/scisignal.2003466
- Sena, C. M., Pereira, A. M., and Seica, R. (2013). Endothelial dysfunction—a major mediator of diabetic vascular disease. *Biochimica Biophysica Acta (BBA)-Molecular Basis Dis.* 1832 (12), 2216–2231. doi:10.1016/j.bbdis.2013.08.006
- Seth, A., Finkbeiner, M., Grischin, J., and Schultz, J. E. (2020). Gsa stimulation of mammalian adenylate cyclases regulated by their hexahelical membrane anchors. *Cell Signal* 68, 109538. doi:10.1016/j.cellsig.2020.109538
- Shi, J., Yang, Y., Cheng, A., Xu, G., and He, F. (2020). Metabolism of vascular smooth muscle cells in vascular diseases. *Am. J. Physiology-Heart Circulatory Physiology* 319, H613–H631. doi:10.1152/ajpheart.00220.2020
- Sikarwar, A. S., Hinton, M., Santhosh, K. T., Dhanaraj, P., Talabis, M., Chelikani, P., et al. (2018). Hypoxia inhibits adenylyl cyclase catalytic activity in a porcine model of persistent pulmonary hypertension of the newborn. *Am. J. Physiol. Lung Cell Mol. Physiol.* 315 (6), L933–L944. doi:10.1152/ajplung.00130.2018
- Sinha, S., and Sprang, S. (2006). Structures, mechanism, regulation and evolution of class III nucleotidyl cyclases. *Rev. Physiology Biochem. Pharmacol.* 157, 105–140. doi:10.1007/112_0603
- Spangler, C. M., Spangler, C., Gottle, M., Shen, Y., Tang, W. J., Seifert, R., et al. (2008a). A fluorimetric assay for real-time monitoring of adenylyl cyclase activity based on terbium norfloxacin. *Anal. Biochem.* 381 (1), 86–93. doi:10.1016/j.ab.2008.06.014
- Spangler, C. M., Spangler, C., and Schaeferling, M. (2008b). Luminescent lanthanide complexes as probes for the determination of enzyme activities. *Ann. N. Y. Acad. Sci.* 1130, 138–148. doi:10.1196/annals.1430.008
- Steer, M. L., and Levitzki, A. (1975). The control of adenylate cyclase by calcium in Turkey erythrocyte ghosts. *J. Biol. Chem.* 250 (6), 2080–2084. doi:10.1016/s0021-9258(19)41685-2
- Storch, U., Straub, J., Erdogmus, S., Gudermann, T., and Mederos y Schnitzler, M. (2017). Dynamic monitoring of G i/o-protein-mediated decreases of intracellular cAMP by FRET-based Epac sensors. *Pflügers Archiv-European J. Physiology* 469, 725–737. doi:10.1007/s00424-017-1975-1
- Streeto, J. M., and Reddy, W. J. (1967). An assay for adenylyl cyclase. *Anal. Biochem.* 21 (3), 416–426. doi:10.1016/0003-2697(67)90316-8
- Sugano, Y., Lai, N. C., Gao, M. H., Firth, A. L., Yuan, J.X.-J., Lew, W. Y., et al. (2011). Activated expression of cardiac adenylyl cyclase 6 reduces dilation and dysfunction of the pressure-overloaded heart. *Biochem. biophysical Res. Commun.* 405 (3), 349–355. doi:10.1016/j.bbrc.2010.12.113
- Sunahara, R. K., Dessauer, C. W., and Gilman, A. G. (1996). Complexity and diversity of mammalian adenylyl cyclases. *Annu. Rev. Pharmacol. Toxicol.* 36 (1), 461–480. doi:10.1146/annurev.pa.36.040196.002333
- Sunahara, R. K., Dessauer, C. W., Whisnant, R. E., Kleuss, C., and Gilman, A. G. (1997). Interaction of G α with the cytosolic domains of mammalian adenylyl cyclase. *J. Biol. Chem.* 272 (35), 22265–22271. doi:10.1074/jbc.272.35.22265
- Syed, A. U., Reddy, G. R., Ghosh, D., Prada, M. P., Nystoriak, M. A., Morotti, S., et al. (2019). Adenylyl cyclase 5-generated cAMP controls cerebral vascular reactivity during diabetic hyperglycemia. *J. Clin. investigation* 129 (8), 3140–3152. doi:10.1172/JCI124705
- Takahashi, T., Tang, T., Lai, N. C., Roth, D. M., Rebollo, B., Saito, M., et al. (2006). Increased cardiac adenylyl cyclase expression is associated with increased survival after myocardial infarction. *Circulation* 114 (5), 388–396. doi:10.1161/CIRCULATIONAHA.106.632513
- Tan, C. M., Kelvin, D. J., Litchfield, D. W., Ferguson, S. S., and Feldman, R. D. (2001). Tyrosine kinase-mediated serine phosphorylation of adenylyl cyclase. *Biochemistry* 40 (6), 1702–1709. doi:10.1021/bi0015818
- Tan, Y.-Q., Li, J., and Chen, H.-W. (2022). Epac, a positive or negative signaling molecule in cardiovascular diseases. *Biomed. Pharmacother.* 148, 112726. doi:10.1016/j.biopha.2022.112726
- Tan, Z., Giamouridis, D., Lai, N. C., Kim, Y. C., Guo, T., Xia, B., et al. (2019). Cardiac-directed expression of adenylyl cyclase catalytic domain (C1C2) attenuates deleterious effects of pressure overload. *Hum. Gene Ther.* 30 (6), 682–692. doi:10.1089/hum.2018.176
- Tang, T., Gao, M. H., Lai, N. C., Firth, A. L., Takahashi, T., Guo, T., et al. (2008). Adenylyl cyclase type 6 deletion decreases left ventricular function via impaired calcium handling. *Circulation* 117 (1), 61–69. doi:10.1161/CIRCULATIONAHA.107.730069
- Tang, T., Gao, M. H., Roth, D. M., Guo, T., and Hammond, H. K. (2004). Adenylyl cyclase type VI corrects cardiac sarcoplasmic reticulum calcium uptake defects in cardiomyopathy. *Am. J. Physiology-Heart Circulatory Physiology* 287 (5), H1906–H1912. doi:10.1152/ajpheart.00356.2004
- Tang, T., Hammond, H. K., Firth, A., Yang, Y., Gao, M. H., Yuan, J.X.-J., et al. (2011). Adenylyl cyclase 6 improves calcium uptake and left ventricular function in aged hearts. *J. Am. Coll. Cardiol.* 57 (18), 1846–1855. doi:10.1016/j.jacc.2010.11.052

- Tang, T., Lai, N. C., Hammond, H. K., Roth, D. M., Yang, Y., Guo, T., et al. (2010). Adenylyl cyclase 6 deletion reduces left ventricular hypertrophy, dilation, dysfunction, and fibrosis in pressure-overloaded female mice. *J. Am. Coll. Cardiol.* 55 (14), 1476–1486. doi:10.1016/j.jacc.2009.11.066
- Tang, T., Lai, N. C., Roth, D. M., Drumm, J., Guo, T., Lee, K., et al. (2006). Adenylyl cyclase type V deletion increases basal left ventricular function and reduces left ventricular contractile responsiveness to beta-adrenergic stimulation. *Basic Res. Cardiol.* 101, 117–126. doi:10.1007/s00395-005-0559-y
- Tang, T., Lai, N. C., Wright, A. T., Gao, M. H., Lee, P., Guo, T., et al. (2013). Adenylyl cyclase 6 deletion increases mortality during sustained β -adrenergic receptor stimulation. *J. Mol. Cell. Cardiol.* 60, 60–67. doi:10.1016/j.yjmcc.2013.04.005
- Tang, W.-J., and Gilman, A. G. (1995). Construction of a soluble adenylyl cyclase activated by Gs alpha and forskolin. *Science* 268 (5218), 1769–1772. doi:10.1126/science.7792604
- Tao, Y.-P., Najafi, L., Shipley, S., Howlett, A., and Klein, C. (1998). Effects of nitric oxide on adenylyl cyclase stimulation in N18TG2 neuroblastoma cells. *J. Pharmacol. Exp. Ther.* 286 (1), 298–304.
- Tarasov, K. V., Chakir, K., Riordon, D. R., Lyashkov, A. E., Ahmet, I., Perino, M. G., et al. (2022). A remarkable adaptive paradigm of heart performance and protection emerges in response to marked cardiac-specific overexpression of ADCY8. *Elife* 11, e80949. doi:10.7554/eLife.80949
- Taussig, R., Tang, W.-J., Hepler, J. R., and Gilman, A. G. (1994). Distinct patterns of bidirectional regulation of mammalian adenylyl cyclases. *J. Biol. Chem.* 269 (8), 6093–6100. doi:10.1016/s0021-9258(17)37574-9
- Tepe, N. M., and Liggett, S. B. (1999). Transgenic replacement of type V adenylyl cyclase identifies a critical mechanism of beta-adrenergic receptor dysfunction in the G alpha q overexpressing mouse. *FEBS Lett.* 458 (2), 236–240. doi:10.1016/s0014-5793(99)01147-3
- Tesmer, J. J., Dessauer, C. W., Sunahara, R. K., Murray, L. D., Johnson, R. A., Gilman, A. G., et al. (2000). Molecular basis for P-site inhibition of adenylyl cyclase. *Biochemistry* 39 (47), 14464–14471. doi:10.1021/bi0015562
- Tesmer, J. J., Sunahara, R. K., Gilman, A. G., and Sprang, S. R. (1997). Crystal structure of the catalytic domains of adenylyl cyclase in a complex with Gsalpha.GTPgammaS. *Science* 278 (5345), 1907–1916. doi:10.1126/science.278.5345.1907
- Tewson, P., Martinka, S., Shaner, N., Berlot, C., Quinn, A. M., and Hughes, T. (2018). Assay for detecting Gai-mediated decreases in cAMP in living cells. *SLAS Discov. Adv. Life Sci. R&D* 23 (9), 898–906. doi:10.1177/2472555218786238
- Thangavel, M., Liu, X., Sun, S. Q., Kaminsky, J., and Ostrom, R. S. (2009). The C1 and C2 domains target human type 6 adenylyl cyclase to lipid rafts and caveolae. *Cell. Signal.* 21 (2), 301–308. doi:10.1016/j.cellsig.2008.10.017
- Timofeyev, V., Myers, R. E., Kim, H. J., Woltz, R. L., Sirish, P., Heiserman, J. P., et al. (2013). Adenylyl cyclase subtype-specific compartmentalization: differential regulation of L-type Ca^{2+} current in ventricular myocytes. *Circulation Res.* 112 (12), 1567–1576. doi:10.1161/CIRCRESAHA.112.300370
- Tobise, K., Ishikawa, Y., Holmer, S., Im, M., Newell, J., Yoshie, H., et al. (1994). Changes in type VI adenylyl cyclase isoform expression correlate with a decreased capacity for cAMP generation in the aging ventricle. *Circulation Res.* 74 (4), 596–603. doi:10.1161/01.res.74.4.596
- Toya, Y., Schwencke, C., and Ishikawa, Y. (1998). Forskolin derivatives with increased selectivity for cardiac adenylyl cyclase. *J. Mol. Cell. Cardiol.* 30 (1), 97–108. doi:10.1006/jmcc.1997.0575
- Tsunematsu, T., Okumura, S., Mototani, Y., Ohnuki, Y., Jin, H., Cai, W., et al. (2015). Coupling of β 1-adrenergic receptor to type 5 adenylyl cyclase and its physiological relevance in cardiac myocytes. *Biochem. Biophys. Res. Commun.* 458 (3), 531–535. doi:10.1016/j.bbrc.2015.01.149
- Tsutsui, K., Florio, M. C., Yang, A., Wirth, A. N., Yang, D., Kim, M. S., et al. (2021). cAMP-dependent signaling restores AP firing in dormant SA node cells via enhancement of surface membrane currents and calcium coupling. *Front. Physiology* 12, 596832. doi:10.3389/fphys.2021.596832
- Tykocki, N. R., Boerman, E. M., and Jackson, W. F. (2017). Smooth muscle ion channels and regulation of vascular tone in resistance arteries and arterioles. *Compr. Physiol.* 7 (2), 485–581. doi:10.1002/cphy.c160011
- Valkovic, A. L., Leckey, M. B., Whitehead, A. R., Hossain, M. A., Inoue, A., Kocan, M., et al. (2018). Real-time examination of cAMP activity at relaxin family peptide receptors using a BRET-based biosensor. *Pharmacol. Res. Perspect.* 6 (5), e00432. doi:10.1002/prp2.432
- Vatner, D. E., Yan, L., Lai, L., Yuan, C., Mouchiroud, L., Pachon, R. E., et al. (2015). Type 5 adenylyl cyclase disruption leads to enhanced exercise performance. *Aging Cell* 14 (6), 1075–1084. doi:10.1111/accel.12401
- Vatner, S. F., Park, M., Yan, L., Lee, G. J., Lai, L., Iwatsubo, K., et al. (2013). Adenylyl cyclase type 5 in cardiac disease, metabolism, and aging. *Am. J. Physiology-Heart Circulatory Physiology* 305 (1), H1–H8. doi:10.1152/ajpheart.00080.2013
- Vila-Petroff, M. G., Younes, A., Egan, J., Lakatta, E. G., and Sollott, S. J. (1999). Activation of distinct cAMP-dependent and cGMP-dependent pathways by nitric oxide in cardiac myocytes. *Circulation Res.* 84 (9), 1020–1031. doi:10.1161/01.res.84.9.1020
- von Hayn, K., Werthmann, R. C., Nikolaev, V. O., Hommers, L. G., Lohse, M. J., and Bunemann, M. (2010). Gq-mediated Ca^{2+} signals inhibit adenylyl cyclases 5/6 in vascular smooth muscle cells. *Am. J. Physiol. Cell Physiol.* 298 (2), C324–C332. ajpcell.00197.2009 [pii]. doi:10.1152/ajpcell.00197.2009
- Wang, T., and Brown, M. (2004). Differential expression of adenylyl cyclase subtypes in human cardiovascular system. *Mol. Cell. Endocrinol.* 223 (1–2), 55–62. doi:10.1016/j.mce.2004.05.012
- Warrier, S., Belevych, A. E., Ruse, M., Eckert, R. L., Zaccolo, M., Pozzan, T., et al. (2005). Beta-adrenergic- and muscarinic receptor-induced changes in cAMP activity in adult cardiac myocytes detected with FRET-based biosensor. *Am. J. Physiology-Cell Physiology* 289 (2), C455–C461. doi:10.1152/ajpcell.00058.2005
- Watson, E. L., Singh, J. C., Jacobson, K. L., and Ott, S. M. (2001). Nitric oxide inhibition of cAMP synthesis in parotid acini: regulation of type 5/6 adenylyl cyclase. *Cell. Signal.* 13 (10), 755–763. doi:10.1016/s0898-6568(01)00204-2
- White, M., Roden, R., Minobe, W., Khan, M. F., Larrabee, P., Wollmering, M., et al. (1994). Age-related changes in beta-adrenergic neuroeffector systems in the human heart. *Circulation* 90 (3), 1225–1238. doi:10.1161/01.cir.90.3.1225
- Willoughby, D., and Cooper, D. M. (2007). Organization and Ca^{2+} regulation of adenylyl cyclases in cAMP microdomains. *Physiol. Rev.* 87 (3), 965–1010. doi:10.1152/physrev.00049.2006
- Wu, G.-C., Lai, H.-L., Lin, Y.-W., Chu, Y.-T., and Chern, Y. (2001). N-glycosylation and residues Asn805 and Asn890 are involved in the functional properties of type VI adenylyl cyclase. *J. Biol. Chem.* 276 (38), 35450–35457. doi:10.1074/jbc.M009704200
- Wu, Y., and Jiang, T. (2022). Developments in FRET- and BRET-based biosensors. *Micromachines* 13 (10), 1789. doi:10.3390/mi13101789
- Wu, Y.-S., Chen, C.-C., Chien, C.-L., Lai, H.-L., Jiang, S.-T., Chen, Y.-C., et al. (2017). The type VI adenylyl cyclase protects cardiomyocytes from β -adrenergic stress by a PKA/STAT3-dependent pathway. *J. Biomed. Sci.* 24 (1), 68–12. doi:10.1186/s12929-017-0367-3
- Yan, L., Park, J. Y., Dillinger, J. G., De Lorenzo, M. S., Yuan, C., Lai, L., et al. (2012). Common mechanisms for calorie restriction and adenylyl cyclase type 5 knockout models of longevity. *Aging Cell* 11 (6), 1110–1120. doi:10.1111/accel.12013
- Yan, L., Vatner, D. E., O'Connor, J. P., Ivessa, A., Ge, H., Chen, W., et al. (2007). Type 5 adenylyl cyclase disruption increases longevity and protects against stress. *Cell* 130 (2), 247–258. doi:10.1016/j.cell.2007.05.038
- Yokochi, A., Itoh, H., Maruyama, J., Zhang, E., Jiang, B., Mitani, Y., et al. (2010). Colforsin-induced vasodilation in chronic hypoxic pulmonary hypertension in rats. *J. Anesth.* 24 (3), 432–440. doi:10.1007/s00540-010-0912-7
- Yokoyama, U., Minamisawa, S., Katayama, A., Tang, T., Suzuki, S., Iwatsubo, K., et al. (2010). Differential regulation of vascular tone and remodeling via stimulation of type 2 and type 6 adenylyl cyclases in the ductus arteriosus. *Circulation Res.* 106 (12), 1882–1892. doi:10.1161/CIRCRESAHA.109.214924
- Yoneyama, M., Sugiyama, A., Satoh, Y., Takahara, A., Nakamura, Y., and Hashimoto, K. (2002). Cardiovascular and adenylate cyclase stimulating effects of colforsin daropate, a water-soluble forskolin derivative, compared with those of isoproterenol, dopamine and dobutamine. *Circulation J.* 66 (12), 1150–1154. doi:10.1253/circj.66.1150
- Younes, A., Lyashkov, A. E., Graham, D., Sheydina, A., Volkova, M. V., Mitsak, M., et al. (2008). Ca^{2+} -stimulated basal adenylyl cyclase activity localization in membrane lipid microdomains of cardiac sinoatrial nodal pacemaker cells. *J. Biol. Chem.* 283 (21), 14461–14468. doi:10.1074/jbc.M707540200
- Zhang, J., Levy, D., Oydanic, M., Bravo, C. A., Yoon, S., Vatner, D. E., et al. (2018). A novel adenylyl cyclase type 5 inhibitor that reduces myocardial infarct size even when administered after coronary artery reperfusion. *J. Mol. Cell. Cardiol.* 121, 13–15. doi:10.1016/j.yjmcc.2018.05.014
- Zhang, X., Szeto, C., Gao, E., Tang, M., Jin, J., Fu, Q., et al. (2013). Cardiotoxic and cardioprotective features of chronic β -adrenergic signaling. *Circulation Res.* 112 (3), 498–509. doi:10.1161/CIRCRESAHA.112.273896
- Zhang, Y., Chen, S., Luo, L., Greenly, S., Shi, H., Xu, J. J., et al. (2023). Role of cAMP in cardiomyocyte viability: beneficial or detrimental? *Circulation Res.* 133 (11), 902–923. doi:10.1161/CIRCRESAHA.123.322652
- Zheng, M., Zhu, W., Han, Q., and Xiao, R. P. (2005). Emerging concepts and therapeutic implications of beta-adrenergic receptor subtype signaling. *Pharmacol. Ther.* 108 (3), 257–268. doi:10.1016/j.pharmthera.2005.04.006



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NLRP3 inflammasome and pyroptosis in cardiovascular diseases and exercise intervention

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NOD-like receptor protein 3 (NLRP3) inflammasome is an intracellular sensing protein complex that possesses NACHT, leucine-rich repeat, and pyrin domain, playing a crucial role in innate immunity. Activation of the NLRP3 inflammasome leads to the production of pro-inflammatory cellular contents, such as interleukin (IL)-1 β and IL-18, and induction of inflammatory cell death known as pyroptosis, thereby amplifying or sustaining inflammation. While a balanced inflammatory response is beneficial for resolving damage and promoting tissue healing, excessive activation of the NLRP3 inflammasome and pyroptosis can have harmful effects. The involvement of the NLRP3 inflammasome has been observed in various cardiovascular diseases (CVD). Indeed, the NLRP3 inflammasome and its associated pyroptosis are closely linked to key cardiovascular risk factors including hyperlipidemia, diabetes, hypertension, obesity, and hyperhomocysteinemia. Exercise compared with medicine is a highly effective measure for both preventing and treating CVD. Interestingly, emerging evidence suggests that exercise improves CVD and inhibits the activity of NLRP3 inflammasome and pyroptosis. In this review, the activation mechanisms of the NLRP3 inflammasome and its pathogenic role in CVD are critically discussed. Importantly, the purpose is to emphasize the crucial role of exercise in managing CVD by suppressing NLRP3 inflammasome activity and proposes it as the foundation for developing novel treatment strategies.

KEYWORDS

cardiovascular disease, exercise, pyroptosis, NLRP3 inflammasome, intervention

1 Introduction

Cardiovascular diseases (CVD) remain a prevalent global health concern, causing a significant burden of illness and mortality, with approximately one-third of all deaths attributed to this condition (Mensah et al., 2019). The common symptoms of CVD include chest pain, shortness of breath, irregular heartbeat, fatigue, and decreased physical stamina (Tutor et al., 2023; Wong and Sattar, 2023). CVD encompass various disorders that affect the heart and blood vessels, including atherosclerosis (AS), obesity, diabetes, hyperhomocysteinemia (HHcy), myocardial infarction (MI), hypertension, heart failure (HF), and diabetic cardiomyopathy (DCM) (Konishi et al., 2022; Haidar and Horwich, 2023). The conventional risk factors that are widely recognized for CVD, such as hypertension, hypercholesterolemia, diabetes mellitus, and cigarette smoking, remain

acknowledged as the main factors responsible for the development and advancement of this condition (Alten et al., 2020).

NOD-like receptor protein 3 (NLRP3) inflammasome is a molecular platform that triggers caspase-1 and facilitates the secretion of interleukin (IL)-1 β and IL-18 in response to cellular infection or stress (Valenzuela et al., 2023). This activation results in the cleavage of gasdermin D (GSDMD) by caspase-1, generating an N-terminal GSDMD fragment (Valenzuela et al., 2023). This fragment induces the formation of membrane pores and triggers inflammatory cell death, namely pyroptosis (Valenzuela et al., 2023). The involvement of NLRP3 inflammasome and pyroptosis has been established in cardiovascular risk factors such as hyperlipidemia, diabetes, hypertension, obesity, and HHcy (Alten et al., 2020). Targeting NLRP3 inflammasome activation and pyroptosis holds great potential for therapeutic interventions against CVD.

Extensive research has demonstrated that exercise plays a crucial role in weight management (Murray et al., 2023), blood pressure (BP) reduction (Tucker et al., 2022), blood sugar (Kar et al., 2019) and lipid regulation (Lee et al., 2020), consequently lowering the risk of CVD (Li et al., 2023). Moreover, moderate exercise improves cardiovascular system function and structure, strengthens the heart muscle, enhances cardio-pulmonary function, promotes blood circulation, and increases the heart's tolerance and overall health (Alten et al., 2020). Furthermore, exercise exhibits a clear anti-inflammatory effect (Lee et al., 2020). It is widely acknowledged that exercise can effectively reduce chronic inflammation by inhibiting the expression of inflammatory factors while increasing the release of anti-inflammatory cytokines (Lee et al., 2020). Previous studies have indicated that exercise can decrease the activation of the NLRP3 inflammasome to substantially inhibit IL-1 β , and IL-18 release (Li et al., 2023; Liu et al., 2023). This review mainly focuses on how exercise can improve CVD by influencing NLRP3 inflammasome or pyroptosis. Additionally, this review will investigate exercise as a therapeutic strategy via NLRP3 inflammasome for managing CVD and address the research questions that need to be explored in the future.1.

2 Overview of NLRP3 inflammasome

Inflammasomes are crucial components of the immune system that play a major role in initiating inflammatory responses (Chang, 2023). They are composed of sensor proteins known as pattern recognition receptors that oligomerize and form a platform for the activation of caspase-1 in response to damage-associated molecular patterns (DAMPs) or pathogen-associated molecular patterns (PAMPs) (Chang, 2023). The nucleotide-binding domain-like receptor (NLR) family all share a central nucleotide-binding domain, and most members have a C-terminal leucine-rich repeat (LRR) domain and a variable N-terminal domain (Swanson et al., 2019). While certain members such as NLRP1, NLRP3, and NLRC4 are recognized as NLRs capable of forming inflammasomes, others like NLRP6 and NLRP12 are considered potential inflammasome sensors (Swanson et al., 2019). The NLRP3 inflammasome, in particular, is essential for the host's immune defense against various bacterial, fungal, and viral infections (Dilucca et al., 2021). However, when dysregulated, it has been implicated in the development of several inflammatory disorders, including CVD (Paerewijck and Lamkanfi, 2022).

The NLRP3 inflammasome is composed of several key components. The NLR protein in the NLRP3 inflammasome contains a conserved nucleotide-binding and oligomerization domain, C-terminal LRRs, and a pyrin domain (PYD) that facilitates multimerization (Xi et al., 2024). Upon activation of the NLRP3 inflammasome, the NLRs oligomerize through their nucleotide-binding and oligomerization domains (Fu and Wu, 2023). This leads to the recruitment of the adaptor protein apoptosis-associated speck-like protein (ASC) through PYD-PYD interactions (Fu and Wu, 2023). ASC then forms large speck-like structures and recruits pro-caspase-1 through caspase recruitment domain (CARD)-CARD interactions. Pro-caspase-1 undergoes autocatalytic cleavage, resulting in the formation of active caspase-1 p10/p20 tetramers (Ruan, 2019). These active caspase-1 tetramers mediate the maturation and secretion of IL-1 β and IL-18 (Ruan, 2019; Fu and Wu, 2023). Additionally, caspase-1 can cleave GSDMD to generate GSDMD n-terminal (NT). GSDMD-NT forms plasma membrane pores, leading to the induction of pyroptosis (Lee et al., 2021; Nie et al., 2021). The canonical activation of inflammasomes is proposed to occur in two steps: priming and assembly (Bockstiegel et al., 2023). Priming involves the initial activation of toll-like receptors by agonists such as lipopolysaccharide (LPS) (Fu and Wu, 2023). This trigger signaling cascades, primarily through the nuclear factor- κ B (NF- κ B) pathway, which leads to the transcriptional upregulation of pro-inflammatory mediators like pro-IL-1 β . Following priming, the activated inflammasome assembles in response to various PAMPs or DAMPs (Fu and Wu, 2023; Krantz et al., 2023). This assembly forms a large molecular platform that activates inflammatory caspases and processes pro-IL-1 β (Huang et al., 2023). The activation of NLRP3 inflammasome requires "priming" with TLR agonists to initiate signaling cascades (primarily nuclear factor- κ B (NF- κ B)-dependent pathway) that ultimately promote a transcriptional response to upregulate pro-inflammatory mediators (Figure 1).

K⁺ efflux is recognized as a crucial upstream signal for NLRP3 inflammasome activation (Zhou et al., 2020). Activation of NLRP3 also requires the mobilization of Ca²⁺ (Diaz-Del-Olmo et al., 2021). Mobilization of Ca²⁺ occurs when extracellular Ca²⁺ moves across channels in the plasma membrane and the Ca²⁺ stored in the endoplasmic reticulum is released into the cytoplasm, which can induce NLRP3 inflammasome activation (Watanabe et al., 2020; Diaz-Del-Olmo et al., 2021). In addition, Cl⁻ was implicated in NLRP3 activation. Cl⁻ channel blockers and elevated levels of extracellular Cl⁻ can inhibit, whereas reduced levels of Cl⁻ can enhance, the activation of NLRP3 (Zhou et al., 2020). Cl⁻ efflux may be downstream of K⁺ efflux and affects ASC polymerization, whereas K⁺ efflux promotes NLRP3 oligomerization (Zhou et al., 2024).

In addition to the ion channels mentioned earlier that can activate the NLRP3 inflammasome, organelle dysfunction can also trigger inflammasome activation (Movahedpour et al., 2023). In a study, sterile lysosomal rupture caused by L-leucyl-L-leucine methyl ester is sufficient to trigger NLRP3 inflammasome activation, whereas suppression of phagosomal acidification or cathepsin B blocks NLRP3 activation (Movahedpour et al., 2023). Mitochondrial dysfunction contributes to the activation of the NLRP3 inflammasome (Kodi et al., 2024). When damaged

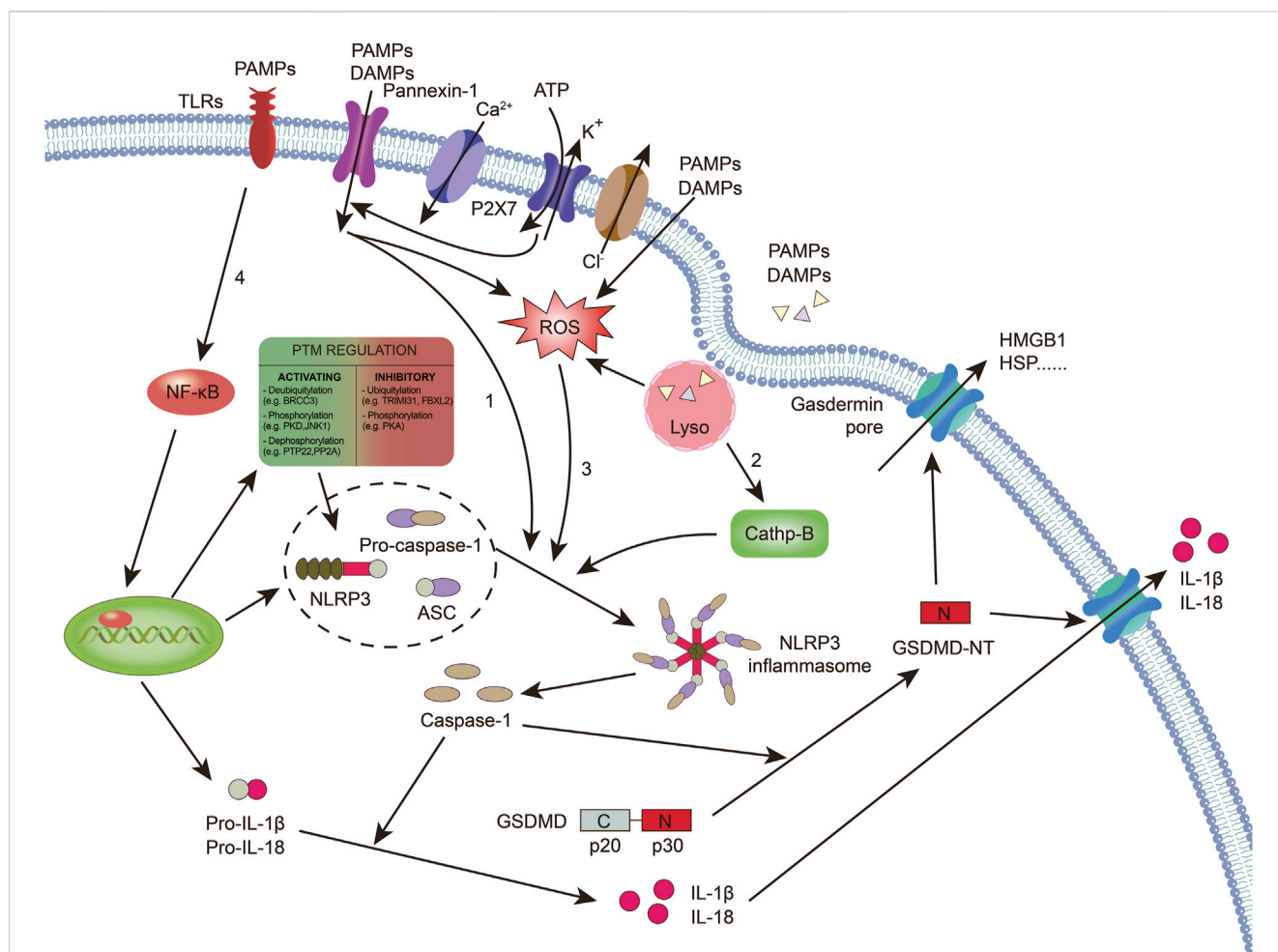


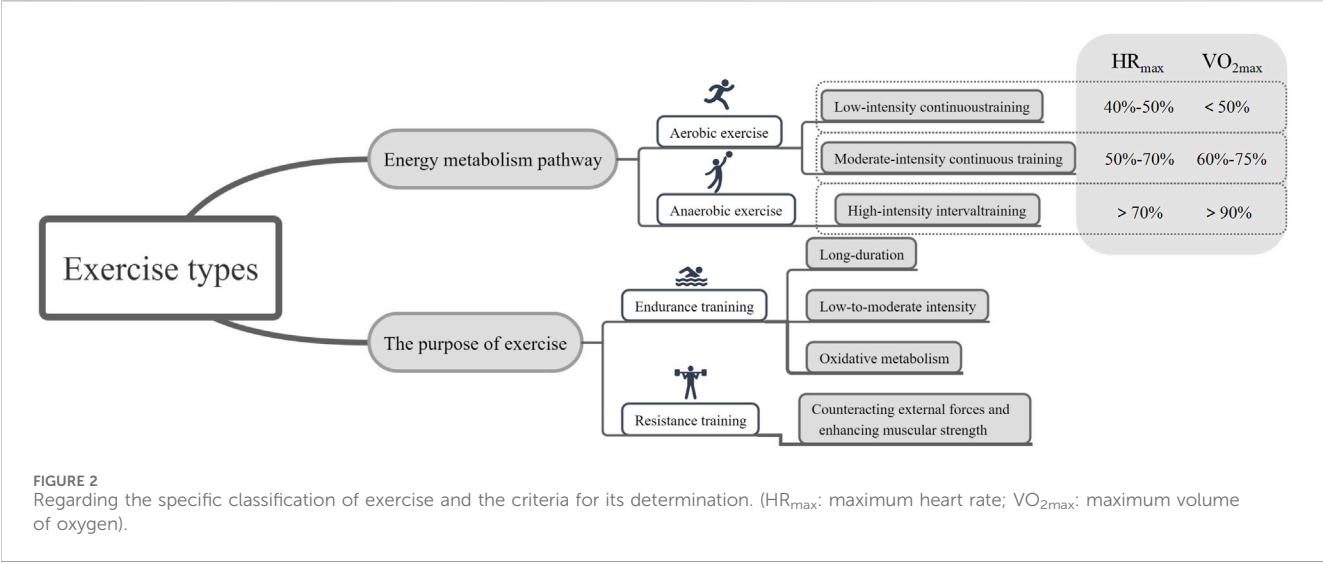
FIGURE 1

Caspase-1-dependent canonical pyroptotic cell death induced by NLRP3 inflammasome activation. The assembly of the NLRP3 inflammasome involves NLRP3 oligomerization and ASC recruitment, triggering the autocleavage of pro-caspase-1. This autocleavage leads to the activation of caspase-1, which converts inactive pro-IL-1 β and pro-IL-18 into their bioactive and secreted forms, namely IL-1 β and IL-18. Additionally, active caspase-1 cleaves GSDMD, generating GSDMD-NT, which forms pores on the plasma membrane, inducing pyroptosis. Various models have been proposed to explain the assembly of the NLRP3 inflammasome. 1): Extracellular ATP can activate the NLRP3 inflammasome through different mechanisms. This includes the activation of the P2X7 receptor leading to the opening of the pannexin-1 pore, allowing the entry of extracellular factors that directly interact with NLRP3. Alternatively, NLRP3 can sense either the efflux of K⁺ or the loss of membrane integrity. 2): Crystalline or particulate agonists can be phagocytosed, resulting in the release of lysosomal cathepsins B and L, which are detected by the NLRP3 inflammasome. 3): NLRP3 agonists such as DAMPs and PAMPs can trigger the production of ROS, which activates the NLRP3 inflammasome. It is important to note that these models are not mutually exclusive but rather interact with each other. 4): The activation of the NLRP3 inflammasome also requires "priming" with TLR agonists, such as LPS. (ASC: apoptosis-associated speck-like protein; TLRs: toll-like receptors; DAMPs: damage-associated molecular patterns; GSDMD: gasdermin D; NT: n-terminal; HSP: heat shock proteins; HMGB-1: high mobility group box-1; IL: interleukin; NLRP3: NOD-like receptor protein 3; NF- κ B: nuclear factor- κ B; PAMPs: pathogen-associated molecular patterns; P2X7: purinergic receptor P2X, ligand-gated ion channel 7; ROS: reactive oxygen species).

organelles accumulate due to deficiencies in autophagic proteins, dysfunctional mitochondria release mitochondrial DNA, leading to NLRP3 inflammasome activation (Kodi et al., 2024). Additionally, mitochondrial ROS (mtROS) can initiate NLRP3 activation (Kodi et al., 2024). Inhibiting autophagy or mitophagy results in the buildup of mitochondrial ROS and subsequent NLRP3 activation (Chen et al., 2022; Lewisluján et al., 2022; Zhang et al., 2022; Lu et al., 2023).

Post-transcriptional modifications of the NLRP3 protein occur in unstimulated cells and during priming and activation to modulate its activation and function (Swanson et al., 2019). In peritoneal macrophages, tripartite motif 31 ubiquitylates NLRP3, targeting it for proteasomal degradation (Chan and Schroder, 2020). The

E3 ubiquitin ligase f-box and leucine-rich repeat protein 2 are suggested to prevent NLRP3 activation by directing it to the proteasome in lung epithelial cells (Han et al., 2020). Additionally, membrane-associated ring-ch-type finger 7, another E3 ubiquitin ligase, inhibits NLRP3 downstream of dopamine-induced D1 receptor signaling (Sandall and Macdonald, 2019). Another regulatory level involves phosphorylation of NLRP3 at Ser291 (or Ser295 in humans) by protein kinase A, leading to K48- and K63-linked ubiquitination and subsequent proteasomal degradation (Ren et al., 2019). Song et al. Demonstrated that following LPS priming, NLRP3 is phosphorylated on Ser194 by c-Jun N-terminal kinase 1, promoting NLRP3 oligomerization upon activation by canonical stimuli (Spalinger et al., 2023). In monocytic



cells, a study revealed that dephosphorylation of NLRP3 at Tyr861 by protein tyrosine phosphatase nonreceptor 22 induces NLRP3 activation (Spalinger et al., 2020). Whether these pathways synergize to tightly control NLRP3 activity remains elusive.

3 Mechanisms of pyroptosis

Recent advances have shed light on the molecular mechanisms of pyroptosis, a form of programmed cell death induced by the agonist of the NLRP3 inflammasome (McKee and Coll, 2020; Yuan et al., 2020; Chen et al., 2022; Zhong et al., 2023). Among these mechanisms, GSDMD has emerged as a crucial mediator of pyroptosis. GSDMD belongs to a family of proteins named GSDMs, which share a pore-forming domain (McKee and Coll, 2020). Cleavage of GSDMD by caspase-1 or caspase-4/5/11 releases GSDMD-NT, the NT domain of GSDMD (McKee and Coll, 2020). GSDMD-NT then forms these pores in the plasma membrane, leading to cell swelling and osmotic lysis (Ren et al., 2019). Other members of the GSDM family also possess pore-forming activity but are not targeted by inflammatory caspases (Chen et al., 2022). The cleavage of GSDMD occurs at a conserved residue called D276, resulting in the separation of GSDMD into two domains: GSDMD-NT (p30) and GSDMD C-terminal domain (p20) (Zhong et al., 2023). GSDMD-NT can interact with lipids in the plasma membrane and assemble into large oligomeric pores (Yuan et al., 2020). This disruption of the cell membrane integrity leads to an increase in intracellular osmotic pressure and the release of inflammatory intracellular contents, including high mobility group box-1 (HMGB-1) and heat shock protein (Yuan et al., 2020; Chan et al., 2023). This process, characterized by caspase-1-dependent cleavage of GSDMD, is known as the pyroptotic pathway (Chan et al., 2023).

4 Overview of exercise

Exercise refers to the movement of the body, typically involving the musculoskeletal system, to maintain health, improve physical

fitness, promote cardiovascular health, and enhance the overall quality of life (Vella et al., 2017). It is a crucial component for maintaining both physical and mental wellbeing (Vella et al., 2017). By selecting forms of exercise that suit individual needs and goals, comprehensive health benefits can be achieved (Vella et al., 2017). The most common classification of exercise is based on the primary energy metabolism pathways, dividing it into aerobic and anaerobic exercise (Paluch et al., 2024) (Figure 2).

Aerobic exercise is a form of activity that generates energy through oxidative metabolism (Mueller et al., 2021). This type of exercise involves relatively low to moderate intensity over an extended period to ensure the body can supply enough oxygen to support energy production (Mueller et al., 2021). Aerobic exercise is characterized by lower intensity, safety, rhythmic, and sustained durations, with relatively minor stress on various organs, reducing the risk of exercise-related injuries (Mueller et al., 2021). Common aerobic exercise programs include low-intensity continuous training (LICT) and moderate-intensity continuous training (MICT) (Troosters et al., 2023). The intensity of exercise is typically measured using parameters such as maximum heart rate and maximum oxygen consumption (Blanks et al., 2019). For maximum heart rate (HR_{max}), low-intensity exercise falls within the 40%–50% range, while moderate-intensity exercise falls within the 50%–70% HR_{max} range (Blanks et al., 2019). Regarding the maximum volume of oxygen (VO_{2max}), values below 60% are considered low intensity, and those between 60% and 75% are considered moderate intensity (Blanks et al., 2019).

Anaerobic exercise involves high-intensity, momentary bursts of muscle activity in an “oxygen-deprived” state (Liao et al., 2022). It is characterized by high-intensity loads and brief durations, making it challenging to sustain for extended periods, and recovery from fatigue is slower (Liao et al., 2022). Anaerobic exercise enhances muscle strength, improves adaptability, and serves as a primary source of muscle growth (Casado et al., 2023). The intensity of anaerobic exercise is relatively high, and the sustainable duration is short, resulting in high-intensity loads that can lead to muscle fatigue and soreness (Casado et al., 2023). Common anaerobic exercise programs include resistance training (RT) and high-

intensity interval training (HIIT) (May et al., 2020). Recently, HIIT has gained popularity as a time-efficient exercise strategy that has been proven to improve cardiovascular risk factors in various populations (May et al., 2020). This training method employs alternating patterns of work and rest to enhance cardiorespiratory endurance, promote fat burning, and provide more efficient training effects in a shorter time (May et al., 2020). High-intensity training is identified by an HR_{max} exceeding 70% or VO_{2max} exceeding 90% (Blanks et al., 2019).

Additionally, based on the primary training goals, exercise can be classified into endurance training (ET) and RT (Consitt et al., 2019). ET involves prolonged, continuous activities at relatively low loads, primarily relying on aerobic metabolism to produce energy (Rothschild and Bishop, 2020). It induces adaptations in the cardiovascular and musculoskeletal systems, supporting overall improvements in exercise capacity and performance (Miko et al., 2020). However, for older individuals who are overweight or obese, especially those with symptoms of osteoarthritis, regular ET may be uncomfortable and painful, necessitating the introduction of alternative forms of exercise (Cutrufello et al., 2020).

RT, on the other hand, offers various health benefits, supports body weight, and avoids imposing impact stress on joints (Schoenfeld et al., 2016). Therefore, RT may be an appealing exercise modality for overweight/obese older individuals. This type of training emphasizes resistance against external forces to enhance muscle strength, endurance, and mass, with each effort specifically targeting the resistance generated, designed to increase muscle strength and explosiveness (Carvalho et al., 2022).

Although most literature suggests that exercise can improve CVD, excessive endurance exercise (EEE) may have many potential adverse effects on cardiac structure and function (O'Keefe et al., 2012). Acutely, EEE can increase myocardial injury markers, lead to chamber dilation, and decrease right ventricular function (Levine, 2014). Chronically, concerns exist that this degree of EEE may lead to detrimental cardiac remodeling and fibrosis, as well as non-lethal arrhythmias, especially an increased risk of atrial fibrillation, and potentially more lethal ventricular arrhythmias (Nath et al., 2023). Recent studies also indicate that despite a more favorable overall profile of coronary heart disease risk in long-distance runners, they may have higher levels of atherosclerosis and coronary heart disease risk (Neumann et al., 2022; O'Keefe et al., 2021). However, the benefits of aerobic exercise on CVD mortality appear to be significantly diminished when running exceeds 30 miles per week or walking exceeds 46 miles per week (Schwartz et al., 2014). Even though that lack of physical activity is more prevalent than EEE in the overall population, the potential adverse effects may be more serious on a societal level for overall health and cardiovascular health (McCullough and Lavie, 2014). These studies also suggest that more is not necessarily better, and even low doses of aerobic exercise, particularly running, seem to confer benefits for long-term cardiovascular health and lifespan.

5 Mechanisms by which exercise regulates NLRP3 inflammasome activation

Exercise has long been acknowledged as an effective intervention in regulating the innate immune response (Pope and Wood, 2020).

In general, LICT and MTCT have positive effects on the immune system, whereas HIIT tends to have the opposite effect (Xian et al., 2022). However, there is limited research on how exercise specifically influences the activity of the NLRP3 inflammasome.

From the perspective of material metabolism, many studies have reported that glucose and lipids can directly activate the NLRP3 inflammasome (Vandanmagsar et al., 2011; Zhang et al., 2021; Baik et al., 2023), while exercise can directly alleviate glucose and lipid metabolism, reducing the levels of blood sugar and lipids, thereby improving CVD. There is currently no literature reporting that exercise can directly regulate the activation pathway of NLRP3 inflammasome through glucose and lipid metabolism (Mardare et al., 2016; Ma et al., 2021). Therefore, it boldly speculates that the effect of exercise on NLRP3 inflammasome activation is likely to be exerted through the regulation of glucose and lipid metabolism.

From the mitochondrial standpoint, since most studies suggest that mitochondria are involved in regulating NLRP3 inflammasome activation (Chen et al., 2019; Hong et al., 2021), it is speculated that exercise adaptation of mitochondria may affect the NLRP3 inflammasome activity. Earlier studies demonstrated that MICT significantly decreased the expression of inflammatory cytokines TNF- α , IL-6, and monocyte chemoattractant protein-1 induced by metabolic disorders, coupled with an upregulation in mitochondrial proteins (Liu et al., 2017; Chen et al., 2018). Conversely, HIIT led to mitochondrial dysfunction and an augmentation in the secretion of pro-inflammatory factors (Memme et al., 2021). Moreover, additional investigations indicated that MICT facilitated mitochondrial biogenesis, bolstered antioxidant capacity, and restrained the overactivation of the NLRP3 inflammasome (Mason et al., 2020; Powers et al., 2020). Furthermore, aerobic exercise mitigated cardiac dysfunction by modulating the expression of proteins implicated in mitochondrial quality and NLRP3/caspase-1/IL-1 β signaling (Salo et al., 1991). In pathological conditions, mitochondrial damage leads to increased production of mtROS (Chen et al., 2020). The elevated mtROS mediates the activation of the NLRP3 inflammasome through thioredoxin interacting protein (TXNIP) and thioredoxin (Zhang et al., 2021). These findings imply that MICT might diminish mitochondrial ROS production through the regulation of mitochondrial quality control, enhancement of mitochondrial function, and the facilitation of damaged mitochondria clearance. Consequently, this could inhibit the NLRP3 inflammasome pathway and alleviate exaggerated inflammatory responses.

From molecular mechanisms in the cell, the key factors in exercise-mediated improvement of CVD through the NLRP3 inflammasome may be related to the regulation of the NF- κ B signaling pathway. Some studies have focused on the NF- κ B pathway as a critical node to explore the molecular mechanisms among exercise and NLRP3 inflammasome (Wang et al., 2019; Zhou et al., 2022). For example, research has demonstrated that aerobic exercise is capable of reducing the expression of nicotinamide adenine dinucleotide phosphate oxidase 4 (NOX4), ROS, TNF- α , IL-18, NF- κ B p65, and the NLRP3 inflammasome (Zhou et al., 2022). These findings suggest that exercise may ameliorate the pathological alterations in diabetes mellitus through the modulation of the NOX4/ROS/NF- κ B/NLRP3 inflammasome signaling cascade (Zhou et al., 2022). In a separate study, Wang

et al., 2019) observed that aerobic exercise suppresses the acetylation of forkhead box transcription factor O1 (FOXO1) in the brain tissue of diabetic rats, which in turn promotes the phosphorylation of FOXO1, thus inhibiting expression of NF- κ B and NLRP3 inflammasome protein. This downregulation contributes to the inhibition of inflammatory responses, indicating that exercise may exert anti-inflammatory effects via the FOXO1/NF- κ B/NLRP3 inflammasome pathway (Wang et al., 2019). Although studies have shown that exercise improves CVD through the NLRP3 inflammasome (Li et al., 2021; Zhou et al., 2022), further research is needed to elucidate the detailed molecular mechanisms by which exercise regulates the NLRP3 inflammasome.

6 Exercise, NLRP3 inflammasome, and CVD

6.1 Exercise improves AS and inhibits NLRP3 inflammasome

The pathogenesis of AS involves multiple processes, including endothelial dysfunction, low-density lipoprotein accumulation and oxidation, monocyte and lymphocyte recruitment, smooth muscle cell migration and proliferation, proinflammatory cytokine activation, and platelet adhesion (Zeng et al., 2019). Notably, the augmented release of inflammatory cytokines primarily attributed to endothelial cells, macrophages, and smooth muscle cell pyroptosis or NLRP3 inflammasome activation significantly contributes to AS formation and development (Zhaolin et al., 2019). Specifically, NLRP3 inflammasome-dependent pyroptosis triggers endothelial dysfunction, thereby acting as a catalyst for AS in these cells (Sun et al., 2017). This highlights the crucial role of NLRP3 inflammasome in promoting the release of inflammatory mediators and contributing to the pathological changes associated with AS.

Exercise can reduce the inflammatory death of local endothelial cells, slowing the development of AS plaques (Xu et al., 2019). A study found that voluntary wheel running, a natural type of aerobic exercise in the murine model could decrease the protein levels of the inflammasome components and markedly inhibit the caspase-1 activity in endothelial cells within the aorta of mice fed with a high-fat diet (HFD) (Lee et al., 2020). Recently, another study has demonstrated that exercise-induced a significant downregulation of m6A modification and methyltransferase-like 14 (Yang et al., 2023). This protein binds to the m6A sites of nuclear paraspeckle assembly transcript 1 (NEAT1) and promotes NEAT1 expression through subsequent YT521-B homology domain-containing 1, which transcriptionally promotes NLRP3 expression and endothelial pyroptosis (Yang et al., 2023). As a result, exercise effectively inhibits NLRP3 expression and endothelial pyroptosis, preventing AS plaque formation (Yang et al., 2023). In addition, it is widely recognized that fibroblast growth factor 21 (FGF21), a well-established negative risk factor for AS expressed in the aorta, exerts inhibitory effects on NLRP3 inflammasome activity, thereby attenuating aortic pyroptosis to prevent AS development (Zeng et al., 2020). Interestingly, aerobic exercise increases FGF21 sensitivity while downregulating the expression of pyroptosis-related proteins mediated by NLRP3 inflammasome in

the aorta (Li et al., 2022). These findings suggest that activated FGF21 may be involved in aerobic exercise inhibiting NLRP3 inflammasome-mediated pyroptosis in the aorta. However, the current study has not investigated whether exercise improved AS via inhibiting NLRP3 inflammasome and pyroptosis in these cells, which needs further research in the future.

6.2 Exercise improves HHcy and inhibits NLRP3 inflammasome

Homocysteine (Hcy), a non-essential amino acid sulfur, is derived from methionine and is used for methylation of DNA/RNA methylation (Veeranki and Tyagi, 2013). HHcy refers to a condition where plasma Hcy levels exceed 15 μ mol/L and has been associated with various diseases especially AS (Winchester et al., 2014). HHcy promotes the generation of ROS through mechanisms such as mixed disulfide formation and auto-oxidation (Veeranki and Tyagi, 2013; Winchester et al., 2014). Recent research has demonstrated that HHcy-induced activation of the NLRP3 inflammasome in macrophages contributes to vascular inflammation and AS by activating caspase-1-mediated pyroptosis (Wang et al., 2017). In addition, acid sphingomyelinase upregulation by Hcy promotes clustering of lipid rafts mediating the assembly of NADPH oxidase complex resulting in increased ROS generation followed by NLRP3 inflammasome activation leading to pyroptosis contributing towards the development of AS (Liu et al., 2022).

Although direct evidence is currently lacking regarding whether exercise specifically modulates NLRP3 inflammasome activation or pyroptosis about HHcy, it has been observed that exercise can lower Hcy levels (Liu et al., 2022). In a folate-restricted model of HHcy, exercise can reduce the increase in plasma Hcy levels by increasing betaine Hcy S-methyltransferase levels in the kidneys and promoting nonclassical remethylation to convert Hcy into methionine (Vincent et al., 2006; Neuman et al., 2013). Moreover, apart from reducing plasma Hcy levels, exercise holds the potential for mitigating Hcy-induced lipid peroxidation and ameliorating reductions in superoxide dismutase and catalase activity, both of which are implicated in the progression of AS (Neuman et al., 2013). These works suggest that exercise might also inhibit NLRP3 inflammasome activation or pyroptosis as a part of its overall impact on reducing HHcy-related complications. Given these findings, further investigation is warranted to explore whether exercise precisely influences NLRP3 inflammasome activation or pyroptosis and their role in HHcy management. Understanding this relationship could provide valuable insights into developing targeted interventions for individuals with elevated Hcy levels and associated CVD.

6.3 Exercise improves obesity and inhibits NLRP3 inflammasome

Obesity, commonly associated with chronic low-grade inflammation, is characterized by the pathological enlargement of adipose tissue (AT) primarily due to excess energy accumulation as fat (Spalding et al., 2008; Wada et al., 2017). Mice with HFD-induced

obesity exhibit increased expression of caspase-1, ASC, and NLRP3. However, knocking out the *Nlrp3* or *Caspase-1* gene suppresses obesity-induced fat depot (Stienstra et al., 2011). Therefore, targeting the activation of NLRP3 inflammasome or pyroptosis represents a promising approach for improving obesity.

A study found that exercise decreased protein expression of inflammasome components (NLRP3 and caspase-1) in bone marrow-derived macrophages (BMDM) and AT isolated from mice with diet-induced obesity (Javaid et al., 2021). Another study investigated perform mice fed either a standard diet or an HFD and subjected to regular ET or RT and discovered that RT attenuated increased NLRP3 expression and reduced levels of IL-18 in isolated AT, while ET effectively reduced the expression of TNF- α and IL-18 in supernatant from AT, suggesting that exercise can reduce inflammasome activation in ATs and achieve systemic downregulation of inflammatory cytokines (Wada et al., 2017). Besides AT, endothelial dysfunction emerges early on in CVD associated with obesity (Stienstra et al., 2011; Wada et al., 2017). A study demonstrated that engaging in voluntary running while on an HFD led to a significant reduction in active caspase-1 levels within the endothelial cells lining the aorta when compared to sedentary mice on the same diet (Li et al., 2023). These findings indicate that voluntary running alleviates impaired blood vessel function via inhibiting NLRP3 inflammasome activation (Li et al., 2023). In addition to endothelial cells, exercise suppressed NLRP3 inflammasome activation as revealed by downregulated IL-1 β and IL-18 in BMDM (Li et al., 2023). This body of research demonstrates that exercise exerts inhibitory effects on NLRP3 inflammasome activation across various cell types to ameliorate obesity.

It's worth noting that a human study found that exercise reduces plasma IL-18 levels in obese individuals, indirectly suggesting the inhibition of the NLRP3 pathway through exercise to improve obesity (Stienstra et al., 2011). Furthermore, 8-week high-intensity and aerobic interval training (three times/week) in men and women with metabolic syndrome resulted in decreased IL-18 mRNA levels in abdominal AT and a numerical decrease in plasma IL-18 concentration (Stienstra et al., 2011; Stienstra et al., 2011; Stienstra et al., 2011; Stienstra et al., 2011). Similarly, a pilot study, conducted among thirty-seven obese individuals demonstrates that exercise intervention, primarily consisting of activities such as walking, jogging, and functional exercise circuits designed to enhance aerobic capacity and speed, leads to significant reductions in ASC mRNA expression levels when compared to the hypocaloric group without any form of exercise participation (Barrón et al., 2020). These findings indicate an inverse relationship between ASC mRNA expression and aerobic interventions. Another randomized controlled trial involving 36 obese inactive subjects further delineated the type of exercise intervention and demonstrated both HIIT and MICT significantly reduced NLRP3 gene expression in serum samples from all subjects, strongly suggesting that diversity intensity interval training can inhibit NLRP3 inflammasome in obese (Armanna et al., 2022).

Therefore, exercise is considered a crucial strategy for reducing inflammation and metabolic disorders associated with obesity. Further research will enhance understanding of the complex relationship between exercise and NLRP3 inflammasome, leading to more effective interventions for managing obesity-related diseases.

6.4 Exercise improves diabetes and inhibits NLRP3 inflammasome

Diabetes is a metabolic disease that poses a significant threat to human health. Type 2 diabetes, a significant risk factor for both microvascular and macrovascular diseases, accounts for 90% of diabetes cases and is a leading cause of death, particularly due to coronary heart disease (Einarson et al., 2018). Inflammatory processes play a crucial role in the development of complications associated with diabetes (Roncero-Ramos et al., 2018). Recently, NLRP3 inflammasome and pyroptosis have emerged as key contributors to insulin resistance (IR) (Roncero-Ramos et al., 2018). Peripheral blood-derived macrophages from drug-naïve patients with type 2 diabetes show increased expression of NLRP3 and ASC along with caspase-1 activation and IL-1 β maturation (Lee et al., 2013). *In vivo* and *in vitro* studies have reported that high glucose stimulates IL-1 β secretion in pancreatic β cells, resulting in their death through the activation of NLRP3 inflammasome (Zhou et al., 2010; Wu et al., 2019). The various modes of activating the NLRP3 inflammasome are fundamental factors influencing its complex effects on the progression of type 2 diabetes (Chen et al., 2021).

Engaging in aerobic exercise can improve IR and reduce the expression of NLRP3 and IL-1 β in individuals with type 2 diabetes in aortic tissue, suggesting its positive impact on IR by inhibiting NLRP3 inflammasome (Hassanpour Soleimani et al., 2021). HMGB1 is a major pro-inflammatory cytokine released as a result of pyroptosis (Bigueti et al., 2019). This work also found that exercise can reduce the secretion of HMGB1, which is significantly increased in individuals with diabetes and contributes to disease progression (Hassanpour Soleimani et al., 2021). Previous studies have shown that exercise has the potential to lower HMGB-1 levels in circulation and tissues of diabetic patients, possibly by inhibiting NLRP3 inflammasome activation and pyroptosis (Haß et al., 2022; Haß et al., 2023). Furthermore, exercise decreased circulating levels of IL-1, which may potentially protect against IL-1-mediated destruction of β -cells (Stumvoll et al., 2005). Although there is currently no direct evidence on whether aerobic exercise specifically affects NLRP3 inflammasome activation or pyroptosis and thus decreases levels of blood glucose, these findings highlight the significant role played by exercise in regulating IR through its impact on NLRP3 inflammasome and pyroptosis.

6.5 Exercise improves DCM and inhibits NLRP3 inflammasome

DCM is a distinct cardiac phenotype observed in diabetic patients characterized by structural changes such as cardiac hypertrophy, cardiomyocyte death, and fibrosis, as well as functional abnormalities (Sun et al., 2021). The molecular mechanisms underlying DCM involve various factors including hyperglycemia, IR, fatty acids, oxidative stress, mitochondrial dysfunction, inflammation, and endothelial dysfunction (Chen et al., 2020). In particular, inflammation is believed to be present in the early stages of diabetes and plays a crucial role in promoting DCM (Luo et al., 2017). Emerging evidence has verified that

NLRP3 inflammasome-mediated cardiomyocyte pyroptosis is a key participant in DCM (Xie et al., 2020). Human diabetic hearts also exhibit elevated activation of the NLRP3 inflammasome and cardiac pyroptosis compared to non-diabetic heart tissues (Zhang et al., 2015; Xie et al., 2020). Moreover, high glucose levels (35 mM) significantly induce increased protein expression of NLRP3, caspase-1, and IL-1 β accompanied by noticeable cardiomyocyte pyroptosis, leading to loss of contractile units and cardiac dysfunction (Zhang et al., 2015). In contrast, silencing the *Nlrp3* gene ameliorates cardiac inflammation and pyroptosis and improves cardiac function by ameliorating cardiac inflammation and pyroptosis both *in vivo* and *in vitro* experiments (Chen et al., 2020).

Exercise intervention has been shown to effectively prevent and treat DCM by modulating the NLRP3 inflammasome. For example, the expressions of NLRP3, caspase-1-p20, caspase-1p20/caspase-1, and IL-1 β were increased in the myocardium of HFD-induced obese mice (Lee et al., 2018). However, treadmill exercise inhibited these parameters (Sun et al., 2021). This demonstrates that exercise training can prevent obesity-induced cardiac inflammasome formation, pyroptosis activation, and pro-inflammatory response (Sun et al., 2021). In addition, recent studies in DCM mice have demonstrated that although exercise has a limited impact on interstitial fibrosis, it can effectively reverse cardiac dysfunction by reducing the activity of NLRP3 inflammasome and inhibiting pyroptosis (Takahashi, 2019; Zhang et al., 2019). Moreover, in DCM rat models, elevated expression levels of the P2X7 receptor, NLRP3, caspase-1, and serum IL-1 β were observed in the myocardium (Wang et al., 2022). However, following a 12-week treadmill running regimen in these rats, improvements were observed in terms of collagen deposition, cell disorder, as well as the expression levels of NLRP3, caspase-1, P2X7 receptor, and IL-1 β within their heart (Zhang et al., 2015). Similarly, aerobic exercise also can inhibit the thioredoxin interacting protein (TXNIP)/NLRP3 inflammasome pathway and alleviate endothelial dysfunction in atherosclerotic coronary arterioles (Hong et al., 2018). These findings suggest that aerobic exercise can effectively mitigate fibrosis in the hearts subjected to an HFD and inhibit the activation of the NLRP3 inflammasome and pyroptosis in the myocardium. The effectiveness of exercise intervention on the NLRP3 inflammasome depends on the duration and intensity of exercise (Hong et al., 2018; Khakroo et al., 2019). Chronic exercise with moderate intensity significantly decreases the expression of the NLRP3 gene and levels of IL-1 β , and IL-18 cytokines in serum (Khakroo et al., 2019). Conversely, chronic exercise with high intensity leads to a significant increase in gene expression of NLRP3 and levels of IL-1 β , and IL-18 cytokines in serum (Khakroo et al., 2019). Therefore, personalized exercise regimens are necessary as there are currently no available guidelines; further research is needed.

6.6 Exercise improves hypertension and inhibits NLRP3 inflammasome

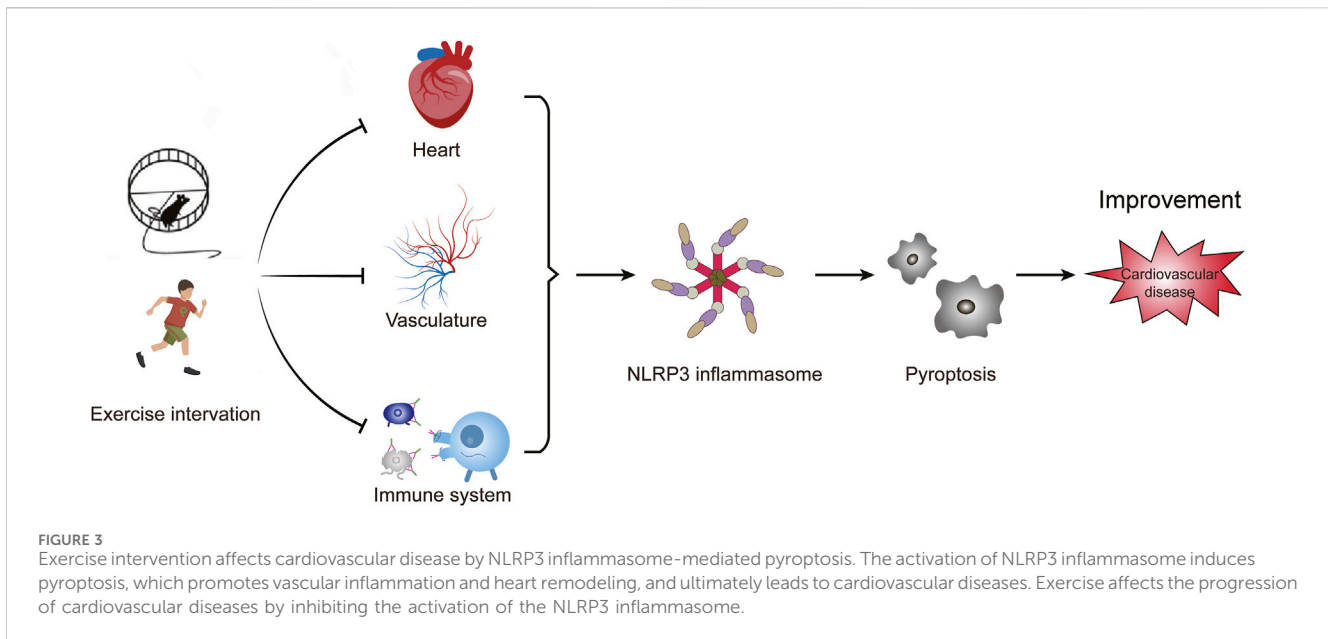
Hypertension is a potentially fatal yet preventable risk factor for CVD and accounts for the majority of cardiovascular mortality (Deussen and Kopaliani, 2023). Persistent inflammation plays a pivotal role in hypertension development, with activation of the

inflammasome and pyroptosis being potential contributors to its onset (De et al., 2021). NLRP3 inflammasome activities have been implicated in various cell types associated with pulmonary hypertension, including pulmonary arterial smooth muscle cells, pulmonary artery endothelial cells, and systemic hypertension (Haß et al., 2023). Additionally, higher serum levels of IL-1 β were observed in patients with hypertension compared to normotensive controls (Wu et al., 2022).

Exercise is commonly suggested as a lifestyle adjustment for individuals with hypertension due to various factors including inhibition of inflammation (Newman and Verdin, 2014; Chakraborty et al., 2018; Kong et al., 2021). β -Hydroxybutyrate (β -OHB) ester is primarily synthesized in the liver and transported to extrahepatic tissues, traditionally recognized as a crucial metabolic fuel during starvation periods (Newman and Verdin, 2014). Contemporary evidence indicates that ketone bodies like β -OHB can maintain physiological homeostasis by inhibiting NLRP3-inflammasome-mediated inflammation (Kong et al., 2021). Recently, a non-targeted metabolomics approach revealed nutritional intervention with β -OHB reversed the high salt-induced adverse effects including renal NLRP3-mediated inflammation, fibrosis, and hypertension (Chakraborty et al., 2018). Interestingly, exercise is associated with increased circulating levels of β -OHB (Luo et al., 2021). Therefore, exercise may raise β -OHB levels and subsequently inhibit renal NLRP3 inflammasome activation, thereby alleviating hypertension and preserving kidney function. Another *in vivo* study provided direct evidence of the impact of exercise training on the downregulation of NF- κ B and NLRP3 pathways in the mesenteric artery of spontaneously hypertensive rats (SHR) (Luo et al., 2021), which revealed that three intensity training intensities from low to high significantly inhibited the expression of NLRP3 inflammasome component and NF- κ B within the mesenteric artery and alleviate BP in SHR rat (Luo et al., 2021). In addition to the SHR rat model, Bal et al., 2022 discovered that regulated aerobic exercise also effectively reduces BP and suppresses protein expression of NLRP3, IL-1 β , and caspase-1 in the heart within the deoxycorticosterone-acetate salt hypertension model. These findings from the diversity hypertension model demonstrate that exercise training effectively attenuates NLRP3 inflammasome activity, highlighting its potential as a therapeutic intervention for hypertension.

6.7 Exercise improves MI and inhibits NLRP3 inflammasome

MI, a common condition, refers to the death of heart muscle cells caused by a blockage in the coronary arteries (Wang et al., 2020). Following a heart attack, inflammatory reactions occur, characterized by the release of inflammatory cells, cytokines, and chemical hormones (Mezzaroma et al., 2011; Wang et al., 2020). These processes contribute to cardiac dysfunction, damage to the heart muscle, and remodeling (Mezzaroma et al., 2011; Wang et al., 2020). Indeed, NLRP3 inflammasome and pyroptosis-mediated inflammation have been reported to contribute to MI progression (Sandanger et al., 2013). Sandanger et al., 2013 first showed an



increase in NLRP3 inflammasome activity in the left ventricle of the heart after MI. Furthermore, it appears that interfering with NLRP3 inflammasome signaling can prevent and mitigate damage caused by MI (Sandanger et al., 2013).

The research has shown that exercise training can delay the onset of ischemic reperfusion injury and MI (Thomas et al., 2016). In other words, exercise acts like ischemia preconditioning, which stimulates beneficial cellular responses. Azam Ahmadi et al., 2022 found HIIT can effectively decrease heart injury and NLRP3 expression in rats with MI. Interestingly, dynamin-related protein 1 (Drp1), an essential mitochondrial fission protein, can activate NLRP3 inflammasome through ROS generation after MI (Ahmadi et al., 2022; Jiang et al., 2022). However, the Azam Ahmadi team's work demonstrates exercise preconditioning (EP) in a short period did not affect Drp1 (Ahmadi et al., 2022), while inhibiting NLRP3 expression. This finding indicates that EP in a short period is required to inhibit NLRP3 inflammasome independently of Drp1; the reduction of NLRP3 can occur through other mechanisms (Jiang et al., 2022).

6.8 Exercise affects HF and regulates NLRP3 inflammasome

HF means that the heart is unable to pump sufficiently to sustain blood flow to meet the needs of the body (Sacco et al., 2014). It represents the advanced stage of various CVD, including myocarditis, MI, cardiomyopathy, hypertension, atrial fibrillation, valvular heart disease, alcohol abuse, and infection (Bracey et al., 2013). Recent studies on calcineurin transgene mice have shown elevated mRNA of NLRP3 and enhanced cleavage of caspase-1 along with cardiac hypertrophy and ventricular dilatation (Bracey et al., 2013; Napodano et al., 2023). Genetic ablation of NLRP3 or administration of IL-1 receptor antagonist has been found to attenuate cardiac inflammation and systolic dysfunction (Bracey et al., 2013; Zhang et al., 2014; Napodano et al., 2023). Additionally,

activation of NLRP3 inflammasome has been observed in LPS-stimulated cardiac fibroblasts and myofibroblasts, suggesting that its potential contribution to myocardial dysfunction through NLRP3 inflammasome or pyroptosis (Zhang et al., 2014; Boza et al., 2016). Although more research is needed to fully understand how the NLRP3 inflammasome or pyroptosis contributes to HF pathogenesis and progression; recent studies indicate that targeting this pathway could lead to new treatments for this debilitating condition (Lu et al., 2022; Xin et al., 2022).

Studies have shown that EP can regulate the NLRP3 inflammasome, reducing downstream inflammatory cytokines and protecting the heart (Li et al., 2020; Ma et al., 2021; Zhang et al., 2022). Li et al., 2020 found that moderate-intensity EP is more effective in protecting cardiac function and inhibiting the expression of TXNIP, NLRP3, NF- κ B p65, and caspase-1 in the heart. As expected with moderate-intensity exercise, 8 weeks of aerobic exercise at this level inhibited protein expression related to myocardial NLRP3/caspase-1/IL-1 β signaling pathways in mice with myocardial hypertrophy (Ma et al., 2021). In addition, Sandanger et al., 2013 show that the NLRP3 inflammasome is upregulated in myocardial fibroblasts post-MI, and maybe a significant contributor to infarct size development during ischemia-reperfusion. These findings highlight the potential therapeutic value of moderate-intensity exercise for cardiovascular health. However, exhaustive exercise (EE), characterized by sustained high-intensity exercise commonly practiced by athletes and soldiers alike, may lead to adverse reactions such as myocardial inflammation. It is reported that EE can increase NLRP3 expression and induce cardiac dysfunction which can be mitigated by EP at different intensities (Zhang et al., 2015). Importantly, moderate-intensity EP has cardioprotective effects on ventricular systolic and diastolic functions, indicating its superior efficacy compared to other intensities (Zhang et al., 2014).

Importantly, a clinical trial with 54 HF patients showed that exercise increased ASC methylation, decreased IL-1 β and ASC

TABLE 1 Types of cells and tissue injury subjected to NLRP3 inflammasome by exercise intervention in various CVD.

| Diseases | Exercise type | Specific exercise intervention | Cell types/ Organ | Injury types | Effects | References |
|--------------|---|---|--|--|---|---------------------------|
| AS | Aerobic exercise | Long-distance walking for at least 60 min at least 5 days a week | Human plasma* | | NLRP3 inflammasome inactivation | Yang et al. (2023) |
| AS | Aerobic exercise | Run on a motorized rodent treadmill 5 days a week | Mouse plasma, heart, and aortic endothelial cells | HFD | NLRP3 inflammasome inactivation | (Yang et al., 2023) |
| AS | Aerobic exercise | Treadmill exercise training for 12 weeks | Serum, the aorta, and the right lobe of the liver | HFD | FGF21 and NLRP3 inflammasome-mediated pyroptosis | Li et al. (2022) |
| HHcy | Aerobic exercise | Wheel-exercised mice had free access to their wheels 24 h/d for 5 days/wk | Plasma, liver tissue, and kidney | AIN-93G semi-purified diet | Exercise reduced circulating Hcy, which may affect NLRP3 inflammasome-mediated pyroptosis | Neuman et al. (2013) |
| Obesity | Endurance training or resistance training | at 0.15 m/s, increasing every 3 min by 0.05 m/s; for 5 times/week for 3 min and 3 series | Mouse adipose tissue and serum | HFD | NLRP3 expression, levels of TNF- α and IL-18 | Mardare et al. (2016) |
| Obesity | Aerobic exercise | Treadmill exercise for 8 weeks | Adipose tissue and bone marrow-derived macrophages | HFD | NLRP3 inflammasome | Javaid et al. (2021) |
| Obesity | Aerobic exercise | Running wheel access | Endothelial cells of the aorta | HFD | Activation of NLRP3 inflammasome | Lee et al. (2013) |
| Obesity | Aerobic exercise and resistance training | High-intensity aerobic interval training and strength training | Human serum* | | IL-18, IL-6, and TNF- α | Stensvold et al. (2012) |
| Obesity | Aerobic exercise | 12 weeks of intervention with exercise | Human serum* | 4 subjects with metabolic syndrome | IL-18 | (Troseid et al., 2009) |
| Obesity | Aerobic exercise | Hypocaloric diet exercise | Human peripheral blood* | | ASC gene expression and IL-1 β | Barrón et al. (2020) |
| Obesity | Aerobic exercise | High-intensity interval and moderate-intensity continuous training | Human serum samples* | | The expression of NLRP3 | Armanna et al. (2022) |
| Diabetes | Aerobic exercise | A treadmill for 6 weeks, 5 sessions per week | Rats adipose tissue and aortic tissue sampling | Streptozotocin | HMGB1 gene expression | Hassanpour et al. (2021) |
| DCM | Aerobic exercise | A motor treadmill running speed was increased until the speed reached 10 m/min | The heart and serum | HFD | The expression of P2X7R, NLRP3, caspase-1 and IL-1 β | Chen et al. (2021) |
| DCM | Aerobic exercise | 1 h of running on a motor-driven treadmill at 15 m/min at a 5° grade, 5 days/week for 15–16 weeks | Heart tissue and isolated coronary arteriole | A western atherogenic diet (0.2% cholesterol, 42% Kcal from fat) | NLRP3 inflammasome inactivation | Hong et al. (2018) |
| Hypertension | Aerobic exercise | High-intensity exercise | Serum and kidneys | High salt-diet | Upregulated β -OHB | Chakraborty et al. (2018) |
| Hypertension | Aerobic exercise | A treadmill for 14 weeks, 5 times a week and 60 min each time | Blood, vascular tissue samples, and mesenteric artery | Wistar Kyoto rats | NF- κ B/NLRP3 inflammatory pathway | Luo et al. (2021) |
| Hypertension | Aerobic exercise | A 45-min running on a horizontal treadmill at 20 m/min, 5 days per week | Systolic blood pressure and the left ventricle tissues | Deoxycorticosterone-acetate and salt administration | NLRP3 inflammasome activation | Bal et al. (2022) |

(Continued on following page)

TABLE 1 (Continued) Types of cells and tissue injury subjected to NLRP3 inflammasome by exercise intervention in various CVD.

| Diseases | Exercise type | Specific exercise intervention | Cell types/ Organ | Injury types | Effects | References |
|----------|---|---|--|---|--|-------------------------|
| MI | Aerobic exercise and anaerobic exercise | 10*1-min running intervals were separated by a 2-min rest | Blood samples from the heart and heart tissue | Isoproterenol | NLRP3 inflammasome | Ahmadi et al. (2022) |
| HF | Aerobic exercise | Low-, middle-, and high-intensity exercise | Serum and myocardial specimens | refer to Bedford's motion load standard | NF-κB p65/ NLRP3 inflammatory | Li et al. (2020) |
| HF | Aerobic exercise | 2, 4, and 8 weeks of moderate-intensity aerobic exercise | Mouse heart tissues | Transverse aortic constriction | NLRP3/caspase-1/IL-1β signaling pathways | Ma et al. (2021) |
| HF | Aerobic exercise | A treadmill for 90 min/day and 6 days/week for 6 weeks at a velocity of 15 cm/s | Mouse hearts and primary neonatal mouse cardiomyocytes | Isoprenaline | NLRP3 inflammasome | Zhang et al. (2022) |
| HF | Aerobic exercise | A progressive, moderate intensity aerobic protocol | Human plasma* | | IL-1β and ASC mRNA gene expression | Butts et al. (2018) |
| HF | Aerobic exercise | Three times a week, 60 min/session, for 8 months | Human serum* | | Levels of TNF-α, IL-1β, IL-8, IF-γ and IL-10 | Rodrigues et al. (2020) |

*Indicated this study with cells or tissue from humans.
ASC, apoptosis-associated speck-like protein; AS, atherosclerosis; DCM, diabetic cardiomyopathy; FGF21, fibroblast growth factor 21; HFD, high-fat diet; HMGB-1, high mobility group box-1; Hcy, homocysteine; HHcy, hyperhomocysteinemia; HF, heart failure; IL, interleukin; β-OHB, β-Hydroxybutyrate; MI, myocardial infarction; NLRP3, NOD-like receptor protein 3; NF-κB, nuclear factor-κB; P2X7, purinergic receptor P2X, ligand-gated ion channel 7; AIN-93G, american institute of nutrition-93 growth).

mRNA levels compared to the control group in plasma, suggesting that exercise may improve HF via epigenetic regulation of ASC (Butts et al., 2018). Recently, our research team has reported for the first time that NLRP3 inflammasome-mediated pyroptosis contributes to the pathogenesis of non-ischemic dilated cardiomyopathy in cellular, murine, and human models (Zeng et al., 2020). Furthermore, numerous studies have investigated exercise intervention in patients with dilated cardiomyopathy and have demonstrated its safety and efficacy in improving exercise capacity and quality of life among these individuals (Stolen et al., 2003; Beer et al., 2008). Consequently, it is meaningful to investigate whether exercise can exert inhibitory effects on NLRP3 inflammasome activation and subsequent pyroptotic cell death in dilated cardiomyopathy. Preliminary data suggest that exercise may effectively inhibit NLRP3 inflammasome activation in dilated cardiomyopathy (unpublished data).

Although these experiments indicate that exercise improves HF by inhibiting NLRP3 inflammasome activation, there is still debate (Butts et al., 2018). For example, previous work reported that exercise training consisting of three sessions per week lasting for 60 min each, including aerobic exercises, strength exercises targeting major muscle groups, and stretching exercises have no effect on serum levels of pro-inflammatory cytokines (TNF-α, IL-1β, IL-8, and monocyte chemoattractant protein-1) of patients with severe Chagas cardiomyopathy (Rodrigues Junior et al., 2020), suggesting that exercise training may benefit patients with severe Chagas cardiomyopathy independent of its impact on inflammasome (Rodrigues Junior et al., 2020).

Most studies suggest that exercise improves HF by inhibiting the activation of NLRP3 inflammasomes. Nevertheless, a minority of research suggests that engaging in activities like EE might potentially worsen the progression of HF. It is imperative for future

investigations to thoroughly examine the role played by NLRP3 inflammasome activation in exercise-induced exacerbation of HF.

7 Summary and prospects

Exercise plays an important regulatory role in the development of various CVD by influencing the activation of the NLRP3 inflammasome (Figure 3). This impact has been observed in different types of cells and tissues, both *in vivo* and *in vitro* models of CVD (Table 1). This review highlights the close relationship between aerobic exercise, NLRP3 inflammasome, and CVD. It suggests that aerobic exercise can alleviate pyroptosis and improve cardiovascular-related diseases by modulating the NLRP3 inflammatory signaling pathway. Different patterns of exercise have varying impacts on the NLRP3 inflammasome; therefore, further research is needed to determine their optimal effects on specific types of cells and their underlying molecular mechanisms involving the NLRP3 inflammasome. Currently, there is a greater emphasis on animal experiments investigating the influence of exercise on NLRP3 inflammasome or cell pyroptosis while human studies are limited with small sample sizes in this area. Conducting comprehensive research on humans would provide a scientifically sound basis for understanding how exercise regulates NLRP3 inflammasome activity and promotes overall health. Hence, there is a need for methodologically rigorous large-scale human studies to determine ideal patterns of exercise.

The investigation of drugs targeting the NLRP3 inflammasome in clinical trials for CVD has been conducted. For example, the canakinumab anti-inflammatory thrombosis outcome study trial demonstrated the efficacy of IL-1-targeting therapy in preventing

atherothrombotic events, indicating the potential of targeting the NLRP3 inflammasome (Ma et al., 2021; Ridker and Rane, 2021). However, the high cost of canakinumab limits its widespread use in the future. Additionally, long-term suppression of IL-1 signaling may have adverse effects such as increased susceptibility to infections and disturbances in immune homeostasis (Hettwer et al., 2022). Common cardiovascular drugs such as statins, beta-blockers, sodium-glucose cotransporter 2 inhibitors, and glucagon-like peptide 1 agonists may modulate NLRP3 inflammasome activity through different mechanisms, exerting protective effects in CVD (Terentes-Printzios et al., 2022). However, these classical medications carry certain side effects on gastrointestinal function and are prone to causing symptoms like hypotension and arrhythmias (Vaiculeviciute et al., 2021). Interestingly, research suggests that exercise effectively modulates NLRP3 inflammasome activation, leading to improvements in AS and other CVD (Vaiculeviciute et al., 2021; Terentes-Printzios et al., 2022). This therapeutic approach is relatively simple, feasible, and considered safe compared to using NLRP3 inhibitors and classical medications for CVD. Therefore, combining medication treatment with exercise intervention offers a potential strategy to improve the quality of life for patients.

Author contributions

PD: Data curation, Writing—original draft, Investigation. YS: Data curation, Writing—original draft, Investigation. YY: Investigation, Resources, Supervision, Writing—review and editing. CZ: Conceptualization, Funding acquisition, Investigation, Supervision, Visualization, Writing—original draft, Writing—review and editing.

References

- Ahmadi, A., Kashef, M., Rajabi, H., and Salehpour, M. (2022). Effects of exercise preconditioning on NLRP3 and mitochondrial fission in isoproterenol-induced myocardial infarcted rats. *Comp. Clin. Pathol.* 32, 289–297. doi:10.1007/s00580-022-03397-3
- Alten, R., Mischkewitz, M., and Nitschmann, S. (2020). Febuxostat or allopurinol in patients with gout: cardiovascular safety of febuxostat and allopurinol in patients with gout and cardiovascular morbidities (CARES). *Internist (Berl.)* 61 (5), 530–532. doi:10.1007/s00108-020-00766-4
- Armanna, F., Ghazalian, F., Shadnough, M., Keyvani, H., and Gholami, M. (2022). Effects of high-intensity interval vs. moderate-intensity continuous training on body composition and gene expression of ACE2, NLRP3, and FNDC5 in obese adults: a randomized controlled trial *Med. J. Islam Repub. Iran.* 36, 161. doi:10.47176/mjiri.36.161
- Baik, S. H., Ramanujan, V. K., Becker, C., Fett, S., Underhill, D. M., and Wolf, A. J. (2023). Hexokinase dissociation from mitochondria promotes oligomerization of VDAC that facilitates NLRP3 inflammasome assembly and activation. *Sci. Immunol.* 8 (84), eade7652. doi:10.1126/sciimmunol.ade7652
- Bal, N. B., Bostanci, A., Sadi, G., Dönmez, M. O., Uludag, M. O., and Demirel, Y. E. (2022). Resveratrol and regular exercise may attenuate hypertension-induced cardiac dysfunction through modulation of cellular stress responses. *Life Sci.* 296, 120424. doi:10.1016/j.lfs.2022.120424
- Barrón, C. E., González, B. K., Rosales, C. G., Mora, J. A., Hernández, C. I., and Martínez, L. E. (2020). Low-grade chronic inflammation is attenuated by exercise training in obese adults through down-regulation of ASC gene in peripheral blood: a pilot study. *Genes. Nutr.* 15, 15. doi:10.1186/s12263-020-00674-0
- Beer, M., Wagner, D., Myers, J., Sandstedt, J., Köstler, H., Hahn, D., et al. (2008). Effects of exercise training on myocardial energy metabolism and ventricular function assessed by quantitative phosphorus-31 magnetic resonance spectroscopy and magnetic resonance imaging in dilated cardiomyopathy. *J. Am. Coll. Cardiol.* 51 (19), 1883–1891. doi:10.1016/j.jacc.2007.09.075
- Bigueti, C. C., Cavalla, F., Silveira, E. V., Tabanez, A. P., Francisconi, C. F., Taga, R., et al. (2019). HGMB1 and RAGE as essential components of tissue integration process in mice. *Front. Immunol.* 10, 709. doi:10.3389/fimmu.2019.00709
- Blanks, A. M., Rodriguez-Miguel, P., Looney, J., Tucker, M. A., Jeong, J., Thomas, J., et al. (2019). Whole body vibration elicits differential immune and metabolic responses in obese and normal weight individuals. *Brain Behav. Immun. Health* 1, 100011. doi:10.1016/j.bbih.2019.100011
- Bockstiegel, J., Engelhardt, J., and Weindl, G. (2023). P2X7 receptor activation leads to NLRP3-independent IL-1 β release by human macrophages. *Cell. Commun. Signal* 21 (1), 335. doi:10.1186/s12964-023-01356-1
- Boza, P., Ayala, P., Vivar, R., Humeres, C., Cáceres, F. T., Muñoz, C., et al. (2016). Expression and function of toll-like receptor 4 and inflammasomes in cardiac fibroblasts and myofibroblasts: IL-1 β synthesis, secretion, and degradation. *Mol. Immunol.* 74, 96–105. doi:10.1016/j.molimm.2016.05.001
- Brace, N. A., Beck, P. L., Muruve, D. A., Hirota, S. A., Guo, J., Jabagi, H., et al. (2013). The Nlrp3 inflammasome promotes myocardial dysfunction in structural cardiomyopathy through interleukin-1 β . *Exp. Physiol.* 98, 462–472. doi:10.1113/expphysiol.2012.068338
- Butts, B., Butler, J., Dunbar, S. B., Corwin, E., and Gary, R. A. (2018). Effects of exercise on ASC methylation and IL-1 cytokines in heart failure. *Med. Sci. Sports Exerc* 50, 1757–1766. doi:10.1249/MSS.0000000000001641
- Carvalho, L., Junior, R. M., Barreira, J., Schoenfeld, B. J., Orazem, J., and Barroso, R. (2022). Muscle hypertrophy and strength gains after resistance training with different volume-matched loads: a systematic review and meta-analysis. *Appl. Physiol. Nutr. Metab.* 47 (4), 357–368. doi:10.1139/apnm-2021-0515
- Casado, A., Foster, C., Bakken, M., and Tjelta, L. I. (2023). Does lactate-guided threshold interval training within a high-volume low-intensity approach represent the "next step" in the evolution of distance running training? *Int. J. Environ. Res. Public Health* 20 (5), 3782. doi:10.3390/ijerph20053782
- Chakraborty, S., Galla, S., Cheng, X., Yeo, J. Y., Mell, B., Singh, V., et al. (2018). Salt-responsive metabolite, β -hydroxybutyrate, attenuates hypertension. *Cell. Rep.* 25, 677–689. doi:10.1016/j.celrep.2018.09.058
- Chan, A. H., Burgener, S. S., Vezirgiannis, K., Wang, X., Acklam, J., Von Pein, J. B., et al. (2023). Caspase-4 dimerisation and D289 auto-processing elicit an interleukin-1 β -converting enzyme. *Life Sci. Alliance* 6 (10), e202301908. doi:10.26508/lsa.202301908

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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- Chan, A. H., and Schroder, K. (2020). Inflammasome signaling and regulation of interleukin-1 family cytokines. *J. Exp. Med.* 217, e20190314. doi:10.1084/jem.20190314
- Chang, M. X. (2023). Emerging mechanisms and functions of inflammasome complexes in teleost fish. *Front. Immunol.* 14, 1065181. doi:10.3389/fimmu.2023.1065181
- Chen, B., Yan, Y., Yang, Y., Cao, G., Wang, X., Wang, Y., et al. (2022). A pyroptosis nanotuner for cancer therapy. *Nat. Nanotechnol.* 17 (7), 788–798. doi:10.1038/s41565-022-01125-0
- Chen, H., Tran, D., Yang, H. C., Nylander, S., Birnbaum, Y., and Ye, Y. (2020). Dapagliflozin and ticagrelor have additive effects on the attenuation of the activation of the NLRP3 inflammasome and the progression of diabetic cardiomyopathy: an AMPK-mTOR interplay. *Cardiovasc Drugs Ther.* 34 (4), 443–461. doi:10.1007/s10557-020-06978-y
- Chen, L., Dong, J., Liao, S., Wang, S., Wu, Z., Zuo, M., et al. (2022). Loss of Sam50 in hepatocytes induces cardiolipin-dependent mitochondrial membrane remodeling to trigger mtDNA release and liver injury. *Hepatology* 76 (5), 1389–1408. doi:10.1002/hep.32471
- Chen, W. K., Tsai, Y. L., Shibui, M. A., Shen, C. Y., Chang-Lee, S. N., Chen, R. J., et al. (2018). Exercise training augments Sirt1-signaling and attenuates cardiac inflammation in D-galactose induced-aging rats. *Aging* 10, 4166–4174. doi:10.18632/aging.101714
- Chen, X., Li, H., Wang, K., Liang, X., Wang, W., Hu, X., et al. (2019). Aerobic exercise ameliorates myocardial inflammation, fibrosis and apoptosis in high-fat-diet rats by inhibiting P2X7 purinergic receptors. *Front. Physiol.* 10, 1286. doi:10.3389/fphys.2019.01286
- Chen, X., Zhang, D., Li, Y., Wang, W., Bei, W., and Guo, J. (2021). NLRP3 inflammasome and IL-1 β pathway in type 2 diabetes and atherosclerosis: friend or foe? *Pharmacol. Res.* 173, 105885. doi:10.1016/j.phrs.2021.105885
- Chen, Y., Hua, Y., Li, X., Arslan, I. M., Zhang, W., and Meng, G. (2020). Distinct types of cell death and the implication in diabetic cardiomyopathy. *Front. Pharmacol.* 11, 42. doi:10.3389/fphar.2020.00042
- Consitt, L. A., Dudley, C., and Saxena, G. (2019). Impact of endurance and resistance training on skeletal muscle glucose metabolism in older adults. *Nutrients* 11 (11), 2636. doi:10.3390/nu11112636
- Cutrufello, P. T., Benson, B. A., and Landram, M. J. (2020). The effect of music on anaerobic exercise performance and muscular endurance. *J. Sports Med. Phys. Fit.* 60 (3), 486–492. doi:10.23736/S0022-4707.19.10228-9
- De, M. C., Pelegrin, P., Baroja, M. A., and Cuevas, S. (2021). Emerging role of the inflammasome and pyroptosis in hypertension. *Int. J. Mol. Sci.* 22, 1064. doi:10.3390/ijms22031064
- Deussen, A., and Kopalani, I. (2023). Targeting inflammation in hypertension. *Curr. Opin. Nephrol. Hypertens.* 32, 111–117. doi:10.1097/MNH.0000000000000862
- Diaz-Del-Olmo, I., Worboys, J., Martin-Sanchez, F., Gritsenko, A., Ambrose, A. R., Tannahill, G. M., et al. (2021). Internalization of the membrane attack complex triggers NLRP3 inflammasome activation and IL-1 β secretion in human macrophages. *Front. Immunol.* 12, 720655. doi:10.3389/fimmu.2021.720655
- Dilucca, M., Ramos, S., Shkarina, K., Santos, J. C., and Broz, P. (2021). Guanylate-binding protein-dependent noncanonical inflammasome activation prevents burkholderia thailandensis-induced multinucleated giant cell formation. *mBio* 12 (4), e0205421. doi:10.1128/mBio.02054-21
- Einarson, T. R., Acs, A., Ludwig, C., and Panton, U. H. (2018). Prevalence of cardiovascular disease in type 2 diabetes: a systematic literature review of scientific evidence from across the world in 2007–2017. *Cardiovasc Diabetol.* 17, 83. doi:10.1186/s12933-018-0728-6
- Fu, J., and Wu, H. (2023). Structural mechanisms of NLRP3 inflammasome assembly and activation. *Annu. Rev. Immunol.* 41, 301–316. doi:10.1146/annurev-immunol-081022-021207
- Haidar, A., and Horwich, T. (2023). Obesity, cardiorespiratory fitness, and cardiovascular disease. *Curr. Cardiol. Rep.* 25 (11), 1565–1571. doi:10.1007/s11886-023-01975-7
- Han, X., Ni, J., Wu, Z., Wu, J., Li, B., Ye, X., et al. (2020). Myeloid-specific dopamine D2 receptor signalling controls inflammation in acute pancreatitis via inhibiting M1 macrophage. *Br. J. Pharmacol.* 177 (13), 2991–3008. doi:10.1111/bph.15026
- Hassanpour Soleimani, S., Abbassi Daloui, A., Abdi, A., and Zilaei Bori, S. (2021). Effect of a 6-week aerobic exercise program on high-mobility group box 1 gene expression in aortic tissue of diabetic rats. *Q. Horizon Med. Sci.* 27, 82–97. doi:10.32598/hms.27.1.2378.1
- Haß, U., Heider, S., Kochlik, B., Herpich, C., Pivovarova-Ramich, O., and Norman, K. (2023). Effects of exercise and omega-3-supplemented, high-protein diet on inflammatory markers in serum, on gene expression levels in PBMC, and after *ex vivo* whole-Blood LPS stimulation in old adults. *Int. J. Mol. Sci.* 24 (2), 928. doi:10.3390/ijms24020928
- Haß, U., Kochlik, B., Herpich, C., Rudloff, S., and Norman, K. (2022). Effects of an omega-3 supplemented, high-Protein diet in combination with vibration and resistance exercise on muscle power and inflammation in old adults: a pilot randomized controlled trial. *Nutrients* 14 (20), 4274. doi:10.3390/nu14204274
- Hettwer, J., Hinterdobler, J., Miritsch, B., Deutsch, M. A., Li, X., Mauersberger, C., et al. (2022). Interleukin-1 β suppression dampens inflammatory leucocyte production and uptake in atherosclerosis. *Cardiovasc Res.* 118 (13), 2778–2791. doi:10.1093/cvr/cvab337
- Hong, J., Kim, K., Park, E., Lee, J., Markowski, M. M., Marrelli, S. P., et al. (2018). Exercise ameliorates endoplasmic reticulum stress-mediated vascular dysfunction in mesenteric arteries in atherosclerosis. *Sci. Rep.* 8 (1), 7938. doi:10.1038/s41598-018-26188-9
- Hong, J., Park, E., Lee, J., Lee, Y., Rooney, B. V., and Park, Y. (2021). Exercise training mitigates ER stress and UCP2 deficiency-associated coronary vascular dysfunction in atherosclerosis. *Sci. Rep.* 11, 15449. doi:10.1038/s41598-021-94944-5
- Huang, L. S., Anas, M., Xu, J., Zhou, B., Toth, P. T., Krishnan, Y., et al. (2023). Endosomal trafficking of two-pore K⁺ efflux channel TWIK2 to plasmalemma mediates NLRP3 inflammasome activation and inflammatory injury. *Elife* 12, e83842. doi:10.7554/eLife.83842
- Javadi, H. M. A., Sahar, N. E., Zhuge, D. L., and Huh, J. Y. (2021). Exercise inhibits NLRP3 inflammasome activation in obese mice via the anti-inflammatory effect of meteorin-like. *Cells* 10, 3480. doi:10.3390/cells10123480
- Jiang, H., Chen, F., Song, D., Zhou, X., Ren, L., and Zeng, M. (2022). Dynamin-related protein 1 is involved in mitochondrial damage, defective mitophagy, and NLRP3 inflammasome activation induced by MSU crystals. *Oxid. Med. Cell. Longev.* 2022, 5064494. doi:10.1155/2022/5064494
- Kar, S., Shahshahan, H. R., Hackfort, B. T., Yadav, S. K., Yadav, R., Kambis, T. N., et al. (2019). Exercise training promotes cardiac hydrogen sulfide biosynthesis and mitigates pyroptosis to prevent high-fat diet-induced diabetic cardiomyopathy. *Antioxidants (Basel)* 8, 638. doi:10.3390/antiox8120638
- Khakroo, A. I., Rahmani, N. F., and Lombardi, G. (2019). The effects of acute and chronic aerobic activity on the signaling pathway of the inflammasome NLRP3 complex in young men. *Med. Kaunas* 55, 105. doi:10.3390/medicina55040105
- Kodi, T., Sankhe, R., Gopinathan, A., Nandakumar, K., and Kishore, A. (2024). New insights on NLRP3 inflammasome: mechanisms of activation, inhibition, and epigenetic regulation. *J. Neuroimmune Pharmacol.* 19 (1), 7. doi:10.1007/s11481-024-10101-5
- Kong, G., Liu, J., Li, R., Lin, J., Huang, Z., Yang, Z., et al. (2021). Ketone metabolite β -hydroxybutyrate ameliorates inflammation after spinal cord injury by inhibiting the NLRP3 inflammasome. *Neurochem. Res.* 46, 213–229. doi:10.1007/s11064-020-03156-2
- Konishi, M., Kojima, S., Uchiyama, K., Yokota, N., Tokutake, E., Wakasa, Y., et al. (2022). Effect of febuxostat on clinical outcomes in patients with hyperuricemia and cardiovascular disease. *Int. J. Cardiol.* 49, 127–133. doi:10.1016/j.ijcard.2021.11.076
- Krantz, M., Eklund, D., Särndahl, E., and Hedbrant, A. (2023). A detailed molecular network map and model of the NLRP3 inflammasome. *Front. Immunol.* 14, 1233680. doi:10.3389/fimmu.2023.1233680
- Lee, A. H., Shin, H. Y., Park, J. H., Koo, S. Y., Kim, S. M., and Yang, S. H. (2021). Fucoxanthin from microalgae *Phaeodactylum tricornutum* inhibits pro-inflammatory cytokines by regulating both NF- κ B and NLRP3 inflammasome activation. *Sci. Rep.* 11 (1), 543. doi:10.1038/s41598-020-80748-6
- Lee, H. M., Kim, J. J., Kim, H. J., Shong, M., Ku, B. J., and Jo, E. K. (2013). Upregulated NLRP3 inflammasome activation in patients with type 2 diabetes. *Diabetes* 62, 194–204. doi:10.2337/db12-0420
- Lee, J., Hong, J., Umetani, M., Lavoy, E. C., Kim, J. H., and Park, Y. (2020). Vascular protection by exercise in obesity: inflammasome-associated mechanisms. *Med. Sci. Sports Exerc* 52, 2538–2545. doi:10.1249/MSS.00000000000002419
- Lee, J., Lee, Y., LaVoy, E. C., Umetani, M., Hong, J., and Park, Y. (2018). Physical activity protects NLRP3 inflammasome-associated coronary vascular dysfunction in obese mice. *Physiol. Rep.* 6, e13738. doi:10.14814/phy2.13738
- Lee, S., Ye, Q., Yang, H., Lee, S., Kim, Y., Lee, N., et al. (2024). Aioeua padiformis extract exhibits anti-inflammatory effects by inhibiting the ATPase activity of NLRP3. *Sci. Rep.* 14 (1), 5237. doi:10.1038/s41598-024-55651-z
- Levine, B. D. (2014). Can intensive exercise harm the heart? The benefits of competitive endurance training for cardiovascular structure and function. *Circulation* 130 (12), 987–991. doi:10.1161/CIRCULATIONAHA.114.008142
- Lewisluján, L. M., McCarty, M. F., Dinicolantonio, J. J., Gálvezruiz, J. C., Rosas-Burgos, E. C., Plascencia-Jatomea, M., et al. (2022). Nutraceuticals/drugs promoting mitophagy and mitochondrial biogenesis may combat the mitochondrial dysfunction driving progression of dry age-related macular degeneration. *Nutrients* 14 (9), 1985. doi:10.3390/nu14091985
- Li, N., Zhang, L., Wang, X., Zhou, Y., and Gong, L. (2023). Exploring exercise-driven inhibition of pyroptosis: novel insights into treating diabetes mellitus and its complications. *Front. Endocrinol. (Lausanne)* 14, 1230646. doi:10.3389/fendo.2023.1230646
- Li, X. H., Liu, L. Z., Chen, L., Pan, Q. N., Ouyang, Z. Y., Fan, D. J., et al. (2022). Aerobic exercise regulates FGF21 and NLRP3 inflammasome-mediated pyroptosis and inhibits atherosclerosis in mice. *PLoS One* 17, e0273527. doi:10.1371/journal.pone.0273527
- Li, Y., Xu, P., Wang, Y., Zhang, J., Yang, M., Chang, Y., et al. (2020). Different intensity exercise preconditions affect cardiac function of exhausted rats through regulating TXNIP/TRX/NF- κ Bp65/NLRP3 inflammatory pathways. *Evid. Based Complement. Altern. Med.* 2020, 5809298. doi:10.1155/2020/5809298

- Liao, C. C., Xu, J. W., Huang, W. C., Chang, H. C., and Tung, Y. T. (2022). Plasma proteomic changes of atherosclerosis after exercise in apoE knockout mice. *Biol. (Basel)* 11 (2), 253. doi:10.3390/biology11020253
- Liu, D., Wang, R., Grant, A. R., Zhang, J., Gordon, P. M., Wei, Y., et al. (2017). Immune adaptation to chronic intense exercise training: new microarray evidence. *BMC Genom* 18, 29. doi:10.1186/s12864-016-3388-5
- Liu, J., Jia, S., Yang, Y., Piao, L., Wang, Z., Jin, Z., et al. (2023). Exercise induced meteorin-like protects chondrocytes against inflammation and pyroptosis in osteoarthritis by inhibiting PI3K/Akt/NF- κ B and NLRP3/caspase-1/GSDMD signaling. *Biomed. Pharmacother.* 158, 114118. doi:10.1016/j.biopha.2022.114118
- Liu, S., Tao, J., Duan, F., Li, H., and Tan, H. (2022). HHcy induces pyroptosis and atherosclerosis via the lipid raft-mediated NOX-ROS-NLRP3 inflammasome pathway in apoE^{-/-} mice. *Cells* 11, 2438. doi:10.3390/cells11152438
- Lu, P., Zheng, H., Meng, H., Liu, C., Duan, L., Zhang, J., et al. (2023). Mitochondrial DNA induces nucleus pulposus cell pyroptosis via the TLR9-NF- κ B-NLRP3 axis. *J. Transl. Med.* 21 (1), 389. doi:10.1186/s12967-023-04266-5
- Lu, Y., Xiang, M., Xin, L., Zhang, Y., Wang, Y., Shen, Z., et al. (2022). Qiliqiangxin modulates the gut microbiota and NLRP3 inflammasome to protect against ventricular remodeling in heart failure. *Front. Pharmacol.* 13, 905424. doi:10.3389/fphar.2022.905424
- Luo, B., Huang, F., Liu, Y., Liang, Y., Wei, Z., Ke, H., et al. (2017). NLRP3 inflammasome as a molecular marker in diabetic cardiomyopathy. *Front. Physiol.* 8, 519. doi:10.3389/fphys.2017.00519
- Luo, B., Li, B., Wang, W., Liu, X., Xia, Y., Zhang, C., et al. (2014). NLRP3 gene silencing ameliorates diabetic cardiomyopathy in a type 2 diabetes rat model. *PLoS One* 9, e104771. doi:10.1371/journal.pone.0104771
- Luo, M., Cao, C., Niebauer, J., Yan, J., Ma, X., Chang, Q., et al. (2021). Effects of different intensities of continuous training on vascular inflammation and oxidative stress in spontaneously hypertensive rats. *J. Cell. Mol. Med.* 25, 8522–8536. doi:10.1111/jcmm.16813
- Ma, J., and Chen, X. (2021). Anti-inflammatory therapy for coronary atherosclerotic heart disease: unanswered questions behind existing successes. *Front. Cardiovasc Med.* 7, 631398. doi:10.3389/fcvm.2020.631398
- Ma, M., Chen, W., Hua, Y., Jia, H., Song, Y., and Wang, Y. (2021). Aerobic exercise ameliorates cardiac hypertrophy by regulating mitochondrial quality control and endoplasmic reticulum stress through M2 AChR. *J. Cell. Physiol.* 236, 6581–6596. doi:10.1002/jcp.30342
- Mardare, C., Kruger, K., Liebisch, G., Seimetz, M., Couturier, A., Ringseis, R., et al. (2016). Endurance and resistance training affect high fat diet-induced increase of ceramides, inflammasome expression, and systemic inflammation in mice. *J. Diabetes. Res.* 2016, 4536470. doi:10.1155/2016/4536470
- Mason, S. A., Trewin, A. J., Parker, L., and Wadley, G. D. (2020). Antioxidant supplements and endurance exercise: current evidence and mechanistic insights. *Redox. Biol.* 35, 101471. doi:10.1016/j.redox.2020.101471
- May, B. K., and Treadwell, R. E. (2020). Increasing exercise intensity: teaching high-intensity interval training to individuals with developmental disabilities using a lottery reinforcement system. *Behav. Anal. Pract.* 13 (4), 826–837. doi:10.1007/s40617-020-00428-9
- McCullough, P. A., and Lavie, C. J. (2014). Coronary artery plaque and cardiotoxicity as a result of extreme endurance exercise. *Mo Med.* 111 (2), 95–98.
- McKee, C. M., and Coll, R. C. (2020). NLRP3 inflammasome priming: a riddle wrapped in a mystery inside an enigma. *J. Leukoc. Biol.* 108, 937–952. doi:10.1002/JLB.3MR0720-513R
- Memme, J. M., Erlich, A. T., Phukan, G., and Hood, D. A. (2021). Exercise and mitochondrial health. *J. Physiol.* 2021, 803–817. doi:10.1113/JP278853
- Mensah, G. A., Roth, G. A., and Fuster, V. (2019). The global burden of cardiovascular diseases and risk factors: 2020 and beyond. *J. Am. Coll. Cardiol.* 74 (20), 2529–2532. doi:10.1016/j.jacc.2019.10.009
- Mezzaroma, E., Toldo, S., Farkas, D., Seropian, I. M., Van Tassell, B. W., Salloum, F. N., et al. (2011). The inflammasome promotes adverse cardiac remodeling following acute myocardial infarction in the mouse. *Proc. Natl. Acad. Sci. U. S. A.* 108, 19725–19730. doi:10.1073/pnas.1108586108
- Miko, H. C., Zillmann, N., Ring-Dimitriou, S., Dorner, T. E., Titze, S., and Bauer, R. (2020). Effects of physical activity on health. *Gesundheitswesen* 82 (S 03), S184–S195. doi:10.1055/a-1217-0549
- Movahedpour, A., Taghvaeifar, R., Asadi-Pooya, A. A., Karami, Y., Tavasolian, R., Khatami, S. H., et al. (2023). Nano-delivery systems as a promising therapeutic potential for epilepsy: current status and future perspectives. *CNS Neurosci. Ther.* 29 (11), 3150–3159. doi:10.1111/cns.14355
- Mueller, S., Winzer, E. B., Duvinage, A., Gevaert, A. B., Edelmann, F., Haller, B., et al. (2021). Effect of high-intensity interval training, moderate continuous training, or guideline-based physical activity advice on peak oxygen consumption in patients with heart failure with preserved ejection fraction: a randomized clinical trial. *Jama* 325, 542–551. doi:10.1001/jama.2020.26812
- Murray, K. O., Mahoney, S. A., Venkatasubramanian, R., Seals, D. R., and Clayton, Z. S. (2023). Aging, aerobic exercise, and cardiovascular health: barriers, alternative strategies and future directions. *Exp. Gerontol.* 173, 112105. doi:10.1016/j.exger.2023.112105
- Napodano, C., Carnazzo, V., Basile, V., Pocino, K., Stefanile, A., Gallucci, S., et al. (2023). NLRP3 inflammasome involvement in heart, liver, and lung diseases—a lesson from cytokine storm syndrome. *Int. J. Mol. Sci.* 24 (23), 16556. doi:10.3390/jms242316556
- Nath, L. C., Elliott, A., La Gerche, A., Weir, J., Forbes, G., Thomas, G., et al. (2023). Associations between posttrace atrial fibrillation and measures of performance, racing history and airway disease in horses. *J. Vet. Intern. Med.* 37 (6), 2573–2583. doi:10.1111/jvim.16878
- Neuman, J. C., Albright, K. A., and Schalinske, K. L. (2013). Exercise prevents hyperhomocysteinemia in a dietary folate-restricted mouse model. *Nutr. Res.* 33, 487–493. doi:10.1016/j.nutres.2013.04.008
- Neumann, F. A., Jagemann, B., Makarova, N., Börschel, C. S., Aarabi, G., Gutmann, F., et al. (2022). Mediterranean diet and atrial fibrillation: lessons learned from the AFHRI case-control study. *Nutrients* 14 (17), 3615. doi:10.3390/nu14173615
- Newman, J. C., and Verdin, E. (2014). β -hydroxybutyrate: much more than a metabolite. *Diabetes Res. Clin. Pract.* 106, 173–181. doi:10.1016/j.diabres.2014.08.009
- Nie, Z., Chen, M., Gao, Y., Huang, D., Cao, H., Peng, Y., et al. (2021). Regulated cell death in urinary malignancies. *Front. Cell. Dev. Biol.* 9, 789004. doi:10.3389/fcell.2021.789004
- O'Keefe, E. L., Sturgess, J. E., O'Keefe, J. H., Gupta, S., and Lavie, C. J. (2021). Prevention and treatment of atrial fibrillation via risk factor modification. *Am. J. Cardiol.* 160, 46–52. doi:10.1016/j.amjcard.2021.08.042
- O'Keefe, J. H., Patil, H. R., Lavie, C. J., Magalski, A., Vogel, R. A., and McCullough, P. A. (2012). Potential adverse cardiovascular effects from excessive endurance exercise. *Mayo Clin. Proc.* 87 (6), 587–595. doi:10.1016/j.mayocp.2012.04.005
- Paerewijck, O., and Lamkanfi, M. (2022). The human inflammasomes. *Mol. Asp. Med.* 88, 101100. doi:10.1016/j.mam.2022.101100
- Paluch, A. E., Boyer, W. R., Franklin, B. A., Laddu, D., Lobelo, F., Lee, D. C., et al. (2024). Resistance exercise training in individuals with and without cardiovascular disease: 2023 update: a scientific statement from the American heart association. *Circulation* 149 (3), e217–e231. doi:10.1161/CIR.0000000000001189
- Pope, B. S., and Wood, S. K. (2020). Advances in understanding mechanisms and therapeutic targets to treat comorbid depression and cardiovascular disease. *Neurosci. Biobehav. Rev.* 116, 337–349. doi:10.1016/j.neubiorev.2020.06.031
- Powers, S. K., Deminice, R., Ozdemir, M., Yoshihara, T., Bomkamp, M. P., and Hyatt, H. (2020). Exercise-induced oxidative stress: friend or foe? *J. Sport Health Sci.* 9, 415–425. doi:10.1016/j.jshs.2020.04.001
- Ren, G., Zhang, X., Xiao, Y., Zhang, W., Wang, Y., Ma, W., et al. (2019). ABRO1 promotes NLRP3 inflammasome activation through regulation of NLRP3 deubiquitination. *EMBO J.* 38 (6), e100376. doi:10.15252/embj.2018100376
- Ridker, P. M., and Rane, M. (2021). Interleukin-6 signaling and anti-interleukin-6 therapeutics in cardiovascular disease. *Circ. Res.* 128 (11), 1728–1746. doi:10.1161/CIRCRESAHA.121.319077
- Ringseis, R., Eder, K., Mooren, F. C., and Krüger, K. (2015). Metabolic signals and innate immune activation in obesity and exercise. *Exerc. Immunol. Rev.* 21, 58–68. doi:10.1111/jpn.12263
- Rodrigues Junior, L. F., Mendes, F. S. N. S., Pinto, V. L. M., da Silva, P. S., Sperandio da Silva, G. M., Pinheiro, R. O., et al. (2020). A cardiac rehabilitation exercise program potentially inhibits progressive inflammation in patients with severe chagas cardiomyopathy: a pilot single-arm clinical trial. *J. Res. Med. Sci.* 25, 18. doi:10.4103/jrms.JRMS_175_18
- Roncero-Ramos, I., Rangel-Zuñiga, O. A., Lopez-Moreno, J., Alcala-Diaz, J. F., Perez-Martinez, P., Jimenez-Lucena, R., et al. (2018). Mediterranean diet, glucose homeostasis, and inflammasome genetic variants: the CORDIOPREV study. *Mol. Nutr. Food Res.* 62 (9), e1700960. doi:10.1002/mnfr.201700960
- Rothschild, J. A., and Bishop, D. J. (2020). Effects of dietary supplements on adaptations to endurance training. *Sports Med.* 50 (1), 25–53. doi:10.1007/s40279-019-01185-8
- Ruan, J. (2019). Structural insight of gasdermin family driving pyroptotic cell death. *Adv. Exp. Med. Biol.* 1172, 189–205. doi:10.1007/978-981-13-9367-9_9
- Sacco, S. J., Park, C. L., Suresh, D. P., and Bliss, D. (2014). Living with heart failure: psychosocial resources, meaning, gratitude and well-being. *Heart Lung* 43, 213–218. doi:10.1016/j.hrtlung.2014.01.012
- Saló, D. C., Donovan, C. M., and Davies, K. J. (1991). HSP70 and other possible heat shock or oxidative stress proteins are induced in skeletal muscle, heart, and liver during exercise. *Free Radic. Biol. Med.* 11, 239–246. doi:10.1016/0891-5849(91)90119-N
- Sandall, C. F., and MacDonald, J. A. (2019). Effects of phosphorylation on the NLRP3 inflammasome. *Arch. Biochem. Biophys.* 670, 43–57. doi:10.1016/j.abb.2019.02.020
- Sandanger, Ø., Ranheim, T., Vinge, L. E., Bliksoen, M., Alfsnes, K., Finsen, A. V., et al. (2013). The NLRP3 inflammasome is up-regulated in cardiac fibroblasts and mediates myocardial ischemia-reperfusion injury. *Cardiovasc. Res.* 99, 164–174. doi:10.1093/cvr/cvt091

- Schoenfeld, B. J., Ogborn, D., and Krieger, J. W. (2016). Effects of resistance training frequency on measures of muscle hypertrophy: a systematic review and meta-analysis. *Sports Med.* 46 (11), 1689–1697. doi:10.1007/s40279-016-0543-8
- Schwartz, R. S., Kraus, S. M., Schwartz, J. G., Wickstrom, K. K., Peichel, G., Garberich, R. F., et al. (2014). Increased coronary artery plaque volume among male marathon runners. *Mo Med.* 111 (2), 89–94. doi:10.1016/j.paid.2014.03.024
- Spalding, K. L., Arner, E., Westermark, P. O., Bernard, S., Buchholz, B. A., Bergmann, O., et al. (2008). Dynamics of fat cell turnover in humans. *Nature* 453, 783–787. doi:10.1038/nature06902
- Spalinger, M. R., Kasper, S., Gottlieb, C., Lang, S., Atrott, K., Vavricka, S. R., et al. (2023). NLRP3 tyrosine phosphorylation is controlled by protein tyrosine phosphatase PTPN22. *J. Clin. Invest.* 133 (4), e169304. doi:10.1172/JCI169304
- Spalinger, M. R., Schwarzfischer, M., and Scharl, M. (2020). The role of Protein tyrosine phosphatases in inflammasome activation. *Int. J. Mol. Sci.* 21 (15), 5481. doi:10.3390/ijms21155481
- Stensvold, D., Slørdahl, S. A., and Wisløff, U. (2012). Effect of exercise training on inflammation status among people with metabolic syndrome. *Metab. Syndr. Relat. Disord.* 10, 267–272. doi:10.1089/met.2011.0140
- Stienstra, R., van Diepen, J. A., Tack, C. J., Zaki, M. H., Perera, D., Neale, G. A., et al. (2011). Inflammasome is a central player in the induction of obesity and insulin resistance. *Proc. Natl. Acad. Sci. U. S. A.* 108, 15324–15329. doi:10.1073/pnas.1100255108
- Stolen, K. Q., Kemppainen, J., Ukkonen, H., Kalliokoski, K. K., Luotolahti, M., Lehtikoinen, P., et al. (2003). Exercise training improves biventricular oxidative metabolism and left ventricular efficiency in patients with dilated cardiomyopathy. *J. Am. Coll. Cardiol.* 41 (3), 460–467. doi:10.1016/s0735-1097(02)02772-9
- Stumvoll, M., Goldstein, B. J., and van Haften, T. W. (2005). Type 2 diabetes: principles of pathogenesis and therapy. *Lancet* 365, 1333–1346. doi:10.1016/S0140-6736(05)61032-X
- Sun, H., Zhao, H., Yan, Z., Liu, X., Yin, P., and Zhang, J. (2021). Protective role and molecular mechanism of action of nesfatin-1 against high glucose-induced inflammation, oxidative stress and apoptosis in retinal epithelial cells. *Exp. Ther. Med.* 22, 833. doi:10.3892/etm.2021.10265
- Sun, H. J., Ren, X. S., Xiong, X. Q., Chen, Y. Z., Zhao, M. X., Wang, J. J., et al. (2017). NLRP3 inflammasome activation contributes to VSMC phenotypic transformation and proliferation in hypertension. *Cell. Death Dis.* 8, e3074. doi:10.1038/cddis.2017.470
- Sun, Y., and Ding, S. (2021). NLRP3 inflammasome in diabetic cardiomyopathy and exercise intervention. *Int. J. Mol. Sci.* 22, 13228. doi:10.3390/ijms222413228
- Swanson, K. V., Deng, M., and Ting, J. P. (2019). The NLRP3 inflammasome: molecular activation and regulation to therapeutics. *Nat. Rev. Immunol.* 19 (8), 477–489. doi:10.1038/s41577-019-0165-0
- Takahashi, M. (2019). Cell-specific roles of NLRP3 inflammasome in myocardial infarction. *J. Cardiovasc. Pharmacol.* 74 (3), 188–193. doi:10.1097/FJC.0000000000000709
- Terentes-Printzios, D., Ioakeimidis, N., Rokkas, K., and Vlachopoulos, C. (2022). Interactions between erectile dysfunction, cardiovascular disease and cardiovascular drugs. *Nat. Rev. Cardiol.* 19 (1), 59–74. doi:10.1038/s41569-021-00593-6
- Thomas, K. N., Cotter, J. D., Williams, M. J., and van Rij, A. M. (2016). Repeated episodes of remote ischemic preconditioning for the prevention of myocardial injury in vascular surgery. *Vasc. Endovasc. Surg.* 50 (3), 140–146. doi:10.1177/1538574416639150
- Troosters, T., Janssens, W., Demeyer, H., and Rabinovich, R. A. (2023). Pulmonary rehabilitation and physical interventions. *Eur. Respir. Rev.* 32 (168), 220222. doi:10.1183/16000617.0222-2022
- Trøseid, M., Lappégård, K. T., Mollnes, T. E., Arnesen, H., and Seljeflot, I. (2009). The effect of exercise on serum levels of interleukin-18 and components of the metabolic syndrome. *Metab. Syndr. Relat. Disord.* 7, 579–584. doi:10.1089/met.2009.0003
- Tucker, W. J., Fegers-Wustrow, I., Halle, M., Haykowsky, M. J., Chung, E. H., and Kovacic, J. C. (2022). Exercise for primary and secondary prevention of cardiovascular disease: jacc focus seminar 1/4. *J. Am. Coll. Cardiol.* 80, 1091–1106. doi:10.1016/j.jacc.2022.07.004
- Tutor, A. W., Lavie, C. J., Kachur, S., Milani, R. V., and Ventura, H. O. (2023). Updates on obesity and the obesity paradox in cardiovascular diseases. *Prog. Cardiovasc. Dis.* 78, 2–10. doi:10.1016/j.pcad.2022.11.013
- Vaiculeviciute, R., Bironaitė, D., Uzielienė, I., Mobasher, A., and Bernotienė, E. (2021). Cardiovascular drugs and osteoarthritis: effects of targeting ion channels. *Cells* 10 (10), 2572. doi:10.3390/cells10102572
- Valenzuela, P. L., Ruilope, L. M., Santos-Lozano, A., Wilhelm, M., Kränkel, N., Fiuza-Luces, C., et al. (2023). Exercise benefits in cardiovascular diseases: from mechanisms to clinical implementation. *Eur. Heart J.* 44 (21), 1874–1889. doi:10.1093/eurheartj/ehad170
- Vandanmagsar, B., Youm, Y. H., Ravussin, A., Galgani, J. E., Stadler, K., Mynatt, R. L., et al. (2011). The NLRP3 inflammasome instigates obesity-induced inflammation and insulin resistance. *Nat. Med.* 17, 179–188. doi:10.1038/nm.2279
- Veeranki, S., and Tyagi, S. C. (2013). Defective homocysteine metabolism: potential implications for skeletal muscle malfunction. *Int. J. Mol. Sci.* 14, 15074–15091. doi:10.3390/ijms140715074
- Vella, C. A., Taylor, K., and Drummer, D. (2017). High-intensity interval and moderate-intensity continuous training elicit similar enjoyment and adherence levels in overweight and obese adults. *Eur. J. Sport Sci.* 17 (9), 1203–1211. doi:10.1080/17461391.2017.1359679
- Vincent, H. K., Bourguignon, C., and Vincent, K. R. (2006). Resistance training lowers exercise-induced oxidative stress and homocysteine levels in overweight and obese older adults. *Obes. (Silver Spring)* 14, 1921–1930. doi:10.1038/oby.2006.224
- Wada, T., Ishikawa, A., Watanabe, E., Nakamura, Y., Aruga, Y., Hasegawa, H., et al. (2017). Eplerenone prevented obesity-induced inflammasome activation and glucose intolerance. *J. Endocrinol.* 235 (3), 179–191. doi:10.1530/JOE-17-0351
- Wang, B., Zhou, R., Wang, Y., Liu, X., Shou, X., Yang, Y., et al. (2020). Effect of high-intensity interval training on cardiac structure and function in rats with acute myocardial infarct. *Biomed. Pharmacother.* 131, 110690. doi:10.1016/j.biopha.2020.110690
- Wang, Q., Hu, J., Liu, Y., Li, J., Liu, B., Li, M., et al. (2019). Aerobic exercise improves synaptic-related proteins of diabetic rats by inhibiting FOXO1/NF-κB/NLRP3 inflammatory signaling pathway and ameliorating PI3K/Akt insulin signaling pathway. *J. Mol. Neurosci.* 69 (1), 28–38. doi:10.1007/s12031-019-01302-2
- Wang, R., Wang, Y., Mu, N., Lou, X., Li, W., Chen, Y., et al. (2017). Activation of NLRP3 inflammasomes contributes to hyperhomocysteinemia-aggravated inflammation and atherosclerosis in apoE-deficient mice. *Lab. Invest.* 97, 922–934. doi:10.1038/labinvest.2017.30
- Wang, T., Li, J., Li, H., Zhong, X., Wang, L., Zhao, S., et al. (2022). Aerobic exercise inhibited P2X7 purinergic receptors to improve cardiac remodeling in mice with type 2 diabetes. *Front. Physiol.* 13, 828020. doi:10.3389/fphys.2022.828020
- Watanabe, S., Usui-Kawanishi, F., Karasawa, T., Kimura, H., Kamata, R., Komada, T., et al. (2020). Glucose regulates hypoxia-induced NLRP3 inflammasome activation in macrophages. *J. Cell. Physiol.* 235 (10), 7554–7566. doi:10.1002/jcp.29659
- Winchester, L., Veeranki, S., Givvimani, S., and Tyagi, S. C. (2014). Exercise mitigates the adverse effects of hyperhomocysteinemia on macrophages, MMP-9, skeletal muscle, and white adipocytes. *Can. J. Physiol. Pharmacol.* 92, 575–582. doi:10.1139/cjpp-2014-0059
- Wong, N. D., and Sattar, N. (2023). Cardiovascular risk in diabetes mellitus: epidemiology, assessment and prevention. *Nat. Rev. Cardiol.* 20 (10), 685–695. doi:10.1038/s41569-023-00877-z
- Wu, J., Zhang, Y. F., Li, J. S., Zhu, G. L., Bi, Z. M., and Li, X. Y. (2019). The effect of high glucose-based peritoneal dialysis fluids on thioredoxin-interacting protein expression in human peritoneal mesothelial cells. *Int. Immunopharmacol.* 66, 198–204. doi:10.1016/j.intimp.2018.11.027
- Wu, Y., Pan, B., Zhang, Z., Li, X., Leng, Y., Ji, Y., et al. (2022). Caspase-4/11-mediated pulmonary artery endothelial cell pyroptosis contributes to pulmonary arterial hypertension. *Hypertension* 79, 536–548. doi:10.1161/HYPERTENSIONAHA.121.17868
- Xi, X., Zhang, R., Chi, Y., Zhu, Z., Sun, R., and Gong, W. (2024). TXNIP regulates NLRP3 inflammasome-induced pyroptosis related to aging via cAMP/PKA and PI3K/Akt signaling pathways. *Mol. Neurobiol.* doi:10.1007/s12035-024-04089-5
- Xian, H., Watari, K., Sanchez-Lopez, E., Offenberger, J., Onyuru, J., Sampath, H., et al. (2022). Oxidized DNA fragments exit mitochondria via mPTP- and VDAC-dependent channels to activate NLRP3 inflammasome and interferon signaling. *Immunity* 55 (8), 1370–1385.e8. doi:10.1016/j.immuni.2022.06.007
- Xie, Y., Huang, Y., Ling, X., Qin, H., Wang, M., and Luo, B. (2020). Chemerin/cmkrl1 axis promotes inflammation and pyroptosis by activating NLRP3 inflammasome in diabetic cardiomyopathy rat. *Front. Physiol.* 11, 381. doi:10.3389/fphys.2020.00381
- Xin, C., Zhang, J., Hao, N., Wang, J., Liu, H., Wei, H., et al. (2022). Irisin inhibits NLRP3 inflammasome activation in HG/HF incubated cardiac microvascular endothelial cells with H/R injury. *Microcirculation* 29 (8), e12786. doi:10.1111/micc.12786
- Xu, M., Duan, Y., and Xiao, J. (2019). Exercise improves the function of endothelial cells by microRNA. *J. Cardiovasc. Transl. Res.* 12, 391–393. doi:10.1007/s12265-018-9855-4
- Yang, Q., Chen, S., Wang, X., Yang, X., Chen, L., Huang, T., et al. (2023). Exercise mitigates endothelial pyroptosis and atherosclerosis by downregulating NEAT1 through N6-methyladenosine modifications. *Arterioscler. Thromb. Vasc. Biol.* 43, 910–926. doi:10.1161/ATVBAHA.123.319251
- Yuan, B., Zhou, X. M., You, Z. Q., Xu, W. D., Fan, J. M., Chen, S. J., et al. (2020). Inhibition of AIM2 inflammasome activation alleviates GSDMD-induced pyroptosis in early brain injury after subarachnoid haemorrhage. *Cell. Death Dis.* 11 (1), 76. doi:10.1038/s41419-020-2248-z
- Zeng, C., Duan, F., Hu, J., Luo, B., Huang, B., Lou, X., et al. (2020). NLRP3 inflammasome-mediated pyroptosis contributes to the pathogenesis of non-ischemic dilated cardiomyopathy. *Redox Biol.* 34, 101523. doi:10.1016/j.redox.2020.101523

- Zeng, C., Wang, R., and Tan, H. (2019). Role of pyroptosis in cardiovascular diseases and its therapeutic implications. *Int. J. Biol. Sci.* 15, 1345–1357. doi:10.7150/ijbs.33568
- Zeng, Z., Zheng, Q., Chen, J., Tan, X., Li, Q., Ding, L., et al. (2020). FGF21 mitigates atherosclerosis via inhibition of NLRP3 inflammasome-mediated vascular endothelial cells pyroptosis. *Exp. Cell. Res.* 393 (2), 112108. doi:10.1016/j.yexcr.2020.112108
- Zhang, L., Yuan, M., Zhang, L., Wu, B., and Sun, X. (2019). Adiponectin alleviates nlrp3-inflammasome-mediated pyroptosis of aortic endothelial cells by inhibiting foxo4 in arteriosclerosis. *Biochem. Biophys. Res.* 514, 266–272. doi:10.1016/j.bbrc.2019.04.143
- Zhang, Q., Yu, W., Lee, S., Xu, Q., Naji, A., and Le, A. D. (2015). Bisphosphonate induces osteonecrosis of the jaw in diabetic mice via NLRP3/caspase-1-dependent IL-1 β mechanism. *J. Bone Min. Res.* 30, 2300–2312. doi:10.1002/jbmr.2577
- Zhang, T., Ding, S., and Wang, R. (2021). Research progress of mitochondrial mechanism in NLRP3 inflammasome activation and exercise regulation of NLRP3 inflammasome. *Int. J. Mol. Sci.* 22 (19), 10866. doi:10.3390/ijms221910866
- Zhang, W., Li, G., Luo, R., Lei, J., Song, Y., Wang, B., et al. (2022). Cytosolic escape of mitochondrial DNA triggers cGAS-STING-NLRP3 axis-dependent nucleus pulposus cell pyroptosis. *Exp. Mol. Med.* 54 (2), 129–142. doi:10.1038/s12276-022-00729-9
- Zhang, W., Xu, X., Kao, R., Mele, T., Kvietys, P., Martin, C. M., et al. (2014). Cardiac fibroblasts contribute to myocardial dysfunction in mice with sepsis: the role of NLRP3 inflammasome activation. *PLoS One* 9, e107639. doi:10.1371/journal.pone.0107639
- Zhang, X., Li, Y., Li, H., Li, W., and Li, Y. (2021). Mitochondrial ROS-induced TXNIP upregulation contributes to NLRP3 inflammasome activation in diabetic cardiomyopathy. *Redox Biol.* 44, 101913. doi:10.1016/j.redox.2021.101913
- Zhao, N., Li, C. C., Di, B., and Xu, L. L. (2020). Recent advances in the NEK7-licensed NLRP3 inflammasome activation: mechanisms, role in diseases and related inhibitors. *J. Autoimmun.* 113, 102515. doi:10.1016/j.jaut.2020.102515
- Zhaolin, Z., Jiaojiao, C., Peng, W., Yami, L., Tingting, Z., Jun, T., et al. (2019). OxLDL induces vascular endothelial cell pyroptosis through miR-125a-5p/TET2 pathway. *J. Cell. Physiol.* 234, 7475–7491. doi:10.1002/jcp.27509
- Zhong, X., Zeng, H., Zhou, Z., Su, Y., Cheng, H., Hou, Y., et al. (2023). Structural mechanisms for regulation of GSDMB pore-forming activity. *Nature* 616 (7957), 598–605. doi:10.1038/s41586-023-05872-5
- Zhou, R., Tardive, I. A., Thorens, B., Choi, I., and Tschopp, J. (2010). Thioredoxin-interacting protein links oxidative stress to inflammasome activation. *Nat. Immunol.* 11, 136–140. doi:10.1038/ni.1831
- Zhou, Y., Tong, Z., Jiang, S., Zheng, W., Zhao, J., and Zhou, X. (2020). The roles of endoplasmic reticulum in NLRP3 inflammasome activation. *Cells* 9 (5), 1219. doi:10.3390/cells9051219
- Zhou, Z., Ying, C., Zhou, X., Shi, Y., Xu, J., Zhu, Y., et al. (2022). Aerobic exercise training alleviates renal injury in db/db mice through inhibiting Nox4-mediated NLRP3 inflammasome activation. *Exp. Gerontol.* 168, 111934. doi:10.1016/j.exger.2022.111934

Glossary

| | |
|--------------------------|---|
| ASC | apoptosis-associated speck-like protein |
| AS | atherosclerosis |
| AT | adipose tissue |
| BMDM | bone marrow-derived macrophages |
| BP | blood pressure |
| CVD | cardiovascular diseases |
| CARD | caspase recruitment domain |
| DAMPs | damage-associated molecular patterns |
| DCM | diabetic cardiomyopathy |
| Drp1 | dynammin-related protein 1 |
| ET | endurance training |
| EP | exercise preconditioning |
| EE | exhaustive exercise |
| EEE | excessive endurance exercise |
| FGF21 | fibroblast growth factor 21 |
| FOXO1 | forkhead box transcription factor O1 |
| GSDMD | gasdermin D |
| NT | n-terminal |
| NOX4 | nicotinamide adenine dinucleotide phosphate oxidase 4 |
| HFD | high-fat diet |
| HMGB-1 | high mobility group box-1 |
| Hcy | homocysteine |
| HHcy | hyperhomocysteinemia |
| HIIT | high-intensity interval training |
| HF | heart failure |
| IL | interleukin |
| LRR | leucine-rich repeat |
| LICT | low-intensity continuous training |
| IR | insulin resistance |
| LPS | lipopolysaccharide |
| β-OHB | β-Hydroxybutyrate |
| MICT | moderate-intensity continuous training |
| MI | myocardial infarction |
| mtROS | mitochondrial ROS |
| HR_{max} | maximum heart rate |
| VO_{2max} | maximum volume of oxygen |
| NLRP3 | NOD-like receptor protein 3 |
| NLR | nucleotide-binding domain-like receptor |
| NF-κB | nuclear factor-κB |
| NEAT1 | nuclear paraspeckle assembly transcript 1 |
| PAMPs | pathogen-associated molecular patterns |

| | |
|--------------|---|
| PYD | pyrin domain |
| P2X7 | purinergic receptor P2X, ligand-gated ion channel 7 |
| ROS | reactive oxygen species |
| RT | resistance training |
| SHR | spontaneously hypertensive rats |
| TGN | trans-golgi network |
| TXNIP | thioredoxin interacting protein |



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Amentoflavone for treating cardiocerebrovascular diseases and neurological disorders

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Amentoflavone (AME) is a flavonoid compound found in over 120 plants. Its extensive pharmacological activity for treating cardiocerebrovascular diseases and neurological disorders have attracted the attention of researchers in recent years. However, owing to the poor solubility and low bioavailability of AME, it has not been developed as a drug for treating these diseases. This review focuses on two aspects of AME: First, it provides a detailed summary and introduction to AME based on its chemical structure, physicochemical properties, plant sources, extraction and purification methods, administration systems, and pharmacokinetic properties. Second, it summarizes the effects of AME on cardiocerebrovascular diseases and neurological disorders, and its specific pharmacological mechanisms. This review aims to promote the use of AME for treating cardiocerebrovascular diseases and neurological disorders. AME exhibits multiple activities, indicating its potential as a natural drug for treating these diseases. Further studies on its pharmacokinetics and toxicology are required to ensure its safety and efficacy.

KEYWORDS

amentoflavone, cardiovascular diseases, cerebrovascular diseases, neurological diseases, pharmacological effects, anti-inflammatory agents, Chinese herbal medicine

1 Introduction

Cardiocerebrovascular diseases refer to a group of conditions that affect the heart, brain, and other tissues. This category includes both cardiovascular and cerebrovascular diseases, as well as a variety of ischemic and hemorrhagic conditions. These diseases are precipitated by factors such as hyperlipidemia, thickened blood, atherosclerosis, and hypertension (Yan and Guo, 2022). Cardiovascular diseases are prevalent among individuals aged over 50 years, causing high morbidity and ranking first as a cause of death (Ma et al., 2021). In 2019, cardiovascular diseases accounted for approximately one-third of global deaths, with the highest number of deaths occurring in China (Roth et al., 2020). According to the 2022 *China Cardiovascular Health and Disease Report*, China currently has 13 million people with stroke, 11.39 million people with coronary heart disease, 8.9 million people with heart failure, 5 million people with cor pulmonale, 4.87 million people with atrial fibrillation, 2.5 million people with rheumatic heart disease, 2 million people with congenital heart disease, 45.3 million people with peripheral artery disease, and

245 million people with hypertension (The Writing Committee Of The Report On Cardiovascular Health And Diseases and Hu, 2023). Neurological disorders are also widely prevalent. Neurological diseases were the second leading cause of death globally in 2015, resulting in approximately 9.4 million deaths, accounting for 16.8% of deaths. From 1990 to 2015, the number of deaths due to neurological diseases increased by 36.7% and disability-adjusted life years increased by 7.4% (Global Burden of Disease Study, 2013 Collaborators, 2015; Zhou et al., 2024). The high morbidity and mortality rates associated with these diseases urgently require effective treatment modalities to curb their progression (*Ginkgo biloba* L.). extract (GBE) has been approved for clinical use for treating various cardiovascular, metabolic, and neurodegenerative disorders (Peng et al., 2024). One of the critical components in GBE is flavonoids, which comprise up to 24% of its total content (Tao et al., 2022).

Amentoflavone (AME) is a flavonoid compound first isolated from the *Selaginella* plant (Okigawa et al., 1971). AME is one of the most common biflavonoid compounds found in *Ginkgo biloba* (Šamec et al., 2022). However, some studies suggest that the AME in *Ginkgo biloba* leaf extracts exhibits no biological activity and AME has been removed from the listing of the active components of such extracts (Xu et al., 2013). Recent research has demonstrated that AME has a wide range of pharmacological properties, including anti-inflammatory (Zhang and Wang, 2013), antioxidant (Li et al., 2020), anti-aging (Park and Kim, 2019), antibacterial (Bajpai et al., 2019), antiviral (Lee et al., 2023), anti-tumor (Qiu S. et al., 2021), antidepressant and anxiolytic (Ishola et al., 2012). AME also has beneficial effects on cardiocerebrovascular diseases and neurological diseases (Li et al., 2021; Saeedan et al., 2023; Sirimangkalakitti et al., 2019).

A bibliometric analysis was conducted on the literature pertaining to AME from 2014 to 2023, providing insights into the research advancements in the field on a global scale. The keyword “Amentoflavone” was queried in the Web of Science database, yielding a total of 394 research papers from 63 countries and 641 institutions, authored by 2,196 individuals. Figure 1A displays the countries with the highest number of publications, providing insight into the level of interest in AME among these nations. Analysis of the data indicates a consistent upward trend in publication output over the past decade, with the number of articles published in recent years significantly surpassing those published in 2014. This trend suggests sustained interest and advancement in the field of AME, as depicted in Figure 1B. Research on the role of AME in cardiocerebrovascular diseases and neurological disorders has gained increasing attention since 2015, emerging as a current research hotspot in the field. Furthermore, the impact of AME on other conditions such as tumors and diabetes should not be underestimated.

This review provides a comprehensive summary of the chemical structure and physicochemical properties of AME, encompassing its botanical sources, extraction and purification techniques, and administration routes. Additionally, it examines the pharmacokinetic characteristics of AME, alongside its therapeutic effects on cardiocerebrovascular diseases and neurological disorders, elucidating the underlying pharmacological mechanisms. The objective of this review is to enhance the understanding of AME

and its potential applications in the treatment of cardiocerebrovascular diseases and neurological disorders.

2 Background

2.1 Chemical structure and physicochemical properties of AME

AME is a biflavonoid compound with the chemical name 8-[5-(5,7-dihydroxy-4-oxo-4H-chromen-2-yl)-2-hydroxyphenyl]-5,7-dihydroxy-2-(4-hydroxyphenyl)-4H-chromen-4-one. It is an apigenin dimer, with multiple double bonds and hydroxyl groups in its molecular structure (Figure 2). The C₂-C₃ double bond is susceptible to hydrogenation, whereas the hydroxyl group is prone to substitution with methoxy groups. Therefore, many hydroxylated derivatives have an amentoflavone nucleus (Yu et al., 2017; Šamec et al., 2022; Xiao et al., 2018). It has been reported that the hydroxyl groups at positions C₇ and C_{4'} are crucial for the anti-inflammatory activity of AME. Substitution of these hydroxyl groups with methoxy groups significantly reduces the anti-inflammatory efficacy (Mangmool et al., 2024). Additionally, the atropisomerism exhibited due to the C₃'-C₈ linkage in the structure of AME may affect its binding to targets, potentially influencing its biochemical activities (Bhattacharya and Mandal, 2024).

The molecular formula of AME is C₃₀H₁₈O₁₀, the molecular weight is 538.46 g/mol, and the melting point is 300°C (Xiong et al., 2021). The cross-conjugated molecular structure has strong characteristic absorption of ultraviolet light (Li, 2011). AME is a planar molecule with a close arrangement between molecules and large intermolecular attraction; therefore, it is poorly soluble in water, but is easily soluble in organic solvents such as ethanol and dimethyl sulfoxide (Xiong et al., 2021). Among the various polymorphs of AME, the amorphous form exhibits higher dissolution rates and solubility, and demonstrates good physical stability during the dissolution process (Zhou et al., 2022). It has been reported that the monomer apigenin from AME can interrupt free radical chain reactions and reduce the photooxidation process by generating resonance-stabilized free radicals (Huvaere et al., 2012). The presence of catechol structures and a double bond adjacent to the carbonyl group, along with hydroxyl groups on a single benzene ring, plays a crucial role in effectively quenching singlet oxygen (Nagai et al., 2005). This indirectly suggests that AME may exhibit strong antioxidant and photostability properties.

2.2 AME sources, and methods of extraction, separation, and purification

2.2.1 Sources

AME is a bioactive compound present in numerous plant species. It was initially isolated from the leaves of (*Selaginella tamariscina* Maxim.), (*Selaginella rupestris* L.), and *Ginkgo biloba*. Subsequently, it has been extracted from over 120 plants species, including (*Celaenodendron mexicanum* L.), (*Cupressus funebris* Endl.), (*Garcinia multiflora* Bl.), and (*Hypericum perforatum* L.) (Xiong et al., 2021). The *Selaginella* genus, comprising 21 species, is

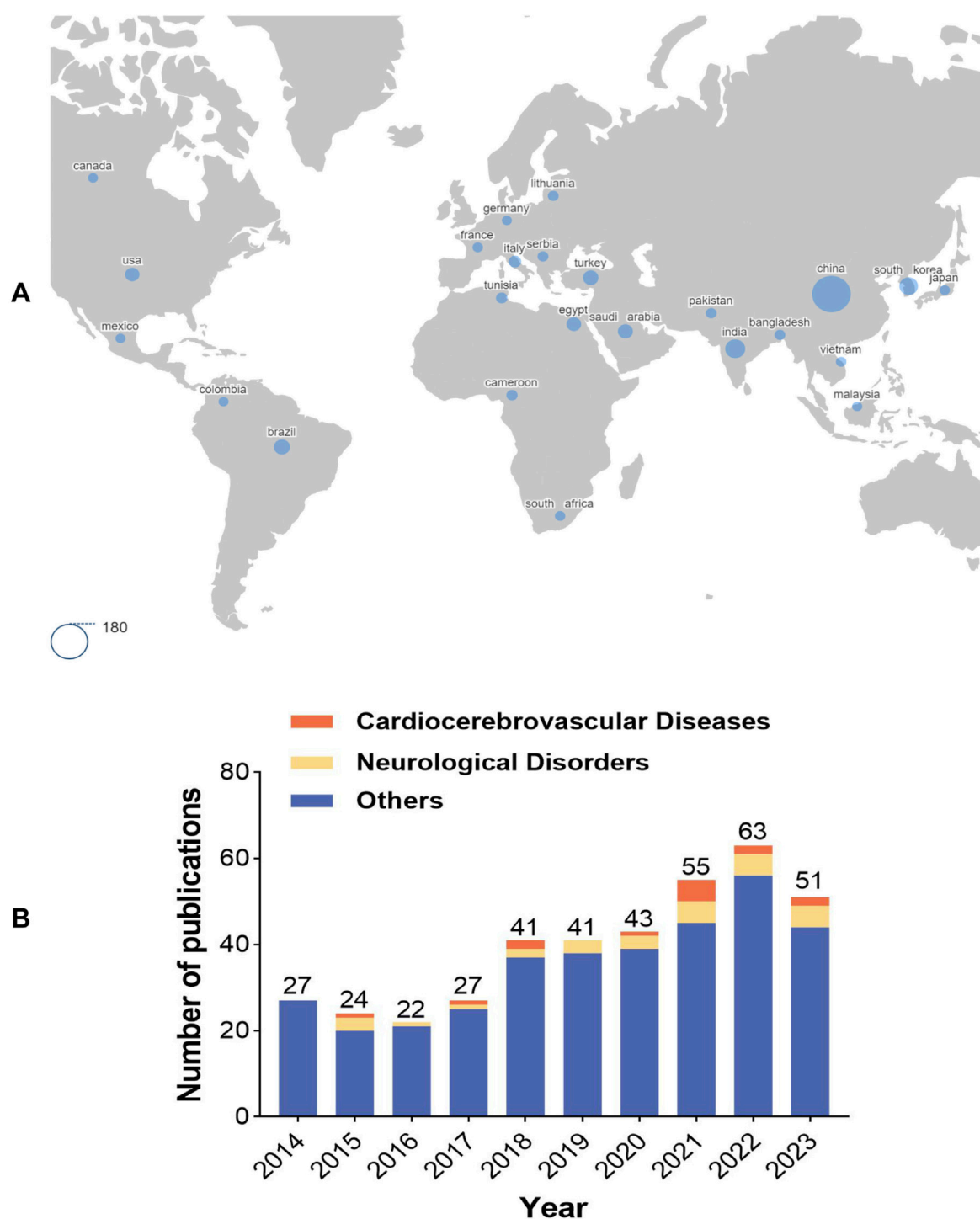


FIGURE 1

Bibliometric analysis of AME (A). Global distribution map of countries involved in AME. The larger the blue circle in the figure is, the greater the number of articles published by the corresponding country. The area of the circle in the lower left corner is equivalent to 180 articles. (B). Number of publications on AME in the past decade. Others represented other research fields of AME, with tumors and diabetes as the main research field.

the most prevalent source of amentoflavone. Other significant botanical families containing this compound include Cupressaceae, Euphorbiaceae, and Clusiaceae. Typically, AME is extracted from the leaves, aerial parts, and whole plants (Yu et al., 2017).

2.2.2 Extraction methods

The principal methods for the extraction of AME include ultrasonic-assisted extraction, microwave-assisted extraction, organic solvent extraction, and semi-bionic extraction (Table 1). Each of these methodologies presents distinct advantages and

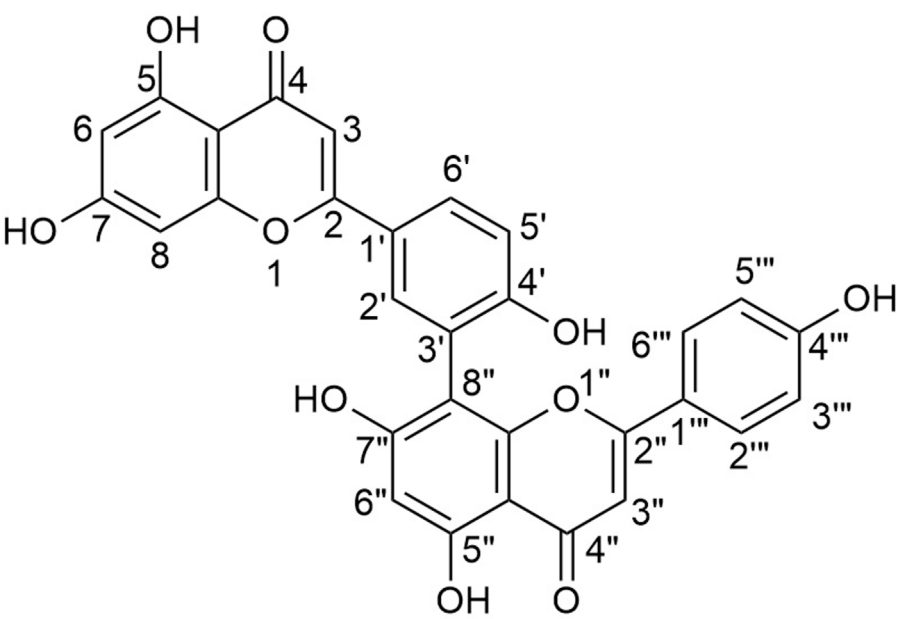


FIGURE 2
Chemical structure of amentoflavone (8-[5-(5,7-dihydroxy-4-oxo-4H-chromen-2-yl)-2-hydroxyphenyl]-5,7-dihydroxy-2-(4-hydroxyphenyl)-4H-chromen-4-one).

TABLE 1 Methods of extracting amentoflavone.

| Source | Extraction method | Extraction yield (%) | Reference |
|-----------------------------------|--|----------------------|---------------------|
| <i>Selaginella tamariscina</i> | Infrared-assisted extraction | 0.290 | Wang et al. (2018a) |
| <i>Selaginella tamariscina</i> | Ultrasonic-assisted ionic liquid extraction | 1.351 | Jiang et al. (2020) |
| <i>Selaginella tamariscina</i> | Ethanol reflux extraction | 1.274 | Wei (2010) |
| <i>Selaginella pulvinata</i> | Ethanol reflux extraction | 1.728 | Luo (2017) |
| <i>Selaginella sinensis</i> | Ionic liquid-microwave-based extraction | 0.196 | Li YY et al. (2019) |
| <i>Selaginella moellendorffii</i> | Deep eutectic solvent extraction | 0.275 | Liu et al. (2022) |
| <i>Selaginella doederleinii</i> | Ionic liquid-microwave-based extraction | 0.650 | Wang et al. (2018b) |
| <i>Selaginella doederleinii</i> | Microwave-assisted extraction | 0.330 | Wang et al. (2018c) |
| <i>Selaginella uncinata</i> | Ultrasonic-assisted extraction | 1.530 | Lai et al. (2018a) |
| <i>Podocarpus nagi</i> | Ethanol reflux extraction | 0.010 | Wang (2017) |
| <i>Cunninghamia lanceolata</i> | Ultrasonic-assisted extraction | 0.319 | Wang et al. (2023) |
| <i>Taxus chinensis</i> | Supercritical-CO ₂ fluid extraction | 0.447 | Ruan et al. (2014) |

disadvantages. For example, ultrasonic-assisted extraction offers benefits such as reduced extraction time, simplicity of operation, and high extraction efficiency. Nevertheless, its limited effective action area renders it unsuitable for industrial-scale production. Conversely, microwave-assisted extraction is noted for its straightforward operation, minimal byproduct formation, high extraction rates, and ease of product purification. However, it necessitates elevated extraction temperatures, which may compromise the integrity of active components. Organic solvent extraction is both cost-effective and straightforward; however, it suffers from low extraction efficiency, environmental pollution, and

potential risks to human health make it less suitable (Li, 2011; Xu et al., 2021). Eutectic solvent (Liu et al., 2022) and infrared-assisted (Wang Y. et al., 2018) extraction techniques are gaining prominence in the extraction of AME due to their environmental sustainability, rapid processing times, and high efficiency.

2.2.3 Separation and purification methods

Currently, the primary methodologies for the separation and purification of AME are silica gel column chromatography, two-step precipitation, polyamide column chromatography, macroporous resin adsorption, and preparative high-performance liquid

TABLE 2 Methods of separating and purifying amentoflavone.

| Source | Separation and purification method | Purity (%) | Recovery (%) | Reference |
|-----------------------------------|--|------------|--------------|--------------------|
| <i>Selaginella tamariscina</i> | Two-step precipitation method | 58.2 | 88.7 | Wei (2010) |
| <i>Selaginella tamariscina</i> | High-speed countercurrent chromatography | 99.2 | 94.7 | Wei (2010) |
| <i>Selaginella tamariscina</i> | Silica gel column chromatography | 97.2 | 50.7 | Wei (2010) |
| <i>Selaginella tamariscina</i> | Low pressure column chromatography | 98.7 | 87.8 | Wei (2010) |
| <i>Selaginella pulvinata</i> | Macroporous adsorption resin (HPD 300) | 52.5 | 62.4 | Luo (2017) |
| <i>Selaginella pulvinata</i> | Two-step precipitation method | 47.9 | 59.7 | Luo (2017) |
| <i>Selaginella uncinata</i> | Macroporous adsorption resin (NKA-9) | N/A | 64.3 | Lai et al. (2018b) |
| <i>Selaginella moellendorffii</i> | Macroporous adsorption resin (D-101) | 80.8 | 62.5 | Fang et al. (2011) |
| <i>Podocarpus nagi</i> | Macroporous adsorption resin (AB-8) | 93.6 | N/A | Wang (2017) |

chromatography (HPLC) (Table 2). Researchers have been actively investigating enhanced techniques for separation and purification. Recently, flash chromatography has emerged as a novel method. Compared to traditional silica gel column chromatography, flash chromatography offers reduced the loading time, minimized dead adsorption, and improved efficiency and product purity. However, it exhibits lower separation capacity and is prone to interference from metal ions during elution. Macroporous resin adsorption offers several advantages, including rapid high-capacity and selective adsorption, as well as high elution efficiency, rendering it suitable for large-scale production. Nonetheless, its desorption efficiency remains suboptimal, and the purification rate is affected by the type of eluent and temperature (Xu et al., 2021). HPLC offers advantages including wide applicability, exceptional quantitative capabilities, and well-recognized methodologies. Nevertheless, it is marked by the necessity for extensive sample preparation, prolonged analysis durations, and expensive instrumentation (Vlasiou, 2023).

2.3 AME dosing forms

AME exhibits significant pharmaceutical potential and promise as a therapeutic agent (Xiong et al., 2021; Li et al., 2021). However, its poor water solubility constrains its release within the body, resulting in incomplete gastrointestinal absorption and low oral bioavailability, thereby limiting its pharmacological efficacy (Ren et al., 2013a). Following oral administration in rats, AME is dominantly distributed in the small intestine, stomach, liver, and large intestine, with minimal distribution to other tissues. To maximize the pharmacological efficacy of AME, it is imperative to enhance its solubility and bioavailability. Key strategies for improving drug solubility and bioavailability include modifying the delivery method, formulation (Table 3), and structural enhancement (Feng et al., 2020).

2.4 Pharmacokinetics of AME

Despite the multiple beneficial biological properties of AME, its pharmacokinetics remain inadequately characterized. A comprehensive understanding of AME’s pharmacokinetics is

crucial for elucidating its *in vivo* mechanisms of action, characterizing its properties, and optimizing drug design and dosing, to maximize therapeutic efficacy. Insights into the fundamental pharmacokinetics of AME can be inferred from studies conducted in animal models (Table 4).

It has been demonstrated that AME is absorbed via passive diffusion in rats (Wei et al., 2017). Furthermore, a study employing the Caco-2 cell model indicated that AME exhibits intestinal absorption. The absorption may involve paracellular passive diffusion and clathrin-mediated endocytosis, while the efflux transporter appears to be uninvolved (Wang B. et al., 2020). The low bioavailability of AME is due to extensive glucuronidation catalyzed by uridine diphosphate glucuronosyltransferase 1 family, polypeptide A (UGT1A1) and UGT1A3 (Gan et al., 2020).

Another study conducted demonstrated that the peak blood concentration in rats was reached at 90 min following oral administration, with a volume of distribution (V/F) of 198.36 ± 17.422 L/kg. These findings suggest that AME is predominant distributed or sequestered in specific tissues and organs within the rat model. (Wang et al., 2015).

In vivo, 34 metabolites of AME were identified, while 24 metabolites were identified *in vitro*. In the *in vivo* study, all metabolites were distributed in feces, with three of the 34 metabolites were also detected in urine, and none were found in bile or plasma. In the *in vitro* study, 20 of the 24 metabolites were distributed in liver microsomes, and 17 of the 24 metabolites were associated with the intestinal microbiota. Of the total metabolites identified both *in vivo* and *in vitro*, 14 were classified as phase I metabolites and 26 as phase II metabolites. The primary metabolic pathways included oxidation, methylation, acetylation, and oxidation methylation (Feng, 2020). It has been reported that the bioactive form of AME is likely conjugated (Liao et al., 2015). Another study found that 90.7% of AME circulates as conjugated metabolites post-administration. In rats, 73.2% and 70.2% of AME in plasma were in conjugated form following intravenous and intraperitoneal injection, respectively (Yu et al., 2017).

The cumulative excretion rates of AME in feces and urine were 23.93% and 0.82%, respectively, indicating that fecal excretion is the predominant route of AME elimination from the body. This high rate of fecal excretion may contribute to the compound’s low bioavailability (Chen et al., 2022).

TABLE 3 Different formulations to improve amentoflavone.

| Route of administration | Formulation | Particle size | Zeta potential | Improved results | Reference |
|-------------------------|---------------------------------|----------------------|---------------------|---|-----------------------|
| Oral | Micro-emulsion formulation | 15.37 ± 0.09 nm | −17.1 ± 0.24mV | Dissolution rate | Ren et al. (2013a) |
| Oral | Micro-powder formulation | 0.08 ± 0.01 μm | N/A | Solubility, dissolution rate | Ren et al. (2013b) |
| Intravenous | Nanoparticulate formulation | 77.3 ± 5.3 nm | −2.92 ± 0.27 mV | Solubility, dispersibility, stability, bioavailability, reduce toxicity | Zhao et al. (2022) |
| Oral | Micelle formulation | 58.8 ± 1.29 nm | 5.26 ± 0.63 mV | Bioavailability | Zhang et al. (2019) |
| Oral | Micelle formulation | 67.33 ± 2.01 nm | −0.84 ± 0.04 mV | Bioavailability | Feng et al. (2020) |
| Oral | Sub-micron particle formulation | Approximately 0.4 μm | N/A | Solubility, dispersibility, stability, dissolution rate | Duan et al. (2022) |
| Oral | Micelle formulation | 25.99 ± 0.10 nm | −27.67 ± 0.25 mV | Solubility, dissolution rate; Bioavailability | Feng et al. (2023) |
| Intranasal | Nanoemulsion formulation | Approximately 37 nm | Approximately −4 mV | Bioavailability | Khafagy et al. (2023) |

3 Biological activity of AME for treating cardiocerebrovascular and neurological diseases

Atherosclerosis is a main pathological basis for cardiovascular and cerebrovascular diseases (Dutta et al., 2023). The cerebrovascular system is closely related to the structure and function of brain tissue. Vascular factors are important in the development of neurological diseases (Iadecola, 2023; Smith et al., 2021). Therefore, this section discusses the effect of AME on cardiocerebrovascular and neurological diseases (Figure 3).

3.1 Potential mechanisms of AME in atherosclerosis

Although there is currently no published literature regarding the use of AME in the treatment of atherosclerosis, its various potential mechanisms of action are promising. The process of atherosclerotic plaque formation can be divided into the following steps (Figure 4): (1) When the vascular wall is damaged, low-density lipoprotein (LDL) enters the intimal layer of the blood vessel through the gaps between endothelial cells (ECs), forming oxidized low-density lipoprotein (ox-LDL). Monocytes migrate into the intimal layer and are activated into macrophages. (2) Ox-LDL binds to receptors on the surface of macrophages, and cholesterol enters the macrophages, where it is esterified. When the intake, esterification, and release of cholesterol are out of equilibrium, intracellular lipid overload leads to the formation of macrophage-derived foam cells, creating fatty streaks in the lesions (Gui et al., 2022). (3) Ox-LDL induces changes in the phenotype of vascular smooth muscle cells (VSMCs) in the tunica media of the arterial wall, leading to abnormal proliferation and migration to the intimal layer. Subsequently, VSMCs engulf ox-LDL to form myogenic foam cells (Yang, 2023; He, 2013), which then form a fibrous plaque. (4) Necrosis and disintegration of macrophage-derived and myogenic foam cells lead to atherosclerotic plaque formation. Inflammatory

cells secrete matrix metalloproteinases (MMPs) to break down collagen fibers in the extracellular matrix, thereby increasing plaque instability. Subsequently, the plaque ruptures, causing bleeding and thrombosis (Carracedo et al., 2019; Libby, 2021; Poznyak et al., 2020). Atherosclerosis is a chronic inflammatory disease involving various inflammatory, free radical, and oxidative stress-related injuries (Meng et al., 2024).

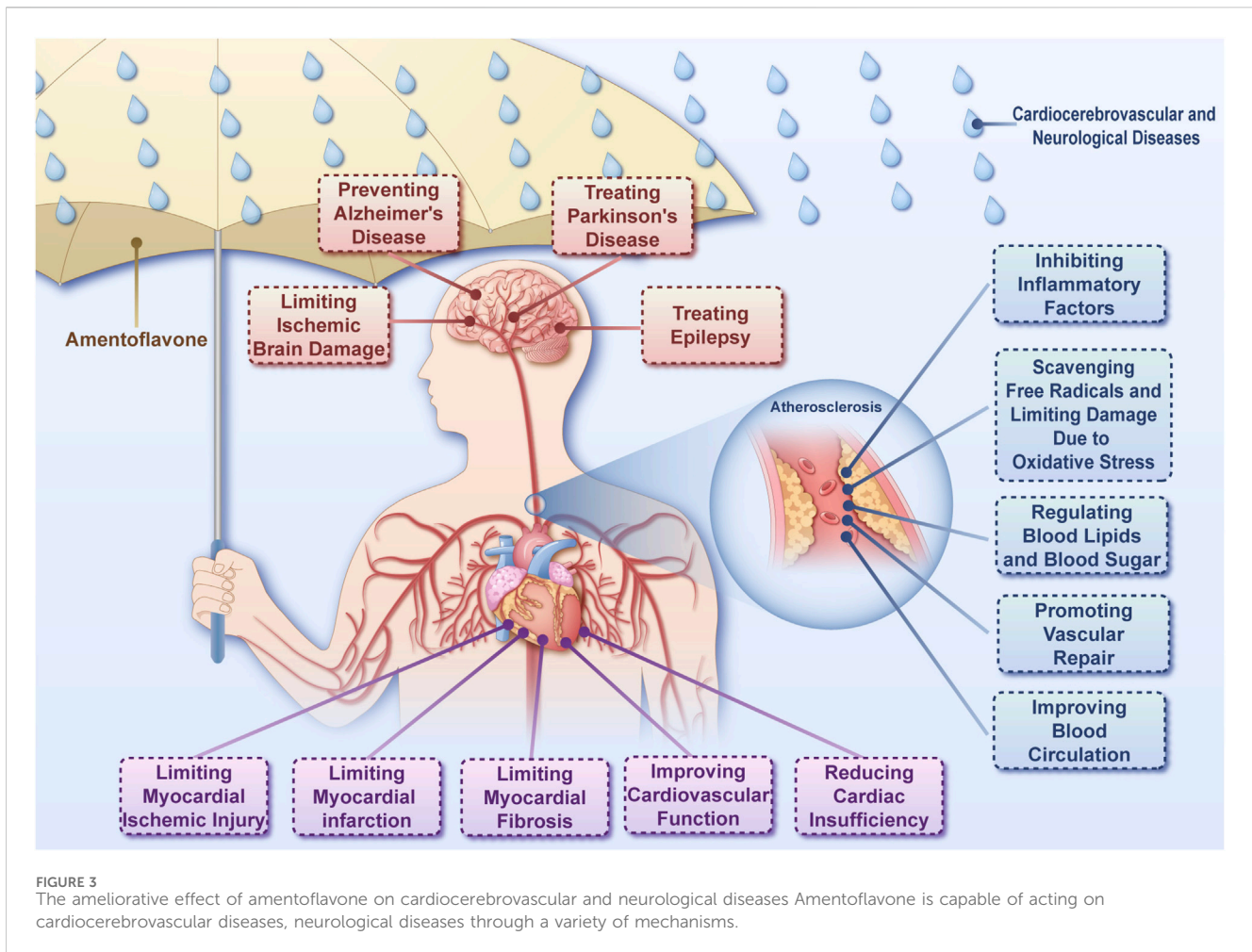
3.1.1 Inhibiting inflammatory factors

When inflammation occurs *in vivo*, IκB kinase phosphorylates inhibitor of NF-κB (IκB) protein, and then IκB protein is separated from the p50 and p65 subunits of nuclear factor-kappa B (NF-κB), so that NF-κB is activated and enters the nucleus for gene transcription and expression of inflammatory factors, inflammatory mediators, chemokines, and adhesion factors (Guo et al., 2024). AME can protect vascular ECs through various mechanisms. AME increases the survival of human umbilical vein endothelial cells (HUVECs) induced by TNF-α in the S phase of cell proliferation. Disruption of the endothelin-1 (ET-1)/nitric oxide (NO) balance in the blood is an indicator of vascular endothelial damage. AME can increase the NO content of HUVECs, reduce the level of ET-1, inhibit the expression of adhesion factors vascular cell adhesion molecule-1 and E-selectin, and inflammatory factors interleukin IL-6 and IL-8, enhance the expression of the inhibitory protein IκBα activated by NF-κB, and reduce the expression of its transcription factor NF-κB in the nucleus, preventing further damage to the vascular endothelium (Zheng et al., 2013). AME downregulates the release of NO from mouse macrophages stimulated by lipopolysaccharide (LPS), and the levels of TNF-α and IL-1β induced by LPS and monosodium urate in THP-1 macrophages, via the NOD-like receptor thermal protein domain associated protein 3 (NLRP3)/apoptosis-associated speck-like protein/cysteiny aspartate specific protease (caspase)-1 signaling pathway (Zhang X. et al., 2021). An experiment was conducted to examine the binding effects of 34 flavonoid products on the NLRP3 inflammasome, using CB-Dock molecular docking for binding predictions. It was found that AME has the strongest

TABLE 4 Pharmacokinetic parameters of amentoflavone.

| Route of administration | Dose | C _{max} | T _{max} | T _{1/2} | AUC _{0-t} | AUC _{0-∞} | CL/F | V/F | MRT _{0-t} | MRT _{0-∞} | F (%) | Reference |
|-------------------------|------------|----------------------|------------------|-------------------|----------------------------|----------------------------|--|-------------------|--------------------|--------------------|-------------|--------------------|
| Oral | 2.8 g/kg | 22.5 ± 1.4 ng/mL | 1.13 ± 0.44 h | 2.06 ± 0.13 h | 125 ± 7 ng/h mL | 133 ± 8 ng/mL h | N/A | N/A | N/A | N/A | N/A | Wang et al. (2014) |
| Intragastric | 60 mg/kg | 0.469 ± 0.046 min/L | 90.0 min | 57.01 ± 2.765 min | 27.7 ± 1.1 mg/L min | 24.9 ± 1.1 mg/L min | 2.4 ± 0.1 L min ⁻¹ kg ⁻¹ | 198.4 ± 17.4 L/kg | 122.8 ± 1.6 min | 126.1 ± 2.3 min | N/A | Wang et al. (2015) |
| Oral | 500 mg/kg | 42.37 ± 11.95 ng/mL | 0.85 ± 0.137 h | 12.33 ± 4.65 h | 194.5 ± 16.9 ng/mL h | 299.2 ± 75.4 ng/mL h | N/A | N/A | 9.58 ± 0.84 h | N/A | 0.06 ± 0.04 | Gan et al. (2020) |
| Intravenous | 10 mg/kg | 17,505 ± 1,532 ng/mL | 0.033 h | 9.36 ± 2.97 h | 10,060.9 ± 1,163.8 ng/mL h | 10,706.6 ± 1,225.9 ng/mL h | N/A | N/A | 3.365 ± 0.34 h | N/A | N/A | Gan et al. (2020) |
| Intravenous | 10 mg/kg | 31.2 ± 27.0 nmol/mL | 0.11 ± 0.02 h | 5.88 ± 1.78 h | 33.0 ± 11.9 nmol/mL h | 35.2 ± 13.9 nmol/mL h | 320 ± 139 mL h ⁻¹ kg ⁻¹ | 2.53 ± 0.65 L/kg | N/A | N/A | N/A | Liao et al. (2015) |
| Intraperitoneal | 10 mg/kg | 6.26 ± 0.33 nmol/mL | 0.83 ± 0.29 h | 3.42 ± 1.45 h | 25.5 ± 1.08 nmol/mL h | 25.7 ± 0.82 nmol/mL h | N/A | N/A | N/A | N/A | 77.4 ± 28.0 | Liao et al. (2015) |
| Oral | 300 mg/kg | 0.06 ± 0.03 nmol/mL | 0.33 ± 0.14 h | 11.3 ± 3.6 h | 0.41 ± 0.08 nmol/mL h | 0.49 ± 0.12 nmol/mL h | N/A | N/A | N/A | N/A | 0.04 ± 0.01 | Liao et al. (2015) |
| Oral | 4.31 mg/kg | 124.61 ± 8.37 ng/mL | 1.5 h | 2.60 ± 1.34 h | 594.48 ± 62.12 µg h/L | 597.84 ± 60.41 µg h/L | 7.27 ± 0.75 L min ⁻¹ kg ⁻¹ | N/A | N/A | N/A | N/A | Shan et al. (2018) |

C_{max}, maximum blood concentration; T_{max}, time to peak concentration; T_{1/2}, biological half-life; AUC(0-t), area under the concentration-time curve; AUC(0-∞), from time zero to all original drug elimination; CL/F, clearance; V/F, apparent volume of distribution; MRT_{0-∞}, mean residence time; MRT_{0-t}, average retention time for a certain period of time; F, bioavailability.

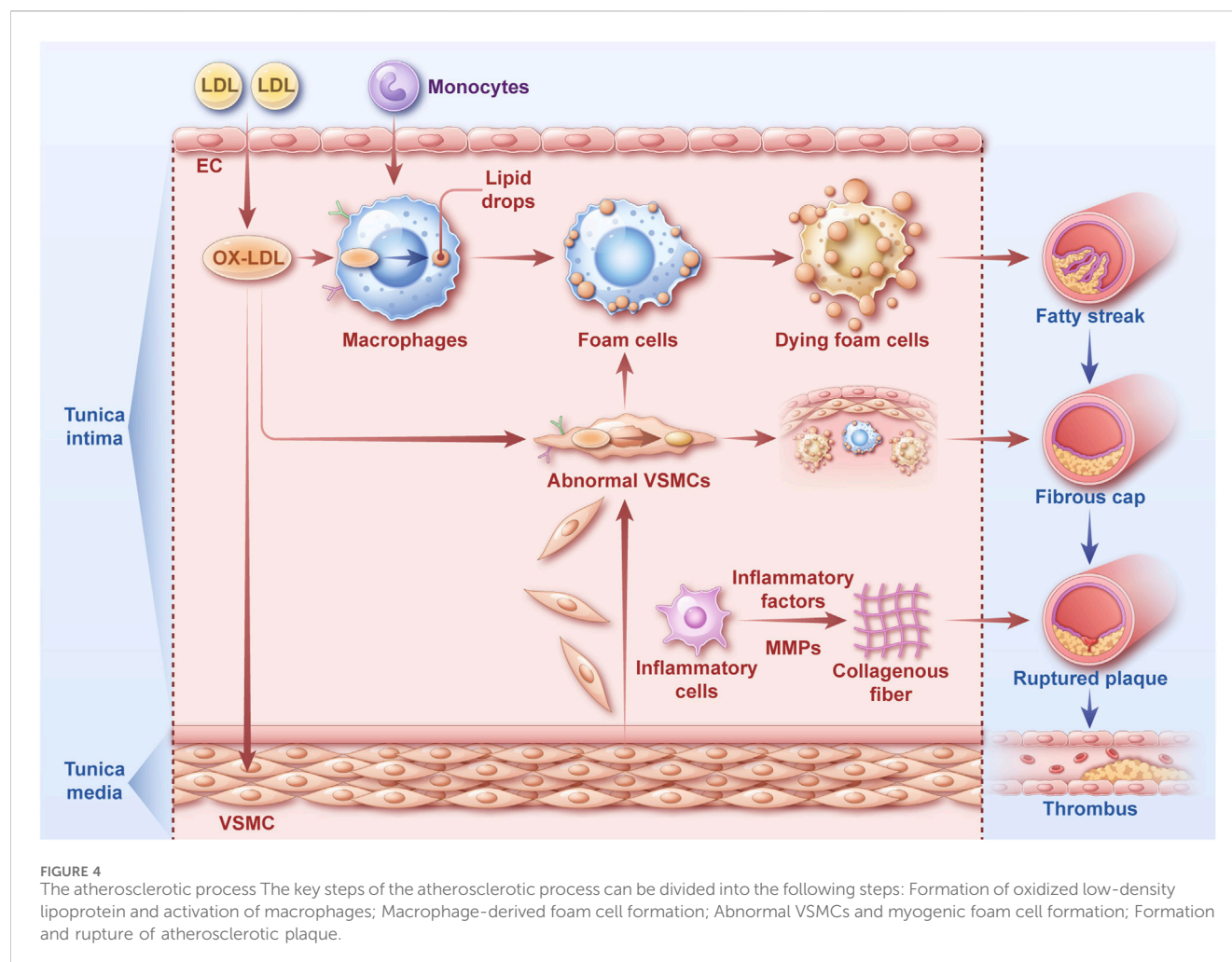


affinity for the NLRP3 inflammasome, surpassing that of its specific inhibitor (CY-09) (Fang H. Y. et al., 2023). Macrophage migration inhibitory factor (MIF) is a crucial pro-inflammatory mediator. AME has been reported to exhibit superior inhibitory activity against MIF compared to ISO-1, a well-established standard MIF inhibitor (Siddiqui et al., 2024).

During inflammation, arachidonic acid is converted to prostaglandin E-2 (PGE-2) by cyclooxygenase-2 (COX-2) through the action of cyclooxygenase-2, leading to pain and inflammation. At a concentration of 50 μ M, AME can inhibit the activity and expression of COX-2 induced by TNF- α in cells, and upregulate the activity of peroxisome proliferator-activated receptor (PPAR) γ , blocking the degradation of I κ B α , and inhibiting the translocation of NF- κ B to inhibit the activation of the NF- κ B signaling pathway (Banerjee et al., 2002). AME isolated from the root of (*Prismatomeris glabra* Mart.) exerts an anti-inflammatory effect by significantly reducing the production of TNF- α , IL-6, and PGE-2 in THP-1-derived macrophages in a dose-dependent manner (Alkadi et al., 2021). AME effectively suppresses the production of NO and PGE-2 in RAW264.7 macrophage cells stimulated by LPS. This inhibitory effect is achieved by inhibiting the kinase activity of extracellular signal-regulated kinase (ERK). Furthermore, AME also inhibits the expression of inflammation-related genes induced by

LPS, including nitric oxide synthase (iNOS), COX-2, and TNF- α (Oh et al., 2013).

Macrophages of different polarization types are involved in distinct atherosclerosis development processes. Specifically, M1-type macrophages are primarily present in early plaques, associated with plaque formation. M2-type macrophages can exert anti-inflammatory effects, promoting the repair of atherosclerosis inflammation. PPARs are a class of ligand-activated transcription factors that play a crucial role in macrophage polarization, regulating macrophage metabolism, suppressing pro-inflammatory genes, and promoting the transformation of M2 macrophage phenotype (Zhuang, 2020). PPAR γ , through its interaction with other transcription factors and the promoter regions of arginase 1 (*Arg1*), found in inflammatory zone (*Fizz1*), and chitinase-like protein 3 (*Ym1*) genes, promotes the expression of *Arg1* and *Fizz1* genes and regulates the M2 polarization level of macrophages. Macrophages lacking PPAR γ exhibit significantly downregulated expression of *Arg1*. AME can inhibit the differentiation of THP-1-derived M0 cells toward M1 cells by activating PPAR- α/γ transcription factors, elevating the mRNA levels of TGF- β and IL-10, reducing TNF- α and IL-6 expression, and upregulating *Arg1* and *Fizz1* protein expression (Qiu F. et al., 2021). These findings suggest that AME



might also reverse the transformation of M1 macrophages toward the M2 type, thereby exerting an anti-inflammatory effect.

3.1.2 Scavenging free radicals and limiting damage due to oxidative stress

NO produced iNOS in activated macrophages is one of the most important inflammatory mediators. iNOS-mediated NO production and the associated production of highly reactive free radicals such as peroxynitrite has a harmful effect (Wang Y. et al., 2020). Therefore, inhibiting NO may be a useful target for addressing oxidative stress.

In vitro antioxidant models have demonstrated that AME exhibits exceptional scavenging and antioxidant capabilities when eliminating (1,1-diphenyl-2 trinitrophenylhydrazine) DPPH free radicals, superoxide anions, and hydroxyl radicals. Additionally, AME possesses the ability to repair and protect DNA from oxidative damage *in vitro*. AME influences the generation and scavenging of OH free radicals, and eliminates free radicals by scavenging hydrogen ion and electron. Therefore AME can treat oxidative damage (Wang, 2013). AME can eliminate DPPH free radicals in a concentration-dependent manner and can alleviate the cell damage caused by ox-LDL in HUVECs. The mechanism might be due to the phenolic hydroxyl group in the molecular structure of AME, which can accept the electron transfer from lipid peroxidation free radicals, forming stable free radicals, thus preventing the

damage of lipid peroxidation free radicals to vascular ECs (Xu et al., 2004). Advanced glycation end-products (AGEs) are associated with various diseases such as atherosclerosis and diabetes, leading to the production of reactive oxygen species (ROS), which subsequently activates the transcription factor NF- κ B and is involved in various inflammatory diseases. Ferchichi et al. extracted various active components from plants, and showed that AME had the strongest *in vitro* ability to effectively resist AGEs and exhibits excellent scavenging capabilities (Ferchichi et al., 2012). Furthermore, AME can block the nuclear translocation of NF- κ B p65, inhibit the phosphorylation of I κ B α and formation of NO in macrophages induced by LPS in a concentration-dependent manner by blocking the degradation of I κ B α to inhibit the formation and transcription activation of the iNOS gene induced by LPS (Woo et al., 2005).

AME reduces LPS-induced oxidative stress damage to HUVECs, increasing the activity of superoxide dismutase (SOD) and reducing the expression levels of NO and malondialdehyde (MDA). A multi-omics study found that glycine, argininosuccinic acid, putrescine, ornithine, spermidine, 5-oxoproline, and dihydrouracil are seven metabolites that might be related to the mechanism by which AME protects ECs (Yao et al., 2016).

The reduced form of thioredoxin (Trx) interacts with the N-terminus of apoptosis signal regulating kinase 1 (ASK1) both

in vitro and *in vivo*, thereby inhibiting ASK1 activity. Under conditions of oxidative stress, the thiol group of the cysteine residue of Trx is oxidized to form intramolecular disulfide bonds, thereby activating ASK1 kinase activity. Therefore, in the complex of Trx with ASK1 protein, Trx1 and thioredoxin reductase (TrxR)-1 proteins are the key molecules regulating ROS-induced ASK1 activation. AME can regulate the ROS/ASK1/p38 mitogen-activated protein kinase (MAPK) pathway by inactivating ASK1 molecules, blocking p38 MAPK signaling, increasing the levels of thioredoxin Trx1 and reductase TrxR-1, and reducing ASK1 and p38 MAPK levels, thereby reducing oxidative stress damage to cells (Li et al., 2020). These studies demonstrate that AME can respond to oxidative stress through various mechanisms.

3.1.3 Regulating blood lipids and blood sugar

The fat accumulation index can serve as a predictive indicator of cardiometabolic diseases and stroke. The fat accumulation index is linearly related to the risk of atherosclerotic cardiovascular disease within 10 years and is an independent risk factor, indicating an integral relationship between body fat and atherosclerotic diseases (Liu et al., 2023). A randomized trial involving over 20 million participants demonstrated that blood lipid components such as LDL, apolipoprotein A, apolipoprotein B, and triacylglycerols can also affect the development of atherosclerosis (Ference et al., 2017). Therefore, controlling the level of blood lipids can alleviate and prevent atherosclerosis.

Obesity leads to the storage of excess energy in the form of triglycerides (TG) in adipose tissue. Dietary fat can lead to increased TG levels in the blood and obesity, whereas lipid absorption disorders can lead to hyperlipidemia and metabolic diseases. The cluster of differentiation 36 (CD36) protein primarily participates in the process of macrophage lipid uptake. When a large amount of ox-LDL appears in the vascular endothelial layer, the CD36 protein is activated to take up ox-LDL into the cell. After intake, ox-LDL is oxidized to new derivatives by linoleic acid, and these new derivatives can activate PPAR γ , thereby promoting the increase in PPAR γ protein expression as a transcription factor of CD36. Increased CD36 expression in turn promotes ox-LDL uptake. AME can reduce the uptake of ox-LDL by THP-1-derived macrophages through the CD36/PPAR γ signaling pathway, thereby inhibiting foam formation induced by ox-LDL (Zhuang, 2020). Feeding AME to mice on a high-fat diet reduced the expression of the lipid absorption-related gene, CD36, by affecting plasma TG levels, thereby affecting the intestinal absorption of lipids (Lee et al., 2022). AME can reduce the body weight, total fat tissue, and serum TG content induced by a high-fat diet in a dose-dependent manner. AME has been demonstrated to reduce blood glucose levels and insulin resistance in rat models. Furthermore, AME influences various stages of 3T3-L1 adipocyte differentiation. Specifically, it modulates ROS production and inhibits the expression of the transcription factor CCAAT/enhancer-binding protein (C/EBP)- β during the mitotic clonal expansion (MCE) phase, thereby impacting MCE. Additionally, AME downregulates the expression of PPAR γ and C/EBP- α during both the early and terminal differentiation phases, consequently affecting lipid droplet formation. (Chen et al., 2016).

Type 2 diabetes is intricately linked to atherosclerosis. Patients with diabetes often have disorders of lipid metabolism and insulin

resistance syndrome. Elevation of blood glucose and lipid levels in patients with type 2 diabetes are risk factors for atherosclerosis, and are positively correlated with the extent of atherosclerosis lesions (Sun et al., 2023; Huang and Sun, 2023). High blood glucose levels disrupt ECs function, causing damage the vascular wall, thereby promoting atherosclerosis progression (Wei and Liang, 2021). AME significantly suppresses the elevation of blood glucose levels and reduces the Homeostatic Model Assessment of Insulin Resistance index. Furthermore, they demonstrated that AME also reduced lipid accumulation in the liver of high fructose and fat diet (HFFD)-fed rats and alleviated the damage caused by lipids to the liver (Qin et al., 2018). AME reduced the expression of genes related to fat production and upregulated the expression of genes related to insulin signaling transmission, thereby having anti-obesity and anti-hyperglycemic effects (Cho et al., 2021).

In vitro hypoglycemic effects of AME using HepG2 cell glucose models and insulin resistance models and found that AME significantly increased glucose metabolism of HepG2 cells and had a synergistic effect on the increased glucose consumption under insulin stimulation. In the insulin resistance HepG2 cell model induced by high insulin, AME also increased the glucose consumption of cells that developed insulin resistance (Zheng et al., 2008). These characteristics suggest that AME may serve as an insulin sensitizer, increasing cell glucose consumption and synergistically working with insulin to reduce blood glucose levels and reduces insulin resistance. In the insulin resistance HepG2 cell model induced by high glucose and high insulin, AME significantly increased the levels of rate-limiting enzymes for glucose oxidative decomposition, such as 6-phosphogluconate kinase, glucose kinase, and pyruvate kinase; reduced glucose synthesis by lowering glucose synthesis kinase-3- β (GSK-3- β) levels; and lowered phosphoenolpyruvate carboxy kinase and glucose-6-phosphate enzyme activity, thereby affecting the glucose biosynthesis pathway. They also demonstrated that AME increases phosphoinositide 3-kinase (PI3K) protein expression, which may improve insulin signal transduction disorders through the PI3K/protein kinase B (Akt) signaling pathway, thereby alleviating insulin resistance (Ke et al., 2013). In type 2 diabetes model (T2DM) rats, AME reduces the release of TNF- α , upregulates glucose transporter 2 expression, enhances the absorption and utilization of blood glucose by the liver and skeletal muscle, and reduces insulin resistance through the PI3K/Akt/mechanistic target of rapamycin (mTOR) and PPAR γ signaling pathways (Zhang et al., 2019).

At a dose of 60 mg/kg AME can repair damaged pancreatic tissues in mice with diabetes and enhanced pancreatic islet β cell function (Zheng et al., 2008). The molecular structure of AME can stably bind to the structure of human islet amyloid polypeptide (hIAPP), thereby interfering with the peptide assembly and abnormal folding process, separating hIAPP fibrils into small oligomers and particles, and reducing the cytotoxicity induced by hIAPP oligomerization (Xu et al., 2022). These results indicate that AME can regulate blood lipids and blood glucose from multiple perspectives and different mechanisms and can alleviate various injuries.

3.1.4 Promoting vascular repair

Endothelial growth factor (VEGF) is a key factor in the early formation of blood vessels and a highly selective mitogen for ECs, promoting the proliferation and migration of ECs to promote

angiogenesis (Wisznia and Schwarz, 2021). AME promotes the proliferation of HUVECs in the mitotic S phase in a dose-dependent manner, and upregulates the expression of VEGF, indicating that it plays a role in repairing vascular endothelial cell damage (Zheng et al., 2011). However, under pathological conditions, VEGF increases the instability of atherosclerotic plaques by promoting angiogenesis and inflammatory infiltration, leading to plaque shedding (Qin et al., 2021). Using two different experimental models, it was found that AME can specifically bind to members of the VEGF family, VEGF-A and placental growth factor-1 (PlGF-1), preventing them from further binding to their receptors and inhibiting endothelial cell migration and capillary-like tube production induced by VEGF-A and PlGF-1, thereby inhibiting the growth and formation of vessels (Tarallo et al., 2011). The abnormal proliferation and migration of VSMCs can lead to vascular lesions, which are the hallmark of atherosclerosis, vascular intimal hyperplasia, and arterial stenosis. AME can inhibit the ox-LDL-induced transformation of VSMCs to foam cells through the CD-36/PPAR γ signaling pathway, and can inhibit VSMC migration, thereby promoting vascular repair (Zhuang, 2020).

3.1.5 Improving blood circulation

The vasodilator effect of AME may be associated with the muscarinic receptor, the β -adrenergic receptor, and the endothelium-derived vasodilator factor. NO is produced by the catalytic action of iNOS on L-arginine. The vasodilator effect of NO is mediated through the increase of guanosine 3',5'-cyclic monophosphate (cGMP) levels in smooth muscle. When the vascular endothelium experiences dysfunction, the release of NO decreases, and the vasoconstriction produced by directly activating vascular smooth muscle further reduces the production of NO (Borow et al., 2015). AME has a significant vasodilator effect; however, after injury to the vascular endothelium of injured rats, its vasodilator effect is inhibited, suggesting that AME may act on the vascular endothelium. The vasodilator effect of AME is also inhibited by NO inhibitors in intact vascular endothelium but is not affected by the addition of propranolol hydrochloride and atropine, suggesting that AME may produce a vasodilator effect by affecting the release of NO from the vascular endothelium (Xun and Yin, 2009). iNOS inhibitors can block the vasodilator effect of AME. Guanylate cyclase inhibitors also block the vasodilator effect of AME. Based on a series of experiments, they concluded that AME activates the Ca²⁺-dependent K⁺ channel of ECs, affecting the NO-cGMP signaling pathway, thereby relaxing the vascular smooth muscle and causing vasodilation (Kang et al., 2004).

After the rupture of atherosclerotic plaques, the activation of platelets and thrombin promotes thrombus formation, leading to vascular blockage and interruption of circulation (Ahmed et al., 2020). Thrombin is a serine protease that plays a significant role in the coagulation cascade, thrombus formation, and platelet activation (Brummel et al., 2002). Therefore, drugs that act on thrombin can alleviate disease progression caused by atherosclerosis. AME in (*St. John's Wort* Diosc.) can inhibit the activity of human thrombin in a dose-dependent manner (Wei et al., 2019). The study compared the effects of 16 major components in *Ginkgo biloba* on thrombin and found that AME has a high affinity for human thrombin and is a strong human thrombin inhibitor. Subsequent molecular docking

experiments revealed that it can primarily bind to key amino acids in the active site of thrombin through salt bridges and hydrogen bonds (Chen et al., 2019). Using an acute rat blood stasis model, it was demonstrated that AME can reduce plasma fibrinogen levels to prolong coagulation time, thereby improving blood circulation (Xi et al., 2020). Furthermore, AME has been shown to inhibit platelet aggregation induced by adenosine diphosphate and arachidonic acid, but has no effect on platelet aggregation induced by thrombin (Zhang et al., 2018). The mechanism underlying these effects warrant further study. These results suggest that AME could serve as a natural drug for targeting thrombin and platelets.

3.2 Effect of AME on cardiocerebrovascular diseases

3.2.1 Limiting myocardial ischemic injury, myocardial infarction and myocardial fibrosis

Myocardial ischemic injury can lead to myocardial infarction, and myocardial fibrosis may occur after infarction, a process that further affects heart function. If left untreated, myocardial ischemia can lead to damage of the heart, ultimately increasing the risk of developing myocardial infarction over time. Patients who have had a myocardial ischemia often have persistently elevated TNF- α levels. (Ridker et al., 2000). Overexpression of TNF- α , IL-1 β , and IL-6 can amplify the harmful effects of inflammation by inducing cell apoptosis. Therefore, inhibiting these inflammatory factors is a promising strategy to prevent the escalation of myocardial ischemic injury. (Kumari et al., 2024).

Ischemic reperfusion injury experiments in mice, have shown that AME significantly reduces the levels of serum myocardial enzymes, specifically lactate dehydrogenase and creatine kinase MS isoenzyme, indicating the protective effects of AME on H9c2 myocardial cells. AME was also shown to significantly reduce the levels of IL-1 β , IL-6, and TNF- α in cell supernatants, inhibiting cell apoptosis after myocardial ischemia-reperfusion injury in rats and reducing the size of the myocardial infarction area (Li et al., 2021). It was discovered that AME improved myocardial ischemia-reperfusion injury by modulating the PI3K/Akt-NF- κ B signaling pathway, enhancing the phosphorylation of Akt and inhibiting the phosphorylation of NF- κ B, reducing cardiac cell apoptosis, and lowering the release of associated inflammatory factors (Li, 2020). In the myocardial infarction model, AME can reduce the values of left ventricular end-systolic diameter and left ventricular end-diastolic diameter, and increase the value of left ventricular ejection fraction, indicating that AME can effectively improve the cardiac function of rats after myocardial infarction. AME can also reduce the expression of carboxy terminal peptide of type I procollagen and the amino terminal peptide of type III procollagen in rats after myocardial infarction, indicating that AME can inhibit myocardial fibrosis after myocardial infarction. The mechanism may involve downregulation of matrix metalloproteinase MMP-2 and transforming growth factor TGF- β 1 expression (Chen et al., 2023).

3.2.2 Reducing cardiac insufficiency

AME can improve adriamycin (DOX)-induced cardiac dysfunction and reduce myocardial injury. Furthermore, AME

can significantly reduce the expression of cell pyroptosis-related proteins, such as NLRP3, cleaved caspase-1, and cleaved gasdermin D, thereby inhibiting myocardial cell pyroptosis without affecting the expression of apoptosis-related proteins. The primary mechanism of AME action is through the inhibition of the stimulator of interferon genes (STING)/NLRP3 inflammasome signaling pathway (Fang G. et al., 2023). The effect of AME on DOX-induced cardiotoxicity was investigated from several perspectives, including pathological characterization, antioxidant stress, mitochondrial function recovery, anti-inflammatory effects, and apoptosis inhibition. It was found that AME increases heart weight to reverse DOX-induced heart atrophy, ameliorates oxidative stress-induced cardiac injury by reducing MDA, inhibits NADPH oxidase (NOX) expression, increases SOD levels, upregulates the expression of myocardial mitochondrial-related genes nuclear respiratory factor-1 (*NRF-1*) and mitochondrial transcription factor A (*TFAM*) to address mitochondrial dysfunction, decreases IL-6 and NF- κ B expression to exert anti-inflammatory effects, upregulates heat shock protein 27 (HSP-27) and downregulates fas ligand (FasL) expression to influence the process of cell apoptosis (Alherz et al., 2022). Through comparing effects of different dietary supplements on rat atria, it was discovered that AME and quercetin might be the pharmacologically active components of GBE in terms of its positive inotropic and chronotropic effects. Moreover, 10–50 μ g/mL of AME can significantly increase the heart rate of rat atria without altering myocardial contractility (Kubota et al., 2002). However, other studies have shown that AME can inhibit the activity of cAMP diesterase, thereby enhancing myocardial contractility and dilating peripheral vessels, further promoting blood flow in the body and reducing pressure on the heart and vessels (Saponara and Bosio, 1998). This suggests that AME is a potential drug for the treatment of heart failure.

3.2.3 Improving cardiovascular function

It was discovered that AME exerts a protective effect on cardiovascular dysfunction through several mechanisms: (1) Ultrasonic electrocardiographic evaluations have shown that AME can inhibit the increase in left ventricular internal diameter and the thickness of the posterior wall during diastole, reduce left ventricular mass, alter the ejection fraction and relative wall thickness, and inhibit the increase in cardiac stiffness and left ventricular wet weight induced by an HFFD. (2) In settings of oxidative stress, AME can modify the levels of oxidative stress markers such as thiobarbituric acid reactive substances, glutathione (GSH), SOD, catalase (CAT) in plasma, and can inhibit the increase in NOX in the heart, thereby reducing the degree of oxidative stress-induced cardiac injury. (3) In rats fed a HFFD, AME inhibits the increased expression of angiotensin (Ang) II receptors, angiotensin-1A receptor and the decreased expression of angiotensin type 2 receptors in the renin-angiotensin system and can significantly inhibit the increase in blood pressure. (4) AME inhibits phenylephrine-induced aortic vasoconstriction and increased acetylcholine-induced vascular relaxation. The mechanism of action of AME may involve the regulation of NOX, thereby modulating angiotensin II cell signaling and oxidative stress (Qin et al., 2018).

3.2.4 Limiting ischemic brain damage

Ischemic stroke is one of the most common brain diseases, accounting for 85% of cerebrovascular diseases (Oliveira et al., 2023). Research has shown that AME can protect the brain from hypoxic-ischemic injury from multiple perspectives. First, administration of AME to rats with hypoxic-ischemic brain injury reduced brain tissue damage in the forebrain by 50%. Second, *in vitro* experiments showed that AME exhibits excellent neuroprotective effects against DNA damage, mitochondrial damage, and NO-induced injury. *In vivo* experiments demonstrated that AME inhibits caspase 3-induced cell pyroptosis in a dose-dependent manner, decreases the expression of iNOS and COX-2, and suppresses the production of inflammatory factors such as IL-1 β and TNF- α stimulated by LPS, thereby reducing the damage caused to microglia by these inflammatory factors (Shin et al., 2006). In a study, a model of left common carotid artery occlusion stroke to investigate the protective effects of AME on cerebral ischemia/reperfusion injury in rats. The findings indicated that after ischemia/reperfusion injury, AME significantly reduced the neurological deficit scores, improved motor coordination ability and spontaneous activity, reduced the levels of TNF- α , IL-1 β , and IL-6, inhibited the NF- κ B signaling pathway, reduced caspase-3 to block cell apoptosis, increased the levels of TNF receptor-associated factor family member-associated NF- κ B activator-binding kinase 1 and interferon β , reduced MDA levels, and increased GSH and CAT levels in the brain. The mechanism of AME may be mediated by high mobility group box protein B1 (HMGB1) through the toll-like receptor-4 (TLR4)/NF- κ B signaling pathway (Saeedan et al., 2023). These experimental phenomena and mechanisms indicate that AME has a good protective effect on ischemic brain injury.

3.3 Effect of AME on neurological diseases

3.3.1 Preventing Alzheimer's disease

Alzheimer's disease (AD) is a neurodegenerative disorder characterized by the accumulation of β -amyloid (A β) peptides, neurofibrillary tangles formed by hyperphosphorylated tau proteins, abnormal oxidative stress damage, inflammatory responses, and neurotransmitter disorders in affected brain regions (Breijyeh and Karaman, 2020). It is commonly treated with acetylcholinesterase (AChE) inhibitors and N-methyl-D-aspartate receptor antagonists. Although these medications can alleviate clinical manifestations, they do not reverse cognitive impairment, and are often accompanied by common side effects, including gastrointestinal symptoms, confusion, dizziness, and headaches. (Chin et al., 2022).

Overproduction of A β leads to the formation of neurofibrillary plaques in the brain, which subsequently accumulate in the blood vessels to cause cerebral amyloid angiopathy (Han et al., 2022). Therefore, inhibiting the production of A β and promoting its clearance are crucial for improving AD. In a study, researchers used the fluorescent dye thiazine 1,3,5-tetracarboxylic acid to investigate the inhibitory effect of various flavonoid compounds on A β aggregation and the structure-activity relationship of promoting A β fibril disaggregation. The results indicated that AME could inhibit the formation of A β ₁₋₄₂ fibrils, had a better

affinity for A β ₁₋₄₂ fibrils than for A β ₁₋₄₀ fibrils. The hydrophilic group of AME was the key group for its function. Furthermore, among flavonoid compounds with different structures, AME with four hydroxyl groups could most effectively inhibit A β fibrillation and promote the disaggregation of pre-formed A β fibrils (Choi et al., 2020). Another study showed that increasing the number of methoxy groups on the AME parent structure weakened the inhibitory activity of AME on A β ₄₀, and the connection of single bonds also affected the inhibitory activity (Sirimangalakitti et al., 2019). A study elucidated how AME inhibits the aggregation of A β ₄₂ peptide at the atomic level. AME bound to aromatic residues in the N-terminal of A β ₄₂ peptide, forming a stable π - π interaction, which destabilized the fibril structure. Subsequently, AME utilized its hydrogen bond donor/acceptor specificity to disrupt the hydrogen bond potential of the fibril peptide backbone, thereby disrupting the fibril structure and promoting A β fibril disaggregation (Windsor et al., 2021).

A study explained the anti-AD function of AME from the perspective of receptor-mediated endocytosis for A β clearance. AME can significantly increase A β uptake by mouse N2a neural cells through class A scavenger receptors, and then enter the lysosome within the cell, where it is degraded by relevant enzymes without causing cytotoxicity. The hydroxyl group in the molecular structure of AME enhances A β uptake, whereas the substitution of methyl groups decreases its uptake effect (Han et al., 2022). Another study identified AME from a library of polyphenolic compounds that can increase the activity and expression of neuropeptidase, an A β degradation enzyme, thereby delaying the progression of AD. Among them, the double-bond chemical structure in the C ring of the AME structure can significantly enhance neuropeptidase activity (Hori et al., 2023).

Research indicated that among various flavonoid compounds, AME exhibited the strongest neuroprotective effect, significantly inhibiting ROS-induced oxidative stress damage in SHSY5Y neuroblastoma cells. AME reduced DNA damage induced by etoposide DNA damage, whereas other flavonoid compounds increased cell death caused by DNA damage. Moreover, at a concentration of 2 μ M, AME was most effective at reducing cytotoxicity induced by the A β ₂₅₋₃₅ peptide in rat PC12 cells, indicating the therapeutic potential of AME in neurodegenerative diseases (Kang et al., 2005). Further studies found that AME could differentially protect SH-SY5Y neural cells from various cytotoxic factors such as hydrogen peroxide, okadaic acid, and A β ₂₅₋₃₅. Specifically, AME was able to inhibit okadaic acid-induced tau protein hyperphosphorylation, restore mitochondrial membrane potential to inhibit cell apoptosis, and protect microtubule structure and mitochondrial function. Furthermore, AME exhibited strong antioxidant capabilities against mitochondrial dysfunction and ROS-induced oxidative stress damage. AME also bound to β -secretase to inhibit its activity, thereby preventing the degradation of A β protein precursor to produce large amounts of A β (Zhang, 2014). In models of AD, AME could activate nuclear factor erythroid 2-related factor-2 (Nrf2) through affecting the adenosine monophosphate-activated protein kinase (AMPK)/GSK-3 β signaling pathway to exert antioxidant stress effects, ameliorating A β -induced neural function deficits and neuronal apoptosis (Chen et al., 2018). Although studies have shown that GSK-3 β is involved in the regulation of Nrf2 by AME, further verification is required to

determine the mechanism by which AME affects GSK-3 β signaling-mediated tau phosphorylation. It was demonstrated through multiple experiments that AME, as a bifunctional chelator, can bind to various A β aggregates with high affinity and reduce their induced cytotoxicity. Additionally, AME can effectively chelate with Cu²⁺ and exhibit strong antioxidant activity, thereby preventing the formation of soluble A β oligomers and ROS-induced by metal ions in the brain (Sun et al., 2020).

Autophagy can degrade and recycle proteins produced by misfolding and damaged organelles, thereby maintaining protein homeostasis. The activation of autophagy in cells can effectively eliminate the accumulation of A β , thereby reducing the progression of AD (Zhang Z. et al., 2021). AME can improve the memory and cognitive impairment induced by A β ₂₅₋₃₅ in the brains of mice and alleviate inflammation, oxidative stress, and the immune response in the hippocampus. Additionally, it can enhance autophagy by binding to multiple amino acid residues of the mTOR protein, thereby inhibiting further phosphorylation of mTOR and exerting a neuroprotective effect against AD (Cao et al., 2021). Experiments were conducted on neuronal cell damage induced by A β ₁₋₄₂. It was found that AME can improve neuronal dysfunction induced by A β ₁₋₄₂ in rats and inhibit NLRP3 inflammasome-induced pyroptosis by regulating AMPK/GSK-3 β signaling, thereby ameliorating the neurotoxicity caused by A β ₁₋₄₂ in AD (Zhao et al., 2019).

Using the scopolamine-induced dementia mouse model, it was discovered that oral administration of AME can inhibit the activity of AChE and increase acetylcholine levels. Furthermore, it can improve the oxidative stress damage induced by scopolamine in mice, mainly by reducing MDA levels and increasing GSH activity, thereby ameliorating cognitive impairment (Ishola et al., 2013a). Additionally, a study collected 50 flavonoid compounds with therapeutic potential for AD. Molecular docking experiments were conducted with the human α 7 nicotinic acetylcholine receptor (α 7nAChR) and found that after binding with AME, it exhibited the best affinity and good stability parameters (Singh et al., 2023). However, the effectiveness of AME as an α 7nAChR agonist for treating AD requires further verification.

3.3.2 Treating Parkinson's disease

In Parkinson's disease (PD), melanin is released as neurons degenerate. Subsequently, it is recognized and ingested by microglia, leading to inflammatory damage. In patients with PD, the breakdown of dopaminergic neurons can be attributed to chronic neuroinflammation, which leads to the production and accumulation of Lewy bodies in the compact region of the substantia nigra in the brain. The main pathological features of PD are damage to dopaminergic neurons in the midbrain substantia nigra and the establishment of diffuse Lewy bodies, which subsequently lead to motor neuron dysfunction. Therefore, taking measures to inhibit neuroinflammation may be beneficial for alleviating PD clinical manifestations (Sharma et al., 2023).

Microglia, analogous to macrophages within the brain, can be activated by various stimuli to release a range of inflammatory mediators, thereby initiating an inflammatory response that may result in neural damage (Cai et al., 2022). AME has been shown to improve motor dysfunction and anxiety-like and depressive-like behaviors in the LPS-induced PD rat model. Mechanistic studies revealed that AME effectively inhibits the expression of pro-

inflammatory factors (TNF- α , IL-1 β , IL-6, iNOS, COX-2) and enhances the expression of anti-inflammatory factors (IL-4, TGF- β , Arg-1, CD206), indicating that AME promotes the polarization of microglia towards the M2 anti-inflammatory phenotype. Furthermore, AME significantly ameliorates the reduction in tyrosine hydroxylase-positive neurons and the increase in α -synuclein expression observed in the model, suggesting that AME exerts neuroprotective effects in the LPS-induced PD model (Liu et al., 2024). It has been discovered that after LPS stimulation of rat astrocytoma cells (C6), AME significantly inhibited the production of nitrites, ROS, MDA, and TNF- α , upregulated GSH levels, and reduced intracellular oxidative stress (Ishola et al., 2013b). Activated ERK1/2 promotes cell death due to oxidative toxicity, but AME can alleviate oxidative stress damage caused by glutamate and ROS in HT22 neuronal cells by maintaining the activity of antioxidant enzymes and inhibiting ERK1/2 activity (Jeong et al., 2014). Molecular docking studies have revealed that AME exhibits strong binding affinity with the glutathione peroxidase 4 (GPX4) protein, which is involved in ferroptosis (Xiong et al., 2024). It was reported that AME can inhibit inflammation induced by ferroptosis in HT22 cells triggered by homocysteine. However, it is important to note that AME treatment results in decreased expression levels of both solute carrier family 7 member 11 (SLC7A11) and GPX4. Given that the SLC7A11/GPX4 signaling pathway requires the upregulation of these proteins to effectively suppress ferroptosis, the observed reduction in their expression following AME treatment may affect its efficacy in preventing ferroptosis (Wang et al., 2024).

In vivo and *in vitro* PD model studies demonstrated that AME can significantly reduce the expression of glial fibrillary acidic protein and Iba1 markers in glial cells under inflammatory conditions, decrease the activation of caspase-3 and p21, and reduce the expression levels of IL-1 β and iNOS and increase the Bcl-2/Bax ratio, through the PI3K/Akt and ERK signaling pathways. In this study, in a *vitro* mouse model induced by methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP), AME was able to protect dopaminergic neurons and reduce striatal fiber loss (Cao et al., 2017). These findings suggest that AME can exert beneficial effects on dopaminergic neurons and glial cells, thereby ameliorating the clinical manifestations of PD. Furthermore, studies have shown a deregulated angiotensin-converting enzyme (ACE)/Ang II/angiotensin II receptor-1 (AT1R) axis is activated at the onset of PD, leading to free radical damage, cell apoptosis, and neuronal disruption. AME can bind to the mitochondrial assembly receptor (MASR) protein and activate the ACE2/Ang (1–7)/MASR, thereby neutralizing the neurodegenerative changes triggered by the ACE/Ang II/AT1R axis (Bhadauriya et al., 2023).

3.3.3 Treating epilepsy

The development of inflammation is closely related to the onset of epilepsy and its clinical progression. Brain inflammation can promote neuronal excitability, reducing the seizure threshold, thereby triggering seizures. Therefore, anti-inflammatory therapy can be used to prevent and treat seizures (Alsaegh et al., 2021). Studies found that AME improve pentetrazole-induced cognitive dysfunction by inhibiting the NLRP3 inflammasome, reducing the susceptibility to seizures and apoptosis in hippocampal neurons of mice. In this study, AME can also inhibit the mRNA expression of NLRP3, ASC, and caspase-1 in BV2 microglia induced by LPS, reducing the expression of

inflammatory factors IL-1 β , IL-18, and TNF- α . However, in the absence of inflammation, administration of AME to mice did not affect the expression of these proteins and inflammatory factors (Rong et al., 2019). Research indicated that AME can inhibit the activation and nuclear translocation of NF- κ B p65 subunit in mice, thereby inhibiting the NF- κ B signaling pathway to reduce the injury of hippocampal CA1 neural cells. AME can also reduce the production of inflammatory mediators NO and PGE-2, and inflammatory factors IL-6 and IL- β in neural cells. Furthermore, histological analysis found that AME can protect neurons after status epilepticus, inhibit the excessive discharge of hippocampal neurons, and shorten the duration of seizures (Zhang et al., 2015). These results indicate that AME has a good protective effect on hippocampal neuronal injury caused by epilepsy and has good anti-inflammatory effects. A study employed various computational methods and found that AME exhibits superior affinity for α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) and voltage-gated sodium ion channels (VGSC) receptors compared to phenytoin. Additionally, physicochemical and pharmacokinetic studies indicate that AME is suitable for oral administration, demonstrating favorable intestinal permeability and the ability to cross the blood-brain barrier, with no significant risk of toxicity (Salaria et al., 2024). These findings suggest that AME is a promising candidate for further research as a potential treatment for epilepsy.

4 Conclusion and future perspective

AME has been reported in multiple studies as the primary active pharmacological component from its plant sources, with a wide range of pharmacological effects (Yu et al., 2017; Li et al., 2021; Zhang et al., 2015). Due to the unique chemical structure of AME, its biological activity can be significantly influenced by multiple factors. Further studies could elucidate how various factors, such as photooxidation, affect the structural stability of AME. Additionally, pharmacokinetic studies have demonstrated that AME undergoes rapid metabolism in the body, resulting in low bioavailability. Therefore, modifying the natural structure or developing suitable drug formulations of AME is crucial for improving its therapeutic properties and enhancing its bioavailability.

The anti-inflammatory, antioxidant, and lipid-lowering effects of AME are notable, and recent studies have demonstrated its application in various cardiac and neurological diseases, suggesting that it may have significant anti-atherosclerotic effects. However, most of the current data on AME are derived from *in vitro* cell experiments, with limited *in vivo* animal testing. More comprehensive animal experiments are required to validate its pharmacological activity. One study has found that AME may have hepatotoxic and nephrotoxic effects (Li D et al., 2019). This warrants further investigation.

AME exhibits significant therapeutic effects on various cardiocerebrovascular diseases and neurological disorders, potentially functioning through multiple mechanisms. All these disease processes are accompanied by inflammatory responses, which are triggered by multiple factors. AME can affect multiple inflammatory mediators and mechanisms, suggesting that it has considerable potential for treating these diseases. However, the specific mechanisms of action involved in the effects of AME on inflammatory diseases require further research. Hemodynamic factors such as hypertension cannot be ignored in the

development of cardiocerebrovascular; however, the mechanisms and targets by which AME counters the effects of these factors remain unclear. Currently, there remains controversy regarding whether AME can exert a therapeutic effect on AD. Furthermore, the specific mechanisms by which AME influences the cellular uptake and clearance of A β also require further validation research.

In summary, AME exhibits multiple activities, indicating its potential as a natural drug for treating cardiocerebrovascular diseases and neurological disorders. Further studies on its pharmacokinetics and toxicology are required to ensure its safety and efficacy.

Author contributions

HZ: Writing–original draft. Y-MB: Writing–original draft. D-ML: Writing–original draft. GW: Writing–review and editing. JG: Funding acquisition, Writing–review and editing. LZ: Funding acquisition, Writing–review and editing.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

- Ahmed, M. U., Kaneva, V., Loyau, S., Nechipurenko, D., Receveur, N., Le Bris, M., et al. (2020). Pharmacological blockade of glycoprotein VI promotes thrombus disaggregation in the absence of thrombin. *Arterioscler. Thromb. Vasc. Biol.* 40 (9), 2127–2142. doi:10.1161/ATVBAHA.120.314301
- Alherz, F. A., El-Masry, T. A., Negm, W. A., and El-Kadem, A. H. (2022). Potential cardioprotective effects of Amentoflavone in doxorubicin-induced cardiotoxicity in mice. *Biomed. Pharmacother.* 154, 113643. doi:10.1016/j.biopha.2022.113643
- Alkadi, K. A. A., Ashraf, K., Adam, A., Shah, S. A. A., Taha, M., Hasan, M. H., et al. (2021). *In vitro* cytotoxicity and anti-inflammatory cytokine activity study of three isolated novel compounds of *Prismatomeris glabra*. *J. Pharm. Bioallied Sci.* 13 (1), 116–122. doi:10.4103/jpbs.JPBS_279_19
- Alsaegh, H. Z., Eweis, H. S. A. E-K., and Kamel, F. O. (2021). Pathophysiological cascade of events leading to epilepsy: role of inflammation. *J. Pharm. Res. Int.* 32 (48), 74–84. doi:10.9734/jpri/2020/v32i4831127
- Bajpai, V. K., Park, I., Lee, J., Shukla, S., Nile, S. H., Chun, H. S., et al. (2019). Antioxidant and antimicrobial efficacy of a biflavonoid, amentoflavone from *Nandina domestica* in vitro and in minced chicken meat and apple juice food models. *Food Chem.* 271, 239–247. doi:10.1016/j.foodchem.2018.07.159
- Banerjee, T., Valacchi, G., Ziboh, V. A., and van der Vliet, A. (2002). Inhibition of TNF α -induced cyclooxygenase-2 expression by amentoflavone through suppression of NF- κ B activation in A549 cells. *Mol. Cell Biochem.* 238 (1–2), 105–110. doi:10.1023/a:1019963222510
- Bhadauriya, P., Varshney, V., and Goyal, A. (2023). Molecular docking-based identification of potential natural neuroprotective molecules for Parkinson's disease. *Chem. Biodivers.* 20 (10), e202300979. doi:10.1002/cbdv.202300979
- Bhattacharya, P., and Mandal, A. (2024). Identification of amentoflavone as a potent SARS-CoV-2 M^{pro} inhibitor: a combination of computational studies and *in vitro* biological evaluation. *J. Biomol. Struct. Dyn.* 1, 1–19. doi:10.1080/07391102.2024.2304676
- Borow, K. M., Nelson, J. R., and Mason, R. P. (2015). Biologic plausibility, cellular effects, and molecular mechanisms of eicosapentaenoic acid (EPA) in atherosclerosis. *Atherosclerosis* 242 (1), 357–366. doi:10.1016/j.atherosclerosis.2015.07.035
- Breijyeh, Z., and Karaman, R. (2020). Comprehensive review on Alzheimer's disease: causes and treatment. *Molecules* 25 (24), 5789. doi:10.3390/molecules25245789
- Brummel, K. E., Paradis, S. G., Butenas, S., and Mann, K. G. (2002). Thrombin functions during tissue factor-induced blood coagulation. *Blood* 100 (1), 148–152. doi:10.1182/blood.v100.1.148
- Cai, Y., Liu, J., Wang, B., Sun, M., and Yang, H. (2022). Microglia in the neuroinflammatory pathogenesis of Alzheimer's disease and related therapeutic targets. *Front. Immunol.* 13, 856376. doi:10.3389/fimmu.2022.856376
- Cao, B., Zeng, M., Zhang, Q., Zhang, B., Cao, Y., Wu, Y., et al. (2021). Amentoflavone ameliorates memory deficits and abnormal autophagy in $\alpha\beta_{25-35}$ -induced mice by mTOR signaling. *Neurochem. Res.* 46 (4), 921–934. doi:10.1007/s11064-020-03223-8
- Cao, Q., Qin, L., Huang, F., Wang, X., Yang, L., Shi, H., et al. (2017). Amentoflavone protects dopaminergic neurons in MPTP-induced Parkinson's disease model mice through PI3K/Akt and ERK signaling pathways. *Toxicol. Appl. Pharmacol.* 319, 80–90. doi:10.1016/j.taap.2017.01.019
- Carracedo, M., Artiaich, G., Arnardottir, H., and Bäck, M. (2019). The resolution of inflammation through omega-3 fatty acids in atherosclerosis, intimal hyperplasia, and vascular calcification. *Semin. Immunopathol.* 41 (6), 757–766. doi:10.1007/s00281-019-00767-y
- Chen, B., Xu, D., Li, Z., Jing, Y., Lin, L., Li, S., et al. (2022). Tissue distribution, excretion, and interaction with human serum albumin of total bioflavonoid extract from *Selaginella doederleinii*. *Front. Pharmacol.* 13, 849110. doi:10.3389/fphar.2022.849110
- Chen, C., Li, B., Cheng, G., Yang, X., Zhao, N., and Shi, R. (2018). Amentoflavone ameliorates $\alpha\beta_{1-42}$ -induced memory deficits and oxidative stress in cellular and rat model. *Neurochem. Res.* 43 (4), 857–868. doi:10.1007/s11064-018-2489-8
- Chen, G., Han, Y., He, W., and Liang, F. (2016). Amentoflavone protects against high fat-induced metabolic dysfunction: possible role of the regulation of adipogenic differentiation. *Int. J. Mol. Med.* 38 (6), 1759–1767. doi:10.3892/ijmm.2016.2772
- Chen, T. R., Wei, L. H., Guan, X. Q., Huang, C., Liu, Z. Y., Wang, F. J., et al. (2019). Biflavones from *Ginkgo biloba* as inhibitors of human thrombin. *Bioorg. Chem.* 92, 103199. doi:10.1016/j.bioorg.2019.103199
- Chen, W. M., Chen, J. M., and Jian, M. H. (2023). Effect of amentoflavone on myocardial fibrosis in rats with acute myocardial infarction. *J. Zunyi Med. Univ.* 46 (07), 646–651+656. doi:10.14169/j.cnki.zunyiixuebao.2023.0103
- Chin, E., Jaqua, E., Safaeipour, M., and Ladue, T. (2022). Conventional versus new treatment: comparing the effects of acetylcholinesterase inhibitors and N-Methyl-D-Aspartate receptor antagonist with aducanumab. *Cureus* 14 (11), e31065. doi:10.7759/cureus.31065

- Cho, S., Lee, H., Han, J., Lee, H., Kattia, R. O., Nelson, Z. V., et al. (2021). *Viburnum stellato-Tomentosum* extract suppresses obesity and hyperglycemia through regulation of lipid metabolism in high-fat diet-fed mice. *Molecules* 26 (4), 1052. doi:10.3390/molecules26041052
- Choi, E. Y., Kang, S. S., Lee, S. K., and Han, B. H. (2020). Polyphenolic biflavonoids inhibit amyloid-beta fibrillation and disaggregate preformed amyloid-beta fibrils. *Biomol. Ther. Seoul* 28 (2), 145–151. doi:10.4062/biomolther.2019.113
- Duan, S., Jia, J. F., Hong, B., Zhou, J., Zhang, Y., Ge, F., et al. (2022). Assessment of amentoflavone loaded sub-micron particle preparation using supercritical antisolvent for its antitumor activity. *Curr. Drug Deliv.* 19 (1), 41–48. doi:10.2174/1567201818666210810142750
- Dutta, S., Singhal, A. K., Suryan, V., and Chandra, N. C. (2023). Obesity: an impact with cardiovascular and cerebrovascular diseases. *Ind. J. Clin. Biochem.* 39, 168–178. doi:10.1007/s12291-023-01157-w
- Fang, G., Li, X., Yang, F., Huang, T., Qiu, C., Peng, K., et al. (2023b). Amentoflavone mitigates doxorubicin-induced cardiotoxicity by suppressing cardiomyocyte pyroptosis and inflammation through inhibition of the STING/NLRP3 signalling pathway. *Phytomedicine* 117, 154922. doi:10.1016/j.phymed.2023.154922
- Fang, H. Y., Zhao, X. N., Zhang, M., Ma, Y. Y., Huang, J. L., and Zhou, P. (2023a). Beneficial effects of flavonoids on cardiovascular diseases by influencing NLRP3 inflammasome. *Inflammopharmacology* 31 (4), 1715–1729. doi:10.1007/s10787-023-01249-2
- Fang, H. Y., Zhuang, R. X., Wang, F. G., Xi, J. J., Zang, H. Q., and Miao, L. B. (2011). Separation and purification of total flavonoids from *Selaginella moellendorffii* hieron by macroporous resin and detected by HPLC. *Chin. Arch. Tradit. Chin. Med.* 29 (12), 2796–2798. doi:10.13193/j.archtcm.2011.12.198.fanghy.052
- Feng, X. (2020). *Study on metabolism of amentoflavone and isoginkgetin in vitro and in vivo preparation and evaluation of their nanomicelles*. Shijiazhuang (Hebei): Hebei Medical University. dissertation.
- Feng, X., Chen, Y., Li, L., Zhang, Y., Zhang, L., and Zhang, Z. (2020). Preparation, evaluation and metabolites study in rats of novel amentoflavone-loaded TPGS/soluplus mixed nanomicelles. *Drug Deliv.* 27 (1), 137–150. doi:10.1080/10717544.2019.1709920
- Feng, Y., Wang, J., Zhang, S., Li, Y., Wang, B., Zhang, J., et al. (2023). Preparation of amentoflavone-loaded DSPE-PEG (2000) micelles with improved bioavailability and in vitro antitumor efficacy. *Biomed. Chromatogr.* 37 (9), e5690. doi:10.1002/bmc.5690
- Ferchichi, L., Derbré, S., Mahmood, K., Touré, K., Guilet, D., Litaudon, M., et al. (2012). Bioguided fractionation and isolation of natural inhibitors of advanced glycation end-products (AGEs) from *Calophyllum flavoramulum*. *Phytochemistry* 78, 98–106. doi:10.1016/j.phytochem.2012.02.006
- Ference, B. A., Ginsberg, H. N., Graham, I., Ray, K. K., Packard, C. J., Bruckert, E., et al. (2017). Low-density lipoproteins cause atherosclerotic cardiovascular disease. 1. Evidence from genetic, epidemiologic, and clinical studies. A consensus statement from the European Atherosclerosis Society Consensus Panel. *Eur. Heart J.* 38 (32), 2459–2472. doi:10.1093/eurheartj/ehx144
- Gan, L., Ma, J., You, G., Mai, J., Wang, Z., Yang, R., et al. (2020). Glucuronidation and its effect on the bioactivity of amentoflavone, a biflavonoid from Ginkgo biloba leaves. *J. Pharm. Pharmacol.* 72 (12), 1840–1853. doi:10.1111/jphp.13247
- Global Burden of Disease Study 2013 Collaborators (2015). Global, regional, and national incidence, prevalence, and years lived with disability for 301 acute and chronic diseases and injuries in 188 countries, 1990–2013: a systematic analysis for the Global Burden of Disease Study 2013. *Lancet* 386 (9995), 743–800. doi:10.1016/S0140-6736(15)60692-4
- Gui, Y., Zheng, H., and Cao, R. Y. (2022). Foam cells in atherosclerosis: novel insights into its origins, consequences, and molecular mechanisms. *Front. Cardiovasc Med.* 9, 845942. doi:10.3389/fcvm.2022.845942
- Guo, Q., Jin, Y., Chen, X., Ye, X., Shen, X., Lin, M., et al. (2024). NF- κ B in biology and targeted therapy: new insights and translational implications. *Signal Transduct. Target Ther.* 9 (1), 53. doi:10.1038/s41392-024-01757-9
- Han, B. H., Cofell, B., Everhart, E., Humpal, C., Kang, S. S., Lee, S. K., et al. (2022). Amentoflavone promotes cellular uptake and degradation of amyloid-beta in neuronal cells. *Int. J. Mol. Sci.* 23 (11), 5885. doi:10.3390/ijms23115885
- He, C. (2013). *The mechanism of Mfn2 Gene to promote cholesterol efflux in vascular smooth muscle cell-derived foam cells*. Wuhan (Hubei): Huazhong University of Science and Technology. dissertation.
- Hori, Y., Watanabe, K., Yassen, A. S. A., Shirotani, K., Tanaka, T., and Iwata, N. (2023). Enhancement of neprilysin activity by natural polyphenolic compounds and their derivatives in cultured neuroglioma cells. *Biol. Pharm. Bull.* 46 (3), 446–454. doi:10.1248/bpb.22-00833
- Huang, Q. Z., and Sun, J. (2023). Correlation analysis of blood glucose and lipid levels with lower extremity vascular disease in patients with type 2 diabetes mellitus. *Mod. Med. Health Res. Electron J.* 7 (18), 16–18. doi:10.3969/j.issn.2096-3718.2023.18.006
- Huvaere, K., Sinnaeve, B., Van Boclaere, J., and Skibsted, L. H. (2012). Flavonoid deactivation of excited state flavins: reaction monitoring by mass spectrometry. *J. Agric. Food Chem.* 60 (36), 9261–9272. doi:10.1021/jf301823h
- Iadecola, C. (2023). The pathobiology of vascular dementia. *Neuron* 80 (4), 844–866. doi:10.1016/j.neuron.2013.10.008
- Ishola, I. O., Chatterjee, M., Tota, S., Tadigopulla, N., Adeyemi, O. O., Palit, G., et al. (2012). Antidepressant and anxiolytic effects of amentoflavone isolated from *Cnestis ferruginea* in mice. *Pharmacol. Biochem. Behav.* 103 (2), 322–331. doi:10.1016/j.pbb.2012.08.017
- Ishola, I. O., Chaturvedi, J. P., Rai, S., Rajasekar, N., Adeyemi, O. O., Shukla, R., et al. (2013b). Evaluation of amentoflavone isolated from *Cnestis ferruginea* Vahl ex DC (Connaraceae) on production of inflammatory mediators in LPS stimulated rat astrocytoma cell line (C6) and THP-1 cells. *J. Ethnopharmacol.* 146 (2), 440–448. doi:10.1016/j.jep.2012.12.015
- Ishola, I. O., Tota, S., Adeyemi, O. O., Agbaje, E. O., Narender, T., and Shukla, R. (2013a). Protective effect of *Cnestis ferruginea* and its active constituent on scopolamine-induced memory impairment in mice: a behavioral and biochemical study. *Pharm. Biol.* 51 (7), 825–835. doi:10.3109/13880209.2013.767360
- Jeong, E. J., Hwang, L., Lee, M., Lee, K. Y., Ahn, M. J., and Sung, S. H. (2014). Neuroprotective biflavonoids of *Chamaecyparis obtusa* leaves against glutamate-induced oxidative stress in HT22 hippocampal cells. *Food Chem. Toxicol.* 64, 397–402. doi:10.1016/j.fct.2013.12.003
- Jiang, Y., Wang, S., Yu, M., Wu, D., Lei, J., Li, W., et al. (2020). Ultrasonic-assisted ionic liquid extraction of two biflavonoids from *Selaginella tamariscina*. *ACS Omega* 5 (51), 33113–33124. doi:10.1021/acsomega.0c04723
- Kang, D. G., Yin, M. H., Oh, H., Lee, D. H., and Lee, H. S. (2004). Vasorelaxation by amentoflavone isolated from *Selaginella tamariscina*. *Planta Med.* 70 (8), 718–722. doi:10.1055/s-2004-827201
- Kang, S. S., Lee, J. Y., Choi, Y. K., Song, S. S., Kim, J. S., Jeon, S. J., et al. (2005). Neuroprotective effects of naturally occurring biflavonoids. *Bioorg Med. Chem. Lett.* 15 (15), 3588–3591. doi:10.1016/j.bmcl.2005.05.078
- Ke, Y. Y., Yuan, P. P., Zhou, Y. Y., Zhang, X., Li, M., Wang, S., et al. (2013). “The intervention experimental study of total flavonoids in *Selaginella tamariscina* (Beauv.) Spring and Amentotaxus Biflavone in HepG2 cells of insulin resistance,” in 2013 China Pharmaceutical Conference and the 13th Chinese pharmacist weekly treatise, 1382–1391.
- Khafagy, E. S., Soliman, G. A., Shahba, A. A., Aldawsari, M. F., Alharthy, K. M., Abdel-Kader, M. S., et al. (2023). Brain targeting by intranasal drug delivery: effect of different formulations of the biflavone “cupressuflavone” from juniperus sabina L. On the motor activity of rats. *Molecules* 28 (3), 1354. doi:10.3390/molecules28031354
- Kubota, Y., Umegaki, K., Tanaka, N., Mizuno, H., Nakamura, K., Kunitomo, M., et al. (2002). Safety of dietary supplements: chronotropic and inotropic effects on isolated rat atria. *Biol. Pharm. Bull.* 25 (2), 197–200. doi:10.1248/bpb.25.197
- Kumari, S., Dhapola, R., Sharma, P., Nagar, P., Medhi, B., and HariKrishnaReddy, D. (2024). The impact of cytokines in neuroinflammation-mediated stroke. *Cytokine Growth Factor Rev.* 78, 105–119. doi:10.1016/j.cytogfr.2024.06.002
- Lai, H. F., Huang, X. X., and Shi, Z. L. (2018a). Optimization of ultrasonic-assisted extraction of amentoflavone from *Selaginella uncinata* (Desv.)spring using response surface methodology. *Food Res. Dev.* 39 (13), 47–51+224. doi:10.3969/j.issn.1005-6521.2018.13.009
- Lai, H. F., Pan, L. W., Lv, G. M., and Huang, X. X. (2018b). Purification of amentoflavone from *Selaginella uncinata* (Desv.)spring by macroporous resin. *Jiangsu Agric. Sci.* 46 (07), 201–204. doi:10.15889/j.issn.1002-1302.2018.07.050
- Lee, H., Cho, S., Kim, S. Y., Ju, J., Lee, S. W., Choi, S., et al. (2022). Amentoflavone-enriched *Selaginella rossii* warb. suppresses body weight and hyperglycemia by inhibiting intestinal lipid absorption in mice fed a high-fat diet. *Life (Basel)* 12 (4), 472. doi:10.3390/life12040472
- Lee, M. M., Cho, W. K., Cha, M. H., Yim, N. H., Yang, H. J., and Ma, J. Y. (2023). The antiviral activity of *Thuja orientalis folium* against Influenza A virus. *Virus Res.* 335, 199199. doi:10.1016/j.virusres.2023.199199
- Li, D. (2020). *Protective effect of amentoflavone against myocardial ischemia-reperfusion injury in rats*. Zunyi (Guizhou): Zunyi Medical University. dissertation.
- Li, W. W., Li, D., Qin, Y., Sun, C. X., Wang, Y. L., Gao, L., et al. (2021). Cardioprotective effects of Amentoflavone by suppression of apoptosis and inflammation on an in vitro and vivo model of myocardial ischemia-reperfusion injury. *Int. Immunopharmacol.* 101 (Pt B), 108296. doi:10.1016/j.intimp.2021.108296
- Li, X. (2011). *Extraction, characterization of Bi-flavonoid from Selaginella doederleinii and its interaction with bovine serum albumin*. Changsha (Hunan): Central South University. dissertation.
- Li, Y. L., Chen, X., Niu, S. Q., Zhou, H. Y., and Li, Q. S. (2020). Protective antioxidant effects of amentoflavone and total flavonoids from *Hedyotis diffusa* on H₂O₂-induced HL-O2 cells through ASK1/p38 MAPK pathway. *Chem. Biodivers.* 17 (7), e2000251. doi:10.1002/cbdv.202000251
- Liao, S., Ren, Q., Yang, C., Zhang, T., Li, J., Wang, X., et al. (2015). Liquid chromatography-tandem mass spectrometry determination and pharmacokinetic analysis of amentoflavone and its conjugated metabolites in rats. *J. Agric. Food Chem.* 63 (7), 1957–1966. doi:10.1021/jf5019615
- Libby, P. (2011). Inflammation during the life cycle of the atherosclerotic plaque. *Cardiovasc. Res.* 117 (13), 2525–2536. doi:10.1093/cvr/cvab303
- Li, D., Sun, C., Yang, J., Ma, X., Jiang, Y., Qiu, S., et al. (2019). Ionic liquid-microwave-based extraction of biflavonoids from *Selaginella sinensis*. *Molecules* 24 (13), 2507. doi:10.3390/molecules24132507

- Liu, C., Li, W. W., Lei, J., Li, G., Lu, D. M., Xu, D. J., et al. (2022). Deep eutectic solvents extraction and optimization of amentoflavone from *Selaginella moellendorffii*. *Sci. Technol. Food Ind.* 43 (16), 176–184. doi:10.13386/j.issn1002-0306.2021100160
- Liu, T. T., Qian, H., Li, Y. P., Wu, W. W., Min, X. W., Yang, H. D., et al. (2023). Study on the correlation between lipid accumulation product and risk levels of atherosclerotic cardiovascular disease. *Chin. Med. Her.* 20 (23), 77–80. doi:10.20047/j.issn1673-7210.2023.23.16
- Liu, Z., Ma, H., Sun, T., Wang, X. Y., and Kong, M. Z. (2024). Neuroprotective role of amentoflavone on LPS-induced Parkinson's disease animal model via inhibition of microglia-mediated inflammation. *J. Biol. Regul. Homeost. Agents* 38 (1), 705–718. doi:10.23812/j.biol.regul.homeost.agents.20243801.58
- Li, Y. Y., Lu, X. Y., Sun, J. L., Wang, Q. Q., Zhang, Y. D., Zhang, J. B., et al. (2019). Potential hepatic and renal toxicity induced by the biflavonoids from *Ginkgo biloba*. *Chin. J. Nat. Med.* 17 (9), 672–681. doi:10.1016/S1875-5364(19)30081-0
- Luo, S. (2017). *Study on the comprehensive quality control mode and preliminary activity of selaginella pulvinata*. Hangzhou (Zhejiang): Zhejiang University of Technology. dissertation.
- Ma, Y. Q., Lan, Y. M., and Zhang, Z. L. (2021). Research progress of correlation between intestinal microecology and cardiovascular and cerebrovascular disease. *J. Northwest Minzu Univ. Nat. Sci. Ed.* 42 (04), 21–27. doi:10.14084/j.cnki.cn62-1188/n.2021.04.007
- Mangmool, S., Duangrat, R., Rujirayunpong, T., and Anantachoke, N. (2024). Anti-inflammatory effects of the Thai herbal remedy Yataprasen and biflavonoids isolated from *Putranjiva roxburghii* in RAW264.7 macrophages. *J. Ethnopharmacol.* 327, 117997. doi:10.1016/j.jep.2024.117997
- Meng, Q. W., Liu, H. J., Yi, H. R., and Liu, Q. B. (2024). Mechanisms of NLRP3 inflammasome in atherosclerosis and advances in targeted inflammatory therapy. *Zhongguo dong mai ying hua za zhi* 32 (01), 79–86. doi:10.20039/j.cnki.1007-3949.2024.01.011
- Nagai, S., Ohara, K., and Mukai, K. (2005). Kinetic study of the quenching reaction of singlet oxygen by flavonoids in ethanol solution. *J. Phys. Chem. B* 109 (9), 4234–4240. doi:10.1021/jp0451389
- Oh, J., Rho, H. S., Yang, Y., Yoon, J. Y., Lee, J., Hong, Y. D., et al. (2013). Extracellular signal-regulated kinase is a direct target of the anti-inflammatory compound amentoflavone derived from *Torreya nucifera*. *Mediat. Inflamm.* 2013, 761506. doi:10.1155/2013/761506
- Okigawa, M., Hwa, C. W., Kawano, N., and Rahman, W. (1971). Biflavones in *selaginella* species. *Phytochemistry* 10, 3286–3287. doi:10.1016/S0031-9422(00)97392-8
- Oliveira, A. R., Jesus, P. A. P., Bulhões, F. V., Martins Netto, E., Oliveira Filho, J., Roevers, L., et al. (2023). Morbimortality and determinants of reperfusion in ischemic stroke. *Rev. Assoc. Medica Bras.* (1992) 70 (1), e20230472. doi:10.1590/1806-9282.20230472
- Park, H. J., and Kim, M. M. (2019). Amentoflavone induces autophagy and modulates p53. *Cell J.* 21 (1), 27–34. doi:10.22074/cellj.2019.5717
- Peng, Y., Chen, Q., Xue, Y. H., Jin, H., Liu, S., Du, M. Q., et al. (2024). *Ginkgo biloba* and its chemical components in the management of Alzheimer's disease. *Am. J. Chin. Med.* 52 (3), 625–666. doi:10.1142/S0192415X24500277
- Poznyak, A., Grechko, A. V., Poggio, P., Myasoedova, V. A., Alfieri, V., and Orekhov, A. N. (2020). The diabetes mellitus-atherosclerosis connection: the role of lipid and glucose metabolism and chronic inflammation. *Int. J. Mol. Sci.* 21 (5), 1835. doi:10.3390/ijms21051835
- Qin, L., Zhao, Y., Zhang, B., and Li, Y. (2018). Amentoflavone improves cardiovascular dysfunction and metabolic abnormalities in high fructose and fat diet-fed rats. *Food Funct.* 9 (1), 243–252. doi:10.1039/c7fo01095h
- Qin, T., Huang, Z. H., and Liu, Y. (2021). Research progress of vascular endothelial growth factor and atherosclerotic plaque. *Chin. Youjiang Med. J.* 49 (06), 465–468. doi:10.3969/j.issn.1003-1383.2021.06.012
- Qiu, F., Zhang, L., Zheng, J., Cao, L., Zhang, Z., and Deng, Y. (2021b). Amentoflavone inhibits M1 polarization of THP-1-derived foam cells by activating PPAR- α /J. *South. Med. Univ.* 41 (3), 344–351. doi:10.12122/j.issn.1673-4254.2021.03.05
- Qiu, S., Zhou, Y., Kim, J. T., Bao, C., Lee, H. J., and Chen, J. (2021a). Amentoflavone inhibits tumor necrosis factor- α -induced migration and invasion through AKT/mTOR/S6k1/hedgehog signaling in human breast cancer. *Food Funct.* 12 (20), 10196–10209. doi:10.1039/d1fo01085a
- Ren, Q. X., Wang, Y. N., Qu, X. Y., Zhen, B. Q., Hong, T., and Zhou, Z. (2013a). Preparation and *in vitro* evaluation of self-microemulsifying drug delivery system containing amentoflavone. *Chin. J. Exp. Tradit. Med. Formulae* 19 (24), 5–9. doi:10.11653/syfy2013240005
- Ren, Q. X., Zhou, Z., and Wang, Q. S. (2013b). Preparation and analytical characterization of micronized amentoflavone by antisolvent freeze-drying method. *J. Int. Pharm. Res.* 40 (02), 237–241. doi:10.13220/j.cnki.jipr.2013.02.009
- Ridker, P. M., Rifai, N., Pfeffer, M., Sacks, F., Lepage, S., and Braunwald, E. (2000). Elevation of tumor necrosis factor- α and increased risk of recurrent coronary events after myocardial infarction. *Circulation* 101 (18), 2149–2153. doi:10.1161/01.cir.101.18.2149
- Rong, S., Wan, D., Fan, Y., Liu, S., Sun, K., Huo, J., et al. (2019). Amentoflavone affects epileptogenesis and exerts neuroprotective effects by inhibiting NLRP3 inflammasome. *Front. Pharmacol.* 10, 856. doi:10.3389/fphar.2019.00856
- Roth, G. A., Mensah, G. A., and Fuster, V. (2020). The global burden of cardiovascular diseases and risks: a compass for global action. *J. Am. Coll. Cardiol.* 76 (25), 2980–2981. doi:10.1016/j.jacc.2020.11.021
- Ruan, X., Yan, L. Y., Li, X. X., Liu, B., Zhang, H., and Wang, Q. (2014). Optimization of process parameters of extraction of amentoflavone, quercetin and ginkgetin from *Taxus chinensis* using supercritical-CO₂ fluid extraction. *Molecules* 19 (11), 17682–17696. doi:10.3390/molecules191117682
- Saeedan, A. S., Abdel-Rahman, R. F., Soliman, G. A., Ogaly, H. A., and Abdel-Kader, M. S. (2023). Amentoflavone attenuates oxidative stress and neuroinflammation induced by cerebral ischemia/reperfusion in rats by targeting HMGB1-mediated TLR4/NF- κ B signaling pathway. *Saudi Pharm. J.* 31 (11), 101798. doi:10.1016/j.jsps.2023.101798
- Salara, P., Subrahmanyeswara Rao, N. N., Dharmeliya, T. M., and Amarendar Reddy, M. (2024). *In silico* investigation of potential phytoconstituents against ligand- and voltage-gated ion channels as antiepileptic agents. *3 Biotech.* 14 (4), 99. doi:10.1007/s13205-024-03948-1
- Šamec, D., Karalija, E., Dahija, S., and Hassan, S. T. S. (2022). Biflavonoids: important contributions to the health benefits of *Ginkgo* (*Ginkgo biloba* L.). *Plants (Basel)* 11 (10), 1381. doi:10.3390/plants11101381
- Saponara, R., and Bosio, E. (1998). Inhibition of cAMP-phosphodiesterase by biflavones of *Ginkgo biloba* in rat adipose tissue. *J. Nat. Prod.* 61 (11), 1386–1387. doi:10.1021/np970569m
- Shan, C. X., Guo, S. C., Yu, S., Shan, M. Q., Li, S. F. Y., Chai, C., et al. (2018). Simultaneous determination of quercetin, afzelin, amentoflavone, hinokiflavone in rat plasma by UPLC-MS-MS and its application to the pharmacokinetics of *Platycladus orientalis* leaves extract. *J. Chromatogr. Sci.* 56 (10), 895–902. doi:10.1093/chromsci/bmy066
- Sharma, P., Kishore, A., De, I., Negi, S., Kumar, G., Bhardwaj, S., et al. (2023). Mitigating neuroinflammation in Parkinson's disease: exploring the role of proinflammatory cytokines and the potential of phytochemicals as natural therapeutics. *Neurochem. Int.* 170, 105604. doi:10.1016/j.neuint.2023.105604
- Shin, D. H., Bae, Y. C., Kim-Han, J. S., Lee, J. H., Choi, I. Y., Son, K. H., et al. (2006). Polyphenol amentoflavone affords neuroprotection against neonatal hypoxic-ischemic brain damage via multiple mechanisms. *J. Neurochem.* 96 (2), 561–572. doi:10.1111/j.1471-4159.2005.03582.x
- Siddiqui, A. R., Mushtaq, M., Sardar, M., Atta, L., Nur-E-Alam, M., Ahmad, A., et al. (2024). Mechanistic insight into the mode of inhibition of dietary flavonoids; targeting macrophage migration inhibitory factor. *Front. Mol. Biosci.* 11, 1414572. doi:10.3389/fmolb.2024.1414572
- Singh, S., Goyal, A., and Agrawal, N. (2023). Molecular docking and dynamic simulation to identify α 7nAChR binding affinity of flavonoids for the treatment of alzheimer's disease. *Chem. Biodivers.* 20 (7), e202300306. doi:10.1002/cbdv.202300306
- Sirimangkalakitti, N., Juliawaty, L. D., Hakim, E. H., Waliana, I., Saito, N., Koyama, K., et al. (2019). Naturally occurring biflavonoids with amyloid β aggregation inhibitory activity for development of anti-Alzheimer agents. *Bioorg. Med. Chem. Lett.* 29 (15), 1994–1997. doi:10.1016/j.bmcl.2019.05.020
- Smith, E. E., Duchesne, S., Gao, F., Saad, F., Whitehead, V., McCreary, C. R., et al. (2021). Vascular contributions to neurodegeneration: protocol of the COMPASS-ND study. *Can. J. Neurol. Sci.* 48 (6), 799–806. doi:10.1017/cjn.2021.19
- Sun, B. R., Wang, Z., and Lin, Z. Y. (2023). Risk factors of carotid atherosclerosis in elderly patients with type 2 diabetes mellitus complicated with coronary heart disease. *Jilin Med. J.* 44 (05), 1183–1185.
- Sun, L., Sharma, A. K., Han, B. H., and Mirica, L. M. (2020). Amentoflavone: a bifunctional metal chelator that controls the formation of neurotoxic soluble A β ₄₂ oligomers. *ACS Chem. Neurosci.* 11 (17), 2741–2752. doi:10.1021/acscchemneuro.0c00376
- Tao, Y., Zhu, F., Pan, M., Liu, Q., and Wang, P. (2022). Pharmacokinetic, metabolism, and metabolomic strategies provide deep insight into the underlying mechanism of *Ginkgo biloba* flavonoids in the treatment of cardiovascular disease. *Front. Nutr.* 9, 857370. doi:10.3389/fnut.2022.857370
- Tarallo, V., Lepore, L., Marcellini, M., Dal Piaz, F., Tudisco, L., Ponticelli, S., et al. (2011). The biflavonoid amentoflavone inhibits neovascularization preventing the activity of proangiogenic vascular endothelial growth factors. *J. Biol. Chem.* 286 (22), 19641–19651. doi:10.1074/jbc.M110.186239
- The Writing Committee Of The Report On Cardiovascular Health And DiseasesHu, S. S. (2023). Report on cardiovascular health and diseases in China 2021: an updated summary. *J. Geriatr. Cardiol.* 20 (6), 399–430. doi:10.26599/1671-5411.2023.06.001
- Vlasiou, M. C. (2023). Cheese and milk adulteration: detection with spectroscopic techniques and HPLC: advantages and disadvantages. *Dairy* 4 (3), 509–514. doi:10.3390/dairy4030034
- Wang, B., Lu, Y., Wang, R., Liu, S., Hu, X., and Wang, H. (2020a). Transport and metabolic profiling studies of amentoflavone in Caco-2 cells by UHPLC-ESI-MS/MS and UHPLC-ESI-Q-TOF-MS/MS. *J. Pharm. Biomed. Anal.* 189, 113441. doi:10.1016/j.jpba.2020.113441
- Wang, G., Li, D., Jiang, F. Q., Xie, X. Y., and He, Y. Q. (2018b). Optimization of microwave-assisted ionic liquid extraction of amentoflavone from *Selaginella doederleinii* Hieron. *Chin. Tradit. Pat. Med.* 40 (08), 1851–1855. doi:10.3969/j.issn.1001-1528.2018.08.040
- Wang, G., Tian, Y. B., Jiang, Y. M., He, Q., Yuan, S. M., and Wei, W. (2018c). Optimization of microwave extraction of amentoflavone from *Selaginella doederleinii* by response surface method. *Sci. Technol. Food Ind.* 39 (07), 146–151. doi:10.13386/j.issn1002-0306.2018.07.027

- Wang, H. (2017). *Study on extraction, isolation, characterization and anti-oxidant activities of flavonoids from Podocarpus nagi*. Jishou (Hunan): Jishou University. dissertation.
- Wang, J. Q., Li, M. M., Fan, B., Cui, W. Y., Wang, Q., Lu, C., et al. (2023). Research progress on extraction, separation and biological activity of natural Bi-flavonoids. *Food Nutr. China*, 1–8. doi:10.19870/j.cnki.11-3716/ts.20231018.001
- Wang, L. (2013). *Protective effect against hydroxyl radical-induced DNA damage and antioxidant mechanism of amentoflavone*. Guangzhou (Guangdong): Guangzhou University of Chinese Medicine. dissertation.
- Wang, X., Zhao, X., Gu, L., Lv, C., He, B., Liu, Z., et al. (2014). Simultaneous determination of five free and total flavonoids in rat plasma by ultra HPLC-MS/MS and its application to a comparative pharmacokinetic study in normal and hyperlipidemic rats. *J. Chromatogr. B Anal. Technol. Biomed. Life Sci.* 953–954, 1–10. doi:10.1016/j.jchromb.2014.01.042
- Wang, Y., Chen, L. Z., Duan, H. T., Wang, L. P., Xu, C., Ling, L., et al. (2018a). Determination of content of amentoflavone in *Selaginella tamariscina* by infrared-assisted extraction method coupled with HPLC. *Chin. J. Clin. Pharm.* 27 (03), 164–166. doi:10.19577/j.1007-4406.2018.03.005
- Wang, Y., Wang, K., and Fu, J. (2020b). HDAC6 mediates macrophage iNOS expression and excessive nitric oxide production in the blood during endotoxemia. *Front. Immunol.* 11, 1893. doi:10.3389/fimmu.2020.01893
- Wang, Y. Z., Zhang, M., Liu, Y., Zhao, H. L., and Zhen, X. K. (2015). Pharmacokinetics study on amentoflavone in rats. *Chin. Tradit. Pat. Med.* 37 (11), 2397–2401. doi:10.3969/j.issn.1001-1528.2015.11.013
- Wang, Z., Wang, B., and Jin, X. (2024). Amentoflavone attenuates homocysteine-induced neuronal ferroptosis-mediated inflammatory response: involvement of the SLC7A11/GPX4 axis activation. *Brain Res. Bull.* 215, 111005. doi:10.1016/j.brainresbull.2024.111005
- Wei, L. N. (2010). *Study on the separation and purification of amentoflavone from selaginella tamariscina* (beauv.) spring. Beijing (Beijing): Beijing University of Chemical Technology. dissertation.
- Wei, B. Y., and Liang, M. H. (2021). Research progress of mechanism of diabetes melitus causes atherosclerosis. *Chin. J. Cardiovasc Rehabil. Med.* 30 (01), 85–87. doi:10.3969/j.issn.1008-0074.2021.01.22
- Wei, D. D., Chen, X., Zhao, J. J., Hu, X. L., Xiong, F., and Wang, H. (2017). Studies on the intestinal absorption kinetics of amentoflavone in rats. *Pharm. Clin. Res.* 25 (04), 282–285. doi:10.13664/j.cnki.pcr.2017.04.002
- Wei, L. H., Chen, T. R., Fang, H. B., Jin, Q., Zhang, S. J., Hou, J., et al. (2019). Natural constituents of *St. John's Wort* inhibit the proteolytic activity of human thrombin. *Int. J. Biol. Macromol.* 134, 622–630. doi:10.1016/j.ijbiomac.2019.04.181
- Windsor, P. K., Plassmeyer, S. P., Mattock, D. S., Bradfield, J. C., Choi, E. Y., Miller, B. R., et al. (2021). Biflavonoid-induced disruption of hydrogen bonds leads to amyloid- β disaggregation. *Int. J. Mol. Sci.* 22 (6), 2888. doi:10.3390/ijms22062888
- Wisznia, S., and Schwarz, Q. (2021). Exploring the intracrine functions of VEGF-A. *Biomolecules* 11 (1), 128. doi:10.3390/biom11010128
- Woo, E. R., Lee, J. Y., Cho, I. J., Kim, S. G., and Kang, K. W. (2005). Amentoflavone inhibits the induction of nitric oxide synthase by inhibiting NF-kappaB activation in macrophages. *Pharmacol. Res.* 51 (6), 539–546. doi:10.1016/j.phrs.2005.02.002
- Xi, C. C., Gu, W., and Zhou, L. L. (2020). The isolation, identification, antioxidation and anticoagulation activity of selaginellin and amentoflavone from *selaginella tamariscina* (beauv.) Spring. *Acta Medica Mediterr.* 36, 2697–2706. doi:10.19193/0393-6384_2020_4_414
- Xiao, L., Chen, Y., Zhang, F., Long, X. Y., Nie, J., and Huang, Z. J. (2018). Chemical constituents of biflavonoids from *Selaginella uncinata*. *J. Pharm. Anal.* 38 (12), 2093–2103. doi:10.16155/j.0254-1793.2018.12.06
- Xiong, L., Liu, Y., Wang, Y., Zhao, H., Song, X., Fan, W., et al. (2024). The protective effect of *Lonicera japonica* Thunb. against lipopolysaccharide-induced acute lung injury in mice: modulation of inflammation, oxidative stress, and ferroptosis. *J. Ethnopharmacol.* 331, 118333. doi:10.1016/j.jep.2024.118333
- Xiong, X., Tang, N., Lai, X., Zhang, J., Wen, W., Li, X., et al. (2021). Insights into amentoflavone: a natural multifunctional biflavonoid. *Front. Pharmacol.* 12, 768708. doi:10.3389/fphar.2021.768708
- Xu, F., Li, J., Mao, Y., Xu, X. J., and He, J. H. (2013). *Ginkgo biloba* leaf extract research progress. *Food Res. Dev.* 16, 124–127. doi:10.3969/j.issn.1005-6521.2013.16.035
- Xu, H., Li, R. H., Xia, Y. S., and Yang, X. Q. (2021). Research status and prospect of extraction and purification methods of flavonoids. *Appl. Chem. Ind.* 50 (06), 1677–1682. doi:10.16581/j.cnki.issn1671-3206.2021.06.010
- Xu, J., Wang, Y., Zheng, T., Huo, Y., and Du, W. (2022). Biflavones inhibit the fibrillation and cytotoxicity of the human islet amyloid polypeptide. *J. Mater Chem. B* 10 (24), 4650–4661. doi:10.1039/d2tb00230b
- Xu, Z., Jia, S. J., Tan, G. S., and Li, Y. J. (2004). Study on pharmacological activity of biflavones from *Selaginella pulvinata* (hook. Et grev.) Maxim. *Chin. J. Mod. Med.* 14, 88–89+100.
- Xun, L., and Yin, M. H. (2009). Experiment study on vasodilative effects of amentoflavone ethylacetate extract of *Selaginella tamariscina*. *J. Med. Sci. Yanbian Univ.* 32 (04), 246–248. doi:10.16068/j.1000-1824.2009.04.011
- Yan, X. D., and Guo, M. L. (2022). Research progress of the effect of flavonoids on cardiovascular and cerebrovascular ischemic diseases. *J. Pharm. Pract. Serv.* 40 (02), 97–102. doi:10.12206/j.issn.1006-0111.202111059
- Yang, Y. T. (2023). *The effect of EQST on atherosclerosis formation in mice and its mechanism*. Guilin (Guangxi): Guangxi Normal University. dissertation.
- Yao, W., Li, H., Liu, Q., Gao, Y., Dai, J., Bao, B., et al. (2016). Cellular metabolomics revealed the cytoprotection of amentoflavone, a natural compound, in lipopolysaccharide-induced injury of human umbilical vein endothelial cells. *Int. J. Mol. Sci.* 17 (9), 1514. doi:10.3390/ijms17091514
- Yu, S., Yan, H., Zhang, L., Shan, M., Chen, P., Ding, A., et al. (2017). A review on the phytochemistry, pharmacology, and pharmacokinetics of amentoflavone, a naturally-occurring biflavonoid. *Molecules* 22 (2), 299. doi:10.3390/molecules22020299
- Zhang, J., Zhou, J., Zhang, T., Niu, Z., Wang, J., Guo, J., et al. (2019). Facile fabrication of an amentoflavone-loaded micelle system for oral delivery to improve bioavailability and hypoglycemic effects in KKAY mice. *ACS Appl. Mater Interfaces* 11 (13), 12904–12913. doi:10.1021/acsami.9b03275
- Zhang, L. J. (2014). *The effects and potential mechanism of total flavonoids of Selaginella Pulvinata and amentoflavone against cognitive impairment*. Changchun (Jiling): Jiling University. dissertation.
- Zhang, Q., Wang, Y. L., Gao, D., Cai, L., Yang, Y. Y., Hu, Y. J., et al. (2018). Comparing coagulation activity of *Selaginella tamariscina* before and after stir-frying process and determining the possible active constituents based on compositional variation. *Pharm. Biol.* 56 (1), 67–75. doi:10.1080/13880209.2017.1421673
- Zhang, X., Liu, Y., Deng, G., Huang, B., Kai, G., Chen, K., et al. (2021a). A purified biflavonoid extract from *Selaginella moellendorffii* alleviates gout arthritis via NLRP3/ASC/Caspase-1 Axis suppression. *Front. Pharmacol.* 12, 676297. doi:10.3389/fphar.2021.676297
- Zhang, Z., Sun, T., Niu, J. G., He, Z. Q., Liu, Y., and Wang, F. (2015). Amentoflavone protects hippocampal neurons: anti-inflammatory, antioxidative, and antiapoptotic effects. *Neural Regen. Res.* 10 (7), 1125–1133. doi:10.4103/1673-5374.160109
- Zhang, Z., and Wang, F. (2013). Progress in research of the biological activity of amentoflavone. *Chin. J. New Drugs.* 22 (23), 2775–2778.
- Zhang, Z., Yang, X., Song, Y. Q., and Tu, J. (2021b). Autophagy in Alzheimer's disease pathogenesis: therapeutic potential and future perspectives. *Ageing Res. Rev.* 72, 101464. doi:10.1016/j.arr.2021.101464
- Zhao, F., Qian, Y., Li, H., Yang, Y., Wang, J., Yu, W., et al. (2022). Amentoflavone-loaded nanoparticles enhanced chemotherapy efficacy by inhibition of AKR1B10. *Nanotechnology* 33 (38), 385101. doi:10.1088/1361-6528/ac7810
- Zhao, N., Sun, C., Zheng, M., Liu, S., and Shi, R. (2019). Amentoflavone suppresses amyloid β 1-42 neurotoxicity in Alzheimer's disease through the inhibition of pyroptosis. *Life Sci.* 239, 117043. doi:10.1016/j.lfs.2019.117043
- Zheng, X. K., Ling, T. L., Wang, X. L., Liu, C. X., Liu, Y. Y., and Feng, W. S. (2011). Effects of total flavonoids and amentoflavone isolated from *Selaginella tamariscina* on human umbilical vein endothelial cells proliferation and VEGF expression. *Chin. Pharm. J.* 46 (13), 998–1002.
- Zheng, X. K., Liu, C. X., Zhai, Y. Y., Li, L. L., Wang, X. L., and Feng, W. S. (2013). Protection effect of amentoflavone in *Selaginella tamariscina* against TNF- α -induced vascular injury of endothelial cells. *Yao Xue Xue Bao* 48 (09), 1503–1509. doi:10.16438/j.0513-4870.2013.09.014
- Zheng, X. K., Wei, Y., Feng, W. S., Li, Y. J., and Zhao, X. M. (2008). "Study on hypoglycemic activity of amentoflavone in vitro," in *Communication between biochemistry and molecular biology of traditional Chinese medicine*, 182–187.
- Zhou, L. N., Wang, R., Wang, J. J., Zhao, K., Zhang, T., Hu, X. L., et al. (2022). Characterization, solubility and stability of amentoflavone polymorphs. *J. Mol. Struct.* 1262, 133101. doi:10.1016/j.molstruc.2022.133101
- Zhou, Y., Yang, Z., Guo, Z., Kang, Y., and Zhou, K. J. (2024). Research progress on the mechanism of *Tianma Gouteng* decoction in preventing and treating nervous system diseases. *Glob. Tradit. Chin. Med.* 17 (01), 157–165. doi:10.3969/j.issn.1674-1749.2024.01.031
- Zhuang, J. L. (2020). *Effects and mechanisms of amentoflavone on lipid accumulation and cell migration*. Guangzhou (Guangdong): Guangzhou University of Chinese Medicine. dissertation.

Glossary

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|------------------------|--|--------------------------|---|
| AME | Amentoflavone | IL | Interleukin |
| AGEs | Advanced glycation end-products | IκB | Inhibitor of NF-κB |
| ASK1 | Apoptosis signal regulating kinase 1 | iNOS | Nitric oxide synthase |
| Akt | Protein kinase B | LPS | Lipopolysaccharide |
| AD | Alzheimer's disease | LDL | Low-density lipoprotein |
| Aβ | β-amyloid | MRT_{0-∞} | Mean residence time |
| AChE | Acetylcholinesterase | MRT_{0-t} | Average retention time for a certain period of time |
| Ang | Angiotensin | MMPs | Matrix metalloproteinases |
| ACE | Angiotensin-converting enzyme | MCE | Mitotic clonal expansion phase |
| AMPK | Adenosine monophosphate-activated protein kinase | MDA | Malondialdehyde |
| AT1R | Angiotensin II receptor-1 | MAPK | Mitogen-activated protein kinase |
| Arg1 | Arginase 1 | mTOR | Mechanistic target of rapamycin |
| AUC(0-t) | Area under the concentration-time curve | MPTP | Methyl-4-phenyl-1,2,3,6-tetrahydropyridine |
| AUC(0-∞) | From time zero to all original drug elimination | MASR | Mitochondrial assembly receptor |
| AMPA | α-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid | MIF | Macrophage migration inhibitory factor |
| α7nAChR | α7 nicotinic acetylcholine receptor | NLRP3 | NOD-like receptor thermal protein domain associated protein 3 |
| C_{max} | Maximum blood concentration | NOX | NADPH oxidase |
| CL/F | Clearance | NF-κB | Nuclear factor-kappa B |
| COX-2 | Cyclooxygenase-2 | NO | Nitric oxide |
| CD36 | Cluster of differentiation 36 | Nrf2 | Nuclear factor erythroid 2-related factor-2 |
| C/EBP | Transcription factor CCAAT enhancer binding protein | NRF-1 | Nuclear respiratory factor-1 |
| CAT | Catalase | ox-LDL | Oxidized low-density lipoprotein |
| cGMP | Guanosine 3',5'-cyclic monophosphate | PGE-2 | Prostaglandin E-2 |
| caspase | Cysteine aspartate specific protease | PPAR | Peroxisome proliferator-activated receptor |
| DPPH | 1,1-diphenyl-2 trinitrophenylhydrazine | PI3K | Phosphoinositide 3-kinase |
| DOX | Adriamycin | PD | Parkinson's disease |
| ECs | Endothelial cells | PIGF-1 | Placental growth factor-1 |
| ET-1 | Endothelin-1 | ROS | Reactive oxygen species |
| ERK | Extracellular signal-regulated kinase | SOD | Superoxide dismutase |
| FasL | Fas ligand | STING | Stimulator of interferon genes |
| F | Bioavailability | SLC7A11 | Solute carrier family 7 member 11 |
| Fizz1 | Found in inflammatory zone | TG | Triglycerides |
| GSK-3-β | Glucose synthesis kinase-3-β | Trx | Thioredoxin |
| GBE | <i>Ginkgo biloba</i> extract | TrxR-1 | Thioredoxin reductase-1 |
| GSH | Glutathione | T_{max} | Time to peak concentration |
| GPX4 | Glutathione peroxidase 4 | T_{1/2} | Biological half-life |
| HMGB1 | High mobility group box protein B1 | T2DM | Type 2 diabetes model |
| HSP-27 | Heat shock protein 27 | TNF | Tumor necrosis factor |
| HFFD | High fructose and fat diet | TGF | Transforming growth factor |
| HPLC | High-performance liquid chromatography | TLR4 | Toll-like receptor-4 |
| HUVECs | Human umbilical vein endothelial cells | TFAM | Mitochondrial transcription factor |
| hIAPP | Human islet amyloid polypeptide | UGT1A | Uridine diphosphate glucuronosyltransferase 1 family, polypeptide A |

| | |
|------------|-----------------------------------|
| V/F | Apparent distribution volume |
| VSMCs | Vascular smooth muscle cells |
| VEGF | Endothelial growth factor |
| VGSC | Voltage-gated sodium ion channels |
| <i>Ym1</i> | Chitinase-like protein 3 |

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