

# SERAFINO ZAPPACOSTA AND THE CEPPELLINI SCHOOL: A PIONEER MODEL FOR NURTURING EDUCATION IN IMMUNOLOGY

EDITED BY: Francesca Di Rosa and Ennio Carbone  
PUBLISHED IN: *Frontiers in Immunology*



IUIS



**frontiers** Research Topics



# frontiers

## Frontiers eBook Copyright Statement

The copyright in the text of individual articles in this eBook is the property of their respective authors or their respective institutions or funders. The copyright in graphics and images within each article may be subject to copyright of other parties. In both cases this is subject to a license granted to Frontiers.

The compilation of articles constituting this eBook is the property of Frontiers.

Each article within this eBook, and the eBook itself, are published under the most recent version of the Creative Commons CC-BY licence.

The version current at the date of publication of this eBook is CC-BY 4.0. If the CC-BY licence is updated, the licence granted by Frontiers is automatically updated to the new version.

When exercising any right under the CC-BY licence, Frontiers must be attributed as the original publisher of the article or eBook, as applicable.

Authors have the responsibility of ensuring that any graphics or other materials which are the property of others may be included in the CC-BY licence, but this should be checked before relying on the CC-BY licence to reproduce those materials. Any copyright notices relating to those materials must be complied with.

Copyright and source acknowledgement notices may not be removed and must be displayed in any copy, derivative work or partial copy which includes the elements in question.

All copyright, and all rights therein, are protected by national and international copyright laws. The above represents a summary only. For further information please read Frontiers' Conditions for Website Use and Copyright Statement, and the applicable CC-BY licence.

ISSN 1664-8714

ISBN 978-2-88966-050-6

DOI 10.3389/978-2-88966-050-6

## About Frontiers

Frontiers is more than just an open-access publisher of scholarly articles: it is a pioneering approach to the world of academia, radically improving the way scholarly research is managed. The grand vision of Frontiers is a world where all people have an equal opportunity to seek, share and generate knowledge. Frontiers provides immediate and permanent online open access to all its publications, but this alone is not enough to realize our grand goals.

## Frontiers Journal Series

The Frontiers Journal Series is a multi-tier and interdisciplinary set of open-access, online journals, promising a paradigm shift from the current review, selection and dissemination processes in academic publishing. All Frontiers journals are driven by researchers for researchers; therefore, they constitute a service to the scholarly community. At the same time, the Frontiers Journal Series operates on a revolutionary invention, the tiered publishing system, initially addressing specific communities of scholars, and gradually climbing up to broader public understanding, thus serving the interests of the lay society, too.

## Dedication to Quality

Each Frontiers article is a landmark of the highest quality, thanks to genuinely collaborative interactions between authors and review editors, who include some of the world's best academicians. Research must be certified by peers before entering a stream of knowledge that may eventually reach the public - and shape society; therefore, Frontiers only applies the most rigorous and unbiased reviews. Frontiers revolutionizes research publishing by freely delivering the most outstanding research, evaluated with no bias from both the academic and social point of view. By applying the most advanced information technologies, Frontiers is catapulting scholarly publishing into a new generation.

## What are Frontiers Research Topics?

Frontiers Research Topics are very popular trademarks of the Frontiers Journals Series: they are collections of at least ten articles, all centered on a particular subject. With their unique mix of varied contributions from Original Research to Review Articles, Frontiers Research Topics unify the most influential researchers, the latest key findings and historical advances in a hot research area! Find out more on how to host your own Frontiers Research Topic or contribute to one as an author by contacting the Frontiers Editorial Office: [researchtopics@frontiersin.org](mailto:researchtopics@frontiersin.org)



# SERAFINO ZAPPACOSTA AND THE CEPPELLINI SCHOOL: A PIONEER MODEL FOR NURTURING EDUCATION IN IMMUNOLOGY

Topic Editors:

**Francesca Di Rosa**, Institute of Molecular Biology and Pathology, Italian National Research Council, Italy

**Ennio Carbone**, University "Magna Graecia" of Catanzaro, Catanzaro, Italy; Department of Microbiology, Cell and Tumor Biology, Karolinska Institutet, Stockholm, Sweden



Image: S-F/Shutterstock.com

We acknowledge the initiation and support of this Research Topic by the International Union of Immunological Societies (IUIS). We hereby state publicly that the IUIS has had no editorial input in articles included in this Research Topic, thus ensuring that all aspects of this Research Topic are evaluated objectively, unbiased by any specific policy or opinion of the IUIS.

**Citation:** Di Rosa, F., Carbone, E., eds. (2020). Serafino Zappacosta and the Ceppellini School: A Pioneer Model For Nurturing Education in Immunology. Lausanne: Frontiers Media SA. doi: 10.3389/978-2-88966-050-6

***This Research Topic is dedicated to the memory  
of Ennio Carbone (1961-2020).***



# Table of Contents

05	<b><i>Editorial: Serafino Zappacosta and the Ceppellini School: A Pioneer Model for Nurturing Education in Immunology</i></b>
	Francesca Di Rosa
08	<b><i>The Ruggero Ceppellini Advanced School of Immunology and the Neapolitan Scientific Renaissance</i></b>
	Antonio Di Giacomo
11	<b><i>Serafino Zappacosta: An Enlightened Mentor and Educator</i></b>
	Ennio Carbone, Mario De Felice, Francesca Di Rosa, Ugo D'Oro, Silvia Fontana, Antonio La Cava, Michele Maio, Giuseppe Matarese, Luigi Racioppi, Giuseppina Ruggiero and Giuseppe Terrazzano
17	<b><i>Ruggero Ceppellini: A Perspective on His Contributions to Genetics and Immunology</i></b>
	Walter Bodmer
21	<b><i>Bone Marrow Transplantation 1957-2019</i></b>
	Elizabeth Simpson and Francesco Dazzi
27	<b><i>Immunology's Coming of Age</i></b>
	Stefan H. E. Kaufmann
40	<b><i>Commentary: Immunology's Coming of Age</i></b>
	Heinz Kohler, Anastas Dimitrov Pashov and Thomas Kieber-Emmons
43	<b><i>Vaccine Evolution and Its Application to Fight Modern Threats</i></b>
	Emanuele Andreano, Ugo D'Oro, Rino Rappuoli and Oretta Finco
48	<b><i>The Mononuclear Phagocytic System. Generation of Diversity</i></b>
	Siamon Gordon and Annette Plüddemann
58	<b><i>Sharing Knowledge With Young and Established Students of Immunology by the Neapolitan Gulf at the Ruggero Ceppellini Advanced School</i></b>
	Francesco Colucci
62	<b><i>Recirculation and Residency of T Cells and Tregs: Lessons Learnt in Anacapri</i></b>
	Silvia Piconese, Silvia Campello and Ambra Natalini
70	<b><i>Appendix: EFIS-EJI Ruggero Ceppellini Advanced School of Immunology Founded by Serafino Zappacosta. List of the activities From Its Foundation in 1991 to 2019</i></b>



# Editorial: Serafino Zappacosta and the Ceppellini School: A Pioneer Model for Nurturing Education in Immunology

**Francesca Di Rosa\***

*Institute of Molecular Biology and Pathology, Consiglio Nazionale delle Ricerche (IBPM-CNR), Rome, Italy*

**Keywords:** education, innate immunity, adaptive immunity, MHC, vaccination

## Editorial on the Research Topic

### Serafino Zappacosta and the Ceppellini School: A Pioneer Model for Nurturing Education in Immunology

This *Frontiers in Immunology Research Topic* is a collection of articles on the activities and the scientific interests of the founders, faculty, and students of the “Scuola Superiore di Immunologia Ruggero Ceppellini” (Ruggero Ceppellini Advanced School of Immunology), an International School of Immunology founded almost 30 years ago following a pioneer idea by Serafino Zappacosta. The school has more recently become known as the EFIS-EJI Ruggero Ceppellini Advanced School of Immunology founded by Serafino Zappacosta. The re-naming of the school followed the sudden death of Zappacosta in 2006 (1). Furthermore, in 2011 the European Federation of Immunological Societies (EFIS) declared the Ceppellini School one of its regularly sponsored activities. Since then, the European Journal of Immunology (EJI, i.e., the EFIS official journal) has regularly reported on the Ceppellini School’s international courses in its “News & EFIS” section [for recent examples see (2–4)].

The opening article of this *Research Topic* is a contribution by Antonio Di Giacomo (Colli Monaldi Hospital, Naples, Italy) who, in 1991, joined Zappacosta (at the time a full professor of immunology at the Federico II University, Naples) in the foundation of the Ceppellini School in Naples, Italy (Di Giacomo). Co-founders were Melchiorre Brai (University of Palermo, Italy), Giovanni B. Ferrara (Federico II University, Naples), Ciro Manzo (Istituto Pascale, Naples), and Alfred Nisonoff (Brandeis University, in Waltham, Massachusetts, USA). The title of Di Giacomo’s article, “The Ruggero Ceppellini Advanced School of Immunology and the Neapolitan Scientific Renaissance,” clearly indicates the strong roots of the Ceppellini School in the city of Naples. Di Giacomo illustrated the pioneer vision of the founders and their strong commitment to the generous educational project of the Ceppellini School, which was summarized in the Latin motto suggested by Zappacosta “*non multa sed multum*” (not many but much, i.e., quality not quantity) (Di Giacomo).

The second article is a tribute to Zappacosta by a group of previous students and collaborators who worked with him in Naples, all of them now well-established immunologists in Italy, Europe, and the USA (Carbone et al.). They reported on the research performed by Zappacosta and his team over more than 30 years on the role of MHC in innate and adaptive immunity, showing how their findings contributed to, and often anticipated, key issues of current literature. Silvia Fontana, one of the authors of this perspective, became the President of the Ceppellini School after Zappacosta’s death, and her strong commitment and passionate work have been essential for the continuation of the School’s educational project. The first author is Ennio Carbone, co-editor of this Research

## OPEN ACCESS

### Edited and reviewed by:

Francesca Granucci,  
University of Milano-Bicocca, Italy

### \*Correspondence:

Francesca Di Rosa  
francesca.dirosa@cnr.it

### Specialty section:

This article was submitted to  
Molecular Innate Immunity,  
a section of the journal  
*Frontiers in Immunology*

**Received:** 29 May 2020

**Accepted:** 09 June 2020

**Published:** 07 August 2020

### Citation:

Di Rosa F (2020) Editorial: Serafino  
Zappacosta and the Ceppellini  
School: A Pioneer Model for Nurturing  
Education in Immunology.  
*Front. Immunol.* 11:1524.  
doi: 10.3389/fimmu.2020.01524

Topic, who sadly died all of a sudden in March 2020 just after he became President of the Ceppellini School. We co-edited this *Research Topic*, but, unfortunately, he could not co-author this editorial. This *Research Topic* is dedicated to his memory.

Zappacosta and colleagues entitled their Advanced Immunology School to Ruggero Ceppellini, an outstanding Italian scientist who gave seminal contributions to the genetics of HLA. Here, Walter Bodmer (University of Oxford, UK) drew a picture of Ruggero Ceppellini and reported about some of his achievements and fruitful insights that inspired his contemporary colleagues and those who followed his path in later years (Bodmer). The inaugural course of the Ceppellini School was on bone marrow transplantation (BMT) in 1992. It was directed by Elizabeth Simpson, at the time working at the Division of Transplantation Biology, MRC Clinical Research Centre, Harrow, Middlesex, UK. In their article for this *Research Topic*, Elizabeth Simpson and Francesco Dazzi (King's College, London, UK) placed the achievements of about six decades of research and clinical experience on BMT in the context of today challenges and discussed how the contributions to the 1992 Ceppellini School course created a remarkable marker point about mid-way between the first BMT in 1957 and current times (Simpson and Dazzi).

In 2006, Stefan Kaufmann, who was Director of the Max-Planck Institute for Infection Biology, Department of Immunology, Berlin, Germany, became Scientific Director of the Ceppellini School. Kaufmann organized and directed several Ceppellini School courses, mostly focused on immune response and vaccination against tuberculosis and other threatening infectious diseases, such as malaria and AIDS (5). Here, Kaufmann gave a historical overview of the most remarkable milestones in immunology, focused on the Nobel laureates' achievements (Kaufmann). This personal and passionate perspective concisely summarized an overwhelming body of work. We also published a commentary by Heniz Kohler (University of Kentucky, Lexington, KY, USA) and colleagues, who integrated Kaufmann's review by emphasizing some additional aspects, for example, the theoretical contribution of the idiotypic network theory by Jerne, the thoughtful work on positive and negative selection (of both T and B cells), and the current successes of therapeutic antibodies (Kohler et al.).

Rino Rappuoli, a distinguished vaccinologist who has been part of the faculty of many Ceppellini School courses over the years (5), is co-author, alongside Emanuele Andreano, Ugo D'Oro, and Oretta Finco, of a mini-review discussing the most promising approaches to vaccinology, going from the genome-based "reverse vaccinology" at the end of last century to the

"reverse vaccinology 2.0" in 2016 and beyond (Andreano et al.). Siamon Gordon (University of Oxford, UK), Stefan Kaufmann, and Fernando Martinez-Estrada (University of Surrey, UK) were the scientific directors of a memorable Ceppellini School course on tissue phagocytes and function held in 2016 at the Stazione Zoologica "Anton Dohrn," a research center in Naples where the Russian scientist Elie Metchnikoff (1845–1916), who first described phagocytosis, worked for a short while (2). The contribution by Siamon Gordon and Annette Plüddemann (University of Oxford, UK) to this *Research Topic* is an inspiring discussion on macrophages diversity and function that highlights key open questions on macrophage heterogeneity and provides insights on its underlying pattern (Gordon and Plüddemann).

The last two articles focus on the fruitful sharing of knowledge between young attendees and senior faculty members of some exemplary Ceppellini School courses (4, 6). One article is by Francesco Colucci (University of Cambridge, UK), a Ceppellini School faculty member who was scientific co-director of the 2014 course on the maternal immune system in pregnancy (Colucci). The other article is by three of the participants to the 2018 Ceppellini School course on T-cell memory, i.e., Silvia Piconese (Sapienza University, Rome, Italy), Silvia Campello (University of Rome Tor Vergata, Rome, Italy), and Ambra Natalini (Sapienza University, and Institute of Molecular Biology and Pathology, CNR, Rome, Italy) (Piconese et al.). Both articles give a flavor of the exceptional learning experiences of participants to the Ceppellini School activities.

In 1991, the foundation of the Ceppellini School was a real breakthrough. After almost 30 years, the Ceppellini School continues to be an attractive pole for hundreds of young and enthusiastic participants from Europe, North and South America, the Middle East, Africa, and India. This *Research Topic* aims to offer some historical background and insightful perspectives on the Ceppellini School. Born from a Zappacosta's utopian idea, the school remains dedicated to strongly engaging new generation of young minds.

## AUTHOR CONTRIBUTIONS

The author confirms being the sole contributor of this work and has approved it for publication.

## ACKNOWLEDGMENTS

I thank all the faculty members of the Ceppellini School, and particularly Giuseppina Ruggiero, for making this *Research Topic* possible.

## REFERENCES

1. La Cava A, De Felice M. In memoriam: Serafino Zappacosta (1935–2006). *Tissue Antigens*. (2006) 68:279. doi: 10.1111/j.1399-0039.2006.00686.x
2. Di Giacomo A. Metchnikoff's legacy: tissue phagocytes and functions. *Eur J Immunol*. (2017) 47:10–13. doi: 10.1002/eji.201770016
3. Carbone E, Di Rosa F, Fridman W, Sautès-Fridman C. EFIS-EJI Ruggero Ceppellini advanced immunology school course: tumour immunology 2017: from tissue microenvironment to immunotherapy. Naples 16–18 October 2017. *Eur J Immunol*. (2018) 48:559–61. doi: 10.1002/eji.201870065
4. Natalini A, Fusco C, Micillo T, Di Rosa F. T cell memory in Capri: a successful course organized by the EFIS-EJI Ruggero Ceppellini advanced school of immunology founded by Serafino



- Zappacosta. *Eur J Immunol.* (2019) 49:361–3. doi: 10.1002/eji.201970035
5. Di Giacomo A. The EFIS-EJI Ruggero Ceppellini Advanced School of Immunology: Malaria, Tuberculosis and HIV/AIDS: novel vaccination strategies against the three major killers. *Eur J Immunol.* (2014) 44:1573–4. doi: 10.1002/eji.201470055
  6. Colucci F. Imagine a world without borders: an immunologist's thoughts on Brexit. *EMBO Rep.* (2016) 17:1241. doi: 10.15252/embr.201643019

**Conflict of Interest:** The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

*Copyright © 2020 Di Rosa. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.*



# The Ruggero Ceppellini Advanced School of Immunology and the Neapolitan Scientific Renaissance

Antonio Di Giacomo\*

UOS Haematology, Clinical Pathology Lab, "V. Monaldi" Hospital, Naples, Italy

In this article the author, cofounder with Serafino Zappacosta and few other knowledgeable scientists of the Ruggero Ceppellini Advanced School of Immunology in 1991, discusses the significance of this initiative not only for the spreading of immunological culture among scientists—including those from disadvantaged Countries—but also for the resurgence of the city of Naples as a cultural pole of attraction for brilliant minds, as it was in its past history. This is a tribute to Serafino Zappacosta's foresightedness and generosity.

**Keywords:** school, Serafino Zappacosta, immunology, renaissance, Naples (Italy)

## OPEN ACCESS

### Edited by:

Francesca Di Rosa,  
Consiglio Nazionale Delle Ricerche  
(CNR), Italy

### Reviewed by:

Peter Katsikis,  
Erasmus University  
Rotterdam, Netherlands  
Paola Nistico',  
Istituti Fisioterapici Ospitalieri  
(IRCCS), Italy

### \*Correspondence:

Antonio Di Giacomo  
digiacomo@itb.it

### Specialty section:

This article was submitted to  
Molecular Innate Immunity,  
a section of the journal  
Frontiers in Immunology

**Received:** 18 April 2019

**Accepted:** 14 June 2019

**Published:** 03 July 2019

### Citation:

Di Giacomo A (2019) The Ruggero  
Ceppellini Advanced School of  
Immunology and the Neapolitan  
Scientific Renaissance.  
Front. Immunol. 10:1494.  
doi: 10.3389/fimmu.2019.01494

Great intellectual achievements are the excellent fruits of rich and stimulating environments, fertile soils that prepare and nurture the mind. Roses do not bloom in a desert.

In this respect, culture constitutes the background for the development of new ideas and discoveries, that in every field of human knowledge represent the tools of advancement and innovation.

Biological sciences, intended as the study of the significance of the processes of life and not only their mechanical aspects, share with the human sciences the vast realm of the thinking mind, since when man experienced what Teilhard de Chardin defined as "the first moment" of self-awareness. Questions started to appear and science was born.

These and other related considerations was I debating in my thoughts when I met professor Serafino Zappacosta in a gray afternoon of November 1988 in the venues of a course on "Immunity in human pathology" that he used to give every 5 years at the Medical Faculty of the University of Naples "Federico II" where he was tenure professor of Immunology.

Indeed I had known him and his fame from before, as a medical student of that Faculty, always attracted by his charismatic personality and by the matter of his teaching, Immunology, at that time still a fast growing science, but I never had the opportunity of joining his group, nor did I try, the development of my career bringing me elsewhere. On that occasion, however, he showed interest in my curriculum and my recent experience abroad in experimental Immunology, and invited me to give seminars in his Institute about experimental cancer immunology, the subject of my studies. A collaboration started at that point, as did a long friendship.

In those years professor Zappacosta was maturing the intention to create a School of Immunology of international relevance in Naples, that would attract renowned scholars of that matter as teachers and an international audience of students coming from all over the world, in particular from developing Countries, in order to spread and promote immunological knowledge among scientists. He proposed me to join this project, along with a restricted group of scientists such as:

Melchiorre Brai, professor of Immunology at the University of Palermo; Giovan Battista Ferrara, professor of Human Genetics at the University of Naples "Federico II"; Albert Nisonoff, professor

of Biology at Brandeis University, Mass. USA; and Dr. Ciro Manzo, head of the Immunology Department of the Istituto Pascale in Naples. I enthusiastically accepted, of course, honored and flattered by his invitation.

The School was founded in June 1991 with the name “Ruggero Ceppellini Advanced School of Immunology,” dedicated to the memory of Ruggero Ceppellini (1917–88), the great Italian immunogeneticist, as a non-profit scientific association whose aim was to foster, encourage and propagate all aspects of knowledge relating to immunology and associated disciplines (genetics, microbiology, oncology) in the scientific community in Italy and in other Countries, through the promotion of scientific research, continuing education and in-service training.

The School's structure consisted in a Council of Directors and a Scientific Advisory Board, composed by a group of scientists each prominent in different areas of Immunology, that proposed and in turn took charge of the courses each year. Technically, the School teaching programmes were conceived and realized according to a “three level” scheme.

Level I courses, the typical refreshing courses, dealing with the so-called continuing education of medical graduates and designed to update the local practitioners on the recent advances having a bearing on their medical thinking and daily operation.

Level II courses, dedicated to young researchers working in Immunology or related fields, wishing to acquire knowledge of a specific topic within the vast area of Immunology. These are 1-week full-immersion activities, often integrated by workshops and small group discussions. Typical audience of these courses has been represented by Ph.D. students coming from all over the world.

Level III courses, short practical laboratory courses, dealing with recent techniques to be applied in research or even in the clinical laboratory, for small groups of graduates.

The School's inaugural ceremonies were held on 11 October, 1992, at Palazzo Serra di Cassano in Naples, the seat of the Istituto Italiano per gli Studi Filosofici, in the occasion of the School's first course, on the immunology of bone marrow transplantation. Many more courses followed, all successful and attended by students from all over the world, but more remarkable was the returning of Naples as the pole of attraction for scientific knowledge and culture after a long period of oblivion.

And this was the focal point of the all thing, the adventure clear to my mind since the beginning, the basic ideal drive that led me to join professor Zappacosta's dream to bring back to Naples the attention of the world's scientific community. Love for science together with love for our land. Naples certainly deserved this tribute as a recognition of its glorious past and its tradition of pole of attraction for excellent minds and inclusive culture.

This was not unexpected in Neapolitan history but determined by peculiar events, both human and geographical that, I believe, led to make this region of the world a favorable one to become a hub for philosophical and intellectual speculation. And science is, as we all know, fundamentally the result of intellectual and philosophical speculation, nothing else.

The same drive that attracted the divine Vergil to the Neapolitan epicurean school of Chiron, his master and philosophical mentor for masterpieces as the Georgics, led a

young and enlightened German emperor in the early Middle Ages to promote Naples as the center of culture, founding the oldest University of Europe and therefore of the western world. The “Studium” established in Naples by Friedrich II Hohenstaufen, emperor and innovator, “*stupor mundi*” as he was called by most historians, was not technically speaking the first one of its kind. The University of Bologna preceded it of many years, but Naples University was the first public institution of a State, born for the political will of a Ruler whose project was that of creating a place where studies were possible without having people leave home, and constituting a center of attraction for scholarly minds. It was an operation of qualified touristic promotion, we might say today, where the goal was the cultural growth of the place that would become in turn economical and social. At that time is ascribed the myth of the four founders, a Jew, an Arab, a Greek and a Latin, not real individuals but cultural influences concurring to the building of the *ars medica*, and the body of laws that regulate the teaching and the practice of medicine as stated in the “*Liber Constitutionum*” in A.D. 1231. Philosophical speculation and observation of the reality, theory and practice, *ratio et observatio* were the leading criteria for the development of scientific rationales and approaches.

Naples has therefore been since the far past the place where different culture met and merged, creating one of the first melting pots of peoples and ideas in history, favored by its geographical position at the center of the Mediterranean and by the efforts of enlightened rulers, like Friedrich II and, more recently, like The Bourbon kings. Starting with Charles III Bourbon, in fact, Naples became along with Vienna and Paris one of the best and most advanced courts in Europe, both for magnificence of arts and for scientific institutions. The first railroad in Italy between Naples and Portici was built in the year 1836, the first Italian scientific museum of mineralogy was established in the city in 1801 and the first volcanic observatory of the world was built on the slopes of Vesuvius in 1841. Naples was also the place where, in 1872 the eminent German biologist Anton Dohrn built the second laboratory of marine biology in Europe, which hosted, among many others, Ilya Metchnikoff, the second Nobel prize winner for medicine. In Naples took also place the seventh Meeting of the Italian Scientists in 1845, with the participation of 16 hundreds scientists, more than 8 hundred of whom from Naples and the south of Italy. Cultural and scientific growth called for economic growth.

Other times have not been so favorable however. Periods of glory alternating with periods of decline, mainly determined by political instability and short-sightedness have determined the cultural oblivion that has especially characterized the scientific life of the city. It looked as though Naples, in spite of sporadic and meritorious efforts operated at different levels by singular initiatives, substantially relied on its traditional and popular image of a place to visit for touristic reasons. An initiative of restoring the glorious role of the past was at this point needed by many people operating in the scientific field. This led to a dreamer such as Professor Zappacosta to enterprise this initiative of creating a novel cultural start that would bring here the best of the world scientists who would in turn attract again an international qualified audience. The project was also highly



philanthropic, since a great deal of attention was dedicated to the students coming from developing Countries who were actively encouraged to attend the courses by granting them bursaries also obtained by international benefactors like the Bill and Melinda Gates Foundation. This was a theme particularly dear to the founders, who envisioned the city to become once again a bridge between the western world and less fortunate areas of the planet that would benefit of this shared knowledge and culture. I recognize in those ideals the reason for my enthusiastic adhesion to Serafino's generous effort and I will be always grateful to him.

The legacy of the Ceppellini School, still vivid in our minds, can be summarized in the motto suggested by Professor Zappacosta "*non multa sed multum*" ("not many but much", i.e., "quality, not quantity"), that best represents the spirit of an institution that is progressively changing the perception of ourselves and of the role of the western culture in the world.

This perfectly fulfills the prophetic perspective of Serafino Zappacosta for a new era of Immunology and Immuno - Oncology in which a more humanistic and philosophical approach should prevail in research. May the Ceppellini School continue to be a beacon for decades to come.

## AUTHOR CONTRIBUTIONS

AD is a co-founder of the School of Immunology.

**Conflict of Interest Statement:** The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

*Copyright © 2019 Di Giacomo. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.*



# Serafino Zappacosta: An Enlightened Mentor and Educator

**Ennio Carbone<sup>1,2</sup>, Mario De Felice<sup>3</sup>, Francesca Di Rosa<sup>4\*</sup>, Ugo D'Oro<sup>5</sup>, Silvia Fontana<sup>3</sup>, Antonio La Cava<sup>6</sup>, Michele Maio<sup>7</sup>, Giuseppe Matarese<sup>3,8</sup>, Luigi Racioppi<sup>8,9</sup>, Giuseppina Ruggiero<sup>10\*</sup> and Giuseppe Terrazzano<sup>10,11</sup>**

<sup>1</sup> Department of Experimental and Clinical Medicine, University "Magna Graecia" of Catanzaro, Catanzaro, Italy, <sup>2</sup> Department of Microbiology, Cell and Tumor Biology, Karolinska Institutet, Stockholm, Sweden, <sup>3</sup> Istituto per l'Endocrinologia e l'Oncologia Sperimentale, Consiglio Nazionale delle Ricerche (IEOS-CNR), Naples, Italy, <sup>4</sup> Institute of Molecular Biology and Pathology, Consiglio Nazionale delle Ricerche (IBPM-CNR), Rome, Italy, <sup>5</sup> GlaxoSmithKline, Siena, Italy, <sup>6</sup> Department of Medicine, University of California, Los Angeles, Los Angeles, CA, United States, <sup>7</sup> Center for Immuno-Oncology, Medical Oncology and Immunotherapy, Department of Oncology, University Hospital of Siena, Siena, Italy, <sup>8</sup> Dipartimento di Medicina Molecolare e Biotecnologie Mediche, Università degli Studi di Napoli "Federico II", Naples, Italy, <sup>9</sup> Division of Hematological Malignancies and Cellular Therapy, Department of Medicine, Duke University School of Medicine, Durham, NC, United States, <sup>10</sup> Dipartimento di Scienze Mediche Traslazionali, Università di Napoli "Federico II", Naples, Italy, <sup>11</sup> Dipartimento di Scienze, Università della Basilicata, Potenza, Italy

## OPEN ACCESS

### Edited by:

Ivan Zanoni,  
Harvard Medical School,  
United States

### Reviewed by:

Francesca Granucci,  
University of Milano Bicocca, Italy  
Diana Boraschi,  
Istituto di Biochimica Delle Proteine  
(IBP), Italy

### \*Correspondence:

Francesca Di Rosa  
francesca.dirosa@cnr.it  
Giuseppina Ruggiero  
giruggie@unina.it

### Specialty section:

This article was submitted to  
Molecular Innate Immunity,  
a section of the journal  
Frontiers in Immunology

**Received:** 17 December 2019

**Accepted:** 27 January 2020

**Published:** 13 February 2020

### Citation:

Carbone E, De Felice M, Di Rosa F,  
D'Oro U, Fontana S, La Cava A,  
Maio M, Matarese G, Racioppi L,  
Ruggiero G and Terrazzano G (2020)  
Serafino Zappacosta: An Enlightened  
Mentor and Educator.  
Front. Immunol. 11:217.  
doi: 10.3389/fimmu.2020.00217

With this article, the authors aim to honor the memory of Serafino Zappacosta, who had been their mentor during the early years of their career in science. The authors discuss how the combination of Serafino Zappacosta's extraordinary commitment to teaching and passion for science created a fostering educational environment that led to the creation of the "Ruggiero Ceppellini Advanced School of Immunology." The review also illustrates how the research on the MHC and the inspirational scientific context in the Zappacosta's laboratory influenced the authors' early scientific interests, and subsequent professional work as immunologists.

**Keywords:** education, MHC, T cells, NK cells, immune response

Serafino Zappacosta, the founder of the "Ruggiero Ceppellini Advanced School of Immunology" (Ceppellini School), epitomized the term "mentor." This term was first used by François Fénelon in the book "Les Aventures de Télémaque" to define an enlightened educator who is endowed with unprejudiced knowledge and wisdom (1). The name came after Mentor, the guardian and educator of Odysseus' son Telemachus who offered him encouragement and support for dealing with personal dilemmas while his father was away fighting in the Trojan War. As a mentor and professor of immunology at the University of Naples "Federico II," Serafino Zappacosta communicated science to his students and close collaborators as a fascinating tool to constantly pursue and advance knowledge, thus nourishing their innate human eagerness to learn. The authors of this article had the privilege to be former members of Zappacosta's laboratory, and wish to offer him this posthumous tribute.

Serafino Zappacosta was a highly cultivated scientist, whose interests spanned from the classics to arts (2). His open-minded vision led him to go beyond the traditional approach to didactics toward new models of education. He conceptualized the idea of an international advanced school of immunology because of his recognition of the quintessential importance in committing educational efforts to nurturing generations of young researchers from all over the world, promoting exchanges between western world and developing countries. The Zappacosta's school model embraced an open and transparent communication that was instrumental to expand the creative potential of independent-minded young investigators, to foster their critical and analytical interests, and to channel their energies into highly valuable scientific directions.

Serafino Zappacosta founded the Ceppellini School in 1991 together with Antonio Di Giacomo (Experimental Immunologist working at the Monaldi Hospital in Naples, Italy), Melchiorre Brai (Professor of Immunology at the University of Palermo, Italy), Giovan Battista Ferrara (Professor of Human Genetics at the Federico II University of Naples, Italy), Albert Nisonoff (Professor of Biology at Brandeis University, in Waltham, Massachusetts, USA), and Ciro Manzo (Head of Immunology Department at the Istituto Pascale in Naples, Italy). The choice of Naples as Ceppellini School headquarters was no accident. This city had experienced in 1799, during the Neapolitan Republic, an unsuccessful attempt to gain freedom from the constraints of a tyrannical monarchy, and to promote a new political organization. This attempt was brutally suppressed. The choice of Naples symbolically reflected the will to propel the freedom of scientific minds according to the School's motto "*non multa sed multum*" ("not many but much," i.e., quality rather than quantity) (3). Over the years, the Ceppellini School has been attracting and engaging large numbers of international young scientists to the field of immunology.

## FOCUSING ON MHC MOLECULES

Serafino Zappacosta had a strong interest in the major histocompatibility complex (MHC) genes and proteins. He shared his fascination for the MHC with his laboratory members, with whom he investigated a variety of topics in the MHC field, including the regulation of HLA expression in tumor cells (4, 5), the association between HLA alleles and diseases in Southern Italy (6, 7), the influences of MHC class I (MHC-I) on tumor killing by NK cells (8), and the cytokine-mediated regulation of MHC-I expression (9). His laboratory also worked in collaboration with many international teams to study HLA polymorphisms, and participated in collaborative workshops on HLA typing, that included the International Histocompatibility Workshop in 1991 in Yokohama, Japan (10). This interest in MHC also led to the name of the Ceppellini School, after the immuno-geneticist Ruggero Ceppellini, who had greatly contributed with his pioneer work to the understanding of the genetic bases of HLA polymorphisms, and coined the term "haplotype" (11).

## MHC POLYMORPHISM AND THE MEDITERRANEAN AREA

Before molecular genetics could rely on modern technology to collect a tremendous amount of information, many data about the genetic background of different human populations were based on the analysis of products of polymorphic loci including HLA. Analyzing the HLA system at the end of the '70s, Zappacosta, together with Mario De Felice, Michele Fiore and Giovan Battista Ferrara (12), found significant differences between people living in Northern Italy and the population of Campania (in Southern Italy). Significant similarities were noticed between Mediterranean and Middle Eastern populations and people from Campania, confirming

that the genetic background of the Italian population is highly mixed. Furthermore, a peculiar association between HLA alleles and congenital adrenal hyperplasia was found by Serafino Zappacosta, Michele Maio, Mario De Felice and Rossella Valentino in the Southern Italian population (6). These studies were performed at the time when Luigi Cavalli-Sforza investigated the selection of advantageous alleles in HLA locus and other polymorphic loci, whereby those findings served to illustrate migration patterns of human populations (13).

Further studies on MHC polymorphism were performed over the years by Giuseppina Ruggiero, Giuseppe Matarese, Giuseppe Terrazzano, and others in the Zappacosta's laboratory. They demonstrated a link between HLA alleles and susceptibility to autoimmune/infectious diseases in Southern Italy (7, 14). Other Zappacosta's team members, including Michele Maio, Luigi Del Vecchio, and Mario De Felice, documented the association between HLA-DR alleles and thyroid carcinoma (15). In the 80's, Michele Maio, Luigi Del Vecchio, Giuseppina Ruggiero, Mario De Felice and others of the Zappacosta's laboratory investigated MHC-I expression as a prognostic factor in breast cancer (5). Antonio Pinto, Michele Maio and others showed that HLA-DR expression by myeloid leukemia cells was modulated by anti-neoplastic drugs (4).

## MHC MOLECULES AND THE REGULATION OF NK CELL RESPONSE

In the late '80, Silvia Fontana, Luigi Racioppi and Ennio Carbone in Zappacosta's team investigated the link between retroviral infection and MHC-I expression by tumor cells, using virus-induced rat thyroid adenocarcinomas as an experimental model (16, 17). At the time, state-of-the-art techniques for these studies included tissue culture, microscopy, immunofluorescence, and cytofluorimetry, that had only become available a few years earlier. Silvia Fontana, Ennio Carbone, and others in the team showed that tumor transformation modulated MHC-I expression by rat tumor cells, and that rat Large Granular Lymphocytes (LGL) killed more effectively tumor cells having low MHC-I expression (8). These observations were puzzling at that time. In 1987—just a few years before these studies—, Bjorkman and colleagues had solved the HLA A2 crystallographic structure (18, 19), and a lot of attention was concentrated on the TCR/MHC-I molecular interaction, and its role in immunity. On the other hand, Ennio Carbone of the Zappacosta's team was fascinated by pioneering studies on the inhibitory signals provided by MHC-I to Natural Killer (NK) cells [the "missing self" hypothesis, formulated by Klas Karre in 1981 (20)]. Together with Antonio La Cava, Giuseppe Terrazzano and others, the Zappacosta group's contributions to the NK field spanned from MHC-I mediated regulation of NK cell cytotoxicity in rat tumor models (8, 21), to NK cell inhibition induced by soluble HLA-I (22), to new findings on the role of NK cells in human tumor immunosurveillance (23), NK/ dendritic cells cross-talk (24, 25), and CD1-mediated inhibition of NK cytotoxicity (26). Giuseppe Terrazzano in the group investigated the effects of IL-10 on MHC-I expression and on the antigen presenting machinery



(9), demonstrating a pathological role of gliadin in the NK cell/dendritic cells cross-talk (27). Sadly, this publication was the last one that included Serafino Zappacosta's co-authorship.

## MHC MOLECULES AND THE REGULATION OF T CELL RESPONSE

In the early 90's, it was well-established that the function of MHC molecules was to bind and present antigenic peptides to T cells (28, 29). The molecular bases of this phenomenon had been largely resolved by several independent studies (30–35). However, looking out of this canonical box, it was possible to hypothesize that, in addition to binding TCR and CD4, MHC class II (MHC-II) molecules might also interact with other cell surface molecules, and in turn regulate the activation of immune cells. Within this context, Zappacosta's group aimed to identify non-canonical functions of HLA-II molecules (36–39). A large panel of monoclonal antibodies (mAbs) directed against different epitopes of HLA-II molecules provided by Soldano Ferrone (40) were instrumental for these studies, that were performed in *in vitro* models of polyclonal T-cell proliferation, induced by either phytohemagglutinin (PHA) or anti-CD3 mAb (41, 42). Although the presence of antigen presenting cells (APC) was required to achieve full T-cell activation, HLA-II antigen presenting function was largely dispensable in these models, thus offering a unique opportunity to evaluate non-canonical functions of HLA-II molecules.

Giuseppina Ruggiero and Luigi Racioppi in Zappacosta's team, in collaboration with Ciro Manzo, initiated these pioneer studies on HLA-II molecules, and demonstrated that the incubation of autologous monocyte-derived macrophages with mAbs directed against non-polymorphic determinants of HLA-II molecules exerted a remarkable inhibitory effect on T cell activation (37, 38, 43). This result suggested that MHC-II molecules expressed on the APC could interact not only with the TCR and CD4, but also with additional ligand(s)—at the time unknown—, expressed on the T cell surface. Interestingly, in 1996 one of these hypothetical ligand was identified by Huard et al. (44), who demonstrated the ability of CD223 (aka LAG-3) to bind MHC-II molecules. In the last two decades, the relevance of LAG-3/MHC-II signaling has been confirmed by several independent studies, being this molecular interaction involved in a variety of immuno-regulatory circuits (45).

D'Oro and Di Rosa from the Zappacosta's team further explored non-canonical functions of MHC-II molecules expressed by activated human T cells. These studies were based on the general hypothesis that HLA-II molecules might transduce intracellular signals, and in turn finely tune T cell response to antigen(s) and cytokines. The results confirmed this possibility, showing that cross-linking of HLA-II on activated T cells was sufficient to induce inositol triphosphates (IP3) accumulation, protein kinase C (PKC) activation, and, ultimately, enhanced T cell proliferation (10, 39). Of note, the ability of MHC-II molecules to transduce intracellular signals has also been recognized in B cells (46–48), and more recently a cell-intrinsic contribution of MHC-II expression has been shown in the B cell development in the bone marrow (49).

As a note, the findings by Zappacosta's group on non-canonical functions of the HLA molecules have relevant, and still largely unexplored, implications in the regulation of the human immune response. For example, high expression of LAG-3 by T regulatory (Treg) cells suggests that LAG-3/MHC-II complexes might play an important role in the bi-directional signaling triggered by Treg/ T effector cell interactions (50, 51). In this sense, an increasing number of studies has pointed to LAG-3/MHC-II interaction as an attractive druggable target to treat autoimmune diseases, stimulate anti-cancer immune response (52), and revert resistance to anti-PD-1 immunotherapy (53).

## MHC AND BEYOND

Giuseppe Matarese devoted his experimental efforts on the innovative hypothesis that nutrient-energy-sensing pathways might represent a powerful tool to control immunological self-tolerance. He showed that leptin, a hormone critically involved in energy balance and body weight regulation, acts as a strong immune-modulator, that influences the susceptibility to infection and autoimmunity (54, 55). Leptin levels inversely correlated with regulatory T cell number in multiple sclerosis patients (56), and a direct link between leptin and regulatory T cell anergy was established (57). This observation, that was further developed in subsequent studies performed by Giuseppe Matarese and his group, in collaboration with Antonio La Cava, was the result of frequent, endless, unforgettable evening lab discussions with Serafino Zappacosta.

After training with Serafino Zappacosta with a focus on the immune-modulating properties of MHC molecules, the team members subsequently developed new hypotheses and investigations in diverse directions, ranging from the study of the fundamental mechanisms of immune regulation and immunological memory, to autoimmunity, cancer immunotherapy and vaccinology (58–66). Serafino Zappacosta kept to enthusiastically follow the progress of the past members of his team after they left to start their independent careers. Many of them remained involved over the years in the Ceppellini School activities, either as faculty members or as components of the board of directors, maintaining the School as an arena of continuous scientific education and dynamic discussion.

## CONCLUSIONS

The review summarizes the legacy left by Serafino Zappacosta to his collaborators who, albeit with different individual perspectives and at a different degree, continued to work on the MHC, looking at these molecules as a window of opportunity to comprehend the complexity of the immune response, rather than merely looking at them as antigen presenting molecules.

After the death of Serafino Zappacosta in 2006, Silvia Fontana became the President of the Ceppellini School, and Ennio Carbone, Giuseppe Matarese, Francesca Di Rosa, Giuseppina Ruggiero, Giuseppe Terrazzano and other previous collaborators of the Zappacosta's team continued to organize advanced international immunology courses, together with the

long-standing Serafino Zappacosta's collaborators and friends Antonio Di Giacomo, who co-founded the School in 1991 (3), Elizabeth Simpson, who organized the first Ceppellini School Course in 1992 on bone marrow transplantation (67), and the newly recruited Ceppellini School Scientific Director Stefan Kaufmann (68). A new type of event, the Serafino Zappacosta Memorial Conferences, was initiated in 2007. Since 2010, this event has been held in the newly inaugurated "Serafino Zappacosta" Auditorium of the Federico II University of Naples. All these activities were made possible by the excellent work of the Ceppellini School Scientific Secretary Tricia Reynolds.

To conclude, the continuation of the activities of the Ceppellini School not only allows an unceasing engagement of new young bright minds to the fascinating field of immunology, but also keeps alive Serafino Zappacosta's dream that intellectual

freedom can be shared without boundaries for the benefit of younger generations.

## AUTHOR CONTRIBUTIONS

All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

## DEDICATION

This article is dedicated to the memory of Luigi Del Vecchio, past member of Zappacosta's team and full professor of Clinical Biochemistry and Molecular Biology, Dipartimento di Medicina Molecolare e Biotecnologie Mediche, Università degli Studi di Napoli "Federico II," Naples, Italy at the time of his premature death in 2018.

## REFERENCES

1. François de Salignac de La Mothe-Fénelon. *Les Aventures de Télémaque* (1699).
2. La Cava A, De Felice M. In memoriam: Serafino Zappacosta (1935–2006). *Tissue Antigens*. (2006) 68:279. doi: 10.1111/j.1399-0039.2006.00686.x
3. Di Giacomo A. The Ruggero Ceppellini advanced school of immunology and the neapolitan scientific renaissance. *Front Immunol*. (2019) 10:1494. doi: 10.3389/fimmu.2019.01494
4. Pinto A, Maio M, Attadia V, Zappacosta S, Cimino R. Modulation of HLA-DR antigens expression in human myeloid leukaemia cells by cytarabine and 5-aza-2'-deoxycytidine. *Lancet*. (1984) 2:867–8. doi: 10.1016/S0140-6736(84)90900-0
5. Iaffaioli RV, Maio M, Ruggiero G, De Felice M, Ungaro A, Del Vecchio L, et al. HLA and prognostic factors in primary breast cancer. *Int J Cancer*. (1985) 35:581–5. doi: 10.1002/ijc.2910350503
6. Zappacosta S, Maio M, de Felice M, Valentino R. The association between congenital adrenal hyperplasia and HLA in Southern Italy. *Ann N Y Acad Sci*. (1985) 458:46–51. doi: 10.1111/j.1749-6632.1985.tb14589.x
7. Ruggiero G, Cosentini E, Zanzi D, Sanna V, Terrazzano G, Matarese G, et al. Allelic distribution of human leucocyte antigen in historical and recently diagnosed tuberculosis patients in Southern Italy. *Immunology*. (2004) 111:318–22. doi: 10.1111/j.1365-2567.2004.01811.x
8. Carbone E, Racioppi L, La Cava A, Portella G, Velotti F, Zappacosta S, et al. NK and LAK susceptibility varies inversely with target cell MHC class I antigen expression in a rat epithelial tumour system. *Scand J Immunol*. (1991) 33:185–94. doi: 10.1111/j.1365-3083.1991.tb03748.x
9. Terrazzano G, Romano M, Turco FM, Salzano CS, Ottaiano A, Venuta S, et al. HLA class I antigen downregulation by interleukin (IL)-10 is predominantly governed by NK-kappaB in the short term and by TAP1+2 in the long term. *Tissue Antigens*. (2000) 55:326–32. doi: 10.1034/j.1399-0039.2000.550406.x
10. D'Oro U, et al. Major Histocompatibility Complex class II molecule transduce activation signal in human T blasts. In: Tsuji K, Aizawa M, and Sasazuki T, editors. *HLA 1991, Proceedings of the Eleventh International Histocompatibility Workshop and Conference held in Yokohama, Japan, 6–13 November, 1991*. (1992). p. 632–5.
11. Bodmer W. Ruggero Ceppellini: a perspective on his contributions to genetics and immunology. *Front Immunol*. (2019) 10:1280. doi: 10.3389/fimmu.2019.01280
12. Zappacosta S, De Felice M, Fiore M, Ferrara GB. The HLA system in the Campania region: a genetic study. *Tissue Antigens*. (1980) 16:286–93. doi: 10.1111/j.1399-0039.1980.tb00308.x
13. Piazza A, Menozzi P, Cavalli-Sforza LL. The HLA-A,B gene frequencies in the world: migration or selection. *Hum Immunol*. (1980) 1:297–304. doi: 10.1016/0198-8859(80)90105-6
14. Montanaro D, Sanna V, Matarese G, Larby BB, Racioppi L, Carrieri PB, et al. The fine specificity of human T cell lines towards myelin basic protein peptides in southern Italian multiple sclerosis patients. *Clin Exp Immunol*. (2001) 123:288–93. doi: 10.1046/j.1365-2249.2001.01457.x
15. Panza N, Del Vecchio L, Maio M, De Felice M, Lombardi G, Minozzi M, et al. Strong association between an HLA-DR antigen and thyroid carcinoma. *Tissue Antigens*. (1982) 20:155–8. doi: 10.1111/j.1399-0039.1982.tb00340.x
16. Fontana S, Del Vecchio L, Racioppi L, Carbone E, Pinto A, Colletta G, et al. Expression of major histocompatibility complex class I antigens in normal and transformed rat thyroid epithelial cell lines. *Cancer Res*. (1987) 47:4178–83.
17. Racioppi L, Carbone E, Grieco M, Del Vecchio L, Berlingieri MT, Fusco A, et al. The relationship of modulation of major histocompatibility complex class I antigens to retrovirus transformation in rat cell lines. *Cancer Res*. (1988) 48:3816–21.
18. Bjorkman PJ, Saper MA, Samraoui B, Bennett WS, Strominger JL, Wiley DC. The foreign antigen binding site and T cell recognition regions of class I histocompatibility antigens. *Nature*. (1987) 329:512–8. doi: 10.1038/329512a0
19. Bjorkman PJ, Saper MA, Samraoui B, Bennett WS, Strominger JL, Wiley DC. Structure of the human class I histocompatibility antigen, HLA-A2. *Nature*. (1987) 329:506–12. doi: 10.1038/329506a0
20. Karre K. Natural killer cell recognition of missing self. *Nat Immunol*. (2008) 9:477–80. doi: 10.1038/ni0508-477
21. La Cava A, Carbone E, Moscarella A, Barcova M, Salzano S, Zappacosta S, et al. A novel strategy of c-myc oncogene in NK activity regulation not related to the W6/32 MHC class-I epitope. *Int J Cancer*. (1994) 58:123–8. doi: 10.1002/ijc.2910580120
22. Carbone E, Terrazzano G, Colonna M, Tuosto L, Piccolella E, Franksson L, et al. Natural killer clones recognize specific soluble HLA class I molecules. *Eur J Immunol*. (1996) 26:683–9. doi: 10.1002/eji.1830260326
23. Carbone E, Ruggiero G, Terrazzano G, Palomba C, Manzo C, Fontana S, et al. A new mechanism of NK cell cytotoxicity activation: the CD40–CD40 ligand interaction. *J Exp Med*. (1997) 185:2053–60. doi: 10.1084/jem.185.12.2053
24. Carbone E, Terrazzano G, Ruggiero G, Zanzi D, Ottaiano A, Manzo C, et al. Recognition of autologous dendritic cells by human NK cells. *Eur J Immunol*. (1999) 29:4022–9. doi: 10.1002/(SICI)1521-4141(199912)29:12<4022::AID-IMMU4022>3.0.CO;2-O
25. Terrazzano G, Pisanti S, Grimaldi S, Sica M, Fontana S, Carbone E, et al. Interaction between natural killer and dendritic cells: the role of CD40, CD80 and major histocompatibility complex class I molecules in cytotoxicity induction and interferon-gamma production. *Scand J Immunol*. (2004) 59:356–62. doi: 10.1111/j.0300-9475.2003.01387.x

26. Carbone E, Terrazzano G, Melián A, Zanzi D, Moretta L, Porcelli S, et al. Inhibition of human NK cell-mediated killing by CD1 molecules. *J Immunol.* (2000) 164:6130–7. doi: 10.4049/jimmunol.164.12.6130
27. Terrazzano G, Sica M, Gianfrani C, Mazzarella G, Maurano F, De Giulio B, et al. Gliadin regulates the NK-dendritic cell cross-talk by HLA-E surface stabilization. *J Immunol.* (2007) 179:372–81. doi: 10.4049/jimmunol.179.1.372
28. Germain RN. Immunology. The ins and outs of antigen processing and presentation. *Nature.* (1986) 322:687–9. doi: 10.1038/322687a0
29. Braciale TJ, Morrison LA, Sweetser MT, Sambrook J, Gething MJ, Braciale VL. Antigen presentation pathways to class I and class II MHC-restricted T lymphocytes. *Immunol Rev.* (1987) 98:95–114. doi: 10.1111/j.1600-065X.1987.tb00521.x
30. Fremont DH, Matsumura M, Stura EA, Peterson PA, Wilson IA. Crystal structures of two viral peptides in complex with murine MHC class I H-2Kb. *Science.* (1992) 257:919–27. doi: 10.1126/science.1323877
31. Zhang W, Young AC, Imarai M, Nathenson SG, Sacchetti JC. Crystal structure of the major histocompatibility complex class I H-2Kb molecule containing a single viral peptide: implications for peptide binding and T-cell receptor recognition. *Proc Natl Acad Sci USA.* (1992) 89:8403–7. doi: 10.1073/pnas.89.17.8403
32. Madden DR, Gorga JC, Strominger JL, Wiley DC. The three-dimensional structure of HLA-B27 at 2.1 Å resolution suggests a general mechanism for tight peptide binding to MHC. *Cell.* (1992) 70:1035–48. doi: 10.1016/0092-8674(92)90252-8
33. Silver ML, Guo HC, Strominger JL, Wiley DC. Atomic structure of a human MHC molecule presenting an influenza virus peptide. *Nature.* (1992) 360:367–9. doi: 10.1038/360367a0
34. Stern LJ, Brown JH, Jardetzky TS, Gorga JC, Urban RG, Strominger JL, et al. Crystal structure of the human class II MHC protein HLA-DR1 complexed with an influenza virus peptide. *Nature.* (1994) 368:215–21. doi: 10.1038/368215a0
35. Jardetzky TS, Brown JH, Gorga JC, Stern LJ, Urban RG, Chi YI, et al. Three-dimensional structure of a human class II histocompatibility molecule complexed with superantigen. *Nature.* (1994) 368:711–8. doi: 10.1038/368711a0
36. Ruggiero G, Manzo C, Fontana S, Scala G, Pirozzi G, Ferrone S, et al. Inhibition by anti-HLA class II monoclonal antibodies of monocyte-dependent T cell proliferation induced by monoclonal antibody OKT3. *Eur J Immunol.* (1987) 17:1585–92. doi: 10.1002/eji.1830171110
37. Racioppi L, Moscarella A, Ruggiero G, Manzo C, Ferrone S, Fontana S, et al. Inhibition by anti-HLA class II monoclonal antibodies of monoclonal antibody OKT3-induced T cell proliferation. Studies at the mRNA level. *J Immunol.* (1990) 145:3635–40.
38. Ruggiero G, Racioppi L, Manzo C, Pirozzi G, D'Oro U, Ferrone S, et al. HLA class II molecules on monocytes regulate T cell proliferation through physical interaction in the CD3 activation pathway. *Eur J Immunol.* (1991) 21:29–33. doi: 10.1002/eji.1830210106
39. Di Rosa F, D'Oro U, Ruggiero G, Racioppi L, Acquaviva A, Ferrone S, et al. HLA class II molecules transduce accessory signals affecting the CD3 but not the interleukin-2 activation pathway in T blasts. *Hum Immunol.* (1993) 38:251–60. doi: 10.1016/0198-8859(93)90552-C
40. Quaranta V, Pellegrino MA, Ferrone S. Serologic and immunochemical characterization of the specificity of four monoclonal antibodies to distinct antigenic determinants expressed on subpopulations of human Ia-like antigens. *J Immunol.* (1981) 126:548–52.
41. Gallagher RB, Cambier JC. Signal transmission pathways and lymphocyte function. *Immunol Today.* (1990) 11:187–9. doi: 10.1016/0167-5699(90)90078-N
42. Samelson LE, Fletcher MC, Ledbetter JA, June CH. Activation of tyrosine phosphorylation in human T cells via the CD2 pathway. Regulation by the CD45 tyrosine phosphatase. *J Immunol.* (1990) 145:2448–54.
43. Manzo C, Ruggiero G, del Vecchio L, Racioppi L, Pirozzi G, Temponi M, et al. Monoclonal antibody OKT3-induced T cell proliferation: differential role of HLA class II determinants expressed by T cells and monocytes. *Cell Immunol.* (1990) 125:79–91. doi: 10.1016/0008-8749(90)90064-X
44. Huard B, Prigent P, Pages F, Bruniquel D, Triebel F. T cell major histocompatibility complex class II molecules down-regulate CD4+ T cell clone responses following LAG-3 binding. *Eur J Immunol.* (1996) 26:1180–6. doi: 10.1002/eji.1830260533
45. Lui Y, Davis SJ. LAG-3: a very singular immune checkpoint. *Nat Immunol.* (2018) 19:1278–9. doi: 10.1038/s41590-018-0257-1
46. Wade WF, Davoust J, Salameo J, Andre P, Watts TH, Cambier JC. Structural compartmentalization of MHC class II signaling function. *Immunol Today.* (1993) 14:539–46. doi: 10.1016/0167-5699(93)90184-M
47. Al-Daccak R, Mooney N, Charron D. MHC class II signaling in antigen-presenting cells. *Curr Opin Immunol.* (2004) 16:108–13. doi: 10.1016/j.coi.2003.11.006
48. Harton J, Jin L, Hahn A, Drake J. Immunological functions of the membrane proximal region of MHC class II molecules. *Fl000Res.* (2016) 5:1–12. doi: 10.12688/fl000research.7610.1
49. Merckenschlager J, Eksmond U, Danelli L, Attig J, Young G, R, Nowosad C, et al. MHC class II cell-autonomously regulates self-renewal and differentiation of normal and malignant B cells. *Blood.* (2019) 133:1108–18. doi: 10.1182/blood-2018-11-885467
50. Zhang Q, Chikina M, Szymczak-Workman AL, Horne W, Kolls JK, Vignali KM, et al. LAG3 limits regulatory T cell proliferation and function in autoimmune diabetes. *Sci Immunol.* (2017) 2:eaa4569. doi: 10.1126/sciimmunol.aah4569
51. Thaker Y, Andrews LP, Workman CJ, Vignali DAA, Sharpe AH. Treg-specific LAG3 deletion reveals a key role for LAG3 in regulatory T cells to inhibit CNS autoimmunity. *J Immunol.* (2018) 200:101.7.
52. Shapiro M, Herishanu Y, Katz BZ, Dezorella N, Sun C, Kay S, et al. Lymphocyte activation gene 3: a novel therapeutic target in chronic lymphocytic leukemia. *Haematologica.* (2017) 102:874–82. doi: 10.3324/haematol.2016.148965
53. Long L, Zhang X, Chen F, Pan Q, Phiphatwatchara P, Zeng Y, et al. The promising immune checkpoint LAG-3: from tumor microenvironment to cancer immunotherapy. *Genes Cancer.* (2018) 9:176–89. doi: 10.18632/genesandcancer.180
54. Lord GM, Matarese G, Howard JK, Baker RJ, Bloom SR, Lechler RI. Leptin modulates the T-cell immune response and reverses starvation-induced immunosuppression. *Nature.* (1998) 394:897–901. doi: 10.1038/29795
55. Sanna V, Di Giacomo A, La Cava A, Lechler RI, Fontana S, Zappacosta S, et al. Leptin surge precedes onset of autoimmune encephalomyelitis and correlates with development of pathogenic T cell responses. *J Clin Invest.* (2003) 111:241–50. doi: 10.1172/JCI200316721
56. Matarese G, Carrieri PB, La Cava A, Perna F, Sanna V, De Rosa V, et al. Leptin increase in multiple sclerosis associates with reduced number of CD4(+)CD25+ regulatory T cells. *Proc Natl Acad Sci USA.* (2005) 102:5150–5. doi: 10.1073/pnas.0408995102
57. De Rosa V, Procaccini C, Cali G, Pirozzi G, Fontana S, Zappacosta S, et al. A key role of leptin in the control of regulatory T cell proliferation. *Immunity.* (2007) 26:241–55. doi: 10.1016/j.immuni.2007.01.011
58. Silberschmidt D, Rodriguez-Mallon A, Mithboakar P, Cali G, Amendola E, Sanges R, et al. *In vivo* role of different domains and of phosphorylation in the transcription factor Nkx2-1. *BMC Dev Biol.* (2011) 11:9. doi: 10.1186/1471-213X-11-9
59. Procaccini C, Carbone E, Di Silvestre D, Brambilla F, De Rosa V, Galgani M, et al. The proteomic landscape of human *ex vivo* regulatory and conventional T cells reveals specific metabolic requirements. *Immunity.* (2016) 44:712. doi: 10.1016/j.immuni.2016.02.022
60. Ali TH, Pisanti S, Ciaglia E, Mortarini R, Anichini A, Garofalo C, et al. Enrichment of CD56(dim)KIR + CD57 + highly cytotoxic NK cells in tumour-infiltrated lymph nodes of melanoma patients. *Nat Commun.* (2014) 5:5639. doi: 10.1038/ncomms6639
61. Racioppi L, Nelson ER, Huang W, Mukherjee D, Lawrence SA, Lento W, et al. CaMKK2 in myeloid cells is a key regulator of the immune-suppressive microenvironment in breast cancer. *Nat Commun.* (2019) 10:2450. doi: 10.1038/s41467-019-10424-5
62. Tulli L, Cattaneo F, Vinot J, Baldari CT, D'Oro U. Src family kinases regulate interferon regulatory factor 1 K63 ubiquitination following activation by TLR7/8 vaccine adjuvant in human monocytes and B cells. *Front Immunol.* (2018) 9:330. doi: 10.3389/fimmu.2018.00330



63. Di Rosa F. Maintenance of memory T cells in the bone marrow: survival or homeostatic proliferation? *Nat Rev Immunol.* (2016) 16:271. doi: 10.1038/nri.2016.31
64. Di Giacomo AM, Covre A, Finotello F, Rieder D, Danielli R, Sigalotti L, et al. Guadecitabine plus ipilimumab in unresectable melanoma: the NIBIT-M4 clinical trial. *Clin Cancer Res.* (2019) 25:7351–62. doi: 10.1158/1078-0432.CCR-19-1335
65. Liu Y, Liu A, Iikuni N, Xu H, Shi FD, La Cava A. Regulatory CD4+ T cells promote B cell anergy in murine lupus. *J Immunol.* (2014) 192:4069–73. doi: 10.4049/jimmunol.1302897
66. Giovazzino A, Leone S, Rubino V, Palatucci AT, Cerciello G, Alfinito F, et al. Reduced regulatory T cells (Treg) in bone marrow preferentially associate with the expansion of cytotoxic T lymphocytes in low risk MDS patients. *Br J Haematol.* (2019) 185:357–60. doi: 10.1111/bjh.15496
67. Simpson E, Dazzi F. Bone marrow transplantation 1957–2019. *Front Immunol.* (2019) 10:1246. doi: 10.3389/fimmu.2019.01246
68. Kaufmann SHE. Immunology's coming of age. *Front Immunol.* (2019) 10:684. doi: 10.3389/fimmu.2019.01214

**Conflict of Interest:** UD is an employee of the GSK group of companies and holds restricted shares of the GSK group of companies.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The reviewer DB declared a shared affiliation, with no collaboration, with several of the authors, MD, FDR, SF, to the handling editor at time of review.

Copyright © 2020 Carbone, De Felice, Di Rosa, D'Oro, Fontana, La Cava, Maio, Matarese, Racioppi, Ruggiero and Terrazzano. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.



# Ruggero Ceppellini: A Perspective on His Contributions to Genetics and Immunology

Walter Bodmer\*

Department of Oncology, Weatherall Institute of Molecular Medicine, University of Oxford, Oxford, United Kingdom

Ruggero Ceppellini, who died at the age of 71 in 1988, was one of the most stimulating and original human geneticists of his generation (1). Ceppellini's outstanding contributions to the genetics of the human blood groups, immunoglobulin allotypes and the HLA system epitomize the study of immunogenetics. By using his considerable skills and insights to unravel the interpretation of the serological data, he made significant contributions to immunology. He is remembered especially for his incisive contributions to the development of the genetics of the HLA system and its nomenclature, including, in particular, his introduction of the term "haplotype," now widely used by geneticists throughout the world, most of whom are unlikely to be aware of its origins.

**Keywords:** Ceppellini, HLA, MLC, haplotype, transplantation

## OPEN ACCESS

### Edited by:

Francesca Di Rosa,  
Istituto di Biologia e Patologia  
Molecolari (IBPM), Consiglio Nazionale  
Delle Ricerche (CNR), Italy

### Reviewed by:

Gerhard Opelz,  
Heidelberg University of Education,  
Germany  
Rita Carsetti,  
Bambino Gesù Children Hospital  
(IRCCS), Italy

### \*Correspondence:

Walter Bodmer  
walter.bodmer@hertford.ox.ac.uk

### Specialty section:

This article was submitted to  
Molecular Innate Immunity,  
a section of the journal  
Frontiers in Immunology

**Received:** 21 February 2019

**Accepted:** 20 May 2019

**Published:** 05 June 2019

### Citation:

Bodmer W (2019) Ruggero Ceppellini:  
A Perspective on His Contributions to  
Genetics and Immunology.  
Front. Immunol. 10:1280.  
doi: 10.3389/fimmu.2019.01280

## EARLY BLOOD GROUP DISCOVERIES

Born during the first world war, Ceppellini was caught up by military service in the second world war and so could not finish his medical studies until after the war had ended. During his service as a sergeant in World War II, Ceppellini was captured by the British and taken as a prisoner of war in Palestine, where the charismatic physician and human geneticist, Chaim Sheba, took him on as a medical orderly because of his medical background. Many years later, when Ceppellini was attending a human genetics meeting in Israel, Chaim Sheba greeted him as "Sergeant Ceppellini." Perhaps it was that brush with genetics that stimulated his interest in the field and led to his appointment, through the influence of Luca Cavalli-Sforza, to a position in the Istituto Sieroterapico Milanese, a blood bank associated with the University of Milan. Cavalli-Sforza, although Ceppellini's junior by 5 years, was already becoming established as a geneticist and was a major influence on Ceppellini's future career.

In 1954, Ceppellini was invited to work in the Institute for the Study of Human Variation at Columbia University in New York, where he came under the influence of L. C. Dunn and made his first significant contribution to immunogenetics. He showed, through a careful family and population based study, that the Rh variant D<sup>u</sup> was actually due to a reduced expression of D when associated in heterozygotes with the combination Cde (2).

Shortly after his return to Italy in 1959, Ceppellini made his second outstanding contribution to the blood grouping field. This was his interpretation of the Lewis b phenotype as an interaction between the secretor and Lewis genes, and his interpretation of the ABO and Lewis blood groups in terms of a form of metabolic sequence involving successive additions of sugars. His model, based entirely on a genetic interpretation of the data, showed remarkable insight and was abundantly confirmed by the studies by Morgan, Watkins, Kabat, and others of the oligosaccharide structures of these blood group determinants and the eventual identification of the two fucosyltransferase genes (3, 4).

## MALARIA, THALASSAEMIA AND THE IMMUNOGLOBULIN (GM) ALLOTYPES

In the early 1950s, Ceppellini had started a systematic study in Sardinia to correlate the distribution of thalassaemia, which was common there in the low lying villages, with the distribution of malaria that had been established in Sardinia by an extensive study of school children. Through this, he provided some of the first clear evidence of the correlation between thalassaemia and resistance to malaria, showing that its frequency was highest in those areas where the incidence of malaria had been greatest (5). His work there stimulated a long and continuing tradition of studies in human genetics in Sardinia, carried forward especially by his close friend and colleague Marcello Siniscalco.

My initial contact with Ruggero Ceppellini was established through Luca Cavalli-Sforza, in the early 1960s, because of my interest in the studies of thalassaemia and malaria as a model of natural selection in human populations. I, and my population genetics colleagues at Stanford University, invited him as a population geneticist to discuss this work. In characteristic manner, just before he was due to arrive, we received a telegram saying that unfortunately, after all, he was not able to come.

Stimulated by his contact with Henry Kunkel during his time in New York, Ceppellini took up the study of the Gm types. Kunkel was a pioneer of the study of the single immunoglobulins produced by myelomas, while Grubb had shown in 1956 (6) that there were inherited serologically detectable differences in the immunoglobulins, which were called Gm allotypes, G for immunoglobulin G and m for marker. By the early 1960s, the basic two chain structure of the immunoglobulins and the distinction between constant and variable regions had been elucidated, so that it became clear that these allotypes were inherited variations in the IgG heavy chain constant regions.

In a major comprehensive and inciteful review of the Gm allotypes, published in Italian in the proceedings of the 1966 meeting of the Italian Genetics Association (7), Ceppellini provided what was at that time the clearest interpretation of the Gm types as a complex genetic system. This was in some ways analogous to the Rhesus blood group system as interpreted by R. A. Fisher, on which he had published earlier with L. C. Dunn. His interpretation was in terms of "haplotypes" (an expression first used in this paper), which determined different combinations of Gm types, and their varying frequencies in different populations. He appreciated the possibility of the creation of new haplotypes by recombination between existing haplotypes and extended some of the formulae, which I had developed for the analysis of two locus-linked systems, to the estimation of haplotype frequencies. This notable paper is hardly ever quoted because of its publication in Italian in a more or less inaccessible journal. It is also, to my mind, odd that, as far as I am aware, this is his only publication on the Gm types. Perhaps that is because it was at this time that he started on his major interest in what became the HLA system.

## HLA, HAPLOTYPES, CELLULAR ASSAYS, AND MONOCLONAL ANTIBODIES

### Early White Cell Agglutination Serology

Jean Dausset pioneered the testing of sera from multiply transfused patients against white blood cells from arbitrary volunteer donors using an agglutination reaction. His aim was to establish whether these reactions could be interpreted to define inherited blood group like determinants on white rather than red blood cells. The initial results, not surprisingly in retrospect given the now known complexity of the HLA system, were very confusing.

Ceppellini suggested to Dausset in 1956 that he should compare the reactions on white cells from pairs of identical (monozygous or MZ) twins with those on pairs of non-identical (dizygous or DZ) twins. Then, if the agglutination reactions reflected inherited determinants, all the reactions should, subject to experimental error, be the same on each of the MZ twin pairs, while this would not necessarily be the case for the DZ twins. Dausset and Brecy published a short note in *Nature* in 1957 (8) confirming this prediction, following which, Dausset described in 1958 a putative first antigen, which he called MAC (9), and for which he shared the Nobel Prize in 1980.

The serology of the white blood cell antigens did not, however, progress any further until Jon van Rood and Rose Payne, independently in 1958, showed that sera from multiparous women contained antibodies against the white cells of their offspring that could be detected by agglutination assays. These were produced by fetal-maternal stimulation, just as in the case of the Rh blood groups and, being limited to the difference between mother and father, were much less complex than sera obtained from multiply transfused individuals. By the early to mid 1960s, Ceppellini had joined the initial group of workers in this field. In the first of the International Histocompatibility Testing workshops, organized by Bernard Amos in 1964, Ceppellini played a major role in analyzing the data and promoting the need for improved reproducibility of the testing techniques as well as for inter-laboratory comparisons of results. [For details of the early history of the HLA field, as described by its pioneers, see (10)].

As a geneticist, Ceppellini appreciated the importance of family studies, and so organized, with his colleagues, the third International Histocompatibility Testing Workshop in Turin in 1967 around the theme of a family study. While by that time it was clear that there were at least two, probably closely linked, loci for the white blood cell determinants being described, this had not been definitively established by family studies. He provided the families and we came with our sera and different technologies to test white blood cells from his family members. Ceppellini expressed to the press his amusement at seeing an erstwhile mathematician sitting at the bench looking down a microscope. The aim of the collaborative on site study was to see whether all the types being defined by the different participating laboratories were inherited together—and they were!

This, then, was the first clear-cut establishment of the HLA system as a set of closely linked genes inherited together in

a way that was analogous to the Rhesus blood groups and immunoglobulin Gm allotypes.

## Haplotype

It was notably at this workshop in 1967 that Ceppellini first really introduced the term “haplotype,” though he had used it the previous year in the Italian Gm allotype review, as already mentioned. His description was as follows:

“If a new term can be introduced without increasing confusion, it is suggested to substitute phenogroup with haplotype (haploid, from  $\alpha\pi\lambda\acute{o}\varsigma$ , single); in fact, the name should convey the concept that the haplotype is not an observed phenone and corresponds to the product of a single gene dose.” My interpretation of this, as given in Cavalli-Sforza and Bodmer [1971, though written in 1969, (11)] was: “**haplotype** (from haploid genotype) for the combination of genetic determinants that leads to a set of antigenic specificities which is controlled by one chromosome and so inherited in coupling.”

The term was originally conceived in the context of a tightly linked cluster of alleles in strong linkage disequilibrium and before the advent of DNA based technology with its almost unlimited number of polymorphisms. However, it can clearly be generalized to refer to the set of variants to be found on any given stretch of DNA on one of the two homologous chromosomes in an individual. That DNA could extend from as little as a single exon, within which there is more than one variable position, to a whole chromosome. The concept therefore becomes vague unless it is related to a defined stretch of DNA.

## Skin Graft Survival and Blood Group Incompatibility

There was an obvious interest in establishing whether the newly identified white blood cell determinants were histocompatibility antigens in the sense of being responsible for graft rejection when not matched. Early data had suggested this was the case. Ceppellini and colleagues exchanged grafts between sibs, parents and offspring, and unrelated individuals in a systematic design. Through a collaboration with van Rood, they showed that skin graft survival times were longer when individuals were matched for the groups defined by van Rood's leukocyte agglutination assay than when they were not (12, 13).

## Mixed Lymphocyte Culture (MLC) Reaction

The Mixed Lymphocyte Culture (MLC) reaction, in which lymphocytes from different individuals when cultured together stimulate a mutual mitotic blast response while lymphocytes from the same individual do not, was discovered by Bach, and independently by Bain, in 1964. This creation of a sort of *in vitro* model for homotransplantation at the cellular level intrigued Ceppellini sufficiently to lead him to invite Fritz Bach to his laboratory to demonstrate his test. Following this, Ceppellini's group developed a “one way” MLC, in which only one of the pair of lymphocytes in a co-culture was able to respond. They then showed that some sera containing anti HLA antibodies were able to block the MLC reaction (14, 15). Ceppellini's group just missed making the important observation made by Bach and Amos (16)

that MLC reactions associated precisely in families with the then serologically defined leukocyte antigens.

This discovery, however, was surely stimulated by Ceppellini's discussion of genetics with Fritz Bach. Van Rood's group was the first to define serological reactions correlating with MLC reactivities, using a rather cumbersome technique involving inhibition of MLC based on Ceppellini's discovery. This laid the foundation for the discovery of the HLA – DR and other Class II determinants using simpler B cell specific serological techniques with the same sources of sera that were used to define the HLA –A, B and C antigens. It is these anti-HLA-DR and related antibodies that explain the MLC inhibition that Ceppellini's group had first observed (see 10 for further details).

## Monoclonal Antibodies, Disease Association, and Nomenclature

Ceppellini became one of the first members of the Basel Institute of Immunology in 1970 and so was amongst the first to realize the importance of monoclonal antibodies, in particular in their application to the HLA system. It was one of his antibodies, produced with Massimo Trucco, that was first used in an International HLA Workshop in 1980. Ceppellini embraced the concept that monoclonal antibodies were the analytic tools of the future and moved from the Basel Institute to Roche as a scientist with the specific objective of producing human monoclonal antibodies. He was, as I recall, quite disappointed in the apparent lack of interest of the Roche pharmaceutical company in the work produced by the outstanding institute that they had created and supported.

In the late 1960s, Ceppellini was, not surprisingly given his background of work on thalassaemia and malaria in Sardinia, one of the first to promote the idea of looking for associations between HLA variants and diseases. It was a follow up of his work in Sardinia that first provided substantial evidence for a role for HLA in malaria resistance (17).

Ceppellini had remarkable insight not only into the genetics and biology of the HLA system but also into the quantitative aspects of its interpretation, as evidenced, for example, by his analysis of the Gm haplotypes. This brought us together in a joint publication from the 1970 International Histocompatibility Testing Workshop on the formal theory of testing the fit of two- and three-locus models of the serological data on HLA at that time and on the analysis of segregation patterns in families (18). My wife, Julia, and I remembered vividly his stay with us in California in 1970 where we finished writing the paper. He had broken his arm skiing, his favorite sport, and he was as lively as ever, but nevertheless, a broken arm did to some extent inhibit his speech! Ceppellini had, much earlier, made a significant contribution to what became a classical method for estimating gene frequencies in a random mating population using iterative gene counting (19).

Ruggero Ceppellini was one of the only professionally trained geneticists among the early workers in the HLA field, apart from myself. Through this we developed a rapport and friendship over a period of more than 20 years.

His clear thinking and forceful contributions to discussions of the WHO International Nomenclature Committee meetings helped enormously in the development of a rational HLA nomenclature based on a proper understanding of the genetics. Ceppellini shares with me and Jon Van Rood the responsibility for the HLA-DR nomenclature and so, eventually, DP and DQ.

## Conclusion

Ruggero Ceppellini unfortunately suffered from periods of depression during the 1970s. His manic periods were easily identified by long and stimulating phone calls in his characteristically deep-throated Italian accent, which would come at any time of the day or night. Indeed, the last communication I had from him, 10 days before he died, was an offer of his lung ascites to culture his tumor cells and a request for references on the genetics of lung cancer.

## REFERENCES

1. Bodmer. In memoriam: Ruggero Ceppellini 1917–1988. *Immunogenetics*. (1989) 29:145–7.
2. Ceppellini R, Dunn LC, Turri M. An interaction between alleles at the Rh locus in man which weakens the reactivity of the Rh(0) factor (D). *Proc Natl Acad Sci USA*. (1955) 41:283–8.
3. Ceppellini R, Siniscalco M. Una nuova ipotesi genetica per il sistema lewis-secretoresi esuoi riflessi nei riguardi di alcune evidenze di linkage con altri loci. *Rivista dell'Istituto Sieroterapico Italiano*. (1955) 30:431–45.
4. Ceppellini R, Dunn LC, Innella F. Immunogenetica II: analisi genetica formale dei caratteri Lewis con particolare riguardo alla natura epistatica della specificità serologica Leb. *Folia Heredit Pathol*. (1959) VIII:261–96.
5. Ceppellini R. Negative correlation between altitude above sea level and incidence of thalassemia in four Rardinian villages. *Cold Spring Harb Symp Quant Biol*. (1955) 2:252.
6. Grubb R. Agglutination of erythrocytes coated with incomplete anti-Rh by certain rheumatoid arthritic sera and some other sera; the existence of human serum groups. *Acta Pathol Microbiol Scand*. (1956) 39:195–7.
7. Ceppellini R. Genetica delle Immunoglobuline. *Atti Associazione Genetica Italiana*. (1967) 12:3–131.
8. Dausset J, Brecy H. Identical nature of the leucocyte antigens detectable in monozygotic twins by means of Iso-Leuco agglutinins. *Nature*. (1957) 180:1430.
9. Dausset J. Iso-leuco-anticorps. *Acta Haematol*. (1958) 20:156–66.
10. Terasaki P. *History of HLA: Ten Recollections*. Los Angeles, CA: UCLA Tissue (1990).
11. Cavalli-Sforza LL, Bodmer WF. (1971). *The Genetics of Human Populations*. San Francisco, CA: W.H. Freeman & Co, Dover Publications.
12. Ceppellini R, Curtoni ES, Mattiuz PL, Leigheb G, Visetti M, Colombi A. Survival of test skin grafts in man: effect of genetic relationship and of blood groups incompatibility. *Ann. N.Y. Acad. Sci.* (1966) 129:421–45.
13. van Rood JJ, van Leeuwen A, Schippers R, Ceppellini PL, Mattiuz S, Curtoni S. Leukocyte groups and their relationship to homo transplantation. *Ann NY Acad Sci*. (1966) 129:467–72.
14. Ceppellini R, Bigliani S, Curtoni ES, Leigheb G. Experimental allotransplantation in man. II. The role of A1, A2, and B antigens. 3. Enhancement by circulating antibody. *Transplant. Proc.* (1969) 1:390.
15. Ceppellini R, Bonnard GD, Coppo F, Miggiano VC, Pospisil M, Curtoni ES, et al. Transplantation antigens: introductory symposium. Mixed leukocyte cultures and HL-A antigens. I. Reactivity of young fetuses, newborns and mothers at delivery. *Transplant.Proc.* (1971) 3:63.
16. Bach FH, Amos DB. Hu-I: Major histocompatibility locus in man. *Science*. (1967) 156:1506.
17. Contu L, Carcassi C, Orrù S, Mulargia M, Arras M, Boero R, et al. HLA-B35 frequency variations correlate with malaria infection in Sardinia. *Tissue Anti*. (1998) 52:452–61.
18. Mattiuz PL, Ihde D, Piazza A, Ceppellini R, Bodmer WF. In: Terasaki PI, editor. *Histocompatibility Testing*. Copenhagen: Munksgaard (1970). p. 193–205.
19. Ceppellini R, Siniscalco M, Smith CA. The estimation of gene frequencies in a random-mating mating population. *Ann Hum Genet*. (1955) 20: 97–115.

**Conflict of Interest Statement:** The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Bodmer. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.





# Bone Marrow Transplantation 1957-2019

Elizabeth Simpson<sup>1\*</sup> and Francesco Dazzi<sup>2</sup>

<sup>1</sup> Division of Immunology & Inflammation, Department of Medicine, Imperial College London, London, United Kingdom,

<sup>2</sup> Division of Cancer Studies, King's College London, London, United Kingdom

Clinical bone marrow transplantation started in 1957 at a time when remarkably little was known about hematopoietic stem cells, immune responses to transplants or the identity of transplant antigens. This review will delineate the substantial increase in knowledge about these three areas gained between then and 1992 when the Ceppellini School course on Bone Marrow Transplantation was held, along with the progress made in clinical application, as well as the stumbling blocks that remained to be overcome by further research to advance knowledge. It will outline the significant progress made between 1992 and the present year, 2019, and the remaining problems.

**Keywords:** transplantation, histocompatibility, graft-vs.-host, graft-vs.-tumour, immunosuppression

## INTRODUCTION

The Scuola Superiore d'Immunologia Ruggero Ceppellini (Ceppellini School) was founded in 1991 in Naples by Professor Serafino Zappacosta, to honor the memory and achievements of Professor Ruggero Ceppellini, a giant in the field of HLA genetics, who led an approach to addressing complex scientific questions through national and international collaboration (**Figure 1**). The Ceppellini school takes this forward by promoting contact and collaboration through residential post-graduate level courses led by international faculty for early career basic scientists and clinicians from advanced and developing regions and countries. It has accelerated Immunology education and influenced the evolution of other international schools of immunology. This issue of *Frontiers in Immunology* is a celebration of the achievements of the school and a tribute to Professor Serafino Zappacosta, Professor of Immunology at the "Federico II" University of Naples, who aimed to create in Southern Italy a pole of attraction for those pursuing immunological studies, and to promote interaction among the scientific and medical communities at the national and international level.

## REVIEW OF THE BONE MARROW TRANSPLANTATION COURSE

In 1992 Bone Marrow Transplantation was the subject of the inaugural course of the Ceppellini School. This topic brought into focus both genetics and immunology, the areas to which Ceppellini's research on hematological disorders and the human major histocompatibility complex, HLA, was pivotal.

This review of bone marrow/hematological stem cell transplantation will focus on how contributions to the 1992 Ceppellini School course on Bone Marrow Transplantation provide a mid-way marker point in the six decades following 1957 when Donnall Thomas first reported on six patients given bone marrow transplants to restore hemopoiesis following ablation by radiation or drug toxicity (1). He was encouraged by Peter Medawar's 1953 report (2) that immunological rejection of skin grafts exchanged between non-genetically identical mice could be abrogated by induction of transplantation tolerance and by Loutit's work showing restoration of hemopoiesis in irradiated mice given spleen cells from the same inbred strain, but not from a different strain (3).

## OPEN ACCESS

### Edited by:

Francesca Di Rosa,  
Istituto di Biologia e Patologia  
Molecolari (IBPM), Consiglio Nazionale  
Delle Ricerche (CNR), Italy

### Reviewed by:

Michaela Semeraro,  
Necker-Enfants Malades  
Hospital, France  
Alois Anton Gratwohl,  
University of Basel, Switzerland

### \*Correspondence:

Elizabeth Simpson  
esimpson@imperial.ac.uk

### Specialty section:

This article was submitted to  
Molecular Innate Immunity,  
a section of the journal  
*Frontiers in Immunology*

**Received:** 12 February 2019

**Accepted:** 16 May 2019

**Published:** 05 June 2019

### Citation:

Simpson E and Dazzi F (2019) Bone  
Marrow Transplantation 1957-2019.  
*Front. Immunol.* 10:1246.  
doi: 10.3389/fimmu.2019.01246



**FIGURE 1** | Naples, where Professor Serafino Zappacosta founded the Scuola Superiore d'Immunologia Ruggero Ceppellini in 1991. Image: Vera Maone, used with permission.

At that time, in the 1950s, there was limited knowledge of the genetics of transplant antigens and the immune responses to them, and all of those first patients died, although transient chimerism was recorded. In 2018, 60 years later, hundreds of thousands of hemopoietic transplants have been carried out, using a variety of sources for stem and precursor cells and an array of pre-conditioning treatments to facilitate graft acceptance in patients. While many recipients survived, cured of hematological malignancies or hematological diseases that would otherwise have killed them, others suffered serious side effects of which graft-vs.-host disease (GVHD) has been the most challenging. This uncomfortable “fact of life” has limited the more widespread use of hemopoietic transplants to treat other conditions that might benefit from “resetting the immune system,” such as autoimmunity and rejection of therapeutic organ transplants.

Advances in pharmacology and the development of less toxic preconditioning regimes have made a series of stepwise improvements, both in graft acceptance and reducing GVHD incidence and severity. These have been built on advances in genetics, particularly with respect to delineation of the major histocompatibility complex (MHC), HLA in humans, along with homologs in species used in preclinical research, mice (H2), dogs (DLA), and non-human primates (SLA). In parallel, increasing knowledge of the immune system has provided insight into factors regulating the quality and quantity of immune responses, and has triggered the development of a range of biologically active pharmaceuticals aimed at controlling over- or under-effective responses in patients. The enrolment of patients into controlled clinical trials is the ultimate way to test safety and efficacy of new treatments: this is now widely embraced.

Our speakers in 1992 included those working on hematopoietic stem cells (Nydia Testa, Maria Grazia Roncarolo, Peter Hoogerbrugge) on identification of major and minor histocompatibility antigens (Robert Lechler, Elizabeth Simpson,

Giovanni Ferrara), on immune responses to transplants (Herman Waldmann, Manlio Ferrarini, Yair Reisner) and on treating patients with hematopoietic disorders with HSC transplants (Jill Hows, Andrea Bacigalupo, Bruno Rotoli, Andrea Velardi, Guido Lucarelli).

## IMMUNE RESPONSES TO TRANSPLANTS

Hemopoietic stem cell transplantation (HSCT) is the forerunner of both cell and gene therapies, which depend on slipping potentially foreign components past homeostatic controls limiting cell numbers and immune responses fine-tuned by evolutionary selection for protection against pathogens. The numbers of cells comprising some tissues can be reduced by irradiation and/or cytotoxic drugs to provide “space” for the introduced cell population to settle and proliferate. The appropriate dose of space-inducing treatment depends on the tissue and on whether total replacement or chimerism is required for therapeutic effect.

However, the immune response remains a formidable barrier, comprised of a moving army of variously armed host cells along with cell-bound and shed molecules, such as antibodies, receptors and cytokines, orchestrated by a complex activatory and inhibitory pathways. For hemopoietic transplants the situation is further complicated by potential two way reactions between recipient and donor: rejection of donor cells is the host-vs.-graft (HVG) response, whereas attack of the host by cells in the donor inoculum is the graft-vs.-host (GVH) response. Graft-vs.-host disease (GVHD) occurs when normal host tissues are attacked, but when this is focused on host tumor cells, the terms graft-vs.-leukemia (GVL) or graft-vs.-tumor (GVT) are used. Separating GVHD from GVL/T has proved difficult. Though most of the target antigens are shared, in principle there could be a set of non-shared tumor antigens. Unfortunately, which patients will develop GvHD and/or GVL cannot yet be accurately predicted because the molecular targets have not been sufficiently identified. The extraordinary diversity of target antigens is amplified by HLA polymorphism as well as that of minor histocompatibility antigens and tumor antigens arising from serial mutation.

Peter Medawar demonstrated that rejection of skin grafts exchanged between genetically dissimilar rabbits or mice showed specificity and memory (4)—hallmarks previously ascribed to antibody responses against pathogens. Mitchison subsequently transferred skin graft rejection with lymphocytes, but not serum, i.e., it was cell and not antibody mediated (5). On the basis that immune responses evolved to discriminate between self and non-self, Medawar designed experiments in which cells from one inbred mouse strain were introduced to immune-incompetent pre-natal or neonatal mice of another strain to induce recognition of them as “self” during development. When skin grafted as adults, most of the injected mice showed prolonged acceptance of test grafts. These experiments were replicated in other mammalian species and in birds (2, 6). Thus, the possibility of inducing transplantation tolerance existed, giving encouragement to both hematologists like Donnell

Thomas and surgeons like Joseph Murray who performed the first kidney transplants in humans.

Inducing tolerance in adult animals, either humans or experimental species, proved more difficult. Making recipients immunoincompetent using irradiation and/or cytotoxic drugs abrogates HVG but can lead to collateral damage to host tissues, and if immunocompetent lymphocytes of donor origin are included in the donor graft, they can cause GVHD. The morbidity and mortality figures during the early years of clinical BMT were daunting but they sparked extensive and focused preclinical experiments in outbred dogs (by Thomas' group) leading to step-wise improvements in the clinical protocols used, including reduced levels of irradiation and the development of less toxic drugs for pre-treatment of recipients. Pretreatment of donor cells was also trialed including removal of contaminating lymphocytes from bone marrow and use of alternative sources, such as mobilized stem cells isolated from peripheral blood or cord blood as a source.

In the 1960s and 1970s basic research studies had probed the composition of lymphocyte populations, leading to the understanding of the different functions of thymus derived (T cells) and bone marrow (B cells) lymphocytes and of T cell subsets, Th helper, and Tc cytotoxic cells. The development of monoclonal antibodies by Milstein and Kohler in 1975 (7) led to the isolation of highly specific reagents for identifying and separating cell types on the basis of cell surface molecules. Separation of cells with characteristic markers using the fluorescence activated cell sorter developed by Len Herzenberg in the 1970s and 1980s (8) was crucial to defining the phenotype and function of hemopoietic and lymphopoietic cell subsets.

Isolation from mouse bone marrow of selected populations containing a high proportion of haemopoietic stem cells (HSC) that could repopulate all lineages was shown by Weissman in the late 1980s and early 1990s (9). However, monoclonal antibodies defining the homologous population in humans have been more difficult to develop for approved clinical use. Even low levels of contaminating T lymphocytes in partially purified sources of hematopoietic stem cells can cause GVHD. If T cells are completely removed leukemic relapse is more likely, although that risk can be significantly reduced by donor lymphocyte infusion (DLI) following T cell depleted hematopoietic stem cell transplantation (10).

The molecular identification of the targets of transplant rejection (histocompatibility antigens) has played a central role in all clinical transplantation, but hematopoietic cell reconstitution presents the greatest challenges because of GVH, but also the greatest rewards, with the development of GVL/GVT effectors.

At the 1992 Ceppellini School BMT course the speakers outlined the key findings underlying the development of the field and presented them for discussion along with recent advances. It was clear that the problem of GVHD remained serious, with questions about identifying the best tolerable genetic match between donor and recipient, and how to minimize and treat this complication. Since 1992 then there have been substantial advances in knowledge of stem cell biology, the genetics of histocompatibility antigens and of the cells, molecules and regulatory circuits of the immune system, permitting further

stepwise improvements, but now, having looked at the immune response, let us consider how transplantation antigen genes and their products were originally identified, especially as key research, including links with a range of human diseases, was carried out in Italy.

## GENETICS, MOLECULAR IDENTITY AND FUNCTION OF TRANSPLANTATION ANTIGENS, MAJOR AND MINOR

George Snell took a systematic genetic approach to enumerating and mapping loci responsible for graft rejection with experiments transplanting skin and tumor grafts between inbred mouse strains, their F1 hybrids and backcross progeny. These were carried out at the Jackson Laboratory in the late 1930s before DNA, genes or chromosomes had been identified as units of inheritance. Instead, co-inheritance of traits mapped them into so-called "linkage groups." Snell numbered his histocompatibility loci, H1, H2, H3, etc., according to their apparent strength, with H2 the strongest, eliciting the fastest graft rejection. It was named the major histocompatibility (H) locus, with the others designated minor H loci. The agglutinating antibodies developed by Peter Gorer following immunization of different mouse strains were found to detect alleles of H2. Snell's 1948 paper (11) summarizes findings defining not only the major but also a number of minor H loci, of which more were found by Snell and his collaborators Bailey and Taylor who isolated and mapped each H locus in congenic and recombinant inbred mouse strains. However, while MHC antigens are highly polymorphic, their minor H counterparts are not. They are either di-allelic or characterized by one expressed and one non-expressed allele.

Ceppellini played a key role within the international consortium (12) in discovering the human homologs of H2 antigens, the human leukocyte antigens (HLA) named from their expression on human peripheral blood lymphocytes, to which agglutinating antibodies from multiparous women were found, directed against paternal alloantigens. Ceppellini's genetic studies in the 1950s on hemoglobinopathies, linking resistance to malaria and thalassemia, was followed by his immunogenetic research on red blood cell antigens, leading him to recommend to Dausset, then working on the elusive human leukocyte antigens using poorly reproducible agglutinating assays, the study of identical twins, whose reactions should be the same if the antigens were genetically controlled. The power of a genetic approach was used by Ceppellini when he HLA typed large families, identifying siblings inheriting the same parental HLA alleles, those with different alleles, and those with one shared allele. He then exchanged skin grafts between them and discovered while the most rapid rejection took place when no HLA alleles were shared, that for even completely HLA matched pairs graft rejection was only delayed by a week or 2 (13). These findings, in parallel with Snell's on mice, were evidence that minor H antigens also existed in humans. Confirmation of this comes from clinical bone marrow transplantation: HLA matched sibling recipients can still suffer life-threatening GVHD directed against minor H allo antigens.

A breakthrough in interpreting the human HLA antibody data accumulated during the international histocompatibility workshops came when viewed as a complex of linked loci, each with a number of alleles, rather than as a single locus. This arrangement had already been observed for the mouse MHC, H2, to consist of two linked polymorphic loci (named H2K and H2D), encoding cell surface molecules expressed on all lymphocytes in peripheral blood (PBL). Other mouse anti-H2 antibodies were found that reacted with B but not T lymphocytes. These were directed against the products of loci within H2 that distinguished alleles associated with the ability to respond to certain haptenated proteins, i.e., those were “Immune Response” (Ir) gene controlled. The Ir genes mapped between H2K and H2D of the mouse MHC complex on chromosome 17 and were named H2A and H2E. H2 studies were easier than those on HLA because of inbred mice, including congenic strains with selected alleles of H2 backcrossed onto a standard strain. Intercrossing could then be carried out to create H2 recombinants, allowing the study of individual loci. In outbred populations, such as humans HLA recombination occurs at a low frequency depending on the chromosomal distance between loci—HLA genes are closely clustered on human chromosome 6.

To resolve a nomenclature clash that had occurred between different laboratories working on human MHC, the first human HLA loci were named HLA-A and -B, and between them a third locus, HLA-C was mapped. The human homologs of mouse Ir genes mapped them just outside (centromeric) the A/B/C region and were named DR, DQ, and DP. Their cell surface molecules, like Ir genes of mice, are not expressed on all somatic cells but on human PBL they were detected on both B cells and activated T cells. Mouse Ir genes and their human homologs were designated MHC class II, to distinguish them from H2K and D, and HLA A, B, and C, that were known as MHC class I.

Understanding the functional homologies of the mouse and human MHC relied on the development of *in vitro* assays of T cell function. Proliferation assays developed by Fritz Bach identified MHC class II alleles as stimulatory in one-way mixed lymphocyte cultures (MLC) between two mis-matched individuals where cells from one were irradiated to prevent them proliferating (14). In parallel studies in other laboratories cytotoxic T cells (Tc) were developed in MLC, initially in mice (15). Tc effector cells in these cultures were directed against MHC class I antigens, while helper T cells, Th, specific for MHC class II were required for optimal responses (16). The same is true for both species.

*In vitro* cultures of lymphocytes were also used to examine the fine specificity of cytotoxic T cells against viral epitopes and minor (H) histocompatibility antigens. The paradigm changing paper on this was published in 1974 by Zinkernagel and Doherty, who showed that *in vitro* re-stimulated spleen cells from mice immune to lymphocytic choriomeningitis virus (LCMV) generated cytotoxic T cells that recognized virus only in association with self MHC, i.e., they were “MHC restricted.” The same was found to be true for mouse and human cytotoxic T cells specific for the male specific minor H antigen, HY (17, 18) and all other minor H antigens. Although the manner in which T cells recognized both self MHC and a foreign antigen was not

understood until the crystal structure of HLA-A2 was solved in 1987, showing a peptide fragment of viral or other origin held in the peptide binding groove of the MHC molecule (19). The generation of minor H specific T cells and clones provided key reagents for mapping and cloning the corresponding genes and identifying their MHC binding peptides. This is also true for a tumor specific antigens (TSA): Thierry Boon led investigation of these by generating TSA specific T cell clones (20), and used these to expression clone a range of melanoma and other tumor antigen genes and explored use of them to immunize patients. This approach also informs treatment of leukemia patients, given HSC.

The *in vitro* T cell correlates of HVG and GVH immune responses against minor H antigens are MHC class II restricted Th helper cells and class I restricted Tc cytotoxic effectors. Unlike *in vitro* responses to mismatched polymorphic MHC antigens that develop in primary cultures, those to minor H antigens require previous *in vivo* exposure to antigen to increase the precursor frequency of T cells specific for the minor H antigen(s). Two MHC matched individuals in an outbred population will differ at many minor H loci, including HY in the case of brother/sister pairs. Immunodominance of the response against one or more minor H antigen occurs. An *in vivo* manifestation of this is stronger GVHD in male recipients of HLA matched female hematopoietic transplants than in female recipients, since HY is immunodominant to some other minor H antigens. However, the strength of each is also a function of the MHC restricting molecule, since it is the combination that determines immunogenicity.

The 1992 Ceppellini course on BMT included presentation and discussion of the research leading up to the discovery of both MHC and minor H antigens in humans and mice. This included methods then currently in use for HLA typing, and how MHC restriction prevailed for the recognition by T cells of all non-MHC antigens, whether minor H, viral and other.

Responses against multiple minor H antigens are strong and can be life threatening in GVHD. In contrast, if they could be separated into their components specificities they could be therapeutic, particularly for selecting those directed against minor H antigens preferentially expressed on tumor cells. An opportunity to create this situation occurs in leukemia patients given HSC transplants. Providing that expression of a minor H antigen is hematopoietic cell specific, cytotoxic T cells directed against the recipient allele of that antigen will target both leukemic cells and residual recipient hematopoietic cells but not those of repopulating donor origin. Such curative T cells would remove leukemic cells without the side effect of damaging other recipient tissues.

The treatment outlined above would require identification of minor H antigens expressed only on hematopoietic cells, for which there are some candidates, together with the isolation and expansion of effector T clones that can be approved for clinical use. Those of donor stem cell origin would be ideal, as they should persist long term in the recipient and mitigate against leukemic relapse. Current research on effector cells transduced with CAR-based or T cell receptor (TCR)-based constructs is relevant to this.



## SOURCES OF HEMATOLOGICAL STEM CELLS (HSC)

HLA typing was recognized early as crucial for allogeneic HSC transplantation, as HLA mismatched grafts were likely to be rejected and/or cause severe GVHD. Use of HLA matched sibling donors reduced but did not remove this risk which was higher when non-sibling HLA matched family donors were used. Use of unrelated HLA matched donors (MUD) increased GVHD risk above that. Haploidentical donors, i.e., those with one of their two HLA haplotypes inherited from a parent, especially the mother, was started in the 1990s and has been substantially increased as methods for abrogating or treating GVHD have improved. Such a procedure has become more common, especially after the development of conditioning regimens involving cyclophosphamide (21) that appear effective at generating the early expansion of regulatory T cells.

Haploidentical transplantation has provided a unique platform for experimental tolerogenic strategies, with several studies providing convincing evidence that, at least when using the most appropriate donor, the outcome can be very good (22). A recent retrospective study (23) has convincingly documented that it is the patient and disease rather than donor features that affect survival of these patients. However, it is important to acknowledge the fact that the technique of haploidentical transplantation exposes patients to delayed immune reconstitution thus potentially limiting some of the benefits.

The use of bone marrow cells and G-CSF mobilized bone marrow cells as sources of HSC have been modified in a number of ways in attempts to reduce GVHD. Complete removal of T cells can be effective, but leukemic patients then have higher relapse rates, due to the removal of GVL effectors. Reduction in the number of contaminating T cells can help, but is difficult to titrate. Mitigating the risk of relapse, donor lymphocyte infusions (10) have been used and these, following HSC transplantation, have provided long-term curative treatment, particularly for chronic myeloid leukemia. Since T cells in these donor inocula are long lived and likely to contain a number of different clones with specificity for several transplantation antigens, mutant leukemic cells are likely to be targeted as they arise, a situation not replicated when targeted molecular therapy is given, directed against a determinant whose expression can be downregulated by mutation.

The use of cord blood as a source of HSC uncontaminated by primed T cells is practical only for child recipients, as single donations rarely contain sufficient stem cells to achieve engraftment. However, recent data suggests the opportunity to use aryl hydrocarbon antagonists to produce a robust expansion of hematopoietic stem and progenitor cells (24).

Autologous HSC avoided the risk of GVH and HVG reactions but its use in treatment of leukemias and other cancers was bedeviled by high relapse rates, as there can be no GVT effect. However, since the early 2000s the introduction of somatic gene therapy for inherited immunodeficiencies has been made possible by the identification of some of the relevant mutant genes, and methods for transducing corrected copies of them into autologous HSC *ex vivo* before transplanting them into the patient pre-treated to provide “space” for the newcomers. There are still issues that can limit the applicability of the gene therapy approach. On one side the modification of HSCs may reduce their capacity to engraft, whilst on the other the modification strategy may require the selection of the gene-corrected cells, thus impacting on the cell yield required to be efficiently transplanted (25).

Based on the notion that it is the graft-vs.-tumor effect that secures long-term eradication of the underlying malignancy, the conditioning regimens used to prepare patients for transplantation have been radically revisited. Whilst radiation was the main component of the pre-transplant conditioning because of its efficacy in eliminating replicating cells, other milder approaches have been used since the end of the 90s. Chemotherapeutic agents are now being used at doses by far lower than before, thus reducing toxicity and eventually reducing the frequency of GvHD that is largely affected, not only by the transplantation antigens but also and perhaps more importantly, by the cytokine storm induced by the tissue damage consequential to chemo/radiotherapy (26). Furthermore, it was shown that the use of cyclophosphamide soon after HSC infusion could mitigate the incidence of GvHD by increasing the number of regulatory T cells (27).

Unfortunately, GvHD remains the most dreadful complication of allografting and when refractory to steroid treatment the associated mortality is dismal. Cellular therapies may provide an alternative to traditional immunosuppressive approaches because they may provide an immunological reprogramming of the patient's inflammatory environment. Important milestones in this direction have been provided by the use of regulatory T cells (28) and mesenchymal stromal cells (MSC). Initially identified as tout-court immunosuppressants (29), MSC have been recently shown as effective at reprogramming the recipient phagocytic system to control unwanted inflammation (30). This has transplanted into very encouraging clinical experience (31).

## HEMATOLOGICAL AND OTHER DISEASE CANDIDATES FOR HSC TRANSPLANTS

Leukemias, bone marrow failures, hemoglobinopathies (e.g., thalassemia, sickle cell disease) and immunodeficiencies have been mentioned, and advances with these could lead to HSC transplant-based treatments for solid tumors and other genetic diseases (e.g., lysosomal storage disease) as well as autoimmunity.



Reducing the incidence and severity of GVHD following HSC transplant remains the biggest challenge for both existing patients and the possibility of extending this treatment to additional diseases.

## REFERENCES

- Thomas ED, Lochte HL, Lu WC, Ferrebee JW. Intravenous infusion of bone marrow in patients receiving radiation and chemotherapy. *N Engl J Med.* (1957) 257:491–6. doi: 10.1056/NEJM195709122571102
- Billingham R, Brent L, Medawar PB. Actively acquired tolerance of foreign cells. *Nature.* (1953) 172:603–6. doi: 10.1038/172603a0
- Barnes DWH, Loutit JF. Protective effects of implantation of spleen tissue. *Proc Roy Soc Med.* (1953) 46:251.
- Medawar PB. The behaviour and fate of skin autografts and homografts in rabbits: a report to the War Wounds Committee of the Medical Research Council. *J Anat.* (1944) 78:176–99.
- Mitchison NA. Passive transfer of transplantation immunity. *Nature.* (1953) 171:267–8. doi: 10.1038/171267b0
- Medawar PB, Brent L, Medawar PB. Quantitative studies on tissue transplantation immunity. III. Actively acquired tolerance. *Phil Trans Roy Soc B.* (1956) 239:357–414. doi: 10.1098/rstb.1956.0006
- Kohler G, Milstein C. Continuous cultures of fused cells secreting antibody of predefined specificity. *Nature.* (1975) 256:495–7. doi: 10.1038/256495a0
- Hardy RR, Roederer M, Leonard Herzenberg (1931–2013): the life of FACS. *Immunity.* (2013) 39:989–91. doi: 10.1016/j.immuni.2013.11.008
- Uchida N, Weissman IL. Searching for haematopoietic stem cells: evidence that Th1/1lo Lin-Sca-1<sup>+</sup> cells are the only stem cells in C57BL/Ka-Thy-1.1 bone marrow. *J Exp Med.* (1992) 175:175–84. doi: 10.1084/jem.175.1.175
- Dazzi F, Szydlo RM, Craddock C, Cross NCP, Kaeda J, Chase A, et al. Comparison of single-dose and escalating-dose regimens of donor lymphocyte infusion for relapse after allografting for chronic myeloid leukemia. *Blood.* (2000) 95:67–71.
- Snell GD. Methods for the study of histocompatibility genes. *J Genet.* (1948) 49:87–108. doi: 10.1007/BF02986826
- Bodmer WF. In memoriam Ruggero Ceppellini 1917–1988. *Immunogenetics.* (1989) 29:145–7. doi: 10.1007/BF00373638
- Ceppellini R, Mattiuz PL, Scudeller G, Visetti M. Experimental allotransplantation in man I. The role of the HLA system in different genetic combinations. *Transplant Proc.* (1969) 1:385–9.
- Amos DB, Bach FH. Phenotypic expressions of the major histocompatibility in man (HL-A), leucocyte antigens and mixed lymphocyte culture reactivity. *J Exp Med.* (1968) 128:623–39. doi: 10.1084/jem.128.4.623
- Simpson E, O'Hopp S, Wunderlich J. Life span of cytotoxic activity & memory activity following allogeneic skin grafting in the mouse. *Transplantation.* (1974) 18:374–7. doi: 10.1097/00007890-197410000-00014
- Janeway CA, Jr., Sharrow SO, Simpson E. T cell populations with different functions. *Nature.* (1975) 253:544–6. doi: 10.1038/253544a0
- Gordon RD, Simpson E, Samelson LE. *In vitro* cell-mediated immune responses to the male specific (H-Y) antigen in mice. *J Exp Med.* (1975) 142:1108–20. doi: 10.1084/jem.142.5.1108
- Goulmy E, Termijtelen A, Bradley BA, van Rood JJ. Y-antigen killing by T cells of women restricted by HLA. *Nature.* (1977) 266:544–5. doi: 10.1038/266544a0
- Borkmann PJ, Saper MA, Samraoui B, Bennett WS, Strominger JL, Wiley DC. The foreign antigen binding site and T cell recognition regions of class I histocompatibility antigens. *Nature.* (1987) 329:512–8. doi: 10.1038/329512a0
- Traversari C, van der Bruggen P, Luescher IF, Lurquin C, Chomez P, Van Pel A, et al. A nonapeptide encoded by human gene MAGE-1 is recognized on HLA-A1 by cytolytic T lymphocytes directed against tumor antigen MZ2-E. *J Exp Med.* (1992) 176:1453–7. doi: 10.1084/jem.176.5.1453
- Chiusolo P, Bug G, Olivieri A, Mats B, Mordini N, Alessandrino PE, et al. A modified post-transplant cyclophosphamide (PT-CY) regimen, following unmanipulated haploidentical bone marrow transplantation, for acute myeloid leukemia: a multicenter study. *Blood.* (2016) 128:1234–49. doi: 10.1016/j.bbmt.2018.01.031
- McCurdy SR, Fuchs EJ. Selecting the best haploidentical donor. *Semin Hematol.* (2016) 53:246. doi: 10.1053/j.seminhematol.2016.08.001
- McCurdy SR, Zhang M-J, St. Martin A, Al Malki MM, Bashey A, Gaballa S, et al. Effect of donor characteristics on haploidentical transplantation with posttransplantation cyclophosphamide. *Blood Adv.* (2018) 2:299. doi: 10.1182/bloodadvances.2017014829
- Wagner JE, Brunstein CG, Boitano AE, DeFor DE, McKenna D, Sumstad D, et al. Phase I/II trial of stem regenin-1 expanded umbilical cord blood hematopoietic stem cells supports testing as a stand-alone graft. *Cell Stem Cell.* (2016) 18:144–55. doi: 10.1016/j.stem.2015.10.004
- Morgan RA, Gray D, Lomova A, Kohn DB. Haematopoietic stem cell therapy: progress and lessons learned. *Cell Stem Cell.* (2017) 21:574–90. doi: 10.1016/j.stem.2017.10.010
- Fozza C, Szydlo RM, Abdel-Rehim MM, Nadal E, Goldman JM, Apperley JF, et al. Factors for graft-versus-host disease after donor lymphocyte infusions with an escalating dose regimen: lack of association with cell dose. *Br J Haematol.* (2007) 136:833–6. doi: 10.1111/j.1365-2141.2007.06501.x
- Luznik L, O'Donnell PV, Ephraim JF. Post-transplantation cyclophosphamide for tolerance induction in HLA-haploidentical BMT. *Semin Oncol.* (2012) 39:683–93. doi: 10.1053/j.seminoncol.2012.09.005
- Brunstein CG, Miller JS, McKenna DH, Hippen KL, DeFor TE, Sumstad D, et al. Umbilical cord blood-derived T regulatory cells to prevent GVHD: kinetics, toxicity profile, and clinical effect. *Blood.* (2016) 127:1044–51. doi: 10.1182/blood-2015-06-653667
- Krampera M, Glennie S, Dyson J, Scott D, Laylor R, Simpson E, et al. Bone marrow mesenchymal stem cells inhibit the response of naive and memory antigen-specific T cells to their cognate peptide. *Blood.* (2003) 101:3722–9. doi: 10.1182/blood-2002-07-2104
- Galleu A, Riffo-Vasquez Y, Trento C, Lomas C, Dolcetti L, Cheung TS, et al. Apoptosis in mesenchymal stromal cells induces *in vivo* recipient-mediated immunomodulation. *Sci Transl Med.* (2017) 9:416. doi: 10.1126/scitranslmed.aam7828
- Galleu A, Milojkovic D, Deplano S, Szydlo R, Loaiza S, Wynn R, et al. Mesenchymal stromal cells for acute graft-versus-host disease: response at 1 week predicts probability of survival. *Br J Haematol.* (2019) 185:89–92. doi: 10.1111/bjh.15749

## AUTHOR CONTRIBUTIONS

ES and FD drafted the review together. ES concentrating on the historical background and FD on the clinical aspects.

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Simpson and Dazzi. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.



# Immunology's Coming of Age

**Stefan H. E. Kaufmann**<sup>1,2\*</sup>

<sup>1</sup> Department of Immunology, Max Planck Institute for Infection Biology, Berlin, Germany, <sup>2</sup> Hagler Institute for Advanced Study, Texas A&M University, College Station, TX, United States

## OPEN ACCESS

### Edited by:

Ennio Carbone,  
Università degli Studi Magna Graecia  
di Catanzaro, Italy

### Reviewed by:

Angelo A. Manfredi,  
Vita-Salute San Raffaele University,  
Italy  
Maria Regina D'Império Lima,  
University of São Paulo, Brazil

### \*Correspondence:

Stefan H. E. Kaufmann  
Kaufmann@mpiib-berlin.mpg.de

### Specialty section:

This article was submitted to  
Molecular Innate Immunity,  
a section of the journal  
Frontiers in Immunology

**Received:** 07 February 2019

**Accepted:** 13 March 2019

**Published:** 03 April 2019

### Citation:

Kaufmann SHE (2019) Immunology's  
Coming of Age.  
Front. Immunol. 10:684.  
doi: 10.3389/fimmu.2019.00684

This treatise describes the development of immunology as a scientific discipline with a focus on its foundation. Toward the end of the nineteenth century, the study of immunology was founded with the discoveries of phagocytosis by Elias Metchnikoff, as well as by Emil Behring's and Paul Ehrlich's discovery of neutralizing antibodies. These seminal studies were followed by the discoveries of bacteriolysis by complement and of opsonization by antibodies, which provided first evidence for cooperation between acquired and innate immunity. In the years that followed, light was shed on the pathogenic corollary of the immune response, describing different types of hypersensitivity. Subsequently, immunochemistry dominated the field, leading to the revelation of the chemical structure of antibodies in the 1960s. Immunobiology was preceded by transplantation biology, which laid the ground for the genetic basis of acquired immunity. With the identification of antibody producers as B lymphocytes and the discovery of T lymphocytes as regulators of acquired immunity, lymphocytes moved into the center of immunologic research. T cells were shown to be genetically restricted and to regulate different leukocyte populations, including B cells and professional phagocytes. The discovery of dendritic cells as major antigen-presenting cells and their surface expression of pattern recognition receptors revealed the mechanisms by which innate immunity instructs acquired immunity. Genetic analysis provided in-depth insights into the generation of antibody diversity by recombination, which in principle was shown to underlie diversity of the T cell receptor, as well. The invention of monoclonal antibodies not only provided ultimate proof for the unique antigen specificity of the antibody-producing plasma cell, it also paved the way for a new era of immunotherapy. Emil Behring demonstrated cure of infectious disease by serum therapy, illustrating how clinical studies can stimulate basic research. The recent discovery of checkpoint control for cancer therapy illustrates how clinical application benefits from insights into basic mechanisms. Last not least, perspectives on immunology progressed from a dichotomy between cellular-unspecific innate immunity and humoral-specific acquired immunity, toward the concept of complementary binarity.

**Keywords:** antibody, cytokine, dendritic cell, immunology, lymphocyte, macrophage, phagocytosis, recombination

## INTRODUCTION

In this treatise, I describe growth and maturation of immunology as a scientific discipline built on both basic research and medical application. Although I emphasize the birth of immunology and early decades of its evolution, I stress that immunology in its full maturity remains equally integrated in both basic and clinical research.

Immunology started in the last quarter of the nineteenth century with two major discoveries. The first of these was Elias Metchnikoff's (1845–1916) identification of phagocytic cells, which engulf and destroy invading pathogens (1). This laid the basis for innate immunity. The second discovery was Emil Behring's (1854–1917) and Paul Ehrlich's (1854–1915) identification of antibodies, which neutralize microbial toxins (1, 2). This became the basis for acquired immunity. These findings also led to the distinction between cellular and humoral immunity. For obvious reasons, humoral immunity was often considered synonymous with acquired immunity, whereas cells were considered tightly linked to innate immunity. This was overlaid by a further segregation between the unique antigen specificity of the acquired arm vs. the non-specific innate arm of the immune response (**Figure 1**). This dichotomous view led to some confusion and controversy and it took some time until it transformed into a perspective of complementary binarity considering innate and acquired immunity as interactive partners. Today the two arms of antigen-specific acquired and antigen-nonspecific innate immunity are best viewed as a ying-yang concept, with highly intertwined, partly overlapping, and mutually beneficial activities. Further highly valuable information on the highlights of immunology in its nascence can be found in the many publications of A. Silverstein of which I only cite his major treatise (3).

From its birth, immunology was at the heart of biomedical research providing both crucial information on basic biological processes and on clinical application. This was recognized by the first ever Nobel Prize in Medicine awarded in 1901 to Emil Behring "for serum therapy in therapeutic medical science," (4) and also by the most recent Nobel Prize 2018 to honor the "discovery of cancer therapy by inhibition of negative immune regulation" by Jim Allison (1948–) and Tasuku Honjo (1942–) (5). Whilst Behring's discovery illustrates how medical application can stimulate basic research, the discoveries of Allison and Honjo epitomize clinical application as the result of in-depth understanding of basic biological mechanisms.

## ACT I: THE FOUNDATION OF IMMUNOLOGY

Immunology emerged as an academic discipline in its own right out of the fertile soil of medical microbiology (6). The discoveries of Louis Pasteur (1822–1895), which confirmed and completed the germ theory of infectious diseases as well as Robert Koch's (1843–1910) meticulous studies on the etiology of infectious diseases, notably tuberculosis, raised a question of fundamental importance: Is the host a helpless prey of pathogenic microbes or is it equipped with an efficient defense mechanism to combat its invaders? Both Pasteur and Koch favored the notion that the host was defenseless. However it was Metchnikoff, at the Pasteur Institute in Paris since 1888, who earlier discovered the critical role of phagocytosis and intracellular killing in host defense (1), and it was Behring and Ehrlich, young independent researchers at Koch's institute for Infectious Diseases in Berlin, who identified antibodies as crucial counterparts to the toxic activities of

bacteria (1, 2). We now know that the outcome of infection depends on close interactions between pathogen and host factors, probably best described by the term infection biology.

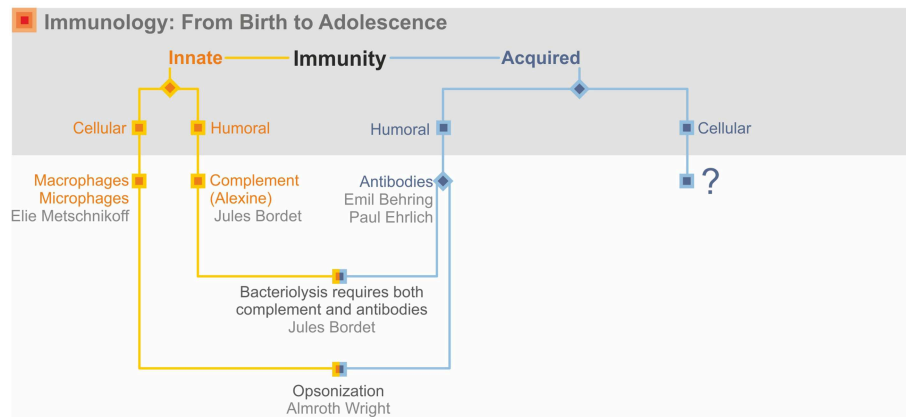
When Koch embarked on the next step in his career in Berlin in 1878, the pathologist Rudolf Virchow (1821–1902) was the most eminent professor at the Charité clinics (6). Virchow is the founder of cellular pathology, which assumes that all diseases are the result of malfunctioning of our body's cells (7). Hence, Koch's ideas on the etiology of infectious diseases seconded by the germ theory of Pasteur were highly criticized by Virchow. Ultimately, Koch's observations, well-supported by experimental evidence, became the accepted paradigm. According to the American physicist and philosopher, Thomas Kuhn (1922–1996), normal science progresses as long as available evidence can be accommodated in the existing paradigm (8). Once anomalies accumulate from scientific research that can no longer be integrated in an existing paradigm, the time is ripe for a paradigm shift (8). Koch and Pasteur introduced a paradigm shift by demonstrating that exogenous invaders can cause certain diseases, beyond those diseases caused by dysfunctional cells. Yet, they both largely overlooked the role of host immunity as important defense mechanism. This paradigm shift was initiated by Metchnikoff, Behring, and Ehrlich. Today we understand infectious diseases as the outcome of a crosstalk between host and pathogen. We also now know that immunology has more roles to play than only pathogen defense, such as surveillance of malignant cells. Moreover, a dysfunctional immune system results in allergy, autoimmunity or chronic inflammation thereby illustrating it as a double-edged sword.

## Phagocytosis

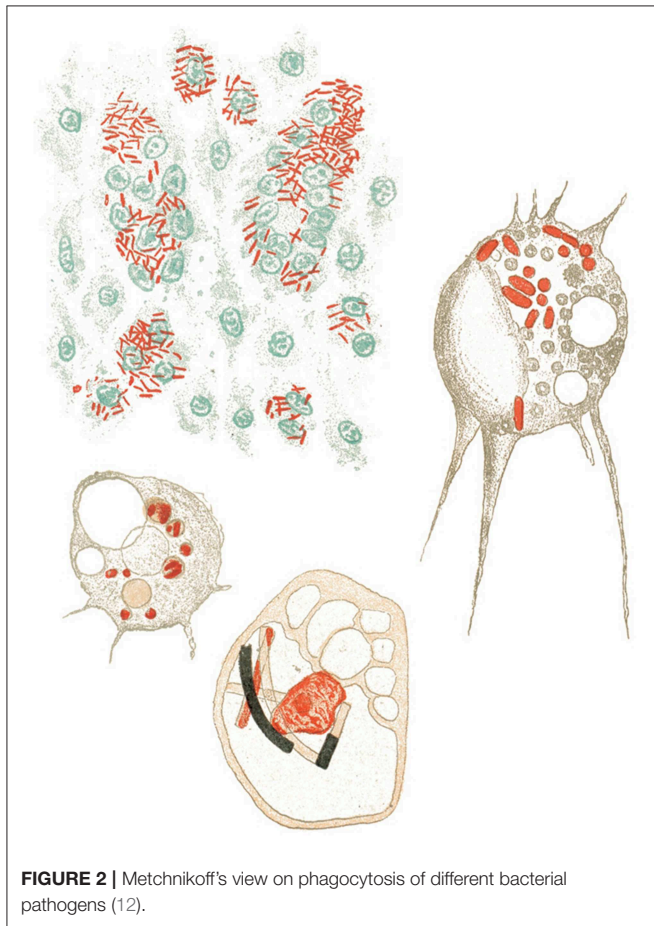
Metchnikoff was born in 1845 in a part of Russia, which now belongs to the Ukraine (9). He studied zoology and soon became a traveling scientist. Notably, when working at the Zoological Station in Naples he studied simple organisms and identified specialized cells dedicated to nutrient uptake. These nutrients could be contained in particles and thus the concept of phagocytosis was conceived as a process of uptake of particles or microbes rich in food. Moreover, in his experiments with starfish larvae in Messina in 1883, Metchnikoff found that phagocytic cells were highly motile and migrated to sites of foreign insult (10). He later wrote about these groundbreaking observations:

"... I fetched from it a few rose thorns and introduced them at once under the skin of some beautiful starfish larvae as transparent as water. I was too excited to sleep that night in the expectation of the result of my experiment and very early the next morning I ascertained that it had fully succeeded. That experiment formed the basis of phagocyte theory to the development of which I devoted the next 25 years of my life ..." (11).

Indeed, Metchnikoff changed his scientific interests from zoology to pathology and in this way became one of the first immunologists. He discovered phagocytes in vertebrates and began analyzing phagocyte functions in infectious diseases, such as anthrax, sepsis, and tuberculosis (**Figure 2**). Based on these



**FIGURE 1** | Immunology's early days.



**FIGURE 2** | Metchnikoff's view on phagocytosis of different bacterial pathogens (12).

which can be easily stained, with a largely polynuclear and fragmented nucleus and faint protoplasm...." (13).

## Serum Therapy and Antibodies

Behring was born in the German province of Prussia, now part of Poland, in 1854 (14). He studied medicine at an army academy and soon became interested in studies on the curative activity of disinfectants in bacterial infections. During his experiments on antiseptic activity of small molecules, together with the Japanese guest researcher Shibasaburo Kitasato (1853–1931) at the Institute for Infectious Diseases in Berlin, he discovered that serum from infected animals contained antibacterial activity that was specific for the infectious agent (15). Essentially, the activity was directed against the bacterial toxin. Whilst the joint paper of Behring and Kitasato mostly focused on tetanus and its toxin, the single-authored paper by Behring published shortly thereafter, described protection against diphtheria and its toxin by antisera (15, 16). Soon these animal experiments were translated into a human study, which revealed that serum therapy protected against diphtheria when given during early stages of infection or even during disease. Behring joined forces with industry to produce large doses of antisera for human use, thus embodying the translational immunologist with great interest in medical application (**Figure 3**). His serum therapy was a breakthrough and honored by the first ever awarded Nobel Prize in Medicine in 1901 (4).

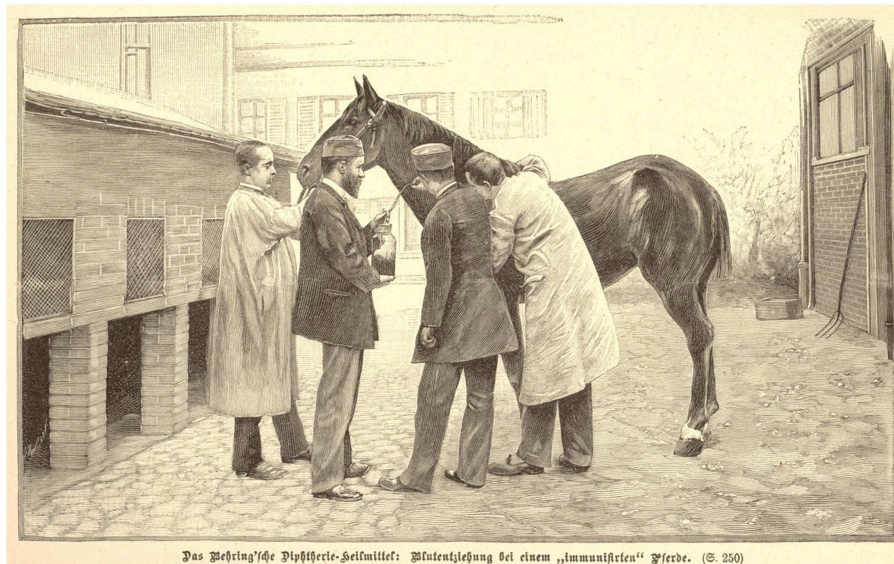
Serum therapy was more than just a curative method. It also provided supportive evidence for the idea that the cause of infectious disease is highly specific and that this specificity is linked to toxins produced by the etiologic pathogen. As a corollary, the cure of the specific disease was accompanied by a specific poison-averting (antitoxic) agent, which circulates in blood and can offer specific protection against the toxin in other individuals (15–17).

Despite all the honors he received, Behring was not fully satisfied with passive vaccination. It took him some 20 years to solve the issue of active vaccination (18). In 1913, at the Congress for International Medicine in Wiesbaden, Germany,

studies, he distinguished macrophages from microphages (which we now call neutrophils) according to the form of their nucleus:

"... I suggest calling all elements macrophages, which generally possess a simple non-polymorphic nucleus that is round or frequently oval. ... as microphages I call smaller amoeboid cells,





**FIGURE 3** | Large-scale production of serum against diphtheria toxin.

Behring gave a remarkable presentation, which the newspaper “Vossische Zeitung” (April 18, 1913) described quite aptly:

“At today’s discussions, Behring appeared as lively as ever and reported on a new protective agent comprising a mixture of diphtheria toxin and anti-toxin. This agent was harnessed for treating individuals at risk prophylactically. It was found that first the agent was completely innocuous, and second that the appearance of true protection could be demonstrated by the formation of sufficiently high abundance of protective agents in the blood of immunized individuals who all remained free of diphtheria” (14).

In order to neutralize the diphtheria toxin, Behring generated antigen-antibody complexes, which stimulated production of toxin-specific antibodies in the immunized host. This was an important, but still suboptimal start toward active vaccination against bacterial toxins. It was the French researcher, Gaston Ramon (1886–1963), who ultimately introduced detoxification by formaldehyde for low-cost production of safe vaccines against diphtheria and tetanus, and aluminum hydroxide as adjuvant for potent immunization (19, 20).

Whilst Behring was a translational immunologist, who contributed significantly to basic immunology, Ehrlich was most interested in the in-depth understanding of basic mechanisms underlying immunity, and contributed profoundly to the clinical development of serum therapy. Indeed it was Ehrlich whose contribution made large-scale production of antisera of reproducible quality possible. By working out “a new and more accurate method for determining the value of the serum and to study the complex relations which govern the neutralization of toxin and antitoxin,” he could show that “... the immunity unit is no longer an arbitrary concept, but is an exactly determinable quantity and one therefore which can be reproduced afresh at

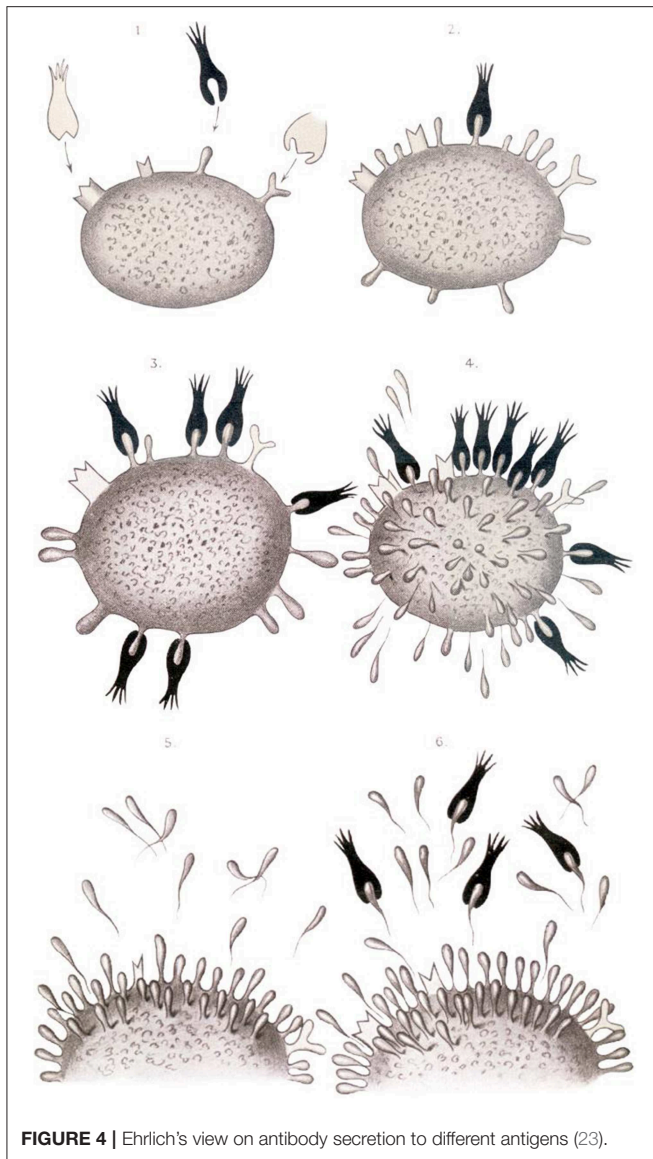
any time ...” (21). Ehrlich was therefore the first to provide the basis for a quality control measure of a biological. At those times, this was urgently needed because of widespread state-controlled compulsory vaccination against smallpox.

Yet, Ehrlich became most famous for basic research of, and stimulating ideas on, how the immune system works. In his MD thesis, Ehrlich described mast cells which, as we now know, are critical effectors of allergy (22). But his most important findings are related to antibodies. He foresaw that antigens, such as toxins, stimulate the production of specific antibodies. Interestingly, similar to Metchnikoff, Ehrlich assumed a nutritional point of view (22). Different cells need different kinds of nutrients and hence Ehrlich postulated specific receptors as being responsible for nutrient uptake. From this he concluded that the cell receptor specific for a given toxin should fulfill similar criteria. Because of the sheer abundance of toxins generated during infection, more specific receptors are produced and are ultimately secreted into the serum (**Figure 4**). In the Croonian Lecture given in 1900 at the Royal Society, Ehrlich reflected on his ideas as follows:

“... the first stage in the toxic action must be regarded as being the union of the toxin .... to a special side chain of the cell protoplasm. ... the side chain involved, so long as the union lasts cannot exercise its normal physiological nutritive function. .... such an excess of side chains is produced that to use a trivial expression, the side chains are present in too great quantity for the cell to carry and are, after a manner of secretion, handed over as superfluous ballast to the blood ...” (23).

Essentially this is the core message of the side chain theory for which Ehrlich is most renowned. But Ehrlich was far more productive. He showed that the milk of breastfeeding mothers carries antibodies beneficial to the suckling infant, thus providing the child with a high degree of immunity (24). He speculated





on the role of tolerance to self and the risk of autoimmunity and coined the well-known term “horror autotoxicus” (24). He revealed several biological features of complement, which was originally discovered by the German scientist, Hans Buchner (1850–1902), and the Belgian researcher, Jules Bordet (1870–1961), who termed it alexine (25). Ultimately, however, the term complement created by Ehrlich prevailed. Bordet and Buchner had already shown that alexine was heat-labile (25–27). Buchner used serum from non-immunized animals, whereas Bordet included serum from immunized animals in his studies and so distinguished the heat-labile alexine from the heat-stable antibodies. Ehrlich, together with his colleague Richard Pfeiffer (1858–1945), further defined the activities of antibodies and complement by mixing untreated and heat-inactivated serum. In his own words, Ehrlich summarized this finding: “The two substances are (i) the specific immune body produced by

immunization and (ii) a substance which usually is thermo-labile, contained even in normal serum” (28).

In 1908 Ehrlich and Metchnikoff were jointly awarded the Nobel Prize in Physiology or Medicine “in recognition of their work on immunity” (29). Bordet was honored “for his discoveries relating to immunity” with the Nobel Prize in 1919 (30).

The interaction of complement and antibodies was the first dent in the dichotomous view of immunity (**Figure 1**). Complement was part of the innate immune response and hence non-specific. But it was humoral. Thus, the exclusive association of innate immunity with cells had become obsolete. More importantly, specific antibodies cooperated with non-specific complement.

The dichotomous view of immunology was further softened by the experiments of the English scientist, Almroth Wright (1861–1947), who showed that antibodies can specifically facilitate phagocytosis of bacteria (31, 32). This is of particular importance for efficient defense against bacterial pathogens which evade phagocytosis, such as encapsulated bacteria (pneumococci, meningococci and gonococci). His finding revealed that for some diseases, specific antibodies are needed to interact with phagocytes for optimal host defense (31, 32). For the first time therefore, specific humoral factors of the acquired immune response (antibodies) were shown to collaborate with non-specific cognates of the cellular innate immune response (macrophage and neutrophils). This was another call for complementary dualism rather than dichotomy between innate and acquired immunity. The findings of Wright caught the interest of George Bernard Shaw (1856–1950), who described the potential of phagocytes for cellular therapy of disease. In Act I of “The Doctor’s Dilemma,” he writes: “There is at bottom only one genuinely scientific treatment for all diseases and that is to stimulate the phagocytes.” During the play, however, the risk of adverse events of such therapy is increasingly recognized and culminates in the question: “Have we overstimulated the phagocytes? Have they not only eaten up the bacilli but attacked and destroyed the red corpuscles, as well?” Adoptive phagocyte therapy never made it into the clinics as an immunologic treatment regimen.

## ACT II: IMMUNOCHEMISTRY AND CLINICAL IMMUNOLOGY

### Immunochemistry

During the first half of the twentieth century, immunologists focused on clinical observations and even more on immunochemistry, which could build on a much broader armamentarium of technical tools. Immunochemistry found its culmination in the discovery of the chemical structure of antibodies (**Figure 4**). This was accomplished independently by the British chemist, Rodney Porter (1917–1985), and the US chemist, Gerald Edelman (1929–2014), in the late 1950s to early 1960s (33, 34). Their work was honored by the Nobel Prize in 1972 (35). The Austrian Karl Landsteiner (1868–1943), first working in Europe and since 1923 in the US, developed the carrier hapten concept by coupling small aromatic molecules to

proteins (36). He showed that the small residue—the hapten—is recognized by antibodies, and therefore serves as epitope, and that the protein serves as carrier to provide the immunogenicity needed for successful stimulation of an antibody response (37, 38). Since the studies of Jacques Miller (1931–), Henry Claman (1930–2016) and others, we know that the antibody response involves B lymphocytes for the recognition of the hapten and T lymphocytes for the recognition of the carrier.

## Hypersensitivity Reactions

Landsteiner is probably best known for the discovery of the ABO major blood group system (39). Working at the time in Vienna, he found that mixing blood of two different individuals resulted in clumping of red blood cells. Based on this finding, he developed a technique for the serologic differentiation of erythrocytes, which allowed him to identify the different blood groups of the ABO system. This discovery was honored by the Nobel Prize in 1930 (40). Ten years later, and together with Alexander Wiener (1907–1976), Landsteiner discovered a second important blood group, called Rhesus (Rh), named after their original discovery with erythrocytes in Rhesus monkeys (41, 42).

Landsteiner's discovery of so-called isoagglutinins—the antibodies responsible for clumping of erythrocytes when mixed with serum from a donor of a different ABO blood group—were criticized by Paul Ehrlich who considered this finding contradictory to his proposed “horror autotoxicus.” Yet, increasing evidence arose that horror autotoxicus, i.e., autoimmune attack against host cells or molecules was not an absolute no-go for the immune system. It became clear that antibodies do not only perform beneficial functions. That aberrant antibody responses could lead to hypersensitivity reactions was first shown by the French clinician Charles Richet (1850–1935) in 1902 (43), who was awarded the Nobel Prize for his research on anaphylaxis in 1913 (44). The term anaphylaxis was coined by Richet to describe harmful reactions, which were later shown by the Japanese immunologist Kimishigi Ishizaka (1925–2018) and his wife Teruko (1926–), to be mediated by antibodies of the IgE isotype (45). One year after Richet's discovery, the French researcher, Maurice Arthus (1862–1945), described a similar yet distinct type of reaction which he induced experimentally by local injection of antigen into the skin of an individual previously immunized with the same antigen (46). In contrast to the reaction described by Richet, this one was mediated by immune complexes and involved complement. With serum therapy against diphtheria and tetanus broadly applied, numerous individuals received serum from horses in which the antiserum had been generated. In 1905, the clinicians, Clemens von Pirquet (1874–1929) from Austria, and Béla Schick (1877–1967) from Hungary, together observed that multiple injections of such serum could result in serum sickness due to the formation of immune complexes (47). They termed this type of reaction “allergy,” which has come to be applied in a broader sense. Yet, another hypersensitivity reaction was first observed by the Japanese physician, Hakaru Hashimoto (1881–1934), in 1912 (48): “Hashimoto's thyroiditis” turned out to be an autoimmune disease partially mediated by IgG antibodies, which facilitate damage by phagocytes and NK cells. This type of hypersensitivity

is also the basis of erythrocyte damage after blood transfusion, e.g., from ABO-disparate donors. At Rockefeller University, Karl Landsteiner together with the American researcher, Merrill Chase (1905–2004), studied the tuberculin reaction first described by Robert Koch and demonstrated that this reaction can be adoptively transferred by cells of an immune animal but not by serum (49). As we know now, the “delayed-type hypersensitivity” reaction mostly involves T lymphocytes.

The four different types of hypersensitivity were categorized by the UK physicians, Philip Gell (1914–2001), and Robin Coombs (1921–2006), in 1963 (50). In this categorization, type I hypersensitivity is the typical IgE-mediated allergy first described by Richet; type II is IgG plus complement-mediated destruction of host cells; type III is mediated by immune complexes such as the Arthus reaction; and type IV is the delayed-type hypersensitivity reaction, including the tuberculin reaction and contact dermatitis. Hashimoto's thyroiditis, originally considered type II, is now known to be a mix of type II and type IV, i.e., it is antibody- and T cell-mediated.

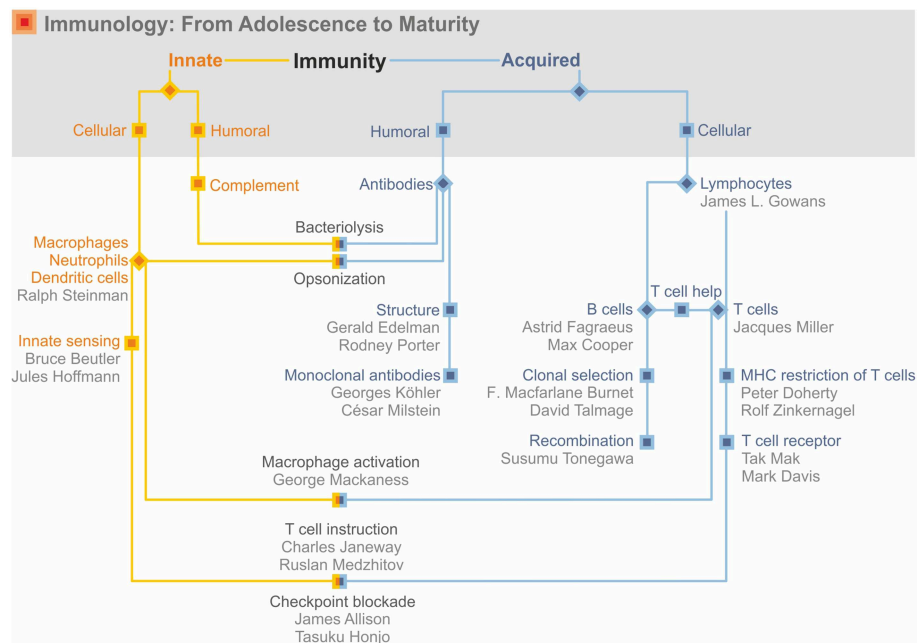
## ACT III: THE RISE OF IMMUNOBIOLOGY

### Transplantation Biology

The 1950s to 1960s witnessed a marked shift in priorities from immunochemistry to immunobiology (Figure 5). In fact, studies on transplant rejection preceded and prepared the ground for immunobiology. The US geneticist George Snell (1903–1996), based on his studies with inbred mouse strains, elegantly demonstrated that distinct genes within the major histocompatibility complex (MHC) were responsible for transplant rejection (51). The French clinician, Jean Dausset (1916–2009), discovered the human MHC, also named human leukocyte antigen (HLA), on the basis of family studies (51). A somewhat more direct link to immunobiology was provided by the Venezuelan-born US scientist, Baruj Benacerraf (1920–2011), who identified the immune response genes within the MHC locus (51). In 1980, Snell, Dausset and Benacerraf were honored by the Nobel Prize “for their discoveries concerning genetically determined structures on the cell surface that regulate immunological reactions” (52). Later the Australian researcher, Peter Doherty (1940–), and the Swiss researcher, Rolf Zinkernagel (1944–), would broaden this perspective by showing that the MHC is crucial for antigen recognition by T lymphocytes, the cells that would become the dominant research target in the second half of the twentieth century.

### Antibody Specificity Revisited

The Australian virologist, Frank Macarlane Burnet (1899–1985), and the UK biologist, Peter Brian Medawar (1915–1987), received the Nobel Prize in 1960 “for their discovery of acquired immunological tolerance” (53). It was they who provided first evidence that the horror autotoxicus, envisaged by Paul Ehrlich, was not prefixed but a matter of education. Medawar had shown that transplant rejection could be prevented by transferring cells from an unrelated donor during neonatal life (54, 55). Cells from the same donor were later accepted by such mice showing that during fetal and neonatal development



**FIGURE 5 |** Immunology: from adolescence to adulthood.

the immune system “learned” to accept self. Indeed it was Burnet who outlined the concept of “self vs. non-self” (55). Although his concept remained speculative and was questioned because of the occurrence of autoimmune diseases, it proved to be a valid theory of immunobiology even though—as with many biological issues—it was not absolute. In fact, impact of “self vs. non-self” on immune tolerance remains a matter of controversial discussions—not the least after the realization that self/non-self discrimination is not only a matter of the acquired but also of the innate immune response (see below). Burnet’s interests were much broader. Originally a virologist who became an immunologist, he readily used tools of virology to interrogate the immune system. He is probably most famous for postulating the “clonal selection” theory, which again had been triggered by Paul Ehrlich (56). Although Ehrlich’s side chain theory held that antibody specificities of all kinds were present before antigen encounter, according to Ehrlich numerous specificities could be expressed by a single cell depending on its requirement for specific nutrients (see **Figure 4**). This assertion, however, was questioned during the area of immunochemistry when a chemical explanation was sought for a biological question. Several researchers including the US Nobel laureate of 1954 and 1963, Linus Pauling (1901–1994), claimed that the structure of the antigen would determine the specificity of its corresponding antibody (57). In the “template hypothesis,” the antigen binding site was the result of a specific chemical formation around a foreign entity. With the understanding that the three-dimensional structure of a protein is strongly determined by its amino acid sequence, this became a matter of impossibility.

The Danish immunologist, Niels Jerne (1911–1994), who received the Nobel Prize in 1984 (58), postulated a more biologically oriented hypothesis, namely that various antibody specificities existed prior to antigen encounter (59). This was then refined by Burnet and independently by the US immunologist David Talmage (1919–2014), who both proposed a selection process for the specific antibody-producing cell (56, 60). Thus, Ehrlich was right in assuming the preexistence of antibody specificities before a foreign antibody arrived, but he was wrong in assuming that one cell would express numerous specificities. Elegant studies by the Australian immunologist, Gustav Nossal (1931–), partly together with US Nobel laureate of 1958 Joshua Lederberg (1925–2008), provided strong evidence that a single cell produces an antibody of unique specificity (61, 62). Under the influence of the specific antigen, the antibody-producing cells expand numerically and produce more antibodies of the same specificity. Hence, interest in antibodies shifted from chemical structure to biological understanding of the generation of specificity, i.e., on the antibody-producing cell.

### Lymphocytes as Masters of Ceremony

The major cell type of the acquired immune response, however, was still missing (**Figure 5**). It was the Australian immunologist, Jacques Miller (1931–), who discovered the role of the thymus in the development of a specific lymphocyte population; this finding led to the identification of T lymphocytes as major regulators of the acquired immune response (63). Independent from Miller, the US transplant immunologist Robert Good (1922–2003) characterized the role of the thymus and other lymphoid organs in the generation of different lymphocyte populations (64, 65). At about the same time, the UK immunologist, James Gowans



(1924–), had shown that the lymphocyte population was able to recirculate through the body and enter the different tissue sites—an important and necessary feature for T lymphocytes which mediate cellular immunity and hence depend on cell–cell contact (66). The producers of antibodies had been identified earlier, namely in 1940 by the Swedish researcher, Astrid Fragaues (1913–1997), as plasma cells (67, 68). Her work as well as that of the US immunologist, Max Cooper (1933–) then led to the revelation that plasma cells are derived from B lymphocytes which develop in the Bursa fabricii in birds and in the bone marrow in mammals (64, 65, 69).

Now the major cells of the acquired immune response had been identified and immunologists increasingly focused on their biological functions (**Figure 5**). Henry Claman (1930–2016) was probably the first to provide compelling evidence that T lymphocytes and B lymphocytes collaborate in the generation of antigen-specific antibodies (70). Av Mitchison (1928–) showed that antibodies were specific for the epitope (Landsteiner's small residues—the haptens) and T cells for the protein carrier (71). The establishment of T lymphocytes and B lymphocytes as responsible cells of acquired cellular and humoral immunity, respectively, and their collaboration in shaping an optimal immune response laid the basis for the golden age of cellular immunity.

Following the footsteps of the founders of immunology, the Australian borne researcher working in the US, George Mackaness (1922–2007), extensively studied immunity against intracellular bacteria. He discovered the cooperation between specific T lymphocytes and mononuclear phagocytes. In this setting, antigen specific T cells stimulate increased antibacterial activities in macrophages which thereby change from a habitat for the intracellular pathogens to the major effectors of cell-mediated immunity against the infection (72).

Transplantation biology and immunobiology converged when Peter Doherty and Rolf Zinkernagel demonstrated that MHC molecules were not only responsible for transplant rejection, but for T-cell recognition of any type of antigen. Antigen recognition by T lymphocytes, therefore, was MHC-restricted and transplant rejection was just one special case (73). Their breakthrough work, honored by the Nobel Prize in 1996, was based on antigen recognition by cytolytic T lymphocytes, which kill virus-infected cells (73, 74). Soon these cells were characterized phenotypically as CD8 T cells, which were MHC I-restricted. CD8 T cell counterparts, the CD4 T cells, were MHC II-restricted and shown to activate other cells of the immune system, notably B cells and macrophages by means of soluble factors, the cytokines. Activation of macrophages increases antibacterial activities, which in turn allows macrophages to control intracellular bacteria, such as the causative agent of tuberculosis. B cell activation leads to the production of antibodies of different isotypes. CD4 T cells were also found to help CD8 T cells become killer T cells. The first molecularly defined T cell cytokine was interleukin-2 (IL-2), which was originally described by the US immunologist, Kendall Smith (1933–) (75). His findings paved the way for the discovery of numerous humoral mediators of T cell immunity. With the identification of many other cytokines, the concept of T helper

1 (TH1) vs. T helper 2 (TH2) cells was developed by the Canadian immunologist Tim Mosmann (1949–) and the US immunologist Bob Coffman (1949–) (76). CD4 T cells of TH1 type contribute to the cellular immune response by activating killer T cells and macrophages. IL-2 was identified as the major mediator of killer T cell activation and interferon- $\gamma$  (IFN- $\gamma$ ), which had already been described earlier as immune IFN was shown to be critical for macrophage activation. In contrast, TH2 cells produce IL-4 and other cytokines, which stimulate B lymphocytes to mature to antibody-producing plasma cells. Early on it was recognized that the immune response is highly regulated and notably that a well-functioning immune response need not only be activated to combat an intruder, but also needs to be downregulated once the intruder had been eliminated. This led to the concept of a highly regulated immune response involving specific T cells with suppressive functions to avoid collateral damage. Early attempts to explain this issue postulated suppressor T cells which, however, did not stand the test of time. The more refined concept of the better defined subsets of regulatory T cells, however, provided compelling evidence for specific T lymphocytes which not only control immune responses after elimination of invading pathogens, but also prevent autoimmunity and maintain homeostasis (77).

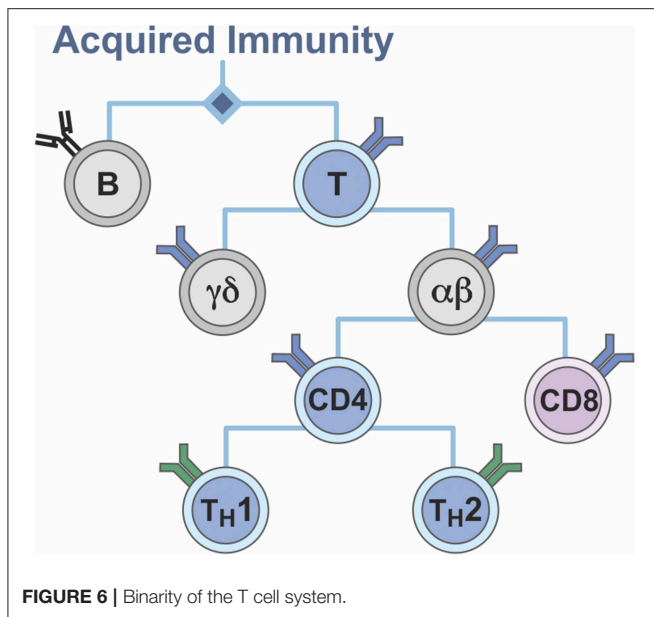
Although the biological functions of T lymphocytes were increasingly better understood, their antigen receptors remained elusive until the 1980s. By using monoclonal antibodies, US immunologists, Pippa Marrack (1945–) and John Kappler (1943–) (in the mouse system) (78), and Ellis Reinherz (1950–) and Stuart Schlossman (1935–) (in the human system) (79), were able to phenotypically identify antigen-specific receptors on T lymphocytes. This was the first hint for the existence of the antigen-specific T cell receptor (TCR). Soon thereafter, genes encoding TCR chains were cloned by Tak Mak (1946–) in Canada and Mark Davis (1952–) in the US (80, 81).

The T lymphocyte system can thus also be viewed as a binary system (**Figure 6**). Lymphocytes segregate into B and T cells; T cells segregate into MHC I- and MHC II-restricted T cells of CD4 or CD8 phenotype, respectively; CD4 T cells separate into TH1 and TH2 cells; the vast majority of T cells express a T cell receptor composed of an  $\alpha$  and a  $\beta$  chain, but a second T cell population exists, which expresses a T cell receptor comprising a  $\gamma$  and a  $\delta$  chain. Again, support was withdrawn for a dichotomous view, in favor of a complementary dualism (**Figure 5**).

## Recombination Generates Diversity

These important findings were preceded by the breakthrough discovery of the Japanese researcher, Susumu Tonogawa (1939–), then in Basel, Switzerland, who elucidated the mechanisms underlying the huge diversity of antibody specificities (82, 83). By then it was generally accepted: a single specific B cell was responsible for antibody production; diversity was generated prior to the first contact with antigen; a single B cell expresses a receptor with a unique specificity; contact with the homologous antigen stimulates selective expansion and differentiation of the specific B cell. Yet, one critical issue remained unsolved, namely that the number of possible antibody specificities exceeded the number of genes present in our





body. The solution to this was identified by Tonegawa as the rearrangement of gene fragments. This recombination allows the generation of more than one million specificities which further increases numerically by additional mechanisms to up to some  $10^9$  specificities. Tonegawa was honored with the Nobel Prize in 1987 “for the discovery of the genetic principle for generation of antibody diversity” (84). Principally, antigen diversity of the T cell receptor is based on similar genetic mechanisms.

### T-Cell Instruction by Antigen-Presenting Cells

In any case, the specificity of the acquired immune response and the multiple roles played by T cells more or less dominated immunobiology in the 1960s to 1990s. An influential researcher in the field of T cell immunology was Charles Janeway (1943–2003) from the US (85), who in a remarkable paper published in 1989 in the Proceedings of the Cold Spring Harbor Symposium, pointed to the widely underestimated role of the innate immune system (86). Prevailing opinion was that innate immune cells, notably macrophages and neutrophils, play an important effector role in host defense, under the guidance of T lymphocytes and their soluble products. Even though it was clear that T cells recognize antigens in the context of MHC presented on the surface of so-called antigen-presenting cells, these cells were viewed more as passive guides than active players. Janeway postulated the presence of pattern recognition receptors on antigen-presenting cells, which sense specific motifs of chemical products of bacteria and viruses and then instruct T cells about the different functions they should perform. Most compelling evidence for such an idea came from studies on the toll-like receptors (TLR) in mammals by the US geneticist Bruce Beutler (1957–), and in insects by the biochemist Jules Hoffmann (1941–) in France (87, 88). This led to the concept that different types of pathogens are sensed by pattern recognition receptors with

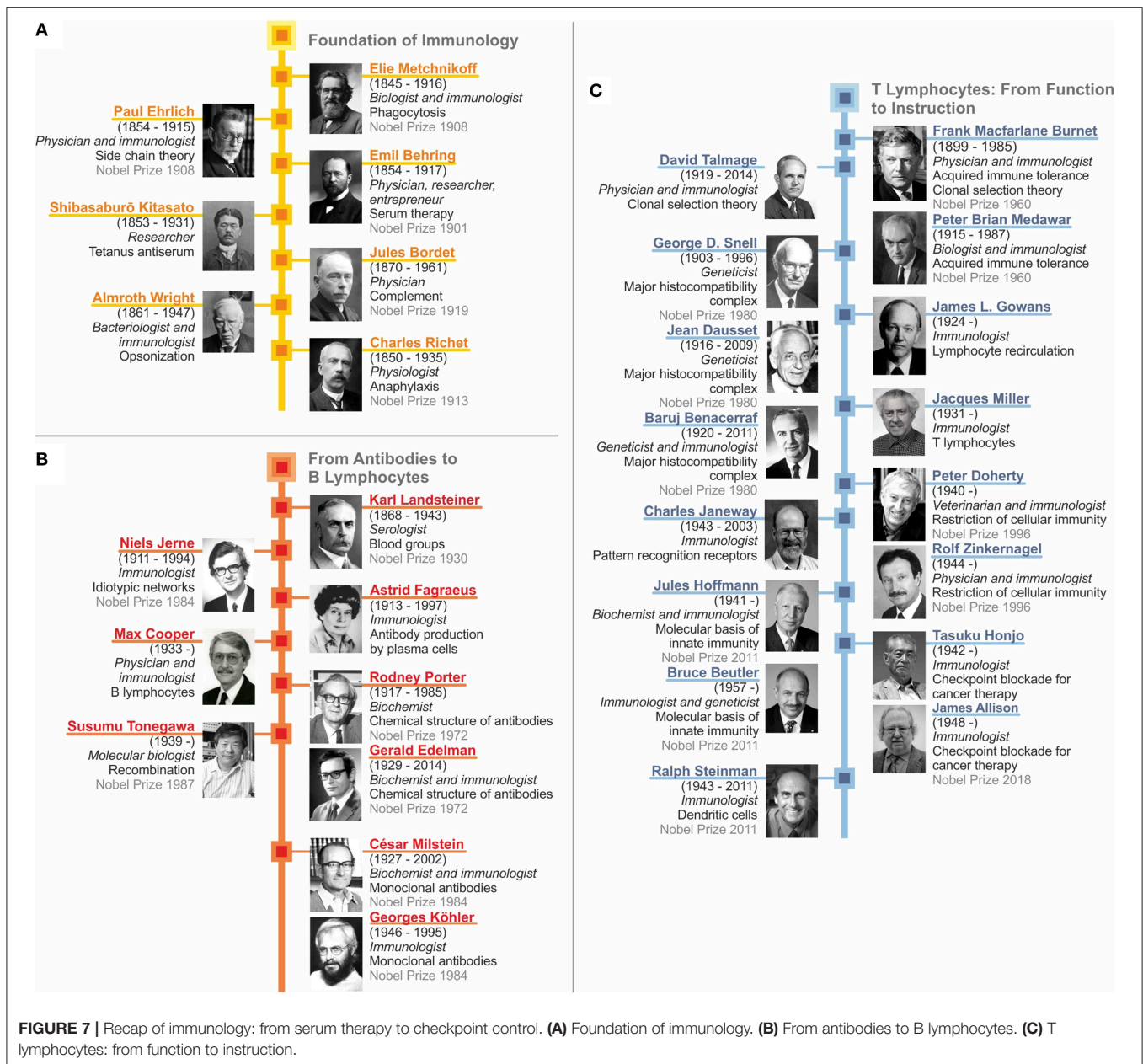
specificity for microbe-associated molecular patterns. Beutler and Hoffmann jointly received the Nobel Prize in 2011 “for their discoveries concerning the activation of innate immunity” (89). The concept of sensing of microbial motifs (so-called pathogen-associated molecular patterns, PAMP) by innate receptors was soon broadened when similar mechanisms were found to be induced by host motifs (so-called danger associated molecular patterns, DAMP) which arise from insult to the host (90). In how far PAMP and DAMP influence immune tolerance by inducing danger associated non-self or self-signals to the induction of an acquired immune response remains a matter of controversial discussion (91, 92).

As early as the 1970s, the Canadian immunologist Ralph Steinman (1943–2011) at Rockefeller University, US, was engaged with defining the critical player in this concept: the dendritic cell (93). He demonstrated that dendritic cells are much more potent antigen presenters than macrophages, and that they are the major instructors of T cells regarding the type of pathogen they will encounter. Steinman was the third to be honored by the Nobel Prize 2011 “for his discovery of the dendritic cell and its role in adaptive immunity” (89). Sadly he could not accept the award in person because he passed away shortly before the ceremony. In conclusion, innate immunity plays a crucial role, from the beginning to the end of an immune response. In the beginning it acts via antigen-presenting cells, which not only stimulate antigen-specific T cells but also serve as instructors for the biological functions T cells have to perform. Toward the end, innate immunity takes care of effector functions, e.g., via professional phagocytes which eliminate invading pathogens.

Instruction of T cell functions strongly depends on cytokines, i.e., humoral factors. Thus, IL-12 induces TH1 cells whereas IL-4 directs TH2 cells. In fact, the first chemically defined cytokine was described by the US immunologist, Charles Dinarello (1943–) as a macrophage-derived product, which accordingly was later named IL-1 (94). IL-1 plays a role in the instruction of TH1 cells and serves as mediator of inflammation.

### From Serum Therapy to Checkpoint Control

B cells stood in the shadow of T lymphocytes during the 1970s. The discovery by the Argentinian researcher, Cesar Milstein (1927–2002), and the German researcher, Georges Köhler (1946–1995), both working in the UK, brought them back to center stage. In 1984, both shared the Nobel Prize “for the production of monoclonal antibodies” (58). Obviously, this discovery had major implications. First, it allowed the ultimate proof for the production of an antibody with single specificity by a single plasma cell and second, it paved the way for a new era of immunotherapy. As a short reminder, the concept of acquired immunity started with antibodies and was intrinsically intertwined with the concept of serum therapy, for which Behring received the Nobel Prize in 1901. Now the tools for more precise passive immunization had been put on the table. This led to the development of a number of monoclonal antibody-based therapies for infectious diseases; currently, the focus of monoclonal antibody therapy is on immunomodulation. Thus,



cytokine-blocking monoclonal antibodies have been introduced in the treatment of chronic inflammatory diseases. Most notable are Infliximab and Adalimumab, which block the critical cytokine tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) in Crohn's disease and rheumatoid arthritis, respectively (95, 96). A second important target of therapeutic monoclonal antibodies are surface-expressed molecules such as CD20 on B lymphocytes, which can be harnessed for treatment of non-Hodgkin's lymphoma such as Rituximab (97).

A major recent breakthrough has been the discovery of monoclonal antibodies which block checkpoint control. What does this mean? Regulation of T cell activity is not only a matter of cytokines but also of costimulatory molecules, which

in addition to TCR recognition of antigen plus MHC as first signal, provide a second signal for T cells in stimulating their effector functions. Eventually, the immune response needs to be dampened. Once it has completed its task, e.g., after the elimination of an infectious agent, it needs to be tuned down to avoid or at least minimize collateral damage. Surface-expressed inhibitory molecules include CTLA-4 and PD-1 on T cells and their counterparts B7 and PD-L1 on antigen-presenting cells (98, 99). These counterparts are also expressed on many tumor cells, which block attack by killer T cells. Blockade of checkpoint control improves T cell responses and thereby allows elimination of certain tumor cells. This finding led to next-generation immunotherapies for certain cancers including

metastatic melanomas and non-small cell lung carcinomas. The highly promising checkpoint blockade for cancer therapy was honored by the Nobel Prize 2018 to the US immunologist Jim Allison (1948–) and Japanese immunologist Tasuku Honjo (1942–) “for their discovery of cancer therapy by inhibition of negative immune regulation (5).”

## SHORT RECAP AND OUTLOOK

As we have seen, immunology as a scientific discipline was kick-started by two seminal discoveries: First, the role of phagocytosis performed by cells and second, the neutralization of bacterial toxins by antibodies. This led to the concept of dichotomous roles of antigen-unspecific innate immunity mediated by cells and antigen-specific acquired immunity mediated by humoral factors. This dichotomous concept converged with the identification of complement and opsonization, which linked innate and acquired immunity. Major early contributors are depicted in **Figure 7A**. The intermediate stage includes the discovery of different forms of clinical hypersensitivity emphasizing that the immune system also embodies detrimental functions. In parallel, immunochemistry reached its climax with the elucidation of the crystal structure of antibodies. Then immunobiology took over with the identification of lymphocytes and their segregation into antibody-producing B cells and plasma cells, as well as T cells, which function as central regulators of immunity (**Figures 7B,C**). TH cells were shown to control B lymphocytes, professional phagocytes and cytolytic T cells. Finally, this dysbalanced perception of acquired immunity dominating innate immunity was rectified by our increasing understanding of how antigen-presenting cells instruct the acquired immune response (**Figure 7C**). Today sufficient knowledge has been accumulated in immunology to devise sophisticated therapeutic approaches, such as checkpoint control for cancer treatment. Yet, in both basic and applied immunology, sufficient challenges persist which guarantee that our discipline will remain as vital as ever.

Importantly, the immune apparatus is increasingly seen as a highly diffuse organ comprising not only bone marrow, thymus and spleen, but also lymph nodes and lymphoid follicles which are spread throughout the body and interconnected by circulating leukocytes and soluble mediators. Accordingly, immune cells are imprinted by their organ of residence to adjust to the special regional needs. Reciprocally, immune cells impact

on the tissue of their main residence. Moreover, our microbiome is increasingly viewed as a human organ vital to health and disease and tightly intertwined with the immune system. As a corollary, dysfunctions of regional immune responses underlie many organ-specific diseases. Future immunology will have to take into account an integrated view on these crosstalks at all levels from organs to tissues to cells to molecules. The enormous advances in high-throughput multi-omics technologies and bioinformatics allow studies on multiple levels of the immune response thus providing a wealth of data which will ultimately result in the construction of molecular multi-networks of the immune response under physiologic and pathologic conditions. Ultimately, this system biology approach will provide a far more comprehensive perspective of immunology which will generate new concepts for prevention and treatment of diseases that are refractory to current intervention strategies due to dysfunctional, insufficient or subverted immunity. Paul Ehrlich's dream of “magic bullets” will take a step closer to reality by the immunology of the future.

## AUTHOR CONTRIBUTIONS

The author confirms being the sole contributor of this work and has approved it for publication.

## ACKNOWLEDGMENTS

A short perspective on the history of immunology is not a comprehensive account of all the different contributions to immunology. And by definition, a perspective is inherently biased. Hence, the views expressed here reflect my personal opinions and should not be taken as consensual reporting on the history of immunology. Wherever a Nobel Prize was awarded in recognition of an important breakthrough in immunology, I focused on the Nobel laureates' work. I am well-aware that with this approach, I may have missed important contributions by others. I apologize to all whose work I have omitted in my attempt to concisely summarize the history of immunology in this short overview. German citations have been translated freely into English by myself. I thank Alan Sher for many helpful comments on the manuscript, Marylu Grossman for excellent editorial support, Souraya Sibaei for excellent secretarial assistance and Diane Schad for superb graphics.

## REFERENCES

1. Kaufmann SH. Immunology's foundation: the 100-year anniversary of the nobel prize to paul ehrlich and elie metchnikoff. *Nat Immunol.* (2008) 9:705–12. doi: 10.1038/ni0708-705
2. Kaufmann SHE. Emil Von behring: translational medicine at the dawn of immunology. *Nat Rev Immunol.* (2017) 17:341–3. doi: 10.1038/nri.2017.37
3. Silverstein AM. *A History of Immunology*, 2 ed. San Diego, CA: Academic Press (2009).
4. Behring EV. *Nobel Prize in Physiology or Medicine 1901*. Available online at: <https://www.nobelprize.org/prizes/medicine/1901/summary/> (accessed March 20, 2019).
5. Allison JP, Honjo T. *Nobel Prize in Physiology or Medicine 2018*. Available online at: <https://www.nobelprize.org/prizes/medicine/2018/summary/> (accessed March 20, 2019).
6. Kaufmann SH, Winau F. From bacteriology to immunology: the dualism of specificity. *Nat Immunol.* (2005) 6:1063–6. doi: 10.1038/ni1105-1063
7. Virchow R. *Die Zellulärpathologie*, Vol. 1. 4 ed. Berlin: August Hirschwald (1871).
8. Kuhn TS. *The Structure of Scientific Revolutions*. Chicago, IL: The University of Chicago Press. (1970).
9. Metchnikoff O. *The Life of Elie Metchnikoff 1845-1916*. Boston, MA; New York, NY: Houghton Mifflin Co. (1921)



10. Metschnikoff E. Eine neue Entzündungstheorie. *Allg Wein Med Ztg.* (1884) 27/29:307–32.
11. Metchnikoff E. My stay in messina (in Russian). *Russk Vedomosti.* (1908) 31:302.
12. Metchnikoff E. *Immunität bei Infektionskrankheiten.* Jena: Verlag von Gustav Fischer (1902), 1–456.
13. Metschnikoff E. Ueber den Kampf der Zellen gegen Erysipel-Kokken. *Archiv für pathologische Anatomie und Physiologie für klinische Med.* (1887) 107:209–49. doi: 10.1007/BF01926053
14. Kaufmann SH. Remembering emil von behring: from tetanus treatment to antibody cooperation with phagocytes. *mBio.* (2017) 8:e00117. doi: 10.1128/mBio.00117-17
15. Behring E, Kitasato S. Ueber das Zustandekommen der Diphtherie-Immunität und der Tetanus-Immunität bei Thieren. *Dt med Wochenschrift.* (1890) 49:1113–4.
16. Behring E. Untersuchungen ueber das Zustandekommen der Diphtherie-Immunität bei Thieren. *Dt med Wochenschrift.* (1890) 50:1145–8.
17. Behring E. *Die Geschichte der Diphtherie (Mit besonderer Berücksichtigung der Immunitätslehre).* Leipzig: Verlag von Georg Thieme (1893).
18. Behring E. Über ein neues Diphtherieschutzmittel. *Dt med Wochenschrift.* (1913) 19:873–6.
19. Ramon G. Sur La production de antitoxins. *C R Acad Sci.* (1925) 181:157–9.
20. Ramon G. Sur le pouvoir floculant et sur le propriétés immunisantes d'une toxine diphthérique rendue anatoxique (Anatoxine). *C R Acad Sci Paris.* (1923) 177:1338–40.
21. Ehrlich, P. The Collected Papers of Paul Ehrlich. In: Himmelweit F, Marquardt M, Dale H, editors. *Immunology and Cancer Research, Vol. II.* London; New York, NY: Pergamon Press. (1957), 86–107.
22. Ehrlich P. The Collected Papers of Paul Ehrlich. In: Himmelweit F, Marquardt M, Dale H, editors. *Histology, Biochemistry and Pathology, Vol. I.* London; New York, NY: Pergamon Press (1956), 19–28.
23. Ehrlich P. The Collected Papers of Paul Ehrlich. In: Himmelweit F, Marquardt M, Dale H, editors. *Immunology and Cancer Research, Vol. II.* London New York: Pergamon Press (1957), 178–195.
24. Ehrlich, P. The Collected Papers of Paul Ehrlich. In: Himmelweit F, Marquardt M, Dale H, editors. *Immunology and Cancer Research, Vol. II.* London; New York, NY: Pergamon Press. (1957), 234–45.
25. Bordet J. *Studies on Immunity.* New York, NY: John Wiley & Sons. (1909).
26. Buchner H. Ueber die bakterientödende Wirkung des zellenfreien Bluterserums. *Centrallblatte für Bakt. u. Parasitenkunde.* (1889) 5:817–23.
27. Buchner H. Ueber die bakterientödende Wirkung des zellenfreien Bluterserums (Schluss). *Centrallblatte für Bakt. u. Parasitenkunde.* (1889) 6:1–11.
28. Ehrlich P. The Collected Papers of Paul Ehrlich. In: Himmelweit F, Marquardt M, Dale H, editors. *Immunology and Cancer Research, Vol. II.* London; New York, NY: Pergamon Press. (1957), 213–23.
29. Mechnikov II, Ehrlich P. *Nobel Prize in Physiology or Medicine 1908.* Available online at: <https://www.nobelprize.org/prizes/medicine/1908/summary/> (accessed March 20, 2019).
30. Bordet J. *Nobel Prize in Physiology of Medicine 1919.* Available online at: <https://www.nobelprize.org/prizes/medicine/1919/summary/> (accessed March 20, 2019).
31. Forsdyke DR. Almroth wright, opsonins, innate immunity and the lectin pathway of complement activation: a historical perspective. *Microbes Infect.* (2016) 18:450–9. doi: 10.1016/j.micinf.2016.04.003
32. Wright AE. *Studies on Immunisation and Their Application to the Diagnosis and Treatment of Bacterial Infections.* London: Constable (1909).
33. Edelman GM. Antibody structure and molecular immunology. *Ann N Y Acad Sci.* (1971) 190:5–25. doi: 10.1111/j.1749-6632.1971.tb13520.x
34. Porter RR. Structural studies of immunoglobulins. *Science.* (1973) 180:713–6. doi: 10.1126/science.180.4087.713
35. Edelman GM, Porter RR. *Nobel Prize in Physiology or Medicine 1972.* Available online at: <https://www.nobelprize.org/prizes/medicine/1972/summary/> (accessed March 20, 2019).
36. Goldman AS, Schmalsteig FC. Karl Otto Landsteiner (1868–1943). Physician–biochemist–immunologist. *J Med Biogr.* (2016). doi: 10.1177/096772016670558. [Epub ahead of print].
37. Landsteiner K. Über heterogenetisches Antigen und Hapten. Xv. Mitteilungen über Antigene. *Biochemische Zeitschrift.* (1921) 119:294–306.
38. Landsteiner K, Jablons B. Ueber die antigeneigenschaften von acetyliertem eiweiss. VI. Mitteilung über die antigene. *Zeitschrift für Immunitätsforschung experimentelle Therapie. Jena. Originale.* (1914) 21:193–201.
39. Landsteiner K. Individual differences in human blood. *Science.* (1931) 73:403–9. doi: 10.1126/science.73.1894.403
40. Landsteiner K. *Nobel Prize in Physiology or Medicine 1930.* Available online at: <https://www.nobelprize.org/prizes/medicine/1930/summary/> (accessed March 20, 2019).
41. Landsteiner K, Wiener AS. An agglutinable factor in human blood recognized by immune sera for rhesus blood. *Proc Soc Exp Biol Med.* (1940) 43:223–24. doi: 10.3181/00379727-43-11151
42. Landsteiner K, Wiener AS. Studies on an agglutinin (Rh) in human blood reacting with anti-rhesus sera and with human isoantibodies. *J Exp Med.* (1941) 74:309–20. doi: 10.1084/jem.74.4.309
43. Lahaie YM, Watier H. Contribution of physiologists to the identification of the humoral component of immunity in the 19th century. *MAbs.* (2017) 9:774–80. doi: 10.1080/19420862.2017.1325051
44. Richet CR. *Nobel Prize in Physiology of Medicine 1913.* Available online at: <https://www.nobelprize.org/prizes/medicine/1913/summary/> (accessed March 20, 2019).
45. Ishizaka, K, Ishizaka T, Hornbrook MM. Physicochemical properties of reaginic antibody. V. Correlation of reaginic activity with gamma-E-globulin antibody. *J Immunol.* (1966) 97:840–53.
46. Arthus NM. *De Lanaphylaxie À L'immunité; Anaphylaxie, Protéotoxies, Evenimations, Anaphylaxie-Immunité, Sérums Antivenimeux.* Paris: Masson (1921).
47. von Pirquet C, Schick B. *Die Serumkrankheit.* Leipzig: Wien: Franz Deuticke (1905).
48. Hashimoto H. Zur Kenntnis Ddr Lymphomatösen Veränderung der Schilddrüse (Struma Lymphomatosa). In: Deutsche Gesellschaft für Chirurgie, editor. *Archiv für Klinische Chirurgie.* Berlin: Springer (1912). p. 219–48.
49. Landsteiner K, Chase MW. Experiments on transfer of cutaneous sensitivity to simple compounds. *Proc Soc Exp Biol Med.* (1942) 49:688–90. doi: 10.3181/00379727-49-13670
50. Coombs RRA, Gell PGH. *The Classification of Allergic Reactions Underlying Disease.* Davis, CA: Clinical Aspects of Immunology. (1963).
51. Cosimi AB. Nobel prizes in medicine in the field of transplantation. *Transplantation.* (2006) 82:1558–62. doi: 10.1097/01.tp.0000249567.11794.c7
52. Benacerraf B, Dausset J, Snell GD. *Nobel Prize in Physiology or Medicine 1980.* Available online at: <https://www.nobelprize.org/prizes/medicine/1980/summary/> (accessed March 20, 2019).
53. Burnet FM, Medawar PB. *Nobel Prize in Physiology or Medicine 1960.* Available online at: <https://www.nobelprize.org/prizes/medicine/1960/summary/> (accessed March 20, 2019).
54. Billingham RE, Brent L, Medawar PB. Actively acquired tolerance of foreign cells. *Nature.* (1953) 172:603–6. doi: 10.1038/172603a0
55. Burnet FM, Fenner F. *The Production of Antibodies.* 2nd ed. New York, NY: Macmillan. (1949).
56. Burnet FM. A Modification of Jerne's theory of antibody production using the concept of clonal selection. *Aust J Sci.* (1957) 20:67–9.
57. Pauling L. A theory of the structure and process of formation of antibodies. *J Am Chem Soc.* (1940) 62:2643–57. doi: 10.1021/ja01867a018
58. Jerne NK, Köhler GJF, Milstein C. *Nobel Prize in Physiology or Medicine 1984.* Available online at: <https://www.nobelprize.org/prizes/medicine/1984/summary/> (accessed March 20, 2019).
59. Jerne NK. The natural-selection theory of antibody formation. *Proc Natl Acad Sci USA.* (1955) 41:849–57. doi: 10.1073/pnas.41.11.849
60. Talmage DW. Allergy and immunology. *Annu Rev Med.* (1957) 8:239–56. doi: 10.1146/annurev.me.08.020157.001323
61. Nossal GJ. One cell-one antibody: prelude and aftermath. *Nat Immunol.* (2007) 8:1015–7. doi: 10.1038/ni1007-1015
62. Nossal GJ, Lederberg J. Antibody production by single cells. *Nature.* (1958) 181:1419–20. doi: 10.1038/1811419a0
63. Miller JFAP. The golden anniversary of the thymus. *Nat Rev Immunol.* (2011) 11:489–95. doi: 10.1038/nri2993



64. Cooper MD, Peterson RDA, Good RA. Delineation of the thymic and bursal lymphoid systems in the chicken. *Nature*. (1965) 205:143–6. doi: 10.1038/205143a0
65. Cooper MD, Raymond DA, Peterson RD, South MA, Good RA. The functions of the thymus system and the bursa system in the chicken. *J Exp Med*. (1966) 123:75–102. doi: 10.1084/jem.123.1.75
66. Gowans JL. The lymphocyte — a disgraceful gap in medical knowledge. *Immunol Today*. (1996) 17:288–91. doi: 10.1016/0167-5699(96)80547-0
67. Fagraeus A. Antibody-producing cells: a survey of four decades of research development - the tenth annual ernest witebsky memorial lecture, 22 April 1980. *Scand J Immunol*. (1981) 13:99–104. doi: 10.1111/j.1365-3083.1981.tb00115.x
68. Fagraeus A. Plasma cellular reaction and its relation to the formation of antibodies *in vitro*. *Nature*. (1947) 159:499. doi: 10.1038/159499a0
69. Cooper MD. The early history of B cells. *Nat Rev Immunol*. (2015) 15:191–7. doi: 10.1038/nri3801
70. Claman HN. On discovering thymus-marrow synergism. *Front Immunol*. (2014) 5:588. doi: 10.3389/fimmu.2014.00588
71. Mitchison NA. The discovery of T cell-B cell cooperation. *Front Immunol*. (2014) 5:377. doi: 10.3389/fimmu.2014.00377
72. Mackaness GB. The monocyte in cellular immunity. *Semin Hematol*. (1970) 7:172–84.
73. Zinkernagel RM, Doherty PC. Mhc-restricted cytotoxic T cells: studies on the biological role of polymorphic major transplantation antigens determining T-cell restriction-specificity, function, and responsiveness. *Adv Immunol*. (1979) 27:51–177. doi: 10.1016/S0065-2776(08)60262-X
74. Doherty PC, Zinkernagel RM. *Nobel Prize in Physiology or Medicine 1996*. Available online at: <https://www.nobelprize.org/prizes/medicine/1996/summary/> (accessed March 20, 2019).
75. Smith KA. Toward a molecular understanding of adaptive immunity: a chronology, Part I. *Front Immunol*. (2012) 3:369–9. doi: 10.3389/fimmu.2012.00369
76. Mosmann TR, Coffman RL. Two types of mouse helper T-cell clone implications for immune regulation. *Immunol Today*. (1987) 8:223–7. doi: 10.1016/0167-5699(87)90171-X
77. Sakaguchi S. Naturally arising Foxp3-expressing Cd25+ Cd4+ regulatory T cells in immunological tolerance to self and non-self. *Nat Immunol*. (2005) 6:345–52. doi: 10.1038/ni1178
78. Kappler JW, Marrack PC. Helper T cells recognise antigen and macrophage surface components simultaneously. *Nature*. (1976) 262:797–9. doi: 10.1038/262797a0
79. Reinherz EL, Meuer SC, Schlossman SF. The delineation of antigen receptors on human T lymphocytes. *Immunol Today*. (1983) 4:5–8. doi: 10.1016/0167-5699(83)90094-4
80. Davis MM. Molecular genetics of the T cell-receptor beta chain. *Annu Rev Immunol*. (1985) 3:537–60. doi: 10.1146/annurev.iy.03.040185.002541
81. Mak TW, Yanagi Y. Genes encoding the human T cell antigen receptor. *Immunol Rev*. (1984) 81:221–33. doi: 10.1111/j.1600-065X.1984.tb01112.x
82. Sakano H, Maki R, Kurosawa Y, Roeder W, Tonegawa S. Two types of somatic recombination are necessary for the generation of complete immunoglobulin heavy-chain genes. *Nature*. (1980) 286:676–83. doi: 10.1038/286676a0
83. Tonegawa S. Nobel lecture in physiology or medicine—1987. Somatic generation of immune diversity. *In vitro Cell Dev Biol*. (1988) 24:253–65. doi: 10.1007/BF02628825
84. Tonegawa S. *Nobel Prize in Physiology or Medicine 1987*. Available online at: <https://www.nobelprize.org/prizes/medicine/1987/summary/> (accessed March 20, 2019).
85. Janeway CA Jr. A trip through my life with an immunological theme. *Annu Rev Immunol*. (2002) 20:1–28. doi: 10.1146/annurev.immunol.20.080801.102422
86. Janeway CA Jr. Pillars article: approaching the asymptote? evolution and revolution in immunology. *Cold Spring Harb Symp Quant Biol*. (1989) 54:1–13. *J Immunol* (2013) 191:4475–87.
87. Beutler B. Microbe sensing, positive feedback loops, and the pathogenesis of inflammatory diseases. *Immunol Rev*. (2009) 227:248–63. doi: 10.1111/j.1600-065X.2008.00733.x
88. Hoffmann JA, Reichhart JM. Drosophila innate immunity: an evolutionary perspective. *Nat Immunol*. (2002) 3:121–6. doi: 10.1038/ni0202-121
89. Beutler BA, Hoffmann JA, Steinman RM. *Nobel Prize in Physiology or Medicine 2011*. Available online at: <https://www.nobelprize.org/prizes/medicine/2011/summary/> (accessed March 20, 2019).
90. Janeway CA Jr, Medzhitov R. Innate immune recognition. *Annu Rev Immunol*. (2002) 20:197–216. doi: 10.1146/annurev.immunol.20.083001.084359
91. Al-Yassin G. Medawar's "Actively Acquired Tolerance" and the danger model: setting the record straight. *Scand J Immunol*. (2018) 88:e12652. doi: 10.1111/sji.12652
92. Fuchs EJ, Matzinger P. Does the danger model shed any light on central tolerance?: a response to Al-Yassin. *Scand J Immunol*. (2018) 88:e12660. doi: 10.1111/sji.12660
93. Steinman RM, Hemmi H. Dendritic cells: translating innate to adaptive immunity. *Curr Top Microbiol Immunol*. (2006) 311:17–58. doi: 10.1007/3-540-32636-7\_2
94. Dinarello CA. Historical insights into cytokines. *Eur J Immunol*. (2007) 37:S34–45. doi: 10.1002/eji.200737772
95. Feldman M, Taylor P, Paleolog E, Brennan FM, Maini RN. Anti-Tnf alpha therapy is useful in rheumatoid arthritis and crohn's disease: analysis of the mechanism of action predicts utility in other diseases. *Transplant Proc*. (1998) 30:4126–7. doi: 10.1016/S0041-1345(98)01365-7
96. Travis S. Advances in therapeutic approaches to ulcerative colitis and crohn's disease. *Curr Gastroenterol Rep*. (2005) 7:475–84. doi: 10.1007/s11894-005-0079-9
97. King KM, Younes A. Rituximab: review and clinical applications focusing on non-hodgkin's lymphoma. *Exp Rev Anticancer Ther*. (2001) 1:177–86. doi: 10.1586/14737140.1.2.177
98. Chamoto K, Al-Habsi M, Honjo T. Role of Pd-1 in immunity and diseases. *Curr Top Microbiol Immunol*. (2017) 410:75–97. doi: 10.1007/82\_2017\_67
99. Wei SC, Duffy CR, Allison JP. Fundamental mechanisms of immune checkpoint blockade therapy. *Cancer Discov*. (2018) 8:1069–86. doi: 10.1158/2159-8290.CD-18-0367

**Conflict of Interest Statement:** The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Kaufmann. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.



# Commentary: Immunology's Coming of Age

Heinz Kohler<sup>1\*</sup>, Anastas Dimitrov Pashov<sup>2</sup> and Thomas Kieber-Emmons<sup>3</sup>

<sup>1</sup> Department of Microbiology and Immunology, University of Kentucky, Lexington, KY, United States, <sup>2</sup> Stephan Angelov Institute of Microbiology, Bulgarian Academy of Sciences, Sofia, Bulgaria, <sup>3</sup> Pathology, University of Arkansas for Medical Sciences, Little Rock, AR, United States

**Keywords:** antibody, idiotype, selection, natural antibody, therapeutic antibody

## A Commentary on

### Immunology's Coming of Age

by Kaufmann, S. H. E. (2019). *Front. Immunol.* 10:684. doi: 10.3389/fimmu.2019.00684

The recent review by Stefan Kaufmann on “Immunology's Coming of Age” is an elegant historical outline of the evolution of Immunology with focusing on a particular perspective of the history of Immunology, that is Nobel Laureate contributions to the discipline. Immunology is a difficult discipline to survey. Even the best attempts would ultimately focus on some selected aspects. As such, it invites comments aiming to complement the presented history in the context of Immunology coming of age. It is the aim of our Commentary to add important research in the field of immunology to demonstrate that it has become a self-containing discipline.

## OPEN ACCESS

### Edited by:

Alessandra Mortellaro,  
San Raffaele Telethon Institute for  
Gene Therapy (SR-Tiget), Italy

### Reviewed by:

Luuk Hilbrands,  
Radboud University Nijmegen,  
Netherlands

### \*Correspondence:

Heinz Kohler  
heinz.kohler@uky.edu

### Specialty section:

This article was submitted to  
Molecular Innate Immunity,  
a section of the journal  
*Frontiers in Immunology*

**Received:** 26 June 2019

**Accepted:** 28 August 2019

**Published:** 12 September 2019

### Citation:

Kohler H, Pashov AD and  
Kieber-Emmons T (2019)  
Commentary: Immunology's Coming  
of Age. *Front. Immunol.* 10:2175.  
doi: 10.3389/fimmu.2019.02175

## INTRODUCTION

Immunology is a rich discipline with successes and failures, with various scholarly works describing its origins and history that lend to our current understanding of immunological principles (1, 2). Still another perspective has been presented recently by Stefan Kaufmann emphasizing notable contributions acknowledged by the awarding of a Nobel Prize to outstanding investigators (3). While touching on extremely important developments, important contextual elements need to be mentioned to complement the presented history as important contributions are not always recognized by a Nobel Prize.

## DISCUSSION

### Antibody Recognition and Diversity

Saying that Immunology is an interdisciplinary science may no longer be entirely true since now it has also its own methods. The most prominent immunological paradigm is the concept of antibody. The specificity of antibodies is still an important question in immunology. Historically, the generation of diversity of antibodies was a hot discussed topic in the middle of the twentieth century initiated by the template hypothesis of Breinl and Haurowitz in 1930 (4), 10 years prior to Pauling's claim, cited in Kaufmann's review, that antibodies were made by folding newly synthesized nascent antibody polypeptide chains around the antigens, which serve as a template. Breinl and Haurowitz “thought that antibodies acquired their specificity for antigen by folding of the newly synthesized nascent polypeptide chain around the antigen” (5). The biochemical properties of antigen-antibody binding interactions were examined in more detail in the late 1930s by John Marrack (6). The biomolecule responsible for these actions was termed antitoxin, precipitin, and agglutinin. It was not known that all three substances were one entity. This was later demonstrated by Elvin A. Kabat showing the heterogeneity of antibodies through ultracentrifugation studies of

horses' sera. Similarly, an equally important milestone in the understanding of Immunological recognition was the x-ray resolution of a Fab antibody fragment (7) not recognized in the review and the founding of the definition of antibody diversity and its biological significance by Kabat (8, 9). This work provided a transforming view of antibody diversity and the molecular basis for antigen recognition (10).

## Idiotypic Hypothesis

Niels Jerne made several important contributions to Immunology. Niels Jerne's antibody selection theory is cited, but his more important contribution in the field of Immunobiology, the Idiotypic Network hypotheses, is not mentioned being essential for a historical record (11). He suggested that antibodies could be recognized as foreign, inducing other antibodies and thereby forming a network. Neglecting idiotype may be seen as more of a cultural aspect since it has not been accepted as a mainstream theory. Nevertheless, it has left a considerable imprint in immunological thinking. Recent reviews in *Frontiers* address the importance of the Idiotypic concept in Immunology (12, 13). It might be argued that the Idiotypic Network hypothesis is the forerunner of present day ideas on the role antibodies plays in integrative Systems Immunology (14).

## Selection

Positive and negative selection (of both T and B cells) as well as the practical and theoretical aspects of intravenous immunoglobulins are important Immunology discoveries. The term "tolerance" was first coined by Ray Owen in reference to a physiological state he observed in dizygotic twin cattle (15) as noted in a review of the historical record of immunological tolerance (16). Just like antibodies, the elucidation of the T cell structure was monumental (17, 18). This facet provided the backdrop of monumental studies by Ellis Reinherz, Philippa Marrack, John Kappler, and James Allison. Checkpoint inhibitors, which are driving Immunotherapy,

owe their existence to the understanding of how T cells in particular function.

## Natural Antibodies

Of no less importance is the regulatory and therapeutic potential of natural antibodies (19). Natural antibodies play an important role in the first line of defense and house keeping (20, 21). For a long period, natural antibodies were merely regarded as insignificant background of immunity. However, an early study in 1925 indicated that natural antibody in normal serum could neutralize bacteria (22).

## Therapeutic Antibodies

With the discovery of immortalizing antibodies by Kohler and Milstein (23) opened a new drug class to treat infections, auto-immunities and other diseases (24, 25). In parallel intravenous immunoglobulin (IVIg) emerged as standard therapy of immunoglobulin deficiencies, auto-immune reactions and in homeostasis (26–28). These translational aspects of Immunology deserve to be noticed.

## CONCLUSION

The History of Immunology began with Edward Jenner's discovery that vaccination protects against smallpox. Many scientists and discoveries have since lent to our understanding of how the immune system fights disease and sometimes causes disease as well to new classes of drugs. As we move closer to individualized medicine scenarios there will be a continuing need to understand and maybe redefine what came before and what will evolve in the discipline Immunology.

## AUTHOR CONTRIBUTIONS

All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

## REFERENCES

1. Silverstein AM. History of Immunology. *eLS* (2001). doi: 10.1038/npg.els.0003078
2. Smith KA. Editorial: a living history of immunology. *Front Immunol.* (2015) 6:502. doi: 10.3389/fimmu.2015.00502
3. Kaufmann SHE. Immunology's coming of age. *Front Immunol.* (2019) 10:684. doi: 10.3389/fimmu.2019.00684
4. Breinl F, Haurowitz F. Chemische Untersuchung des Präzipitates aus Hamoglobin und Anti-Hamoglobin-Serum und Bemerkungen über die Natur der Antikörper. *F Hoppe-Seyler Z. Physiol Chem.* (1930) 192:45. doi: 10.1515/bchm2.1930.192.1-3.45
5. Eisen HN, Schlesinger S. Remembrance of immunology past: conversations with Herman Eisen. *Annu Rev Immunol.* (2015) 33:1–28. doi: 10.1146/annurev-immunol-111214-122349
6. Marrack J. Nature of antibodies. *Nature.* (1934) 133:292–3. doi: 10.1038/133292b0
7. Rudikoff S, Potter M, Segal DM, Padlan EA, Davies DR. Crystals of phosphorylcholine-binding Fab-fragments from mouse myeloma proteins: preparation and x-ray analysis. *Proc Natl Acad Sci USA.* (1972) 69:3689–92. doi: 10.1073/pnas.69.12.3689
8. Kabat EA. Heterogeneity and structure of antibody-combining sites. *Ann N Y Acad Sci.* (1970) 169:43–54. doi: 10.1111/j.1749-6632.1970.tb55968.x
9. Wu TT, Kabat EA. An analysis of the sequences of the variable regions of Bence Jones proteins and myeloma light chains and their implications for antibody complementarity. *J Exp Med.* (1970) 132:211–50. doi: 10.1084/jem.132.2.211
10. Hood LE. Wu and Kabat 1970: a transforming view of antibody diversity. *J Immunol.* (2008) 180:7055–6. doi: 10.4049/jimmunol.180.11.7055
11. Jerne NK. Towards a network theory of the immune system. *Ann Immunol.* (1974) 125C:373–89.
12. Kieber-Emmons T, Monzavi-Karbassi B, Pashov A, Saha S, Murali R, Kohler H. The promise of the anti-idiotypic concept. *Front Oncol.* (2012) 2. doi: 10.3389/fonc.2012.00196
13. Lemke H. Antigen receptor-intrinsic non-self: the key to understanding regulatory lymphocyte-mediated idiotypic control of adaptive immune responses. *Crit Rev Immunol.* (2016) 36:13–56. doi: 10.1615/CritRevImmunol.2016016606
14. Li S, Roupheal N, Duraisingham S, Romero-Steiner S, Presnell S, Davis C, et al. Molecular signatures of antibody responses derived from a systems biology study of five human vaccines. *Nat Immunol.* (2014) 15:195–204. doi: 10.1038/ni.2789

15. Owen RD. Immunogenetic consequences of vascular anastomoses between bovine twins. *Science*. (1945) 102:400–1. doi: 10.1126/science.102.2651.400
16. Schwartz RH. Historical overview of immunological tolerance. *Cold Spring Harb Perspect Biol*. (2012) 4:a006908. doi: 10.1101/cshperspect.a006908
17. Yanagi Y, Yoshikai Y, Leggett K, Clark SP, Aleksander I, Mak TW. A human T cell-specific cDNA clone encodes a protein having extensive homology to immunoglobulin chains. *Nature*. (1984) 308:145–9. doi: 10.1038/308145a0
18. Hedrick SM, Cohen DI, Nielsen EA, Davis MM. Isolation of cDNA clones encoding T cell-specific membrane-associated proteins. *Nature*. (1984) 308:149–53. doi: 10.1038/308149a0
19. Briles DE, Nahm M, Schroer K, Davie J, Baker P, Kearney J, Barletta R. Antiphosphocholine antibodies found in normal mouse serum are protective against intravenous infection with type 3 streptococcus pneumoniae. *J Exp Med*. (1981) 153:694–705. doi: 10.1084/jem.153.3.694
20. Silverman GJ. Protective natural autoantibodies to apoptotic cells: evidence of convergent selection of recurrent innate-like clones. *Ann N Y Acad Sci*. (2015) 1362:164–75. doi: 10.1111/nyas.12788
21. Maddur MS, Lacroix-Desmazes S, Dimitrov JD, Kazatchkine MD, Bayry J, Kaveri SV. Natural antibodies: from first-line defense against pathogens to perpetual immune homeostasis. *Clin Rev Allergy Immunol*. (2019) 2019:8746–9. doi: 10.1007/s12016-019-08746-9
22. Mackie TJ. Non-specific stimulation of a natural antibody. *J Hyg (Lond)*. (1925) 24:176–88. doi: 10.1017/S0022172400008676
23. Kohler G, Milstein C. Continuous cultures of fused cells secreting antibody of predefined specificity. *Nature*. (1975) 256:495–7. doi: 10.1038/256495a0
24. Steplewski Z, Thurin M, Kieber-Emmons T. Antibodies: at the nexus of antigens and cancer vaccines. *J Infect Dis*. (2015) 212:S59–66. doi: 10.1093/infdis/jiu638
25. Shepard HM, Phillips GL, Feldmann M. Developments in therapy with monoclonal antibodies and related proteins. *Clin Med (Lond)*. (2017) 17:220–32. doi: 10.7861/clinmedicine.17-3-220
26. Kaveri SV, Lacroix-Desmazes S, Mouthon L, Kazatchkine MD. Human natural autoantibodies: lessons from physiology and prospects for therapy. *Immunologist* (1998) 6:227–33.
27. Kazatchkine MD, Kaveri SV. Immunomodulation of autoimmune and inflammatory diseases with intravenous immune globulin. *N Engl J Med*. (2001) 345:747–55. doi: 10.1056/NEJMr993360
28. Vas J, Grönwall C, Silverman G. Fundamental roles of the innate-like repertoire of natural antibodies in immune homeostasis. *Front Immunol*. (2013) 4:4. doi: 10.3389/fimmu.2013.00004

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Kohler, Pashov and Kieber-Emmons. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.





# Vaccine Evolution and Its Application to Fight Modern Threats

Emanuele Andreano<sup>1,2,3</sup>, Ugo D'Oro<sup>2</sup>, Rino Rappuoli<sup>2,3,4</sup> and Oretta Finco<sup>2\*</sup>

<sup>1</sup> Department of Life Sciences, University of Siena, Siena, Italy, <sup>2</sup> GlaxoSmithKline, Siena, Italy, <sup>3</sup> vAMRes Lab, Toscana Life Sciences, Siena, Italy, <sup>4</sup> Faculty of Medicine, Imperial College, London, United Kingdom

Before the development of the first vaccine, infectious diseases were a major cause of death around the globe with life expectancy estimated to be <50 years. Three measures have helped to drastically reduce the burden of infectious diseases but only vaccines have proven to be able to eradicate infectious agents. Herein, we describe new methodologies that have paved the way for what is currently known as modern vaccinology and the use of vaccines to tackle antimicrobial resistance, the biggest global threat of our time.

**Keywords:** vaccines, infectious diseases, antimicrobial resistance (AMR), vaccine development, vaccinology

## OPEN ACCESS

### Edited by:

Ennio Carbone,  
Università degli Studi Magna Graecia  
di Catanzaro, Italy

### Reviewed by:

Sven Hammerschmidt,  
University of Greifswald, Germany  
Paola Italiani,  
Italian National Research Council  
(CNR), Italy

### \*Correspondence:

Oretta Finco  
oretta.x.finco@gsk.com

### Specialty section:

This article was submitted to  
Molecular Innate Immunity,  
a section of the journal  
Frontiers in Immunology

**Received:** 08 May 2019

**Accepted:** 09 July 2019

**Published:** 25 July 2019

### Citation:

Andreano E, D'Oro U, Rappuoli R and  
Finco O (2019) Vaccine Evolution and  
Its Application to Fight Modern  
Threats. *Front. Immunol.* 10:1722.  
doi: 10.3389/fimmu.2019.01722

## THE BURDEN OF INFECTIOUS DISEASES BEFORE ANTIBIOTICS AND VACCINE INTERVENTION

Infectious diseases have always had a devastating impact on humankind. Some of the most catastrophic pandemics of our history include the Justinian plague (542-546 AD), which had a tragic toll of 100 million deaths, the bubonic plague (1347-50 AD), also known as the “Black Death,” which erased one-third of the entire human population (1, 2), and more recently the “Spanish” influenza in 1918 which caused ~50–100 million deaths worldwide reducing the European population by half (3–5). Before the introduction of effective preventive and therapeutic strategies, life expectancy was estimated to be <50 years and bacterial infections were the imperative toll setting this limit (6). This scenario changed with the introduction of three measures that helped to dramatically reduce the death burden caused by infectious diseases. The measures include hygiene, antibiotics, and vaccination (7, 8). The introduction of penicillin in 1929 (9), and its first use in humans a decade later (10), led to a dramatic reduction of mortality caused by infectious diseases. Unfortunately, in 1940 the first case of a penicillin resistant *E. coli* strain was documented and by the late 1960s over 80% of *S. aureus* strains acquired the same resistance (10–12). Therefore, despite the use of antibiotics resulted to be an outstanding first line of defense to treat infections, pathogens have shown to quickly acquire resistance phenotypes after only few years from their introduction (13). Vaccines, on the other hand, have only rarely shown to induce resistant phenotypes as they usually aim to elicit a multi-targets immune response and their prophylactic use reduces the likelihood of spreading resistant-conferring mutations (14). Indeed the smallpox vaccine introduced in 1796, and subsequently manufactured from infected calf skin (15), has led to the eradication of this infectious agent in 1988 (16, 17). Therefore, despite the fact that antibiotics and vaccines are pivotal interventions against infectious diseases, vaccination has been the sole intervention capable of eradicating an infectious agent and, given its potential, it can also be considered as the most appropriate solution against future global threats represented by infectious diseases (18–20).

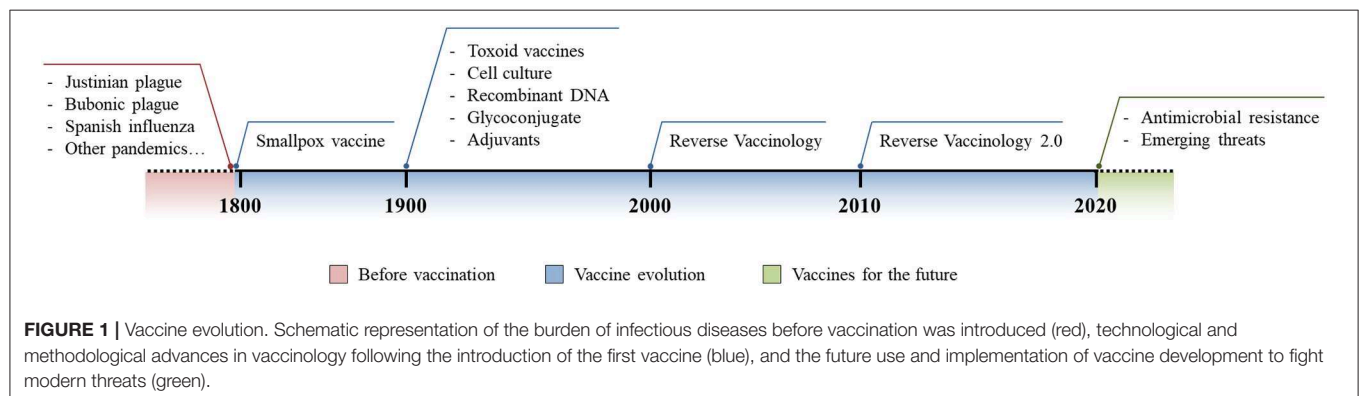
## REVERSE VACCINOLOGY AND THE DEVELOPMENT OF MODERN VACCINES

Since Edward Jenner first vaccinated an 8 year old boy in 1796 by inoculating fresh cowpox lesion matter (21), enormous leaps forward have been made in the field of vaccine development. Empirical approaches like attenuation and inactivation of microorganisms were the first steps forward to modern vaccinology (22). Recently, new technologies such as glycoconjugates and the introduction of novel vaccine adjuvants changed the field of vaccines, however the biggest change came with the first sequencing of the *Haemophilus influenzae* whole genome in 1995, a discovery that allowed the birth of “Reverse Vaccinology,” a genome-based approach to vaccine development (23, 24). This approach, following the sequencing and analysis of the *Neisseria meningitidis* serogroup B strain whole genome, allowed the identification of novel candidates and the development of a four-component meningococcus B vaccine (4CMenB) (25, 26). This recently licensed vaccine has already shown incredible effectiveness in the UK with 82.9% protection against all MenB strains in infants (27). The evolution of vaccine development further moved forward with the advancement of new methodologies and technological breakthroughs. Indeed, in 2016 the “reverse vaccinology 2.0” entered the stage. With this approach, the human immune system is analyzed at a single cell level allowing the characterization of the antibody response like never before (28). The gain of knowledge acquired by this approach allows to rapidly identify highly immunogenic antigens to develop novel and more efficacious vaccine candidates. The RSV fusion protein (F) case is a major example of the phenomenal power of the reverse vaccinology 2.0. Indeed, human B cells were directly isolated from RSV convalescent donors and cultured to naturally produce human monoclonal antibodies (humAbs). Among all the antibodies screened for RSV neutralization *in vitro*, the humAbs named D25 resulted in the most potent antibody with a median half-maximal inhibitory concentration (IC<sub>50</sub>) of 2.1 ng/ml (100–150 times more than palivizumab, the only monoclonal antibody approved by the FDA for RSV prevention in infants) (29). Interestingly, D25 was not capable of binding to the RSV F-protein in its post-fusion conformation, the only vaccine candidate available at the time against RSV (30). Then, McLellan and coworkers had the brilliant

intuition to test D25 complex with the RSV F-protein to perform structural studies. This experiment was paramount in solving the crystal structure of RSV F-protein in its pre-fusion conformation (preF) which in turn led to the design of a stabilized RSV preF molecule (30, 31). Following the production of a soluble preF reagent, numerous human neutralizing antibodies have been identified allowing a deep characterization of the antigen surface and the identification of two preF-specific antigenic sites that have shown incredible high neutralization potency (32). The effectiveness of the preF antigen has already been proven in different animal models (mice, rhesus macaques, and calves) further supporting the potential of RSV preF as an ideal vaccine candidate against this pneumovirus (10, 13). The power of reverse vaccinology 2.0 has allowed, in <5 years since preF stabilization, to start clinical trials that are currently on-going to develop the first vaccine against RSV (7). This approach, which has found broad applicability to fight viral infections, could also be considered as a key stratagem to tackle bacterial infections.

## USE OF PEPTIDE-ANTIGEN DERIVED FOR GERMLINE TARGETING VACCINOLOGY

The production of germline-targeting (GT) antigens for vaccine development is another pivotal example that underlies the outstanding potential of reverse vaccinology 2.0. Indeed, the combined knowledge acquired by the identification and characterization of novel antigens plus the functional/genetic analysis of human monoclonal antibodies naturally produced by infected or vaccinated human donors, can be used to design antigen-derived peptides, capable of tailoring the antibody immune response. In case of highly variable pathogens such as HIV, the use of the whole antigen can result in a strain specific response, while the development of GT-antigens can lead to the elicitation of broadly neutralizing antibodies (bnAbs) capable of clearing multiple infective strains. This is a two-step approach which, using different rationally designed immunogens, aims to: (1) prime the germline precursor B cell of antibodies previously shown to possess broadly neutralizing activity; (2) shepherding the bnAb population by driving their maturation affinity toward the highly immunogenic epitope of interest. GT-vaccinology has



been used to elicit a specific class of HIV-1 gp120 CD4-binding site specific bnAbs known as VRC01, through the use of engineered outer domain germline-targeting (eOD-GT) peptides (33). The interest to prime VRC01-bnAbs arises from their ability to mimic the CD4-binding to the gp120 receptor binding site and their capability to potentially neutralize (median IC<sub>50</sub> 40 ng/mL) up to 98% of a large panel of global HIV-1 isolates (34, 35). An in-depth analysis of the VRC01 genetic features has shown peculiarities in this class of bnAbs. They classically derive from an extensively mutated (32–48%) VH1-2\*02 heavy chain germline which pairs with light chains, presenting a rare five amino acid long CDR3 motif (usually QQYEF) (36). These analyses were paramount for the development of novel and potentially therapeutic candidates to fight HIV infections. Examples of the use of VRC01-bnAbs as a therapeutic tool are the monoclonal antibodies named VRC-HIVMAB060-00-AB (VRC01) and a FC-modified version of this latter named VRC01LS. These two bnAbs are currently under clinical investigation (NCT02568215, NCT02716675, and NCT02599896) evaluating safety and efficacy in reducing acquisition of HIV-1 infection (37–40). In addition to monoclonal antibody development and application, the knowledge acquired from these studies and the ability to selectively expand this class of bnAbs upon immunization (41), have allowed the development of specific peptides as vaccine candidates capable of shepherding the immune system toward a VRC01-like antibody response. The most promising candidate is the tailored immunogen named eOD-GT8 60-subunit self-assembling nanoparticle (eOD-GT8 60mer) (36, 42) which has shown superior affinity and breadth of binding to germline-reverted VRC01-like bnAbs (41).

The HIV case described above further confirms the outstanding power of reverse vaccinology 2.0. Indeed, in only 3 years since its design and stabilization (43), the eOD-GT8 60mer antigen is under investigation in a phase I clinical trial in healthy adults aimed at assessing safety, tolerability and immunogenicity of this germline-targeting immunogen (NCT03547245).

## VACCINES FOR THE FUTURE: THE FIGHT AGAINST ANTIMICROBIAL RESISTANCE

Despite antibiotics being the only lifesaving tool in fighting acute bacterial infection, as Stanley Falkow said (3), they are creating some problems of their own. In fact, the improper and excessive use of antibiotics has pressured bacteria to acquire antibiotic resistant phenotypes and this problem is currently growing out of control. Bacteria have shown several mechanisms to acquire antibiotic resistance and examples include the expression of  $\beta$ -lactamases, efflux pumps, modification of the cellular surface, and gene mutations to alter those molecules that are targeted by antibiotics (4). This phenomenon, known as antimicrobial resistance (AMR), is arguably one of the biggest threats that our world is facing today. Indeed, up to 700,000 deaths each year are AMR-related and these have been estimated to increase up to 10 million by 2050, exceeding the 8.2 million deaths per year caused by cancer today (8, 44). A solution to this alarming threat would be the prevention of antibiotic resistant bacteria

infections through vaccination, a strategy that has already proven its great value to humanity (6). Several reasons suggest that vaccines would be a promising solution against AMR. First, antibiotics have shown to rapidly become obsolete and resistance emerges soon after their introduction, while vaccines allow long-lasting protection against infections and resistance has only rarely evolved after vaccination (13). Second, while antibiotics only hit a few metabolic target vaccines, based on the selected strategy, they can elicit a broad multi-target immune response reducing the probability of the evolution of resistant mutations. Furthermore, although major investments have been made to enrich antibiotic R&D pipelines, the discovery of innovative antimicrobial targets are running dry since the 1970s. Therefore, given the incredibly high pace with which pathogens are capable of developing resistance to new classes of antibiotics, focusing our attention exclusively on antibiotic R&D will not be sufficient (13, 45). In a marked contrast, thanks to incredible technological advancements of the last few decades, vaccine R&D pipelines are promising for the development of innovative and highly effective vaccines which can have an important contribution in controlling AMR (13, 18). Finally, antibiotics can only be used to treat individuals already infected, while successful vaccination campaigns can prevent the occurrence of infection, reducing the spread of the infectious agent and protecting the whole population through herd immunity (8, 20, 46). Vaccine evolution has allowed us to address several unmet medical needs and, given all of the reasons stated above, it should be considered a key solution in fighting emerging threats such as AMR (**Figure 1**).

## CONCLUSIONS

Since their introduction, vaccines have helped save billions of lives all over the world. Empirical approaches were not sufficient to support the development of vaccines against pathogens for which no preventive strategies or treatments were available. Methodological and technological advancements have introduced the world to modern vaccinology approaches which have unlocked the possibility to develop novel vaccines against virtually any pathogen. The RSV and HIV case studies reported herein, are clear examples of how innovative technologies and their corollary applications have paved the way for new experimental approaches capable of tackling and possibly addressing these unmet global medical needs. Vaccines have provided the basis for a global and sustainable public health in the past and they can potentially continue to do so by addressing major and upcoming global threats like AMR.

## AUTHOR CONTRIBUTIONS

All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

## FUNDING

This work has received funding under the European Research Council (ERC) advanced grant agreement number 787552 (vAMRes).

## REFERENCES

- WHO. Chapter 3. WHO Report on Global Surveillance of Epidemic-Prone Infectious Diseases. (2000), 26p.
- Schmid BV, Büntgen U, Easterday WR, Ginzler C, Walløe L, Bramanti B, et al. Climate-driven introduction of the Black Death and successive plague reintroductions into Europe. *Proc Natl Acad Sci USA*. (2015) 112:3020–5. doi: 10.1073/pnas.1412887112
- Taubenberger JK, Morens DM. 1918 influenza: the mother of all pandemics. *Emerg Infect Dis*. (2006) 12:15–22. doi: 10.3201/eid1209.05-0979
- Johnson NP, Mueller J. Updating the accounts: global mortality of the 1918–1920 “Spanish” influenza pandemic. *Bull Hist Med*. (2002) 76:105–15. doi: 10.1353/bhm.2002.0022
- Morens DM, Taubenberger JK, Harvey HA, Memoli MJ. The 1918 influenza pandemic: lessons for 2009 and the future. *Crit Care Med*. (2010) 38(4 Suppl.):e10–e20. doi: 10.1097/CCM.0b013e3181ceb25b
- Rappuoli R, Pizza M, Del Giudice G, De Gregorio E. Vaccines, new opportunities for a new society. *Proc Natl Acad Sci USA*. (2014) 111:12288–93. doi: 10.1073/pnas.1402981111
- Rappuoli R. Twenty-first century vaccines. *Philos Trans R Soc B Biol Sci*. (2011) 366:2756–8. doi: 10.1098/rstb.2011.0075
- Tagliabue A, Rappuoli R. Changing priorities in vaccinology: antibiotic resistance moving to the top. *Front Immunol*. (2018) 9:1068. doi: 10.3389/fimmu.2018.01068
- Fleming A. On the antibacterial action of cultures of a penicillium, with special reference to their use in the isolation of *B. influenzae*. *Br J Exp Pathol*. (1929) 10:226–36.
- Abraham EP, Chain E. An enzyme from bacteria able to destroy penicillin. 1940. *Rev Infect Dis*. (1988) 10:677–8.
- Lobanovska M, Pilla G. Penicillin's discovery and antibiotic resistance: lessons for the future? *Yale J Biol Med*. (2017) 90:135–45.
- Lowy FD. Antimicrobial resistance: the example of *Staphylococcus aureus*. *J Clin Invest*. (2003) 111:1265–73. doi: 10.1172/JCI18535
- Kennedy DA, Read AF. Why does drug resistance readily evolve but vaccine resistance does not? *Proc Biol Sci*. (2017) 284:20162562. doi: 10.1098/rspb.2016.2562
- Bloom DE, Black S, Salisbury D, Rappuoli R. Antimicrobial resistance and the role of vaccines. *Proc Natl Acad Sci USA*. (2018) 115:12868–71. doi: 10.1073/pnas.1717157115
- Smith KA. Edward Jenner and the small pox vaccine. *Front Immunol*. (2011) 2:21. doi: 10.3389/fimmu.2011.00021
- Fenner F, Henderson DA, Arita I, Jezek Z, Ladnyi ID. *Smallpox and Its Eradication*. Geneva: WHO (1988).
- Bhattacharya S. The World Health Organization and global smallpox eradication. *J Epidemiol Commun Health*. (2008) 62:909–12. doi: 10.1136/jech.2006.055590
- Mishra RP, Oviedo-Orta E, Prachi P, Rappuoli R, Bagnoli F. Vaccines and antibiotic resistance. *Curr Opin Microbiol*. (2012) 15:596–602. doi: 10.1016/j.mib.2012.08.002
- Jansen KU, Knirsch C, Anderson AS. The role of vaccines in preventing bacterial antimicrobial resistance. *Nat Med*. (2018) 24:10–9. doi: 10.1038/nm.4465
- Lipsitch M, Siber GR. How can vaccines contribute to solving the antimicrobial resistance problem? *mBio*. (2016) 7:e00428–16. doi: 10.1128/mBio.00428-16
- Riedel S. Edward Jenner and the history of smallpox and vaccination. *Proc (Bayl Univ Med Cent)*. (2005) 18:21–5. doi: 10.1080/08998280.2005.11928028
- Plotkin S. History of vaccination. *Proc Natl Acad Sci USA*. (2014) 111:12283–7. doi: 10.1073/pnas.1400472111
- Fleischmann RD, Adams MD, White O, Clayton RA, Kirkness EF, Kerlavage AR, et al. Whole-genome random sequencing and assembly of *Haemophilus influenzae* Rd. *Science*. (1995) 269:496–512. doi: 10.1126/science.7542800
- Rappuoli R. Reverse vaccinology. *Curr Opin Microbiol*. (2000) 3:445–50. doi: 10.1016/S1369-5274(00)00119-3
- Vernikos G, Medini D. Bexsero® chronicle. *Pathog Global Health*. (2014) 108:305–16. doi: 10.1179/204773214Y.0000000162
- Pizza M, Scarlato V, Masignani V, Giuliani MM, Arico B, Comanducci M, et al. Identification of vaccine candidates against serogroup B meningococcus by whole-genome sequencing. *Science*. (2000) 287:1816–20. doi: 10.1126/science.287.5459.1816
- Parikh SR, Andrews NJ, Beebejaun K, Campbell H, Ribeiro S, Ward C, et al. Effectiveness and impact of a reduced infant schedule of 4CMenB vaccine against group B meningococcal disease in England: a national observational cohort study. *Lancet*. (2016) 388:2775–82. doi: 10.1016/S0140-6736(16)31921-3
- Rappuoli R, Bottomley MJ, D'Oro U, Finco O, De Gregorio E. Reverse vaccinology 2.0: human immunology instructs vaccine antigen design. *J Exp Med*. (2016) 213:469–81. doi: 10.1084/jem.20151960
- Kwakkenbos MJ, Diehl SA, Yasuda E, Bakker AQ, van Geelen CM, Lukens MV, et al. Generation of stable monoclonal antibody-producing B cell receptor-positive human memory B cells by genetic programming. *Nat Med*. (2010) 16:123–8. doi: 10.1038/nm.2071
- McLellan JS, Chen M, Leung S, Graepel KW, Du X, Yang Y, et al. Structure of RSV fusion glycoprotein trimer bound to a prefusion-specific neutralizing antibody. *Science*. (2013) 340:1113–7. doi: 10.1126/science.1234914
- McLellan JS, Chen M, Joyce MG, Sastry M, Stewart-Jones GB, Yang Y, et al. Structure-based design of a fusion glycoprotein vaccine for respiratory syncytial virus. *Science*. (2013) 342:592–8. doi: 10.1126/science.1243283
- Rossey I, McLellan JS, Saelens X, Schepens B. Clinical potential of prefusion RSV F-specific antibodies. *Trends Microbiol*. (2018) 26:209–19. doi: 10.1016/j.tim.2017.09.009
- Jardine J, Julien JP, Menis S, Ota T, Kalyuzhnyi O, McGuire A, et al. Rational HIV immunogen design to target specific germline B cell receptors. *Science*. (2013) 340:711–6. doi: 10.1126/science.1234150
- Huang J, Kang BH, Ishida E, Zhou T, Griesman T, Sheng Z, et al. Identification of a CD4-binding-site antibody to HIV that evolved near-pan neutralization breadth. *Immunity*. (2016) 45:1108–21. doi: 10.1016/j.immuni.2016.10.027
- Sok D, Burton DR. HIV Broadly neutralizing antibodies: taking good care of the 98. *Immunity*. (2016) 45:958–60. doi: 10.1016/j.immuni.2016.10.033
- Jardine JG, Kulp DW, Havenar-Daughton C, Sarkar A, Briney B, Sok D, et al. HIV-1 broadly neutralizing antibody precursor B cells revealed by germline-targeting immunogen. *Science*. (2016) 351:1458–63. doi: 10.1126/science.aad9195
- Cohen YZ, Caskey M. Broadly neutralizing antibodies for treatment and prevention of HIV-1 infection. *Curr Opin HIV AIDS*. (2018) 13:366–73. doi: 10.1097/COH.0000000000000475
- Mayer KH, Seaton KE, Huang Y, Grunenberg N, Isaacs A, Allen M, et al. Safety, pharmacokinetics, and immunological activities of multiple intravenous or subcutaneous doses of an anti-HIV monoclonal antibody, VRC01, administered to HIV-uninfected adults: results of a phase 1 randomized trial. *PLoS Med*. (2017) 14:e1002435. doi: 10.1371/journal.pmed.1002435
- Huang Y, Zhang L, Ledgerwood J, Grunenberg N, Bailer R, Isaacs A, et al. Population pharmacokinetics analysis of VRC01, an HIV-1 broadly neutralizing monoclonal antibody, in healthy adults. *mAbs*. (2017) 9:792–800. doi: 10.1080/19420862.2017.1311435
- Gaudinski MR, Coates EE, Houser KV, Chen GL, Yamshchikov G, Saunders JG, et al. Safety and pharmacokinetics of the Fc-modified HIV-1 human monoclonal antibody VRC01LS: a phase 1 open-label clinical trial in healthy adults. *PLoS Med*. (2018) 15:e1002493. doi: 10.1371/journal.pmed.1002493
- Abbott RK, Lee JH, Menis S, Skog P, Rossi M, Ota T, et al. Precursor frequency and affinity determine B cell competitive fitness in germinal centers, tested with germline-targeting HIV vaccine immunogens. *Immunity*. (2018) 48:133–46.e6. doi: 10.1016/j.immuni.2017.11.023
- Briney B, Sok D, Jardine JG, Kulp DW, Skog P, Menis S, et al. Tailored immunogens direct affinity maturation toward HIV neutralizing antibodies. *Cell*. (2016) 166:1459–70.e11. doi: 10.1016/j.cell.2016.08.005
- Jardine JG, Ota T, Sok D, Pauthner M, Kulp DW, Kalyuzhnyi O, et al. HIV-1 VACCINES. Priming a broadly neutralizing antibody response to HIV-1 using a germline-targeting immunogen. *Science*. (2015) 349:156–61. doi: 10.1126/science.aac5894



44. The Review on Antimicrobial Resistance. *Tackling Drug-Resistant Infections Globally: Final Report and Recommendations*. (2016). Available online at [https://amr-review.org/sites/default/files/160518\\_Final%20paper\\_with%20cover.pdf](https://amr-review.org/sites/default/files/160518_Final%20paper_with%20cover.pdf) (accessed May 7, 2018).
45. O'Neil J. *Tackling Drug-Resistant Infections Globally: Final Report and Recommendations*. (2016). Wellcome Trust and UK Government.
46. Mallory ML, Lindesmith LC, Baric RS. Vaccination-induced herd immunity: successes and challenges. *J Allergy Clin Immunol*. (2018) 142:64–6. doi: 10.1016/j.jaci.2018.05.007

**Conflict of Interest Statement:** RR, OF, and UD'O are full-time employees of GSK group of companies. EA participated in a postgraduate program at GSK.

Copyright © 2019 Andreano, D'Oro, Rappuoli and Finco. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.



# The Mononuclear Phagocytic System. Generation of Diversity

Siamon Gordon<sup>1,2\*</sup> and Annette Plüddemann<sup>3</sup>

<sup>1</sup> College of Medicine, Graduate Institute of Biomedical Sciences, Chang Gung University, Taoyuan City, Taiwan, <sup>2</sup> Sir William Dunn School of Pathology, University of Oxford, Oxford, United Kingdom, <sup>3</sup> Nuffield Department of Primary Care Health Sciences, University of Oxford, Oxford, United Kingdom

We are living through an unprecedented accumulation of data on gene expression by macrophages, reflecting their origin, distribution, and localization within all organs of the body. While the extensive heterogeneity of the cells of the mononuclear phagocyte system is evident, the functional significance of their diversity remains incomplete, nor is the mechanism of diversification understood. In this essay we review some of the implications of what we know, and draw attention to issues to be clarified in further research, taking advantage of the powerful genetic, cellular, and molecular tools now available. Our thesis is that macrophage specialization and functions go far beyond immunobiology, while remaining an essential contributor to innate as well as adaptive immunity.

## OPEN ACCESS

### Edited by:

Francesca Di Rosa,  
Consiglio Nazionale Delle Ricerche  
(CNR), Italy

### Reviewed by:

Antonio Sica,  
University of Eastern Piedmont, Italy  
Andreas Wack,  
Francis Crick Institute,  
United Kingdom

### \*Correspondence:

Siamon Gordon  
siamon.gordon@path.ox.ac.uk

### Specialty section:

This article was submitted to  
Molecular Innate Immunity,  
a section of the journal  
Frontiers in Immunology

**Received:** 29 May 2019

**Accepted:** 26 July 2019

**Published:** 09 August 2019

### Citation:

Gordon S and Plüddemann A (2019)  
The Mononuclear Phagocytic System.  
Generation of Diversity.  
Front. Immunol. 10:1893.  
doi: 10.3389/fimmu.2019.01893

**Keywords:** mononuclear phagocyte, macrophage, tissue-specific function, monocyte, plasticity, macrophage heterogeneity, macrophage receptors

## INTRODUCTION

Participation in several Ceppellini workshops by one of the authors (SG) provided an opportunity to examine and present to young investigators some aspects of the unique features of the macrophage, a cell type with an ancient origin in eukaryotic evolution. SG's attachment to the macrophage family has extended over 50 years, rejuvenated over every decade as methodological advances brought new insights and information. However, their biological role in the multicellular organism has remained incomplete, eclipsed as accessory to the specific recognition, and effector functions of lymphoid cells. Metchnikoff already appreciated their professional phagocytic capacity, their digestive proficiency, and potential role in antimicrobial defense (1), while Ehrlich and Wright (2) drew attention to the role of antibodies and opsonins, which enhance phagocytic uptake by monocytes, macrophages, and polymorphonuclear neutrophils (PMN). The discovery of complement and, decades later, the plasma membrane receptors for the Fc domain of IgG specific antibodies and for C3 activated by the classical and alternative pathways, initiated pioneering studies by many investigators [reviewed by Taylor et al. (3)]. Horteaga recognized the special properties of microglia in the Central Nervous System (CNS) (4). The discovery of Dendritic cells (DC) by Steinman and Cohn (5, 6), demonstrated their superior role in antigen capture, processing, and presentation to naïve lymphocytes of peptides, in association with the highly polymorphic Major Histocompatibility (MHC) antigens, thus inducing specific T and B lymphocyte activation and expansion (7). DC-like cells can be readily produced in culture of mouse bone marrow or human monocytes in Granulocyte Macrophage Colony-Stimulating Factor (GM-CSF; CSF-2) and IL-4 (8). To some extent DC eclipsed the role of macrophages in adaptive immunity, although their role in innate immunity was secured by the discovery of Toll-like Receptors (TLR) (9). The discovery and characterization of cytokines produced by and acting

on macrophages, such as Tumor necrosis factor(TNF) (10) and IL-1 (11), prepared the way for anti-TNF therapy (12), to ameliorate destructive immunopathologies such as rheumatoid arthritis. Activation of macrophages by cellular immunity, characterized by Mackaness (13), was shown to be antigen dependent, but non-specific, and lead to the characterization of Interferon (IFN) gamma (14) as the sole mediator of classical activation produced by antigen-specific T lymphocytes and Natural Killer (NK) cells. After setting the stage above, further relevant milestones of macrophage history will be introduced in subsequent sections. Selected historic figures important in the present understanding of tissue macrophage diversity are shown in **Figure 1**.

## THE MONONUCLEAR PHAGOCYTE SYSTEM, A DISPERSED ORGAN

Metchnikoff recognized migratory and sessile, fixed tissue phagocytic cells in his early studies of invertebrate development, by microscopy, and intravital labeling. Direct observation of their recruitment to foreign particles injected *in vivo* lead to further studies in many vertebrate species on their role in host defense against bacteria. Tissue macrophages were subsequently shown to be widely distributed as a system of related cells during development, in the adult steady state, during inflammation, and infection. Aschoff introduced the term Reticulo-Endothelial System (RES), hallmarked by the efficient clearance of particles from the circulation, and extravascular space (15). The imprecise RES nomenclature was replaced by that of the Mononuclear Phagocyte System (MPS) (16), to distinguish mononuclear monocytes and macrophages from PMN, while sharing their highly active capacity as phagocytes. Although widely used till the present day, this terminology is not perfect, since other cell types phagocytose dying cells, and some macrophage-related cells are poorly or even non-phagocytic (17). The diverse cells of the MPS cannot all be characterized by single antigen markers or unique functions expressed at all stages of cell differentiation or activation. Nevertheless, their origin and diversification have common features which point to the valid concept of a specific, dispersed myeloid lineage.

During mammalian development, macrophages derive from haematopoietic precursors in para-aortic regions of the embryo, the yolk sac and fetal liver, seeding organs such as the brain and other tissues before birth (18, 19). A paradigm shift over

recent decades has shown that after birth, in the absence of inflammation, resident macrophages in adult tissues derive from embryonic macrophages which can persist, and gradually turn over locally throughout adult life (20–22). This is especially the case for microglia in the (CNS) and Langerhans cells in the epidermis. The bone marrow, which develops as the main haematopoietic organ perinatally, and functions throughout adult life (23), serves to replenish resident tissue macrophages, for example in the gut (24), where macrophages turn over more actively, and provides blood monocytes (25) in response to increased demand, for example during inflammation and infection (26). The chemokines and receptors which mediate distribution of monocytes and macrophages in the fetus and adult are not completely defined, nor the adhesion molecules which determine organ-specific localization. Chemokines of resident macrophages include fractalkine and its receptor, CX3CR1 (27), and inflammatory, and immune monocyte recruitment mediated by CCL2 and its receptor, CCR2 [**Figure 2**; (29, 30)]. Apart from these and related chemokines, recent studies have uncovered macrophage axonal guidance by semaphorins, and plexinA (31, 32). While resident macrophage populations, for example in the peritoneal cavity, persist locally, they can be induced by inflammation, to enter lymphatic vessels for delivery to lymph nodes (33), or to enter neighboring organs such as liver, by sterile local injury (34). Blood monocytes of bone marrow origin may remain inside the circulation and interact with the luminal surface of vascular endothelium (35), become part of sinus-lining endothelium, as Kupffer cells, or diapedese into tissues. Such recruited monocytes are transient in blood (24–48 h) and shorter lived (4–7 days) after migration into tissues, compared with resident macrophages of yolk sac origin e.g., microglia, which can be extremely long-lived. Other reservoirs of precursors and mature macrophages are found in splenic red pulp (36) or in secondary haematopoietic organs, such as liver.

While the dual origin of tissue resident macrophages is now widely accepted, there is still uncertainty about the relative contribution of the bone marrow in the adult steady state. In mouse liver, for example, early studies by van Furth and Cohn (37), before their embryonic origin was appreciated, argued for a major contribution of recently dividing bone marrow-derived blood monocytes to resident Kupffer cell populations. The pendulum has swung to yolk sac origin, perhaps too far, as acknowledged by more recent studies (38). The Geissmann group, investigating the origin of murine osteoclasts, showed that after initial perinatal formation of multinucleated cells in bone, monocytes of bone marrow origin are recruited and continue to fuse with osteoclasts throughout adult life (39).

## GROWTH AND DIFFERENTIATION

Studies by Metcalf (**Figure 1**) on colony forming cells and lineage-specific growth factors contributed greatly to our understanding of haematopoietic stem cell growth and differentiation *in vitro* (40). Lineage tracing by several groups (41–43) built on studies by Stanley on CSF-1 [reviewed by Chitu and Stanley (44)] and on GM-CSF (45), the major

**Abbreviations:** ADGR, adhesion 7-transmembrane G-protein coupled receptor; BAI-1, brain-specific angiogenesis inhibitor-1; CSF1-R, macrophage colony stimulating factor receptor; DC, Dendritic cells; EMR, epidermal growth factor-like module-containing, mucin-like hormone receptor-like receptor; ITAM, immunoreceptor tyrosine-based activation motif; GM-CSF, granulocyte macrophage colony stimulating factor; GPCR, G-protein coupled receptor; IL, Interleukin; iPSC, induced pluripotent stem cell; MARCO, macrophage receptor with collagenous domain; MHC, major histocompatibility complex; MPS, Mononuclear phagocyte system; NK, natural killer cell; PD-1, Programmed cell death-1; PMN, polymorphonuclear leukocyte; RES, reticulendothelial system; scRNA seq, single cell ribonucleic acid sequencing; Sirp, signal regulatory protein alpha; SR-A, scavenger receptor A; Tie2, Tyrosine-protein kinase receptor for angiopoietins-2; TLR, toll-like receptor; TNF, tumor necrosis factor; TREM-2, Triggering receptor expressed on myeloid cells-2.



**FIGURE 1** | Historic figures associated with macrophages, related cells and their specialized functions.

growth/differentiation factors for monocytes, macrophages, and DC. After initial description by von Kolliker in 1873 (**Figure 1**) (46), Loutit (47) produced proof of the bone marrow origin of osteoclasts; CSF-1 –deficient osteopetrotic op/op mice (48) lacked many, but not all tissue macrophage populations (49). Residual tissue macrophage populations such as microglia, for example, may depend on IL-34, a second ligand for the CSF-1 Receptor, since patients with profound human CSF-1 R deficiency have grossly abnormal CNS development attributed to the absence of microglia (50). Collin et al. have identified mutations which affect monocyte and DC growth and differentiation in humans; bone marrow transplantation and adoptive transfer of haematopoietic stem cells provide clinical and experimental models of monocytopoietic differentiation *in vivo* (51). Recent studies by Olsson et al. (52) and Yanez et al. (53) have demonstrated a binary origin of monocytes in the mouse, exploiting single cell and population RNA seq analysis and adoptive cell transfer.

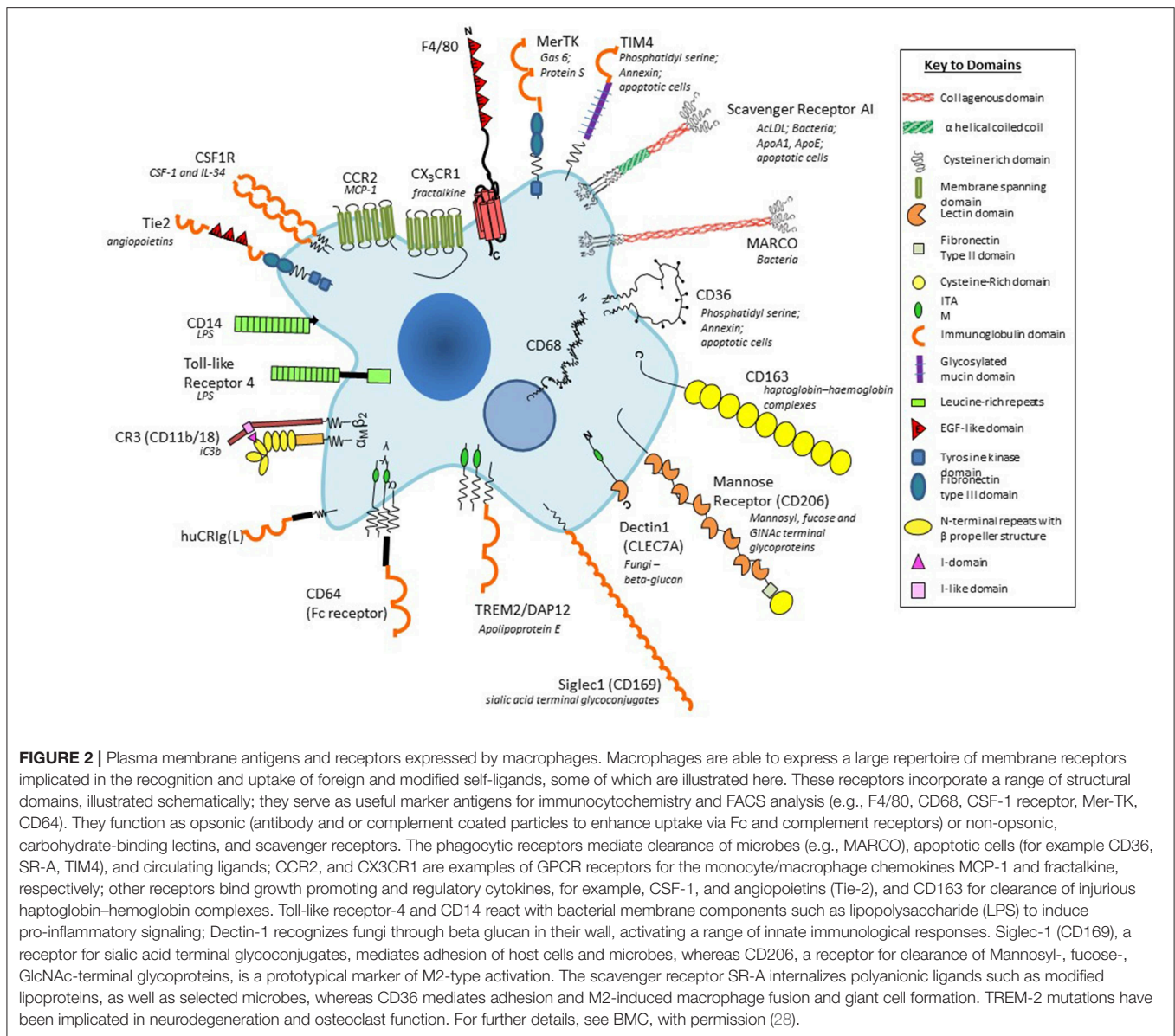
In spite of these basic discoveries, we need more quantitative information on the number of monocytes, macrophages, and DC in human tissues, and their life span *in vivo*. Yona et al. traced the relationship of human monocytes in the steady state and the kinetic response of monocyte subpopulations to endotoxin administration *in vivo* (54). The subset of monocytic precursors which gives rise to osteoclasts remains to be determined;

osteoclasts can be readily produced *in vitro* by culture of monocytes in CSF-1, and Rank Ligand (46), which should facilitate such studies.

## TISSUE DISTRIBUTION AND ORGAN-SPECIFIC PROPERTIES

In the mouse, we have benefited from the availability of monoclonal antibody markers such as F4/80 to detect macrophages in the developing embryo, in the adult steady state and following a wide range of models of inflammation, infection, malignancy, and atherosclerosis. In addition, we used a panel of mab to identify tissue-specific heterogeneity of marker expression (3). **Figure 2** provides a schematic cartoon of these and additional macrophage plasma membrane receptors (55). With the aid of these reagents we identified substantial morphologic and antigenic heterogeneity of resident murine macrophages in different tissue environments such as CNS, spleen and bone marrow (28). Further studies demonstrated heterogeneous antigen expression of monocyte-derived macrophages in BCG-induced granulomata (56), as well as in multinucleated macrophage giant cells (57) and osteoclasts (58). Knowledge of the *in situ* phenotypes of human tissue macrophages is still limited.





## HETEROGENEITY OF TISSUE MACROPHAGES: ANTIGEN MARKERS

The F4/80 antigen (EMR1/ADGRE1), discovered by Austyn and Gordon (59), was used by Hume and others (60) to define monocytes, and macrophages in the mouse. F4/80 is mainly expressed on the plasma membrane, with minimal endocytosis, and is stable to aldehyde fixation; immunocytochemistry therefore can provide exquisite detail of plasma membrane processes in tissue macrophages, suggestive of potential interactions with neighboring cells. Regional variation in morphology and dendritic processes is particularly notable in the brain (61). F4/80, a member of a leukocyte 7-transmembrane, adhesion G protein-coupled receptor family, has been implicated in peripheral tolerance (62), but natural ligands have not

been identified. It is also expressed by eosinophils in mouse and human; EMR1 has been identified in other species (63), but expression is transient in human monocyte-derived macrophages. A related molecule, EMR2 (CD312), discovered by Lin and Stacey (64), is expressed by human myeloid cells in blood and tissues, binds chondroitin sulfate B/dermatan sulfate and has been implicated in a human genetic syndrome, vibratory urticaria (65), associated with mast cell degranulation. EMR2 undergoes autoproteolytic cleavage of its extracellular domain to generate an N-terminal polypeptide agonist of GPCR activation.

The F4/80 antigen is expressed during mouse development from midgestation (19) and has been particularly useful in studies of microglia (61). It is also well-expressed in the adult mouse on resident tissue macrophages in the peritoneal cavity, red pulp of spleen, epidermal Langerhans cells, lamina propria

of the gut, and Kupffer cells; expression is low on alveolar macrophages in lung, and absent or minimal in white pulp and T-cell rich areas. F4/80 is absent on osteoclasts, metallophilic macrophages in the splenic marginal zone and on subcapsular sinus macrophages in lymph nodes, which express the pan-macrophage endo/lysosomal marker, CD68. Bone marrow-derived monocytes and tissue macrophages recruited to sites of inflammation, infection and malignancy in the mouse express F4/80 strongly.

SIGLEC-1 (CD169, sialoadhesin) is a macrophage-specific sialic acid-recognition lectin discovered by Crocker on bone marrow stromal macrophages, at the center of haematopoietic islands (66). It is strongly expressed by marginal metallophilic cells in mouse spleen and by subcapsular sinusoidal macrophages in lymph nodes. It has been implicated in retention and release of monocytes from bone marrow into the circulation. Other lectins widely expressed by macrophages, especially after alternative activation by IL-4/-13, include the macrophage mannose receptor (CD206) (67), and Dectin-1 (CLEC7A), identified as a receptor for fungal  $\beta$ -glucan by Brown and Gordon (68) and Taylor et al. (3). Scavenger receptors implicated in clearance of apoptotic cells (69), non-opsonic microbial phagocytosis and lipoprotein endocytosis (70), include SRA-I/II, constitutively present on many tissue macrophages (71) and the structurally related collagenous receptor, MARCO (72), which is constitutively expressed by macrophages in the outer marginal zone of rodent spleen (73), but is induced on many tissue macrophages by microbial Toll-like receptor stimulation.

In addition to the above antigens, macrophages express plasma membrane receptors (28) involved in opsonic recognition of IgG antibodies (FcR), complement components (e.g., CD11b/18), and other opsonins such as milk fat globulin. Other adhesion molecules include various integrins and CD44; plasma membrane receptors that mediate apoptotic cell clearance include an adhesion GPCR BAI-1 (17) and immunoreceptor tyrosine-based activation motif (ITAM) receptors, Tyro, Axl, and MerTK (74). Immunoregulatory receptors include TREM 1 and 2, SIRP  $\alpha$ , and PD-1. These and receptors for growth factors, cytokines and chemokines have served as useful reagents for FACS, lineage and functional analysis, contributing to our knowledge of macrophage heterogeneity in mouse and human. CD11b expression, for example, is well-expressed on microglia and peritoneal macrophages whereas it is downregulated on alveolar macrophages and Kupffer cells *in situ*.

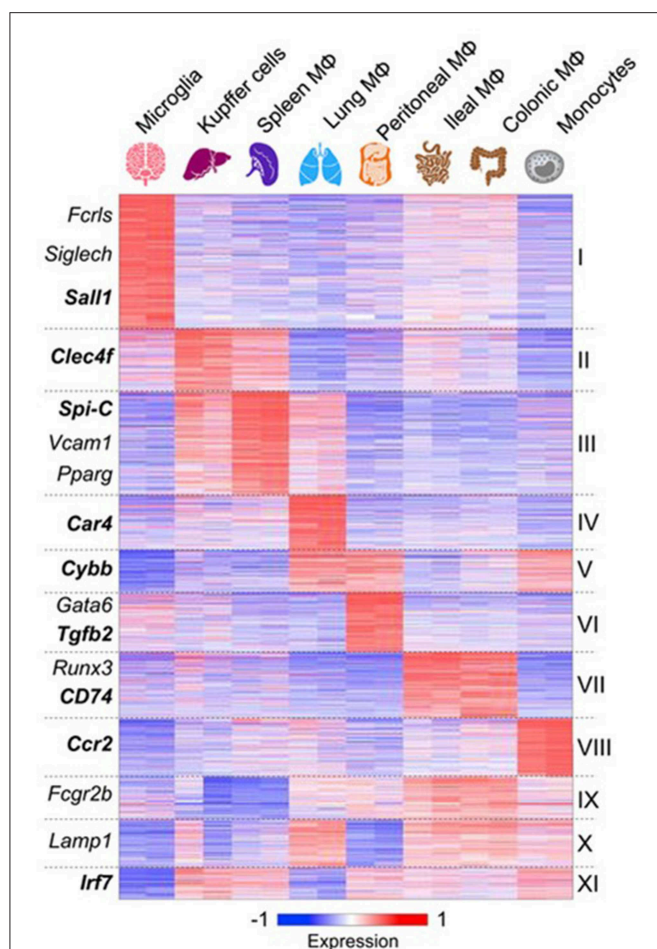
## GENE EXPRESSION

Advances in analysis of macrophage mRNA expression by bulk and single cell sequencing have begun to provide a great deal of new information which has not yet been fully validated by protein expression *in situ* (75–77). However, important conclusions can already be drawn. These studies confirm that macrophages from different tissues are biosynthetically highly active, expressing a large number of diverse, yet canonical macrophage genes. However, tissue macrophages from different organs also express distinctive antigen and mRNA signatures (77) (Figure 3).

Recent publications have reported scRNA-seq analysis of blood mononuclear cells (80), embryonic and adult cell populations, including human placenta (81, 82), which contains both fetal, and maternal macrophages. Improved methods of *in situ* protein expression (83, 84) are required to validate heterogeneity of genomic and epigenomic expression by macrophages isolated from different tissues. Spatial reconstruction of immune niches has been proposed by combining photoactivatable reporters and sc RNA-seq (NICHE-seq) (85). Consortia of investigators are contributing to a human tissue atlas (86), which has already lead to discovery of novel cell types and functions. Open access to data will extend knowledge of variation in gene expression by macrophages from different sources. This will illustrate developmental, physiologic, and pathologic expression and functions of resident and monocyte-derived macrophages, as well as indicating the cells with which they interact locally. Striking results have already been reported on the overriding effect of phagocytosis of apoptotic cells on gene expression by macrophages in different sites. These have used *in vivo* models in gut (87), for example, and include parabiotic experiments (88). The microbiome of the gut does not only affect the macrophage phenotype in its local microenvironment, but also systemically (89, 90), through release of microbial products.

## POLARIZATION AND PLASTICITY OF MACROPHAGES

We used selected membrane markers to examine the phenotype of mouse peritoneal and human monocyte-derived macrophages in culture, following exposure to Th1 and -2 associated cytokines. In the mouse, IL-4, and subsequently IL-13, was shown to enhance expression and function of mannose receptors (CD206), whereas Interferon gamma selectively downregulated this marker (91). Since MHC class II expression was upregulated by both types of cytokine, we termed this process, alternative, and classical activation, respectively. The terminology M2 and M1 was introduced to include other prototypic stimuli such as immune complexes and macrophage expressed-signatures of selected marker genes (92, 93). We found, using a range of *in vitro* and *in vivo* models, that transglutaminase 2 expression, which is not specific for macrophages, was a consistent marker of alternative macrophage activation in humans and mice (94). Subsequent studies by many investigators showed that macrophage polarization involved a spectrum of changes in gene expression (95); to be a useful concept, we proposed that the term alternative activation should be restricted to the prototypical Th-2 cytokines, IL-4, and IL-13 and their common and specific plasma membrane receptors (96). Microarray analysis of macrophage populations using a range of activation and regulatory stimuli, indicates that modules of genes can be identified as signatures to distinguish among different forms of activation. Further analyses of single cell RNA, and protein expression of gene signatures by yolk sac- and bone marrow-derived macrophages and their correlation with distinct functions such as cytotoxicity, and tissue repair, are required to refine polarization in individual organs.



**FIGURE 3 |** Macrophages express canonical and tissue-specific mRNA signatures. From (77) for further details, with permission. See also (78), ImmGen Consortium (79).

Both classical and alternative macrophage activation can be divided into two distinct phases, an initial priming step by the appropriate cytokine, and completion by a phagocytic or microbial stimulus which induces further changes in gene expression and serves to localize macrophage effector activity. Microbial uptake enhances cytotoxic and pro-inflammatory activity of interferon-primed, classically activated macrophages, whereas uptake of apoptotic cells by IL-4 treated macrophages, enhances anti-inflammatory gene expression by alternatively primed macrophages (97). In experimental models, LPS can induce paradoxical enhancement of JNK activation following Scavenger receptor ligation of IL-4-primed macrophages, suggesting that the outcome will depend on the nature of the phagocytic receptor involved (98).

Priming of macrophages can also induce an adaptive enhancement of microbial phagocytosis and innate immune function. For example, LPS or microbial stimulation upregulates MARCO expression enabling subsequent enhanced uptake of *Neisseria meningitidis* via this receptor (99, 100). This observation harks back to the earlier studies of Mackaness on

macrophage activation by BCG and *Listeria monocytogenes*, shown to be antigen dependent, but non-specific for the inducing organism (13). Netea et al. have extended this phenomenon, an example of “trained immunity” (101, 102), and have implicated epigenetic mechanisms in its imprinting.

These concepts are important in attempts to reverse polarization, for example of tumor associated macrophages, for potential immunotherapy. Evidence that the macrophage phenotype *in vivo* is plastic and reversible by adoptive transfer to different tissue microenvironments is sketchy. Van de Laar et al. have shown that yolk sac macrophages, fetal liver and adult monocytes efficiently colonize the empty alveolar niche of *Csf2rb*<sup>-/-</sup> mice, unlike mature liver peritoneal or colon macrophages (103). We have found that once macrophages have differentiated terminally, for example to a resident peritoneal phenotype, they cannot be induced to express adhesion receptors characteristic of other terminally differentiated macrophages such as those found in bone marrow haemopoietic clusters. Furthermore, experiments need to distinguish between changes in cell populations and individual cells. However, the phenomenon of induced pluripotency (104) indicates that transcription factors and chromatin conformation can enable true plasticity and the ability to give rise to embryonic stem cells, able to generate different somatic cell types, including macrophages (105) and microglia (106) *de novo*.

## GENERATION OF DIVERSITY IN TISSUE MACROPHAGES

The evidence that resident embryo or bone marrow-derived populations of tissue macrophages, distributed throughout organs in the steady state, acquire distinct phenotypes as well as expressing core macrophage properties, raises a fascinating problem of origin of their diversity. The extent of adaptation by monocytes recruited by infection to different tissue environments, for example in granuloma formation, requires further characterization. In order to establish a testable hypothesis to account for the generation of diversity, we have to keep in mind several properties which distinguish macrophages from T and B lymphocytes, in which antigen receptor gene rearrangement and clonal selection have provided unexpected solutions to account for repertoire diversity and antigen specificity. Macrophages express a broader range of receptors than lymphocytes to distinguish foreign, modified-self and self-ligands; these include proteins and peptides, carbohydrates, nucleic acids, and lipids. Macrophage receptors can be viewed as “hard wired,” unlike the more selective, antigen-specific receptors of adaptive lymphocytes. Tissue macrophages are terminally differentiated, capable of only a limited degree of proliferative capacity once they enter tissues. Clonal selection can therefore be ruled out. We do not know the size of the macrophage repertoire, but it must be substantial to accommodate interactions with other cell types within the body, including macrophages themselves, as well as so-called “pattern recognition receptors” for exogenous and endogenous ligands. Many investigators acknowledge that the local tissue as well as



exogenous micro-environment must play a specifying role in inducing or selecting expression of a particular constellation of surface receptors and gene products [for example (75, 107)]. In addition, macrophages can recognize a host of intracellular ligands in their cytosolic, biosynthetic, secretory, or endocytic compartments. However, chromatin conformation, transcription factors and enhancers, in addition to epigenetic mechanisms, must also determine the programme of differential gene expression, and modulation of the macrophage phenotype (108–112). T'Jonck et al. have discussed the role of niche signals and transcription factors involved in tissue resident macrophage development in detail (113).

These considerations leave many questions as to how, when and where, and specifically by which intrinsic and environmental mechanisms, diversity is achieved. Surprisingly little consideration has been given to the nature of the diverse ligands in the extracellular matrix of different tissues (114); nor the role of various epithelia, endothelia, mesenchymal, and neuro-endocrine cells, all of which interact with macrophages as a result of their unique migration and organ distribution (83, 84, 115, 116).

## TISSUE-SPECIFIC FUNCTIONS OF MACROPHAGES

Tissue macrophages express general, prototypic, functions throughout the body which contribute to homeostasis, recognition and responses to intrinsic and external perturbation, restoring physiologic stability, and contributing to repair after injury. In different organs they adapt to different micro-environments with variations on the themes of clearance of particles and soluble ligands, digestion or storage in lysosomes, constitutive, and induced biosynthesis, and secretion. They interact with living or dying cells and microbes, blood and lymph, undergoing metabolic adaptation, and altering adhesion to extracellular matrix as they migrate, through different locations over time. In the process, they may respond to injurious stimuli by autophagy, cell growth or death. Nevertheless, we can already discern remarkable variations in organ-specific functions to which they contribute; these include a central role in haematopoietic turnover, and haem degradation (117, 118), lymphoid trafficking of immune cells (33); mucosal physiology, for instance in the gut (119, 120); remodeling in the CNS (107, 121, 122); neural- adipose tissue metabolism (123), and adipose- sympathetic nervous interactions (124); and electrical activity in the heart (125). Current studies in single cell RNA

and protein expression by tissue macrophages will provide more examples of trophic and defense functions, contributing to embryonic development, anatomic, physiologic, and pathologic processes. Returning to our earlier discussion of how such diversity might be generated, it seems likely that encounters with different ligands in their tissue microenvironment can exploit pre-existing or induce novel sensors to activate adaptive changes in transcription and epigenetic modification; this begs the question of the extent and mechanisms of initial tissue-specific receptor diversification. While differentiation can generate a core panel of recognition molecules on and within the macrophage, it may be necessary to postulate further induction, feedback amplification, or selection by as yet unknown somatic gene expression mechanisms. Investigating the details of osteoclast and DC development *in vivo* and *in vitro* may provide further clues to novel molecular mechanisms.

## CONCLUSIONS

Recent progress in molecular and cellular biology have brought exciting insights into view, enabling us to characterize monocyte/macrophage heterogeneity *in situ*. Understanding the themes of their functions within multicellular organisms across a range of evolutionary stages will make it possible to discover a unifying pattern extending far beyond innate or adaptive, cellular and humoral immunity. The challenge will be to imagine the properties underlying the genes and molecules which can lead us to such knowledge. Finally, we need to consider the implications of monocyte/macrophage heterogeneity for therapy. Factors to be taken into account for macrophage-directed immunotherapy include the expression of target antigens on distinct subpopulations, the route of administration, risk of off-target effects and species differences. Similarly, for potential adoptive cell therapy, the origin, differentiation, proliferative capacity and activation status have to be defined, as well as genetic compatibility.

## AUTHOR CONTRIBUTIONS

SG conceived and wrote the manuscript. AP reviewed and edited the manuscript and designed the figures.

## ACKNOWLEDGMENTS

We acknowledge stimulating discussions with numerous colleagues and fellow macrophage enthusiasts.

## REFERENCES

1. Teti G, Biondo C, Beninati C. The phagocyte, metchnikoff, and the foundation of immunology. *Microbiol Spectr.* (2016) 4:MCHD-0009-2015. doi: 10.1128/microbiolspec.MCHD-0009-2015
2. Kaufmann SHE. Immunology's coming of age. *Front Immunol.* (2019) 10:684. doi: 10.3389/fimmu.2019.00684
3. Taylor PR, Martinez-Pomares L, Stacey M, Lin HH, Brown GD, Gordon S. Macrophage receptors and immune recognition. *Annu Rev Immunol.* (2005) 23:901–44. doi: 10.1146/annurev.immunol.23.021704.115816
4. Sierra A, De Castro F, Del Rio-Hortega J, Rafael Iglesias-Rozas J, Garrosa M, Kettenmann H. The “big-bang” for modern glial biology: translation and comments on pio del rio-hortega 1919 series of papers on microglia. *Glia.* (2016) 64:1801–40. doi: 10.1002/glia.23046



5. Steinman RM, Cohn ZA. Identification of a novel cell type in peripheral lymphoid organs of mice. I morphology, quantitation, tissue distribution. *J Exp Med.* (1973) 137:1142–62. doi: 10.1084/jem.137.5.1142
6. Steinman RM, Moberg CL, Zanvil alexander cohn 1926-1993. *J Exp Med.* (1994) 179:1–30. doi: 10.1084/jem.179.1.1
7. Moberg CL. An appreciation of ralph marvin steinman (1943–2011). *J Exp Med.* (2011) 208:2337–42. doi: 10.1084/jem.20112294
8. Austyn JM. Dendritic cells in the immune system—history, lineages, tissues, tolerance, and immunity. *Microbiol Spectr.* (2016) 4:MCHD-0046-2016. doi: 10.1128/microbiolspec.MCHD-0046-2016
9. Medzhitov R. TLR-mediated innate immune recognition. *Semin Immunol.* (2007) 19:1–2. doi: 10.1016/j.smim.2007.02.001
10. Beutler B, Cerami A. The biology of cachectin/TNF—a primary mediator of the host response. *Annu Rev Immunol.* (1989) 7:625–55. doi: 10.1146/annurev.iv.07.040189.003205
11. Mantovani A, Dinarello CA, Molgora M, Garlanda C. Interleukin-1 and related cytokines in the regulation of inflammation and immunity. *Immunity.* (2019) 50:778–95. doi: 10.1016/j.immuni.2019.03.012
12. Feldmann M, Maini RN. Lasker clinical medical research award. TNF defined as a therapeutic target for rheumatoid arthritis and other autoimmune diseases. *Nat Med.* (2003) 9:1245–50. doi: 10.1038/nm939
13. Mackaness GB. The immunological basis of acquired cellular resistance. *J Exp Med.* (1964) 120:105–20. doi: 10.1084/jem.120.1.105
14. Lazear HM, Schoggins JW, Diamond MS. Shared and distinct functions of type I and type III interferons. *Immunity.* (2019) 50:907–23. doi: 10.1016/j.immuni.2019.03.025
15. Aschoff L. Das reticulo-endotheliale system. *Ergeb Inn Med Kinderheilk.* (1924) 26:1–118. doi: 10.1007/978-3-642-90639-8\_1
16. Van Furth R, Cohn ZA, Hirsch JG, Humphrey JH, Spector WG, Langevoort HL. The mononuclear phagocyte system: a new classification of macrophages, monocytes, and their precursor cells. *Bull World Health Organ.* (1972) 46:845–52.
17. Morioka S, Maueroed K, Ravichandran KS. Living on the edge: efferocytosis at the interface of homeostasis and pathology. *Immunity.* (2019) 50:1149–62. doi: 10.1016/j.immuni.2019.04.018
18. Perry VH, Andersson PB, Gordon S. Macrophages and inflammation in the central nervous system. *Trends Neurosci.* (1993) 16:268–73. doi: 10.1016/0166-2236(93)90180-T
19. Morris L, Graham CF, Gordon S. Macrophages in haemopoietic and other tissues of the developing mouse detected by the monoclonal antibody F4/80. *Development.* (1991) 112:517–26.
20. Guillems M, Ginhoux F, Jakubczik C, Naik SH, Onai N, Schraml BU, et al. Dendritic cells, monocytes and macrophages: a unified nomenclature based on ontogeny. *Nat Rev Immunol.* (2014) 14:571–8. doi: 10.1038/nri3712
21. Hoeffel G, Ginhoux F. Ontogeny of tissue-resident macrophages. *Front Immunol.* (2015) 6:486. doi: 10.3389/fimmu.2015.00486
22. Lavin Y, Mortha A, Rahman A, Merad M. Regulation of macrophage development and function in peripheral tissues. *Nat Rev Immunol.* (2015) 15:731–44. doi: 10.1038/nri3920
23. Dzierzak E, De Pater E. Regulation of blood stem cell development. *Curr Top Dev Biol.* (2016) 118:1–20. doi: 10.1016/bs.ctdb.2016.01.001
24. Faria AMC, Reis BS, Mucida D. Tissue adaptation: implications for gut immunity and tolerance. *J Exp Med.* (2017) 214:1211–26. doi: 10.1084/jem.20162014
25. Ziegler-Heitbrock L, Ancuta P, Crowe S, Dalod M, Grau V, Hart DN, et al. Nomenclature of monocytes and dendritic cells in blood. *Blood.* (2010) 116:e74–80. doi: 10.1182/blood-2010-02-258558
26. Geissmann F, Jung S, Littman DR. Blood monocytes consist of two principal subsets with distinct migratory properties. *Immunity.* (2003) 19:71–82. doi: 10.1016/S1074-7613(03)00174-2
27. Longman RS, Diehl GE, Victorio DA, Huh JR, Galan C, Miraldi ER, et al. CX(3)CR1(+) mononuclear phagocytes support colitis-associated innate lymphoid cell production of IL-22. *J Exp Med.* (2014) 211:1571–83. doi: 10.1084/jem.20140678
28. Gordon S, Plüddemann A. Tissue macrophages: heterogeneity and functions. *BMC Biol.* (2017) 15:53. doi: 10.1186/s12915-017-0392-4
29. Charo IF, Ransohoff RM. The many roles of chemokines and chemokine receptors in inflammation. *N Engl J Med.* (2006) 354:610–21. doi: 10.1056/NEJMra052723
30. Gerard C, Rollins BJ. Chemokines and disease. *Nat Immunol.* (2001) 2:108–15. doi: 10.1038/84209
31. Kong Y, Janssen BJ, Malinauskas T, Vangoor VR, Coles CH, Kaufmann R, et al. Structural basis for plexin activation and regulation. *Neuron.* (2016) 91:548–60. doi: 10.1016/j.neuron.2016.06.018
32. Seiradake E, Jones EY, Klein R. Structural perspectives on axon guidance. *Annu Rev Cell Dev Biol.* (2016) 32:577–608. doi: 10.1146/annurev-cellbio-111315-125008
33. Zhang Y, Roth TL, Gray EE, Chen H, Rodda LB, Liang Y, et al. Migratory and adhesive cues controlling innate-like lymphocyte surveillance of the pathogen-exposed surface of the lymph node. *Elife.* (2016) 5:e18156. doi: 10.7554/eLife.18156
34. Wang J, Kubes P. A reservoir of mature cavity macrophages that can rapidly invade visceral organs to affect tissue repair. *Cell.* (2016) 165:668–78. doi: 10.1016/j.cell.2016.03.009
35. Auffray C, Fogg D, Garfa M, Elain G, Join-Lambert O, Kayal S, et al. Monitoring of blood vessels and tissues by a population of monocytes with patrolling behavior. *Science.* (2007) 317:666–70. doi: 10.1126/science.1142883
36. Swirski FK, Nahrendorf M, Etzrodt M, Wildgruber M, Cortez-Retamozo V, Panizzi P, et al. Identification of splenic reservoir monocytes and their deployment to inflammatory sites. *Science.* (2009) 325:612–6. doi: 10.1126/science.1175202
37. Van Furth R, Cohn ZA. The origin and kinetics of mononuclear phagocytes. *J Exp Med.* (1968) 128:415–35. doi: 10.1084/jem.128.3.415
38. Guillems M, Mildner A, Yona S. Developmental and functional heterogeneity of monocytes. *Immunity.* (2018) 49:595–613. doi: 10.1016/j.immuni.2018.10.005
39. Jacome-Galarza CE, Percin GI, Muller JT, Mass E, Lazarov T, Eitler J, et al. Developmental origin, functional maintenance and genetic rescue of osteoclasts. *Nature.* (2019) 568:541–5. doi: 10.1038/s41586-019-1105-7
40. Metcalf D. Growth and differentiation factors. *Microbiol Spectr.* (2016) 4:MCHD-0004-2015. doi: 10.1128/microbiolspec.MCHD-0004-2015
41. Ginhoux F, Guillems M. Tissue-resident macrophage ontogeny and homeostasis. *Immunity.* (2016) 44:439–49. doi: 10.1016/j.immuni.2016.02.024
42. Gomez Perdiguero E, Klapproth K, Schulz C, Busch K, Azzoni E, Crozet L, et al. Tissue-resident macrophages originate from yolk-sac-derived erythromyeloid progenitors. *Nature.* (2015) 518:547–51. doi: 10.1038/nature13989
43. Yona S, Kim KW, Wolf Y, Mildner A, Varol D, Breker M, et al. Fate mapping reveals origins and dynamics of monocytes and tissue macrophages under homeostasis. *Immunity.* (2013) 38:79–91. doi: 10.1016/j.immuni.2012.12.001
44. Chitu V, Stanley ER. Regulation of embryonic and postnatal development by the CSF-1 receptor. *Curr Top Dev Biol.* (2017) 123:229–75. doi: 10.1016/bs.ctdb.2016.10.004
45. Dougan M, Dranoff G, Dougan SK. GM-CSF, IL-3, and IL-5 family of cytokines: regulators of inflammation. *Immunity.* (2019) 50:796–811. doi: 10.1016/j.immuni.2019.03.022
46. Martin TJ. Historically significant events in the discovery of RANK/RANKL/OPG. *World J Orthop.* (2013) 4:186–97. doi: 10.5312/wjo.v4.i4.186
47. Nisbet NW, Menage J, Loutit JF. Osteogenesis in osteopetrotic mice. *Calcif Tissue Int.* (1982) 34:37–42. doi: 10.1007/BF02411206
48. Wiktor-Jedrzejczak W, Bartocci A, Ferrante AW Jr, Ahmed-Ansari A, Sell KW, Pollard JW, et al. Total absence of colony-stimulating factor 1 in the macrophage-deficient osteopetrotic (op/op) mouse. *Proc Natl Acad Sci USA.* (1990) 87:4828–32. doi: 10.1073/pnas.87.12.4828
49. Witmer-Pack MD, Hughes DA, Schuler G, Lawson L, McWilliam A, Inaba K, et al. Identification of macrophages and dendritic cells in the osteopetrotic (op/op) mouse. *J Cell Sci.* (1993) 104(Pt 4):1021–9.
50. Oosterhof N, Chang IJ, Karimiani EG, Kuil LE, Jensen DM, Daza R, et al. Homozygous mutations in CSF1R cause a pediatric-onset leukoencephalopathy and can result in congenital absence of microglia. *Am J Hum Genet.* (2019) 104:936–47. doi: 10.1016/j.ajhg.2019.03.010

51. Collin M, Bigley V. Monocyte, macrophage, and dendritic cell development: the human perspective. *Microbiol Spectr.* (2016) 4:MCHD-0015-2015. doi: 10.1128/microbiolspec.MCHD-0015-2015
52. Olsson A, Venkatasubramanian M, Chaudhri VK, Aronow BJ, Salomonis N, Singh H, et al. Single-cell analysis of mixed-lineage states leading to a binary cell fate choice. *Nature.* (2016) 537:698–702. doi: 10.1038/nature19348
53. Yanez A, Coetzee SG, Olsson A, Muench DE, Berman BP, Hazelett DJ, et al. Granulocyte-monocyte progenitors and monocyte-dendritic cell progenitors independently produce functionally distinct monocytes. *Immunity.* (2017) 47:890–902 e4. doi: 10.1016/j.immuni.2017.10.021
54. Patel AA, Zhang Y, Fullerton JN, Boelen L, Rongvaux A, Maini AA, et al. The fate and lifespan of human monocyte subsets in steady state and systemic inflammation. *J Exp Med.* (2017) 214:1913–23. doi: 10.1084/jem.20170355
55. Gordon S. Phagocytosis: an immunobiological process. *Immunity.* (2016) 44:463–75. doi: 10.1016/j.immuni.2016.02.026
56. Keshav S, Chung P, Milon G, Gordon S. Lysozyme is an inducible marker of macrophage activation in murine tissues as demonstrated by *in situ* hybridization. *J Exp Med.* (1991) 174:1049–58. doi: 10.1084/jem.174.5.1049
57. Milde R, Ritter J, Tennent GA, Loesch A, Martinez FO, Gordon S, et al. Multinucleated giant cells are specialized for complement-mediated phagocytosis and large target destruction. *Cell Rep.* (2015) 13:1937–48. doi: 10.1016/j.celrep.2015.10.065
58. Pereira M, Petretto E, Gordon S, Bassett JHD, Williams GR, Behmoaras J. Common signalling pathways in macrophage and osteoclast multinucleation. *J Cell Sci.* (2018) 131:jcs216267. doi: 10.1242/jcs.216267
59. Austyn JM, Gordon S. F4/80, a monoclonal antibody directed specifically against the mouse macrophage. *Eur J Immunol.* (1981) 11:805–15. doi: 10.1002/eji.1830111013
60. Hume DA, Gordon S. Mononuclear phagocyte system of the mouse defined by immunohistochemical localization of antigen F4/80. identification of resident macrophages in renal medullary and cortical interstitium and the juxtaglomerular complex. *J Exp Med.* (1983) 157:1704–9. doi: 10.1084/jem.157.5.1704
61. Lawson LJ, Perry VH, Dri P, Gordon S. Heterogeneity in the distribution and morphology of microglia in the normal adult mouse brain. *Neuroscience.* (1990) 39:151–70. doi: 10.1016/0306-4522(90)90229-W
62. Lin HH, Faunce DE, Stacey M, Terajewicz A, Nakamura T, Zhang-Hoover J, et al. The macrophage F4/80 receptor is required for the induction of antigen-specific effector regulatory T cells in peripheral tolerance. *J Exp Med.* (2005) 201:1615–25. doi: 10.1084/jem.20042307
63. Waddell LA, Lefevre L, Bush SJ, Raper A, Young R, Lisowski ZM, et al. ADGRE1 (EMR1, F4/80) is a rapidly-evolving gene expressed in mammalian monocyte-macrophages. *Front Immunol.* (2018) 9:2246. doi: 10.3389/fimmu.2018.02246
64. Lin HH, Stacey M. G protein-coupled receptors in macrophages. *Microbiol Spectr.* (2016) 4:MCHD-0028-2016. doi: 10.1128/microbiolspec.MCHD-0028-2016
65. Boyden SE, Desai A, Cruse G, Young ML, Bolan HC, Scott LM, et al. Vibratory urticaria associated with a missense variant in ADGRE2. *N Engl J Med.* (2016) 374:656–63. doi: 10.1056/NEJMoa1500611
66. Klaas M, Crocker PR. Sialoadhesin in recognition of self and non-self. *Semin Immunopathol.* (2012) 34:353–64. doi: 10.1007/s00281-012-0310-3
67. Martinez-Pomares L. The mannose receptor. *J Leukoc Biol.* (2012) 92:1177–86. doi: 10.1189/jlb.0512231
68. Brown GD, Gordon S. Immune recognition. a new receptor for beta-glucans. *Nature.* (2001) 413:36–7. doi: 10.1038/35092620
69. Gordon S, Plüddemann A. Macrophage clearance of apoptotic cells: a critical assessment. *Front Immunol.* (2018) 9:127. doi: 10.3389/fimmu.2018.00127
70. Neyen C, Plüddemann A, Roversi P, Thomas B, Cai L, Van Der Westhuyzen DR, et al. Macrophage scavenger receptor A mediates adhesion to apolipoproteins A-I and E. *Biochemistry.* (2009) 48:11858–71. doi: 10.1021/bi9013769
71. Plüddemann A, Mukhopadhyay S, Gordon S. The interaction of macrophage receptors with bacterial ligands. *Expert Rev Mol Med.* (2006) 8:1–25. doi: 10.1017/S1462399406000159
72. Mukhopadhyay S, Chen Y, Sankala M, Peiser L, Pikkarainen T, Kraal G, et al. MARCO, an innate activation marker of macrophages, is a class A scavenger receptor for *Neisseria meningitidis*. *Eur J Immunol.* (2006) 36:940–9. doi: 10.1002/eji.200535389
73. Kraal G, Mebius R. New insights into the cell biology of the marginal zone of the spleen. *Int Rev Cytol.* (2006) 250:175–215. doi: 10.1016/S0074-7696(06)50005-1
74. Rothlin CV, Carrera-Silva EA, Bosurgi L, Ghosh S. TAM receptor signaling in immune homeostasis. *Annu Rev Immunol.* (2015) 33:355–91. doi: 10.1146/annurev-immunol-032414-112103
75. Amit I, Winter DR, Jung S. The role of the local environment and epigenetics in shaping macrophage identity and their effect on tissue homeostasis. *Nat Immunol.* (2016) 17:18–25. doi: 10.1038/ni.3325
76. Becher B, Schlitzer A, Chen J, Mair F, Sumatoh HR, Teng KW, et al. High-dimensional analysis of the murine myeloid cell system. *Nat Immunol.* (2014) 15:1181–9. doi: 10.1038/ni.3006
77. Lavin Y, Winter D, Blecher-Gonen R, David E, Keren-Shaul H, Merad M, et al. Tissue-resident macrophage enhancer landscapes are shaped by the local microenvironment. *Cell.* (2014) 159:1312–26. doi: 10.1016/j.cell.2014.11.018
78. Gautier EL, Shay T, Miller J, Greter M, Jakubick C, Ivanov S, et al. Gene-expression profiles and transcriptional regulatory pathways that underlie the identity and diversity of mouse tissue macrophages. *Nat Immunol.* (2012) 13:1118–28. doi: 10.1038/ni.2419
79. Immgenconsortium. Open-source ImmGen: mononuclear phagocytes. *Nat Immunol.* (2016) 17:741. doi: 10.1038/ni.3478
80. Villani AC, Satija R, Reynolds G, Sarkizova S, Shekhar K, Fletcher J, et al. Single-cell RNA-seq reveals new types of human blood dendritic cells, monocytes, and progenitors. *Science.* (2017) 356:eaa4573. doi: 10.1126/science.aaa4573
81. Suryawanshi H, Morozov P, Straus A, Sahasrabudhe N, Max KEA, Garzia A, et al. A single-cell survey of the human first-trimester placenta and decidua. *Sci Adv.* (2018) 4:eaau4788. doi: 10.1126/sciadv.aau4788
82. Vento-Tormo R, Efremova M, Botting RA, Turco MY, Vento-Tormo M, Meyer KB, et al. Single-cell reconstruction of the early maternal-fetal interface in humans. *Nature.* (2018) 563:347–53. doi: 10.1038/s41586-018-0698-6
83. Goltsev Y, Samusik N, Kennedy-Darling J, Bhate S, Hale M, Vazquez G, et al. Deep profiling of mouse splenic architecture with CODEX multiplexed imaging. *Cell.* (2018) 174:968–81 e15. doi: 10.1016/j.cell.2018.07.010
84. Keren L, Bosse M, Marquez D, Angostari R, Jain S, Varma S, et al. A Structured Tumor-Immune Microenvironment in Triple Negative Breast Cancer Revealed by Multiplexed Ion Beam Imaging. *Cell.* (2018) 174:1373–87 e19. doi: 10.1016/j.cell.2018.08.039
85. Medaglia C, Giladi A, Stoler-Barak L, De Giovanni M, Salame TM, Biram A, et al. Spatial reconstruction of immune niches by combining photoactivatable reporters and scRNA-seq. *Science.* (2017) 358:1622–6. doi: 10.1126/science.aao4277
86. Regev A, Teichmann SA, Lander ES, Amit I, Benoist C, Birney E, et al. The human cell atlas. *Elife.* (2017) 6:e27041. doi: 10.7554/eLife.27041
87. Cummings RJ, Barbet G, Bongers G, Hartmann BM, Gettler K, Muniz L, et al. Different tissue phagocytes sample apoptotic cells to direct distinct homeostasis programs. *Nature.* (2016) 539:565–9. doi: 10.1038/nature20138
88. A-Gonzalez N, Quintana JA, Garcia-Silva S, Mazariegos M, Gonzalez De La Aleja A, Nicolas-Avila JA, et al. Phagocytosis imprints heterogeneity in tissue-resident macrophages. *J Exp Med.* (2017) 214:1281–96. doi: 10.1084/jem.20161375
89. Abdel-Haq R, Schlachetzki JCM, Glass CK, Mazmanian SK. Microbiome-microglia connections via the gut-brain axis. *J Exp Med.* (2019) 216:41–59. doi: 10.1084/jem.20180794
90. Erny D, Hrabé De Angelis AL, Jaitin D, Wieghofer P, Staszewski O, David E, et al. Host microbiota constantly control maturation and function of microglia in the CNS. *Nat Neurosci.* (2015) 18:965–77. doi: 10.1038/nn.4030
91. Stein M, Keshav S, Harris N, Gordon S. Interleukin 4 potentially enhances murine macrophage mannose receptor activity: a marker of alternative immunologic macrophage activation. *J Exp Med.* (1992) 176:287–92. doi: 10.1084/jem.176.1.287
92. Mantovani A, Sozzani S, Locati M, Allavena P, Sica A. Macrophage polarization: tumor-associated macrophages as a paradigm for polarized

- M2 mononuclear phagocytes. *Trends Immunol.* (2002) 23:549–55. doi: 10.1016/S1471-4906(02)02302-5
93. Mosser DM, Edwards JP. Exploring the full spectrum of macrophage activation. *Nat Rev Immunol.* (2008) 8:958–69. doi: 10.1038/nri2448
  94. Martinez FO, Helming L, Milde R, Varin A, Melgert BN, Draijer C, et al. Genetic programs expressed in resting and IL-4 alternatively activated mouse and human macrophages: similarities and differences. *Blood.* (2013) 121:e57–69. doi: 10.1182/blood-2012-06-436212
  95. Murray PJ, Allen JE, Biswas SK, Fisher EA, Gilroy DW, Goerdt S, et al. Macrophage activation and polarization: nomenclature and experimental guidelines. *Immunity.* (2014) 41:14–20. doi: 10.1016/j.immuni.2014.06.008
  96. Martinez FO, Gordon S. The M1 and M2 paradigm of macrophage activation: time for reassessment. *F1000Prime Rep.* (2014) 6:13. doi: 10.12703/P6-13
  97. Bosurgi L, Cao YG, Cabeza-Cabrero M, Tucci A, Hughes LD, Kong Y, et al. Macrophage function in tissue repair and remodeling requires IL-4 or IL-13 with apoptotic cells. *Science.* (2017) 356:1072–6. doi: 10.1126/science.aai8132
  98. Guo M, Hartlova A, Gierlinski M, Prescott A, Castellvi J, Losa JH, et al. Triggering MSR1 promotes JNK-mediated inflammation in IL-4-activated macrophages. *EMBO J.* (2019) 38:e100299. doi: 10.15252/embj.2018100299
  99. Bowdish DM, Loffredo MS, Mukhopadhyay S, Mantovani A, Gordon S. Macrophage receptors implicated in the “adaptive” form of innate immunity. *Microbes Infect.* (2007) 9:1680–7. doi: 10.1016/j.micinf.2007.09.002
  100. Locati M, Mantovani A, Sica A. Macrophage activation and polarization as an adaptive component of innate immunity. *Adv Immunol.* (2013) 120:163–84. doi: 10.1016/B978-0-12-417028-5.00006-5
  101. Arts RJ, Netea MG. Adaptive characteristics of innate immune responses in macrophages. *Microbiol Spectr.* (2016) 4:MCHD-0023-2015. doi: 10.1128/microbiolspec.MCHD-0023-2015
  102. Netea MG, Schlitzer A, Placek K, Joosten LAB, Schultze JL. Innate and adaptive immune memory: an evolutionary continuum in the host's response to pathogens. *Cell Host Microbe.* (2019) 25:13–26. doi: 10.1016/j.chom.2018.12.006
  103. Van De Laar L, Saelens W, De Prijck S, Martens L, Scott CL, Van Isterdael G, et al. Yolk sac macrophages, fetal liver, and adult monocytes can colonize an empty niche and develop into functional tissue-resident macrophages. *Immunity.* (2016) 44:755–68. doi: 10.1016/j.immuni.2016.02.017
  104. Theunissen TW, Jaenisch R. Mechanisms of gene regulation in human embryos and pluripotent stem cells. *Development.* (2017) 144:4496–509. doi: 10.1242/dev.157404
  105. Mukherjee C, Hale C, Mukhopadhyay S. A simple multistep protocol for differentiating human induced pluripotent stem cells into functional macrophages. *Methods Mol Biol.* (2018) 1784:13–28. doi: 10.1007/978-1-4939-7837-3\_2
  106. Haenseler W, Sansom SN, Buchrieser J, Newey SE, Moore CS, Nicholls FJ, et al. A highly efficient human pluripotent stem cell microglia model displays a neuronal-co-culture-specific expression profile and inflammatory response. *Stem Cell Reports.* (2017) 8:1727–42. doi: 10.1016/j.stemcr.2017.05.017
  107. Gosselin D, Skola D, Coufal NG, Holtman IR, Schlachetzki JCM, Sajti E, et al. An environment-dependent transcriptional network specifies human microglia identity. *Science.* (2017) 356:ea3222. doi: 10.1126/science.aal3222
  108. Glass CK. Genetic and genomic approaches to understanding macrophage identity and function. *Arterioscler Thromb Vasc Biol.* (2015) 35:755–62. doi: 10.1161/ATVBAHA.114.304051
  109. Heinz S, Benner C, Spann N, Bertolino E, Lin YC, Laslo P, et al. Simple combinations of lineage-determining transcription factors prime cis-regulatory elements required for macrophage and B cell identities. *Mol Cell.* (2010) 38:576–89. doi: 10.1016/j.molcel.2010.05.004
  110. Hoeksema MA, Glass CK. Nature and nurture of tissue-specific macrophage phenotypes. *Atherosclerosis.* (2019) 281:159–67. doi: 10.1016/j.atherosclerosis.2018.10.005
  111. Ivashkiv LB. Epigenetic regulation of macrophage polarization and function. *Trends Immunol.* (2013) 34:216–23. doi: 10.1016/j.it.2012.11.001
  112. Zhu YP, Thomas GD, Hedrick CC. 2014 Jeffrey M. Hoeg award lecture: transcriptional control of monocyte development. *Arterioscler Thromb Vasc Biol.* (2016) 36:1722–33. doi: 10.1161/ATVBAHA.116.304054
  113. T'Jonck W, Williams M, Bonnardel J. Niche signals and transcription factors involved in tissue-resident macrophage development. *Cell Immunol.* (2018) 330:43–53. doi: 10.1016/j.cellimm.2018.02.005
  114. Hynes RO. The extracellular matrix: not just pretty fibrils. *Science.* (2009) 326:1216–9. doi: 10.1126/science.1176009
  115. Pearce OMT, Delaine-Smith RM, Maniati E, Nichols S, Wang J, Bohm S, et al. Deconstruction of a metastatic tumor microenvironment reveals a common matrix response in human cancers. *Cancer Discov.* (2018) 8:304–19. doi: 10.1158/2159-8290.CD-17-0284
  116. Zhang F, Wei K, Slowikowski K, Fonseka CY, Rao DA, Kelly S, et al. Defining inflammatory cell states in rheumatoid arthritis joint synovial tissues by integrating single-cell transcriptomics and mass cytometry. *Nat Immunol.* (2019) 20:928–942. doi: 10.1038/s41590-019-0378-1
  117. Haldar M, Murphy KM. Origin, development, and homeostasis of tissue-resident macrophages. *Immunol Rev.* (2014) 262:25–35. doi: 10.1111/imr.12215
  118. Haldar M, Kohyama M, So AY, Kc W, Wu X, Briseno CG, et al. Heme-mediated SPI-C induction promotes monocyte differentiation into iron-recycling macrophages. *Cell.* (2014) 156:1223–34. doi: 10.1016/j.cell.2014.01.069
  119. Gabanyi I, Muller PA, Feighery L, Oliveira TY, Costa-Pinto FA, Mucida D. Neuro-immune interactions drive tissue programming in intestinal macrophages. *Cell.* (2016) 164:378–91. doi: 10.1016/j.cell.2015.12.023
  120. Muller PA, Kosco B, Rajani GM, Stevanovic K, Berres ML, Hashimoto D, et al. Crosstalk between muscularis macrophages and enteric neurons regulates gastrointestinal motility. *Cell.* (2014) 158:300–13. doi: 10.1016/j.cell.2014.04.050
  121. Hammond TR, Marsh SE, Stevens B. Immune signaling in neurodegeneration. *Immunity.* (2019) 50:955–74. doi: 10.1016/j.immuni.2019.03.016
  122. Hong S, Beja-Glasser VF, Nfonoyim BM, Frouin A, Li S, Ramakrishnan S, et al. Complement and microglia mediate early synapse loss in Alzheimer mouse models. *Science.* (2016) 352:712–6. doi: 10.1126/science.aad8373
  123. Wolf Y, Boura-Halfon S, Cortese N, Haimon Z, Sar Shalom H, Kuperman Y, et al. Brown-adipose-tissue macrophages control tissue innervation and homeostatic energy expenditure. *Nat Immunol.* (2017) 18:665–74. doi: 10.1038/ni.3746
  124. Pirzgalska RM, Seixas E, Seidman JS, Link VM, Sanchez NM, Mahu I, et al. Sympathetic neuron-associated macrophages contribute to obesity by importing and metabolizing norepinephrine. *Nat Med.* (2017) 23:1309–18. doi: 10.1038/nm.4422
  125. Hulsmans M, Clauss S, Xiao L, Aguirre AD, King KR, Hanley A, et al. Macrophages facilitate electrical conduction in the heart. *Cell.* (2017) 169:510–22 e20. doi: 10.1016/j.cell.2017.03.050

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Gordon and Plüddemann. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.



# Sharing Knowledge With Young and Established Students of Immunology by the Neapolitan Gulf at the Ruggero Ceppellini Advanced School

Francesco Colucci<sup>1,2\*</sup>

<sup>1</sup> Department of Obstetrics and Gynaecology, National Institute for Health Research Cambridge Biomedical Research Centre, University of Cambridge School of Clinical Medicine, University of Cambridge, Cambridge, United Kingdom, <sup>2</sup> Centre for Trophoblast Research, University of Cambridge, Cambridge, United Kingdom

**Keywords:** pregnancy, immunity, transplantation, cancer, collaborations

## INTRODUCTION

In his *Origin*, Charles Darwin led the foundations to debunk the long-held belief that man and animals derive from separate lineages, landing the final blow in *The Descent of Man*. The discovery in the mid-1980's that fertilized mammal eggs must have male components to generate healthy offspring had similarly dramatic consequences on other religious beliefs, as discussed in "*Genetics: immaculate misconception*" (1). In the Catholic calendar, the 8th of December is dedicated to the Virgin Mary. The occasion was celebrated with loud fireworks cracking during the second night of the 25th course of the EFIS-EJI Ruggero Ceppellini Advanced School of Immunology, held at Castellammare di Stabia, near Naples, 7th–9th December 2014. A faculty of 13 gathered together with 60 attendants from 19 countries to discuss the theme *Maternal Immune System in Pregnancy*. While the conclusions of the course were not quite as dramatic as Darwin's and Surani's, new exciting concepts were discussed that had already emerged at a previous meeting held in Cambridge in 2013 to celebrate the 60th anniversary of Peter Medawar's famous article on the "immunological paradox" of pregnancy (2). I had the honor of directing both events, together with Ashley Moffett, and learned a great deal.

This brief article is a report on the activities during that 25th course, as well as an opportunity to celebrate the importance of the Ceppellini School to connect young immunologists with leader scientists in their fields, as well as to spur new collaborations. With the generous support of the EFIS-EJI, the Bill and Melinda Gates Foundation, and the International Union of Immunological Societies, a record number of travel fellowships was offered to 13 participants from African countries, including South Africa, Kenya, Nigeria, Gabon, and Cameroon. This was appropriate because it is in Sub-Saharan Africa (SSA) that maternal morbidity and mortality is highest due to pregnancy complications, such as the hypertensive disorder of pregnancy pre-eclampsia, still birth or intrauterine growth restriction (3).

## ACTIVITIES DURING THE 2014 COURSE

On the first day of the course, Silvia Fontana Zappacosta talked about the ethos and history of the School founded by her late husband Serafino Zappacosta. One of the remits of the School is to "foster wider interest for immunology and to attract to the discipline young scientists, also from disadvantaged countries" (4). I introduced the course with a brief synopsis of each lecturer's topic and told the story of my own connection to the Ceppellini School. My late maternal uncle Tommaso (Tommi) Meo trained with Ceppellini himself in the 60's and 70's in Turin

## OPEN ACCESS

### Edited by:

Ennio Carbone,  
University of Catanzaro, Italy

### Reviewed by:

Gerard Chaouat,  
INSERM U976 Immunologie,  
Dermatologie, Oncologie, France

### \*Correspondence:

Francesco Colucci  
fc287@medschl.cam.ac.uk

### Specialty section:

This article was submitted to  
Molecular Innate Immunity,  
a section of the journal  
Frontiers in Immunology

**Received:** 18 November 2019

**Accepted:** 09 January 2020

**Published:** 28 January 2020

### Citation:

Colucci F (2020) Sharing Knowledge With Young and Established Students of Immunology by the Neapolitan Gulf at the Ruggero Ceppellini Advanced School. *Front. Immunol.* 11:43. doi: 10.3389/fimmu.2020.00043



and Basel. Ceppellini made seminal contributions for the advancement of our understanding of immunogenetics (5). Among the factors determining pregnancy outcome are immune system genes—that is combinations of certain variants of genes coding for Human Leukocyte Antigens (HLA) and Killer-cell Immunoglobulin-like Receptors (KIR) (6). How odd that part of my research today was the subject of my uncle's science with Ceppellini and that he so excitedly narrated to us on his summer visits back in our native Southern Italy. John Trowsdale (University of Cambridge) reviewed the “ABC” of KIR and HLA, explaining how the system may have evolved to deploy the two A and B haplotypes that code for KIR receptors on Natural Killer (NK) cells to bind HLA-C on fetal trophoblast cells. Ashley Moffett (University of Cambridge) discussed how the KIR and HLA systems may have evolved and can be used to study population history (7). Annette Nakimuli (Makerere University and Mulago Hospital, Kampala, Uganda) discussed the diverse KIR and HLA genes that cause susceptibility to or protection from pregnancy disorders in Europeans and Africans (8). Allison Elliott (London School of Hygiene and tropical Medicine and Uganda Virus Research Institute, Entebbe, Uganda) presented fascinating data on the impact of helminth infection during pregnancy and the outcomes in the offspring. Angela Santoni (University of Rome La Sapienza) reviewed leukocyte trafficking and the changes occurring during pregnancy. On the second day, Elizabeth Simpson (Imperial College, London, UK) gave a historical background on multiple histocompatibility antigens and how the maternal immune system is aware of fetal antigens yet does not mount an immune response against the fetus. Tamara Tilburgs (Harvard University, Cambridge, US and now at the Cincinnati Children's Hospital, US) discussed the delicate balance that the maternal immune system must strike between fetal tolerance and antiviral immunity. Jakob Michaelsson (Karolinska Institutet, Stockholm, Sweden) reminded the audience that the fetus also has its immune system that may engage with maternal antigens, with consequences on micro-chimerism. Marise Alegre (University of Chicago, US) revised the evidence that tolerance can be induced experimentally to transplants. Anthony De Tomaso (University of California Santa Barbara, US) talked about the strange and fascinating life of a basal chordate that uses allorecognition to regulate stem cell parasitism. In the third and last day, Ennio Carbone (Karolinska Institute, Stockholm, Sweden and University of Catanzaro, Italy) opened the lectures with an overview on tumor immunology. Tom Gajewski (University of Chicago, US) followed up highlighting the immune pathways in the tumor microenvironment that may be operating also at the maternal-fetal interface, with the engagement of several inhibitory checkpoints. The course ended with my closing lecture on mouse models of immunogenetics of pregnancy.

In the typical spirit of the Ceppellini School, the presentations were enriched by ample discussions and debates in which both faculty and students participated actively. The search for elusive pathogenic T cells in pregnancy complications was discussed as it was the antigen specificity of these effector T cells, which most likely are HLA-C-restricted. Another theme was the importance of studying human populations in which the prevalence of

pregnancy complications is highest. New technology that can help visualize lymphocytes at the maternal-fetal interface were discussed, including imaging approaches. Finally, various routes of vertical transmission were considered, including through maternal monocytes and fetal placental macrophages (i.e., Hofbauer cells).

## FOSTERING YOUNG IMMUNOLOGISTS AND FACILITATING COLLABORATIONS

There were plenty of opportunities for the participants to interact among each other and with the faculty members over lunches, coffee breaks, and the poster session. Several collaborations stemmed from this course and continue till today. Both Anthony De Tomaso and Allison Elliott, two of the members of the faculty at this course came to spend a year as visiting Fellows of King's College, Cambridge, where myself and Ashley Moffett are also Fellows. Annette Nakimuli and Ashley Moffett have strengthened their collaboration and have since initiated a series of initiatives within the Cambridge-Africa partnership to improve patients care in the Department of Obstetrics and Gynecology at the Makerere University, including several trips from Cambridge obstetricians to visit Uganda. For example, Catherine Aiken, also at our Department, now mentors Imelda Namagembe's PhD thesis in Uganda, that focuses on improving maternal health.

Despite not present at the course, Stephen Tukwasibwe, then a research assistant in the same hospital of faculty member Annette Nakimuli, became interested in the immunogenetics of pregnancy. Having worked successfully on the genetics of resistance to malaria and secured a Wellcome Trust PhD grant, Stephen started his thesis at Makerere University under the supervision of Annette Nakimuli and my co-mentorship, to test the hypothesis that *Plasmodium* may have selected for those genetic variants that may protect from malaria but expose women to pregnancy complications in SSA. Stephen has since visited Cambridge several times working at the Pathology Department as part of his thesis. One of the participants, Iva Filipovic from Serbia, was completing her MSc degree at Imperial College, London, during the course and was very keen to learn more on immunology of pregnancy. She secured a PhD Studentship from the University of Cambridge Center for Trophoblast Research and came to work on her PhD as a graduate student of King's College and in my laboratory to study the gene expression profile of innate lymphoid cells in the uterus of mice (9). She is currently working as a post-doc at the Karolinska Institute and I look forward to seeing her future successes.

## NEW CONCEPTS AND RECENT PROGRESS IN THE FIELD

Peter Medawar in 1953 famously proposed three mechanisms underlying placental tolerance: (i) anatomical separation of mother and fetus; (ii) antigenic immaturity of the fetus; (iii) immunological unresponsiveness of the mother. Bearing in mind these proposals were formulated in light of the progress made during those days in transplantation immunology, and with

unimaginable less knowledge of the details of the human immune system than we have today, it is perhaps not surprising that none of these three mechanisms have been fully substantiated—although they have influenced generations of immunologists of reproduction. On the contrary, we know that the placenta is not such a tight barrier and cells can mix in both directions. We also know that the fetus is not antigenically immature and the mother is not unresponsive. Indeed, pregnant women can make both T cells and antibodies that recognize fetal antigens (e.g., anti-D antibodies in Rhesus incompatibility).

One major conceptual shift in the immunology of pregnancy is the understanding that pregnant women are not immunosuppressed. Changes in the immune system during pregnancy may however be responsible for the greater morbidity and mortality of mothers and infants infected with certain pathogens (10). The emergence of new epidemics has attracted the attention of investigators who are now addressing the mechanisms of vertical transmission of certain pathogens, e.g., Zika virus (11, 12). That microbes are integral part of human health and disease has become established in the recent past, perhaps best illustrated by the influence of the gut microbiota on the immunotherapy of cancer (13)—one of themes of the 2019 course (*Microbes, Immunity and Cancer*) of the Ceppellini School (14). Transplantation immunology also may be influenced by microbes (15, 16), however the search for a placental microbiome has so far been elusive (17). Yet, maternal infections may have repercussions on neuropsychiatric disorders (18) and the development of the immune system in the offspring. Clinical trials are ongoing to evaluate the effectiveness of vaccinating mothers to prevent children's allergies (19, 20).

There are obvious selective disadvantages in a strategy that would suppress the immune system of pregnant women to allow the implantation and growth of the placenta. The placenta evolved much later than the immune system and it is reasonable to think that placentation and immunity have co-evolved agreeably, rather than embarking in a deleterious conflict. One illustrative example may be the interactions of maternal KIR on uterine NK cells with fetal HLA-C molecules on the placental cells, which may engage in a molecular

conversation that, rather than leading to allorecognition-driven rejection, may in fact contribute to uterine vascular remodeling and placental growth (6). Adding to the complexity of the maternal-fetal interactions is the heterogeneity of immune cells, revealed recently by single-cell RNA-sequencing (21) and mass cytometry (22). Mass cytometry has been applied to study also the fluctuations in blood immune cells throughout pregnancy (23, 24). Multiple populations of innate lymphoid cells (9, 21, 25), regulatory T cells (26), and macrophages (27) compose the diverse immune cell landscape operating at the maternal-fetal interface, which varies during the stages of pregnancy and it is therefore difficult to decipher precisely. New technology such as three-dimensional organoid cell cultures (28) may help to determine some of the mechanisms underlying placentation (29). Advances in typing polymorphic KIR and HLA genes (30, 31) may also help to shed light on the immunogenetics of pregnancy. Although the interactions of maternal KIR with fetal HLA-C may be a pivotal one to activate uterine NK cells and determine the outcome of pregnancy (6), the importance of the interaction of NK cell receptors with self HLA class I molecules is emerging, in a process known as NK-cell education. We have shown recently that NK-cell education in the uterus may follow different rules than in the blood (32) and that NK-cell education reduces the risk of pregnancy complications in women genetically programmed to engage the inhibitory NKG2A receptor on NK cells (33). The next grand challenge is to precisely decipher the multiple and changing interactions between mother and fetus in the decidua, to eventually manipulate them in order to improve the outcome of pregnancy (29).

## AUTHOR CONTRIBUTIONS

The author confirms being the sole contributor of this work and has approved it for publication.

## FUNDING

Work in the Colucci laboratory is funded by the Wellcome Trust (Grant Number 200841/Z/16/Z).

## REFERENCES

- Surani MA. Genetics: immaculate misconception. *Nature*. (2002) 416:491–3. doi: 10.1038/416491a
- Colucci F, Moffett A, Trowsdale J. Medawar and the immunological paradox of pregnancy: 60 years on. *Eur J Immunol*. (2014) 44:1883–5. doi: 10.1002/eji.201470065
- Nakimuli A, Chazara O, Byamugisha J, Elliott AM, Kaleebu P, Mirembe F, Moffett A. Pregnancy, parturition and preeclampsia in women of African ancestry. *Am J Obstet Gynecol*. (2014) 210:510–20.e1. doi: 10.1016/j.ajog.2013.10.879
- Di Giacomo A. The Ruggero Ceppellini advanced school of immunology and the neapolitan scientific renaissance. *Front Immunol*. (2019) 10:1494. doi: 10.3389/fimmu.2019.01494
- Bodmer W. Ruggero Ceppellini: a perspective on his contributions to genetics and immunology. *Front Immunol*. (2019) 10:1280. doi: 10.3389/fimmu.2019.01280
- Moffett A, Colucci F. Co-evolution of NK receptors and HLA ligands in humans is driven by reproduction. *Immunol Rev*. (2015) 267:283–97. doi: 10.1111/imr.12323
- Parham P, Moffett A. Variable NK cell receptors and their MHC class I ligands in immunity, reproduction and human evolution. *Nat Rev Immunol*. (2013) 13:133–44. doi: 10.1038/nri3370
- Nakimuli A, Chazara O, Hiby SE, Farrell L, Tukwasibwe S, Jayaraman J, et al. A KIR B centromeric region present in Africans but not Europeans protects pregnant women from pre-eclampsia. *Proc Natl Acad Sci USA*. (2015) 112:845–50. doi: 10.1073/pnas.1413453112
- Filipovic I, Chiossone L, Vacca P, Hamilton RS, Inegner T, Doisne JM, et al. Molecular definition of group 1 innate lymphoid cells in the mouse uterus. *Nat Commun*. (2018) 9:4492. doi: 10.1038/s41467-018-06918-3
- Kourtis AP, Read JS, Jamieson DJ. Pregnancy and infection. *N Engl J Med*. (2014) 371:1077. doi: 10.1056/NEJMr1213566
- Musso D, Ko AI, Baud D. Zika virus infection - after the pandemic. *N Engl J Med*. (2019) 381:1444–57. doi: 10.1056/NEJMr1808246

12. Pierson TC, Diamond MS. The emergence of Zika virus and its new clinical syndromes. *Nature*. (2018) 560:573–81. doi: 10.1038/s41586-018-0446-y
13. Zitvogel L, Ma Y, Raoult D, Kroemer G, Gajewski TF. The microbiome in cancer immunotherapy: diagnostic tools and therapeutic strategies. *Science*. (2018) 359:1366–70. doi: 10.1126/science.aar6918
14. Colucci F, Carbone E. Microbes, immunity and cancer in capri – another successful course of the EFIS-EJI Ruggero Ceppellini advanced school of immunology founded by Serafino Zappacosta. *Eur J Immunol*. (2019) 49:2123–6. doi: 10.1002/eji.201970125
15. Lei YM, Chen L, Wang Y, Stefká AT, Molinero LL, Theriault B, et al. The composition of the microbiota modulates allograft rejection. *J Clin Invest*. (2016) 126:2736–44. doi: 10.1172/JCI85295
16. McIntosh CM, Chen L, Shaiber A, Eren AM, Alegre ML. Gut microbes contribute to variation in solid organ transplant outcomes in mice. *Microbiome*. (2018) 6:96. doi: 10.1186/s40168-018-0474-8
17. de Goffau MC, Lager S, Sovio U, Gaccioli F, Cook E, Peacock SJ, et al. Human placenta has no microbiome but can contain potential pathogens. *Nature*. (2019) 572:329–34. doi: 10.1038/s41586-019-1451-5
18. Estes ML, McAllister AK. Maternal immune activation: implications for neuropsychiatric disorders. *Science*. (2016) 353:772–7. doi: 10.1126/science.aag3194
19. Luty AJ, Elliott AM. Tackling neglect: treating schistosomiasis in pregnancy. *Lancet Infect Dis*. (2016) 16:137–9. doi: 10.1016/S1473-3099(15)00379-5
20. Namara B, Nash S, Lule SA, Akurut H, Mpairwe H, Akello F, et al. Effects of treating helminths during pregnancy and early childhood on risk of allergy-related outcomes: follow-up of a randomized controlled trial. *Pediatr Allergy Immunol*. (2017) 28:784–92. doi: 10.1111/pai.12804
21. Vento-Tormo R, Efremova M, Botting RA, Turco MY, Vento-Tormo M, Meyer KB, et al. Single-cell reconstruction of the early maternal-fetal interface in humans. *Nature*. (2018) 563:347–53. doi: 10.1038/s41586-018-0698-6
22. Huhn O, Ivarsson MA, Gardner L, Hollinshead MS, Stinchcombe JC, Chen P, et al. Distinctive phenotypes and functions of innate lymphoid cells in human decidua during early pregnancy. *Nat Commun*. (2019). doi: 10.17863/CAM.47154. [Epub ahead of print].
23. Aghaeepour N, Ganio EA, McIlwain D, Tsai AS, Tingle M, Van Gassen S, et al. An immune clock of human pregnancy. *Sci Immunol*. (2017) 2:eaan2946. doi: 10.1126/sciimmunol.aan2946
24. Le Gars M, Seiler C, Kay AW, Bayless NL, Starosvetsky E, Moore L, et al. Pregnancy-induced alterations in NK cell phenotype and function. *Front Immunol*. (2019) 10:2469. doi: 10.3389/fimmu.2019.02469
25. Doisne JM, Balmas E, Boulenouar S, Gaynor LM, Kieckbusch J, Gardner L, et al. Composition, development, and function of uterine innate lymphoid cells. *J Immunol*. (2015) 195:3937–45. doi: 10.4049/jimmunol.1500689
26. Salvany-Celades M, van der Zwan A, Benner M, Setrajic-Dragos V, Bougleux Gomes HA, Iyer V, et al. Three types of functional regulatory T cells control T cell responses at the human maternal-fetal interface. *Cell Rep*. (2019) 27:2537–47.e5. doi: 10.1016/j.celrep.2019.04.109
27. Jiang X, Du MR, Li M, Wang H. Three macrophage subsets are identified in the uterus during early human pregnancy. *Cell Mol Immunol*. (2018) 15:1027–37. doi: 10.1038/s41423-018-0008-0
28. Turco MY, Gardner L, Kay RG, Hamilton RS, Prater M, Hollinshead MS, et al. Trophoblast organoids as a model for maternal-fetal interactions during human placentalation. *Nature*. (2018) 564:263–7. doi: 10.1038/s41586-018-0753-3
29. Colucci F. The immunological code of pregnancy. *Science*. (2019) 365:862–3. doi: 10.1126/science.aaw1300
30. Jiang W, Johnson C, Simecek N, Lopez-Alvarez MR, Di D, Trowsdale J, et al. qKAT: a high-throughput qPCR method for KIR gene copy number and haplotype determination. *Genome Med*. (2016) 8:99. doi: 10.1186/s13073-016-0358-0
31. Norman PJ, Hollenbach JA, Nemat-Gorgani N, Marin WM, Norberg SJ, Ashouri E, et al. Defining KIR and HLA class I genotypes at highest resolution via high-throughput sequencing. *Am J Hum Genet*. (2016) 99:375–91. doi: 10.1016/j.ajhg.2016.06.023
32. Sharkey AM, Xiong S, Kennedy PR, Gardner L, Farrell LE, Chazara O, et al. Tissue-specific education of decidual NK cells. *J Immunol*. (2015) 195:3026–32. doi: 10.4049/jimmunol.1501229
33. Shreeve N, Traherne JA, Sovio U, Hawkes DA, Huhn O, Jayaraman J, et al. NKG2A educates uterine NK cells to optimise pregnancy in humans and mice. *Immunity*. (2019). doi: 10.2139/ssrn.3477575. [Epub ahead of print].

**Conflict of Interest:** The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2020 Colucci. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.



# Recirculation and Residency of T Cells and Tregs: Lessons Learnt in Anacapri

Silvia Piconese<sup>1,2</sup>, Silvia Campello<sup>3</sup> and Ambra Natalini<sup>4,5\*</sup>

<sup>1</sup> Dipartimento di Scienze Cliniche Internistiche, Anestesiologiche e Cardiovascolari, Sapienza Università di Roma, Rome, Italy, <sup>2</sup> Laboratory Affiliated to Istituto Pasteur Italia – Fondazione Cenci Bolognetti, Rome, Italy, <sup>3</sup> Department of Biology, University of Rome Tor Vergata, Rome, Italy, <sup>4</sup> Institute of Molecular Biology and Pathology, National Research Council (CNR), Rome, Italy, <sup>5</sup> Dipartimento di Medicina Molecolare (DMM), Sapienza Università di Roma, Rome, Italy

## OPEN ACCESS

### Edited by:

Francesca Granucci,  
University of Milano-Bicocca, Italy

### Reviewed by:

Clémence Granier,  
Assistance Publique Hôpitaux De  
Paris, France  
Claudia Ida Brodskyn,  
Gonçalo Moniz Institute (IGM), Brazil

### \*Correspondence:

Ambra Natalini  
ambra.natalini@uniroma1.it

### Specialty section:

This article was submitted to  
Molecular Innate Immunity,  
a section of the journal  
Frontiers in Immunology

**Received:** 10 January 2020

**Accepted:** 26 March 2020

**Published:** 05 May 2020

### Citation:

Piconese S, Campello S and  
Natalini A (2020) Recirculation  
and Residency of T Cells and Tregs:  
Lessons Learnt in Anacapri.  
Front. Immunol. 11:682.  
doi: 10.3389/fimmu.2020.00682

“Location, location, and location”: according to this mantra, the place where living beings settle has a key impact on the success of their activities; in turn, the living beings can, in many ways, modify their environment. This idea has now become more and more true for T cells. The ability of T cells to recirculate throughout blood or lymph, or to stably reside in certain tissues, turned out to determine immunity to pathogens, and tumors. If location matters also for human beings, the inspiring environment of Capri Island has contributed to the success of the EFIS-EJI Ruggero Ceppellini Advanced School of Immunology focused on “T cell memory,” held in Anacapri from October 12, 2018 to October 15, 2018. In this minireview, we would like to highlight some novel concepts about T cell migration and residency and discuss their implications in relation to recent advances in the field, including the mechanisms regulating compartmentalization and cell cycle entry of T cells during activation, the role of mitochondrial metabolism in T cell movement, and the residency of regulatory T cells.

**Keywords:** T cells, Tregs, cell migration, cell cycle, recirculation

## INTRODUCTION

This minireview is inspired by the EFIS-EJI Ruggero Ceppellini Advanced School of Immunology about “T cell memory” 2018 (1) and will expand in further detail two hot topics discussed during the course: T cell migration and residency.

T cell differentiation and function are strictly related to their distribution within different lymphoid and non-lymphoid compartments. In physiological conditions, naive T cells recirculate through secondary lymphoid organs (SLOs), increasing the opportunity to encounter the antigen. After infection, vaccination, or tumor growth, the draining lymphoid compartments undergo dramatic changes, promoting naive T cells’ interaction with antigen-presenting cells and subsequent T cell activation. Activated T cells undergo a strong proliferation (so-called clonal expansion) and deep changes in their metabolism (2, 3). The process culminates with T cell differentiation and the generation of short-lived effectors and long-lived memory cells (4–6). Effector T cells migrate broadly, reaching the site of infection or tumor growth where they exert their effector functions before dying. Memory cells persist in the body, circulating between blood and lymphoid or non-lymphoid tissues as conventional memory T cells, or residing in peripheral tissues as resident memory T cells (Trm) (7). Trm represent a first-line defense against tissue



damage and pathogen invasion (8, 9). However, the functional distinction between Trm and conventional effector/memory T cells needs to be clarified. Moreover, it is now clear that some technical caveats may hinder an appropriate and complete analysis of these cells (10). A better understanding of the immunological and metabolic signals dictating the switch between T cell recirculation and residency is needed. Here, we will focus on some emerging concepts regarding this topic: first, the relation between the cell cycle phase and migration during T cell activation; second, the role of mitochondria relocation for T cell movements and compartmentalization; finally, the features of residency of a well-known tissue-infiltrating T cell population, i.e., the regulatory T cells (Tregs).

## T CELL RECIRCULATION AND CELL CYCLE

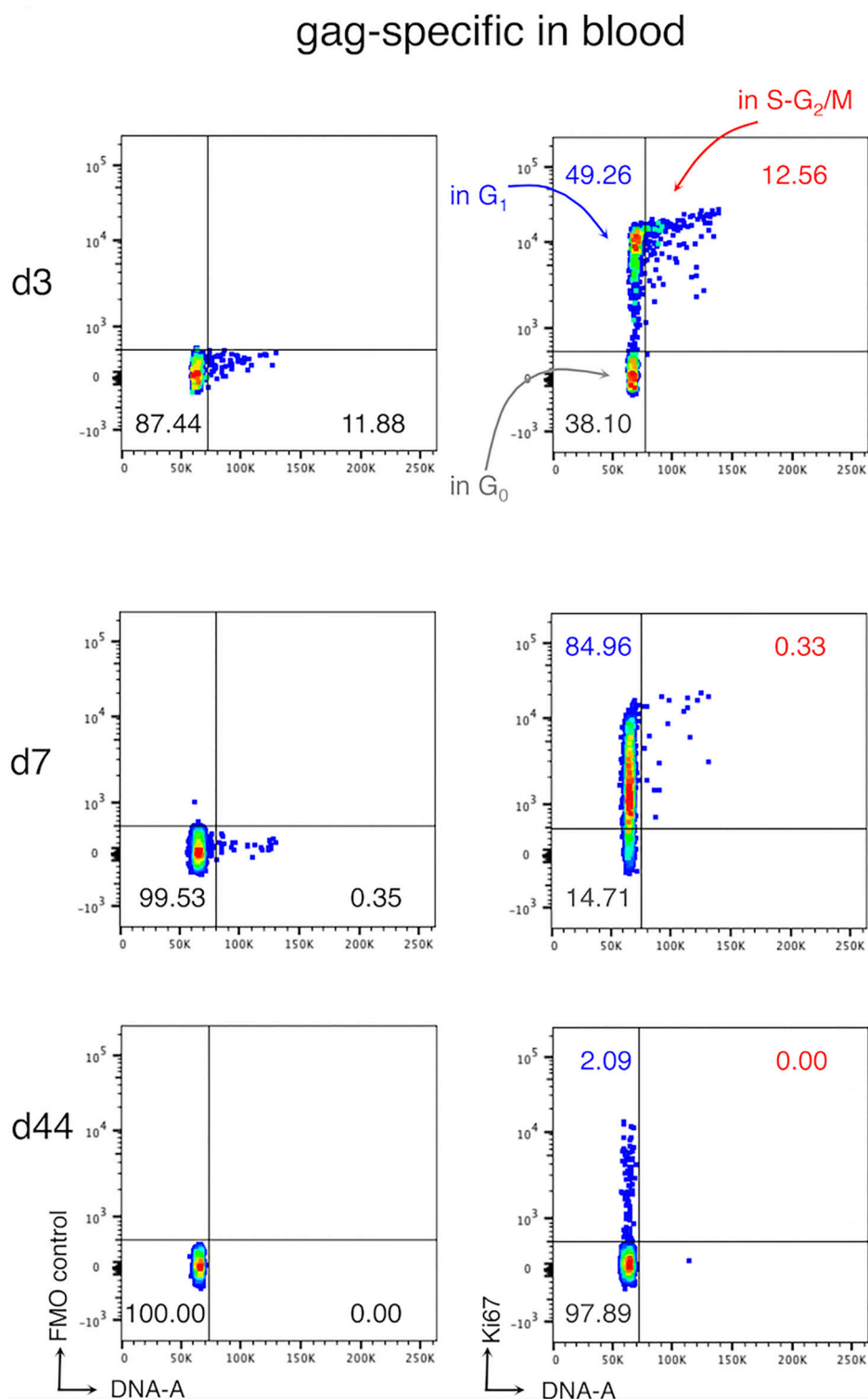
After development in the thymus, naive T cells reach the blood circulation, and continuously circulate between blood and SLOs. This journey is finely regulated by the expression of specific homing molecules. Indeed, the L-selectin CD62L expressed by naive T cells mediates their entry into lymph nodes (LNs) by binding ligands expressed on high endothelial venules (HEVs). This binding overcomes blood shear forces, leading to T cell rolling on HEVs (11). At this stage, the interaction between the CC chemokine ligand 21 (CCL21) expressed on HEVs and the CC chemokine receptor 7 (CCR7) on T cells activates the integrin lymphocyte function-associated antigen 1 (LFA1). Activated LFA1 binds the intracellular adhesion molecule 1 (ICAM-1), mediating T cell arrest on the endothelium. Consequently, T cells migrate across the blood vessels and enter the tissue (12). Once in the LN, naive T cells are guided in the paracortical region, also known as T cell zone. In this area, naive T cells interact with dendritic cells (DCs), scanning for the presence of the cognate antigen. It has been estimated that one DC can be scanned simultaneously by up to 500 naive T cells (13, 14). Migration in this area is regulated by a gradient of chemokines and local factors. The chemokine CCL19, produced within the T cell zone, increases T cell motility and promotes T cell–DC interactions by binding CCR7 on the T cell surface (15). Furthermore, after immunization, naive CD8 T cells upregulate CCR5, which binds CCL3 and CCL4 produced at the site of the CD4 T cell–DC interaction in the immunogen-draining LNs (16).

Hence, migration in the SLOs seems to be not only a stochastic process but rather a finely regulated mechanism which increases the probability of antigen recognition. In the case that this rare event occurs, T cells undergo a series of dramatic changes. Resting naive T cells are activated by the integration of three signals: antigen recognition (signal 1), co-stimulation (signal 2), and cytokines, released at the site of T cell–DC interaction (signal 3) (17). This process culminates with the extensive proliferation of antigen-specific T cells, named clonal expansion. T cell expansion is driven by T cell–DC interaction within specialized niches in SLOs and is controlled by several factors which promote the rapid entry of T cell in the cell cycle (18–20). The final goal of this process is to increase the number of

T cells capable of eliminating the antigen. It has been estimated that, in the first week of a typical primary T cell response, CD8 T cells can increase their number to about 100 times or more (21). At this point, deregulation of the cell cycle could deeply affect the ability to develop a proper T cell response. For example, a reduced clonal expansion could lead to a decreased number of effector and memory T cells, with consequent loss of protection. Furthermore, it has been hypothesized that the inability to mount an effective primary T cell response in old age and the vaccination failure occurring in elderly persons could be correlated with defects of T cell clonal expansion (22, 23).

Expanding T cells modulate the expression of homing molecules, preparing themselves to reach the peripheral tissue, the site of antigen entry. Retention in SLOs is controlled by the sphingosine-1-phosphate (S1P) receptor expression on T cells. S1P is a lipid molecule that is more concentrated in the blood and in the lymph than in tissues (24). S1P receptor expression is increased in naive T cells, leading to egress from SLOs. Activated T cells upregulate CD69, which prevents S1P receptor expression, holding T cells in the SLOs until the completion of differentiation into effector cells, which can take a few days (25). Once completely differentiated, effector T cells downregulate CD69, and migrate along the S1P gradient. Effector T cells also downregulate CD62L and express chemokine receptors that guide them to the site of infection (26).

The kinetic of expansion and migration is poorly defined. Indeed, although it is known that clonal expansion starts in SLOs, the location where activated T cells progress and/or complete their cell cycle is still unclear. To date, the few tools available for the analysis of dividing antigen-specific CD8 T cells, such as cell-labeling dyes and anti-Ki67 antibody, show some important limitations. Indeed, cell-labeling dyes do not allow evaluating whether cells found in one organ proliferated locally or rather migrated in this organ after division (19, 27). Ki67 is a nuclear protein expressed by cells in all the phases of the cell cycle (G1, S, G2, and M), except for those in G0 (or quiescent). Hence, Ki67 analysis alone does not distinguish proliferating cells (in S-G2-M) from those in G1, which may remain for a long time in G1, or even revert to G0 (or quiescent) without dividing (28, 29). We recently set up a new flow cytometric method for the cell cycle analysis of CD8 T cells, which was based on the combination of Ki67 expression and DNA content analyses and allowed us to discriminate between cells in the G0, G1, and S-G2/M phases. By using this method together with a novel gating strategy for the analysis of actively responding T cells, we demonstrated that, at early times after vaccination in mice, cycling antigen-specific CD8 T cells (cells in the S-G2-M phases) were present in the blood, which is usually not considered a site of proliferation (**Figure 1**) (30). This finding questions the general view by which activated T cells proliferate locally in SLOs and only after completing their cell cycle and differentiation enter the blood circulation, reaching the infection site. In addition, studies on cancer patients have shown that antitumor CD8 T cells increase Ki67 expression after checkpoint inhibitor treatment, suggesting that unleashed T cells can actively cycle in the blood after therapy (31, 32).



**FIGURE 1 |** Cell cycle analysis of antigen-specific CD8 T cells in the blood after vaccination. Female Balb/c mice were primed and boosted with viral vectors expressing the model antigen gag of HIV-1. At days (d) 3, 7, and 44, post-boost blood was collected and blood cells were analyzed with our new method. The figure shows a typical ki67/DNA staining profile of gag-specific CD8 T cells in the blood. Fluorescence Minus One (FMO) controls (*left*) and Ki67 staining (*right*) are shown, as indicated; the *numbers* represent the percentages of cells in the corresponding quadrant. Figure adapted from (30).

## MITOCHONDRIAL DYNAMICS IN MEMORY T CELLS AND T CELL MIGRATION

In the past, immunologists did not take seriously into account T cell mitochondria since they are poorly represented within a T cell, and T cells are mainly considered as relying on glycolysis for their principal functions. In recent decades, a large body of evidence emerged on the crucial role that the mitochondria, their metabolism, and their morphological dynamics have on these cells. Nowadays, the pivotal role of mitochondrial morphology changes in almost all processes that are essential for a correct T cell development and function is clear and evident (33). Thus, these less attractive organelles suddenly became “main characters” for several immunologists in recent years.

Mitochondria, the cellular energetic hubs, are highly motile organelles, continuously fusing and fragmenting (a.k.a. fission) their network under the control of the so-called mitochondria-shaping proteins (34) (**Figure 2**). Drp1 and Dyn2 are the main players controlling fission in concert (35), while mitofusins 1 and 2 and Opa1 are the principal proteins orchestrating mitochondria fusion (36, 37). The balance between these opposing events, at every time or cell demand, determines organelle morphology, which acts as an intracellular signal that instructs different metabolic pathways, reflecting the different physiological functions of the cell. For instance, an elongated network sustains oxidative phosphorylation (OXPHOS) for a correct assembly of the electron transport chain (ETC) complexes, and an optimal ATP production, besides diluting the matrix content (38). A fragmented network, instead, promotes aerobic glycolysis and mitophagy or accelerates cell proliferation in response to nutrient excess and cellular dysfunction (38). Mitochondrial morphology directly regulates T cell differentiation *in vitro* by affecting the engagement of these alternative metabolic routes upon activation. Mitochondrial fusion-dependent fatty acid oxidation with a predominance of OXPHOS is a hallmark of a memory cell signature, while an effector cell subtype mostly relies on fission-dependent

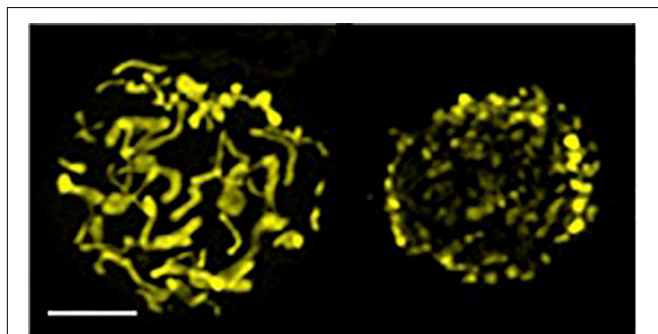
glycolysis (39, 40). Thus, mitochondrial dynamics controls T cell fate. Evidence *in vivo* of these findings, together with the molecular mechanisms explaining how mitochondrial dynamics can orchestrate these metabolic shifts and T cell fate, came soon after. Indeed, our lab showed that mitochondrial fragmentation, favoring glycolysis in effector T cells, is dependent on the Erk1-mediated activation of Drp1. Further and interestingly, an additional—but not mutually exclusive—transcriptional mechanism sustains the metabolic shifts in T cell differentiation. Upon T cell receptor (TCR) engagement, in T cells with an elongated mitochondria, the extracellular calcium uptake is exacerbated [presumably because of an inability of the unfragmented mitochondria to reach the immunological synapse and to buffer calcium (41)], this leading to alterations on the mTOR–cMyc axis, decrease of cMyc expression, and related defective transcription of glycolytic enzymes, cMyc being known as a promoting factor in the transcription of glycolytic enzymes upon T cell activation (42). The consequence is a prominent oxidative metabolism and a memory-like phenotype for these T cells (43). Thus, in sum, memory T cell differentiation is driven by ERK1- and cMyc-dependent mitochondria morphological changes.

More interestingly, for this review's purpose, the capability of memory T cells to reach the tissues and being resident, rather than to recirculate in the periphery, crucially relies on the ability of these cells to (trans)migrate and extravasate into and from the blood vessels. These basic processes also strictly depend on mitochondrial dynamics. Polarized T cells need to accumulate their mitochondria at the uropod during migration, to fuel the ATP-consuming myosin II cell motor. Drp1-dependent fragmentation of the mitochondria is essential to allow this organelle relocation, while unbalancing the morphology toward an elongated mitochondrial network strongly impairs T cell chemotaxis (44). *In vivo* extravasation and invasion of T cells are regulated likewise. During their trans-migration across an endothelial layer, lymphocytes squeeze and insert their nuclei into a subendothelial pseudopodium (45), a process heavily relying on the activity of the myosin motor (46) and requiring Drp1-dependent mitochondria fragmentation (43). Consistently, *in vivo* Drp1 removal from T cells inhibits their extravasation from the blood toward SLOs, and toward “danger sites” (43).

Noteworthy is that Drp1 knockout (KO) T cells are deficient in cell migration, even though their metabolism is shifted toward an OXPHOS-based metabolism, ideally producing more ATP to fuel the myosin II, which should drive a higher migration rate. This apparent paradox underlines the cell's need to better modulate the relocation of the mitochondria for a local, subcellular production of mitochondrial ATP rather than for a general mitochondria functionality.

Overall, these findings shed light on a new and crucial role for mitochondrial dynamics in T cell differentiation and function, paving the way for new, and important therapeutic opportunities through pharmacological or genetic manipulation of mitochondria-shaping proteins, also based on memory T cells.

It needs to be considered that forcing mitochondrial fusion during *in vitro* T cell expansion promotes the differentiation of naive T cells toward a memory phenotype, this conferring



**FIGURE 2 |** Elongated and fragmented mitochondria morphology in T cells. Confocal z-stack acquisition and 2D reconstruction of an elongated (*left*) or fragmented (*fissed, right*) mitochondrial network of Jurkat single cells transfected with mtYFP (scale bar, 5  $\mu$ m). Picture modified from (34).

a higher survival to these cells. However, we observed that T cell migration strictly depends on optimal fragmentation of the mitochondrial network; thus, an unbalance toward mitochondria fusion in memory T cells would inhibit their (trans)migratory capability, therefore impinging on their “choice” to be resident or to recirculate. This observation suggests that a one-way or “chronical” modulation of the activity of mitochondria-shaping proteins could hardly result in successful therapeutic strategies, with this highlighting the actual complexity of the topic. Finally, also in a T cell terminal differentiation into senescence, in which cell migration and proliferation are fatally altered, mitochondria structure, and function result impaired as well (47).

## TISSUE REGULATORY T CELLS: RESIDENT OR RECIRCULATING?

Most of the available information about resident T cells come from the study of CD8 Trm, and a growing body of data demonstrates their key role in response to pathogens, in antitumor immunity, in mucosal defense, in vaccine efficacy, and so forth [reviewed in (10)]. Less clear are the identity and functions of CD4 Trm in different contexts, probably because in tissues the CD4 T cell population may comprise variable proportions of Tregs displaying completely different immune functions. Tregs represent a class of CD4 T cells defined by the expression of Foxp3 and exerting non-redundant immunosuppressive and tissue repair functions. In several non-lymphoid tissues, Treg subtypes have been identified that show tissue-specific profiles, differentiate locally in response to variable signals, and perform specialized functions [reviewed in (48)].

Whether tissue Tregs are truly resident cells is still a matter of investigation. Parabiosis experiments have demonstrated that Treg chimerism was lower in the adipose tissue and intestine compared to the spleen, blood, and liver (49–51). When Tregs were further classified into central or effector cells, the latter were found more resistant to recirculation (52, 53); however, this event was transient (52), and upon parabiosis disconnection, the chimerism of both effector and central Tregs decayed in a few weeks (52). These results suggest that, at least in certain tissues, effector Tregs may be continuously replenished from circulating Tregs, which locally differentiate and proliferate (54).

When effector Tregs were further subdivided according to the expression of the CD49b integrin, it was possible to distinguish circulating Tregs: indeed, compared to other districts, the blood and highly vascularized tissues (liver and lung) contained a high frequency of CD49b<sup>+</sup> effector Tregs that displayed a significantly higher rate of exchange between parabiotic mice (55). It could be hypothesized that CD49b<sup>+</sup> Tregs may be devoted to continuous tissue patrolling through blood circulation, being able to promptly reach damaged or inflamed tissues (55), while the CD49b<sup>−</sup> cells may show a certain degree of stable residency and exert on-site repair/regenerative functions in physiological settings. For instance, Tregs localize to the epithelial stem cell niche and promote hair growth at the steady state (56). Resident Tregs may exist in the heart protecting from fortuitous inflammation and tissue damage (57). Such tiny and highly

specialized Treg populations are settled in locations that are poorly accessible to the circulation and, thus, probably may have acquired better capacities to survive and self-renew locally.

Tregs, or certain Treg subsets, share with Trm some phenotypical markers. For instance, Tregs express CD69 at a higher level in non-lymphoid than in lymphoid tissues (58–60). The expression of CD103 by effector Tregs was established several years ago (61), and CD103<sup>+</sup> Tregs have been observed at the steady state in several tissues including the lung (58) and the dermis (62). CD39 is a well-recognized marker of Tregs from lymphoid organs (63) and maintained at high levels in tissues like VAT (64). Notably, one of the key transcription factors for the acquisition of a residency program, Blimp1 (65), plays a well-recognized function in the instruction of the effector program in Treg (66). Therefore, in tissues, effector Tregs possess the whole armamentarium that may be needed to establish residency. In this context, a recent paper has shown that the majority of lung-resident CD4 T cells are indeed composed of Tregs that play tissue-protective functions (58).

More elusive is the extent of Treg residency in human tissues. Tregs can be found in several healthy human tissues such as the intestine, skin, adipose tissue, and skeletal muscle (48). In healthy human skin, arginase 2 expression was found as a feature of resident Tregs (67). Whether Tregs can establish long-term residency in these tissues and whether this process may be modified in pathologic conditions remain unclear. Recent analyses in human lung transplant recipients have demonstrated that, contrary to conventional T cells, most Tregs in the bronchoalveolar lavage were of recipient origin (68): this result underscores the dominance of Treg colonization from the blood over persistent Treg residency, at least in this context. According to the mouse data mentioned above (55), it could be suggested that the lung, as a highly vascularized tissue, may be particularly prone to Treg replenishment from the blood and that Treg residency may be more stringent in less vascularized tissues.

The balance between Treg residency and recirculation may have key implications during tissue modifications occurring in chronic inflammation and cancer. Tumor Tregs display a gene signature that combines tissue-specific and tumor-specific genes [reviewed in (69)], and a “core signature” is shared among Tregs infiltrating diverse human cancers (70). In human melanoma, Tregs express a higher level of arginase 2 than in healthy skin (67), suggesting that tumor Tregs may co-opt and enforce signals that preexisted in Tregs resident in the normal parenchyma. In human breast cancer and colon cancer, tumor Tregs were much more similar to the corresponding healthy tissue Tregs than to circulating Tregs (71, 72). However, the analysis of the TCR repertoire of tumor and tissue Tregs led to conflicting results in different tumor types (70–72), and whether tumor Tregs derive from the amplification of Treg clones populating normal tissues, rather than from circulating cells, remains to be ascertained. A deeper understanding of the tumor Treg complexity will be key to designing Treg-targeted therapies that would spare physiological functions of tissue Tregs.



## DISCUSSION

T cell heterogeneity comprises not only a great variety of T cell subpopulations with different functions but also a considerable diversity of migratory patterns. These patterns are strongly related to the function that these cells will exert in a specific tissue. After activation, changes in T cell migratory capacity occur simultaneously with cell expansion and differentiation into effectors and memory cells. Noteworthy is the evidence that cycling antigen-specific T cells are present in the blood in the acute phase of the response, suggesting a very dynamic interplay between cell cycle and migration (30–32). Nevertheless, how clonal expansion and migration are related is still unclear. Interestingly, the elderly show an altered T cell clonal expansion and a worse T cell response to infections and vaccination. However, only a few studies have focused on the possible impact of aging on T cell recirculation (73, 74), and a possible relation is still unclear.

Whether T cells recirculate or reside in one tissue strongly depends on their metabolism: indeed, mitochondrial dynamics regulate T cell migration and differentiation (39, 40, 44). Metabolism could also dictate the survival of certain Trm, i.e., resident Tregs, which exert important tissue homeostatic

functions (48). However, in some pathological conditions such as tumors, whether infiltrating Tregs derive from the resident population or are mobilized from the circulating pool remains unclear (70, 72). This review highlights novel concepts of T cell compartmentalization and opens new interesting perspectives regarding the regulation of this process both in physiological and in pathological conditions.

## AUTHOR CONTRIBUTIONS

SP conceived the review structure. SC prepared the figure. All authors wrote the manuscript.

## FUNDING

This work was supported by the Associazione Italiana per la Ricerca sul Cancro Grant IG-2017 19784 to SP, and Grant IG-2017 19826 to SC, Ministry of Education, University and Research (MIUR), Progetti di Ricerca di Interesse Nazionale (PRIN) Grant 2017 Prot. 2017K7FSYB to SP, and Istituto Pasteur Italia-Fondazione Cenci Bolognietti Call 2019 under 45 to SP.

## REFERENCES

- Natalini A, Fusco C, Micillo T, Di Rosa F. T cell memory in capri: a successful course organized by the EFIS-EJI ruggero ceppellini advanced school of immunology founded by serafino zappacosta. *Eur J Immunol.* (2019) 49:361–3. doi: 10.1002/eji.201970035
- Jones RG, Thompson CB. Revving the engine: signal transduction fuels T cell activation. *Immunity.* (2007) 27:173–8. doi: 10.1016/j.immuni.2007.07.008
- Murali-Krishna K, Altman JD, Suresh M, Sourdive DJ, Zajac AJ, Miller JD, et al. Counting antigen-specific CD8 T cells: a reevaluation of bystander activation during viral infection. *Immunity.* (1998) 8:177–87. doi: 10.1016/S1074-7613(00)80470-7
- Gasper DJ, Tejera MM, Suresh M. CD4 T-cell memory generation and maintenance. *Crit Rev Immunol.* (2014) 34:121–46. doi: 10.1615/critrevimmunol.2014010373
- Harty JT, Badovinac VP. Shaping and reshaping CD8+ T-cell memory. *Nat Rev Immunol.* (2008) 8:107–19. doi: 10.1038/nri2251
- Kalia V, Sarkar S, Ahmed R. CD8 T-cell memory differentiation during acute and chronic viral infections. *Adv Exp Med Biol.* (2010) 684:79–95. doi: 10.1007/978-1-4419-6451-9\_7
- Sathaliyawala T, Kubota M, Yudanin N, Turner D, Camp P, Thome JJ, et al. Distribution and compartmentalization of human circulating and tissue-resident memory T cell subsets. *Immunity.* (2013) 38:187–97. doi: 10.1016/j.immuni.2012.09.020
- Gebhardt T, Wakim LM, Eidsmo L, Reading PC, Heath WR, Carbone FR. Memory T cells in nonlymphoid tissue that provide enhanced local immunity during infection with herpes simplex virus. *Nat Immunol.* (2009) 10:524–30. doi: 10.1038/ni.1718
- Schenkel JM, Fraser KA, Beura LK, Pauken KE, Vezys V, Masopust D. T cell memory. Resident memory CD8 T cells trigger protective innate and adaptive immune responses. *Science.* (2014) 346:98–101. doi: 10.1126/science.1254536
- Masopust D, Soerens AG. Tissue-resident T cells and other resident leukocytes. *Annu Rev Immunol.* (2019) 37:521–46. doi: 10.1146/annurev-immunol-042617-053214
- Gallatin WM, Weissman IL, Butcher EC. A cell-surface molecule involved in organ-specific homing of lymphocytes. *Nature.* (1983) 304:30–4. doi: 10.1038/304030a0
- Walling BL, Kim M. LFA-1 in T cell migration and differentiation. *Front Immunol.* (2018) 9:952. doi: 10.3389/fimmu.2018.00952
- Bousso P, Robey E. Dynamics of CD8+ T cell priming by dendritic cells in intact lymph nodes. *Nat Immunol.* (2003) 4:579–85. doi: 10.1038/ni928
- Miller MJ, Hejazi AS, Wei SH, Cahalan MD, Parker I. T cell repertoire scanning is promoted by dynamic dendritic cell behavior and random T cell motility in the lymph node. *Proc Natl Acad Sci USA.* (2004) 101:998–1003. doi: 10.1073/pnas.0306407101
- Kaiser A, Donnadieu E, Abastado JP, Trautmann A, Nardin A. CC chemokine ligand 19 secreted by mature dendritic cells increases naive T cell scanning behavior and their response to rare cognate antigen. *J Immunol.* (2005) 175:2349–56. doi: 10.4049/jimmunol.175.4.2349
- Castellino F, Huang AY, Altan-Bonnet G, Stoll S, Scheinecker C, Germain RN. Chemokines enhance immunity by guiding naive CD8+ T cells to sites of CD4+ T cell-dendritic cell interaction. *Nature.* (2006) 440:890–5. doi: 10.1038/nature04651
- Gutcher I, Becher B. APC-derived cytokines and T cell polarization in autoimmune inflammation. *J Clin Invest.* (2007) 117:1119–27. doi: 10.1172/JCI31720
- Curtsinger JM, Johnson CM, Mescher MF. CD8 T cell clonal expansion and development of effector function require prolonged exposure to antigen, costimulation, and signal 3 cytokine. *J Immunol.* (2003) 171:5165–71. doi: 10.4049/jimmunol.171.10.5165
- van Stipdonk MJ, Lemmens EE, Schoenberger SP. Naive CTLs require a single brief period of antigenic stimulation for clonal expansion and differentiation. *Nat Immunol.* (2001) 2:423–9. doi: 10.1038/87730
- Williams MA, Tyznik AJ, Bevan MJ. Interleukin-2 signals during priming are required for secondary expansion of CD8+ memory T cells. *Nature* (2006) 441:890–3. doi: 10.1038/nature04790
- Badovinac VP, Haring JS, Harty JT. Initial T cell receptor transgenic cell precursor frequency dictates critical aspects of the CD8(+) T cell response to infection. *Immunity.* (2007) 26:827–41. doi: 10.1016/j.immuni.2007.04.013
- Renkema KR, Li G, Wu A, Smithey MJ, Nikolich-Zugich J. Two separate defects affecting true naive or virtual memory T cell precursors combine to reduce naive T cell responses with aging. *J Immunol.* (2014) 192:151–9. doi: 10.4049/jimmunol.1301453
- Effros RB. Role of T lymphocyte replicative senescence in vaccine efficacy. *Vaccine.* (2007) 25:599–604. doi: 10.1016/j.vaccine.2006.08.032

24. Takahama Y. Journey through the thymus: stromal guides for T-cell development and selection. *Nat Rev Immunol.* (2006) 6:127–35. doi: 10.1038/nri1781
25. Shiow LR, Rosen DB, Brdickova N, Xu Y, An J, Lanier LL, et al. CD69 acts downstream of interferon- $\alpha$ /beta to inhibit S1P1 and lymphocyte egress from lymphoid organs. *Nature.* (2006) 440:540–4. doi: 10.1038/nature04606
26. Yang S, Liu F, Wang QJ, Rosenberg SA, Morgan RA. The shedding of CD62L (L-selectin) regulates the acquisition of lytic activity in human tumor reactive T lymphocytes. *PLoS One.* (2011) 6:e22560. doi: 10.1371/journal.pone.0022560
27. Nguyen XD, Eichler H, Dugrillon A, Piechaczek C, Braun M, Kluter H. Flow cytometric analysis of T cell proliferation in a mixed lymphocyte reaction with dendritic cells. *J Immunol Methods.* (2003) 275:57–68. doi: 10.1016/s0022-1759(03)00002-4
28. Di Rosa F. Two niches in the bone marrow: a hypothesis on life-long T cell memory. *Trends Immunol.* (2016) 37:503–12. doi: 10.1016/j.it.2016.05.004
29. Di Rosa F. Maintenance of memory T cells in the bone marrow: survival or homeostatic proliferation? *Nat Rev Immunol.* (2016) 16:271. doi: 10.1038/nri.2016.31
30. Simonetti S, Natalini A, Folgori A, Capone S, Nicosia A, Santoni A, et al. Antigen-specific CD8 T cells in cell cycle circulate in the blood after vaccination. *Scand J Immunol.* (2019) 89:e12735. doi: 10.1111/sji.12735
31. Kamphorst AO, Pillai RN, Yang S, Nasti TH, Akondy RS, Wieland A, et al. Proliferation of PD-1+ CD8 T cells in peripheral blood after PD-1-targeted therapy in lung cancer patients. *Proc Natl Acad Sci USA.* (2017) 114:4993–8. doi: 10.1073/pnas.1705327114
32. Wieland A, Kamphorst AO, Adsay NV, Masor JJ, Sarmiento J, Nasti TH, et al. T cell receptor sequencing of activated CD8 T cells in the blood identifies tumor-infiltrating clones that expand after PD-1 therapy and radiation in a melanoma patient. *Cancer Immunol Immunother.* (2018) 67:1767–76. doi: 10.1007/s00262-018-2228-7
33. Simula L, Nazio F, Campello S. The mitochondrial dynamics in cancer and immune-surveillance. *Semin Cancer Biol.* (2017) 47:29–42. doi: 10.1016/j.semcancer.2017.06.007
34. Corrado M, Mariotti FR, Trapani L, Taraborrelli L, Nazio F, Cianfanelli V, et al. Macroautophagy inhibition maintains fragmented mitochondria to foster T cell receptor-dependent apoptosis. *EMBO J.* (2016) 35:1793–809. doi: 10.15252/embj.201593727
35. Lee JE, Westrate LM, Wu H, Page C, Voeltz GK. Multiple dynamin family members collaborate to drive mitochondrial division. *Nature.* (2016) 540:139–43. doi: 10.1038/nature20555
36. Cipolat S, Martins de Brito O, Dal Zilio B, Scorrano L. OPA1 requires mitofusin 1 to promote mitochondrial fusion. *Proc Natl Acad Sci USA.* (2004) 101:15927–32. doi: 10.1073/pnas.0407043101
37. Eura Y, Ishihara N, Yokota S, Mihara K. Two mitofusin proteins, mammalian homologues of FZO, with distinct functions are both required for mitochondrial fusion. *J Biochem.* (2003) 134:333–44. doi: 10.1093/jb/mvg150
38. Kingate C, Charoenkwan K, Kumfu S, Chattipakorn N, Chattipakorn SC. Possible roles of mitochondrial dynamics and the effects of pharmacological interventions in chemoresistant ovarian cancer. *Ebiomedicine.* (2018) 34:256–66. doi: 10.1016/j.ebiom.2018.07.026
39. Buck MD, O'Sullivan D, Klein Geltink RI, Curtis JD, Chang CH, Sanin DE, et al. Mitochondrial dynamics controls T cell fate through metabolic programming. *Cell.* (2016) 166:63–76. doi: 10.1016/j.cell.2016.05.035
40. van der Windt GJ, Everts B, Chang CH, Curtis JD, Freitas TC, Amiel E, et al. Mitochondrial respiratory capacity is a critical regulator of CD8+ T cell memory development. *Immunity.* (2012) 36:68–78. doi: 10.1016/j.immuni.2011.12.007
41. Baixauli F, Martin-Cofreces NB, Morlino G, Carrasco YR, Calabia-Linares C, Veiga E, et al. The mitochondrial fission factor dynamin-related protein 1 modulates T-cell receptor signalling at the immune synapse. *EMBO J.* (2011) 30:1238–50. doi: 10.1038/emboj.2011.25
42. Wang R, Dillon CP, Shi LZ, Milasta S, Carter R, Finkelstein D, et al. The transcription factor Myc controls metabolic reprogramming upon T lymphocyte activation. *Immunity.* (2011) 35:871–82. doi: 10.1016/j.immuni.2011.09.021
43. Simula L, Pacella I, Colamatto A, Procaccini C, Cancila V, Bordini M, et al. Drp1 controls effective T cell immune-surveillance by regulating T cell migration, proliferation, and cMyc-dependent metabolic reprogramming. *Cell Rep.* (2018) 25:3059–73.e10. doi: 10.1016/j.celrep.2018.11.018
44. Campello S, Lacalle RA, Bettella M, Manes S, Scorrano L, Viola A. Orchestration of lymphocyte chemotaxis by mitochondrial dynamics. *J Exp Med.* (2006) 203:2879–86. doi: 10.1084/jem.20061877
45. Barzilai S, Yadav SK, Morrell S, Roncato F, Klein E, Stoler-Barak L, et al. Leukocytes breach endothelial barriers by insertion of nuclear lobes and disassembly of endothelial actin filaments. *Cell Rep.* (2017) 18:685–99. doi: 10.1016/j.celrep.2016.12.076
46. Jacobelli J, Estin Matthews M, Chen S, Krummel MF. Activated T cell trans-endothelial migration relies on myosin-IIA contractility for squeezing the cell nucleus through endothelial cell barriers. *PLoS One.* (2013) 8:e75151. doi: 10.1371/journal.pone.0075151
47. Henson SM, Lanna A, Riddell NE, Franzese O, Macaulay R, Griffiths SJ, et al. p38 signaling inhibits mTORC1-independent autophagy in senescent human CD8(+) T cells. *J Clin Invest.* (2014) 124:4004–16. doi: 10.1172/JCI75051
48. Panduro M, Benoist C, Mathis D. Tissue treigs. *Annu Rev Immunol.* (2016) 34:609–33. doi: 10.1146/annurev-immunol-032712-095948
49. Kolodin D, van Panhuys N, Li C, Magnuson AM, Cipolletta D, Miller CM, et al. Antigen- and cytokine-driven accumulation of regulatory T cells in visceral adipose tissue of lean mice. *Cell Metab.* (2015) 21:543–57. doi: 10.1016/j.cmet.2015.03.005
50. Korn LL, Hubbeling HG, Porrett PM, Yang Q, Barnett LG, Laufer TM. Regulatory T cells occupy an isolated niche in the intestine that is antigen independent. *Cell Rep.* (2014) 9:1567–73. doi: 10.1016/j.celrep.2014.11.006
51. Lynch L, Michelet X, Zhang S, Brennan PJ, Moseman A, Lester C, et al. Regulatory iNKT cells lack expression of the transcription factor PLZF and control the homeostasis of T(reg) cells and macrophages in adipose tissue. *Nat Immunol.* (2015) 16:85–95. doi: 10.1038/ni.3047
52. Luo CT, Liao W, Dadi S, Toure A, Li MO. Graded Foxo1 activity in Treg cells differentiates tumour immunity from spontaneous autoimmunity. *Nature.* (2016) 529:532–6. doi: 10.1038/nature16486
53. Smigiel K, Richards E, Srivastava S, Thomas KR, Dudda JC, Klonowski KD, et al. CCR7 provides localized access to IL-2 and defines homeostatically distinct regulatory T cell subsets. *J Exp Med.* (2014) 211:121–36. doi: 10.1084/jem.20131142
54. Mackay LK, Kallies A. Transcriptional regulation of tissue-resident lymphocytes. *Trends Immunol.* (2017) 38:94–103. doi: 10.1016/j.it.2016.11.004
55. Fan X, Moltedo B, Mendoza A, Davydov AN, Faire MB, Mazutis L, et al. CD49b defines functionally mature Treg cells that survey skin and vascular tissues. *J Exp Med.* (2018) 215:2796–814. doi: 10.1084/jem.20181442
56. Ali N, Zirak B, Rodriguez RS, Pauli ML, Truong HA, Lai K, et al. Regulatory T cells in skin facilitate epithelial stem cell differentiation. *Cell.* (2017) 169:1119–29.e11. doi: 10.1016/j.cell.2017.05.002
57. Emmerson A, Trevelin SC, Mongue-Din H, Becker PD, Ortiz C, Smyth LA, et al. Nox2 in regulatory T cells promotes angiotensin II-induced cardiovascular remodeling. *J Clin Invest.* (2018) 128:3088–101. doi: 10.1172/JCI97490
58. Ichikawa T, Hirahara K, Kokubo K, Kiuchi M, Aoki A, Morimoto Y, et al. CD103(hi) Treg cells constrain lung fibrosis induced by CD103(lo) tissue-resident pathogenic CD4 T cells. *Nat Immunol.* (2019) 20:1469–80. doi: 10.1038/s41590-019-0494-y
59. Vasanthakumar A, Moro K, Xin A, Liao Y, Gloury R, Kawamoto S, et al. The transcriptional regulators IRF4, BATF and IL-33 orchestrate development and maintenance of adipose tissue-resident regulatory T cells. *Nat Immunol.* (2015) 16:276–85. doi: 10.1038/ni.3085
60. Yu L, Yang F, Zhang F, Guo D, Li L, Wang X, et al. CD69 enhances immunosuppressive function of regulatory T-cells and attenuates colitis by prompting IL-10 production. *Cell Death Dis.* (2018) 9:905. doi: 10.1038/s41419-018-0927-9
61. Huehn J, Siegmund K, Lehmann JC, Siewert C, Haubold U, Feuerer M, et al. Developmental stage, phenotype, and migration distinguish naive- and effector/memory-like CD4+ regulatory T cells. *J Exp Med.* (2004) 199:303–13. doi: 10.1084/jem.20031562
62. Suffia I, Reckling SK, Salay G, Belkaid Y. A role for CD103 in the retention of CD4+CD25+ Treg and control of Leishmania major infection. *J Immunol.* (2005) 174:5444–55. doi: 10.4049/jimmunol.174.9.5444

63. Deaglio S, Dwyer KM, Gao W, Friedman D, Usheva A, Erat A, et al. Adenosine generation catalyzed by CD39 and CD73 expressed on regulatory T cells mediates immune suppression. *J Exp Med.* (2007) 204:1257–65. doi: 10.1084/jem.20062512
64. Han JM, Patterson SJ, Speck M, Ehses JA, Levings MK. Insulin inhibits IL-10-mediated regulatory T cell function: implications for obesity. *J Immunol.* (2014) 192:623–9. doi: 10.4049/jimmunol.1302181
65. Mackay LK, Minnich M, Kragten NA, Liao Y, Nota B, Seillet C, et al. Hobit and Blimp1 instruct a universal transcriptional program of tissue residency in lymphocytes. *Science.* (2016) 352:459–63. doi: 10.1126/science.aad2035
66. Cretney E, Xin A, Shi W, Minnich M, Masson F, Miasari M, et al. The transcription factors Blimp-1 and IRF4 jointly control the differentiation and function of effector regulatory T cells. *Nat Immunol.* (2011) 12:304–11. doi: 10.1038/ni.2006
67. Lowe MM, Boothby I, Clancy S, Ahn RS, Liao W, Nguyen DN, et al. Regulatory T cells use arginase 2 to enhance their metabolic fitness in tissues. *JCI Insight.* (2019) 4:e129756. doi: 10.1172/jci.insight.129756
68. Snyder ME, Finlayson MO, Connors TJ, Dogra P, Senda T, Bush E, et al. Generation and persistence of human tissue-resident memory T cells in lung transplantation. *Sci Immunol.* (2019) 4:eaav5581. doi: 10.1126/sciimmunol.aav5581
69. Chao JL, Savage PA. Unlocking the complexities of tumor-associated regulatory T cells. *J Immunol.* (2018) 200:415–21. doi: 10.4049/jimmunol.1701188
70. Zheng C, Zheng L, Yoo JK, Guo H, Zhang Y, Guo X, et al. Landscape of infiltrating T cells in liver cancer revealed by single-cell sequencing. *Cell.* (2017) 169:1342–56.e16. doi: 10.1016/j.cell.2017.05.035
71. De Simone M, Arrigoni A, Rossetti G, Gruarin P, Ranzani V, Politano C, et al. Transcriptional landscape of human tissue lymphocytes unveils uniqueness of tumor-infiltrating T regulatory cells. *Immunity.* (2016) 45:1135–47. doi: 10.1016/j.immuni.2016.10.021
72. Plitas G, Konopacki C, Wu K, Bos PD, Morrow M, Putintseva EV, et al. Regulatory T cells exhibit distinct features in human breast cancer. *Immunity.* (2016) 45:1122–34. doi: 10.1016/j.immuni.2016.10.032
73. Cane S, Ponnappan S, Ponnappan U. Altered regulation of CXCR4 expression during aging contributes to increased CXCL12-dependent chemotactic migration of CD4(+) T cells. *Aging Cell.* (2012) 11:651–8. doi: 10.1111/j.1474-9726.2012.00830.x
74. Richner JM, Gmyrek GB, Govero J, Tu Y, van der Windt GJ, Metcalf TU, et al. Age-dependent cell trafficking defects in draining lymph nodes impair adaptive immunity and control of west Nile virus infection. *PLoS Pathog.* (2015) 11:e1005027. doi: 10.1371/journal.ppat.1005027

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2020 Piconese, Campello and Natalini. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# APPENDIX: EFIS-EJI Ruggero Ceppellini Advanced School of Immunology Founded by Serafino Zappacosta. List of the Activities From Its Foundation in 1991 to 2019

*Note: for ongoing activities see [www.ceppellini.it](http://www.ceppellini.it).*

## Table of Contents

<b>71</b>	<b><i>Level 1 Courses</i></b>
<b>72</b>	<b><i>Level 2 Courses</i></b>
<b>100</b>	<b><i>Level 3 Courses</i></b>
<b>101</b>	<b><i>Other Events Till 2006</i></b>
<b>104</b>	<b><i>Serafino Zappacosta Conferences</i></b>



## LEVEL 1 COURSES

Level 1 courses are immunology refresher courses aimed at updating Italian physicians, pharmacists, or professionals in other medical disciplines. Notably, the Ruggero Ceppellini Advanced School of Immunology started organizing these courses long before Continuing Medical Education (CME) became mandatory for MDs in Italy in 2002.

### **L'Immunità in patologia umana 95/96 - Immunity in human pathology 95/96**

National Tumor Institute "Giovanni Pascale" of Naples, Naples, November 1995-March 1996

### **L'Immunità in patologia umana 1999 - Immunity in human pathology 1999**

Federico II University Medical School, Naples February - May 1999

### **L'Immunità in patologia umana 2002 - Immunity in human pathology 2002**

Azienda Ospedaliera V Monaldi, Naples, November-December 2002

### **L'Immunità in patologia umana 2012 - Immunity in human pathology 2012**

Federico II University Medical School, Naples, September - December 2012

### **Immunodeficienze in Pediatria - Immunodeficiency in Pediatrics**

Federico II University Medical School, Naples, February - April 2015

## LEVEL 2 COURSES

Level 2 courses are the typical activities of the Ceppellini School. They are International Advanced Immunology courses dedicated to young researchers (PhD students, post-docs, etc.) from all over the world, particularly from developing countries, wishing to acquire in-depth knowledge on a specific topic from leader scientists in the field.

### “Immunology of Bone Marrow Transplantation”

Palazzo Serra di Cassano, Istituto Italiano per gli Studi Filosofici, Naples,  
12-16 October 1992

This Course followed immediately the School's inaugural ceremony at the Istituto Italiano per gli Studi Filosofici. Therefore, the course took place in an atmosphere of emotion and hope and benefited from the direction of Elizabeth Simpson, who played the role of enthusiastic organizer, continuing since then to collaborate lovingly with the School in both the didactic and the organizational aspects.

#### *The Course Director*

Elizabeth Simpson  
*Division of Transplantation Biology, MRC Clinical Research Centre, Harrow, Middlesex, UK*

#### *The Faculty*

Andrea Bacigalupo  
*Divisione di Ematologia ed Immunologia Clinica, Ospedale San Martino, Genova, Italy*  
Giovanni B Ferrara  
*Servizio di Immunogenetica, Istituto Nazionale Tumori, Genova, Italy*  
Manlio Ferrarini  
*Servizio di Immunologia Clinica, Istituto Nazionale Tumori, Genova, Italy*  
Peter M Hoogerbrugge  
*Department of Gene Therapy, Institute for Applied Radiobiology and Immunology, Rijswijk, The Netherlands*  
Jill M Hows  
*Department of Haematology, Southmead Hospital, Bristol, UK*  
Robert I Lechler  
*Department of Immunology, Royal Postgraduate Medical School, Hammersmith Hospital, London, UK*  
Guido Lucarelli  
*Divisione di Ematologia e Centro Trapianto Midollo Osseo di Muraglia, Ospedale di Pesaro, Pesaro, Italy*  
Yair Reisner  
*Department of Biophysics & Membrane Research, The Weizmann Institute of Science, Rehovot, Israel*  
Maria Grazia Roncarolo  
*DNAX Research Institute, Palo Alto, CA, USA*  
Bruno Rotoli  
*Divisione di Ematologia Clinica, Facoltà di Medicina e Chirurgia, Università di Napoli Federico II, Napoli, Italy*  
Elizabeth Simpson  
*Division of Transplantation Biology, MRC Clinical Research Centre, Harrow, Middlesex, UK*  
Nydia G Testa  
*Department of Experimental Haematology, Paterson Institute for Cancer Research, Manchester, UK*  
Andrea Velardi  
*Istituto di Clinica Medica, Policlinico Monteluce, Università di Perugia, Perugia, Italy*  
Herman Waldmann  
*Immunology Division, Department of Pathology, Cambridge University, Cambridge, UK*  
Serafino Zappacosta  
*Cattedra di Immunologia, Dipartimento di Biologia e Patologia Cellulare e Molecolare, Università di Napoli Federico II, Napoli, Italy*

*The Organizing Committee: Antonio Di Giacomo, Azienda Ospedaliera V Monaldi, Napoli, Italy.*

*Secretariat: Tricia Reynolds, Scuola Superiore d'Immunologia Ruggero Ceppellini, Napoli, Italy; Vivien Tikerpae, Division of Transplantation Biology, MRC Clinical Research Centre, Harrow, Middlesex, UK*

*Sponsorships: Istituto Italiano per gli Studi Filosofici, Naples, Italy; Becton Dickinson Italia SpA, Milano; Boehringer Mannheim Italia SpA, Milano; Heraeus SpA, Cavenago Brianza MI; ICN Biomedical SpA, Cassina de' Pecchi FI; Italfarmaco SpA, Milano; Janssen Farmaceutici SpA, Roma; Lagitre Srl, Milano; M-Medical Srl, Firenze; Microglass Srl, Napoli; Società Prodotti Antibiotici SpA, Milano; Zambon Group SpA, Milano; and the Azienda Autonoma di Soggiorno Cura e Turismo of Naples.*

## **“Progress and Perspectives in Vaccination”**

Palazzo Serra di Cassano, Istituto Italiano per gli Studi Filosofici, Naples,  
16–20 May 1994

The Course dealt with the most recent aspects of vaccination. Basic knowledge about the immune response as well as new vaccine technology were covered. The Course included an Opening and a Closing Session addressed to the general public on “The Impact of Vaccination on Human Welfare and Society”.

### *The Course Director*

Gino Doria  
*Laboratorio di Immunologia, ENEA-CRE Casaccia, Roma, Italy*

### *The Faculty*

Francisco E Baralle  
*International Centre for Genetic Engineering and Biotechnology, UNIDO, Trieste, Italy*  
Antonio Cassone  
*Dipartimento di Batteriologia e Micologia Medica, Istituto Superiore di Sanità, Roma, Italy*  
Andrea Crisanti  
*Istituto di Parassitologia, Università di Roma La Sapienza, Roma, Italy*  
Ferdinando Dianzani  
*Istituto di Virologia, Università di Roma La Sapienza, Roma, Italy*  
Manfred P Dierich  
*Institut für Hygiene der Leopold-Franzens-Universität, Innsbruck, Austria*  
Soldano Ferrone  
*Department of Microbiology and Immunology, New York Medical College, Valhalla, NY, USA*  
Daniela Frasca  
*Laboratorio di Immunologia, ENEA-CRE Casaccia, Roma, Italy*  
Michele E Grandolfo  
*Dipartimento di Epidemiologia e Biostatistica, Istituto Superiore di Sanità, Roma, Italy*  
Helmut Hahn  
*Institut für Medizinische Mikrobiologie und Infektionsimmunologie, Freie Universität Berlin, Berlin, Germany*  
Richard J Hodes  
*National Institute on Aging, NIH, Bethesda, MD, USA*  
Ada M Kruisbeek  
*Afdeling Immunologie, Het Nederlands Kanker Instituut, Amsterdam, The Netherlands*  
Andrew J McMichael  
*Institute of Molecular Medicine, John Radcliffe Hospital, Oxford University, Oxford, UK*  
Filippo Palumbo  
*Osservatorio Epidemiologico Regionale, Regione Campania, Napoli, Italy*  
Giorgio Parmiani  
*Divisione di Oncologia Sperimentale D, Istituto Nazionale Tumori, Milano, Italy*  
Marcello Piazza  
*Istituto di Malattie Infettive, Università di Napoli Federico II, Napoli, Italy*  
Rino Rappuoli  
*Istituto Ricerche Immunobiologiche Siena, Siena, Italy*  
Angela Vegnente  
*Dipartimento di Pediatria, Università di Napoli Federico II, Napoli, Italy*  
Marc E Weksler  
*Division of Geriatrics, Department of Medicine, Cornell University Medical College, New York, NY, USA*  
Serafino Zappacosta  
*Cattedra di Immunologia, Dipartimento di Biologia e Patologia Cellulare e Molecolare, Università di Napoli Federico II, Napoli, Italy*

*The Organizing Committee:* Antonio Di Giacomo, Azienda Ospedaliera V Monaldi, Napoli, Italy; Ciro Manzo & Armando Tripodi, Istituto Nazionale Tumori Fondazione Pascale, Napoli, Italy; Luigi Racioppi & Serafino Zappacosta, Università di Napoli Federico II, Napoli, Italy.

*Secretariat:* Donatella Capone & Carla Corradini, Scuola Superiore d'Immunologia Ruggero Ceppellini, Napoli, Italy; Margherita Foggia & Francesco Scerbo, Istituto Nazionale Tumori Fondazione Pascale, Napoli, Italy; Amanda Wren, Nature Classified, Macmillan Magazines Ltd, London, UK.

*Sponsorships:* Istituto Italiano per gli Studi Filosofici, Naples, Italy; Istituto Nazionale Tumori Fondazione Pascale Naples, Italy; Biocine Srl, Siena; Bio-Rad SpA, Milano, and Microg

## **“T-Cell Activation, Anergy and Immunosuppressive Drug Action”**

Istituto Nazionale Tumori Fondazione Pascale, Naples,  
17-21 October 1994

This Course covered the main aspects of T cell activation, central to all mechanisms of the immune response, and it included an Opening session on “The continuing education of physicians and researchers”, addressed to a local audience.

### *The Course Director*

Stefan C Meuer  
*Abteilung Angewandte Immunologie, Deutsches Krebsforschungszentrum Heidelberg, Heidelberg, Germany*

### *The Faculty*

V Enrico Avvedimento  
*Dipartimento di Medicina Sperimentale e Clinica, Università di Reggio Calabria, Catanzaro, Italy*  
Patrick A Baeuerle  
*Lehrstuhl für Biochemie, Albert-Ludwigs-Universität, Freiburg, Germany*  
Doreen A Cantrell  
*Lymphocyte Activation Laboratory, Imperial Cancer Research Fund, London, UK*  
Dino Collavo  
*Cattedra di Immunologia, Facoltà di Medicina e Chirurgia, Università di Padova, Padova, Italy*  
Mario Condorelli  
*Dipartimento di Cardiologia e Cardiochirurgia, Università di Napoli Federico II, Napoli, Italy*  
Bernhard Fleischer  
*Abteilung Medizinische Mikrobiologie und Immunologie, Bernhard-Nocht-Institut für Tropenmedizin, Hamburg, Germany*  
Gerardo Marotta  
*Istituto Italiano per gli Studi Filosofici, Napoli, Italy*  
Polly Matzinger  
*Laboratory for Cellular and Molecular Immunology, National Institute of Allergy and Infectious Diseases, NIH, Bethesda, MD, USA*  
Stefan C Meuer  
*Abteilung Angewandte Immunologie, Deutsches Krebsforschungszentrum Heidelberg, Heidelberg, Germany*  
Frank Momburg  
*Abteilung Molekulare Immunologie, Deutsches Krebsforschungszentrum Heidelberg, Heidelberg, Germany*  
Luigi Racioppi  
*Cattedra di Immunologia, Dipartimento di Biologia e Patologia Cellulare e Molecolare, Università di Napoli Federico II, Napoli, Italy*  
Stephen Shaw  
*Experimental Immunology Branch, National Cancer Institute, NIH, Bethesda, MD, USA*  
Craig B Thompson  
*Department of Medicine & Molecular Genetics & Cell Biology, Howard Hughes Medical Institute, University of Chicago, Chicago, IL, USA*  
Serafino Zappacosta  
*Cattedra di Immunologia, Dipartimento di Biologia e Patologia Cellulare e Molecolare, Università di Napoli Federico II, Napoli, Italy*

*The Organizing Committee:* Antonio Di Giacomo, Azienda Ospedaliera V Monaldi, Napoli, Italy; Ciro Manzo & Armando Tripodi, Istituto Nazionale Tumori Fondazione Pascale, Napoli, Italy; Serafino Zappacosta, Università di Napoli Federico II, Napoli, Italy.

*Secretariat:* Tricia Reynolds (Chairperson) & Carla Corradini, Scuola Superiore d'Immunologia Ruggero Ceppellini, Napoli, Italy; Margherita Foggia & Francesco Scerbo, Istituto Nazionale Tumori Fondazione Pascale, Napoli, Italy; Amanda Wren, Nature Classified, Macmillan Magazines Ltd, London, UK.

*Sponsorships:* Istituto Italiano per gli Studi Filosofici, Naples, Italy; Istituto Nazionale Tumori Fondazione Pascale Naples, Italy; Dianova GmbH, Hamburg, Germany, and from Hermann Biermann GmbH, Bad Neuheim, Germany.



## **“Immunity to Intracellular Bacteria & Parasites”**

Positano, Hotel Le Agavi, near Salerno,  
20-25 May 1995

The objective of this Course was to spread information on infectious disease pathogenesis and immunity. About 35% of the world population still die of infections. Thus, there is a great demand for improvement. A prerequisite is a sound understanding of basic mechanisms of pathogenesis and immunity to infections. Leading scientists in these fields contributed to make this course a success.

### *The Course Director*

Helmut Hahn  
*Institut für Medizinische Mikrobiologie und Infektionsimmunologie,  
Freie Universität Berlin, Berlin, Germany*

### *The Faculty*

Peter Andersen  
*Bacterial Vaccines Department, Statens Seruminstitut, Copenhagen, Denmark*

Ingo B Autenrieth  
*Institut für Medizinische Mikrobiologie der Universität Würzburg, Würzburg, Germany*

Gregory J Bancroft  
*Department of Clinical Sciences, London School of Hygiene and Tropical Medicine, London, UK*

Dov L Boros  
*Department of Immunology and Microbiology, Wayne State University School of Medicine, Detroit, MI, USA*

Stefan Brocke  
*Department of Neurology and Neurological Sciences, Stanford University Medical Center, Beckman Center for Molecular and Genetic Medicine, Stanford, CA, USA*

Trinad Chakraborty  
*Institut für Medizinische Mikrobiologie der Justus-Liebig-Universität, Giessen, Germany*

Eric Y Denkers  
*Immunology & Cell Biology Section, National Institute of Allergy & Infectious Diseases, NIH, Bethesda, MD, USA*

Stefan Ehlers  
*Institut für Medizinische Mikrobiologie und Infektionsimmunologie,  
Freie Universität Berlin, Berlin, Germany*

Ralph van Furth  
*Department of Infectious Diseases, University Hospital, Leiden, The Netherlands*

Klas Kärre  
*Microbiology and Tumorbiology Center, Karolinska Institutet, Stockholm, Sweden*

Stefan H E Kaufmann  
*Institut für Immunologie der Universität Ulm, Ulm and Max-Planck-Institut für Infektionsbiologie, Berlin, Germany*

Peter G Kremsner  
*Institut für Tropenmedizin, Berlin, Germany and International Research Laboratory of the Albert Schweitzer Hospital, Lambaréné, Gabon*

F Y Liew  
*Department of Immunology, Western Infirmary, Glasgow University, Glasgow, UK*

Ralf Lucas  
*Département de Pathologie, Faculté de Médecine, Université de Genève, Genève, Switzerland*

Martin E A Mielke  
*Institut für Medizinische Mikrobiologie und Infektionsimmunologie,  
Freie Universität Berlin, Berlin, Germany*

Heidrun Moll  
*Zentrum für Infektionsforschung der Universität Würzburg, Würzburg, Germany*

Lorenzo Moretta  
*Istituto Nazionale per la Ricerca sul Cancro, Genova, Italy*

Robert J North  
*Trudeau Institute, Saranac Lake, NY, USA*

Armelle Phalipon  
*Unité de Pathogénie Microbienne Moléculaire, Institut Pasteur, Paris, France*

*The Organizing Committee:* Antonio Di Giacomo, Azienda Ospedaliera V Monaldi, Napoli, Italy; Stefan Ehlers & Martin E A Mielke, Freie Universität Berlin, Berlin, Germany; Luigi Racioppi & Serafino Zappacosta, Università di Napoli Federico II, Napoli, Italy.

*Secretariat:* Tricia Reynolds, Scuola Superiore d'Immunologia Ruggero Ceppellini, Napoli, Italy.

*Sponsorships:* Istituto Italiano per gli Studi Filosofici, Naples, Italy; Dipartimento di Biologia e Patologia Cellulare e Molecolare of the Federico II University of Naples; Bio-Rad Srl, Segrate MI; Microglass Srl, Napoli; Bayer (Germany); Lederle Pharma (Wolfartshausen, Germany); and Takeda Pharma (Aachen, Germany); Assessorato al Turismo e Spettacolo of the Campania Region.

## **“Mechanisms and Manipulation of Autoimmunity”**

Istituto Nazionale Tumori Fondazione Pascale, Naples,  
23–27 June 1996

This Course was devoted to the presentation and discussion of the essential aspects of autoimmunity, of remarkable interest from both the speculative and the clinical standpoints. After a review of the immunological tolerance and of the antigen presentation mechanisms, the course focussed on several clinical aspects, from the genetic susceptibility to recent therapeutic approaches to autoimmune diseases.

### *The Course Director*

Robert I Lechler  
*Department of Immunology, Royal Postgraduate Medical School, Hammersmith Hospital, London, UK*

### *The Faculty*

Anne Cooke  
*Department of Pathology, Immunology Division, University of Cambridge, Cambridge, UK*  
Rikard Holmdahl  
*Department of Cell and Molecular Biology, Section for Medical Inflammation Research, Lund University, Lund, Sweden*  
Eric J Jenkinson  
*Department of Anatomy, Medical School, University of Birmingham, Birmingham, UK*  
Peter Lane  
*Basel Institute for Immunology, Basel, Switzerland*  
Robert I Lechler  
*Department of Immunology, Royal Postgraduate Medical School, Hammersmith Hospital, London, UK*  
David Lo  
*Department of Immunology IMM-25, The Scripps Research Institute, La Jolla, CA, USA*  
James McCluskey  
*Department of Clinical Immunology, Flinders Medical Centre, Bedford Park, South Australia*  
Francesco Sinigaglia  
*Preclinical Research Division, Roche Milano Ricerche, Milano, Italy*  
Hans Stauss  
*Department of Immunology, Royal Postgraduate Medical School, Hammersmith Hospital, London, UK*  
Brigitta Stockinger  
*National Institute for Medical Research, London, UK*  
David C Wraith  
*Department of Pathology & Microbiology, School of Medical Sciences, Bristol, UK*

*The Organizing Committee:* Silvia Fontana, *Centro di Endocrinologia e Oncologia Sperimentale, CNR, Napoli, Italy*; Ciro Manzo & Armando Tripodi, *Istituto Nazionale Tumori Fondazione Pascale, Napoli, Italy*; Guido Rossi, *Università di Napoli Federico II, Napoli, Italy*.

*Secretariat:* Margherita Foggia, Tiziana Foggia & Francesco Scerbo, *Istituto Nazionale Tumori Fondazione Pascale, Napoli, Italy*; Amanda Wren, *Nature Classified, Macmillan Magazines Ltd, London, UK*.

*Sponsorships:* Istituto Italiano per gli Studi Filosofici, Naples, Italy; Istituto Nazionale Tumori Fondazione Pascale, Naples, Italy; Amersham Italia Srl, Milano; Dasit SpA, Cornaredo MI; International PBI SpA, Milano; Microglass Srl, Napoli; and M-Medical Srl, Firenze.

## “HLA & Tumours”

Istituto Nazionale Tumori Fondazione Pascale, Naples,  
2-6 December 1996

This Course was aimed at the presentation and discussion of some immunological aspects of tumor-host interaction, including the control exerted by the innate and adaptive immune responses. Therefore, the attention was focussed on the potential therapeutic approaches based on exploiting anti-tumour immunity.

### *The Course Directors*

Soldano Ferrone

*Department of Microbiology & Immunology, New York Medical College, Valhalla, NY, USA*

Ciro Manzo

*Divisione di Oncologia Sperimentale C, Immunologia, Istituto Nazionale Tumori Fondazione Pascale, Napoli, Italy*

### *The Faculty*

Ennio Carbone

*Cattedra di Immunologia, Dipartimento di Biologia e Patologia Cellulare e Molecolare, Università di Napoli Federico II, Napoli, Italy*

Marco Colonna

*Basel Institute for Immunology, Basel, Switzerland*

Pierre G. Coulie

*Unité de Génétique Cellulaire, Université Catholique de Louvain, Brussels, Belgium*

Giovanni B Ferrara

*Laboratorio di Immunogenetica, Istituto Scientifico Tumori e Centro di Biotecnologie Avanzate, Genova, Italy*

Soldano Ferrone

*Department of Microbiology & Immunology, New York Medical College, Valhalla, NY, USA*

Patrizio Giacomini

*Laboratorio di Immunologia, Istituto Nazionale Tumori Regina Elena, Roma, Italy*

John Guardioli

*Istituto Internazionale di Genetica e Biofisica del CNR, Napoli, Italy*

Klas Kärre

*Microbiology & Tumorbiology Center, Karolinska Institutet, Stockholm, Sweden*

Jim Kaufman

*Institute for Animal Health, Compton, nr. Newbury, Berks, UK*

Rolf Kiessling

*Microbiology & Tumorbiology Center, Karolinska Institutet, Stockholm, Sweden*

Michele Maio

*Unità di Immunoterapia Avanzata, Centro di Riferimento Oncologico, INRCCS, Aviano PN, Italy*

Francesco M. Marincola

*Surgery Branch, National Cancer Institute, National Institutes of Health, Bethesda, MD, USA*

Lorenzo Moretta

*Laboratorio di Immunopatologia, Istituto Scientifico Tumori e Centro di Biotecnologie Avanzate, Genova, Italy*

Licia Rivoltini

*Divisione di Oncologia Sperimentale D, Istituto Nazionale Tumori, Milano, Italy*

Barbara Seliger

*III Medizinische Klinik, Abteilung für Innere Medizin-Hämatologie,*

*Johann-Gutenberg-Universität Mainz, Mainz, Germany*

John Trowsdale

*Human Immunogenetics Laboratory, Imperial Cancer Research Fund, London, UK*

*The Organizing Committee: Antonio Di Giacomo, Azienda Ospedaliera V Monaldi, Napoli, Italy; Silvia Fontana, Centro di Endocrinologia e Oncologia, Sperimentale, CNR, Napoli, Italy; Giuseppina Ruggiero, Università di Napoli Federico II, Napoli, Italy; Armando Tripodi, Istituto Nazionale Tumori Fondazione Pascale, Napoli, Italy.*

*Secretariat: Maria de Manes & Alessandra Saioni, Effe Erre Congressi, Napoli, Italy; Anna Maria Masci & José Terrazzano, Università di Napoli Federico II, Napoli, Italy.*

*Sponsorships: Istituto Italiano per gli Studi Filosofici, Naples, Italy; Istituto Nazionale Tumori Fondazione Pascale, Naples, Italy; Dako SpA, Milano; Labscience Italia Srl, Torre del Greco NA; and Microglass Srl, Napoli.*

# Cytokines in Immunity

Città della Scienza, Naples,  
3-7 November 1997

This Course dealt with the structural and functional properties of cytokines and their receptors in the complex regulatory circuits of immunity. The timeliness of such Course in a period characterised by an explosive outbreak of information both on the physiological and on the pathological implications of the immune response was witnessed by the enthusiastic participation of a very distinguished faculty and of a large international audience.

## *The Course Director*

Abul K Abbas  
*Department of Pathology, Immunology Research Division, Brigham and Women's Hospital, Boston, MA, USA*

## *The Faculty*

Abul K Abbas  
*Department of Pathology, Immunology Research Division, Brigham and Women's Hospital, Boston, MA, USA*  
Gregory J Bancroft  
*Department of Clinical Sciences, London School of Hygiene & Tropical Medicine, London, UK*  
Flavia Bazzoni  
*Istituto di Patologia Generale, Università di Verona, Verona, Italy*  
Fionula M Brennan  
*Cytokine Biology Group, The Mathilda and Terence Kennedy Institute of Rheumatology, London, UK*  
Margaret J Dallman  
*Department of Biology (Immunology), Imperial College of Science, Technology and Medicine, London, UK*  
Jo van Damme  
*Katholieke Universiteit Leuven, Rega Institute, Leuven, Belgium*  
Gino Doria  
*Cattedra di Immunologia, Dipartimento di Biologia, Università di Roma Tor Vergata, Roma, Italy*  
Olivera J Finn  
*Immunology Program, Department of Molecular Genetics & Biochemistry, University of Pittsburgh Cancer Institute, Pittsburgh, PA, USA*  
Alberto Mantovani  
*Istituto Ricerche Farmacologiche Mario Negri, Milano, Italy*  
Andreas Radbruch  
*Deutsches Rheuma-Forschungszentrum Berlin, Berlin, Germany*  
Sergio Romagnani  
*Istituto di Medicina Interna e Immunoallergologia, Università di Firenze, Firenze, Italy*  
Francesco Sinigaglia  
*Preclinical Research Division, Roche Milano Ricerche, Milano, Italy*  
Jacques Thèze  
*Unité d'Immunogénétique Cellulaire, Institut Pasteur, Paris, France*

*The Organizing Committee:* Antonio Di Giacomo, Azienda Ospedaliera V Monaldi, Napoli, Italy; Giuseppina Ruggiero, José Terrazzano & Serafino Zappacosta, Università di Napoli Federico II, Napoli, Italy.

*Secretariat:* Maria de Manes, Alessandra Saioni & Daniela Giampaolo, Effe Erre Congressi, Napoli, Italy, Amanda Wren, Nature Classified, Macmillan Magazines Ltd, London, UK.

*Sponsorships:* IDIS Foundation-Città della Scienza; Istituto Italiano per gli Studi Filosofici, Naples, Italy; Istituto Nazionale Tumori Fondazione Pascale, Naples, Italy; the Campania Region (Research Service); the Federico II University of Naples; Amersham Italia Srl, Milano; Becton Dickinson Italia SpA, Milano; Corning Costar Italia Srl, Conrorezzo MI; DBA Italia Srl, Segrate MI; Dia-Chem Srl, Napoli; EG&G SpA, Milano; Eppendorf Srl, Milano; Immucor Italia Srl, Novesasco di Opera MI; Internationl PBI SpA, Milano; Microglass Srl, Napoli; Perkin Elmer Italia SpA, Monza MI; Primm Srl, Milano; Schering-Plough SpA, Milano; Sigma Aldrich Srl, Milano; Tema Ricerca Srl, Bologna.



## **“Dendritic Cell Physiology”**

Hotel Le Agavi, Positano, near Salerno,  
20–24 May 1999

This Course was dedicated to one of the the most central topics of immunology, the interest in which was enhanced by the potential applications of dendritic cells in all interventions on the immune system. The Course benefited highly from the enthusiastic role played by Paola Ricciardi Castagnoli, who was able to gather the best faculty available, running then an impressive sequence of lectures. A Round Table on “In vivo Veritas: Integration of in vitro and in vivo Models of Dendritic Cell Physiology” was organised by Ian McConnell & Elizabeth Simpson and run in the afternoon of the last day of the Course.

### *The Course Director*

Paola Ricciardi Castagnoli  
*Dipartimento di Biotecnologie e Bioscienze, Università di Milano Bicocca, Milano, Italy*

### *The Faculty*

Sebastian Amigorena  
*INSERM U520, Institut Curie, Paris, France*  
Francine Brière  
*Centre de Recherche Schering-Plough, Dardilly, France*  
Thomas Bocker  
*Abteilung Innere Medizin, Max-Planck-Institut für Immunologie, Freiburg, Germany*  
Ennio Carbone  
*Microbiology and Tumorbiology Center, Karolinska Institutet, Stockholm, Sweden*  
Carl G Figdor  
*Tumor Immunology Laboratory, University Hospital Nijmegen, Nijmegen, The Netherlands*  
Olivera J Finn  
*Department of Molecular Genetics & Biochemistry, University of Pittsburgh School of Medicine, Pittsburgh, PA, USA*  
Giampiero Girolomoni  
*Laboratorio di Immunologia, Istituto Dermatologico dell'Immacolata, Roma, Italy*  
Francesca Granucci  
*Centro per lo Studio della Farmacologia Cellulare e Molecolare del CNR, Milano, Italy*  
Antonio Lanzavecchia  
*Basel Institute for Immunology, Basel, Switzerland*  
Charles R Maliszewski  
*Immunex Corporation, Seattle, WA, USA*  
Eugene Maraskovsky  
*Oncology Unit, Ludwig Institute for Cancer Research, Melbourne, Australia*  
Ian McConnell  
*Centre for Veterinary Science, University of Cambridge, Cambridge, UK*  
Anne O'Garra  
*Immunology Division, DNAX Research Institute of Molecular and Cellular Biology, Inc, Palo Alto, CA, USA*  
Maria Rescigno  
*Centro per lo Studio della Farmacologia Cellulare e Molecolare del CNR, Milano, Italy*  
Paola Ricciardi Castagnoli  
*Dipartimento di Biotecnologie e Bioscienze, Università di Milano Bicocca, Milano, Italy*  
Elizabeth Simpson  
*MRC Clinical Sciences Centre, Imperial College School of Medicine, Hammersmith Hospital, London, UK*  
Silvano Sozzani  
*Laboratorio di Immunologia e Biologia Cellulare, Istituto Ricerche Farmacologiche Mario Negri, Milano, Italy*  
Ralph M Steinman  
*Laboratory of Cellular Physiology and Immunology, The Rockefeller University, New York, NY, USA*

*The Organizing Committee:* Silvia Fontana, *Centro di Endocrinologia e Oncologia Sperimentale, CNR, Napoli, Italy;* Giuseppina Ruggiero, José Terrazzano & Serafino Zappacosta, *Università di Napoli Federico II, Napoli, Italy.*

*Secretariat:* Maria de Manes, Alessandra Saioni & Daniela Giampaolo, *Effe Erre Congressi, Napoli, Italy;* Nevin Bayoumi, *Nature Classified, Macmillan Magazines Ltd, London, UK.*

*Sponsorships:* Istituto Italiano per gli Studi Filosofici, Naples, Italy; Università di Napoli Federico II; the Campania Region Education Service, the Mayor of Positano; Immunex Corporation, Seattle, WA, USA; Miltenyi Biotech GmbH, Bergisch Gladbach, Germany; Roche SpA, Milano; Tema Ricerca Srl, Bologna, and The Ares Sero Group, Geneva, Switzerland.

# **“Escape From Immune Surveillance of Tumours and Micro-organisms: Emerging Mechanisms and Shared Strategies”**

Centro S. Ignazio, Naples,  
23–27 March 2000

The Course was aimed at discussing in a number of systems the molecular mechanisms behind the immunological escape of tumours, viruses, bacteria and parasites, accounting for their ability to evade detection by T cells, NK cells and antibodies. In particular, selection of antigenic loss variants, defects in antigen presentation, immune suppressive cytokines, defects in signal transducing molecules, induction of apoptosis in T cells and loss of cytokine receptors were discussed.

## *The Course Directors*

Soldano Ferrone  
*Department of Immunology, Roswell Park Cancer Institute, Buffalo, N Y, USA*  
Rolf Kiessling  
*Immune and Genetherapy Laboratory, Cancer Center Karolinska, Karolinska Hospital, Stockholm, Sweden*

## *The Faculty*

J Dave Barry  
*Wellcome Centre for Molecular Parasitology, University of Glasgow, Glasgow, UK*  
Sven Bergström  
*Department of Microbiology, Umeå Universitet, Umeå, Sweden*  
Pierre G Coulie  
*Unité de Génétique Cellulaire, Université Catholique de Louvain, Brussels, Belgium*  
Soldano Ferrone  
*Department of Immunology, Roswell Park Cancer Institute, Buffalo, N Y, USA*  
Federico Garrido  
*Departamento de Analisis Clinicos, Hospital Universitario Virgen de las Nieves, Granada, Spain*  
Klas Kärre  
*Microbiology & Tumorbiology Center, Karolinska Institutet, Stockholm, Sweden*  
Rolf Kiessling  
*Immune and Genetherapy Laboratory, Cancer Center Karolinska, Karolinska Hospital, Stockholm, Sweden*  
Maria Grazia Masucci  
*Microbiology & Tumorbiology Center, Karolinska Institutet, Stockholm, Sweden*  
Andrew J McMichael  
*MRC Human Immunology Unit, Institute of Molecular Medicine, John Radcliffe Hospital, Oxford, UK*  
Lorenzo Moretta  
*Dipartimento di Oncologia Clinica e Sperimentale, Istituto Nazionale per la Ricerca sul Cancro, Genova, Italy*  
Giorgio Parmiani  
*Unità d'Immunoterapia dei Tumori Umani, Istituto Nazionale Tumori, Milano, Italy*  
Graham Pawelec  
*Tübingen Ageing and Tumour Immunology Group, Section for Transplantation Immunology and Immunohaematology, University of Tübingen, Tübingen, Germany*  
Barbara Seliger  
*III Medizinische Klinik, Abteilung für Innere Medizin - Hämatologie, Johann-Gutenberg-Universität-Mainz, Mainz, Germany*  
Steffen Stenger  
*Insitut für Klinische Mikrobiologie, Immunologie und Hygiene, Friedrich-Alexander-Universität-Erlangen-Nürnberg, Erlangen, Germany*  
John Trowsdale  
*Department of Pathology, Immunology Division, Cambridge University, Cambridge, UK*  
Raymond M Welsh  
*Department of Pathology, University of Massachusetts Medical Center, Worcester, MA, USA*

*The Organizing Committee: Silvia Fontana, Centro di Endocrinologia e Oncologia Sperimentale, CNR, Napoli, Italy; Giuseppina Ruggiero, José Terrazzano & Serafino Zappacosta, Università di Napoli Federico II, Napoli, Italy.*

*Secretariat: Nevin Bayoumi, Nature Classified, Macmillan Magazines Ltd, London, UK; Valeria Lamorgese, Scuola Superiore d'Immunologia Ruggero Ceppellini, Napoli, Italy; Alessandra Saioni & Daniela Giampaolo, Effe Erre Congressi, Napoli, Italy.*

*Sponsorships: Istituto Italiano per gli Studi Filosofici, Naples, Italy; Università di Napoli Federico II; Swedish Cancer Society of Stockholm; Becton Dickinson Italia SpA, Milano; Microglass Scientific Apparatus snc, Napoli; PBI International SpA, Milano; Valter Occhiena Srl, Torino.*

# **“Remembering Environmental Experiences: The Physiological Basis of Memory in the Immune and Nervous Systems”**

Cala Moresca Hotel Club, Capo Miseno, near Naples,  
28 June 28- 2 July 2001

The course reviewed the mechanisms generating and maintaining memory at the cellular and biochemical levels in both the immune and nervous systems, to show similarities and differences between these two interfaces of the host with the external environment. Issues such as the need for repetitive stimulation to maintain memory, the role played by changes in cellular differentiation in providing effective memory, and the biochemistry of the memory state were reviewed both in the immunology and the neurobiology fields, looking for common themes in the biology of learning.

This activity of the Ceppellini School was dedicated to the memory of Alfred Nisonoff, one of the most relevant immunologists of past Century and one of the founders of the School, suddenly deceased on March 12, 2001.

## *The Course Directors*

Ronald N Germain  
*Lymphocyte Biology Section, Laboratory of Immunology, National Institute of Allergy and Infectious Diseases,  
National Institutes of Health, Bethesda, MD, USA*  
Daniele Piomelli  
*360 Med Surge II, University of California, Irvine, CA, USA*

## *The Faculty*

Rafi Ahmed  
*Emory Vaccine Center, Emory University School of Medicine, Atlanta, GA, USA*  
Martin Bachmann  
*Cytos Biotechnology AG, Zürich Schlieren, Switzerland*  
Thomas J Carew  
*Department of Neurobiology and Behavior, University of California, Irvine, CA, USA*  
Tamás F Freund  
*Institute of Experimental Medicine, Hungarian Academy of Sciences, Budapest, Hungary*  
Ronald N Germain  
*Lymphocyte Biology Section, Laboratory of Immunology, National Institute of Allergy and Infectious Diseases,  
National Institutes of Health, Bethesda, MD, USA*  
Gian Luigi Gessa  
*Dipartimento di Neuroscienze, Università di Cagliari, Cagliari, Italy*  
David Gray  
*Institute of Cell, Animal and Population Biology, University of Edinburgh, Ashworth Laboratories, Edinburgh, UK*  
Antonio Lanzavecchia  
*Istituto di Ricerca in Biomedicina, Bellinzona, Switzerland*  
Adrian F Ochsenbein  
*Abteilung Onkologie, Inselspital Bern, Bern, Switzerland*  
Daniele Piomelli  
*360 Med Surge II, University of California, Irvine, CA, USA*  
Andreas Radbruch  
*Deutsches Rheuma-Forschungszentrum Berlin, Berlin, Germany*  
James H Schwartz  
*Center for Neurobiology and Behavior, Columbia University College of Physicians and Surgeons, New York, NY, USA*

*The Organizing Committee:* Silvia Fontana, *Centro di Endocrinologia e Oncologia Sperimentale, CNR, Napoli, Italy;*  
Silvestro Formisano, Veronica Sanna & José Terrazano, *Università di Napoli Federico II, Napoli, Italy.*

*Secretariat:* Nevin Bayoumi, *NatureJobs, Macmillan Magazines Ltd, London, UK;* Pina Ippolito, *Scuola Superiore d'Immunologia Ruggero Ceppellini, Napoli, Italy;* Daniela Giampaolo & Ornella Spada, *Effe Erre Congressi, Napoli, Italy;* Sarah Green, *British Society for Immunology, London, UK.*

*Sponsorships:* Istituto Italiano per gli Studi Filosofici, Naples, Italy; Università di Napoli Federico II; the Campania Region Education Service; Microglass Scientific Apparatus snc, Napoli.

## **“Physiology of the Mucosal Immune Response”**

Cala Moresca Hotel Club, Capo Miseno, near Naples,  
18-22 October 2001

The Course discussed the most recent advances in the mucosal immune response, particularly the intestinal immune response. Emphasis was given to the physiological aspects of the mucosal immune system, but the course covered also vaccine development and some of the diseases characterised by dysregulated mucosal immunity.

Like the preceding course held in the same year, the Course was dedicated to the memory of Alfred Nisonoff.

### *The Course Directors*

Allan McI Mowat  
*Department of Immunology and Bacteriology, University of Glasgow,  
Western Infirmary, Glasgow, UK*  
Paul Garside  
*Department of Immunology and Bacteriology, University of Glasgow,  
Western Infirmary, Glasgow, UK*

### *The Faculty*

Paul Bland  
*Division of Molecular & Cellular Biology, University of Bristol, Bristol, UK*  
Richard S Blumberg  
*Gastroenterology Division, Brigham and Women's Hospital, Harvard University Medical School, Boston, MA, USA*  
Per Brandtzaeg  
*Laboratory for Immunohistochemistry and Immunopathology (LIIPAT), Institute of Pathology, University of Oslo, Rikshospitalet, Oslo, Norway*  
Paul Garside  
*Department of Immunology and Bacteriology, University of Glasgow, Western Infirmary, Glasgow, UK*  
Adrian Hayday  
*Peter Gorer Department of Immunobiology, GKT Guy's Hospital, London, UK*  
Martin F Kagnoff  
*University of California at San Diego, Department of Medicine 0623D, La Jolla, CA, USA*  
Nils Lycke  
*Department of Clinical Immunology, University of Göteborg, Göteborg, Sweden*  
Stefan C Meuer  
*Institut für Immunologie, Ruprecht-Karls-Universität Heidelberg, Heidelberg, Germany*  
Fiona Powrie  
*Sir William Dunn School of Pathology, Oxford University, Oxford, UK*  
Rino Rappuoli  
*IRIS, Chiron SpA, Siena, Italy*  
Paola Ricciardi Castagnoli  
*Dipartimento di Biotecnologie e Bioscienze, Università di Milano-Bicocca, Milano, Italy*  
Jo Viney  
*Immunex Corporation, Seattle, WA, USA*

*The Organizing Committee: Salvatore Auricchio, Silvestro Formisano, & Riccardo Troncone, Università di Napoli Federico II, Napoli, Italy; Francesca Di Rosa, Istituto Internazionale di Genetica e Biofisica, CNR, Napoli, Italy.*

*Secretariat: Nevin Bayoumi, NatureJobs, Macmillan Magazines Ltd, London, UK; Pina Ippolito, Scuola Superiore d'Immunologia Ruggero Ceppellini, Napoli, Italy; Daniela Giampaolo & Ornella Spada, Effe Erre Congressi, Napoli, Italy; Sarah Green, British Society for Immunology, London, UK.*

*Sponsors: European Laboratory for the Investigation of Food-Induced Diseases of the Università di Napoli Federico II; Istituto Italiano per gli Studi Filosofici, Naples, Italy; Research Service of the Campania Region.*



# **“The Immune System in the Protection and Susceptibility to Tuberculosis”**

Cala Moresca Hotel Club, Capo Miseno, near Naples, and Naples,  
13-16 September 2002

The Course reviewed the most recent advances in the immunological aspects of tuberculosis research and treatment. Emphasis was given to the role of innate and acquired immunity to the pathogen, to the molecular biology of mycobacteria, as well as to novel vaccination strategies.

The 10th Anniversary of the Ceppellini School of Immunology was celebrated with a Special Session on Sept 13 in Naples.

## *The Course Directors*

Vittorio Colizzi

*Dipartimento di Biologia, Università di Roma Tor Vergata, Roma, Italy*

Helmut Hahn

*Institut für Infektionsmedizin, Freie Universität Berlin, Berlin, Germany*

Stefan H E Kaufmann

*Abteilung Immunologie, Max-Planck-Institut für Infektionsbiologie, Campus Charité Mitte, Berlin, Germany*

## *The Faculty*

C John Clements

*Medical Officer, Department of Vaccines and Biologicals, World Health Organization, Geneva, Switzerland*

Vittorio Colizzi

*Dipartimento di Biologia, Università di Roma Tor Vergata, Roma, Italy*

Gennaro De Libero

*Experimental Immunology Department of Research, Basel University Hospital, Basel, Switzerland*

Francesco Dieli

*Dipartimento di Biopatologia, Università di Palermo, Palermo, Italy*

Brigitte Gicquel

*Unité de Génétique Mycobactérienne, Institut Pasteur, Paris, France*

Helmut Hahn

*Institut für Infektionsmedizin, Freie Universität Berlin, Berlin, Germany*

Stefan H E Kaufmann

*Abteilung Immunologie, Max-Planck-Institut für Infektionsbiologie, Campus Charité Mitte, Berlin, Germany*

Yukari Carol Manabe

*Division of Infectious Diseases, Johns Hopkins School of Medicine, Baltimore, MD, USA*

Francesca Mariani

*Istituto di Neurobiologia e Medicina Molecolare, CNR, Roma, Italy*

John Joe McFadden

*School of Biomedical and Life Sciences, University of Surrey, Guildford, Surrey, UK*

Robert J North

*Biomedical Research Laboratories, Trudeau Institute, Saranac Lake, USA*

Ian M Orme

*Mycobacteria Research Laboratory, Department of Microbiology, Colorado State University, CO, USA*

Francesco Perna

*Cattedra di Pneumologia, Dipartimento di Medicina Clinica e Sperimentale, Università di Napoli Federico II*

Gaby E Pfyffer

*Swiss National Centre for Mycobacteria, Department of Medical Microbiology, University of Zürich, Switzerland*

Fabrizio Poccia

*Laboratorio di Immunopatologia, I NM I Lazzaro Spallanzani, Roma, Italy*

Sabine Rüscher-Gerdes

*Mykobakteriologie, Research Center Borstel, Borstel, Germany*

Alessandro Sanduzzi

*Dipart. di Medicina Clinica e Sperimentale, Università di Napoli Federico II, Azienda Ospedal. Monaldi, Napoli*

Timo Ulrichs

*Abteilung Immunologie, Max-Planck-Institut für Infektionsbiologie, Berlin, Germany*

*The Organizing Committee: Francesca Di Rosa, Istituto Internazionale di Genetica e Biofisica A Buzzati-Traverso, CNR, Napoli; Silvestro Formisano & Giuseppina Ruggiero, Università di Napoli Federico II, Napoli; Pina Ippolito, Scuola Superiore d'Immunologia Ruggero Ceppellini, Napoli.*

*Secretariat: Daniela Giampaolo & Roberta De Marco, Effe Erre Congressi, Napoli; Silvana Aprile, Istituto Italiano per gli Studi Filosofici, Napoli; Stacey Sheekey, Sales Support Manager, Current Trends, Elsevier Science, London, UK.*

*Sponsorships: Assessorato alla Ricerca of the Campania Region; Federico II University of Naples; Istituto Italiano per gli Studi Filosofici, Naples; UNESCO; Lazzaro Spallanzani Institute, Rome.*

## **“Mucosal Immunity 2: Mucosal Infection and Inflammation”**

Cala Moresca Hotel Club, Capo Miseno, near Naples,  
10-14 October 2002

This Course was intended to move forward the first Ruggero Ceppellini Course on Basic Mechanisms of Mucosal Immunity held in 2001, by exploring the mechanisms involved in the local immune defence of mucosal surfaces against infection, as well as those responsible for chronic inflammatory conditions of the intestine. The Course focussed on the interactions between epithelial cells and pathogenic/commensal organisms and the resulting effects on local inflammatory cells. The pathogenic mechanisms of chronic inflammatory bowel disease, as well as of coeliac disease, were discussed.

### *The Course Director*

Allan Mcl Mowat

*Depart of Immunology and Bacteriology, University of Glasgow, Western Infirmary, Glasgow, Scotland, UK*

### *The Faculty*

Sean P Colgan

*Department of Anaesthesia, Brigham and Women's Hospital, Boston, MA, USA*

Sander van Deventer

*Department of Gastroenterology & Hepatology, AMC C2 - 330 Academic Medical Centre, Amsterdam, The Netherlands*

Charles O Elson

*Division of Gastroenterology and Hepatology, University of Alabama at Birmingham, Birmingham, AL, USA*

Luigi Greco

*Dipartimento di Pediatria, Università di Napoli Federico II, Napoli, Italy*

Richard K Grencis

*Department of Immunology, Stopford Building, University of Manchester, Manchester, UK*

David A van Heel

*Wellcome Trust Centre for Human Genetics, University of Oxford, Oxford, UK*

Denise Kelly

*Rowett Research Institute, Aberdeen, Scotland, UK*

Oliver Liesenfeld

*Institute for Infection Medicine, Department of Medical Microbiology and Immunology of Infection, Benjamin Franklin Medical Center, Free University of Berlin, Berlin, Germany*

Francesco Lizza

*Dipartimento di Medicina Sperimentale e Clinica, Cattedra di Gastroenterologia, Università Magna Græcia, Catanzaro, Italy*

Giovanni Monteleone

*Dipartimento di Medicina Interna, Università di Roma Tor Vergata, Roma, Italy*

Allan Mcl Mowat

*Department of Immunology and Bacteriology, University of Glasgow, Western Infirmary, Glasgow, Scotland, UK*

Ludwig M Sollid

*Rikshospitalet, University of Oslo, Oslo, Norway*

Riccardo Troncone

*Dipartimento di Pediatria, Università di Napoli Federico II, Napoli, Italy*

Mary Jo Wick

*Department of Clinical Immunology, University of Göteborg, Göteborg, Sweden*

*The Organizing Committee: Francesca Di Rosa, Istituto Internazionale di Genetica e Biofisica A Buzzati-Traverso, CNR, Napoli; Silvestro Formisano, Salvatore Auricchio & Riccardo Troncone, Università di Napoli Federico II, Napoli, Pina Ippolito, Scuola Superiore d'Immunologia Ruggero Ceppellini, Napoli.*

*Secretariat: Daniela Giampaolo and Associates, Effe Erre Congressi, Napoli; Stacey Sheekey, Sales Support Manager, Current Trends, Elsevier Science, London, UK; Doreen Shand; British Society for Immunology, London, UK.*

*Sponsorships: European Laboratory for the Investigation of Food Induced Diseases of the Federico II University of Naples; Assessorato alla Ricerca of the Campania Region; Istituto Italiano per gli Studi Filosofici, Naples; Società Italiana di Storia Patria di Terra di Lavoro; Microtech srl, Naples.*

## **“The Stem Cell: From Theory to Clinics”**

Lloyd's Baia Hotel, Vietri sul Mare, near Salerno,  
16-20 October 2003

The recent discovery of stem cells in many human tissues has raised high hope for the cure of many genetic and acquired diseases. This course addressed the basic concepts of stem cells, including topics such as human embryonic stem cell lines, stem cell plasticity, molecular control of stem cell proliferation, differentiation and plasticity, stem cell manipulation in vitro. The Course included a round table discussion on “Ethical issues in human embryonic cell research”.

### *The Course Directors*

Anna Rita Migliaccio

*Laboratorio di Biochimica Clinica, Istituto Superiore di Sanità, Roma, Italy*

Piero Musiani

*Sezione di Anatomia Patologica, Dipartimento di Oncologia e Neuroscienze, Università di Chieti-Pescara G D'Annunzio, Ospedale Clinizzato SS. Annunziata, Chieti, Italy*

### *The Faculty*

Dirk W van Bekkum

*Crucell B V, Leiden, The Netherlands*

Emer Clarke

*StemCell Technologies Inc, Vancouver, BC, Canada*

Seth Corey

*Department of Pediatrics, UT-MD Anderson Cancer Center, Houston, TX, USA*

Tariq Enver

*Weatherall Institute of Molecular Medicine, University of Oxford, John Radcliffe Hospital, Oxford, UK*

Willem E Fibbe

*Department of Haematology, Leiden University Medical Centre, Leiden, The Netherlands*

Rosaria Giordano

*Centro Trasfusionale e di Immunologia dei Trapianti, Ospedale Maggiore Policlinico, Milano, Italy*

Luigi Giaccaia

*Instrumentation Laboratory SpA, Roma, Italy*

Jacques A Hatzfeld

*Lab de Biologie des Cellules Souches Somatiques Humaines, UPR 1983 du CNRS, IFC1, Villejuif, France*

Fulvio Mavilio

*Dipartimento di Scienze Biomediche, Università di Modena, Modena, Italy*

Anna Rita Migliaccio

*Laboratorio di Biochimica Clinica, Istituto Superiore di Sanità, Roma, Italy*

Giovanni Migliaccio

*Laboratorio di Biologia Cellulare, Istituto Superiore di Sanità, Roma, Italy*

Piero Musiani

*Dipart. di Oncologia e Neuroscienze, Università di Chieti-Pescara G D'Annunzio, Chieti, Italy*

Maria Grazia Pallavicini

*Mount Zion Cancer Center, University of California at San Francisco, San Francisco, CA, USA*

Graziella Pellegrini

*Laboratorio di Cellule Staminali Epiteliali, Fondazione Banca degli Occhi del Veneto, Ospedale Civile SS Giovanni e Paolo, Venezia, Italy*

Wanda Piacibello

*Dipartimento di Scienze Oncologiche, Istituto per la Ricerca sul Cancro, Università di Torino, Candiolo TO*

Raffaele Prodomo

*Facoltà di Giurisprudenza, Seconda Università di Napoli, S Maria Capua Vetere CE, Italy*

Rodolfo Quarto

*Dipartimento di Oncologia, Biologia e Genetica, Università di Genova, Genova, Italy*

Peter Wernet

*Institute of Transplantation Diagnostics and Cell Therapeutics, Heinrich-Heine Universität, Düsseldorf,*

*The Organizing Committee: Francesca Di Rosa, Institute of Genetics and Biophysics A Buzzati Traverso, CNR, Napoli, Italy; Maurizio Bifulco, Università di Salerno, Fisciano SA, Italy; Silvestro Formisano, & Francesco Salvatore, Università di Napoli Federico II, Napoli, Italy.*

*Secretariat: Pina Ippolito & Riccardo Zappacosta, Ceppellini School Secretarial Office.*

*Sponsorships: CEINGE Biotecnologie Avanzate Scarl, Naples; Federico II University of Naples; Assessorato alla Ricerca of the Campania Region; Istituto Nazionale Tumori Fondazione Pascale, Naples; Istituto Italiano per gli Studi Filosofici, Naples; Microtech srl, Naples, from Becton Dickinson SpA, Buccinasco (Milan) and from Instrumentation Laboratory, Rome.*

## **“Innate Immunity in Self and Infectious Non-Self Recognition”**

Cala Moresca Hotel Club, Capo Miseno, near Naples,  
10-14 March 2005

The course reviewed the most advanced knowledge about innate immune mechanisms at the genetic, cellular and molecular levels. Addressed topics included the innate immune cells and receptors in viral, bacterial, fungal and parasitic infections; the genomic analysis of innate immunity receptor families, the pathogen recognition by insect vectors of human infectious diseases, the molecules recognising pathogen Toll and non-Toll-like, the signalling events leading to NK cell activation, the NK cell-associated receptors and their role in immune regulation, the interactions between innate and adaptive immunity, the possibility of recognizing cancer with innate receptors, and new therapeutic approaches to infectious diseases.

### *The Course Directors*

Gregory J Bancroft  
*Immunology Unit, Department of Infectious & Tropical Diseases, London School of Hygiene and Tropical Medicine, London, UK*  
Roberto Biassoni  
*Dipartimento di Medicina Molecolare, Istituto Giannina Gaslini, Genova, Italy*  
Ennio Carbone  
*Dipartimento di Medicina Clinica e Sperimentale, Università Magna Græcia, Catanzaro, Italy*

### *The Faculty*

Siamon Gordon  
*Sir William Dunn School of Pathology, University of Oxford, UK*  
Klas Kärre  
*Microbiology and Tumor Biology Center, Karolinska Institutet, Stockholm, Sweden*  
Paul M Kaye  
*Immunology and Infection Unit, Department of Biology and The Hull York Medical School, The University of York, York, U K*  
Elena A Levashina  
*European Molecular Biology Laboratory, Heidelberg, Germany and UPR 9022 du CNRS, Institut de Biologie Moléculaire et Cellulaire (IBMC), Strasbourg, France*  
Miguel López-Botet  
*Molecular Immunopathology Unit, Department of Health and Experimental Sciences (DCEXS), Universitat Pompeu Fabra, Barcelona, Spain*  
Alberto Mantovani  
*Cattedra di Patologia Generale, Università di Milano and Dipartimento di Immunologia e Biologia Cellulare, Istituto di Ricerche Farmacologiche Mario Negri, Milano, Italy*  
Lorenzo Moretta  
*Centro di Eccellenza per le Ricerche Biomediche, Università di Genova and Dipartimento di Medicina Sperimentale, Istituto Giannina Gaslini, Genova, Italy*  
Anne O'Garra  
*Division of Immunoregulation, National Institute for Medical Research, London, UK*  
Peter Parham  
*Department of Structural Biology, Stanford University School of Medicine, Stanford, CA, USA*  
Angela Santoni  
*Dipartimento di Medicina Sperimentale e Patologia, Università di Roma La Sapienza, Roma, Italy*  
John Trowsdale  
*Department of Pathology, University of Cambridge, Cambridge, UK*  
Andrea Velardi  
*Divisione di Ematologia ed Immunologia Clinica, Dipartimento di Medicina Clinica e Sperimentale, Università di Perugia, Perugia, Italy*

*The Organizing Committee:* Silvestro Formisano, Giuseppina Ruggiero & José Terrazzano, *Università di Napoli Federico II, Napoli, Italy*; Pina Ippolito, *Scuola Superiore d'Immunologia Ruggero Ceppellini, Napoli, Italy*.

*Secretariat:* Alessandra Saioni, Mia Liotti and Associates, *Effe Erre Congressi, Napoli, Italy*; Sille Opstrup, *NatureJobs, Macmillan Magazines Ltd, London, UK*.

*Sponsorships:* Magna Græcia University of Catanzaro; Federico II University of Naples; Microtech srl, Naples, Instrumentation Laboratory SpA, Rome, Italy, and Becton-Dickinson SpA, Buccinasco Milan.



## **"The recrudescence of an old infectious disease: Tuberculosis"**

Congress Center University of Naples "Federico II", Naples  
2-5 May 2007

The course reviewed an infectious disease of increasing global concern, namely tuberculosis. In recent years, strains of *Mycobacterium tuberculosis* have developed resistance to classical drug therapies which limit its pathological burden. The course reviewed this recent health care emergency with a focus on the genetics of *Mycobacterium*, the emergence of new drug-resistant strains, and the proteomics of the *M. tuberculosis* cell wall. The immune response to *M. tuberculosis* was discussed in-depth by reviewing state-of-the-art research on mechanisms involved in immune control of *M. tuberculosis* infection. In addition, novel anti-*M. tuberculosis* vaccines and drug development strategies were discussed.

### *The Course Director*

Stefan H.E. Kaufmann  
*Max Planck Institute for Infection Biology, Department of Immunology, Berlin, Germany*

### *The Faculty*

Fabio Bagnoli  
*Dipartimento di Energetica Università di Firenze Firenze, Italy*

Eric Boettger  
*Danish Centre for Experimental Parasitology, The Royal Veterinary and Agricultural University Frederiksberg C, Denmark*

Roland Brosch  
*Unité de Génétique Moléculaire Bactérienne, Institut Pasteur, Paris Cedex, France*

Vittorio Colizzi  
*Dipartimento di Biologia, Università Tor Vergata, Roma, Italy*

Mamadou Daffé  
*Institute of Pharmacology and Structural Biology (CNCS), Toulouse Cedex, France*

Francesco Dieli  
*Dipartimento di Biopatologia e Metodologie Biomediche, Università di Palermo, Palermo, Italy*

Hazel Dockrell  
*Department of Infectious and Tropical Diseases, London School of Hygiene & Tropical Medicine, London, UK*

Brigitte Gicquel  
*Unit of Mycobacterial Genetics, Institut Pasteur, Paris, France*

Eileen Hoal  
*DST/NRF Centre of Excellence for Biomedical Tuberculosis Research MRC Centre for Molecular and Cellular Biology; Division of Molecular Biology and Human Genetics, Faculty of Health Sciences, Stellenbosch University, Tygerberg, South Africa*

Mark Jacobsen  
*Department of Immunology, Max Planck Institute for Infection Biology, Berlin, German*

Stefan H.E. Kaufmann  
*Max Planck Institute for Infection Biology, Department of Immunology, Berlin, Germany*

Stefan Niemann  
*National Reference Center for Mycobacteria, Forschungszentrum Borstel, Borstel, Germany*

Rino Rappuoli  
*Novartis Vaccines, Siena, Italy*

Paul van Helden  
*Division of Molecular Biology and Human Genetics Stellenbosch University, Tygerberg, South Africa*

D. van Soolingen  
*National Mycobacteria Reference Laboratory, National Institute of Public Health and the Environment, 3720 Bilthoven, The Netherlands*

Francois Spertini  
*IAL, Faculté de Biologie et Médecine University of Losanne, Losanne, Suisse*

*The Organizing Committee:* Antonio Di Giacomo, Azienda Ospedaliera V Monaldi, Napoli, Italy; Ennio Carbone, Magna Grecia University, Catanzaro; Enrico Avvedimento, Guido Rossi & Silvestro Formisano, Università di Napoli Federico II, Napoli, Italy.

*Secretariat:* Effe Erre Congressi, Napoli, Italy.

*Sponsorships:* European Federation of Immunological Societies (EFIS)- The European Journal of Immunology (EJI); Dipartimento di Biologia e Patologia Cellulare e Molecolare, Università di Napoli Federico II; Magna Grecia University, Catanzaro; Istituto Italiano per gli Studi Filosofici; International Doctorate Program in Molecular Oncology and Endocrinology; Research Service of the Campania Region.

## **“Tumour Immune Escape 2008”**

Circolo dei Forestieri, Sorrento near Naples,  
16-18 October, 2008

The Course brought together scientists with various points of view on anti-cancer immune surveillance. During the course, several observations on the role of immunity in controlling tumor progression were reviewed and discussed. Mechanisms involved in immune subversion by established tumors were discussed, as well as novel anti-cancer vaccines and drug development strategies.

### *The Course Directors*

Soldano Ferrone  
*Hilman Cancer Center, Pittsburgh, USA*  
Barbara Seliger  
*Martin Luther University Institute of Medical Immunology, Halle, Germany*

### *The Faculty*

Hinrich Abken  
*Tumor Genetics and Cell Biology Klinik für Innere Medizin Köln University, Köln, Germany*  
Thomas Blankenstein  
*Max-Delbrück-Zentrum für Molekulare Medizin Berlin-Buch, Germany*  
Vincenzo Bronte  
*Department of Oncology and Surgical Sciences, Padova, Italy*  
Edgardo D. Carosella  
*CEA-DSV-DRM Hopital Saint-Louis Institut Universitaire d'Hématologie, Paris, France*  
Federica Cavallo  
*Molecular Biotechnology Center, Department of Clinical and Biological Sciences, University of Turin, Torino, Italy*  
Soldano Ferrone  
*Hilman Cancer Center, Pittsburgh, USA*  
Rolf Kiessling  
*Cancer Center Karolinska Immune and Gene therapy Laboratory (IGT) Karolinska Hospital, Stockholm, Sweden*  
Joël LeMaoult  
*CEA-DSV-DRM Hopital Saint-Louis Institut Universitaire d'Hématologie, Paris, France*  
Barbara Seliger  
*Martin Luther University Institute of Medical Immunology, Halle, Germany*  
Andrea Velardi  
*Sezione di Ematologia e Immunologia Clinica University of Perugia, Perugia, Italy*  
Theresa Whiteside  
*University of Pittsburgh Cancer Institute Pittsburgh, USA*

*The Organizing Committee:* Antonio Di Giacomo, Azienda Ospedaliera V Monaldi, Napoli, Italy; Ennio Carbone, Magna Grecia University, Catanzaro; Enrico Avvedimento, Guido Rossi & Silvestro Formisano, Università di Napoli Federico II, Napoli, Italy.

*Secretariat:* Effe Erre Congressi, Napoli, Italy.

*Sponsorships:* European Federation of Immunological Societies (EFIS)- The European Journal of Immunology (EJI); Dipartimento di Biologia e Patologia Cellulare e Molecolare, Università di Napoli Federico II; Magna Grecia University, Catanzaro; Istituto Italiano per gli Studi Filosofici; International Doctorate Program in Molecular Oncology and Endocrinology; Research Service of the Campania Region.

## EFIS-EJI course on “The role of B Cells in the Physiology and Pathology of the Immune System”

Circolo dei Forestieri, Sorrento near Naples,  
5-7 November 2009

This Course dealt with recent advancements in the B cell field. Topics included B cell development, B cell diversification and memory formation, B cell involvement in autoimmune diseases.

### *The Course Directors*

Antonio La Cava  
*Lupus Research Laboratory, University of California Los Angeles, California, USA*  
Andreas Radbruch  
*German Rheumatology Research Center, Leibniz Research Institute, Berlin, Germany*

### *The Faculty*

Marina Botto  
*Rheumatology Section, Faculty of Medicine, Imperial College, London, UK*  
Rita Carsetti  
*Ospedale Pediatrico Bambino Gesù, Rome, Italy*  
Falk Hiepe  
*Charité University of Medicine, Berlin, Germany*  
Antonio La Cava  
*Lupus Research Laboratory, University of California Los Angeles, California, USA*  
Ian MacLennan  
*MRC Centre for Immune Regulation, University of Birmingham, Birmingham, UK*  
Fritz Melchers  
*Max Planck Institute for Infection Biology, Berlin, Germany*  
Andreas Radbruch  
*German Rheumatology Research Center, Leibniz Research Institute, Berlin, Germany*  
Antonius Rolink  
*Developmental and Molecular Immunology, University of Basel, Switzerland*  
Elisabetta Traggiai  
*Institute G. Gaslini, Genova, Italy*  
Jean-Claud Weill  
*Paris Descartes University, Site Necker-Enfants Malade, Paris, France*

*The Organizing Committee:* Silvia Fontana (President, *Scuola Superiore d'Immunologia Ruggero Ceppellini*), Centro di Endocrinologia e Oncologia Sperimentale, CNR, Napoli, Italy; Ennio Carbone, Magna Graecia University, Catanzaro; Guido Rossi & Silvestro Formisano, Università di Napoli Federico II, Napoli, Italy; Antonio La Cava, Lupus Research Laboratory, University of California Los Angeles, California, USA; Tricia Reynolds, Intercultural Relations Center, Naples, Italy.

*Secretariat:* Effe Erre Congressi, Napoli, Italy.

*Sponsorships:* European Federation of Immunological Societies (EFIS) - The European Journal of Immunology (EJI); the Federico II University of Naples; the Magna Graecia University of Catanzaro; the Research Service of Campania Region; the Istituto Italiano per gli Studi Filosofici, Naples, Italy; the Department of Cellular and Molecular Biology and Pathology, Federico II University of Naples; the International Doctorate Program in Molecular Oncology and Endocrinology of Naples; the International Doctorate Program in Molecular Oncology, Immunology and Development of New Therapy of Catanzaro, Italy.

## EFIS-EJI course on “Innovative strategies for vaccines: malaria, tuberculosis, HIV”

Circolo dei Forestieri, Sorrento near Naples,  
4-6 November 2010

The Course highlighted the current views and state of the art of vaccine production against three dangerous infectious diseases, i.e. malaria, tuberculosis, HIV/AIDS, that taken together are responsible for millions of deaths globally. Future developments in the area were discussed.

### *The Course Directors*

Stefan H.E. Kaufmann  
*Max Planck Institute for Infection Biology, Department of Immunology, Berlin, Germany*  
Rino Rappuoli  
*Novartis Vaccines, Siena, Italy*  
Giuseppe del Giudice  
*Novartis Vaccines, Siena, Italy*

### *The Faculty*

W. Ripley Ballou  
*Clinical Research & Translational Science, GSK Vaccine, Rixensart, Belgium*  
Giuseppe del Giudice  
*Novartis Vaccines, Siena, Italy*  
Peter Andersen  
*Department of Infectious Disease Immunology, Statens Serum Institute, Copenhagen, Denmark*  
Susan W Barnett  
*Novartis Vaccines and Diagnostics, Cambridge, MA, USA*  
Bertram L Jacobs  
*Arizona State University, Tempe, Arizona, USA*  
Stefan H.E. Kaufmann  
*Max Planck Institute for Infection Biology, Department of Immunology, Berlin, Germany*  
Kai Matuschewski  
*Max Planck Institute for Infection Biology, Parasitology Unit, Berlin, Germany*  
Martin OC Ota  
*Bacterial Diseases Programme, MRC Laboratories, Banjul, Gambia*  
Rino Rappuoli  
*Novartis Vaccines, Siena, Italy*  
Eleanor M Riley  
*Department of Infectious and Tropical Diseases, London School of Hygiene and Tropical Medicine, London, UK*  
Alexander von Gabain  
*Intercell AG, Vienna, Austria*  
Barry Walker  
*National Institute for Biological Standards and Controls, Potters Bar, Hertfordshire, UK.*  
Hedda Wardemann  
*Molecular Immunology Research Group, Max Planck Institute for Infection Biology, Berlin, Germany*  
Fidel Zavala  
*Johns Hopkins Malaria Research Institute and Department of Molecular Microbiology and Immunology, Bloomberg School of Public Health, Baltimore, Maryland, USA*

*The Organizing Committee:* Silvia Fontana (President, *Scuola Superiore d'Immunologia Ruggero Ceppellini*), *Centro di Endocrinologia e Oncologia Sperimentale, CNR, Napoli, Italy*; Antonio Di Giacomo (*Azienda Ospedaliera V Monaldi, Napoli, Italy*); Ennio Carbone, *Magna Grecia University, Catanzaro, Italy*; Guido Rossi & Silvestro Formisano, *Università di Napoli Federico II, Napoli, Italy*; Tricia Reynolds, *Intercultural Relations Center, Naples, Italy*.

*Secretariat:* Effe Erre Congressi, Napoli, Italy.

*Sponsorships:* European Federation of Immunological Societies (EFIS)- The European Journal of Immunology (EJI); Dipartimento di Biologia e Patologia Cellulare e Molecolare, Università di Napoli Federico II; Magna Grecia University, Catanzaro; Novartis, Siena, Italy; Intercell AG, Austria; GlaxoSmithkline, Belgium.



## EFIS-EJI course on “Innovative strategies to prevent transplant rejection”

Circolo dei Forestieri, Sorrento near Naples,  
26-29 October, 2011

The Course reviewed and discussed the current views on basic concepts of Immunology of transplantation, as well as clinical perspectives in transplantation and new mechanisms of tolerance induction.

### *The Course Directors*

Robert Lechler  
*Medical Research Council Centre for Transplantation, King's College London, London, UK*  
Giovanna Lombardi  
*Medical Research Council Centre for Transplantation, King's College London, London, UK*  
Randolph J Noelle  
*Medical Research Council Centre for Transplantation, King's College London, London, UK*

### *The Faculty*

Stephen Cobbold  
*Sir William Dunn School of Pathology, University of Oxford, Oxford, UK*  
Anthony Dorling  
*Medical Research Council Centre for Transplantation, King's College London, London, UK*  
Maria P. Hernandez-Fuentes  
*Medical Research Council Centre for Transplantation, King's College London, London, UK*  
Rachel Hilton  
*Medical Research Council Centre for Transplantation, King's College London, London, and NIHR Biomedical Research Centre at Guy's and St Thomas' Hospital and King's College London, London, UK.*  
Claudia Kemper  
*Division of Immunology, Infection and Inflammatory Diseases, King's College London, Medical Research Council Centre for Transplantation, Guy's Hospital, London, UK*  
Robert Lechler  
*Medical Research Council Centre for Transplantation, King's College London, London, UK*  
Giovanna Lombardi  
*Medical Research Council Centre for Transplantation, King's College London, London, UK*  
Randolph J Noelle  
*Medical Research Council Centre for Transplantation, King's College London, London, UK*  
Alberto Sanchez-Fueyo  
*Liver Transplant Unit, Hospital Clinic Barcelona, IDIBAPS, CIBEREHD, University of Barcelona, Barcelona, Spain.*  
Elizabeth Simpson  
*Division of Immunology and Inflammation, Imperial College, London, UK*  
Richard D Smith  
*Department of Public Health and Policy, London School of Hygiene and Tropical Medicine, London, UK*  
Terry B Strom  
*The Transplant Institute, Beth Israel Deaconess Medical Center, Harvard Medical School, Boston, MA, USA*  
Laurence A Turka  
*The Transplant Institute, Beth Israel-Deaconess Medical Center, Harvard Medical School, Boston, MA, USA.*  
Kathryn Wood  
*Nuffield Department of Surgery, University of Oxford, Oxford, UK*

*The Organizing Committee:* Silvia Fontana (President, Scuola Superiore d'Immunologia Ruggero Ceppellini), Centro di Endocrinologia e Oncologia Sperimentale, CNR, Napoli, Italy; Antonio Di Giacomo, Azienda Ospedaliera V Monaldi, Napoli, Italy; Ennio Carbone, Magna Grecia University, Catanzaro, Italy; Silvestro Formisano, Università di Napoli Federico II, Napoli, Italy; Tricia Reynolds, Intercultural Relations Center, Naples, Italy.

*Secretariat:* Effe Erre Congressi, Napoli, Italy.

*Sponsorships:* European Federation of Immunological Societies (EFIS)- The European Journal of Immunology (EJI); Dipartimento di Biologia e Patologia Cellulare e Molecolare, Università di Napoli Federico II; Magna Grecia University, Catanzaro; Miltenyi, UK; Miltenyi, Germany; Novartis, UK.

## EFIS-EJI course on “Innate Immunity 2012: from evolution to revolution”

Circolo dei Forestieri, Sorrento near Naples,  
31 October-4 November 2012

Innate immunity is essential for anti-pathogen protection, but it also contributes to the pathogenesis of many diseases. The Course reviewed and discussed the enormous body of work that recently revolutionized the field, including for example the discovery of innate receptors, the increasing knowledge on macrophage heterogeneity, etc.

### *The Course Directors*

Klas Kärre  
*Karolinska Institutet, Stockholm, Sweden*  
Lorenzo Moretta  
*Istituto Giannina Gaslini, Genova, Italy*  
Ennio Carbone  
*Magna Grecia University, Catanzaro, Italy*

### *The Faculty*

Niels Borregaard  
*The Granulocyte Research Laboratory, Department of Hematology, National University Hospital, University of Copenhagen, Denmark*  
Ennio Carbone  
*Magna Grecia University, Catanzaro, Italy*  
Susanna Cardell  
*Dept of Microbiology and Immunology, Institute of Biomedicine, University of Goteborg, Sweden*  
Jonathan Ewbank  
*Centre d'Immunologie de Marseille-Luminy, Marseille, France*  
Siamon Gordon  
*Sir William Dunn School of Pathology, University of Oxford, Oxford, UK*  
Francesca Granucci  
*Department of Biotechnology and Bioscience, University of Milano-Bicocca, Milan, Italy*  
Dieter Kabelitz  
*Institute of Immunology, University of Kiel, Germany*  
Klas Kärre  
*Karolinska Institutet, Stockholm, Sweden*  
Ed Lavelle  
*Adjuvant Research Group, School of Biochemistry and Immunology, Trinity Biomedical Sciences Institute, Trinity College, Dublin, Ireland*  
Alberto Mantovani  
*Istituto Clinico Humanitas, University of Milan, Italy*  
Lorenzo Moretta  
*Istituto Giannina Gaslini, Genova, Italy*  
Christian Münz  
*Viral Immunobiology, Institute of Experimental Immunology, University of Zürich, Switzerland*  
Jean-Marc Reichhart  
*UPR 9022 CNRS, Strasbourg, France*  
Francesco Tedesco  
*Department of Life Sciences, University of Trieste, Italy*  
Andrea Velardi  
*Bone Marrow Transplantation Programme, University of Perugia, Italy*

*The Organizing Committee:* Silvia Fontana (President, *Scuola Superiore d'Immunologia Ruggero Ceppellini*), Centro di Endocrinologia e Oncologia Sperimentale, CNR, Napoli, Italy; Antonio Di Giacomo, *V Monaldi Hospital, Napoli, Italy*; Ennio Carbone, *Magna Grecia University, Catanzaro*; Tricia Reynolds, *Intercultural Relations Center, Naples, Italy*.

*Secretariat:* Effe Erre Congressi, Napoli, Italy.

*Sponsorships:* European Federation of Immunological Societies (EFIS)- The European Journal of Immunology (EJI); Gender Equality and Career Development Committee, International Union of Immunological Societies (IUIS); The Magna Grecia University of Catanzaro, Italy; the Department of Cellular and Molecular Biology and Pathology, Federico II University of Naples; the International Doctorate Program in Molecular Oncology, Immunology and Development of New Therapy of Catanzaro, Italy.

# EFIS-EJI Course on “Novel Vaccination Strategies Against the Three Major Killers: the Latest News on Malaria, Tuberculosis, HIV/AIDS and Vaccine Development in general”

Restoring Ancient Stabiae–Vesuvian Institute, Castellammare di Stabia, near Naples,  
16 - 20 Oct 2013

This Course was intended to move forward the 2010 Ruggero Ceppellini Course on “Innovative strategies for vaccines: malaria, tuberculosis, HIV”, by dissecting host–pathogen interactions, discussing the recent improvements on vaccine design, and exploring new methods for identification of biomarkers of protective responses, among other topics.

## *The Course Directors*

Stefan HE Kaufmann

*Max Planck Institute for Infection Biology, Department of Immunology, Berlin, Germany*

Francesca Chiodi

*Department of Microbiology, Tumor and Cell Biology, Karolinska Institutet, Stockholm, Sweden*

## *The Faculty*

W. Ripley Ballou

*Clinical Research & Translational Science, GSK Vaccine, Rixensart, Belgium*

Francesca Chiodi

*Department of Microbiology, Tumor and Cell Biology, Karolinska Institutet, Stockholm, Sweden*

Mark Cotton

*Faculty of Medicine and Health Sciences, Stellenbosch University, Tygerberg, South Africa*

Willem Hanekom

*University of Cape Town, South Africa*

Ali Harandi

*Department of Microbiology and Immunology, Sahlgrenska Academy at University of Gothenburg, Sweden*

Stefan HE Kaufmann

*Max Planck Institute for Infection Biology, Department of Immunology, Berlin, Germany*

Elena A Levashina

*Max Planck Institute for Infection Biology, Department of Vector Biology, Berlin, Germany*

Kai Matuschewski

*Max Planck Institute for Infection Biology, Parasitology Unit, Berlin, Germany*

Ndung'u Thumbi

*HIV Pathogenesis Programme, Doris Duke Medical Research Institute, and the KwaZulu-Natal Research Institute for Tuberculosis and HIV (K-RITH), University of KwaZulu-Natal, Durban, South Africa*

Anne O'Garra

*Division of Immunoregulation, MRC National Institute for Medical Research and Faculty of Medicine, Imperial College, London, UK*

Rino Rappuoli

*Novartis Vaccines and Diagnostics, Siena, Italy*

Federica Sallusto

*Institute for Research in Biomedicine, University of Lugano (USI), Bellinzona, Switzerland*

Marita Troye-Blomberg

*The Wenner-Gren Institute, Stockholm University, Stockholm, Sweden*

January Weiner

*Max Planck Institute for Infection Biology, Department of Immunology, Berlin, Germany*

Robin A Weiss

*Division of Infection & Immunity, University College London, London, UK*

Chris Wilson

*Director Discovery & Translational Science, Bill and Melinda Gates Foundation, Seattle, WA, USA*

*The Organizing Committee: Silvia Fontana (President, Scuola Superiore d'Immunologia Ruggero Ceppellini), Centro di Endocrinologia e Oncologia Sperimentale, CNR, Napoli, Italy; Antonio Di Giacomo, V Monaldi Hospital, Napoli, Italy; Tricia Reynolds, Intercultural Relations Center, Naples, Italy.*

*Secretariat: Effe Erre Congressi, Napoli, Italy.*

*Sponsorships: European Federation of Immunological Societies (EFIS)- The European Journal of Immunology (EJI); Educational Committee, International Union of Immunological Societies (IUIS); The Bill & Melinda Gates Foundation; the Department of Cellular and Molecular Biology and Pathology, Federico II University of Naples; The Journal of Internal Medicine; Novartis vaccines and Diagnostics; Mabtech.*

## EFIS-EJI course on “The Maternal Immune System in Pregnancy”

Restoring Ancient Stabiae–Vesuvian Institute, Castellammare di Stabia, near Naples,  
6-9 Dec 2014

This course reviewed current views on immunological mechanisms allowing pregnant mothers to tolerate their fetus and at the same time to display anti-pathogen protection. Topics included the influence of variation of immune system genes (e.g. HLA, KIR) on human reproduction, and the role of fetal immune system, among others.

### *The Course Directors*

Francesco Colucci  
*Department of Obstetrics and Gynaecology, University of Cambridge, UK*  
Ashley Moffett (Cambridge)  
*Department of Pathology, University of Cambridge, UK*

### *The Faculty*

Maria-Luisa Alegre  
*Section of Rheumatology, Department of Medicine, University of Chicago, IL, USA*  
Ennio Carbone  
*Università Magna Graecia, Catanzaro, Italy and Karolinska Institutet, Stockholm, Sweden*  
Francesco Colucci  
*Department of Obstetrics and Gynaecology, University of Cambridge, UK*  
Anthony W De Tomaso  
*Department of Molecular, Cellular and Developmental Biology, University of California, Santa Barbara, CA, USA*  
Alison Elliot  
*MRC/UVRI Uganda Research Unit on AIDS, Entebbe, Uganda and London School of Hygiene and Tropical Medicine, London, UK*  
Thomas F Gajewski  
*Department of Pathology, University of Chicago Medical Center, IL, USA*  
Jacob Michaëlsson  
*Center for Infectious Medicine, Karolinska Institutet, Stockholm, Sweden*  
Ashley Moffett (Cambridge)  
*Department of Pathology, University of Cambridge, UK*  
Annette Nakimuli  
*Department of Obstetrics and Gynaecology, Makerere University, Kampala, Uganda*  
Angela Santoni  
*Department of Molecular Medicine, Sapienza University, Rome, Italy*  
Elizabeth Simpson  
*Division of Immunology and Inflammation, Imperial College, London, UK*  
Tamara Tilburgs  
*Department of Stem Cells and Regenerative Biology, Harvard University, Cambridge, MA, USA*  
John Trowsdale  
*Division of Immunology, Department of Pathology, University of Cambridge, UK*

*The Organizing Committee: Silvia Fontana (President, Scuola Superiore d'Immunologia Ruggero Ceppellini), Centro di Endocrinologia e Oncologia Sperimentale, CNR, Napoli, Italy; Antonio Di Giacomo, V Monaldi Hospital, Napoli, Italy; Tricia Reynolds, Scientific Secretariat, Scuola Superiore d'Immunologia Ruggero Ceppellini.*

*Secretariat: Effe Erre Congressi, Napoli, Italy.*

*Sponsorships: European Federation of Immunological Societies (EFIS)- The European Journal of Immunology (EJI); the Educational Committee and the Gender Equality and Career Development Committee, International Union of Immunological Societies (IUIS); The Bill & Melinda Gates Foundation; the Department of Cellular and Molecular Biology and Pathology, Federico II University of Naples.*



## EFIS-EJI Course on “Treg Biology and Metabolism”

Grand Hotel Oriente, Naples,  
5-6 November 2015

Regulatory T cell (Treg) biology was reviewed, particularly in the context of anti-transplant immunity. Treg metabolic programs, and their influence on Treg function were some of the topics discussed in this Course.

### *The Course Directors*

Fiona Powrie  
*Kennedy Institute of Rheumatology, University of Oxford, Oxford, UK*  
Giovanna Lombardi  
*MRC Centre for Transplantation, King's College London, London, UK*  
Giuseppe Matarese  
*Laboratorio di Immunologia, Istituto di Endocrinologia e Oncologia Sperimentale, CNR, Napoli, Italy*

### *The Faculty*

Hogbo Chi  
*Department of Immunology, St Jude Children's Research Hospital, Memphis, TN, USA*  
Marika Falcone  
*IRCSS San Raffaele Hospital, Milan, Italy*  
Giovanna Lombardi  
*MRC Centre for Transplantation, King's College London, London, UK*  
Graham Lord  
*MRC Centre for Transplantation, King's College London, London, UK*  
Federica Marelli-Berg  
*William Harvey Research Institute, Barts, and London School of Medicine and Dentistry, London, UK*  
Giuseppe Danilo Dorata  
*Department of Pharmacological and Biomolecular Sciences, University of Milan, Milan, Italy*  
Erika L. Pearce  
*Department of Immunometabolism, Max Planck Institute for Immunobiology and Epigenetics, Freiburg, Germany*  
Fiona Powrie  
*Kennedy Institute of Rheumatology, University of Oxford, Oxford, UK*

*The Organizing Committee:* Silvia Fontana (President, *Scuola Superiore d'Immunologia Ruggero Ceppellini*), *Istituto di Endocrinologia e Oncologia Sperimentale, CNR, Napoli, Italy*; Antonio Di Giacomo, *V Monaldi Hospital, Napoli, Italy*; Tricia Reynolds, *Scientific Secretariat, Scuola Superiore d'Immunologia Ruggero Ceppellini*.

*Secretariat:* Effe Erre Congressi, Napoli, Italy.

*Sponsorships:* European Federation of Immunological Societies (EFIS)- The European Journal of Immunology (EJI); the Gender Equality and Career Development Committee, International Union of Immunological Societies (IUIS); the Department of Translational Medicine, Federico II University of Naples; European Research Council (ERC); .M&M Biotech; Space Import & Export srl; Seahorse Bioscience; Euroclone SpA.

## EFIS-EJI Course on “Metchnikoff’s Legacy: tissue Phagocytes and Functions”

Stazione Zoologica “Anton Dohrn”, Naples,  
12-14 October 2016

This Course brought together scientists with various perspectives on macrophages, their heterogeneity, and their role in immune responses, e.g. against *M. tuberculosis*. The pioneer studies of Metchnikoff, who first described phagocytosis, were mentioned and discussed in the light of current literature in the field.

### *The Course Directors*

Siamon Gordon  
*Sir William Dunn School of Pathology, University of Oxford, Oxford, UK*  
Stefan H E Kaufmann  
*Max-Planck-Institut für Infektionsbiologie, Berlin, Germany*  
Fernando Martinez Estrada  
*University of Surrey, Guildford, Surrey, UK*

### *The Faculty*

Vincenzo Bronte  
*Sezione di Immunologia Department of Medicine Verona University Hospital Verona, Italy*  
Cecilia Garlanda  
*Laboratory of Experimental Immunopathology, Istituto Clinico Humanitas – IRCCS, Rozzano, Milan, Italy*  
Diego Gomez-Nicola  
*Biological Sciences, University of Southampton Southampton General Hospital, Southampton, UK*  
Siamon Gordon  
*Sir William Dunn School of Pathology, University of Oxford, Oxford, UK*  
Muzlifah Haniffa  
*Wellcome Trust, Institute of Cellular Medicine Newcastle University, Newcastle upon Tyne, UK*  
Branka Horvat  
*International Center for Infectiology University of Lyon, Lyon, France*  
Stefan H E Kaufmann  
*Max-Planck-Institut für Infektionsbiologie, Berlin, Germany*  
Elzbieta Kolaczowska  
*Department of Evolutionary Immunobiology, Institute of Zoology Jagiellonian University, Krakow, Poland*  
Foo Y (Eddy) Liew  
*Institute of Infection, Immunity and Inflammation College of Medical, Veterinary and Life Sciences University of Glasgow, Glasgow, Scotland, UK*  
Fernando Martinez Estrada  
*University of Surrey, Guildford, Surrey, UK*  
Giacchino Natoli,  
*European Institute of Oncology IFOM-IEO, Milan, Italy*  
Kodi S Ravichandran  
*Depart. of Microbiology, Immunology, and Cancer Biology, University of Virginia, Charlottesville, VA, USA*  
Maria Rescigno  
*European Institute of Oncology, Department of Experimental Oncology, Milan, Italy*  
Anna Katharina Simon  
*Human Immunology Unit, The Weatherall Institute of Molecular Medicine (WIMM), University of Oxford, John Radcliffe Hospital, Oxford, UK*  
Quentin Sattentau  
*Sir William Dunn School of Pathology University of Oxford, Oxford, UK*  
Miguel Soares  
*Gulbenkian Institute Oeiras, Portugal*  
Giuseppe (Gio) Teti  
*Department of Pediatric, Gynecological, Microbiological and Biomedical Sciences, University of Messina, Italy*

*The Organizing Committee: Silvia Fontana (President, Scuola Superiore d’Immunologia Ruggero Ceppellini), Istituto di Endocrinologia e Oncologia Sperimentale, CNR, Napoli, Italy; Antonio Di Giacomo, V Monaldi Hospital, Napoli, Italy; Tricia Reynolds, Scientific Secretariat, Scuola Superiore d’Immunologia Ruggero Ceppellini.*

*Secretariat: Effe Erre Congressi, Napoli, Italy.*

*Sponsorships: European Federation of Immunological Societies (EFIS)- The European Journal of Immunology (EJI); Gender Equality and Career Development Committee, International Union of Immunological Societies (IUIS); the Department of Translational Medicine, Federico II University of Naples; The Wellcome Trust; Biolegend; GlaxoSmithKline.*

## EFIS-EJI Course on “Tumour Immunology: from Tissue Microenvironment to Immunotherapy”

Complesso dei SS. Marcellino e Festo, Università di Napoli “Federico II”,  
16-18 October, 2017

The Course offered an overview of recent advancements in our knowledge about tumor/host interaction. New avenues for investigation of anti-cancer immunity have recently opened, with enormous translational potential.

### *The Course Directors*

Catherine Sautès-Fridman

*Immunopathology Department, Cordeliers Research Centre, Université Paris Descartes, Paris, France*

Wolf Fridman

*Cancer and Immune Escape Laboratory, Cordeliers Research Centre, Université Paris Descartes, Paris, France*

Ennio Carbone

*University of Magna Græcia, Catanzaro, Italy*

### *The Faculty*

Vincenzo Bronte

*Sezione di Immunologia Department of Medicine Verona University Hospital Verona, Italy*

Ennio Carbone

*University of Magna Græcia, Catanzaro, Italy*

Federica Cavallo

*Molecular Biotechnology Center, University of Turin, Torino, Italy*

Nadine Cerf-Bensussan

*Laboratory of Intestinal Immunity, INSERMU1163-Institut Imagine & Université Paris Descartes-Sorbonne Paris Cité, Paris, France*

Soldano Ferrone

*Department of Surgery Massachusetts General Hospital, Harvard Medical School, Boston, MA, USA*

Wolf Fridman

*Cancer and Immune Escape Laboratory, Cordeliers Research Centre, Université Paris Descartes, Paris, France*

Michele Maio

*Division of Medical Oncology and Immunotherapy, Department of Oncology, University Hospital of Siena, Siena, Italy*

Alberto Mantovani

*Istituto Clinico Humanitas, Milan, Italy*

Lorenzo Moretta

*Bambino Gesù Children's Hospital, IRCCS, Rome, Italy*

Dimitrios Mougiakakos

*Department of Hematology and Oncology University Hospital of*

*Erlangen-Nuremberg, Nuremberg, Germany*

Ugur Sahin

*TRON Translational Oncology Johannes Gutenberg-University Mainz, Germany*

Catherine Sautès-Fridman

*Immunopathology Department, Cordeliers Research Centre, Université Paris Descartes, Paris, France*

Hergen Spits

*Academic Medical Center University of Amsterdam Amsterdam, Netherlands*

Zlatko Trajanoski

*Division for Bioinformatics-Biocenter Medical University of Innsbruck Innsbruck, Austria*

*The Organizing Committee: Silvia Fontana (President, Scuola Superiore d'Immunologia Ruggero Ceppellini); Giuseppina Ruggiero, University of Naples Federico II, Naples, Italy; Francesca Di Rosa, Institute of Molecular Biology and Pathology, National Research Council of Italy (CNR), Rome, Italy; Francesco Colucci, University of Cambridge, Cambridge, UK.*

*Secretariat: Roberta Saioni, Fuori Rotta Eventi & Congressi, Italy.*

*Sponsorships: European Federation of Immunological Societies (EFIS)- The European Journal of Immunology (EJI); Gender Equality and Career Development Committee, International Union of Immunological Societies (IUIS); International Centre for Genetic Engineering and Biotechnology (ICGEB); Université Paris Descartes; Network Italiano per la Bioterapia dei Tumori (NIBIT); Servier; Institute de Recherche Pierre Fabre; Innate Pharma; Dipartimento Medicina Sperimentale e Clinica, University of Magna Græcia, Catanzaro, Italy.*

## EFIS-EJI Course on “T Cell Memory”

“Osservatorio Cultura Ricerca Formazione Divulgazione” (OCRFD) Congress Center,  
Italian National Research Council (CNR),  
Anacapri, Capri island, near Naples, 12-15 October 2018

The Course offered an overview of key cellular and molecular signals required for a durable T cell response, focussing on emerging themes in the field. Fundamental questions and translational implications were discussed. Participants had plenty of opportunity for scientific interactions and networking in a friendly atmosphere. Course was held in the charming CNR congress center in Anacapri.

### *The Course Directors*

Francesca Di Rosa  
*Institute of Molecular Biology and Pathology, National Research Council of Italy (CNR), Rome, Italy*  
Stephen Schoenberger  
*La Jolla Institute for Allergy and Immunology, La Jolla, CA, USA*

### *The Faculty*

Vincenzo Barnaba  
*Department of Internal Medicine and Medical Specialties, Sapienza University of Rome, Italy*  
Francesca Di Rosa  
*Institute of Molecular Biology and Pathology, National Research Council of Italy (CNR), Rome, Italy*  
Peter Katsikis  
*Department of Immunology, Erasmus University Medical Center, Rotterdam, The Netherlands*  
David Masopust  
*Department of Microbiology and Immunology, University of Minnesota, Minneapolis, MN, USA*  
Polly Matzinger  
*National Institutes of Health (NIH), Bethesda, MD, USA*  
Luigia Pace  
*Italian Institute for Genomic Medicine, Turin, Italy*  
Stephen Schoenberger  
*La Jolla Institute for Allergy and Immunology, La Jolla, CA, USA*  
Rene van Lier  
*Sanquin Blood Supply Foundation, Amsterdam, The Netherlands*  
Andrew Weinberg  
*Laboratory of Basic Immunology, Earle A. Chiles Research Institute Providence Health & Services, Portland, OR, USA*  
Dietmar Zehn  
*The Technical University of Munich, Freising, Germany*  
  
*The Organizing Committee:* Silvia Fontana, *President, Scuola Superiore d'Immunologia Ruggero Ceppellini, Napoli, Italy;*  
Giuseppe Matarese & Giuseppina Ruggiero, *Università di Napoli Federico II, Napoli, Italy* Ennio Carbone, *Magna Grecia University, Catanzaro;* Francesco Colucci, *University of Cambridge, Cambridge, UK.*

*Secretariat:* Joanna Cyran, *Coordinator, Scuola Superiore d'Immunologia Ruggero Ceppellini;* Roberta Saioni, *Fuori Rotta Eventi e Congressi, Italy.*

*Sponsorships:* European Federation of Immunological Societies (EFIS)- The European Journal of Immunology (EJI); Gender Equality and Career Development Committee, International Union of Immunological Societies (IUIS); the Andrew and Mary Weinberg Foundation; the Immunotherapy Foundation; the Company of Biologists; ACEA Biosciences; GlaxoSmithKline; M&M Biotech; Euroclone; Biotechne; Biolegend; Miltenyi Biotech; Agilent Technologies; Aurogene; Tema.



## EFIS-EJI Course on “Microbes, Immunity and Cancer”

“Osservatorio Cultura Ricerca Formazione Divulgazione” (OCRFD) Congress Center,  
Italian National Research Council (CNR),  
Anacapri, Capri island, near Naples, 8-10 October 2019

World leaders discussed fundamental and clinically relevant aspects of the interactions between the microbiome and the immune system. These interactions influence both how certain cancers develop and how cancer patients respond to new immunotherapies.

### *The Course Directors*

Francesco Colucci  
*University of Cambridge, Cambridge, UK*  
Ennio Carbone  
*University of Magna Græcia, Catanzaro, Italy*  
Guido Kroemer  
*Centre de Recherche des Cordeliers, National Cancer Institute, Université Paris Descartes, Paris, France*  
Giorgio Trinchieri  
*National Institute of Health, Bethesda, MD, USA*  
Laurence Zitvogel  
*Gustave Roussy Cancer Campus, Paris, France*

### *The Faculty*

Petter Brodin  
*Karolinska Institutet, Stockholm, Sweden, Karolinska University Hospital, Stockholm, Sweden*  
Jolande De Vries  
*Radboud University Medical Center, Nijmegen, The Netherlands, Nijmegen, The Netherlands*  
Sebastian Kobold  
*Klinikum der Universität München, LMU Munich, Germany*  
Guido Kroemer  
*Centre de Recherche des Cordeliers, Paris, France, Gustave Roussy Comprehensive Cancer Center, Paris, France*  
Nicola Segata  
*Erasmus University, University of Trento, Trento, Italy*  
Giorgio Trinchieri  
*Center for Cancer Research, National Cancer Institute, National Institutes of Health, Bethesda, USA*  
Laurence Zitvogel  
*Gustave Roussy Comprehensive Cancer Center, Paris, France*

*The Organizing Committee:* Silvia Fontana e Riccardo Zappacosta, *Scuola Superiore d'Immunologia Ruggero Ceppellini, Napoli, Italy*; Giuseppina Ruggiero, *Università di Napoli Federico II, Napoli, Italy*.

*Secretariat:* Roberta Saioni, *Fuori Rotta Eventi e Congressi, Italy*.

*Sponsorships:* European Federation of Immunological Societies (EFIS)- The European Journal of Immunology (EJI); The European Academy of Tumor Immunology (EATI); Gender Equality and Career Development Committee, International Union of Immunological Societies (IUIS); The Department of Experimental and Clinic Medicine (DMSC), University of Catanzaro, Catanzaro, Italy.

## LEVEL 3 COURSES

Level 3 Courses are practical, laboratory courses, dealing with recent techniques, and devised for small groups of graduates.

### **"Molecular Analysis of T Cell Repertoires by CDR3 Length Heterogeneity"**

Federico II University Medical School, Naples, Italy,  
8-10 May 1998

This laboratory course was aimed to practically show, to a limited number of students, the molecular analysis of T-cell receptor (TCR) repertoire, based on the assessment of the length heterogeneity of CDR3 regions, specifically involved in antigen recognition. The course was inaugurated on May 8, 1998, by a lecture session dealing with the significance of the variations of the TCR repertoires in human pathology and the diagnostic relevance of molecular approaches to such studies.

#### *The Course Director*

Jack Gorski,  
*Blood Research Institute, BloodCenter of Wisconsin, Milwaukee, WI, USA*

#### *The Faculty*

Alfredo Ciccodicola  
*International Institute of Genetics and Biophysics, Italian National Research Council (CNR), Naples, Italy*  
Raffaele De Palma  
*Laboratorio di Medicina Molecolare-IRCCS Fondazione S. Maugeri, Pavia*  
*Dipartimento di Internistica Clinica e Sperimentale-Seconda Università di Napoli, Napoli*  
Claudia Giachino  
*Laboratorio di Medicina Molecolare-IRCCS Fondazione S. Maugeri, Pavia*  
Jack Gorski  
*Blood Research Institute, Blood Center of Southeastern Wisconsin-Medical Complex of Wisconsin, Milwaukee, WI, USA*  
Philippe Kourilsky  
*Pasteur Institute, Paris, France*  
Antonio Lanzavecchia  
*Basel Institute for Immunology, Basel, Switzerland*

The laboratory activity took place from 9 to 10 May, 1998, at the Genome Research and Sequencing Laboratory, Servizio di Tecnologie Biomolecolari, Area di Ricerca del CNR, Naples and included amplification of the V-D-J regions of TCR gene transcripts by primers specific to each family of such molecules, sequence gel visualization of the obtained products and computer analysis of the data by especially devised softwares. Experiments were carried out under the supervision of Laleh Ansari (Milwaukee), Alfredo Ciccodicola (Naples), Raffaele De Palma (Naples & Pavia), Anna Maria Masci (Naples), Giuliana Soldati (Naples), Maryam Yassai (Milwaukee)

*The Organizing Committee:* Alfredo Ciccodicola, *International Institute of Genetics and Biophysics, Italian National Research Council (CNR), Naples, Italy*; Raffaele De Palma & Luigi Racioppi, *Università di Napoli Federico II*; Guido Sacerdoti, *Seconda Università di Napoli*.

*Secretariat:* Ceppellini School Director's Office, Napoli, Italy.

*Sponsorships:* The International Institute of Genetics and Biophysics and Area di Ricerca, CNR Naples; The Post-Graduate School of Allergology and Clinical Immunology of the Seconda Università di Napoli, Naples, Italy; Perkin Elmer Italia SpA, Milan, Italy.

## OTHER EVENTS TILL 2006

### Inaugural ceremony of the “Ruggero Ceppellini Advanced School of Immunology”

Palazzo Serra di Cassano, Naples,  
October 11, 1992

The School's inaugural ceremony was held at the seat of the Istituto Italiano per gli Studi Filosofici, on the occasion of the School's first course, dealing with the immunology of bone marrow transplantation. Talks entitled as follows were delivered in that memorable session:

“Plato, Jerne, Ceppellini: Speculation and Experiment in Immunology”  
by Jan Klein  
*Max Planck Institute for Biology, Tübingen, Germany*

“The Ruggero Ceppellini Legacy”  
by Giovanni B Ferrara  
*Federico II University of Naples, Italy*

“Immunology: Basic Principles and Challenges”  
by Alfred Nisonoff  
*Brandeis University, Waltham, MA, USA*

“The Advanced School of Immunology: Aims and Ideals”  
by Serafino Zappacosta & Antonio Di Giacomo  
*SZ: Federico II University of Naples, Italy; AD: Colli Monaldi Hospital, Naples, Italy*

“Concluding Address”  
by Gerardo Marotta  
*Istituto Italiano per gli Studi Filosofici, Naples, Italy*

### Conference “Migration Flows and Emerging Pathologies: The Role of Immunology”

Palazzo Serra di Cassano, Naples, April 29, 2005

A Conference held on the Occasion of the European Day of Immunology, coorganised with the Italian Society for Immunology, Clinical Immunology and Allergology (SIICA).

#### *Chairman*

Serafino Zappacosta  
*Cattedra di Immunologia, Dipartimento di Biologia e Patologia Cellulare e Molecolare, Università di Napoli Federico II, Napoli, Italy*

#### *Invited Speakers*

Helmut Hahn  
*Institut für Medizinische Mikrobiologie und Infektionsimmunologie, Freie Universität Berlin, Berlin, Germany*

Eleanor M Riley  
*Department of Infectious and Tropical Diseases, London School of Hygiene and Tropical Medicine, London, UK*

Stephan Becker  
*Institut für Virologie, Marburg, Germany*

Giuseppe Scala, *Università Magna Græcia, Catanzaro, Italy*

## **Symposium “Interface between Innate and Adaptive Immunity: Conversation between Tissues and T Cells”**

Ischia, near Naples,  
April 27, 2004

A Ceppellini School Plenary Symposium within the 3rd National Conference of the Italian Society for Immunology, Clinical Immunology and Allergology (SIICA).

### *Invited Speakers*

Polly Matzinger  
*Laboratory for Cellular and Molecular Immunology, National Institute of Allergy and Infectious Diseases, NIH,  
Bethesda, MD, USA*  
Daniele D'Ambrosio  
*BioXell, Milan, Italy*

## **Workshop “Meeting the Challenges of Clinical Organ Transplantation”**

Azienda Ospedaliera V Monaldi, Naples, June 16, 2000

A Workshop on the biological bases and the clinical perspectives of organ transplantation, in collaboration with the Azienda Ospedaliera V Monaldi, under the auspices of the Second University of Naples and of the Health Service of the Campania Regional Government.

### *Chairmen*

Robert Lechler  
*Medical Research Council Centre for Transplantation, King's College London, London, UK*  
Maurizio Cotrufo  
*Seconda Università di Napoli, Ospedale Mondaldi, Napoli, Italy*

### *Invited Speakers*

Maurizio Cotrufo  
*Seconda Università di Napoli, Ospedale Mondaldi, Napoli, Italy*  
Andrew JT George  
*Section of Molecular Immunology, Department of Medicine, Imperial College London, Hammersmith Hospital, London, UK*  
Robert Lechler  
*Medical Research Council Centre for Transplantation, King's College London, London, UK*  
Anthony N Warrens  
*Renal and Transplantation Medicine, Barts and The London School of Medicine and Dentistry, Queen Mary University of London, London, UK.*  
Kathryn Wood  
*Nuffield Department of Surgery, University of Oxford, Oxford, UK*  
Serafino Zappacosta  
*Cattedra di Immunologia, Dipartimento di Biologia e Patologia Cellulare e Molecolare, Università di Napoli Federico II, Napoli, Italy*



## Conference “Emergence of Infectious Diseases: An Evolutionary Perspective”

Palazzo Serra di Cassano, Naples,  
27-29 May 1998

A Conference aimed at a reappraisal of the biological and social significance of emerging and reemerging infectious diseases. The Conference included two panel discussions, one on “Socioeconomic and Historical Aspects of Infection in Developing vs Developed Countries” (A. Caprioli, Rome, Italy; P. Conforti, Rome, Italy; D. Greco, Rome, Italy; J. A. Louis, Geneva and Epalinges, Switzerland; I. Luzzi, Rome, Italy), and the other on “New Views on HIV Infection” (M. Clerici, Milan, Italy; C. F. Perno, Rome, Italy; O. Perrella, Naples, Italy). Under the patronage of: The World Health Organization; The Health Minister of the Italian Republic; The Istituto Superiore di Sanità, Rome, Italy; The Mayor of Naples; The Provincia di Napoli.

### Chairmen

Jan Klein  
*Max Planck Institute for Biology, Tübingen, Germany*  
Serafino Zappacosta  
*Cattedra di Immunologia, Dipartimento di Biologia e Patologia Cellulare e Molecolare, Università di Napoli Federico II, Napoli, Italy*

### Invited Speakers

Martin Achtman  
*Max Planck Institut for Molecular Genetics, Berlin, Germany*  
Donato Greco  
*Laboratorio di Epidemiologia e Biostatistica, Istituto Superiore di Sanità, Rome, Italy*  
Eduardo A Groisman  
*Howard Hughes Medical Institute, Washington University School of Medicine, St Louis, MO, USA*  
Sunetra Gupta  
*The Wellcome Trust Centre for the Epidemiology of Infectious Diseases, University of Oxford, Oxford, UK*  
Helmut Hahn  
*Institut für Medizinische Mikrobiologie und Infektionsimmunologie, Freie Universität Berlin, Berlin, Germany*  
Adrian V S Hill  
*Institute of Molecular Medicine, John Radcliffe Hospital, Oxford University, Oxford, UK*  
Jonathan C Howard  
*Institute for Genetics, University of Cologne, Germany*  
Jan Klein  
*Max Planck Institute for Biology, Tübingen, Germany*  
Jack A Louis  
*World Health Organization, Geneva and University of Lausanne, Epalinges, Switzerland*  
Andrew J S Macpherson  
*Institute of Experimental Immunology, University Hospital Zurich, Zurich, Switzerland*  
Stephen M Ostroff  
*National Center for Infectious Diseases, Center for Disease Control and Prevention, Atlanta, GA, USA*  
Rino Rappuoli  
*Istituto Ricerche Immunobiologiche Siena, Chiron SpA, Siena, Italy*  
Margaret A Riley  
*Yale University, New Haven, CT, USA*  
Thomas S Whittam  
*The Pennsylvania State University, PA, USA*  
Serafino Zappacosta  
*Cattedra di Immunologia, Dipartimento di Biologia e Patologia Cellulare e Molecolare, Università di Napoli Federico II, Napoli, Italy*

*The Organizing Committee:* Antonio Di Giacono Azienda Ospedaliera V Monaldi, Napoli, Italy, Donato Greco, Istituto Superiore di Sanità, Rome, Italy; Jack A Louis, World Health Organization, Geneva and University of Lausanne, Epalinges, Switzerland; Giuseppina Ruggiero, Università di Napoli Federico II, Napoli, Italy.

*Secretariat:* Effe Erre Congressi, Napoli, Italy.

# SERAFINO ZAPPACOSTA CONFERENCES

These conferences were initiated in 2007, in honor of Serafino Zappacosta (1935-2006). Since 2010, most of these events have been held in the newly inaugurated "Serafino Zappacosta" Auditorium of the Federico II University of Naples.

## I Serafino Zappacosta Conference

### **"Xeno-transplantation: biological advances, clinical possibilities, philosophical and ethical concerns"**

*Invited Speaker: Robert J. Lechler  
King's College London, Guy's Hospital, London, UK*

Gerardo Marotta, chairman of the Istituto Italiano per gli Studi Filosofici, introduced this first conference. He discussed the basis of the very intriguing interplay between Science and Philosophy that the long-standing collaboration between the Scuola Superiore d'Immunologia Ruggero Ceppellini and the Istituto Italiano per gli Studi Filosofici has been proposing to the scientific community since many years. The invited speaker gave an intriguing overview on key issues in xenotransplantation.

Palazzo Serra di Cassano, Istituto Italiano per gli Studi Filosofici, Naples, February 1st, 2007

## II Serafino Zappacosta Conference

### **"1908-2008, Science and culture in Naples: the Metchnikoff heritage a century after the Nobel Prize"**

*Invited Speaker: Helmut Hahn  
Berlin Medical Association and Koch-Metchnikoff Forum, Berlin, Germany*

### **"Phagocytosis 100 years later: Imaging proteins, lipids and charges"**

*Invited Speaker: Sergio Grinstein  
Hospital for Sick Children, Toronto, Canada*

This conference proposed a fruitful discussion on the role of the Stazione Zoologica Anton Dohrn, a pioneer neapolitan institution, for the development of Metchnikoff theories on cell-mediated processes in immune recognition.

Stazione Zoologica Anton Dohrn, Naples, June 16, 2008

## III Serafino Zappacosta Conference

### **"Small RNAs, transcription and epigenetic modifications"**

*Invited Speaker: V. Enrico Avvedimento  
Dipartimento di Biologia e Patologia Cellulare e Molecolare, Università di Napoli, Federico II, Napoli*

### **"Proteins and microRNAs controlling NK cell activity"**

*Invited Speaker: Ofer Mandelboim  
The Hebrew University of Jerusalem, Jerusalem, Israel*

This conference was focused on the emerging interplay between small interfering RNAs and immune-regulation.

Palazzo Serra di Cassano, Istituto Italiano per gli Studi Filosofici, Naples, June 4, 2009

## IV Serafino Zappacosta Conference

### **"Allelic exclusion in the immune system"**

*Invited Speaker: Yehudit Bergman*

*Department of Developmental Biology and Cancer Research, The Hebrew University Medical School, Jerusalem, Israel*

This conference focused on the role of genetic processes in the generation and development of adaptive humoral immune response.

Aula "Serafino Zappacosta", Dipartimento di Biologia e Patologia Cellulare e Molecolare, Università di Napoli "Federico II", June 30, 2010

## V Serafino Zappacosta Conference

### **"Paroxysmal Nocturnal Haemoglobinuria: Stem cells, Complement, Autoimmunity"**

*Invited Speaker: Lucio Luzzatto*

*Università di Firenze, Istituto Toscano Tumori, Firenze*

This conference discussed the intriguing involvement of autoimmune selection processes in the emergence and dominance of Bone Marrow defective progenitors in the pathogenesis of Paroxysmal Nocturnal Haemoglobinuria.

Aula "Serafino Zappacosta", Dipartimento di Biologia e Patologia Cellulare e Molecolare, Università di Napoli "Federico II", January 27, 2012

## VI Serafino Zappacosta Conference

### **"The immune contexture of human tumours"**

*Invited Speaker: Catherine Sautès-Fridman*

*Immunology, Cancer and Inflammation Department, Université Paris Descartes, Paris, France*

This conference highlighted the relevance of immune contexture in the priming and differentiation of an effective anti-tumor immune response. It also addressed the prognostic potential of the analysis of immune infiltrates in solid tumors.

Aula "Serafino Zappacosta", Dipartimento di Biologia e Patologia Cellulare e Molecolare, Università di Napoli "Federico II", September 12, 2014

## VII Serafino Zappacosta Conference

### **"What triggers an immune response?"**

*Invited Speaker: Polly Matzinger*

*Laboratory of Immunogenetics, National Institute of Allergy and Infectious Diseases, NIH, Bethesda, USA*

Ennio Carbone & Giuseppe Matarese, two past students of prof Zappacosta and current members of the Board of Directors of the Ceppellini School of Immunology introduced this conference. They highlighted that the Ceppellini School represents a precious heritage after the death of Serafino Zappacosta. The invited speaker, a world leader in immunology, offered her provocative ideas on immune tolerance and discussed the reasoning behind them.

Complesso dei SS. Marcellino e Festo, Università di Napoli "Federico II", September 28, 2016

## VIII Serafino Zappacosta Conference

### **Epigenetic cancer immuno-modeling to improve the efficacy of checkpoint-based immunotherapy**

*Invited Speaker: Michele Maio*

*Center for Immuno-Oncology, Medical Oncology and Immunotherapy, Siena University Hospital, Siena*

The invited speaker gave an historical overview on Immuno-therapy, highlighting that recent advancements in this field have provided (by now) well-established therapeutic tools, powerfully improving clinical management of human tumors.

Auditorium "Gaetano Salvatore", Università di Napoli "Federico II", March 12, 2019

# Advantages of publishing in Frontiers



## OPEN ACCESS

Articles are free to read  
for greatest visibility  
and readership



## FAST PUBLICATION

Around 90 days  
from submission  
to decision



## HIGH QUALITY PEER-REVIEW

Rigorous, collaborative,  
and constructive  
peer-review



## TRANSPARENT PEER-REVIEW

Editors and reviewers  
acknowledged by name  
on published articles

## Frontiers

Avenue du Tribunal-Fédéral 34  
1005 Lausanne | Switzerland

**Visit us:** [www.frontiersin.org](http://www.frontiersin.org)

**Contact us:** [info@frontiersin.org](mailto:info@frontiersin.org) | +41 21 510 17 00



## REPRODUCIBILITY OF RESEARCH

Support open data  
and methods to enhance  
research reproducibility



## DIGITAL PUBLISHING

Articles designed  
for optimal readership  
across devices



## FOLLOW US

@frontiersin



## IMPACT METRICS

Advanced article metrics  
track visibility across  
digital media



## EXTENSIVE PROMOTION

Marketing  
and promotion  
of impactful research



## LOOP RESEARCH NETWORK

Our network  
increases your  
article's readership