

# EMERGING BIOMARKERS IN PERSONALIZED THERAPY OF UROLOGIC TUMORS

EDITED BY: Rodolfo Montironi, Matteo Santoni, Alessia Cimadamore,  
Antonio Lopez-Beltran and Liang Cheng

PUBLISHED IN: Frontiers in Oncology





# frontiers

## Frontiers Copyright Statement

© Copyright 2007-2019 Frontiers Media SA. All rights reserved.

All content included on this site, such as text, graphics, logos, button icons, images, video/audio clips, downloads, data compilations and software, is the property of or is licensed to Frontiers Media SA ("Frontiers") or its licensees and/or subcontractors. The copyright in the text of individual articles is the property of their respective authors, subject to a license granted to Frontiers.

The compilation of articles constituting this e-book, wherever published, as well as the compilation of all other content on this site, is the exclusive property of Frontiers. For the conditions for downloading and copying of e-books from Frontiers' website, please see the Terms for Website Use. If purchasing Frontiers e-books from other websites or sources, the conditions of the website concerned apply.

Images and graphics not forming part of user-contributed materials may not be downloaded or copied without permission.

Individual articles may be downloaded and reproduced in accordance with the principles of the CC-BY licence subject to any copyright or other notices. They may not be re-sold as an e-book.

As author or other contributor you grant a CC-BY licence to others to reproduce your articles, including any graphics and third-party materials supplied by you, in accordance with the Conditions for Website Use and subject to any copyright notices which you include in connection with your articles and materials.

All copyright, and all rights therein, are protected by national and international copyright laws.

The above represents a summary only. For the full conditions see the Conditions for Authors and the Conditions for Website Use.

ISSN 1664-8714

ISBN 978-2-88945-929-2

DOI 10.3389/978-2-88945-929-2

## About Frontiers

Frontiers is more than just an open-access publisher of scholarly articles: it is a pioneering approach to the world of academia, radically improving the way scholarly research is managed. The grand vision of Frontiers is a world where all people have an equal opportunity to seek, share and generate knowledge. Frontiers provides immediate and permanent online open access to all its publications, but this alone is not enough to realize our grand goals.

## Frontiers Journal Series

The Frontiers Journal Series is a multi-tier and interdisciplinary set of open-access, online journals, promising a paradigm shift from the current review, selection and dissemination processes in academic publishing. All Frontiers journals are driven by researchers for researchers; therefore, they constitute a service to the scholarly community. At the same time, the Frontiers Journal Series operates on a revolutionary invention, the tiered publishing system, initially addressing specific communities of scholars, and gradually climbing up to broader public understanding, thus serving the interests of the lay society, too.

## Dedication to Quality

Each Frontiers article is a landmark of the highest quality, thanks to genuinely collaborative interactions between authors and review editors, who include some of the world's best academicians. Research must be certified by peers before entering a stream of knowledge that may eventually reach the public - and shape society; therefore, Frontiers only applies the most rigorous and unbiased reviews.

Frontiers revolutionizes research publishing by freely delivering the most outstanding research, evaluated with no bias from both the academic and social point of view. By applying the most advanced information technologies, Frontiers is catapulting scholarly publishing into a new generation.

## What are Frontiers Research Topics?

Frontiers Research Topics are very popular trademarks of the Frontiers Journals Series: they are collections of at least ten articles, all centered on a particular subject. With their unique mix of varied contributions from Original Research to Review Articles, Frontiers Research Topics unify the most influential researchers, the latest key findings and historical advances in a hot research area! Find out more on how to host your own Frontiers Research Topic or contribute to one as an author by contacting the Frontiers Editorial Office: [researchtopics@frontiersin.org](mailto:researchtopics@frontiersin.org)

# EMERGING BIOMARKERS IN PERSONALIZED THERAPY OF UROLOGIC TUMORS

Topic Editors:

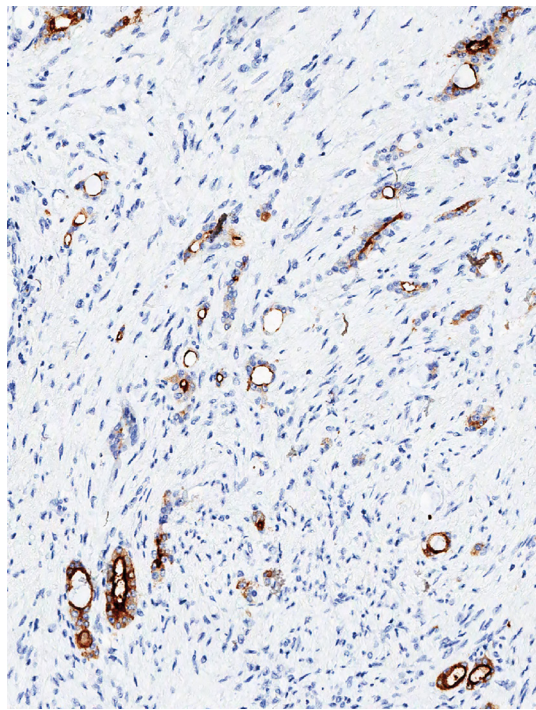
**Rodolfo Montironi**, Polytechnic University of the Marche Region, Italy

**Matteo Santoni**, Macerata Hospital, Italy

**Alessia Cimadamore**, Polytechnic University of the Marche Region, Italy

**Antonio Lopez-Beltran**, Cordoba University, Spain

**Liang Cheng**, Indiana University School of Medicine, United States



PSMA expression in prostate cancer glands following radiotherapy.

Image by Dr. Alessia Cimadamore.

The identification of effective biomarkers has becoming a major focus in cancer research, mainly due to the necessity of selecting potentially responsive patients in order to improve their outcomes, as well as to reduce the toxicity and costs related to ineffective treatments. In genitourinary tumors, the lack of biomarkers does not allow for the development of personalized strategies for a single patient, thus representing a major goal in this field. This eBook includes the description of these emerging techniques and identify the most promising biomarkers in genitourinary tumors.

**Citation:** Montironi, R., Santoni, M., Cimadamore, A., Lopez-Beltran, A., Cheng, L., eds. (2019). Emerging Biomarkers in Personalized Therapy of Urologic Tumors. Lausanne: Frontiers Media. doi: 10.3389/978-2-88945-929-2

# Table of Contents

- 04 Editorial: Emerging Biomarkers in Genitourinary Tumors**  
Rodolfo Montironi, Matteo Santoni, Alessia Cimadamore,  
Antonio Lopez-Beltran and Liang Cheng
- 06 Microbiome and Cancers, With Focus on Genitourinary Tumors**  
Alessia Cimadamore, Matteo Santoni, Francesco Massari, Silvia Gasparrini,  
Liang Cheng, Antonio Lopez-Beltran, Rodolfo Montironi and Marina Scarpelli
- 12 The Identification of Immunological Biomarkers in Kidney Cancers**  
Antonio Lopez-Beltran, Vanessa Henriques, Alessia Cimadamore,  
Matteo Santoni, Liang Cheng, Thomas Gevaert, Ana Blanca,  
Francesco Massari, Marina Scarpelli and Rodolfo Montironi
- 25 New Prostate Cancer Targets for Diagnosis, Imaging, and Therapy: Focus  
on Prostate-Specific Membrane Antigen**  
Alessia Cimadamore, Monica Cheng, Matteo Santoni, Antonio Lopez-Beltran,  
Nicola Battelli, Francesco Massari, Andrea B. Galosi, Marina Scarpelli and  
Rodolfo Montironi
- 36 Recent Advances in Liquid Biopsy in Patients With Castration Resistant  
Prostate Cancer**  
Vincenzo Di Nunno, Lidia Gatto, Matteo Santoni, Alessia Cimadamore,  
Antonio Lopez-Beltran, Liang Cheng, Marina Scarpelli, Rodolfo Montironi  
and Francesco Massari
- 45 Urinary Markers in Bladder Cancer: An Update**  
Giorgio Santoni, Maria B. Morelli, Consuelo Amantini and Nicola Battelli
- 54 Circulating Tumor Cells in Renal Cell Carcinoma: Recent Findings and  
Future Challenges**  
Matteo Santoni, Alessia Cimadamore, Liang Cheng, Antonio Lopez-Beltran,  
Nicola Battelli, Francesco Massari, Marina Scarpelli, Andrea Benedetto Galosi,  
Sergio Bracarda and Rodolfo Montironi
- 60 Emerging Prognostic Biomarkers in Testicular Germ Cell Tumors: Looking  
Beyond Established Practice**  
Michal Chovanec, Costantine Albany, Michal Mego, Rodolfo Montironi,  
Alessia Cimadamore and Liang Cheng
- 67 Emerging Biomarkers in Bladder Cancer Identified by Network Analysis of  
Transcriptomic Data**  
Matteo Giulietti, Giulia Occhipinti, Alessandra Righetti, Massimo Bracci,  
Alessandro Conti, Annamaria Ruzzo, Elisabetta Cerigioni, Tiziana Cacciamani,  
Giovanni Principato and Francesco Piva
- 75 Emerging Molecular Technologies in Genitourinary Tumors**  
Francesca Giunchi, Alessia Cimadamore and Michelangelo Fiorentino



# Editorial: Emerging Biomarkers in Genitourinary Tumors

Rodolfo Montironi<sup>1\*</sup>, Matteo Santoni<sup>2</sup>, Alessia Cimadamore<sup>1</sup>, Antonio Lopez-Beltran<sup>3</sup> and Liang Cheng<sup>4</sup>

<sup>1</sup> Section of Pathological Anatomy, School of Medicine, United Hospitals, Polytechnic University of the Marche Region, Ancona, Italy, <sup>2</sup> Oncology Unit, Macerata Hospital, Macerata, Italy, <sup>3</sup> Department of Pathology and Surgery, Faculty of Medicine, Cordoba University, Cordoba, Spain, <sup>4</sup> Department of Pathology and Laboratory Medicine, Indiana University School of Medicine, Indianapolis, IN, United States

**Keywords:** genitourinary tumors, microbiome, renal cell carcinoma, prostate cancer, bladder cancer, liquid biopsy, PSMA, immunotherapy

## Editorial on the Research Topic

### Emerging Biomarkers in Genitourinary Tumors

This is a contemporary update in the field of *Emerging Biomarkers in Genitourinary Tumors*. This series of papers, published in Frontiers in Oncology, section Genitourinary Oncology, by internationally renowned researchers, covers five major topics: (1). Identification of immunological biomarkers in genitourinary cancers; (2). New prostate cancer targets for imaging and therapy; (3) Liquid molecular biomarkers in genitourinary tumors; (4) Emerging biomarkers in testicular germ cell tumors; and (5) Future perspectives on molecular biomarkers in genitourinary tumors: toward a personalized approach to diagnosis, prognosis and prediction of response to therapy.

The first two papers are related to the Identification of immunological biomarkers in genitourinary cancers. In particular, the first, by Cimadamore et al., deals with the biological relationship between the gut microbiome and the immune system, in particular cancer development and treatment. As an example, *Akkermansia muciniphila* is a commensal associated with excellent clinical outcomes in renal cell carcinoma and non-small cell lung cancer. Interesting results have emerged on the microbiome in prostate cancer (PCa) patients, with specific bacteria as potential biomarkers in risk stratification. Abnormal gut microbiome composition could also have an influence on primary resistance to PD-1 blockade in mice xenografts and patients with cancer. The contribution by Lopez-Beltran et al. deals with the identification of novel immunological biomarkers in kidney cancers. Robust and reliable biomarkers are crucial for patient's selection for treatment with immunomodulatory drugs. PD-L1 expression is predictive of better response from both PD-1 and PD-L1 inhibitors in a variety of tumor types including RCC. A single biomarker for patient selection may not be feasible, given that immune responses are dynamic and evolve over time. A multidisciplinary approach is very much needed to fully develop the current and future value of immune checkpoint inhibitors in clinical practice.

The third paper of the whole series, by Cimadamore et al., deals with New PCa targets for imaging and therapy, focusing on Prostate-Specific Membrane Antigen (PSMA). This contribution reviews the current role of PSMA as a marker for PCa diagnosis, imaging and therapy. PSMA is expressed in the epithelial cells of the prostate and is strongly upregulated in PCa, with elevated expression correlating with androgen independence, metastasis and progression. PSMA has been found to be an active target of investigation by several approaches, including the successful use of small molecule inhibitors, RNA aptamer conjugates, PSMA-based immunotherapy, and PSMA-targeted prodrug therapy. The next three papers deals

## OPEN ACCESS

### Edited and reviewed by:

Michael Kattan,  
Cleveland Clinic Lerner College of  
Medicine, United States

### \*Correspondence:

Rodolfo Montironi  
r.montironi@univpm.it

### Specialty section:

This article was submitted to  
Genitourinary Oncology,  
a section of the journal  
Frontiers in Oncology

**Received:** 14 March 2019

**Accepted:** 10 April 2019

**Published:** 26 April 2019

### Citation:

Montironi R, Santoni M,  
Cimadamore A, Lopez-Beltran A and  
Cheng L (2019) Editorial: Emerging  
Biomarkers in Genitourinary Tumors.  
Front. Oncol. 9:326.  
doi: 10.3389/fonc.2019.00326

with Liquid molecular biomarkers in genitourinary tumors. The first of the three, by Di Nunno et al., is related to recent advances in liquid biopsy in patients with castration resistant PCa. The selection of patients more likely to benefit from a specific therapeutic approach still remains a key issue as well as the early identification of patients with aggressive disease which could benefit from a more aggressive treatment strategy. They review the literature to explore current knowledge on liquid biopsy in PCa focusing on possible future applications. In particular they focus on circulating DNA and circulating tumor cells as a promising and attractive approach despite to date practical applications of these techniques are few and not validated. The paper by Santoni et al. is an updates on urine markers in superficial and non-superficial bladder cancer (BCa). There is a growing evidence toward the use of minimally invasive “liquid biopsy” to identify new biomarkers. DNA- and RNA-based markers in body fluids such as blood and urine are promising potential markers in diagnostic, prognostic, predictive and monitoring BCa. However, proteomic and genomic data must to be validated in well-designed multicenter clinical studies, before to be employed in clinic oncology. The paper by Santoni et al. deals with recent findings and future challenges of circulating tumor cells (CTCs) in renal cell carcinoma (Santoni et al.). Renal Cell Carcinoma (RCC) may absolutely benefit from the development of non-invasive and reliable biomarkers, allowing early and timely personalized treatment changes. The introduction of CTC analysis within daily clinical practice for patients with RCC seems still so far at the moment. However, the advances obtained in the last 5 years in isolating and analyzing CTCs bring optimism about the future therapeutic landscape in RCC patients.

The contribution by Chovanec et al. deals with Emerging biomarkers in testicular germ cell tumors (GCTs). The ability to predict prognosis and treatment response in GCTs did not improve for many years. Clinical trials with novel targeting agents that were conducted in refractory GCT patients have proven to have negative outcomes. Novel biomarkers have emerged in the field of GCT oncology. Since then, oncology has exploded with various molecular biomarkers to further refine the prognosis and treatment of malignancies. This review summarizes the current knowledge in the research of novel biomarkers in GCTs.

The remaining two papers deal with Future perspectives on molecular biomarkers in genitourinary tumors: toward a personalized approach to diagnosis, prognosis and prediction of response to therapy. The paper by Giulietti et al. is related to emerging biomarkers in BCa identified by network analysis

of transcriptomic data. Such complex gene interaction networks can be revealed by a recently developed systems approach called Weighted Gene Co-expression Network Analysis (WGCNA). In this review, the authors focused on the studies where the WGCNA approach has been applied to analyze gene expression data deriving from BCa samples. The paper by Giunchi et al. is a perspective article on emerging molecular technologies in genitourinary tumors. In particular, they deal with wide spectrum mutational analyses using next generation sequencing (NGS) platforms that will soon represent the standard-of-care technologies for the assessment of genetic variants in genitourinary tumors. They also deal with genome-wide transcriptome analyses which include gene expression profiling, miRNA and non-coding RNA profiling and RNA sequencing. Toward the end of the contribution they refer to patient-derived xenografts (PDX), i.e., mouse models where disaggregated cells or little fragments of human tumors are implanted into immunodeficient mice. The establishment of a PDX allows treating and monitoring the response to treatment of the original tumor *in vivo* in the mouse, instead of the patient, providing the best therapeutic selection at the same time.

## CONCLUSIONS

The identification of effective biomarkers has becoming a major focus in cancer research, mainly due to the necessity of selecting potentially responsive patients in order to improve their outcomes, as well as to reduce the toxicity and costs related to ineffective treatments. This Research Topic aims to include the description of these emerging techniques and identify the most promising biomarkers in genitourinary tumors.

## AUTHOR CONTRIBUTIONS

RM and MS: Conception and design. AC: Drafting the manuscript. AL-B and LC: Critical revision of the manuscript and review of the literature.

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Montironi, Santoni, Cimadamore, Lopez-Beltran and Cheng. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.





# Microbiome and Cancers, With Focus on Genitourinary Tumors

Alessia Cimadamore<sup>1\*</sup>, Matteo Santoni<sup>2</sup>, Francesco Massari<sup>3</sup>, Silvia Gasparrini<sup>1</sup>, Liang Cheng<sup>4</sup>, Antonio Lopez-Beltran<sup>5</sup>, Rodolfo Montironi<sup>1\*</sup> and Marina Scarpelli<sup>1</sup>

<sup>1</sup> Section of Pathological Anatomy, School of Medicine, United Hospitals, Polytechnic University of the Marche Region, Ancona, Italy, <sup>2</sup> Oncology Unit, Macerata Hospital, Macerata, Italy, <sup>3</sup> Division of Oncology, S. Orsola-Malpighi Hospital, Bologna, Italy, <sup>4</sup> Department of Pathology and Laboratory Medicine, Indiana University School of Medicine, Indianapolis, IN, United States, <sup>5</sup> Department of Pathology and Surgery, Faculty of Medicine, Cordoba, Spain

**Keywords:** microbiome, renal cell carcinoma, immunotherapy, resistance, PD-1 blockade, antibiotic therapy, prostate cancer

## INTRODUCTION

Every individual is characterized by a specific “enterotype,” based on the major components of her/his microbiome (i.e., collection of host and microorganism genomes and environmental conditions in an ecosystem) of the gut influenced by diet and geography. This is also influenced by the effects of the organisms present in the infancy as well as the type and pattern of the individual immune system (1).

In the last few years, a close biological relationship has emerged among the microbiome of the gut, the metabolism of the body, as well as the immune system including cancer development. With the increasing availability of high-throughput sequencing, single-cell transcriptomics, and mass spectrometry for a very precise characterization of single enteric, neoplastic, and immune cells, and more extensive databases of organisms already sequenced, experimental exploration of this network has become possible. These advances have also included “culturomics” to make an ever-expanding portion of the microbiota investigable, and sophisticated bioinformatics implements in order to achieve data deconvolution and combination. In 2008, the Human Microbiome Project (HMP) started to characterize the microbial communities from 300 healthy individuals, providing one of the broadest microbial genome databases targeting different body sites: nares, oral cavity, skin, gastrointestinal tract, breast, and urogenital tract (2, 3) (**Figure 1**).

## MICROBIOME AND CANCER

In 2017, many experimental studies had been published to demonstrate the importance of single bacterial species on the intestine, the individual immune response and cancer progression, and response to therapy. One of the first unexpected pieces of evidence was that secondary tumor deposits in patients with colorectal cancers include bacteria, such as *Fusobacterium* species, including *Bacteroides*, *Prevotella*, and *Selenomonas* species, and its associated microbiome. These findings demonstrated the microbiome stability between paired primary and metastatic tumors. Antibiotic treatment of *Fusobacterium*-positive colon cancers mice-xenografts reduces tumor growth, cancer cell proliferation along with *Fusobacterium* load, which favors the hypothesis that *Fusobacterium* species is associated with neoplastic progression (5, 6).

## OPEN ACCESS

### Edited by:

Ja Hyeon Ku,  
Seoul National University, South Korea

### Reviewed by:

Sergio Bracarda,  
Azienda Ospedaliera S.Maria, Italy

### \*Correspondence:

Alessia Cimadamore  
alessiacimadamore@gmail.com  
Rodolfo Montironi  
r.montironi@univpm.it

### Specialty section:

This article was submitted to  
Genitourinary Oncology,  
a section of the journal  
Frontiers in Oncology

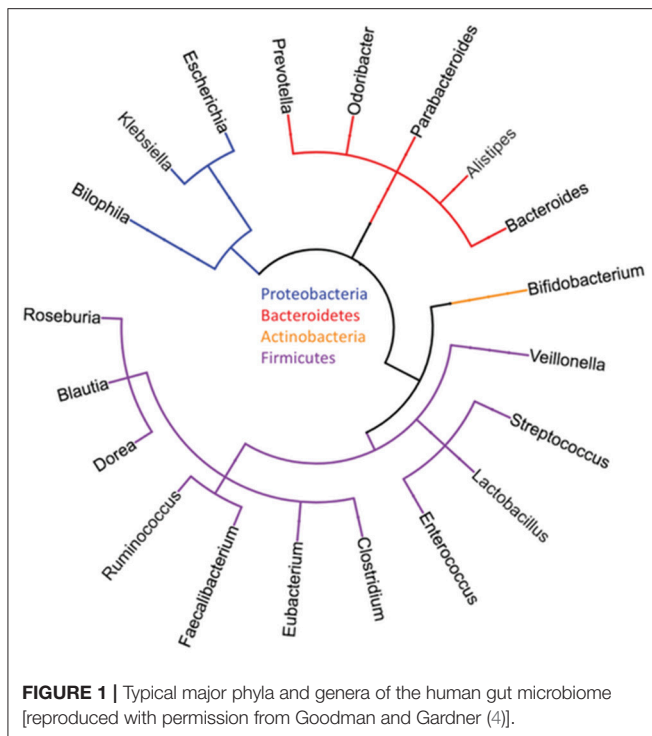
**Received:** 13 August 2018

**Accepted:** 04 March 2019

**Published:** 26 March 2019

### Citation:

Cimadamore A, Santoni M, Massari F,  
Gasparrini S, Cheng L,  
Lopez-Beltran A, Montironi R and  
Scarpelli M (2019) Microbiome and  
Cancers, With Focus on Genitourinary  
Tumors. *Front. Oncol.* 9:178.  
doi: 10.3389/fonc.2019.00178



In breast cancer, bacterium *Methylobacterium radiotolerans* was found relatively enriched compared to normal adjacent tissue from the same patient. Furthermore, bacterial DNA load was reduced in cancer samples vs. healthy tissue and correlated inversely with advanced disease (7, 8). In the distal esophagus, the impact of the microbiome in the pathogenesis of reflux-related disorders and in the development of intestinal metaplasia is well-demonstrated. Patients with esophagitis and Barrett's esophagus have a greater proportion of gram-negative anaerobes/microaerophiles with respect to the normal controls. This altered microbiome may promote Barrett's metaplasia and progression to adenocarcinoma (9, 10). The compositions of bacteria community and the throat biodiversity in laryngeal carcinoma patients compared to a control population were different and might be a risk factor for laryngeal carcinoma (11).

The most clinical-affecting evidence regarding cancer microbiome is its contribution to therapy resistance. In pancreatic cancer the most common species identified belong to the Enterobacteriaceae and Pseudomonadaceae families. Enterobacteriaceae express a bacterial enzyme cytidine deaminase (CDD) isoform that confer resistance to gemcitabine. Supporting this, co-treatment with the antibiotic ciprofloxacin abrogate the gemcitabine resistance in colon cancer mouse models (12).

On the other side, there is evidence that corroborates the hypothesis of a protection role of microbiome toward neoplastic changes. Hence, results show that individuals with microbiota linked to a plant diet are the ones with a lower incidence of cancer of the colon (13). Such a diet stimulates bacteria to produce short-chain fatty acids (SCFAs), particularly butyrate, propionate, and acetate. These fatty acids show an anti-inflammatory property

through the induction of T-regulatory cells of colonic tissues. Connections of microbiome, production of short-chain fatty acids, and the immune system become more interesting when researchers started to explore the influence of the microbiome in relation to immunotherapy drugs response.

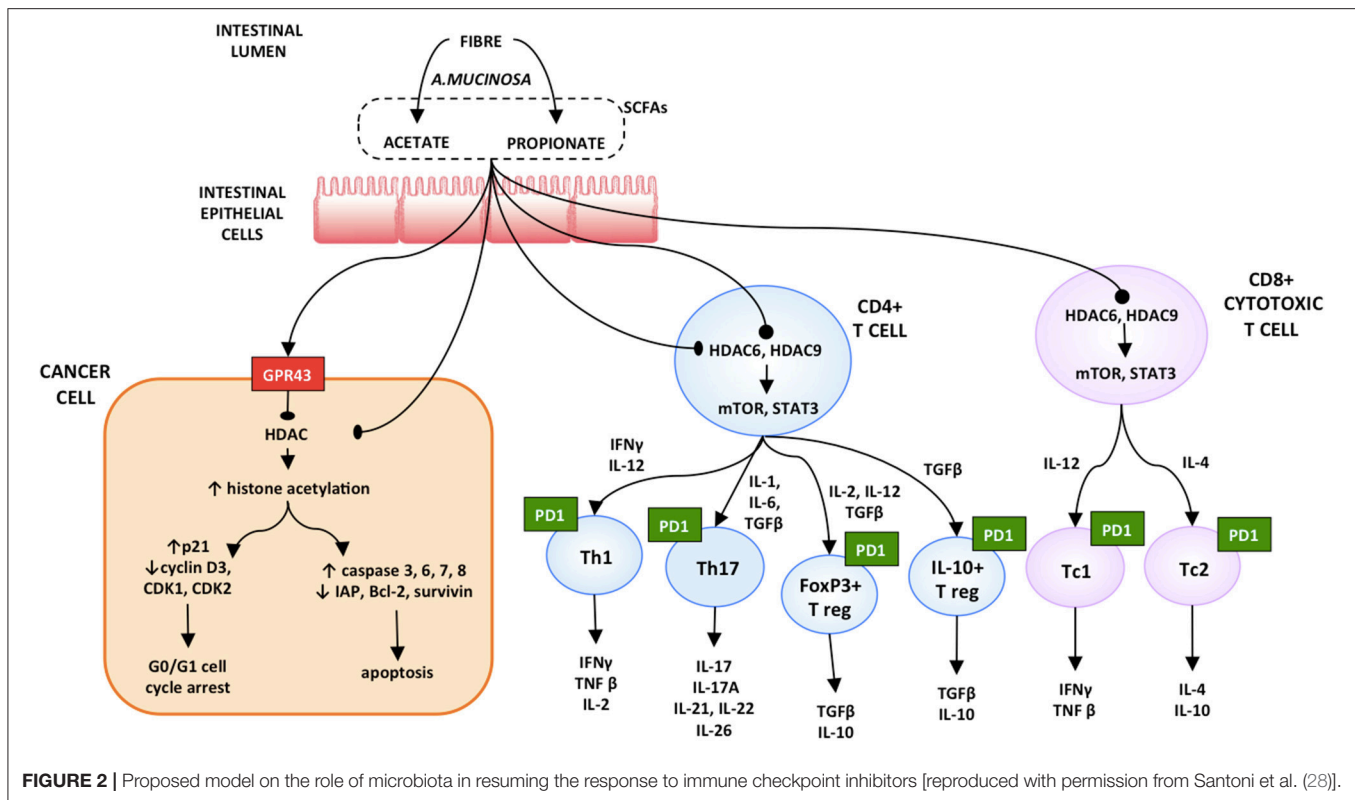
## MICROBIOME AND IMMUNOTHERAPY DRUGS RESPONSE

Immunotherapy based on and programmed death-ligand 1 (PD-L1)- and programmed death 1 (PD-1)-targeted antibodies has profoundly modified the prognostic and therapeutic landscape for many types of tumors, with demonstrated efficacy against renal cell carcinoma (RCC), non-small cell lung cancer (NSCLC), and melanoma. PD-L1 tissue expression is a poor prognostic factor as well as a predictor of good responses from both PD-1 and PD-L1 inhibitors in urothelial carcinoma (UC) and RCC (14). In a recent meta-analysis on the expression of PD-1 and PD-L1 in solid tumors, as a predictive biomarker of benefit from PD-1/PD-L1 axis inhibitors, odds ratios of objective response in PD-L1-positive patients compared with PD-L1-negative patients was 2.34 for RCC and 2.20 for bladder cancer (15). Liu et al. also confirmed that "patients with higher ratios of PD-L1-positive cells responded significantly better to both PD-1 and PD-L1 antibodies than those with lower ratios of PD-L1-positive cells" (16).

Each PD-1/PD-L1 drug approved by FDA is associated with a PD-L1, a immunohistochemistry (IHC)-based tissue assay. IHC-based PD-L1 assay is basically utilized to potentially predict the response to anti-PD-1 or/and anti-PD-L1 therapies. A fraction of patients with a negative IHC assay can show a response. This means that identification and utilization of other biomarkers is of great importance for a better selection of patients who might respond to such therapies.

Primary resistance to Immune checkpoint inhibitors (ICIs) has been linked to different factors, including poor intrinsic antigenicity of malignant cells, lack of priming by potentially immunogenic pretreatment with radio-, or/and chemotherapy (17), poor antigen presentation at the time of the priming phase (18), immunosuppression exerted locally by extracellular metabolites (19), and functional exhaustion of lymphocytes infiltrating the tumor (20, 21). On the contrary, high mutational burden and high immunogenic antigenicity of malignant cells are in favor of a better response to ICIs (22, 23). Recently, Routy et al. demonstrated that abnormal gut microbiome composition could have an influence on primary resistance to PD-1 blockade in mice xenografts and patients with cancer. In particular, they showed that the clinical benefit of ICIs in patients with cancer at an advanced stage is inhibited by antibiotic therapy (ATB) (24). They tested the effect of ATB on patients with advanced UC, RCC, or NSCLC, who had received PD-1/PD-L1mAb following one or several previous therapies. Progression-free survival (PFS) and overall survival (OS) were significantly shorter in the ATB-treated cohort when either all patients were combined together or when individual cancer types were investigated. In univariate and multivariate analyses, ATB was a





predictor factor for resistance to PD-1 blockade, not dependent from traditional prognostic markers in RCC and NSCLC. To evaluate the composition of the microbiota of the gut, they used quantitative metagenomics with analysis of the data in a reference catalog of 9.9 million genes. The greatest richness of the samples, analyzed at the levels of metagenomic species (MGS) and gene count, was correlated with the clinical response. This was defined by the lack of progression of disease 6 months following the initiation of ICIs.

*Akkermansia muciniphila* (*A. muciniphila*) was the commensal associated most significantly with excellent clinical outcomes in both RCC and NSCLC. When analyzing memory T cell responses from the peripheral blood of the patients, stimulated against microbiota following initiation of PD-1 blockade, the only immune response linked with the clinical benefit at the time of immunotherapy was the Th1 and Tc1 cell reactivity against *A. muciniphila*. This immunomodulatory effect might be explained by the production of SCFAs, such as propionate and acetate, by *A. muciniphila*. These short-chain fatty acids are ligands of the two orphan G-protein-coupled receptors 41 and 43 (GPR41 and GPR43). The former regulates the tumor cells apoptosis induced by SCFA and so exerts a tumor suppressor activity. Furthermore, propionate produced by the bacterium inhibits histone deacetylases and thus increases the histone hyperacetylation.

The inhibition of the expression of Histone Deacetylases (HDACs) has several effects, ranging from a pro-apoptotic activity to a pro-inflammatory response. By opening cell

chromatin and thus increasing the DNA accessibility to transcription factors, the histone hyperacetylation induces overexpression of caspases 6, 7, and 8, including caspase 3, and reduces the inhibitor of apoptosis (IAP) family expression (25). The inhibition of expression the HDACs activates the mTOR-S6K and STAT3 pathways. All this stimulates Th17, Th1, FoxP3+, and IL-10+ T cells, as well as the production of IL-10, IFN- $\gamma$ , and IL-17 in CD8+ T cells in both Tc1- and Tc17-cell subsets (26). Moreover, propionate promotes T-cell migration by increasing the expression of intercellular adhesion molecule 1 (ICAM-1) and E-selectin on endothelial cells (27, 28) (**Figure 2**).

Derosa et al. demonstrated the effect of ATB in patients with RCC and NSCLC treated with anti-PDL1 mAb monotherapy or combination therapy. In patients with RCC, ATB compared with no ATB was linked with an increased risk of progressive disease, shorter PFS, and shorter OS. Similar rates were also obtained in the NSCLC cohort (29). Researchers are now planning to transfer fecal bacteria from patients who respond to treatment with checkpoint inhibitors into the intestine of non-responder patients. This process is currently called “fecal microbiome transplant.” Microbiota composition might also be manipulated by the application of foods and prebiotics. The prevalence of a subspecies selected by diet rather than others could modify the population predisposition to a specific disease and the response to therapy of cancer patients (30). Fecal microbiota transplant, although not being probiotic, could be considered a fermented food, given the microbes, and nutrients present.

**TABLE 1 |** Metagenomic human studies identifying microbiota associated with cancer tissues [reproduced with permission from Goodman and Gardner (4)].

Tissue type	Species differential	References
Colorectal cancer	<i>Fusobacterium</i> , <i>Selenomonas</i> , and <i>Leptotrichia</i> species increased in cancer tissues	(5, 6)
Breast cancer	<i>Alistipes</i> , <i>Sphingomonas</i> , and <i>Methylobacterium</i> increased in cancer tissue	(7, 8)
Esophageal cancer	<i>Streptococcus</i> , <i>Prevotella</i> , and <i>Veillonella</i> species increased in cancer tissues	(9, 10)
Head and neck cancer	<i>Fusobacterium</i> , <i>Prevotella</i> , and <i>Gemella</i> species increased in cancer tissues; <i>Streptococcus</i> and <i>Rothia</i> species decreased in cancer tissues	(11)
Pancreatic cancer	Enterobacteriaceae, Pseudomonadaceae, Moraxellaceae, and Enterococcaceae increased in cancer tissues	(12)
Prostate cancer	<i>Propionibacterium acnes</i> increased in cancer tissues	(32–37)

## MICROBIOME IN BLADDER AND PROSTATE CANCER

Of great interest is the urinary microbiota profile investigated by Wu et al. They analyzed DNA from urine pellet collected from male patients with urothelial carcinoma and non-neoplastic controls. They observed enrichment of some bacterial genera (such as *Sphingobacterium*, *Anaerococcus*, and *Acinetobacter*) and decrease of others (such as *Roseomonas*, *Proteus*, and *Serratia*) in the group with cancer in comparison with the control group. Patients with high risk of recurrence and progression had an enrichment of *Herbaspirillum*, *Porphyrobacter*, and *Bacteroides*. This means these bacteria can be considered as potential biomarkers in risk stratification (31).

In the last year, interesting results have emerged by investigations on the microbiome in PCa patients. The microflora of tumor, peri-tumor, and benign prostate tissue samples have recently been characterized by massive ultradeep pyrosequencing. Interestingly, differences in microbial populations among paired tumor/peri-tumor and non-tumor prostate tissues have been detected. This finding generates the hypothesis that the distribution of bacterial microbes varies according to the nature of tissue within the same gland. This suggests a pathophysiological association between the local microbial niche and composition, and the tumor itself (32, 33) (Table 1).

A case-control pilot study has been conducted by Golombos et al. to demonstrate the impact of the gut microbiota on PCa pathogenesis. They performed a computational genomics analysis on stool samples of men with benign prostatic conditions

and men with intermediate or high risk clinically localized PCa. Biologically significant abundance differences of bacteria species and 23 metabolic differentially abundant pathways were identified between the two cohorts (34). Likewise, analyses on the urinary microbiome showed a prevalence of uropathogens and pro-inflammatory bacteria differentially abundant in PCa patients compared to healthy subjects in urine collected from men prior to biopsy for PCa (35).

Liss et al. developed a microbiome-derived risk profile for PCa, derived from altered metabolic pathways, comparing the taxonomic composition of samples (64 with PCa and 41 without) of rectal swab collected 2 weeks before prostate biopsy (36). Even though the differences between the two groups are not impressive, these results are hypothesis-generating and pave the way to further evaluate the manipulation of aberrant microbiomes to reduce PCa risk (3).

The composition of the microbiota in the gut is influenced by oral androgen receptor axis-targeted therapies (ATT) in prostate cancer patients. Results on fecal microbiota profile shows the abundance of species linked to response to anti-PD-1 immunotherapy, including *Ruminococcaceae* spp., and *A. muciniphila*, and an greater representation of bacterial gene pathways that are involved in steroid biosynthesis as well as steroid hormone biosynthesis in the fecal microbiota of men under ATT (4, 37). Additional studies are needed to evaluate whether the gut microbiota can influence clinical responses to ATT, and modulate the anticancer effects of future therapies, including immunotherapy.

As regards to genitourinary tumors, there are only few trials ongoing (38, 39). One to take into consideration is a prospective study on prostate cancer and breast cancer patients who are undergoing two different standards of care radiation regimens. Exposure to radiation can impact immune cells that are present in the blood as well as the underlying microbiota. The aim of this study is to study microbial changes and how these changes correlate with alteration in immune mediators (i.e., lymphocytes, cytokines) present in blood samples before, during, and after radiation, by collecting stool specimens at baseline, end of radiation therapy and during the follow up (ClinicalTrials.gov Identifier: NCT03383107).

## CONCLUSIONS

The host and the microbiota share a complex balanced relationship that can be overthrown in a state of dysbiosis consequential to environmental changes. Alteration of this balance could lead to promotion of inflammatory diseases and cancer. There is evidence showing that the activity of microbiota in the restoration of response to immune checkpoint inhibitors involves both the immune and cancer cells. Stimulating recall Th1 responses against *A. muciniphila* improves immunosurveillance in cancer patients. Microbiome composition has the potential to be a novel biomarker of response to ICIs and a therapeutic opportunity for unresponsive patients. In patients with RCC, antibiotic therapy was linked to an increased risk of progressive disease, shorter PFS, and shorter

OS. Pioneer studies on bladder and prostate cancer patients' microbiome pave the way to the investigation of a possible novel prognostic, diagnostic, and therapeutic tool.

## AUTHOR CONTRIBUTIONS

RM and MSc conception and design. AC drafting the manuscript. FM, MSa and SG review of the literature. LC and AL-B critical revision of the manuscript.

## REFERENCES

- Arumugam M, Raes J, Pelletier E, Le Paslier D, Yamada T, Mende DR, et al. Enterotypes of the human gut microbiome. *Nature*. (2011) 473:174–80. doi: 10.1038/nature09944
- Lloyd-Price J, Mahurkar A, Rahnavard G, Crabtree J, Orvis J, Hall AB, et al. Strains, functions and dynamics in the expanded human microbiome project. *Nature*. (2017) 550:61–6. doi: 10.1038/nature23889
- Cavarretta I, Mancini N, Salonia A. Analysis of the enteric microbiome: first tentative steps towards a comprehensive work-up of prostate cancer? *Eur Urol*. (2018). 74:583–584. doi: 10.1016/j.eururo.2018.07.009
- Goodman B, Gardner H. The microbiome and cancer. *J Pathol*. (2018) 244:667–76. doi: 10.1002/path.5047
- Bullman S, Pedamallu CS, Sicinska E, Clancy TE, Zhang X, Cai D, et al. Analysis of *Fusobacterium* persistence and antibiotic response in colorectal cancer. *Science*. (2017) 358:1443–8. doi: 10.1126/science.aal5240
- Kostic AD, Gevers D, Pedamallu CS, Michaud M, Duke F, Earl AM, et al. Genomic analysis identifies association of *Fusobacterium* with colorectal carcinoma. *Genome Res*. (2012) 22:292–8. doi: 10.1101/gr.126573.111
- Xuan C, Shamonki JM, Chung A, Dinome ML, Chung M, Sieling PA, et al. Microbial dysbiosis is associated with human breast cancer. *PLoS ONE*. (2014) 9:e83744. doi: 10.1371/journal.pone.0083744
- Chan AA, Bashir M, Rivas MN, Duvall K, Sieling PA, Pieber TR, et al. Characterization of the microbiome of nipple aspirate fluid of breast cancer survivors. *Sci Rep*. (2016) 6:28061. doi: 10.1038/srep28061
- Yang L, Lu X, Nossa CW, Francois F, Peek RM, Pei Z. Inflammation and intestinal metaplasia of the distal esophagus are associated with alterations in the microbiome. *Gastroenterology*. (2009) 137:588–97. doi: 10.1053/j.gastro.2009.04.046
- Snider EJ, Freedberg DE, Abrams JA. Potential role of the microbiome in barrett's esophagus and esophageal adenocarcinoma. *Dig Dis Sci*. (2016) 61:2217–25. doi: 10.1007/s10620-016-4155-9
- Gong HL, Shi Y, Zhou L, Wu CP, Cao PY, Tao L, et al. The composition of microbiome in larynx and the throat biodiversity between laryngeal squamous cell carcinoma patients and control population. *PLoS ONE*. (2013) 8:e66476. doi: 10.1371/journal.pone.0066476
- Geller LT, Barzily-Rokni M, Danino T, Jonas OH, Shental N, Nejman D, et al. Potential role of intratumor bacteria in mediating tumor resistance to the chemotherapeutic drug gemcitabine. *Science*. (2017) 357:1156–60. doi: 10.1126/science.aah5043
- Smith PM, Howitt MR, Panikov N, Michaud M, Gallini CA, Bohlooly-Y M, et al. The microbial metabolites, short-chain fatty acids, regulate colonic Treg cell homeostasis. *Science*. (2013) 341:569–73. doi: 10.1126/science.1241165
- Mann SA, Lopez-Beltran A, Massari F, Pili R, Fiorentino M, Koch MO, et al. Targeting the programmed cell death-1 pathway in genitourinary tumors: current progress and future perspectives. *Curr Drug Metab*. (2017) 18:700–11. doi: 10.2174/1389200218666170518162500
- Khunger M, Hernandez AV, Pasupuleti V, Rakshit S, Pennell N, Stevenson JG, et al. Programmed cell death 1 (PD-1) ligand (PD-L1) expression in solid tumors as a predictive biomarker of benefit from PD-1/PD-L1 axis inhibitors: a systematic review and meta-analysis. *JCO Precision Oncol*. (2017) 1:1, 1–15. doi: 10.1200/PO.16.00030
- Liu J, Zhang C, Hu J, Tian Q, Wang X, Gu H, et al. Effectiveness of anti-PD-1/PD-L1 antibodies in urothelial carcinoma patients with different

## FUNDING

The authors have no relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript. This includes employment, consultancies, honoraria, stock ownership or options, expert testimony, grants, or patents received or pending, or royalties. No writing assistance was utilized in the production of this manuscript.

- PD-L1 expression levels: a meta-analysis. *Oncotarget*. (2018) 9:12400–7. doi: 10.18632/oncotarget.24249
- Carbone DP, Reck M, Paz-Ares L. First-line nivolumab in stage iv or recurrent non-small-cell lung cancer. *N Engl J Med*. (2017) 376:2415–26. doi: 10.1056/NEJMoa1613493
  - Schumacher TN, Schreiber RD. Neoantigens in cancer immunotherapy. *Science*. (2015) 348:69–74. doi: 10.1126/science.aaa4971
  - Spranger S, Bao R, Gajewski TF. Melanoma-intrinsic  $\beta$ -catenin signalling prevents anti-tumour immunity. *Nature*. (2015) 523:231–5. doi: 10.1038/nature14404
  - Smyth MJ, Ngiew SF, Ribas A, Teng MW. Combination cancer immunotherapies tailored to the tumour microenvironment. *Nat Rev Clin Oncol*. (2016) 13:143–58. doi: 10.1038/nrclinonc.2015.209
  - Koyama S, Akbay EA, Li YY, Herter-Sprie GS, Buczkowski KA, Richards WG, et al. Adaptive resistance to therapeutic PD-1 blockade is associated with upregulation of alternative immune checkpoints. *Nat Commun*. (2016) 7:10501. doi: 10.1038/ncomms10501
  - Rizvi NA, Hellmann MD, Snyder A. Cancer immunology. Mutational landscape determines sensitivity to PD-1 blockade in non-small cell lung cancer. *Science*. (2015) 348:124–8. doi: 10.1126/science.aaa1348
  - Riaz N, Havel JJ, Kendall SM, Makarov V, Walsh LA, Desrichard A, et al. Recurrent SERPINB3 and SERPINB4 mutations in patients that respond to Anti-CTLA4 immunotherapy. *Nat Genet*. (2016) 48:1327–9. doi: 10.1038/ng.3677
  - Routy B, Le Chatelier E, Derosa L, Duong CPM, Alou MT, Daillère R, et al. Gut microbiome influences efficacy of PD-1-based immunotherapy against epithelial tumors. *Science*. (2018) 359:91–7. doi: 10.1126/science.aan3706
  - Tang Y, Chen Y, Jiang H, Robbins GT, Nie D. G-protein-coupled receptor for short-chain fatty acids suppresses colon cancer. *Int J Cancer*. (2011) 128:847–56. doi: 10.1002/ijc.25638
  - Park J, Kim M, Kang SG, Jannasch AH, Cooper B, Patterson J, et al. Short-chain fatty acids induce both effector and regulatory T cells by suppression of histone deacetylase and regulation of the mTOR-S6K pathway. *Mucosal Immunol*. (2015) 8:80–93. doi: 10.1038/mi.2014.44
  - Miller SJ, Zaloga GP, Hoggatt AM, Labarrere C, Faulk WP. Short-chain fatty acids modulate gene expression for vascular endothelial cell adhesion molecules. *Nutrition*. (2005) 21:740–8. doi: 10.1016/j.nut.2004.11.011
  - Santoni M, Piva F, Conti A, Santoni A, Cimadamore A, Scarpelli M, et al. Re: Gut microbiome influences efficacy of PD-1-based immunotherapy against Epithelial Tumors. *Eur Urol*. (2018) 74:521–2. doi: 10.1016/j.eururo.2018.05.033
  - Derosa L, Hellmann MD, Spaziano M, Halpenny D, Fidelle M, Rizvi H, et al. Negative association of antibiotics on clinical activity of immune checkpoint inhibitors in patients with advanced renal cell and non-small-cell lung cancer. *Ann Oncol*. (2018) 29:1437–44. doi: 10.1093/annonc/mdy103
  - Reid G. Microbes in food to treat and prevent disease. *Exp Rev Precision Med Drug Dev*. (2018) 3:79–81. doi: 10.1080/23808993.2018.1429217
  - Wu P, Zhang G, Zhao J, Chen J, Chen Y, Huang W, et al. Profiling the urinary microbiota in male patients with bladder cancer in China. *Front Cell Infect Microbiol*. (2018) 8:167. doi: 10.3389/fcimb.2018.00167
  - Cavarretta I, Ferrarese R, Cazzaniga W, Saita D, Lucianò R, Ceresola ER, et al. The microbiome of the prostate tumor microenvironment. *Eur Urol*. (2017) 72:625–31. doi: 10.1016/j.eururo.2017.03.029



33. Fassi Fehri L, Mak TN, Laube B, Brinkmann V, Ogilvie LA, Mollenkopf H, et al. Prevalence of *Propionibacterium acnes* in diseased prostates and its inflammatory and transforming activity on prostate epithelial cells. *Int J Med Microbiol.* (2011) 301:69–78. doi: 10.1016/j.ijmm.2010.08.014
34. Golombos DM, Ayangbesan A, O'Malley P, Lewicki P, Barlow L, Barbieri CE, et al. The role of gut microbiome in the pathogenesis of prostate cancer: a prospective, pilot study. *Urology.* (2018) 111:122–8. doi: 10.1016/j.urology.2017.08.039
35. Shrestha E, White JR, Yu SH, Kulac I, Ertunc O, De Marzo AM, et al. Profiling the urinary microbiome in men with positive versus negative biopsies for prostate cancer. *J Urol.* (2018) 199:161–71. doi: 10.1016/j.juro.2017.08.001
36. Liss MA, White JR, Goros M, Gelfond J, Leach R, Johnson-Pais T, et al. Metabolic biosynthesis pathways identified from fecal microbiome associated with prostate cancer. *Eur Urol.* (2018) 74:575–82. doi: 10.1016/j.eururo.2018.06.033
37. Sfanos KS, Sauvageot J, Fedor HL, Dick JD, De Marzo AM, Isaacs WB. A molecular analysis of prokaryotic and viral DNA sequences in prostate tissue from patients with prostate cancer indicates the presence of multiple and diverse microorganisms. *Prostate.* (2008) 68:306–20. doi: 10.1002/pros.20680
38. Markowski MC, Boorjian SA, Burton JP, Hahn NM, Ingersoll MA, Maleki Vareki S, et al. The microbiome and genitourinary cancer: a collaborative review. *Eur Urol.* (2019). 75, 637–646. doi: 10.1016/j.eururo.2018.12.043
39. Sfanos KS, Markowski MC, Peiffer LB, Ernst SE, White JR, Pienta KJ, et al. Compositional differences in gastrointestinal microbiota in prostate cancer patients treated with androgen axis-targeted therapies. *Prostate Cancer Prostatic Dis.* (2018) 21:539–48. doi: 10.1038/s41391-018-0061-x

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Cimadamore, Santoni, Massari, Gasparrini, Cheng, Lopez-Beltran, Montironi and Scarpelli. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.



# The Identification of Immunological Biomarkers in Kidney Cancers

Antonio Lopez-Beltran<sup>1\*</sup>, Vanessa Henriques<sup>2</sup>, Alessia Cimadamore<sup>3</sup>, Matteo Santoni<sup>4</sup>, Liang Cheng<sup>5</sup>, Thomas Gevaert<sup>6,7</sup>, Ana Blanca<sup>8</sup>, Francesco Massari<sup>9</sup>, Marina Scarpelli<sup>3</sup> and Rodolfo Montironi<sup>3</sup>

<sup>1</sup> Department of Pathology and Surgery, Faculty of Medicine, Cordoba University, Cordoba, Spain, <sup>2</sup> Pathology Service, Champalimaud Clinical Center, Lisbon, Portugal, <sup>3</sup> Section of Pathological Anatomy, United Hospital, School of Medicine, Polytechnic University of the Marche Region, Ancona, Italy, <sup>4</sup> Oncology Unit, Macerata Hospital, Macerata, Italy, <sup>5</sup> Department of Pathology and Laboratory Medicine, Indiana University School of Medicine, Indianapolis, IN, United States, <sup>6</sup> Laboratory of Experimental Urology, Organ Systems, KU Leuven, Leuven, Belgium, <sup>7</sup> Department of Pathology, AZ Kline, Brasschaat, Belgium, <sup>8</sup> Instituto Maimonides de Investigación Biomédica de Córdoba, Córdoba, Spain, <sup>9</sup> Division of Oncology, S. Orsola-Malpighi Hospital, Bologna, Italy

## OPEN ACCESS

### Edited by:

Fabio Grizzi,  
Humanitas Research Hospital, Italy

### Reviewed by:

Elena Ranieri,  
University of Foggia, Italy  
Sanja Štifter,  
University of Rijeka, Croatia

### \*Correspondence:

Antonio Lopez-Beltran  
em1lobea@uco.es

### Specialty section:

This article was submitted to  
Genitourinary Oncology,  
a section of the journal  
Frontiers in Oncology

**Received:** 13 August 2018

**Accepted:** 01 October 2018

**Published:** 02 November 2018

### Citation:

Lopez-Beltran A, Henriques V,  
Cimadamore A, Santoni M, Cheng L,  
Gevaert T, Blanca A, Massari F,  
Scarpelli M and Montironi R (2018)  
The Identification of Immunological  
Biomarkers in Kidney Cancers.  
Front. Oncol. 8:456.  
doi: 10.3389/fonc.2018.00456

The recent approval of several agents have revolutionized the scenario of therapeutic management of metastatic renal cell carcinoma (RCC) allowing us to reach important clinical end points with extended patients' survival. Actually, every new drug approved has represented an important step forward to the improvement of patient's survival. On the other hand, we now understand that RCC includes a large group of tumor entities, each of them with different genetic and mutational alterations, but also showing different clinical behavior; a reason behind the needs of subtype specific personalized approach to therapy of RCC. Immunotherapy is gradually becoming a key factor in the therapeutic algorithm for patients with locally advanced or metastatic RCC. Due to the combination of potent treatment success and potentially deadly adverse effects from immune checkpoint inhibitors (ICI), gathering prognostic and predictive information about FDA-indicated tumors seems to be prudent. Robust and reliable biomarkers are crucial for patient's selection of treatments with immunomodulatory drugs. PD-L1 expression is a poor prognostic factor and predictive of better responses from both PD-1 and PD-L1 inhibitors in a variety of tumor types including RCC. Each FDA approved PD-1/PD-L1 drug is paired with a PD-L1 Immunohistochemistry (IHC) assay. Thus, there is need for improved knowledge and application of PD-1/PD-L1 IHC biomarkers in daily practice. IHC staining appears in membranous fashion. The atezolizumab approved IHC assay is unique in that only immune cell staining is quantified for the use of this assay in RCC. A single biomarker for patient selection may not be feasible, given that immune responses are dynamic and evolve over time. Biomarker development for ICI drugs will likely require integration of multiple biologic components like PD-L1 expression, TILs and mutational load. New methodological approaches based on digital pathology may be relevant since they will allow recognition of the biomarker and to objectively quantitate its expression, and therefore might produce objective and reproducible cut-off assessment. Multidisciplinary approach is very much needed to fully develop the current and future value of ICI in clinical practice.

**Keywords:** renal cell carcinoma, PD-L1, immunotherapy, RCC subtypes, immunological biomarker, predictive biomarker, tumor mutation load

## INTRODUCTION

The recent approval of several agents have revolutionized the scenario of therapeutic management of metastatic renal cell carcinoma (RCC) allowing us to reach important clinical end points with extended patients' survival (1).

The first generation of immune checkpoint inhibitors (anti-CTLA-4 and anti-PD-1/PD-L1) targeted natural immune homeostasis pathways to drive anti-tumor immune responses. These agents led to unprecedented results in patients with previously incurable metastatic disease and therefore became first-line therapies for some advanced cancers (2–12). Since these agents are efficacious in only a minority of patients, however, newer strategies are becoming available that target additional immunomodulatory mechanisms to activate patients' own anti-tumor immune responses. Emerging targets include co-inhibitory and co-stimulatory markers of the innate and adaptive immune system.

In this review, we will discuss: (1) Pathologic and molecular subtypes of RCC; (2) Current landscape of targeted therapy in renal cell carcinoma; (3) Overview of immunotherapy in renal cell carcinoma; (4) Predictive immunological biomarkers in renal cell carcinoma; (5) Gene expression as predictive biomarkers in renal cell carcinoma; (6) The current status of PD-L1 immunohistochemistry; (7) MMR-deficiency and mutational load in RCC; and (8) Biomarkers of acquired resistance. Finally, we briefly highlight likely future perspectives of predictive biomarkers of immunotherapy in RCC.

## PATHOLOGIC AND MOLECULAR SUBTYPES OF RENAL CELL CARCINOMA

Clear cell RCC (ccRCC) accounts for about 75% of kidney cancer while the other 25% are classified as non-clear cell renal cell carcinoma (nccRCC) (13). Over a dozen pathological subtypes are now recognized by the most recent World Health Organization classification of Tumors of the Urinary System and Male Genital Organs (13). These subtypes include papillary renal cell carcinoma (pRCC) (20%) and chromophobe renal cell carcinoma (chRCC) (5%), which are the most frequent nccRCC subtypes; hereditary leiomyomatosis and renal cell associated -carcinoma, collecting duct carcinoma, renal medullary carcinoma, MiT family translocation carcinoma, succinate dehydrogenase-deficient RCC, mucinous tubular and spindle cell carcinoma, tubulocystic RCC, Acquired cystic disease-associated RCC, clear cell papillary RCC, and RCC unclassified represent less common subtypes (13).

Several genomic changes have been found in ccRCC, mostly epigenetic reprogramming and oncogenic metabolism pathways alterations (13–18) with other common genetic changes in genes controlling cellular oxygen pathway (e.g., VHL) and the maintenance of chromatin structure (e.g., PBRM1) (19–22). TCGA analysis of a ccRCC cohort found similar genomic changes and reported recurrent alterations in the PI(3)K/AKT pathway and several epigenetic changes in DNA methylation (22). Molecular stratification of ccRCC revealed 2 different

subtypes: clear cell type A (ccA) and B (ccB), with ccA patients having a markedly better prognosis (23, 24). A second TCGA study focussed on papillary RCC (pRCC) and found that type 1 and type 2 pRCC are distinctly different diseases based on molecular features and that type 2 pRCC is a heterogeneous disease with at least three different subgroups (25). A third TCGA project focussed on the chromophobe RCC (ChRCC) and found gene expression changes related to mitochondrial function and recurrent structural breakpoints within TERT promoter region (26). Recently, a multilevel molecular characterization of the 3 TCGA RCC databases revealed nine major genomic RCC subtypes, each one being distinct in terms of altered pathways and patient survival (16). Overlapping and subtype-specific genomic changes were observed, and good correlation with histologic subtypes was noticed. These molecular classes show substantial molecular diversity represented within each major histologic type, but importantly, actionable alterations also included PI3K and immune checkpoint pathways (16).

## CURRENT LANDSCAPE OF TARGETED THERAPY IN RENAL CELL CARCINOMA

The better knowledge of molecularly altered pathways of RCC has led to the development of new classes of drugs rising the targeting therapy era (1, 12, 16–23, 27). Angiogenesis, the hallmark of RCC, is the final target of several TKi (Sunitinib, Axitinib, Sorafenib and pazopanib) (1–12). After angiogenesis, the finding that, the deregulation of the PI3K–Akt–mTOR pathway, activated at different levels of the signaling cascade, drives RCC progression has led to the development of the mTOR inhibitors everolimus and temsirolimus. The association between everolimus and lenvatinib (a VEGFR1, VEGFR2 and VEGFR3 FGFR1, FGFR2, FGFR3, FGFR4, PDGFR, RET and KIT inhibitor) has been recently explored in a phase II clinical trial which demonstrated a better progression free survival (PFS) for patients receiving the combination of these two drugs compared to those who received everolimus monotherapy (10). Recently, also the mesenchymal-epithelial transition and multi-tyrosine kinases inhibitor cabozantinib has been included in clinical practice (1–12).

These drugs have led to an improvement in overall survival (OS) (sunitinib, pazopanib, cabozantinib, temsirolimus) and PFS (sunitinib, axitinib, cabozantinib, sorafenib, pazopanib, everolimus and temsirolimus) showing a safety profile with a remarkable clinical activity in a disease which has always been poor of active treatments (12, 15).

## OVERVIEW OF IMMUNOTHERAPY IN RENAL CELL CARCINOMA

Targeting drugs have significantly changed the course of RCC, but it's likely that a new classes of agents, the immune-checkpoint inhibitors (ICI), are destined to feed this new paradigm in RCC treatment (12, 28–51).

Programmed Death Receptor 1/Programmed Death Receptor Ligand 1 (PD 1/PD-L1) and Cytotoxic T Lymphocytes Antigen 4



(CTLA-4) inhibitors are agents able to target specific pathways related to immune-response which are often hyper-activated by tumor cell interaction (46). By inhibition of these targets, ICI could reactivate a specific immune response against tumor cells (**Figure 1**) (52). The observation that, RCC is related to a high mutation burden and so maybe to a high antigens expression, has led to test these drugs in different stages of the disease. Checkmate 025 was the first large phase III clinical trials comparing the PD-1 inhibitor nivolumab to everolimus in patients with locally advanced or metastatic RCC progressed to at least one VEGF/VEGFR inhibitor (11). This study met its primary endpoints showing an OS benefit in patients receiving nivolumab. Furthermore, patients treated with immunotherapy showed a higher overall response rate (ORR) compared to everolimus with an important percentage of patients achieving long lasting response (11). It is not surprising that the important results achieved in this trial have move to explore immunotherapy in other setting, such as adjuvant/neo-adjuvant stage and as first line therapy (12). Two different strategies been adopted: (1) the combination between an immune-checkpoint inhibitor and a VEGF inhibitor has been evaluated in a phase II trials. Indeed, in Immotion150 305 patients with locally advanced/mRCC and untreated RCC were randomized to receive: atezolizumab (an anti PD-L1 inhibitor) and bevacizumab, atezolizumab alone or sunitinib (41). The association arm resulted in a longer PFS compared to atezolizumab (6.1 months) and sunitinib arms with a higher percentage of ORR in combination arm (41). Of note, patients with PD-L1 positive expression ( $\geq 1\%$ ) showed a longer PFS (14.7 months) and higher ORR (46%) in atezolizumab arm; (14) and (2) the combination between two immune-checkpoint inhibitors have been recently tested in a large phase III trial: The Checkmate 214. In this study patients were randomized to receive the nivolumab (anti PD-1) and Ipilimumab (Anti CTLA 4) combination or sunitinib as first line therapy (42). In ESMO 2017, Escudier et al. (42) presented primary results after 17.5 months of follow up showing that the combination between ipilimumab-nivolumab resulted in higher ORR and complete response rate in intermediate/poor risk patients. Of note, patients with intermediate/poor risk disease and PD-L1 expression  $\geq 1\%$  showed higher ORR and PFS compared to sunitinib, while patients with favorable category of risk (showing lower PD-L1 expression) displayed a longer PFS and a higher ORR with sunitinib (42) (**Table 1**).

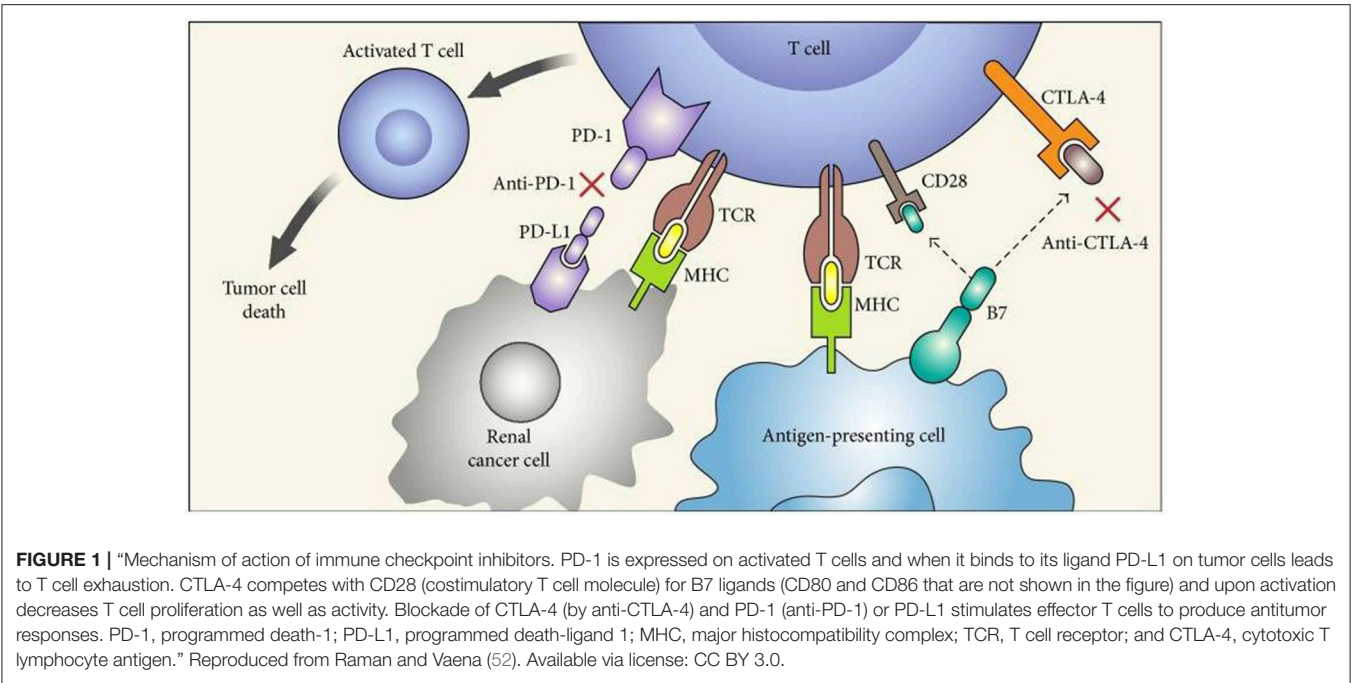
These encouraging results suggest that we are about to enter a new era for the management of metastatic RCC since the data provided from these trials might represent only the tip of the iceberg, and therefore, we could expect more therapeutic novelties to come. Nonetheless, even if immunotherapy provides a new hope for patients with metastatic RCC, the “old” targeting therapy is far from being abandoned. Indeed, Checkmate 025 showed that nivolumab is better than everolimus, but there are other agents showing to be extremely effective after VEGF/VEGFR inhibitors progression, thus, the decision of second line treatment should be weighted on the basis of the clinical outcome pursued as well as patient preference and toxicity profile (53). Though in first line setting immunotherapy

showed interesting results, it's probably that the positive effect could be restricted to patients with specific clinical features, such as intermediate/poor risk disease while patients with favorable profile could benefit from a standard therapeutic approach (46). It is probably that the worst clinical profile of the disease could be related to a high mutational burden of tumor cells, therefore resulting in a higher antigens expression. Preliminary data seems to indicate that these patients with high mutational load present a higher percentage of tumors with positive PD-L1 expression. Future studies will help us to better understand the role of PD-L1 as prognostic and predictive response factor since to date we have highly diverging information. Indeed, a meta-analysis of six published studies revealed that a higher level of PD-L1 expression increased the risk of death by representing therefore a negative prognostic factor (44). Differently to what was expected, the improved OS with nivolumab was not correlated with PD-L1 expression in Checkmate 025 while patients with positive PD-L1 expression seems to show more clinical benefit from immune-checkpoint inhibitors in Immotion150 and Checkmate 214 (**Table 2**).

## PREDICTIVE IMMUNOLOGICAL BIOMARKERS IN RENAL CELL CARCINOMA

Due to the increasing role of immunotherapy in clinical practice, the research of predictive response factors remains a critical but still unmet issue. A PD-L1 assessment on tumor cells through Dako PD-L1 IHC 28-8 pharmDx test was performed in both the Checkmate 025 and 214 trials. In Checkmate 025 nivolumab efficacy was not influenced by PD-L1 expression. However, patients expressing PD-L1 more than 1% ( $n = 181$ ) showed a worse OS in both treatment arms thus suggesting a prognostic role more than a predictive one. On the other hand, an exploratory analysis of the Checkmate214 showed a PFS benefit favoring combination only in patients expressing PD-L1 (1% or greater). Survival and ORR advantages were maintained in all PD-L1 categories. However, patients with higher PD-L1 expression showed greater benefit with the immune-combination. Taking together, these results seem to confirm that PD-L1 IHC expression does not act as predictor of response in patients with metastatic ccRCC receiving ICI immunotherapy (12, 15, 28, 46, 54). Furthermore, intratumoral eterogeneity of PD-L1 expression is another issue to take into consideration. As demonstrated by López et al. a multisite tumor sampling strategy identified a greater number of positive cases compared to current tumor sampling protocols and a different pattern of PD-L1 expression with positive and negative regions in the same tumor (55).

As seen in other neoplastic diseases in which immunotherapy has been successfully tested, tumor mutational burden and non-synonymous mutation expression have been related to higher neo-antigens tumor expression and to favorable immunotherapy response. A rationale supporting additional research of this variable in RCC derives from the evidence that immunotherapy is associated to higher clinical benefit in worst risk categories of



**TABLE 1** | Results obtained in selected trials exploring immune check point inhibitors in metastatic/locally advanced RCC using different combination of drugs.

Study name with experimental and comparator arms	Setting	N ITT	N PD-L1+	OS ITT	HR	OS PD-L1+	PFS ITT	HR	PFS PD-L1+	HR	ORR ITT	ORR PD-L1+	CR
IMMOTION150													
Atezolizumab + Bevacizumab	Untreated patients with locally advanced or metastatic renal cell carcinoma	101	164	NR	NR	NR	NR	NR	NR	NR	32%	46%	NR
Atezolizumab		103		NR		NR		NR		25%	28%	NR	
Sunitinib		101		NR		NR		NR		29%	27%	NR	
CHECKMATE 214													
Ipilimumab + Nivolumab	Untreated patients with locally advanced or metastatic renal cell carcinoma	550	204	NR	NR	NR	11.6	0.82	22.8*	0.48	NR	58%*	9.4%*
Sunitinib		546	224	NR	NR	NR	8.4		5.9*		NR	25%*	1.2%*

NR, not reported; PFS: progression free survival; OS, overall survival; ORR, overall response rate; CR, complete response. \*Intermediate/poor risk patients with PD-L1 expression ≥1%.

RCC, a clinical category of RCC in which high mutational load is present (30–32). Indeed, considering the subgroup analysis of the Checkmate 025 study and the significantly better results of nivolumab-ipilimumab combination in intermediate/poor risk patients in the Checkmate 214, it seems likely that tumors with worst clinical features are those that better respond to immune-checkpoint inhibitors and this may be due to a higher mutational load resulting in higher neo-antigen content. Unfortunately, differently than expected, mutational load does not seem to correlate with MSKCC or IMDC prognostic criteria (31). Moreover, no difference has been observed between clear cell and sarcomatoid components of different tumor samples,

suggesting that the level of mutational load is not a variable associated to worst clinical features of the disease, hypothesis that clearly needs further investigation (33). Concerning the correlation between mutational burden and response to ICI immunotherapy in ccRCC, de Velasco et al. carried out a whole exomes and transcriptomes sequencing of 9 patients with metastatic RCC receiving nivolumab. They discovered that RCC had relatively few non-synonymous mutations and neo-antigens and, surprisingly, that among patients receiving nivolumab non-synonymous mutations were significantly higher in non-responder patients ( $n = 6$ ) compared to responder patients ( $n = 3$ ) (34). Of note, they found a

**TABLE 2 |** Summary of assays and response rates in immune checkpoint inhibitor trials.

Drug		Antibody for PD-L1 IHC assay	Definition of PD-L1 positivity
Nivolumab (SA)	PD-1	Rabbit 28-8 (Dako)	PD-L1 $\geq 5\%$ (TC)
Atezolizumab (SA)	PD-L1	Rabbit SP142 (Ventana)	IHC 1/2/3 (IC)
Nivolumab/ Ipilimumab (C)	PD-1/CTLA-4	Rabbit 28-8 (Dako)	PD-L1 $\geq 1\%$ (TC)
Atezolizumab/ Bevacizumab (C)	PD-L1/antiVEGF	Rabbit SP142 (Ventana)	IHC 1/2/3 (IC)

Locally advanced or in metastatic renal cell carcinoma.

TC, tumor cells; IC, immune cells in the microenvironment; SA, single agent; C, combination of agents; IHC 1/2/3: IHC1 is  $\geq 1\%$ , IHC2 is  $\geq 5\%$ , IHC3 is  $\geq 10\%$ .

very impressive expression of immune-mediated genes (PD-L1, PD-L2, CTLA4, PD-1, PRF1, GZMA, BTLA, CD8A) in a single patient with PD-L1 expression less of 5% but  $>1\%$  who showed an impressive complete response to nivolumab. Although no final conclusion could be resumed from this study due to the small number of patients explored it is probably that tumors mutational burden and non-synonymous mutations play a different role in ccRCC as compared to other disease, however, further large prospective trials might be necessary to confirm this hypothesis.

It is worth mentioning the IMmotion 151 trial, a randomized Phase III study of Atezolizumab plus Bevacizumab vs. Sunitinib in untreated metastatic RCC (35). Primary endpoints included PFS in PD-L1 positive patients and OS in an intention to treat analysis. The IMmotion 151 trial met its primary PFS endpoint in the PD-L1 positive patients with atezolizumab + bevacizumab compared to sunitinib with fewer high grade adverse reactions. This data does support atezolizumab + bevacizumab as first line therapy in metastatic clear cell renal cell (35, 56).

## GENE EXPRESSION AS PREDICTIVE IMMUNOLOGICAL BIOMARKERS IN RENAL CELL CARCINOMA

Regarding gene expression, several data seem to correlate the expression of specific classes of genes (especially DNA repair genes) to immunotherapy outcomes (30). Reportedly, the most frequent event involved in ccRCC is the loss of chromosome 3p, which is associated with the development of VHL, PBRM1, BAP1 and SETD2 alterations in about 90% of ccRCC cases. Together with KDM5C, PTEN, MTOR and TP53, these represent the eight most frequently altered genes in ccRCC (22, 23, 36). However, second most frequently mutated sub-network included AID1A, SMARCA4, and PBAF SWI/SNF chromatin remodeling complex. When mutations occur in chromatin regulators PBRM1, BAP1, and SETD2, several related genes showed altered expression as compared to VHL mutation (22, 23, 36). In particular, chromatin modification pathways interact with several genes involved in hormonal activity (*ESR1*), *RAS oncogene*, transcriptional output (*HIF1A*, *JUN*, *FOS*, and *SP1*),

TGF-beta and especially DNA repair (*BAP1*) and immune-mediated signaling (*NFKB1* and *IL-6*) (22, 23, 36). To date, no data about the correlation between gene expressions (especially DNA repair gene alterations and immune-related genes) are available, but this appears to be an attractive hypothesis to test mainly focused on the detection of predictive markers and the better understanding of mechanisms related to immune response in ccRCC (22, 23, 36).

The fundamental role of the gene alterations of PBRM1, BAP1 and SETD2 has been recently enforced by the findings presented at ASCO Annual Meeting 2018. The Spanish Oncology Genitourinary Group (SOGUG) presented the results of an observational prospective study collecting samples from 77 RCC patients treated with mTOR inhibitors everolimus or temsirolimus (79 and 21% of cases, respectively) (37). The study analysis included both IHC for p-S6, p-S6K1, p-AKT, p21, BAP1, and PBRM1 and NGS (next generation sequencing) for mutational analysis on key genes of mTOR pathway in RCC. Among enrolled patients, 87% had ccRCC histology; 60% had intermediate, 39% good prognosis, and 1% poor prognosis (MSKCC). No association between p-S6, p-S6K1, p-AKT, and p21 staining and response to temsirolimus/everolimus was reported. However, negative IHC expression for BAP1 and PBRM1 was associated with better mTOR inhibitor response (OR = 4.0, 95%CI = 1.4–11.9,  $p = 0.011$  and OR = 3.9, 95%CI = 1.2–12.8,  $p = 0.025$ ).

On the other hand, Bossé et al. reported on the prognostic value of genetic alterations resulting in loss of function (defined by the presence of pathogenic gene variant or 2 copy deletion) of VHL, PBRM1, BAP1, SETD2, TP53, and KDM5C, which are frequently mutated in metastatic RCC, in patients with ccRCC stratified by IMDC risk classification and treated with 1st line VEGFR tyrosine kinase inhibitors (38). Tumor samples were analyzed by NGS or whole exome sequencing (TCGA). Three hundred and eight patients were included; 21% of them with IMDC good risk features, 54% intermediate and 17% poor risk (8% unknown). The presence of gene alterations in VHL, PBRM1, SETD2, BAP1, TP53, and KDM5C was, respectively, 77, 43, 29, 19, 11, and 11%. Gene alterations in BAP1 were associated with worst OS (HR 1.7; 95%CI 1.1–2.5,  $p = 0.01$ ), while alterations in PBRM1 and KDM5C were correlated with longer OS. Patients with tumors PBRM1 wild type and harboring gene alterations in BAP1 had worse OS (37 vs. 50 months, HR 1.9, 95% CI 1.2–2.8,  $p = 0.004$ ). Interestingly, when IMDC stratified criteria were applied the genomic profile was prognostic only in patients with intermediate risk.

The advances in understanding the molecular landscape of RCC parallel with the progresses in the histopathological characterization of this neoplasm. The 2016 WHO classification of the tumors of the kidney (13) has identified new renal entities including hereditary leiomyomatosis and renal cell carcinoma syndrome-associated RCC, succinate dehydrogenase-deficient RCC, tubulocystic RCC, acquired cystic disease-associated RCC, and clear cell papillary RCC. The list of histologic categories includes also emerging entities, such as RCC associated with ALK gene rearrangements and thyroid-like follicular RCC (13). A more accurate identification of the different histological tumor



categories represents another fundamental step forward for the selection of molecularly targetable approaches for patients with RCC thus enabling the possibility to selectively target the gene drivers of specific tumor variants (13). More recently, Chen et al. (16) surveyed 894 RCC cases for expression of genes involved in immune checkpoint pathways, including PD1 and PDL1 genes. Clear cell RCC subtypes had relatively high expression of several genes representing targets for immunotherapy, including PDCD1 (PD1), CD247 (CD3), PDCD1LG2 (PDL2), CTLA4 (CD152), TNFRSF9 (CD137), and TNFRSF4 (CD134). In addition, analysis of gene expression signatures and of DNA methylation signatures suggested greater levels of immune cell infiltrates, including T cells, within clear cell RCC relative to other RCC subtypes (16, 17).

Within clear cell-enriched (CC-e) RCC genomic subtypes, differential expression of specific checkpoint-related genes was observed mostly involving differences between CC-e.3 and CC-e.2 groups (more aggressive and less aggressive ccRCC categories, respectively). Compared to CC-e.2, CC-e.3 showed increased promoter methylation of miR-21 (MIR21) with corresponding decreased levels of the miR-21 target PTEN. In cancer, PTEN has an established role in intrinsic cellular control of PD-L1 (16, 17). Some other genes—including PDCD1, CTLA4, and TLR9—were associated with worse patient survival within ccRCC-associated cases; PDL1 expression was correlated with better patient survival, though this association was confounded by copy loss of 9p region associated with aggressive clear cell RCC and worse prognosis (18). In summary, better understanding the predictive and prognostic significance of PD1/PD-L1 expression and the identification of molecularly defined subtypes correlated with survival and response to therapy, represent quick steps toward implementing precision medicine in RCC via reducing the distance to the goal of identifying the best approach for a single RCC patient (28, 29, 40) (Table 3).

## PD-L1 IMMUNOHISTOCHEMISTRY IN RCC

Immunocheckpoint inhibitors (ICI) have marked a new paradigm in the treatment of RCC. The anti-PD1 drug nivolumab has been the first ICI drug to obtain approval by the FDA and European Commission for the treatment of RCC, and showed a significant OS benefit in patients with RCC that progressed following antiangiogenic therapy compared with everolimus (mTOR inhibitor) (26). Several other ICI compounds are currently under investigation for the treatment of RCC, alone or in combination with TKIs or other drugs (57–59).

Predictive biomarker research to select RCC patients eligible for ICI has mainly focussed on the PD1-PD-L1 axis detected by means of IHC. Low-to-no expression of PD-L1 on IC (immune cells) and TC (tumor cells) correlated with a trend toward lower response (PFS and OS) to the anti-PD-L1 drug atezolizumab compared with moderate to high PD-L1 expression levels (60). Updated analysis further confirmed the association between high PD-L1 expression and improved OS with atezolizumab treatment (57). For the anti-PD1 drug nivolumab, early data suggested a positive correlation between PD-L1 expression on TC and ORR

(61–64). Data from the Checkmate 025 trial showed that higher levels of PD-L1 expression are associated with poorer survival in RCC, but did not support PD-L1 as a marker predictive of treatment benefit in RCC; a benefit was observed, however, with nivolumab irrespective of PD-L1 expression (62). Furthermore, PD-L1 seems to be a dynamic biomarker since prior exposure to VEGF and mTOR inhibitors modulates its expression which can be largely variable after therapy (64, 65). Notably, a significant number of patients with PD-L1+ RCC do not respond to PD-1 pathway blockade, suggesting that additional intra-tumoral factors may influence treatment outcome (64, 65). Based on recent data, PD-L1 could be a prognostic biomarker for the adverse clinic-pathologic features of RCC but may not be discriminant enough to be a predictive biomarker (64, 66, 67). Furthermore, it was found that PD-L1 staining is almost exclusively observed in the high-grade component of a tumor and additionally a discordant expression of PD-L1 between primary tumors and their metastases was detected in ~20% of cases (68). Similar heterogeneity has been observed between primary and metastatic tumor based on molecular analysis (69).

Other possible biomarkers like PD-L2 and CTLA4 are reported in literature, thus far without straightforward predictive value (57). Increased amounts of CD3<sup>+</sup>/CD8<sup>+</sup> tumor-infiltrating T-cells have been reported after nivolumab treatment, but further research is needed to determine the biomarker-potential of this observations (70).

Recent data from a gene expression study on a small cohort of PDL1+ RCC patients treated with nivolumab identified a metabolic gene profile in the non-responding subgroup and overexpression of immunologic factors in the responding subgroup (71). Increasing mutational burden and neo-antigen formation have been associated with increased responsiveness to ICI in several other malignancies and recent data showed increased frequency of genomic alterations in RCC post-VEGFR therapy (72). These findings might explain the observed benefit of nivolumab post-VEGFR therapy and seem to correlate with the observation of lower response rates to nivolumab monotherapy in front line studies (70). A recent multilevel molecular analysis on the integrated TCGA RCC database showed relatively high expression of several genes representing targets for immunotherapy in ccRCC-associated molecular subtypes compared to other RCC subtypes, with additional differences within the several clear cell-enriched RCC genomic subtypes (16). These data also suggested greater levels of IC infiltrates within ccRCC relative to other RCC types (16). TCGA data suggest the hypothesis that clear cell-enriched RCC genomic subtypes would be most responsive to targeted immune checkpoints, hypothesis that awaits validation in prospective cohort series (16).

Several technical and biochemical issues are involved to explain the observed ambiguity of PD-L1 expression as predictor of response to ICI therapy in RCC. Differences in anti-PD-L1 antibody-clones, staining assays, tissue characteristics and scoring systems are amongst the major technical obstacles to overcome. The knowledge that PD-L1 expression is not binary, but instead shows a continuum with significant intratumour heterogeneity and therapy-induced changes, might

**TABLE 3 |** Prognostic and predictive biomarkers in Renal Cell Carcinoma.

Biomarker	Results	Association with	References
IHC expression of p-S6, p-S6K1, p-AKT, and p21	NA	No association with response to temsirolimus/everolimus	(37)
Negative IHC expression for BAP1	OR = 4.0, 95% CI = 1.4–11.9, $p = 0.011$	Better mTOR inhibitor response	(37)
Negative IHC expression for PBRM1	OR = 3.9, 95% CI = 1.2–12.8, $p = 0.025$	Better mTOR inhibitor response	(37)
Gene alterations in BAP1	HR 1.7; 95% CI 1.1–2.5, $p = 0.01$	Worse OS	(38)
Gene alterations in PBRM1	HR = 0.6; 95%CI 0.4–0.8, $p = 0.001$	Better OS	(38)
Gene alterations in KDM5C	HR = 0.4; 95%CI 0.2–0.8, $p = 0.007$	Better OS	(38)
SETD2, TP53, and VHL	NA ( $p > 0.4$ )	Not associated with prognosis	(38)
PBRM1 wild type + gene alterations BAP1	37 vs. 50 months, HR 1.9, 95% CI 1.2–2.8, $p = 0.004$	Worse OS	(38)
PDCD1, CTLA4, and TLR9	NA	Worse OS	(16)
9p deletion	HR 4.323; $p = 0.021$ HR 4.603; $p = 0.007$	High risk of recurrence and RCC-specific mortality	(18)

even represent a bigger challenge for being an ideal biomarker (73, 74). The recent report on the presence of compensatory inhibitory pathways (VISTA) in the setting of immunotherapy in metastatic prostate further underlines the complexity to predict the therapeutic response based on a single biomarker like PD-L1 (74). Recent concordance studies on non-small cell lung cancer have shown only minimal differences in staining patterns between most of the different validated and commercially available anti-PD-L1 antibody clones (73, 75–78). These findings are encouraging, although clinical cross-validation data between the different assays are not available at this moment. High concordances between the different assays and between the pathologists within a single assay were only found for PD-L1 scoring TC and not in immune cells IC (75, 76). Concerning RCC this could be a critical point since PD-L1 expression in IC is used as a companion biomarker for some FDA-approved anti-PD-L1 drugs.

## MMR-DEFICIENCY AND MUTATIONAL LOAD IN RCC

Renal cell carcinoma are not considered to belong to the HNPCC (hereditary non-polyposis colon cancer) spectrum, but in sporadic RCC loss of MMR proteins is frequently observed, especially of MLH1 and MSH2 (79–81). Variable MMR gene alterations have been reported as underlying mechanisms, but others did not detect microsatellite instability (MSI) caused by either promoter hypermethylation or alteration of the coding region of MMR studied genes (81–83). The reduced MMR protein expression by IHC has been linked to RCC subtypes and might contribute to the respective different biological behavior (84). As addressed earlier in this review, MMR-deficiency is more and more recognized as an important biologic event in genitourinary cancers. MMR deficiency can occur in patients with Lynch syndrome (HNPCC) and in patients with sporadic

MMR-deficient tumors (84). MMR-deficient tumors exhibit a higher rate of mutations (high mutational burden), which can result in the formation of neo-antigens to enhance the antitumor immune response (85). Furthermore, MMR-deficient tumors express different immune checkpoint ligands indicating that their active immune microenvironment is counterbalanced by immune inhibitory signals that resist tumor destruction (86). Recently reported data showing a better clinical response to the anti-PD-1 drug pembrolizumab in MMR-deficient patients support the hypothesis that MMR-deficient tumors respond better to anti-PD-1 therapy than do MMR-proficient tumors (87).

In RCC cancers, data on the relation between MMR-status and response to immunotherapy are still emerging (88, 89). Based on the promising results in patients with MMR-deficient cancers, FDA has recently approved pembrolizumab for the treatment of adult and pediatric patients with un-resectable or metastatic MMR-deficient solid tumors, irrespective of the tumor origin. In this context, MMR-deficient/MSI-H solid genitourinary tumors could be important candidates for anti PD-1 treatment. The reality might however be much more complex; for instance, several clinical trials have shown that some MMR-deficient tumors do not respond to immunotherapy, while mutations in other genes have also been linked to high mutational burden and upregulation of immune checkpoints (85). From a methodological point of view, there is an ongoing discussion and evolution in literature concerning the methodology to get reliable data on mutational load and MSI in a context of cost-efficiency and optimal logistics. Whole exome sequencing, T-cell receptor sequencing and targeted NGS can be used to assess mutational load (90) and promising data on novel platforms to detect MSI (e.g., MSI-Sensor and MANTIS) have recently been published (91). The detection of MMR-deficient tumors and the selection of those patients that will really benefit from immunotherapy remains an ongoing and challenging task.

## BIOMARKERS OF ACQUIRED RESISTANCE

Despite the durable responses observed with immune checkpoint inhibition, nearly all patients will progress. A number of mechanisms have been identified including neo-antigen loss, upregulation of alternative immune checkpoints, loss of antigen presentation, and defective interferon signaling (92–94). A recent whole exome sequencing study on paired tumor samples prior to treatment with ICI and at the time of progression ( $n = 4$ , 2 treated with nivolumab/ipilimumab, 2 treated with nivolumab) was reported by Anagnostou et al. (92). Although they found an increase in total number of candidate neo-antigens, a subset of them was actually eliminated at the time of acquired resistance. In the four patients, there were 18, 10, 7, and 6 neo-antigens lost, and all of them had higher predicted MHC binding affinity. There were no copy number alterations of CD274 which encodes for PD-L1, PDCD1 encoding for PD-1, CTLA-4, JAK1, or JAK2. There were no genetic alterations in HLA or  $\beta$ 2-microglobulin. They also evaluated clonal T-cell reactivity in three of these patients using peripheral blood mononuclear cells loaded with predicted neo-antigens cultured with purified T-cells. All patients showed clonal T-cell expansion to lost peptides and either no affinity or lower affinity for the wild type of the predicted neo-antigen (92). Neo-antigen loss and growth of a subclone lacking the neo-antigen eliciting the immune response are both potential explanations of this resistance mechanism, although the power of available information is limited. This mechanism of resistance underscores the rationale for using neo-antigen profiling as a predictive biomarker of benefit and also underscores the dynamic nature of these biomarkers.

Defects in the interferon- $\gamma$  signaling pathway have also been identified as a major mechanism of resistance. Interferon- $\gamma$  signaling plays a crucial role in the anticancer immune response. It has been shown to upregulate PD-L1 expression on TC and IC, to increase MHC Class I expression and promote antigen presentation, and recruit effector cells (92–97). It results in the downstream stimulation of JAK/STAT signaling pathway and expression of a number of anti-cancer genes (98). Mutations in JAK1/2 render cells insensitive to interferon- $\gamma$  signaling, which results in escape from PD-L1 pathway inhibition and impairs the antitumor immune response. This has been identified as a mechanism of both primary and secondary resistance (99–101). Interferon- $\gamma$  signaling has been demonstrated to increase expression of immune inhibitory molecules, such as indoleamine-2,3-deoxygenase (IDO) that can limit the anti-tumor response (101). Inhibition of IDO production is the subject of an ongoing clinical trial in combination with PD-1 immune checkpoint inhibition. Defects in antigen presentation, such as mutations in the  $\beta$ -2 microglobulin gene, have also been identified as a mechanism of resistance (100). Beta-2 microglobulin is essential for MHC class I molecule surface expression and a defect can block CD8-Tcell recognition. HLA loss is another potential mechanism of immune evasion and determining copy number alterations have been difficult due to the polymorphic nature of the locus. McGrahan et al. developed a computational tool using NGS data to determine HLA loss of heterozygosity in

100 early stage NSCLC patients. Interestingly, 40% of patients displayed HLA loss of heterozygosity and phylogenetic analysis shows that this is likely a later evolutionary event (102). TIM-3, LAG-3, and TIGIT are known alternative immune checkpoints that play a role in T-cell exhaustion and are expressed on tumor infiltrating lymphocytes (93). Koyama and colleagues identified TIM-3 to be upregulated in a murine model of NSCLC at the time of resistance to anti-PD-1 therapy and demonstrated a survival advantage with treatment using a TIM-3 blocking antibody. The authors additionally identified two patients with biopsies performed at the time of progression to anti-PD-1 therapy with increased TIM-3 expression (103). Novel therapeutic approach directed at these alternative immune checkpoints are the subject of ongoing clinical trials and are of potential relevance in RCC.

## FUTURE PERSPECTIVE

The complex interplay of signaling pathways and inflammatory mediators seems to be crucial for RCC development and response to therapy (43, 44, 53). Immune cells including neutrophils, lymphocytes and macrophages have been implicated in promoting metastatic spread, tumor angiogenesis, in primary and acquired drug resistance, as well as in the formation of pre-metastatic niches (43, 44, 53). On this scenario, the checkpoint molecules have gained wide interest since the introduction of anti-CTLA-4 and anti-PD-1/PD-L1 agents into daily oncology practice (45). Beyond PD-1 and CTLA-4, a variety of molecules are emerging as potentially future therapeutic immunotargets in RCC (46). This list includes the V-domain immunoglobulin containing suppressor of T-cell activation (VISTA), which has been recently shown to exert its inhibitory activity by acting as a ligand on antigen presenting cells and as a receptor on T cells (104–106), chemokine receptors (45), the soluble lymphocyte-activation gene-3 (LAG-3), 4-1BB, B and T lymphocyte attenuator, and OX40 (CD134) (47).

Nowadays, there is not a clear-cut knowledge of the underlying mechanisms of immuno-checkpoint inhibitors-induced tumor response. To address this issue, Wei et al. investigated the effects of anti-PD-1 and anti-CTLA-4 inhibitors in human melanoma and murine tumor models (48). They first revealed that these agents are able to target distinct tumor-infiltrating T cell subpopulations. In particular, PD-1 blockade promotes the expansion of specific exhausted-like CD8-T cell population, while CTLA-4 blockade induces both an ICOS<sup>+</sup> Th1-like CD4 effector subset and exhausted-like CD8-T cells (48). This evidence favors the combined use of current and probably future checkpoint inhibitors in cancer patients. These combinations, seems to be characterized by a tolerable safety profile (107). Tumor responsiveness may vary according to the mutational load and the expression of immunotargets in the tumor environment, which is variable in the different phases of RCC development and progression (49, 108–110). Based on this evidence, assessing the expression of PD-1/PD-L1 or other emerging immunotargets only at the diagnosis of metastatic disease may not reflect tumor dynamicity.



To improve the feasibility and reduce the clinical impact of re-biopsy, assessing biomarkers on circulating tumor cells (CTCs) or exosomes (111) may represent a not invasive strategy that can be performed several times during cancer therapy in order to reflect the changes occurred in the tumor environment. An early identification of validated biomarkers would be crucial to definitively place immunotherapy into the era of precision medicine and to optimize the cost-effectiveness of ICI agents in cancer patients (50, 112). In addition, the recent paper by Routy et al. showed that primary resistance to ICI can be correlated with abnormal gut microbiome composition (51). In this study, the effectiveness of PD-1 blockade resulted enhanced by transplanting fecal microbiota from responder cancer patients into germ-free or antibiotic-treated mice (51), thus representing another step forward on the way to personalized and precision immunotherapy in cancer patients.

Another factor that results of great relevance to improve the efficacy of ICI in RCC patients is the comprehension of the immunological effects of TKIs and mTOR inhibitors (53, 113). Actually, these agents can indirectly exert their anti-tumor activity by targeting immune cells in the RCC microenvironment (53), and this should be considered in order to combine or sequence them with currently available and probably future immunotherapies. For instance, sunitinib has been shown to inhibit the colony forming units driven by GM-CSF and FLT3 ligand FLT3L (114) as well as dendritic cell antigen-presentation (115) (by decreasing the secretion of cytokines and the expression of MHC and CD1a molecules), to suppress the myeloid-derived suppressor cells (MDSCs are involved in RCC progression and drug resistance), to enhance tumor cell sensitivity to NK cell killing (116) and to reduce the total count of CD3 and CD4T cells and regulatory T cells (117, 118). On the other hand, pazopanib showed lower inhibitory potency and affinity against FLT3 and c-kit compared to sunitinib (119). Interestingly, we previously showed that axitinib can increase the surface NKG2D ligand expression, thus promoting NK cell recognition and degranulation in A-498 RCC cells in a ROS-dependent manner (120). At present, few evidences are available on the immunomodulatory effects of cabozantinib and lenvatinib, recently introduced into RCC clinical practice.

## EXPERT OPINION AND CONCLUSIONS

Optimizing the combination between immunotherapy and target agents as well as the possible favorable sequence of treatment between these two classes of drugs remain open questions at this moment but ongoing studies support this as of great future potential. On this way we have only limited data provided from Immotion150 which demonstrated that association between a PD-L1 inhibitor and bevacizumab is feasible, well-tolerated, and results in an effective clinical benefit from our patients. Of relevance, is to note that most studies explored immunotherapy in patients with ccRCC and the role of ICI still remains unknown in mRCC. Though, there are several questions that need to be answered, current data support that immunotherapy represents a revolution for the management of RCC resulting in a dynamic and evolving scenario in which more novelties will be shortly made available. Because the potentially deadly

adverse effects from immune checkpoint inhibitors, gathering predictive information in RCC seems to be prudent. However, recent scientific insights indicate that a single biomarker for patient selection may not be feasible, given that immune responses are dynamic and evolve over time (121). Biomarker development for ICI drugs will require integration of multiple biologic components like PD-L1 expression, TILs, mutational load, and probably many others now considered emergent biomarkers.

## EXECUTIVE SUMMARY

- Immunotherapy is gradually becoming a key factor in the therapeutic algorithm for patients with renal cell cancers at different stages of disease.
- The increasing knowledge on the genomic landscape of renal cell carcinoma supports stratification of patients for targeted therapies.
- A single biomarker for patient selection may not be feasible, given that immune responses are complex, dynamic and evolve over time.
- Biomarker development for ICI drugs will require integration of multiple biologic components like PD-L1 expression, TILs and mutational load.

## NEXT STEPS

- New methodological approaches likely based on digital pathology may be relevant since they allow objectively recognizing and quantitation of the biomarker and therefore might produce objective and reproducible cut-offs useful in patient's therapeutic stratification.
- Radiologic derived biomarkers, such as artificial intelligence derived, radiopharmaceutical, and liquid biopsy derived biomarkers, are likely to enter the biomarker-field in the next coming years.
- Large-scale biomarker-driven prospective trials with consensus methodologies on biomarker assessment and scoring are needed to reach clinical validation of different biomarkers, needed for a reliable single-patient appointment to the appropriate immunotherapy.
- Multidisciplinary approaches are needed to fully develop the current and future value of ICI in clinical practice.
- Better understanding of solid tumor genomics shows that also for RCC, combining targeted therapy with ICI has the potential to improve cancer outcomes, and that reliable biomarkers will be crucial for a stringent patient selection in trials of targeted and checkpoint inhibitor drugs and to apply novel therapeutic strategies aimed at restoring effective antitumor immunity in patients with cancer.

## AUTHOR CONTRIBUTIONS

AL-B, RM, MSc, and LC: conception and design; AL-B and VH: drafting the manuscript; AB, AC, and VH: review of the literature; AL-B, LC, RM, MSc, MSa, FM, VH, and TG: critical revision of the manuscript.

## REFERENCES

- Graham J, Heng DY, Brugarolas J, Vaishampayan U. Personalized management of advanced kidney cancer. *Am Soc Clin Oncol Educ Book* (2018) 38:330–41. doi: 10.1200/EDBK\_201215
- Motzer RJ, Hutson TE, Tomczak P, Michaelson MD, Bukowski RM, Rixe O, et al. Sunitinib versus interferon alfa in metastatic renal-cell carcinoma. *N Engl J Med.* (2007) 356:115–24. doi: 10.1056/NEJMoa065044
- Motzer RJ, Hutson TE, Cella D, Reeves J, Hawkins R, Guo J, et al. Pazopanib versus sunitinib in metastatic renal-cell carcinoma. *N Engl J Med.* (2013) 369:722–31. doi: 10.1056/NEJMoa1303989
- Sternberg CN, Hawkins RE, Wagstaff J, Salman P, Mardiak J, Barrios CH, et al. A randomised, double-blind phase III study of pazopanib in patients with advanced and/or metastatic renal cell carcinoma: final overall survival results and safety update. *Eur J Cancer* (2013) 49:1287–96. doi: 10.1016/j.ejca.2012.12.010
- Choueiri TK, Escudier B, Powles T, Tannir NM, Mainwaring PN, Rini BI, et al. Cabozantinib versus everolimus in advanced renal-cell carcinoma. *N Engl J Med.* (2015) 373:1814–23. doi: 10.1056/NEJMoa1510016
- Motzer RJ, Escudier B, Tomczak P, Hutson TE, Michaelson MD, Negrier S, et al. Axitinib versus sorafenib as second-line treatment for advanced renal cell carcinoma: overall survival analysis and updated results from a randomised phase 3 trial. *Lancet Oncol.* (2013) 14:552–62. doi: 10.1016/S1470-2045(13)70093-7
- Motzer RJ, Escudier B, Oudard S, Hutson TE, Porta C, Bracarda S, et al. Phase 3 trial of everolimus for metastatic renal cell carcinoma: final results and analysis of prognostic factors. *Cancer* (2010) 116:4256–65. doi: 10.1002/cncr.25219
- Escudier B, Eisen T, Stadler WM, Szczylik C, Oudard S, Staehler M, et al. Sorafenib for treatment of renal cell carcinoma: final efficacy and safety results of the phase III treatment approaches in renal cancer global evaluation trial. *J Clin Oncol.* (2009) 27:3312–8. doi: 10.1200/JCO.2008.19.5511
- Hutson TE, Escudier B, Esteban E, Bjarnason GA, Lim HY, Pittman KB, et al. Randomized phase III trial of temsirolimus versus sorafenib as second-line therapy after sunitinib in patients with metastatic renal cell carcinoma. *J Clin Oncol.* (2014) 32:760–7. doi: 10.1200/JCO.2013.50.3961
- Motzer RJ, Hutson TE, Glen H, Michaelson MD, Molina A, Eisen T, et al. Lenvatinib, everolimus, and the combination in patients with metastatic renal cell carcinoma: a randomised, phase 2, open-label, multicentre trial. *Lancet Oncol.* (2015) 16:1473–82. doi: 10.1016/S1470-2045(15)00290-9
- Motzer RJ, Escudier B, McDermott DF, George S, Hammers HJ, Srinivas S, et al. Nivolumab versus everolimus in advanced renal-cell carcinoma. *N Engl J Med.* (2015) 373:1803–13. doi: 10.1056/NEJMoa1510665
- Santoni M, Massari F, Di Nunno V, Conti A, Cimadamore A, Scarpelli M, et al. Immunotherapy in renal cell carcinoma: latest evidence and clinical implications. *Drugs Context* (2018) 7:212528. doi: 10.7573/dic.212528
- Moch H, Cubilla AL, Humphrey PA, Reuter VE, Ulbright TM. The 2016 WHO classification of tumours of the urinary system and male genital organs-part A: renal, penile, and testicular tumours. *Eur Urol.* (2016) 70:93–105. doi: 10.1016/j.eururo.2016.02.029
- Montironi R, Lopez-Beltran A, Cheng L. Editorial: emerging biomarkers in genitourinary tumors. *Curr Drug Metab.* (2017) 18:690–91. doi: 10.2174/138920021808171016103101
- Gevaert T, Montironi R, Lopez-Beltran A, Van Leenders G, Allory Y, De Ridder D, et al. Genito-urinary genomics and emerging biomarkers for immunomodulatory cancer treatment. *Semin Cancer Biol.* (2017) 52:216–27. doi: 10.1016/j.semcancer.2017.10.004
- Chen F, Zhang Y, Senbabaoglu Y, Ciriello G, Yang L, Reznik E, et al. Multilevel genomics-based taxonomy of renal cell carcinoma. *Cell Rep.* (2016) 14:2476–89. doi: 10.1016/j.celrep.2016.02.024
- Shuch B, Amin A, Armstrong AJ, Eble JN, Ficarra V, Lopez-Beltran A, et al. Understanding pathologic variants of renal cell carcinoma: distilling therapeutic opportunities from biologic complexity. *Eur. Urol.* (2015) 67:85–97. doi: 10.1016/j.eururo.2014.04.029
- El-Mokadem I, Fitzpatrick J, Bondad J, Rauchhaus P, Cunningham J, Pratt N, et al. Chromosome 9p deletion in clear cell renal cell carcinoma predicts recurrence and survival following surgery. *BJC* (2014) 111:1381–90. doi: 10.1038/bjc.2014.420
- Dalglish GL, Furge K, Greenman C, Chen L, Bignell G, Butler A, et al. Systematic sequencing of renal carcinoma reveals inactivation of histone modifying genes. *Nature* (2010) 463:360–3. doi: 10.1038/nature08672
- Guo G, Gui Y, Gao S, Tang A, Hu X, Huang Y, et al. Frequent mutations of genes encoding ubiquitin-mediated proteolysis pathway components in clear cell renal cell carcinoma. *Nat Genet.* (2011) 44:17–9. doi: 10.1038/ng.1014
- Varela I, Tarpey P, Raine K, Huang D, Ong CK, Stephens P, et al. Exome sequencing identifies frequent mutation of the SWI/SNF complex gene PBRM1 in renal carcinoma. *Nature* (2011) 469:539–42. doi: 10.1038/nature09639
- Brannon AR, Reddy A, Seiler M, Arreola A, Moore DT, Pruthi RS, et al. Molecular stratification of clear cell renal cell carcinoma by consensus clustering reveals distinct subtypes and survival patterns. *Genes Cancer* (2010) 1:152–63. doi: 10.1177/1947601909359929
- Brooks SA, Brannon AR, Parker JS, Fisher JC, Sen O, Kattan MW, et al. ClearCode34: a prognostic risk predictor for localized clear cell renal cell carcinoma. *Eur Urol.* (2014) 66:77–84. doi: 10.1016/j.eururo.2014.02.035
- Cancer Genome Atlas Research Network. Comprehensive molecular characterization of papillary renal-cell carcinoma. *N Engl J Med.* (2016) 374:135–45. doi: 10.1056/NEJMoa1505917
- Davis CF, Ricketts CJ, Wang M, Yang L, Cherniack AD, Shen H, et al. The somatic genomic landscape of chromophobe renal cell carcinoma. *Cancer Cell* (2014) 26:319–30. doi: 10.1016/j.ccr.2014.07.014
- Escudier B, Porta C, Schmidinger M, Rioux-Leclercq N, Bex A, Khoo V, et al. Renal cell carcinoma: ESMO clinical practice guidelines for diagnosis, treatment and follow-up. *Ann Oncol.* (2016) 27(Suppl. 5):v58–68. doi: 10.1093/annonc/mdw328
- Bindea G, Mlecnik B, Tosolini M, Kirilovsky A, Waldner M, Obenauf AC, et al. Spatiotemporal dynamics of intratumoral immune cells reveal the immune landscape in human cancer. *Immunity* (2013) 39:782–95. doi: 10.1016/j.immuni.2013.10.003
- Mann SA, Lopez-Beltran A, Massari F, Pili R, Fiorentino M, Koch MO, et al. Targeting the programmed cell death-1 pathway in genitourinary tumors: current progress and future perspectives. *Curr Drug Metab.* (2017) 18:700–11. doi: 10.2174/1389200218666170518162500
- Motzer RJ, Tannir NM, McDermott DF, Arén Frontera O, Melichar B, Choueiri TK, et al. Nivolumab plus Ipilimumab versus sunitinib in advanced renal-cell carcinoma. *N Engl J Med.* (2018) 378:1277–90. doi: 10.1056/NEJMoa1712126
- Rizvi NA, Hellmann MD, Snyder A, Kvistborg P, Makarov V, Havel JJ, et al. Cancer immunology. Mutational landscape determines sensitivity to PD-1 blockade in non-small cell lung cancer. *Science* (2015) 348:124–8. doi: 10.1126/science.aaa1348
- de Velasco G, Miao D, Voss MH, Hakimi AA, Hsieh JJ, Tannir NM, et al. Tumor mutational load and immune parameters across metastatic renal cell carcinoma risk groups. *Cancer Immunol Res.* (2016) 4:820–2. doi: 10.1158/2326-6066.CIR-16-0110
- Yarchoan M, Hopkins A, Jaffee EM. Tumor mutational burden and response rate to PD-1 inhibition. *N Engl J Med.* (2017) 377:2500–1. doi: 10.1056/NEJMc1713444
- Malouf GG, Ali SM, Wang K, Balasubramanian S, Ross JS, Miller VA, et al. Genomic characterization of renal cell carcinoma with sarcomatoid dedifferentiation pinpoints recurrent genomic alterations. *Eur Urol.* (2016) 70:348–57. doi: 10.1016/j.eururo.2016.01.051
- de Velasco G, Miao D, Shukla S. Integrated genomic correlates of response to PD-1 inhibitor nivolumab in metastatic renal cell carcinoma (mRCC). *J Clin Oncol.* (2016) 34(Suppl. 2):545. doi: 10.1200/jco.2016.34.2\_suppl.545
- Motzer RJ, Powles T, Atkins MB, Escudier B, McDermott DF, Suarez C. IMmotion151: A randomized phase III study of atezolizumab plus bevacizumab vs sunitinib in untreated metastatic renal cell carcinoma (mRCC). *JCO* (2018) 36(6\_Suppl.):578. doi: 10.1200/JCO.2018.36.6\_suppl.578
- Cancer Genome Atlas Research Network. Comprehensive molecular characterization of clear cell renal cell carcinoma. *Nature* (2013) 499:43–9. doi: 10.1038/nature12222
- Garcia-Donas J, Roldan JM, Lainez N, Castellano DE, González EE, Climent MA, et al. Comprehensive molecular and immunohistochemical analysis of

- advanced renal cell carcinoma patients treated with mTOR inhibitors. *J Clin Oncol.* (2018) 36:4559. doi: 10.1200/JCO.2018.36.15\_suppl.4559
38. Bossé D, Xie W, Ged Y, Hahn AW, Bergerot PG, Ho TH, et al. Alterations in key clear cell renal cell carcinoma (RCC) genes to refine patient prognosis. *J Clin Oncol.* (2018) 36:4516. doi: 10.1200/JCO.2018.36.15\_suppl.4516
  39. Mosillo C, Ciccarese C, Bimbatti D, Fantinel E, Volta AD, Bisogno I, et al. Renal cell carcinoma in one year: going inside the news of 2017—a report of the main advances in RCC cancer research. *Cancer Treat Rev.* (2018) 67:29–33. doi: 10.1016/j.ctrv.2018.02.009
  40. Santoni M, Massari F, Piva F, Carrozza F, Di Nunno V, Cimadamore A, et al. Tivozanib for the treatment of renal cell carcinoma. *Expert Opin Pharmacother.* (2018) 19:1–5. doi: 10.1080/14656566.2018.1480722
  41. Atkins MB, McDermott DF, Powles T, Motzer RJ, Rini BI, Fong L. IMmotion150: a phase II trial in untreated metastatic renal cell carcinoma (mRCC) patients (pts) of atezolizumab (atezo) and bevacizumab (bev) vs and following atezo or sunitinib (sun). *J Clin Oncol.* (2017) 35:4505. doi: 10.1200/JCO.2017.35.15\_suppl.4505
  42. Escudier B, Tannir N, McDermott DF, Frontera OA, Melichar B, Plimack ER, et al. CheckMate 214: efficacy and safety of nivolumab + ipilimumab (N+I) v sunitinib (S) for treatment-naïve advanced or metastatic renal cell carcinoma (mRCC), including IMDC risk and PD-L1 expression subgroups. *Ann Oncol.* (2017) 28:v605–49. doi: 10.1093/annonc/mdx440
  43. Santoni M, Massari F, Amantini C, Nabissi M, Maines F, Burattini L, et al. Emerging role of tumor-associated macrophages as therapeutic targets in patients with metastatic renal cell carcinoma. *Cancer Imm Immunother.* (2013) 62:1757–68. doi: 10.1007/s00262-013-1487-6
  44. Lolli C, Basso U, Derosa L, Scarpi E, Sava T, Santoni M, et al. Systemic immune-inflammation index predicts the clinical outcome in patients with metastatic renal cell cancer treated with sunitinib. *Oncotarget* (2016) 7:54564–71. doi: 10.18632/oncotarget.10515
  45. Massari F, Santoni M, Ciccarese C, Santini D, Alfieri S, Brunelli M, et al. PD-1/PD-L1 blockade alone or in combination in renal cell carcinoma: current studies and future promises. *Cancer Treat Rev.* (2015) 41:114–23. doi: 10.1016/j.ctrv.2014.12.013
  46. Kucharczyk J, Matrana MR, Santoni M, Massari F, Scarpelli M, Cheng L, et al. Emerging immunotargets in metastatic renal cell carcinoma. *Curr Drug Targets* (2016) 17:771–6. doi: 10.2174/1389450117666151209115753
  47. Massari F, Santoni M, Ciccarese C, Santini D. The immunocheck-points in modern oncology: the next 15 years. *Expert Opin Biol Ther.* (2015) 15:917–21. doi: 10.1517/14712598.2015.1035251
  48. Wei SC, Levine JH, Cogdill AP, Zhao Y, Anang NAS, Andrews MC, et al. Distinct cellular mechanisms underlie anti-CTLA-4 and anti-PD-1 checkpoint blockade. *Cell* (2017) 170:1120–33. doi: 10.1016/j.cell.2017.07.024
  49. Piva F, Santoni M, Scarpelli M, Briganti A, Berardi R, Montorsi F, et al. “Anti-programmed cell death protein 1 (PD-1) antibody nivolumab leads to a dramatic and rapid response in papillary renal cell carcinoma with sarcomatoid and rhabdoid features” [Geynisman DM, *Eur Urol* 68 (2015) 912–915]. Role of *PBMR1* variants in modulating the expression of PD-1 and PD-L1 in renal cell carcinoma. *Eur Urol.* (2016) 70:e72–4. doi: 10.1016/j.eururo.2015.07.008
  50. Montironi R, Santoni M, Tartari F, Lopez-Beltran A, Cheng L, Berardi R, et al. Testing PD-1/PD-L1 expression in cancer therapy: pathological insights and economic sustainability. *Arch Pathol Lab Med.* (2016) 140:501–2. doi: 10.5858/arpa.2015-0529-LE
  51. Rout B, Le Chatelier E, Derosa L, Duong CPM, Alou MT, Daillière R, et al. Gut microbiome influences efficacy of PD-1-based immunotherapy against epithelial tumors. *Science* (2018) 359:91–7. doi: 10.1126/science.aan3706
  52. Raman R, Vaena D. Immunotherapy in metastatic renal cell carcinoma: a comprehensive review. *Biomed Res Int.* (2015) 2015:367354. doi: 10.1155/2015/367354
  53. Santoni M, Berardi R, Amantini C, Burattini L, Santini D, Santoni G, et al. Role of natural and adaptive immunity in renal cell carcinoma response to VEGFR-TKIs and mTOR inhibitor. *Int J Cancer* (2014) 134:2772–7. doi: 10.1002/ijc.28503
  54. Alsaab HO, Sau S, Alzhrani R, Tatiparti K, Bhise K, Kashaw SK, et al. PD-1 and PD-L1 checkpoint signaling inhibition for cancer immunotherapy: mechanism, combinations, and clinical outcome. *Front Pharmacol.* (2017) 8:561. doi: 10.3389/fphar.2017.00561
  55. López JI, Pulido R, Cortés JM, Angulo JC, Lawrie CH. Potential impact of PD-L1 (SP-142) immunohistochemical heterogeneity in clear cell renal cell carcinoma immunotherapy. *Pathol Res Pract.* (2018) 214:1110–4. doi: 10.1016/j.prp.2018.06.003
  56. Drug Combo Bests Sunitinib in RCC. *Cancer Discov.* (2018) 8:OF4. doi: 10.1158/2159-8290.CD-NB2018-023
  57. Atkins MB, Clark JI, Quinn DI. Immune checkpoint inhibitors in advanced renal cell carcinoma: experience to date and future directions. *Ann Oncol.* (2017) 28:1484–94. doi: 10.1093/annonc/mdx151
  58. Ghatlani P, Zibelman M, Geynisman DM, Plimack ER. Checkpoint inhibitors for the treatment of renal cell carcinoma. *Curr Treat Options Oncol.* (2017) 18:7. doi: 10.1007/s11864-017-0458-0
  59. Yang JC, Hughes M, Kammula U, Royal R, Sherry RM, Topalian SL, et al. Ipilimumab (Anti-CTLA4 antibody) causes regression of metastatic renal cell cancer associated with enteritis and hypophysitis. *J Immunother.* (2007) 30:825–30. doi: 10.1097/CJI.0b013e318156e47e
  60. McDermott DF, Sosman JA, Sznol M, Massard C, Gordon MS, Hamid O, et al. Atezolizumab, an anti-programmed death-ligand 1 antibody, in metastatic renal cell carcinoma: long-term safety, clinical activity, and immune correlates from a phase Ia study. *J Clin Oncol.* (2016) 34:833–42. doi: 10.1200/JCO.2015.63.7421
  61. Topalian SL, Hodi FS, Brahmer JR, Gettinger SN, Smith DC, McDermott DF, et al. Safety, activity, and immune correlates of anti-PD-1 antibody in cancer. *N Engl J Med.* (2012) 366:2443–54. doi: 10.1056/NEJMoa1200690
  62. Motzer RJ, Rini BI, McDermott DF, Redman BG, Kuzel TM, Harrison MR, et al. Nivolumab for metastatic renal cell carcinoma: results of a randomized phase II trial. *J Clin Oncol.* (2015) 33:1430–7. doi: 10.1200/JCO.2014.59.0703
  63. Taube JM, Klein A, Brahmer JR, Xu H, Pan X, Kim JH, et al. Association of PD-1, PD-1 ligands, and other features of the tumor immune microenvironment with response to anti-PD-1 therapy. *Clin Cancer Res.* (2014) 20:5064–74. doi: 10.1158/1078-0432.CCR-13-3271
  64. Mazza C, Escudier B, Albiges L. Nivolumab in renal cell carcinoma: latest evidence and clinical potential. *Ther Adv Med Oncol.* (2017) 9:171–81. doi: 10.1177/1758834016679942
  65. Hirayama Y, Gi M, Yamano S, Tachibana H, Okuno T, Tamada S, et al. Anti-PD-L1 treatment enhances antitumor effect of everolimus in a mouse model of renal cell carcinoma. *Cancer Sci.* (2016) 107:1736–44. doi: 10.1111/cas.13099
  66. Xu F, Xu L, Wang Q, An G, Feng G, Liu F. Clinicopathological and prognostic value of programmed death ligand-1 (PD-L1) in renal cell carcinoma: a meta-analysis. *Int J Clin Exp Med.* (2015) 8:14595–603.
  67. Choueiri TK, Figueroa DJ, Fay AP, Signoretti S, Liu Y, Gagnon R, et al. Correlation of PD-L1 tumor expression and treatment outcomes in patients with renal cell carcinoma receiving sunitinib or pazopanib: results from COMPARE, a randomized controlled trial. *Clin Cancer Res.* (2015) 21:1071–7. doi: 10.1158/1078-0432.CCR-14-1993
  68. Callea M, Albiges L, Gupta M, Cheng SC, Genega EM, Fay AP, et al. Differential expression of PD-L1 between primary and metastatic sites in clear-cell renal cell carcinoma. *Cancer Immunol Res.* (2015) 3:1158–64. doi: 10.1158/2326-6066.CIR-15-0043
  69. Serie DJ, Joseph RW, Chevillier JC, Ho TH, Parasramka M, Hilton T, et al. Clear cell type A and B molecular subtypes in metastatic clear cell renal cell carcinoma: tumor heterogeneity and aggressiveness. *Eur Urol.* (2017) 71:979–85. doi: 10.1016/j.eururo.2016.11.018
  70. Choueiri TK, Fishman MN, Escudier B, McDermott DF, Drake CG, Kluger H, et al. Immunomodulatory activity of nivolumab in metastatic renal cell carcinoma. *Clin Cancer Res.* (2016) 22:5461–71. doi: 10.1158/1078-0432.CCR-15-2839
  71. Ascierto ML, McMiller TL, Berger AE, Danilova L, Anders RA, Netto GJ, et al. The intratumoral balance between metabolic and immunologic gene expression is associated with anti-PD-1 response in patients with renal cell carcinoma. *Cancer Immunol Res.* (2016) 4:726–33. doi: 10.1158/2326-6066.CIR-16-0072
  72. Pal SK, Sonpavde G, Agarwal N, Vogelzang NJ, Srinivas S, Haas NB, et al. Evolution of circulating tumor DNA profile from first-line to subsequent



- therapy in metastatic renal cell carcinoma. *Eur Urol.* (2017) 72:557–64. doi: 10.1016/j.eururo.2017.03.046
73. Kerr KM, Tsao MS, Nicholson AG, Yatabe Y, Wistuba II, Hirsch FR, IASLC Pathology Committee. Programmed death-ligand 1 immunohistochemistry in lung cancer: in what state is this art? *J Thorac Oncol.* (2015) 10:985–9. doi: 10.1097/JTO.0000000000000526
  74. Gao J, Ward JE, Pettaway CA, Shi LZ, Subudhi SK, Vence LM, et al. VISTA is an inhibitory immune checkpoint that is increased after ipilimumab therapy in patients with prostate cancer. *Nat Med.* (2017) 23:551–5. doi: 10.1038/nm.4308
  75. Rimm DL, Han G, Taube JM, Yi ES, Bridge JA, Flieder DB, et al. A prospective, multi-institutional, pathologist-based assessment of 4 immunohistochemistry assays for PD-L1 expression in non-small cell lung cancer. *JAMA Oncol.* (2017) 3:1051–8. doi: 10.1001/jamaoncol.2017.0013
  76. Hirsch FR, McElhinny A, Stanforth D, Ranger-Moore J, Jansson M, Kulangara K, et al. PD-L1 immunohistochemistry assays for lung cancer: results from phase 1 of the blueprint PD-L1 IHC assay comparison project. *J Thorac Oncol.* (2017) 12:208–22. doi: 10.1016/j.jtho.2016.11.2228
  77. Gaule P, Smithy JW, Toki M, Rehman J, Patell-Socha F, Cougot D, et al. A quantitative comparison of antibodies to programmed cell death 1 ligand 1. *JAMA Oncol.* (2017) 3:256–9. doi: 10.1001/jamaoncol.2016.3015
  78. Ratcliffe MJ, Sharpe A, Midha A, Barker C, Scott M, Scorer P, et al. Agreement between programmed cell death ligand-1 diagnostic assays across multiple protein expression cutoffs in non-small cell lung cancer. *Clin Cancer Res.* (2017) 23:3585–91. doi: 10.1158/1078-0432.CCR-16-2375
  79. Feng C, Ding G, Jiang H, Ding Q, Wen H. Loss of MLH1 confers resistance to PI3K $\beta$  inhibitors in renal clear cell carcinoma with SETD2 mutation. *Tumour Biol.* (2015) 36:3457–64. doi: 10.1007/s13277-014-2981-y
  80. Deguchi M, Shiina H, Igawa M, Kaneuchi M, Nakajima K, Dahiya R. DNA mismatch repair genes in renal cell carcinoma. *J Urol.* (2003) 169:2365–71. doi: 10.1097/01.ju.0000065668.19267.b4
  81. Rubio-Del-Campo A, Salinas-Sánchez AS, Sánchez-Sánchez F, Giménez-Bachs JM, Donate-Moreno MJ, Pastor-Navarro H, et al. Implications of mismatch repair genes hMLH1 and hMSH2 in patients with sporadic renal cell carcinoma. *BJU Int.* (2008) 102:504–9. doi: 10.1111/j.1464-410X.2008.07581.x
  82. Leach FS, Koh M, Sharma K, McWilliams G, Talifero-Smith L, Codd A, et al. Mismatch repair gene mutations in renal cell carcinoma. *Cancer Biol Ther.* (2002) 1:530–6. doi: 10.4161/cbt.1.5.171
  83. Yoo KH, Won KY, Lim SJ, Park YK, Chang SG. Deficiency of MSH2 expression is associated with clear cell renal cell carcinoma. *Oncol Lett.* (2014) 8:2135–9. doi: 10.3892/ol.2014.2482
  84. Stoehr C, Burger M, Stoehr R, Bertz S, Ruemmele P, Hofstaedter F, et al. Mismatch repair proteins hMLH1 and hMSH2 are differently expressed in the three main subtypes of sporadic renal cell carcinoma. *Pathobiology* (2012) 79:162–8. doi: 10.1159/000335642
  85. Nebot-Bral L, Brandao D, Verlingue L, Rouleau E, Caron O, Despras E, et al. Hypermutated tumours in the era of immunotherapy: the paradigm of personalised medicine. *Eur J Cancer* (2017) 84:290–303. doi: 10.1016/j.ejca.2017.07.026
  86. Llosa NJ, Cruise M, Tam A, Wicks EC, Hechenbleikner EM, Taube JM, et al. The vigorous immune microenvironment of microsatellite instable colon cancer is balanced by multiple counter-inhibitory checkpoints. *Cancer Discov.* (2015) 5:43–51. doi: 10.1158/2159-8290.CD-14-0863
  87. Le DT, Uram JN, Wang H, Bartlett BR, Kemberling H, Eyring AD, et al. PD-1 blockade in tumors with mismatch repair 1 deficiency. *N Engl J Med.* (2015) 372:2509–20. doi: 10.1056/NEJMoa1500596
  88. Iyer G, Audenet F, Middha S, Carlo MI, Regazzi AM, Funt S, et al. Mismatch repair (MMR) detection in urothelial carcinoma (UC) and correlation with immune checkpoint blockade (ICB) response. *J Clin Oncol.* (2017) 35(15\_Suppl.):4511. doi: 10.1200/JCO.2017.35.15\_suppl.4511
  89. Teo MY, Seie K, Ostrovskaya I, Regazzi AM, Kania BE, Moran MM, et al. DNA damage repair and response (DDR) gene alterations (alt) and response to PD1/PDL1 blockade in platinum-treated metastatic urothelial carcinoma (mUC). *J Clin Oncol.* (2017) 35(15\_Suppl.):4509. doi: 10.1200/JCO.2017.35.15\_suppl.4509
  90. Johnson DB, Frampton GM, Rioth MJ, Yusko E, Xu Y, Guo X, et al. Targeted next generation sequencing identifies markers of response to PD-1 blockade. *Cancer Immunol Res.* (2016) 4:959–67. doi: 10.1158/2326-6066.CIR-16-0143
  91. Kautto EA, Bonneville R, Miya J, Yu L, Krook MA, Reeser JW, et al. Performance evaluation for rapid detection of pan-cancer microsatellite instability with MANTIS. *Oncotarget* (2017) 8:7452–63. doi: 10.18632/oncotarget.13918
  92. Anagnostou V, Smith KN, Forde PM, Niknafs N, Bhattacharya R, White J, et al. Evolution of neoantigen landscape during immune checkpoint blockade in nonsmall cell lung cancer. *Cancer Discov.* (2017) 7:264–76. doi: 10.1158/2159-8290.CD-16-0828
  93. Anderson AC, Joller N, Kuchroo VK. Lag-3, tim-3, and TIGIT: co-inhibitory receptors with specialized functions in immune regulation. *Immunity* (2016) 44:989–1004. doi: 10.1016/j.immuni.2016.05.001
  94. Sharma P, Hu-Lieskovan S, Wargo JA, Ribas A. Primary, adaptive, and acquired resistance to cancer immunotherapy. *Cell* (2017) 168:707–23. doi: 10.1016/j.cell.2017.01.017
  95. Blank C, Brown I, Peterson AC, Spiotto M, Iwai Y, Honjo T, et al. PD-L1/B7H-1 inhibits the effector phase of tumor rejection by T cell receptor (TCR) transgenic CD8<sup>+</sup> T cells. *Cancer Res.* (2004) 64:1140–5. doi: 10.1158/0008-5472.CAN-03-3259
  96. Dunn GP, Koebel CM, Schreiber RD. Interferons, immunity and cancer immunoediting. *Nat Rev Immunol.* (2006) 6:836–48. doi: 10.1038/nri1961
  97. Garcia-Diaz A, Shin DS, Moreno BH, Saco J, Escuin-Ordinas H, Rodriguez GA, et al. Interferon receptor signaling pathways regulating PD-L1 and PD-L2 expression. *Cell Rep.* (2017) 19:1189–201. doi: 10.1016/j.celrep.2017.04.031
  98. Bach EA, Aguet M, Schreiber RD. The IFN gamma receptor: a paradigm for cytokine receptor signaling. *Annu Rev Immunol.* (1997) 15:563–91. doi: 10.1146/annurev.immunol.15.1.563
  99. Shin DS, Zaretsky JM, Escuin-Ordinas H, Garcia-Diaz A, Hu-Lieskovan S, Kalbasi A, et al. Primary resistance to PD-1 blockade mediated by JAK1/2 mutations. *Cancer Discov.* (2017) 7:188–201. doi: 10.1158/2159-8290.CD-16-1223
  100. Zaretsky JM, Garcia-Diaz A, Shin DS, Escuin-Ordinas H, Hugo W, Hu-Lieskovan S, et al. Mutations associated with acquired resistance to PD-1 blockade in melanoma. *N Engl J Med.* (2016) 375:819–29. doi: 10.1056/NEJMoa1604958
  101. Spranger S, Koblisch HK, Horton B, Scherle PA, Newton R, Gajewski TF. Mechanism of tumor rejection with doublets of CTLA-4, PD-1/PD-L1, or IDO blockade involves restored IL-2 production and proliferation of CD8(+) T cells directly within the tumor microenvironment. *J Immunother Cancer* (2014) 2:3. doi: 10.1186/2051-1426-2-3
  102. McGranahan N, Rosenthal R, Hiley CT, Rowan AJ, Watkins TBK, Wilson GA, et al. Allele-specific HLA loss and immune escape in lung cancer evolution. *Cell* (2017) 171:1259–1271.e11. doi: 10.1016/j.cell.2017.10.001
  103. Koyama S, Akbay EA, Li YY, Herter-Sprie GS, Buczkowski KA, Richards WG, et al. Adaptive resistance to therapeutic PD-1 blockade is associated with upregulation of alternative immune checkpoints. *Nat Commun.* (2016) 7:10501. doi: 10.1038/ncomms10501
  104. Montironi R, Santoni M, Massari F, Lopez-Beltran A, Cheng L, Berardi R, et al. Emerging immunotargets in genitourinary tumors. *Curr Drug Targets* (2016) 17:748–9. doi: 10.2174/138945011707160412185542
  105. Montironi R, Santoni M, Cheng L, Lopez-Beltran A, Massari F, Matrana MR, et al. An overview of emerging immunotargets of genitourinary tumors. *Curr Drug Targets* (2016) 17:750–6. doi: 10.2174/1389450117666151209144649
  106. Slovin SF. The need for immune biomarkers for treatment prognosis and response in genitourinary malignancies. *Biomark Med.* (2017) 11:1149–59. doi: 10.2217/bmm-2017-0138
  107. Ciccamese C, Alfieri S, Santoni M, Santini D, Bergamini C, Licitra L, et al. New toxicity profile for novel immunotherapy agents: focus on checkpoint inhibitors. *Expert Opin Drug Metab Toxicol.* (2016) 12:57–75. doi: 10.1517/17425255.2016.1120287
  108. Iacovelli R, Nolè F, Verri E, Paglino C, Santoni M, Cossu Rocca M, et al. Prognostic role of PD-L1 expression in renal cell carcinoma. A systematic review and meta-analysis. *Target Oncol.* (2016) 11:143–8. doi: 10.1007/s11523-015-0392-7



109. Piva F, Giulietti M, Occhipinti G, Santoni M, Massari F, Sotte V, et al. Computational analysis of the mutations in BAP1, PBRM1 and SETD2 genes reveals the impaired molecular processes in renal cell carcinoma. *Oncotarget* (2015) 6:32161–8. doi: 10.18632/oncotarget.5147
110. Santoni M, Santini D, Massari F, Conti A, Iacovelli R, Burattini L, et al. Heterogeneous drug target expression as possible basis for different clinical and radiological response to the treatment of primary and metastatic renal cell carcinoma: suggestions from bench to bedside. *Cancer Metastasis Rev.* (2014) 33:321–31. doi: 10.1007/s10555-013-9453-5
111. Ciccarese C, Santoni M, Massari F, Cheng L, Lopez-Beltran A, Scarpelli M, et al. Present and future of personalized medicine in adult genitourinary tumors. *Fut Oncol.* (2015) 11:1381–8. doi: 10.2217/fon.15.30
112. Tartari F, Santoni M, Burattini L, Mazzanti P, Onofri A, Berardi R. Economic sustainability of anti-PD-1 agents nivolumab and pembrolizumab in cancer patients: recent insights and future challenges. *Cancer Treat Rev.* (2016) 48:20–4. doi: 10.1016/j.ctrv.2016.06.002
113. Santoni M, Rizzo M, Burattini L, Farfariello V, Berardi R, Santoni G, et al. Present and future of tyrosine kinase inhibitors in renal cell carcinoma: analysis of hematologic toxicity. *Recent Pat Antinfect Drug Discov.* (2012) 7:104–10. doi: 10.2174/157489112801619719
114. Pessina A, Albella B, Bueren J, Brantom P, Casati S, Gribaldo L, et al. Prevalidation of a model for predicting acute neutropenia by colony forming unit granulocyte/macrophage (CFU-GM) assay. *Toxicol In Vitro.* (2001) 6:729–40. doi: 10.1016/S0887-2333(01)00085-6
115. Hipp MM, Hilf N, Walter S, Werth D, Brauer KM, Radsak MP, et al. Sorafenib, but not sunitinib, affects function of dendritic cells and induction of primary immune responses. *Blood* (2008) 111:5610–20. doi: 10.1182/blood-2007-02-075945
116. Huang Y, Wang Y, Li Y, Guo K, He Y. Role of sorafenib and sunitinib in the induction of expressions of NKG2D ligands in nasopharyngeal carcinoma with high expression of ABCG2. *J Cancer Res Clin Oncol.* (2011) 137:829–37. doi: 10.1007/s00432-010-0944-2
117. Powles T, Chowdhury S, Bower M, Saunders N, Lim L, Shamash J, et al. The effect of sunitinib on immune subsets in metastatic clear cell renal cancer. *Urol Int.* (2011) 86:53–9. doi: 10.1159/000319498
118. Adotevi O, Pere H, Ravel P, Haicheur N, Badoual C, Merillon N, et al. A decrease of regulatory T cells correlates with overall survival after sunitinib-based antiangiogenic therapy in metastatic renal cancer patients. *J Immunother.* (2010) 33:991–8. doi: 10.1097/CJL.0b013e3181f4c208
119. Kumar R, Crouthamel M-C, Rominger DH, Gontarek RR, Tummino PJ, Levin RA, et al. Myelosuppression and kinase selectivity of multikinase angiogenesis inhibitors. *Br J Cancer* (2009) 101:1717–23. doi: 10.1038/sj.bjc.6605366
120. Morelli MB, Amantini C, Santoni M, Soriani A, Nabissi M, Cardinali C, et al. Axitinib induces DNA damage response leading to senescence, mitotic catastrophe, and increased NK cell recognition in human renal carcinoma cells. *Oncotarget* (2015) 6:36245–59. doi: 10.18632/oncotarget.5768
121. Ritprajak P, Azuma M. Intrinsic and extrinsic control of expression of the immunoregulatory molecule PD-L1 in epithelial cells and squamous cell carcinoma. *Oral Oncol.* (2015) 51:221–8. doi: 10.1016/j.oraloncology.2014.11.014

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Lopez-Beltran, Henriques, Cimadamore, Santoni, Cheng, Gevaert, Blanca, Massari, Scarpelli and Montironi. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.



# New Prostate Cancer Targets for Diagnosis, Imaging, and Therapy: Focus on Prostate-Specific Membrane Antigen

Alessia Cimadamore<sup>1\*</sup>, Monica Cheng<sup>2</sup>, Matteo Santoni<sup>3</sup>, Antonio Lopez-Beltran<sup>4</sup>, Nicola Battelli<sup>3</sup>, Francesco Massari<sup>5</sup>, Andrea B. Galosi<sup>6</sup>, Marina Scarpelli<sup>1</sup> and Rodolfo Montironi<sup>1\*</sup>

<sup>1</sup> Section of Pathological Anatomy, School of Medicine, United Hospitals, Polytechnic University of the Marche Region, Ancona, Italy, <sup>2</sup> Department of Radiology and Imaging Sciences, Indiana University School of Medicine, Indianapolis, IN, United States, <sup>3</sup> Oncology Unit, Macerata Hospital, Macerata, Italy, <sup>4</sup> Department of Pathology and Surgery, Faculty of Medicine, Cordoba, Spain, <sup>5</sup> Division of Oncology, S. Orsola-Malpighi Hospital, Bologna, Italy, <sup>6</sup> Institute of Urology, School of Medicine, United Hospitals, Marche Polytechnic University, Ancona, Italy

## OPEN ACCESS

### Edited by:

Scott T. Tagawa,  
Cornell University, United States

### Reviewed by:

Jaspreet Singh Batra,  
Johns Hopkins Medicine,  
United States  
Daniele Baiz,  
Plymouth University, United Kingdom

### \*Correspondence:

Alessia Cimadamore  
alessiacimadamore@gmail.com  
Rodolfo Montironi  
r.montironi@univpm.it

### Specialty section:

This article was submitted to  
Genitourinary Oncology,  
a section of the journal  
Frontiers in Oncology

**Received:** 02 July 2018

**Accepted:** 10 December 2018

**Published:** 21 December 2018

### Citation:

Cimadamore A, Cheng M, Santoni M, Lopez-Beltran A, Battelli N, Massari F, Galosi AB, Scarpelli M and Montironi R (2018) New Prostate Cancer Targets for Diagnosis, Imaging, and Therapy: Focus on Prostate-Specific Membrane Antigen. *Front. Oncol.* 8:653. doi: 10.3389/fonc.2018.00653

The rising incidence rate of the cancer in the prostate gland has increased the demand for improved diagnostic, imaging, and therapeutic approaches. Prostate-specific membrane antigen (PSMA), with folate hydrolase and carboxypeptidase and, internalization activities, is highly expressed in the epithelial cells of the prostate gland and is strongly upregulated in prostatic adenocarcinoma, with elevated expression correlating with, metastasis, progression, and androgen independence. Recently, PSMA has been an active target of investigation by several approaches, including the successful utilization of small molecule inhibitors, RNA aptamer conjugates, PSMA-based immunotherapy, and PSMA-targeted prodrug therapy. Future investigations of PSMA in prostate cancer (PCa) should focus in particular on its intracellular activities and functions. The objective of this contribution is to review the current role of PSMA as a marker for PCa diagnosis, imaging, and therapy.

**Keywords:** prostate cancer, prostate-specific membrane antigen, PSMA, small molecule inhibitors, RNA aptamer conjugates, PSMA-based immunotherapy, PSMA-targeted prodrug therapy, positron emission tomography

## INTRODUCTION

Prostate-specific membrane antigen (PSMA) is a type 2 integral membrane glycoprotein with folate hydrolase and carboxypeptidase, and internalization activities. This internalization capability is increased up to 3-fold when PSMA is linked to anti-PSMA antibodies. PSMA expression is highest in prostate tissue (secretory acinar epithelium), but detectable levels of PSMA protein are also found in the kidney (proximal tubules), the small bowel (i.e., jejunal brush border), neuroglia (Schwann cells and astrocytes), and salivary glands (1, 2). Notably, PSMA is highly expressed in prostate cancer cells and the vessels of various non-prostatic solid tumors (it is not expressed in the normal vasculature) (3).

With the rise and evolution of several targeted approaches to examine prostate cancer using PSMA, the aim of this contribution is to review the current role of PSMA as a marker for PCa diagnosis, imaging, and therapy.

## EXPRESSION AND ROLE OF PSMA IN PCA

PSMA was originally discovered using the monoclonal antibody 7E11 obtained from the cell membrane of the LNCaP cell line (4). It has been shown by immunohistochemistry that expression of PSMA at the tissue level increases through the progression from normal prostate cells to high-grade prostatic intraepithelial neoplasia (HGPIN) and to PCa (3) (**Figure 1**). There exists a strong positive correlation between PSMA expression and Gleason score. Elevated PSMA expression is strongly correlated with a high serum PSA. These indications are associated with increased tumor angiogenesis and lack of ets-related gene (ERG) expression which leads to reduced vitamin D and androgen receptor expression (5). PSMA expression is regulated by the androgen receptor (AR). PSMA expression increases dramatically during androgen-deprivation therapy (6).

Downregulation of PSMA expression by AR may be associated to the presence of an enhancer region although no androgen response elements have been identified (7).

PSMA expression is significantly correlated with prostate growth and differentiation (8). In particular, *in vitro* expression of PSMA is associated with an increased cellular folate content. This induces a proliferative property to cells expressing PSMA (9, 10). In addition, PSMA stimulates PCa cell proliferation, migration and survival through the phospho-p38 (P-p38) MAPK pathway in LNCaP cancer cells (11). Guo et al. demonstrated that PSMA knockdown in a LNCaP cell line was associated with not only the inhibition of the pathway of phosphatidylinositol 3-kinase/Akt signaling but also decreased cell proliferation, migration and survival (12).

PSMA is involved in the development of PCa metastases. Xu et al. evaluated four prostate cancer cell lines (i.e., DU145, LNCap, PC-3, and 22RV1) for metastasis-related genes potentially involved in PCa metastasis regulated by PSMA. In their study, *CDH6*, *MMP3*, and *MTSS1* were seen as PSMA-related genes. Their expression was inversely related with the stage of cancer, thus suggesting their possible involvement in the suppression of PCa metastasis by PSMA (13).

## PSMA-BASED IMAGING IN PATIENTS WITH PCA

Conventional imaging techniques, such as ultrasound, CT, bone scintigraphy and Magnetic Resonance Imaging (MRI), are at present utilized to detect primary PCa and its metastatic deposits. However, the limitation of such traditional imaging techniques and modalities is their low sensitivity in the detection of recurrent or/and metastatic PCa. Improved imaging modalities are needed to optimize the management of the patients with PCa.

Positron Emission Tomography (PET) and single photon emission computed tomography (SPECT) with emerging radiopharmaceuticals provide more accurate staging for primary cancer, detection of metastatic disease, and restaging of tumor recurrence. PSMA has received considerable attention as a useful marker for imaging purposes in patients with PCa (14, 15).

Several PSMA-based approaches have been developed, including antibodies, nanobodies, and small molecule inhibitors.

### Antibodies and Nanobodies

Indium-111 capromab pendetide ( $^{111}\text{In}$ -capromab, ProstaScint®) was the first monoclonal antibody against PSMA used in PCa immunoscintigraphy. Correlation of scan results with tissue specimens showed that  $^{111}\text{In}$ -capromab detected soft tissue metastases, with an average negative predictive value of 70%, sensitivity of 60%, and positive predictive value of 60% (16–18). However,  $^{111}\text{In}$ -capromab lacks sensitivity because it recognizes an intracellular epitope of PSMA, thereby targeting only apoptotic/necrotic or damaged cells.

Unlike  $^{111}\text{In}$ -capromab, J591 is an antibody against the extracellular domain of PSMA.  $^{111}\text{In}$ -labeled J591 has been evaluated against conventional imaging techniques in the evaluation of bone metastases.  $^{111}\text{In}$ -labeled J591 identifies 93.7% of skeletal lesions detected by a conventional imaging technique. Thirteen out of Eighteen bone deposits detected only with  $^{111}\text{In}$ -labeled J591 were successively confirmed to be metastases (19). In a more recent study, J591 has been radiolabeled with  $^{89}\text{Zr}$  (20) and  $^{64}\text{Cu}$  (21) for PET imaging and demonstrate robust targeting of skeletal, nodal and soft tissue metastasis (22).

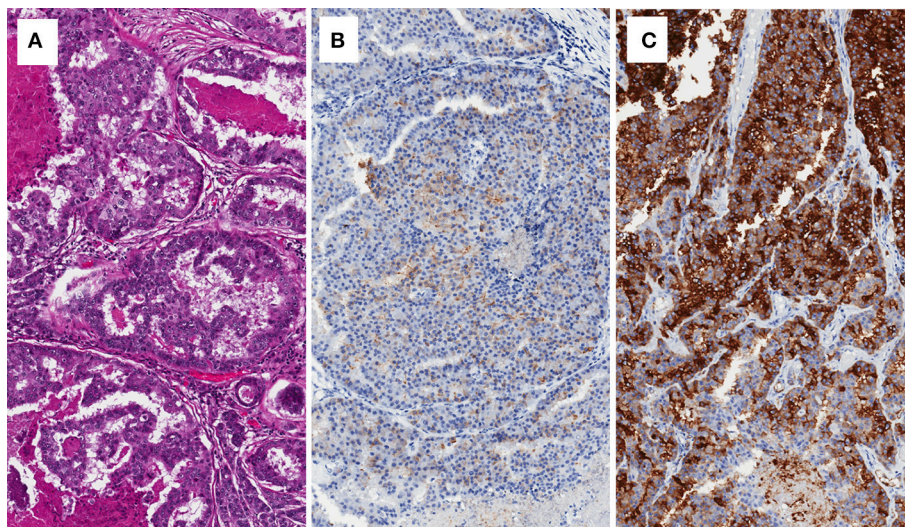
A new strategy in the development of high-contrast nuclear imaging is the utilization of specific antibody fragments, called nanobodies. Nanobodies contain antibody-derived smaller fragments (typically the variable domain alone of heavy chain antibodies) that largely retain the specific antigen binding properties of the original antibodies, but with more rapid pharmacokinetics and lower immunogenic potential. Evazalipour et al. compared the properties of different nanobodies radiolabeled with 99 m-Technetium ( $^{99\text{m}}\text{Tc}$ ) in PSMA<sup>+</sup> LNCaP and PSMA<sup>-</sup> PC3 cell lines and in PSMA<sup>-</sup> and PSMA<sup>+</sup> tumor-bearing xenografts through SPECT/micro-CT imaging and tissue analysis. Among the evaluated molecules, nanobody PSMA30 resulted in an important compound for future applications in PCa imaging trials (23).

Interesting results were also obtained with minibodies, i.e., IAB2M, an 80-kDa minibody genetically engineered from the parent antibody J591 that targets the extracellular domain of PSMA. A phase I dose-escalation study in patients with metastatic prostate cancer demonstrated PET imaging with  $^{89}\text{Zr}$ -Df-IAB2M is feasible and well tolerated, and targets both bone and soft-tissue disease (24).

### Small Molecules

The identification of the functional (25) and structural (26) homology between N-acetylaspartylglutamate peptidase or NAAALDASE (for which a number of enzymatic inhibitors had been identified) (27, 28) and PSMA has been a major step forward for the development of PSMA-targeted radiotracers. Generally, small molecule PSMA inhibitors consist of zinc binding compounds linked to a glutamate isostere or glutamate. Phosphonate-, phosphate-, and phosphoramidates (1) and ureas (2) constitute the two main families of compounds. Based on NAAALDASE homology, several compounds have been





**FIGURE 1** | Brain metastasis of prostate cancer with cribriform pattern (A), showing low expression of PSA (B), and intense expression of PSMA (C).

developed and labeled with  $^{123}\text{I}$  (20, 29, 30),  $^{99\text{m}}\text{Tc}$  (21, 31),  $^{18}\text{F}$  (32),  $^{111}\text{In}$  (33), and  $^{68}\text{Ga}$  (34).

$^{123}\text{I}$ -MIP-1072 and  $^{123}\text{I}$ -MIP-1095 were the first small molecule inhibitors of PSMA adopted in the clinic. SPECT/CT using these compounds showed a rapid detection of PCa deposits in the bone, soft tissue, and prostate gland of men with metastatic PCa (35). A phase I trial on  $^{131}\text{I}$ -MIP-1095 in men with mCRPC is now active (NCT03030885).

Among the emerging PSMA small molecule inhibitors, *N*-(*N*-(*S*)-1,3-dicarboxypropyl) carbamoyl)-4-( $^{18}\text{F}$ )fluorobenzyl-L-cysteine ( $^{18}\text{F}$ -DCFBC) is under evaluation in several ongoing studies. Using  $^{18}\text{F}$ -DCFBC, PSMA<sup>+</sup> PC-3 PIP xenografts were early visualized with little radioactivity in the PSMA<sup>-</sup> isogenic PC-3 flu xenografts. After 2 h, the PC-3 PIP xenografts remained visible, with clearance of background radioactivity from kidneys, liver and blood (36, 37).

The use of  $^{18}\text{F}$ -DCFBC has been investigated in a few patients with Gleason scores between 7 and 9 and with radiological evidence of metastatic PCa. Bone scans or CT identified 21 lesions (5 bone and 16 lymph node lesions), while 32 lesions were visible with  $^{18}\text{F}$ -DCFBC PET. Ten of Eleven additional lesions were located in the bone and were suggestive of early bone deposits, indicating the potential of  $^{18}\text{F}$ -DCFBC PET in this subpopulation (38). Currently, the use of  $^{18}\text{F}$ -DCFBC PET/CT is under evaluation in a study enrolling patients scheduled for surgical prostate (Group 1), or with biochemical recurrence after surgery or radiotherapy (Group 2), or in metastatic PCa patients (Group 3) (NCT02190279). In addition, another ongoing phase I/II study is assessing the potential of  $^{18}\text{F}$ -DCFBC PET in the detection of primary PCa, nodal and bone metastases in men at initial diagnosis (NCT01496157) (Table 1).

As for the PSMA inhibitor  $^{18}\text{F}$ -DCFpyL (2-(3-{1-carboxy-5-((6-(( $^{18}\text{F}$ )fluoro-pyridine-3-carbonyl)-amino)-pentyl}-ureido)-pentanedioic acid), Chen et al. evaluated its use in immunocompromised mice utilizing isogenic PSMA PC3 PIP

and PSMA- PC3 flu xenografts, suggesting that this agent could be viable and effective in this setting (32). A phase I study is now assessing the biodistribution and pharmacokinetic of  $^{18}\text{F}$ -DCFpyL in patients with advanced PCa (NCT02151760).

The early distinction between local disease and metastasis is crucial in the management of patients with PCa.  $^{18}\text{F}$ -choline can distinguish lesions with moderate to good sensitivity, but its activity is limited to patients with a PSA >1 ng/mL (39). The results obtained by  $^{68}\text{Ga}$ -labeled PSMA inhibitors showed a high potential in the detection of small recurrent PCa lesions in patients with low levels of serum PSA (40–42). Indeed,  $^{68}\text{Ga}$ -labeled PSMA inhibitors are characterized by accumulation in small metastatic deposits and a rapid clearance from the tissue in the background (43). Recently, a comparison between PET/CT and PET/MRI hybrid systems using a  $^{68}\text{Ga}$ -labeled PSMA compound for the detection of recurrent PCa has been performed. The results showed that Ga-PSMA PET/MRI was far more accurate in the detection of PCa and, at the same time, associated with lower radiation exposure (34).

Beyond  $^{68}\text{Ga}$ -labeled compounds,  $^{99\text{m}}\text{Tc}$ -labeled inhibitors of PSMA have shown great promise in the detection of PCa lesions. Presently, a phase II study is testing  $^{99\text{m}}\text{Tc}$ -MIP-1404 PSMA inhibitor in patients with high-risk PCa scheduled for radical prostatectomy (RP) surgery including extended pelvic lymph node (LN) dissection compared to histopathology (NCT01667536). Results are expected from the completed phase 3 trial proSPECT-AS (NCT02615067). Primary outcome measures of the study are sensitivity and specificity of  $^{99\text{m}}\text{Tc}$ -MIP-1404 SPECT/CT image assessments to correctly detect clinically significant prostate cancer when compared to histopathology following either RP or prostate biopsy in men with newly diagnosed PCa whose biopsy indicates a histopathologic Gleason Score of  $\leq 3 + 4$ .

Furthermore, BAY1075553 [2-PMPA analogs (2*S*, 4*S*)-2- $^{18}\text{F}$ -fluoro-4-(phosphonomethyl) pentanedioic acid] has



**TABLE 1 |** Current completed trial on PSMA-based imaging.

NCT Identifier	Study phase	Tracer and technique	Study outcomes
NCT01496157	Phase 1 Phase 2	18F-DCFBC PET	PET detection of primary and sextant localization of PCa and detection of metastatic disease at initial staging
NCT02048150	Phase 1	MDX1201-A488	Assess the best dose given before robotic assisted laparoscopic RP to aid in visualization of the prostate
NCT02151760	Phase 1	18F-DCFPyL	Compare diagnostic accuracy of 18-DCFPyL to CT and bone scintigraphy for the detection of metastatic PCa
NCT01667536	Phase 2	99mTc-MIP-1404	Assessment of the diagnostic accuracy in detection PCa within the prostate and metastatic PCa
NCT03558711	Phase 1	18F-PSMA	Safety of administration
NCT02611882	Phase 1 Phase 2	Ga-68 HBED-CC PSMA	SE and SP for the detection of nodal metastasis in high-risk pre-RP patients, of metastatic disease in patients with BCR after RP or radiation therapy, comparison to conventional imaging in CRPC patients.
NCT03486886	Not Applicable	PSMA -PET/CT	Evaluation of detection Yield performance and reproducibility in mPCa Patients
NCT02796807	Phase 2	68Ga-HBED-CC- PSMA PET/CT	Correlation Between SUV on 68Ga-HBED-CC-PSMA and GS in PCa
NCT02488070	Phase 1 Phase 2	68Ga-PSMA PET/CT	Average SUVmax of Ga68 PSMA Uptake Outside the Expected Normal Biodistribution
NCT01359189	Phase 1	ProxiScan	Evaluation of a Transrectal Scintigraphic Detector (ProxiScan™) for Detection of Primary PCa
NCT02918357	Phase 2 Phase 3	Ga-68 labeled PSMA-11 PET	Sensitivity and PPV of for the detection of metastases a per-patient and per-region-basis confirmed by histopathology
NCT02916537	Phase 1	CTT1057 PET/MR	Evaluation of the safety, pharmacokinetics, and [18F] radiation dosimetry
NCT00712829	Phase 1	123-I-MIP-1072 123-I-MIP-1095	Evaluating the safety, pharmacokinetics, tissue distribution
NCT01615406	Phase 1	99mTc MIP 1404	Comparison study of 99mTc-MIP-1404 (SPECT)/CT imaging to histology
NCT01261754	Phase 1	99mTc-MIP-1404 and MIP-1405	Safety, pharmacokinetics, biodistribution in mPCa patients; newly diagnosed, high-risk PCa and healthy subjects
NCT01572701	Phase 1	99mTc-MIP-1404	Measure activity counts in tissue samples post-surgery, Intensity of 99mTc-MIP-1404 Uptake with Respect to PSMA expression in Men With PCa Undergoing RP and/or PLND
NCT02190279	Early phase 1	18F-DCFBC PET/CT	Assess the ability to identify sites of localized, recurrent and metastatic PCa
NCT00992745	Phase 1	123-I-MIP-1072	Estimate the imaging SE and SP of 123I MIP 1072 compared to 111In capromab pendetide in mPCa
NCT02349022	Phase 2	[89Zr]Df-IAB2M	Compare SE/SP/PPV/NPV/Accuracy of [111In] capromab pendetide SPECT/CT to [89Zr]-Df-IAB2M PET/CT as confirmed by pathology
NCT02615067	Phase 3	99mTc-MIP-1404 SPECT/CT	Safety and Efficacy of 99mTc-MIP-1404 SPECT/CT Imaging to Detect Clinically Significant PCa in Men With Biopsy Proven Low-Grade PCa Candidates for AS (proSPECT-AS)
ACTRN1261700005358	Phase 2	Ga-68 PSMA-PET/CT	Compare the diagnostic accuracy of Ga-68 PSMA-PET/CT to that of conventional imaging for detecting nodal or distant metastatic disease.

Source: <https://clinicaltrials.gov>.

PSMA, Prostate specific membrane antigen; RP, radical prostatectomy; MDX1201-A488, Anti-PSMA monoclonal antibody; PCa, prostate cancer; SE, sensitivity; SP, specificity; BCR, biochemical recurrence; CRPC, castration resistant prostate cancer; GS, Gleason score; PPV, positive predictive value; 123-I-MIP-1072, MIP 1404, MIP-1405, small molecule inhibitor of PSMA; PLND, Pelvic Lymph Node Dissection; NPV, negative predictive value; AS, active surveillance.

demonstrated high uptake in PSMA<sup>+</sup> LNCaP tumor xenografts (44). The phase I study showed that BAY1075553 was able to detect primary PCa, lymph node and bone metastases, although its high uptake with degenerative bone lesions may limit its use in assessing bone disease (45).

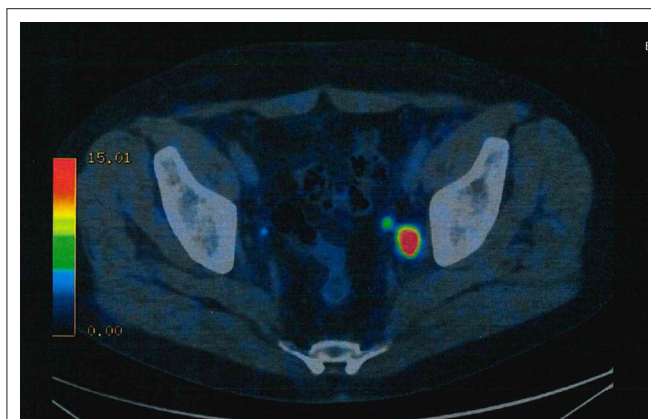
Worth mentioning is the registrational phase II/III OSPREY study (NCT02981368) that evaluated the diagnostic accuracy of 18F-DCFPyL PET/CT relative to histopathology, for detecting PCa in pelvic lymph nodes in patients with high risk localized

prostate cancer who are planned for RP with lymphadenectomy, and in patients with locally recurrent or metastatic disease willing to undergo biopsy.

## Imaging at Diagnosis of PCa

A number of recent studies has dealt with the use of PSMA-based imaging for the purpose of diagnosing primary PCa (**Figure 2**).

Fendler et al. assessed the accuracy of <sup>68</sup>Ga-PSMA-11 PET/CT in identifying PCa at the initial diagnosis in men with



**FIGURE 2 |**  $^{68}\text{Ga}$ -PSMA ligand PET/CT exhibits solitary left iliac radiotracer-positive lymph node.

biopsy-proven PCa (46). They found that the optimal  $\text{SUV}_{\text{max}}$  cutoff for distinction of histopathology-positive segments from histopathology-negative segments is of 6.5. With this approach they obtained 67% sensitivity, 92% specificity, 97% positive predictive value, and 72% accuracy.

Woythal et al. (47) evaluated the association of intraprostatic  $^{68}\text{Ga}$ -PSMA PET/CT features and PSMA immunohistochemical expression in 31 patients who underwent RP and preoperative  $^{68}\text{Ga}$ -PSMA-11 PET/CT.  $^{68}\text{Ga}$ -PSMA-11 PET/CT demonstrated sensitivity and specificity of 87 and 97%, respectively, in the detection of PCa. However, there was no correlation between Gleason Score (GS) and the  $\text{SUV}_{\text{max}}$  of PCa.

On the other hand, Uprimny et al. (48) found that PCa with a GS of 6, 7a (3 + 4) and 7b (4 + 3) showed lower  $^{68}\text{Ga}$ -PSMA-11 uptake, with  $\text{SUV}_{\text{max}}$  of 5.9, 8.3, and 8.2, respectively, compared to men with a GS greater than 7 (median  $\text{SUV}_{\text{max}}$ : 21.2). In addition, men with a PSA of 10.0 ng/mL or above it showed a greater uptake than those patients with PSA levels below 10.0 ng/mL.

The correlation of intraprostatic PSMA uptake with clinical parameter, such as PSA value, GS and d'Amico risk score, was analyzed by Koerber et al. in 104 patients with newly diagnosed PCa (49). Results of this study indicated that men with higher PSA, higher d'Amico risk score and higher GS had greater intensity of PSMA uptake on PET/CT.

The comparison between the multiparametric Magnetic Resonance Imaging (mpMRI) and  $^{68}\text{Ga}$ -PSMA-11 PET/CT findings showed a concordance in the detection of intraprostatic tumor lesions, with the highest GS of 89.55%. By giving additional molecular imaging information to the mpMRI features, this method can be improved to avoid false-negative results or understaging tumors, in particular the detection of those with the highest GSs. In addition, PSMA PET/ MRI may prove useful in finding lower rates of indolent cancer detection and a great number of intermediate- and high-risk tumors.

## Imaging at Staging of PCa

For pre-operative staging, current guidelines recommend at least abdomino-pelvic cross-sectional imaging (MRI or CT) and a

bone scan, for intermediate- and high-risk PCa (50) only. In a prospective study 30 patients with intermediate- and high-risk PCa underwent preoperative  $^{68}\text{Ga}$ -PSMA PET/CT followed by RP and extended pelvic LN dissection. Using pathology as reference,  $^{68}\text{Ga}$ -PSMA PET/CT showed a sensitivity of 64% for the evaluation of LN metastasis, with a 95% specificity, 88%, positive predictive value, and 82% negative predictive value (51).

In a prospective, phase II, single center study, Gorin et al. analyzed the diagnostic value of PSMA targeted  $^{18}\text{F}$ -DCFPyL PET/CT in the preoperative staging of 25 patients considered to be at high risk for having metastatic PCa, despite a negative conventional staging result. With this technique, they obtained a sensitivity and specificity of 71 and 88%, respectively, per patient analysis and 66 and 92% per LN packet analysis (52).

The retrospective study conducted by Maurer et al. (53) involved a 130 men with intermediate and high risk PCa staged with  $^{68}\text{Ga}$ -PSMA-PET/magnetic resonance tomography or PET/CT. The sensitivity, specificity and accuracy of  $^{68}\text{Ga}$ -PSMA-PET were 65.9, 98.9, and 88.5%, and those of morphological imaging were 43.9, 85.4, and 72.3%, respectively. Such figures are higher than those for traditional imaging techniques and other alternative PET tracers. Hence, the addition of  $^{68}\text{Ga}$ -PSMA PET to traditional approaches has the potential to replace current standard imaging, enabling more complete and accurate primary staging.

## Imaging at Biochemical Recurrence of PCa

In men with biochemical recurrence (BCR) after RP or radiotherapy the detection rate of  $^{68}\text{Ga}$ -PSMA PET/CT increases with higher pre-scan PSA value. In the post-RP patients the rate of  $^{68}\text{Ga}$ -PSMA PET/CT was 11.3, 26.6, 53.3, 79.1, and 95.5% for serum PSA levels of 0.01 to <0.2 ng/mL, 0.2 to <0.5 ng/mL, 0.5 to <1 ng/mL, 1 to <2 ng/mL, and  $\geq 2$  ng/mL, respectively. In the post-radiotherapy patients, the rate was 33.3% for PSA 0.01 to <0.5 ng/mL, 71.4% for PSA 0.5 to <1 ng/mL, 93.3% for PSA 1 to <2 ng/mL, and 100% for PSA  $\geq 2$  ng/mL (54). Such figures are in agreement with the meta-analysis data by Perera et al. (55). In that study, on per-patient analysis, the sensitivity and specificity of  $^{68}\text{Ga}$ -PSMA-11 PET were both 86%. On per-lesion analysis, the sensitivity and specificity were 80 and 97%.  $^{68}\text{Ga}$ -PSMA PET positivity increased with a shorter PSA doubling time.

Higher rates have been reported by Raucher et al. (56) in a cohort of men with PSA value between 0.2 and 1 ng/mL after RP. The rate of detection was 55% in men with "very low" serum PSA (0.2–0.5 ng/mL) and of 74% in patients with "low" PSA (0.5–1.0 ng/mL). In such investigation the most relevant predictors for  $^{68}\text{Ga}$ -PSMA-ligand PET/CT positivity in multivariable analysis were concurrent androgen deprivation therapy and serum PSA value. Identification of the sites of recurrent disease is of great importance, thus avoiding unnecessary localized treatments in patients of systemic recurrence and avoid the side effects of systemic treatments in men with localized recurrence (57, 58).

**Table 1** summarizes the completed trials on PSMA and imaging. For additional trials please visit: <https://clinicaltrials.gov>.

$^{18}\text{F}$ -fluciclovine (Axumin<sup>®</sup>) ( $^{18}\text{F}$ -FACBC) is an amino-acid targeting radiotracer and not a PSMA based PET/CT agent (59).

The sensitivity of 18F-fluciclovine PET for identifying recurrent disease changes with PSA levels, with reported detection rates in the post-prostatectomy biochemical failure setting of 72.0% (for PSA values of less than 1 ng/mL) 83.3% (for PSA 1–2 ng/mL), and 100% for PSA levels of 2 or more ng/mL (60). In patients with pathologically enlarged lymph nodes, presence of true-positive lesions was noted in 29% patients with 18F-fluciclovine vs. 7% patients with CT (61, 62). A prospective study compared overall detection rate of 18F-FACBC and 11C-Choline PET/CT on 28 patients with biochemical relapse after RP. Anti-3-18F-FACBC PET/CT detected 60% additional tumor lesions including 5 (17.8%) additional patients (63).

## PSMA-Targeting Strategies for PCa Therapy

PSMA has been widely utilized as a target antigen due to its constitutive or induced internalization property as well as to its high expression in PCa. Several strategies, including peptides, monoclonal antibodies and aptamers, have been utilized as nanoparticles or prodrugs to improve targeting efficiency in PCa cells. The discovery and development of anticancer aptamers may prove to be relevant contribution to PCa molecular imaging.

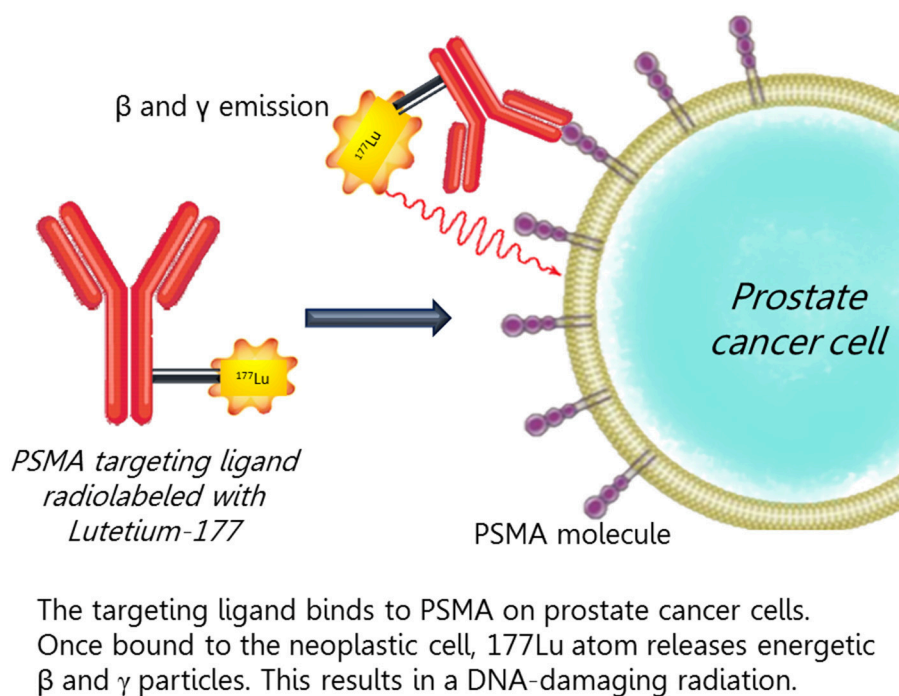
Aptamers are short DNA, RNA or peptide oligomers able to assume a specific and stable three-dimensional shape *in vivo* (64). Their high affinity and specificity, similar to antibodies, is achieved by a three-dimensional conformation complementary to the target surface. At this regard, Lupold et al. identified two RNA aptamers (A9 and A10) characterized

with high binding affinity to PSMA, leading to the inhibition of its NAALADase/glutamate carboxypeptidase II activity (65). Successively, Xu et al. conjugated A10 aptamer on the surface of micelles, showing high drug uptake in PSMA<sup>+</sup> cancer cells both *in vitro* and *in vivo* investigations (66).

PSMA can be used as target for delivery of therapeutic agents such as in antibody-drug conjugated (ADC) therapy. PSMA ADC is a fully human anti-PSMA monoclonal antibody conjugated to monomethyl auristatin E through a valine-citrulline linker.

Wang and his group assessed the antitumor activity of PSMA ADC in PCa cell lines *in vitro* and in a novel *in vivo* model of taxane-refractory human PCa. They observed that *in vitro* cytotoxic activity was efficient for PCa cells with increased PSMA expression (>105 molecules/cell; IC<sub>50</sub> 0.022 nmol/L). In addition, PSMA ADC showed high *in vivo* activity in treating xenograft tumors that have progressed on previous docetaxel therapy (67).

Petrylak et al. (68) reported data from a phase II trial based on PSMA-ADC at 2.5 mg/kg in patients with taxane-refractory metastatic castration-resistant PCa (CRPCa). Thirty-Nine Percent of the patients had been treated with both cabazitaxel and docetaxel, while 58% had received both enzalutamide and abiraterone. Dosing was started at 2.5 mg/kg and adjusted at 2.3 mg/kg for tolerability. The study demonstrated that PSA decline of 30% or more was observed in 36% (2.3 mg/kg) and 16% (2.5 mg/kg). Circulating tumor cell (CTC) decline of ≥50% was seen in 74% patients in both 2.3 and 2.5 mg/kg. Duration of therapy on 2.3 mg/kg was far longer than on 2.5 mg/kg, as well as the rate of



**FIGURE 3 |** The targeting ligand binds to PSMA on prostate cancer cells. Once bound to the neoplastic cell, <sup>177</sup>Lu atom releases an energetic beta and gamma particles that results in a DNA-damaging radiation.

serious adverse events (37 vs. 59%). Notably, PSA and CTC decline was associated with higher PSMA expression + CTC level, while PSA responses alone were correlated with lower neuroendocrine (NE) marker expression, thus suggesting that NE differentiation may have a role in this context. On the basis of such results, this study has been further extended (see NCT02020135).

Phage display technology has been used by researchers in the identification of peptide sequences, which can bind to PSMA and, at the same time, inhibit its enzymatic activity. Denmeade

et al. conjugated a PSMA-specific peptide to an inhibitor (i.e., Thapsigargin) of the sarcoplasmic/endoplasmic reticulum calcium adenosine triphosphate (SERCA) pump. The type of pump shares the catalytic properties of ion-motive ATPases of the P-type family. It transports calcium ions from the cytoplasm into the sarco-endoplasmic reticulum. Its activity is needed for viability by all types of cells. The conjugate remains inactive until the PSMA-specific peptide is cleaved, thereby starting SERCA inhibition. In xenograft models, thapsigargin induced tumor regression at doses that appeared to be minimally toxic

**TABLE 2 |** Selection of trials of PSMA-based therapy (Selection based on active and completed trials).

NCT identifier	Study phase	Drug	Study objectives (Number of patients)	Study results
NCT01695044 NCT02020135	Phase 2 (Extension Study)	PSMA ADC	Assess total serum PSA response, CTC response, overall radiologic response in mCRPC pts (119 pts- completed 17 pts) in two groups: (1) CHT-experienced and (2) CHT naïve.	-PSA response: >30% Decrease in PSA: 29% (1); 32% (2). >50% Decrease in PSA: 11% (1); 21% (2). -CTC response >30% Decrease in CTC: 81% (1), 92% (2). >50% Decrease in CTC: 74%(1),85%(2). -Overall radiologic response Stable disease 61% (1),69% (2); Progressive disease: 13% (1), 9% (2); Partial response: 0 (1), 6% (2)
NCT01414283	Phase 1	PSMA ADC	Determine the maximum tolerated dose of PSMA ADC (13 weeks) (52 pts)	No results posted
NCT01414296	Phase 1 Extended 39-Week	PSMA ADC	Safety and tolerability of PSMA ADC as measured by all adverse events in mCRPC patients (10 pts)	No results posted
NCT00705835	Phase 1	Rs-PSMA	Safety, tolerability, and immune response of vaccine therapy with increasing dose levels of rsPSMA protein (14 pts)	No results posted
NCT00015977	Phase 2	PSMA peptide vaccine	Immunization with PSMA peptide vaccine followed by injection of Interleukin-12 in Metastatic PCa patients, determinate disease response (13 pts)	No results posted
NCT01140373	Phase 1	Autologous T cells targeted to PSMA	Safety and tolerability using increasing doses of engineered autologous T cells targeted to PSMA after cyclophosphamide in CMPC patients (13 pts)	No results posted
NCT02202447	Phase 1	EC1169	Safety, pharmacokinetic profile and preliminary efficacy of PSMA Targeting-Tubulysin Conjugate EC1169 in Patients With Recurrent Metastatic CRPC (40 pts)	No results posted
NCT00694551	Not Applicable	Polypeptide vaccines: PSMA27-35- PSMA687-701	Pilot immunotherapy study of combination PSMA and TARP peptide with Poly IC-LC adjuvant in patients with elevated PSA after initial definitive treatment (29 pts)	Adverse events (Grade 3 or higher): 0 pts PSA doubling: 19/29 pts; No PSA doubling: 10/29 pts
NCT00916123	Phase 1	177Lu-J591	Effectiveness of 177Lu-J591 antibody in combination with docetaxel chemotherapy against metastatic CRPC (15 pts)	No results posted

For a full list of trials please visit: <https://clinicaltrials.gov> PSMA ADC, Prostate Specific Membrane Antigen Antibody Drug Conjugate; CHT, chemotherapy; PSA, prostatic specific antigen; CTC, Circulating tumor cells; 177Lu- J591, Anti-prostate-specific Membrane Antigen Monoclonal Antibody; CMPC, Castrate Metastatic Prostate Cancer; TARP, T-cell receptor  $\gamma$  alternate reading frame protein; CRPC, castrate-resistant prostate cancer; Rs-PSMA, Recombinant Soluble PSMA.



(69). Based on such findings, a phase I study is evaluating the Thapsigargin prodrug G-202 in patients with advanced PCa and other solid tumors (NCT01056029).

PSMA can be used in immunotherapy and radiotherapy approaches. “Adoptive immunotherapy based on infusion of designer T cells engineered with chimeric antigen receptors (CARs)” to potentiate their antitumor activity could serve as a highly specific modality for the treatment of cancer. Thus, PSMA $\times$ CD3 diabody is able to retarget human CD4<sup>+</sup> and CD8<sup>+</sup> lymphocytes to lyse PSMA-expressing C4-2 PCa cells. Other 1st and 2nd generation anti-PSMA designer T cells have shown their activity in both *in vitro* and *in vivo* studies (70). More recently, mouse-human chimeric IgG1 of 2C9 (KM2777) has been fused with C-terminus interleukin-2 (IL-2). In a xenograft tumor model using PSMA-expressing PCa cells, this fusion, KM2812, showed evident antitumor activity, with complete regression in some cases (71). Bispecific antibodies have been utilized in human clinical trials. A phase I trial has studying the safety of adoptive transfer of autologous T cells targeted to PSMA for the treatment of castrate metastatic PCa patients (NCT01140373). Vaccine is another very important area that utilizes PSMA as a target to increase cellular and humoral immune responses against tumor cells (72).

Concerning the potential role of PSMA targeted antivascular radiotherapy, Bandekar et al. has evaluated liposomes loaded with the  $\alpha$ -particle generator 225Ac to kill in a selective manner PSMA positive PCa cells. In such study, anti-PSMA-targeted liposomes have been able to kill PSMA positive cells, including the endothelial cells expressing PSMA, thus suggesting their use for selective antivascular radiotherapy (73).

## Therapy of Metastatic Castration Resistant PCa (CRPCa) With Radiopharmaceuticals

Recently, studies have explored the role of PSMA-based treatments with radiopharmaceuticals of metastatic castration resistant PCa (Figure 3). The first antibody-based radiotherapeutic was Yttrium-90 (90Y) capromab. Phase 1 (74) and Phase 2 (75) studies were unsuccessful for significant toxicity and lack of efficacy.

J591 was the first humanized monoclonal antibody directed against the extracellular domain of PSMA (76). The PSMA antibody-based radiotherapeutic Lutetium-177 J591 (177Lu-J591) showed acceptable toxicity with evident targeting of known metastatic sites in a phase 1 trial. In phase II, almost 60% of men showed a decline in serum PSA levels, 10.6% of them experiencing a  $>50\%$  decline in PSA (77). Myelosuppression associated with treatment was reversible. “Fractionated dosing allowed for higher cumulative doses with less toxicity” (78). However, there are limitations to the use of PSMA antibody-based radiotherapeutic. Lutetium-177 J591, i.e., slow diffusion of the antibodies into solid tumor lesions and hematotoxicity caused by a long circulation time in the blood (79). Retrospective studies have caveats such as treatments that have been done outside of clinical trials,

unrecognized and unmeasured covariates that might influence final results.

A recent systematic review showed that 43% of the men showed a maximum decline of serum PSA of  $\geq 50\%$  following treatment with 177Lu-PSMA radioligand therapy (RLT). In particular, 177Lu-PSMA RLT gave objective remission and decline of PSA of  $\geq 50\%$ , more often than with third-line treatments (enzalutamide and cabazitaxel). Median survival was longer after 177Lu-PSMA RLT than after third-line treatment, the difference being not significant from statistically point of view (80).

A promising available option for patients with mCRPC is the recently investigated compound PSMA-DKFZ-617, a small molecule peptide, rather than an antibody, chemically conjugated with 177Lutetium that binds with high affinity to PSMA. Unlike antibodies such as J591, it shows more rapid plasma clearance, higher affinity binding to PSMA and lower toxicity. Interesting outcomes came up from a single-arm, single-center, phase 2 trial (ACTRN12615000912583) recently published by Hofman et al. (81, 82). Patients with mCRPC and progressive disease after standard treatments underwent screening PSMA and FDG-PET/CT to confirm high PSMA-expression. After four cycles of intravenous [<sup>177</sup>Lu]-PSMA-617, 17 (57%) of 30 patients (95% CI 37–75) obtained a PSA decrease of 50% or more, objective response in nodal or visceral disease in 14 (82%) of 17 patients with measurable disease and reported minor toxic effects and improvement in pain severity. Phase 3 trial of 177Lu-PSMA-617 (NCT03511664) is currently recruiting. A multicenter randomized trial comparing LuPSMA with cabazitaxel chemotherapy (NCT03392428) is ongoing.

**Table 2** summarizes a selection of active and completed trials on PSMA and therapy. For a full list of trials please visit: <https://clinicaltrials.gov>.

## CONCLUSION

PSMA represents an attractive target for the detection and treatment of patients with PCa. PSMA immunohistochemical evaluation should be further investigated as a predictive marker in men with metastatic PCa, to guide clinicians in the selection of the most appropriate imaging technique and therapy in individual patients. The choice of emerging PSMA-targeted tracers and therapeutic agents requires further investigation in order to identify the most specific compound for the distinct sites and phases of the disease (83–85). As our understanding of the role of PSMA in prostate carcinogenesis advances and molecular techniques become more refined, PSMA-based strategies will have a crucial role in the evolving diagnostic and therapeutic landscape of patients with PCa.

## AUTHOR CONTRIBUTIONS

RM and MSc conception and design. AC and MSa drafting the manuscript. MC, FM, and AG review of the literature. NB and AL-B critical revision of the manuscript.

## REFERENCES

- Pinto JT, Suffoletto BP, Berzin TM, Qiao CH, Lin S, Tong WP, et al. Prostate-specific membrane antigen: a novel folate hydrolase in human prostatic carcinoma cells. *Clin Cancer Res.* (1996) 2:1445–51.
- Carter RE, Feldman AR, Coyle JT. Prostate-specific membrane antigen is a hydrolase with substrate and pharmacologic characteristics of a neuropeptidase. *Proc Natl Acad Sci USA.* (1996) 93:749–53. doi: 10.1073/pnas.93.2.749
- Ghosh A, Heston WD. Tumor target prostate specific membrane antigen (PSMA) and its regulation in prostate cancer. *J Cell Biochem.* (2004) 91:528–39. doi: 10.1002/jcb.10661
- Horoszewicz JS, Kawinski E, Murphy GP. Monoclonal antibodies to a new antigenic marker in epithelial prostatic cells and serum of prostatic cancer patients. *Anticancer Res.* (1987) 7:927–35.
- Kasperzyk JL, Finn SP, Flavin R, Fiorentino M, Lis R, Hendrickson WK, et al. Prostate-specific membrane antigen protein expression in tumor tissue and risk of lethal prostate cancer. *Cancer Epidemiol Biomarkers Prev.* (2013) 22:2354–63. doi: 10.1158/1055-9965.EPI-13-0668
- Wright GL Jr, Grob BM, Haley C, Grossman K, Newhall K, Petrylak D, et al. Upregulation of prostate-specific membrane antigen after androgen-deprivation therapy. *Urology* (1996) 48:326–34.
- Noss KR, Wolfe SA, Grimes SR. Upregulation of prostate specific membrane antigen/folate hydrolase transcription by an enhancer. *Gene* (2002) 285:247–56. doi: 10.1016/S0378-1119(02)00397-9
- Chang SS, Gaudin PB, Reuter VE, Heston WD. Prostate-specific membrane antigen: present and future applications. *Urology* (2000) 55:622–9. doi: 10.1016/S0090-4295(99)00600-7
- Yao V, Bacich DJ. Prostate specific membrane antigen (PSMA) expression gives prostate cancer cells a growth advantage in a physiologically relevant folate environment *in vitro*. *Prostate* (2006) 66:867–75. doi: 10.1002/pros.20361
- Yao V, Berkman CE, Choi JK, O'Keefe DS, Bacich DJ. Expression of prostate-specific membrane antigen (PSMA), increases cell folate uptake and proliferation and suggests a novel role for PSMA in the uptake of the non-polyglutamated folate, folic acid. *Prostate* (2010) 70:305–16. doi: 10.1002/pros.21065
- Zhang Y, Guo Z, Du T, Chen J, Wang W, Xu K, et al. Prostate specific membrane antigen (PSMA): a novel modulator of p38 for proliferation, migration, and survival in prostate cancer cells. *Prostate* (2013) 73:835–41. doi: 10.1002/pros.22627
- Guo Z, Lai Y, Du T, Zhang Y, Chen J, Bi L, et al. Prostate specific membrane antigen knockdown impairs the tumorigenicity of LNCaP prostate cancer cells by inhibiting the phosphatidylinositol 3-kinase/Akt signaling pathway. *Chin Med J.* (2014) 127:929–36.
- Xu L, Wang Z, Li XF, He X, Guan LL, Tuo JL, et al. Screening and identification of significant genes related to tumor metastasis and PSMA in prostate cancer using microarray analysis. *Oncol Rep.* (2013) 30:1920–8. doi: 10.3892/or.2013.2656
- Miyahira AK, Pienta KJ, Morris MJ, Bander NH, Baum RP, Fendler WP, et al. Meeting report from the Prostate Cancer Foundation PSMA-directed radionuclide scientific working group. *Prostate* (2018) 78:775–89. doi: 10.1002/pros.23642
- Fanti S, Minozzi S, Antoch G, Banks I, Briganti A, Carrio I, et al. Consensus on molecular imaging and theranostics in prostate cancer. *Lancet Oncol.* (2018) 19:e696–708. doi: 10.1016/S1470-2045(18)30604-1
- Bermejo CE, Coursey J, Basler J, Austenfeld M, Thompson I. Histologic confirmation of lesions identified by ProstateScint scan following definitive treatment. *Urol Oncol.* (2003) 21:349–52. doi: 10.1016/S1078-1439(02)00253-3
- Lau HY, Kindrachuk G, Carter M, Prestage K, Webber D, Stauffer E, et al. Surgical confirmation of ProstateScint abnormalities in two patients with high risk prostate cancer. *Can J Urol.* (2001) 8:1199–202.
- Hinkle GH, Burgers JK, Olsen JO, Williams BS, Lamatrice RA, Barth RF, et al. Prostate cancer abdominal metastases detected with indium-111 capromab pendetide. *J Nucl Med.* (1998) 39:650–2.
- Pandit-Taskar N, O'Donoghue JA, Morris MJ, Wills EA, Schwartz LH, Gonen M, et al. Antibody mass escalation study in patients with castration-resistant prostate cancer using <sup>111</sup>In-J591: lesion detectability and dosimetric projections for 90Y radioimmunotherapy. *J Nucl Med.* (2008) 49:1066–74. doi: 10.2967/jnumed.107.049502
- Foss CA, Mease RC, Fan H, Wang Y, Ravert HT, Dannals RF, et al. Radiolabeled small-molecule ligands for prostate-specific membrane antigen: *in vivo* imaging in experimental models of prostate cancer. *Clin Cancer Res.* (2005) 11:4022–8. doi: 10.1158/1078-0432.CCR-04-2690
- Lu G, Maresca KP, Hillier SM, Zimmerman CN, Eckelman WC, Joyal JL, et al. Synthesis and SAR of (99m)Tc/Re-labeled small molecule prostate specific membrane antigen inhibitors with novel polar chelates. *Bioorg Med Chem Lett.* (2013) 23:1557–63. doi: 10.1016/j.bmcl.2012.09.014
- Pandit-Taskar N, O'Donoghue JA, Beylgeril V, Lyashchenko S, Ruan S, Solomon SB, et al. (89)Zr-huJ591 immuno-PET imaging in patients with advanced metastatic prostate cancer. *Eur J Nucl Med Mol Imaging* (2014) 4:2093–105. doi: 10.1007/s00259-014-2830-7
- Evazalipour M, D'Huyvetter M, Tehrani BS, Abolhassani M, Omidfar K, Abdoli S, et al. Generation and characterization of nanobodies targeting PSMA for molecular imaging of prostate cancer. *Contrast Media Mol Imaging* (2014) 9:211–20. doi: 10.1002/cmmi.1558
- Pandit-Taskar N, O'Donoghue JA, Ruan S, Lyashchenko SK, Carrasquillo JA, Heller G, et al. First-in-human imaging with <sup>89</sup>Zr-Df-IAB2M anti-PSMA minibody in patients with metastatic prostate cancer: pharmacokinetics, biodistribution, dosimetry, and lesion uptake. *J Nucl Med.* (2016) 57:1858–64. doi: 10.2967/jnumed.116.176206
- Tiffany CW, Lapidus RG, Merion A, Calvin DC, Slusher BS. Characterization of the enzymatic activity of PSM: comparison with brain NAALADase. *Prostate* (1999) 39:28–35. doi: 10.1002/(SICI)1097-0045(19990401)39:1<28::AID-PROS5>3.0.CO;2-A
- Luthi-Carter R, Barczak AK, Speno H, Coyle JT. Molecular characterization of human brain N-acetylated alpha-linked acidic dipeptidase (NAALADase). *J Pharmacol Exp Ther.* (1998) 286:1020–5.
- Jackson PF, Cole DC, Slusher BS, Stetz SL, Ross LE, Donzanti BA, et al. Design, synthesis, and biological activity of a potent inhibitor of the neuropeptidase N-acetylated alpha-linked acidic dipeptidase. *J Med Chem.* (1996) 39:619–22. doi: 10.1021/jm950801q
- Jackson PF, Slusher BS. Design of NAALADase inhibitors: a novel neuroprotective strategy. *Curr Med Chem.* (2001) 8:949–57. doi: 10.2174/0929867013372797
- Maresca KP, Hillier SM, Femia FJ, Keith D, Barone C, Joyal JL, et al. A series of halogenated heterodimeric inhibitors of prostate specific membrane antigen (PSMA) as radiolabeled probes for targeting prostate cancer. *J Med Chem.* (2009) 52:347–57. doi: 10.1021/jm800994j
- Hillier SM, Maresca KP, Femia FJ, Marquis JC, Foss CA, Nguyen N, et al. Preclinical evaluation of novel glutamate-urea-lysine analogues that target prostate-specific membrane antigen as molecular imaging pharmaceuticals for prostate cancer. *Cancer Res.* (2009) 69:6932–40. doi: 10.1158/0008-5472.CAN-09-1682
- Kularatne SA, Zhou Z, Yang J, Post CB, Low PS. Design, synthesis, and preclinical evaluation of prostate-specific membrane antigen targeted (99m)Tc-radioimaging agents. *Mol Pharm.* (2009) 6:790–800. doi: 10.1021/mp9000712
- Chen Y, Pullambhatla M, Byun Y, Foss CA, Nimmagadda S, Senthambizhelvan S, et al. 2-(3-[1-Carboxy-5-((6-(18F)fluoropyridine-3-carbonyl)-amino)-pentyl]-ureido)-pentanedioic acid, (18F)DCFPyL, a PSMA-based PET imaging agent for prostate cancer. *Clin Cancer Res.* (2011) 17:7645–53. doi: 10.1158/1078-0432.CCR-11-1357
- Banerjee SR, Pullambhatla M, Shallal H, Lisok A, Mease RC, Pomper MG. A modular strategy to prepare multivalent inhibitors of prostate-specific membrane antigen (PSMA). *Oncotarget* (2011) 2:1244–53. doi: 10.18632/oncotarget.415
- Afshar-Oromieh A, Haberkorn U, Schlemmer HP, Fenchel M, Eder M, Eisenhut M, et al. Comparison of PET/CT and PET/MRI hybrid systems using a <sup>68</sup>Ga-labelled PSMA ligand for the diagnosis of recurrent prostate cancer: initial experience. *Eur J Nucl Med Mol Imaging* (2014) 4:887–97. doi: 10.1007/s00259-013-2660-z

35. Barrett JA, Coleman RE, Goldsmith SJ, Vallabhajosula S, Petry NA, Cho S, et al. First-in-man evaluation of two high-affinity PSMA-avid small molecules for imaging prostate cancer. *J Nucl Med.* (2013) 54:380–7. doi: 10.2967/jnumed.112.111203
36. Mease RC, Foss CA, Pomper MG. PET imaging in prostate cancer: focus on prostate-specific membrane antigen. *Curr Top Med Chem.* (2013) 13:951–62. doi: 10.2174/1568026611313080008
37. Mease RC, Dusich CL, Foss CA, Ravert HT, Dannals RF, Seidel J, et al. N-(N-(S)-1,3-Dicarboxypropyl)carbamoyl-4- (18F)fluorobenzyl-L-cysteine, (18F)DCFBFC: a new imaging probe for prostate cancer. *Clin Cancer Res.* (2008) 14:3036–43. doi: 10.1158/1078-0432.CCR-07-1517
38. Cho SY, Gage KL, Mease RC, Senthambhavan S, Holt DP, Kwanisai-Jeffrey A, et al. Biodistribution, tumor detection and Radiation dosimetry of 18F-DCFBFC, a low molecular weight inhibitor of PSMA, in patients with metastatic prostate cancer. *J Nucl Med.* (2012) 53:1883–91. doi: 10.2967/jnumed.112.104661
39. Giovacchini G, Picchio M, Coradeschi E, Bettinardi V, Gianolli L, Scattoni V, et al. Predictive factors of ((11)C)choline PET/CT in patients with biochemical failure after radical prostatectomy. *Eur J Nucl Med Mol Imaging* (2010) 37:301–9. doi: 10.1007/s00259-009-1253-3
40. Afshar-Oromieh A, Haberkorn U, Eder M, Eisenhut M, Zechmann C. ((68)Ga)Gallium-labelled PSMA ligand as superior PET tracer for the diagnosis of prostate cancer: comparison with (18)F-FECH. *Eur J Nucl Med Mol Imaging* (2012) 39:1085–6. doi: 10.1007/s00259-012-2069-0
41. Afshar-Oromieh A, Malcher A, Eder M, Eisenhut M, Linhart HG, Hadaschik BA, et al. PET imaging with a ((68)Ga)gallium-labelled PSMA ligand for the diagnosis of prostate cancer: biodistribution in humans and first evaluation of tumour lesions. *Eur J Nucl Med Mol Imaging* (2013) 40:486–95. doi: 10.1007/s00259-012-2298-2
42. Eder M, Schafer M, Bauder-Wust U, Hull WE, Wangler C, Mier W, et al. (68)Ga-Complex lipophilicity and the targeting property of a urea-based PSMA inhibitor for PET imaging. *Bioconjug Chem.* (2012) 23:688–97. doi: 10.1021/bc200279b
43. Eder M, Eisenhut M, Babich J, Haberkorn U. PSMA as a target for radiolabelled small molecules. *Eur J Nucl Med Mol Imaging* (2013) 40:819–23. doi: 10.1007/s00259-013-2374-2
44. Graham K, Kettschau G, Gromov A, Friebe M, Gekeler V, Dinkelborg L. (18F)-Labeled PET Tracer BAY1075553, Small Molecule Inhibitor of PSMA for Molecular Imaging of Prostate Cancer. World Molecular Imaging Conference; September 7–10; San Diego, CA (2011) p. P235
45. Langsteger W, Kunit T, Haim S, Nader M, Valencia R, Lesche R, et al. BAY 1075553 PET/CT in the assessment of prostate cancer: safety, tolerability and biodistribution-Phase I first in human study results. *J Nucl Med.* (2012) 53:1125.
46. Fendler WP, Schmidt DE, Wenter V, Thierfelder KM, Zach C, Stief C, et al. 68GaPSMA PET/CT detects the location and extent of primary prostate cancer. *J Nucl Med.* (2016) 57:1720–5. doi: 10.2967/jnumed.116.172627
47. Woythal N, Arsenic R, Kempkensteffen C, Miller K, Janssen JC, Huang K, et al. Immunohistochemical validation of PSMA expression measured by 68Ga-PSMA PET/CT in primary prostate cancer. *J Nucl Med.* (2018) 59:238–43. doi: 10.2967/jnumed.117.195172
48. Uprimny C, Kroiss AS, Decristoforo C, Fritz J, von Guggenberg E, Kendler D, et al. 68Ga-PSMA-11 PET/CT in primary staging of prostate cancer: PSA and Gleason score predict the intensity of tracer accumulation in the primary tumour. *Eur J Nucl Med Mol Imaging* (2017) 44:941–9. doi: 10.1007/s00259-017-3631-6
49. Koerber SA, Utzinger MT, Kratochwil C, Kesch C, Haefner MF, Katayama S, et al. 68Ga-PSMA-11 PET/CT in newly diagnosed carcinoma of the prostate: correlation of intraprostatic PSMA uptake with several clinical parameters. *J Nucl Med.* (2017) 58:1943–8. doi: 10.2967/jnumed.117.190314
50. Mottet N, Bellmunt J, Bolla M, Briers E, Cumberbatch MG, De Santis M, et al. EAU-ESTRO-SIOG guidelines on prostate cancer. Part 1: screening, diagnosis, and local treatment with curative intent. *Eur Urol.* (2017) 71:618–29. doi: 10.1016/j.eururo.2016.08.003
51. van Leeuwen PJ, Emmett L, Ho B, Delprado W, Ting F, Nguyen Q, et al. Prospective evaluation of 68Gallium-prostate-specific membrane antigen positron emission tomography/computed tomography for preoperative lymph node staging in prostate cancer. *BJU Int.* (2017) 119:209–15. doi: 10.1111/bju.13540
52. Gorin MA, Rowe SP, Patel HD, Vidal I, Mana-Ay M, Javadi MS, et al. Prostate specific membrane antigen targeted 18F-DCFPyL positron emission tomography/computerized tomography for the preoperative staging of high risk prostate cancer: results of a prospective, phase ii, single center study. *J Urol.* (2018) 199:126–32. doi: 10.1016/j.juro.2017.07.070
53. Maurer T, Gschwend JE, Rauscher I, Souvatzoglou M, Haller B, Weirich G, et al. Diagnostic efficacy of (68)gallium-PSMA positron emission tomography compared to conventional imaging for lymph node staging of 130 consecutive patients with intermediate to high risk prostate cancer. *J Urol.* (2016) 195:1436–43. doi: 10.1016/j.juro.2015.12.025
54. Meredith G, Wong D, Yaxley J, Coughlin G, Thompson L, Kua B, Gianduzzo T. The use of 68 Ga-PSMA PET CT in men with biochemical recurrence after definitive treatment of acinar prostate cancer. *BJU Int.* (2016) 118(Suppl 3):49–55. doi: 10.1111/bju.13616
55. Perera M, Papa N, Christidis D, Wetherell D, Hofman MS, Murphy DG, et al. Sensitivity, specificity, and predictors of positive 68Ga-prostate-specific membrane antigen positron emission tomography in advanced prostate cancer: a systematic review and meta-analysis. *Eur Urol.* (2016) 70:926–37. doi: 10.1016/j.eururo.2016.06.021
56. Rauscher I, Düwel C, Haller B, Rischpler C, Heck MM, Gschwend JE, et al. Efficacy, predictive factors, and prediction nomograms for Ga-labeled prostate-specific membrane antigen–ligand positron-emission tomography/computed tomography in early biochemical recurrent prostate cancer after radical prostatectomy. *Eur Urol.* (2018) 73:656–61. doi: 10.1016/j.eururo.2018.01.006
57. Rauscher I, Maurer T, Beer AJ, Graner FP, Haller B, Weirich G, et al. Value of 68GaPSMA HBED-CC PET for the assessment of lymph node metastases in prostate cancer patients with biochemical recurrence: comparison with histopathology after salvage lymphadenectomy. *J Nucl Med.* (2016) 57:1713–9. doi: 10.2967/jnumed.116.173492
58. Cimadamore A, Scarpelli M, Cheng L, Lopez-Beltran A, Montorsi F, Montironi R. Re: Isabel Rauscher, Charlotte Düwel, Bernhard Haller, et al. Efficacy, predictive factors, and prediction nomograms for 68Ga-labeled prostate-specific membrane antigen–ligand positron-emission tomography/computed tomography in early biochemical recurrent prostate cancer after radical prostatectomy. *Eur Urol* 2018;73:656–61: Clinical Significance of Prostate-specific Membrane Antigen Immunohistochemistry and Role of the Uropathologists. *Eur Urol.* (2018) 74:e141–4. doi: 10.1016/j.eururo.2018.07.034
59. Parent EE, Schuster DM. Update on 18F-Fluciclovine PET for prostate cancer imaging. *J Nucl Med.* (2018) 59:733–9. doi: 10.2967/jnumed.117.204032
60. Akin-Akintayo OO, Jani AB, Odewole O, Tade FI, Nieh PT, Master VA, et al. Change in salvage radiotherapy management based on guidance with FACBC (fluciclovine) PET-CT in postprostatectomy recurrent prostate cancer. *Clin Nucl Med.* (2017) 42:e22–8. doi: 10.1097/RLU.0000000000001379
61. Odewole OA, Tade FI, Nieh PT, Savir-Baruch B, Jani AB, Master VA, et al. Recurrent prostate cancer detection with anti-3-[18F]FACBC PET-CT: comparison with CT. *Eur J Nucl Med Mol Imaging* (2016) 43:1773–83. doi: 10.1007/s00259-016-3383-8
62. Han S, Woo S, Kim YJ, Suh CH. Impact of 68Ga-PSMA PET on the management of patients with prostate cancer: a systematic review and meta-analysis. *Eur Urol.* (2018) 74:179–90. doi: 10.1016/j.eururo.2018.03.030
63. Nanni C, Schiavina R, Brunocilla E, Borghesi M, Ambrosini V, Zanoni L, et al. 18F-FACBC compared with 11C-choline PET/CT in patients with biochemical relapse after radical prostatectomy: a prospective study in 28 patients. *Clin Genitourin Cancer* (2014) 12:106–10. doi: 10.1016/j.clgc.2013.08.002
64. Ireson CR, Kelland LR. Discovery and development of anticancer aptamers. *Mol Cancer Ther.* (2006) 5:2957–62. doi: 10.1158/1535-7163.MCT-06-0172
65. Lupold SE, Hicke BJ, Lin Y, Coffey DS. Identification and characterization of nuclease-stabilized RNA molecules that bind human prostate cancer cells via the prostate-specific membrane antigen. (2002) *Cancer Res.* 62:4029–33.
66. Xu W, Siddiqui IA, Nihal M, Pilla S, Rosenthal K, Mukhtar H, et al. Aptamer-conjugated and doxorubicin-loaded unimolecular micelles for targeted therapy of prostate cancer. *Biomaterials* (2013) 34:5244–53. doi: 10.1016/j.biomaterials.2013.03.006

67. Wang X, Ma D, Olson WC, Heston WD. *In vitro* and *in vivo* responses of advanced prostate tumors to PSMA ADC, an auristatin-conjugated antibody to prostate-specific membrane antigen. *Mol Cancer Ther.* (2011) 10:1728–39. doi: 10.1158/1535-7163.MCT-11-0191
68. Petrylak DP, Smith DC, Appleman LJ, Fleming MT, Hussain A, Dreicer R, et al. *A Phase 2 Trial of Prostate-Specific Membrane Antigen Antibody Drug Conjugate (PSMA ADC) in Taxane-Refractory Metastatic Castration-Resistant Prostate Cancer (mCRPC)*. 2014 ASCO Annual Meeting; abstract 5023.
69. Denmeade SR, Mhaka AM, Rosen DM, Brennen WN, Dalrymple S, Dach I, et al. Engineering a prostate-specific membrane antigen-activated tumor endothelial cell prodrug for cancer therapy. *Sci Transl Med.* (2012) 4:140ra86. doi: 10.1126/scitranslmed.3003886
70. Ma Q, Safar M, Holmes E, Wang Y, Boynton AL, Junghans RP. Anti-prostate specific membrane antigen designer T cells for prostate cancer therapy. *Prostate* (2004) 61:12–25. doi: 10.1002/pros.20073
71. Sugimoto Y, Hirota M, Yoshikawa K, Sumitomo M, Nakamura K, Ueda R, et al. The therapeutic potential of a novel PSMA antibody and its IL-2 conjugate in prostate cancer. *Anticancer Res.* (2014) 34:89–97.
72. Yeku O, Slovin SF. Immune therapy for prostate cancer. *Cancer J.* (2016) 22:334–41. doi: 10.1097/PPO.0000000000000223
73. Bandekar A, Zhu C, Jindal R, Bruchertseifer F, Morgenstern A, Sofou S. Anti-prostate-specific membrane antigen liposomes loaded with 225Ac for potential targeted antivasculature  $\alpha$ -particle therapy of cancer. *J Nucl Med.* (2014) 55:107–14. doi: 10.2967/jnumed.113.125476
74. Deb N, Goris M, Trisler K, Fowler S, Saal J, Ning S, et al. Treatment of hormone-refractory prostate cancer with 90Y-CYT-356 monoclonal antibody. *Clin Cancer Res.* (1996) 2:1289–97.
75. Kahn D, Austin JC, Maguire RT, Miller SJ, Gerstbrein J, Williams RD. A phase II study of [90Y]yttrium-capromabpendetide in the treatment of men with prostate cancer recurrence following radical prostatectomy. *Cancer Biother Radiopharm.* (1999) 14:99–111. doi: 10.1089/cbr.1999.14.99
76. Bander NH, Trabulsi EJ, Kostakoglu L, Yao D, Vallabhajosula S, Smith-Jones P, et al. Targeting metastatic prostate cancer with radiolabeled monoclonal antibody J591 to the extracellular domain of prostate specific membrane antigen. *J Urol.* (2003) 170:1717–21. doi: 10.1097/01.ju.0000091655.77601.0c
77. Tagawa ST, Milowski MI, Morris M. Phase II trial of 177Lu-radiolabeled anti-prostate specific membrane antigen (PSMA) monoclonal antibody J591(177Lu-J591) in patients (pts) with metastatic castrate resistant prostate cancer (metCRPC). *J Clin Oncol.* (2008) 28 (Suppl. 15):5140. doi: 10.1200/jco.2008.26.15\_suppl.5140
78. Tagawa ST, Vallabhajosula S, Osborne J, Goldsmith K, Petrillo L, Tyrell GS, et al. Phase I trial of fractionated-dose 177Lu-radiolabeled anti-prostate specific membrane antigen (PSMA) monoclonal antibody J591(177Lu-J591) in patients with metastatic castration-resistant prostate cancer. *J Clin Oncol.* (2010) 28:4667. doi: 10.1200/jco.2010.28.15\_suppl.4667
79. Tagawa ST, Milowsky MI, Morris M, Vallabhajosula S, Christos P, Akhtar NH, et al. Phase II study of Lutetium-177-labeled anti-prostate-specific membrane antigen monoclonal antibody J591 for metastatic castration-resistant prostate cancer. *Clin Cancer Res.* (2013) 19:5182–91. doi: 10.1158/1078-0432.CCR-13-0231
80. von Eyben FE, Roviello G, Kiljunen T, Uprimny C, Virgolini I, Kairemo K, Joensuu T. Third-line treatment and 177Lu-PSMA radioligand therapy of metastatic castration-resistant prostate cancer: a systematic review. *Eur J Nucl Med Mol Imaging* (2018) 45:496–508. doi: 10.1007/s00259-017-3895-x
81. Hofman MS, Violet J, Hicks RJ, Ferdinandus J, Thang SP, Akhurst T, et al. [177Lu]-PSMA-617 radionuclide treatment in patients with metastatic castration-resistant prostate cancer (LuPSMA trial): a single-centre, single-arm, phase 2 study. *Lancet Oncol.* (2018) 19:825–33. doi: 10.1016/S1470-2045(18)30198-0
82. Bradley CA. [177Lu]PSMA-617 radionuclide therapy shows promise. *Nat Rev Urol.* (2018) 15:468. doi: 10.1038/s41585-018-0029-6
83. Cimadamore A, Gasparrini S, Scarpelli M, Doria A, Mazzucchelli R, Massari F, et al. Epigenetic modifications and modulators in prostate cancer. *Crit Rev Oncog.* (2017) 22:439–50. doi: 10.1615/CritRevOncog.2017020964
84. Gasparrini S, Cimadamore A, Mazzucchelli R, Scarpelli M, Massari F, Raspollini MR, et al. Pathology and molecular updates in tumors of the prostate: towards a personalized approach. *Expert Rev Mol Diagn.* (2017) 17:781–9. doi: 10.1080/14737159.2017.1341314
85. Kularatne SA, Thomas M, Myers CH, Gagare P, Kanduluru AK, Crian CJ, et al. Evaluation of Novel Prostate-Specific Membrane Antigen-Targeted Near-Infrared Imaging Agent for Fluorescence-Guided Surgery of Prostate Cancer. *Clin Cancer Res.* (in press) doi: 10.1158/1078-0432.CCR-18-0803

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Cimadamore, Cheng, Santoni, Lopez-Beltran, Battelli, Massari, Galosi, Scarpelli and Montironi. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.





# Recent Advances in Liquid Biopsy in Patients With Castration Resistant Prostate Cancer

Vincenzo Di Nunno<sup>1</sup>, Lidia Gatto<sup>1</sup>, Matteo Santoni<sup>2</sup>, Alessia Cimadamore<sup>3\*</sup>, Antonio Lopez-Beltran<sup>4</sup>, Liang Cheng<sup>5</sup>, Marina Scarpelli<sup>3</sup>, Rodolfo Montironi<sup>3</sup> and Francesco Massari<sup>1\*</sup>

<sup>1</sup> Division of Oncology, S.Orsola-Malpighi Hospital, Bologna, Italy, <sup>2</sup> Oncology Unit, Macerata Hospital, Macerata, Italy,

<sup>3</sup> Section of Pathological Anatomy, School of Medicine, United Hospital, Polytechnic University of the Marche Region,

Ancona, Italy, <sup>4</sup> Department of Pathology and Surgery, Faculty of Medicine, Cordoba, Spain, <sup>5</sup> Department of Pathology and Laboratory Medicine, Indiana University School of Medicine, Indianapolis, IN, United States

## OPEN ACCESS

### Edited by:

Masaki Shiota,  
Kyushu University, Japan

### Reviewed by:

Takeshi Yuasa,  
Japanese Foundation For Cancer  
Research, Japan  
Daniel C. Danila,  
Memorial Sloan Kettering Cancer  
Center, United States

### \*Correspondence:

Alessia Cimadamore  
alessiacimadamore@gmail.com  
Francesco Massari  
fmassari79@gmail.com

### Specialty section:

This article was submitted to  
Genitourinary Oncology,  
a section of the journal  
Frontiers in Oncology

**Received:** 11 July 2018

**Accepted:** 03 September 2018

**Published:** 24 September 2018

### Citation:

Di Nunno V, Gatto L, Santoni M,  
Cimadamore A, Lopez-Beltran A,  
Cheng L, Scarpelli M, Montironi R and  
Massari F (2018) Recent Advances in  
Liquid Biopsy in Patients With  
Castration Resistant Prostate Cancer.  
Front. Oncol. 8:397.  
doi: 10.3389/fonc.2018.00397

Management of localized and advanced prostate cancer benefits from several therapeutic options with a surprising improvement in terms of clinical outcome. The selection of patients more likely to benefit from a specific approach still remains a key issue as well as the early identification of patients with aggressive disease which could benefit from a more aggressive treatment strategy. The lack of reliable bio-marker in castration resistant setting able to monitor response to treatment and early inform about tumor progression is an emerging issue. Accordingly, circulating DNA and circulating tumor cells appears a promising and attractive approach despite to date practical applications of these techniques are few and not validated. The aim of this review of the literature is to explore current knowledge on liquid biopsy in prostate cancer focusing on possible future applications.

**Keywords:** prostate cancer, metastatic castration resistant prostate cancer, CTCs, liquid biopsy, circulating DNA

## INTRODUCTION

Prostate Cancer (PCa) represents the most common adult malignancies ranking as one of the major cause of cancer related death in men (1). Management of the disease accounts various options in both localized and advanced stages. Each options are generally evaluated according to different variables related to patients (performance status, comorbidities, disease related symptoms, and patients' preferences) and tumor features (biological aggressiveness and site and number of metastases). Thus, the management of localized stages could range from a first instance no invasive approach (watchful waiting or active surveillance approach) to a radical approach by surgery, external radiation treatment, a combination of both of them (radiation treatment in case of positive surgical margins) or also brachytherapy (which consists on the prostate implantation of sealed radiotherapy sources) with or without an adjuvant androgen deprivation therapy (ADT) (2–8).

Similarly, advanced stages of the disease count different therapeutic options. As first approach ADT represents the cornerstone of advanced prostate cancer due to the high sensitivity of tumor cells to hormone deprivation. The addition of further treatment including anti-androgens abiraterone acetate or docetaxel can improve the outcome of patients with metastatic castration sensitive prostate cancer (mCSPC) (9–15).

After a first period of hormone deprivation sensitivity, tumor cells develop several mechanisms which lead to overcome the hormone inhibition leading to metastatic castration resistant prostate cancer (mCRPC). In this setting, several different agents have demonstrated to be effective treatment: new hormonal agents (abiraterone, enzalutamide, apalutamide), chemotherapy (docetaxel, cabazitaxel), radiometabolic drugs (Radium 223), and Sipuleucel-T immunotherapy (16–25).

## RATIONAL FOR LIQUID BIOPSY IN PROSTATE CANCER

The availability of several active therapeutic options has led to different emerging needs in clinical practice requiring the development of reliable markers able to monitor response to treatment and help clinicians to select patients more likely to benefit from one approach rather than another.

Prostate-Specific Antigen (PSA) represents a reliable and useful biomarker adopted for early detection and early diagnosis of disease recurrence progression. However, it does not give information about biological features of the disease and it loses its predictive rule in mCRPC setting (26).

Liquid biopsy is an emerging technique which purposes is the detection of tumor cells/tumor DNA from patients' peripheral blood.

There are several issues which make the development of liquid biopsies in prostate cancer an attractive approach: (1) the low invasiveness; (2) the early detection of more aggressive tumors since early phases; (3) the early diagnosis of residual tumors or micro-metastases after surgery; (4) the monitoring of tumor response/progression to systemic treatment in advanced setting of the disease and especially in mCRPC; (5) the prediction of tumor sensitivity/resistance to systemic treatments; (6) the acquisition of an accurate genetic assessment of the disease focusing on key alterations which are related to tumor resistance. In particular, several genomic alterations seem to be attractive target due to their correlation to treatment resistance and/or sensitivity to specific treatments (27–30). Some of the more attractive targets are:

- **Phosphate and tensin homolog (PTEN) loss.** PTEN loss results in PI3K/AKT activation which has been associated to worst survival due to higher tumor proliferation and resistance to hormonal treatment. The inhibition of the PI3K/AKT/mTOR pathway could be an interesting target in this subgroup of patients which could be associated to an Androgen Receptor (AR) inhibition (31, 32).
- **MYC amplification** is generally acquired in metastatic phases of the disease and is correlated to poor prognosis and higher Gleason score. Furthermore, more than one evidences seem to correlate the combination of MYC amplification and PTEN loss to worst prognosis and increase risk of tumor related death (33, 34).
- **Androgen Receptor (AR) mutations** and in particular AR splice variant 7 (AR-V7) is known to be related to resistance to hormonal treatments including also new hormonal agents abiraterone and enzalutamide (35).

- **TMPRSS2-ERG gene fusion** leads to ETS-related gene (ERG) and steroidogenic enzyme AKR1C3 co-overexpression which promotes AR signaling and represents a promising target in prostate cancer (36, 37).
- **DNA repair genes deficiency** and in particular genes related to the identification of single strand breaks (such as PARP1 and PARP2) as well as the identification of the alterations of non-homologous recombination system genes (such as BRCA1, BRCA2, PALB2, MRE11, Check2, RAD51, XRCC2/3) appears an attractive approach for two reasons. First, tumors with repair genes deficiency are related to more aggressive features and poorer survival. Second, therapeutic implications related to these genomic assessments involve a possible sensitivity to platinum cytotoxic therapy. The development of PARP inhibitors represents another possible target for the management of advanced prostate cancer which has already been evaluated in small trials and is currently under clinical investigation (38–43).

Due to these issues, the development of reliable techniques able to perform liquid biopsy appears a promising and suggestive approach (**Table 1**). Here we performed a review of the main techniques adopted or under investigation focusing our attention on approaches based on circulating tumor cell (CTC) and circulating DNA (ct-DNA) detection. **Table 2** summarizes the current methods available for CTC detection as well as the percentage of detection (See also below).

## CIRCULATING TUMOR CELLS IN PROSTATE CANCER

To date, the CellSearch system assay is the only FDA approved method for the detection of CTCs in prostate cancer (**Table 2**). This device consists of different components including a CellPrep system which is a semi-automated sample preparation system and a CellSearch Epithelial Cell Kit. This last component involves ferro-fluids coated with epithelial cell-specific EpCAM antibodies and a mixture of antibodies directed against cytokeratins 8, 18, 19, CD45 conjugated to allophycocyanin and DAPI (nuclear dye 4', 6 diamidino-2-phenylindole for fluorescent cells label). After an incubation period in which CTC are isolated from peripheral blood and enriched in EpCAM composed ferro-fluids the MagNest Cell Presentation Device (a device composed of a chamber with two magnets) orients labeled cells for analysis in a CellSpotter Analyzer (a four-color semi-automated fluorescence microscope) for the CTCs enumeration (44).

Initial studies carried out on patients with different solid tumor demonstrated a promising activity with this method and regarding patients with PCa detection of CTCs was possible in 57% of patients (44).

Further study aimed to investigate the clinical value of CTCs detected by CellSearch assay showed that CTCs baseline levels were an independent prognostic factor for overall survival (OS) (49).

In 2008, de Bono et al. identified a correlation between CTCs number and median overall survival. In this study carried out on 231 mCRPC two distinct subgroup of patients were identified:

**TABLE 1** | An overview of ongoing clinical studies evaluating CTCs/ctDNA in prostate cancer patients.

Trial	Patients enrolled	Study description/outcomes
NCT03284684	Patients undergoing surgery for non-metastatic solid tumors: colon, breast and prostate.	Change in concentration of total mutant circulating DNA. Change in proportion of mutant circulating DNA. Change in integrity index of circulating DNA for ACTB gene. Change in integrity index of circulating DNA for KRAS gene.
NCT02449837	Patients undergoing radiation treatments for one of six cancer types including PC.	To measure CTCs levels to evaluate the change pre- and post-treatment. Change in CTC levels from Baseline to Post-RT treatment and the correlation with local tumor response or pathological evaluation
NCT01961713	Subjects with prostate cancer diagnosed on prostate biopsy who undergo radical prostatectomy	To evaluate the relationship between pre-operative CTC quantity and pathologic stage in men with early stage prostate cancer undergoing prostatectomy. To examine the relationship between persistent CTCs and biochemical recurrence after radical prostatectomy for localized prostate cancer
NCT02997709	Men with intermediate to high risk prostate cancer who are candidates for radiotherapy (RT)	Comparison of Pre- and Post-Treatment Quantitative Imaging Parameters to Changes in Circulating Tumor Cells Over Time in Prostate Cancer Patients Receiving Radiation Therapy (RT) with or without Androgen Deprivation Therapy per standard of care
NCT02853097	Prostate cancer patients at various points throughout androgen deprivation therapy and at the initiation of androgen deprivation therapy, enzalutamide, abiraterone and docetaxel.	To document the appearance of androgen receptor isoform splice variant 7 (AR-V7) expression over the course of therapy in castration-resistant prostate cancer (CRPC). To determine whether detectable AR-V7 is associated with a shortened duration of treatment benefit of abiraterone or enzalutamide.
NCT03089099	mCRPC	To determine whether sequentially analyzing the expression of molecular markers in high volume circulating tumor cells in metastatic castration-resistant prostate cancer patients can predict the therapeutic effects and outcomes of these patients.
NCT03488706	Prostate cancer screening with PSA is plagued by high rate of unnecessary prostate biopsies, especially in the “gray zone” (PSA levels: 4.00 ng/ml e 10.99 ng/ml)	Circulating tumor cells detection Using a circulating-tumor-cell (CTC) test to detect prostate cancer in patients in the PSA “gray zone” level
NCT03236688	mCRPC	Demonstrate detection of ARv7 splice variant transcripts from exosomes in the circulation of MCRPC patients pre and post treatment with selective Androgen pathway inhibitors (i.e., abiraterone and enzalutamide)
NCT02771769	Patients with planned prostate biopsy	Multi-center prospective study in which blood samples will be taken from 1500 male patients aged between 21–80 scheduled for prostate biopsy. Analysis of cell-free cancer DNA extracted from these samples will be undertaken to determine whether copy number instability scores derived from the cfDNA correlates with PSA screening levels and prostate biopsy results (i.e., Gleason score) in these patients
NCT02723526	Patients with newly Diagnosed Metastatic Hormone-Sensitive Prostate Cancer	To determine whether sequentially analyzing the expression of tumor markers in circulating tumor cells in newly diagnosed metastatic hormone-sensitive prostate cancer patients can predict the outcome of these patients.
NCT02742259	Metastatic prostate cancer to the bone	Confirmation of the clinical utility of the cutoff level for the Prostate Cancer Assay for prognosis of progression free survival (PFS) in comparison to the predicate device, CellSearch CTC Assay
NCT02456571	Metastatic PC	To explore the prevalence of expression of four immune checkpoint biomarkers on circulating tumor cells (CTCs) from men with metastatic prostate cancer
NCT02735252	Metastatic PCa.	Develop a first-in-man CTC-based molecular taxonomy of CRPC. Comparison of median PFS to CTC-based AR-v7 status.
NCT02099864	Advanced PCa patients receiving enzalutamide therapy.	Correlation between PSA response and gene expression signatures, DNA copy number alterations, mutations. Assess the association for changes in CTC counts from baseline and maximal PSA observed while on study.

one (Unfavorable group) which showed a CTCs number of 5 or more and the other (favorable) with < 5 CTCs per 7.5 mL of blood. Overall survival was significantly better in favorable group (21.7 vs. 11.5 months). Moreover, patients who presented a significant decrease of CTCs number during or after treatment (moving from unfavorable to favorable groups) significantly improved their survival compared to patients who continued to present a CTCs number of 5 or more CTCs. According to the results of de Bono et al, a meta-analysis of 10 studies confirmed the prognostic rule of CTCs in patients with prostate cancer (50).

Furthermore, pre-planned analyses of large phase III trials: SWOG 20421 (docetaxel with or without atrasentain in mCRPC patients), COU-AA-301 (in which a score composed by LDH levels and CTCs divided patients in 3 different subgroups with favorable, intermediate and poor prognosis) and AFFIRM (enzalutamide in patients with mCRPC progressed to chemotherapy) confirmed the prognostic rule of CTCs as independent factor related to OS (51–53).

Unfortunately, none of these studies demonstrated an association between CTCs number and response to treatment and so the role of CTCs in this setting still remains unclear.

**TABLE 2 |** An overview of CTCs detection techniques.

Method	Mechanism	CTCs detection rate/ other outcomes	Limitations
CELLSEARCH System (44)	A 7.5 mL sample of blood is placed in a special tube, centrifuged to separate solid blood components from plasma, then placed in the CELLTRACKS® AUTOPREP® System. Cells binds ferro-fluid nanoparticles presenting antibodies targeting epithelial adhesion molecules, then CTCs are magnetically separated from other blood cells. CTCs are stained with cytokeratin monoclonal antibodies, DAPI (a DNA stain) and leukocytes which may have contaminated the sample are marked by antibody targeting CD45. Stained CTCs are then placed onto Cell-Spotter Analyzer (a four-color semi-automated fluorescence microscope) for the CTCs enumeration CTCs+: DAPI+, cytokeratine +, CD45—cells	CTCs detected in 57% of patients with prostate cancer	- Low CTCs detection rate in non-metastatic prostate cancer - Conflicting results about correlation between CTCs number and treatment response.
CELLCOLLECTOR EPISPOT (45)	Cell-Collector is based on a sterile stainless steel medical wire, covered with 2 µm gold and a hydrogel layer which is covalently coupled with antibodies against the EpCAM protein and pan-keratins. CD45 staining (performed to exclude unspecific leucocytes) CTCs +: CTCs identified as pan-keratin positive, leukocyte marker CD45 negative. EPISPOT on an EpCAM-independent enrichment method (i.e., leukocyte depletion) and enables the identification of viable PSA-secreting tumor cells CTCs+: PSA+ cells.	Combining Cellsearch, CellCollector and Epispot assay, detection rate of CTCs was 81.3%	- Experimental approach. - This approach does not offer a characterization of CTCs. - Impact on prognosis and predictive value under investigation.
Microfluidic capture of CTCs (46).	Considering the expression of PSA (up-regulated by AR) and PSMA (down regulated by AR) they classified CTCs in AR on, AR mixed and AR positive according to the expression of PSA (+ in AR on and mixed) and PSMA (+ in AR off and mixed).	CTCs detection rate: 80%	- Experimental approach - Under investigation for detection of anti-androgen resistance mechanisms.
EPCAM cells enrichment and sequencing (47).	The recovered cells, enriched with CTCs, were deposited into dense arrays of subnanoliter wells and imaged by automated epifluorescence imaging. Enrichment was obtained through Illumina MagSweeper CTCs expressing EpCAM. Individual EpCAM (+) CD45 (–) CTCs were recovered by robotic micromanipulation for whole genome amplification using multiple displacement amplification.	Mutation concordance between CTCs and primary or metastatic tumor tissue: 86%	- Experimental approach. - High cost. - Loss of concordance between CTCs mutation and primary/metastatic tumor tissues.
ADNAtest (48).	Is a device able to isolate MUC1-negative and EpCAM positive CTCs. After CTCs isolation, cells are lysed and RNA is extracted for downstream analyses with RT-PCR. Of note this device adopts primers against EGFR, PSA and PSMA making a sample positive if one of these genes are expressed.	CTCs detection rate: 62%	- Experimental approach - High cost - Few data about the application of this approach in localized/ non metastatic prostate cancer.

Moreover, another possible issue which could partially explain the failure of this approach in clinical practice is the low detection rate of CTCs in non metastatic patients which ranges only from 5 to 27% (54). To avoid this problem, Kuske et al combined three different methods for the detection of CTCs before and after prostatectomy in non metastatic patients with PC, CellSearch system assay, CellCollector (a system capturing EpCAM-positive CTCs by an antibody-coated needle introduced in arm vein) and EPISPOT (a system able to enrich CTCs by negative depletion of leukocytes and detects circulating prostate cancer cells thanks to their active secretion of PSA) (45). CTCs were detected in 37, 54.9 and 58.7% of patients using CellSearch, CellCollector, and EPISPOT, respectively. The cumulative positivity rate of the three CTC assay was 81.3% and despite it is not a validated approach, it represents an attractive early method able to estimate the risk of tumor recurrence or persistence after surgery.

A combined analysis of COU-AA-301 and IMMC-38 trials showed that an increase of 30% in CTCs count from baseline was independently associated to worst OS in patients treated with

abiraterone and chemotherapy (53). To sustain the correlation between CTCs count changes and survival, another analysis performed on 119 patients with CRPC treated at the Royal Marsden Hospital suggested that a decrease of 30% in CTC counts from baseline was associated to improved survival (55).

The only CTCs enumeration resulted in an independent prognostic factor with an unclear role in terms of early diagnosis of disease recurrence/persistence after surgery as well as a predictive response factor. Another interesting approach consists in a characterization of CTCs resulting in a genetic assessment and in a detection of target altered pathways.

AR protein has been extensively investigated in prostate cancer CTCs. Through a FISH based assay AR gene amplification detection in CTCs was possible in 40% of cases, a percentage comparable to the AR amplification described in bone metastases biopsy analyses (47). Further investigations demonstrated that patients with higher cytoplasm expression of AR resulting in a reduction of nuclear translocation was significantly associated to better response to docetaxel (56). By a microfluidic capture



of CTCs Miyamoto et al. evaluated dynamic changes in CTCs AR expression. In particular, considering the expression of PSA (up-regulated by AR) and PSMA (down regulated by AR) they classified CTCs in AR on, AR mixed and AR positive according to the expression of PSA (+ in AR on and mixed) and PSMA (+ in AR off and mixed). Moreover, Authors identified that AR status changed from “on” to “off” during ADT while patients treated with abiraterone acetate with an increase of AR-on CTCs or baseline level of AR-mixed more than 10% were significantly associated to worse overall survival (46). The technology developed by Miyamoto et al was also adopted for the detection of anti-androgen resistance mechanisms in CTCs demonstrating higher activation of Wnt signaling and considerable heterogeneity in signaling pathways, expression of AR gene mutations and splicing variants (57).

Due to the important role of AR-V7 in mCRPC (35), several studies have focused on the detection of this splice variants on CTCs. An EpCAM assay demonstrated that CTCs-ARV7+ detection was associated to resistance to enzalutamide and abiraterone but no to docetaxel and cabazitaxel and that the detection of these CTCs was independently associated to worse clinical outcome compared to patients with CTCs-ARV7- cells (58–60). Other studies modified the CTCs and ARV7 detection method in order to evaluate the AR-V7 cellular localization (61) and the presence of other splice variants of AR (62). Particularly, not only AR-V7 but also other splice variants of the AR protein were significantly associated to worse progression free survival. Moreover 6 of 17 poor responders to treatment were AR-V7 negative, but carried other AR perturbations (62).

About other pathways detected in CTCs, the PTEN loss assessed by FISH and Epic Sciences test (an assay which adopted a fiberoptic array scanning techniques for the detection of DAPI, CD45, cytokeratins stained cells) has been associated to worse clinical outcomes (63, 64) while the detection of TMPRSS2-ERG fusion gene performed by microfluidic device and by the use of a RT-PCR analysis failed to show a predictive response value to abiraterone acetate in mCRPC patients (47).

Next Generation Sequencing (NGS) involves a series of different techniques able to perform a whole genome sequencing of tumor cells. The possibility to obtain a complete genomic assessment from CTCs appears a novel and promising approach investigated in different studies.

In 2014, Lohr et al evaluated a method able to perform a CTCs isolation, enrichment (thruEp-CAM expressing CTCs), genomic amplification and sequencing in metastatic PC (65). They demonstrated that a complete mapping of the standard exome was possible in CTCs. NGS analysis of CTCs and tumor sample of a single patient with advanced prostate cancer showed a concordance of 86% from the mutations isolated in CTCs and genomic anomalies identified in primary or metastatic tumors (66). Despite NGS performed to CTCs represents an attractive approach, to date no validated or prospective studies have been carried out and so this method is still under investigation.

Another interesting issue is the detection of whole blood RNA, without enriching for CTC. In 2012, Ross et al assessed a whole blood RNA transcript based model as prognostic factor in patients with PC. After the analysis of blood collected from

62 men with mCRPC, they identified a six gene model (genes considered were: ABL2, SEMA4D, ITGAL, C1QA, TIMP1, CDKN1A which are genes involved mainly in immunity regulation) able to divide patients in two risk groups with different mOS (67). In the same year, Olmos et al carried out a validation study of a nine-gene signature as prognostic factor (68). Design of the study consisted in a derivation set in which patients with mCRPC and patients in Active Surveillance were used as case and control groups respectively. After genomic assessment 94 patients were divided in four distinct prognostic groups. Thus nine altered genes HMBS, TMCC2, SLC4A1, STOM, GABARAPL2, RIOK3, TERF2IP, TFDPI isolated in prognostic groups with worst survival (composed of only mCRPC patients) were validated in a validation set of patients with mCRPC. More recently, an assessment of 5 key genes (KLK3, KLK2, HOXB13, GRHL2, FOXA1) obtained after reverse transcription polymerase chain reaction (RT-PCR) demonstrated to be a reliable prognostic marker compared to CellSearch system count (48). Isolation of two or more of the selected genes were possible in 53% (51/97) patients with mCRPC. AdnaTest is a technique adopted for CTCs enrichment and consists of a device able to isolate MUC1-negative and EpCAM positive CTCs. After CTCs isolation, cells are lysed and RNA is extracted for downstream analyses with RT-PCR. Of note this device adopts primers against EGFR, PSA, and PSMA making a sample positive if one of these genes are expressed. Sensitivity of KLK2, KLK3, HOXB13, GRHL2, and FOXA1 genes detection by this method is similar to DDPCR (direct detection PCR) and both of these techniques showed a higher sensitivity compared to CellSearch system (69).

Concerning the several devices utilized for CTCs detection, enrichment and evaluation, only CellSearch has been approved from FDA. However, despite a large range of potential applications (such as diagnosis, evaluation of treatment response, early detection of tumor relapse, and progression) CTCs detection by CellSearch is not commonly adopted in clinical practice. This mainly due to a low sensitivity of the method as well as a conflicting relationship between CTCs and treatment response evaluation. Several other approaches are under investigation. It is likely that CTCs evaluation will be an important factor able to improve our decisions in clinical practice (48, 69).

## CIRCULATING DNA IN PROSTATE CANCER

The evidence that cell-free DNA could be detected in peripheral blood is a well known issue, and its application in clinical practice has been investigated only in last years. Regarding cancer patients, the unique composition of tumors' ctDNA presenting several genomic mutations (especially single base-pair substitution) which are not detectable in ctDNA originating from normal cells make tumor ctDNA an ideal markers of the disease. Moreover, the possible correlation between mutations detected on ctDNA and genomic mutations of primary or metastatic

tumors make ctDNA a unique markers able to provide key information by a no-invasive approach.

Regarding PC, ctDNA could be detected in peripheral blood and detection of known driver aberrations can be obtained in more than 97% of cases. Moreover, changings in ctDNA genomic mutations could be detected by repeated analyses of ctDNA with high grade of concordance with genomic assessment of primary tumors or metastases (70, 71).

The quantitative assessment of ctDNA has been related to prognosis of patients with PC in different studies (72, 73). In particular, Romanel A et al examined AR status of mCRPC patients starting abiraterone acetate. They detected a 45% of patients (tot number 97) with AR point mutations (T878A/L702H) before the first administration of abiraterone who showed a significant worse overall survival (73). Similarly, other studies confirmed the prognostic role of AR genomic alterations as prognostic markers raising the acquisition of ctDNA examination as a possible to monitor response to hormonal agents and to achieve an early diagnosis of progressive disease (74–77).

As known, mutation in DNA repair genes is acquiring an increasing interest in PC due to the association by these mutation and more aggressive tumor features and to the possible benefit derived from a PARP targeted treatment. Mutations in repair genes are common in prostate cancer. In a DNA assessment of 692 men with metastatic PC a total of 84 germline DNA repair gene mutations (BRCA2, ATM, CHEK2, BRCA1, RAD51D, and PALB2) were found in 82 men (78). This study demonstrated that incidence of germline mutations of DNA-repair gene were common (as detected in 11.8% of all patients) in metastatic patients regardless to age and family history of prostate cancer.

PARP inhibition is one of the important strategy currently under investigation in patients with metastatic prostate cancer. In a phase II trial 50 mCRPC patients received the PARP inhibitor Olaparib (41). 17 (33%) patients showed an objective response while NGS sequencing showed that 16 patients presented homozygous deletions, deleterious mutations, or both in DNA repair genes (BRCA1/2, ATM, Fanconi's anemia genes and CHECK2). A subsequent analysis of tumors DNA highlighted that patients with an overall reduction of 50% or more of ctDNA were associated with better OS and PFS (79). In ASCO 2018, Clarke et al presented the results of a phase II study comparing the administration of Abiraterone with Olaparib or placebo in 171 patients with mCRPC (80). This trial met its primary endpoint showing a better radiological PFS in patients receiving olaparib. Of note Authors researched homologous recombination repair mutations by a NGS approach in tumor samples and plasma. Sequencing was possible on 91 of 136

patients and positive results (defined as discovery mutated patients) was obtained in 13 patients. By germline analysis and tumor sample analyses detection of homologous recombination repair mutations were identified in 3 patients (38 tumor samples analyzed of 68 total samples) and 7 patients (by a germline analysis of 102 patients). Results of this study raising the dosage and analyses of ctDNA as possible approach for the detection of keys DNA-repair gene mutations. Other larger prospective trials are needed to explore the role of ctDNA in this setting.

Another promising target gene is represented by PTEN loss which has show to be a predictive biomarker of response to treatments targeting PI3K/AKT pathway. Hyper-activation of the PI3K/Akt/mTOR resulting from PTEN loss is related to decreased AR transcription output and stability and vice versa. The addition of ipatasertib (an Akt inhibitor) to abiraterone acetate increased radiological PFS of patients with mCRPC and PTEN loss previously treated with docetaxel based therapy and progressed during at last one previous hormonal therapy (81).

Extracellular vesicles are membrane-enclosed structures that are released from all cells in the body. These vesicles contain several substances such as proteins, lipids, RNA, and DNA and are considered a very promising tumor-related biomarkers. Recently, it has been demonstrated that large extracellular vesicles isolated from plasma of patients with prostate cancer cells are an important source of chromosomal DNA which reflects faithfully genetic aberration of the cell of origin, including copy number variations of genes frequently altered in metastatic prostate cancer (such as MYC and PTEN) (82). The study of extracellular vesicles represents a novel and promising approach for biomarkers development in prostate cancer however further studies are needed to explore the effective value of this method.

## CONCLUSION

The surprising potential of CTCs or tumors' ctDNA detection, characterization and genomic assessment have start a revolution which probably will give important results in next years. Despite to date application of these techniques are few probably that better knowledge of genomic anomalies of PC and their correlation with the clinical course of the disease as well as their potential relationship with specific targeted treatments will increase the attention on this issue.

## AUTHOR CONTRIBUTIONS

RM and FM: conception and design; VD and LG: drafting the manuscript; MSa and AC: review of the literature; LC, MSc, and AL-B: critical revision of the manuscript.

## REFERENCES

1. Siegel RL, Miller KD, Jemal A. Cancer statistics 2018. *CA Cancer J Clin.* (2018) 68:7–30. doi: 10.3322/caac.21442
2. Wilt TJ, Jones KM, Barry MJ, Andriole GL, Culin D, Wheeler T, et al. Follow-up of prostatectomy versus observation for early prostate cancer. *N Engl J Med.* (2017) 377:132–42. doi: 10.1056/NEJMoa1615869
3. Bill-Axelson A, Holmberg L, Garmo H, Rider JR, Taari K, Busch C, et al. Radical prostatectomy or watchful waiting in early prostate cancer. *N Engl J Med.* (2014) 370:932–42. doi: 10.1056/NEJMoa1311593
4. Parker C. Active surveillance: towards a new paradigm in the management of early prostate cancer. *Lancet Oncol.* (2004) 5:101–6. doi: 10.1016/S1470-204501384-1

5. Hamdy FC, Donovan JL, Lane JA, Mason M, Metcalfe C, Holding P, et al. 10-Year outcomes after monitoring, surgery, or radiotherapy for localized prostate cancer. *N Engl J Med.* (2016) 375:1415–24. doi: 10.1056/NEJMoa1606220
6. Donovan JL, Hamdy FC, Lane JA, Mason M, Metcalfe C, Walsh E, et al. Patient-reported outcomes after monitoring, surgery, or radiotherapy for prostate cancer. *N Engl J Med.* (2016) 375:1425–37. doi: 10.1056/NEJMoa1606221
7. Mason MD, Parulekar WR, Sydes MR, Brundage M, Kirkbride P, Gospodarowicz M, et al. Final report of the intergroup randomized study of combined androgen-deprivation therapy plus radiotherapy versus androgen-deprivation therapy alone in locally advanced prostate cancer. *J Clin Oncol.* (2015) 33:2143–50. doi: 10.1200/JCO.2014.57.7510
8. Messing EM, Manola J, Yao J, Kiernan M, Crawford D, Wilding G, et al. Immediate versus deferred androgen deprivation treatment in patients with node-positive prostate cancer after radical prostatectomy and pelvic lymphadenectomy. *Lancet Oncol.* (2006) 7:472–9. doi: 10.1016/S1470-204570700-8
9. Schmitt B, Bennett C, Seidenfeld J, Samson D, Wilt T. Maximal androgen blockade for advanced prostate cancer. *Cochrane Database Syst Rev.* (2000) 22:CD001526. doi: 10.1016/j.beem.2008.01.004
10. Samson DJ, Seidenfeld J, Schmitt B, Hasselblad V, Albertsen PC, Bennett CL, et al. Systematic review and meta-analysis of monotherapy compared with combined androgen blockade for patients with advanced prostate carcinoma. *Cancer* (2002) 95:361–76. doi: 10.1002/cncr.10647
11. Gravis G, Boher JM, Joly F, Soulie M, Albiges L, Priou F, et al. Androgen Deprivation Therapy (ADT) plus docetaxel versus ADT alone in metastatic non castrate prostate cancer: impact of metastatic burden and long-term survival analysis of the randomized phase 3 GETUG-AFU15 Trial. *Eur Urol.* (2016) 70:256–62. doi: 10.1016/j.eururo.2015.11.005
12. James ND, Sydes MR, Clarke NW, Mason MD, Dearnaley DP, Spears MR, et al. Addition of docetaxel, zoledronic acid, or both to first-line long-term hormone therapy in prostate cancer (STAMPEDE): survival results from an adaptive, multiarm, multistage, platform randomised controlled trial. *Lancet* (2016) 387:1163–77. doi: 10.1016/S0140-673601037-5
13. Kyriakopoulos CE, Chen YH, Carducci MA, Liu G, Jarrard DF, Hahn NM, et al. Chemohormonal therapy in metastatic hormone-sensitive prostate cancer: long-term survival analysis of the randomized phase III E3805 CHAARTED trial. *J Clin Oncol.* (2018) 36:1080–7. doi: 10.1200/JCO.2017.75.3657
14. James ND, de Bono JS, Spears MR, Clarke NW, Mason MD, Dearnaley DP, et al. Abiraterone for prostate cancer not previously treated with hormone therapy. *N Engl J Med.* (2017) 377:338–51. doi: 10.1056/NEJMoa1702900
15. Fizazi K, Tran N, Fein L, Matsubara N, Rodriguez-Antolin A, Alekseev BY, et al. Abiraterone plus prednisone in metastatic, castration-sensitive prostate cancer. *N Engl J Med.* (2017) 377:352–60. doi: 10.1056/NEJMoa1704174
16. Tannock IF, de Wit R, Berry WR, Horti J, Pluzanska A, Chi KN. Docetaxel plus prednisone or mitoxantrone plus prednisone for advanced prostate cancer. *N Engl J Med.* (2004) 351:1502–12. doi: 10.1056/NEJMoa040720
17. Petrylak DP, Tangen CM, Hussain MH, Lara PN, Jones JA, Taplin ME. Docetaxel and estramustine compared with mitoxantrone and prednisone for advanced refractory prostate cancer. *N Engl J Med.* (2004) 351:1513–20. doi: 10.1056/NEJMoa041318
18. De Bono JS, Logothetis CJ, Molina A, Fizazi K, North S, Chu L. Abiraterone and increased survival in metastatic prostate cancer. *N Engl J Med.* (2011) 364:1995–2005. doi: 10.1056/NEJMoa1014618
19. Ryan CJ, Smith MR, de Bono JS, Molina A, Logothetis CJ, de Souza P, et al. Abiraterone in metastatic prostate cancer without previous chemotherapy. *N Engl J Med.* (2013) 368:138–48. doi: 10.1056/NEJMoa1209096
20. Beer TM, Armstrong AJ, Rathkopf DE, Loriot Y, Sternberg CN, Higano CS, et al. Enzalutamide in metastatic prostate cancer before chemotherapy. *N Engl J Med.* (2014) 371:424–33. doi: 10.1056/NEJMoa1405095
21. Scher HI, Fizazi K, Saad F, Taplin ME, Sternberg CN, Miller K, et al. Increased survival with enzalutamide in prostate cancer after chemotherapy. *N Engl J Med.* (2012) 367:1187–97. doi: 10.1056/NEJMoa1207506
22. Smith MR, Saad F, Chowdhury S, Oudard S, Hadaschik BA, Graff JN, et al. Apalutamide treatment and metastasis-free survival in prostate cancer. *N Engl J Med.* (2018) 378:1408–18. doi: 10.1056/NEJMoa1715546
23. De Bono JS, Oudard S, Ozguroglu M, Hansen S, Machiels JP, Kocak I, et al. Prednisone plus cabazitaxel or mitoxantrone for metastatic castration-resistant prostate cancer progressing after docetaxel treatment: a randomised open-label trial. *Lancet* (2010) 376:1147–54. doi: 10.1016/S0140-673661389-X
24. Parker C, Nilsson S, Heinrich D, Helle SI, O'Sullivan JM, Fosså SD, et al. Alpha emitter radium-223 and survival in metastatic prostate cancer. *N Engl J Med.* (2013) 369:213–23. doi: 10.1056/NEJMoa1213755
25. Kantoff PW, Higano CS, Shore ND, Berger ER, Small EJ, Penson DF, et al. Sipuleucel-T immunotherapy for castration-resistant prostate cancer. *N Engl J Med.* (2010) 363:411–22. doi: 10.1056/NEJMoa1001294
26. Scher HI, Morris MJ, Larson S, Heller G. Validation and clinical utility of prostate cancer biomarkers. *Nat Rev Clin Oncol.* (2013) 10:225–34. doi: 10.1038/nrclinonc.2013.30
27. Cancer Genome Atlas Research Network. The molecular taxonomy of primary prostate cancer. *Cell* (2015) 163:1011–25. doi: 10.1016/j.cell.2015.10.025
28. Robinson D, Van Allen EM, Wu YM, Schultz N, Lonigro RJ, Mosquera JM, et al. Integrative clinical genomics of advanced prostate cancer. *Cell* (2015) 161:1215–28. doi: 10.1016/j.cell.2015.05.001
29. Massari F, Di Nunno V, Comito F, Cubelli M, Ciccarese C, Iacovelli R, et al. Circulating tumor cells in genitourinary tumors. *Ther Adv Urol.* (2017) 10:65–77. doi: 10.1177/1756287217742564
30. Ciccarese C, Montironi R, Fiorentino M, Martignoni G, Brunelli M, Iacovelli R, et al. Circulating tumor cells: a reliable biomarker for prostate cancer treatment assessment? *Curr Drug Metab.* (2017) 18:692–9. doi: 10.2174/1389200218666170518163549
31. Mulholland DJ, Tran LM, Li Y, Cai H, Morim A, Wang S, et al. Cell autonomous role of PTEN in regulating castration-resistant prostate cancer growth. *Cancer Cell* (2011) 19:792–804. doi: 10.1016/j.ccr.2011.05.006
32. De Bono JS, De Giorgi U, Massard C, Bracarda S, Nava Rodrigues D, Kocak I, et al. PTEN loss as a predictive biomarker for the Akt inhibitor ipatasertib combined with abiraterone acetate in patients with metastatic castration-resistant prostate cancer (mCRPC). *Ann Oncol.* (2016) 27(Suppl. 6):vi243–65. doi: 10.1093/annonc/mdw372.02
33. Anderson PD, McKissic SA, Logan M, Roh M, Franco OE, Wang J, et al. Nkx3.1 and Myc crossregulate shared target genes in mouse and human prostate tumorigenesis. *J Clin Invest.* (2012) 122:1907–19. doi: 10.1172/JCI58540
34. Kirschner AN, Wang J, van der Meer R, Anderson PD, Franco-Coronel OE, Kushner MH, et al. PIM kinase inhibitor AZD1208 for treatment of MYC-driven prostate cancer. *J Natl Cancer Inst.* (2014) 107:dju407. doi: 10.1093/jnci/dju407
35. Antonarakis ES, Lu C, Wang H, Luber B, Nakazawa M, Roeser JC, et al. AR-V7 and resistance to enzalutamide and abiraterone in prostate cancer. *N Engl J Med.* (2014) 371:1028–38. doi: 10.1056/NEJMoa1315815
36. Stone L. Prostate cancer: mastering transcription: TMPRSS2-ERG and the cis-regulatory landscape. *Nat Rev Urol.* (2017) 14:579. doi: 10.1038/nrurol.2017.141
37. Brenner JC, Ateeq B, Li Y, Yocum AK, Cao Q, Asangani IA, et al. Mechanistic rationale for inhibition of poly(ADP-ribose) polymerase in ETS gene fusion-positive prostate cancer. *Cancer Cell* (2011) 19:664–78. doi: 10.1016/j.ccr.2011.04.010
38. Aparicio AM, Harzstark AL, Corn PG, Wen S, Araujo JC, Tu SM, et al. Platinum-based chemotherapy for variant castrate-resistant prostate cancer. *Clin Cancer Res.* (2013) 19:3621–30. doi: 10.1158/1078-0432.CCR-12-3791
39. Sternberg CN, Petrylak DP, Sartor O, Witjes JA, Demkow T, Ferrero JM, et al. Multinational, double-blind, phase III study of prednisone and either satraplatin or placebo in patients with castrate-refractory prostate cancer progressing after prior chemotherapy: the SPARC trial. *J Clin Oncol.* (2009) 27:5431–8. doi: 10.1200/JCO.2008.20.1228
40. Cerrato A, Morra F, Celetti A. Use of poly ADP ribose polymerase [PARP] inhibitors in cancer cells bearing DDR defects: the rationale for their inclusion in the clinic. *J Exp Clin Cancer Res.* (2016) 35:179. doi: 10.1186/s13046-016-0456-2
41. Mateo J, Carreira S, Sandhu S, Miranda S, Mossop H, Perez-Lopez R, et al. DNA-repair defects and olaparib in metastatic prostate cancer. *N Engl J Med.* (2015) 373:1697–708. doi: 10.1056/NEJMoa1506859



42. Schiewer MJ, Goodwin JF, Han S, Brenner JC, Augello MA, Dean JL, et al. Dual roles of PARP-1 promote cancer growth and progression. *Cancer Discov.* (2012) 2:1134–49. doi: 10.1158/2159-8290.CD-12-0120
43. Ciccicarese C, Massari F, Iacovelli R, Fiorentino M, Montironi R, Di Nunno V et al. Prostate cancer heterogeneity: discovering novel molecular targets for therapy. *Cancer Treat Rev.* (2017) 54:68–73. doi: 10.1016/j.ctrv.2017.02.001
44. Allard WJ, Matera J, Miller MC, Repollet M, Connely MC, Rao C et al. Tumor cells circulate in the peripheral blood of all major carcinomas but not in healthy subjects or patients with nonmalignant diseases. *Clin Cancer Res.* (2004) 10: 6897–904. doi: 10.1158/1078-0432.CCR-04-0378
45. Kuske A, Gorges TM, Tennstedt P, Tiebel AK, Pompe R, Preißer F, et al. Improved detection of circulating tumor cells in non-metastatic high-risk prostate cancer patients. *Sci Rep.* (2016) 6:39736. doi: 10.1038/srep39736
46. Miyamoto DT, Lee RJ, Stott SL, Ting DT, Wittner BS, Ulman M, et al. Androgen receptor signaling in circulating tumor cells as a marker of hormone-ally responsive prostate cancer. *Cancer Discov.* (2012) 2:995–1003. doi: 10.1158/2159-8290.CD-12-0222
47. Leversha MA, Han J, Asgari Z, Danila DC, Lin O, Gonzalez-Espinoza R, et al. Fluorescence *in situ* hybridization analysis of circulating tumor cells in metastatic prostate cancer. *Clin. Cancer Res.* (2009) 15:2091–7. doi: 10.1158/1078-0432.CCR-08-2036
48. Danila DC, Anand A, Schultz N, Heller G, Wan M, Sung CC, et al. Analytic and clinical validation of a prostate cancer-enhanced messenger RNA detection assay in whole blood as a prognostic bio- marker for survival. *Eur Urol.* (2014) 65:1191–7. doi: 10.1016/j.eururo.2013.07.006
49. Danila DC, Heller G, Gignac GA, Gonzalez-Espinoza R, Anand A, Tanaka E, et al. Circulating tumor cell number and prognosis in progressive castration-resistant prostate cancer. *Clin Cancer Res.* (2007) 13:7053–8. doi: 10.1158/1078-0432.CCR-07-1506
50. Zheng Y, Zhang C, Wu J, Cheng G, Yang H, Hua L et al. Prognostic value of circulating tumor cells in castration resistant prostate cancer: a meta-analysis. *Urol J.* (2016) 13:2881–8. doi: 10.22037/uj.v13i6.3592
51. Goldkorn A, Ely B, Quinn DI, Tangen CM, Fink LM, Xu T et al. Circulating tumor cell counts are prognostic of overall survival in SWOG S0421: a phase III trial of docetaxel with or without atrasentan for metastatic castration-resistant prostate cancer. *J Clin Oncol.* (2014) 32: 1136–42. doi: 10.1200/JCO.2013.51.7417
52. Fleisher M, Danila DC, Fizazi K, Hirmand M, Selby B, Phung D et al. Circulating tumor cell (CTC) enumeration in men with metastatic castration-resistant prostate cancer (mCRPC) treated with enzalutamide post- chemotherapy (phase 3 AFFIRM study). *J Clin Oncol.* (2015) 33:5035. doi: 10.1200/jco.2015.33.15\_suppl.5035
53. Lorente D, Olmos D, Mateo J, Dolling D, Bianchini D, Seed G et al. Circulating tumor cell increase as a biomarker of disease progression in metastatic castration-resistant prostate cancer patients with low baseline CTC counts. *Ann Oncol.* (2018) 29:1554–60. doi: 10.1093/annonc/mdy172
54. Thalgot M, Rack B, Horn T, Heck MM, Eiber M, Kübler H, et al. Detection of circulating tumor cells in locally advanced high-risk prostate cancer during neoadjuvant chemotherapy and radical prostatectomy. *Anticancer Res.* (2015) 35:5679–85.
55. Olmos D, Arkenau HT, Ang JE, Ledaki I, Attard G, Carden CP, et al. Circulating tumour cell (CTC) counts as intermediate end points in castration-resistant prostate cancer (CRPC): a single- centre experience. *Ann Oncol.* (2009) 20:27–33. doi: 10.1093/annonc/mdn544
56. Darshan MS, Loftus MS, Thadani-Mulero M, Levy BP, Escuin D, Zhou XK, et al. Taxane-induced blockade to nuclear accumulation of the androgen receptor predicts clinical responses in metastatic prostate cancer. *Cancer Res.* (2011) 71:6019–29. doi: 10.1158/0008-5472.CAN-11-1417
57. Miyamoto DT, Zheng Y, Wittner BS, Lee RJ, Zhu H, Broderick KT, et al. RNA-Seq of single prostate CTCs implicates noncanonical Wnt signaling in antiandrogen resistance. *Science* (2015) 349:351–6. doi: 10.1126/science.aab0917
58. Antonarakis ES, Lu C, Luber B, Wang H, Chen Y, Nakazawa M, et al. Androgen receptor splice variant 7 and efficacy of taxane chemotherapy in patients with metastatic castration-resistant prostate cancer. *JAMA Oncol.* (2015) 1:582–91. doi: 10.1001/jamaoncol.2015.1341
59. Onstenk W, Sieuwerts AM, Kraan J, Van M, Nieuweboer AJ, Mathijssen RH, et al. Efficacy of cabazitaxel in castration- resistant prostate cancer is independent of the presence of AR-V7 in circulating tumor cells. *Eur Urol.* (2015) 68:939–45. doi: 10.1016/j.eururo.2015.07.007
60. Antonarakis ES, Lu C, Luber B, Wang H, Chen Y, Zhu Y, et al. Clinical significance of androgen receptor splice variant-7 mRNA detection in circulating tumor cells of men with metastatic castration-resistant prostate cancer treated with first- and second- line abiraterone and enzalutamide. *J Clin Oncol.* (2017) 35:2149–56. doi: 10.1200/JCO.2016.70.1961
61. Scher HI, Lu D, Schreiber NA, Louw J, Graf RP, Vargas HA, et al. Association of AR-V7 on circulating tumor cells as a treatment-specific biomarker with outcomes and survival in castration- resistant prostate cancer. *JAMA Oncol.* (2016) 2:1441–9. doi: 10.1001/jamaoncol.2016.1828
62. De Laere B, van Dam PJ, Whittington T, Mayrhofer M, Diaz EH, Van den Eynden G, et al. Comprehensive profiling of the androgen receptor in liquid biopsies from castration-resistant prostate cancer reveals novel intra-AR structural variation and splice variant expression patterns. *Eur Urol.* (2017);72:192–200. doi: 10.1016/j.eururo.2017.01.011
63. Werner SL, Graf RP, Landers M, Valenta DT, Schroeder M et al. Analytical validation and capabilities of the epic CTC platform: enrichment-free circulating tumour cell detection and characterization. *J Circ Biomark.* (2015) 4:3. doi: 10.5772/60725
64. Punnoose EA, Ferraldeschi R, Szafer-Glusman E, Tucker EK, Mohan S, Flohr P, et al. PTEN loss in circulating tumour cells correlates with PTEN loss in fresh tumour tissue from castration-resistant prostate cancer patients. *Br J Cancer* (2015) 113:1225–33. doi: 10.1038/bjc.2015.332
65. Lohr JG, Adalsteinsson VA, Cibulskis K, Choudhury AD, Rosenberg M, Cruz-Gordillo P, et al. Whole-exome sequencing of circulating tumor cells provides a window into metastatic prostate cancer. *Nat Biotechnol.* (2014) 32:479–84. doi: 10.1038/nbt.2892
66. Jiang R, Lu YT, Ho H, Li B, Chen JF, Lin M, et al. A comparison of isolated circulating tumor cells and tissue biopsies using whole- genome sequencing in prostate cancer. *Oncotarget* (2015) 6:44781–93. doi: 10.18632/oncotarget.6330
67. Ross RW, Galsky MD, Scher HI, Magidson J, Wassmann K, Lee GS, et al. A whole-blood RNA transcript-based prognostic model in men with castration-resistant prostate cancer: a prospective study. *Lancet Oncol.* (2012) 13:1105–13. doi: 10.1016/S1470-204570263-2
68. Olmos D, Brewer D, Clark J, Danila DC, Parker C, Attard G, et al. Prognostic value of blood mRNA expression signatures in castration-resistant prostate cancer: a prospective, two-stage study. *Lancet Oncol.* (2012) 13:1114–24. doi: 10.1016/S1470-204570372-8
69. Danila DC, Samoila A, Patel C, Schreiber N, Herkal A, Anand A, et al. Clinical validity of detecting circulating tumor cells by AdnaTest assay compared with direct detection of tumor mRNA in stabilized whole blood, as a biomarker predicting overall survival for metastatic castration-resistant prostate cancer patients. *Cancer J.* (2016) 22:315–20. doi: 10.1097/PPO.0000000000000220
70. Ulz P, Belic J, Graf R, Auer M, Lafer I, Fischereider K, et al. Whole- genome plasma sequencing reveals focal amplifications as a driving force in metastatic prostate cancer. *Nat Commun.* (2016) 7:12008. doi: 10.1038/ncomms12008
71. Wyatt AW, Annala M, Aggarwal R, Beja K, Feng F, Youngren J, et al. Concordance of circulating tumor DNA and matched meta- static tissue biopsy in prostate cancer. *J Natl Cancer Inst.* (2018) 110:78–86. doi: 10.1093/jnci/djx118
72. Carreira S, Romanel A, Goodall J, Grist E, Ferraldeschi R, Miranda S, et al. Tumor clone dynamics in lethal prostate cancer. *Sci Transl Med.* (2014); 6:254ra125. doi: 10.1126/scitranslmed.3009448
73. Romanel A, Gasi Tandefelt D, Conteduca V, Jayaram A, Casiraghi N, Wetterskog D, et al. Plasma AR and abiraterone-resistant prostate cancer. *Sci Transl Med.* (2015) 7:312re10. doi: 10.1126/scitranslmed.aac9511
74. Conteduca V, Wetterskog D, Sharabiani MTA, Grande E, Fernandez-Perez MP, Jayaram A, et al. Androgen receptor gene status in plasma DNA associates with worse outcome on enzalutamide or abiraterone for castration-resistant prostate cancer: a multi-institution correlative biomarker study. *Ann Oncol.* (2017) 28:1508–16. doi: 10.1093/annonc/mdx155
75. Lallous N, Volik SV, Awrey S, Leblanc E, Tse R, Murillo J, et al. Functional analysis of androgen receptor mutations that confer anti- androgen resistance identified in circulating cell-free DNA from prostate cancer patients. *Genome Biol.* (2016) 17:10. doi: 10.1186/s13059-015-0864-1
76. Wyatt AW, Azad AA, Volik SV, Annala M, Beja K, McConeghy B, et al. Genomic alterations in cell-free DNA and enzalutamide resistance



- in castration-resistant prostate cancer. *JAMA Oncol.* (2016) 2:1598–606. doi: 10.1001/jamaoncol.2016.0494
77. Azad AA, Volik SV, Wyatt AW, Haegert A, Le Bihan S, Bell RH, et al. Androgen receptor gene aberrations in circulating cell-free DNA: biomarkers of therapeutic resistance in castration-resistant prostate cancer. *Clin Cancer Res.* (2015) 21:2315–24. doi: 10.1158/1078-0432.CCR-14-2666
  78. Pritchard CC, Mateo J, Walsh MF, De Sarkar N, Abida W, Beltran H, et al. Inherited DNA-repair gene mutations in men with meta- static prostate cancer. *N Engl J Med.* (2016) 375:443–53. doi: 10.1056/NEJMoa1603144
  79. Goodall J, Mateo J, Yuan W, Mossop H, Porta N, Miranda S, et al. Circulating cell-free DNA to guide prostate cancer treatment with PARP inhibition. *Cancer Discov.* (2017) 7:1006–17. doi: 10.1158/2159-8290.CD-17-0261
  80. Clarke N, Wiechno P, Alekseev B, Sala N, Jones R, Kocak I. Olaparib combined with abiraterone in patients with metastatic castration-resistant prostate cancer: a randomised, double-blind, placebo-controlled, phase 2 trial. *Lancet Oncol.* (2018) 19:975–86. doi: 10.1016/S1470-2045(18)30365-6
  81. de Bono JS, De Giorgi U, Nava Rodrigues D, Massard C, Bracarda S, Font A, et al. Randomized phase II study of Akt blockade with or without Ipatasertib in Abiraterone-treated patients with metastatic prostate cancer with and without PTEN loss. *Clin Cancer Res.* (2018). doi: 10.1158/1078-0432.CCR-18-0981. [Epub ahead of print].
  82. Vagner T, Spinelli C, Minciaccchi VR, Balaj L, Zandian M, Conley A, et al. Large extracellular vesicles carry most of the tumour DNA circulating in prostate cancer patient plasma. *J Extracell Vesicles* (2018) 7:1505403. doi: 10.1080/20013078.2018.1505403

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Di Nunno, Gatto, Santoni, Cimadamore, Lopez-Beltran, Cheng, Scarpelli, Montironi and Massari. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.



# Urinary Markers in Bladder Cancer: An Update

Giorgio Santoni<sup>1\*</sup>, Maria B. Morelli<sup>1,2</sup>, Consuelo Amantini<sup>2</sup> and Nicola Battelli<sup>3</sup>

<sup>1</sup>Immunopathology Laboratory, School of Pharmacy, University of Camerino, Camerino, Italy, <sup>2</sup>Immunopathology Laboratory, School of Biosciences, Biotechnology and Veterinary Medicine, University of Camerino, Camerino, Italy, <sup>3</sup>Oncology Unit, Macerata Hospital, Macerata, Italy

## OPEN ACCESS

### Edited by:

Rodolfo Montironi,  
Università Politecnica delle Marche,  
Italy

### Reviewed by:

Simona Di Francesco,  
Independent Researcher, Chieti, Italy  
Riccardo Autorino,  
Virginia Commonwealth University,  
United States

### \*Correspondence:

Giorgio Santoni  
giorgio.santoni@unicam.it

### Specialty section:

This article was submitted to  
Genitourinary Oncology,  
a section of the journal  
Frontiers in Oncology

**Received:** 29 June 2018

**Accepted:** 16 August 2018

**Published:** 07 September 2018

### Citation:

Santoni G, Morelli MB, Amantini C  
and Battelli N (2018) Urinary Markers  
in Bladder Cancer: An Update.  
Front. Oncol. 8:362.  
doi: 10.3389/fonc.2018.00362

Bladder cancer (BC) is one of the most common cancer worldwide. It is classified in muscle invasive (MIBC) and muscle non-invasive (NMIBC) BC. NMIBCs frequently recur and progress to MIBCs with a reduced survival rate and frequent distant metastasis. BC detection requires unpleasant and expensive cystoscopy and biopsy, which are often accompanied by several adverse effects. Thus, there is an urgent need to develop novel diagnostic methods for initial detection and surveillance in both MIBCs and NMIBCs. Multiple urine-based tests approved by FDA for BC detection and surveillance are commercially available. However, at present, sensitivity, specificity and diagnostic accuracy of these urine-based assays are still suboptimal and, in the attempt to improve them, novel molecular markers as well as multiple-assays must be translated in clinic. Now there is growing evidence toward the use of minimally invasive “liquid biopsy” to identify biomarkers in urologic malignancy. DNA- and RNA-based markers in body fluids such as blood and urine are promising potential markers in diagnostic, prognostic, predictive and monitoring urological malignancies. Thus, circulating cell-free DNA, DNA methylation and mutations, circulating tumor cells, miRNA, lncRNA and mRNAs, cell-free proteins and peptides, and exosomes have been assessed in urine specimens. However, proteomic and genomic data must be validated in well-designed multicenter clinical studies, before to be employed in clinic oncology.

**Keywords:** urinary biomarkers, bladder cancer, liquid biopsy, microRNA, exosomes

## INTRODUCTION

Bladder cancer (BC) represents the 9th and 4th most common cancer worldwide and in men in the USA, respectively (1, 2). Its main histological type is urothelial carcinoma (UC). About 70–80% of BC is diagnosed as non-muscle invasive BC (NMIBC) and 20–30% as muscle invasive (MIBC). Because 10–30% of patients with NMIBC progress to invasive disease (3–8), early diagnosis and early detection of recurrence are very important. BC diagnosis requires cystoscopy and biopsy, which are unpleasant and costly procedures (9). It is necessary to develop new diagnostic methods less invasive and expensive for BC diagnosis and surveillance. The Food and Drug Administration (FDA) has approved the use of multiple urine-based tests that are commercially available. However, none of these tests has been routinely used and incorporated in the American Urological Association or in the European Association of Urology clinical guidelines for BC treatment (10). In this mini-review we discuss the clinical implementation by the use of novel molecular approaches and liquid biopsy in BC.

At present, the gold standard methods for BC diagnosis are urine cytology and cystoscopy. Cytopathology of urine specimens is the widely used non-invasive test for detection and surveillance of BC (11–13). Cytology is very specific (about 86%), but it is low sensitive (48%) limiting its use in low-grade BC (14–16). Diagnostic accuracy of urinary cytology is subjective, depending on cytopathologist expertise (17). Thus, new molecular-based urinary tests for reducing or substituting, the endoscopy frequency in BC recurrence patients, are required (18, 19).

Advanced technology utilizes patients' urine as samples instead of primary BC tissues to identify novel predictive biomarkers. At present, the major problem is to translate the extensive proteomic and genomic data in clinical practice and to validate the expression of these biomarkers in well-designed multicenter clinical studies (20).

## PROTEOMIC AND PEPTIDOMIC ANALYSIS

Proteomic analyses have opened a new horizon for cancer biomarker discovery (21). At present, seven tests are available: FDA approved six on seven of these tests, and the last one is in agree with the Clinical Laboratory Improvement Act standards. NMP22, NMP22 BladderChek, and UroVysion have FDA approval for BC diagnosis and surveillance; immunocytology (uCyt+), BTA-TRAK, and BTA-STAT have been approved only for surveillance (22–26).

In order to improve sensitivity, specificity and diagnostic accuracy in BC diagnosis, novel protein markers, waiting to be approved, are used experimentally. BCLA-1 and BCLA-4 are nuclear matrix proteins specifically targeting BC tissues, with no interference with infection, smoking, catheterization or cystitis (27). In patients with hematuria, aurora A kinase (AURKA) discriminates between low-grade BC vs. normal patients (28). The Aura Tek FDP Test<sup>TM</sup> in urine can detect BC recurrence (29). The activated leukocyte cell adhesion molecule (ALCAM), a cell adhesion molecule (30), positively correlates with tumor stage and overall survival (OS), after adjusting for patients, clinical features and Bacillus Calmette-Guerin treatment (31). Nicotinamide N-methyltransferase is high in BC patients and correlate with histological grade (32). Apurinic/aprimidinic endonuclease 1/redox factor-1 (APE/Ref-1) levels are higher in BC, respect to non-BC, and correlate with tumor grade and stage; moreover it is high also in patients with recurrence history of BC (33). The cytokeratin-20 (CK20) urine RT-PCR assay shows 78–87% sensitivity and 56–80% specificity for urothelial BC detection, with improved diagnostic accuracy in tumor progression (34) but it has poor performance for low-grade tumors. Higher levels of CK8 and CK18 was detected in the urine by UBC Rapid test in high- vs low-grade BC (35).

As multiple markers for BC detection, increased urinary levels of apolipoprotein A1, A2, B, C2, C3, E (APOA1, APOA2, APOB, APOC2, APOC3, APOE) were found in BC relative to healthy controls (36, 37). A signature of 4 urinary fragments of uromodulin, collagen  $\alpha$ -1 (I), collagen  $\alpha$ -1 (III), and membrane-associated progesterone receptor component 1 seems

to discriminate MIBCs from NMIBCs (38). Other panel employs IL-8, MMP-9/10, ANG, APOE, SDC-1,  $\alpha$ 1AT, PAI-1, VEGFA, and CA9 to diagnose BC starting from urine samples (39). The advantage of these multi-urinary protein biomarkers was evident in high- and low-grade and high- and low-stage disease (39). The combination of urinary markers such as midkine (MDK) and synuclein G or MDK, ZAG2 and CEACAM1 (40), angiogenin and clusterin (41) evaluated by immunoassay and urine cytology increases the sensitivity and specificity in NMIBC diagnosis (40). Increased CK20 and Insulin Like Growth Factor II (IGFII) levels were detected in the urine sediments of NMIBC patients compared to controls (42). Increased levels of urinary HAI-1 and Epcam evaluated by ELISA, are prognostic biomarkers in high-risk NMIBC patients (43). Urinary survivin evaluated by chemiluminescence enzyme immunoassay correlates with tumor stage, lymph node and distant metastases and represents a potential marker for preliminary BC diagnosis (44). Snail overexpression represents an independent prognostic factor for tumor recurrence in NMIBC (45). Finally, specific glycoproteins were identified by glycan-affinity glycoproteomics nanoplateforms in the urine of low- and high-grade NMIBC; among these, increased urinary CD44 levels were evidenced in high-grade MIBC (46).

Urinary metabolomics signature could also be useful in early BC. By ultra-performance liquid chromatography time and mass spectrometry, imidazole-acetic acid was evidenced in BC (47). Moreover, acid trehalose, nicotinuric acid, AspAspGlyTrp peptide were upregulated; inosinic acid, ureidosuccinic acid and GlyCysAlaLys peptide were downregulated in BC, but not in normal cohort (48). A metabolite panel with indolylacryloylglycine, N2-galacturonil-L-lysine and aspartyl-glutamate permits to discriminate high- vs. low-grade BC (49). In addition, the alteration of phenylalanine, arginine, proline and tryptophan metabolisms was evidenced by UPLC-MS in NMIBC (50).

## CIRCULATING TUMOR AND CELL-FREE DNA

Tumors release DNA fragments into circulation, called circulating tumor DNA (ctDNA) containing tumor-specific mutations, variations of copy number and alterations in DNA methylation status. This ctDNA reflects the heterogeneity of tumor subclones. In BC patients, ctDNA is detectable in over 70% of urine samples (51) and it allows to discriminate between BC patients and control subjects (52). CtDNA measures about 180 and 200 base pairs. It is easily accessible, but it is rapidly cleared from circulation following systemic therapy (53). PCR-based approaches, and more recently, digital-PCR and genome sequencing, represent the methods of choice for cell-free DNA (cfDNA) analysis.

## DNA Methylation

The methylation status of tumor-related genes represents a very important epigenetic alteration affecting cancer initiation and progression. Hyper- and hypo-methylated regions are

identified in BC and in premalignant lesions. Alterations in DNA methylation status are chemically stable, develop early during tumorigenesis and can be assessed in circulating cfDNA fragments and in cells shed into the urine (54). A significant prevalence of methylated genes, for example APC and cyclin D2, was found in the urine from malignant vs. benign cases (55). Hyper-methylation in GSTP1 and RAR $\beta$ 2 and APC genes has been identified in the urine from BC patients (56). The evaluation of Twist Family BHLH Transcription Factor 1 (TWIST1) and NID2 genes methylation status in urine permits to differentiate primary BC patients from controls with 90% sensitivity and 93% specificity (57). In addition, the evaluation of the methylation status of NID2 and TWIST1 or CFTR, SALL3 and TWIST1 genes in urinary cells in combination with cytology, has been found to increase sensitivity and high negative predictive value in BC patients (58, 59). The analysis of 1,370 loci specific DNA methylation patterns seem to permit to distinguish NMIBC from MIBC (60). Sun and coworkers demonstrated higher recurrence predictivity than urine cytology and cystoscopy (80 vs. 35 vs. 15%) by using SOX-1, IRAK3, and Li-MET genes methylation status from urine sediments of BC patients (54). POU4F2 and PCDH17 methylation levels in urine distinguish BC from normal controls with 90% sensitivity and 94% specificity (61). Promoter hyper-methylation of HS3ST2, SEPTIN9 and SLIT2 genes combined with FGFR3 mutation showed 97.6% sensitivity and 84.8% specificity for diagnosis, surveillance and risk stratification in low- or high-risk NMIBC patients (62). Finally, the methylation status of p14ARF, p16INK4A, RASSF1A, DAPK, and APC tumor suppressor genes has been found to correlate with BC grade and stage (63).

Altogether, although promising results were obtained, accuracy of urinary methylated DNA is variable and results still await validation studies and complementary markers for clinical implementation (64, 65). In this regard, the recent introduction of the methylation-sensitive High Resolution Melting and Methylated CpG Island Recovery methods could further increase the sensitivity for the detection of methylome in BC urine (Table 1) (72, 73).

## cfDNA, Mutation and Microsatellite Alterations

Since tumor-derived DNA can be released into circulation and mutations in cfDNA can be detected in various biological fluids, their use as non-invasive cancer biomarkers has been proposed. Urinary TERT promoter mutations, that occur early in urothelial neoplasia, FGFR3 mutation and telomere length correlate with high-risk BC recurrence (66, 67). TERT, evaluated by telomeric repeat amplification protocol, in combination with FGF3 and OTX1 shows high sensitivity in NMIBCs as well as in pT1 tumors and in high-grade BC (68). In addition, increased FGFR3 and PIK3CA mutated DNA levels in urine has been found to be indicative of progression and metastasis in NMIBC (69). Microsatellite analysis in circulating DNA of BC patients targets highly polymorphic, short tandem repeats. Loss of heterozygosity (LOH) analysis is more sensitive than urine cytology (97 vs. 79%), particularly for low-grade BC diagnosis. It also significantly

improves the detection of low-grade and low-stage BC, with 95% sensitivity for G1-G2 grades and 100% for pTis and pTa tumors (Table 1) (74).

## Histone Tail Modifications

The levels of histone methylation are lower in advanced tumors respect to controls and correlated to poor survival. Thus, increased levels of HAK20me3 were evidenced in a MIBC subset (70); furthermore high H3K27me3 levels correlate with worse survival after cystectomy in pT1-3 and pN- BC patients (71). H2AFX1 gene methylation was detected in paraffin-embedded BC and its expression correlated with increased recurrence rates (Table 1) (75).

## URINARY TUMOR RNA

Several RNA classes, messenger RNAs (mRNAs), microRNAs (miRs) and long non-coding RNAs (lncRNAs), have been recognized as potential non-invasive cancer biomarkers (76). Altered levels of circulating RNAs in cancer, which returned to normal following surgery have been reported (77), suggesting release of RNA molecules from tumors.

### miRNAs (miRNAs)

miRNAs are short (21–23 nucleotides length) non-coding RNAs regulating gene expression by pairing to the 3' untranslated region (UTR) of their target mRNA. Several miRNAs have been found to play an important role in tumorigenesis, progression and metastasis of cancer cells (78, 79). Urine seems to be a good source for miRNA detection for its content of cell-free nucleic acid in supernatant or sediments (80). However, the diagnostic significance in the detection of miRs in urine as respect to blood of BC patients is controversial (81). MiR-126 urinary levels were found to be enhanced in BC compared to healthy controls (82). Urine miR-146a-5p is significantly increased in high-grade BC (77). Low miR-200c expression correlates with tumor progression in NMIBCs (83). Chen et al. detected 74 miRNAs, of which 33 upregulated and 41 downregulated in BC compared to healthy patients (84). The most interesting are let-7miR, miR-1268, miR-196a, miR-1, miR-100, miR-101, and miR-143 (84). MiR-200 was identified as epithelial–mesenchymal transition regulator in BC cells by targeting Zinc Finger E-Box Binding Homeobox 1 (ZEB1), ZEB2 and Epidermal growth factor receptor (EGFR) (85). Some miRNAs have been associated with hemolysis including miR-451a, miR-16, miR-486-5p, and miR-92a (86). Eissa et al. by screening BC patients with negative cystoscopy, identified miR-96 and miR-210 in BC (87). Sapre et al., by using a panel of 12 miRNA, reduced the cystoscopy rates by 30% by increasing sensitivity and specificity (88). MiR-125b, miR-30b, miR-204, miR-99a, and miR-532-3p were downregulated in BC patient's urine supernatant, with miR-125 levels (95.7% specificity, 59.3% sensitivity) (89). MiR-9, miR-182 and miR-200b correlated with MIBC aggressiveness, recurrence-free and OS (90). MiR-145 distinguishes NMIBCs from non-BCs (91). MiR-144-5p inhibited BC proliferation, affecting CCNE1, CCNE2, CDC25A, PKMYT1 target genes (92). Cell-free urinary miR-99a and miRNA-125b were found to be downregulated



**TABLE 1 |** Urinary tumor-derived DNAs as biomarkers in BCs.

Urinary tumor-derived DNA	Gene	Application	References
CfDNA	TERT and FGFR3	Recurrence	(66, 67)
	TERT, FGFR3/OTX1	BC diagnosis	(68)
	FGFR3 and PIK3CA	Progression/metastasis	(69)
Histone modifications	HAK20me3	Poor survival	(70)
	H3K27me3	Poor survival	(71)
DNA methylation status	GSTP1 and RARb2 and APC	BC diagnosis	(56)
	Twist1 and NID2	BC diagnosis	(57)
	SOX-1, IRAK3, and Li-MET	Recurrence	(54)
	POU4F2 and PCDH17	BC diagnosis	(61)
	HS3ST2, SEPTIN9, SLIT2/FGFR3	surveillance, low vs. high risk	(62)
	NID2 and Twist1	BC diagnosis	(58)
	CFTR, SALL3/TWIST1	BC diagnosis	(59)
	p14ARF, p16INK4A, RASSF1A	BC grade and stage	(63)
	DAPK and APC		

BC, Bladder cancer; CfDNA, circulating-free DNA.

in the urine supernatants of BC patients (sensitivity 86.7%; specificity 81.1%) (93). Urinary levels of miR-618 and miR-125b-5p in MIBC patients were increased in comparison to controls (94). Multiple miRNA assay shows higher diagnostic performance than single RNA assay (95). By whole genome analysis increased miR-31-5p, miR-191-5p and miR-93-5p levels were identified in the urine of BC patients as compared to controls (96).

Recently, a miRNA profile, identified in urine by next-generation sequencing (NGS) analysis, has been capable to stratify different BC subtypes (97). In NMIBC G1/G2 patients a miR-205-5p upregulation compared to controls was observed. Among NMIBC G3, upregulation of miR-21-5p, miR-106b-3p, miR-486-5p, miR-151a-3p, miR-200c-3p, miR-185-5p, miR-185-5p and miR-224-5p and downregulation of miR-30c-2-5p and miR-10b-5p were observed. In MIBCs, miR-205-5p, miR-451a, miR-25-3p and miR-7-1-5p were upregulated, while miR-30a-5p was downregulated compared to controls (97). The application of NGS have increased the diagnostic accuracy. However results obtained in NGS were only partially overlapping with that obtained by qRT-PCR (98) (Table 2).

## Long Non Coding RNAs (lncRNAs)

Long non coding RNAs (lncRNAs) regulate gene expression or epigenetic levels. Several findings show lncRNA changes in cancers suggesting a role in the promotion of tumor development and progression (105, 106). The use of lncRNAs as non-invasive BC marker has recently interested (107). Circulating urothelial carcinoma antigen 1 (UCA1) levels in urinary sediments represents a potential diagnostic marker for UC, with 81% sensitivity and 92% specificity (108). Du et al. describe high uc004cox.4 lncRNA level association with poor recurrence-free survival in NMIBCs (102). The retrotransposome, long interspaced element-1 (LINE-1) has been found to be hypo-methylated and its expression was associated with long recurrence-free and tumor specific survival in BC (109) (Table 2).

## Messenger RNAs (mRNAs)

Circulating messenger RNAs (mRNAs) were detected in cancer patients, although the majority of circulating mRNAs are degraded by RNases (110). Given their role in intracellular protein translation, their presence reflects the status of intracellular processes and they are potential cancer biomarkers. Urine Ubiquitin Conjugating Enzyme E2C (UBE2C) mRNA levels were higher in BC patients, compared to normal and hematuria specimens (111). The expression of isoleucine glutamine motif-containing GTPase-activating proteins (IQGAP3) mRNA in urine was found higher in BC than in controls (112). Further analysis of IQGAP3, with respect to tumor invasiveness and grade also yielded a high diagnostic accuracy, suggesting that IQGAP3 can be used to discriminate BC from non-BC patients with hematuria (112).

In regard to mRNAs extracted by exfoliated urinary cells, the Xpert BC Monitor measuring ABL1, corticotropin releasing hormone (CRH), IGF2, uroplakin 1B (UPK1B), annexin A10 (ANXA10) mRNAs by RT-PCR, increased the overall sensitivity over urinary cytology in low-grade and pTa disease (113).

In addition, the presence of carbonic anhydrase 9 (CAIX) splice variant mRNA in the urine, increased the diagnostic performance for BC (90% sensitivity and 72% specificity) (114). The downregulation of N-Myc downstream-regulated gene 2 (NDRG2) mRNA levels in the urine of BC patients correlated with tumor grade and stage (99) (Table 2).

## Transfer RNA Fragments (tRFs)

Elevated levels of transfer RNA fragments (tRF) are found in cancer (115). tRF are 14-32 base long single-stranded RNA derived from mature or precursor tRNA. They are grouped into 3 classes (tRF-1, -3, and -5) and, depending of their cleavage site within a mature RNA, they are further divided in 5 subclasses. The first identified tRF in NMIBCs was miR720/3007a (101) (Table 2).

**TABLE 2 |** Urinary tumor-derived RNAs as biomarkers in BCs.

Urinary tumor-derived RNAs	RNA/Protein	Application	References
mRNA	CK20, IGF-II	BC diagnosis	(42)
	ABL1, CRH, IGF2, UPK1B and ANXA10	BC diagnosis	(78)
	NDRG2	Tumor grade and stage	(99)
miRNA	miR-146a	BC diagnosis	(77)
	miR-126	BC diagnosis	(82)
	miR-200c	Tumor progression	(83)
	let-7, miR-1268, -196a, -1, -101, -143	BC diagnosis	(84)
	miR-451a, -16, -486, -92a	Hemolysis	(86)
	miR-96, -210	BC diagnosis	(87)
	miR-125b, -30b, -204a, -99a, -532	BC diagnosis	(89)
	miR-9, -182, -200b	aggressiveness, recurrence	(90)
	miR-145	BC diagnosis	(91)
	miR-99a, -125b	BC diagnosis	(93)
	miR-618, -1255b	BC diagnosis	(94)
	miR-21, -106b, -486, -151a, -200c	NMIBC diagnosis	(97)
	-185, -224, 30c-2, -10b		
	miR-205, -451a, -25, -7-1, -30a	MIBC diagnosis	(97)
	miR-31, -191, -93	BC diagnosis	(96)
miRNA/EVs and Exosomes	miR-375, -146a	BC diagnosis	(100)
miRNA/tRF	miR720/3007a	BC diagnosis	(101)
lncRNA	uc004cox.4	Recurrence	(102)
lncRNA/exosomes	HOX-AS, ANRIL, and linc-RoR	BC diagnosis	(103, 104)

BC, Bladder cancer; NMIBC, muscle non-invasive BC; MIBC, muscle invasive; mRNA, messenger RNA; miR, microRNA; EVs, extracellular vesicles; tRF, transfer RNA fragments; lncRNA, Long non-coding RNA.

## EXTRACELLULAR VESICLES (EVs) AND EXOSOMES

Extracellular Vesicles (EVs) enrichment was found in BC patient urine. EVs, analyzed by MS based proteomics, demonstrated specific protein and miRNAs pattern in BC patients (116). By using a microarray platform and RT-PCR analysis, miR-375, and miR146a have been found to specifically identify high-grade and low-grade BC, respectively (100). The application of nanowires anchored into a microfluidic substrate will enable the efficiency of EV collection, thus permitting to identify EV harboring miRNAs (117).

Exosomes are membrane vesicles secreted in nearly all body fluids at elevated levels in cancer patients relative to healthy subjects (118, 119). They realize intercellular communication through transferring distinct biologically active molecules (RNAs, DNA, and proteins), thus influencing the therapeutic responses. The HOX transcript antisense RNA (HOTAIR) together with other lncRNA, such as HOX-AS-2, ANRIL, and linc-RoR, were augmented in urinary exosomes from high-grade MIBC patients (103). Loss of HOTAIR expression in BC cells alters the expression of SNA1, TWIST1, ZEB1, ZO1, MMP-1, Laminin Subunit Beta 3 (LAMB3), and Laminin Subunit Gamma 2 (LAMC2) epithelial-to mesenchymal transition genes. Moreover, the tumor-associated calcium-signal transducer 2 (TACSTD2) was found in BC exosomes by proteomic analysis (104). EVs can also promote BC progression by delivering the

protein EGF-like repeat and discoidin I-like domain-containing protein-3 (120).

Exosomes in urine also contain miRNAs, in particular miR-1224-3p, miR-135b, and miR15b; in particular, miR-126/miR-152 ratio correlated with positive BC diagnosis (121) (Table 2).

Although EVs and exosomes represent an interesting source of cancer biomarkers, the lack of accurate isolation and detection methods affects their utilization in practice. In the next future, the development of sensitive capture platforms for exosomes, likely increases their introduction into clinic.

## URINARY MICROBIOME

Dysbiosis of urinary microbiome has been suggested to be involved in bladder tumorigenesis. Recently, Wu et al. by analyzing DNA extracted by urine pellets, observed specific enrichment of *Acinetobacter*, *Anaerococcus*, and *Sphingobacterium* in BC cohort as respect to controls (122). Moreover, the increase of *Herbaspirillum*, *Porphyrobacter*, and *Bacteroides* in high-risk BC patients suggested that these genera may represent new potential biomarkers (122).

## CONCLUSIONS AND PERSPECTIVES

We provide the state of art into the use of urinary biomarkers as tool to aid diagnosis of BC. Urine cytology, utilized for

decades, shows poor sensitivity, particularly for low-grade tumors. The addition of immunoassay and FISH analysis has provided an additional diagnostic armamentarium to determine which patients may need further evaluation. At present, there are growing evidence toward the use of “Liquid Biopsy” to identify urinary biomarkers such as circulating cell-free DNA, DNA methylation, miRNA, cell-free proteins/peptides and exosomes, useful for discriminating NMIBC from MIBC (123). The potential introduction of “smart toilets” working with a more advanced “nano-sensor” able to detect RNA and proteins in urine is close to reality, more that we think (124). However, now in clinical reality, there is an urgent need to validate the recently discovered extensive proteomic and

genomic, epigenomic, transcriptomic and metabolomic data as urinary biomarkers in well-designed multicenter clinical studies (125, 126).

## AUTHOR CONTRIBUTIONS

GS, MM conception and design. GS drafting the manuscript. CA and NB critical revision of the manuscript.

## FUNDING

The authors are grateful to AIRC IG 2014 and Post-doctoral Fellowships 2018 Fondazione Veronesi.

## REFERENCES

- Siegel RL, Miller KD, Jemal A. Cancer statistics. *CA Cancer J Clin.* (2015) 65:5–29. doi: 10.3322/caac.21254
- van Rhijn BW, Burger M, Lotan Y, Solsona E, Stief CG, Sylvester RJ, et al. Recurrence and progression of disease in non-muscle-invasive bladder cancer: from epidemiology to treatment strategy. *Eur Urol.* (2009) 56:430–42. doi: 10.1016/j.eururo.2009.06.028
- Prout GR, Barton BA, Griffin PP, Friedell GH. Treated history of noninvasive grade 1 transitional cell carcinoma. *J Urol.* (1992) 148:1413–9. doi: 10.1016/S0022-5347(17)36924-0
- Herr HW. Tumor progression and survival of patients with high grade, noninvasive papillary (TaG3) bladder tumors: 15-year outcome. *J Urol.* (2000) 163:60–2. doi: 10.1016/s0022-5347(05)67972-4
- Sylvester RJ, van der Meijden AP, Oosterlinck W, Witjes JA, Bouffuoux C, Denis L, et al. Predicting recurrence and progression in individual patients with stage Ta T1 bladder cancer using EORTC risk tables: a combined analysis of 2596 patients from seven EORTC trials. *Eur Urol.* (2006) 49:466–7. doi: 10.1016/j.eururo.2005.12.031
- Johnson MI, Merrilees D, Robson WA, Lennon T, Masters J, Orr KE, et al. Oral ciprofloxacin or trimethoprim reduces bacteriuria after flexible cystoscopy. *BJU Int.* (2007) 100:826–9. doi: 10.1111/j.1464-410X.2007.07093.x
- Soloway MS. Bladder cancer: lack of progress in bladder cancer—what are the obstacles? *Nat Rev Urol.* (2013) 10:5–6. doi: 10.1038/nrurol.2012.219
- Türkölmez K, Tokgöz H, Reşorlu B, Köse K, Bedük Y. Muscle-invasive bladder cancer: predictive factors and prognostic difference between primary and progressive tumors. *Urology* (2007) 70:477–81. doi: 10.1016/j.urol.2007.05.008
- Burke DM, Shackley DC, O'Reilly PH. The community-based morbidity of flexible cystoscopy. *BJU Int.* (2002) 89:347–9. doi: 10.1046/j.1464-4096.2001.01899.x
- Zuiverloon TCM, de Jong FC, Theodorescu D. Clinical decision making in surveillance of non-muscle-invasive bladder cancer: the evolving roles of urinary cytology and molecular markers. *Oncology* (Williston Park) (2017) 31:855–62.
- Tëtü B. Diagnosis of urothelial carcinoma from urine. *Mod Pathol.* (2009) 22 (Suppl 2):S53–9. doi: 10.1038/modpathol.2008.193
- Sapre N, Hong MK, Huang JG, Pedersen J, Ryan A, Anderson P, et al. Bladder cancer biorepositories in the “-omics” era: integrating quality tissue specimens with comprehensive clinical annotation. *Biopreserv Biobank* (2013) 11:166–72. doi: 10.1089/bio.2012.0062
- Yafi FA, Brimo F, Steinberg J, Aprikian AG, Tanguay S, Kassouf W. Prospective analysis of sensitivity and specificity of urinary cytology and other urinary biomarkers for bladder cancer. *Urol Oncol.* (2015) 33:66.e25–31. doi: 10.1016/j.urolonc.2014.06.008
- Lotan Y, Roehrborn CG. Sensitivity and specificity of commonly available bladder tumor markers versus cytology: results of a comprehensive literature review and meta-analyses. *Urology* (2003) 61:109–18. doi: 10.1016/S0090-4295(02)02136-2
- Simon MA, Lokeshwar VB, Soloway MS. Current bladder cancer tests: unnecessary or beneficial? *Crit Rev Oncol Hematol.* (2003) 47:91–107. doi: 10.1016/S1040-8428(03)00074-X
- van der Aa MN, Steyerberg EW, Sen EF, Zwarthoff EC, Kirkels WJ, van der Kwast TH, et al. Patients' perceived burden of cystoscopic and urinary surveillance of bladder cancer: a randomized comparison. *BJU Int.* (2008) 101:1106–10. doi: 10.1111/j.1464-410X.2007.07224.x
- Shariat SF, Karam JA, Lotan Y, Karakiewicz PI. Critical evaluation of urinary markers for bladder cancer detection and monitoring. *Rev Urol.* (2008) 10:120–35.
- Lokeshwar VB, Habuchi T, Grossman HB, Murphy WM, Hautmann SH, Hemstreet GP, et al. Bladder tumor markers beyond cytology: international Consensus Panel on bladder tumor markers. *Urology* (2005) 66 (6 Suppl 1):35–63. doi: 10.1016/j.urol.2005.08.064
- Owens CL, VandenBussche CJ, Burroughs FH, Rosenthal DL. A review of reporting systems and terminology for urine cytology. *Cancer Cytopathol.* (2013) 121:9–14. doi: 10.1002/cncy.21253
- Ralla B, Stephan C, Meller S, Dietrich D, Kristiansen G, Jung K. Nucleic acid-based biomarkers in body fluids of patients with urologic malignancies. *Crit Rev Clin Lab Sci.* (2014) 51:200–31. doi: 10.3109/10408363.2014.914888
- Di Meo A, Pasic MD, Yousef GM. Proteomics and peptidomics: moving toward precision medicine in urological malignancies. *Oncotarget* (2016) 7:52460–74. doi: 10.18632/oncotarget.8931
- Kim WT, Cho NH, Ham WS, Lee JS, Ju HJ, Kwon YU, et al. Comparison of the efficacy of urine cytology, Nuclear Matrix Protein 22 (NMP22), and Fluorescence *in Situ* Hybridization (FISH) for the diagnosis of bladder cancer. *Korean J Urol.* (2009) 50:6–11. doi: 10.4111/kju.2009.50.1.6
- Hajdinjak T. UroVysion FISH test for detecting urothelial cancers: meta-analysis of diagnostic accuracy and comparison with urinary cytology testing. *Urol Oncol.* (2008) 26:646–51. doi: 10.1016/j.urolonc.2007.06.002
- Horstmann M, Patschan O, Hennenlotter J, Senger E, Feil G, Stenzl A. Combinations of urine-based tumour markers in bladder cancer surveillance. *Scand J Urol Nephrol.* (2009) 43:461–6. doi: 10.3109/00365590903296837
- Todenhöfer T, Hennenlotter J, Esser M, Mohrhardt S, Tews V, Aufderklamm S, et al. Combined application of cytology and molecular urine markers to improve the detection of urothelial carcinoma. *Cancer Cytopathol.* (2013) 121:252–60. doi: 10.1002/cncy.21247
- He H, Han C, Hao L, Zang G. ImmunoCyt test compared to cytology in the diagnosis of bladder cancer: a meta-analysis. *Oncol Lett.* (2016) 12:83–8. doi: 10.3892/ol.2016.4556
- Deininger S, Hennenlotter J, Rausch S, Docktor K, Neumann E, da Costa IA, et al. No influence of smoking status on the performance of urine markers for the detection of bladder cancer. *J Cancer Res Clin Oncol.* (2018) 144:1367–73. doi: 10.1007/s00432-018-2639-z

28. de Martino M, Shariat SF, Hofbauer SL, Lucca I, Taus C, Wiener HG, et al. Aurora A Kinase as a diagnostic urinary marker for urothelial bladder cancer. *World J Urol.* (2015) 33:105–10. doi: 10.1007/s00345-014-1267-8
29. Siemens DR, Morales A, Johnston B, Emerson L. A comparative analysis of rapid urine tests for the diagnosis of upper urinary tract malignancy. *Can J Urol.* (2003) 10:1754–8.
30. Rosso O, Piazza T, Bongarzone I, Rossello A, Mezzananza D, Canevari S, et al. The ALCAM shedding by the metalloprotease ADAM17/TACE is involved in motility of ovarian carcinoma cells. *Mol Cancer Res.* (2007) 5:1246–53. doi: 10.1158/1541-7786.MCR-07-0060
31. Egloff SA, Du L, Loomans HA, Starchenko A, Su PF, Ketova T, et al. Shed urinary ALCAM is an independent prognostic biomarker of three-year overall survival after cystectomy in patients with bladder cancer. *Oncotarget* (2017) 8:722–41. doi: 10.18632/oncotarget.13546
32. Pozzi V, Di Ruscio G, Sartini D, Campagna R, Seta R, Fulvi P, et al. Clinical performance and utility of a NNMT-based urine test for bladder cancer. *Int J Biol Mark.* (2018) 33:94–101. doi: 10.5301/ijbm.5000311
33. Choi S, Shin JH, Lee YR, Joo HK, Song KH, Na YG, et al. Urinary APE1/Ref-1: A Potential bladder cancer biomarker. *Dis Mark.* (2016) 2016:7276502. doi: 10.1155/2016/7276502
34. Mi Y, Zhao Y, Shi F, Zhang M, Wang C, Liu X. Diagnostic accuracy of urine cytokeratin 20 for bladder cancer: a meta-analysis. *Asia Pac J Clin Oncol.* (2018) doi: 10.1111/ajco.13024. [Epub ahead of print].
35. Ecke TH, Weiß S, Stephan C, Hallmann S, Barski D, Otto T, et al. UBC® Rapid Test for detection of carcinoma *in situ* for bladder cancer. *Tumor Biol.* (2017) 39:1010428317701624. doi: 10.1177/1010428317701624
36. Chen YT, Chen CL, Chen HW, Chung T, Wu CC, Chen CD, et al. Discovery of novel bladder cancer biomarkers by comparative urine proteomics using iTRAQ technology. *J Proteome Res.* (2010) 5:5803–15. doi: 10.1021/pr100576x
37. Chen YT, Chen HW, Domanski D, Smith DS, Liang KH, Wu CC, et al. Multiplexed quantification of 63 proteins in human urine by multiple reaction monitoring-based mass spectrometry for discovery of potential bladder cancer biomarkers. *J Proteomics* (2012) 75:3529–45. doi: 10.1016/j.jpro.2011.12.031
38. Schiffer E, Vlahou A, Petrolekas A, Stravodimos K, Tauber R, Geschwend JE, et al. Prediction of muscle-invasive bladder cancer using urinary proteomics. *Clin Cancer Res.* (2009) 15:4935–43. doi: 10.1158/1078-0432.CCR-09-0226
39. Masuda N, Ogawa O, Park M, Liu AY, Goodison S, Dai Y, et al. Meta-analysis of a 10-plex urine-based biomarker assay for the detection of bladder cancer. *Oncotarget* (2018) 9:7101–11. doi: 10.18632/oncotarget.23872
40. Soukup V, Kalousová M, Capoun O, Sobotka R, Breyt Z, Pešl M, et al. Panel of urinary diagnostic markers for non-invasive detection of primary and recurrent urothelial urinary bladder carcinoma. *Urol Int.* (2015) 95:56–64. doi: 10.1159/000368166
41. Shabayek MI, Sayed OM, Attaia HA, Awida HA, Abozeed H. Diagnostic evaluation of urinary angiogenin (ANG) and clusterin (CLU) as biomarker for bladder cancer. *Pathol Oncol Res.* (2014) 20:859–66. doi: 10.1007/s12253-014-9765-y
42. Salomo K, Huebner D, Boehme MU, Herr A, Brabetz W, Heberling U, et al. Urinary transcript quantitation of CK20 and IGF2 for the non-invasive bladder cancer detection. *J Cancer Res Clin Oncol.* (2017) 143:1757–69. doi: 10.1007/s00432-017-2433-3
43. Snell KIE, Ward DG, Gordon NS, Goldsmith JC, Sutton AJ, Patel P, et al. Exploring the roles of urinary HAI-1, EpCAM & EGFR in bladder cancer prognosis & risk stratification. *Oncotarget* (2018) 9:25244–53. doi: 10.18632/oncotarget.25397
44. Yang Y, Xu J, Zhang Q. Detection of urinary surviving using a magnetic particles-based chemiluminescence immunoassay for the preliminary diagnosis of bladder cancer and renal cell carcinoma combined with LAPTM4B. *Oncol Lett.* (2018) 15:7923–33. doi: 10.3892/ol.2018.8317
45. Santi R, Cai T, Nobili S, Galli IC, Amorosi A, Comperat E, et al. Snail immunohistochemical overexpression correlates to recurrence risk in non-muscle invasive bladder cancer: results from a longitudinal cohort study. *Virchows Arch.* (2018) 472:605–13. doi: 10.1007/s00428-018-2310-8
46. Azevedo R, Soares J, Gaiteiro C, Peixoto A, Lima L, Ferreira D, et al. Glycan affinity magnetic nanoplateforms for urinary glycomarkers discovery in bladder cancer. *Talanta* (2018) 184:347–55. doi: 10.1016/j.talanta.2018.03.028
47. Shao CH, Chen CL, Lin JY, Chen CJ, Fu SH, Chen YT, et al. Metabolite marker discovery for the detection of bladder cancer by comparative metabolomics. *Oncotarget* (2017) 8:38802–10. doi: 10.18632/oncotarget.16393
48. Shen C, Sun Z, Chen D, Su X, Jiang J, Li G, et al. Developing urinary metabolomic signatures as early bladder cancer diagnostic markers. *OMICS* (2015) 19:1–11. doi: 10.1089/omi.2014.0116
49. Liu X, Cheng X, Liu X, He L, Zhang W, Wang Y, et al. Investigation of urinary metabolic variations and the application in bladder cancer biomarker discovery. *Int J Cancer* (2018) 143:408–18. doi: 10.1002/ijc.31323
50. Loras A, Trassiera M, Sanjuan-Herráez D, Martínez-Bisbal MC, Castell JV, Quintás G, et al. Bladder cancer recurrence surveillance by urine metabolomics analysis. *Sci Rep.* (2018) 8:9172. doi: 10.1038/s41598-018-27538-3
51. Goessl C, Müller M, Straub B, Miller K. DNA alterations in body fluids as molecular tumor markers for urological malignancies. *Eur Urol.* (2002) 41:668–76. doi: 10.1016/S0302-2838(02)00126-4
52. Brisuda A, Pazourkova E, Soukup V, Horinek A, Hrbáček J, Capoun O, et al. Urinary cell-free DNA quantification as non-invasive biomarker in patients with bladder cancer. *Urol Int.* (2016) 96:25–31. doi: 10.1159/000438828
53. Qin Z, Ljubimov VA, Zhou C, Tong Y, Liang J. Cell-free circulating tumor DNA in cancer. *Chin J Cancer* (2016) 35:36. doi: 10.1186/s40880-016-0092-4
54. Su SF, de Castro Abreu AL, Chihara Y, Tsai Y, Andreu-Vieyra C, Daneshmand S, et al. A panel of three markers hyper- and hypomethylated in urine sediments accurately predicts bladder cancer recurrence. *Clin Cancer Res.* (2014) 20:1978–89. doi: 10.1158/1078-0432.CCR-13-2637
55. Pu RT, Laitala LE, Clark DP. Methylation profiling of urothelial carcinoma in bladder biopsy and urine. *Acta Cytol.* (2006) 50:499–506. doi: 10.1159/000326003
56. Hauser S, Kogej M, Fechner G, VON Pezold J, Vorreuther R, Lümmen G, et al. Serum DNA hypermethylation in patients with bladder cancer: results of a prospective multicenter study. *Anticancer Res.* (2013) 33:779–84.
57. Renard I, Joniau S, van Cleynebreugel B, Collette C, Naômé C, Vlassenbroeck I, et al. Identification and validation of the methylated TWIST1 and NID2 genes through real-time methylation-specific polymerase chain reaction assays for the noninvasive detection of primary bladder cancer in urine samples. *Eur Urol.* (2010) 58:96–104. doi: 10.1016/j.eururo.2009.07.041
58. Fantony JJ, Longo TA, Gopalakrishna A, Owusu R, Lance RS, Foo WC, et al. Urinary NID2 and TWIST1 methylation to augment conventional urine cytology for the detection of bladder cancer. *Cancer Biomark.* (2017) 18:381–7. doi: 10.3233/CBM-160261
59. van der Heijden AG, Mengual L, Ingelmo-Torres M, Lozano JJ, van Rijt-van de Westerloo CCM, Baixauli M, et al. Urine cell-based DNA methylation classifier for monitoring bladder cancer. *Clin Epigenetics.* (2018) 10:71. doi: 10.1186/s13148-018-0496-x
60. Wolff EM, Chihara Y, Pan F, Weisenberger DJ, Siegmund KD, Sugano K, et al. Unique DNA methylation patterns distinguish noninvasive and invasive urothelial cancers and establish an epigenetic field defect in premalignant tissue. *Cancer Res.* (2010) 70:8169–78. doi: 10.1158/0008-5472.CAN-10-1335
61. Wang Y, Yu Y, Ye R, Zhang D, Li Q, An D, et al. An epigenetic biomarker combination of PCDH17 and POU4F2 detects bladder cancer accurately by methylation analyses of urine sediment DNA in Han Chinese. *Oncotarget* (2016) 7:2754–64. doi: 10.18632/oncotarget.6666
62. Roperch JP, Grandchamp B, Desgrandchamps F, Mongiat-Artus P, Ravary V, Ouzaid I, et al. Promoter hypermethylation of HS3ST2, SEPTIN9 and SLIT2 combined with FGFR3 mutations as a sensitive/specific urinary assay for diagnosis and surveillance in patients with low or high-risk non-muscle-invasive bladder cancer. *BMC Cancer* (2016) 16:704. doi: 10.1186/s12885-016-2748-5
63. Pietrusinski M, Kępczyński Ł, Jędrzejczyk A, Borkowska E, Traczyk-Borszyńska M, et al. Detection of bladder cancer in urine sediments



- by a hypermethylation panel of selected tumor suppressor genes. *Cancer Biomark.* 2017;18:47–59. doi: 10.3233/CBM-160673
64. Peng M, Chen C, Hulbert A, Brock MV, Yu F. Non-blood circulating tumor DNA detection in cancer. *Oncotarget* (2017) 8:69162–73. doi: 10.18632/oncotarget.19942
  65. Bosschier J, Lutz C, Segerink LI, Vis AN, Zwarthoff EC, A van Moorselaar RJ, et al. The diagnostic accuracy of methylation markers in urine for the detection of bladder cancer: a systematic review. *Epigenomics* (2018) 10:673–87. doi: 10.2217/epi-2017-0156
  66. Kinde I, Munari E, Faraj SF, Hruban RH, Schoenberg M, Bivalacqua T, et al. TERT promoter mutations occur early in urothelial neoplasia and are biomarkers of early disease and disease recurrence in urine. *Cancer Res.* (2013) 73:7162–7. doi: 10.1158/0008-5472.CAN-13-2498
  67. Hosen I, Rachakonda PS, Heidenreich B, de Verdier PJ, Ryk C, Steineck G, et al. Mutations in TERT promoter and FGFR3 and telomere length in bladder cancer. *Int J Cancer* (2015) 137:1621–9. doi: 10.1002/ijc.29526
  68. Beukers W, van der Keur KA, Kandimalla R, Vergouwe Y, Steyerberg EW, Boormans JL, et al. FGFR3, TERT and OTX1 as a urinary biomarker combination for surveillance of patients with bladder cancer in a large prospective multicenter study. *J Urol.* (2017) 197:1410–8. doi: 10.1016/j.juro.2016.12.096
  69. Christensen E, Birkenkamp-Demtröder K, Nordentoft I, Høyer S, van der Keur K, van Kessel K, et al. Liquid biopsy analysis of FGFR3 and PIK3CA hotspot mutations for disease surveillance in bladder cancer. *Eur Urol.* (2017) 71:961–9. doi: 10.1016/j.eururo.2016.12.016
  70. Schneider AC, Heukamp LC, Rogenhofer S, Fechner G, Bastian PJ, von Ruecker A, et al. Global histone H4K20 trimethylation predicts cancer-specific survival in patients with muscle-invasive bladder cancer. *BJU Int.* (2011) 108:E290–6. doi: 10.1111/j.1464-410X.2011.10203.x
  71. Liu J, Li Y, Liao Y, Mai S, Zhang Z, Liu Z, et al. High expression of H3K27me3 is an independent predictor of worse outcome in patients with urothelial carcinoma of bladder treated with radical cystectomy. *Biomed Res Int.* (2013) 2013:390482. doi: 10.1155/2013/390482
  72. Hussmann D, Hansen LL. Methylation-Sensitive High Resolution Melting (MS-HRM). *Methods Mol Biol.* (2018) 1708:551–71. doi: 10.1007/978-1-4939-7481-8\_28
  73. Tommasi S, Besaratinia A. A versatile assay for detection of aberrant DNA methylation in bladder cancer. *Methods Mol Biol.* (2018) 1655:29–41. doi: 10.1007/978-1-4939-7234-0\_3
  74. Seripa D, Parrella P, Gallucci M, Gravina C, Papa S, Fortunato P, et al. Sensitive detection of transitional cell carcinoma of the bladder by microsatellite analysis of cells exfoliated in urine. *Int J Cancer* (2001) 95:364–9. doi: 10.1002/1097-0215(20011120)95:6<364::AID-IJC1064>3.0.CO;2-V
  75. García-Baquero R, Puerta P, Beltran M, Alvarez-Mújica M, Alvarez-Ossorio JL, Sánchez-Carbayo M. Methylation of tumor suppressor genes in a novel panel predicts clinical outcome in paraffin-embedded bladder tumors. *Tumour Biol.* (2014) 35:5777–86. doi: 10.1007/s13277-014-1767-6
  76. Bryzgunova OE, Laktionov PP. Extracellular nucleic acids in urine: sources, structure, diagnostic potential. *Acta naturae* (2015) 7:48–54.
  77. Sasaki H, Yoshiike M, Nozawa S, Usuba W, Katsuoka Y, Aida K, et al. expression level of urinary microRNA-146a-5p is increased in patients with bladder cancer and decreased in those after transurethral resection. *Clin Genitourin Cancer* (2016) 14:e493–9. doi: 10.1016/j.clgc.2016.04.002
  78. Sethi S, Sethi S, Bluth MH. Clinical Implication of micrornas in molecular pathology: an update for 2018. *Clin Lab Med.* (2018) 38:237–51. doi: 10.1016/j.clm.2018.02.003
  79. Liu X, Wu Y, Wu Q, Wang Q, Yang Z, Li L. MicroRNAs in biofluids are novel tools for bladder cancer screening. *Oncotarget* (2017) 8:32370–9. doi: 10.18632/oncotarget.16026
  80. Fuessel S, Lohse-Fischer A, Vu Van D, Salomo K, Erdmann K, Wirth MP. quantification of micrornas in urine-derived specimens. *Methods Mol Biol.* (2018) 1655:201–26. doi: 10.1007/978-1-4939-7234-0\_16
  81. Xiao S, Wang J, Xiao N. MicroRNAs as noninvasive biomarkers in bladder cancer detection: a diagnostic meta-analysis based on qRT-PCR data. *Int J Biol Markers* (2016) 31:e276–85. doi: 10.5301/jbm.5000199
  82. Hanke M, Hoefig K, Merz H, Feller AC, Kausch I, Jocham D, et al. A robust methodology to study urine microRNA as tumor marker: microRNA-126 and microRNA-182 are related to urinary bladder cancer. *Urol Oncol.* (2010) 28:655–61. doi: 10.1016/j.urolonc.2009.01.027
  83. Wiklund ED, Gao S, Hulf T, Sibbritt T, Nair S, Costea DE, et al. MicroRNA alterations and associated aberrant DNA methylation patterns across multiple sample types in oral squamous cell carcinoma. *PLoS ONE* (2011) 6:e27840. doi: 10.1371/journal.pone.0027840
  84. Chen YH, Wang SQ, Wu XL, Shen M, Chen ZG, Chen XG, et al. Characterization of microRNAs expression profiling in one group of Chinese urothelial cell carcinoma identified by Solexa sequencing. *Urol Oncol.* (2013) 31:219–27. doi: 10.1016/j.urolonc.2010.11.007
  85. Braicu C, Cojocneanu-Petric R, Chira S, Truta A, Floares A, Petrut B, et al. Clinical and pathological implications of miRNA in bladder cancer. *Int J Nanomed.* (2015) 10:791–800. doi: 10.2147/IJN.S72904
  86. Pritchard CC, Kroh E, Wood B, Arroyo JD, Dougherty KJ, Miyaji MM, et al. Blood cell origin of circulating microRNAs: a cautionary note for cancer biomarker studies. *Cancer Prev Res.* (2012) 5:492–7. doi: 10.1158/1940-6207.CAPR-11-0370
  87. Eissa S, Matboli M, Essawy NO, Kotb YM. Integrative functional genetic-epigenetic approach for selecting genes as urine biomarkers for bladder cancer diagnosis. *Tumour Biol.* (2015) 36:9545–52. doi: 10.1007/s13277-015-3722-6
  88. Sapre N, Macintyre G, Clarkson M, Naeem H, Cmero M, Kowalczyk A, et al. A urinary microRNA signature can predict the presence of bladder urothelial carcinoma in patients undergoing surveillance. *Br J Cancer* (2016) 114:454–62. doi: 10.1038/bjc.2015.472
  89. Pospisilova S, Pazourkova E, Horinek A, Brisuda A, Svoboda I, Soukup V, et al. MicroRNAs in urine supernatant as potential non-invasive markers for bladder cancer detection. *Neoplasia* (2016) 63:799–808. doi: 10.4149/neo\_2016\_518
  90. Pignot G, Cizeron-Clairac G, Vacher S, Susini A, Tozlu S, Vieillefond A, et al. MicroRNA expression profile in a large series of bladder tumors: identification of a 3-miRNA signature associated with aggressiveness of muscle-invasive bladder cancer. *Int J Cancer* (2013) 132:2479–91. doi: 10.1002/ijc.27949
  91. Yun SJ, Jeong P, Kim WT, Kim TH, Lee YS, Song PH, et al. Cell-free microRNAs in urine as diagnostic and prognostic biomarkers of bladder cancer. *Int J Oncol.* (2012) 41:1871–8. doi: 10.3892/ijo.2012.1622
  92. Matsushita R, Seki N, Chiyomaru T, Inoguchi S, Ishihara T, Goto Y, et al. Tumour-suppressive microRNA-144-5p directly targets CCNE1/2 as potential prognostic markers in bladder cancer. *Br J Cancer* (2015) 113:282–9. doi: 10.1038/bjc.2015.195
  93. Zhang DZ, Lau KM, Chan ES, Wang G, Szeto CC, Wong K, et al. Cell-free urinary microRNA-99a and microRNA-125b are diagnostic markers for the non-invasive screening of bladder cancer. *PLoS ONE* (2014) 9:e100793. doi: 10.1371/journal.pone.0100793
  94. Tölle A, Jung M, Rabenhorst S, Kilic E, Jung K, Weikert S. Identification of microRNAs in blood and urine as tumour markers for the detection of urinary bladder cancer. *Oncol Rep.* (2013) 30:1949–56. doi: 10.3892/or.2013.2621
  95. Chen L, Cui Z, Liu Y, Bai Y, Lan F. MicroRNAs as biomarkers for the diagnostics of bladder cancer: a meta-analysis. *Clin Lab.* (2015) 61:1101–8.
  96. Juracek J, Peltanova B, Dolezel J, Fedorko M, Pacik D, Radova L, et al. Genome-wide identification of urinary cell-free microRNAs for non-invasive detection of bladder cancer. *J Cell Mol Med.* (2018) 22:2033–8. doi: 10.1111/jcmm.13487
  97. Pardini B, Cordero F, Naccarati A, Viberti C, Birolo G, Oderda M, et al. MicroRNA profiles in urine by next-generation sequencing can stratify bladder cancer subtypes. *Oncotarget* (2018) 9:20658–69. doi: 10.18632/oncotarget.25057
  98. Matullo G, Naccarati A, Pardini B. MicroRNA expression profiling in bladder cancer: the challenge of next-generation sequencing in tissues and biofluids. *Int J Cancer* (2016) 138:2334–45. doi: 10.1002/ijc.2989
  99. Zhang M, Ren B, Li Z, Niu W, Wang Y. Expression of N-Myc Downstream-Regulated Gene 2 in bladder cancer and its potential utility as a urinary diagnostic biomarkers. *Med Sci Monit.* (2017) 23:4644–9. doi: 10.12659/MSM.901610

100. Andreu Z, Otta Oshiro R, Redruello A, López-Martín S, Gutiérrez-Vázquez C, Morato E, et al. Extracellular vesicles as a source for non-invasive biomarkers in bladder cancer progression. *Eur J Pharm Sci.* (2017) 98:70–9. doi: 10.1016/j.ejps.2016.10.008
101. Armstrong DA, Green BB, Seigne JD, Schned AR, Marsit CJ. MicroRNA molecular profiling from matched tumor and bio-fluids in bladder cancer. *Mol Cancer* (2015) 14:194. doi: 10.1186/s12943-015-0466-2
102. Du L, Duan W, Jiang X, Zhao L, Li J, Wang R, et al. Cell-free lncRNA expression signatures in urine serve as novel non-invasive biomarkers for diagnosis and recurrence prediction of bladder cancer. *J Cell Mol Med.* (2018) 22:2838–45. doi: 10.1111/jcmm.13578
103. Berrondo C, Flax J, Kucharov V, Siebert A, Osinski T, Rosenberg A, et al. Expression of the long non-coding rna hotair correlates with disease progression in bladder cancer and is contained in bladder cancer patient urinary exosomes. *PLoS ONE* (2016) 11:e0147236. doi: 10.1371/journal.pone.0147236
104. Chen CL, Lai YF, Tang P, Chien KY, Yu JS, Tsai CH et al. Comparative and targeted proteomic analyses of urinary microparticles from bladder cancer and hernia patients. *J Proteome Res.* (2012) 11:5611–29. doi: 10.1021/pr3008732
105. Martens-Uzunova ES, Böttcher R, Croce CM, Jenster G, Visakorpi T, Calin GA. Long noncoding RNA in prostate, bladder, and kidney cancer. *Eur Urol.* (2014) 65:1140–51. doi: 10.1016/j.eururo.2013.12.003
106. Terracciano D, Ferro M, Terreri S, Lucarelli G, D'Elia C, Musi G, et al. Urinary long noncoding RNAs in nonmuscle-invasive bladder cancer: new architects in cancer prognostic biomarkers. *Transl Res.* (2017) 184:108–17. doi: 10.1016/j.trsl.2017.03.005
107. Fan Y, Shen B, Tan M, Mu X, Qin Y, Zhang F, et al. Long non-coding RNA UCA1 increases chemoresistance of bladder cancer cells by regulating Wnt signaling. *FEBS J.* (2014) 281:1750–8. doi: 10.1111/febs.12737
108. Peter S, Borkowska E, Drayton RM, Rakhit CP, Noon A, Chen W, et al. Identification of differentially expressed long noncoding RNAs in bladder cancer. *Clin Cancer Res.* (2014) 20:5311–21. doi: 10.1158/1078-0432.CCR-14-0706
109. Neuhausen A, Florl AR, Grimm MO, Schulz WA. DNA methylation alterations in urothelial carcinoma. *Cancer Biol Ther.* (2006) 5:993–1001. doi: 10.4161/cbt.5.8.2885
110. Deligezer U, Erten N, Akisik EE, Dalay N. Circulating fragmented nucleosomal DNA and caspase-3 mRNA in patients with lymphoma and myeloma. *Exp Mol Pathol.* (2006) 80:72–6. doi: 10.1016/j.yexmp.2005.05.001
111. Kim WT, Jeong P, Yan C, Kim YH, Lee IS, Kang HW, et al. UBE2C cell-free RNA in urine can discriminate between bladder cancer and hematuria. *Oncotarget* (2016) 7:58193–202. doi: 10.18632/oncotarget.11277
112. Kim WT, Kim YH, Jeong P, Seo SP, Kang HW, Kim YJ, et al. Urinary cell-free nucleic acid IQGAP3: a new non-invasive diagnostic marker for bladder cancer. *Oncotarget* (2018) 9:14354–65. doi: 10.18632/oncotarget
113. Pichler R, Fritz J, Tulchiner G, Klinglmair G, Soleiman A, Horninger W, et al. Increased accuracy of a novel mRNA-based urine test for bladder cancer surveillance. *BJU Int.* (2018) 121:29–37. doi: 10.1111/bju.14019
114. Malentacchi F, Vinci S, Melina AD, Kuncova J, Villari D, Nesi G, et al. Urinary carbonic anhydrase IX splicing messenger RNA variants in urogenital cancers. *Urol Oncol.* (2016) 34:292.e9–292.e16. doi: 10.1016/j.urolonc.2016.02.01
115. Haussecker D, Huang Y, Lau A, Parameswaran P, Fire AZ, Kay MA. Human tRNA-derived small RNAs in the global regulation of RNA silencing. *RNA* (2010) 16:673–95. doi: 10.1261/rna.2000810
116. Lee J, McKinney KQ, Pavlopoulos AJ, Niu M, Kang JW, Oh JW, et al. Altered proteome of extracellular vesicles derived from bladder cancer patients urine. *Mol Cells* (2018) 41:179–87. doi: 10.14348/molcells.2018.2110
117. Yasui T, Yanagida T, Ito S, Konakade Y, Takeshita D, Naganawa T, et al. Unveiling massive numbers of cancer-related urinary-microRNA candidates via nanowires. *Sci Adv.* (2017) 3:e1701133. doi: 10.1126/sciadv.1701133
118. Yu S, Cao H, Shen B, Feng J. Tumor-derived exosomes in cancer progression and treatment failure. *Oncotarget* (2015) 6:37151–68. doi: 10.18632/oncotarget.6022
119. Rosell R, Wei J, Taron M. Circulating MicroRNA signatures of tumor-derived exosomes for early diagnosis of non-small-cell lung cancer. *Clin Lung Cancer* (2009) 10:8–9. doi: 10.3816/CLC.2009.n.001
120. Beckham CJ, Olsen J, Yin PN, Wu CH, Ting HJ, Hagen FK, et al. Bladder cancer exosomes contain EDIL-3/Del1 and facilitate cancer progression. *J Urol.* (2014) 192:583–92. doi: 10.1016/j.juro.2014.02.035
121. Huang X, Liang M, Dittmar R, Wang L. Extracellular microRNAs in urologic malignancies: chances and challenges. *Int J Mol Sci.* (2013) 14:14785–99. doi: 10.3390/ijms140714785
122. Wu P, Zhang G, Zhao J, Chen J, Chen Y, Huang W, et al. Profiling the urinary microbiota in male patients with bladder cancer in China. *Front Cell Infect Microbiol.* (2018) 8:167. doi: 10.3389/fcimb.2018.00167
123. Ward DG, Bryan RT. Liquid biopsies for bladder cancer. *Transl Androl Urol.* (2017) 6:331–5. doi: 10.21037/tau.2017.03.08
124. Wald C. Diagnostics: a flow of information. *Nature* (2017) 551:S48–50. doi: 10.1038/551S48a
125. Piao XM, Byun YJ, Kim WJ, Kim J. Unmasking molecular profiles of bladder cancer. *Invest Clin Urol.* (2018) 59:72–82. doi: 10.4111/icu.2018.59.2.72
126. Soria F, Droller MJ, Lotan Y, Gontero P, D'Andrea D, Gust KM, et al. An up-to-date catalog of available urinary biomarkers for the surveillance of non-muscle invasive bladder cancer. *World J Urol.* (2018). doi: 10.1007/s00345-018-2380-x. [Epub ahead of print].

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Santoni, Morelli, Amantini and Battelli. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.



# Circulating Tumor Cells in Renal Cell Carcinoma: Recent Findings and Future Challenges

Matteo Santoni<sup>1†</sup>, Alessia Cimadamore<sup>2†</sup>, Liang Cheng<sup>3</sup>, Antonio Lopez-Beltran<sup>4</sup>, Nicola Battelli<sup>1</sup>, Francesco Massari<sup>5</sup>, Marina Scarpelli<sup>1</sup>, Andrea Benedetto Galosi<sup>6</sup>, Sergio Bracarda<sup>7</sup> and Rodolfo Montironi<sup>1\*</sup>

<sup>1</sup> Oncology Unit, Macerata Hospital, Macerata, Italy, <sup>2</sup> Section of Pathological Anatomy, School of Medicine, United Hospitals, Polytechnic University of the Marche Region, Ancona, Italy, <sup>3</sup> Department of Pathology and Laboratory Medicine, Indiana University School of Medicine, Indianapolis, IN, United States, <sup>4</sup> Department of Pathology and Surgery, Faculty of Medicine, Córdoba, Spain, <sup>5</sup> Division of Oncology, S. Orsola-Malpighi Hospital, Bologna, Italy, <sup>6</sup> Department of Urology, School of Medicine, United Hospitals, Marche Polytechnic University, Ancona, Italy, <sup>7</sup> Medical Oncology, Department of Oncology, Azienda Ospedaliera S. Maria, Terni, Italy

**Keywords:** circulating tumor cells, diagnosis, isolation techniques, prognosis, renal cell carcinoma, circulating tumor microemboli

## INTRODUCTION

Renal cell carcinoma (RCC) is the most common tumor of the kidney. After diagnosis, 20–30% of patients will relapse, with a high probability of death from cancer-related causes. The development of non-invasive biomarkers will allow the identification of patients with a high risk of recurrence after radical or partial nephrectomy and will improve the assessment of tumor response to targeted therapy or immunotherapy. The search for non-invasive diagnostic techniques represents one of the most difficult challenges for cancer researchers. The contemporary scenario includes a variety of strategies that share the aim of maximally reducing the impact of the diagnosis on patients' quality of life (QoL). In this context, liquid biopsy offers a promising perspective for cancer diagnosis and monitoring, with several advantages compared to traditional diagnostic procedures (1). Indeed, it can be more frequently performed and allows for better tracking of tumors and mutations over a period of time (1). Moreover, it has been showed that genomic profiles of liquid biopsies can match very closely with the corresponding tumors (2).

The liquid biopsy of circulating tumor cells (CTCs), which belong to the larger family of circulating rare cells (CRC), has been validated and approved by the US Food and Drug Administration (FDA) as a useful prognostic tool in a variety of cancer types (3). This is based not only on the ability of CTCs to be a mirror of tumor heterogeneity but also on the possibility to combine the genetic and transcriptomic status of single CTCs (4) with epigenome analyses (5). Although CTCs have received great attention based on their potential in evaluating the status of localized and metastatic diseases, their clinical implementation is not yet widespread.

In the last decade, several studies have investigated the clinical and pathological significance of CTC numbers and characteristics in patients with urogenital cancers (6, 7). In view of the increasing number of dedicated clinical trials, genitourinary tumors represent the next urgent field of application of molecular diagnostics and drug discovery after gastro-intestinal and thoracic oncology. Among these tumors, the absence of reliable predictive biomarkers in RCC prevents the proper selection of patients who will benefit from any one of the three main drug categories approved for treating this disease: (1) anti-vascular endothelial growth factor (VEGF) monoclonal antibodies (i.e., bevacizumab) or tyrosine kinase inhibitors (i.e., sunitinib, sorafenib, pazopanib, axitinib, cabozantinib, tivozanib, and lenvatinib); (2) immune checkpoint inhibitors [i.e., anti-programmed death(PD)-1 Nivolumab alone or in combination with anti-cytotoxic T-lymphocyte antigen (CTLA)-4]; and (3) Mammalian target of rapamycin (mTOR) inhibitors

## OPEN ACCESS

### Edited by:

Ronald M. Bukowski,  
Cleveland Clinic, United States

### Reviewed by:

Elena Ranieri,  
University of Foggia, Italy

### \*Correspondence:

Rodolfo Montironi  
r.montironi@univpm.it

<sup>†</sup>These authors have contributed  
equally to this work

### Specialty section:

This article was submitted to  
Genitourinary Oncology,  
a section of the journal  
Frontiers in Oncology

**Received:** 02 October 2018

**Accepted:** 14 March 2019

**Published:** 05 April 2019

### Citation:

Santoni M, Cimadamore A, Cheng L, Lopez-Beltran A, Battelli N, Massari F, Scarpelli M, Galosi AB, Bracarda S and Montironi R (2019) Circulating Tumor Cells in Renal Cell Carcinoma: Recent Findings and Future Challenges. *Front. Oncol.* 9:228. doi: 10.3389/fonc.2019.00228

(i.e., everolimus and temsirolimus). In this manuscript we describe the emerging data on the role of CTCs in the diagnosis and treatment of RCC, focusing on their future applicability in daily clinical practice.

## INTRA- AND INTER-TUMOR HETEROGENEITY IN RCC

Intratumoral heterogeneity—in terms of somatic mutations, chromosome aberrations, and tumor gene expression—is a characteristic feature of clear cell RCC (8–11). These alterations are primarily centered around the *Von Hippel-Lindau* (*VHL*) gene and include LOH at 3p and epigenetic silencing. *VHL* inactivation is a crucial event in the majority of clear cell RCC and represents the only ubiquitous event, in contrast from the other genetic and/or epigenetic aberrations reported which are just subclonal (12–14). In this setting, Xu et al. firstly revealed by single-cell exome sequencing in a single patient that <30% of gene alterations are common to multiple cells within tumor tissue, whilst the majority are only cell-specific (15). In the same view, activated drug target pathways have been shown to be considerably variable within the same tumor and between primary RCC and lung metastases (16).

The rate of patients with late-relapsing disease (>5 y after radical or partial nephrectomy) (17–19) and the common intravenous tumor embolization (20) are just two of a series of clear signs suggesting that CTCs may provide fundamental information to optimize RCC diagnosis and evaluate tumor response to therapy and progression.

## CTC ISOLATION AND CHARACTERIZATION IN RCC

The collection, identification, enrichment, and analysis of CTC require the use of different methods, including the following: (1) Epithelial or non-epithelial marker-dependent isolation; (2) RT-PCR-based methods; (3) and morphological- and cell size-based detection (21).

The first method consists of the detection of CTCs through epithelial markers, such as the epithelial cell adhesion molecule (EpCAM, **Figure 1**). EpCAM is a transmembrane glycoprotein involved in cell signaling, migration, proliferation, and differentiation (22). The number of CTCs that can be isolated by EpCAM is usually low (23). This is based on the biological behavior of clear cell RCC, which often transdifferentiates through a process named “epithelial-to-mesenchymal transition (EMT),” a morphological transformation that is phenotypic of RCC cells (24, 25) and that leads to the loss of their epithelial antigens and the acquisition of mesenchymal features (i.e., vimentin expression). Detecting EMT markers on CTCs provides fundamental information on the status of the disease, considering the straight association between EMT and the prognosis of RCC patients (24) as well as its role in the acquisition of invasive properties and resistance to anti-VEGF TKIs (24). More recently, antibodies directed against membrane carbonic anhydrase 9 (CA9/CAIX) and CD147 [a widely expressed

membrane glycoprotein involved in matrix metalloproteinase induction, cell adhesion and T cell activation (26)] have been developed to increase the number of selected CTCs in RCC patients (**Figure 1**). Indeed, Liu et al. reported that while EpCAM was found only in about 18% of clear cell RCC tumors, CAIX, and CD147 were present in more than 97% of samples (27).

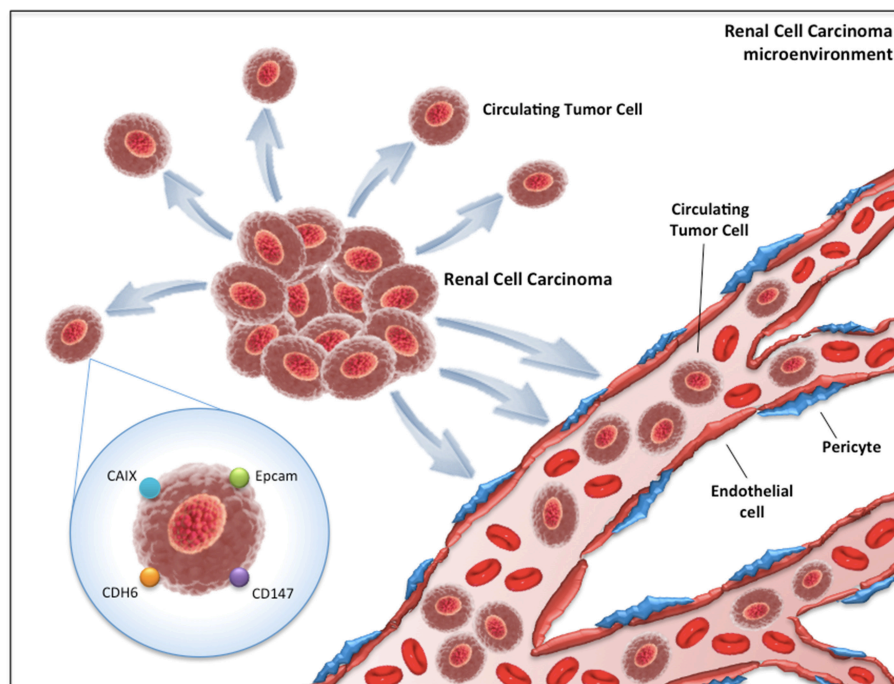
The second method is based on the RT-PCR approach. The three main targets of this technique are *CAIX*, *VHL* and *Cadherin-6* (*CDH-6*) (**Figure 1**). *VHL* gene alterations detected in tumor samples have reported concordance with those identified on peripheral blood in about 75% of cases (28). On the other hand, *CDH-6* gene expression by RT-PCR has been observed in 45% of clear cell RCC blood samples (29).

The third method is built upon size-based blood filtration combined with morphological and genetic analyses. The need to associate different techniques derives from the evidence that cytomorphological classification alone is not sufficient to detect CTCs in RCC patients (30). Interestingly, the use of these combined methods has enabled the detection of the presence of circulating clusters with a core of cancer cells surrounded by an external coating of endothelial cells (31).

The survival mechanisms underlying the circulation and migration of CTCs depend on multiple factors. Their biological characteristics, genetic alterations, epithelial mesenchymal transition, and cancer stem cell properties are internal factors that influence their survival. Great importance is now also being attributed to the external factors in the bloodstream microenvironment, consisting of platelets, immune cells, cytokines, and circulating tumor microemboli (CTM) (32).

CTM are composed by cell clusters, from two to more than 50 CTCs, together with leukocytes, cancer-associated fibroblasts, endothelial cells, and platelets (33). CTM demonstrated high metastatic potential by inhibiting apoptosis, promoting cell clonal proliferation, conferring resistance to shear stress, and protecting the innermost cells from immune surveillance from the identification of NK cells and their cytolytic activity (34). Testing the expression of Ki-67 on CTCs and CTMs has revealed a sharp contrast between CTCs (Ki-67 positive) and CTMs (all negative for Ki-67), even in patients with Ki67 (+) CTCs (35). This gives rise to the hypothesis that CTMs, in comparison to CTCs, may have the capability to remain inactive and “dormant” for long time periods, inhibiting apoptosis and cell destruction in the bloodstream. They may also have the capacity to confer resistance to cytotoxic drugs for cells within CTM, as observed by Hou et al. in patients with non-small-cell lung cancer (36). Colorectal cancer patients with CTMs in their blood have a shorter survival period than patients with only CTCs detected (37). Similar results were obtained in blood samples from gastric cancer patients, in which the presence of CTMs was an independent predictor of shorter PFS and OS in stage IV patients in multivariate analysis (38). Detection of CTM has emerged as a valuable tool to improve the prognostic significance of liquid biopsy (39) (**Figure 2**). Comparing the epigenomes of CTC clusters and single CTCs, Gkoutela et al. demonstrated that CTC clusters are enriched with binding sites for several key transcription factors (TF)—such as OCT4, NANOG, and SOX2—that confer to clusters stem cell features, whereas single





**FIGURE 1 |** Circulating tumor cells (CTCs) in renal cell carcinoma microenvironment. CAIX, Carbonic anhydrase 9; CDH6, Cadherin-6; Epcam, Epithelial cell adhesion molecule.

CTC have enrichment in different sets of TF. Moreover, stem cell TF binding is lost when clusters are dissociated into single cells via  $\text{Na}^+/\text{K}^+$  ATPase inhibitors such as digitoxin. In xenograft mouse models generated using *ex vivo* expanded CTC lines, administration of digitoxin significantly inhibited the capacity of CTC clusters to generate metastases (40, 41). These results open an area of new research and new therapeutic targets.

## NEW METHODS OF DETECTION

New methods of isolation and analysis such as subtraction enrichment (SE) combined with immunostaining-fluorescence *in situ* hybridization (iFISH) are being developed to better characterize CTCs. With this method, independent of cell size variation, and free of hypotonic damage as well as anti-EpCAM perturbing, it is possible to karyotype chromosome ploidy of CTCs and phenotype multi-protein expression. Among different cancers, an aneuploid chromosome 8 (tetraploid or polyploid) identified a positive CTC (42–44). This allows for efficient enrichment, identification, and characterization of both large and small size CTCs as well as CTM in various biofluid samples including cerebrospinal fluid (CSF). Unlike conventional methodologies, SE-iFISH enables the characterization of different heterogeneous CTC subtypes classified by both chromosome ploidy and the expression of biomarkers (44). Broncy et al. recently applied single-cell genetic analysis after isolation by SizE of Tumor/Trophoblastic cells (ISET<sup>®</sup>) in order to assess the specificity and sensitivity of cytopathology. They performed single cell analysis targeted to VHL mutations in all the 205 CRCs identified by cytopathology in the blood of 29 ccRCC patients after ISET filtration. They found a complete (100%) specificity of

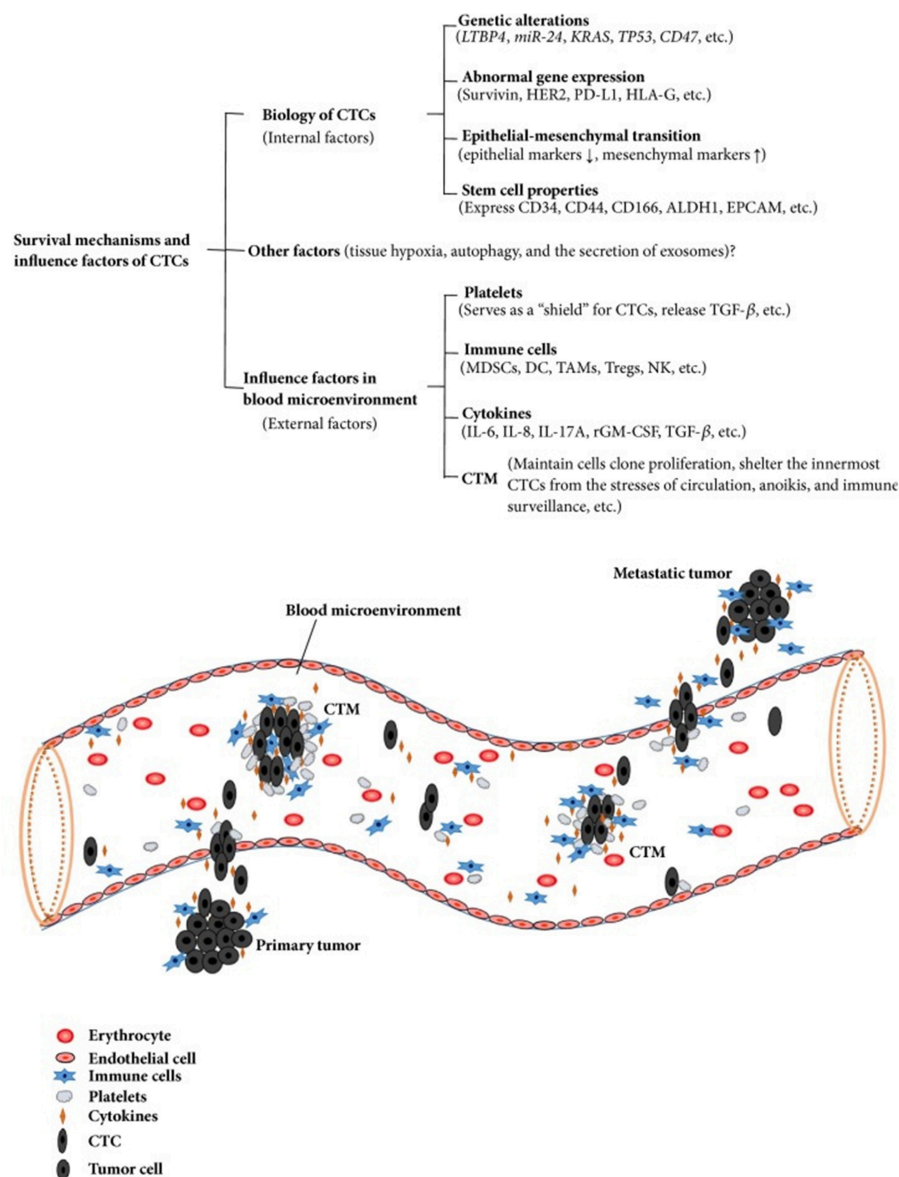
the cytopathological approach in the identification of circulating cancer cells (CCC) with a low sensitivity (35%) compared to genetic analysis (72%) (45, 46).

A novel straight microfluidic chip technology to focus and capture CTCs has been applied in head and neck cancer patients. The microchip is designed based on inertial migration of cells in a straight microchannel and allow to isolate single CTCs, CTCs clusters, and CTM by a size-based method, with high recovery efficiencies and low background cell contamination (47, 48).

## FUTURE PERSPECTIVES

In future years, technical advances should aim to isolate a greater number of CTCs in metastatic patients than in patients with localized disease and to find the same mutations present in the correspondent histologic sample or, especially, in the metastatic cohort. In our opinion, this could be realized by improving the connection between cytomorphological and genetic analyses, thus overcoming the limits of present techniques, which can be challenging and time-consuming. Indeed, the specificity of CAIX and CDH-6 for CTCs in RCC patients is poor and could be increased only by the design of studies focused on comparing CTCs isolated from patients with clear cell and from patients with benign kidney diseases or healthy volunteers.

We hope that in future years whole genome, transcriptome, and proteome analyses of single cells could lead to an increase in our knowledge of tumor heterogeneity and acquired drug resistance. In the localized RCC setting, CTCs could have potential as a surveillance biomarker for disease recurrence. Earlier detection of metastatic RCC, prior to the onset of symptoms, may lead to improved clinical outcomes. In patients



**FIGURE 2 |** Survival mechanisms and influence factors of CTCs. Copyright © 2018 Wang et al. (32). This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

with metastatic disease, CTC analysis could be used to select patients for biomarker-guided clinical trials. As in colorectal cancers, the mutational profile of metastatic RCC could evolve after treatment progression, developing an acquired resistance to therapy potentially investigable in a non-invasive way with CTCs. Moreover, the introduction of data on CTCs within the TNM classification represents another step forward on the route of personalized medicine for RCC patients.

## CONCLUSIONS

RCC may benefit from the development of non-invasive and reliable biomarkers, enabling early and timely personalized

treatment changes. The introduction of CTC analysis within daily clinical practice for patients with RCC seems still far at the moment. However, the advances obtained in the last 5 years in isolating and analyzing CTCs bring optimism about the future therapeutic landscape in RCC patients.

## AUTHOR CONTRIBUTIONS

RM and MSc: conception and design; MSa, AC, and FM: drafting the manuscript; NB and ABG: review of the literature; LC, SB, and AL-B: critical revision of the manuscript.

## REFERENCES

- Crowley E, Di Nicolantonio F, Loupakis F, Bardelli A. Liquid biopsy: monitoring cancer-genetics in the blood. *Nat Rev Clin Oncol.* (2013) 10:472–84. doi: 10.1038/nrclinonc.2013.110
- Gingras I, Salgado R, Ignatiadis M. Liquid biopsy: will it be the “magic tool” for monitoring response of solid tumors to anticancer therapies? *Curr Opin Oncol.* (2015) 27:560–7. doi: 10.1097/CCO.0000000000000223
- Karachaliou N, Mayo-de-Las-Casas C, Molina-Vila MA, Rosell R. Real-time liquid biopsies become a reality in cancer treatment. *Ann Transl Med.* (2015) 3:36. doi: 10.3978/j.issn.2305-5839.2015.01.16
- Klein CA, Seidl S, Petat-Dutter K, Offner S, Geigl JB, Schmidt-Kittler O, et al. Combined transcriptome and genome analysis of single micrometastatic cells. *Nat. Biotechnol.* (2002) 20:387–92. doi: 10.1038/nbt0402-387
- Zhao C, Hu S, Huo X, Zhang Y. Dr.seq2: a quality control and analysis pipeline for parallel single cell transcriptome and epigenome data. *PLoS ONE.* (2017) 12:e0180583. doi: 10.1371/journal.pone.0180583
- Montironi R, Lopez-Beltran A, Cheng L, Cimadamore A, Zizzi A, Galosi A, et al. EAU Section of Urothology (ESUP): urothologists are fast moving forward to include blood (liquid) biopsy in their routine armamentarium. *Eur Urol Today.* (2018) 2018:26–27.
- Bergerot PG, Hahn AW, Bergerot CD, Jones J, Pal SK. The role of circulating tumor DNA in renal cell carcinoma. *Curr Treat Options Oncol.* (2018) 19:10. doi: 10.1007/s11864-018-0530-4
- Santoni M, Santini D, Massari F, Conti A, Iacovelli R, Burattini L, et al. Heterogeneous drug target expression as possible basis for different clinical and radiological response to the treatment of primary and metastatic renal cell carcinoma: suggestions from bench to bedside. *Cancer Metastasis Rev.* (2014) 33:321–31. doi: 10.1007/s10555-013-9453-5
- Piva F, Santoni M, Matrana MR, Satti S, Giulietti M, Occhipinti G, et al. BAP1, PBRM1 and SETD2 in clear cell renal cell carcinoma: molecular diagnostics and possible targets for personalized therapies. *Expert Rev Mol Diagn.* (2015) 15:1201–10. doi: 10.1586/14737159.2015.1068122
- Massari F, Ciccarese C, Santoni M, Brunelli M, Piva F, Modena A, et al. Metabolic alterations in renal cell carcinoma. *Cancer Treat Rev.* (2015) 41:767–76. doi: 10.1016/j.ctrv.2015.07.002
- Piva F, Giulietti M, Occhipinti G, Santoni M, Massari F, Sotte V, et al. Computational analysis of the mutations in BAP1, PBRM1 and SETD2 genes reveals the impaired molecular processes in renal cell carcinoma. *Oncotarget.* (2015) 6:32161–8. doi: 10.18632/oncotarget.5147
- Gerstung M, Beisel C, Rechsteiner M, Wild P, Schraml P, Moch H, et al. Reliable detection of subclonal single-nucleotide variants in tumour cell populations. *Nat. Commun.* (2012) 3:811. doi: 10.1038/ncomms1814
- Gerlinger M, Rowan AJ, Horswell S, Math M, Larkin J, Endesfelder D, et al. Intratumor heterogeneity and branched evolution revealed by multiregion sequencing. *N Engl J Med.* (2012) 366:883–92. doi: 10.1056/NEJMoa1113205
- Gerlinger M, Horswell S, Larkin J, Rowan AJ, Salm MP, Varela I, et al. Genomic architecture and evolution of clear cell renal cell carcinomas defined by multiregion sequencing. *Nat Genet.* (2014) 46:225–33. doi: 10.1038/ng.2891
- Xu X, Hou Y, Yin X, Bao L, Tang A, Song L, et al. Single-cell exome sequencing reveals single-nucleotide mutation characteristics of a kidney tumor. *Cell.* (2012) 148:886–95. doi: 10.1016/j.cell.2012.02.025
- Kim KT, Lee HW, Lee HO, Song HJ, Jeong da E, Shin S, et al. Application of single-cell RNA sequencing in optimizing a combinatorial therapeutic strategy in metastatic renal cell carcinoma. *Genome Biol.* (2016) 17:80. doi: 10.1186/s13059-016-0945-9
- Santoni M, Conti A, Porta C, Procopio G, Sternberg CN, Basso U, et al. Sunitinib, pazopanib or sorafenib for the treatment of patients with late-relapsing (>5 years) metastatic renal cell carcinoma. *J Urol.* (2015) 193:41–7. doi: 10.1016/j.juro.2014.07.011
- Santoni M, Buti S, Conti A, Porta C, Procopio G, Sternberg CN, et al. Prognostic significance of host immune status in patients with late relapsing renal cell carcinoma treated with targeted therapy. *Targeted Oncol.* (2015) 10:517–22. doi: 10.1007/s11523-014-0356-3
- Santoni M, Conti A, Procopio G, Porta C, Ibrahim T, Barni S, et al. Bone metastases in patients with metastatic renal cell carcinoma: are they always associated with poor prognosis? *J Exp Clin Cancer Res.* (2015) 34:10. doi: 10.1186/s13046-015-0122-0
- Gu L, Li H, Wang Z, Wang B, Huang Q, Lyu X, et al. A systematic review and meta-analysis of clinicopathologic factors linked to oncologic outcomes for renal cell carcinoma with tumor thrombus treated by radical nephrectomy with thrombectomy. *Cancer Treat Rev.* (2018) 69:112–20. doi: 10.1016/j.ctrv.2018.06.014
- van der Toom EE, Verdone JE, Gorin MA, Pienta KJ. Technical challenges in the isolation and analysis of circulating tumor cells. *Oncotarget.* (2016) 7:62754–66. doi: 10.18632/oncotarget.11191
- Maetzel D, Denzel S, Mack B, Canis M, Went P, Benk M, et al. Nuclear signalling by tumour-associated antigen EpCAM. *Nat Cell Biol.* (2009) 11:162–71. doi: 10.1038/ncb1824
- Gradilone A, Iacovelli R, Cortesi E, Raimondi C, Gianni W, Nicolazzo C, et al. Circulating tumor cells and “suspicious objects” evaluated through CellSearch® in metastatic renal cell carcinoma. *Anticancer Res.* (2011) 31:4219–21.
- Montironi R, Santoni M, Scarpelli M, Piva F, Lopez-Beltran A, Cheng L, et al. Re: epithelial-to-mesenchymal transition in renal neoplasms. *Eur Urol.* (2015) 68:736–7. doi: 10.1016/j.eururo.2015.06.031
- Piva F, Giulietti M, Santoni M, Occhipinti G, Scarpelli M, Lopez-Beltran A, et al. Epithelial to mesenchymal transition in renal cell carcinoma: implications for cancer therapy. *Mol Diagn Ther.* (2016) 20:111–7. doi: 10.1007/s40291-016-0192-5
- Hanna SM, Kirk P, Holt OJ, Puklavec MJ, Brown MH, Barclay AN. A novel form of the membrane protein CD147 that contains an extra Ig-like domain and interacts homophilically. *BMC Biochem.* (2003) 4:17. doi: 10.1186/1471-2091-4-17
- Liu S, Tian Z, Zhang L, Hou S, Hu S, Wu J, et al. Combined cell surface carbonic anhydrase 9 and CD147 antigens enable high-efficiency capture of circulating tumor cells in clear cell renal cell carcinoma patients. *Oncotarget.* (2016) 7:59877–91. doi: 10.18632/oncotarget.10979
- Ashida S, Okuda H, Chikazawa M, Tanimura M, Sugita O, Yamamoto Y, et al. Detection of circulating cancer cells with von Hippel-Lindau gene mutation in peripheral blood of patients with renal cell carcinoma. *Clin Cancer Res.* (2000) 6:3817–22.
- Li G, Passebosch-Faure K, Gentil-Perret A, Lambert C, Genin C, Tostain J. Cadherin-6 gene expression in conventional renal cell carcinoma: a useful marker to detect circulating tumor cells. *Anticancer Res.* (2005) 25:377–81.
- El-Heliebi A, Kroneis T, Zöhrer E, Haybaeck J, Fischereder K, Kampel-Kettner K, et al. Are morphological criteria sufficient for the identification of circulating tumor cells in renal cancer? *J. Transl. Med.* (2013) 11:214. doi: 10.1186/1479-5876-11-214
- Kats-Ugurlu K, Roodink I, de Weijert M, Tiemessen D, Maass C, Verrijp K, et al. Circulating tumour tissue fragments in patients with pulmonary metastasis of clear cell renal cell carcinoma. *J Pathol.* (2009) 219:287–93. doi: 10.1002/path.2613
- Wang W-C, Zhang X-F, Peng J, Li X-F, Wang A-L, Bie Y-Q, et al. Survival mechanisms and influence factors of circulating tumor cells. *Biomed Res Int.* (2018) 2018:6304701. doi: 10.1155/2018/6304701
- Krebs MG, Metcalf RL, Carter L, Brady G, Blackhall FH, Dive C. Molecular analysis of circulating tumour cells—biology and biomarkers. *Nat Rev Clin Oncol.* (2014) 11:129–44. doi: 10.1038/nrclinonc.2013.253
- Palumbo JS, Talmage KE, Massari JV, La Jeunesse CM, Flick MJ, Kombrinck KW, et al. Platelets and fibrin(ogen) increase metastatic potential by impeding natural killer cell-mediated elimination of tumor cells. *Blood.* (2005) 105:178–85. doi: 10.1182/blood-2004-06-2272
- Li J, Sharkey CC, Wun B, Liesveld JL, King MR. Genetic engineering of platelets to neutralize circulating tumor cells. *J Control Release.* (2016) 228:38–47. doi: 10.1016/j.jconrel.2016.02.036
- Hou JM, Krebs MG, Lancashire L, Sloane R, Backen A, Swain RK, et al. Clinical significance and molecular characteristics of circulating tumor cells and circulating tumor microemboli in patients with small-cell lung cancer. *J Clin Oncol.* (2012) 30:525–32. doi: 10.1200/JCO.2010.33.3716
- Zhang D, Zhao L, Zhou P, Ma H, Huang F, Jin M, et al. Circulating tumor microemboli (CTM) and vimentin+ circulating tumor cells (CTCs) detected by a size-based platform predict worse prognosis in advanced colorectal cancer patients during chemotherapy. *Cancer Cell Int.* (2017) 17:6. doi: 10.1186/s12935-016-0373-7

38. Zheng X, Fan L, Zhou P, Ma H, Huang S, Yu D, et al. Detection of circulating tumor cells and circulating tumor microemboli in gastric cancer. *Transl Oncol.* (2017) 10:431–41. doi: 10.1016/j.tranon.2017.02.007
39. Umer M, Vaidyanathan R, Nguyen NT, Shiddiky MJA. Circulating tumor microemboli: Progress in molecular understanding and enrichment technologies. *Biotechnol Adv.* (2018) 36:1367–89. doi: 10.1016/j.biotechadv.2018.05.002
40. Gkoutela S, Castro-Giner F, Szczerba BM, Vetter M, Landin J, Scherrer R, et al. Circulating tumor cell clustering shapes dna methylation to enable metastasis seeding. *Cell.* (2019) 176:98–112.e14. doi: 10.1016/j.cell.2018.11.046
41. Yu M. Metastasis stemming from circulating tumor cell clusters. *Trends Cell Biol.* (2019) 29:P275–6. doi: 10.1016/j.tcb.2019.02.001
42. Lin PP. Aneuploid CTC and CEC. *Diagnostics.* (2018) 8:E26. doi: 10.3390/diagnostics8020026
43. Ye Z, Ding Y, Chen Z, Li Z, Ma S, Xu Z, et al. Detecting and phenotyping of aneuploid circulating tumor cells in patients with various malignancies. *Cancer Biol Ther.* (2018) 20:546–51. doi: 10.1080/15384047.2018.1538000
44. Ge F, Zhang H, Wang DD, Li L, Lin PP. Enhanced detection and comprehensive *in situ* phenotypic characterization of circulating and disseminated heteroploid epithelial and glioma tumor cells. *Oncotarget.* (2015) 6:27049–64. doi: 10.18632/oncotarget.4819
45. Broncy L, Paterlini-Bréchet P. Circulating tumor cells for the management of renal cell carcinoma. *Diagnostics.* (2018) 8:63. doi: 10.3390/diagnostics8030063
46. Broncy L, Njima BB, Méjean A, Bérout C, Romdhane KB, Ilie M, et al. Single-cell genetic analysis validates cytopathological identification of circulating cancer cells in patients with clear cell renal cell carcinoma. *Oncotarget.* (2018) 9:20058–74. doi: 10.18632/oncotarget.25102
47. Kulasinghe A, Zhou J, Kenny L, Papautsky I, Punyadeera C. Capture of circulating tumour cell clusters using straight microfluidic chips. *Cancers.* (2019) 11:E89. doi: 10.3390/cancers11010089
48. Zhou J, Kulasinghe A, Bogseth A, O'Byrne K, Punyadeera C, Papautsky I. Isolation of circulating tumor cells in non-small-cell-lung cancer patients using a multi-flow microfluidic channel. *Nat Microsyst Nanotechnol.* (2018) 5:8. doi: 10.1038/s41378-019-0045-6

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Santoni, Cimadamore, Cheng, Lopez-Beltran, Battelli, Massari, Scarpelli, Galosi, Bracarda and Montironi. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.





# Emerging Prognostic Biomarkers in Testicular Germ Cell Tumors: Looking Beyond Established Practice

Michal Chovanec<sup>1,2</sup>, Costantine Albany<sup>2</sup>, Michal Mego<sup>1</sup>, Rodolfo Montironi<sup>3</sup>, Alessia Cimadamore<sup>3</sup> and Liang Cheng<sup>4,5\*</sup>

<sup>1</sup> 2nd Department of Oncology, Faculty of Medicine, Comenius University and National Cancer Institute, Bratislava, Slovakia, <sup>2</sup> Division of Hematology and Oncology, Indiana University Melvin and Bren Simon Cancer Center, Indianapolis, IN, United States, <sup>3</sup> Section of Pathological Anatomy, Polytechnic University of the Marche Region, School of Medicine, United Hospitals, Ancona, Italy, <sup>4</sup> Department of Pathology and Laboratory Medicine, Indiana University School of Medicine, Indianapolis, IN, United States, <sup>5</sup> Department of Urology, Indiana University School of Medicine, Indianapolis, IN, United States

## OPEN ACCESS

### Edited by:

Fabio Grizzi,  
Humanitas Research Hospital, Italy

### Reviewed by:

Riccardo Autorino,  
Virginia Commonwealth University,  
United States  
Ewa Rajpert-De Meyts,  
Rigshospitalet, Denmark  
Rodolfo A. Rey,  
Centro de Investigaciones  
Endocrinológicas "Dr. César Bergadá"  
(CEDIE), Argentina

### \*Correspondence:

Liang Cheng  
liang\_cheng@yahoo.com

### Specialty section:

This article was submitted to  
Genitourinary Oncology,  
a section of the journal  
Frontiers in Oncology

**Received:** 12 September 2018

**Accepted:** 14 November 2018

**Published:** 28 November 2018

### Citation:

Chovanec M, Albany C, Mego M,  
Montironi R, Cimadamore A and  
Cheng L (2018) Emerging Prognostic  
Biomarkers in Testicular Germ Cell  
Tumors: Looking Beyond Established  
Practice. *Front. Oncol.* 8:571.  
doi: 10.3389/fonc.2018.00571

Testicular germ cell tumors are unique among solid cancers. Historically, this disease was deadly if progressed beyond the stage I. The implementation of cisplatin-based chemotherapy regimens has drastically changed the clinical outcome of metastatic testicular cancer. Several biomarkers were established to refine the prognosis by International Germ Cell Collaborative Group in 1997. Among these, the most significant were primary tumor site; metastatic sites, such as non-pulmonary visceral metastases; and the amplitude of serum tumor markers  $\alpha$ -fetoprotein,  $\beta$ -chorionic gonadotropin, and lactate dehydrogenase. Since then, oncology has experienced discoveries of various molecular biomarkers to further refine the prognosis and treatment of malignancies. However, the ability to predict the prognosis and treatment response in germ cell tumors did not improve for many years. Clinical trials with novel targeting agents that were conducted in refractory germ cell tumor patients have proven to have negative outcomes. With the recent advances and developments, novel biomarkers emerge in the field of germ cell tumor oncology. This review article aims to summarize the current knowledge in the research of novel prognostic biomarkers in testicular germ cell tumors.

**Keywords:** testis, testicular germ cell tumors, molecular genetics, biomarkers, liquid biopsy

## INTRODUCTION

Testicular germ cell tumors (GCT) are unique in terms of molecular landscape, pathogenesis, clinical presentation, and response to chemotherapy (1). The exceptional position of GCT among the solid cancers can be perhaps attributed to their developmental origin in primordial germ cells. While the cure rate of patients with metastatic disease exceeds 80% (2), the ones failing the initial and salvage chemotherapy die of their disease in young age. About 40–80% of patients with relapsed GCT fail the salvage chemotherapy, resulting in the loss of 35 years of life on average (3–5). The utility of biomarkers to risk-stratify the treatment is well-established in GCT. Markers of the risk of relapse in the stage I disease, such as tumor size of >4 cm and rete testis invasion for seminoma, and lymphovascular invasion and predominance of embryonal carcinoma for non-seminoma, are currently used to risk-stratify the patients for surveillance or adjuvant treatment (6–9). International Germ Cell Cancer Collaborative Group (IGCCCG) presented the risk-stratification model for metastatic disease in 1997 using biomarkers such as primary tumor

site, metastatic sites, the amplitude of serum  $\alpha$ -fetoprotein (AFP),  $\beta$ -chorionic gonadotropin (HCG), and lactate dehydrogenase (LDH) (10). These criteria are based on patient series collected retrospectively between 1975 and 1990. Since then, the treatment strategy was optimized, and outcomes improved as reported from high volume centers (2, 11, 12). Further refining of IGCCCG criteria is expected soon in the updated version of the IGCCCG classification (**Figure 1**).

New reports on novel biomarkers are scarce since the introduction of the commonly used GCT biomarkers over three decades ago. The utility of novel molecular biomarkers in numerous solid cancers has significantly moved the advancement of oncology. Malignancies, such as lung cancer, melanoma, and kidney cancer, were previously considered untreatable, but now the array of molecular markers renders these diseases treatable with targeting agents ultimately prolonging lives of patients with incurable cancer (13, 14). Such advancement seemingly evades testicular GCT due to lack of known drugable targets. While the overall cure rate of GCT patients is excellent, ones refractory to standard chemotherapy lack the possibility to receive novel effective treatments and their prognosis is dismal. The biology of GCTs is unique, therefore translational research to uncover the biological implications is essential in the pursuit of treatment targets that may improve the prognosis of platinum refractory GCT patients. This article aims to summarize the current knowledge on the emerging biomarkers in GCT.

## BRIEF OVERVIEW OF MOLECULAR LANDSCAPE IN TESTICULAR GERM CELL TUMORS

Understanding why we lack a significant predictive biomarker in GCT requires a look into their molecular landscape and developmental origins. The origin of GCT particularly show how different their biology is compared to other solid cancers. The data from The Cancer Genome Atlas (TCGA) show a rather quiet mutational landscape in GCT compared to other solid tumors (15).

Several genomewide studies suggested driver mutations in only three genes (*KIT*, *KRAS*, and *NRAS*) in 4–31% of seminoma, and up to 14% of non-seminoma patients (16–19). Since these mutations were discovered in a minority of patients, a single universal mutational driver is not a feasible explanation in the development of GCT. Rather, a polygenic nature of testicular cancer was proposed, where the number of low frequency susceptibility genes (up to 50 risk loci reported until present) seems to produce an increased risk for the GCT (20). A recent paper by Shen et al. conducted a comprehensive molecular characterization of available tissue from 137 GCT patients. The authors confirmed findings of previously known mutated genes (*KIT*, *KRAS*, and *NRAS*) and provided yet additional evidence of low mutational burden with frequency of 0.5 mutations per megabase (15).

Despite the unimpressive mutational characteristics, GCT share a unique epigenetic landscape. GCT subtypes are an example of developmental processes from pluripotent embryonic

stem cells toward certain degrees of differentiation to somatic tissues. The mapping of GCT methylome is perhaps the most comprehensively assessed to this date. The global DNA-methylation status clearly correlates with the state of differentiation in the histological GCT subtypes. Seminomas, which show the lowest degree of differentiation are typically unmethylated or severely hypomethylated tumors. Embryonal carcinomas show low to intermediate levels of global DNA methylation and well-differentiated yolk sac tumors, and teratomas show high levels of DNA methylation. Thus, the significant histological variability complies with the epigenetic heterogeneity. These findings also comply with the epigenetic landscape of healthy tissues where differentiated somatic tissues show hypermethylated pattern (21–23). Non-CpG methylation, acetylation, and methylation of histones are also mechanisms likely involved in the biology of GCT. They are, however, poorly understood in present time. microRNA (miR) signaling research on the other hand seems to provide promising results toward increasing the knowledge about molecular biology of GCT. While the miR signaling is generally complex and is a subject of innumerable interactions, the clusters of miR discussed later in this paper provide a significant biomarker potency to further refine the management of GCT.

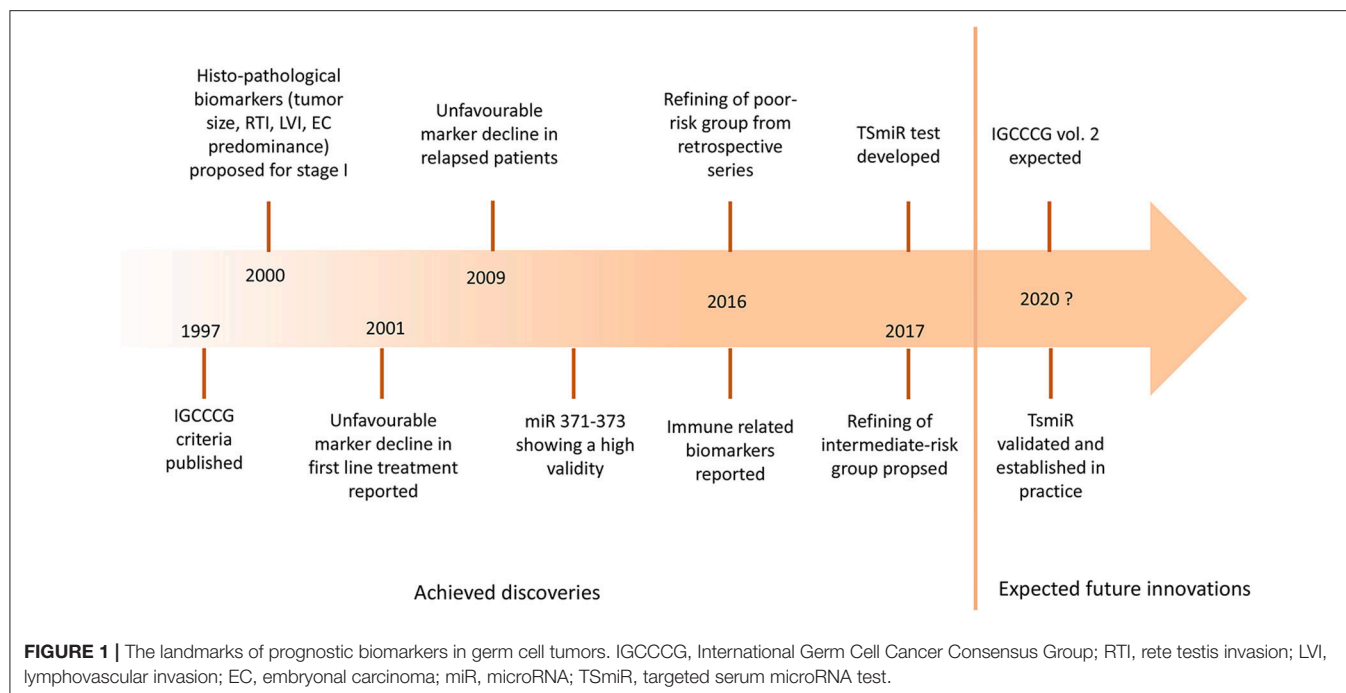
The unique germline origin of GCT is underlined with the overexpression of markers of pluripotency such as NANOG, OCT3/4 or a tissue stem cell factor KIT and its' ligand (24–30). The expressions of these markers have been linked to epigenetic regulation with DNA methylation and histone acetylation (30–34).

## EMERGING BIOMARKERS IN GERM CELL TUMORS

### Clinical Biomarkers

IGCCCG vol. 2 will bring a long-awaited update for risk stratification of treatment of GCT based on clinical characteristics. The advent of clinical biomarkers is rather slow since the original publication of the IGCCCG criteria. Several other risk assessment criteria were proposed that considered a more detailed look into clinical characteristics in GCT patients. Adra et al. published results of their retrospective analysis of 273 patients with a poor risk disease treated at Indiana University (35). Primary mediastinal non-seminoma (PMNSGCT), brain metastases and increasing age were significant predictors of mortality (HR = 4.63, 3.30, and 1.06, respectively). Multiple criteria for a poor risk disease carried a significantly worse prognosis compared to a single criterion (35).

Necchi et al. proposed an improved model for intermediate risk patients in the two-institutional initiative using PMNSGCT, brain metastases, pulmonary metastases, and age at diagnosis as risk factors. According to the results, a number of intermediate risk patients would suffice from treatment with BEPx3, whereas the current standard remains BEPx4 (11). While the refining of prognosis based on clinical criteria may have reached its limits, authors from Memorial Sloan Kettering Cancer



Center have proposed a novel prognostic marker based on a marker decline after the first course of chemotherapy (36). Patients who had unfavorable (slower) marker decline after the initiation if chemotherapy had reportedly worse outcomes compared to patients with favorable marker decline (72 vs. 95% for 2-year overall survival;  $P < 0.01$ ) (36). These findings were subsequently replicated in independent studies (37, 38).

Furthermore, the prognostic significance of tumor marker decline was reported also in patients with relapse (39–41). Fizazi et al. conducted a randomized phase III study in poor risk GCT. Patients receiving first cycle of BEP had an assessment of serum markers prior to second cycle and ones with an unfavorable decline were randomized to receive either remaining three cycles of standard BEP or dose-intensified chemotherapy regimen. Based on this biomarker-based strategy, a significant advantage was reported for 5-year progression-free survival (PFS) (60 vs. 48%,  $P = 0.037$ ), but not for 5-year overall survival (OS) (70 vs. 61%,  $P = 0.012$ ) (42, 43). Interestingly, in cases of progression, patients from this study relapsed predominantly in brain (54% of all relapses) (44).

## Molecular Biomarkers From Immunohistochemistry Studies

Immunohistochemistry studies have started to emerge in recent years to supplement the clinical biomarkers in predicting the prognosis of GCT. The higher expression of DNA repair enzyme poly (ADP-ribose) polymerase (PARP) was reported in GCT tissue compared to normal testicular tissue. However, no association with clinical characteristics nor the survival difference was reported in regard to levels of expression (45).

Kalavaska et al. published two studies examining the prognostic value of carbonic anhydrase nine assessed from plasma and from tumor tissue (46, 47). Levels of this marker of hypoxia and aggressive tumor behavior correlated in plasma and in tumor tissue. High expression in tumor was associated with shorter PFS; however, the clinically more useful utility of plasmatic assessment failed to be prognostic in GCT (46, 47). The hepatocyte growth factor (HGF) and its receptor c-MET were investigated by immunohistochemistry in tumors and in cell-line culture. c-MET is a known proto-oncogene involved in tumor progression and metastasis. Authors of this study reported an abundant immunohistochemical expression in both seminomas and non-seminomas, particularly in epithelial structures of well-differentiated subtypes such as teratomas, yolk sac tumors, and choriocarcinomas. Upon the activation of c-MET in an NT2 cell line (embryonal carcinoma), the cells acquired a more robust ability to proliferate, migrate, and invade. This may create the rationale for further research; however, the clinical significance of this finding is currently unknown (48).

## Immune-Related Biomarkers

The discovery of novel immune-related biomarkers, programmed-death receptor and its ligand (PD-1 and PD-L1) in various cancers, led to a confirmation of active PD-1/PD-L1 signaling also in GCT by Fankhauser et al. (49). The authors conducted an immunohistochemistry study and showed a frequent PD-L1 expression in 479 GCT tissue samples. Both seminomas and non-seminomas exhibited a significant expression of PD-L1 (in 73% and 64% of patients, respectively) (49).

Another research team led by Mardiak et al. performed a similar study and scored the PD-L1 expression semi-quantitatively with multiplicative quick score. The scores were correlated with clinical outcome. Patients with low levels of PD-L1 expression had significantly better PFS (HR = 0.40;  $P = 0.008$ ) and OS (HR = 0.43;  $P = 0.040$ ) (50). Furthermore, the expression of PD-L1 on tumor infiltrating lymphocytes (TIL) proved to be highly predictive of outcome in a reverse manner. Patients with high PD-L1 expression on TIL had significantly better prognosis than patients with low PD-L1 TIL (51). The prognostic significance of TIL was earlier reported by Bols et al., who also performed the phenotyping of immune-cell infiltrates (52). However, the abundant expression of PD-L1 does not seem to be predictive of response to treatment with immune-checkpoint inhibitors.

A phase II study with anti-PD1 agent pembrolizumab provided data about insufficient anti-tumor activity in refractory patients with GCT (53). While several case reports documented possible responses to immune-check point inhibitor, these are likely due to concomitant treatment with chemotherapy (54–56). Another phase II study with anti-PD-L1 agent avelumab is currently ongoing, which will shed more light on single agent immunotherapy in refractory GCT (NCT03403777).

Currently, there is a level of uncertainty in predicting response according to PD-L1 expression levels. While several cancer types have proven to be sensitive to PD-1/PD-L1 blockade based on PD-L1 expression, PD-L1 negative tumors were described to respond to such treatment as well. On the other hand, the expression of PD-L1 in tumor and TIL in GCT signifies a vivid immunogenic microenvironment but fails to respond to immunotherapy according to our present knowledge. As such, PD-1/PD-L1 axis seems to be only a part of the involved immune machinery and we are lacking a deeper understanding. Shen et al. recently published findings of comprehensive molecular characterization of GCT and did not discover a significant neoantigen signal in GCT, thus the insufficient activity of immune check-point inhibitors in GCT may be partly explained by this fact and the presence of very low mutational load (15).

Two independent studies published simultaneously examined the role of a simple marker of proinflammatory macroenvironment, a systemic-immune infiltration index (SII) (57, 58). SII is calculated from total counts of neutrophils, lymphocytes, and platelets. Fankhauser et al. reported numerous markers associated with poor prognosis in GCT, including low hemoglobin and albumin, high leukocytes, neutrophils, CRP, neutrophil to lymphocyte ratio, and SII (58). At the same time, our study showed that high SII was associated with poor prognosis in two independent cohorts of GCT patients. We also evaluated a combined prognostic value of SII and PD-L1 expression on TIL. As a result, we identified patients who never experienced death nor a relapse if they exhibited low SII and high PD-L1 on TIL (57). Both studies reported the prognostic significance of SII being independent from the standard IGCCCG risk criteria. SII can be easily calculated from complete blood count performed prior to treatment and offers a simple tool to predict outcome in metastatic GCT. Poor prognosis in patients exhibiting high levels of SII also suggests

that proinflammatory pathways likely unleashed by an aggressive tumor microenvironment may point to an unsuccessful struggle of the host immune system to overcome the tumor growth. Furthermore, signaling of proinflammatory cytokines, such as IFN- $\alpha$ 2, IL-2R $\alpha$ , or IL-16, was reported to be associated with poor risk clinical characteristics and inferior survival in GCT patients (59).

Nilius et al. recently reported that high expression of  $\beta$ -1,4-galactosyltransferase-I (B4GALT1) in peripheral T-lymphocytes is a marker of lower risk of relapse in GCT patients treated with salvage high-dose chemotherapy and peripheral stem cell transplant (HR = 0.66; 95% CI 0.45–0.97;  $P = 0.02$ ) (60). T-cells were collected before the high-dose chemotherapy using the non-myeloablative chemotherapy and granulocyte growth factor (60). B4GALT1 is important for interaction and adhesion of immune cells and its role in disease control in stage I lung cancer has been established (61). This study supported their hypothesis of the importance of activated peripheral T cells in *in vitro* experiments by lectin stimulation of mononuclear cells with Conavalin A. As a result, B4GALT1 was upregulated, particularly in CD4<sup>+</sup> cells and an antiinflammatory cytokine IL10 was significantly expressed. Interestingly, higher levels of IL10 from patient T cells were also associated with better outcome in GCT (60). Activated T cells, thus, seem to play an important role in cancer control.

## Liquid Biopsies and Epigenetic Biomarkers

Sensitive and specific biomarkers indicating the presence of cancer that are assessed from peripheral blood represent an attractive and convenient approach in the diagnosis malignancies. Researchers recently published an array of articles showing that certain clusters of miR are highly informative of the presence of viable cancer in GCT patients (62–70). Serum examination for miR371-373 showed sensitivity of 98–100%, exceeding the sensitivity of the commonly used serum tumor markers AFP and HCG (71, 72). The targeted serum miRNA test (TSmiR) was developed and it seems to be very effective in predicting viable GCT after orchiectomy in clinical stage I patients or after chemotherapy in metastatic disease (72). The clinical utility of the TSmiR test is therefore very promising and clinicians may be expecting this novel biomarker to be implemented in the common practice in the near future (73). One possible utility of these highly sensitive miRNAs seems to be predicting the presence of a microscopic disease in clinical stage I GCTs. As such, these are likely to change the outlook over adjuvant treatment vs. surveillance. Another valuable input would be predicting the presence of viable cancer in post-chemotherapy residual masses, thus refining the need to perform often difficult surgeries in this setting. However, TSmiR does not identify teratoma components which still represent a diagnostic dilemma in the residual disease. Establishing the novel clinical practice stems from our ability to validate the utility of TSmiR in larger prospective cohorts of patients.

Majewski et al. assessed five patients with stage I seminoma and evaluated a possible role of liquid biopsy in identifying the presence of the tumor. The study showed promising



results and identified candidate genes in whole blood prior to orchiectomy. This series is, however, too small to draw any conclusions and a larger study is suggested for validation (74).

A global DNA hypermethylation was proposed as one of the acting mechanisms in cisplatin resistance, the most frustrating challenge for oncologists treating GCT patients. *In vitro* epigenetic studies suggested that treatment with DNA demethylating agents may restore the sensitivity to cisplatin (75–77). In a study by Beyrouthy et al., a GCT cell-line treated with decitabine was resensitized to cisplatin (78). Based on these findings, Albany et al. performed a series of experiments in cell-line culture and patient-derived xenograft mouse model using a second-generation inhibitor of DNA-methyltransferase guadecitabine. Upon treatment of platinum resistant xenografts, a significant growth inhibition and even complete tumor regression was registered (79). An ongoing phase I trial using guadecitabine in combination with cisplatin in refractory GCT will shed more light on clinical significance of these promising findings (NCT02429466).

## REFERENCES

- Cheng L, Albers P, Berney DM, Feldman DR, Daugaard G, Gilligan T, et al. Testicular cancer. *Nat Revs Dis Primers* (2018) 4:29. doi: 10.1038/s41572-018-0029-0
- Albany C, Adra N, Snaveley AC, Cary C, Masterson TA, Foster RS. Multidisciplinary clinic approach improves overall survival outcomes of patients with metastatic germ-cell tumors. *Ann Oncol.* (2018) 29:341–6. doi: 10.1093/annonc/mdx731
- Mardiak J, Sálek T, Sycová-Milá Z, Obertová J, Hlavatá Z, Mego M. Paclitaxel plus ifosfamide and cisplatin in second-line treatment of germ cell tumors: a phase II study. *Neoplasma* (2005) 52:497–501.
- Kondagunta GV, Bacik J, Donadio A, Bajorin D, Marion S, Sheinfeld J. Combination of paclitaxel, ifosfamide, and cisplatin is an effective second-line therapy for patients with relapsed testicular germ cell tumors. *J Clin Oncol.* (2005) 23:6549–55. doi: 10.1200/JCO.2005.19.638
- Adra N, Abonour R, Althouse SK, Albany C, Hanna NH, Einhorn LH. High-dose chemotherapy and autologous peripheral-blood stem-cell transplantation for relapsed metastatic germ cell tumors: the indiana university experience. *J Clin Oncol.* (2017) 35:1096–102. doi: 10.1200/JCO.2016.69.5395
- Warde P, Specht L, Horwich A, Oliver T, Panzarella T, Gospodarowicz M. Prognostic factors for relapse in stage I seminoma managed by surveillance: a pooled analysis. *J Clin Oncol.* (2002) 20:4448–52. doi: 10.1200/JCO.2002.01.038
- Mortensen MS, Lauritsen J, Gundgaard MG, Agerbæk M, Holm NV, Christensen IJ. A nationwide cohort study of stage I seminoma patients followed on a surveillance program. *Eur Urol.* (2014) 66:1172–8. doi: 10.1016/j.eururo.2014.07.001
- Albers P, Siener R, Kliesch S, Weissbach L, Krega S, Sparwasser C. Risk factors for relapse in clinical stage I nonseminomatous testicular germ cell tumors: results of the German testicular cancer study group trial. *J Clin Oncol.* (2003) 21:1505–12. doi: 10.1200/JCO.2003.07.169
- Sweeney CJ, Hermans BP, Heilman DK, Foster RS, Donohue JP, Einhorn LH. Results and outcome of retroperitoneal lymph node dissection for clinical stage I embryonal carcinoma-predominant testis cancer. *J Clin Oncol.* (2000) 18:358–62. doi: 10.1200/JCO.2000.18.2.358
- International Germ Cell Consensus Classification: a prognostic factor-based staging system for metastatic germ cell cancers. International

## CONCLUSION

The investigation for biomarkers in testicular cancer has been insufficient in the past, but with emerging data our knowledge it is built up with an increasing consistency. Such consistency is essential to generate experimental data and perform laboratory research which will ultimately lead to development of novel drugs with a promise to overcome the resistance to cisplatin.

## AUTHOR CONTRIBUTIONS

MC and LC contributed to conception and design. MC drafted the manuscript. CA, MM, RM, and AC contributed critical revision of the manuscript.

## FUNDING

This work was supported by the Slovak Research and Development Agency under contract No. APVV-15-0086 and Scientific Grant Agency under contract number VEGA 1/0043/18 for MC.

- germ cell cancer collaborative group. *J Clin Oncol.* (1997) 15:594–603. doi: 10.1200/JCO.1997.15.2.594
- Necchi A, Pond GR, Nicolai N, Giannatempo P, Raggi D, Adra N. A suggested prognostic reclassification of intermediate and poor-risk nonseminomatous germ cell tumors. *Clin Genitourin Cancer* (2017) 15:306–312 e303. doi: 10.1016/j.clgc.2016.07.022
- Ku K, Ibrahim S, Adra N, Althouse S, Hanna NH, Einhorn LH, et al. A retrospective analysis of patients with metastatic germ cell tumor (GCT) treated at Indiana University (IU) from 2000 to 2012. *J Clin Oncol.* (2015) 33:4539. doi: 10.1200/jco.2015.33.15
- Cheng L, Lopez-Beltran A, Massari F, MacLennan GT, Montironi R. Molecular testing for BRAF mutations to inform melanoma treatment decisions: a move toward precision medicine. *Mod Pathol.* (2018) 31:24–38. doi: 10.1038/modpathol.2017.104
- Cheng L, Alexander RE, MacLennan GT, Cummings OW, Montironi R, Lopez-Beltran A. Molecular pathology of lung cancer: key to personalized medicine. *Mod Pathol.* (2012) 25:347–69. doi: 10.1038/modpathol.2011.215
- Shen H, Shih J, Hollern DP, Wang L, Bowlby R, Tickoo SK. Integrated molecular characterization of testicular germ cell tumors. *Cell Rep.* (2018) 23:3392–406. doi: 10.1016/j.celrep.2018.05.039
- McIntyre A, Summersgill B, Grygalewicz B, Gillis AJ, Stoop J, van Gurp RJ. Amplification and overexpression of the KIT gene is associated with progression in the seminoma subtype of testicular germ cell tumors of adolescents and adults. *Cancer Res.* (2005) 65:8085–9. doi: 10.1158/0008-5472.CAN-05-0471
- Kemmer K, Corless CL, Fletcher JA, McGreevey L, Haley A, Griffith D. KIT mutations are common in testicular seminomas. *Am J Pathol.* (2004) 164:305–13. doi: 10.1016/S0002-9440(10)63120-3
- Litchfield K, Summersgill B, Yost S, Sultana R, Labreche K, Dudakia D. Whole-exome sequencing reveals the mutational spectrum of testicular germ cell tumours. *Nat Commun.* (2015) 6:5973. doi: 10.1038/ncomms6973
- Cheng L, Lyu B, Roth LM. Perspectives on testicular germ cell neoplasms. *Hum Pathol.* (2017) 59:10–25. doi: 10.1016/j.humpath.2016.08.002
- Litchfield K, Levy M, Orlando G, Loveday C, Law PJ, Migliorini G. Identification of 19 new risk loci and potential regulatory mechanisms influencing susceptibility to testicular germ cell tumor. *Nat Genet.* (2017) 49:1133–40. doi: 10.1038/ng.3896
- Liu J, Shi H, Li X, Chen G, Larsson C, Lui WO. miR2233p regulates cell growth and apoptosis via FBXW7 suggesting an oncogenic role in human testicular

- germ cell tumors. *Int J Oncol.* (2017) 50:356–64. doi: 10.3892/ijo.2016.3807
22. Kremensky M, Kremenska Y, Ohgane J, Hattori N, Tanaka S, Hashizume K. Genome-wide analysis of DNA methylation status of CpG islands in embryoid bodies, teratomas, and fetuses. *Biochem Biophys Res Commun.* (2003) 311:884–90. doi: 10.1016/j.bbrc.2003.10.078
  23. Fukushima S, Yamashita S, Kobayashi H, Takami H, Fukuoka K, Nakamura T. Genome-wide methylation profiles in primary intracranial germ cell tumors indicate a primordial germ cell origin for germinomas. *Acta Neuropathol.* (2017) 133:445–62. doi: 10.1007/s00401-017-1673-2
  24. Cheng L, Sung MT, Cossu-Rocca P, Jones TD, MacLennan GT, De Jong J. OCT4: biological functions and clinical applications as a marker of germ cell neoplasia. *J Pathol.* (2007) 211:1–9. doi: 10.1002/path.2105
  25. Cheng L. Establishing a germ cell origin for metastatic tumors using OCT4 immunohistochemistry. *Cancer* (2004) 101:2006–10. doi: 10.1002/cncr.20566
  26. Looijenga LH, Stoop H, de Leeuw HP, de Gouveia Brazao CA, Gillis AJ, van Roozendaal KE. POU5F1 (OCT3/4) identifies cells with pluripotent potential in human germ cell tumors. *Cancer Res.* (2003) 63:2244–50.
  27. Chabot B, Stephenson DA, Chapman VM, Besmer P, Bernstein A. The proto-oncogene c-kit encoding a transmembrane tyrosine kinase receptor maps to the mouse W locus. *Nature* (1988) 335:88–9. doi: 10.1038/335088a0
  28. Huang E, Nock K, Beier DR, Chu TY, Buck J, Lahm HW. The hematopoietic growth factor KL is encoded by the Sl locus and is the ligand of the c-kit receptor, the gene product of the W locus. *Cell* (1990) 63:225–33. doi: 10.1016/0092-8674(90)90303-V
  29. Yarden Y, Kuang WJ, Yang-Feng T, Coussens L, Munemitsu S, Dull TJ. Human proto-oncogene c-kit: a new cell surface receptor tyrosine kinase for an unidentified ligand. *EMBO J.* (1987) 6:3341–51. doi: 10.1002/j.1460-2075.1987.tb02655.x
  30. Mirabello L, Kratz CP, Savage SA, Greene MH. Promoter methylation of candidate genes associated with familial testicular cancer. *Int J Mol Epidemiol Genet.* (2012) 3:213–27.
  31. Freberg CT, Dahl JA, Timoskainen S, Collas P. Epigenetic reprogramming of OCT4 and NANOG regulatory regions by embryonal carcinoma cell extract. *Mol Biol Cell* (2007) 18:1543–53. doi: 10.1091/mbc.e07-01-0029
  32. Almstrup K, Nielsen JE, Mlynarska O, Jansen MT, Jørgensen A, Skakkebaek NE. Carcinoma in situ testis displays permissive chromatin modifications similar to immature foetal germ cells. *Br J Cancer* (2010) 103:1269–76. doi: 10.1038/sj.bjc.6605880
  33. Nettersheim D, Biermann K, Gillis AJ, Steger K, Looijenga LH, Schorle H. NANOG promoter methylation and expression correlation during normal and malignant human germ cell development. *Epigenetics* (2011) 6:114–22. doi: 10.4161/epi.6.1.13433
  34. Villasante A, Piazzolla D, Li H, Gomez-Lopez G, Djabali M, Serrano M. Epigenetic regulation of Nanog expression by Ezh2 in pluripotent stem cells. *Cell Cycle* (2011) 10:1488–98. doi: 10.4161/cc.10.9.15658
  35. Adra N, Althouse SK, Liu H, Brames MJ, Hanna NH, Einhorn LH. Prognostic factors in patients with poor-risk germ-cell tumors: a retrospective analysis of the Indiana University experience from 1990 to 2014. *Ann Oncol.* (2016) 27:875–9. doi: 10.1093/annonc/mdw045
  36. Mazumdar M, Bajorin DE, Bacik J, Higgins G, Motzer RJ, Bosl GJ. Predicting outcome to chemotherapy in patients with germ cell tumors: the value of the rate of decline of human chorionic gonadotrophin and alpha-fetoprotein during therapy. *J Clin Oncol.* (2001) 19:2534–41. doi: 10.1200/JCO.2001.19.9.2534
  37. Fizazi K, Culine S, Kramar A, Amato RJ, Bouzy J, Chen I. Early predicted time to normalization of tumor markers predicts outcome in poor-prognosis nonseminomatous germ cell tumors. *J Clin Oncol.* (2004) 22:3868–76. doi: 10.1200/JCO.2004.04.008
  38. Motzer RJ, Nichols CJ, Margolin KA, Bacik J, Richardson PG, Vogelzang NJ. Phase III randomized trial of conventional-dose chemotherapy with or without high-dose chemotherapy and autologous hematopoietic stem-cell rescue as first-line treatment for patients with poor-prognosis metastatic germ cell tumors. *J Clin Oncol.* (2007) 25:247–56. doi: 10.1200/JCO.2005.05.4528
  39. Mego M, Rejlekova K, Reckova M, Sycova-Mila Z, Obertova J, Rajec J. Kinetics of tumor marker decline as an independent prognostic factor in patients with relapsed metastatic germ-cell tumors. *Neoplasma* (2009) 56:398–403. doi: 10.4149/neo\_2009\_05\_398
  40. Massard C, Kramar A, Beyer J, Hartmann JT, Lorch A, Pico JL. Tumor marker kinetics predict outcome in patients with relapsed disseminated non-seminomatous germ-cell tumors. *Ann Oncol.* (2013) 24:322–8. doi: 10.1093/annonc/mds504
  41. Murphy BA, Motzer RJ, Mazumdar M, Vlamis V, Nisselbaum J, Bajorin D. Serum tumor marker decline is an early predictor of treatment outcome in germ cell tumor patients treated with cisplatin and ifosfamide salvage chemotherapy. *Cancer* (1994) 73:2520–6.
  42. Fizazi K, Pagliaro L, Laplanche A, Fléchon A, Mardiak J, Geoffrois L. Personalised chemotherapy based on tumour marker decline in poor prognosis germ-cell tumours (GETUG 13): a phase 3, multicentre, randomised trial. *Lancet Oncol.* (2014) 15:1442–50. doi: 10.1016/S1470-2045(14)70490-5
  43. Fizazi K, Fléchon A, Teuff GL, Mardiak J, Pagliaro LC, Geoffrois L, et al. Mature results of the GETUG 13 phase III trial in poor-prognosis germ-cell tumors (GCT). *J Clin Oncol.* (2016) 34:4504. doi: 10.1200/JCO.2016.34.15\_suppl.4504
  44. Loriot Y, Pagliaro L, Fléchon A, Mardiak J, Geoffrois L, Kerbrat P. Patterns of relapse in poor-prognosis germ-cell tumours in the GETUG 13 trial: Implications for assessment of brain metastases. *Eur J Cancer* (2017) 87:140–6. doi: 10.1016/j.ejca.2017.09.029
  45. Mego M, Cierna Z, Svetlovská D, Macak D, Machalekova K, Miskovska V. PARP expression in germ cell tumours. *J Clin Pathol.* (2013) 66:607–12. doi: 10.1136/jclinpath-2012-201088
  46. Kalavská K, Cierna Z, Chovanec M, Takacova M, Svetlovská D, Miskovska V. Prognostic value of intratumoral carbonic anhydrase IX expression in testicular germ cell tumors. *Oncol Lett.* (2017) 13:2177–85. doi: 10.3892/ol.2017.5745
  47. Kalavská K, Chovanec M, Zatošvicova M, Takacova M, Gronesova P, Svetlovská D. Prognostic value of serum carbonic anhydrase IX in testicular germ cell tumor patients. *Oncol Lett.* (2016) 12:2590–8. doi: 10.3892/ol.2016.5010
  48. Scheri KC, Leonetti E, Laino L, Gigantino V, Gesualdi L, Grammatico P. c-MET receptor as potential biomarker and target molecule for malignant testicular germ cell tumors. *Oncotarget* (2018) 9:31842–31860. doi: 10.18632/oncotarget.25867
  49. Fankhauser CD, Curioni-Fontecedro A, Allmann V, Beyer J, Tischler V, Sulser T. Frequent PD-L1 expression in testicular germ cell tumors. *Br J Cancer* (2015) 113:411–3. doi: 10.1038/bjc.2015.244
  50. Cierna Z, Mego M, Miskovska V, Machalekova K, Chovanec M, Svetlovská D. Prognostic value of programmed-death-1 receptor (PD-1) and its ligand 1 (PD-L1) in testicular germ cell tumors. *Ann Oncol.* (2016) 27:300–5. doi: 10.1093/annonc/mdv574
  51. Chovanec M, Cierna Z, Miskovska V, Machalekova K, Svetlovská D, Kalavská K. Prognostic role of programmed-death ligand 1 (PD-L1) expressing tumor infiltrating lymphocytes in testicular germ cell tumors. *Oncotarget* (2017) 8:21794–805. doi: 10.18632/oncotarget.15585
  52. Bols B, Jensen L, Jensen A, Braendstrup O. Immunopathology of in situ seminoma. *Int J Exp Pathol.* (2000) 81:211–7. doi: 10.1046/j.1365-2613.2000.00151.x
  53. Adra N, Einhorn LH, Althouse SK, Ammakannavar NR, Musapatika D, Albany C. Phase II trial of pembrolizumab in patients with platinum refractory germ cell tumors: a hoosier cancer research network study GU14–206. *Ann Oncol.* (2017) 29:209–14. doi: 10.1093/annonc/mdx680
  54. Zschäbitz S, Lasitschka F, Hadaschik B, Hofheinz RD, Jentsch-Ullrich K, Grüner M. Response to anti-programmed cell death protein-1 antibodies in men treated for platinum refractory germ cell cancer relapsed after high-dose chemotherapy and stem cell transplantation. *Eur J Cancer* (2017) 76:1–7. doi: 10.1016/j.ejca.2017.01.033
  55. Zschäbitz S, Lasitschka F, Jäger D, Grulich C. Activity of immune checkpoint inhibition in platinum refractory germ-cell tumors. *Ann Oncol.* (2016) 27:1356–60. doi: 10.1093/annonc/mdw146
  56. Shah S, Ward JE, Bao R, Hall CR, Brockstein BE, Luke JJ. Clinical response to anti-PD1 immunotherapy in a patient with non-seminomatous germ cell tumor and evaluation of the immune landscape in testicular cancer. *J Clin Oncol.* (2016) 34:e16040. doi: 10.1200/JCO.2016.34.15\_suppl.e16040
  57. Chovanec M, Cierna Z, Miskovska V, Machalekova K, Kalavská K, Rejlekova K, et al. Systemic immune-inflammation index in germ-cell tumours. *Br J Cancer* (2018) 118:831–8. doi: 10.1038/bjc.2017.460

58. Fankhauser CD, Sander S, Roth L, Gross O, Eberli D, Sulser T. Systemic inflammatory markers have independent prognostic value in patients with metastatic testicular germ cell tumours undergoing first-line chemotherapy. *Br J Cancer* (2018) 118:825–30. doi: 10.1038/bjc.2017.467
59. Svetlovska D, Miskovska V, Cholujova D, Gronesova P, Cingelova S, Chovanec M. Plasma cytokines correlated with disease characteristics, progression-free survival, and overall survival in testicular germ-cell tumor patients. *Clin Genitourin Cancer* (2017) 15:411–416 e412. doi: 10.1016/j.clgc.2017.01.027
60. Nilius V, Killer MC, Timmesfeld N, Schmitt M, Moll R, Lorch A. High beta-1,4-Galactosyltransferase-I expression in peripheral T-lymphocytes is associated with a low risk of relapse in germ-cell cancer patients receiving high-dose chemotherapy with autologous stem cell reinfusion. *Oncoimmunology* (2018) 7:e1423169. doi: 10.1080/2162402X.2017.1423169
61. Lu Y, Wang L, Liu P, Yang P, You M. Gene-expression signature predicts postoperative recurrence in stage I non-small cell lung cancer patients. *PLoS ONE* (2012) 7:e30880. doi: 10.1371/journal.pone.0030880
62. Voorhoeve PM, le Sage C, Schrier M, Gillis AJ, Stoop H, Nagel R. A genetic screen implicates miRNA-372 and miRNA-373 as oncogenes in testicular germ cell tumors. *Adv Exp Med Biol.* (2007) 604:17–46. doi: 10.1007/978-0-387-69116-9\_2
63. Cheung HH, Davis AJ, Lee TL, Pang AL, Nagrani S, Rennert OM. Methylation of an intronic region regulates miR-199a in testicular tumor malignancy. *Oncogene* (2011) 30:3404–15. doi: 10.1038/onc.2011.60
64. Dieckmann KP, Spiekermann M, Balks T, Flor I, Löning T, Bullerdiek J. MicroRNAs miR-371–3 in serum as diagnostic tools in the management of testicular germ cell tumours. *Br J Cancer* (2012) 107:1754–60. doi: 10.1038/bjc.2012.469
65. Syring I, Bartels J, Holdenrieder S, Kristiansen G, Müller SC, Ellinger J. Circulating serum miRNA (miR-367–3p, miR-371a–3p, miR-372–3p and miR-373–3p) as biomarkers in patients with testicular germ cell cancer. *J Urol* (2015) 193:331–7. doi: 10.1016/j.juro.2014.07.010
66. Spiekermann M, Belge G, Winter N, Ikogho R, Balks T, Bullerdiek J. MicroRNA miR-371a–3p in serum of patients with germ cell tumours: evaluations for establishing a serum biomarker. *Andrology* (2015) 3:78–84. doi: 10.1111/j.2047-2927.2014.00269.x
67. Dieckmann KP, Spiekermann M, Balks T, Ikogho R, Anheuser P, Wosniok W. MicroRNA miR-371a–3p - a novel serum biomarker of testicular germ cell tumors: evidence for specificity from measurements in testicular vein blood and in neoplastic hydrocele fluid. *Urol Int.* (2016) 97:76–83. doi: 10.1159/000444303
68. Dieckmann KP, Radtke A, Spiekermann M, Balks T, Matthies C, Becker P, et al. Serum levels of MicroRNA miR-371a–3p: a sensitive and specific new biomarker for germ cell tumours. *Eur Urol.* (2017) 71:213–20. doi: 10.1016/j.eururo.2016.07.029
69. van Agthoven T, Eijkenboom WMH, Looijenga LHJ. microRNA-371a–3p as informative biomarker for the follow-up of testicular germ cell cancer patients. *Cell Oncol.* (2017) 40:379–88. doi: 10.1007/s13402-017-0333-9
70. Flor I, Spiekermann M, Löning T, Dieckmann KP, Belge G, Bullerdiek J. Expression of microRNAs of C19MC in different histological types of testicular germ cell tumour. *Cancer Genom Proteom* (2016) 13:281–9.
71. Gillis AJ, Rijlaarsdam MA, Eini R, Dorssers LC, Biermann K, Murray MJ. Targeted serum miRNA (TSmiR) test for diagnosis and follow-up of (testicular) germ cell cancer patients: a proof of principle. *Mol Oncol.* (2013) 7:1083–92. doi: 10.1016/j.molonc.2013.08.002
72. Nazario Leao RR, van Agthoven T, Figueiredo A, Jewett MAS, Fadaak K, Sweet J, et al. Serum miRNA to predict post-chemotherapy viable disease in testicular non-seminomatous germ cell tumor patients. *J Clin Oncol.* (2018) 36:abstr 546. doi: 10.1200/JCO.2018.36.6
73. Belge G, Dieckmann KP, Spiekermann M, Balks T, Bullerdiek J. Serum levels of microRNAs miR-371–3: a novel class of serum biomarkers for testicular germ cell tumors? *Eur Urol.* (2012) 61:1068–9. doi: 10.1016/j.eururo.2012.02.037
74. Majewski M, Nestler T, Kagler S, Richardsen I, Ruf CG, Matthies C, et al. Liquid biopsy using whole blood from testis tumor and colon cancer patients-a new and simple way? *Health Phys.* (2018) 115:114–20. doi: 10.1097/HP.0000000000000867
75. Koul S, Houldsworth J, Mansukhani MM, Donadio A, McKiernan JM, Reuter VE. Characteristic promoter hypermethylation signatures in male germ cell tumors. *Mol Cancer* (2002) 1:8. doi: 10.1186/1476-4598-1-8
76. Koul S, McKiernan JM, Narayan G, Houldsworth J, Bacik J, Dobrzynski DL. Role of promoter hypermethylation in Cisplatin treatment response of male germ cell tumors. *Mol Cancer* (2004) 3:16. doi: 10.1186/1476-4598-3-16
77. Honorio S, Agathangelou A, Wernert N, Rothe M, Maher ER, Latif F. Frequent epigenetic inactivation of the RASSF1A tumour suppressor gene in testicular tumours and distinct methylation profiles of seminoma and nonseminoma testicular germ cell tumours. *Oncogene* (2003) 22:461–6. doi: 10.1038/sj.onc.1206119
78. Beyrouthy MJ, Garner KM, Hever MP, Freemantle SJ, Eastman A, Dmitrovsky E. High DNA methyltransferase 3B expression mediates 5-aza-deoxycytidine hypersensitivity in testicular germ cell tumors. *Cancer Res.* (2009) 69:9360–6. doi: 10.1158/0008-5472.CAN-09-1490
79. Albany C, Hever-Jardine MP, von Herrmann KM, Yim CY, Tam J, Warzecha JM. Refractory testicular germ cell tumors are highly sensitive to the second generation DNA methylation inhibitor guadecitabine. *Oncotarget* (2017) 8:2949–59. doi: 10.18632/oncotarget.13811

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Chovanec, Albany, Mego, Montironi, Cimadamore and Cheng. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.



# Emerging Biomarkers in Bladder Cancer Identified by Network Analysis of Transcriptomic Data

Matteo Giulietti<sup>1\*</sup>, Giulia Occhipinti<sup>1</sup>, Alessandra Righetti<sup>1</sup>, Massimo Bracci<sup>2</sup>, Alessandro Conti<sup>3</sup>, Annamaria Ruzzo<sup>4</sup>, Elisabetta Cerigioni<sup>5</sup>, Tiziana Cacciamani<sup>6</sup>, Giovanni Principato<sup>1</sup> and Francesco Piva<sup>1</sup>

<sup>1</sup> Department of Specialistic Clinical and Odontostomatological Sciences, Polytechnic University of Marche, Ancona, Italy, <sup>2</sup> Department of Clinical and Molecular Sciences, Polytechnic University of Marche, Ancona, Italy, <sup>3</sup> Department of Urology, Bressanone/Brixen Hospital, Bressanone, Italy, <sup>4</sup> Department of Biomolecular Sciences, University of Urbino "Carlo Bo", Fano, Italy, <sup>5</sup> Unit of Pediatric and Specialistic Surgery, United Hospitals, "G.Salesi", Ancona, Italy, <sup>6</sup> Department of Life and Environmental Science, Polytechnic University of Marche, Ancona, Italy

## OPEN ACCESS

### Edited by:

Matteo Santoni,  
Polytechnic University of Marche, Italy

### Reviewed by:

Simona Di Francesco,  
Independent Researcher, Chieti, Italy  
Francesco Massari,  
Azienda Ospedaliera Universitaria  
Integrata Verona, Italy

### \*Correspondence:

Matteo Giulietti  
m.giulietti@univpm.it

### Specialty section:

This article was submitted to  
Genitourinary Oncology,  
a section of the journal  
Frontiers in Oncology

**Received:** 22 June 2018

**Accepted:** 25 September 2018

**Published:** 12 October 2018

### Citation:

Giulietti M, Occhipinti G, Righetti A,  
Bracci M, Conti A, Ruzzo A,  
Cerigioni E, Cacciamani T,  
Principato G and Piva F (2018)  
Emerging Biomarkers in Bladder  
Cancer Identified by Network Analysis  
of Transcriptomic Data.  
Front. Oncol. 8:450.  
doi: 10.3389/fonc.2018.00450

Bladder cancer is a very common malignancy. Although new treatment strategies have been developed, the identification of new therapeutic targets and reliable diagnostic/prognostic biomarkers for bladder cancer remains a priority. Generally, they are found among differentially expressed genes between patients and healthy subjects or among patients with different tumor stages. However, the classical approach includes processing these data taking into consideration only the expression of each single gene regardless of the expression of other genes. These complex gene interaction networks can be revealed by a recently developed systems biology approach called Weighted Gene Co-expression Network Analysis (WGCNA). It takes into account the expression of all genes assessed in an experiment in order to reveal the clusters of co-expressed genes (modules) that, very probably, are also co-regulated. If some genes are co-expressed in controls but not in pathological samples, it can be hypothesized that a regulatory mechanism was altered and that it could be the cause or the effect of the disease. Therefore, genes within these modules could play a role in cancer and thus be considered as potential therapeutic targets or diagnostic/prognostic biomarkers. Here, we have reviewed all the studies where WGCNA has been applied to gene expression data from bladder cancer patients. We have shown the importance of this new approach in identifying candidate biomarkers and therapeutic targets. They include both genes and miRNAs and some of them have already been identified in the literature to have a role in bladder cancer initiation, progression, metastasis, and patient survival.

**Keywords:** WGCNA, bladder cancer, tumor biomarkers, gene expression, heterogeneity

## INTRODUCTION

Bladder cancer (BCa) is the ninth most prevalent malignant disease globally, with more than 400,000 new cases diagnosed each year, especially in males and elderly, and the thirteenth most common cause of cancer death. Five-year survival rate for early stage BCa patients reaches 95.7%, while for metastatic patients it is just 5% (1). Recent advancements in therapy include immunotherapy with PD-1 antibody pembrolizumab and the PD-L1 antibody atezolizumab



which have yielded better results compared to chemotherapy (2). The most common BCa type is transitional cell carcinoma, also called urothelial carcinoma, while squamous cell carcinoma and adenocarcinomas are rare. For the majority of new BCa patients, non-muscle-invasive BCa (NMIBC) is diagnosed. This low-grade tumor often recurs and in about 20% of cases it progresses to high grade, i.e., to the muscle-invasive BCa (MIBC) which is more likely to develop metastases (3). In addition, molecular data, including chromosomal aberrations, mutation rates, presence of mutated tumor suppressor genes, gene, and miRNA expression levels have led to the definition of specific molecular subtypes. This depicts BCa as a molecularly and clinicopathologically heterogeneous disease. Diagnosis can be based on cystoscopy, microscopy, voided urinary cytology, blood detection in the urine and assessment of urine-based tumor markers, such as complement factor H-related protein (BTA test) and nuclear matrix protein 22 (NMP22) (4). However, the sensitivity and specificity of these markers decrease in the presence of inflammatory cells and other contaminating cells in the sample (5). Therefore, the identification of new biomarkers could improve diagnostic or prognostic performance of BCa tests. Also small extracellular vesicles released by BCa are currently being investigated, since they are known to be involved in cancer growth, progression and metastatic spread and the molecules contained in these vesicles may be potential BCa biomarkers (6, 7).

An effective method to discover biomarkers is transcriptome profiling, by microarrays or more recently by RNA-seq, that allows the determination of differentially expressed genes and non-coding RNAs under different conditions. For example, the comparison of the expression levels of thousands of genes in healthy vs. cancer tissue samples, or among different tumor stages or tumors under different treatments, has led to the identification of several differentially expressed genes. They could represent candidate therapeutic targets or biomarkers for tumor onset, progression, or prognosis. However, over the past few years, a major drawback has emerged regarding differential expression analysis. In particular, differentially expressed genes are treated individually and this does not allow the identification of co-regulation mechanisms among them. Highlighting these gene interactions is important, because they are often altered in complex genetic disorders like cancer (8–10). This emerging systems biology method focuses on the analysis of gene regulation alterations allowing a better understanding of cancer onset and its progression and the identification of critical cancer driver genes. Moreover, these clusters of co-expressed genes, that constitute the regions of a complex gene network, often correspond to cellular pathways (8–10).

Currently, a widely used approach to process gene expression data and investigate network alterations is the Weighted Gene Co-expression Network Analysis (WGCNA), that draws gene networks where the connections among pairs of genes are identified and weighted based on their correlated expression levels across multiple samples (11). Briefly, after processing the expression profiles into weighted connections, WGCNA can identify the network topology and, by using the topological overlap dissimilarity as the measure of distance among genes,

it allows the identification of sub-networks, called modules [for further details see (12)]. Therefore, only highly co-expressed genes (i.e., connected with strong weights in the network) can compose the gene modules. It is also possible to relate these modules to clinical traits of interest. For example, regarding the comparison of two distinct networks deriving from tumor and normal gene expression data, WGCNA can identify the modules and genes belonging to them that reflect the regulatory alterations related to transcriptional changes. In particular, the most interconnected genes in a module (hub genes) are often functionally important and thus they could play a key role in cancer and represent candidate diagnostic and prognostic biomarkers or potential therapeutic targets (13–15).

## EMERGING BCA BIOMARKERS IDENTIFIED BY THE WGCNA METHOD

In this review, we focused on all studies where the WGCNA method was applied to analyze gene expression data deriving from BCa samples (Table 1).

Recently, WGCNA was applied on publicly available microarray gene expression data deriving from 93 BCa patients in order to identify genes related to different tumor stages (Ta, T1, T2, T3, and T4) and therefore to suggest potential prognostic biomarkers (16). A network module was highly correlated with tumor progression and further processing was performed in order to identify which cellular pathways were represented by the module genes. In general, since not all genes of a pathway are present in a module, the term “enrichment” is used to indicate how significantly a pathway overlaps with a module. A functional enrichment analysis is usually performed on a list of genes in order to reveal these enriched pathways. The identified module was significantly enriched in genes belonging to important biological processes (Table 1). Within this module, four hub genes were identified: COL3A1, COL5A2, FBN1, and POSTN and, by using independent expression datasets, the COL3A1 (Collagen type III  $\alpha 1$  chain) gene was validated as both a diagnostic and prognostic biomarker. In fact, it was upregulated in BCa tissues compared to normal ones and its high expression strongly correlated with tumor progression, shorter overall survival (OS) and disease-free survival (DFS) times. While there are few literature data about COL5A2 and FBN1 genes, the role of POSTN (periostin) in BCa has been widely investigated. In particular, high grade BCa showed very low levels of POSTN gene and the artificial restoration of its expression suppressed cell invasiveness and metastasis (22). It was also shown that the decrease of invasiveness and the inhibition of the epithelial-to-mesenchymal transition (EMT) were due to bladder-specific upregulation of the E-cadherin expression by periostin (23).

Lately, microarray gene expression data of 165 primary BCa samples were analyzed by WGCNA in order to identify genes correlated with TNM staging and OS (17). In total, 11 modules correlated with TNM staging and they were enriched in genes belonging to cell proliferation associated pathways (Table 1). A filtering step using protein-protein interaction

**TABLE 1** | Hub genes and miRNAs detected by different authors using WGCNA method.

Comparison	Number and source of samples	Expression data source	Genes	Involved pathways	References
Different BCa stages	93 BCa tissue samples	NCBI GEO microarray dataset GSE31684	COL3A1, COL5A2, FBN1, POSTN	Extracellular matrix (ECM) organization, ECM-receptor interaction, regulation of actin cytoskeleton, cell adhesion, focal adhesion	(16)
Different BCa stages	165 primary BCa tissue samples	NCBI GEO microarray dataset GSE13507	AURKB, BUB1B, CCNB2, CDC45, CENPA, CEP55, KIF2C, KIF4A, KIF15, NUSAP1, PRC1, UBE2C (only diagnostic); AEBP1, CDC25B, COL5A2, MMP11, TK1, TPX2 (only prognostic); CDCA3, CDCA8, CENPF, FOXM1, TOP2A (both diagnostic and prognostic)	Cell cycle, nuclear division, chromosome segregation, organelle fission	(17)
SCCB vs. normal	75 tumor cells and 18 normal cells from the tissue of one patient	NCBI SRA single cell RNA-seq SRP078083	ARHGAP15, BCAR3, CACNA2D3, CENPH, CTNND2, DOHH, GCC2, HERC2, LINC00189, NLK, OR9Q1, PCSK6, POU2F3, SCN2A, TUBGCP2	Spliceosome complex, VEGF, MAPK, neurotrophic signaling, and cell cycle pathways	(18)
BCa vs. normal	9 BCa and 9 normal tissue samples *	NCBI GEO microarray dataset GSE3167	ATF7, CER1, CYP1A2, GDF9, KCNIP1, LRRC15, PTPRJ, TRPM3	Response to stimulus, regulation of localization, gamma-hexachlorocyclohexane degradation, fatty acid metabolism, adherens junction, tryptophan metabolism, and Wnt signaling pathways	(19)
NMIBC vs. MIBC	8 NMIBC and 11 MIBC tissues*	NCBI GEO microarray dataset GSE37317	ATP2A2, BCAP31, BRD2, CYP3A5, DCAF8, LRRC37A2, MEIS3P1, POLR2A, PURA, SRPK2, TRAK1, UBE2I, UPF3A, VPS13D, WDFY3, ZZEF1	Regulation of mitotic cell cycle, cell cycle phase, organelle organization, negative regulation of programmed cell death, DNA replication, DNA recombination, mRNA splicing, cellular localization, B cell receptor signaling pathway, Ras pathway	(20)
BCa vs. normal	418 BCa and 19 normal tissue samples	TCGA miRNA-Seq dataset BLCA	miR-1-1, miR-1-2, miR-28, miR-133a-1, miR-133a-2, miR-133b, miR-139, miR-143, miR-145, miR-195, miR-548ba, miR-3199-2, miR-6507	Cell proliferation, regulation of cell growth, regulation of actin cytoskeleton, proteoglycans in cancer, focal adhesion, Wnt, PI3K-Akt, MAPK, and p53 signaling pathways	(21)

BCa, bladder cancer; SCCB, squamous cell carcinoma of bladder; MIBC, muscle-invasive BCa; NMIBC, non-muscle-invasive BCa. \*Note that WGCNA has been performed using too few samples, indeed at least 20 samples are required for a robust WGCNA analysis and to obtain reliable results.

network information collected in STRING tool (<https://string-db.org/>) resulted in 17 genes with a potential role in BCa (Table 1). Moreover, 11 hub genes could be considered as prognostic biomarkers, since their lower expression was associated with better OS of BCa patients. Some identified hub genes had been previously investigated. In particular, the high expression of AURKB (Aurora Kinase B), implicated in cancer through development of aneuploidy and chromosomal instability, correlated with advanced BCa stages (24). Similarly, the mitotic checkpoint protein BUB1B, known to contribute to chromosomal instability, was over-expressed in advanced BCa stages and correlated with high cell proliferation (25). High expression levels of CDC25B (Cell division cycle 25B) were associated with advanced stages, recurrence and poor prognosis in BCa patients (26). Cyclin B2 (CCNB2) expression level was higher in cancer than in normal bladder mucosa

and its downregulation inhibited cell migration, invasion, and metastatic abilities (27). Also the role of FOXM1 (Forkhead box M1) has been widely investigated, indeed it was found to be a reliable prognostic biomarker for MIBC (28) and its high expression level correlated with TNM stage, histological grade, metastases, and poor prognosis in BCa patients, whereas its down-regulation through miR-24-1 inhibited cell proliferation, migration, and invasion (29, 30). Since CCNB2 and FOXM1 suppression resulted in BCa inhibition, they can be considered as reliable therapeutic targets and deserve further exploration. Moreover, MMP11 (Matrix metalloproteinase-11) overexpression correlated with very aggressive phenotypes (advanced pT status, nodal metastasis, high histological grade) and with unfavorable clinical outcomes (31). Serum concentration of TK1 (Thymidine kinase 1) gene was associated with tumor stage, degree of invasion, and metastasis (32). TOP2A

(Topoisomerase-IIA) free DNA in urine has been identified as a diagnostic biomarker and its levels can also distinguish NMIBC from MIBC (33). Its over-expression has also been associated with high-grade and high-stage BCa and with high rates of recurrence in NMIBC (34). Moreover, TOP2A protein levels have been identified as a predictor of DFS times (35). TPX2 (microtubule nucleation factor) gene was upregulated in tumor tissues compared to normal bladder samples and it was strongly associated with pT status, high histological grade, lymph node metastasis, and shorter OS time. Indeed its overexpression promoted proliferation and tumorigenicity and suppressed apoptosis (36). UBE2C (Ubiquitin conjugating enzyme E2) upregulation was associated with high BCa stages, presence of lymphovascular invasion, progression to MIBC, and it has been suggested as a biomarker of unfavorable prognosis (37, 38).

In an experiment where single-cell transcriptomics was applied to squamous cell carcinoma of urinary bladder (SCCB), 75 tumor cells and 18 normal cells were isolated from cancer and normal control fresh resected tissues from one patient (18). Then, gene expression analysis was performed by single-cell RNA-seq. WGCNA analysis identified five large modules enriched in genes belonging to several cancer-associated pathways (Table 1). While some of the identified hub genes (Table 1) have rarely been reported in previous cancer studies, some of them have already been suggested as BCa biomarkers. For example, CTNND2 (Catenin delta 2) gene is frequently amplified in BCa, with high copy numbers (39). Moreover, copy number variations of PCSK6 (Proprotein convertase subtilisin/kexin type 6) gene have been reported to be a prognostic marker for NMIBC progression (40). Intriguingly, POU2F3 (POU class 2 homeobox 3) is a transcription factor expressed in stratified squamous epithelia and related to squamous epithelial stratification (41), so it could play a role in squamous cell BCa (SCCB).

Microarray gene expression profiles of 9 normal bladder and 9 transitional cell carcinoma tissue samples have been analyzed by the WGCNA method (19). A differential co-expression network analysis was carried out and eight hub genes were identified (Table 1). Interestingly, among the identified hub genes, the rs762551 polymorphism in CYP1A2 (a cytochrome P450 family member) gene was associated with decreased BCa risk (42). Molecular mechanisms explaining this association are still undetermined, however since it lies in the first intron of the gene, it could alter pre-mRNA splicing processing at 5'UTR level, transcription regulation, or protein folding (43–51).

Recently, different gene expression profiles between NMIBC and MIBC have been investigated using public microarray expression data from 8 and 11 snap frozen cancer tissues, respectively (20). WGCNA analysis showed significant correlations between three modules and the tumor stage. In particular, these modules were enriched in genes mainly involved in cell cycle (Table 1). Among them, 16 hub genes have been identified (Table 1). Therefore, they can be involved in BCa progression from NMIBC to MIBC phenotype. Many hub genes have been previously suggested as candidate diagnostic or prognostic BCa biomarkers. For example, TRAK1 (trafficking kinesin protein 1) gene has been identified as a favorable prognostic marker, since its low level expression was associated

with poorer survival (52). Polymorphisms in CYP3A5 (a cytochrome P450 family member) can define a subset of BCa patients who better respond to cabazitaxel and temsirolimus, in terms of lower toxicity and higher efficacy (53, 54).

However, regarding the last two studies (19, 20), it should be noted that WGCNA was performed using sample sizes that were too small, that is 8, 9, or 11 samples for each condition. Indeed, at least 20 samples are required for a robust WGCNA analysis and to obtain reliable results.

Recently, also miRNA expression data of 418 BCa and 19 normal tissue samples collected in The Cancer Genome Atlas (TCGA) have been investigated (21). After the selection of differentially expressed miRNAs, WGCNA analysis allowed the identification of a module closely related to BCa progression. Thirteen downregulated miRNAs (Table 1) also had a prognostic value since their low expression levels were associated with poorer OS in BCa patients. Interestingly, the predicted targets of these miRNAs were found to be significantly enriched in cancer-related pathways (Table 1). It has been shown that miR-1-1 and miR-133a are under-expressed in BCa cells and act as tumor suppressive miRNAs since, when overexpressed, they inhibited cell proliferation and invasion and increased apoptosis (55). In particular, miR-1 can exert its tumor suppressive function by targeting both coding genes, for example CCL2 (56), and non-coding RNAs, including UCA1 (57). MiR-133a can also induce apoptosis through silencing of GSTP1 in BCa cell lines (58) and, along with miR-1, it can inhibit BCa cell proliferation and increase apoptosis by targeting TAGLN2 mRNA (59). Moreover, miR-133a and miR-145 suppress cancer cell proliferation by directly regulating FSCN1 expression (60). MiR-133b plays a key role in proliferation and apoptosis by silencing Bcl-W and AKT1 genes (61) and its downregulation is associated with BCa progression and poor prognosis (62). Also miR-139 is a tumor suppressive miRNA since it can inhibit BCa cell proliferation by targeting the BMI1 oncogene (63) and cell migration and invasion by silencing matrix metalloprotease 11 (MMP11) (64). It has been observed that miR-143 can inhibit tumor cell proliferation, it is associated with BCa resistance to gemcitabine (65) and, along with miR-145, is a good prognostic biomarker for BCa patient survival (66). Interestingly, the polymorphism rs353293 in the common promoter of miR-143 and miR-145 is associated with BCa risk (67). Finally, miR-195 suppressed cancer cell proliferation by silencing GLUT3 (68), CDK4 (69), and CDC42 (70) genes. Therefore, these tumor suppressive miRNAs can be considered for *in vivo* delivery of therapeutics in bladder cancer, even though there are still challenges to the development of miRNA delivery strategies without toxicity induction.

## FURTHER ANALYSES ON IDENTIFIED HUB GENES AND MIRNAS

We performed a Gene Ontology analysis using all hub genes identified by WGCNA in the reviewed studies. Overall, they were found to belong to related biological processes. In particular, we highlighted pathways such as cell cycle, mitosis, mitotic spindle organization, kinetochore assembly, and nuclear division. All

these processes are involved in the uncontrolled cell proliferation in cancers. In addition, by using the transcription factor binding data generated by the ENCODE project consortium ([www.encodeproject.org](http://www.encodeproject.org)), we identified the E2F4 and FOXM1 transcription factors as the master regulators of hub gene expression, since they resulted as being significantly over-represented in the promoters of hub genes ( $p = 5.892 \times 10^{-9}$  and  $p = 3.220 \times 10^{-8}$ , respectively). Moreover, in order to investigate whether relationships exist between these miRNAs and the hub genes listed in **Table 1**, we performed a comprehensive literature search and identified which hub genes are targets of the hub miRNAs. In order to increase the analysis stringency, we considered only the experimentally assessed miRNA targets. Results reported in **Table 2** show that nearly all hub miRNAs target at least one hub gene, therefore these miRNAs could be involved in the deregulation of hub genes. Alternatively, hub miRNAs and genes could be deregulated due to the alteration of master regulators, such as transcription factors.

## CONCLUSIONS

WGCNA is a recently developed method for the analysis of gene expression data able to propose candidate therapeutic targets or diagnostic/prognostic biomarkers. Here, we reviewed all studies where WGCNA has been applied for the analysis of expression data from BCa. They include analyses of gene and miRNA expression data. Notably, neither lncRNA expression or splicing isoform-specific RNA expression have yet been investigated in BCa by WGCNA, as recently carried out, for example, in pancreatic cancer (15) and in clear cell renal cell carcinoma (ccRCC) (86), respectively. In addition, expression data of circular RNAs and RNAs in exosomes have not been analyzed by WGCNA in any cancer type. Recently, this network strategy has also been applied to proteomic and metabolomic data, although, due to the low coverage of proteomic and metabolomic analytical methods, WGCNA needed to be modified (87). Moreover, since RNA-seq data simultaneously allow the gene expression measure and mutation analysis, it would be interesting to perform an

**TABLE 2 |** Experimentally validated targets (among BCa hub genes previously identified by WGCNA) of hub miRNAs recently identified in BCa (21).

Hub miRNAs	Targeted hub genes	Validation assay	Cell lines	References
miR-1-1/2	CENPF FBN1 HERC2 KIF2C KIF4A UBE2I	Proteomics (pSILAC)	HeLa (human cervical cancer)	(71)
miR-28	PTPRJ	PAR-CLIP	HCT116 (human colon cancer)	(72)
miR-133a-1/2	CDC4A8	PAR-CLIP	C8166 (HIV-1 infected human T cells) and TZM-bl (HIV-1 infected human epithelial cells)	(73)
miR-139	FBN1	Microarray	T24 and KK47 (human bladder cancer)	(58)
	MMP11	Luciferase reporter assay, Microarray, qRT-PCR, Western blot	T24 and BOY (human bladder cancer)	(64)
miR-143	BRD2	PAR-CLIP	HCT116 (human colon cancer)	(72)
	CDC25B			
	COL3A1	<i>In situ</i> hybridization, qRT-PCR, Western blot	NF-38 (human normal gastric fibroblasts) and CaF-38 (human cancer-associated fibroblasts)	(74)
miR-195	COL5A2	Microarray	USSCs (human unrestricted somatic stem cells)	(75)
	CEP55	PAR-CLIP	HEK293S (human embryonic kidney) and MCF7 (breast cancer)	(76–79)
	PTPRJ	HITS-CLIP	Jijoye (EBV-transformed B cells)	(80)
miR-6507	PURA			
	TRAK1	HITS-CLIP	HEK293S (human embryonic kidney) and fresh-frozen human heart tissues	(81, 82)
	UBE2I	Microarray, qRT-PCR	PBMCs (peripheral blood mononuclear blood cells)	(83)
	CDC25B	PAR-CLIP	HCT116 (human colon cancer)	(72)
	CDC4A8			
	DCAF8	PAR-CLIP	HEK293S (human embryonic kidney)	(76)
	PRC1			
	CENPA	PAR-CLIP	hESCs (human embryonic stem cells)	(84)
	CEP55	PAR-CLIP	BC-1 and BC-3 (KSHV-infected primary effusion lymphoma)	(85)

pSILAC, pulsed Stable Isotope Labeling with Aminoacids in Cell culture; PAR-CLIP, PhotoActivatable Ribonucleoside-enhanced CrossLinking and ImmunoPrecipitation; HITS-CLIP, High-Throughput Sequencing of RNA isolated by CrossLinking and ImmunoPrecipitation.



evaluation of the mutation effects on the gene co-expression module assignment of a gene.

Although WGCNA requires expression data from at least 20 samples in order to obtain reliable results, only 8, 9, or 11 samples for each condition were analyzed in two reviewed studies (19, 20). Generally, in order to overcome the problem of a small sample size, researchers use more than one expression dataset, particularly during analysis of microarray data. However, systematic and technical differences between different microarray platforms and datasets (called batch effects) could emerge. The correct data pre-processing is needed since WGCNA is sensitive to batch effects. Moreover, the presence of outliers (samples with very different expression profiles from the bulk) may affect WGCNA results, thus their removal is a critical step (12). Unfortunately, sometimes researchers tend to reject few outliers because of the small sample size. A further critical element for WGCNA analysis is the highly variable gene expression among samples due to tumor heterogeneity. To overcome this problem, large sample size expression datasets should be used and, in particular, datasets that include samples isolated from different points of a single cancer tissue should be preferred. Moreover, we suggest performing sample clustering based on expression data before WGCNA analysis, in order to process less heterogeneous samples, as recently carried out for ccRCC (88).

High BCa heterogeneity, different microarray platforms, specific setting of WGCNA algorithm and different patient therapies, lifestyle, stages, age, and sex could explain the

little overlap between key genes identified among the studies. However, since these hub genes are highly co-expressed, they could highlight a common mechanism of transcriptional and post-transcriptional regulation, thus revealing mechanistic insights into cancer development. In particular, we identified the hub gene FOXM1 as the master regulator of the other genes identified by WGCNA. Hub genes could also be related at a functional level, since they belong to similar pathways and are regulated by a few common hub miRNAs. Furthermore, these miRNAs are known to be involved in the same pathways of hub genes, thus supporting the role in BCa of genes and miRNAs identified by WGCNA. Finally, although currently no hub gene has sufficient validity for clinical practice, many of them are already known to serve as biomarkers in BCa. Therefore, these genes are worth being further explored, since they can shed light into the molecular mechanisms of BCa, thus leading to the definition of novel personalized therapies. For example, regarding biomarkers for chemotherapy efficacy and toxicity, WGCNA identified the cytochrome P450 family member CYP3A5, already found to be useful in defining BCa patients with better responses to treatments.

## AUTHOR CONTRIBUTIONS

MG and FP conception and design. MG, GO, ARi, and FP drafting the manuscript. GO, ARi, MB, and AC review of the literature. ARu, EC, TC, and GP critical revision of the manuscript.

## REFERENCES

- Antoni S, Ferlay J, Soerjomataram I, Znaor A, Jemal A, Bray F. Bladder cancer incidence and mortality: a global overview and recent trends. *Eur Urol.* (2017) 71:96–108. doi: 10.1016/j.eururo.2016.06.010
- Galsky MD. Bladder cancer in 2017: Advancing care through genomics and immune checkpoint blockade. *Nat Rev Urol.* (2018) 15:71–2. doi: 10.1038/nrurol.2017.199
- Sanli O, Dobruch J, Knowles MA, Burger M, Alemozaffar M, Nielsen ME, et al. Bladder cancer. *Nat Rev Dis Primers* (2017) 3:17022. doi: 10.1038/nrdp.2017.22
- Lotan Y, Elias K, Svatek RS, Bagrodia A, Nuss G, Moran B, et al. Bladder cancer screening in a high risk asymptomatic population using a point of care urine based protein tumor marker. *J Urol.* (2009) 182:52–7; discussion 58. doi: 10.1016/j.juro.2009.02.142
- Leiblich A. Recent developments in the search for urinary biomarkers in bladder cancer. *Curr Urol Rep.* (2017) 18:100. doi: 10.1007/s11934-017-0748-x
- Giulietti M, Santoni M, Cimadamore A, Carrozza F, Piva F, Cheng L, et al. Exploring small extracellular vesicles for precision medicine in prostate cancer. *Front Oncol.* (2018) 8:221. doi: 10.3389/fonc.2018.00221
- Nawaz M, Camussi G, Valadi H, Nazarenko I, Ekstrom K, Wang X, et al. The emerging role of extracellular vesicles as biomarkers for urogenital cancers. *Nat Rev Urol.* (2014) 11:688–701. doi: 10.1038/nrurol.2014.301
- Barabasi AL, Oltvai ZN. Network biology: understanding the cell's functional organization. *Nat Rev Genet.* (2004) 5:101–13. doi: 10.1038/nrg1272
- de la Fuente A. From “differential expression” to “differential networking” - identification of dysfunctional regulatory networks in diseases. *Trends Genet.* (2010) 26:326–33. doi: 10.1016/j.tig.2010.05.001
- Zeng T, Sun SY, Wang Y, Zhu H, Chen L. Network biomarkers reveal dysfunctional gene regulations during disease progression. *FEBS J.* (2013) 280:5682–95. doi: 10.1111/febs.12536
- Langfelder P, Horvath S. WGCNA: an R package for weighted correlation network analysis. *BMC Bioinformatics* (2008) 9:559. doi: 10.1186/1471-2105-9-559
- Oldham MC, Konopka G, Iwamoto K, Langfelder P, Kato T, Horvath S, et al. Functional organization of the transcriptome in human brain. *Nat Neurosci.* (2008) 11:1271–82. doi: 10.1038/nn.2207
- Giulietti M, Occhipinti G, Principato G, Piva F. Identification of candidate miRNA biomarkers for pancreatic ductal adenocarcinoma by weighted gene co-expression network analysis. *Cell Oncol.* (2017) 40:181–92. doi: 10.1007/s13402-017-0315-y
- Giulietti M, Occhipinti G, Principato G, Piva F. Weighted gene co-expression network analysis reveals key genes involved in pancreatic ductal adenocarcinoma development. *Cell Oncol.* (2016) 39:379–88. doi: 10.1007/s13402-016-0283-7
- Giulietti M, Righetti A, Principato G, Piva F. LncRNA co-expression network analysis reveals novel biomarkers for pancreatic cancer. *Carcinogenesis* (2018) 39:1016–25. doi: 10.1093/carcin/bgy069
- Yuan L, Shu B, Chen L, Qian K, Wang Y, Qian G, et al. Overexpression of COL3A1 confers a poor prognosis in human bladder cancer identified by co-expression analysis. *Oncotarget* (2017) 8:70508–20. doi: 10.18632/oncotarget.19733
- Li S, Liu X, Liu T, Meng X, Yin X, Fang C, et al. Identification of biomarkers correlated with the TNM staging and overall survival of patients with bladder cancer. *Front Physiol.* (2017) 8:947. doi: 10.3389/fphys.2017.00947
- Zhang X, Zhang M, Hou Y, Xu L, Li W, Zou Z, et al. Single-cell analyses of transcriptional heterogeneity in squamous cell carcinoma of urinary bladder. *Oncotarget* (2016) 7:66069–76. doi: 10.18632/oncotarget.11803
- Deng SP, Zhu L, Huang DS. Mining the bladder cancer-associated genes by an integrated strategy for the construction and analysis of differential co-expression networks. *BMC Genomics* (2015) 16(Suppl. 3):S4. doi: 10.1186/1471-2164-16-S3-S4

20. Gaballah HH. Integration of gene coexpression network, GO enrichment analysis for identification gene expression signature of invasive bladder carcinoma. *Transcriptomics* (2016) 4:126. doi: 10.4172/2329-8936.1000126
21. Zhao F, Ge YZ, Zhou LH, Xu LW, Xu Z, Ping WW, et al. Identification of hub miRNA biomarkers for bladder cancer by weighted gene coexpression network analysis. *Oncol Targets Ther.* (2017) 10:5551–9. doi: 10.2147/OTT.S146479
22. Kim CJ, Yoshioka N, Tambe Y, Kushima R, Okada Y, Inoue H. Periostin is down-regulated in high grade human bladder cancers and suppresses *in vitro* cell invasiveness and *in vivo* metastasis of cancer cells. *Int J Cancer* (2005) 117:51–8. doi: 10.1002/ijc.21120
23. Kim CJ, Sakamoto K, Tambe Y, Inoue H. Opposite regulation of epithelial-to-mesenchymal transition and cell invasiveness by periostin between prostate and bladder cancer cells. *Int J Oncol.* (2011) 38:1759–66. doi: 10.3892/ijo.2011.997
24. Bufo P, Sanguedolce F, Tortorella S, Cormio L, Carrieri G, Pannone G. Expression of mitotic kinases phospho-aurora A and aurora B correlates with clinical and pathological parameters in bladder neoplasms. *Histol Histopathol.* (2010) 25:1371–7. doi: 10.14670/HH-25.1371
25. Yamamoto Y, Matsuyama H, Chochi Y, Okuda M, Kawauchi S, Inoue R, et al. Overexpression of BUBR1 is associated with chromosomal instability in bladder cancer. *Cancer Genet Cytogenet.* (2007) 174:42–7. doi: 10.1016/j.cancergencyto.2006.11.012
26. Zhang Z, Zhang G, Kong C. High expression of Cdc25B and low expression of 14-3-3sigma is associated with the development and poor prognosis in urothelial carcinoma of bladder. *Tumour Biol.* (2014) 35:2503–12. doi: 10.1007/s13277-013-1331-9
27. Lei CY, Wang W, Zhu YT, Fang WY, Tan WL. The decrease of cyclin B2 expression inhibits invasion and metastasis of bladder cancer. *Urol Oncol.* (2016) 34:237 e1–10. doi: 10.1016/j.urolonc.2015.11.011
28. Rinaldetti S, Wirtz RM, Worst TS, Eckstein M, Weiss CA, Breyer J, et al. FOXM1 predicts overall and disease specific survival in muscle-invasive urothelial carcinoma and presents a differential expression between bladder cancer subtypes. *Oncotarget* (2017) 8:47595–606. doi: 10.18632/oncotarget.17394
29. Liu D, Zhang Z, Kong CZ. High FOXM1 expression was associated with bladder carcinogenesis. *Tumour Biol.* (2013) 34:1131–8. doi: 10.1007/s13277-013-0654-x
30. Inoguchi S, Seki N, Chiyomaru T, Ishihara T, Matsushita R, Mataka H, et al. Tumour-suppressive microRNA-24-1 inhibits cancer cell proliferation through targeting FOXM1 in bladder cancer. *FEBS Lett.* (2014) 588:3170–9. doi: 10.1016/j.febslet.2014.06.058
31. Li WM, Wei YC, Huang CN, Ke HL, Li CC, Yeh HC, et al. Matrix metalloproteinase-11 as a marker of metastasis and predictor of poor survival in urothelial carcinomas. *J Surg Oncol.* (2016) 113:700–7. doi: 10.1002/jso.24195
32. Zhang J, Jia Q, Zou S, Zhang P, Zhang X, Skog S, et al. Thymidine kinase 1: a proliferation marker for determining prognosis and monitoring the surgical outcome of primary bladder carcinoma patients. *Oncol Rep.* (2006) 15:455–61. doi: 10.3892/or.15.2.455
33. Kim YH, Yan C, Lee IS, Piao XM, Byun YJ, Jeong P, et al. Value of urinary topoisomerase-IIA cell-free DNA for diagnosis of bladder cancer. *Investig Clin Urol.* (2016) 57:106–12. doi: 10.4111/icu.2016.57.2.106
34. Kim EJ, Lee YS, Kim YJ, Kim MJ, Ha YS, Jeong P, et al. Clinical implications and prognostic values of topoisomerase-II alpha expression in primary non-muscle-invasive bladder cancer. *Urology* (2010) 75:1516 e9–13. doi: 10.1016/j.urolgy.2009.08.055
35. Raspollini MR, Luque RJ, Menendez CL, Bollito E, Brunelli M, Martignoni G, et al. T1 high-grade bladder carcinoma outcome: the role of p16, topoisomerase-IIalpha, survivin, and E-cadherin. *Hum Pathol.* (2016) 57:78–84. doi: 10.1016/j.humpath.2016.06.022
36. Yan L, Li S, Xu C, Zhao X, Hao B, Li H, et al. Target protein for Xklp2 (TPX2), a microtubule-related protein, contributes to malignant phenotype in bladder carcinoma. *Tumour Biol.* (2013) 34:4089–100. doi: 10.1007/s13277-013-1000-z
37. Morikawa T, Kawai T, Abe H, Kume H, Homma Y, Fukayama M. UBE2C is a marker of unfavorable prognosis in bladder cancer after radical cystectomy. *Int J Clin Exp Pathol.* (2013) 6:1367–74.
38. Frstrup N, Birkenkamp-Demtroder K, Reinert T, Sanchez-Carbayo M, Segersten U, Malmstrom PU, et al. Multicenter validation of cyclin D1, MCM7, TRIM29, and UBE2C as prognostic protein markers in non-muscle-invasive bladder cancer. *Am J Pathol.* (2013) 182:339–49. doi: 10.1016/j.ajpath.2012.10.017
39. Zheng M, Simon R, Mirlacher M, Maurer R, Gasser T, Forster T, et al. TRIO amplification and abundant mRNA expression is associated with invasive tumor growth and rapid tumor cell proliferation in urinary bladder cancer. *Am J Pathol.* (2004) 165:63–9. doi: 10.1016/S0002-9440(10)63275-0
40. Yamamoto Y, Suehiro Y, Suzuki A, Nawata R, Kawai Y, Inoue R, et al. Germline DNA copy number variations as potential prognostic markers for non-muscle invasive bladder cancer progression. *Oncol Lett.* (2017) 14:1193–9. doi: 10.3892/ol.2017.6233
41. Zhang Z, Huettner PC, Nguyen L, Bidder M, Funk MC, Li J, et al. Aberrant promoter methylation and silencing of the POU2F3 gene in cervical cancer. *Oncogene* (2006) 25:5436–45. doi: 10.1038/sj.onc.1209530
42. Sun WX, Chen YH, Liu ZZ, Xie JJ, Wang W, Du YP, et al. Association between the CYP1A2 polymorphisms and risk of cancer: a meta-analysis. *Mol Genet Genomics* (2015) 290:709–25. doi: 10.1007/s00438-014-0956-8
43. Piva F, Giulietti M, Burini AB, Principato G. SpliceAid 2: a database of human splicing factors expression data and RNA target motifs. *Hum Mutat.* (2012) 33:81–5. doi: 10.1002/humu.21609
44. Piva F, Giulietti M, Nocchi L, Principato G. SpliceAid: a database of experimental RNA target motifs bound by splicing proteins in humans. *Bioinformatics* (2009) 25:1211–3. doi: 10.1093/bioinformatics/btp124
45. Giulietti M, Piva F, D'Antonio M, D'Onorio De Meo P, Paoletti D, Castrignano T, et al. SpliceAid-F: a database of human splicing factors and their RNA-binding sites. *Nucleic Acids Res.* (2013) 41:D125–31. doi: 10.1093/nar/gks997
46. Piva F, Giulietti M, Nardi B, Bellantuono C, Principato G. An improved *in silico* selection of phenotype affecting polymorphisms in SLC6A4, HTR1A and HTR2A genes. *Hum Psychopharmacol.* (2010) 25:153–61. doi: 10.1002/hup.1100
47. Piva F, Giulietti M, Baldelli L, Nardi B, Bellantuono C, Armeni T, et al. Bioinformatic analyses to select phenotype affecting polymorphisms in HTR2C gene. *Hum Psychopharmacol.* (2011) 26:365–72. doi: 10.1002/hup.1214
48. Piva F, Giulietti M, Occhipinti G, Santoni M, Massari F, Sotte V, et al. Computational analysis of the mutations in BAP1, PBRM1 and SETD2 genes reveals the impaired molecular processes in renal cell carcinoma. *Oncotarget* (2015) 6:32161–8. doi: 10.18632/oncotarget.5147
49. Giulietti M, Milantoni SA, Armeni T, Principato G, Piva F. ExportAid: database of RNA elements regulating nuclear RNA export in mammals. *Bioinformatics* (2015) 31:246–51. doi: 10.1093/bioinformatics/btu620
50. Piva F, Giulietti M, Armeni T, Principato G. Cross-link immunoprecipitation data to detect polymorphisms lying in splicing regulatory motifs: a method to refine single nucleotide polymorphism selection in association studies. *Psychiatr Genet.* (2012) 22:88–91. doi: 10.1097/YPG.0b013e32834c0bd1
51. Giulietti M, Grillo G, Liuni S, Pesole G. A guideline for the annotation of UTR regulatory elements in the UTRsite collection. *Methods Mol Biol.* (2015) 1269:339–48. doi: 10.1007/978-1-4939-2291-8\_21
52. Xie JY, Chen PC, Zhang JL, Gao ZS, Neves H, Zhang SD, et al. The prognostic significance of DAPK1 in bladder cancer. *PLoS ONE* (2017) 12:e0175290. doi: 10.1371/journal.pone.0175290
53. Duran I, Hagen C, Arranz JA, Apellaniz-Ruiz M, Perez-Valderrama B, Sala N, et al. SNPs associated with activity and toxicity of cabazitaxel in patients with advanced urothelial cell carcinoma. *Pharmacogenomics* (2016) 17:463–71. doi: 10.2217/pgs.15.186
54. Mbatshi LC, Gassiot M, Pourquier P, Goberna A, Mahammedi H, Mourey L, et al. Association of NR1I2, CYP3A5 and ABCB1 genetic polymorphisms with variability of temsirolimus pharmacokinetics and toxicity in patients with metastatic bladder cancer. *Cancer Chemother Pharmacol.* (2017) 80:653–9. doi: 10.1007/s00280-017-3379-5

55. Yamasaki T, Yoshino H, Enokida H, Hidaka H, Chiyomaru T, Nohata N, et al. Novel molecular targets regulated by tumor suppressors microRNA-1 and microRNA-133a in bladder cancer. *Int J Oncol.* (2012) 40:1821–30. doi: 10.3892/ijo.2012.1391
56. Wang W, Shen F, Wang C, Lu W, Wei J, Shang A. MiR-1-3p inhibits the proliferation and invasion of bladder cancer cells by suppressing CCL2 expression. *Tumour Biol.* (2017) 39:1010428317698383. doi: 10.1177/1010428317698383
57. Wang T, Yuan J, Feng N, Li Y, Lin Z, Jiang Z, et al. Hsa-miR-1 downregulates long non-coding RNA urothelial cancer associated 1 in bladder cancer. *Tumour Biol.* (2014) 35:10075–84. doi: 10.1007/s13277-014-2321-2
58. Uchida Y, Chiyomaru T, Enokida H, Kawakami K, Tatarano S, Kawahara K, et al. MiR-133a induces apoptosis through direct regulation of GSTP1 in bladder cancer cell lines. *Urol Oncol.* (2013) 31:115–23. doi: 10.1016/j.urolonc.2010.09.017
59. Yoshino H, Chiyomaru T, Enokida H, Kawakami K, Tatarano S, Nishiyama K, et al. The tumour-suppressive function of miR-1 and miR-133a targeting TAGLN2 in bladder cancer. *Br J Cancer* (2011) 104:808–18. doi: 10.1038/bjc.2011.23
60. Chiyomaru T, Enokida H, Tatarano S, Kawahara K, Uchida Y, Nishiyama K, et al. miR-145 and miR-133a function as tumor suppressors and directly regulate FSCN1 expression in bladder cancer. *Br J Cancer* (2010) 102:883–91. doi: 10.1038/sj.bjc.6605570
61. Chen XN, Wang KF, Xu ZQ, Li SJ, Liu Q, Fu DH, et al. MiR-133b regulates bladder cancer cell proliferation and apoptosis by targeting Bcl-w and Akt1. *Cancer Cell Int.* (2014) 14:70. doi: 10.1186/s12935-014-0070-3
62. Chen X, Wu B, Xu Z, Li S, Tan S, Liu X, et al. Downregulation of miR-133b predict progression and poor prognosis in patients with urothelial carcinoma of bladder. *Cancer Med.* (2016) 5:1856–62. doi: 10.1002/cam4.777
63. Luo H, Yang R, Li C, Tong Y, Fan L, Liu X, et al. MicroRNA-139-5p inhibits bladder cancer proliferation and self-renewal by targeting the Bmi1 oncogene. *Tumour Biol.* (2017) 39:1010428317718414. doi: 10.1177/1010428317718414
64. Yonemori M, Seki N, Yoshino H, Matsushita R, Miyamoto K, Nakagawa M, et al. Dual tumor-suppressors miR-139-5p and miR-139-3p targeting matrix metalloproteinase 11 in bladder cancer. *Cancer Sci.* (2016) 107:1233–42. doi: 10.1111/cas.13002
65. Wang H, Li Q, Niu X, Wang G, Zheng S, Fu G, et al. miR-143 inhibits bladder cancer cell proliferation and enhances their sensitivity to gemcitabine by repressing IGF-1R signaling. *Oncol Lett.* (2017) 13:435–40. doi: 10.3892/ol.2016.5388
66. Aygeris M, Mavridis K, Tokas T, Stravodimos K, Fragoulis EG, Scorilas A. Uncovering the clinical utility of miR-143, miR-145 and miR-224 for predicting the survival of bladder cancer patients following treatment. *Carcinogenesis* (2015) 36:528–37. doi: 10.1093/carcin/bgv024
67. Wu J, Huang Q, Meng D, Huang M, Li C, Qin T. A functional rs353293 polymorphism in the promoter of miR-143/145 is associated with a reduced risk of bladder cancer. *PLoS ONE* (2016) 11:e0159115. doi: 10.1371/journal.pone.0159115
68. Fei X, Qi M, Wu B, Song Y, Wang Y, Li T. MicroRNA-195-5p suppresses glucose uptake and proliferation of human bladder cancer T24 cells by regulating GLUT3 expression. *FEBS Lett.* (2012) 586:392–7. doi: 10.1016/j.febslet.2012.01.006
69. Lin Y, Wu J, Chen H, Mao Y, Liu Y, Mao Q, et al. Cyclin-dependent kinase 4 is a novel target in microRNA-195-mediated cell cycle arrest in bladder cancer cells. *FEBS Lett.* (2012) 586:442–7. doi: 10.1016/j.febslet.2012.01.027
70. Zhao C, Qi L, Chen M, Liu L, Yan W, Tong S, et al. microRNA-195 inhibits cell proliferation in bladder cancer via inhibition of cell division control protein 42 homolog/signal transducer and activator of transcription-3 signaling. *Exp Ther Med.* (2015) 10:1103–8. doi: 10.3892/etm.2015.2633
71. Selbach M, Schwanhauss B, Thierfelder N, Fang Z, Khanin R, Rajewsky N. Widespread changes in protein synthesis induced by microRNAs. *Nature* (2008) 455:58–63. doi: 10.1038/nature07228
72. Krell J, Stebbing J, Carissimi C, Dabrowska AF, de Giorgio A, Frampton AE, et al. TP53 regulates miRNA association with AGO2 to remodel the miRNA-mRNA interaction network. *Genome Res.* (2016) 26:331–41. doi: 10.1101/gr.191759.115
73. Whisnant AW, Bogerd HP, Flores O, Ho P, Powers JG, Sharova N, et al. In-depth analysis of the interaction of HIV-1 with cellular microRNA biogenesis and effector mechanisms. *MBio* (2013) 4:e000193. doi: 10.1128/mBio.00193-13
74. Naito Y, Sakamoto N, Oue N, Yashiro M, Sentani K, Yanagihara K, et al. MicroRNA-143 regulates collagen type III expression in stromal fibroblasts of scirrhous type gastric cancer. *Cancer Sci.* (2014) 105:228–35. doi: 10.1111/cas.12329
75. Bakhshandeh B, Soleimani M, Paylakhi SH, Ghaemi N. A microRNA signature associated with chondrogenic lineage commitment. *J Genet.* (2012) 91:171–82.
76. Hafner M, Landthaler M, Burger L, Khorshid M, Hausser J, Berninger P, et al. Transcriptome-wide identification of RNA-binding protein and microRNA target sites by PAR-CLIP. *Cell* (2010) 141:129–41. doi: 10.1016/j.cell.2010.03.009
77. Farazi TA, Ten Hoeve JJ, Brown M, Mihailovic A, Horlings HM, van de Vijver MJ, et al. Identification of distinct miRNA target regulation between breast cancer molecular subtypes using AGO2-PAR-CLIP and patient datasets. *Genome Biol.* (2014) 15:R9. doi: 10.1186/gb-2014-15-1-r9
78. Memczak S, Jens M, Elefsinioti A, Torti F, Krueger J, Rybak A, et al. Circular RNAs are a large class of animal RNAs with regulatory potency. *Nature* (2013) 495:333–8. doi: 10.1038/nature11928
79. Kishore S, Jaskiewicz L, Burger L, Hausser J, Khorshid M, Zavolan M. A quantitative analysis of CLIP methods for identifying binding sites of RNA-binding proteins. *Nat Methods* (2011) 8:559–64. doi: 10.1038/nmeth.1608
80. Riley KJ, Rabinowitz GS, Yario TA, Luna JM, Darnell RB, Steitz JA. EBV and human microRNAs co-target oncogenic and apoptotic viral and human genes during latency. *EMBO J.* (2012) 31:2207–21. doi: 10.1038/emboj.2012.63
81. Karginov FV, Hannon GJ. Remodeling of Ago2-mRNA interactions upon cellular stress reflects miRNA complementarity and correlates with altered translation rates. *Genes Dev.* (2013) 27:1624–32. doi: 10.1101/gad.215939.113
82. Spengler RM, Zhang X, Cheng C, McLendon JM, Skeie JM, Johnson FL, et al. Elucidation of transcriptome-wide microRNA binding sites in human cardiac tissues by Ago2 HITS-CLIP. *Nucleic Acids Res.* (2016) 44:7120–31. doi: 10.1093/nar/gkw640
83. Devadas K, Biswas S, Haleyurisetty M, Ragupathy V, Wang X, Lee S, et al. Identification of host micro RNAs that differentiate HIV-1 and HIV-2 infection using genome expression profiling techniques. *Viruses* (2016) 8:E12. doi: 10.3390/v8050121
84. Lipchina I, Elkabetz Y, Hafner M, Sheridan R, Mihailovic A, Tuschl T, et al. Genome-wide identification of microRNA targets in human ES cells reveals a role for miR-302 in modulating BMP response. *Genes Dev.* (2011) 25:2173–86. doi: 10.1101/gad.17221311
85. Gottwein E, Corcoran DL, Mukherjee N, Skalsky RL, Hafner M, Nusbaum JD, et al. Viral microRNA targetome of KSHV-infected primary effusion lymphoma cell lines. *Cell Host Microbe* (2011) 10:515–26. doi: 10.1016/j.chom.2011.09.012
86. Hamilton MJ, Girke T, Martinez E. Global isoform-specific transcript alterations and deregulated networks in clear cell renal cell carcinoma. *Oncotarget* (2018) 9:23670–80. doi: 10.18632/oncotarget.25330
87. Pei G, Chen L, Zhang W. WGCNA application to proteomic and metabolomic data analysis. *Methods Enzymol.* (2017) 585:135–58. doi: 10.1016/bs.mie.2016.09.016
88. Wu P, Liu JL, Pei SM, Wu CP, Yang K, Wang SP, et al. Integrated genomic analysis identifies clinically relevant subtypes of renal clear cell carcinoma. *BMC Cancer* (2018) 18:287. doi: 10.1186/s12885-018-4176-1

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The handling Editor declared a shared affiliation, though no other collaboration, with several of the authors MG, GO, ARI, MB, TC, GP and FP.

Copyright © 2018 Giulietti, Occhipinti, Righetti, Bracci, Conti, Ruzzo, Cerigioni, Cacciamani, Principato and Piva. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.



# Emerging Molecular Technologies in Genitourinary Tumors

Francesca Giunchi<sup>1</sup>, Alessia Cimadamore<sup>2</sup> and Michelangelo Fiorentino<sup>1\*</sup>

<sup>1</sup> Laboratory of Oncologic Molecular Pathology, S. Orsola-Malpighi Teaching Hospital University of Bologna, Bologna, Italy,

<sup>2</sup> Section of Pathological Anatomy, Polytechnic University of the Marche Region, School of Medicine, United Hospitals, Ancona, Italy

**Keywords:** next generation sequencing, targeted gene sequencing, NanoString System, patient-derived xenografts, organoids, renal cell carcinoma, prostate carcinoma, bladder carcinoma

## INTRODUCTION

Diagnostic molecular pathology of genito-urinary (GU) tumors is facing new technological challenges in the era of genome-wide analyses and patient-derived animal tumor models. In view of the increasing number of dedicated clinical trials, GU tumors represent the next urgent field of application of molecular diagnostics and drug discovery after gastro-intestinal and thoracic oncology.

## OPEN ACCESS

### Edited by:

Liang Cheng,  
Indiana University - Purdue University  
Indianapolis, United States

### Reviewed by:

Gregor Mikuz,  
Innsbruck Medical University, Austria  
Maria Rosaria Raspollini,  
Azienda Ospedaliero-Universitaria  
Careggi, Italy

### \*Correspondence:

Michelangelo Fiorentino  
fiorentinomichelangelo@gmail.com

### Specialty section:

This article was submitted to  
Genitourinary Oncology,  
a section of the journal  
Frontiers in Oncology

**Received:** 31 August 2018

**Accepted:** 11 October 2018

**Published:** 30 October 2018

### Citation:

Giunchi F, Cimadamore A and  
Fiorentino M (2018) Emerging  
Molecular Technologies in  
Genitourinary Tumors.  
Front. Oncol. 8:489.  
doi: 10.3389/fonc.2018.00489

## DNA-BASED GENOME-WIDE ANALYSES

Wide spectrum mutational analyses using next generation sequencing (NGS) platforms will soon represent the standard-of-care technologies for the assessment of genetic variants in solid tumors (1). These technologies apply successfully to archival pathology specimens, cytological samples and even liquid biopsies (blood or pleural effusions) (2). Mutational analyses can be wider (whole exome sequencing, WES) or restricted to selected genes or amplicons (targeted gene sequencing TGS). Both approaches are used to identify single or multiple genetic variants as predictive biomarkers of response to targeted oncologic therapies. At least the following three genome-wide mutational analyses will become routine diagnostic tests for GU tumors in the immediate future. Analysis of *BRCA1* and *BRCA2* germ-line mutations will be required to assess inherited prostate cancer risk and to predict response to treatment with poly(ADP-ribose) polymerase (PARP) inhibitors and even next-generation anti-androgens (3, 4). Given the complexity of the *BRCA1* and *BRCA2* mutations the NGS sequencing is the ideal method for their assessment. Similarly, deep sequencing of the DNA mismatch repair genes will be required in patients with familial prostate and colorectal cancer for suspected Lynch syndrome (3). Mutations in homologous recombination repair genes (*ATM/BRCA1/2* specifically) is enriched in men with advanced clinical stage ( $\geq$  cT3) and higher Gleason grade groups ( $\geq$  3) (5). Patients with metastatic castration-resistant prostate cancer whose tumors harbor homologous recombination DNA repair gene alterations, experience a different response to PARP inhibitor therapy. In particular, patients with cancer harboring DNA repair alterations in genes other than *BRCA2* are often non-responders (6). The assessment of tumor mutation burden defined as the number of mutations per mega-base of tumor cell DNA is becoming the most relevant candidate biological predictor of response to immunotherapies targeting the PD-1/PD-L1 axis (7). Tumor mutation load can be achieved either by WES or by TGS using NGS dedicated panels covering at least 2 mega-bases of tumor DNA. Assessment of tumor mutation load is also prognostically relevant in metastatic renal cell cancer and in muscle-invasive bladder cancer (8–10). Finally, epigenetic changes, including CpG island hypermethylation can be investigated using genome-wide methylation NGS panels in the attempt to better stratify high-grade and low-grade disease (11).



## RNA-BASED GENOME-WIDE ANALYSES

Genome-wide transcriptome analyses include gene expression profiling, miRNA and non-coding RNA profiling and RNA sequencing. In particular, RNA sequencing with high-throughput NGS platforms starting from RNA libraries allows simultaneous analysis of differential gene expression, allele-specific expression, splicing variants, and gene rearrangements (12). These analyses can also be done on RNA and DNA contained in small extracellular vesicles (EVs) that could be found in blood, urine, and other biological fluids (13). RNA abundance and sequence can be also investigated by array hybridization using platforms such as the NanoString System (14). Immediate clinical application of RNA sequencing to GU tumor include primarily the following fields of interest. The study of tumor immune micro-environment through the expression analysis of immune response genes is becoming important to assess tumor response to immune check-point inhibitors and BCG in bladder cancer (15, 16). The new molecular classification of muscle-invasive bladder cancer is largely based on gene expression profiling (17). Recognition of the molecular subtypes has prognostic and therapeutic implications for patients with advanced urothelial cancer. The assessment in the tumor tissue of the AR-V7 splicing variant of the androgen receptor (AR) gene is a predictor of poor response to anti-androgens and good response to chemo-therapy in castration-resistant prostate cancer (CRPC). The presence of AR splicing variants can be successfully investigated by RNA sequencing in prostate cancer tissue samples (18).

## PATIENT-DERIVED ANIMAL MODELS

Patient-derived xenografts (PDX) are mouse models where disaggregated cells or little fragments of human tumors are implanted into immunodeficient mice. The establishment of a PDX allows treating and monitoring the response to treatment of the original tumor *in vivo* in the mouse, instead of the patient, providing the best therapeutic selection at the same time (19). This procedure is ethically and commercially valuable since it spares pointless drug toxicity to the patient while saving money for oncological treatments that would be ineffective. Successful PDX establishment for monitoring response to treatment has been described in GU tumors (20). In CRPC there are available examples of PDX for treatment with abiraterone and enzalutamide as well as for a number of drugs in pre-clinical phase of development (21). In papillary type kidney cancer harboring *MET* mutations, there is evidence of successful treatment of PDX with Cabozantinib and other *MET* inhibitors

(22, 23). PDX created using human bladder tumor tissues have been utilized to assess response rates to cisplatin or PI3K inhibitors (24). The success of PDX establishment is highly variable and depends on several tumor-related or animal-related factors. For instance, in a meta-analysis on bladder cancer, the tumor engraftment rate varied between 20 and 100% (24). In addition, several flaws can affect the reliability of PDX as surrogate models of original patients' tumors. Tumor histological appearance may change in the PDX frequently toward squamous or sarcomatoid or neuroendocrine differentiation. Cancer cell proliferative rates in PDX may increase as well as cancer mutations may turn out enriched or underestimated (25). On the other hand, host mice for PDX can be selected to be totally immunodeficient or "humanized" by forcing in the animals the expression of cytokines or injecting in the mouse bloodstream human bone marrow stem cells to re-create the tumor inflammatory microenvironment. Humanized PDX have been established for several tumor types but not yet for GU cancers (26).

Organoids are 3D cell-cultures recapitulating the natural complex environmental organization of a normal or a cancer tissue. They differ from the cell-lines that grow flat in 2D and lack the signal trafficking and the organization of a tissue (27). Organoids can be constructed from human cancer cells or tissues and can be utilized for testing the response to drugs (28). Compared to PDX, organoids are more amenable to grow but they are transient in nature and represent a methodological choice in-between cell-lines and animal xenografts. Organoid models have been created to trait rare phenotypes or genotypes of prostate cancer and to test their potential response to drugs, or to track evolution of bladder cancer (29, 30).

Patient-derived models are increasingly used to address questions in GU oncology. There are still limitations to the reliability of these models to actually guide patients' therapy. In addition, these model technologies require dedicated infrastructures (such as bio-banks, laboratories, and animal facilities) and experienced professionals. There are also several ethical restrictions to the use of model systems in different countries. Notwithstanding, PDX and organoids represent a fascinating opportunity to enhance cancer drug discovery and to provide more therapeutic options to cancer patients.

## AUTHOR CONTRIBUTIONS

MF: Conception and design; FG: Drafting the manuscript and review of the literature; AC: Critical revision of the manuscript.

## REFERENCES

- Ballester LY, Luthra R, Kanagal-Shamanna R, Singh RR. Advances in clinical next-generation sequencing: target enrichment and sequencing technologies. *Expert Rev Mol Diagn.* (2016) 16:357–72. doi: 10.1586/14737159.2016.1133298
- Van Allen EM, Wagle N, Stojanov P, Perrin DL, Cibulskis K, Marlow S. Whole-exome sequencing and clinical interpretation of formalin-fixed, paraffin-embedded tumor samples to guide precision cancer medicine. *Nat Med.* (2014) 20:682–8. doi: 10.1038/nm.3559
- Giri VN, Knudsen KE, Kelly WK, Abida W, Andriole GL, Bangma CH. Role of genetic testing for inherited prostate cancer risk: philadelphia prostate cancer consensus conference 2017. *J Clin Oncol.* (2018) 36:414–24. doi: 10.1200/JCO.2017.74.1173

4. Antonarakis ES, Lu C, Luber B, Liang C, Wang H, Chen Y. Germline DNA-repair gene mutations and outcomes in men with metastatic castration-resistant prostate cancer receiving first-line abiraterone and enzalutamide. *Eur Urol.* (2018) 74:218–25. doi: 10.1016/j.eururo.2018.01.035
5. Marshall CH, Fu W, Wang H, Baras AS, Lotan TL, Antonarakis ES. Prevalence of DNA repair gene mutations in localized prostate cancer according to clinical and pathologic features: association of Gleason score and tumor stage. *Prostate Cancer Prostatic Dis.* (2018) doi: 10.1038/s41391-018-0086-1. [Epub ahead of print].
6. Lu E, Thomas GV, Chen Y, Wyatt AW, Lloyd P, Youngren J, et al. DNA repair gene alterations and parp inhibitor response in patients with metastatic castration-resistant prostate cancer. *J Natl Compr Canc Netw.* (2018) 16:933–7. doi: 10.6004/jnccn.2018.7020
7. Colli LM, Machiela MJ, Myers TA, Jessop L, Yu K, Chanock SJ. Burden of nonsynonymous mutations among TCGA cancers and candidate immune checkpoint inhibitor responses. *Cancer Res.* (2016) 76:3767–72. doi: 10.1158/0008-5472.CAN-16-0170
8. de Velasco G, Miao D, Voss MH, Hakimi AA, Hsieh JJ, Tannir NM. Tumor mutational load and immune parameters across metastatic renal cell carcinoma risk groups. *Cancer Immunol Res.* (2016) 4:820–2. doi: 10.1158/2326-6066.CIR-16-0110
9. Choudhury NJ, Kiyotani K, Yap KL, Campanile A, Antic T, Yew PY, et al. Low T-cell receptor diversity, high somatic mutation burden, and high neoantigen load as predictors of clinical outcome in muscle-invasive bladder cancer. *Eur Urol Focus* (2016) 2:445–52. doi: 10.1016/j.euf.2015.09.007
10. Cimdamore A, Gasparrini S, Santoni M, Cheng L, Lopez-Beltran A, Battelli N. Biomarkers of aggressiveness in genitourinary tumors with emphasis on kidney, bladder, and prostate cancer. *Expert Rev Mol Diagn.* (2018) 18:645–55. doi: 10.1080/14737159.2018.1490179
11. Olkhov-Mitsel E, Savio AJ, Kron KJ, Pethe VV, Hermanns T, Fleshner NE. Epigenome-Wide DNA methylation profiling identifies differential methylation biomarkers in high-grade bladder cancer. *Transl Oncol.* (2017) 10:168–77. doi: 10.1016/j.tranon.2017.01.001
12. Kukurba KR, Montgomery SB. RNA sequencing and analysis. *Cold Spring Harb Protoc.* (2015) 2015:951–69. doi: 10.1101/pdb.top084970
13. Giulietti M, Santoni M, Cimdamore A, Carrozza F, Piva F, Cheng L, et al. Exploring small extracellular vesicles for precision medicine in prostate cancer. *Front Oncol.* (2018) 8:221. doi: 10.3389/fonc.2018.00221
14. Veldman-Jones MH, Brant R, Rooney C, Geh C, Emery H, Harbron CG. Evaluating robustness and sensitivity of the nanostring technologies ncounter platform to enable multiplexed gene expression analysis of clinical samples. *Cancer Res.* (2015) 75:2587–93. doi: 10.1158/0008-5472.CAN-15-0262
15. Tretiakova M, Fulton R, Kocherginsky M, Long T, Ussakli C, Antic T, et al. Concordance study of PD-L1 expression in primary and metastatic bladder carcinomas: comparison of four commonly used antibodies and RNA expression. *Mod Pathol.* (2018) 31:623–32. doi: 10.1038/modpathol.2017.188
16. Kamat AM, Briggman J, Urbauer DL, Svatek R, Noguera-González GM, Anderson R, et al. Cytokine panel for response to intravesical therapy (CyPRIT): nomogram of changes in urinary cytokine levels predicts patient response to bacillus Calmette-Guérin. *Eur Urol.* (2016) 69:197–200. doi: 10.1016/j.eururo.2015.06.023
17. Robertson AG, Kim J, Al-Ahmadie H, Bellmunt J, Guo G, Cherniack AD, et al. Comprehensive molecular characterization of muscle-invasive bladder cancer. *Cell* (2017) 171:540–56.e25. doi: 10.1016/j.cell.2017.09.007
18. Kohli M, Ho Y, Hillman DW, Van Etten JL, Henzler C, Yang R. Androgen receptor variant AR-V9 is coexpressed with AR-V7 in prostate cancer metastases and predicts abiraterone resistance. *Clin Cancer Res.* (2017) 23:4704–15. doi: 10.1158/1078-0432.CCR-17-0017
19. Verma B, Ritchie M, Mancini M. Development and applications of patient-derived xenograft models in humanized mice for oncology and immunology drug discovery. *Curr Protoc Pharmacol.* (2017) 78:14.41.1–12. doi: 10.1002/cpph.26
20. Inoue T, Terada N, Kobayashi T, Ogawa O. Patient-derived xenografts as in vivo models for research in urological malignancies. *Nat Rev Urol.* (2017) 14:267–83. doi: 10.1038/nrurol.2017.19
21. Lawrence MG, Obinata D, Sandhu S, Selth LA, Wong SQ, Porter LH. Patient-derived models of abiraterone- and enzalutamide-resistant prostate cancer reveal sensitivity to ribosome-directed therapy. *Eur Urol.* (2018) 74:562–72. doi: 10.1016/j.eururo.2018.06.020
22. Zhao H, Nolley R, Chan AMW, Rankin EB, Peehl DM. Cabozantinib inhibits tumor growth and metastasis of a patient-derived xenograft model of papillary renal cell carcinoma with MET mutation. *Cancer Biol Ther.* (2017) 18:863–71. doi: 10.1080/15384047.2016.1219816
23. Schuller AG, Barry ER, Jones RD, Henry RE, Frigault MM, Beran G. The MET Inhibitor AZD6094 (Savolitinib, HMPL-504) induces regression in papillary renal cell carcinoma patient-derived xenograft models. *Clin Cancer Res.* (2015) 21:2811–9. doi: 10.1158/1078-0432.CCR-14-2685
24. Bernardo C, Costa C, Sousa N, Amado F, Santos L. Patient-derived bladder cancer xenografts: a systematic review. *Transl Res.* (2015) 166:324–31. doi: 10.1016/j.trsl.2015.02.001
25. Hollingshead MG, Stockwin LH, Alcoser SY, Newton DL, Orsburn BC, Bonomi CA, et al. Gene expression profiling of 49 human tumor xenografts from in vitro culture through multiple in vivo passages - strategies for data mining in support of therapeutic studies. *BMC Genomics* (2014) 15:393. doi: 10.1186/1471-2164-15-393
26. Morton JJ, Bird G, Keysar SB, Astling DP, Lyons TR, Anderson RT, et al. XactMice: humanizing mouse bone marrow enable microenvironment reconstitution in a patient-derived xenograft model of head and neck cancer. *Oncogene* (2016) 35:290–300. doi: 10.1038/onc.2015.94
27. Sachs N, Clevers H. Organoid cultures for the analysis of cancer phenotypes. *Curr Opin Genet Dev.* (2014) 24:68–73. doi: 10.1016/j.gde.2013.11.012
28. Weeber F, Ooft SN, Dijkstra KK, Voest EE. Tumor organoids as a pre-clinical cancer model for drug discovery. *Cell Chem Biol.* (2017) 24:1092–100. doi: 10.1016/j.chembiol.2017.06.012
29. Puca L, Bareja R, Prandi D, Shaw R, Benelli M, Karthaus WR. Patient derived organoids to model rare prostate cancer phenotypes. *Nat Commun.* (2018) 9:2404. doi: 10.1038/s41467-018-04495-z
30. Lee SH, Hu W, Matulay JT. Tumor evolution and drug response in patient-derived organoid models of bladder cancer. *Cell* (2018) 173:515–28.e17. doi: 10.1016/j.cell.2018.03.017

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Giunchi, Cimdamore and Fiorentino. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# Advantages of publishing in Frontiers



## OPEN ACCESS

Articles are free to read  
for greatest visibility  
and readership



## FAST PUBLICATION

Around 90 days  
from submission  
to decision



## HIGH QUALITY PEER-REVIEW

Rigorous, collaborative,  
and constructive  
peer-review



## TRANSPARENT PEER-REVIEW

Editors and reviewers  
acknowledged by name  
on published articles

## Frontiers

Avenue du Tribunal-Fédéral 34  
1005 Lausanne | Switzerland

**Visit us:** [www.frontiersin.org](http://www.frontiersin.org)

**Contact us:** [info@frontiersin.org](mailto:info@frontiersin.org) | +41 21 510 17 00



## REPRODUCIBILITY OF RESEARCH

Support open data  
and methods to enhance  
research reproducibility



## DIGITAL PUBLISHING

Articles designed  
for optimal readership  
across devices



## FOLLOW US

[@frontiersin](https://twitter.com/frontiersin)



## IMPACT METRICS

Advanced article metrics  
track visibility across  
digital media



## EXTENSIVE PROMOTION

Marketing  
and promotion  
of impactful research



## LOOP RESEARCH NETWORK

Our network  
increases your  
article's readership